



UNIVERSITI PUTRA MALAYSIA

***INFLUENCE OF MATURITY STAGES, STORAGE TEMPERATURES AND
DURATIONS ON CHILLING INJURY, ANTIOXIDANT RESPONSES AND
QUALITY OF GINGER (*Zingiber officinale* Roscoe)***

TAN XUE YI

FP 2017 20



**INFLUENCE OF MATURITY STAGES, STORAGE TEMPERATURES AND
DURATIONS ON CHILLING INJURY, ANTIOXIDANT RESPONSES AND
QUALITY OF GINGER (*Zingiber officinale* Roscoe)**

By

TAN XUE YI

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirement for the Degree of Master of Science**

March 2017



COPYRIGHT

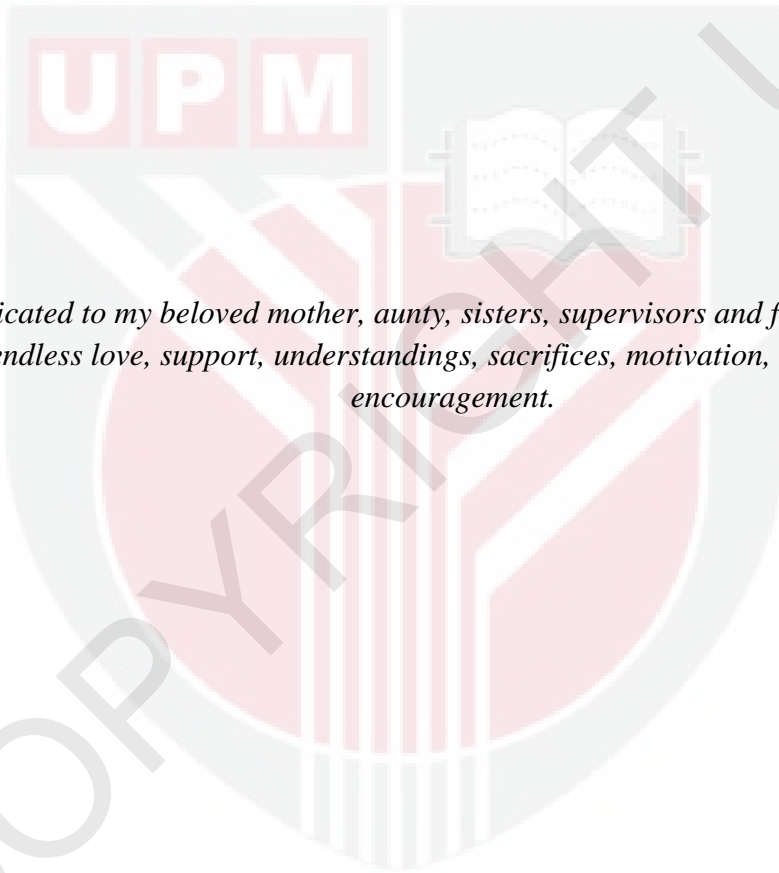
All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

Dedicated to my beloved mother, aunty, sisters, supervisors and family for their endless love, support, understandings, sacrifices, motivation, advice and encouragement.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment
of the requirement for the degree of Master of Science

**INFLUENCE OF MATURITY STAGES, STORAGE TEMPERATURES AND
DURATIONS ON CHILLING INJURY, ANTIOXIDANT RESPONSES AND
QUALITY OF GINGER (*Zingiber officinale* Roscoe)**

By

TAN XUE YI

March 2017

Chairman: Siti Hajar Ahmad, PhD
Faculty: Agriculture

Ginger (*Zingiber officinale* Roscoe) harnesses an incredible healing power for a host of ailments and used as spice throughout the world. It contains bioactive compounds such as phenolics, terpenes, flavonoids and curcumin. However, storage life and usage of ginger is limited as it is susceptible to chilling injury (CI). Storage of rhizomes below 12 °C causes CI and improper maturity stages at harvest caused reduction of ginger quality, decrease storage life and increase fiber and sprouting of ginger. Thus, the objective of the first experiment was to characterize CI of ginger as affected by maturity stages (7, 9 and 11-months after planting), storage temperatures (5, 15 and 25 °C) and durations (0, 8, 16, 24 and 32 days). Weight loss due to loss of moisture content was significantly higher in the ginger stored at 25 °C. Eleven-month ginger stored at 25 °C showed 192% and 149% higher weight loss than 7 and 9-months ginger, respectively, after 32 days of storage. CI indices as measured by pulp translucency, skin peeling and browning on ginger were severe at 5 °C, less at 15 °C while no symptom occurred after 32 days storage at 25 °C. Browning at 5 °C storage caused ginger pulp colour changed from yellow to light brown with reduction in pulp firmness. However, the pulp firmness for ginger stored at 15 °C still maintained after 32 days of storage. There was similar soluble solids concentration peak at storage day 24 and ascorbic acid contents dropped as storage durations were extended for all treatments. Thus, based on the results of the first experiment, 7 and 9-months ginger were selected for the second experiment.

The objective of the second experiment was to determine phytochemicals and chemical marker contents, antioxidant and browning enzyme activities of ginger under different maturity stages, storage temperatures and durations. Seven and nine months ginger rhizomes were stored at 5, 15 and 25 °C for 24 days. The total phenolic contents (TPC), total flavonoid contents (TFC) and DPPH radical scavenging activity for 9-

months ginger was highest at 29.71 mg GAE/g FW, 44.94 mg QE/g FW and 74.63%, respectively, under 15 °C storage at day 24. There were also significant interaction effects between storage temperatures x storage durations x maturity stages on 6-gingerol and 6-shogaol contents. Results showed a similar trend for all treatments whereby 6-gingerol and 6-shogaol increased sharply until day 16 followed by rapid reduction. Nine-month ginger at 15 °C storage showed highest 6-gingerol and 6-shogaol contents, with 50.93 mg/g and 3.11 mg/g, respectively, after 24 days of storage. Also, the results indicated polyphenol oxidase and phenylalanine ammonia lyase enzyme activities were higher in 7-months ginger stored at 5 °C and the activities increased with increasing storage durations compared to 9-months ginger. Thus, the 9-months ginger under 15 °C storage were selected as optimum maturity stage and storage temperature since these ginger exhibited minimum CI with high phytochemical and chemical marker contents, and antioxidant activity after 24 days storage.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PENGARUH PERINGKAT KEMATANGAN, SUHU DAN TEMPOH
SIMPANAN TERHADAP KECEDEeraan DINGIN, TINDAKBALAS
ANTIOKSIDAN DAN KUALITI HALIA (*Zingiber officinale* Roscoe)**

Oleh

TAN XUE YI

Mac 2017

Pengerusi: Siti Hajar Ahmad, PhD
Fakulti: Pertanian

Halia (*Zingiber officinale* Roscoe) telah dikenal pasti sebagai salah satu herba yang berpotensi yang telah digunakan sebagai rempah dan ubat-ubatan tradisional di seluruh dunia. Halia mengandungi sebatian bioaktif dan antioksidan seperti fenolik, terpena, flavonoid dan kukumin. Tahap penyimpanan dan penggunaan halia adalah terhad kerana ia adalah mudah terdedah kepada kecederaan dingin. Penyimpanan rizom di bawah 12 °C menyebabkan kecederaan dingin seperti lutcahaya pada isi, pengelupasan kulit rizom, tisu lembut dan pemerangan pulpa dan peringkat kematangan yang tidak sesuai pada musim menuai menyebabkan pengurangan kualiti rizom, tempoh penyimpanan dan meningkatkan serat dan percambahan rizom. Oleh itu, objektif eksperimen yang pertama adalah untuk mencirikan kecederaan dingin rizom halia yang dipengaruhi oleh peringkat kematangan (7, 9 dan 11 bulan selepas penanaman), suhu penyimpanan (5, 15 dan 25 °C) dan tempoh penyimpanan (0, 8, 16, 24 dan 32 hari). Penurunan berat rizom halia akibat kehilangan kandungan lembapan adalah jauh lebih tinggi dalam rizom yang disimpan pada suhu 25 °C. Halia yang berumur 11 bulan disimpan pada 25 °C menunjukkan penurunan berat rizom sebanyak 192% dan 149% lebih tinggi daripada halia 7 dan 9 bulan, selepas 32 hari penyimpanan. Indeks kecederaan dingin seperti lutcahaya pada isi, pengelupasan kulit dan pemerangan pada rizom adalah lebih teruk pada suhu 5 °C, kurang pada 15 °C manakala tiada gejala berlaku selepas 32 hari penyimpanan pada suhu 25 °C. Indeks pemerangan pada suhu penyimpanan 5 °C meningkat apabila jangka masa penyimpanan meningkat dan warna isi rizom berubah dari kuning ke perang muda dengan pengurangan kekerasan rizom. Pemanjangan jangka masa penyimpanan menyebabkan kelembutan dalam isi rizom. Manakala, penyimpanan halia pada 15 °C, mengekalkan kekerasan isi halia selepas 32 hari penyimpanan untuk semua peringkat kematangan halia. Kepekatan pepejal terlarut didapati sama dalam penyimpanan hari 24 dan kandungan asid askorbik menurun apabila jangka masa penyimpanan telah dipanjangkan untuk semua rawatan.

Oleh itu, berdasarkan keputusan eksperimen pertama, 7 dan 9 bulan halia telah dipilih untuk eksperimen kedua.

Objektif eksperimen kedua adalah untuk menentukan peningkatan fitokimia dan kandungan penanda kimia, aktiviti antioksidan dan enzim pemerangan dalam rizom halia. Halia yang berumur 7 dan 9 bulan disimpan pada suhu 5, 15 dan 25 °C selama 24 hari. Jumlah kandungan fenolik (TPC), flavonoid (TFC) dan aktiviti pemerangkapan radikal DPPH untuk halia 9 bulan adalah paling tinggi pada 29.71 mg GAE/g FW, 44.94 mg QE/g FW dan 74.63% masing-masing, di bawah suhu penyimpanan 15 °C pada hari ke-24. Terdapat juga kesan interaksi yang signifikan antara suhu penyimpanan x jangka masa penyimpanan x peringkat kematangan halia untuk kandungan 6-gingerol dan 6-shogaol dalam halia. Keputusan menunjukkan trend yang sama untuk semua suhu penyimpanan dan tahap kematangan halia di mana 6-gingerol dan 6-shogaol meningkat dengan ketara sehingga penyimpanan ke-16 hari diikuti dengan pengurangan yang ketara. Halia berumur 9 bulan yang disimpan pada suhu 15 °C menunjukkan kepekatan 6-gingerol dan 6-shogaol tertinggi, dengan 50.93 mg/g dan 3.11 mg/g, masing-masing, pada hari ke-24. Keputusan juga menunjukkan aktiviti enzim polifenol oksidase (PPO) dan fenilalanina amonia liase (PAL) adalah lebih tinggi pada halia berumur 7 bulan pada suhu penyimpanan 5 °C dan aktiviti meningkat dengan peningkatan jangka masa penyimpanan berbanding halia pada 9 bulan. Oleh itu, halia 9 bulan di bawah suhu penyimpanan 15 °C telah dipilih sebagai peringkat kematangan yang optimum dan suhu penyimpanan yang optimum kerana halia menunjukkan kecederaan dingin yang minimum dengan kandungan fitokimia dan kandungan penanda kimia, dan aktiviti antioksida yang tinggi selepas penyimpanan 24 hari.

ACKNOWLEDGEMENTS

First and foremost I would like to take this opportunity to express my utmost appreciation to my dedicated supervisor, Associate Professor Dr. Siti Hajar Ahmad. Thank you for giving me the opportunity to gain invaluable experiences in conducting my Master study. Also, thank you for the productive discussions, valuable teaching and shared a lot of her expertise, research insight and ideas for my research. I am truly grateful for your patience, encouragement, enthusiasm, advice, support and effort in guiding me to accomplish my Master study. I have been extremely lucky to have supervisor who cared so much about my research.

In addition, I owed my indebtedness and gratitude to Professor Nazamid Saari and Dr. Mohd Firdaus Ismail for their encouragement, insightful comments and give suggestions and feedbacks in improving my research. This thesis could not have been done without their strong supervision.

I would like to dedicate my heartfelt thanks to my beloved and wonderful family members, who deserve special attention for their unconditional loves, supports and encouragement that have kept me going on until the end. They have been there from the start until the finishing end of my Master study.

I would like to thank UPM for providing Graduate Research Fellowship (GRF) and research facilities to conduct my Master study. I consider it an honour to work with all the administrative and technical staffs of the Faculty Agriculture especially Mr Azhar Othman. I would also like to say thank you very much to Mrs. Nur Indah Shukor, Mrs. Nor Elliza Tajidin, Mrs. Bunga Raya Ketaren, Mrs. Munirah Mohamad, Miss Esther Yap, Miss Fatin Nabilah and all the juniors who helped me during my Master study duration. My master study would not success without the support of all these people.

I certify that a Thesis Examination Committee has met on 24 March 2017 to conduct the final examination of Tan Xue Yi on her thesis entitled "Influence of Maturity Stages, Storage Temperatures and Durations on Chilling Injury, Antioxidant Responses and Quality of Ginger (*Zingiber officinale* Roscoe)" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Mohamad bin Osman, PhD

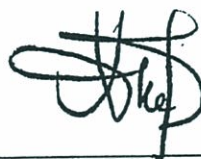
Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Mahmud bin Tengku Muda Mohamed, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Zora Singh, PhD

Professor
Curtin University
Australia
(External Examiner)



NOR AINI AB. SHUKOR, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 2 June 2017

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree Master of Science. The members of Supervisory Committee were as follows:

Siti Hajar Binti Ahmad, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Nazamid Saari, PhD

Professor
Faculty of Food Science
Universiti Putra Malaysia
(Member)

Mohd. Firdaus Ismail, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____

Date: _____

Name and Matric No.: Tan Xue Yi, GS42352

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of the thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of Chairman of

Supervisory Committee: Siti Hajar Binti Ahmad, PhD

Signature: _____

Name of Member of

Supervisory Committee: Nazamid Saari, PhD

Signature: _____

Name of Member of

Supervisory Committee: Mohd.Firdaus Ismail, PhD

TABLE OF CONTENTS

ABSTRACT	Page i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiv
LIST OF APPENDICES	xvii
LIST OF ABBREVIATIONS	xviii
 CHAPTER	
1 GENERAL INTRODUCTION	1
2 LITERATURE REVIEW	
2.1 Ginger	3
2.1.1 Botany and morphology	5
2.2 Maturity stages	6
2.3 Postharvest storage	6
2.4 Chilling injury symptoms and mechanism	7
2.5 Postharvest physical and physiological quality characteristics	8
2.6 Phytochemical contents and antioxidant activity	12
2.7 Chemical markers (gingerol and shogaol)	14
3 POSTHARVEST QUALITY OF CHILLING INJURED GINGER RHIZOMES (<i>Zingiber officinale</i> ROSCOE CV. BENTONG) AS AFFECTED BY MATURITY STAGES, STORAGE TEMPERATURES AND DURATIONS	
3.1 Introduction	16
3.2 Materials and Methods	
3.2.1 Plant materials and sample preparation	17
3.2.2 Physical quality characteristics measurement	
3.2.2.1 Weight loss	18
3.2.2.2 Pulp firmness	18
3.2.2.3 Chilling injury indices	18
3.2.2.4 Degree of browning	18
3.2.2.5 Colour evaluation	19
3.2.2.6 Titratable acidity	19
3.2.2.7 Soluble solids concentration	20
3.2.2.8 Ascorbic acids determination	20
3.2.3 Experimental design and analysis	21
3.3 Results and discussion	
3.3.1 Weight loss	21
3.3.2 Pulp firmness	26

3.3.3	Chilling injury indices	29
3.3.4	Degree of browning	36
3.3.5	Colour evaluation	40
3.3.6	Titrateable acidity	48
3.3.7	Soluble solids concentration	50
3.3.8	Ascorbic acids determination	52
3.4	Conclusions	55
4	PHYTOCHEMICALS, CHEMICAL MARKERS AND BROWNING ENZYME ACTIVITIES OF GINGER (<i>Zingiber officinale</i> ROSCOE) UNDER DIFFERING MATURITY STAGES, STORAGE TEMPERATURES AND DURATIONS	
4.1	Introduction	57
4.2	Materials and methods	
4.2.1	Plant materials and sample preparation	59
4.2.2	Chemical quality characteristics measurement	
4.2.2.1	Total phenolic content	59
4.2.2.2	Total flavonoid content	60
4.2.2.3	2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging activity	60
4.2.2.4	High performance liquid chromatography (HPLC) analysis	60
4.2.2.5	Polyphenol oxidase (PPO) enzyme activity	61
4.2.2.6	Phenylalanine ammonia lyase (PAL) enzyme activity	61
4.2.3	Experimental design and analysis	62
4.3	Results and Discussion	
4.3.1	Total phenolic content	63
4.3.2	Total flavonoid content	66
4.3.3	2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging activity	68
4.3.4	Quantification of chemical markers (6-gingerols and 6-shogaols)	71
4.3.5	Polyphenol oxidase (PPO) enzyme activity	77
4.3.6	Phenylalanine ammonia lyase (PAL) enzyme activity	80
4.4	Conclusions	83
5	SUMMARY, CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE RESEARCH	84
	REFERENCES	86
	APPENDICES	114
	BIODATA OF STUDENT	119
	LIST OF PUBLICATIONS	120

LIST OF TABLES

Table	Page
2.1 Time series data on hectareage and production of ginger by states in Malaysia from 2011-2013.	4
3.1 Main and interaction effects of maturity stages (7, 9 and 11-months after planting), storage temperatures (5, 15 and 25 °C) and storage durations (0, 8, 16, 24 and 32 days) on weight loss and pulp firmness of ginger rhizomes.	22
3.2 Correlation coefficients (r) between weight loss, pulp firmness, chilling injury indices (pulp translucency (PT), skin peeling (SP)), degree of browning (DOB) and colour values (lightness = L*, chromaticity = C* and hue = h°), titratable acidity (TA), soluble solids concentrations (SSC) and ascorbic acids (AA) of ginger rhizomes in three maturity stages (7, 9 and 11-months after planting), three storage temperature (5, 15 and 25 °C) and at five storage durations (0, 8, 16, 24 and 32 days).	30
3.3 Main and interaction effects of maturity stages (7, 9 and 11-months after planting), storage temperatures (5, 15 and 25 °C) and storage durations (0, 8, 16, 24 and 32 days) on chilling injury indices (pulp translucency, skin peeling) and degree of browning (DOB) of ginger rhizomes.	32
3.4 Main and interaction effects of maturity stages (7, 9 and 11-months after planting), storage temperatures (5, 15 and 25 °C) and storage durations (0, 8, 16, 24 and 32 days) on colour values (lightness = L*, chromaticity = C* and hue = h°) of ginger rhizomes.	41
3.5 Main and interaction effects of maturity stages (7, 9 and 11-months after planting), storage temperatures (5, 15 and 25 °C) and storage durations (0, 8, 16, 24 and 32 days) on titratable acidity (TA), soluble solids concentrations (SSC) and ascorbic acids of ginger rhizomes.	49
4.1 Gradient elution program used in separation of 6-gingerol and 6-shogaol.	61
4.2 Main and interaction effects of maturity stages (7 and 9-months after planting), storage temperatures (5, 15 and 25 °C) and storage durations (0, 8, 16 and 24 days) on total phenolic contents (TPC), total flavonoids contents (TFC), 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity, and 6-gingerols contents and 6-shogaols contents of ginger rhizomes.	63

- 4.3 Correlation coefficients (r) between total phenolic contents (TPC), total flavonoid contents (TFC), DPPH free radical scavenging activity, 6-gingerols contents, 6-shogaols contents, polyphenol oxidase (PPO) activity and phenylalanine ammonia-lyase (PAL) activity of ginger rhizomes. 77
- 4.4 Main and interaction effects of maturity stages (7, 9 and 11-months after planting), storage temperatures (5, 15 and 25 °C) and storage durations (0, 8, 16, 24 and 32 days) on polyphenol oxidase (PPO) and phenylalanine ammonia lyase (PAL) enzyme activities of ginger rhizomes. 78



LIST OF FIGURES

Figure		Page
2.1	World production of ginger from 2010-2014 (FAO, 2015).	4
2.2	Top 5 countries and Malaysia in ginger production on 2014 (FAO, 2015).	4
2.3	Production of ginger in Malaysia from 2010-2014 (FAO, 2015).	4
2.4	Summary of hectareage of main spices in Malaysia (Anonymous, 2014).	4
2.5	Ginger plant (FAO, 2002).	5
2.6	Ginger rhizome (Mishra et al., 2012)	5
2.7	<i>Zingiber officinale</i> Roscoe cv. Bentong (A) and <i>Zingiber officinale</i> var. Rubrum (B) grown in Malaysia (Mansor et al., 2005).	6
2.8	Chemical constituents of ginger (Rahmani et al., 2014).	15
3.1	Relationships between storage durations (0, 8, 16, 24 and 32 days) and percentage of weight loss of 7 (A), 9 (B) and 11-months (C) ginger rhizomes stored at different storage temperatures (5, 15 and 25 °C). Solid line indicates a significant regression trend at $P = 0.05$, ($n = 12$).	23
3.2	Relationships between storage durations (0, 8, 16, 24 and 32 days) and pulp firmness of 7 (A), 9 (B) and 11-months (C) ginger rhizomes stored at different storage temperatures (5, 15 and 25 °C). The solid line indicates a significant regression trend at $P = 0.05$, ($n = 12$).	27
3.3	Pulp translucency of 7 (A), 9 (B) and 11-months (C) ginger stored at 5, 15 and 25 °C during 0, 8, 16, 24 and 32 days of storage. Vertical line indicates pooled LSD at $P = 0.05$, ($n = 12$).	33
3.4	Skin peeling of 7 (A), 9 (B) and 11-months (C) ginger stored at 5, 15 and 25 °C during 0, 8, 16, 24 and 32 days of storage. Vertical line indicates pooled LSD at $P = 0.05$, ($n = 12$).	34
3.5	Relationships between storage durations (0, 8, 16, 24 and 32 days) and degree of browning of 7 (A), 9 (B) and 11-months (C) ginger rhizome stored at different storage temperatures (5, 15 and 25 °C). The solid line indicates a significant regression trend at $P = 0.05$, ($n = 12$).	38

3.6	Relationships between storage durations and lightness of ginger rhizomes harvested at different maturity stages (7, 9 and 11-months). The solid line indicates a significant regression trend indicates no significant difference at $P = 0.05$, ($n = 12$).	42
3.7	Relationships between storage durations and lightness of ginger rhizomes stored at different storage temperatures (5, 15 and 25 °C). The solid line indicates a significant regression trend indicates no significant difference at $P = 0.05$, ($n = 12$).	42
3.8	Relationships between storage durations (0, 8, 16, 24 and 32 days) and hue of 7 (A), 9 (B) and 11 (C) months ginger rhizome stored at different storage temperatures (5, 15 and 25 °C). Solid line indicates a significant regression trend at $P = 0.05$, ($n = 12$).	44
3.9	Relationships between storage durations (0, 8, 16, 24 and 32 days) and chromaticity of 7 (A), 9 (B) and 11 (C) months ginger rhizomes stored at different storage temperatures (5, 15 and 25 °C). The solid line indicates a significant regression trend indicates no significant difference at $P = 0.05$, ($n = 12$).	46
3.10	Relationships between storage durations (0, 8, 16, 24 and 32 days) and soluble solids concentrations of 7 (A), 9 (B) and 11 (C) months ginger rhizomes stored at different storage temperatures (5, 15 and 25 °C). The solid line indicates a significant regression trend at $P = 0.05$, ($n = 12$).	51
3.11	Relationships between storage durations (0, 8, 16, 24 and 32 days) and ascorbic acids of 7 (A), 9 (B) and 11 (C) months ginger rhizome stored at different storage temperatures (5, 15 and 25 °C). The solid line indicates a significant regression trend at $P = 0.05$, ($n = 12$).	53
4.1	The relationships between storage durations (0, 8, 16 and 24 days) and total phenolic contents (TPC) of 7 (A) and 9-months (B) ginger rhizomes stored at different storage temperatures (5, 15 and 25 °C). The solid line indicates a significant regression trend at $P = 0.05$, ($n = 12$).	64
4.2	The relationships between storage durations (0, 8, 16 and 24 days) and total flavonoids contents (TFC) of 7 (A) and 9-months (B) ginger rhizomes stored at different storage temperatures (5, 15 and 25 °C). The solid line indicates a significant regression trend and the dotted line indicates no significant difference at $P = 0.05$, ($n = 12$).	67
4.3	The relationships between storage durations and DPPH free radical scavenging activity of ginger rhizomes harvested at different maturity stages (7 and 9-months). The solid line indicates a significant regression trend at $P = 0.05$, ($n = 12$).	69

4.4	The relationships between storage durations and DPPH free radical scavenging activity of ginger rhizomes of ginger rhizomes stored at different storage temperatures (5, 15 and 25 °C). The solid line indicates a significant regression trend at $P = 0.05$, ($n = 12$).	69
4.5	High Performance Liquid Chromatography (HPLC) chromatogram of (A) different maturity ginger rhizome (7 and 9-months) and (B) 9-months ginger stored at different storage temperatures (5, 15 and 25 °C) during day 16 of storage.	72
4.6	The relationships between storage durations (0, 8, 16 and 24 days) and 6-gingerols contents of 7 (A) and 9-months (B) ginger rhizomes stored at different storage temperatures (5, 15 and 25 °C). The solid line indicates a significant regression trend at $P = 0.05$, ($n = 12$).	73
4.7	The relationships between storage durations (0, 8, 16 and 24 days) and 6-shogaols contents of 7 (A) and 9-months (B) ginger rhizomes stored at different storage temperatures (5, 15 and 25 °C). The solid line indicates a significant regression trend at $P = 0.05$, ($n = 12$).	75
4.8	The relationships between storage durations and polyphenol oxidase (PPO) activity of ginger rhizomes harvested at different maturity stages (7 and 9-months). The solid line indicates a significant regression trend at $P = 0.05$, ($n = 12$).	79
4.9	The relationships between storage durations and polyphenol oxidase (PPO) activity of ginger rhizomes stored at different storage temperatures (5, 15 and 25 °C). The solid line indicates a significant regression trend, and the dotted line indicates no significant regression trend at $P = 0.05$, ($n = 12$).	79
4.10	The relationships between storage durations (0, 8, 16 and 24 days) and phenylalanine ammonia lyase activity (PAL) of 7 (A) and 9-months (B) ginger rhizomes stored at different storage temperatures (5, 15 and 25 °C). The solid line indicates a significant regression trend at $P = 0.05$, ($n = 12$).	81

LIST OF APPENDICES

Appendix		Page
1	Analyses of variance of 7, 9 and 11 months ginger rhizome stored at 5, 15 and 25 °C at different storage durations (0, 8, 16, 24 and 32 days) on weight loss and pulp firmness.	114
2	Analyses of variance of 7, 9 and 11 months ginger rhizome stored at 5, 15 and 25 °C at different storage durations (0, 8, 16, 24 and 32 days) on CI indices (pulp translucency and skin peeling) and degree of browning (DOB).	114
3	Analyses of variance of 7, 9 and 11-months ginger rhizome stored at 5, 15 and 25 °C at different storage durations (0, 8, 16, 24 and 32 days) on colour evaluation (L^* = lightness, C^* = chromaticity and h° = hue angle).	115
4	Analyses of variance of 7, 9 and 11-months ginger rhizome stored at 5, 15 and 25 °C at different storage durations (0, 8, 16, 24 and 32 days) on titratable acidity (TA), soluble solids concentration (SSC) and ascorbic acids.	115
5	Analyses of variance of 7 and 9-months ginger rhizome stored at 5, 15 and 25 °C at different storage durations (0, 8, 16 and 24 days) on total phenolic content (TPC), total flavonoid content (TFC) and DPPH free radical scavenging activity.	116
6	Analyses of variance of 7 and 9-months ginger rhizome stored at 5, 15 and 25 °C at different storage durations (0, 8, 16 and 24 days) on 6-gingerols and 6-shogaols.	116
7	Analyses of variance of 7 and 9-months ginger rhizome stored at 5, 15 and 25 °C at different storage durations (0, 8, 16 and 24 days) on polyphenol oxidase (PPO) and phenylalanine ammonia lyase (PAL) enzymes activities.	117
8	Symptoms on ginger at 5, 15 and 25 °C storage.	118

LIST OF ABBREVIATIONS

°C	Degree Celsius
g	Gram
cm	Centimeter
rpm	Revolutions per minute
min	Minute
%	Percentage
MS	Maturity stage
ST	Storage temperature
SD	Storage duration
kg	Kilogram
mm	Millimetre
mm/min	Millimetre per minute
N	Newton
CI	Chilling injury
mL	Millilitre
uL	Microlitre
DOB	Degree of browning
L*	Lightness
C*	Chromaticity
h°	Hue angle
NaOH	Sodium hydroxide
TA	Titrateable acidity
SSC	Soluble solids concentration
HPO ₃	Metaphosphoric acid
ANOVA	Analysis of variance

LSD	Least significant differences
RCBD	Randomized complete block design
HSD	Honestly significant differences
FW	Fresh weight
FAO	Food and Agriculture Organization
ug/cm ² /hr/kPa	Microgram per centimetre square per hour per kilopascal
PPO	Polyphenol oxidase
PAL	Phenylalanine ammonia lyase
TPC	Total phenolic contents
TFC	Total flavonoid contents
DPPH	2,2-diphenyl-1-picrylhydrazyl
nm	Nanometre
GAE	Gallic acid equivalent
w/w	Weight over weight
FC	Folin-Ciocalteu
Na ₂ CO ₃	Sodium carbonate
NaNO ₃	Sodium nitrate
AlCl ₃	Aluminium chloride
QE	Quecertin equivalent
HPLC	High performance liquid chromatography
ppm	Part per million
FW/min	Fresh weight per minute
PVP-30	Polyvinylpyrrolidone
FW/h	Fresh weight per hour
RH	Relative humidity
DOA	Department of Agriculture
mg/g FW	Milligram per gram fresh weight

MOA	Ministry of Agriculture
N	Nitrogen
P ₂ O ₅	Phosphorus pentoxide
K ₂ O	Potassium oxide
TE	Trace element
H ₃ PW ₁₂ O ₄₀	Phosphotungstic
H ₃ PMo ₁₂ O ₄₀	Phosphomolybdic
W ₈ O ₂₃	Tungstene
Mo ₈ O ₂₃	Molybdene
MARDI	Malaysian Agricultural Research and Development Institute
AIP	2-aminoindan-2-phosphonic acid
AOA	α-aminoxyacetic acid
AOPP	α-aminoxy-β-phenylpropionic acid

CHAPTER 1

GENERAL INTRODUCTION

Ginger (*Zingiber officinale* Roscoe) is one of the potential herbs under the 10th Malaysia Plan. It is one of the most profitable crops cultivated in Asia and India, which accounted for 40% of the world's ginger production. Malaysia is the 13th producer of ginger in the world, producing 10,775 tonnes of ginger in 2014. In Malaysia, ginger is mostly planted in Pahang, Sabah, Sarawak, Selangor and Johor. Pahang produced the largest amount of ginger (7,281MT) followed by Sarawak (2,599 MT) in 2014 (Anonymous, 2014). There are two main types of ginger, white and red. The 'Bentong' ginger, a white ginger with large rhizomes, juicy flesh, low fibre content, strong pungency, pale white pulp and light brown peel, is widely cultivated; whereas, the 'Bara' ginger, a red ginger with small rhizomes, yellow to the pinkish cross-section, small size and more pungency compared to other ginger (Mansor et al., 2005; DOA, 2015). A previous study reported that Bentong ginger has the highest gingerol contents (0.808 mg/g), which is an important constituent in ginger, compared to 'China', 'Tanjung Sepat' and 'Bara' gingers (DOA, 2015).

Ginger rhizomes are highly demanded in the local and international markets due to their excellent uses for medicinal purposes, food, health care and beauty. It is also used as a spice and flavouring agent for cooking due to its unique aroma. Ginger harnesses unusual and extraordinary healing power proven for a host of ailments such as arthritis, migraine, morning sickness, anticancer, anti-inflammatory, digestion and boosts immune system, headache and promotes healthy heart and cholesterol levels. Ginger contains high bioactive and antioxidant compounds such as terpenes, flavonoids and phenolics (Grzanna et al., 2005; Ghasemzadeh et al., 2010). The phenolic components which are 6-shogaol, 6-gingerol, 8-gingerol, 8-gingerol, 10-gingerol and curcumin have been identified as major antioxidant components in ginger (Yeh et al., 2014). Gingerol, a phenolic compound acts as a secondary metabolite and responsible for pungency and flavour in ginger. Gingerol has been reported to be an antioxidant, and possessed analgesic, antipyretic, chemopreventive and anti-inflammatory properties (Weiss, 2002). Shogaol, a phenolic compound with a pungent characteristic, is the degradation products of gingerol (Chen et al., 1986). Gingerol degradation to the shogaol occurs under high temperature and acidic conditions. Shogaol has been reported to possess anticancer, antioxidants, regulatory roles in cellular defense mechanism and impact the pungent characteristic to dried ginger (Afzal et al., 2001).

However, ginger is susceptible to chilling injury (CI). Maturity stages, varieties, environment and storage temperatures, and durations influence the occurrence of CI in the rhizome. Storage at ambient temperature leads to high moisture loss, shriveling and sprouting of rhizome during storage below 12 °C, causing CI on the rhizome. The CI caused pitting and sunken lesions on the ginger surface, browning, shriveling, softening, and postharvest decay. The shelf life is shortened and rhizomes degradation reduced the eating quality by the loss of flavour, aroma and pungency. Also, delaying

harvest caused reduction of rhizome quality, decrease storage life and increase fibre and sprouting on rhizomes.

The high demand for ginger locally (18,600 MT) requires its importation from countries like China, India and Thailand. However, imported ginger is usually damaged and low quality due to the improper handling temperatures during transportation and storage in the wholesale market. Ginger rhizomes stored at temperatures below 5 °C during transportation and stockpiling at the wholesale market resulted in CI (browning, translucency, tissue softening), loss of flavour and odour, reduction in phytochemicals and reduction of shelf life. However, information on the maturity stages, and storage temperatures and durations on the postharvest quality and phytochemical contents of ginger are lacking. Therefore, the objectives of this study were to determine the main and interaction effects of maturity stages, and storage temperatures and durations on ginger rhizomes (i) postharvest quality and CI, and (ii) phytochemicals and chemical markers contents, antioxidant activity as well as the browning enzymes.

REFERENCES

- Abbasi, N. A., Zahoor, M., Khan, H. A. and Qureshi, A. A. (2012). Effect of Encapsulated Calcium Carbide Application at Different Growth Stages on Potato (*Solanum tuberosum* L.) Growth, Yield and Tuber Quality. *Pakistan Journal of Botany*, 35, 601-610.
- Abbott, J. A. (2004). Textural Quality Assessment for Fresh Fruits and Vegetables. In *Quality of Fresh and Processed Foods* (pp. 265-279). US: Springer.
- Aeschbach, R., Loliger, J., Scott, B. C., Murcis, A., Buler, J., Halliwell, B. and Agnoma, O. I. (1994). Antioxidant Actions of Thymol, Carbacrol, 6-gingerol, Zingerone and Hydroxytyrosol. *Food and Chemical Toxicology*, 32, 31–36.
- Afoakwa E. O. and Sefa-Dedeh, S. (2001). Chemical Composition and Quality Changes Occurring in *Dioscorea dumetorum* Pax Tubers after Harvest. *Food Chemistry*, 75, 85-91.
- Afzal, M., Al-Hadidi, D., Menon, M., Pesek, J. and Dhimi, M. S. (2001). Ginger: An Ethnomedical, Chemical and Pharmacological Review. *Drug Metabolism and Drug Interaction*, 18, 159-90.
- Agati, G., Azzarello, E., Pollastri, S. and Tattini, M. (2012). Flavonoids as Antioxidants in Plants: Location and Functional Significance. *Plant Science*, 196, 67-76.
- Ajibola, V. O., Babatunde, O. A. and Suleiman, S. (2009). The Effect of Storage Method on the Vitamin C Content in some Tropical Fruit Juices. *Trends in Applied Science Research Journal*, 4, 79-84.
- Akamine, E. K. (1962). Storage of Fresh Ginger Rhizomes. Bulletin 130 Hawaii Agricultural Experimental Station, University of Hawaii, U. S. A.
- Akowi, G. A., Ismail, Z., Norhayati, I. and Sadikun, A. (2005). The Effects of Different Extraction Solvents of Varying Polarities of Polyphenols of *Orthosiphon stamineus* and Evaluation of the Free Radical-scavenging Activity. *Food Chemistry*, 93, 311-317.
- Alam, P. (2013). Densitometric HPTLC Analysis of 8-gingerol in *Zingiber officinale* Extract and Ginger-containing Dietary Supplements, Teas And Commercial Creams. *Asian Pacific Journal of Tropical Biomedicine*, 3(8), 634–638.
- Ali, B. H., Blunden, G, Tanira, M. O and Nemmar, A. (2008). Some phytochemical, Pharmacological and Toxicological Properties of Ginger (*Zingiber Officinale* Roscoe): A Review of Recent Research. *Food and Chemical Toxicology*, 46(2), 409–420.

- Allakhverdiev, S. I., Kreslavski, V. D., Klimov, V. V., Los, D. A., Carpentier, R. and Mohanty, P. (2008). Heat Stress: An Overview of Molecular Responses in Photosynthesis. *Photosynthesis Research*, 98, 541–550.
- Anonymous. (2002a). A Concise Guide to Spices, Herbs, Seeds, and Extractives. American Spice Trade Association.
- Anonymous. (2002b). Compendium of Medicinal Plants Used in Malaysia (Vol. 2). Kuala Lumpur: HMRC-IMR.
- Anonymous. (2004). Ginger – Postharvest Care and Market Preparation. Postharvest Handling Technical Bulletin 23, National Agricultural Research Institute, U.S.A.
- Anonymous. (2014). Herbs and Spices Statistics Malaysia 2014. Department of Agriculture, Malaysia.
- AOAC. (1990). Official Methods of Analysis, 15th ed. Association of Official Analytical Chemists, Washington, D. C.
- Araji, S., Grammer, T. A., Stephen, R., Anderson, D., Mikulic-Petkovsek, M., Veberic, R., Phu, M. L., Solar, A., Leslie, C. A., Dandekar, A. M. and Escobar, M. A. (2014). Novel Roles for the Polyphenol Oxidase Enzyme in Secondary Metabolism and the Regulation of Cell Death in Walnut *Plant Physiology*, 164(3), 1191-1203.
- Aroca, R., Irigoyen, J. and Sanchez-Diaz, M. (2003). Drought Enhances Maize Chilling Tolerance. II. Photosynthetic Traits and Protective Mechanisms against Oxidative Stress. *Plant Physiology*, 117, 540–549.
- Arts, I. C., Jacobs, D. R. and Gross, M. (2002). Dietary Catechins and Cancer Incidence among Postmenopausal Women: The Iowa Women's Health Study (United States). *Cancer Causes Control*, 13, 373-82.
- Asada, K. (2006). Production and Scavenging of Reactive Oxygen Species in Chloroplasts and Their Functions. *Plant Physiology*, 141, 391–396.
- Atanda, S.A. Pessu P. O., Agoda S., Isong I. U. and Ikotun. (2011). The Concepts and Problems of Post-harvest Food Losses in Perishable Crops. *African Journal of Food Science*, 5(11), 603-6013.
- Ayala-Zavala, J. F., Wang, S. Y., Wang, C. Y. and Gonzalez-Aguilar, G. A. (2004). Effect of Storage Temperatures on Antioxidant Capacity and Aroma Compounds in Strawberry Fruit. *LWT Food Science and Technology*, 37, 687–695.
- Baker, E. A, Bukovac, M. J. and Hunt, G. M. (1982). Composition of Tomato Fruit Cuticle as Related to Fruit Growth and Development. In: Cutler DF, Alvin KL, Price CE, eds. The plant cuticle. London: Academic Press: 33–44.

- Baliga, M. S., Haniadka, R., Pereira, M. M., D'Souza, J.J., Pallaty, P. L. (2011). Update on the Chemopreventive Effects of Ginger and its Phytochemicals. *Critical Reviews in Food Science Nutrition*, 51, 499-523.
- Bareja, B. G. (2013). About Water Loss in Plants: Transpiration and Guttation. Retrieved on 15 August 2016 from <http://www.cropsreview.com/transpiration.html>.
- Bargel, H. and Neinhuis, C. (2005). Tomato (*Lycopersicon esculentum* Mill.) Fruit Growth and Ripening as Related to the Biomechanical Properties of Fruit Skin and Isolated Cuticle. *Journal of Experimental Botany*, 56 (413): 1049-1060.
- Barrett, D. M., Beaulieu, J. C. and Shewfelt, R. (2010). Color, Flavor, Texture, and Nutritional Quality of Fresh-Cut Fruits and Vegetables: Desirable Levels, Instrumental and Sensory Measurement, and the Effects of Processing. *Critical Reviews in Food Science and Nutrition*, 50, 369-389.
- Bartosz, G. (1997). Oxidative Stress in Plants. *Acta Physiologiae Plantarum*, 19(1), 47-64.
- Bauchot, A. D., Hallett, I. C., Redgwell, R. J. and Lallu, N. (1999). Cell Wall Properties of Kiwifruit Affected by Low Temperature Breakdown. *Postharvest Biology and Technology*, 16(3), 245-255.
- Benjamin, N. and Montgomery, M.W. (1973). Polyphenol Oxidase of Royal Ann Cherries: Purification and Characterization. *Journal of Food Science*, 38, 799-806.
- Ben-Yehoshua, S. and Rodov, V. (2003). Transpiration and Water Stress. In: Bartz, J.A., Brecht, J.K. (Eds.), *Postharvest Physiology and Pathology of Vegetables*, 2nd ed. (pp. 111-159). Basel, New York: Marcel Dekker, Inc.
- Bernalte, M. J., Sabio, E., Hernandez, M. T. and Gervasini C. (2003). Influence of Storage Delay on Quality of 'Van' Sweet Cherry. *Postharvest of Biology and Technology*, 23, 303-312.
- Berner, M., Krug, D., Bihlmaier, C., Vente, A., Müller, R. and Bechthold, A. (2006). Genes and Enzymes Involved in Caffeic Acid Biosynthesis in Actinomyces *Saccharothrix espanaensis*. *Journal of Bacteriology*, 188, 2666-2673.
- Bhardwaj, R. L. and Pandey, S. (2011). Juice Blends - A way of Utilization of Under-utilized Fruits, Vegetables, and Spices: A Review. *Critical Review Food Science Nutrition*, 51, 563-570.
- Bhattacharai, S., Tran, V. H. and Duke, C. C. (2001). The Stability of Gingerol and Shogaol in Aqueous Solutions. *Journal of Pharmaceutical Science*, 90, 1658-1664.
- Bolwell, G.P., Mavandad, M., Millar, D., Edwards, K., Schuch, W. and Dixon, D. (1988). Inhibition of mRNA Levels and Activities by Trans-cinnamic Acid in Elicitor-Induced Bean Cells. *Phytochemistry*, 27, 2109-2117.

- Bors, W., Michel, C. and Saran, M. (1994). Flavonoid Antioxidants: Rate Constants for Reactions with Oxygen Radicals. *Methods Enzymology*, 234, 420-429.
- Brand-Williams, W., Cuvelier, M. E. and Berset, C. (1995). Use of a Free Radical Method to Evaluate Antioxidant Activity. *Food Science and Technology*, 28, 25-30.
- Bravo, L. (1998). Polyphenols: Chemistry, Dietary Sources, Metabolism and Nutritional. *Nutrition Review*, 56, 317-333.
- Brecht, J. K. and Nunes, M. C. N. (2010). Time-temperature Combinations that Induce Chilling Injury of Mangoes. Report, University of Florida.
- Brummell, D. A. (2006). Cell Wall Disassembly in Ripening Fruit. *Functional Plant Biology*, 33, 103-119.
- Bugaud, C., Daribo, M. O., Beate, M. P., Telle, N. and Dubois, C. (2009). Relative Importance of Location and Period of Banana Bunch Growth in Carbohydrate Content and Mineral Composition of Fruit. *Fruit*, 64(2), 63-74.
- Burdon, J. and Clark, C. (2001). Effect of Postharvest Water Loss on 'Hayward' Kiwifruit Water Status. *Postharvest Biology and Technology*, 22, 215-225.
- Busch, J. M. (1999). Enzymic Browning in Potatoes: A Simple Assay for a Polyphenol Oxidase Catalysed Reaction. *Biochemical Education*, 27, 171-173.
- Bushra, S., Farooq, A. and Muhammad, A. (2009). Effect of Extraction Solvent/Technique on the Antioxidant Activity of Selected Medicinal Plant Extracts. *Molecules*, 14, 2167-2180.
- Caillet, S., Salmieri, S. and Lacriox, M. (2006). Evaluation of Free Radical-scavenging Properties of Commercial Grape Phenol Extracts by a Fast Colorimetric Method. *Food Chemistry*, 95, 1-8.
- Cantos, E., Espín, J. C. and Tomás-Barberán, F. A. (2001). Effect of Wounding on Phenolic Enzymes in Six Minimally Processed Lettuce Cultivars Upon Storage. *Journal of Agricultural and Food Chemistry*, 49(1), 322-330.
- Cantos, E., Tudela, J. A., Gil, M. A. and Espin, J. C. (2002). Phenolic Compounds and Related Enzymes are Not Rate-limiting in Browning Development of Fresh cut Potatoes. *Journal of Agriculture and Food Chemistry*, 50, 3015-3023.
- Cantwell, M., Orozco, W., Rubatzky, V. and Hernandez, L. (1992). Postharvest Handling and Storage of Jicama Roots. *Acta Horticulture*, 318, 333-343.
- Carnelossi, M. A. G., Silva, E. O., Campos, R. S. and Puschmann, R. (2005). Physiological Responses of Minimally Processed Cabbage Leaves. *Brazilian Horticulture*, 23(2), 215-220.

- Carvalho, A. V. and Lima, L. D. O. (2002). Kiwifruit Quality of Minimally Processed and Subjected to Treatment with Ascorbic Acid, Citric Acid and Calcium Chloride. *Agricultural Research Brazilian*, 37(5), 679-685.
- Castleman, M. (2001). *The New Healing Herbs*, 2nd edition. USA: Rodale Press, Emmaus, Pa.
- Chan, Z. and Tian, S. (2006). Induction of H₂O₂-metabolizing Enzymes and Total Protein Synthesis by Antagonistic Yeast and Salicylic Acid in Harvested Sweet Cherry Fruit. *Postharvest Biology and Technology*, 39, 314-320.
- Chandrasekara, A. and Kumar, T. J. (2016). Roots and Tuber Crops as Functional Foods: A Review on Phytochemical Constituents and Their Potential Health Benefits. *International Journal of Food Science*, 2016(3), 15.
- Chen, C. C. and Paull, R. E. (2000). Sugar Metabolism and Pineapple Flesh Translucency. *Journal of American Society for Horticulture Science*, 125, 558-562.
- Chen, C. C., Kuo, M.C., Wu, C.M., and Ho, C.T. (1986). Pungent Compounds of Ginger (*Zingiber officinale* Roscoe) Extracted by Liquid Carbon Dioxide. *Journal of Agriculture Food Chemistry*, 34, 447-480.
- Chen, C. R. and Ramaswamy, H. S. (2002). Colour and Texture Change Kinetics in Ripening Bananas. *LWT- Food Science and Technology*, 35, 318-328.
- Chen, J. Y., He, L. H., Jiang, Y. M., Wang, Y., Joyce, D.c., Ji, Z. L. and Lu, W. J. (2008) Role of Phenylalanine Ammonia Lyase in Heat Pretreatment-Induced Chilling Tolerance in Banana Fruit. *Plant Physiology*, 132(2), 318-328.
- Chen, L. S., Li, P. and Cheng, L. (2008). Effects of High Temperature Coupled with High Light on the Balance between Photooxidation and Photoprotection in the Sun-exposed Peel of Apple. *Planta*, 228, 745-775.
- Chew, Y. L., Chan, E. W. L., Tan, P. L., Lim, Y. Y., Stanslas, J. and Goh, J. K. (2011). Assessment of Phytochemical Content, Polyphenolic Composition, Antioxidant and Antibacterial Activities of Leguminosae Medicinal Plants in Peninsular Malaysia. *BMC Complementary and Alternative Medicine*, 11(12), 1-10.
- Choi, E. J., Lee, K. A., Kim, B. S. and Ku, K. K. (2012). Effect of Pre-treatment and Storage Conditions on the Quality Characteristics of Ginger Paste. *Preventive Nutrition and Food Science*, 17(1), 46-52.
- Christie, P. J., Alfenito, M. R. and Walbot, V. (1994). Impact of Low Temperature Stress on General Phenylpropanoid and Anthocyanin Pathways: Enhancement of Transcript Abundance and Anthocyanin Pigmentation in Maize Seedlings. *Planta* 194, 541-549.

- Chu, Y., Chang, C. and Hsu, H. 2000. Flavonoid Content of Several Vegetables and Their Antioxidant Activity. *Journal of Science and Food Agriculture*, 80, 561-566.
- Chung, I. M., Kim, J. J., Lim, J. D., Yu, C. Y., Kim, S. H. and Hahn, S. J. (2006). Comparison of Resveratrol, SOD activity, Phenolic Compounds and Free Amino Acid in *Rehmannia glutinose* under Temperature and Water Stress. *Environmental and Experimental Botany*, 56, 44–53.
- Cisneros-Zevallos L. (2003). The Use of Controlled Postharvest Abiotic Stresses as a Tool for Enhancing the Nutraceutical Content and Adding-value of Fresh Fruits and Vegetables. *Journal of Food Science*, 68, 1560–1565.
- Concellon, A., Anon, M. C. and Chaves, A. R. (2004). Characterization and Changes in Polyphenol Oxidase from Eggplant Fruit (*Solanum melongena* L.) during Storage at Low Temperature. *Food Chemistry*, 88, 17–24.
- Conforti, F., Statti, G., Uzunov, D. and Menichini, F. (2006). Comparative Chemical Composition and Antioxidant Activities of Wild and Cultivated *Laurus nobilis* L. Leaves and *Foeniculum vulgare* subsp. *piperitum* (Ucria) coutinho seeds. *Biological and Pharmaceutical Bulletin*, 29 (10), 2056-2064.
- Connell, D.W. and Sutherland, M.D. (1969). A Re-examination of Gingerol, Shogaol, and Zingerone, the Pungent Principles of Ginger (*Zingiber officinale* Roscoe). *Australian Journal of Chemistry*, 22, 1033-1043.
- Correa, C. R., Li, L., Aldini, G., Carini, M., Chen, C. Y. O., Chun, H. K., Cho, S. M., Park, K. M., Russell, R. M., Blumberg, J. B. and Yeum, K. J. (2010). Composition and Stability of Phytochemicals in Five Varieties of Black Soybeans (*Glycine max*). *Food Chemistry*, 123, 1176-1184.
- Coesteng, M. Y. and Lee, C. Y. (1987). Changes in Apple Polyphenoloxidase and Polyphenol Concentrations in Relation to Degree of Browning. *Journal of Food Science*, 52, 985- 989.
- Crisosto, C. H., Mitchell, F. G. and Ju, Z. (1999). Susceptibility to Chilling Injury of Peach, Nectarine, and Plum Cultivars Grown in California. *Horticultural Science*, 34, 1116–1118.
- Croteau, R., Kutchan, T. M. and Lewis, N. G. (2000). Natural Products (Secondary Metabolites). In *Biochemistry and Molecular Biology of Plants* (eds. Buchanan, B., Gruissem, W. and Jones, R.L. pp. 1250-1318). USA: American Society of Plant Physiologists, Beltsville, MD.
- Cvirkova, M., Sukhova, L. S., Eder, J. and Korablova, N. P. (1994). Possible Involvement of ABA, Ethylene and Phenolic Acids in Potato Tuber Dormancy. *Plant Physiology and Biochemistry*, 32, 685-691.
- Dandago, M. A. and Gungula, D.T. (2011). Effects of Various Storage Methods on the Quality and Nutritional Composition of Sweet Potato (*Ipomera batatas* L.) in Yola Nigeria. *International Food Research Journal*, 18, 271-278.

- Davey, M. W., Van, M., Montagn, D., Inze, M., Sanmartin, A., Kanallis, A. and Smirnoff, N. (2000). Plant L-ascorbic acid: Chemistry Function Metabolism, Bioavailability and Effects of Processing. *Journal of the Science and Food Agriculture*, 80, 3825-860.
- Deepaa, M. and Wong, C. W. (2012). Characterization of Polyphenol Oxidase in Sweet Potato (*Ipomoea batatas* L.). *Journal for the Advancement of Science and Arts*, 3(1), 14-30.
- Deighton, N., Brennan, R., Finn, C. and Davies, H. V. (2000). Antioxidant Properties of Domesticated and Wild Rubus Species. *Journal of Science Food and Agriculture*, 80, 1307–1131.
- Dixon, R. A. and Paiva, N. L. (1995). Stress-induced Phenylpropanoid Metabolism. *Plant Cell*, 7, 1085-1097.
- DOA. (2015). Department of Agriculture, Perak. Retrieved on 9 January 2016 from <http://www.pertanianperak.gov.my/index.php/en/maklumat-pertanian/maklumat-tanaman/herba-landskap/187-panduan-menanam-halia>.
- Donega, M. A., Tessmer, M. A, Mooz, E. D., Dall'orto, L. T. C., Sasaki, F. C. and Kluge, R. A. (2013). Fresh Cut Yam Stored under Different Temperatures. *Brazilian Horticulture*, 31, 248-254.
- Doughari, J. H. (2012). Phytochemicals: Extraction Methods, Basic Structures and Mode of Action as Potential Chemotherapeutic Agents (pp. 1-32). Nigeria: Federal University of Technology, Yola.
- Douglas, M., Heyes, J. and Smallfield, B. (2005). Herbs, Spices and Essential Oils: Postharvest Operations in Developing Countries. FAO, New Zealand: NZ Institute for Corp and Food Research Ltd.,
- Dugasani, S., Pichika, M. R., Nadarajah, V. D., Balijepalli, M. K., Tandra, S. and Korlakunta, J. N. (2010). Comparative Antioxidant and Anti-inflammatory Effects of 6-gingerol, 8-gingerol, 10-gingerol and 6-shogaol. *Journal of Ethnopharmacol*, 127, 515–520.
- Dumas, Y., Dadomo, M., Di Lucca, G. and Grolier, P. (2003). Effects of Environmental Factors and Agricultural Techniques on Antioxidant Content of Tomatoes. *Journal of Science and Food Agriculture*, 83(5), 369–382.
- Eitenmiller, R. R., Landen, W. O., and Ye, L. (2008). *Vitamin analysis for the health and food sciences (2nd ed.)*. Boca Raton, USA: CRC Press, Taylor and Francis Group.
- Elattar, T. M. and Virji, A. S. (2000). The Inhibitory Effect of Curcumin, Genistein, Quercetin and Cisplatin on the growth of Oral Cancer Cells In Vitro. *Anticancer Research*, 20, 1733–1738.

- Elavarthi, S. and Martin, B. (2010). Spectrophotometric Assays for Antioxidant Enzymes in Plants. *Methods in Molecular Biology*, 639, 273–281.
- Emese, J. and Nagymate, P. F. (2008). The Stability of Vitamin C in Different Beverages. *British Food Journal*, 110(3), 296-309.
- Ezz, T. M., Ritenour, M. A. and Brecht, J. K. (2004). Hot Water and Elevated CO₂ Effects on Proline and Other Compositional Changes in Relation to Postharvest Chilling Injury of ‘Marsh’ Grapefruit. *Journal of the American Society for Horticultural Science*, 129(129), 576-582.
- FAO. (2002). Food and Agriculture Organization of the United Nations. Ginger Grading and Packing. Retrieved on 15 January 2016 from https://www.researchgate.net/profile/Tadesse_Teferra2/publication/279334607_Ginger_Zingiber_Officinale_Rosec_Production_Postharvest_Handling_Processing_and_Marketing_A_Comprehensive_Extension_Package_Manual/links/559256d008ae15962d8e5b1f.pdf?origin=publication_list.
- FAO. (2014). Food and Agriculture Organization of the United Nations. FAOSTAT. Retrieved on 5 January 2016 from <http://faostat3.fao.org/browse/Q/QC/E>.
- FAO. (2016). Food and Agriculture Organization of the United Nations. Factors Affecting Storability of Roots and Tubers. Retrieved 9 October 2016, from <http://www.fao.org/docrep/x5415e/x5415e02.htm>.
- Farag, S.E. D. A., Aziz N. H., and Attia E. S. A. (1995). Effect of Irradiation on the Microbiological Status and Flavouring Materials of Selected Spices. *Zeitschrift fuer Lebensmittel Untersuchung und Forschung*, 201(3), 282-288.
- Farhan, H., Rammal, H., Hijazi, A., Hamad, H., Daher, A., Reda, M. and Badran, B. (2012). In Vitro Antioxidant Activity of Ethanolic and Aqueous Extracts from Crude *Malva parviflora* L. Grown in Lebanon. *Asian Journal of Pharmaceutical and Clinical Research*, 5(3), 542-648.
- Ferris, R., Wheeler, T. R., Ellis, R. H. And Hadley, P. (1999). Seed Yield after Environmental Stresses in Soybean Grown under Elevated CO₂. *Crop Science*, 39, 710–718.
- Fishman, M. L., Levaj, B., Gillespie, D. and Scorza, R. (1993). Changes in the Physico-chemical Properties of Peach Fruit Pectin during on-tree Ripening and Storage. *Journal of American Society for Horticultural Science*, 118, 343-349.
- Friedman, M. (1996). Food Browning and its Prevention: An Overview. *Journal of Agriculture and Food Chemistry*, 44, 631-651.
- Friedrich, J. E. (2001). Liquid Chromatography of Non-volatile Acids. In: Current Protocols in Food Analytical Chemistry. United States: John Wiley & Sons, Inc.
- Fu, Z. Z., Xu, P. P., He, S. L., Jaime, A. Silva, T. and Tanaka, M. (2011). Dynamic Changes in Enzyme Activities and Phenolic Content during In-vitro Rooting of

Tree Peony (*Paeonia suffruticosa* Andr.) Plantlets. *Maejo International Journal of Science and Technology*, 5(2), 252-265.

- Gemma, C., Mesches, M.H., Sepesi, B., Choo, K., Holmes, D. B. and Bickford, P. C. (2002). Diets Enriched in Foods with High Antioxidant Activity Reverse Age-Induced Decreases in Cerebellar Beta-Adrenergic Function and Increases in Proinflammatory Cytokines. *Journal of Neuroscience*, 22, 6114-6120
- Gennaro, L., Leonardi, C., Esposito, F., Salucci, M., Madani, G., Quaglia, G. and Fogliano, V. (2002). Flavonoid and Carbohydrate Contents in Tropea Red Onions: Effects of Homelike Peeling and Storage. *Journal of Agricultural and Food Chemistry*, 50, 1904–1910.
- Georgelis, N. (2002). High Fruit Sugar Characterization, Inheritance and Linkage of Molecular Markers in Tomato. Master thesis University of Florida, USA.
- Geta, E. and Kifle, A. (2011). Production, Processing and Marketing of Ginger in Southern Ethiopia. *Journal of Horticulture and Forestry*, 3(7), 207-213.
- Ghasemnezhad, M., Marsh, K., Shilton, R., Babalar, M. and Woolf, A. (2008). Effect of Hot Water Treatment on Chilling Injury and Heat Damage in ‘Satsuma’ Mandarins: Antioxidant enzymes and Vacuolar ATPase, and Pyrophosphatase. *Postharvest Biology and Technology*, 48, 364-371.
- Ghasemzadeh, A., Jaafar, H. Z .E., Rahmat, S., Wahab, P. E. M. and Halim, M. R. A. (2010). Effect of Different Light Intensities on Total Phenolics and Flavonoids Synthesis and Antioxidant Activities in Young Ginger Varieties (*Zingiber officinale* Roscoe). *International Journal of Molecular Science*, 11, 3885–3897.
- Ghasemzadeh, A., Jaafar, H. Z. E., Ashkani, S., Rahmat, A., Juraimi, A. S. Puteh, A. and Mohamed, M. T. M. (2016). Variation in Secondary Metabolite Production as Well as Antioxidant and Antibacterial Activities of *Zingiber zerumbet* (L.) at Different Stages of Growth. *BMC Complementary and Alternative Medicine*, 16, 104-116.
- Ghasemzadeh, A., Nasiri, A., Jaafar, H. Z., Baghdadi, A. and Ahmad, I. (2014). Changes in Phytochemical Synthesis, Chalcone Synthase Activity and Pharmaceutical Qualities of Sabah snake grass (*Clinacanthus nutans* L.) in Relation to Plant Age. *Molecules*, 19(11), 17632–17648.
- Ghosh, R. (2015). Enzymatic Responses of Ginger Plants to Pythium Infection after SAR Induction . *Journal of Plant Pathology and Microbiology*, 6, 7.
- Gilmour, S. J., Sebolt, A. M., Salazar, M. P., Everard, J. D. and Thomashow, M. F. (2000). Overexpression of the Arabidopsis CBF3 Transcriptional Activator Mimics Multiple Biochemical Changes Associated with Cold Acclimation. *Plant Physiology*, 124, 1854–1865.
- Gonzalez-Aguilar, G. A., Ayala-Zavala, J. F., Olivas, G. I., de la Rosa, L. A. and Alvarez-Parrilla, E. (2010). Preserving Quality of Fresh-cut Products Using Safe Technologies. *Journal of Consumer Protection and Food safety*, 5, 65-72.

- Grace, S. C. (2005). Phenolics as Antioxidants. In *Antioxidants and Reactive Oxygen Species in Plants* (ed. Smirnoff, N. pp. 141-168). UK: Blackwell Publishing, Oxford.
- Gross, K. C., Wang, C. Y. and Salveit, M. (2002). The Commercial Storage of Fruits, Vegetables and Florist and Nursery of Crops. Beltsville Area, US: Agriculture Research Service.
- Gruenwald, J. (2004). PDR for Herbal Medicine 3rd ed. Montvale, NJ, U.S.A.: Thomson PDR.
- Grzanna, R., Lindmark, L. and Frondoza, C. G. (2005). Ginger - An Herbal Medicinal Product with Broad Anti-Inflammatory Actions. *Journal of Medicinal Food*, 8(2), 125-132.
- Gulcin, I. (2012). Antioxidant Activity of Food Constituents: An Overview. *Archives of Toxicology*, 86, 345-391.
- Ha, S. K., Moon, E., Ju, M. S., Kim, D. H., Ryu, J. H., Oh, M. S. and Kim, S. Y. (2012). 6-Shogaol, A Ginger Product, Modulates Neuroinflammation: A New Approach to Neuroprotection. *Neuropharmacology*, 63(2), 211-23.
- Hagenmaier, R. D. and Baker, R. A. (1993). Reduction in Gas Exchange of Citrus Fruit by Wax Coatings. *Journal of Agriculture and Food Chemistry*, 41, 283-287.
- Häkkinen, S. H., Kärenlampi, S. O., Mykkänen, H. M. and Törrönen, A. R. (2000). Influence of Domestic Processing and Storage on Flavonol Contents in Berries. *Journal of Agriculture and Food Chemistry*, 48, 2960-2965.
- Halachmy, I. B. and Mannheim, C. H. (1991). Modified Atmosphere Packaging of Fresh Mushrooms. *Packaging Technology and Science*, 4(5), 279-286.
- Han, X., Shen, T. and Lou, H. (2007). Dietary Polyphenols and Their Biological Significance. *International Journal for Molecular Science*, 8, 950-988.
- Hardenburg, R.E., Watada, A.E. and Wang, C.Y. (1986). The Commercial Storage of Fruits, Vegetables, and Florist, and Nursery Stocks. Washington: Agriculture Handbook.
- Hasna, O. and Afidah, A. (2009). Antioxidant Activity and Phenolic Content of *Paederia foetida* and *Syzygium aqueum*. *Molecules*, 14, 970-978.
- Hausman, J. F., Evers, D., Thiellement, H. and Jouve, L. (2000). Compared Responses of Poplar Cuttings and In Vitro Raised Shoots to Short-term Chilling Treatments. *Plant Cell Reports*, 19, 954-960.
- He, X. H., Bernart, M. W., Lian, L. Z. and Lin, L. Z. (1998). High-Performance Liquid Chromatography-Electrospray Mass Spectrometric Analysis of Pungent Constituents of Ginger. *Journal of Chromatography A*, 796, 327-334.

- Heinonen, M., Lehtonen, P. J. and Hopla, A. (1998). Antioxidant Activity of Berry and Fruit Wines and Liquor. *Journal of Agricultural Food Chemistry*, 48, 25–31.
- Hernandez, Y., Lobo, G. and Gonzalez, M. (2009). Factor Affecting Simple Extraction in the Liquid Chromatography Determination of Organic Acids in Papaya and Pineapple. *Food Chemistry*, 114, 734-741.
- Heyes, J. A., Blaikie, F. H., Downs, C. G. and Sealey, D. F. (1994). Textural and Physiological Chnges during Pepino (*Solanum muricatum*) Ripening. *Horticultural Science*, 58, 1-15.
- Hiatt, A. N., Taylor, L. S. and Mauer, L. J. (2010). Influence of Simultaneous Variations in Temperature and Relative Humidity on Chemical Stability of Two Vitamin C Forms and Implications for Shelf Life Models. *Journal of Agricultural and Food Chemistry*, 58, 3532–3540.
- Hodges, D. M., Lester, G. E., Munro, K. D. and Toivonen, P. M. (2004). Oxidative Stress: Importance for Postharvest Quality. *Horticultural Science*, 39, 924-929.
- Holderbaum, D. F., Kon, T., Kudo, T. and Guerra, M. P. (2010). Enzymatic Browning, Polyphenol Oxidase Activity, and Polyphenols in Four Apple Cultivars: Dynamics during Fruit Development. *Horticultural Science*, 45(8), 1150-1154.
- Holland, N., Menezes, H. C. and Lafuente, M. T. (2002). Carbohydrates as Related to Heat Induced Chilling Tolerance and Respiratory Rate of Fortune Mandarin Fruit Harvested at Different Maturity Stages. *Postharvest Biology and Technology*, 25(2), 181-191.
- Hollman, P. C., Tijburg, L. B. and Yang, C. S. (1997). Bioavailability of Flavonoids from Tea. *Critical Review on Food Science and Nutrition*, 37, 719-738.
- Holtmann, S., Clarke, A. H., Scherer, H. and Hohn, M. (1989). The Anti-Motion Sickness Mechanism of Ginger. A Comparative Study with Placebo and Dimenhydrinate. *Acta Oto-Laryngologica*, 108 (3-4), 168–174.
- Hossain, M. A. and Shah, M. D. (2015). A Study on the Total Phenols Content and Antioxidant Activity of Essential Oil and Different Solvent Extracts of Endemic Plant *Merremia borneensis*. *Arabian Journal of Chemistry*, 8(1), 66-71.
- Howard, L. A., Wong, A. D., Perry, A. K. and Klein, B. P. (1999). β -carotene and Ascorbic Acid Retention in Fresh and Processed Vegetables. *Journal of Food Science*, 64(5), 929–936.
- Huang, D., Ou, B. and Prior, R. L. (2005). The Chemistry Behind Antioxidant Capacity Assays. *Journal of Agricultural and Food Chemistry*, 53, 1841-1856.
- Inoue, K. (2011). Emerging Roles of the Chloroplast Outer Envelope Membrane. *Trends Plant Science*, 16, 550–557.

- Inze, D. and Montagu, M. V. (1995). Oxidative Stress in Plants. *Current Opinion in Biotechnology*, 6(2), 153-158.
- Ioannou, I. and Ghoul, M. (2013). Prevention of Enzymatic Browning in Fruit and Vegetables. *European Scientific Journal*, 9(30), 310-341.
- Ippoushi, K., Azuma, K., Ito, H., Horie, H. and Higashio, H. (2003). 6-Gingerol Inhibits Nitric Oxide Synthesis in Activated J774. Mouse Macrophages and Prevents Peroxynitrite-Induced Oxidation And Nitration Reactions. *Life Science*, 73(26), 3427-3437.
- Ishizuka, M., Yamada, F., Tanara, Y., Takeuchi, Y. and Imaseki, H. (1991). Sequential Induction of mRNAs for Phenylalanine Ammonialyase in Slice of Potato Tuber., *Plant Cell Physiology*, 32(1), 57-64.
- Jaitak, V., Sharma, K., Kalia, K., Kumar, N., HP, S., Kaul, V. and Singh, B. (2010). Antioxidant Activity of *Potentilla fulgens*: An Alpine Plant of Western Himalaya. *Journal of Food Composition and Analysis*, 23, 142-147.
- Jakes, J. S. (2007). Ginger as "Beverage of Champion". *Journal Plant Nutrition*, 38(6), 45-56.
- Jayashree, E. and Visvanathan, R. (2011). Mechanical and Thermal Properties of Ginger (*Zingiber officinale* Rosc.). *Journal of Spices and Aromatic Crops*, 20(2), 60-65.
- Jayashree, E., Kandiannan, K., Prasath, D., Rashid, P., Sasikumar, B., Senthil, K. C. M., Srinivasan, V., Suseela, B. R. and Thankamani, C. K. (2015). Ginger. Kerala, India: Indian Institute of Spices Research Kozhikode.
- Jiang, Y., Zhang, Z., Joyce, D. C. and Ketsa, S. (2002). Postharvest Biology and Handling of Longan Fruit (*Dimocarpus longan* Lour). *Postharvest Biology and Technology*, 26, 241-252.
- Jones, H. D. (1984). Phenylalanine Ammonia Lyase: Regulation of Its Induction, and its Role in Plant Development. *Phytochemistry*, 23, 1349-1355.
- Junqueira, M. J., Simões, A. N., Sediya, T., Correa, P. C. and Puschmann, R. (2014). Biochemical and Bioactive Phytonutrients Changes in Tissues of Two Cultivars of Fresh-Cut Cassava in Stick Form under Refrigerated Storage. *Rural Science*, 44(7), 1284-1290.
- Kader, A. (2006). Changes in Ascorbic Acid Contents in Stored Sweet Potatoes. Personal correspondence.
- Kader, A. A. (2002). Mangosteen – Recommendations for Maintaining Postharvest Quality. "Department of Pomology, University of California, Davis.
- Kader, A. A. (2007). Controlled Atmosphere. In: Mitcham, E. J. and Elkins, R. B. (eds.), Pear Production Handling Manual (pp. 175-177). USA: University of California Agriculture and Natural Resource Publication 3483.

- Kader, A. A. (2008). Flavor Quality of Fruits and Vegetables. *Journal of Food Science and Agriculture*, 88, 1863-1868.
- Kader, A. A. and Rolle, R. S. (2004). The Role of Post-harvest Management in Assuring the Quality and Safety of Horticultural Produce. Rome: Food and Agriculture Organization of the United Nations.
- Kalt, W., Forney, C. H., Martin, A. and Prior, R. L. (1999). Antioxidant Capacity, Vitamin C, Phenolics, and Anthocyanins after Fresh Storage of Small Fruits. *Journal of Agriculture and Food Chemistry*, 47, 4638-4644.
- Kaur, A. P. and Singh, K. (2011). Free Radicals, Antioxidant Activities in Fruit Crops and Their Importance as Phytochemicals. *The Asian Journal of Horticulture*, 6, 496-509.
- Kays, S. J. (1999). Preharvest Factors Affecting Appearance. *Postharvest Biology and Technology*, 15, 233-247.
- Kilcast, D. (2004). Texture in Foods: Solid Foods. Cambridge: Woodhead Pub. Ltd.
- Kim, B. J., Kim, J. H., Kim, H. P. and Heo, M. Y. (1997). "Biological Screening of 100 Plant Extracts for Cosmetic Use. II- Antioxidative Activity and Free Radical Scavenging Activity". *Journal of Cosmetic Science*, 19, 299-307.
- Koc, E., Dslek, C. and Ustun, A. S. (2010). Effect of Cold on Protein, Proline, Phenolic Compounds and Chlorophyll Content of Two Pepper (*Capsicum annuum* L.) Varieties. *Journal of Science*, 23(1), 1-6.
- Koo, K. L., Ammit, A. J., Tran, V. H., Duke, C. C. and Roufogalis, B. D. (2001). Gingerols and Related Analogues Inhibit Arachidonic Acid-induced Human Platelet Serotonin Release and Aggregation. *Thrombosis Research*, 103(5), 387-97.
- Koukounaras, A., Siomos, A. S., and Sfakiotakis, E. (2007). Postharvest CO₂ and Ethylene Production and Quality of Rocket (*Eruca sativa* Mill.) Leaves as Affected by Leaf Age and Storage Temperature. *Postharvest Biology and Technology*, 46, 167-173.
- Kozukue, N., Kozukue, E., Kishiguchi, M. and Lee, S. W. (1978). Studies on Keeping Quality of Vegetables and Fruits. III. Changes in Sugar and Organic Acid Contents Accompanying the Chilling Injury of Eggplant Fruits. *Science Horticulture*, 8, 19-26.
- Kranner, I., Beckett, R. P., Wornik, S., Zorn, M. and Pfeifhofer, H. W. (2002). Revival of a Resurrection Plant Correlates with its Antioxidant Status. *Plant Journal*, 31, 13-24.
- Kratsch, H. A. and Wise, R. R. (2000). The Ultrastructure of Chilling Stress. *Plant, Cell and Environment*, 23, 337-350.

- Kundu, J. K. and Surh, Y. J. (2009). Molecular Basis of Chemoprevention with Dietary Phytochemicals: Redox-regulated Transcription Factors as Relevant Targets. *Phytochemistry Review*, 8(2), 333-347.
- Lai, T. C., Huang, C. L., Chan, C. F., Lien, C. Y. and Wayne, C. L. (2013). Studies of Sugar Composition and Starch Morphology of Baked Sweet Potatoes (*Ipomoea batatas* (L.) Lam). *Journal of Food Science and Technology*, 50(6), 1193-1199.
- Laidler, K. J. and Peterman, B. F. (1983). Temperature Effects in Enzyme Kinetics. In Contemporary Enzyme Kinetics and Mechanism, (D.L. Purich, ed. pp. 149-171). New York: Academic Press.
- Langner, E., Greifenberg, S. and Gruenwald, J. (1998). Ginger: History and Use. *Advances in Therapy*, 15(1), 25-44.
- Lattanzio, V. (2003). Bioactive Polyphenols: Their Role in Quality and Storability of Fruit and Vegetables. *Journal of Applied Botany*, 77, 128-146.
- Lattanzio, V., Cardinali, A. and Palmieri, S. (1994). The Role of Phenolics in the Postharvest Physiology of Fruits and Vegetables: Browning Reactions and fungal Diseases. *Italian Journal of Food Science*, 1, 3-22.
- Lazan, H., Selamat, M. K. and Ali, Z. M. (1995). β -Galactosidase, Polygalacturonase and Pectinesterase in Different Softening and Cell Wall Modification during Papaya Fruit Ripening. *Physiologia Plantarum*, 95, 106-112.
- Lee, S. K. and Kader, A. A. (2000). Preharvest and Postharvest Factors Influencing Vitamin C Content of Horticultural Crops. *Postharvest Biology and Technology*, 20, 207-220.
- Lehninger, A. L., Nelson, D. L. and Cox, M. M. (1993). Principles of Biochemistry, Second Ed., (pp. 198-229). New York: Worth Publishers.
- Leja, M., Mareczek, A. and Ben, J. (2003). Antioxidant Properties of Two Apple Cultivars during Long-term Storage. *Food Chemistry*, 80, 303-307.
- Leubner-Metzger, G. and Amrhein, N. (1994). Phenylalanine Ammonia-lyase Analogues: Potent Inhibitors of Phenylalanine Ammonia-lyase are Weak Inhibitors of Phenylalanine-tRNA Synthetases. *Nature for Science*, 49, 781-790.
- Levine, R.L., Mosoni, L., Berlett, B.S. and Stadtman, E.R. (1996). Methionine Residues as Endogenous Antioxidants in Proteins. *Proceeding of the National Academic of Science, USA*, 93, 15036-15040.
- Levy, D. (1992). Osmotic Potential of Potatoes Subjected to a Single Cycle of Water Deficit. *Potato Research*, 35, 17-24.
- Li, M., Huang, Z. L., Tan, S. M., Mao, X. Y., Lin, H. Q. and Long, T. (2000). "Comparison on the Activities and Isoenzymes of Polyphenol Oxidase and Indoleacetic Acid Oxidase of Difficult and Easy to Root Eucalyptus Species". *Forest Research*, 13, 493-500.

- Liu, Q. and Yao, H. (2007). Antioxidant Activities of Barley Seeds Extracts. *Food Chemistry*, 102, 732–737.
- Liu, W., Zu, Y. G., Fu, Y. J., Kong, Y., Ma, W., Yang, M., Li, J. and Wu, N. (2010). Variation in Contents of Phenolic Compounds during Growth and Post-harvest Storage of Pigeon Pea Seedlings. *Food Chemistry*, 121, 732–739.
- Lo'pez-Ga'lviz, G., Saltveit, M. E., Cantwell, M. I. (1996). Wound-induced Phenylalanine Ammonia Lyase Activity: Factors Affecting Its Induction and Correlation with the Quality of Minimally Processed Lettuce. *Postharvest Biology and Technology*, 9, 223-233.
- Lownds, N. K., Banaras, M. and Bosland, P. W. (1994). Postharvest Water Loss and Storage Quality of Nine Pepper (Capsicum) Cultivars. *Horticulture Science*, 29, 191-193.
- Luengwilai, K., Beckles, D. M. and Saltveit, M. E. (2012). Chilling Injury of Harvested Tomato (*Solanum lycopersicum* L.) cv. Micro-Tom Fruit is Reduced by Temperature Pre-treatments. *Postharvest Biology and Technology*, 63, 123-128.
- Luo, Y. and Barbosa-Canovas. (1996). Preservation of apple Slices Using Ascorbic Acid and 4-hexylresorcinol. *International Food Science and Technology*, 2, 315-321.
- Maalekuu, K., Elkind, Y., Leikin-Frenkel, A., Lurie, S. and Fallik, E. (2006). The Relationship between Water Loss, Lipid Content, Membrane Integrity and LOX Activity in Ripe Pepper Fruit after Storage. *Postharvest Biology and Technology*, 42, 248-255.
- Mahmood, W. A., Sultan, S. H. and Hamza, S. R. (2009). Extraction and Characterization of Polyphenol Oxidase from Apricot, Apple, Eggplant and Potato. *Journal of Agriculture*, 37(4), 28-36.
- Maizura, M., Aminah, A. and Wan Aida, W. M. (2011). Total Phenolic Content and Antioxidant Activity of Kesum (*Polygonum minus*), Ginger (*Zingiber officinale*) and Turmeric (*Curcuma longa*) Extract. *International Food Research Journal*, 18, 529-534.
- Manassokorn, C. and Kosiyachinda, S. (1985). Temperature and Storage Conditions Affecting Storage Life of Ginger Rhizomes (*Zingiber officinale*). Bangkok, Thailand: Kasetsart University.
- Mannapperuma, J. D., Singh, R. P. and Montero, M. E. (1991). Simultaneous Gas Diffusion and Chemical Reaction in Foods Stored in Modified Atmospheres. *Journal of Food Engineering*, 14(3), 167-183.
- Manolopouloul, E. and Varzakas, T. (2011). Effect of Storage Conditions on the Sensory Quality, Colour and Texture of Fresh-Cut Minimally Processed

Cabbage with the Addition of Ascorbic Acid, Citric Acid and Calcium Chloride. *Food and Nutrition Science*, 2, 956-963.

- Mansor, P., Muhamad Ghawas, M. and Sentoer, K. (2005). Halia (*Zingiber officinale* Ross). In: Penanaman Tumbunhan Ubatan dan Beraroma. (Musa, Y., Muhammad Ghawas, M. and Mansor, P., ed). (pp.29-35). Serdang, Malaysia: MARDI.
- Marais, J. P. J., Deavours, B., Dixon, R.A. and Ferreira, D. (2006). The Stereochemistry of Flavonoids, in *The Science of Flavonoids*, Grotewold, E., Ed.. New York: Springer Science.
- Marangoni, A. G., Duplessis, P. M. and Yada, R. Y. (1997). Kinetic Model for Carbon Partitioning in *Solanum tuberosum* Tubers Stored at 2 °C and the Mechanism for Low Temperature Stress-induced Accumulation of Reducing Sugars. *Biophysical Chemistry*, 65, 211-220.
- Marangoni, A. G., Duplessis, P. M., Lencki, R. W. and Yada, R. Y. (1996). Low-temperatures Stress Induces Transient Oscillations in Sucrose Metabolism in *Solanum tuberosum*. *Biophysical Chemistry*, 61, 177-184.
- Maria, C. D. V., Liane, K. Y., Fabiana, F. S., Juan, S. A., Ricardo, A. K. and Angelo, P. J. (2005). Quality of Minimally Processed Beet Roots Stored in Different Temperatures. *Brazilian Archives of Biology and Technology*, 48(2), 503-510.
- Marshall, M. R., Kim, J. and Wei, C. I. (2000). Enzymatic Browning in Fruits, Vegetable and Seafoods. *Journal of Food and Agriculture Organization*, 41, 259-312.
- Mavandad, M., Edwards, R., Liang, X., Lamb, C. and Dixon, R. (1990). Effects of Trans-cinnamic Acid on Expression of the Bean Phenylalanine Ammonia-lyase Gene Family. *Plant Physiology*, 94, 671-680.
- Mayer, A. M. (1987). Polyphenoloxidase in Plants - Recent Progress. *Phytochemistry*, 26, 11-20.
- McGuire, R. G. (1992). Reporting of Objective Color Measurements. *Horticultural Science*, 27, 1254-1255.
- Medlicott, A. P., Sigrist, J. M. and Sy, O. (1990). Ripening of Mangoes Following Low Temperature Storage. *Journal of American Society Horticulture Science*, 115, 430-434.
- Mercado-Silva, E. and Cantwell, M. (1997). Quality Changes in Jicama Roots Stored at Chilling and Nonchilling Temperatures. *Journal of Food Quality*, 21, 211-221.
- Middleton, E. J. (1998). Effect of Plant Flavonoids on Immune and Inflammatory Cell Function. *Advances in Experimental Medicine and Biology*, 439, 175-182.

- Miller, H. E., Rigelhof, F., Marquart, L., Prakash, A. and Kanter, M. (2000). Antioxidant Content of Whole Grain Breakfast Cereals, Fruits and Vegetables. *Journal of American College of Nutrition*, 19(3), 312-319.
- Mirdehghan, S. H. and Ghotbi, F. (2014). Effects of Salicylic Acid, Jasmonic Acid and Calcium Chloride on Reducing Chilling Injury of Pomegranate (*Punica granatum* L.) Fruit. *Journal of Agricultural Science technology*, 16, 163-173.
- Mishra, R. K., Kumar, A. and Kumar, A. (2012). Pharmacological Activity of *Zingiber officinale*. *International Journal of Pharmaceutical and Chemical Sciences*, 1(3), 1422-1427.
- MOA. (2013). Ministry of Agriculture and Agro-based Industry Malaysia. Retrieved on 12 February 2016 from <http://www.moa.gov.my/sayur-sayuran>.
- Moellering, E. R., Muthan, B. and Benning, C. (2010). Freezing Tolerance in Plants Requires Lipid Remodeling at the Outer Chloroplast Membrane. *Science*, 330, 226-228.
- Molisch, H. (1996). The Freezing of Plants at Temperatures above the Ice Point. Vienna, Austria: The Imperial Academy Of Sciences.
- Moretti, C. L., Mattos, L. M., Calbo, A. G. and Sargent, S. A. (2010). Climate Changes and Potential Impacts on Postharvest Quality of Fruit and Vegetable Crops: A Review. *Food Research International*, 43, 1824–1832.
- Morison, J. I. L. and Lawlor, D. W. (1999). Interactions between Increasing CO₂ Concentration and Temperature on Plant Growth. *Plant, Cell and Environment*, 22, 659–82.
- Murata, M., Noda, I. and Homma, S. (1995). Enzymatic Browning of Apples on the Market: Relationship between Browning, Polyphenol Content, and Polyphenol Oxidase. *The Japanese Society for Food Science and Technology*, 42, 820–826.
- Murata, N., Allakhverdiev, S. I. and Nishiyama, Y. (2012) The Mechanism of Photoinhibition In Vivo: Re-evaluation of the Roles of Catalase, α -tocopherol, Non-photochemical Quenching, and Electron Transport. *Biochimica et Biophysica Acta*, 1817(8), 1127–1133.
- Naik, G. H., Priyadersini, K. L., Satav, J. G., Bansvalikar, M. M., Sohoni, P. P. and Biyani, M. K. (2003). Comparative Antioxidant Activity of Individual Herbal Component Used in Ayurvedic Medicine. *Phytochemistry*, 63, 97–104.
- Naziri, D., Quaye, W., Siwoku, B., Wanlapatt, S., Tu, V. P. and Bennett, B. (2014). Not All Those Who Wander are Lost: A Comparative Analysis 69 of Postharvest Losses in Cassava Value Chains in Ghana, Nigeria, Thailand and Vietnam. Paper submitted for the 14th Congress of the European Association of Agricultural Economists "Agri-food and Rural Innovations for Healthier Societies", Ljubljana, Slovenia.

- Nguyen, T. B. T., Ketsa, S. and van Doorn, W. G. (2003). Relationship between Browning and the Activities of Polyphenol Oxidase and Phenylalanine Ammonia Lyase in Banana Peel during Low Temperature Storage. *Postharvest Biology and Technology*, 30, 187-193.
- Nunes, M. N., Emond, M. C., Brecht, J. P., Dea, J. K. and Proulx, S. (2007). Quality Curves for Mango Fruit (cv. Tommy Atkins and Palmer) Stored at Chilling and Nonchilling Temperatures. *Journal of Food Quality*, 30, 104-120.
- Oktaý, M., Kufrevioglu, I., Kocacalıskan, I. and Sakiroglu, H. (2003). Polyphenol Oxidase from Amasya Apple. *Journal of Food Science*, 60, 494-496.
- Ong, H. C. (1996). *Nelumbo nucifera* (pp.131-133) In: Flach, M. and F. Rumawas (Eds.). Plant Resources of South-East Asia No.9: Plants yielding non-seed carbohydrates. Wageningen, The Netherlands: Pudoc Science Publication.
- Onyenekwe, P. (2000). Assessment of Oleoresin and Gingerol Contents in Gamma Irradiated Ginger Rhizomes. *Food Nahrung*, 44(2), 130–2.
- Oyetade, O. A., Oyeleke, G. O., Adegoke, B. M. and Akintunde, A. O. (2012). Stability Studies on Ascorbic Acid (Vitamin C) from Different Sources. *Journal of Applied Chemistry*, 2(4), 20-24.
- Ozoglu, H. and Bayindirli, A. (2002). Inhibition of Enzymic Browning in Cloudy Apple Juice with Selected Antibrowning Agents. *Food Control*, 13, 213–221.
- Palma, T., Marangoni, A. G. and Stanley, D. W. (1995). Environmental Stress Affect Tomato Microsomal Membrane Function Differently than Natural Ripening and Senescence. *Postharvest Biology and Technology*, 6, 257-273.
- Pantastico, E.B., Ali Azizan, M., Abdullah. H., Acedo, A.L., Dasuki, I.M. and Kosiyachinda, S. (1990). Physiological Disorders of Banana Fruit. In: Hassan, Abd., Pantastico, Er.B. (Eds.), *Banana: Fruit Development, Postharvest Physiology, Handling and Marketing in ASEAN*. ASEAN Food Handling Bureau, Kuala Lumpur, Malaysia, pp. 85–103
- Park, C., Kim, G. Y., Kim, G. D., Choi, B. T., Park, Y. M., and Choi, Y. H. (2006). Induction of G2/M Arrest and Inhibition of Cyclooxygenase-2 Activity by Curcumin in Human Bladder Cancer T24 Cells. *Oncology Report*, 15, 1225-1231
- Paull, R. E. (1994). Response of Tropical Horticultural Commodities to Insect Disinfestation Treatments. *Horticulture Science*, 29, 988–991.
- Paull, R. E. (1999). Effect of Temperature and Relative Humidity of Fresh Commodity Quality. *Postharvest Biology and Technology*, 15, 263-277.
- Paull, R. E. and Chen, C. C. (2015). *Ginger Postharvest Quality-Maintenance Guidelines*. Manoa: University of Hawaii.

- Paull, R. E. and Chen, N. J. (1989). Waxing and Plastic Wraps Influence Water Loss from Papaya Fruit during Storage and Ripening. *Journal of American Society for Horticultural Science*, 114, 937-942.
- Paull, R. E., Chen, N. J., and Goo, T. T. C. (1988). Compositional Changes in Ginger Rhizomes During Storage. *Journal of the American Society for Horticultural Science*, 113(4), 584-588.
- Peiser, G., Lo'pez-Ga'lvez, G., Cantwell, M. and Saltveit, M. E. (1998). Phenylalanine Ammonia Lyase Inhibitors Control Browning of Cut Lettuce. *Postharvest Biology and Technology*, 14, 171-177.
- Perez-Tello, G. O., Silva-Espinoza, B. A., Vargas-Arispuro, I., Briceno-Torres, B. O. and Martinez-Tellez, M. A. (2001). Effect of Temperature on Enzymatic and Physiological Factors Related to Chilling Injury in Carambola Fruit (*Averrhoa carambola* L.). *Biochemical and Biophysical Research Communications*, 287, 846-851.
- Policegoudra, R. S. and Aradhya, S. M. (2007). Biochemical Changes and Antioxidant Activity of Mango Ginger (*Curcuma amada* Roxb.) Rhizomes During Postharvest Storage at Different Temperatures. *Postharvest Biology and Technology*, 46, 189-194.
- Policegoudra, R. S., Divakar, S. and Aradhya, S. M. (2007). Identification of Difurocumenonol, A Novel Antimicrobial Compound from Mango Ginger (*Curcuma amada* Roxb.) Rhizome. *Journal of Application Microbiology*, 102, 1594-1602.
- Prior, R. L. and Cao, G. (2000). Antioxidant Phytochemicals in Fruits and Vegetables. *Horticulture Sciences*, 35, 588-592.
- Prior, R. L., Cao, G., Martin, A., Sofic, E., McEwen, J., O'Brien, C., Lischner, N., Ehlenfeldt, M., Kalt, W. and Krewer, G. (1998). Antioxidant Capacity as Influenced by Total Phenolic and Anthocyanin Content, Maturity, and Variety of Vaccinium species. *Journal of Agriculture and Food Chemistry*, 46(7), 2686-2693.
- Prior, R. L., Wu, X. and Schaich, K. (2005). Standardized Methods for the Determination of Antioxidant Capacity and Phenolics in Food and Dietary Supplement. *Journal of Agriculture and Food Chemistry*, 53, 4290-4303.
- Proulx, E., Cecilia, M., Nunes, N., Emond, J. P. and Brecht, J. K. (2005). Quality Attributes Limiting Papaya Postharvest Life at Chilling and Non-chilling Temperatures. *Florida State Horticultural Society*, 118, 389-395.
- Purnomo, H., Jaya, F. and Widjanarko, S. B. (2010). The Effects of Type and Time of Thermal Processing on Ginger (*Zingiber officinale* Roscoe) Rhizome Antioxidant Compounds and its Quality. *International Food Research Journal*, 17, 335-347.

- Pyo, Y. H, Lee, T. C., Longedra, L. and Rosen, R. T. (2004). Antioxidant Activity and Phenolic Compounds of Swiss chard (-Vulgaris Subspecies Cyclo) Oleoresins. *Food Chemistry*, 85, 19-26.
- Qidwai, W., Alim, S. R., Dhanani, R. H., Jehangir, S., Nasrullah, A. and Raza, A. (2003). Use of Folk Remedies among Patients in Karachi Pakistan. *Journal of Ayub Medical College, Abbottabad*, 15(2), 31–33.
- Queiroz, C., Lopes, M. L. M., Fialho, E. and Valente-Mesquita, V. L. (2008). Polyphenol Oxidase: Characteristics and Mechanisms of Browning Control. *Food Reviews International*, 24(4), 361-375.
- Rahmani, A. H., Shabrmi, F. M. A. I. and Aly, S. M. (2014). Active Ingredients of Ginger as Potential Candidates in the Prevention and Treatment of Diseases via Modulation of Biological Activities. *International Journal of Physiology, Pathophysiology and Pharmacology* 6(2), 125–136.
- Rastovski, A. (1987). Storage Losses. Storage of Potatoes - Post-Harvest Behavior, Store Design, Storage Practice, Handling (pp177-180). Netherland: Pudoc, Wageningen.
- Ratnambal, M. J., Gopalam, A., and Nair, M. K. (1987). Quality Evaluation in Ginger (*Zingiber officinale* Rosc.) in Relation to Maturity. *Journal of Plantation Crops*, 15(2), 108-117.
- Ravi, V. and Aked, J. (1996). Review on Tropical Root and Tuber Crops. II. Physiological Disorders in Freshly Stored Roots and Tubers. *Critical Review in Food Science and Nutritional*, 36, 711–731.
- Ravindran, P. and Babu, K. N. (2004). Ginger: The Genus Zingiber. New York, USA: CRC Press.
- Reid, M. S. (2004). Ginger, Shell Ginger, Torch Ginger: Recommendations for Maintaining Postharvest Quality. Retrieved on 27 January 2016 from University of California. <http://postharvest.ucdavis.edu/pformentals/Ginger/>
- Reihaneh, A. G. and Mehdi, G. D. (2010). Studies on Physicochemical Properties of Tomato Powder as Affected by Different Dehydration Methods and Pretreatments. *World Academy of Science Engineering and Technology*, 69, 117–129.
- Riteniour, M. A. and Saltveit, M. E. (1996). Identification of A Phenylalanine Ammonia-Lyase Inactivating Factor in Harvested Iceberg Lettuce (*Lactuca Sativa* L.), *Physiologia Plantarum*, 97(2), 327-331.
- Rivera, F., Pelayo-Zaldivar, C., Leon, F. D. D., Buentello, B., Castillo-Rivera, M. (2007). Cold-conditioning Treatment Reduces Chilling Injury in Mexican Limes (*Citrus aurantifolia* S.) Stored at Different Temperature. *Journal of Food Quality*, 30, 121-134.

- Rivero, R. M., Ruiz, J. M., Garcia, P. C., Lopez-Lefebvre, L. R., Sanchez, E. and Romero, L. (2001). Resistance to Cold and Heat Stress: Accumulation of Phenolic Compounds in Tomato and Watermelon Plants. *Plant Science*, 160, 315-321.
- Robards, K., Prenzler, P., Tucker, G., Swatsitang, P. and Glover, W. (1999). Phenolic Compounds and Their Role in Oxidative Processes in Fruits. *Food Chemistry*, 66, 401-436.
- Robert, C. (1998). A Comparative Grammar of the Dravidian or South-Indian Family of Languages. *Asian Educational Services*.
- Sadler, G. D and Murphy, P. A. (2010). pH and Titratable Acidity. In. "Food Analysis". ed. by. S. S. Nielsen (pp. 219-238). New York: Springer Science + Business Media, LLC.
- Saltveit, M. E. (2000). Wound Induced Changes in Phenolic Metabolism and Tissue Browning are Altered by Heat Shock. *Postharvest Biology and Technology*, 21, 61-69.
- Samim, W. and Banks, N. H. (1993). Colour Changes in Bruised Apple Fruit Tissue. *New Zealand Journal of Crop and Horticultural Science*, 21, 367-372.
- Sams, C. E. (1999). Preharvest Factors Affecting Postharvest Texture. *Postharvest Biology and Technology*, 15, 249-254.
- Sapers, G. M. (1993). Browning of Foods: Control by Sulfites. Antioxidants and Other Means. *Food Technology*, 47(10), 75-84.
- Sarikurkcü, C., Arisoy, K., Tepe, B., Cakir, A., Abali, G. and Mete, E. (2009). Studies on the Antioxidant Activity of Essential Oil and Different Solvent Extracts of *Vitex agnus castus* L. Fruits from Turkey. *Food and Chemistry Toxicology*, 47(10), 2479-2483.
- Sasaki, F. F., Del Aguila, J. S., Gallo, C. R., Ortega, E. M. M., Jacomino, A. P. and Kluge, R. A. (2006). Physiological, Qualitative and Microbiological Changes During Storage Pumpkin Minimally Processed into Different Types Of Cuts. *Brazilian Horticulture*, 24, 170-174.
- Sayyari, M., Babalar, M., Kalantari, S., Serrano, M. and Valero, D. (2009): Effect of Salicylic Acid Treatment on Reducing Chilling Injury in Stored Pomegranates. *Postharvest Biology and Technology*, 53, 152-154.
- Scandalios, J. G. (2001). Oxygen Stress and Superoxide Dismutase. *Plant Physiology*, 101, 7-12.
- Schijlen, E. G. W. M., de Vos, C. H. R., Martens, S., Jonker, H. H., Rosin, F. M., Molthoff, J. W., Tikunov, Y. M., Angenent, G. C., van Tunen, A. J. and Bovy, A. G. (2007). RNAi Silencing of Chalcone Synthase, the First Step in the Flavonoid Biosynthesis Pathway, Leads to Parthenocarpic Tomato Fruits. *Plant Physiology*, 144, 1520-1530.

- Schuhbaum, H., Burgermeister, J., Paper, D.H. and Franz, G. (2000) Anti-inflammatory Activity of *Zingiber officinale* extracts. *Pharmaceutical and Pharmacological Letters*, 10 (2), 82–85.
- Sergio, T. F., Engil, I. P.P., Gomez, A. C. S., Brackmann, A., Nicoloso, F. and Bisognin, D. A. (2012). Processing Quality of Potato Tubers Produced During Autumn and Spring and Stored At Different Temperatures. *Journal of the Brazilian Association for Horticultural Science*, 30(1), 91-98.
- Serrano, M., Guillen, F., Martinez-Romero, D., Castillo, S. and Valero, D. (2005). Chemical Constituents and Antioxidant Activity of Sweet Cherry at Different Ripening Stages. *Journal of Agriculture and Food Chemistry*, 53, 2741-2745.
- Shahidi, F. and Naczk, M. (2004). Phenolic Compounds in Fruits and Vegetables (pp. 131–156). Tulsa, OK, USA: Phenolics in Food and Nutraceutical, CRC Press.
- Sharififar, F., Nudeh-Dehghn, G. and Mirtajaldini, M. (2008): Major Flavonoids with Antioxidant Activity from *Teucrium polium* L. *Food and Chemistry*, 112, 885-888.
- Shewfelt, R. L. (1990). Sources of Variation in the Nutrient Content of Agricultural Commodities from the Farm to the Consumer. *Journal of Food Quality*, 13, 37-54.
- Shewfelt, R. L. (2009). Measuring Quality and Maturity in: Florkowski, W. J., Shewfelt, R., Brueckner, B. and Prussia, S. E. (eds). *Postharvest Handlings: A Systems Approach* (2nd ed). San Diego, CA: Academic Press.
- Shewfelt, R. L. and del Rosario, B. A. (2000). The Role of Lipid Peroxidation in Storage Disorders of Fresh Fruits and Vegetables. *Horticulture Science*, 35, 575-579.
- Shibairo, S. I., Upadhyaya, M. K. and Toivonen, P. M. A. (1997). Postharvest Moisture Loss Characteristics of Carrot (*Daucus carota* L.) Cultivars during Short-term Storage. *Horticulture Science*, 71(1-2), 1-12.
- Shukla, Y. and Singh, M. (2007). Cancer Preventive Properties of Ginger: A Brief Review. *Food and Chemical Toxicology*, 45(5), 683–690.
- Shukor, A. R., Aziz, A. B. and Ahmad Shokri, A. (1986). Physico-chemical Changes of Fresh Ginger Rhizomes as Influenced by Storage Temperature and Duration. *Mardi Research Bulletin*, 4(3), 243-248.
- Simoes, A. N., Puiatti, M., Salomao, L. C. C., Mosquim, P. R. and Puschmann, R. (2010). Effect in the Quality of Intact and Minimally Processed Leaves of Collard Greens Stored at Different Temperatures. *Brazilian Horticulture*, 28(1), 81-86.

- Simões, N. A., Ventrella, M. C., Moretti, C. L., Cernelossi, M. A. G. and Puschmann, R. (2015). Anatomical and Physiological Evidence of White Blush on Baby Carrot Surfaces. *Postharvest Biology and Technology*, 55, 45-52.
- Simon, G. (2002). Bioactive Compounds from Natural Sources-Isolation Characterization and Biological Properties. UK: John Wiley and Sons, Ltd.
- Singh, P. K. and Rao, K. M. (2012). Phytochemicals in Vegetables and Health Protective Effects. *Asian Journal of Agriculture and Rural Development*, 2(2), 177-183.
- Singh, S. P., Singh, Z. and Swinny, E. E. (2009). Postharvest Nitric Oxide Fumigation Delays Fruit Ripening and Alleviates Chilling Injury during Cold Storage of Japanese Plums (*Prunus salicina* Lindell). *Postharvest Biology and Technology*, 53, 101-108.
- Siti Fairuz, Y., Mahmud, T. M. M., Farinazleen M. G. and Ahmad, S. H. (2012). A Survey on Contamination of *E. coli* O157:H7 on Butterhead Lettuce. *Trans. Malaysian Society Plant Physiology*, 20, 167-169.
- Sivakumar, D. and Marisa, M. W. (2012). Papaya Fruit Quality Management during the Postharvest Supply Chain. *Food Review International*.
- Solecka, D. and Kacperska, A. (2003). Phenylpropanoid Deficiency Affects the Course of Plant Acclimation to Cold. *Physiology Plant*, 119, 253-262.
- Soliva-Fortuny, R.C., Biosca-Biosca, M., Grigelmo-Miguel, N. and Martín-Belloso, O. (2002). Browning, Polyphenol Oxidase Activity and Headspace Gas Composition during Storage of Minimally Processed Pears Using Modified Atmosphere Packaging. *Journal of Science and Food Agriculture*, 82, 1490-1496.
- Solomos, T. and Laties, G. G. (1975). The Mechanism of Ethylene and Cyanide Action in Triggering the Rise in Respiration in Potato Tubers. *Plant Physiology*, 55, 73-78.
- Srivastava, A., Akoh, C. C., Yi, W., Fischer, J. and Krewer, G. (2007). Effect of Storage Conditions on the Biological Activity of Phenolic Compounds of Blueberry Extract Packed in Glass Bottles. *Journal of Agriculture and Food Chemistry*, 55, 2705-2713.
- Steinmetz, K. A. and Potter, J. D. (1991) Vegetables, Fruits, and Cancer. Epidemiology, *Cancer Causes Control*, 2, 325-337.
- Suhaj, M. (2006). Spice Antioxidants Isolation and Their Antiradical Activity: A Review. *Journal of Food Composition and Analysis*, 19, 531-537.
- Sweetlove, L. J., Beard, K. F., Nunes-Nesi, A., Fernie, A. R. and Ratcliffe, R. G. (2010). Not Just a Circle: Flux Modes in the Plant TCA Cycle. *Trends in plant science*, 15(8), 462-470.

- Taga, M. S., Miller, E. E. and Pratt, D. E. (1984). Chia Seeds as a Source of Natural Lipid Antioxidants. *Journal of the American Oil Chemist's Society*, 61, 928-993.
- Tang, M., Bie, Z. L., Wu, M. Z. and Feng, J. X. (2010). Changes in Organic Acids and Acid Metabolism Enzymes in Melon Fruit during Development. *Horticultural Science*, 123(3), 360-365.
- Thompson, D. S., Davies, W. J. and Ho, L. C. (1998). Regulation of Tomato Fruit Growth by Epidermal Cell Wall Enzymes. *Plant, Cell and Environment*, 21, 589-599.
- Thumdee, S., Manenoi, A., Chen., N. and Paull, R. (2010). Papaya Fruit Softening: Role of Hydrolases. *Tropical Plant Biology*, 3, 98-109.
- Tijssens, L. M. M., Hertog, M. L. A. T. M., Vanschaik, A. C. R. and De Jager, A. (1999). Modeling the Firmness of Elstar Apples During Storage and Transport. *International of Horticultural Science*, 48, 363-372.
- Toivonen, P. M. A. and Brummell, D. A. (2008). Biochemical Bases of Appearance and Texture Changes in Fresh-cut Fruit and Vegetables. *Postharvest Biology and Technology*, 48(1), 1-14.
- Tomas-Barberan, F. A. and Espin, J. C. (2001). Phenolic Compounds and Related Enzymes as Determinants of Quality in Fruits and Vegetables. *Journal of Science of Food and Agriculture*, 81, 853-876.
- Toor, R. K. and Savage, G. P. (2006). Changes in Major Antioxidant Components of Tomatoes during Post-harvest Storage. *Food Chemistry*, 99, 724-727.
- Trichopoulou, A., Vasilpoulou, E. and Hollman, P. (2000). Nutritional Composition and Flavonoid Content of Edible Wild Greens and Green Pies: A Potential Rich Source of Antioxidant Nutrients in the Mediterranean Diet. *Food Chemistry*, 70, 319-323.
- Tripathi, S., Suzuki, J. Y., Carr, J. B. and Gonsalves, D. (2011). Nutritional Composition of Rainbow Papaya, the First Commercialized Transgenic Fruit Crop. *Journal of Food Composition and Analysis*, 24(2), 140-147.
- Upadhyay, I. P., Noomhorm, A. and Ilangantileke, S. G. (1994). Effects of Gamma Irradiation and Hot Water Treatment on the Shelf Life and Quality of Thai mango cv Red. *The Australian Centre for International Agricultural Research*, 348-351.
- Valente, A. and Albuquerque, T. G. (2011). Ascorbic Acid Content in Exotic Fruits: A Contribution to Produce Quality Data for Food Composition Databases. *Food Research International*, 44(7), 2237-2242.
- Valero, E., Varon, R. And Garcia-Carmona, F. (1988). Characterization of Polyphenol Oxidase from Airen Grapes. *Journal of Food Science*, 53, 1482-1485.

- Van der Sluis, A. A., Dekker, M., de Jager, A. and Jongen, W. M. (2001). Activity and Concentration of Polyphenolic Antioxidants in Apple: Effect of Cultivar, Harvest Year, and Storage Conditions. *Journal of Agriculture and Food Chemistry*, 49(8), 3606–3613.
- Vaughn, K. C. and Duke, S. O. (1984). Function of Polyphenol Oxidase in Higher Plants. *Physiology Plant*, 60, 106–112.
- Veraverbeke, E. A., Verboven, P., Lammertyn, J., Cronje, P., Baerdemaeker, J. D. (2006). Thermographic Surface Quality Evaluation of Apple. *Journal of Food Engineering*, 77, 162-168.
- Vilas-Boas, E. V. B. (1999). Physiological Aspects of Fruit Development (pp. 75). Lavras, Brazil: Federal University of Lavras.
- Vitti, M. C. D., Kluge, R. A., Yamamoto, L. K. and Jacomino, A. P. (2003). Beet Behavior Minimally Processed in Different Cutting Thicknesses. *Agricultural Research Brazilian*, 21, 623-626.
- Voss, D. H. (1992). Relating Colorimeter Measurement of Plant Color to the Royal Horticultural Society Colour Chart. *Horticulture Science*, 27(12), 1256-1260.
- Vuorinen, H., Maata, K. and Torronen, R. (2000). Content of the Flavonols Myricetin, Quercetin, and Kaempferol in Finnish Berry Wines. *Journal of Agriculture and Food Chemistry*, 48, 2675–2680.
- Wajahatullah, K., Balkrishnan, P. and Donald, L. S. (2002). Chitosan and Chitin Oligomers Increase Phenylalanine Ammonia-lyase and Tyrosine ammonia-lyase Activities in Soybean Leaves. *Plant Physiology*, 160, 859-863.
- Walker, J. R. L. and Ferrar, P. H. (1998). Diphenol Oxidase, Enzyme-Catalysed Browning and Plant Disease Resistance. *Biotechnology and Genetic Engineering Reviews*, 15, 457-498.
- Wang S.Y. and Zheng, W. (2001). Effect of Plant Growth Temperature on Antioxidant Capacity in Strawberry. *Journal of Agriculture and Food Chemistry*, 49, 4977-4982.
- Wang, C. Y. (1992). Chilling Injury of Tropical Horticultural Commodities. *Horticultural Science*, 27, 569.
- Wang, C. Y. (1993). Alleviation of Chilling Injury in Horticultural Crops (pp. 282-283). Bacon Raton, Florida: CRC Press
- Wang, C.Y. (1990). Chilling Injury of Horticultural Crops. Bacon Raton, Florida: CRC Press.
- Wang, C. Y. (1982). Physiological and Biochemical Responses of Plants to Chilling Stress. *Horticultural Science*, 17, 173–186.

- Wang, H. and Helliwell, K. (2001). Determination of Flavonols in Green and Black Tea Leaves and Green Tea Infusions by High-Performance Liquid Chromatography. *Food Research International*, 34, 223-227.
- Wang, S. Y., Bunce, J. A. and Maas, J. (2003). Elevated Carbon Dioxide Increases Contents of Antioxidant Compounds in Field-grown Strawberries. *Journal of Agriculture and Food Chemistry*, 51(15), 4315-4320.
- Wedding, R. T. (1989). Malic Enzymes of Higher Plants. *Plant Physiology*, 90(2), 367-371.
- Weidner, S., Kordala, E., Brosowska-Arendt, W., Karamać, M., Kosińska, A. and Amarowicz, R. (2009). Phenolic Compounds and Properties of Antioxidants in Grapevine Roots Followed by Recovery. *Polish Journal of Botany*, 78, 279-286.
- Weiss, E. A. (2002). Spice Crops. Oxon, UK: CAB International publishing.
- Westby, A. (2002). Cassava Utilization, Storage and Small-scale Processing. UK: Natural Resources Institute, University of Greenwich.
- Whitaker, J. R. (1994). Principles of Enzymology for the Food Sciences, Second Ed., (pp. 271-556). New York: Marcel Dekker.
- Whitaker, J. R. and Lee, C. Y. (1995). Enzymatic Browning and its Prevention. ACS Symposium Series, 600, 3-7.
- Wilcox, B. F. 2006. Fundamentals of Food Chemistry 1st Ed. Vol. 1-2. Yola: Paraclete Publishers Nigeria.
- Wills, R., McGlasson, B., Graham, D. and Joyce, D. (2007). Postharvest: An Introduction to the Physiology and Handling of Fruit, Vegetables and Ornamentals (2 edn). Sydney: University of New South Wales Press.
- Wismer, W. V., Marangoni, A. G. and Yada, R. Y. (1995). Low Temperature Sweetening in Roots and Tubers. *Horticultural Review*, 27, 203-231.
- Wismer, W. V., Worthing, W. M., Yada, R. Y. and Marangoni, A. G. (2000). Membrane Lipid Dynamics and Lipid Peroxidation in the Early Stages of Low-temperature Sweetening in Tubers of *Solanum tuberosum*. *Physiologia Plantarum*, 102, 396-410.
- Wohlmuth, H. (2008). Phytochemistry and Pharmacology of Plants from the Ginger Family, Zingiberaceae. Australia: Southern Cross University.
- Woolf, A. B., Wexler, A., Prusky, D., Kobiler, E., and Lurie, S. (2000). Direct Sunlight Influences Postharvest Temperature Responses and Ripening of Five Avocado Cultivars. *Journal of the American Society for Horticultural Science*, 125, 370-376.

- Wu, J. J. and Yang, J. S. (1994). Effect of Irradiation on the Volatile Compounds of Ginger Rhizome (*Zingiber officinale* Roscoe). *Journal of Agriculture and Food Chemistry*, 42, 2574-2577.
- Wu, T. S., Wu, C., Wu, P. L., Chern, C. Y., Leu, Y. L. and Chan, Y. Y. (1998). Structure and Synthesis of (n)-dehydroshogaols from *Zingiber officinale*. *Phytochemistry*, 48, 889–891.
- Yang, J., Gadi, R. and Thomson, T. (2011). Antioxidant Capacity, Total Phenols, and Ascorbic Acid Content of Noni (*Morinda citrifolia*) Fruits and Leaves at Various Stages of Maturity. *Micronesica*, 41, 167–176.
- Yanuriati, A., Savage, G. P., Rowe, R. N. (1999). The Effects of Ethanol Treatment on the Metabolism, Shelf Life and Quality of Stored Tomatoes at Different Maturities and Temperatures. *Journal of Science of Food and Agriculture*, 79, 995–1002.
- Yeh, H. Y., Chuang, C. H., Chen, H. C., Wan, C. J, Chen, T. and Lin L. Y. (2014). Bioactive Components Analysis of Two Various Gingers (*Zingiber officinale* Roscoe) and Antioxidant Effect of Ginger Extracts. *Food and Science Technololgy*, 55, 329-334.
- Yelenosky, G. (1975). Cold Hardening in Citrus Stems. *Plant Physiology*, 56, 540-543.
- Young, T. E., Juvik, J. A. and Sullivan, J. G. (1993). Accumulation of the Components of Total Soluble Solids in Ripening Fruits of Tomato. *Journal of the American Society of Horticultural Science*, 118(2), 286-292.
- Yu, Z. F., Zhao, Y. X., Li, N., Xia, Z. H. and Peng, G. X. (2002). Analysis of Substrates of Browning Enzymes in Fresh Cut Lotus Rhizomes. *Basic Research in Cardiology*, 33, 41-44.
- Yun, S. D. and Lee, S. K. (1998). Quality of Red Ginseng Processed from the stored Roots. *International Society for Horticultural Science*, 464, 534-536.
- Zawistowsky, J., Biliaderis, C. G. and Eskin, N. A. M. (1991). "Polyphenol Oxidase," *Oxidative Enzymes in Foods* (pp. 217-273). London: Elsevier.
- Zhan, J. F. and He, C. L. (2006). Comparison of Several Storage Methods of Lotus Root. *Journal of Shaanxi University of Science and Technology*, 24, 66-68.
- Zhang, X., Iwaoka, W. T., Huang, A. S., Nakamoto, S. T. and Wong, R. (1994). Gingerol Decreases after Processing and Storage of Ginger. *Journal of Food Science*, 59(6), 1338-1343.
- Zhang, Z., Yang, M. J., and Pawliszyn, J. (1994). Solid-Phase Microextraction: A Solvent-Free Alternative for Sample Preparation. *Analytical Chemistry*, 66, 844-853.

Zhuang, H., Hildebrand, D. F. and Barth, M. M. (1997). Temperature Influenced Lipid Peroxidation and Deterioration in Broccoli Buds during *Postharvest Storage*. *Postharvest Biology and Technology*, 10, 49-58.

Zo'n, J. and Amrhein, N. (1992). Inhibitors of Phenylalanine Ammonia-lyase: 2-aminoindan-2-phosphonic Acid and Related Compounds. *European Journal of Organic Chemistry*, 6, 625-628.

