

UNIVERSITI PUTRA MALAYSIA

EFFECTS OF XYLANASE AND CELLULASE SUPPLEMENTATION ON EARLY GROWING STAGE, CAECAL BACTERIA POPULATION AND FERMENTATION PROFILES OF BROILER FED WITH PALM KERNEL MEAL

SHARMILA BT AHMAD

FP 2014 81



EFFECTS OF XYLANASE AND CELLULASE SUPPLEMENTATION ON EARLY GROWING STAGE, CAECAL BACTERIA POPULATION AND FERMENTATION PROFILES OF BROILER FED WITH PALM KERNEL MEAL



By

SHARMILA BT AHMAD

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master Science

December 2014

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysiain fulfilment of the requirement for the degree of Master Science

EFFECTS OF XYLANASE AND CELLULASE SUPPLEMENTATION ON EARLY GROWING STAGE, CAECAL BACTERIA POPULATION AND FERMENTATION PROFILES OF BROILER FED WITH PALM KERNEL MEAL

By

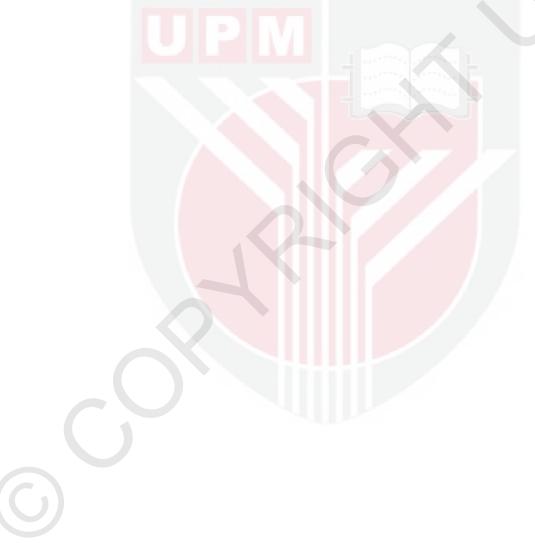
SHARMILA BT AHMAD

December 2014

Chairman: Anjas Asmara Samsudin, PhD

Faculty: Agriculture

The optimum inclusion of palm kernel meal in broiler diets is reported about 20% without having any detrimental effect on the performance and health of broilers. The addition of enzyme in palm kernel mealdiet for broilers isnecessary in promoting growth performances of the broilers through increase in feed efficiency and Since, previous study only focusing on their effect on growth digestibility. performance; therefore, the present study was conducted to study the effect of enzyme supplementation in palm kernel mealdiet on caecal fermentation profiles, early growth performances and caecal bacteria population of broilers. The *in-vitro* analysis was carried out by incubating the formulated diets with caecal digesta of broilers for 72 hours at 39±5°C. The gas production and end products of the fermentation were examined. The gas production rate of unsupplemented and enzyme-supplemented palm kernel meal diet observed was similar in trends. However, it was found that the amount of acetate was significantly (P<0.05) reduced with enzyme supplementation, whilst the *in-vitro* dry matter degradability of both, starter and finisher diet were improved (P<0.05), with more outstanding result observedfollowing xylanase supplementation at 11.09 and 17.59%, respectively.In the feeding trial, 75 broiler chicks were randomly allocated to three dietary treatment groups receiving diet containing 20% of palm kernel meal with or without xylanase and cellulase supplementation. The individual body weight and feed intakewere recorded weekly, and at day 35, broilers were slaughtered for ceacal digesta collection. Both enzyme addition did not improve the average daily gain of broiler at starter phase (1- 21 days), but had improved at grower phase (22-35 days), where broilers fed with xylanase supplementation had improved their body weight by 7% compared to control diet (61.98 g/bird). The similar result was also observed in the final body weight of broilers, where xylanase-supplemented group was the highest followed by control and cellulase-supplemented group which values is 1495.2, 1436.8 and 1367.8 kg, respectively. In the enumeration of caecal bacteria population, lactic acid bacteria and Salmonella spp. population were reduced, by 4.1 and 1.9% in xylanase supplemented group, and 8.3 and 7.9% in cellulase supplemented group, respectively. In contrast, the caecal bacteria population quantified using real-time PCR analysis revealed that enzyme supplementation had reduced the number of total bacteria, lactic acid bacteria, enterococcus, enterobacteria, *Escherichia coli*, bifidobacteria and *Salmonellaspp.*, respectively compared to unsupplemented palm kernel meal diet. Apart from that, significant negative correlation (P<0.01) was observed between the population of coliform bacteria (-0.97) and *Salmonella* spp. (-0.84) with n-butyrate in caecal digesta of broilers. The current data show that n-butyrate have a toxic effect on coliform bacteria and *Salmonella* spp. In conclusion, enzyme supplementation in 20% palm kernel meal diet was only effective in improving the body weight of broilers at growing phase but not in the starter phase. So, reducing the amount of palm kernel meal in the starter broiler diet is recommended; since, body weight of broilers fed 20% palm kernel meal diet was not improved, even with enzyme addition.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Sarjana Sains

KESAN SUPLEMENTASI XILANASE DAN SELULASE TERHADAP PERINGKAT AWAL TUMBESARAN, POPULASI BAKTERIA DAN PROFIL FERMENTASI SEKUM AYAM PEDAGING YANG DIBERI MIL ISIRONG SAWIT

Oleh

SHARMILA BT AHMAD

Disember 2014

Chairman: Anjas Asmara Samsudin, PhD

Fakulti: Pertanian

Tahap kemasukan optimum mil isirong sawit dalam diet ayam pedaging dilaporkan sebanyak 20% tanpa memberi kesan mudarat terhadap prestasi tumbesaran dan kesihatan ayam pedaging. Penambahan enzim dalam diet mil isirong sawit adalah penting bagi meningkatkan prestasi tumbesaran ayam pedaging melalui peningkatan kecekapan dan pencernaan makanan. Oleh kerana kajian sebelum ini hanya memberi tumpuan kesannya terhadap prestasi tumbesaran ayam pedaging, maka kajian ini dijalankan untuk mengkaji kesan suplemen enzim dalam diet mil isirong sawit terhadap profil fermentasi sekum, peringkat awal tumbesaran dan populasi bakteria sekum ayam pedaging. Analisis *in-vitro* telah dijalankan dengan mengikubasi rumusan diet berserta digesta sekum ayam pedaging selama 72 jam pada suhu 39±5°C. Kadar pengeluaran gas dan produk akhir fermentasi diperiksa. Kadar pengeluaran gas diet mil isirong sawit dengan suplemen enzim atau tidak adalah sama. Walau bagaimanapun, didapati jumlah asid asetik telah berkurang secara nyata (P<0.05) oleh diet dengan suplemen enzim, manakala kadar penguraian bahan kering diet bagi ransum pemula dan pembesar telah meningkat secara ketara (P<0.05), dengan kadar penguraian yang paling tinggi oleh diet dengan suplemen xilanase pada kadar 11.09 dan 17.59%. Dalam ujian perbandingan makanan, sebanyak 75 ekor anak ayam pedaging secara rawaknya dibahagikan kepada tiga kumpulan diet menerima diet 20% mil isirong sawit tanpa atau dengan suplemen enzim xilanase dan selulase. Berat badan dan pengambilan makanan individu telah direkodkan setiap minggu, dan pada hari ke-35, ayam disembelih bagi pengumpulan digesta sekum. Penambahan enzim tidak meningkatkan purata berat badan harian ayam pedaging pada fasa permulaan (1-21 hari), tetapi telah meningkat pada fasa pembesaran (22-35 hari) oleh ayam pedaging yang diberi makan diet dengan suplemen enzim xilanase sebanyak 7% berbanding dengan diet tanpa enzim (61.98 g/ayam). Di akhir eksperimen, berat badan ayam pedaging yang diberi makan diet dengan suplemen xilanase adalah yang tertinggi, diikuti oleh ayam pedaging yang diberi diet tanpa enzim, seterusnya dengan suplemen selulase, masing-masing sebanyak 1495.2,

1436.8 dan 1367.8 kg. Dalam penghitungan populasi bakteria sekum, bakteria asid laktik dan Salmonella spp. telah berkurang sebanyak 4.1 dan 1.9% dengan suplemen xilanase manakala berkurang sebanyak 8.3 dan 7.9% dengan suplemen selulase.Sebaliknya analisa populasi bakteria sekum melalui kaedah 'PCR' menunjukkan bahawa diet dengan suplemen enzim telah mengurangkan bilangan populasi bakteria total, bakteria asid laktik, enterokokus, enterobakteria, Escherichia coli, bifidobakteria dan Salmonella spp. berbanding diet tanpa suplemen enzim. Selain itu, korelasi negatif yang signifikan (P<0.01) didapati antara populasi bakteria koliform (-0,97) dan Salmonella spp. (-0,84) dengan asid n-butirik dalam digesta sekum ayampedaging.Data semasa menunjukkan bahawa asid n-butirikmempunyai kesan toksik terhadap bakteria koliform dan Salmonella spp. Kesimpulannya, suplemen enzim dalam 20% diet mil isirong sawit hanya dapat meningkatkan berat badan ayam pedaging pada fasa pembesaran tetapi tidak pada fasa permulaan.Oleh itu, adalah disyorkan untuk mengurangkan jumlah mil isirong sawit dalam diet fasa awal ayam pedaging, kerana berat badan ayam pedaging diberi makan 20% mil isirong sawit tidak meningkat walaupun dengan suplemen enzim.

ACKNOWLEDGEMENTS

First and foremost I offer my sincerest gratitude toward my supervisor, Dr. Anjas Asmara Samsudin, and co-supervisors, Prof. Madya Dr. Azhar Kasim and also Dr. Mohd Hezmee Mohd Nor, whose had supported me throughout my work and thesis with patience, knowledge, guidance and support. I attribute the level of my Master Degree to their encouragement and effort and without them this thesis, too, would not have been written or completed.

In the various laboratory that I have engaged for a few month, I would like to give my deepest thanks and gratitude to all lecturers and staffs of Department of Animal Science, Faculty of Agriculture, Universiti Putra Malaysia for their advises and technical assistance during the laboratory work. I would also like to extend my appreciation and thank to Dr. Muhammad Fasaleh Jahromi, from animal production laboratory of Institute Tropical Agriculture, UPM for assisting me in molecular work of determining bacterial quantification using qPCR analysis. Not to forget, I would also like to thank Agro-Biotechnological Institute (ABI), MOSTI, Malaysia for letting me use their laboratory facilities such qPCR machine.

Last, but not least, I would like to express my special thank to my beloved parents, Mr. Ahmad Daud and Mrs. Salmiah Mohd Amin, family and friends whose had encouraged, advised and supported me from the initial of the project until it completed. Lastly, I offer my regards and blessings to all of those who directly or indirectly supported me in any respect during the completion of the project. I certify that a Thesis Examination Committee has met on 22 December 2014 to conduct the final examination of Sharmila binti Ahmadon her thesis entitled "Effect of Xylanase and Cellulase Supplementation on Early Growing Stage, Caecal Bacterial Population and Fermentation Profiles of Broiler Fed with Palm Kernel Meal " in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Ismail bin Idris, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Dahlan bin Ismail, PhD Professor

Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

Wan Zuhainis binti Saad, PhD

Senior Lecturer Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Internal Examiner)

Wan Zahari bin Moha<mark>med, PhD</mark>

Professor Universiti Malaysia Kelantan Malaysia (External Examiner)



ZULKARNAIN ZAINAL, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 26 February 2015

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Anjas Asmara Samsudin, PhD

Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Azhar Kasim, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

Mohd Hezmee Mohd Noor, PhD

Senior Lecturer Faculty of Veterinary Universiti Putra Malaysia (Member)

BUJANG BIN KIM HUAT, PhD Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research And Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals modules, proceedings, popular writings, seminars papers, manuscripts, posters reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _

Name and Matric No.: Sharmila binti Ahmad (GS32836)

Declaration by Member of Supervisory Committee

This is to confirm that:

Ċ,

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:	 Signature:	
Name of	Name of	
Chairman of	Member of	
Supervisory	Supervisory	
Committee: -	 Committee:	
Committee: - Signature: Name of Member of Supervisory Committee: -	Committee:	

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	V
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiv
LIST OF FIGURES	XV
LIST OF ABBREVIATIONS	xvi

CHAPTER 1

INTRODUCTION

2	LITE	ERATURE REVIEW	5
	2.1	Poultry industry in Malaysia	5
		2.1.1 Constraints/ limitations in poultry industry	5
	2.1.1	.1 Cost of feeding and feed availability	5
	2.2	Fiber digestion in poultry	6
	2.3	Poultry caecal development in relation to fiber	6
		feeding	
	2.4	Palm kernel meal (PKM)	7
		2.4.1 Production of PKM	7
		2.4.2 Nutritional content of PKM	10
		2.4.3 The use of PKM in poultry diets	11
		2.4.4 Improving the nutritional value of PKM for	12
		poultry	
	2.5	The use of non-starch polysaccharides (NSPs)	15
		enzymes in poultry diets	
		2.5.1The use of xylanase in poultry diets	16
		2.5.2The use of cellulase in poultry diets	17
	2.6	Bacteria involvement in the fermentation of feed in	18
		poultry caecum	
		2.6.1 Caeca and feed digestion in poultry	18
		2.6.2 Bacterial population in poultry caeca	19
		2.6.3 Caecal bacteria involve with caecal	20
	ferme	entation	
		2.6.3.1 Lactic acid bacteria (LAB)	20
		2.6.3.2 Fiber/NSPs degrading bacteria	21
	2.7	Effect of enzymes supplementation on gut	21
		microflora of poultry	
	2.8	Gastrointestinal tract microorganism identification	22
	2.9	Method used to study and indentified the caecal	23
		bacterial community	
		2.9.1 Culture dependent-method	23
		2.9.2 Culture independent-method	23

Х

	3.0	Conclusion	24
3	GEN	ERAL MATERIALS AND METHODS	25
	3.1	Proximate analysis	25
		3.1.1 Determination of dry matter (DM)	25
		3.1.2 Determination of ash	25
		3.1.3 Analysis of crude protein (CP)	25
		3.1.4 Analysis of ether extract (EE)	26
		3.1.5 Analysis of crude fiber (CF)	26
	3.2 Q	uantification of volatile fatty acids	26
		3.2.1Preparation of standard	26
		3.2.1.1 External standard	26
		3.2.1.2 Internal standard	27
		3.2.2Volatile fatty acids determination using gas	27
		chromatography	
	3.3	Quantification of ammonia	27
		3.3.1Solution preparation	27
		3.3.1.1 Phenol solution	27
		3.3.1.2 Sodium nitroprusside	28
		3.3.1.3 Alkaline reagent	28
		3.3.1.4 Sodium hypochlorite	28
		3.3.1.5 Oxidizing solution	28
		3.3.2Preparation of standard	28 28
		3.3.3Quantification of ammonia using spectrophotometer	20
	3.4	Analysis of amino acid (AAs)	29
	5.4	3.4.1 Preparation of feed samples	29 29
		3.4.1.1 Acid hydrolysis	29
		3.4.1.2 Perfomic oxidation	30
		3.4.1.3 Alkaline hydrolysis	30
	3.5	Preparation of enzymes	30
	3.6	Preparation of formulated diets	30
	3.7	Statistical analysis	32
4		ITRODETERMINATION OF CAECAL	33
		MENTATION PROFILES OF PKM DIET IN	
		PONSE TO XYLANASE AND CELLULASE	
		PLEMENTATION	22
	4.1	Introduction	33
	4.2	Materials and methods	34
		4.2.1 Preparation of caecal digesta	34 34
	1	4.2.2Preparation of basal media 2.2.1 Media	34 34
	4.	4.2.2.2 Basal solution	34 34
	Δ	2.2.3Reducing agent	34
	4.	4.2.2.4 Bicarbonate solution	35
	Δ	2.2.5Vitamin/phosphate solution	35
	-т.	4.2.2.4 Bicarbonate solution	35
		4.2.3 Substrate preparation	35
		4.2.4Method of incubation	36
			-

. .

	4.2.5Samples collection for quantification of VFAs	36
	and ammonia	26
	4.3 Results	36
	4.3.1 <i>In-vitro</i> fermentation profiles	36
	4.3.1.1 Volatile fatty acids and ammonia production	36
	4.3.1.2Gas production profiles	38
	4.3.2 Fermentation kinetics	39
	4.4 Discussion	41
	4.4.1 <i>In-vitro</i> fermentation profiles and gas	41
	production	
	4.4.2 <i>In-vitro</i> fermentation kinetics and IVDMD	42
	4.5 Conclusion	42
5	EFFECT OF XYLANASE AND CELLULASE	43
5	SUPPLEMENTATION ON EARLY GROWTH	+3
	PERFORMANCE, VOLATILE FATTY ACIDS AND	
	CAECAL BACTERIA POPULATION OF BROILER	
	CHICKENS FED WITH PKM DIET	10
	5.1 Introduction	43
	5.2 Materials and methods	44
	5.2.1 Location	44
	5.2.2 Birds and diets	44
	5.2.3 Microbiological analysis	45
	5.2.3.1Preparation of agar	45
	5.2.3.2Preparation of bacterial enumeration	46
	5.3 Results	46
	5.3.1 Early growth performance	46
	5.3.2 Caecal microbial count	48
	5.3.3 Caecal VFAs profiles	48
	5.3.4 Correlation between caecal bacteria and VFAs	49
	5.4 Discussion	50
	5.4.1 Early growth performance	50
	5.4.2 Caecal bacterial enumeration	51
	5.4.3 Caecal VFAs profiles	52
	5.4.4 Correlation between caecal bacteria population	53
	and VFAs	55
	5.5 Conclusion	54
6	QUANTIFICATION OF CAECAL BACTERIA POPULATION IN RESPONSE TO XYLANASE AND CELLULASE SUPPLEMENTATION USING QUANTITAVTIVE REAL TIME-PCR ANALYSIS (qPCR)	55
		55
		55
	6.2 Materials and methods	55
	6.2.1 Preparation of caecal sample	55
	6.2.2 Extraction of caecal DNA	56
	6.2.3 Primers and SYBR Green of qPCR analysis	56
	6.2.4 Preparation of standard curve	58
	6.3 Results	59

xii

	6.4 Discussion		60
	6.5 Conclusion		61
7	GENERAL DISCUS	SSION	63
8	GENERAL CONCL	USION AND	65
	RECOMMENDATI	ON	
REFERE	NCES		66
APF	ENDICES		83
BIO	DATA OF STUDENT		86
LIS	OF PUBLICATIONS		87



G

LIST OF TABLES

Table		Page
2.1	Nutrient composition of solvent extracted and expeller pressed PKM	10
2.2	Amino acid contents and mineral contents of PKM	11
2.3	Recommended levels of PKM in broiler diets	12
2.4	Proximate analysis of raw PKM and fermented PKM of different fungal strain	14
2.5	Proximate composition (% dry matter) of raw and treated- PKM	14
3.1	Time and gradients pump setting for mobile phases used in HPLC	29
3.2	Ingredient composition and nutrient level (%) of PKM diet	31
4.1	<i>In-vitro</i> fermentation profiles of PKM diet supplemented with or without enzyme	37
4.2	<i>In-vitro</i> fermentation kinetics and IVDMD of PKM diet supplemented with or without enzyme	40
5.1	Growth performance (means \pm SE) of broiler fed with 20% PKM diet supplemented with or without enzyme	47
5.2	The bacterial population in the caecal digesta $(\log_{10}CFU/g digesta)$ fed 20% PKM diet supplemented with or without enzyme	
5.3	The VFAs production in caecal digesta of broiler fed 20% PKM diet supplemented with or without enzyme	49
5.4	Correlation between VFAs and caecal bacteria population of broiler	49
6.1	The sequence of primers used targeting total bacteria, LAB, bifidobacteria, enterobacteria, <i>E.coli</i> and <i>Salmonella</i> spp.	57
6.2	Effect of xylanase and cellulase supplementation on cell counts of caecal bacteria (log10 of copy number/g DNA extract)	59

LIST OF FIGURES

Figure		Page
2.1	Oil palm fruit and its shells	7
2.2	The different type of oil palm fruits; <i>E. guineensis</i> fo. <i>dura</i> , <i>E. guineensis</i> var. <i>pisifera</i> and <i>E. guineensis</i> fo. <i>tenera</i>	8
2.3	Extraction of palm kernel oil	9
	Gas production profiles from fermentation of PKMdiet for starter and finisher supplemented or unsupplemented with xylanase or cellulase	38

LIST OF ABBREVIATIONS

ADF	Acid detergent fiber
ADFCR	Average daily feed conversion ratio
ADFI	Average daily feed intake
ADG	Average daily gain
ADL	Acid detergent lignin
AME	Apparent metabolizable energy
CF	Crude fiber
CFCR	Cumulative feed conversion ratio
CFI	Cumulative feed intake
CFU	Colony forming unit
CP	Crude protein
СРСРКО	
CPK0 CPO	Crude palm kernel oil
	Crude palm oil
Cq	Quantification cycle
CWG	Cumulative weight gain
DDGE	Denaturing gradient gel electrophoresis
DF	Dietary fiber
DNA	Deoxyribonucleic acid
E	Amplification efficiency
EE	Ether extract
g	Gram
GIT	Gastrointestinal tract
HCl	Hydrochloric acid
HFFS	High fiber feedstuff
hr	Hour
IVDMD	In-vitro dry matter degradability
LAB	Lactic acid bacteria
MPOB	Malaysian palm oil board
MU	Million units
NaOH	Sodium hydroxide
NDF	Neutral detergent fiber
NFE	Nitrogen free extract
NH ₃	Ammonia
NSP	Non-starch polysaccharide
NTC	No-template control
РКМ	Palm kernel meal
POS	Palm oil sludge
PPF	Palm press fiber
ppm	Part per million
qPCR	Quantitative real-time polymerase chain reaction
rRNA	Ribosomal ribonucleic acid
SBM	Soybean meal
spp	Subspecies
SSF	Solid state fermentation
TGGE	Temperature gradient gel electrophoresis
T-RFLP	Terminal restriction fragment length polymorphism
VFA	Volatile fatty acid
	······································

wt Weight X.L.D Xylose-lysine-Deoxycholate





CHAPTER 1

INTRODUCTION

The used of agriculture by-products such as palm kernel meal (PKM) in broilers diet has been widely practiced worldwide. It could spare conventional feed ingredients such as maize and soybeans because it is relatively cheap, easily available (Onuh *et al.*, 2010) and virtually has no competition between man and farm animals (Kperegbeyi and Ikperite, 2011). The PKM contains moderate amount of protein and energy (Alimon, 2004); however, the nutrient value of PKM is considered very low, due to poor amino acid contents particularly the essential amino acids such as lysine and methionine (Sundu *et al.*, 2006) besides the high fiber content in the form of non-starch polysaccharides (NSPs) (Sekoni *et al.*, 2008) contributed to its low inclusion in the broiler diet.

It has been reported that, the optimum inclusion of PKM in broilers diet varied from one study to another. Sekoni *et al.* (2008) reported that PKM can be added in the diet of broilers up to 20%, without having any detrimental effect on their growth performances. In contrast, in a study conducted by Adrizal *et al.*(2011) found that the inclusion can be varied between 20 to 30%. In another study, Saenphoom *et al.*(2013) reported that 20% PKM can only be included in the grower diet of broilers, whilst they observed that 5% PKM can be added in the starter diet of broilers without impairing their growth performances. However, there is still lack of information on the optimum utilization of PKM in thestarter diet of broilers.

The used of high fiber feedstuff such PKM in the broiler diets can impair the digestibility of the diet due to fact that broilers have a limited capacity to assimilate dietary fiber because they lack of fiber digestive enzymic activity in their gastrointestinal tract. The digestibility of feed, particularly apparent nitrogen and calcium retention is found to decrease with the inclusion of PKM, although the feed intake is increased (Sundu *et al.*, 2005). Reducing in apparent nitrogen digestibility for example, indicates that the protein of PKM is entrapped in the cell walls, making it unavailable and difficult for digestive enzymes to digest it and to be absorbed by broilers. Furthermore, contamination of PKM with the nut shell also contributed to the poor digestibility of PKM by broilers. Since the digestibility of PKM is very low, efforts are made to improve the digestibility and utilization of PKM by broilers. Previously, several studies have shown that the supplementation of exogenous enzyme in the PKM diet for example, could improve its nutritive quality and make it more available to the broilers through the enhance in the digestibility of nutrients entrapped in the cell wall of PKM (Chong *et al.*, 2008; Sekoni *et al.*, 2008).

The research on feeding of high fiber feedstuffs to broilers is typically focusing on their detrimental effect on the digestibility of feed and performance of broilers. However, there is still lack of understanding on how fiber is actually degraded in the intestinal tract, particularly in the caeca, and the potential of feeding high fiber feedstuffs such as PKM to broilers. It has been reported that, dietary fibers are fermented in the caeca by caecal microorganisms which produces end products such as volatile fatty acids (VFAs) that play an important role in controlling the proliferation of pathogenic bacteria such as *Salmonella* spp. (van der Weilan, 2000). The assessment of high fiber feedstuffs fermentability by caecal and/or faecal microorganism can be measured by measuring the gas production (Menke *et al.*, 1979), and a study conducted by McLean *et al.* (2004) showed that supplementation of enzymes in the high fiber diet had influenced the fermentation characteristic of the diet tested. Thus, it is also important to examine the extent of fiber fermentation in the caeca of broilers in response to enzyme supplementation, and it effects on microbial population in the caeca.

In general, the basis of inclusion of enzyme in PKM diet of broilers is to reduce the effect of non-starch polysaccharides in PKM especially to promote their growth performance through the increases in broilers' feed efficiency and digestibility. However, there is no report on their effect on the fermentation profiles of PKM diet in the caeca of broilers and on caecal bacteria population. The use of exogenous enzymes could improved the nutritional value of broilers diet, facilitate the growth of beneficial gut microflora of broilers and consequently, improve the utilization of feeds by maximizing the conversion of dietary nutrients into broilers meat (Bedford and Coweison, 2012). Thus, this study was conducted to study the effect of dietary enzymes supplementation on early growth performance, caecal fermentation profiles and caecal bacteria population of broilers fed with PKM diet.

1.1 RESEARCH HYPOTHESIS

The supplementation of xylanase and cellulase in PKM diet fed to broilers will influence the caecal fermentation characteristics of PKM diet, will improve the early growth performances of broilers, and will also reduce the population of pathogenic bacteria in the caecum.

1.2 OBJECTIVES

- i. To examine the fermentation profiles of caeca in response to xylanase and cellulase supplementation.
- ii. To determine the effect of xylanase and cellulase supplementation in PKMdiet on the early growthperformance of the broilers.
- iii. To determine the effect of xylanase and cellulasesupplementation in PKMdiet on caecal bacteria counts and population through culture method and real-time PCR analysis.
- iv. To determine the effect of xylanase and cellulase supplementation in PKMdiet on the VFAs production of caecal digesta.



REFERENCES

- A'dilah, M.M., Alimon, A.R. (2011). Improving the nutritive value of palm kernel cake (pkc) through chemical pre-treatment and fungal fermentation. Proceedings of the 32nd Malaysian Society of Animal Production (MSAP) Annual Conference, Tawau, Malaysia, June 3-5, 2011.
- Adeola, O., Jendza, J.A., Southern, L.L., Powell, S. and Owusu-Asiedu, A. (2010). Contribution of exogenous dietary carbohydrases to the metabolizable energy value of corn distillers grains for broiler chickens. *Poultry Science*. 89: 1947-1954.
- Adesehinwa, A.O.K. (2007). Utilization of palm kernel cake as a replacement for maize in diets of growing pigs: Effects on performance, serum metabolites, nutrient digestibility and cost of feed conversion. *Bulgarian Journal of Agriculture Science*. 13: 593-600.
- Adrizal, A., Yusrizal, Y., Fakhri, S., Haris, W., Ali, E. and Angel, C.R. (2011). Feeding native laying hens diets containing palm kernel meal with or without enzyme supplementations: 1. Feed conversion ratio and egg production. *Journal Applied Poultry Research*. 20: 40-49.
- Ahmad Borhan, A.N., Venugopal, R., Amiruddin, N. and Simeh, M.A. (2005). Palm kernel cake marketing: Constraints and prospects. *Oil Palm Industry Economic Journal*. 5(2): 37-46.
- Ahrens, F., Schön, J., Schmitz, M. (1991). A discontinuous *in-vitro* technique for measuring hind guts fermentation in pigs. In M.W.A. Verstegen, J. Huisman, and L.A. den Hortog. *Digestive Physiology in Pigs*. (pp. 226-230). No.54. Pudoc, Wageningen, the Netherlands: EAAP Publication.
- Alimon, A.R. (2004). The nutritive value of palm kernel cake for animal feed. *Palm Oil Developments*.40: 12-16.
- Aneato, M., Chioma, G.O. and Omeshi, D.J. (2009). Palm kernel cake as substitute for maize in broiler finisher diet. *International Journal of Poultry Science*. 8(12): 1206-1208.
- Angle, C.R., Saylor, W., Viera, S.L. and Ward, N. (2011). Effects of a monocomponent protease on performance and protein utilization in 7 to 22 day-old broiler chickens. *Poultry Science*. 90: 2281-2286.
- Anisson, E.F., Hill,K.J. and Kenworthy,R. (1968). Volatile fatty acids in the digestive tract of the fowl. *British Journal of Nutrition*.22: 207-216.
- Ao, T., Cantor, H.A., Pescatore, A.J., Ford, M.J., Pierce, J.L. and Dawson, K.A. (2009). Effect of enzyme supplementation and acidification of diets on nutrient digestibility and growth performance of broiler chicks. *Poultry Science*. 88: 111-117.

- AOAC (1997). Official methods of analysis, 16th Edition. Association of Official Analytical Chemists, Arlington, VA, USA. ISBN 0-935584-54-4.
- Apajalahti, J.H.A., Kettunen, A. and Graham, H. (2004). Characteristics of gastrointestinal microbial communities, with special reference to the chickens. *World's Poultry Science Journal*. 60: 223-232.
- Bach Knudsen, K.E. (2001). The nutritional significance of "dietary fiber" analysis. *Animal Feed Science and Technology*. 90: 3-20.
- Barnes, E.M., Mead, G.C., Barnum, D.A. and Harry, E.G. (1972). The intestinal flora of the chicken in the period 2 to 6 weeks of age, with particular reference to the anaerobic bacteria. *British Poultry Science*. 13: 311-326.
- Barnes, E.M. (1979). The intestinal microflora of poultry and game birds during life and after storage. *Journal of Applied Bacteriology*.46: 407-419.
- Bartosch, S., Fite, A., MacFarlane, G.T., McMurdo, M.E. (2005). Characterization of bacterial communities in feces from healthy elderly volunteers and hospitalized elderly patients by using real-time PCR and efficient of antibiotic treatment on the fecal microbiota. *Applied Environmental and Microbiology*. 70: 3575-3581.
- Beckmann, L., Simon, O. and Vahjen, W. (2006). Isolation and identification of mixed linked β-glucan degrading bacteria in the intestine of broiler chickens and partial characterization of respective 1, 3-1, 4-β-glucanase activities. *Journal of Basic Microbiology*. 46: 175-185.
- Bedbury, H.P. and Duke, G.E. (1983). Cecal microflora of turkeys fed low or high fiber diets: Enumeration, identification and determination of cellulolytic activity. *Poultry Science*. 62: 675-682.
- Bedford, M.R. and Morgan, A.J. (1996). The use of enzymes in poultry diets. *World's Poultry Science Journal*. 52:61-68.
- Bedford, M.R. (2000). Exogenous enzymes in monogastric nutrition: Their current value and future benefits. *Animal Feed Science and Technology*. 86: 1-13.
- Bedford, M.R. (2002). The role of carbohydrases in feedstuff digestion. In J. McNab and K.N. Boorman. *Poultry Feedstuffs-Supply, Composition and Nutritive Value*. (pp. 319-336). England: CABI Publishers.
- Bedford, M.R. and Cowieson, A.J. (2012). Exogenous enzymes and their effects on intestinal microbiology. *Animal Feed Science and Technology*. 173: 76-85.
- Bhat, M.K. (2000). Cellulases and related enzymes in biotechnology. *Biotechnology Advanced*. 18: 355-383.
- Brenes, B., Smith, M., Guenter, Marquardt, W. (1993). Effect of enzyme supplementation on the performance and digestive tract size of broiler chickens fed wheat- and barley-based diets. *Poultry Science*. 72: 1731-1739.

- Castillo, M., Martín-Orúe, S.M., Manzanilla, E.G., Badiola, I., Martín, M. and Gasa, J. (2006). Quantification of total bacteria, enterobacteria and *Lactobacilli*populations in pig digesta by real-time PCR. *Veterinary Microbiology*.114: 165-170.
- Cheng Hai, T. (2011). The palm oil industry in Malaysia: From seed to frying pan. <u>http://www.studymode.come/essays/The-Palm-Oil-Industry-In-Malaysia-</u>776923.html.
- Chenost, M. and Kayouli, C. (1997). Rough utilization in warm climates. *Animal Production and Health, FAO.* 135.
- Chesson, A. (1993). Feed enzyme. Animal Feed Science and Technology. 45: 65-79.
- Chin, F.Y. (2001). Palm kernel cake (pkc) as a supplement for fattening and dairy cattle in Malaysia, In *Forage Development in Southeast Asia; Strategies and Impact*, 7th meeting of the Regional Working Group on Grazing and Feed Resources, Universiti of Sam Rapulangi, Manado, Indonesia, July 2-7, 2001. Manado. Moog, F.A., Reynolds, S.G. and Maaruf, K. Ed; FAO, 2001.
- Choct, M.R., Hughes, R.J., Wang, J., Bedford, M.R., Morgan, A.J. and Anisson, G. (1996). Increased small intestinal fermentation is partly responsible for the antinutritive activity of non-starch polysaccharides in chickens. *British Poultry Science*. 37:609-6211.
- Choct, M.R., Hughes, R.J. and Bedford, M.R. (1999). Effects of a xylanase on individual bird variation, starch digestion throughout the intestine, ileal and cecal volatile fatty acid production in chickens fed wheat. *British Poultry Science*.40: 419-422.
- Chong, C.H., Zulkifli, I. and Blair, R. (2008). Effects of dietary inclusion of palm kernel cake and palm oil, and enzyme supplementation on performance of laying hens. *Asian-Australasian Journal of Animal Science*. 21(7): 1053-1058.
- Coates, M.E., Ford, J.E. and Harrison, G.F. (1968). Intestinal synthesis of vitamins of the B complex in chicks. *British Journal of Nutrition*. 22: 493-498.
- Coles, L.T., Moughan, P.J. and Darragh, A.J. (2005). *In-vitro* digestion and fermentation methods, including gas production techniques, as applied to nutritive evaluation of foods in the hindgut of humans and other simple-stomached animal. *Animal Feed Science and Technology*. 123-124: 421-444.
- Coloe, P.J., Bagust, T.J. and Ireland, L. (1984). Development of the normal gastrointestinal microflora of specific pathogen-free chicken. *Journal of Hygiene Cambridge*. 92: 79-87.
- Cornu, A., Besle, J.M., Mosoni, P. and Grenet, E. (1994). Lignin-carbohydrate complexes in forages: Structure and consequences in the ruminal degradation of cell-wall carbohydrates. *Reproduction and Nutrition Development*. 34: 385-398.

- Coweison, A.J. (2005). Factors that affect the nutritional value of maize for broilers. *Animal Feed Science and Technology*. 119 (3-4): 293-305.
- Cowieson, A.J., Singh, D.N. and Adeola, O. (2006). Prediction of ingredient quality and the effect of a combination of xylanase, amylase, protease and phytase in the diets of broiler chicks. 1. Growth performance and digestible nutrient intake. *British Poultry Science*. 47(4): 477-489.
- Cox, N.A., McHan, F. and Bailey, J.S. (1994). Effect of butyric or lactic acid on the *in-vitro* colonization of *Salmonella typhimurium*. *Journal of Applied Poultry Research*. 3: 315-318.
- Cummings, J.H. and MacFarlane, G.T. (1991). The control and consequences of bacterial fermentation in the human colon. *Journal Applied Bacteriology*. 70: 443-459.
- Cummings, J.H. and Macfarlane, G.T. (1997). Role of intestinal bacteria in nutrient metabolism. *Journal of Clinical Nutrition*. 16:3-11.
- Dairo, F.A.S. and Fasuyi, A.O. (2008). Evaluation of fermented palm kernel meal and fermented copra meal proteins as substitute for soybean meal protein in laying hens diets. *Journal of Central European Agriculture*. 9: 33-47.
- Dänicke, S., Vahjen, W., Simon, O. and Jeroch, H. (1998). Effects of dietary fat type and xylanase supplementation to rye-based broiler diets on selected bacterial groups adhering to the intestinal epithelium, on transit time of feed, and on nutrient digestibility. *Poultry Science*. 78: 1292-1299.
- Donalson, L.M., Kim, W.K., Chalova, V.I., Herrera, P., McReynolds, J.L., Gotcheva, V.G., Vidonovič, D., Woodward, C.L., Kubena, L.F., Nisbet, D.J. and Ricke, S.C. (2008). *In-vitro* fermentation response of laying hen cecal bacteria to combinations of fructooligosaccharides prebiotics with alfalfa or a layer ration. *Poultry Science*. 87: 1263-1275.
- Duke, G.E., Ecclestone, E., Kirkwood, S., Louis, C.F. and Bedbury, H.P. (1984). Cellulose digestion by domestic turkeys fed low or high fibre diets. *Journal of Nutrition*. 114: 95-102.
- Dunkley, D.K., Dunkley, C.S., Njongmeta, N.L., Callaway, T.R., Hume, M.E., Kubena, L.F., Nisbet, D.J. and Ricke, S.C. (2007). Comparison of *in-vitro* fermentation and molecular microbial profiles of high fiber substrates incubated with chicken cecal inocula. *Poultry Science*. 86: 801-810.
- Dusterhoft, E.M. and Voragen, A.G.J. (1991). Non-starch polysaccharides from sunflower (*Helianthus annus*) and palm kernel (*Elaeis guineensis*) meal preparation of cell wall material and extraction of polysaccharides fractions. *Journal of Science and Food Agriculture*. 55: 411-422.

- Ekinchi, M.S., McCrae, S.I. and Flint, H.J. (1997). Isolation and overexpression of a gene encoding an extracellular β- (1, 3-1, 4)-glucanase from *Streptoccocus Bovis*. *JB1 Applied Environment and Microbiology*. 63: 3752-3756.
- Elaeis guineensis, the palm. Retrieved 21st June 2014 from<u>http://www.etawau.com/OilPalm/Elaeis_guineens.htm</u>.
- El-Deek, A.A., Al-harthi, M. and Yakout, H.M. (2008). Use of enzymes to supplements diets containing date waste meal for Lohmann white layers. *International Journal ofPoultry Science*. 7(4): 397-407.
- Engberg, R.M., Hedemann, M.S., Steenfeldt, S. and Jensen, B.B. (2004). Influence of whole wheat and xylanase on broiler performance and microbial composition and activity in the digestive tract. *Poultry Science*. 83: 925-938.
- Ezema, C. and Eze, D.C. (2009). Performance and economic benefit of broilers fed palm kernel cake-based diet supplemented with probiotic. *International Journal of Poultry Science*. 8(10): 1003-1005.
- Ezieshi, E.V. and Olomu, J.M. (2008). Nutritional evaluation of palm kernel meal types: 2. Effects on live performance and nutrition retention in broiler chicks diets. *African Journal of Biotechnology*. 7: 1171-1175.
- Fonty, G. and Gouet, P.H. (1989). Fiber-degrading microorganisms in the monogastric digestive tract. *Animal Feed Science and Technology*. 23: 91-107.
- Frahm, E. and Obst, U. (2009). Application of the fluorogenic probe technique (TaqMan PCR) to the detection of *Enterococcus* spp. and *Escherichia coli* in water samples. *Journal Microbiology Methods*. 52: 123-131.
- Francesh, M. and Geraert, P.A. (2009). Enzyme complex containing carbohydrases and phytase improves growth performance and bone mineralization of broilers fed reduced nutrient corn-soyabean-based diet. *Poultry Science*. 88: 1915-1924.
- Friesen, O.D., Guenter, W., Marquardt, R.R. and Rotter, B.A. (1992). The effect of enzyme supplementation on the apparent metabolizable energy and nutrient digestibilities of wheat, barley, oats, and rye for the young broiler chick. *Poultry Science*. 71: 1710–1721.
- Geier, M.S., Torok, V.A., Allison, G.E., Ophel-Keller, K. and Hughes, R.J. (2009). Indigestible carbohydrates alter intestinal microbiota but do not influence the performance of broiler chickens. *Journal of Applied Microbiology*. 106: 1540-1548.
- Gerard, H., Ducatelle, R. and Immerseel, F.V. (2011). An update on alternatives to antimicrobial growth promoters for broilers. *The Veterinary Journal*. 187: 182-188.
- Goa F, Jiang, Y., Zhuo, G.H. and Han, Z.K. (2008). The effects of xylanase supplementation on performance, characteristics of the gastrointestinal tract,

blood parameters and gut microflora in broilers fed on wheat-based diets. *Animal Feed Science and Technology*. 142: 173-184.

- Goldstein, D.L. (1989). Absorption by the cecum of wild birds: Is there interspecific variation. *Journal of Experimental Zoology Supplement*. 3: 103-110.
- Gong, J., Forster, R.J., Yu, H., Chambers, J.R., Wheatcroft, R., Sabour, P.M. and Chen, S. (2002). Molecular analysis of bacterial populations in the ileum of broiler chickens and comparison of the bacteria in the cecum. *FEMSMicrobiology Ecology*. 41: 171-179.
- Graham, H. and Pettersson, D. (1992). A note on the effect of beta-glucanase and a multi-enzyme on production in broiler chicks fed a barley-based diet. *Journal of Agricultural Research*. 22: 39-42.
- Groot, J.C.J., Williams, B.A., Debersaques, F.M.A. and Lantiga, A. (1996). Multiphasic analysis of gas production kinetics for *in-vitro* fermentation of ruminant feeds. *Animal Feed Science and Technology*. 64: 77-99.
- Gunawardana, P., Roland, D.A., Bryant, M.M. (2009). Effect of dietary energy, protein, and a versatile enzyme on hen performance, egg solids, egg composition, and egg quality of Hy-Line W36 hens during second cycle, phase two. *Journal Applied Poultry Research*. 18: 43-53.
- Guo, F.C., Williams, B.A., Kwakkel, R.P. and Verstegen, M.W.A. (2003). *In-vitro* fermentation characteristics of two mushroom species, and herb, and their polysaccharide fractions, using chicken cecal contents as inoculum. *Poultry Science*.82: 1608-1615.
- Han, Y.K., Jin, Y.H., Kim, J.H. and Thacker, P.F. (2010). Influence of enzyme and/or lysolecithin on performance, nutrient digestibility and egg quality for laying hens. *Trends inAnimal Veterinary Science*. 1(1): 28-35.
- Haruna, U. and Hamidu, B.M. (2004). Economic analysis of turkey production in the wewstern agricultural zone of Bauchi, *In economic analysis of broiler production at miango plateau state, Nigeria*, Proceedings of the 9th Annual Conference of Animal Science Nigeria, Sept. 4-6, 2004.
- Hassanien, H.H.M., Sanaa and Elnagar, H.M. (2011). Comparison difference levels of phytase enzyme supplementation on laying hen performances, egg quality and some blood parameters. *Asian Journal of Poultry Science*. 5: 77-85.
- Hishamuddin, M.A. (2001). Malaysia palm kernel cake as animal feed. *Palm Oil Developments*. 34: 4-6.
- Iji, P.A., Khumalo, K., Slippers, S. and Gous, R.M. (2004). Intestinal function and body growth of broiler chickens on maize-based diets supplemented with mimosa tannins and a microbial enzyme. *Journal of Science Food and Agriculture*. 84 (12): 1451-1458.

- Iluyemi, F.B., Hanafi, M.M., Radziah, O. and Kamarudin, M.S. (2006). Fungal solid state culture of palm kernel cake. *Bioresources Technology*. 97: 477-482.
- Jaafar, M.D. and Jarvis, M.C. (1992). Mannans of oil palm kernels. *Phytochemistry*. 31: 463-464.
- Jahromi, M.F., Liang, J.B., Rosfarizan, M., Goh, Y.M., Shokryazdan, P. and Ho, Y.W. (2013). Lovastatin-enriched rice straw enhances biomass quality and suppresses ruminal methanogenesis. *BioMed Research International*. 2013: 1-10.
- Jamroz, D., Wiliczkiewicz, A., Orda, J. and Skorupiňska, J. (1994). Ileal and postileal fermentation of cereal carbohydrates in chicken and duck Wein. *TierarztlMonatsschr.* 81: 80-84.
- Jamroz, D., Jakobsen, K., Bach Knudsen, K.E., Wilczkiewicz, A. and Orda, J. (2002). Digestibility and energy value of the non-starch polysaccharides in young chickens, ducks and geese, fed diets containing high amounts of barley. *Comparative Biochemistry and Physiology*. 131: 657-668.
- Jonsson, E. and Hemmingson, S. (1991). Establishment in the piglet gut of *Lactobacilli* capable of degrading mixed-linked β-glucan. *Journal of Applied Bacteriology*. 70: 512-516.
- Jørgensen, H., Zhao, X.Q., Knudsen, K.E. and Eggum, B.O. (1996). The influence of dietary fiber source and level on the development of the gastrointestinal tract, digestibility and energy metabolism in broiler chickens. *BritishJournal of Nutrition.* 75: 379-395.
- Juanpere, J., Pérez-Vendrell, A.M. and Brufau, J. (2004). Effect of microbial phytase on broilers fed barley-based diets in the presence or not of endogenous phytase. *Animal Feed Science and Technology*. 115: 265-279.
- Józefiak, D., Rutkowski, A. and Martin, S.A. (2004). Carbohydrate fermentation in the avian ceaca. *Animal Feed Science and Technology*. 113: 1-15.
- Józefiak, D., Rutkowski, A., Jensen, B.B. and Engberg, R.M. (2006). The effect of βglucanase supplementation of barley- and oat-based diets on growth performance and fermentation in broiler chicken gastrointestinal tract. *British Poultry Science*. 47: 57-64.
- Józefiak, D., Rutkowski, A., Kaczmarek, S., Jensen, B.B., Engberg, R.M. and Højberg, O. (2010). Effect of β-glucanase and xylanase supplementation of barley and rye-based diets on ceacal microbiota of broiler chickens. *BritishPoultry Science*. 51(4): 546-557.
- Kaur, B. and Fatimah, M.A. (2007). Marketing of poultry in Malaysia: Structural issues and challenges. 50 years of Malaysian Agricultures. *International Issues Challenges and Directions*, pp. 585-613. Universiti Putra Malaysia.

- Khattak, F.M., Pasha, T.N., Hayat, Z. and Mahmud, A. (2006). Enzymes in poultry nutrition. *Journal Animal and Plant Science*. 16: 1-2.
- Khattak, A.B., Zeb, A., Bibi, N., Khalil, S.A. and Khattak, M.S. (2007). Influence of germination techniques on phytic acid and polyphenols content of chickpea (*Cicerarietinum L.*) sprouts. *Food Chemistry*. 104: 1074-1079.
- Kperegbeyi, J.I. and Ikperite, S.E. (2011). The effectiveness of replacing maize with palm kernel cake in broilers' starter diets. *Journal of Environmental Issues and Agriculture in Development Country*. 3 (1): 145-149.
- Kwan, Y.M. and Ricke, S.C. (1998). Introduction of acid resistance of *Salmonella typhimurium* by exposure to short chain fatty acids. *Applied Environmental and Microbiology*. 64: 3458-3463.
- Lan, Y., Xun, S., Tamingga, S., Williams, B.A., Verstegen, M.W.A. and Erdit, G. (2004). Real-time PCR detection of lactic acid bacteria in cecal contents of *Eimeria tenella*-infected broilers fed soybean oligosaccharides and soluble soybean polysaccharides. *Poultry Science*. 83: 1696-1702.
- Lan, Y., Williams, B.A., Tamingga, S., Boer, H., Akkermans, A., Erdi, G. and Verstegen, M.W.A. (2005). *In-vitro* fermentation kinetics of some non-digestible carbohydrates by the caecal microbial community of broilers. *Animal Feed Science and Technology*.123-124: 687-702.
- Lattimer, J.M., Cooper, S.R., Freeman, D.W. and Lalman, D.L. (2007). Effect of *Aspergillus* yeast culture on *in-vitro* fermentation of high-concentrate or high fiber diet using equine fecal inoculums in a Daisy ¹¹ incubator. *Journal of Animal Science*. 85: 2848-2491.
- Lawal, T.E., Iyayi, E.A., Adeniyi, B.A. and Adaromoye, O.A. (2010). Biodegradation of palm kernel cake with multi-enzymes complexes from fungi and its feeding value for broilers. *International Journal of Poultry Science*. 9(7): 695-701.
- Leeson, S., Namkung, H., Antongiovanni, M. and Lee, E.H. (2005). Effect of butyric acid on the performance and carcass yield of broiler chickens. *Poultry Science*. 88: 1418-1422.
- Leslie, M.A., Moran Jr, E.T. and Bedford, M.R. (2007). The effect of phytase and glucanase on the ileal digestible energy of corn and soyabean meal fed to broilers. *Poultry Science*. 86: 2350-2357.
- Lou, D., Yang, F., Yang, X., Yoa, J., Shi, B. and Zhou, Z. (2009). Effects of xylanase on performance, blood parameters, intestinal morphology, microflora and digestive enzyme activities of broilers fed wheat-based diets. *Asian-Australasian Journal of Animal Sciences*. 22(9): 1288-1295.

- Lu, J., Idris, U., Harmon, B., Hofacre, C., Maurer, J.J. and Lee, M.D. (2003). Diversity and succession of the intestinal bacterial community of the maturing broiler chicken. *Applied Environmental and Microbiology*. 69: 6816-6824.
- Lui, N., Ru, Y.J., Tang, D.F., Xu, T.S. and Patridge, G.G. (2011). Effects of corn distillers dried grains with soluble and xylanase on growth performance and digestibility of diet components in broilers. *Animal Feed Science and Technology*. 163: 260-266.
- Luo, D., Yang, F., Yang, X., Yoa, J., Shi, B. and Zhou, Z. (2009). Effects of xylanase on performance, blood parameters, intestinal morphology, microflora and digestive enzyme activities of broilers fed wheat-based diets. *Asian-Australasian Journal of Animal Science*.22: 1288-1295.
- Macfarlane, S. and Macfarlane, G.T. (2003). Regulation of short-chain fatty acid production, In *Short-chain fatty acids*. Proceedings of the Nutrition Society, Edinburgh, United Kingdom, May 27-30, 2003.
- Marini, A.M., Daud, M.J., Noraini, S., Jame'ah, H. and Engku Azahan, E.A. (2005). Performance of locally isolated microorganisms in degrading palm kernel cake (pkc) and improving the nutritional value of fermented pkc. *Journal of Tropical Agriculture and Food Science*. 33(2): 311-319.
- Marounek, M., Suchorska, O. and Savka, O. (1999). Effect of substrate and feed antibiotics on *in-vitro* production of volatile fatty acids in caecal contents of chickens. *Animal Feed Science and Technology*. 80: 223-230.
- Martínez, M., Savón, L., Dihigo, L.E., Hernandez, Y., Oramas, A., Sierra, F., Montejo, A., Cucto, M. and Herrera, F.R. (2010). Cecal and blood fermentative indicators in broiler chickens fed *morus alba* foliage meal in the ration. *Cuban Journal of Agricultural Science*. 44(1): 49-53.
- Mateos, G.G., Jiménez-Moreno, E., Serrano, M.P. and Lázaro, R.P. (2012). Poultry response to high levels of dietary fiber sources varying in physical and chemical characteristics. *Journal of Applied Poultry Research*. 21: 156-174.
- McDaniel, A.C., Martin, S.A., McCann, J.S. and Parks, A.H. (1993). Effect of *Aspergillus oryzae* fermentation extract on *in-vitro* equine cecal fermentation. *Journal of Animal Science*. 71: 2164-2172.
- McHan, F. and Shotts, E.B. (1993). Effect of short-chain fatty acids on the growth of *Salmonella typhimurium* in an *in-vitro* system. *Avian Diseases*. 37: 396-398.
- McLean, J., Nevison, I., Bertin, G. and Acamovic, T. (2004). The feasibility of using nutritional modifications to replace drugs in poultry feeds. *Journal of Applied Poultry Research*. 11(4): 437-452.
- McNab, J.M. (1973). The avian ceca: A review. *World's Poultry Science Journal*. 29: 251-263.

- Mead, G.C. (1997). Bacteria in the gastrointestinal tract of birds. In R.I. Mackie, B.A. White, and R.E. Isaacson. *Gastrointestinal microbiology*, vol. 2. (pp. 216-240). New York: Chapman and Hall Publishers.
- Meimandipour, A., Shuhaimi, M., Hair-Bejo, M., Azhar, K., Kabeir, B.M., Rasti, B. and Yazid, A.M. (2009). *In-vitro* fermentation of broiler cecal content: The role of *Lactobacilli* and pH value on the composition of microbiota and end products fermentation. *Applied Microbiology*. 49: 415-420.
- Menke, K.H., Raab, L., Salewski, A., Steingass, H., Fritz, D. and Schneider, W. (1979). The estimation of digestibility and metabolizable energy content of ruminant's feedstuffs from gas production when they are incubated with rumen liquor *in-vitro*. *Journal of Agricultural Science in Cambridge*. 93: 217-222.
- Menke, K.H. and Steingass, H. (1988). Estimation of energetic feed value obtained from chemical analysis and *in-vitro* gas production using rumen fluid. *Animal Research Development*. 28: 7-55.
- Metzler, B., Bauer, B. and Mosenthin, R. (2005).Microflora management in the gastrointestinal tract of piglets.*Asian-Australasian Journal of Animal Science*.18: 1353-1362.
- Miller, M.R. (1976). Cecal fermentation in mallards in relation to diet. *Condor*. 78(1): 107-111.
- Mirnawati, Rizal, Y., Marlida, Y. and Kompiang, P. (2010). The role of humic acid in palm kernel cake fermented by *Aspergillus niger* for poultry ration. *Pakistan Journal of Nutrition.* 9(2): 182-185.
- Mustafa, M.F., Alimon, A.R., Wan Zahari, M.W., Ismail, I. and Hair-Bejo, M. (2004). Nutrient digestibility of palm kernel cake for Muscovy ducks. *Asian-Australasian Journal of Animal Science*. 17(4): 514-17.
- Nam, H.M., Srinivasan, V., Gillespie, B.E., Murinda, S.E. and Oliver, S.P. (2005). Application of SYBR green real-time PCR assay for specific detection of *Salmonella* spp. in dairy farm environmental samples.*International Journal of Food Microbiology*. 102: 161-171.
- Navidshad, B., Liang, J.B. and Jahromi, M.F. (2012). Correlation coefficients between different methods of expressing bacterial quantification using real time PCR. *International Journal of Molecular Sciences*. 13: 2119-2132.
- Ng, W.K. and Chong, K.K. (2002). The nutritive value of palm kernel meal and the effect of enzyme supplementation in practical diets for red hybrid tilapia (*Oreochromis* sp.). *Asian Fisheries Science*. 15: 167-176.
- Ng, W.K., Lim, H.K., Lim, S.W. and Ibrahim, O. (2002). Nutritive value of palm kernel meal pretreated with enzyme or fermented with *Trichoderma koningii* (*Oudemans*) as a dietary ingredient for red hybrid tilapia (*Oreochromis* sp.). *Aqua Research.* 33: 1119-1207.

- Ng, W.K. (2004). Researching the use of palm kernel cake in aquaculture feeds. *Palm Oil Developments*. 41: 19-21.
- Nian, F., Guo, Y.M., Ru, Y.J., Peron, A. and Li, F.D. (2011). Effects of xylanase supplementation on the net energy for production, performance and gut microflora of broilers fed corn/soyabean-based diet. *Asian-Australasian Journal of Animal Science*. 24(9): 1282-1287.
- O'Mara, F.P., Mulligan, F.J., Cronin, E.J., Rath, M. and Caffery, P.J. (1999). The nutritive value of palm kernel meal measured *in-vivo* and using rumen fluid and enzymatic techniques. *Livestocks Production Science*. 60: 305-316.
- Olukosi, O.A., Bedford, M.R. and Adeola, O. (2007). Xylanase in diets for growing pigs and broiler chicks. *Canadian Journal of Animal Science*.87: 227-235.
- Oluwafemi, R.A. and Akpodiete, O.J. (2010). Carcass characteristics and meat quality of weaner pigs fed palm kernel cake based ration. *Electronic Journal of Environment Agriculture and Food Chemistry*. 9(1): 123-128.
- Onuh, S.O., Ortserga, D.D. and Okoh, J.J. (2010). Response of broilers chickens to palm kernel cake and maize offal mixed in different ratios. *Pakistan Journal of Nutrition*. 9(6): 516-519.
- Onwudike, O.C. (1996). Oil palm (*Elaeis guineensis* Jacq.). In E. and Nwokolo, J. Smartt. *Legume and Oilseeds in Nutrition*. (pp. 318-330). England: Chapman and Hall Publishers.
- Paloheimo, M., Piironen, J. and Vehmaanperä, J. (2010). Xylanases and cellulases as feed additives. In Enzymes in M.R. Bedford and G.G. Partridge. *Farm Animal Nutrition*.(pp. 12–53). London, UK: CAB International, 2nd edition.
- Parson, T.R., Maita, Y. and Laili, C.A. (1984). Mannual of chemical and biological methods for seawater analysis. Pergamon Press, New York.
- Ponte, P.I.P., Ferriera, L.M.A., Soares, M.A.C., Aguiar, M.A.N.M., Lemos, J.P.C., Mendes, I. and Fontes, C.M.G.A. (2004). Use of cellulases and xylanases to supplements diets containing alfalfa for broiler chicks: Effects on bird performance and skin color. *Journal Applied Poultry Research*. 13: 412-420.
- Ponte, P.I.P., Lordela, M.M.S., Guerreiro, C.I.PD., Soares, M.C., Mourõo, J.L., Crespo, J.P., Crespo, D.G., Prates, J.A.M., Ferreira, L.M.A. and Fontes, C.M.G.A. (2008). Corn β -glucanase activity limits the effectiveness of a recombinant cellulase used to supplement a barley-based feed for free range broilers. *British Poultry Science*. 49(3): 347-359.
- Poulsen, L.K., Licht, T.R., Rang, C., Krogfelt, K.A. and Molin, S. (1995). Physiological state of *Escherichia coli* BJ4 growing in the large intestines of streptomycin-treated mice. *Journal of Bacteriology*. 177: 5840-5845.

- Preston, C.M., McCracken, K.J. and Bedford, M.R. (2001). Effect of wheat content, fat source and enzyme supplementation on diet metabolisability and broiler performance. *British Poultry Science*. 425(5): 625-632.
- Rahim. F., Sabrina, Rusmawati and Syibli, F. (2007). Broiler small intestine villi response to feed containing palm kernel cake which fermented with *Rhizopus*. *Journal of IndonesiaTropical Animal Agriculture*. 32(4): 251-256.
- Rahner, R. (1901). Bakteriologische Mitteilungen ueber die Darmbakterien der Huehner. Zentbl. Bakteriol. Parasitendkd. 80: 239-244.
- Ramin, M., Alimon, A.R. and Ivan, M. (2010). Effects of fungal treatment on the *in-vitro* digestion of palm kernel cake. *Livestock Research for Rural Development*. 22(4).
- Redig, P. (1989). The avian ceca: Obligate combustion chambers or facultative afterburners. The conditioning influence of the diets. *Journal of Experiment Zoology and Supplied.* 3: 66-69.
- Rehman, H.U., Vahjen, W., Awad, W.A. and Zentek, J. (2007). Indigenous bacteria and bacterial metabolic products in the gastrointestinal tract of broiler chickens. *Archive of Animal Nutrition*. 61: 319-335.
- Rhule, S.W.A. (1996). Growth rate and carcass characteristics of pigs fed on diets containing palm kernel cake. *Animal Feed Science and Technology*. 61: 167-172.
- Ricke, S.C., Woodward, C.L., Kwon, Y.M., Kubena, L.F. and Nisbet, D.J. (2004).
 Limiting avian gastrointestinal tract *Salmonella* colonization by cecal anaerobic bacteria and a potential role for methanogens. In R.C. Beier, S.D Pillai, T.D Phillips and R.L Ziprin. *Preharvest and Postharvest Food safety: Contemporary Issues and Future Directions*. (pp. 141-150). Ames, Iowa: Blackwell Publishing Professional.
- Rodríguez, R., Martínez, M., Valdivié, M., Cisneros, M., Cárdenas, M. and Sarduy, L. (2006). Morphometry of the gastrointestinal tract and its accessory organs in laying hens fed feedstuffs containing proteinic sugarcane meal. *Cuban Journal of Agricultural Science*. 40:343.
- Rosin, E.A., Blank, G., Slominski, B.A. and Holley, R.A. (2007). Enzyme supplements in broiler chicken diets: *In-vitro* and *in-vivo* effects on bacterial growth. *Journal of Science and Food Agriculture*. 87: 1009-1020.
- Saengkerdsub, S., Kim, W., Anderson, R.C, Nisbet, D.J. and Ricke, C. (2006). Effects of nitrocompounds and feedstuffs on *in-vitro* methane production in chickens cecal contents and rumen fluid. *Anaerobe*. 12: 85-92.
- Saenphoom, P., Liang, J.B., Hol, Y.W., Loh, T.C. and Rosfarizan, M. (2011). Effect of enzyme treatment on chemical composition and production of reducing sugars in palm (*Elaeis guineenis*) kernel expeller. *African Journal of Biotechnology*. 10(68): 15372-15377.

- Saenphoom, P., Liang, J.B., Hol, Y.W., Loh, T.C. and Rosfarizan, M. (2013). Effects of enzyme treated palm kernel expeller on metabolizable energy, growth performance, villus height and digesta viscosity in broiler chickens. *Asian-Australasian Journal of Animal Sciences*. 26(4): 537-544.
- Salanitro, J.P., Blake, I.G., Muirhead, P.A., Maglio, M. and Goodman, J.R. (1978). Bacteria isolated from the duodenum, ileum and caecum of young chicks. *Applied Environment and Microbiology*. 35: 782-790.
- Saleh, F., Tahir, M., Ohtsuka, A. and Hayashi, K. (2005). A mixture of pure cellulase, hemicellulase and pectinase improves broiler performance. *British Poultry Science*. 46(5): 602-606.
- Salyers, A.A, Vercelloti, J.R., West, S.E.H. and Wilkins, T.D. (1977). Fermentation of mucus and plant polysaccharides by strains of bacteroides from the human colon. *Applied Environmental and Microbiology*.37: 319-322.
- Salyers, A.A., Gherardini. F. and O'Brien, M. (1981). Utilization of xylan by two species human colonic bacteroides. *Applied Environmental and Microbiology*. 41(4): 1065-1068.
- Saw, H.Y., Janaun, J., Kumaresan, S. and Chu, C.M. (2012). Characterization of the physical properties of palm kernel cake. *International Journal of Food Properties*. 15: 536-548.
- Sayyazadeh, H., Rahimi, G. and Rezai, M. (2006). Influence of enzyme supplementation of maize, wheat and barley-based diets on the performance of broiler chickens. *Pakistan Journal of Biological Science*. 9(4): 616-621.
- Sekoni, A.A., Omage, J.J., Bawa, G.S. and Esuga, P.M. (2008). Evaluation of enzyme (Maxigrain®) treatment of graded levels of palm kernel meal (pkm) on nutrient retention. *Pakistan Journal of Nutrition*. 7(4): 614-619.
- Selle, P.H., Ravindran, V. and Patridge, G.G. (2009). Beneficial effects of xylanase and/or phytase inclusions on ileal amino acid digestibility, energy utilization, mineral retention and growth performance in wheat-based broiler diets. *Animal Feed Science and Technology*. 153: 303-313.
- Setthapukdee, C., Jalaludin, S. and Mahyuddin, M. (1991). Effect of Supplementing grass and urea on the performance and carcass characteristics of lambs fed palm kernel cake-based diet.Proceedings of the 3rd International Symposium on the Nutrition of Herbivores, Penang, Malaysia, Aug. 25-30, 1991.
- Shakila, S., Reddy, P.S., Reddy, P.V.V.S., Ramana, J.V. and Ravi, A. (2012). Effect of palm kernel meal on the performance of broilers. *Tamilnadu Journal of Veterinary and Animal Science*. 8(4): 227-234.
- Shakouri, M.D., Iji, P.A., Mikkelson, L.L. and Cowieson, A.J. (2009). Intestinal function and gut microflora of broiler chickens as influenced by cereal grains and

microbial enzyme supplementation. *Journal of Animal Physiology and Animal Nutrition*. 93: 647-658.

- Shanmugavelu, S., Ruzickova, G., Zrustova, J. and Brooker, J.D. (2006). A fermentation assay to evaluate the effectiveness of antimicrobial agents on gut microflora. *Journal of Microbiological Methods*. 67: 93-101.
- Shapiro, S.K. and Sarles, W.B. (1949). Microorganisms in the intestinal tract of normal chickens. *Journal of Bacteriology*. 58: 531-544.
- Siew, W.L. (1989). Authenticity of palm kernel oil by fatty acid and triglyceride compositions. *PORIM Bulletin*. 19: 19-23.
- Silva, S.S.P. and Smithard, R.R. (2002). Effect of enzyme supplementation of a ryebased diet on xylanase activity in the small intestine of broilers, on intestinal crypt proliferation and nutrient digestibility and growth performance of the birds. *British Poultry Science*. 43: 274-282.
- Sohail, S.S., Bryant, M.M., Roland, D.A., Apajalahti, J.H.A. and Pierson, E.E.M. (2003).Influence of Avizyme 1500 on performance of commercial leghorns. *Journal of Applied Poultry Research*. 12 (1): 284-290.
- Soltan, M.A. (2009).Growth performance, immune response and carcass traits of broiler chicks fed on graded levels of palm kernel cake without or with enzyme supplementation. *Livestock Research for Rural Development*. 21:1-15.
- Stanley, E.G. (1990). Health and nutritional benefits from lactic acid bacteria. *FEMS Microbiology Revolution.* 87: 175-188.
- Steenfeldt, S., Hammershoj, M., Mullertz, A. and Jensen, F. (1998). Enzymes supplementation of wheat-based diet for broilers. 2. Effect on apparent metabolizable energy content and nutrient digestibility. *Animal Feed Science and Technology*. 75: 45-64.
- Stef, L., Dumitrescu, G., Drinceanu, D., Stef, D., Călin, J., Nicole, C., Cosmin, P. and Ioana, S. (2010). Impact of wheat non-starch polysaccharide on the intestinal wall and microflora in broilers. *Animal Science Biotechnology*. 43(1): 111-116.
- Strydom, J. and Cohen, S.A. (1994). Comparison of amino acid analyses by phenylisothiocyanate and 6-aminoquinoly-N-hydroxysuccinimidyl carbamate precolumn derivatization. *Analytical Biochemistry*. 222: 19-18.
- Sulabo, R.C., Ju, W.S. and Stein, H.H. (2013). Amino acid digestibility and concentration of digestible and metabolizable energy in copra meal, palm kernel expellers and palm kernel meal fed to growing pig. *Journal of Animal Science*. 91: 1391-1399.
- Sundu, B.A., Kumar, A. and Dingle, J. (2004). The effect of commercial enzymes on chicks fed high copra meal and palm kernel meal diets. Proceedings Seminar berkelanjutan, Indonesia. M.H. Husin. Ed.; Tadulako University Press: Indonesia.

- Sundu, B., Kumar, A. and Dingle, J. (2005). Response of birds fed increasing levels palm kernel meal supplemented with enzymes. *Australia Poultry Science Symposium*. 12: 63-75.
- Sundu, B., Kumar, A. and Dingle, J. (2006). Palm kernel meal in broiler diets: Effect on chicken performance and health. *World's Poultry Science Association*. 62 (2): 316-325.
- Tahir, M., Saleh, F., Ohtsuka, A. and Hayashi, K. (2008). An effective combination of carbohydrases that enables reduction of dietary protein in broilers. Importance of hemicellulase. *Poultry Science*. 87: 713-718.
- Tang, T.S. (2001). Quality and characteristics of Malaysian palm kernel cake/expeller. *Palm Oil Developments*. 34: 1-3.
- Tapia, L., López, J.L., Bueno, J.R. and Riva, R. (2000). Hojas de Leucaena leucocephala en la alimentación de patos. *Revolution Production Animal.* 12:41.
- Temmerman, R., Huys, G. and Swings, J. (2004). Identification of lactic acid bacteria: Culture-dependent and culture-independent methods. *Trends in Food Science and Technology*. 348-359.
- Tortuero, F., Brenas, A. and Riperez, J. (1975). The influence of intestinal (ceaca) flora on serum and egg yolk cholesterol levels in laying hens. *Poultry Science*. 54: 1935-1938.
- Vahjen, W., Gläser, K., Schäfer, K. and Simon, O. (1998). Influence of xylanasesupplemented feed on the development of selected bacterial groups in the intestinal tract of broiler chicks. *Journal of Agriculture Science Cambridge*. 130: 489-500.
- van der Wielan, P.W.J.J., Biesterveld, S., Notermans, S., Hofstra, H., Urlings, B.A. and vanKapen, F. (2000). Role of volatile fatty acids in development of the cecal microflora in broiler chickens during growth. *Applied Environmental and Microbiology*. 66: 2536-2540.
- van der Wielen, P. (2002). Dietary strategies to influences the gastrointestinal microflora of young animals and its potential to improve intestinal health. In M. C. Blok, H.A. Vahl, L. de Lange, A.E. van de Braak, G. Hemke, and M. Hessing *Nutrition and Health of the Gastrointestinal Tract*. (pp. 37-60). Wageningen, The Netherlands: Wageningen Academic Publishers.
- Van Immerseel, F., Rusell, J.B., Flythe, M.D., Gantois, J., Timbermont, L., Pasmans, F., Haesebrouck, F. and Ducatelle, R. (2006). The use of organic acids to combat Salmonella in poultry: a mechanistic explanation of the efficacy. *Avian pathology*. 35(3): 182-188.
- Varastegani, A. and Dahlan, I. (2014). Influence of dietary fiber levels on feed utilization and growth performance in poultry. *Journal of animal Science Advances*. 4(6): 422-429.

- Wang, J.J., Garlich, J.D. and Shih, J.C.H. (2005). Beneficial effects of versazyme, a keratinase feed additive, on body weight, feed conversion, and breast yield of broiler chickens. *Poultry Science Research*. 15:544-550.
- Wang, R.F., Cao, W.W. and Cerniglia, C.E. (1996). PCR detection and quantitation of predominant anaerobic bacteria in human and animal fecal samples. *Applied Environmental and Microbiology*. 62: 1242–1247.
- Watkins, B.A. and Kratzer, F.H. (1983). Effect of oral dosing of *Lactobacillus* strains on gut colonization and liver biotin in broiler chicks. *Poultry Science*. 62: 2088-2094.
- Wenk, C. (2001). The role of dietary fibre in the digestive physiology of the pig. *Animal Feed Science and Technology*. 90: 21-33.
- Williams, B.A., Marlou, W.B., Boer, H., Martin, W.A. and Tamminga, S. (2005). An *in-vitro* batch culture method to assess potential fermentability of feed ingredients for monogastric diets. *Animal Feed and Technology*. 123-124: 445-462.
- Wise, M.G. and Siragusa, G.R. (2007). Quantitative analysis of the intestinal bacterial community in one- to three-week-old commercially reared broiler chickens fed conventional or antibiotic-free vegetable-based diets. *Journal of Applied Microbiology*. 102: 1138-1149.
- Wong, C.M.V.L., Lau, S.Y.L., Abdullah, N. and Elaine, R.D.T. (2010). Bioconversion of palm kernel cake (pkc) to value added feed using fibrolytic bacteria and fungi. Paper presented at the National Biotechnology Seminar, PWTC, Kuala Lumpur, Malaysia. May 24-26, 2010.
- Wong, H.K. and Wan Zahari, M. (2011). Utilization of oil palm by-products as ruminant feed in Malaysia. *Journal of Oil Palm Research*. 23: 1029-1035.
- Yang, Z.B., Yang, W.R., Jiang, S.Z., Zhang, G.G., Zhang, Q.Q. and Siow, K.C. (2010). Effects of a thermotolerant multi-enzyme product on nutrient and energy utilization of broilers fed mash or crumbled corn-soybean meal diets. *Journal of Applied Poultry Research*. 19: 38-45.
- Yusof, B. and Weng, C.K. (2004). The oil palm and its sustainability. *Journal of Oil Palm Research*. 16(1): 1-10.
- Zhang, H.Q., Ye, H., Wang, W.C., Dong, Z.M., Lin, J. and Yang, L. (2013). Effect of dietary fiber on growth performance, enzyme activities, microflora and digesta physiochemical characteristics of Lionhead gees. *Journal of Agriculture and Environment*. 11(3&4): 1653-1658.
- Zhu, X.Y., Zhong, T., Pandya, Y. and Joegger, R.D. (2002). 16S rRNA-based analysis of microbiota from the ceacum of broiler chickens. *Applied and Environment Microbiology*. 68: 124-137.

Zulkifli, I., Ginson, J., Liew, P.K. and Gilbert, J. (2003). Growth performance and Newcastle disease antibody titers of broiler chickens fed palm-based diets and their response to heat stress during fasting. *Archiv für Geflügelkunde*. 67: 125-130.

