

UNIVERSITI PUTRA MALAYSIA

ISOLATION OF SERRATIA MARCESCENS (ISOLATE 30) AND PARTIAL PURIFICATION OF ITS ACRYLAMIDE-DEGRADING AMIDASE

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By

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PARTIAL PURIFICATION AND CHARACTERIZATION OF AMIDASE FROM ACRYLAMIDE-DEGRADING BACTERIA STRAIN 30

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May 2005

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A total of 217 bacterial isolates were isolated by an enrichment procedure from Malaysian soils which had been treated and non-treated with acrylamide. From the preliminary screening using MTT (3-[4,5-dimethylthiazol-2-yl]-,5diphenyltetrazoliumbromide) assay, 10 bacterial isolates were found to be capable of utilizing acrylamide as a nitrogen source. Formation of the blue colour of formazan and high absorbance were the criteria in the selection of bacteria. These ten isolates were labeled as Isolate 5, 9, 10, 15, 30, 45, 60, 110, 115 and 145. The isolates were subjected to secondary screening using MTT assay, Direct Plate Count and High Perfomance Liquid Chromatography (HPLC). From the results obtained, five bacterial isolates, Isolate 9, 10, 15, 30 and 60 gave the highest colony count in CFU/ mL and absorbance reading from the MTT assay. These five bacterial isolates were then selected for further of examined for their ability to degrade acrylamide by HPLC. Isolate 30 showed the best degradation of acrylamide with 99.84% degradation after 48 hours incubation in



enrichment culture with acrylamide as a substrate at a final concentration of 100 mg/L. Isolate 60 showed the second highest degradation of acrylamide at 70.53%, followed by Isolate 10 at 50.8% degradation and isolates 15 and 9 at 16.24% and 14.58% respectively. Control without bacteria accounted only 0.16% acrylamide degradation.

Isolate 30 was selected for optimization of growth media with six different parameters. These parameters were temperature, pH, different types of carbon sources, different concentrations of carbon source, different amides as the substrate and effect of different concentrations of selected amide substrate. From the results obtained, Isolate 30 showed an optimum growth temperature at 27 ° C. Its highest colony count \log_{10} cfu/ml was directly proportional to the growth of bacteria. This isolate showed the highest growth at pH 7.5 where this type of bacteria grew well near neutrality and slightly alkaline media. For the carbon sources optimization, glucose was selected due to the high colony count. Isolate 30 was found to grow best at the glucose concentration of 1%. Among the amide substrates, acrylamide was found to be the best sole nitrogen source which can support bacterial growth in enrichment cultures and the bacteria showed the highest colony count when 400 mg/L acrylamide was provided.

From the macroscopic and microscopic observations of Isolate 30, the bacteria was gram-negative rod, might become differentiated into a convex, pigmented and relatively opaque centre and an effuse, colourless, almost transparent periphery with an irregular crenated edge. Biochemical tests using Microbact 24E



(12A + 12B) showed that the Isolate 30, which had been isolated from Tanjung Karang, Selangor gave 99.83% similarity to Serratia marcescens.

Amidase (EC 3.5.1.4) produced by Serratia marcescens strain 30 was partially purified and characterized. The purification procedure used, including ionexchange chromatography, Mono-QTM strong-anion exchanger, ultrafiltration and gel filtration, yielded a partially purified amidase fraction having an overall purification factor of 13.83 fold and a yield of 48.33%. Indophenol blue method was used for the amidase assay and amidase characterization studies was done to determine the K_m and Vmax value, the effect of temperature and pH on amidase activity and the effect of heavy metals on amidase activity. $K_{\rm m}$ values for acrylamide were determined by incubating amidase at 50° C with various concentrations of acrylamide (0.25 to 20mM) in phosphate buffer (50mM; pH 7.5). By using non-linear regression analysis, the Km and Vmax values were 0.9376mM acrylamide and 60.92 µmol ammonia disappeared/min respectively. The amidase exhibited maximal activity at 50° C and pH value at 7.5. The result of effects of heavy metals on amidase activity showed that AgNO3 and CuCl2 inhibited amidase activity while HBO3 enhanced amidase activity. The enzyme is a monomer with an apparent molecular weight of 111 kDa.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PENULENAN SEPARA DAN PENCIRIAN AMIDASE DARIPADA AKRILAMIDA YANG BOLEH DIURAI OLEH BAKTERIA STRAIN 30

OLEH

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Mei 2005

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Sejumlah 217 isolat bacteria telah dipencilkan menggunakan prosedur pengayaan daripada tanah-tanah di Malaysia yang telah dirawat dan tidak dirawat dengan akrilamida. Daripada keputusan penyaringan permulaan menggunakan ujian MTT, 10 isolat bakteria berupaya menggunakan akrilamida sebagai sumber nitrogen. Pembentukan warna biru formazan dan bacaan absorban tertinggi adalah kriteria dalam pemilihan bakteria. Sepuluh isolat ini adalah Isolat 5, 9, 10, 15, 30, 45, 60, 110, 115 dan 145. Isolat-isolat ini telah dilakukan penyaringan kedua menggunakan asai MTT, kaedah pengiraan koloni bakteria dan "High Perfomance Liquid Chromatography (HPLC)". Daripada keputusan yang diperolehi, lima isolat bakteria, Isolat 9, 10, 15, 30 dan 60 telah memberi bacaan tertinggi dalam pengiraan koloni bakteria dalam CFU/mL dan bacaan absorban daripada asai MTT. Kelima-lima isolat bakteria dipilih untuk analisis lanjutan degradasi akrilamida menggunakan HPLC. Isolat 30 menunjukkan degradasi akrilamida tertinggi dengan 99.84% degradasi selepas 48 jam pengeraman di dalam larutan pengayaan dengan akrilamida sebagai substrat dan kepekatan akrilamida terakhir adalah 100 mg/L. Isolat 60 menunjukkan degradasi kedua tertinggi akrilamida dengan 70.53%, diikuti dengan 50.8% degradasi



oleh Isolat 10, 16.24% dan 14.58% oleh Isolat 15 dan 9. Pengawalan tanpa bakteria menunjukkan hanya 0.16% degradasi akrilamida.

Isolat 30 dipilih untuk lanjutan optimasi media pertumbuhan termasuk enam parameter berbeza. Parameter-parameter ini adalah suhu, pH, sumber karbon berbeza, substrat amida berbeza dan kesan kepekatan berbeza bagi substrat amida terpilih. Daripada keputusan yang diperoleh, Isolat 30 menunjukkan suhu optimum pada 27°C, koloni bakteria tertinggi log₁₀ CFU/mL adalah berkadar kepada pertumbuhan bakteria. Isolat ini menunjukkan pertumbuhan bakteria tertinggi pada pH 7.5 di mana bakteria tumbuh dengan baik berhampiran sifat neutral dan sedikit media beralkali. Untuk optimasi sumber karbon, glukosa telah dipilih berdasarkan bacaan optimum koloni bakteria log₁₀ CFU/mL. Bakteria strain 30 telah dijumpai tumbuh paling baik pada kepekatan glukosa 1%. Di kalangan substrat amida, akrilamida telah ditemui sebagai satu-satunya sumber nitrogen terbaik di mana pertumbuhan bakteria dapat disokong di dalam media pengayaan dan bakteria menunjukkan pengiraan koloni

Daripada pemerhatian makroskopik dan mikroskopik ke atas Isolat 30, bakteria ini adalah gram negatif berbentuk rod, berupaya membeza kepada konveks, berpigmen dan agak legap di tengahnya dan efusi, tidak berwarna, kebanyakan jernih di pinggir dengan sisi berlekuk. Ujian Biokimia menggunakan Microbact 24E (12A+B) menunjukkan Isolat 30, yang dipencilkan dari Tanjung Karang, Selangor memberi 99.83% persamaan dengan bakteria daripada *Serratia marcescens*.

Amidase (EC 3.5.1.4) yang dihasilkan oleh Serratia marcescens strain 30 telah dilakukan penulenan separa dan pencirian. Prosedur penulenan yang digunakan termasuk kromatografi penukar ion, penukar anion- kuat Mono-QTM, penurasan ultra dan penurasan gel menghasilkan fraksi amidase separa tulen yang mempunyai keseluruhan factor penulenan melebihi 13.83 ikatan dan kadar hasil 48.33%. Kaedah "Indophenol blue" digunakan untuk kajian asai dan pencirian amidase telah dijalankan untuk kajian pencirian amidase bagi menentukan K_m dan V_{max} , kesan suhu dan pH ke atas aktiviti amidase dan kesan logam berat ke atas aktiviti amidase. Nilai K_m untuk akrilamida dapat ditentukan dengan eraman amidase pada suhu 50°C dengan kepekatan akrilamida pelbagai (0.25 hingga 20 mM) di dalam penimbal fosfat (50mM; pH 7.5). Dengan menggunakan analisis regresi tidak linear, nilai Km dan V_{mak} adalah 0.9376 mM akrilamida dan 60.92 µmol ammonia dihasilkan/ min. Kemunculan aktiviti amidase yang maksimum adalah pada 50° C dan nilai pH 7.5. Daripada keputusan kesan logam berat pada aktiviti amidase menunjukkan AgNO3 dan CuCl₂ merencat aktiviti amidase manakala HBO₃ meningkatkan aktiviti amidase. Enzim ini adalah monomer yang muncul dengan berat molekul 111 kDa.



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I certify that an Examination Committee met on 24th May 2005 to conduct the final examination of Nina Suhaity Azmi on her Master of Science thesis entitled "Isolation of *Serratia marcescens* (Isolate 30) and Partial Purification of its Acrylamide-Degrading Amidase" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

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DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

NINA SUHAITY BINTI AZMI

Date: 19th July 2005



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LIST OF ABBREVIATIONS

CFU	colony forming unit
DDT	dichlorodiphenyltrichloroethane
GPS	Global Positioning System
HPLC	High Performance Liquid Chromatography
IARC	International Agency for Research on Cancer
K _m	Michaelis Menten Constant
MARDI	Malaysian Agricultural Research and Development
	Institute
mAu*s	milliabsorbance unit
MTT	3-[4,5-dimethylthiazol-2-yl]-,5-
	diphenyltetrazoliumbromide
PAM	Polyacrylamide
PBM	Phosphate buffered medium
V _{max}	maximum initial velocity
WHO	World Health Organization



CHAPTER 1

INTRODUCTION

Manufactured chemicals present in our environment present possible health hazards. The United States Environmental Protection Agency (USEPA) categorized chemicals by their known or suspected effects as carcinogens, mutagens or as developmental, reproductive, acute, chronic or neurological toxins, and many chemicals fall into more than one of these categories. Over 10,000 pounds of carcinogens, agents known to increase cancer risks, have been released into the soil of more than 50% of the United States. Obviously, exposure to these and other chemicals released into the environment can have both acute and cumulative effects on health (Skipper and Turco, 1995).

Acrylamide is an important industrial chemical, used primarily in the production of polymers and copolymers. The widespread use and indiscriminate discharge of acrylamide and polyacrylamide has led to the contamination of soil (Lande *et al.*, 1979; Nawaz *et al.*, 1992) and plants (Nishikawa *et al.*, 1978; Nawaz *et al.*, 1992). There is abundant evidence, however, that most chemicals are degraded or dissipated in our not-so-fragile environment, despite efforts by environmental ethicists and the media to convince us otherwise (Skipper and Turco, 1995). But for most scientists



involved in reduction of environmental contaminants, there is indeed room for improvement in virtually all spheres.

The desire to minimize health effects has led to large expenditures to combat pollution. Bioremediation is one technology which shows great promise in terms of cost-effective pollution abatement. Bioremediation is the use of microorganisms or plants to detoxify an environment, mostly by transforming or degrading pollutants. The utility of bioremediation in the degradation of pollutants in the environment has been successfully demonstrated for several chemical compounds. Biodegradation is an important removal mechanism for chemicals released in large quantities to aquatic and terrestrial environments. It results in a decrease in the mass or load of chemicals present in the environment and is important in preventing the accumulation and persistence of chemicals (Shimp *et al.*, 1990; Babu *et al.*, 1996).

Biodegradation is the major route of removal acrylamide from soils. In aerobic soils, the chemical is 74- 94 % degraded in 14 days. Depending on the soil type, estimated half- lives range from 21- 36 hours. This study attempts to isolate and identify the bacteria, which utilize acrylamide as a carbon source. Microbial amidases (amidohydrolase, EC 3.5.1.4) catalyze the hydrolysis of aliphatic amides to their corresponding carboxylic acids and ammonia. Although numerous microorganisms catabolize aliphatic amides, acrylamide, because of its inhibitory effect on sulfhydryl proteins, inhibits their growth. Therefore, few microorganisms capable of degrading



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acrylamide have been isolated (Nawaz et al., 1994).

The objectives of this study are firstly, to isolate and screen acrylamide-degrading bacteria from previously treated and untreated Malaysian soil. Secondly, to identify and characterized the best acrylamide degrading bacterium by measuring acrylamide degradation by the bacterium in the shortest period and lowest residue left. Thirdly, is to partially purify the enzyme(s) involved in acrylamide degradation from the chosen bacterium.



CHAPTER 11

LITERATURE REVIEW

2.1 Acrylamide

Acrylamide (C.A.S. 79-06-1) is an odorless, free flowing white crystalline powder used as a chemical intermediate in the production and synthesis of polyacrylamides. These high-molecular weight polymers can be modified to develop nonionic, anionic, or cationic properties for specific uses. Acrylamide, an aliphatic amide, is extensively used in numerous industrial processes.

Global production of acrylamide has been estimated to be over 200,000 tons (ca. 180, 000 metric tons) (Nawaz *et al.*, 1994). Synonyms for acrylamide are acrylamide monomer, acrylic amide, propenamide, 2-propenamide, acrilamida (DOT Spanish), acrylamide (DOT French), acrylamide solution, acrylic acid amide (50%), acrylic amide (50%), ethylene carboxamide, ethylenecarboxamide, propenamide (50%), propenoic acid, amide, RCRA Waste Number U007, UN 2074, and vinyl amide (WHO, 1985).

2.2 Physical and chemical properties of acrylamide

Acrylamide occurs in crystalline form and in aqueous solution. The solid monomer is a colorless to white, free flowing crystal that is soluble in water, methanol, ethanol, dimethyl ether, and acetone, are insoluble in benzene and heptane. The chemical formula of acrylamide is C_3H_3NO and its molecular

