



UNIVERSITI PUTRA MALAYSIA

USE OF *Lactobacillus plantarum* AND *Propionibacterium freudenreichii* ssp. *Shermanii* AS INOCULANTS IN CORN SILAGE FERMENTATION

NORAFIZAH BINTI HAJI ABDUL RAHMAN

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By

NORAFIZAH BINTI HAJI ABDUL RAHMAN

This thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in fulfilment of the requirement for the Degree of Master of Science

January 2017

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January 2017

Chairman : Associate Professor Mohd Ridzwan Bin Abd Halim, PhD

Faculty : Agriculture

Bacterial inoculant as feed additives have been shown to be beneficial in silage fermentation in many parts of the world. However, there is still not much evidence of the use of bacterial additives in Malaysia and the impact of using them to produce quality feed for ruminant industry is rarely documented. Thus, this study was conducted to determine the effectiveness of using two bacterial additives for corn silage fermentation in Malaysia. The two bacterial species used in this study were *Lactobacillus plantarum* (B1) and *Propionibacterium freudenreichii* (B2). These bacteria were applied singly and as a mixture of both bacteria (B3).

Three experiments were conducted where the first experiment was to determine the suitable bacterial growth condition and inoculation size. The preferable growth media for B1 was found to be MRS agar (18-24 hours) while for B2, it was Reinforced Clostridium Difficile Agar (48-56 hours). The wavelength for OD measurement of B1 was 490 nm and 419 nm for B2. Suitable inoculation size for B1 and B2 applied individually was at 1×10^5 cfu/g, while suitable inoculation size for B3 is 0.5×10^5 cfu/g.

The second experiment was conducted to determine the effect of B1 and B2 applied singly or as a mixture to the fermentation characteristic and quality of corn silage. The physical characteristic of the silage measured were pH, mould and temperature while the quality measurement included crude protein (CP), dry matter (DM) and fibre content. The fermentation

products measured were water soluble carbohydrate (WSC) and fatty acid concentrations (lactic, propionic, acetic and butyric acids). A good silage quality should have a low pH, acetic acid, and butyric acid, but high lactic acid content.

Whole corn plant was harvested at dough stage (32 - 37% of dry matter) and cut into 2-3 cm particle size using a forage chopper. Two kilograms of chopped materials were placed on polyethylene sheet and sprayed with 10 ml of prepared bacteria solution. The forage were mixed thoroughly and ensiled in four replicates for each treatment in 4 L laboratory silo. Bacterial combination (B3) ended with the lowest pH (3.33), while the highest was B2 (pH 3.45). The pH of silage treated with bacteria showed a faster reduction compared to untreated silage. The use of *L. plantarum* alone rapidly reduced pH in the early stages of silage fermentation. Bacterial additives did not reduce mould infection and silage treated with B2 produced higher mould percentage than control (B0). Bacterial treatment did not reduce the fibre content as compared to control. Both bacteria in this research showed low activity on digesting fibre. Bacterial treatments gave a significant effect ($p < 0.01$) on WSC, lactic acid (LA) and acetic acid (AA) while there was no significant effect on propionic acid and butyric acid. Fermentation with *L. plantarum* alone appeared to produce high LA concentration among all treatments. The LA production was 12 times higher than AA production showing that it promoted the homolactic fermentation to be dominant in the silage. However, when *L. plantarum* was mixed with *P. freudenreichii*, the fermentation tended to be dominated by *P. freudenreichii*.

The effect of bacterial additive was observed at 5 different ensiling period (3, 6, 9, 18 and 27 days) and the quality was determined in the third experiment. The time course study in fermentation indicated that the ensilation process was completed at day 6 even without using bacterial additives.

This research showed that silage produced with *L. plantarum* or *P. freudenreichii* either alone or mixed together produced a desirable silage property but it was not significantly better than control. It was speculated that naturally occurring bacteria was already adequate in the uninoculated corn silage because of the high amount of fermentable sugar in the corn itself which provided the essential source of energy for bacteria to carry out successful preservation during fermentation. Hence, the nutrient qualities of corn silage produced with the addition of the two selected bacteria strains were shown to be the same as uninoculated corn silage.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Sarjana Sains

**PENGGUNAAN *Lactobacillus plantarum* DAN *Propionibacterium
freudenreichii* ssp. *Shermanii* SEBAGAI INOKULAN DALAM
FERMENTASI SILAJ JAGUNG**

Oleh

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Penggunaan bakteria inokulan sebagai bahan tambah makanan ternakan terbukti berfaedah membantu proses fermentasi silaj di kebanyakan negara luar. Namun begitu, maklumat-maklumat saintifik berkaitan kesan dan impak penggunaan bakteria sebagai aditif untuk industri ruminan di Malaysia masih lagi terhad dan perlu dikaji dengan lebih mendalam. Sehubungan itu, kajian ini dilaksanakan bertujuan untuk mengetahui keberkesanan penggunaan dua bakteria inokulan terpilih untuk fermentasi silaj jagung di Malaysia. Dua spesies bakteria yang digunakan di dalam kajian ini adalah *Lactobacillus plantarum* (B1) and *Propionibacterium freudenreichii* (B2). Kedua bakteria ini digunakan secara bersendirian dan juga digabungkan keduanya (B3).

Tiga eksperimen dilaksanakan dalam kajian ini. Eksperimen pertama adalah bertujuan untuk mengetahui kriteria asas untuk pertumbuhan bakteria dan kuantiti inokulasi yang bersesuaian untuk digunapakai untuk fermentasi silaj. Media yang sesuai untuk B1 adalah MRS (18-24 jam) manakala bagi B2 adalah media *Reinforced Clostridium Difficile* (48-56 jam). Gelombang OD 490 nm adalah bersesuaian untuk mengesan B1 manakala B2 dikesan pada 419 nm. Saiz inokulasi yang bersesuaian adalah 1×10^5 cfu/g bagi B1 dan B2 manakala kuantiti separuh daripadanya (0.5×10^5 cfu/g) adalah memadai bagi B3.

Dalam eksperimen kedua, kesan penggunaan bakteria B1 dan B2 secara individu atau gabungan keduanya (B3) dianalisis berdasarkan ciri-ciri fermentasi dan kandungan nutrisi silaj. Ciri-ciri fizikal fermentasi yang diukur adalah pH, peratusan kulapuk dan suhu manakala kualiti nutrisi yang ditentukan ialah kandungan protein kasar (CP), berat kering (DM), dan serat. Selain itu, produk yang terhasil daripada proses fermentasi seperti kandungan karbohidrat larut air (WSC) dan asid lemak (asid laktik, propionik, asetik dan butirik) turut dianalisis. Fermentasi yang baik akan menghasilkan silaj dengan pH yang rendah, kandungan asid laktik yang tinggi di samping mempunyai kandungan asid asetik dan butirik yang rendah.

Rumpun jagung telah dituai di peringkat *dough* (kandungan berat kering 32-37%) dan dikisar kecil (2-3 cm) dengan menggunakan pericik foder. Sebanyak 10 ml larutan bakteria yang telah disediakan disembur pada 2 kg foder yang telah dikisar dan kemudiannya digaul rata. Foder yang terawat kemudiannya disimpan untuk fermentasi di dalam balang (silo) berkapasiti 4 L. Di akhir proses fermentasi, B3 mencatatkan pH terendah (3.33), manakala B2 mencatatkan pH tertinggi (3.45). Silaj yang dirawat dengan bakteria menunjukkan penurunan pH yang lebih pantas berbanding silaj kawalan. Penggunaan *L. plantarum* secara individu menunjukkan kadar penurunan pH yang pantas semasa di awal tempoh fermentasi. Penggunaan bakteria didapati tidak mengurangkan pertumbuhan kulapuk malahan penggunaan B2 telah menunjukkan hasil kulapuk yang lebih banyak berbanding kawalan. Bakteria terpilih daripada kajian ini menunjukkan kadar serat yang setara berbanding kawalan (B0). Kedua bakteria yang digunakan di dalam kajian ini telah menunjukkan aktiviti penguraian serat yang rendah. Kombinasi bakteria (B3) tidak mengurangkan kadar serat manakala perlakuan dari bakteria lain tidak mengurangkan serat secara signifikan berbanding kawalan. Perlakuan bakteria memberikan kesan yang signifikan pada $p < 0.01$ kepada karbohidrat larut air (WSC), asid laktik (LA) dan asid asetik manakala tiada perbezaan yang signifikan bagi asid propionik dan asid butirik. Penggunaan *L. plantarum* telah menghasilkan 12 kali kandungan laktik asid lebih tinggi berbanding asetik asid. Ini menunjukkan bahawa bakteria ini telah menyebabkan proses fermentasi menjadi homolaktik. Namun begitu, apabila digabungkan bersama *P. freudenreichii*, fermentasi telah beralih menjadi heterolaktik.

Kesan penggunaan bakteria sebagai aditif turut direkodkan pada 5 tempoh fermentasi yang berbeza di dalam eksperimen ketiga. Perbandingan kajian di antara tempoh fermentasi telah menunjukkan bahawa proses fermentasi telahpun lengkap pada hari ke 6 walaupun tanpa penggunaan bakteria sebagai aditif.

Oleh yang demikian, hasil daripada kajian ini telah membuktikan bahawa dengan menggunakan *L. plantarum* mahupun *P. freudenreichii* sebagai aditif sama ada secara bersendirian mahupun percampuran antara keduanya telah menghasilkan silaj yang berkualiti tetapi tidak mempunyai perbezaan yang signifikan berbanding kawalan. Disebabkan kandungan karbohidrat yang tinggi di dalam foder jagung, diandaikan bahawa bakteria

semulajadi di dalam silaj telah mempunyai substrat yang mencukupi untuk mengendalikan fermentasi dengan sempurna tanpa perlu pertambahan bakteria lain sebagai aditif. Sehubungan itu, penggunaan dua bakteria terpilih ini telah menghasilkan kualiti silaj yang setara dengan silaj kawalan.



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APPROVAL

I certify that a Thesis Examination Committee has met on 12th January 2017 to conduct the final examination of **Norafizah Haji Abdul Rahman** on her thesis entitled “**USE OF *Lactobacillus plantarum* AND *Propionibacterium freudenreichii* ssp. *Shermanii* AS INOCULANTS IN CORN SILAGE FERMENTATION**” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

μ	micro
AA	acetic acid
ADF	acid detergent fibre
BA	butyric acid
cfu	colony forming unit
CP	crude protein
DM	dry matter
GC	gas chromatography
LA	lactic acid
LAB	lactic acid bacteria
M	molar
MJ/kg	megajoules per kilogram
MS	mass spectrometry
MW	molecular weight
NDF	neutral detergent fibre
nm	nanometer
nVFA	non volatile fatty acid
O. D.	optical density
PA	propionic acid
PAB	propionic acid bacteria
rpm	rotation per minute
v/v	volume per volume
VFA	volatile fatty acid
w/v	weight per volume
WSC	water soluble carbohydrate

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CHAPTER 1

INTRODUCTION

Silages can be defined as forages, crop residues, or agricultural or industrial by products that have been preserved naturally for the use as animal feeds. It is conserved by reducing the pH through natural anaerobic fermentation or induced by additives and stored in closed containers. Success in silage-making depends on adequate levels of water soluble carbohydrate to be fermented to form lactic acid under anaerobic condition which eventually leads to a low pH environment that is required in preserving the forage (Henderson, 1993).

Silage is the product of a series of fermentation processes by which cut forage with high moisture content (60-75%) is allowed to ferment to preserve the quality usually by lactic acid bacteria (Kung, 2001). The acidic environment induced by lactic acid bacteria in anaerobic condition will resist further breakdown of plant material during storage (Woolford, 1984). The objective is to retain the nutrients present in the original forage and to be acceptable by livestock.

Most studies have shown that silage is more palatable than other conserved feed such as hay (Erdman et al., 2011; Holt et al., 2013; Arriola et al., 2015). Silage can be kept for months or years and can be used at any time as and when required, especially during periods of drought or floods. In a tropical country like Malaysia, silage production is preferred as it is less dependent on weather conditions for sun-drying to produce hay. Silage kept in trenches in Malaysia has been shown to be in good condition even after five years of storage (Chin and Idris, 1999). Later reports also showed that silage can be kept in a good conditions from 90 to 361 days (Kung et al., 2007a; Mohd-Setapar et al., 2012).

There are three basic conditions to make ensilation possible; i.e., fermentable sugars to support the bacterial activity, suitable conditions for the desirable bacteria to be active and suitable composition and correct numbers of the desirable microorganisms in the raw material (Aksu et al., 2006; Filya and Sucu, 2010; Queiroz et al., 2013). With a better knowledge on silage improvement nowadays, a technology to improve silage quality and recovery has been developed by using bacterial inoculant as additives.

Two different types of microorganism are involved in silage fermentation, one group is desirable while the other is undesirable. The example of desirable microorganisms are lactic acid producing bacteria (LAB), propionic acid producing bacteria (PAB) and some other fungal species that undergo homofermentative or heterofermentative process during fermentation. The desirable bacteria can improve the fermentation, shelf life, and quality of silages (Filya et al., 2007; Pedroso et al., 2010), while in the event of accidental soil incorporation, a long aerobic phase or slow

acidification, the microbial communities in the silage will be dominated by *Clostridia*, yeasts, moulds and by chance incorporated pathogenic microorganisms such as *Listeria* spp. can cause anaerobic spoilage on silage (Dunière et al., 2013).

Lactic acid bacteria (LAB) are the most popular bacterial additives for silage fermentation. Studies on LAB showed that the bacteria can rapidly reduce pH and enhance silage quality (Kung et al., 2007b; Meeske and Basson, 1998; Ranjit and Kung, 2000). However, several microorganisms that are not LAB have been used as silage inoculants specifically for the purpose of improving aerobic stability (Kleinschmit and Kung, 2006; Kristensen et al., 2010). Propionic acid bacteria (PAB) was also being chosen for the ability to convert lactic acid and glucose to acetic and propionic acids that are more antifungal than lactic acid, thus it can stabilize silage longer after exposing to air and increase the aerobic stability. Flores-Galarza et al. (1985) and Dawson et al. (1998) reported that addition of *P. shermanii* prevented the growth of moulds and markedly reduced the initial population of yeast in high moisture corn where the final pH was greater than 4.5. Bolsen et al. (1996) reported more propionic acid, lower yeasts and moulds, and greater aerobic stability in corn silage (pH of 3.6) treated with *Propionibacteria*.

Studies on microbial additives for silage have been conducted in other countries and they have indicated significant benefits (Aksu et al., 2004; Huisden et al., 2009; Nkosi et al., 2010). Using bacteria as an additive enhanced nutrient recovery and improved fermentation quality. Research by Weinberg et al. (1995) proved the efficiency of using bacterial additives in silage production compared to non-treated silage (without bacterial additives) for corn and alfalfa silage.

Due to the lack of good quality feed and grazing area for ruminant production in Malaysia, intensive research on the use of indigenous feed resources are required to reduce the burden of feed imports. Furthermore, Malaysia also faces periods of feed limitation, usually due to drought or floods in certain parts of the country. The high humidity environment in Malaysia also causes the plant materials like corn to be easily contaminated by certain microorganisms like mould. This results in shortage of fresh feeds available to animals. Hence, strategies should be developed to sustain the animal production industry. Forage conservation is a practical consideration to ensure continuous supply of feeds throughout the year.

The use of microbial additives for silage making in Malaysia is not widely practised. Therefore, the information on the benefits and the effectiveness of microbial additives in silage making is needed to make appropriate recommendations for feed conservation under local conditions. Corn (*Zea mays*) was selected as the silage crop for this study as it is known to contain higher amounts of water soluble carbohydrates compared to tropical forage grasses. Soluble carbohydrates would be a source of nutrients for microbial fermentation for production of organic acids.

Thus, the aim of this study was to determine the nutrient quality of the corn silage produced by using hetero and homofermentative bacterial species applied singly or as a mixture in the corn silage fermentation. Specific objectives were:

- a. To determine the optimum growth conditions for culture and multiplication of *L. plantarum* and *P. freudenrinchii* ssp. *Shermanii*.
- b. To determine the effect of bacterial inoculants on corn silage characteristics and fermentation profiles after 4 weeks of ensilation.
- c. To determine the effect of bacterial inoculants on corn silage characteristics and fermentation profiles at five duration of fermentation from 5 to 27 days of ensilation.

REFERENCES

- Abdullah, A., Chee, W.C., 1999. Foraj Untuk Ternakan, First Edit. ed. Perpustakaan Negara Malaysia, Serdang, Malaysia.
- Acosta Aragón, Y., Jatkauskas, J., Vrotniakienė, V., 2012. The effect of a silage inoculant on silage quality, aerobic stability, and meat production on farm scale. *ISRN Vet. Sci.* 2012, 345927. doi:10.5402/2012/345927
- Adesogan, A.T., Krueger, N., Salawu, M.B., Dean, D.B., Staples, C.R., 2004. The influence of treatment with dual purpose bacterial inoculants or soluble carbohydrates on the fermentation and aerobic stability of bermudagrass. *J. Dairy Sci.* 87, 3407–16. doi:10.3168/jds.S0022-0302(04)73476-1
- Adlam, K., Kunkel, D., 2013. Microbe Wikipedia [WWW Document]. URL https://microbewiki.kenyon.edu/index.php/Lactobacillus_plantarum_and_its_biological_implications (accessed 10.13.15).
- Adogla-Bessa, T., Owen, E., 1995. Ensiling of whole-crop wheat with cellulase-hemicellulase based enzymes. 1. Effect of crop growth stage and enzyme on silage composition and stability. *Anim. Feed Sci. Technol.* 55, 335–347.
- Aksu, T., Baytok, E., Bolat, D., 2004. Effects of a bacterial silage inoculant on corn silage fermentation and nutrient digestibility. *Small Rumin. Res.* 55, 249–252. doi:10.1016/j.smallrumres.2003.12.012
- Aksu, T., Baytok, E., Karlı, M.A., Muruz, H., 2006. Effects of formic acid, molasses and inoculant additives on corn silage composition, organic matter digestibility and microbial protein synthesis in sheep. *Small Rumin. Res.* 61, 29–33. doi:10.1016/j.smallrumres.2004.12.013
- Amer, S., Hassanat, F., Berthiaume, R., Seguin, P., Mustafa, a. F., 2012. Effects of water soluble carbohydrate content on ensiling characteristics, chemical composition and in vitro gas production of forage millet and forage sorghum silages. *Anim. Feed Sci. Technol.* 177, 23–29. doi:10.1016/j.anifeedsci.2012.07.024
- AOAC, 2000. Official Methods of Analysis. Gaithersburg, USA.
- Arriola, K.G., Kim, S.C., Adesogan, A.T., 2011a. Effect of applying inoculants with heterolactic or homolactic and heterolactic bacteria on the fermentation and quality of corn silage. *J. Dairy Sci.* 94, 1511–6. doi:10.3168/jds.2010-3807
- Arriola, K.G., Kim, S.C., Staples, C.R., Adesogan, A.T., 2011b. Effect of applying bacterial inoculants containing different types of bacteria to corn silage on the performance of dairy cattle. *J. Dairy Sci.* 94, 3973–9. doi:10.3168/jds.2010-4070

- Arriola, K.G., Queiroz, O.C.M., Romero, J.J., Casper, D., Muniz, E., Hamie, J., Adesogan, A.T., 2015. Effect of microbial inoculants on the quality and aerobic stability of bermudagrass round-bale haylage. *J. Dairy Sci.* 98, 478–85. doi:10.3168/jds.2014-8411
- Bal, M. a, Coors, J.G., Shaver, R.D., 1997. Impact of the maturity of corn for use as silage in the diets of dairy cows on intake, digestion, and milk production. *J. Dairy Sci.* 80, 2497–503. doi:10.3168/jds.S0022-0302(97)76202-7
- Bal, M.A., Shaver, R.D., Jirovec, A.G., Shinnors, K.J., Coors, J.G., 2000. Crop Processing and Chop Length of Corn Silage : Effects on Intake , Digestion , and Milk Production by Dairy Cows 3563, 1264–1273.
- Ball, D., Collins, M., Lacefield, G., Martin, N., Mertens, D., Olson, K., Putnam, D., Undersander, D., Wolf, M., 2001. Understanding forage quality. American Farm Bureau Federation Publication.
- Bayatkouhsar, J., Tahmasebi, a. M., Naserian, a. a., 2011. The effects of microbial inoculation of corn silage on performance of lactating dairy cows. *Livest. Sci.* 142, 170–174. doi:10.1016/j.livsci.2011.07.007
- Begot, C., Desnier, I., Daudin, J.D., Labadie, J.C., Lebert, A., 1996. Recommendations for calculating growth parameters by optical density measurements. *J. Microbiol. Methods* 25, 225–232.
- Beresford, T., Williams, A., 2004. The Microbiology of Cheese Ripening. *Cheese Chem. Phys. Microbiol.* 1, Chemistry, Physics and Microbiology.
- Bisen, P.S., Debnath, M., Prasad, G.B.K.S., 2012. *Microbes: Concept and Applications*, 1st ed. Wiley-Blackwell, New Jersey.
- Black, J.G., Black, L.J., 2014. Growth and Culturing Bacteria, in: *Microbiology: Principles and Explorations*. Wiley-Blackwell, p. 146.
- Bolsen, K., Bonilla, D.R., Huck, G.L., Hart-Thakur, R.A., Young, M.A., 1996. Effect of a Propionic Acid Bacterial, in: *Cattlemen's Day, 1996*, Kansas State University. Manhattan, KS, pp. 78–81.
- Bolsen, K.K., Bolsen, R.E., 2015. The Silage Triangle and Important Practices in Managing Bunker , Trench , and Drive-over Pile Silos [WWW Document]. www.dairyweb.ca. doi:28.10.2015
- Bolsen, K.K., Lin, C., Brent, B.E., Feyerherm, A.M., Urban, J.E., Aimutis, W.R., 1992. Effect of Silage Additives on the Microbial Succession and Fermentation Process of Alfalfa and Corn Silages. *J. Dairy Sci.* 75, 3066–3083. doi:10.3168/jds.S0022-0302(92)78070-9
- Cai, Y., Benno, Y., Ogawa, M., Kumai, S., 1999. Effect of applying lactic acid bacteria isolated from forage crops on fermentation characteristics and aerobic deterioration of silage. *J. Dairy Sci.* 82,

520–6. doi:10.3168/jds.S0022-0302(99)75263-X

- Cai, Y., Kumai, S., 1994. The Proportion of Lactate Isomers in Farm Silage and the Influence of Inoculation with Lactic Acid Bacteria on the Proportion of L-Lactate in Silage. *Nihon Chikusan Gakkaiho* 65, 788–795. doi:10.2508/chikusan.65.788
- Chaia, A.P., Saad, A.M.S. De, Holgado, A.P.D.R., Oliver, G., 1995. Short-chain fatty acids modulate growth of lactobacilli mixed culture fermentations with propionibacteria in 26, 365–374.
- Chin, F.Y., Idris, A.B., 1999. Silage making activities of the department of veterinary services Malaysia, in: *FAO Electronic Conference on Tropical Silage*. Kuala Lumpur, pp. 1–3.
- Chizzotti, F.H.M., Pereira, O.G., Filho, S.C.V., Chizzotti, M.L., Rodrigues, R.T.S., Tedeschi, L.O., Silva, T.C., 2015. Does sugar cane ensiled with calcium oxide affect intake, digestibility, performance, and microbial efficiency in beef cattle? *Anim. Feed Sci. Technol.* 203, 23–32. doi:10.1016/j.anifeedsci.2014.12.014
- Coral, J., 2008. Propionic acid production by *Propionibacterium* sp. Using Low Cost Carbon Resources in Submerged Fermentation. Federal University of Parana.
- Dawson, T.E., Rust, S.R., Yokoyama, M.T., 1998. Improved Fermentation and Aerobic Stability of Ensiled , High Moisture Corn with the Use of *Propionibacterium acidipropionici* 1. *J. Dairy Sci.* 81, 1015–1021. doi:10.3168/jds.S0022-0302(98)75663-2
- Dehghani, M.R., Weisbjerg, M.R., Hvelplund, T., Kristensen, N.B., 2012. Effect of enzyme addition to forage at ensiling on silage chemical composition and NDF degradation characteristics. *Livest. Sci.* 150, 51–58. doi:10.1016/j.livsci.2012.07.031
- Driehuis, F., Oude Elferink, S.J.W.H., Van Wikselaar, P.G., 2001. Fermentation characteristics and aerobic stability of grass silage inoculated with *Lactobacillus buchneri*, with or without homofermentative lactic acid bacteria. *Grass Forage Sci.* 56, 330–343. doi:10.1046/j.1365-2494.2001.00282.x
- Dubois, M., Gilles, K. a., Hamilton, J.K., Rebers, P. a., Smith, F., 1956. Colorimetric method for determination of sugars and related substances. *Anal. Chem.* 28, 350–356. doi:10.1021/ac60111a017
- Dunière, L., Sindou, J., Chaucheyras-Durand, F., Chevallier, I., Thévenot-Sergentet, D., 2013. Silage processing and strategies to prevent persistence of undesirable microorganisms. *Anim. Feed Sci. Technol.* 182, 1–15. doi:10.1016/j.anifeedsci.2013.04.006
- Emde, R., Schink, B., 2007. Microbiology in a poised-potential amperometric culture system 153, 506–512.

- Erdman, R. a, Piperova, L.S., Kohn, R. a, 2011. Corn silage versus corn silage:alfalfa hay mixtures for dairy cows: effects of dietary potassium, calcium, and cation-anion difference. J. Dairy Sci. 94, 5105–10. doi:10.3168/jds.2011-4340
- Felipe, L., Cassoli, L.D., Carlos, L., Júnior, R., Carolina, A., Rodrigues, D.O., Machado, P.F., Quatro, E.D.E.C.C.E.M., Maturidade, E.D.E., 2008. In Situ Degradability of Corn Stover and Elephant-Grass Harvested at Four Stages of Maturity. Sci. Angiculture 65, 595–603.
- Filya, I., 2004. Nutritive value and aerobic stability of whole crop maize silage harvested at four stages of maturity. Anim. Feed Sci. Technol. 116, 141–150. doi:10.1016/j.anifeedsci.2004.06.003
- Filya, I., 2003. The effect of *Lactobacillus buchneri*, with or without homofermentative lactic acid bacteria, on the fermentation, aerobic stability, and ruminal degradability of wheat, sorghum, and maize silages. J. Appl. Microbiol. 95, 1080–1086. doi:10.1046/j.1365-2672.2003.02081.x
- Filya, I., Ashbell, G., Hen, Y., Weinberg, Z., 2000. The effect of bacterial inoculants on the fermentation and aerobic stability of whole crop wheat silage. Anim. Feed Sci. Technol. 88, 39–46. doi:10.1016/S0377-8401(00)00214-5
- Filya, I., Muck, R.E., Contreras-Govea, F.E., 2007. Inoculant effects on alfalfa silage: fermentation products and nutritive value. J. Dairy Sci. 90, 5108–14. doi:10.3168/jds.2006-877
- Filya, I., Sucu, E., 2010. The effects of lactic acid bacteria on the fermentation, aerobic stability and nutritive value of maize silage. Grass Forage Sci. 65, 446–455. doi:10.1111/j.1365-2494.2010.00763.x
- Flores-Galarza, R. A., Glatz, B. A., Bern, C. J., & Van Fossen, L.D., 1985. Preservation of high-moisture corn by microbial fermentation. J. Food Prot. 48, 407–415.
- Furuichi, K., Katakura, Y., Ninomiya, K., Shioya, S., 2007. Maximum production of 1,4-dihydroxy-2-naphthoic acid by fed-batch and anaerobic/aerobic culture of *Propionibacterium freudenreichii* ET-3. IFAC Proc. Vol. 10, 121–126.
- Giraud, E., Brauman, A., Keleke, S., Lelong, B., Raimbault, M., Biotechnologie, L. De, Orstom, C., Montferrand, D., Cedex, F.-M., 1991. Microbiology Biotechnology A /, p // ed Isolation and physiological study of an amylolytic strain of *Lactobacillus plantarum* 379–383.
- Harrison, J.H., Soderlund, S.D., Loney, K. a, 1989. Effect of inoculation rate of selected strains of lactic acid bacteria on fermentation and in vitro digestibility of grass-legume forage. J. Dairy Sci. 72, 2421–2426.

doi:10.3168/jds.S0022-0302(89)79376-0

Henderson, N., 1993. Silage additives. *Anim. Feed Sci. Technol.* 45, 35–56. doi:10.1016/0377-8401(93)90070-Z

Higginbotham, G.E., Mueller, S.C., Bolsen, K.K., DePeters, E.J., 1998. Effects of Inoculants Containing Propionic Acid Bacteria on Fermentation and Aerobic Stability of Corn Silage. *J. Dairy Sci.* 81, 2185–2192. doi:10.3168/jds.S0022-0302(98)75797-2

Hildebrand, B., Boguhn, J., Rodehutsord, M., 2011. Effect of maize silage to grass silage ratio and feed particle size on ruminal fermentation in vitro. *Animal* 5, 528–36. doi:10.1017/S1751731110002211

Holt, M.S., Neal, K., Eun, J.-S., Young, a J., Hall, J.O., Nestor, K.E., 2013. Corn silage hybrid type and quality of alfalfa hay affect dietary nitrogen utilization by early lactating dairy cows. *J. Dairy Sci.* 96, 6564–76. doi:10.3168/jds.2013-6689

Hugenschmidt, S., Schwenninger, S.M., Gnehm, N., Lacroix, C., 2010. Screening of a natural biodiversity of lactic and propionic acid bacteria for folate and vitamin B12 production in supplemented whey permeate. *Int. Dairy J.* 20, 852–857. doi:10.1016/j.idairyj.2010.05.005

Huisden, C.M., Adesogan, A.T., Kim, S.C., Ososanya, T., 2009. Effect of applying molasses or inoculants containing homofermentative or heterofermentative bacteria at two rates on the fermentation and aerobic stability of corn silage. *J. Dairy Sci.* 92, 690–7. doi:10.3168/jds.2008-1546

Jaichumjai, P., Valyasevi, R., Assavanig, A., Kurdi, P., 2010. Isolation and characterization of acid-sensitive *Lactobacillus plantarum* with application as starter culture for Nham production. *Food Microbiol.* 27, 741–8. doi:10.1016/j.fm.2010.03.014

Johnson, L.M., Harrison, J.H., Davidson, D., Mahanna, W.C., Shinnors, K., 2003. Corn silage management: effects of hybrid, chop length, and mechanical processing on digestion and energy content. *J. Dairy Sci.* 86, 208–31. doi:10.3168/jds.S0022-0302(03)73601-7

Johnson, L.M., Harrison, J.H., Davidson, D., Mahanna, W.C., Shinnors, K., Linder, D., 2002a. Corn silage management: effects of maturity, inoculation, and mechanical processing on pack density and aerobic stability. *J. Dairy Sci.* 85, 434–44. doi:10.3168/jds.S0022-0302(02)74092-7

Johnson, L.M., Harrison, J.H., Davidson, D., Swift, M., Mahanna, W.C., Shinnors, K., 2002b. Corn silage management III: effects of hybrid, maturity, and processing on nitrogen metabolism and ruminal fermentation. *J. Dairy Sci.* 85, 2928–47. doi:10.3168/jds.S0022-0302(02)74380-4

- Johnson, L.M., Harrison, J.H., Davidson, D., Swift, M., Mahanna, W.C., Shinnars, K., 2002c. Corn silage management II: effects of hybrid, maturity, and mechanical processing on digestion and energy content. *J. Dairy Sci.* 85, 2913–27. doi:10.3168/jds.S0022-0302(02)74379-8
- Keady, T.W., J., 1996. A review of the effects of molasses treatment of unwilted grass at ensiling on silage fermentation, digestibility and intake, and on animal performance. *Irish J. Agric. Food Res.* 35, 141–150.
- Khuntia, A., Chaudhary, L.C., 2002. Performance of male crossbred calves as influenced by substitution of grain by wheat bran and the addition of lactic acid bacteria to diet. *Asian-Australasian J. Anim. Sci.* 15, 188–194.
- Kleinschmit, D.H., Kung, L., 2006. The effects of *Lactobacillus buchneri* 40788 and *Pediococcus pentosaceus* R1094 on the fermentation of corn silage. *J. Dairy Sci.* 89, 3999–4004. doi:10.3168/jds.S0022-0302(06)72443-2
- Kristensen, N.B., Sloth, K.H., Højberg, O., Spliid, N.H., Jensen, C., Thøgersen, R., 2010. Effects of microbial inoculants on corn silage fermentation, microbial contents, aerobic stability, and milk production under field conditions. *J. Dairy Sci.* 93, 3764–74. doi:10.3168/jds.2010-3136
- Krizsan, S.J., Westad, F., Adnøy, T., Odden, E., Aakre, S.E., Randby, a T., 2007. Effect of volatile compounds in grass silage on voluntary intake by growing cattle. *Animal* 1, 283–92. doi:10.1017/S1751731107683773
- Kung Jr, L., 1993. A review on Silage Additives and Enzyme, in: 59th Minneapolis Nutrition Conference. Minneapolis; MN, pp. 121–135. doi:2/4/2014
- Kung, L., 2001. Silage fermentation and additives. *Sci. Tehcnology Feed Ind.*
- Kung, L., Chen, J.H., Kreck, E.M., Knutsen, K., Agricultural, D., Station, E., Biochemistry, A., 1993. Effect of Microbial Inoculants on the Nutritive Value of Corn Silage for Lactating Dairy Cows1 t d. *J. Dairy Sci.* 76, 3763–3770.
- Kung, L., Robinson, J.R., Ranjit, N.K., Chen, J.H., Golt, C.M., Pesek, J.D., 2000. Microbial populations, fermentation end-products, and aerobic stability of corn silage treated with ammonia or a propionic acid-based preservative. *J. Dairy Sci.* 83, 1479–86. doi:10.3168/jds.S0022-0302(00)75020-X
- Kung, L., Schmidt, R.J., Ebling, T.E., Hu, W., 2007a. The effect of *Lactobacillus buchneri* 40788 on the fermentation and aerobic

- stability of ground and whole high-moisture corn. J. Dairy Sci. 90, 2309–14. doi:10.3168/jds.2006-713
- Kung, L., Schmidt, R.J., Ebling, T.E., Hu, W., 2007b. The effect of *Lactobacillus buchneri* 40788 on the fermentation and aerobic stability of ground and whole high-moisture corn. J. Dairy Sci. 90, 2309–14. doi:10.3168/jds.2006-713
- Lemee, R., Lortal, S., Cesselin, B., Heijenoort, J. Van, 1994. Autolysis of *Propionibacterium freudenreichii* Involvement of an N-Acetylglucosaminidase in Autolysis of *Propionibacterium freudenreichii* CNRZ 725.
- Lind, H., Jonsson, H., Schnürer, J., 2005. Antifungal effect of dairy propionibacteria--contribution of organic acids. Int. J. Food Microbiol. 98, 157–65. doi:10.1016/j.ijfoodmicro.2004.05.020
- Liu, Q.H., Shao, T., Bai, Y.F., 2016. The effect of fibrolytic enzyme , *Lactobacillus plantarum* and two food antioxidants on the fermentation quality , alpha-tocopherol and beta-carotene of high moisture napier grass silage ensiled at different temperatures. Anim. Feed Sci. Technol. 221, 1–11. doi:10.1016/j.anifeedsci.2016.08.020
- Lynch, J.P., O'Kiely, P., Waters, S.M., Doyle, E.M., 2012. Conservation characteristics of corn ears and stover ensiled with the addition of *Lactobacillus plantarum* MTD-1, *Lactobacillus plantarum* 30114, or *Lactobacillus buchneri* 11A44. J. Dairy Sci. 95, 2070–80. doi:10.3168/jds.2011-5013
- Meeske, R., Basson, H., 1998. The effect of a lactic acid bacterial inoculant on maize silage. Anim. Feed Sci. Technol. 70, 239–247. doi:10.1016/S0377-8401(97)00066-7
- Mehnaz, S., Kowalik, T., Reynolds, B., Lazarovits, G., 2010. Growth promoting effects of corn (*Zea mays*) bacterial isolates under greenhouse and field conditions. Soil Biol. Biochem. 42, 1848–1856. doi:10.1016/j.soilbio.2010.07.003
- Merry, R.J., Davies, D.R., 1999. Propionibacteria and their role in the biological control of aerobic spoilage in silage. Lait 79, 149–164. doi:10.1051/lait:1999112
- Mhere, O., 2002. Forage Production and Conservation Manual Growing and ensiling.
- Mills, J. a, Kung, L., 2002. The effect of delayed ensiling and application of a propionic acid-based additive on the fermentation of barley silage. J. Dairy Sci. 85, 1969–75. doi:10.3168/jds.S0022-0302(02)74273-2
- Mohd Najib, M., 1993. Forage conservation for livestock smallholders in Malaysia, in: Third Meeting of the Regional Working Group on Grazing and Feed Resources of Southeast Asia. Khon Kaen,

Thailand, pp. 103–109.

- Mohd-Setapar, S.H., Abd-Talib, N., Aziz, R., 2012. Review on Crucial Parameters of Silage Quality. APCBEE Procedia 3, 99–103. doi:10.1016/j.apcbee.2012.06.053
- Muck, R.E., 1996. A lactic acid bacterial strain to improve aerobic stability of silages. Madison, Wisconsin.
- Muck, R.E., Kung, L.J., 1997. Effects of silage additives on ensiling, Silage: Field to Feedbunk, in: North American Conference (NRAES). pp. 187–199.
- Muller, V., 2001. Bacterial Fermentation. Encycl. Life Sci.
- Nkosi, B.D., Meeske, R., van der Merwe, H.J., Groenewald, I.B., 2010. Effects of homofermentative and heterofermentative bacterial silage inoculants on potato hash silage fermentation and digestibility in rams. Anim. Feed Sci. Technol. 157, 195–200. doi:10.1016/j.anifeedsci.2010.03.008
- Patrick Wincker, Falentin, H., Lortal, S., 2008. *Propionibacterium freudenreichii* ssp *shermanii* ATCC9614: A bacterium used in the production of Emmental [WWW Document]. Inst. Genomic. URL <http://www.genoscope.cns.fr/spip/Propionibacterium-freudenreichii,467.html#gaignaire> (accessed 10.13.15).
- Pedroso, A.F., Adesogan, A.T., Queiroz, O.C.M., Williams, S.K., 2010. Control of *Escherichia coli* O157:H7 in corn silage with or without various inoculants: efficacy and mode of action. J. Dairy Sci. 93, 1098–1104. doi:10.3168/jds.2009-2433
- Playne, M.J., 1985. Determination of ethanol, volatile fatty acids, lactic and succinic acids in fermentation liquids by gas chromatography. J. Sci. Food Agric. 36, 638–644. doi:10.1002/jsfa.2740360803
- Queiroz, O.C.M., Arriola, K.G., Daniel, J.L.P., Adesogan, a T., 2013. Effects of 8 chemical and bacterial additives on the quality of corn silage. J. Dairy Sci. 96, 5836–43. doi:10.3168/jds.2013-6691
- Ranjit, N.K., Kung, L., 2000. The effect of *Lactobacillus buchneri*, *Lactobacillus plantarum*, or a chemical preservative on the fermentation and aerobic stability of corn silage. J. Dairy Sci. 83, 526–35. doi:10.3168/jds.S0022-0302(00)74912-5
- Ranjit, N.K., Taylor, C.C., Kung Jr, L., 2002. Effect of *Lactobacillus buchneri* 40788 on the fermentation, aerobic stability and nutritive value of maize silage. Grass Forage Sci. 57, 73–81. doi:10.1046/j.1365-2494.2002.00304.x
- Rowghani, E., Zamiri, M.J., Khorvash, M., Abdollahipanah, A., 2008. The effects of *Lactobacillus plantarum* and *Propionibacterium*

acidipropionici on corn silage fermentation , ruminal degradability and nutrient digestibility in sheep. Iran. J. Vet. Res. 9, 308–315.

Seglar, B., 2003. Fermentation Analysis and Silage Quality Testing. Sci. York 119–136.

Sheperd, A.C., Kung, L., 1996. An Enzyme Additive for Corn Silage: Effects on Silage Composition and Animal Performance. J. Dairy Sci. 79, 1760–1766.

Sinclair, L. a., Jackson, M. a., Huntington, J. a., Readman, R.J., 2005. The effects of processed, urea-treated whole-crop wheat or maize silage and supplementation of whole-crop wheat on the performance of dairy cows. Livest. Prod. Sci. 95, 1–10. doi:10.1016/j.livprodsci.2004.10.013

Staples, C.R., 2003. Corn Silage for Dairy Cows, Sciences-New York. Florida.

Sun, Z.H., Liu, S.M., Tayo, G.O., Tang, S.X., Tan, Z.L., Lin, B., He, Z.X., Hang, X.F., Zhou, Z.S., Wang, M., 2009. Effects of cellulase or lactic acid bacteria on silage fermentation and in vitro gas production of several morphological fractions of maize stover. Anim. Feed Sci. Technol. 152, 219–231. doi:10.1016/j.anifeedsci.2009.04.013

Sutton, J.D., Abdalla, A.L., Phipps, R.H., Cammell, S.B., Humphries, D.J., 1997. The effect of the replacement of grass silage by increasing proportions of urea-treated whole-crop wheat on food intake and apparent digestibility and milk production by dairy cows. Anim. Sci. J. 65, 343–351. doi:http://dx.doi.org/10.1017/S1357729800008547

Tabacco, E., Piano, S., Revello-Chion, a, Borreani, G., 2011a. Effect of *Lactobacillus buchneri* LN4637 and *Lactobacillus buchneri* LN40177 on the aerobic stability, fermentation products, and microbial populations of corn silage under farm conditions. J. Dairy Sci. 94, 5589–98. doi:10.3168/jds.2011-4286

Tabacco, E., Righi, F., Quarantelli, a, Borreani, G., 2011b. Dry matter and nutritional losses during aerobic deterioration of corn and sorghum silages as influenced by different lactic acid bacteria inocula. J. Dairy Sci. 94, 1409–19. doi:10.3168/jds.2010-3538

Taylor, K. a. C.C., 1996. A simple colorimetric assay for muramic acid and lactic acid. Appl. Biochem. Biotechnol. 56, 49–58. doi:10.1007/BF02787869

Thierry, A., Deutsch, S.-M., Falentin, H., Dalmaso, M., Cousin, F.J., Jan, G., 2011. New insights into physiology and metabolism of *Propionibacterium freudenreichii*. Int. J. Food Microbiol. 149, 19–27. doi:10.1016/j.ijfoodmicro.2011.04.026

Thierry, A., Maillard, M., Yvon, M., 2002. Conversion of L -Leucine to

Isovaleric Acid by *Propionibacterium freudenreichii* TL 34 and ITGP23 68, 608–615. doi:10.1128/AEM.68.2.608

- Todar, K., 2012. Diversity of Metabolism in Procaryotes, in: Online Textbook of Bacteriology. www.textbookofbacteriology.net, Madison, Wisconsin, p. 8. doi:12102015
- Van Soest, P.J., Roberston, J.B., Lewis, B.A., 1991. Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. *J. Dairy Sci.* 74, 3583–3597.
- Voet, D., Voet, J.G., 2004. *Biochemistry*, 3rd Editio. ed. John Wiley & Sons, Inc., United States of America.
- Weinberg, Z.G., Ashbell, G., Bolsen, K.K., Pahlow, G., Hen, Y., Azrieli, a, 1995. The Effect of a Propionic-Acid Bacterial Inoculant Applied at Ensiling, with or without Lactic-Acid Bacteria, on the Aerobic Stability of Pearl-Millet and Maize Silages. *J. Appl. Bacteriol.* 78, 430–436.
- Weinberg, Z.G., Chen, Y., 2013. Effects of storage period on the composition of whole crop wheat and corn silages. *Anim. Feed Sci. Technol.* 185, 196–200. doi:10.1016/j.anifeedsci.2013.08.009
- Weinberg, Z.G., Muck, R.E., Weimer, P.J., Chen, Y., Gamburg, M., 2004. Lactic acid bacteria used in inoculants for silage as probiotics for ruminants. *Appl. Biochem. Biotechnol.* 118, 1–9. doi:10.1385/ABAB:118:1-3:001
- Woolford, M.K., 1984. *The Silage Fermentation*. Marcel Dekker, Inc, Madison Avenue, New York.
- Yitbarek, M., Tamir, B., 2014. Silage Additives: Review. *Open J. Appl. Sci.* 258–274.
- Yuan Kun, L., 2013. *Microbial Biotechnology: Principle and Applications*, Third. ed. Singapore.
- Yuan, X., Guo, G., Wen, A., Desta, S.T., Wang, J., Wang, Y., Shao, T., 2015. The effect of different additives on the fermentation quality, in vitro digestibility and aerobic stability of a total mixed ration silage. *Anim. Feed Sci. Technol.* 207, 41–50. doi:10.1016/j.anifeedsci.2015.06.001