



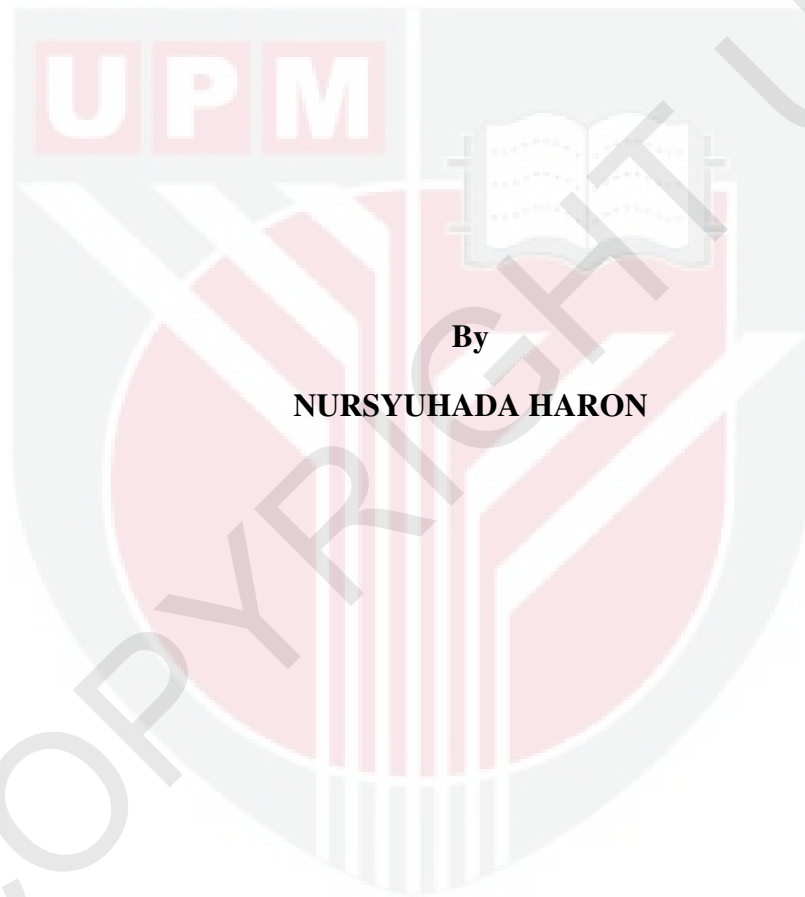
**UNIVERSITI PUTRA MALAYSIA**

***PREVENTIVE EFFECTS OF CONSUMING NUTRITIONAL SUPPLEMENT  
OF *Morinda citrifolia* L. ON BAX AND BCL-2 IN EARLY LEUKEMIC  
RATS***

**NURSYUHADA HARON**

**FPV 2012 7**

**PREVENTIVE EFFECTS OF CONSUMING NUTRITIONAL SUPPLEMENT  
OF *Morinda citrifolia* L. ON BAX AND BCL-2 IN EARLY LEUKEMIC RATS**



By  
**NURSYUHADA HARON**

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirements for the Degree of Master of Science**

**May 2012**



*This thesis is dedicated with love and gratitude to my parents,*

*Haron Marjunid and Sa'diah Saiman;*

*my brothers, Mohd Syukri Syafiq Haron and Mohd Fahrurrazi Haron;*

*my sister in law, Noor Hazilah Sukri;*

*and my dearest nephew, Muaz Mohd Syukri Syafiq*



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

**PREVENTIVE EFFECTS OF CONSUMING NUTRITIONAL SUPPLEMENT  
OF *Morinda citrifolia* L. ON BAX AND BCL-2 IN EARLY LEUKEMIC RATS**

By

**NURSYUHADA HARON**

**May 2012**

**Chairman : Hazilawati Hamzah, PhD**

**Faculty : Veterinary Medicine**

The most common cancer in Malaysia includes leukaemia and lymphoma. Cancer has been linked with the failure of apoptosis process regulated by Bcl-2 family genes, consisting of pro apoptotic genes such as Bax and anti apoptotic genes such as Bcl-2. The ratio of Bcl-2:Bax determines susceptibility towards apoptosis. Meanwhile, *Morinda citrifolia* fruit is known to exhibit anticancer properties. Thus, the objectives of this study were to determine the anticarcinogenic and preventive effects of *M. citrifolia* fruit in rats with early leukaemia/lymphoma induced by *N*-Methyl-*N*-Nitrosourea (MNU). Male Sprague Dawley rats were divided into four groups; Group 1 (control group), Group 2 (*M. citrifolia* fruit treated group), Group 3 (MNU treated group) and Group 4 (leukaemia/lymphoma prevention group treated with *M. citrifolia* fruit). The rats in the Group 3 and Group 4 were injected with four intraperitoneal injections of the

MNU mixture, at a dose of 60 mg/kg of body weight per injection, twice a week for two consecutive weeks with a total dose of 240 mg/kg of body weight. Rats in Group 1 and Group 2 were injected with normal saline following the same procedure. Ground dried *M. citrifolia* fruit at the dose of 3000 mg/kg of body weight were supplemented daily to the rats in Groups 2 and 4 for 12 weeks. Blood samples were taken at week 0, week 4 and week 8 of experimental study via cardiac puncture. At week 12, all rats were sacrificed and blood samples were taken via posterior vena cava. RNA were extracted from the blood and the relative transcription of Bcl-2 and Bax were quantified using Eva Green quantitative real time reverse transcription-PCR. For haemogram and serum biochemical analyses, blood samples were taken at week 4, week 8 and week 12 of the experimental study and analysed using automated blood analysers. For histopathological examination, organs were taken after sacrifice at the end of the experimental period. The organs were spleen, lymph nodes, liver, kidneys, lung and heart. The organs were processed using standard methods and stained with hematoxylin and eosin (H&E). Quantitative real time RT-PCR assays (qRT-PCR) for quantification of Bax and Bcl-2 genes which were successfully developed using an Eva Green dye showed that the ratio of Bcl-2:Bax in the rats injected with MNU was significantly higher compared to other groups. Significant reduction of Bcl-2:Bax ratio in Group 4 at week 12 showed that *M. citrifolia* fruit limits the progression of leukaemia/lymphoma through regulation of apoptosis gene transcription. Intraperitoneal injections of MNU at a total dose of 240 mg/kg of body weight successfully induced leukaemia without lymphocytosis in Group 3 at 50% and 75% after 8 weeks and 12 weeks of injection, respectively. Meanwhile, supplementation of dried *M. citrifolia* fruit did not significantly reduce the percentages of leukaemia without lymphocytosis in Group 4 at

20% and 60% after 8 weeks and 12 weeks of injection, respectively. All blood results obtained from the automated analysers in rats injected with MNU were similar to the control. All organs were macroscopically normal. Histopathology results showed that the rats in Group 3 developed 100%, 50% and 25% early lymphoma lesions in the spleen, mesenteric lymph nodes and other lymph nodes respectively. Meanwhile, histopathology results showed that the rats in Group 4 developed 100%, 80% and 20% early lymphoma lesions in the spleen, mesenteric lymph nodes and other lymph nodes respectively. Lesion scoring results showed that supplementation of *M. citrifolia* fruit at the dose of 3000 mg/kg of body weight did not significantly reduced the incidence of leukaemia and lymphoma Group 4.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai  
memenuhi keperluan untuk Ijazah Master Sains

**KESAN PENCEGAHAN PEMAKANAN TAMBAHAN *Morinda citrifolia* L. KE  
ATAS BAX DAN BCL-2 DALAM TIKUS LEUKEMIA AWAL**

Oleh

**NURSYUHADA HARON**

**Mei 2012**

**Pengerusi : Hazilawati Hamzah, PhD**

**Fakulti : Perubatan Veterinar**

Antara kanser yang selalu terdapat di Malaysia termasuklah leukemia dan limfoma. Kanser telah dikaitkan dengan kegagalan proses apoptosis yang di kawal atur oleh kumpulan gen-gen Bcl-2, terdiri daripada gen pro apoptotik seperti Bax dan anti apoptotik seperti Bcl-2. Kadar Bcl-2:Bax menentukan kecenderungan kepada apoptosis. Manakala buah *Morinda citrifolia* diketahui mempunyai ciri-ciri antikanser. Maka, objektif kajian ini adalah untuk menentukan kesan-kesan antikarsinogenik dan pencegahan buah *M. citrifolia* dalam tikus dengan leukemia/limfoma awal yang diaruh oleh *N*-Methyl-*N*-Nitrosourea (MNU). Tikus Sprague Dawley jantan telah dibahagikan kepada empat kumpulan; kumpulan 1 (kumpulan kawalan), kumpulan 2 (kumpulan rawatan *M. citrifolia*), kumpulan 3 (kumpulan rawatan MNU) dan kumpulan 4 (kumpulan pencegahan leukemia/limfoma dan rawatan *M. citrifolia*). Kumpulan 3 dan kumpulan 4 telah disuntik dengan empat suntikan campuran MNU secara

intraperitoneum, pada dos 60 mg/kg berat badan setiap suntikan, dua kali seminggu untuk dua minggu berturut-turut dengan jumlah dos 240 mg/kg berat badan. Tikus dalam kumpulan 1 dan kumpulan 2 disuntik dengan normal saline mengikut prosedur yang sama. Buah *M. citrifolia* yang kering pada dos 3000 mg/kg berat badan diberi makan kepada kumpulan 2 dan 4 sehingga akhir eksperimen. Sampel darah diambil pada minggu 0, minggu ke 4 dan minggu ke 8 kajian melalui tusukan jantung. Pada minggu ke 12, semua tikus dikorbankan dan sampel darah diambil melalui vena kava posterior. RNA diekstrak daripada darah dan transkripsi relatif Bcl-2 dan Bax ditentukan menggunakan PCR masa nyata. Untuk hemogram dan analisis biokimia serum, sampel darah diambil pada minggu ke 4, minggu ke 8 dan minggu ke 12 dan dianalisa menggunakan penganalisa darah automatik. Untuk pemeriksaan histopatologikal, organ-organ diambil setelah dikorbankan pada akhir tempoh eksperimen. Organ-organ yang diambil adalah limpa, nodus limfa, hati, buah pinggang, paru-paru dan jantung. Organ-organ diproses mengikut kaedah standard dan diwarnakan dengan hematoxilin dan eosin (H&E). Asai kuantitatif PCR masa nyata (qRT-PCR) untuk pengkuantitian gen Bax dan Bcl-2 yang telah berjaya dibangunkan menggunakan pewarna Eva Green menunjukkan bahawa kadar Bcl-2:Bax dalam tikus yang disuntik MNU adalah lebih tinggi ketara berbanding kumpulan lain. Penurunan ketara kadar Bcl-2:Bax dalam kumpulan 4 pada minggu ke 12 menunjukkan bahawa buah *M. citrifolia* yang kering menghalang progresi leukemia/limfoma melalui regulasi proses transkripsi gen apoptosis. Suntikan intraperitoneum MNU pada jumlah dos 240 mg/kg berat badan berjaya mengaruh leukemia tanpa limfositosis pada 50% dan 75% setelah 8 minggu dan 12 minggu menerima suntikan, masing-masing. Manakala, pemakanan tambahan buah *M. citrifolia* kering tidak memberikan penurunan ketara pada peratusan leukemia tanpa



limfositosis dalam kumpulan 4 pada 50% dan 75% setelah 8 minggu dan 12 minggu menerima suntikan, masing-masing. Keputusan analisa darah yang didapati daripada penganalisa automatik dalam kumpulan MNU tidak berbeza tererti dengan kumpulan kawalan. Tikus dalam kumpulan 3 membentuk 100% limfoma pada limpa, 50% limfoma pada limfa nodus mesentrik dan 25% limfoma pada nodus limfa yang lain. Tikus dalam kumpulan 4 pula membentuk 100% limfoma pada limpa, 80% limfoma pada limfa nodus mesentrik dan 20% limfoma pada nodus limfa yang lain. Keputusan menunjukkan bahawa penambahan buah *M. citrifolia* yang kering pada dos 3000 mg/kg berat badan tidak berjaya menurunkan ketara insiden leukemia dan limfoma.

## ACKNOWLEDGEMENT

In the name of Allah, the Most Gracious, the Merciful

Alhamdulillah, praise to Allah the Almighty, as I have finally completed this thesis. First of all, I owe my deepest gratitude to my dear supervisor, Dr. Hazilawati Hamzah, for her tremendous support, guidance and motivation given to me throughout the course of this study. I am extremely grateful for her immense knowledge, ideas, suggestions and insightful comments as it have enabled me to finish the research and thesis writing successfully.

Special thanks to Prof. Dr. Noordin Mohamed Mustapha for guiding me in this study. I would also like to show my sincere appreciation to Dr. Mohd Rosly Shaari and Dr. Shanmugavelu A/L Sithambaram for their great advice on the experimental design in animal study, and also for providing *M. citrifolia* fruit and excellent research facilities for the animal experiment at the Animal House, MARDI, Serdang. Additionally, I would also like to sincerely thank the staff of Animal House, MARDI, Serdang; Mr. Subramaniam a/l Kunjiraman, Mr. Ahmad Tarmizi Salimin, Mr. Kasambo and Mrs. Rukumani, for giving me their generous help and assistance throughout the study.

My sincere thanks and appreciation also goes to Mr. Abdullah Mison, Mr. Mohamed Halmi Othman, Miss Daarulmuqaamah Masaud, Mr. Arman Addelan and Mrs. Siti Khadijah Muhamad from the Haematology and Clinical Biochemistry Laboratory, Faculty of Veterinary Medicine, UPM and Biologic Laboratory, Faculty of Veterinary

Medicine, UPM for their help, sincere encouragement and providing me necessary facilities for the period of study.

I would also like to express my heart-felt gratitude to Dr. Tan Sheau Wei from the Molecular Medicine Laboratory, Institute of Bioscience, UPM for her sincere cooperation in providing me the essential equipment for my study and also sharing her knowledge modestly, which has help me to complete the research smoothly.

I am deeply indebted to all my labmates; Farah, Azwa, Dr. Nurul Huda, Dr. Hutheyfa, Dr. Hemn, Dr. Abdul Hamid, Yusnaini and Hakimi for continuously assisting me during the research. I am also thankful to my best friends; Syazani, Salmiah, Husna , Munira, Fajar, Nadia, Faedah, Nuyue and Yanti for always being there for me and keeps me going towards my dreams. Their endless support and encouragement has been my strength throughout my study.

I would like to thank Universiti Putra Malaysia (UPM) for providing me the Graduate Research Fellowship (GRF) and an excellent study environment, and also to the Ministry of Higher Education (MOHE) for providing the Fundamental Research Grant Scheme (FRGS) (04-04-10-912FR) for the research project.

Last but not least, I want to thank my parents and my brothers who taught me the value of hard work by their own example. They rendered me enormous support during the whole tenure of my research.

I certify that a Thesis Examination Committee has met on 31 May 2012 to conduct the final examination of Nursyuhada binti Haron on her thesis entitled "Preventive Effects of Consuming Nutritional Supplement of *Morinda citrifolia* L. on Bax and Bcl-2 in Early Leukemic rats" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the degree of Master of Science.

Members of the Examination Committee were as follows:

**Mohd Zamri Saad, PhD**

Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Chairman)

**Md. Zuki Abu Bakar, PhD**

Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Internal Examiner)

**Md Sabri Mohd Yusoff, PhD**

Senior Lecturer  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Internal Examiner)

**Mohd Effendy Abd. Wahid, PhD**

Professor  
Institute of Marine Biotechnology  
Universiti Malaysia Terengganu  
(External Examiner)

---

**ZULKARNAIN ZAINAL, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 27 AUGUST 2012

The thesis was submitted to the senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Hazilawati Hamzah, PhD**

Senior Lecturer  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Chairman)

**Noordin Mohamed Mustapha, PhD**

Professor  
Faculty of Veterinary Medicine  
University Putra Malaysia  
(Member)

**Shanmugavelu A/L Sithambaram, PhD**

Director  
Strategic Livestock Research Centre  
Malaysian Agriculture Research and Development  
(Member)

---

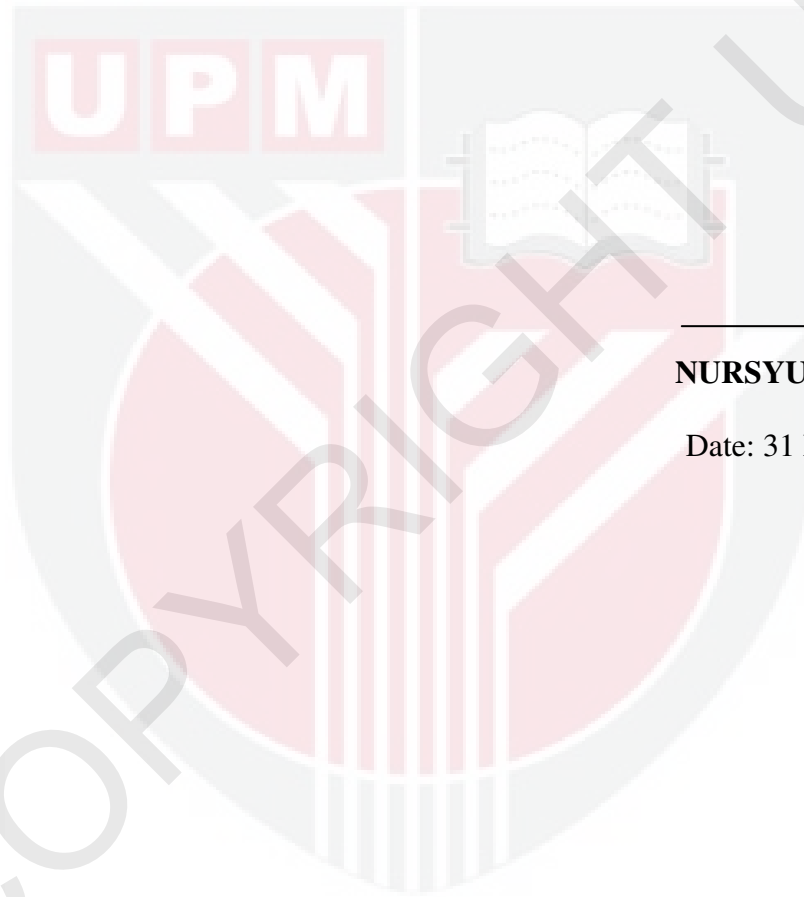
**BUJANG BIN KIM HUAT, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.



---

**NURSYUHADA HARON**

Date: 31 May 2012

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iv
<b>ACKNOWLEDGEMENT</b>	vii
<b>APPROVAL</b>	ix
<b>DECLARATION</b>	xi
<b>LIST OF TABLES</b>	xvi
<b>LIST OF FIGURES</b>	xviii
<b>LIST OF ABBREVIATIONS</b>	xxi
<b>CHAPTER</b>	
<b>1. INTRODUCTION</b>	<b>1</b>
<b>2. LITERATURE REVIEW</b>	<b>7</b>
Apoptosis	7
Caspase activation	8
Apoptotic signaling pathway	9
Bcl-2 family proteins	10
Cancer	12
Apoptosis and cancer	13
Carcinogenesis	13
Chemical carcinogenesis in animal models	15
<i>N</i> -Methyl- <i>N</i> -Nitrosourea	16
Leukaemia	17
Acute myeloid leukaemia (AML)	19
Acute lymphoid leukaemia (ALL)	19
Chronic lymphocytic leukaemia (CLL)	20
Chronic myeloid leukaemia (CML)	21
Lymphoma	22
Diagnosis of leukaemia and lymphoma	24
Haematology	24
Bone marrow cells analysis	26
Biochemical analysis	26
Histopathology of lymphoma	27
Gene expression study	28
Quantitative real time PCR assay	28
Detection chemistries of quantitative real time PCR	30

Double stranded DNA binding dyes	31
Flourescently labeled sequence-specific probes	33
Quantitative real time PCR quantification methods	33
Absolute quantification	34
Relative quantification	34
Standard curve method	35
$2^{-\Delta\Delta C_q}$ (Comparative $C_q$ ) method	36
Cancer treatment	37
Cancer chemoprevention	38
Anticancer agents from natural products	39
<i>Morinda citrifolia</i>	41
Chemical compositions of plant	42
Medicinal and anticancer properties of plant	43
<b>3. MATERIALS AND METHODS</b>	47
Animals and experimental design	47
Chemical preparation to induce leukaemia-lymphoma, dosage and administration	48
Preparation of dried <i>Morinda citrifolia</i> fruits, dosage and administration	48
Determination of average body weight and daily feed intake	50
Blood collection for RNA extractions, haemogram and serum biochemical analyses	50
Quantitative real time RT-PCR Assay of Bcl-2 and Bax genes	51
Total RNA extraction	51
Reverse transcription of RNA	53
Primers	53
Optimisations of assay	55
Determination of optimal annealing temperatures	55
Melting curve analysis	56
Detection of PCR products by gel electrophoresis	56
Development of standard curves	57
Validation of reference genes	58
Validation of the $2^{-\Delta\Delta C_q}$ (Comparative $C_q$ ) method	58
Relative Quantification of Bcl-2 and Bax genes	59
Gene transcription analysis	60
Statistical analysis	61
Blood parameters analyses	61
Haemogram	61
Serum biochemical analyses	62



Statistical analysis	63
Animal euthanasia	63
Gross and histopathological examinations of organs	63
Gross examination	63
Histopathological examination	63
Determination of lymphoma/leukaemia incidence	65
Statistical analysis	65
<b>4. RESULTS</b>	<b>67</b>
Average body weight and daily feed intake of rats	67
Development of quantitative real time RT-PCR assays for quantification of Bcl-2 and Bax mRNA transcripts in blood	71
Determination of optimal annealing temperatures	71
Development of standard curves	78
Validation of reference genes	83
Validation of the $2^{-\Delta\Delta C_q}$ (comparative $C_q$ ) method	83
Relative transcription levels of Bcl-2 and Bax transcripts in the blood	86
Bcl-2:Bax ratio	88
Haemogram and serum biochemical analyses	90
Leukon and erythron parameters	90
Serum biochemical parameters	96
Gross and histopathological examinations of organs	100
Organ weight and gross lesions	100
Histopathological findings	101
Percentages of lymphoma in organs	101
Lesion scoring for splenic lymphoma/leukaemia	102
Lesion characteristics	102
Lesion scoring results	103
Lesion scoring for nodal lymphoma	114
<b>5. DISCUSSION</b>	<b>123</b>
Average body weight and daily feed intake of rats	123
Development of quantitative real time RT-PCR assays of Bcl-2 and Bax genes	124
Relative transcription of Bcl-2 and Bax genes	128
Haemogram and serum biochemical analyses	132
Gross and histopathological examinations of organs	136

<b>6.</b>	<b>GENERAL DISCUSSION AND CONCLUSION</b>	140
	<b>REFERENCES</b>	145
	<b>APPENDICES</b>	167
	<b>BIODATA OF STUDENT</b>	175
	<b>LIST OF PUBLICATIONS</b>	175



## LIST OF TABLES

Table		Page
1	Experimental design	49
2	Sequence of primers used in the quantitative real time RT-PCR assay	54
3	Lymphoma/leukaemia lesion scoring characteristics for all organs	66
4	The average daily feed intake of rats in all groups during experimental period	69
5	The average dosage of dried <i>M. citrifolia</i> fruit consumed by the rats in Group 2 and 4	70
6	C <sub>q</sub> values at different annealing temperatures obtained from the gradient PCR	73
7	Optimal annealing temperature and melting temperature of the primers	73
8	The average C <sub>q</sub> values at different amount of templates and the $\Delta C_q$ values between target genes (Bcl-2 and Bax) and reference genes (GAPDH and Beta Actin)	84
9	The leukon parameters in all groups at week 4 of experimental period	91
10	The leukon parameters in all groups at week 8 of experimental period	91
11	The leukon parameters in all groups at week 12 of experimental period	92
12	The percentages of leukaemic rat in all groups at week 4, 8 and 12 of experimental period	92
13	The erythron parameters in all groups at week 4 of experimental period	94
14	The erythron parameters in all groups at week 8 of experimental period	94
15	The erythron parameters in all groups at week 12 of experimental period	95
16	The serum biochemical parameters of experiment groups at week 4 of experimental period	97

17	The serum biochemical parameters of experiment groups at week 8 of experimental period	98
18	The serum biochemical parameters of experiment groups at week 12 of experimental period	99
19	Relative organ weights in all groups	100
20	Lesion scores of lymphoma lesions in the haematopoietic organs of all groups analysed using Kruskal-Wallis test	112
21	Results of Mann-Whitney U test for comparisons between groups for lymphoma/ leukaemia lesions in the haematopoietic organs	113

## LIST OF FIGURES

Figure		Page
1	Typical real-time PCR results	30
2	<i>Morinda citrifolia</i> fruit	41
3	Mean body weight of rats in all groups taken every week during the experimental period	67
4	Specificity of GAPDH primer	74
5	Specificity of Beta actin primer	75
6	Specificity of Bcl-2 primer	76
7	Specificity of Bax primer	77
8	Real time RT-PCR assay for GAPDH	79
9	Real time RT-PCR assay for Beta actin	80
10	Real time RT-PCR assay for Bcl-2	81
11	Real time RT-PCR assay for Bax	82
12	Validation of the $2^{-\Delta\Delta C_q}$ (Comparative $C_q$ ) method for Bcl-2 gene	85
13	Validation of the $2^{-\Delta\Delta C_q}$ (Comparative $C_q$ ) method for Bax gene	85
14	The relative transcription levels of Bcl-2 normalised to the transcription levels of reference genes, GAPDH and Beta actin	87
15	The relative transcription levels of Bax normalised to the transcription levels of reference genes, GAPDH and Beta actin	88
16	The means ratio of Bcl-2:Bax in all groups at different week	89
17	Photomicrograph of leukaemic cells in Group 3	93
18	The percentages of lymphoma in spleen, mesenteric lymph nodes and other lymph nodes in Group 3 and Group 4 rats at week 12 of experimental period	101

19	Photomicrographs of spleen of rat necropsied at week 12 of experimental period showing population of small lymphocytes with normal and mild lesion scores	105
20	Photomicrographs of spleen of rat necropsied at week 12 of experimental period showing population of small lymphocytes with moderate lesion score	106
21	Photomicrographs of spleen of rat necropsied at week 12 of experimental period showing population of large lymphocytes in the splenic cord of the red pulp with normal and mild lesion scores	107
22	Photomicrographs of spleen of rat necropsied at week 12 of experimental period showing population of large lymphocytes in the splenic cord of the red pulp with moderate lesion score	108
23	Photomicrographs of spleen of rat necropsied at week 12 of experimental period showing population of small lymphocytes in the venous sinuses (VS) of the red pulp at the subcapsular area (SC) with normal and mild lesion scores	109
24	Photomicrographs of spleen of rat necropsied at week 12 of experimental period showing population of small lymphocytes in the venous sinuses (VS) at the red pulp of the spleen and subcapsular area (SC) of the spleen with mild and moderate lesion scores	110
25	Photomicrograph of spleen of rat necropsied at week 12 of experimental period showing the presence of pleomorphic megakaryocytes and pleomorphic neoplastic lymphocytes in the red pulp	111
26	Photomicrograph of lymph node of rat necropsied at week 12 of experimental period showing normal lesion score	115
27	Photomicrographs of lymph node of rat necropsied at week 12 of experimental period showing mild lesion score	116
28	Higher magnification of Figure 27(A)	117
29	Photomicrographs of lymph nodes of rat necropsied at week 12 of experimental period showing moderate lesion score	118
30	Higher magnification of Figure 29(A)	119
31	Photomicrograph of lymph nodes of rat necropsied at week 12 of	120

experimental period showing severe lymphoma lesion

- |    |  |     |
|----|--|-----|
| 32 | Higher magnification of Figure 31(A) in the cortical and paracortical area | 121 |
| 33 | Higher magnification of Figure 31(A) in the medullary area                 | 122 |



## LIST OF ABBREVIATIONS

%	percentage
°C	degree Celcius
Alb	albumin
ALL	Acute lymphoid leukaemia
ALP	alkaline phosphatase
ALT	alanine aminotransferase
AML	acute myeloid leukaemia
AST	alanine aminotransferase
bp	base pair
CAM	complementary and alternative medicine
CBC	complete blood count
cDNA	complementary deoxyribonucleic acid
CK	creatinine kinase
CL	chloride
CLL	chronic lymphocytic leukaemia
CML	chronic myeloid leukaemia
dH <sub>2</sub> O	distilled water
DNA	deoxyribonucleic acid
dNTP	deoxyribonucleotides
dsDNA	double stranded DNA
EDTA	ethylenediaminetetraacetic acid
HL	Hodgkin's lymphoma



IL	interleukin
K	potassium
L	litre
LDH	lactic dehydrogenase
M	molar
mA	miliampere
MC	<i>Morinda citrifolia</i>
MCH	mean corpuscular haemoglobin
MCHC	mean corpuscular haemoglobin
MCV	mean corpuscular volume
mg	miligram
MNU	<i>N</i> -Methyl- <i>N</i> -nitrosourea
mRNA	messenger ribonucleic acid
MTP	mitochondrial transmembrane potential
Na	sodium
NHL	non-Hodgkin's lymphoma
NK	natural killer
OD	optical density
PAHs	polycyclic aromatic hydrocarbons
PBS	phosphate buffered saline
PCR	polymerase chain reaction
PCR	polymerase chain reaction
PCV	packed-cell volume
pH	<i>puissance hydrogen</i>

PTCL	peripheral T-cell lymphoma
qPCR	quantitative real-time PCR
qRT-PCR	quantitative real time reverse transcriptase-polymerase chain reaction
RBC	red blood cells
RNA	ribonucleic acid
RNAase	ribonuclease
rpm	revolutions per minute
RS	Reed-Sternberg
TAE	Tris-acetate-EDTA-buffer
Taq	<i>Thermus aquaticus</i>
TNJ	Tahitian Noni Juice Liquid Dietary Supplement
ug	microgram
ul	microlitre
uM	micromolar
v/v	volume/volume
w/v	weight/volume
WBC	white blood cells
WHO	World Health Organization

## CHAPTER 1

### INTRODUCTION

*Morinda citrifolia*, or locally known as mengkudu, is believed to originate from Polynesia. Apart from growing widely in Pacific Islands such as Tahiti and Hawaii (McClatchey, 2002), this plant is also found in other tropical countries such as Malaysia and Indonesia. Nowadays, *M. citrifolia* has been grown commercially as it offered a variety of interesting medicinal values. *M. citrifolia* is used regularly as folk medicine to treat common illnesses among the Polynesians. It was reported that the fruit juice is able to stimulate significant body's immune responses and therefore was mainly consumed for health maintenance (Wang *et al.*, 2002).

Cancer is one of the major killer diseases in human, caused by an abnormal proliferation of cells in the body. The abnormal cells can block the circulations and invade other tissues thus causing devastating damage to normal organ functions which may lead to death (Bozzone, 2009). There are many types of cancer, usually named from the type of organs or cells which were involved. According to National Cancer Registry, 67,792 new cases of cancer were reported among Malaysians in Peninsular Malaysia from the year of 2003 to 2005 (Lim *et al.*, 2008).

Leukaemia, cancer of the blood cells, is one of the threatening cancers that attacked not only adult human being but the children. Leukaemia falls at the fifth place for the ten most frequent cancers among Malaysians in Peninsular Malaysia from the year of 2003

to 2005. The disease was most frequent in both males and females at the age of 0 to 14 years old with 47.6% and 45.4% of incidence respectively (Lim *et al.*, 2008). Generally, leukaemia can be divided into acute and chronic leukaemia based on the time taken for the disease to progress. Leukaemia is also further divided into lymphocytic or myeloid leukaemia based on the progenitor of the blood cells, either myeloid lineages or lymphoid lineages (Bain, 2005).

Lymphoma is a cancer of the lymphatic system that starts from the lymphocytes. In the case of lymphoma, blood cells called lymphocytes grow abnormally and proliferate continuously without control. Lymphoma usually begins in a lymph node, but it can also begin in the stomach, intestines, skin or any other organ (Chua *et al.*, 2009). According to the World Health Organization (WHO), there are three major categories of lymphoid neoplasms; B cell neoplasms, T cell and natural killer (NK) cell neoplasms and Hodgkin's lymphoma (Jaffe *et al.*, 2001). A study done by Chai *et al.* (1999) shown that the spectrum of lymphoma seen in East Malaysia was similar to West Malaysia except for the very low prevalence of peripheral T-cell lymphoma (PTCL) in Sarawak (3.3%). Another recent statistical data shown that lymphoma, along with leukaemia, is listed in the ten most frequent cancers that occur among males and females in peninsular Malaysia from the year 2003 to 2005 (Lim *et al.*, 2008).

One of the leading causes of cancer is exposure to carcinogens. Carcinogens can be defined as a group of substances that able to alter the genetic structure of the cells, thus causing the cells to multiply uncontrollably and becomes malignant cells. Epidemiological studies indicate that approximately 80% of human cancer is caused by

exposure to chemical carcinogens in tobacco smoke, in the diet, and in the work place (Srivastava, 1999). Benzene, ionizing radiation, aromatic hydrocarbons and organic solvents, electromagnetic fields, infectious agents and pesticides were suspected to induce the carcinogenesis in human (Descatha *et al.*, 2005).

*N*-Methyl-*N*-Nitrosourea (MNU) is a highly reliable carcinogen and mutagen. MNU is an alkylating agent, and exhibits its toxicity by transferring its methyl group to nucleobases in nucleic acids. Acute exposure to MNU in humans can result in skin and eye irritation, headache, nausea, and vomiting. MNU is reasonably anticipated to be a human carcinogen based on sufficient evidence of carcinogenicity in experimental animals (IARC, 1972, 1978, 1987) and was shown to be found in food products (Fiddler, 1975).

Research had shown that cancer can be linked with the failure of cells to undergo a process called apoptosis which is a process where cells were eliminated from the system. The programmed cell death was triggered when cells reach some physiologic conditions in the biological systems (Hanahan and Weinberg, 2000). Among the functions of apoptosis were to remove the abnormal and defect cells during the process of development, differentiation and proliferation of cells (Faddeel *et al.*, 1999), and to maintain the number of cells (Rathmell and Thompson, 2002). However, there are also other types of cell death that might be of importance but nevertheless, the evasion of cells from apoptosis is one of the important factors in the development of cancer.

Apoptosis is regulated by two sets of genes called pro apoptotic and anti apoptotic genes. Pro apoptotic genes such as Bax and Bak promote the process of apoptosis, while the anti apoptotic genes such as Bcl-2 and Bcl-xl inhibit the apoptosis process. The relative abundance of pro apoptotic and anti apoptotic proteins determines the susceptibility of the cell to programmed death. Research has shown that haematological malignancies such as leukaemia and lymphoma are the result of inactivation of the apoptotic genes and over expression of the pro apoptotic genes (Lowe and Lin, 2000).

Currently, the most common treatment for cancers includes surgery, chemotherapy and radiotherapy. However, some of the procedures can give unwanted side effects such as hair loss, nausea and vomiting. Thus many cancer patients decide on Complementary and Alternative Medicine (CAM) for the treatment of cancer. CAM refers to a term for medical products and practices that are not part of a standard care. Complimentary medicine is a non standard treatment used with standard treatments; meanwhile Alternative medicine refers to treatments being used instead of the standard treatments (Cassileth *et al.*, 2001).

In 1997, 43% of Americans used CAM particularly among the elders (Astin *et al.*, 2000; Flaherty *et al.*, 2001). One of the commonly used treatments for CAM is medicinal herb that has been used traditionally for treatment of diseases. Current research has shown the significant of nutritive and non nutritive plant-based dietary factors that can prevent the process of carcinogenesis effectively.

*M. citrifolia* has many potential in acting as an anticancer agent. Hirazumi and Furusawa (1999) showed that a better survival time and curative effects occurred when Noni-ppt (derived from *M. citrifolia* fruits) was combined with sub optimal doses of the standard chemotherapeutic agents such as adriamycin (adria), 5-flourouracil (5-FU), cisplatin (CDDP) and vincristine (VCR) in tumour bearing mice. The findings supported the hypothesis that *M. citrifolia* can act as a potential therapeutic agent to improve the effects of anticancer drugs by incorporating the fruits of *M. citrifolia* in patient's diet.

This research seeks to address the effects of *M. citrifolia* in the regulation of apoptotic genes as the failure of cells to undergo apoptosis process which contribute to the development of leukaemia/lymphoma. Based on the current issues to reduce the incidence of cancer and search for a potential anti cancer agent, it is worth looking on natural sources especially from local source as a promising approach towards the prevention of cancer. Therefore, the present study hypothesizes that *M. citrifolia* fruit will prevent tumourigenesis induced by MNU via down regulating the transcription of anti apoptic gene, Bcl-2 and up regulating the transcription of pro apoptotic gene, Bax, thus decreasing the ratio of Bcl-2:Bax transcripts.

The anticarcinogenic effects of *M. citrifolia* fruit in rats with early leukaemia/lymphoma induced using MNU were investigated through these following objectives:

1. to evaluate the effects of *M. citrifolia* fruit in alleviating the carcinogenic effects of MNU in rats via haemogram, serum biochemistry and histopathological examination of the organs, and

2. to investigate the effects of *M. citrifolia* fruit in regulating the transcription of Bax and Bcl-2 in early leukaemic rats by determining the ratio of Bcl-2:Bax mRNA transcripts in the blood.





## REFERENCES

- Abbott, I.A. (1992). *La'au Hawai'i: Traditional Hawaiian Uses of Plants*. Honolulu, Hawaii: Bishop Museum Press.
- Abbott, I.A. and Shimazu, C. (1985). The geographic origin of the plants most commonly used for medicine by Hawaiians. *Journal of Ethnopharmacology*. **14**: 213-222.
- Abdel-Rahman, M.K. (2011). Can apricot kernels fatty acids delay the atrophied hepatocytes from progression to fibrosis in dimethylnitrosamine (DMN)-induced liver injury in rats? *Lipids Health*. **10**: 114.
- Adams, J.M. (2003). Ways of dying: multiple pathways to apoptosis. *Genes and Development*. **17**: 2481-2495.
- Adams, J.M. and Cory, S. (2002). The Bcl-2 family: Regulators of the cellular life-or-death switch. *Nature Reviews Cancer*. **2**: 647-656.
- Akihisa, T., Yasukawa, K. and Tokuda, H. (2003). Potentially cancer chemopreventive and anti-inflammatory terpenoids from natural sources. In Rahman, A. *Studies in Natural Products Chemistry* (pp.73-126). Amsterdam: Elsevier.
- Alvero, A.B. and Mor, G. (2008). Apoptosis and cancer: methods and protocols. In F. Richard, and M. Aubert. *Flow cytometric detection of caspase-3*. (pp.47-57). New Jersey: Humana Press Inc.
- Anetor, J.I., Anetor, G.O., Udah, D.C. and Adeniyi, F.A.A. (2008). Chemical carcinogenesis and chemoprevention: Scientific priority area in rapidly industrializing developing countries. *African Journal of Environmental Science and Technology*. **2**: 150-156.
- Appelbaum, F.M. (2007). The acute leukaemias. In L. Goldman, and D. Ausiell. *Cecil Medicine* (pp194). Philadelphia: Saunders Elsevier.
- Assis, G.F., Ceolin, D.S., Marques, M.E., Salvadori, D.M. and Ribeiro, D.A. (2005). Cigarette smoke affects apoptosis in rat tongue mucosa: role of bcl-2 gene family. *Journal of Molecular Histology*. **36**: 486-489.
- Astin, J.A., Pelletier, K.R., Marie, A. and Haskell, W.L. (2000). Complementary and alternative medicine use among elderly persons: one-year analysis of a Blue Shield Medicare supplement. *The Journals of Gerontology Series A: Biological Sciences*. **55**: M4-9.
- Bain, B.J. (2005). Diagnosis from the blood smear. *The New England Journal of Medicine*. **353**: 498-507.

- Balakrishnan, S., Seshadri, T.R. and Venkataramani, B. (1961). Special chemical component of commercial woods and related plant materials: path X- Heart wood of *Morinda citrifolia*, Linn. *Journal of Scientific and Industrial Research*. **20**: 331-333.
- Banudevi, S., Elumalai, P., Arunkumar, K., Gunadharini, D.N., Sharmila, G. and Arunakaran, J. (2011). Chemopreventive effects of zinc on prostate carcinogenesis induced by *N*-methyl-*N*-nitrosourea and testosterone in adult male Sprague-Dawley rats. *Journal of Cancer Research and Clinical Oncology*. **137**: 677-86.
- Bengtsson, M., Karlsson, H.J., Westman, G. and Kubista, M. (2003). A new minor groove binding asymmetric cyanine reporter dye for real-time PCR. *Nucleic Acids Research*. **31**: 45.
- Beremblum, I. and Shubik, P. (1947). The role of croton oil applications, associated with a single painting of a carcinogen, in tumour induction of the mouse's skin. *British Journal of Cancer*. **1**: 383-391.
- Bertram, J.S. (2001). The molecular biology of cancer. *Molecular Aspects of Medicine*. **21**: 167-223.
- Besa, E.C. and Woermann, E.C. (2010). *Chronic Myelogenous Leukaemia*. Retrieved October 2011, from <http://emedicine.medscape.com/article/199425-overview>.
- Birgegård, G., Gascon, P. and Ludwig, H. (2006). Evaluation of anaemia in patients with multiple myeloma and lymphoma: findings of the European cancer anaemia survey. *European Journal of Haematology*. **77**: 378-86.
- Bishop, J.M. (1987). The molecular genetics of cancer. *Science*. **235**: 305-311.
- Boatright, K.M. and Salvesen, G.S. (2003). Mechanisms of caspase activation. *Current Opinion in Cell Biology*. **15**: 725-731.
- Bollag, W. (1970). Vitamin A and vitamin A acid in the prophylaxis and therapy of epithelial tumours. *Internationale Zeitschrift für Vitamin for schung*. **40**: 299-314.
- Boone, C.W. and Wattenberg, L.W. (1994). Current Strategies of Cancer Chemoprevention: 13th Sapporo Cancer Seminar. *Cancer Research*. **54**: 3315-3318.
- Bozzone, D.M. (2009). *Cancer genetics (the biology of cancer)*. New York: Chelsea House Publications.
- Brambrink, M.A., Schneider, A., Noga, H., Astheimer, A., Gotz, B., Korner, I., Heimann, A., Welschof, M. and Kempfski, O. (2000). Tolerance-inducing dose of 3-nitropropionic acid modulates bcl-2 and bax balance in the rat brain: a potential

mechanism of chemical preconditioning. *Journal of Cerebral Blood Flow and Metabolism*. **20**: 1425-1436.

Budavari, S.O., Neil, M.J., Smith, A. and Heckelman, P.E. (1989). *The Merck Index. An Encyclopedia of Chemicals, Drugs and Biologicals*. Rathway, New Jersey: Merck & Co Inc.

Bustin, S.A. (2000). Absolute quantification of mRNA using real-time reverse transcription polymerase chain reaction assays. *Journal of Molecular Endocrinology*. **25**: 248-256.

Bustin, S.A., Benes, V., Garson, J.A., Helleman, J., Hugget, J., Kubista, M., Mueller, R., Nolan, T., Pfaffl, M.W., Shipley, G.L., Vandesompele, J. and Wittwer, C.T. (2009). The MIQE guidelines: minimum information for publication of quantitative real-time PCR experiments. *Clinical Chemistry*. **55**: 4.

Calabrese, E.J. (2001). Apoptosis: biphasic dose responses. *Critical Reviews in Toxicology*. **31**: 607-613.

Cassileth, B.R., Schraub, S., Robinson, E. and Vickers, A. (2001). Alternative medicine use worldwide. *Cancer*. **91**: 1390-1393.

Chabner, B.A. and Longo, L. (2005). *Cancer chemotherapy and biotherapy: principles and practice*. Baltimore: Lippincott Williams and Wilkins.

Chai, S. P., Peh, S. C., Kim, L. H., Lim, M. Y. and Gudum, H. R. (1999). The pattern of lymphoma in east Malaysian patients as experienced in the University Hospital, Kuala Lumpur. *The Malaysian Journal of Pathology*. **21**: 45-50.

Chamilova, M., Kubatka, P., Kalicka, K., Adamekova, E., Bojkova, B., Ahlers, I., Ahlersova, E. (2002). Chemoprevention of *N*-Methyl-*N*-Nitrosourea Induced Mammary Carcinogenesis with Raloxifene and Melatonin: Metabolic Changes in Female Rats. *Acta Veterinaria Borneo*. **71**: 235-242.

Chang, Y-C., Hsu, J-D., Lin, W-L., Lee, Y-J. and Wang, C-J. (2012). High incidence of acute promyelocyticleukemiaspecifically induced by *N*-nitroso-*N*-methyl-urea (NMU) in Sprague Dawley rats. *Archives of Toxicology*. **86**: 315-327.

Chen, J., Rider, D.A. and Ruan, R. (2006). Identification of valid housekeeping genes and antioxidant enzyme gene expression change in the aging rat liver. *Journal of Gerontology: Biological Sciences*. **21A**: 20-27.

Cheng, J., Yim, O-S., Low, P-S., Tay, S.K.H., Yap, E.P.H. and Lai, P-S. (2011). Detection of hemi/homozygotes through heteroduplex formation in high-resolution melting analysis. *Analytical Biochemistry*. **410**: 158-160.

Chial, H. (2008). Proto-oncogenes to oncogenes to cancer. *Nature Education*. **1**: 1.

- Chua, S.C., Rozalli, F.I. and O'Connor, S.R. (2009). Imaging features of primary extranodal lymphomas. *Clinical Radiology*. **64**: 574-588.
- Čikoš\*, Š., Bukovská, A. and Koppel, J. (2007). Relative quantification of mRNA: comparison of methods currently used for real-time PCR data analysis. *BMC Molecular Biology*. **8**: 113.
- da Silva Franchi, C.A., Bacchi, M.M., Padovani, C.R. and Camargo, J.L.V. (2003). Thymic lymphomas in Wistar rats exposed to *N*-methyl-*N*-nitrosourea (MNU). *Cancer Science*. **94**: 240-243.
- Danial, N.N. and Korsmeyer, S.J. (2004). Cell death: critical control points. *Cell*. **116**: 205–219.
- Davis, C.D. and Milner, J. (2004). Frontiers in nutrigenomics, proteomics, metabolomics and cancer prevention. *Mutation Research*. **551**: 51-64.
- Deprez, R.H.L., Fijnvandraat, C.A., Ruijter, J.M. and Moorman, A.F.M. (2001). Sensitivity and accuracy of quantitative real-time polymerase chain reaction using SYBR green I depends on cDNA synthesis conditions. *Analytical Biochemistry*. **307**: 63–69.
- Derveaux, S., Vandesompele, J. and Hellemans, J. (2010). How to do successful gene expression analysis using real-time PCR. *Methods*. **50**: 227-230.
- Descatha, A., Jenabian, A., Conso, F. and Ameille, J. (2005). Occupational Exposures and Haematological Malignancies: Overview on Human Recent Data. *Cancer Causes and Control*. **16**: 39-953.
- DeVita, V.T., Hellman, S. and Rosenberg, S. (1997). *Cancer Principles and Practice of Oncology, 5th edition*. Philadelphia: Lippincot-Raven Publishers.
- Dighiero, G. and Hamblin, T.J. (2008). Chronic lymphocytic leukaemia. *Lancet*. **371**: 1017-1029.
- Dixon, A.R., McMillen, H., Etkin, N.L. (1999). Ferment this: the transformation of Noni, a traditional Polynesian medicine (*Morinda citrifolia*, Rubiaceae). *Ecological Botony*. **53**: 51-68.
- Doctors, C.V., Rizzari, C., Sala, A., Chiesa, R., and Citterio, M. (2004). *Acute Lymphoblastic Leukaemia*. Retrieved August 2011, from <http://www.oncologychannel.com/leukaemias/diagnosis.html>
- Drake, M., Kettle, P., Lynch, T., Morris, C., Page, W., Shiels, A. and Venkatramen, L. (2007). The Role of PET/CT in Lymphoma Part I. *Imaging*. **1**: 16-19.
- Dualatabad, C.D., Mulla, G.M. and Mirajikar, A.M. (1989). Riconoleic acid in *Morinda citrifolia* seed oil. *Oil Technologist's Association of India*. **21**: 26-27.

- Dussault, A.A. and Pouliot, M. (2006). Rapid and simple comparison of messenger RNA levels using real-time PCR. *Biological Procedures Online*. **8**: 1-10.
- Eichhorst, B. and Hallek, M. (2007). Revision of the guidelines for diagnosis and therapy of chronic lymphocytic leukaemia (CLL). *Best Practice and Research Clinical Haematology*. **20**: 469-477.
- Elter, T., Hallek, M. and Engert, A. (2006). Fludarabine in chronic lymphocytic leukaemia. *Expert Opinion on Pharmacotherapy*. **7**: 1641-1651.
- Ernst, E. (1995). Complementary cancer treatments: Hope or hazard? *Journal of Clinical Oncology*. **7**: 259-263.
- Ernst, E., Resch, K.L., Mills, S., Hill, R., Mitchell, A., Willoughby, M. and White, A. (1995). Complementary medicine - a definition. *British Journal of General Practice*. **45**: b506-506.
- Fadeel, B., Gleiss, B., Hogstrand, K., Chandra, J., Wiedmer, T., Sims, P.J., Henter, J.I., Orrenius, S. and Samali, A. (1999). Phosphatidylserine exposure during apoptosis is a cell-type-specific event and does not correlate with plasma membrane phospholipid scramblase expression. *Biochemical and Biophysical Research*. **266**: 504-511.
- Falleni, M., Pellegrini, C., Marchetti, A., Roncalli, M., Nosotti, M. and Paleschi, A., Santambrogio, L., Coggi, G. and Bosari, S. (2005). Quantitative evaluation of the apoptosis regulating genes Survivin, Bcl-2 and Bax in inflammatory and malignant pleural lesions. *Lung Cancer*. **48**: 211-216.
- Farber, J.L. (1982). Biology of disease: membrane injury and calcium homeostasis in the pathogenesis of coagulative necrosis. *Laboratory Investigation*. **47**: 114-123.
- Farine, J.P., Legal, L., Moreteau, B. and Le Quere, J.L. (1996). Volatile components of ripe fruits of *Morinda citrifolia* and their effects on *Drosophila*. *Phytochemistry*. **41**: 433-438.
- Feher, M. and Schmidt, J.M. (2003). Property distributions: Differences between drugs, natural products, and molecules from combinatorial chemistry. *Journal of Chemical Information and Modeling*. **43**: 218-227.
- Ferlay, J., Shin, H.-R., Bray, F., Forman, D., Mathers, C. and Parkin, D. M. (2010). Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *International Journal of Cancer*. **127**: 2893-2917.
- Fiddler, W. (1975). The occurrence and determination of n-nitroso compounds. *Toxicology and Applied Pharmacology*. **31**: 352-360.

- Flaherty, J.H., Takahashi, R., Teoh J., Kim, J.I., Habib, S., Ito, M. and Matsushita S. (2001). Use of alternative therapies in older outpatients in the United States and Japan: prevalence, reporting patterns, and perceived effectiveness. *The Journals of Gerontology Series A: Biological Sciences and Medical Sciences*. **56**: M650-655.
- Fleige, S. and Pfaffl, M.W. (2006). RNA integrity and the effect on the real-time qRT-PCR performance. *Molecular Aspects of Medicine*. **27**: 126-139.
- Fong, S.T., Johnson, A., Ho, C-T. and Csiszar, K. (2001). *Extracts of Morinda citrifolia (noni) exhibit selective anti-tumor activity against breast and colon carcinoma cell lines*. Poster presented at: Building Bridges with Traditional Knowledge Summit, Honolulu, Hawaii.
- Fu, L.Y., Jia, H.L., Dong, Q.Z., Wu, J.C., Zhao, Y., Zhou, H.J., Ren, N., Ye, Q.H. and Lun, X.Q. (2009). Suitable reference genes for real-time PCR in human HBV-related hepatocellular carcinoma with different clinical prognoses. *BMC Cancer*. **9**: 49.
- Galiveti, C.R., Rozhdestvensky, T.S., Brosius, J., Lehrach, H. and Konthur, Z. (2010). Application of housekeeping npcRNAs for quantitative expression analysis of human transcriptome by real-time PCR. *RNA*. **16**: 450-461.
- Garber, M.E., Troyanskaya, O.G., Schluens K., Petersen, S., Thaesler, Z., Pacyna-Gengelbach, M., van de Rijn, M., Rosen, G.D., Perou, C.M., Whyte, R.I., Altman, R.B., Brown, P.O., Botstein, D. and Petersen, I. (2001). Diversity of gene expression in adenocarcinoma of the lung. *Proceedings of the National Academy of Sciences of the United States of America*. **98**: 13784-13789.
- Gentle, A., Anastasopoulos, F. and McBrien, N. A. (2001). High-resolution semi-quantitative real-time PCR without the use of a standard curve. *BioTechniques*. **31**: 502-508.
- Gerlach, J. (1996). Native or introduced plant species? *Phelsuma*. **4**: 70-74.
- Giboney, P.T. (2005). Mildly Elevated Liver Transaminase Levels in the Asymptomatic Patient. *American Family Physician*. **71**: 1105-1110.
- Giglio, S., Monis, P. T. and Saint, C. P. (2003). Demonstration of preferential binding of SYBR Green I to specific DNA fragments in real-time multiplex PCR. *Nucleic Acids Research*. **31**: 136.
- Gonzalez, M.S., De Brasia, C.D., Bianchinia, M., Gargalloa, P., Moiraghib, P., Bengióa, R. and Larripaa, I.B. (2010). BAX/BCL-XL gene expression ratio inversely correlates with disease progression in chronic myeloid leukaemia. *Blood Cells, Molecules, and Diseases*. **45**: 192-196.
- Granziero, L., Ghia, P., Circosta, P., Gottardi, D., Strola, G., Geuna, M., Montagna, L., Piccoli, P., Chilosi, M., and Caligaris-Cappio, F. (2001). Survivin is expressed on

CD40 stimulation and interfaces proliferation and apoptosis in B-cell chronic lymphocytic leukaemia. *Blood*. **97**: 2777-2783.

Green, D.R. and Evan, G.I. (2002). A matter of life and death. *Cancer Cell*. **1**: 19-30.

Green, D.R. and Reed, J.C. (1998). Mitochondria and apoptosis. *Science*. **281**: 1309-1312.

Greer, J.P. and Wintrobe, M.M. (2008). *Wintrobe's clinical hematology*. Philadelphia: Lippincott Williams & Wilkins.

Guppy, H.B. (1917). *Plants, seeds and currents in the West Indies and Azores*. Covent Garden, London, England: Williams & Norgate.

Hanahan, D. and Weinberg, R.A. (2000). The hallmarks of cancer. *Cell*. **100**: 57-70.

Hancock, B.W., Selby, P.J., MacLennan, K., Armitage, J.O. (2000). *Malignant lymphoma*. London: Arnold Publishers.

Hazilawati, H., Huthyfa, A.H., Rosly, S.M., Jasni, S., Noordin, M.M. and Shanmugavelu, S. (2010). Haematological parameters of leukaemic rats supplemented with *Morinda citrifolia*. *The Malaysian Medical Journal*. **65**: 125-126.

Hehlmann, R., Hochhaus, A. and Baccarani, M. (2007). Chronic myeloid leukaemia. *Lancet*. **370**: 342-350.

Heid, C. A., Stevens, J., Livak, K. J. and Williams, P. M. (1996). Real time quantitative PCR. *Genome Research*. **6**: 986-994.

Heinicke, R. (1985). The pharmacologically active ingredients of Noni. *Bulletin of the National Tropical Botanical garden*.

Hengartner, M.O. (2000). The biochemistry of apoptosis. *Nature*. **407**: 770-776.

Hengartner, M.O. and Horvitz, H.R. (1994). *C. elegans* cell survival gene ced-9 encodes a functional homolog of the mammalian proto-oncogene bcl-2. *Cell*. **76**: 665-76.

Henkel, T., Brunne, R.M., Muller, H. and Reichel, F. (1999). Statistical investigation into the structural complementarity of natural products and synthetic compounds. *Angewandte Chemie International Edition*. **38**: 43-647.

Heywood, R. (1983). Long term toxicity. In M. Balls, R.J. Riddell and A.N. Worden (Eds.), *Animals and alternatives in toxicity testing* (pp. 79-89). London: Academic Press.

Higuchi, R., Dollinger, G., Walsh, P. S. and Griffith, R. (1992). Simultaneous amplification and detection of specific DNA sequences. *Biotechnology (NY)*. **10**: 413-417.

- Higuchi, R., Fockler, C., Dollinger, G. and Watson, R. (1993). Kinetic PCR analysis: real-time monitoring of DNA amplification reactions. *Biotechnology (NY)*. **11**: 1026-1030.
- Hingorjo, M.R. and Syed, S. (2008). Presentation, staging and diagnosis of lymphoma: a clinical perspective. *Journal of Ayub Medical College Abbottabad*. **20**: 100-103.
- Hiramastu, T., Imoto, M., Koyano, T. and Umezawa, K. (1993). Induction of normal phenotypes in ras transformed cells by damnacanthol from *Morinda citrifolia* L. *Cancer letters*. **73**: 161-166.
- Hirazumi, A. and Furusawa, E. (1999). An Immunomodulatory polysaccharide rich substance from the fruit juice of *Morinda citrifolia* (Noni) with antitumour activity. *Phytotherapy Research*. **13**: 380-387.
- Hirazumi, A., Furusawa, E., Chou, S.C. and Hokama, Y. (1996). Immunomodulation contributes to the anticancer activity *Morinda citrifolia* (Noni) fruit juice. *Proceedings of the Western Pharmacology Society*. **39**: 7-9.
- Hirazumi, A., Furusawa, E., Chou, S.C. and Hokama, Y. (1994). Anticancer activity of *Morinda citrifolia* (Noni) on intraperitoneally implanted Lewis lung carcinoma in syngenic mice. *Proceedings of the Western Pharmacology Society*. **37**: 145-146.
- Hiwasa, T., Arase, Y., Chen, Z., Kita, K., Umezawa, K., Ito, H. and Suzuki, N. (1999). Stimulation of ultraviolet-induced apoptosis of human fibroblast UVR-1 cells by tyrosine kinase inhibitors. *FEBS Letters*. **444**: 173-176.
- Hoffman, R., Furie, B., Benz, E.J., McGlave, P., Silberstein, L.E. and Shattil, S.J. (2008). *Hematology: Basic Principles and Practice*. Philadelphia: Elsevier Churchill Livingstone.
- Hong, W.K. and Lippman, S.M. (1995). Cancer chemoprevention. *Journal of The National Cancer Institute Monographs*. **17**: 49-53.
- Hornick, C.A., Myers, A., Halina, S.K., Anthony, C.T. and Woltering, E.A. (2003). Inhibition of angiogenic initiation and disruption of newly established human vascular networks by juice from *Morinda citrifolia* (noni). *Angiogenesis*. **6**: 143-149.
- HSDB.(2001). Hazardous Substances Data Base. National Library of Medicine. Retrieved Mei 2011, from <http://toxnet.nlm.nih.gov/cgi-bin/sis/htmlgen?HSDB>.
- Hugget, J., Dheda, K., Bustin, S. and Zumla, A. (2005). Real-time RT-PCR normalization; strategies and considerations. *Genes and Immunity*. **6**: 279-284.
- Huggins, C.B., Grand, L. and Ueda, N. (1982). Specific induction of erythroleukaemia and myelogenous leukaemia in Sprague-Dawley rats. *Proceedings of the National Academy of Sciences of the United States of America*. **79**: 5411-5414.



- Hutheyfa, A.H., Hazilawati. H., Rosly S.M., Jasni, S., Noordin, M.M. and Shanmugavelu, S. (2011). Histopathological features of peripheral T-cell lymphoma in Sprague Dawley rats induced with *N*-methyl-*N*-nitrosourea. *Pertanika Journal of Tropical Agricultural Science*. **34**: 351–361.
- Hutheyfa. (2011). Master thesis, Universiti Putra Malaysia.
- IARC. (1972). Some Inorganic Substances, Chlorinated Hydrocarbons, Aromatic Amines, *N*-Nitroso Compounds and Natural Products. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, vol. 1. Lyon, France: International Agency for Research on Cancer. pp. 184.
- IARC. (1978). Some *N*-Nitroso Compounds IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, vol. 17. Lyon, France: International Agency for Research on Cancer. pp. 365.
- IARC. (1987). Overall Evaluations of Carcinogenicity. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, supplement 7. Lyon, France: International Agency for Research on Cancer. pp. 440.
- Ihrig, J., Lill, R., Muhlenhoff, U. (2006). Application of the DNA-specific dye Eva Green for the routine quantification of DNA in microplates. *Analytical Biochemistry*. **359**: 265-267.
- Jäättelä, M. (1999). Escaping cell death: survival proteins in cancer. *Experimental Cell Research*. **248**: 30-43.
- Jaffe, E.S. (1998). Histopathology of the non-Hodgkin's lymphomas and Hodgkin's disease. In G. P. Canellos, T.A. Lister and J. L. Sklar (Eds.), *The Lymphomas* (pp77-106). Philadelphia: WB Saunders Co.
- Jaffe, E.S., Harris, N.L., Stein, H. and Vardiman, J.W. (2001). *World Health Organization Classification of Tumours, Pathology and Genetics of Tumours of Haematopoietic and Lymphoid Tissues*. Lyon: IARC Press.
- Jain, M., Nijhawan, A., Tyagi, A.K. and Khurana, J.P. (2006). Validation of housekeeping genes as internal control for studying gene expression in rice by quantitative real-time PCR. *Biochemical and Biophysical Research Communications*. **345**: 646-651.
- Jain, R.K. and Srivastava, S.D. (1992). Two new anthraquinones in the roots of *Morinda citrifolia*. *Proceedings of the National Academy of Sciences of India*. **62A**: 11-13.
- Jordan, J.J. (2000). Real-time detection of PCR products and microbiology. *New Technologies for Life Sciences: A Trends Guide*. **8**: 61-66.
- Joshi, V V. and Frei, J.V. (1970). Effects of dose and schedule of methyl nitrosourea on incidence of malignant lymphoma in adult female mice. *Journal of the National Cancer Institute*. **45**: 335.

- Kang, C.D., Jang, J.H., Kim, K.W., Lee, H.J., Jeong, C. S., Kim, C.M., Kim, S.H. and Chung, B.S. (1998). Activation of c-jun N-terminal kinase/stress-activated protein kinase and the decreased ratio of Bcl-2 to Bax are associated with the auto-oxidized dopamine-induced apoptosis in PC12 cells. *Neuroscience Letters*. **256**: 37-40.
- Kantarjian, H. and O'Brien, S. (2007). The chronic leukaemias. In L. Goldman and D. Ausiello (Eds), *Cecil Textbook of Medicine*. Philadelphia: Saunders Elsevier.
- Kapoor, M. and Chan, G.Z. (2001). Malignancy and renal disease. *Critical Care Clinics*. **17**: 571-598.
- Karsai, A., Muller, S., Platz, S. and Hauser, M. T. (2002). Evaluation of a homemade SYBR green I reaction mixture for real-time PCR quantification of gene expression. *Biotechniques*. **32**: 790-792.
- Kas, K. (2004). On the technicalities of discovering and applying protein biomarkers for cancer prevention. *European Journal of Cancer Prevention*. **13**: 437-446.
- Kerr, J.F.R., Wyllie, A.H. and Currie, A.R. (1972). Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *British Journal of Cancer*. **26**: 239-257.
- Kiensey, B. (2012). *Morinda citrifolia*-noni. Hawaiian Plants and Tropical Flowers: A Guide to the Flowers and Plants of Hawaii. Retrieved July 7, 2012, from <http://wildlifeofhawaii.com/flowers/518/morinda-citrifolia-noni>
- Kjeldsberg, C. and Perkins, S.L. (2009). *Practical Diagnosis of Hematologic Disorders*. Chicago: American Society for Clinical Pathology (ASCP) Press.
- Klein, D. (2002). Quantifications using real-time PCR technology: applications and limitations. *Trends in molecular medicine*. **8**: 257-260.
- Klein, D., Leutenegger, C. M., Bahula, C., Gold, P., Hofmann-Lehmann, R., Salmons, B., Lutz, H. and Gunzburg, W. H. (2001). Influence of preassay and sequence variations on viral load determination by a multiplex real-time reverse transcriptase-polymerase chain reaction for feline immunodeficiency virus. *Journal of Acquired Immune Deficiency Syndromes*. **26**: 8-20.
- Koestner, A.W., Ruecker, F.A., and Koestner, A. (1977). Morphology and pathogenesis of tumors of the thymus and stomach in Sprague-Dawley rats following intragastric administration of MNU. *International Journal of Cancer*. **20**: 418-426.
- Kolonel, L.N., Altshuler, D. & Henderson, B.E. (2004). The multiethnic cohort study: exploring genes, lifestyle and cancer risk. *Nature Reviews Cancer*. **4**: 519-527.

- Korsmeyer, S. J., Shutter, J. R., Veis, D. J., Merry, D. E. and Oltvai, Z. N. (1993). Bcl-2/Bax: a rheostat that regulates an anti-oxidant pathway and cell death. *Seminars in Cancer Biology*. **4**: 327-332.
- Krajewski, S., Blomqvist, C., Franssila, K., Krajewska, M., Wasenius, V-M., Niskanen, E. and Reed, J. (1995). Reduced expression of pro-apoptotic gene Bax is associated with poor response rates to combination chemotherapy and shorter survival in women with metastatic breast adenocarcinoma. *Cancer Research*. **55**: 4471-4478.
- Krajewski, S., Krajewska, M., Shabaik A., Miyashita, T., Wang, H. G. and Reed, J. C. (1994). Immunohistochemical determination of *in vivo* distribution of Bax, a dominant inhibitor of Bcl-2. *American Journal of Pathology*. **145**: 323-336.
- Krauss, B.H. (1993). *Plants in Hawaiian Culture*. Honolulu, Hawaii: The University of Hawaii Press.
- Kristinsson, S.Y., Dickman, P.W., Wilson, W.H., Caporaso, N., Björkholm, M. and Landgren, O. (2009). Improved survival in chronic lymphocytic leukaemia in the past decade: a population-based study including 11,179 patients diagnosed between 1973-2003 in Sweden. *Haematologica*. **94**: 1259-1265.
- Kroemer, G, Zamzami, N and Susin, SA (1997). Mitochondrial control of apoptosis. *Immunology Today*. **18**: 44-51.
- Kuhne, B. S. and Oschmann, P. (2002). Quantitative real-time RT-PCR using hybridization probes and imported standard curves for cytokine gene expression analysis. *Biotechniques*. **33**: 1080-1084.
- Lanasa, M.C. (2010). Novel insights into the biology of CLL. *Hematology*. **2010**: 70-76.
- Lang-rollin, I. 2005. Apoptosis and conformational change of Bax induced by proteosomal inhibition of PC12 cells are inhibited by bcl-xL and bcl-2. *Apoptosis*. **10**: 809-820.
- Larionov, A., Krause, A. and Miller, W. (2005). A standard curve based-method for relative real time PCR data processing. *BMC Bioinformatics*. **6**: 62.
- Lee, T.H., Kim, W.R., Benson, J.T., Therneau, T.M. and Melton, L.J. (2008). Serum aminotransferase activity and mortality risk in a United States community. *Hepatology*. **47**: 880-887.
- Legal, L., David, J.R. and Jallon, J.M. (1994). Molecular Basis *Morinda citrifolia* (L): toxicity on drosophila. *Journal of Chemical Ecology*. **20**: 1931-1943.
- Letilovic, T., Vrhovac, R., Verstovsek, S., Jaksic, B. and Ferrajoli A. (2006). Role of angiogenesis in chronic lymphocytic leukaemia. *Cancer*. **107**: 925-934.

- Levand, O. and Larson, H.O. (1979). Some chemical constituents of *Morinda citrifolia*. *Planta Medica*. **36**: 186–187.
- Lim, G.C.C., Rampal, Y. and Halimah, Y. (2008). Cancer Incidence in Peninsular Malaysia, 2003 - 2005. National Cancer Registry, Kuala Lumpur.
- Lippman, S. M. and Hong, W. K. (2002). Cancer prevention by delay. *Clinical Cancer Research*. **8**: 314–346.
- Liss, B. and Roeper, J. (2004). Correlating function and gene expression of individual basal ganglioneurons. *TRENDS in Neurosciences*. **27**: 475-481.
- Liu, D., Pan, F., Li, B., Han, X., Li, W., Shi, Y., Pang, Z. and Zhang, Q. (2011). Intervention of nicotine on MNU-induced bladder cancer in rats. *Journal Of Huazhong University of Science and Technology (Medical Sciences)*. **31**: 103-106.
- Liu, G., Bode, A., Ma, W.Y, Sang, S., Ho, C.T. and Dong, Z. (2001). Two novel glycosides from fruits of *Morinda citrifolia* (Noni) inhibit AP-1 trans activation and cell transformation in the mouse epidermal JB6 cell line. *Cancer Research*. **61**: 5749-5756.
- Liu, W. and Saint, D.A. (2002). A new quantitative method of real time reverse transcription polymerase chain reaction assay based on simulation of polymerase chain reaction kinetics. *Analytical Biochemistry*. **302**: 52-59.
- Livak, K. (1997). ABI Prism 7700 Sequence Detection System, User Bulletin 2. PE Applied Biosystems, Foster City, CA.
- Livak, K.J. and Schmittgen, T.D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods*. **25**: 402-408.
- Lowe, S.W. and Lin, A.W. (2000). Apoptosis in cancer. *Carcinogenesis*. **21**: 485-495.
- Ludwig, H., Pohl, G. and Osterborg, A. (2004). Anemia in multiple myeloma. *Clinical Advances in Hematology and Oncology*. **2**: 233-241.
- Lutz, W.K. (2002). Differences in individual susceptibility to toxic effects of chemicals determine the dose-response relationship and consequences of setting exposure standards. *Toxicology Letters*. **126**: 155-158.
- Malinen, E., Kassinen, A., Rintitila, T. and Palva, A. (2003). Comparison of real-time PCR with SYBR Green I or 5'-nuclease assays and dot-blot hybridization with rDNA-targeted oligonucleotide probes in quantification of selected faecal bacteria. *Microbiology*. **149**: 269-277.
- Mao, F., Leung, W-Y. and Xin, X. (2007). Characterization of EvaGreen and the implications of its physicochemical properties for qPCR applications. *BMC Biotechnology*. **7**: 76.

- Mathinavan, N. Surendiran, G., Srinivasa, K., Sagadevan, E. and Malarvizhi, K. (2005). Review on the current scenario of Noni research: Taxonomy, distribution, chemistry, medicinal and therapeutic values of *Morinda citrifolia*. *International Journal of Noni Research*. **1**: 1-16.
- McClatchey, W. (2002). From Polynesian Healers to Health Food Stores: Changing perspectives of *Morinda citrifolia* (Rubiaceae). *International Journal of Noni Research*. **1**: 110-120.
- Meirow, D. and Nugent, D. (2001). The effects of radiotherapy and chemotherapy on female reproduction. *Human Reproduction Update*. **7**: 535-543.
- Mellino, G. and Vaux, D. (2010). *Cell death*. West Sussex : Wiley-Blackwell.
- Melo, J.V., Hughes, T.P. and Apperley, J.F. (2003). Chronic myeloid leukaemia. *Hematology American Society of Hematology Education Program*. **2003**: 132-152.
- Minamoto, T., Mai, M. and Ronai, Z. (2000). K-ras mutation: early detection in molecular diagnosis and risk assessment of colorectal, pancreas, and lung cancers-a review. *Cancer Detection and Prevention*. **24**: 1-12.
- Mizoguchi, M., Naito, H., Kurata, Y., Shibata, M-A., Tsuda, H., Wild, C. P., Montesano, R. and Fukushima, S. (1993). Influence of aging on multi-organ carcinogenesis in rats induced by *N*-Methyl-*N*-Nitrosourea. *Japanese Journal of Cancer Research*. **4**: 139-146.
- Monis, P. T, Giglio, S. and Saint, C. P. (2005). Comparison of SYTO9 and SYBR Green I for real-time polymerase chain reaction and investigation of the effect of dye concentration on amplification and DNA melting curve analysis. *Analytical Biochemistry*. **340**: 24-34.
- Montesano, R. and Hall, J. (2001). Environmental Causes of Human Cancers. *European Journal of Cancer*. **37**: 67-87.
- Moon, R.C and Mehta, R.G. (1989). Chemoprevention of experimental carcinogenesis in animals. *Preventive Medicine*. **18**: 576-581.
- Moorthy, N.K. and Reddy, G.S. (1970). Preliminary phytochemical and pharmacological study of *Morinda citrifolia*, Linn. *Antiseptic*. **67**: 167-171.
- Morris, T., Robertson, B. and Gallagher, M. (1996). Rapid reverse transcription-PCR detection of hepatitis C virus RNA in serum by using the TaqMan fluorogenic detection system. *Journal of Clinical Microbiology*. **34**: 2933-2936.
- Morrison, T.B., Weis, J.J. and Wittwer, C.T. (1998). Quantification of low-copy transcripts by continuous SYBR Green I monitoring during amplification. *Biotechniques*. **24**: 954-962.

- Morse, M.A. and Stoner, G.D. (1993). Cancer chemoprevention: principles and prospects. *Carcinogenesis*. **14**: 1737-1746.
- Morton, D., Bailey, K.L., Stout, C.L., Weaver, R.J., White, K.A., Lorenzen, M.J. and Ball, D.J. (2008). *N*-Methyl-*N*-Nitrosourea (MNU): A Positive Control Chemical for p53+/- Mouse Carcinogenicity Studies. *Toxicologic Pathology*. **36**: 926.
- Motadi, L.R., Misso, N.L., Dlamini, Z. and Bhoola, K.D. (2007). Molecular genetics and mechanisms of apoptosis in carcinomas of the lung and pleura: therapeutic targets. *International Immunopharmacology*. **7**: 1934-1947.
- Mughal, T.I., Goldman, J.M. and Mughal, S.T. (2006). *Understanding leukaemias, lymphomas, and myelomas*. London: Taylor & Francis.
- Murakami, A., Ohigashi, H. and Koshimizu, K. (1996). Anti-tumor promotion with food phytochemicals: a strategy for cancer chemoprevention. *Bioscience, Biotechnology and Biochemistry*. **60**: 1-8.
- Narayanan, B. A., Condon, M. S., Bosland, M. C., Narayanan, N. K. and Reddy, B. S. (2003). Suppression of *N*-methyl-*N*-nitrosourea/testosterone-induced rat prostate cancer growth by celecoxib: effects on cyclooxygenase-2, cell cycle regulation, and apoptosis mechanism(s). *Clinical Cancer Research*. **9**: 3503-3513.
- Newman, D.J, Cragg, G.M. and Snader, K.M. (2003). Natural products as sources of new drugs over the period 1981-2002. *Journal of Natural Products*. **66**: 1022-1037.
- Nolan, T., Hands, R.E. and Bustin, S.A. (2006). Quantification of mRNA using real-time RT-PCR. *Nature Protocols*. **1**: 1559-1582.
- OECD Guideline 408. (1998). Repeated Dose 90-Day Oral Toxicity Study in Rodents. *OECD Guidelines for Testing of Chemicals*. Retrieved August 20, 2010 from <http://www.oecd.org>.
- Ogiu, T., Fukami, H. and Nishimura, M. (1992). DNA strand breaks and death of thymocytes induced by *N*-methyl-*N*-nitrosourea. *Journal of Cancer Research and Clinical Oncology*. **118**: 23-29.
- Ohta, T., Tokishita, S. and Yamagata, H. (2001). Ethidium bromide and SYBR Green I enhance the genotoxicity of UV-irradiation and chemical mutagens in *E. coli*. *Mutation Research*. **492**: 91-97.
- Okazaki, Y., Tanaka, Y., Tonogi, M. and Yamane, G. (2002). Investigation of environmental factors for diagnosing malignant potential in oral epithelial dysplasia. *Oral Oncology*. **38**: 562-573.
- Oltvai, Z. N., Millman, C. L. and Korsmeyer, S. J. (1993). Bcl-2 heterodimerizes *in vivo* with a conserved homolog, Bax that accelerates programmed cell death. *Cell*. **74**: 609-619.

- Packham, G. and Stevenson, F.K. (2005). Bodyguards and assassins: Bcl-2 family proteins and apoptosis control in chronic lymphocytic leukaemia. *Immunology*. **114**: 441-449.
- Parikh, R.R., Gildener-Leapman, N., Narendran, A., Lin, H.Y., Lemanski, N., Bennett, J.A., Jacobson, H.I. and Andersen, T.T. (2005). Prevention of *N*-methyl-*N*-nitrosourea-induced breast cancer by alpha-fetoprotein (AFP)-derived peptide, a peptide derived from the active site of AFP. *Clinical Cancer Research*. **11**: 8512-8520.
- Parker, R.G. (1997). Iatrogenic carcinogenesis: clinical implications. *The Cancer Journal*. **3**: 316-327.
- Paul, T.R., Uppin, S.G., Uppin, S.M., Jacob, R.T., Rao, D.R. and Rajappa, S.J. (2010). Evaluation of cytopenias occurring in Imatinib treated Chronic Myeloid Leukaemia (CML) patients. *Indian Society of Haematology and Transfusion Medicine*. **B**: 56-61.
- Peerzada, N., Renaud, S. and Ryan, P. (1990). Vitamin C and elemental composition of some bush fruits. *Journal of Plant Nutrition*. **13**: 787-793.
- Perazella, M.A., Eisen, R.N., Frederick, W.G. and Brown, E. (1993). Renal failure and severe hypokalemia associated with acute myelomonocytic leukaemia. *American Journal of Kidney Diseases*. **22**: 462-467.
- Perry, M.C. (2008). *The chemotherapy source book*. Baltimore: Lippincott Williams and Wilkins.
- Perry, M.C., Anderson, C.M., Doll, D.C., Malhotra, V. and Shahab, N. (2004). *Companion Handbook to The Chemotherapy Sourcebook*. Baltimore: Lippincott Williams and Wilkins.
- Peter, M.E. and Krammer, P.H. (2003). The CD95 (APO-1/Fas) DISC and beyond. *Cell Death Differ*. **10**: 26-35.
- Pettman, B. and Henderson, C. E. (1998). Neuronal cell death. *Neuron*. **20**: 633-647.
- Pfaffl, M. W., Vandesompele, J. and Kubista, M. (2009). Data analysis software. In J. Logan, K. Edwards, and N. Saunders (Eds), *Real-Time PCR : Current Technology and Applications*. (pp 65-83). Norwich: Caister Academic Press.
- Pfaffl, M.W. (2001). A new mathematical model for relative quantification in real-time RT-PCR. *Nucleic Acids Research*. **29**: 2002-2007.
- Ponchel, F., Carmel, T., Bransfield, K., Leong, F. T., Douglas, S. H., Field, S. L., Bell, S. M., Combaret, V., Puisieux, A., Mighell, A. J., Robinson, P. A., Inglehearn, C. F., Isaacs, J. D. and Markham, A. F. (2003). Real-time PCR based on SYBR-Green

I fluorescence: An alternative to the TaqMan assay for a relative quantification of gene rearrangements, gene amplifications and micro gene deletions. *BMC Biotechnology*. **3**: 18.

Qu, W., Bortner, C.D., Sakurai, T., Hobson, M.J. and Waalkes, M.P. (2002). Acquisition of apoptotic resistance in arsenic-induced malignant transformation: role of the JNK signal transduction pathway. *Carcinogenesis*. **23**: 151-159.

Rasmussen, R. (2001). Quantification on the Light Cycler. In S. Meuer, C. Wittwer and K. Nakagawara (Eds), *Rapid cycle real-time PCR, methods and applications* (pp 21-34). Heidelberg: Springer.

Rathmell, J.C. and Thompson, C.B. (2002). Pathways of apoptosis in lymphocyte development, homeostasis, and disease. *Cell*. **109**: S97-S107.

Reed, J. C. (2000). Mechanisms of apoptosis. *The American Journal of Pathology*. **157**: 1415-1430.

Reed, J.C. (1994). Bcl-2 and regulation of programmed cell death. *The Journal of Cell Biology*. **124**: 1-6.

Reed, J.C. (1997). Bcl-2 family proteins: strategies for overcoming chemoresistance in cancer. *Advances in Pharmacology*. **41**: 501-532.

Reese, J.S., Allay, E. and Gerson, S.L. (2001). Overexpression of human O6-alkylguanine DNA alkyltransferase (AGT) prevents MNU induced lymphomas in heterozygous p53 deficient mice. *Oncogene*. **20**: 5258-263.

Regier, N. and Frey, B. (2010). Experimental comparison of relative RT-qPCR quantification approaches for gene expression studies in poplar. *BMC Molecular Biology*. **11**: 57.

Reis, L., Eisner, M. and Kosary, C. (2005). SEER cancer statistics review 1975-2002. National Cancer Institute. <http://www.seer.cancer.gov>

Ren, S. and Lien, E. J. (1997). Products and their derivatives as cancer chemopreventive agents. *Progress in Drug Research*. **48**: 147-171.

Renehan, A.G., Booth, C. and Potten, C.S. (2001). What is apoptosis, and why it is important? *British Medical Journal*. **322**: 1536-1538.

Ribera, J.M. and Oriol, A. (2009). Acute lymphoblastic leukaemia in adolescents and young adults. *Hematology/ Oncology Clinics of North America*. **23**: 1033-1042.

Riedl, S.J. and Salvesen, G.S. (2007). The apoptosome: signalling platform of cell death. *Nature Reviews Molecular Cell Biology*. **8**: 405-413.



- Riedl., S. J. and Shi., Y. (2004). Molecular mechanisms of caspase regulation during apoptosis. *Nature Reviews Molecular Cell Biology*. **5**: 897-907.
- Robertson, L.E., Plunkett, W., McConnell K. and McDonnell, T.J. (1996). Bcl-2 expression in chronic lymphocytic leukaemia and its correlation with the induction of apoptosis and clinical outcome. *Leukaemia*. **10**: 456-459.
- Roitt, I., Brostoff, J. and Male, D. (2001). *Immunology*. London: Mosby.
- Rosly, S.M., Shanmugavelu, S., Murugaiyah, M., Hadijah, H., Ahmad Tarmizi, S., Noridayusni, Y. and Subramaniam, K. (2011). Subchronic Oral Toxicity Study of *Morinda citrifolia* (Mengkudu) in Sprague Dawley Rats. *Pertanika Journal of Tropical Agricultural Science*. **34**: 341–349.
- Ruwanpura, S. M., McLachlan, R.I., Stanton, P.G., Loveland, K.L. and Meachem, S.J. (2008). Pathways involved in testicular germ cell apoptosis in immature rats after FSH suppression. *Journal of Endocrinology*. **197**: 35-43.
- Sathiya, M. and Muthuchelian, K. (2009). Significance of immunologic markers in the diagnosis of lymphoma. *Academic Journal of Cancer Research*. **2**: 40-50.
- Saunders, N.A. (2009). An introduction to Real-Time PCR. In J. Logan., K. Edwards. and N. Saunders (Eds), *Real-Time PCR: Current Technology and Applications* (pp.20-35). Caister Academic Press, London.
- Schelman, W.R., Andres, R.D., Sipe, K.J., Kang, E., Weyhenmeyer, J.A. (2004). Glutamate mediates cell death and increases the Bax to Bcl-2 ratio in a differentiated neuronal cell line. *Molecular Brain Research*. **128**: 160–169.
- Schmittgen, T.D., Zakrajsek, B.A., Mills, A.G., Gorn, V., Singer, M.J. and Reed, M.W. (2000). Quantitative reverse transcription-polymerase chain reaction to study mRNA decay: comparison of endpoint and real-time methods. *Analytical Biochemistry*. **285**: 194-204.
- Sedlak, T.W., Oltvai, Z.N., Yang, E., Wang, K., Boise, L.H., Thompson, C.B. and Korsmeyer, S. J. (1995). Multiple Bcl-2 family members demonstrate selective dimerizations with Bax. *Proceedings of the National Academy of Sciences of the United States of America*. **92**: 834-7838.
- Segal, I., Rassekh, S. R., Bond, M. C., Senger, C., Schreiber, R. A. (2010). Abnormal liver transaminases and conjugated hyperbilirubinemia at presentation of acute lymphoblastic leukaemia. *Pediatric Blood Cancer*. **55**: 434-439.
- Seidemann, J. (2002). Noni- a questionable magic fruit from the South Seas. *Zeitschrift-Fur-Phytotherapie*. **23**: 62-67.
- Senthikumar, S., Yogeeta, S.K., Subashini, R. and Devaki, T. (2006). Attenuation of cyclophosphamide induced toxicity by squalene in experimental rats. *Chemico Biological Interactions*. **160**: 252-60.

- Shain, E.B. and Clemens, J.M. (2008). A new method for robust quantitative and qualitative analysis of real-time PCR. *Nucleic Acids Research*. **36**: 91.
- Simons, J.W. (1995). Genetic, epigenetic, dysgenetic, and non-genetic mechanisms in tumorigenesis. *Critical Reviews in Oncogenesis*. **6**: 261-273.
- Simonsen, J.L. (1920). Note on the constituents of *Morinda citrifolia*. *Journal of the Chemical Society*. **117**: 561-564.
- Singh, J. and Tiwari, R.D. (1976). Flavone glycosides from the flowers of *Morinda citrifolia*. *Journal of the Indian Chemical Society*. **53**: 424.
- Sjöström, J. and Bergh, J. (2001). How apoptosis is regulated, and what goes wrong in cancer. *British Medical Journal*. **322**: 1538-1539.
- Souaze, F., Ntodou-Thome, A., Tran, C.Y., Rostene, W. and Forgez, P. (1996). Quantitative RT-PCR: limits and accuracy. *BioTechniques*. **21**: 280-285.
- Sporn, M.B., Dunlop, N.M., Newton, D.L. and Smith, J.M. (1976). Prevention of chemical carcinogenesis by vitamin A and its synthetic analogs (retinoids). *Federation Proceedings*. **35**: 1332-1338.
- Srivastava, D.N. (2006). *Review on the Anti-Inflammatory and Analgesic Activity of Morinda citrifolia (NONI) in Experimental Animals*. Paper presented at: First National Symposium on Noni Research, Hyderabad.
- Srivastava, H.S. (1999). Atmospheric Carcinogens: How Safe We Are! *Enviro News - Newsletter of ISEB India*. **5**: 4.
- Srivastava, M. and Singh, J. (1993). A new anthraquinone glycoside from *Morinda citrifolia*. *International Journal of Pharmacology*. **31**: 182-184.
- Stahlberg, A., Zoric, A., Aman, P. and Kubista, M. (2005). Quantitative real-time PCR for cancer detection: the lymphoma case. *Expert Review of Molecular Diagnostics*. **5**: 221-230.
- Stern, M. and Sun, P. (2009). Renal complications of cancer. In M. D. Stubblefield (Ed.), *Cancer Rehabilitation: Principles and Practice* (pp. 80-90). New York: Demos Medical Publishing.
- Stone, R.M., O'Donnell, M.R. and Sekeres, M.A. (2004). Acute myeloid leukaemia. *Hematology*. **2004**: 98-117.
- Studzinski, G.P. (1999). *Apoptosis: A practical approach*. Oxford: Oxford University Press.
- Sung, H.J., Kim, S.J., Lee, J.H., Lee, G., Lee, K.A., Choi, C.W., Kim, B.S. and Kim, J.S. (2007). Persistent anemia in a patient with diffuse large B cell lymphoma: pure

red cell aplasia associated with latent Epstein-Barr virus infection in bone marrow. *Journal of Korean Medical Science*. **22**: S167-S170.

- Swierzewski, S.J. (1999). *Leukaemia diagnosis*. Retrieved Mei 2011, from <http://www.oncologychannel.com/leukaemias/diagnosis.shtml>
- Tabrah, F.L. and Eveleth, B.M. (1966). Evaluation of the effectiveness of ancient Hawaiian medicine. *Hawaii Medical Journal*. **25**: 223-230.
- Taft, R., Pang, K.C., Mercer, T.R., Dinger, M. and Mattick, J.S. (2010). Non-coding RNAs: regulators of disease. *Journal of Pathology*. **220**: 126-139.
- Tait, S.W.G. and Green, D.R. (2010). Mitochondria and cell death: outer membrane permeabilization and beyond. *Nature Reviews Molecular Cell Biology*. **11**: 621-632.
- Tanghapazham, R. L., Rajeshkumar, N. V., Sharma, A., Warren, J., Singh, A. K., Ives, J. A., Gaddipati, J. P., Maheshwari, R. K. and Bonas, W. B. (2006). Effect of homeopathic treatment on gene expression in Copenhagen rat tumor tissues. *Integrative Cancer Therapies*. **5**: 350-355.
- Tannock, I.F. (1998). Conventional cancer therapy: promise broken or promise delayed? *Lancet*. **3519**: 16.
- Thomas, A., Pepper, C., Hoy, T. and Bentley, P. (2000). Bcl-2 and Bax expression and chlorambucil-induced apoptosis in the T-cells and leukaemic B-cells of untreated B-cell chronic lymphocytic leukaemia patients. *Leukaemia Research*. **24**: 813-821.
- Thordarson, G., Slusher, N., Leong, H., Ochoa, D., Rajkumar, L., Guzman, R., Nandi, S. and Talamantes, F. (2004). Insulin-like growth factor (IGF)-I obliterates the pregnancy-associated protection against mammary carcinogenesis in rats: evidence that IGF-I enhances cancer progression through estrogen receptor-alpha activation via the mitogen-activated protein kinase pathway. *Breast Cancer Research*. **6**: R423-R436.
- Tiwari, R.D. and Singh, J. (1977). Structural study of the anthraquinone glycoside from the flowers of *Morinda citrifolia*. *Journal of the Indian Chemical Society*. **54**: 429-430.
- Traynor, J., Mactier, R., Geddes, C.C. and Fox, J.G. (2006). How to measure renal function in clinical practice. *British Medical Journal*. **333** :733-737.
- Trümper L.H., Brittinger, G., Diehl, V. and Harris, N.L. (2004). Non-Hodgkin's lymphoma: a history of classification and clinical observations. In P.M. Mauch, J.O. Armitage, B. Coiffier., R. Dalla-Favera and N.L. Harris (Eds.), *Non-Hodgkin's Lymphomas* (pp. 3-19). Philadelphia, PA: Lippincott Williams & Wilkins.

- Tsao, A., Kim, E.S. and Hong, W.K. (2004). Chemoprevention of cancer. *A Cancer Journal for Clinicians*. **54**: 150-180.
- Tsubura, A., Lai, Y.C., Miki, H., Sasaki, T., Uehara, N., Yuri, T. and Yoshiziwa, K. (2011). Animal Models of *N*-Methyl-*N*-nitrosourea-induced mammary cancer and retinal degeneration with special emphasis on therapeutic trials. *In vivo*. **25**: 11-22.
- Tules, S.D. (2006). New development in lymphoma and Hodgkin's disease research. Nova Science Publishers: New York.
- Tuzuner, N. and Bennett, J.M. (1994). Reference standards for bone marrow cellularity. *Leukaemia research*. **18**: 645-647.
- Valasek, M.A. and Repa, J.J. (2005). The power of real-time PCR. *Advances in Pyhsiology Education*. **29**: 151-159.
- Van de Putte, G., Holm, R., Lie, A. K., Baekelandt, M. and Kristensen, G. B. (2005). Markers of apoptosis in stage IB squamous cervical carcinoma. *Journal of Clinical Pathology*. **58**: 590-594.
- Vandesompele, J., De Preter, K., Pattyn, F., Poppe, B., Van Roy, N., De Paepe, A., Speleman, F. (2002). Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biology*. **3**: RESEARCH0034.
- VanGuilder, H. D., Vrana, K. E. and Freeman, W. M. (2008). Twenty-five years of quantitative PCR forgene expression analysis. *BioTechniques*. **44**: 619-626.
- Varga, A. and James, D. (2006). Real-time RT-PCR and SYBR Green I melting curve analysis for the identification of Plum pox virus strains C, EA, and W: effect of amplicon size, melt rate, and dye translocation. *Journal of Virological Methods*. **132**: 146-153.
- Venugopal, M., Callaway, A. and Snyderwine, E. G. (1999). 2-Amino-1-methyl-6-phenylimidazo[4,5-b]pyridine (PhIP) retards mammary gland involution in lactating Sprague–Dawley rats. *Carcinogenesis*. **20**: 1309-1314.
- Walker, H.K., Hall, W.D. and Hurst, J.W. (1990). *Clinical Methods: The History, Physical, and Laboratory Examinations*. Boston: Butterworth-Heinemann.
- Wallace, J. (2000). Humane Endpoints for Animals Used in Biomedical Research and Testing. *Institute for Laboratory Animal Research Journal*. **41**: 2.
- Wang, M. and Su, C. (2001). Cancer preventive effect of *Morinda citrifolia* (Noni). *Annals of the New York Academy of Sciences*. **1952**: 161-168.
- Wang, M.Y., Brett, J.W., Jarakae, J., Diane, N. and Gary, A. (2002). *Morinda citrifolia* (Noni): a literature review and recent advances in Noni research. *Acta Pharmacology Sinaca*. **12**: 1127-1141.

- Wang, M-Y., Nowicki, D., Anderson, G., Jensen, J. and West, B. (2008). Liver protective effects of *Morinda citrifolia* (Noni). *Plant Foods for Human Nutrition*. **65**: 59-56.
- Wang, T. and Brown, M. J. (1999). mRNA quantification by real time TaqMan polymerase chain reaction: validation and comparison with RNase protection. *Analytical Biochemistry*. **269**: 198-201.
- Wedemeyer, H., Hofmann, W. P., Lueth, S., Malinski, P., Thimme, R., Tacke, F., Wiegand, J. (2010). ALT screening for chronic liver diseases: scrutinizing the evidence. *Z Gastroenterol*. **48**: 46-55.
- Whistler, W.A. (1991). Polynesian plant introductions. In P.A. Cox and S.A. Banack (Eds.), *Islands, Plants, and Polynesians* (pp. 41-66). Portland: Dioscorides Press.
- Whitby, L.G., Percy-Robb, I.W. and Smith, A. F. (1984). *Lecture notes on Clinical Chemistry*. London: Blackwell Science Publication.
- White, P. and Mamaroneck, A. (2005). Leukemia Facts and Statistics from Leukemia, Lymphoma, Myeloma, Facts 2010-2011. *Journal Leukemia and Lymphoma Society*. **13**: 91-105.
- Williams, D.H., Stone, M.J., Hauck, P.R., Rahman, S.K. (1989). Why are secondary metabolites (natural products) biosynthesized? *Journal of Natural Products*. **52**: 1189-1208.
- Williams, G.M. (2001). Mechanisms of chemical carcinogenesis and application to human cancer risk assessment. *Toxicology*. **161**: 3-10.
- Wittwer, C. T., Herrmann, M. G., Moss, A. A. and Rasmussen, R. P. (1997). Continuous fluorescence monitoring of rapid cycle DNA amplification. *Biotechniques*. **22**: 176-181.
- Wolbach, S.B. and Howe, P.R. (1925). Tissue changes following deprivation of fat-soluble A vitamin. *The Journal of Experimental Medicine*. **42**: 753-77.
- Wong, M.L. and Medrano, J.F. (2005). Real-time PCR for mRNA quantitation. *Biotechniques*. **39**: 75-85.
- Wyllie, A.H., Kerr, J.F.R. and Currie, A.R. (1980). Cell death: the significance of apoptosis. *International Review of Cytology*. **68**: 251-306.
- Yamagiwa, K. and Ichikawa, K. (1915). Experimentelle Studie über die Pathogenese der Epithelialgeschwülste. *Mitt. Med. Fak. Kaiserl. Univ. Tokio*. **15**: 295-344.
- Yamamoto, N. and Okamoto, T. (1995). A rapid detection of PCR amplification product using a new fluorescent intercalator; the pyrylium dye, P2. *Nucleic Acids Research*. **23**: 1445-1446.

Yang, E. and Korsmeyer, S.J. (1996). Molecular thanatopsis: a discourse on the BCL2 family and cell death. *Blood*. **88**: 386-401.

Yin, X.M., Oltvai, Z.N. and Korsmeyer, S.J. (1994). BH1 and BH2 domains of Bcl-2 are required for inhibition of apoptosis and heterodimerization with Bax. *Nature*. **369**: 321-323.

Zhou, S.B., Wang, G.J., Zhu, Y. and Chen, B.Q. (2000). Effect of dietary fatty acids on colon tumorigenesis induced by methyl nitrosourea in rats. *Biomedical and Environmental Sciences*. **13**: 105-116.

Zimmerman, K.C. and Green, D.R. (2001). How cells die: Apoptosis pathways. *The Journal of Allergy and Clinical Immunology*. **108**: S99-S103.

