



UNIVERSITI PUTRA MALAYSIA

***LACCASE PRODUCTION BY LOCALLY-ISOLATED *Trichoderma Sp.*
IS1037 USING RUBBERWOOD DUST FOR BIOBLEACHING OF SODA
BAMBOO PULPS***

SAJA MAHDEY JABER

FBSB 2017 30



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By

SAJA MAHDEY JABER

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia
Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

July 2017

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DEDICATION

This thesis is dedicated to my parents and my husband for their love and supports me. Without them, none of this would have been possible.

Saja Mahdey Jaber
July 2017



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

LACCASE PRODUCTION BY LOCALLY-ISOLATED *Trichoderma* Sp. IS1037 USING RUBBERWOOD DUST FOR THE BIOBLEACHING OF SODA BAMBOO PULPS

By

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July 2017

Chairman : Associate Professor Umi Kalsom Md Shah, PhD
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Conventional pulp bleaching of lignocellulosic materials using chlorine and its compounds leads to the generation of toxic compounds including chlorolignins. Therefore a study on biobleaching of lignocellulosic material was carried out as an alternative for pulp bleaching purpose. This study focused on the use of laccase from *Trichoderma* sp. IS1037 for bamboo pulp biobleaching. The laccase enzyme was produced from a local fungi isolate *Trichoderma* sp. IS1037. The possibility of using crude laccase for bleaching bamboo soda pulp by combining it with bleaching chlorine agents was also studied. A total of five native fungi was screened to determine the potential producer of laccase enzyme. The fungi isolate was identified by utilizing the Biolog Kit and molecular assay. Optimization of culture condition for laccase production was carried out using seven selected parameters known as carbon source, nitrogen source, temperature, pH, agitation speed, concentration of CuSO₄, and surfactant. The crude laccase produced was used for the pretreatment of bleaching process on unbleached bamboo soda pulp. Optimization of temperature, pH and reaction time on crude laccase pretreatment of bamboo pulp was investigated using response surface method (RSM) followed by chemical bleaching sequences D₁E_pD₂. In order to evaluate the decrease in lignin content, adsorbable organic halogens (AOX), hexenuronic acid (HexA) and chemical oxygen demand (COD) were tested. Solid phase extraction was conducted to extract AOX from effluent after bleaching process and HPLC was applied to determine AOX concentration. HexA was conducted using UV spectrophotometer. The result from screening demonstrated that laccase produced from locally isolated fungi identified as *Trichoderma* sp. IS1037 under optimized condition was 65.0 U/ml which is 11 fold higher than the production of the control culture 5.8 via submerged fermentation strategy. The optimum condition for laccase production was 30 °C, 4.5 and 150 rpm for temperature, initial pH media and agitation speed, respectively, using

rubberwood dust as a carbon source together with organic nitrogen source (peptone, yeast extract and malt extract). The effects of copper sulphate and Tween 80 were found significant in improving the extracellular laccase production. In pretreatment of bamboo pulp using laccase under optimized condition has decreased lignin by 64% and hemicellulose by 50% content was achieved at pH, temperature and reaction time of 4, 50 °C and 4 hours, respectively. XRD and FTIR analysis showed an increase in relative crystallinity of cellulose and delignification due to hydroxylation and exfoliation of amorphous regions through laccase pretreatment. In addition, there were significant result found in the reduction of AOX, COD, HexA and lignin by the enzymatic pretreatment. The AOX content of bamboo pulp was reduced by 29%, 34% for biobleaching and modified sample respectively. Moreover, HexA results revealed decrement in content with 1.44% to 1.42% for biobleaching and modified sample respectively. Therefore, the pretreatment of bamboo pulp by laccase has considerable benefit on reducing the organic compound of AOX, COD, HexA and chemical composition in different sequence bleaching. The outcomes presented in this study could contribute to the future study on the production of laccase enzyme from *Trichoderma* sp.IS1037 for various industrial applications including pulp and paper industry.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENGELUARAN LACCASE DARIPADA *Trichoderma* sp. IS1037 PENCILAN TEMPATON MENGGUNAKAN HABUK KAYU GETAH UNTUK BIO-PELUNTUR SODA PULPA BULUH

Oleh

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Julai 2017

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Pelunturan pulpa bahan lignoselulosa konvensional dan sebatianya membawa kepada penghasilan sebatian toksik termasuk klorolignins. Oleh itu kajian mengenai bio-peluntur dan bahan lignoselulosa telah dijalankan sebagai alternatif untuk tujuan pelunturan pulpa. Kajian ini tertumpu kepada penggunaan enzim laccase daripada *Trichoderma* sp. IS1037 untuk bio-pelunturan pulpa buluh. Enzim laccase dihasilkan daripada isolate kulat *Trichoderma* sp. IS1037. Kemungkinan menggunakan laccase mentah untuk pelunturan pulpa soda buluh dengan menggabungkan ia dengan agen klorin juga dikaji.

Dalam kajian ini, lima kulat tempatan telah disaring untuk menentakar kulat yang berpotensi mengeluarkan enzim laccase. Pencilan kulat telah dikenal pasti dengan menggunakan Kit Biolog dan cerakin molekul. Pengoptimuman keadaan kultur untuk pengeluaran laccase telah dijalankan dengan menggunakan tujuh parameter terpilih yang dikenali sebagai sumber karbon, sumber nitrogen, suhu, pH, kelajuan pergolakan, kepekatan CuSO₄, dan surfaktan. Laccase mentah yang dihasilkan telah digunakan untuk rawatan awal proses pelunturan pada pulpa buluh soda yang belum diluntur. Pengoptimuman suhu, pH dan masa tindak balas kepada rawatan awal laccase mentah ke atas pulpa buluh dikaji menggunakan kaedah gerak balas permukaan (RSM) diikuti dengan urutan pelunturan kimia D₁EpD₂. Untuk menilai penurunan kandungan lignin, halogen organik boleh serap (AOX), asid hexenuronik (Hexa) dan permintaan oksigen kimia (COD) telah diujid. Pengekstrakan fasa pepejal telah dijalankan untuk mengeluarkan AOX daripada pelunturan efluen selepas proses pelunturan dan HPLC telah dijalankan untuk menentukan kepekatan AOX. HexA ditentukan dengan menggunakan spektrofotometer UV. Hasil daripada saringan menunjukkan laccase yang dihasilkan dari kulat asing tempatan dikenal pasti sebagai *Trichoderma* sp. IS1037 pada keadaan optimum adalah 65.0 U / ml iaitu 11

kali ganda lebih tinggi daripada pengeluaran daripada kultur kawalan 5.8 melalui strategi fermentasi tenggelam. Keadaan optimum untuk pengeluaran laccase adalah 30 ° C, 4.5 dan 150 rpm untuk suhu, media pH awal dan kelajuan pergolakan, masing-masing, dengan menggunakan habuk kayu getah sebagai sumber karbon bersama-sama dengan sumber organik nitrogen (pepton, ekstrak yis dan ekstrak malt). Kesan kuprum sulfat dan Tween 80 didapati ketara dalam meningkatkan pengeluaran laccase luarsel. Dalam rawatan awal pulpa buluh, menggunakan laccase bawah keadaan optimum telah menurunkan lignin sebanyak 64% dan hemiselulosa dengan kandungan 50% telah dicapai pada pH, suhu dan masa tindak balas 4, 50 ° C dan 4 jam, masing-masing. Analisis XRD dan FTIR menunjukkan peningkatan dalam penghabluran relatif selulosa dan delignifikasi kerana penghidroksilan dan pengelupasan kawasan amorfus melalui rawatan awal laccase. Di samping itu, terdapat hasil yang ketara dijumpai di dalam pengurangan AOX, COD, Hexa dan lignin daripada prarawatan bio-pelunturan itu. Kandungan AOX pulpa buluh telah dikurangkan sebanyak 29%, 34%, untuk bio-pelunturan dan sampel yang diubahsuai, masing-masing. Selain itu, keputusan Hexa menunjukkan penurunan dalam kandungan dengan 1.42%, dan 1.14%, untuk bio-pelunturan dan sampel yang diubahsuai, masing-masing. Oleh itu, rawatan awal pulpa bambu oleh laccase mempunyai manfaat yang besar dalam mengurangkan kompaun organik AOX, COD, Hexa dan komposisi kimia dalam urutan pelunturan yang berbeza. Hasil dibentangkan dalam kajian ini boleh menyumbang kepada kajian masa depan tentang pengeluaran enzim laccase dari *Trichoderma* sp. IS1037 untuk pelbagai aplikasi industri termasuk industri pulpa dan kertas.

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Saja Mahdey Jaber
July 2017

I certify that a Thesis Examination Committee has met on 18 July 2017 to conduct the final examination of Saja Mahdey Jaber on her thesis entitled "Laccase Production by Locally-Isolated *Trichoderma* Sp. IS1037 Using Rubberwood Dust for Biobleaching of Soda Bamboo Pulps" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

ABTS	2, 2-azino-bis (3-ethylbenzothiazoline-6-sulphonic acid)
AOX	Absorbed organic halogen
COD	Chemical oxygen demand
CTMP	Chemi-thermo-mechanical pulping
D	Sodium chlorite
ECF	elemental chlorine-free
E _p	Alkaline
FTIR	Fourier Transform Infrared Spectroscopy
HexA	Hexeneuronic acid
HPLC	High Performants Liquid Chromatography
KH ₂ PO ₄	Potassium Dihydrogen Phosphate
L	Liter
LiP	Lignin peroxidase
LMS	Laccase mediator system
LWM	Low molecular weight
mL	Milliliter
MnP	Manganese peroxidase
MgSO ₄ .7H ₂ O	Magnesium Sulfate Heptahydrate
NaOH	Sodium Hydroxide
Nm	Nanometer
mg/mL	milligram/milliliter
Rpm	revolutions per minute
RSM	Response surface method
SEM	Scanning Electron Microscopy
SmF	Submerged fermentation
SSF	Solid state fermentation
U/g	Unite/gram
U/mL	Unite/milliliter
µL	Microliter
XRD	X-ray Diffraction

CHAPTER 1

INTRODUCTION

1.1 General Background

Nowadays, laccase has become increasingly important as an eco-friendly technology in the pulp and paper industry and lead to greater profitability for the industry. With the growing demands for laccase applications in industrial processes, there should be expanding research of microorganisms with high laccase activities and discovering novel laccases with enhanced stability and superior catalytic properties (Desai and Nityanand, 2011; Kiiskinen *et al.* 2004). Although studies about laccase activity in lignin degradation have been reported, laccases cannot catalase the lignin degradation and model compounds without combination with redox mediator compounds, which is increase the range of compounds for oxidation such as non-phenolic substrates. In addition, Laccase–mediator systems enhance the interaction between the laccase and non-phenolic fractions of lignin in wood pulp due to ability to diffuse into cellulose fibres (Barreca *et al.*, 2004; Galli and Gentili, 2004; Bajpai *et al.*, 2006; Fillat and Roncero, 2010).

The paper and pulp industry involves the process of bleaching pulp which currently uses mostly chemicals such as chlorine dioxide and alkaline extraction (Zhao *et al.* , 2010) which is responsible for generating adsorbed organic halides (AOX) and other chlorine compounds due to chlorine reacts with lignin (Comlekcioglu *et al.*, 2014). Traditionally, using a high amount of chlorine in the bleaching process results high brightness paper and large quantity of harmful substances in paper mill effluents into rivers and ponds, which is a potential hazard and contributes to environmental pollution (Morselli *et al.*, 2014; Yao *et al.*, 2016). The best way to solve this problem is to adopt an effective bleaching method with low pollution and that is enzymatic bleaching (Pei *et al.*, 2016; Gangwar *et al.*, 2016). Enzyme technology is gaining global recognition due to the fact which it is high catalytic efficiency, high substrate specificity, shorter reaction time, low energy input, mild reaction conditions and environmentally friendly (Hanana *et al.*, 2015; Wong, 2016). Therefore, using laccase treatment in bleaching processes is recommended for the pulp and paper industry to significantly reduce the consumption of chlorine dioxide and generation organic halides, at the same time achieve enhanced paper quality.

Various agricultural wastes have been used as fermentation substrates including wheat bran, rice straw, bagasse, corncobs, banana peels, sunflower receptacle, apricot seed shell, and saw dust in order to produce laccase enzyme (Jesus *et al.*, 2015; Hendro *et al.*, 2012; Emre and Özfer, 2013). However, there have been some limited studies that used rubber wood dust as a carbon source for laccase production.

Laccases (EC 1.10.3.2) are derived from a family of blue multi-copper oxidases usually associated with polymerisation–depolymerisation reactions that induce phenols and aromatic or aliphatic amines to be oxidised to the corresponding reactive radicals using oxygen as electron acceptor (Riva, 2006). Besides plants, they are also distributed in fungi, bacteria and insects (Janusz, *et al.*, 2013; Arca-Ramos *et al.*, 2015). In fungi, there has been successful isolation of these enzymes from Ascomycetes, Deuteromycetes, and Basidiomycetes (Brijwani *et al.*, 2010), which secrete laccases that contribute to lignin degradation and function essentially in the carbon cycle in the degradation of lignocellulosic material. Many achievements have been reported, but for most of the industrial processes, there is still an obstacle to wider application of laccases due to the high cost of production (Osma *et al.*, 2011; Liu *et al.*, 2013), which significantly affects production efficiency. Therefore, many researches have been carried out and attempted to produce laccases more efficiently, such as by optimization of fermentation conditions, isolation, and identification of new potential high-level laccase producing strains.

The production of laccase in fungi is low and is unable to meet the demand for practical applications in industry and biotechnology. As a result, there has been much work done to develop a laccase production process that is both cost-effective and efficient. Towards this end, attention has been focused on producing laccase from fungi by utilizing aromatic compounds including xylidine, ferulic acid, and veratric acid which have been found to enhance laccase production during the fermentation process (Yang, *et al.*, 2013; De Souza *et al.*, 2004). Copper has also been known to promote laccase activity in the majority of fungi, in *Trametes versicolor* (Collins and Dobson, 1997) *Pleurotus sajor-caju*, (Soden and Dobson, 2001) *Ceriporiopsis subvermispora* (Alvarez, *et al.*, 2009) *Pleurotus ostreatus* (Faraco, *et al.*, 2003) and *Trametes pubescens*, (Galhaup, *et al.*, 2002).

Applying a statistical process such as Response Surface Method (RSM) is very useful in the optimisation of the biobleaching process of unbleached soda pulp, by determining the effects and interactions of pH, temperature, and reaction time on laccase pretreatment, as this is crucial for the reduction of chemical composition content, especially lignin in bamboo. An RSM comprises an empirical modelling system that assesses how a group of experimentally controllable variables is related to an observed response. Limited research has been carried out to examine the benefits of RSM in optimizing laccase pre-treatment of the chemical composition in the bamboo bleaching process.

With increasing scarcity of natural resources, and environmental pollution has raised to demand for natural fiber non wood in paper industry as alternative raw materials for this intent in the future. Bamboo (*Gigantochloa scortechini*) is a hard non wood plant with superior physical and mechanical properties, it's a rapid growth rate and can give high yields. Therefore, bamboo has long been widely used for constructional purpose and for the pulp and paper in industry (Liu *et al.*, 2012; Abdul Khalil *et al.*, 2014). On the other hand, the natural bamboo fibers have great properties such as hygroscopic, which is mean it is able to obtain water from the

environment and hold onto it (Erakhrumen and Ogunsanwo 2009). This is important because in the chemical pulping processes, cooking liquor needs to penetrate throughout the chips of a sample such as bamboo (Moradbak *et al.* 2016). However, not much research has been focused on the use of crude laccase on soda bamboo pulp bleaching. Therefore, to assess the suitability of utilizing cured laccase, it is suggested that there be a sequential pretreatment before standard elemental chlorine-free (ECF) bleaching that acts on bamboo soda pulp to determine the optical properties prior to and following an accelerated ageing process of the resulting paper sheets.

1.2 Problem Statement

The increasing volume and chemical complexity such as chlorine in the bleaching methods from pulp and paper industry is seriously represented environmental problem today. Chlorine dioxide is the most important bleaching chemical in the elemental chlorine-free (ECF) bleaching of pulp. It has good selectivity for lignin removal, but there will be some adsorbable organic halogen (AOX) formed in the bleaching process (Pei *et al.*, 2016; Zhu *et al.*, 2016). As a result of reaction between residual lignin from wood fibres and chlorine/alkaline compounds used for bleaching could be discharging these chemical compounds and other toxic as wastewater reason carcinogenic and mutagenic (Gangwar *et al.* 2016) beside that removal of these compounds are costly. To overcome this problem, there is option of cost effective strategy through using enzymatic stage integrated into a bleach sequence (Moldes *et al.*, 2010; Gangwar *et al.* 2016). Many researchers have use enzyme in pulping and bleaching process such as xylanase and laccase (Moldes *et al.*, 2010; Xu *et al.*, 2013; Gangwar *et al.* 2015; Pei *et al.*, 2016; Zhu *et al.*, 2016) with different type of non-wood.

Laccases are widely distributed multicopper oxidases, and have been isolated from fungal strains belonging to Ascomycetous, Deuteromyctous and Basidiomycetous family. Many of the previous studies have been focused on lignin degrading enzymes from white rot-fungi such as *Phanerochaete chrysosporium*, *Trametes versicolor*, and *Echinodontium taxodii* 2538 (Shi, *et al.*, 2014; Kong *et al.*, 2015) however, studying the lignolytic enzymes from *Trichoderma* spp. recently there has been a growing interest due to exhibit better lignolytic systems with characterization of their substrates, copper content or amino acid sequence, for use in various biotechnological applications (D'Souza *et al.*, 1999; Chakroun *et al.*, 2010; Khambhaty *et al.*, 2015). Thus, it was of importance to know *Trichoderma* strains with phenol oxidase activity has high efficiency for selective lignin degradation in the biobleaching process, causing modifications in lignin structures and significant decreases organic compounds ratios. In addition, pretreatment with laccase can improve reduction of chlorine dioxide and the other bleaching chemicals usage. Beside that improved paper properties such as brightness, whiteness, burst index, tensile and tear index values (Thakur *et al.* 2012; Andreu and Vidal 2013; Song *et al.*, 2016).

1.3 Objectives

This current study was conducted to determine the potential of locally available fungi-produced laccase enzyme with optimum culture conditions of production and to evaluate the effectiveness of laccase as the pulp biobleaching agent. The specific objectives of this study were:

- 1- To screen and select the potential fungal from local isolate with ability to produced laccase.
- 2- To determine the optimum culture conditions for laccase production from *Trichoderma* sp. in submerged fermentation and to characterize the laccase produced.
- 3- To optimize the enzymatic ECF bleaching of soda bamboo using response surface method (RSM) viz pH, temperature, and reaction time.
- 4- To evaluate the effect of laccase pretreatment on bamboo fibre to reduce the chemical dosage in (ECF) bleaching sequences.

REFERENCES

- Abdul Khalil, H. P. S., Noriman, N. Z., Ahmad, M. N., Ratnam, M. M., Fuaad, N. A. (2007). Polyester composites filled carbon black and activated carbon from bamboo (*Gigantochloa scortechinii*): Physical and mechanical properties. *Journal of Reinforced Plastics and Composites*. 26. 3
- Abdul Khalil, H. P. S., Bhat, I. U. H., Jawaid, M., Zaidon, A., Hermawan, D., Hadi, Y. S. (2012). Bamboo fibre reinforced biocomposites: A review. *Materials and Design*. 42: 353-368.
- Abdul Khalil, H. P. S., Hossain, S.MD., Rosamah, E., Norulaini, N.N.A, Peng, C.L., Asniza, M., Davoudpour, Y., Zaidul, I.S.M.(2014). High-Pressure Enzymatic Hydrolysis to Reveal Physicochemical and Thermal Properties of Bamboo Fiber Using a Supercritical Water Fermenter. *BioResources* 9(4), 7710-7720.
- Abd Razak, O, Hashim, M.N. (1992). Vegetative propagation of selected Malaysian Bamboo, In *Towards the Management, Conservation, Marketing and Utilization of Bamboo*, proceedings of the National Bamboo seminar 1, Kuala Lumpur, Malaysia, Nov 2-4. Wan Razali, W.M. and Aminuddin, M.(Eds.), pp.46-57. Forest Research Institute Malaysia.
- Ahmadu, A. F. (2012). *Bioethanol Production Using Lignocellulose Biomass Hydrolysate from Indigenous Fungi Fermentation*, Master's Thesis. Institute of Bioscience, Universiti Putra Malaysia.
- Ahmed, I. E., Nora M. E., Aymen S. Y., Magdy A. A. (2015). Laccase production by *Pleurotus ostreatus* and its application in synthesis of gold nanoparticles. *Biotechnology Reports* 5: 31–39.
- Agematu, H., Tsuchida, T., Kominato, K. (1993). Enzymatic dimerization of penicillin X. *Journal of Antibiotics*. 46 (1): 141–148.
- Alcalde, M. (2007). Laccases: biological functions, molecular structure and industrial applications. In: Polaina J and MacCabe AP. *Industrial Enzymes*. 461–476.
- Andreu, G., Vidal, T. (2013). Laccase from *Pycnoporus cinnabarinus* and phenolic compounds: Can the efficiency of an enzyme mediator for delignifying kenaf pulp be predicted? *Bioresource Technology* 131 (2013) 536–540.
- Alves, E., Lucas, G. C., Pozza, E. A., Alves, M. C. (2013). *Scanning Electron Microscopy for Fungal Sample Examination*. Laboratory Protocols in Fungal Biology Part of the series Fungal Biology. 133-150.
- Alwani, M. S., Khalil, H. P. S. A., Sulaiman, O., Islam, M. N., Dungani, R. (2014). An approach to using agricultural waste fibres in biocomposites application: Thermogravimetric analysis and activation energy study. *BioResources*. 9(1): 218- 230.
- Amanda, F., Josep F. C., Teresa, V. (2010). A new approach to the biobleaching of flax pulp with laccase using natural mediators. *Bioresource Technology*. 101: 4104–4110.

- Andreu, G., Vidal, T. (2013). Laccase from *Pycnoporus cinnabarinus* and phenolic compounds: Can the efficiency of an enzyme mediator for delignifying kenaf pulp be predicted? *Bioresource Technology*. 131: 536–540.
- Ang, K. N. I. (2007). *Production of Laccase by a Locally Isolated Fungus for Biodegradation of Selected Agro Wastes*. Master's Thesis, Biotechnology and Biomolecular Sciences, University Putra Malaysia.
- Ang, T. N., Ngoh, G. C., Chua A. S. M. (2011). A quantitative method for fungal ligninolytic enzyme screening studies. *Asia-Pacific Journal. Chemistry Engineering*. 6(4): 589-595.
- Ang TN, Ngoh GC, Chua ASM (2010). A quantitative method for fungal ligninolytic enzyme screening studies. *Asia-Pacific Journal of Chemical Engineering*. 6 (4): 589–595
- Angel, T.M. (2002). Molecular biology and structure-function of lignin-degrading heme peroxidases. *Enzyme Microbiology and Technology*. 30:425–444.
- Anita, S., Somvir, B., Narsi, R. B., Namita, S. (2010). Laccase production By *Aspergillus heteromorphus* using distillery spent wash and lignocellulosic biomass, *Journal of Hazardous Materials*. 176: 1079-1082.
- Anwar, Z., Gulfranz, M., Irshad, M. (2014). Agro-industrial lignocellulosic biomass a key to unlock the future bio-energy. *Journal of Radiation Research and Applied Sciences*. 7 (2): 163–173.
- Aracri, E., Colom, F. J., Vidal, T. (2009). Application of laccase-natural mediator systems to sisal pulp: An effective approach to biobleaching or functionalizing pulp fibres? *Bioresource Technology*. 100: 5911–5916.
- Arca-Ramos, A, Eibes, G. Feijoo, G. Lema, J. M., Moreira, M.T. (2015). Potentiality of a ceramic membrane reactor for the laccase-catalyzed removal of bisphenol A from secondary effluents. *Applied Microbiology and Biotechnology*. 1–10.
- Arora, D. S., Gill, P. K. (2000). Laccase production by some white rot fungi under different nutritional conditions. *Bioresource Technology*. 73: 283-285.
- Arora, D.S. Sharma, R.K. (2010). Ligninolytic Fungal Laccases and Their Biotechnological Applications. *Appl Biochem Biotechnol*. 160: 1760–1788.
- Asgher, M., Bhatti, H. N., Ashraf, M., Legge, R. L. (2008). Recent developments in biodegradation of industrial pollutants by white rot fungi and their enzyme system. *Biodegradation*. 19 (6) : 771–783.
- Assavaning, A., Amornikitticharoen, B., Ekpaisal, N., Meevootisom, V., Flegel, T. W. (1992). Isolation, characterization and function of laccase from *Trichoderma*. *Applied Microbiology and Biotechnology*. 38 (2): 198- 202.
- Asgher, M., Ahmad, Z., Iqbal, H. M. N. (2013). Alkali and enzymatic delignification of sugarcane bagasse to expose cellulose polymers for saccharification and bio-ethanol production. *Industrial Crops and Products*. 44: 488-495
- Babot, E.D., Rico A., Rencoret, J., Kalum, L., Lund, H., Romero, J., del R ó. J. C., Martínez, Á. T., Gutiérrez, A. (2011). Towards industrially-feasible delignification and pitch removal by treating paper pulp with *Myceliophthora*

- thermophile laccase and a phenolic mediator. *Bioresource Technology*. 102:6717–6722.
- Badar, S. Farooqi, I.H. (2012). Pulp and Paper Industry—Manufacturing Process, Wastewater Generation and Treatment. In *Environmental Protection Strategies for Sustainable Development*. Malik, A., Grohmann, E., Eds.; Springer: Dordrecht, The Netherlands, 397–436.
- Bajpai, P. (2004). Biological bleaching of chemical pulps. *Critical Reviews in Biotechnology*. 24: 51-58.
- Bajpai, P., Anand, A., Sharma, N., Mishra, S.P., Bajpai, P.K., Lachenal, D. (2006). Enzymes improve ECF bleaching of pulp. *Bioresources*. 1: 34–44.
- Bajpai, P. (2010). Recent developments in cleaner production in *Environmentally Friendly production of pulp and paper*, P. Bajpai (ed.), John Wiley & Sons, Inc., Hoboken, NJ, pp. 264-344.
- Bajpai, P. (2012). *Biotechnology for Pulp and Paper Processing*. Chapter 2. Springer. 1-9.
- Bakkiyaraj, S., Aravindan, R., Arrivukkarasan, S., Viruthagiri, T. (2013). Enhanced laccase production by *Trametes hirsuta* using wheat bran under submerged fermentation. *International Journal of ChemTech Research*. 5(3): 1224-1238.
- Baldrian, P. (2006). Fungal laccases—Occurrence and properties. *FEMS Microbiology Reviews*. 30(2): 215-242.
- Balaraju, K., Kyungseok, P., Shamarao, J., Kaviyarasan, V. (2010). Production of cellulase and laccase enzymes by *Oudemansiella radicata* using agro wastes under solid-state and submerged conditions. *Research in Biotechnology*. 1: 21-28.
- Balsiger, J., Bahdan, J., and Whiteman, A. (2000). *The Utilization, Processing and Demand for Rubberwood as a Source of Wood Supply*. APFC-Working Paper No. APFSOS/WP/50, FAO, Bangkok.
- Barreca, A. M., Sjögren, B., Fabbrini, M., Galli, C., Gentili, P. (2004). Catalytic efficiency of some mediators in laccase-catalyzed alcohol oxidation. *Biocatalysis And Biotransformation*. 22: 105–112.
- Barreto Xavier, A. M. R., Mora Tavares, A. P., Ferreira, R., Amado, F. (2007). *Trametes versicolor* growth and laccase induction with by-products of pulp and paper industry. *Electronic Journal of Biotechnology*. 10: 444-451.
- Bermek, H., Yazici, H., Oztürk, H., Tamerler, C., Jung, H., Li, K., Brown, K. M., Ding, H., Xu, F. (2004). Purification and characterization of manganese peroxidase from wood degrading fungus *Trichophyton rubrum* LSK-27. *Enzyme and Microbial Technology*. 35: 87-92.
- Bezerra, M. A., Ricardo, E. S., Eliane, P. O., Leonardo, S. V., Luciane, A. E. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry *Talanta*. 76: 965–977.
- Bertrand, G. (1894). Sur le latex de l'arbre à laque. *Bull. Soc. Chim.* 11: 717–721.
- Berthet, S., Thevenin, J., Baratiny, D., Demont-Caulet, N., Debeaujon, I., Bidzinski, P., Leple, J.-C., Huis, R., Hawkins, S., Gomez, L.-D., Lapiere, C., Jouanin,

- L. (2012). Role of plant laccases in lignin polymerization. In: L.J., Lapiere, C. (Eds.). *Advances in Botanical Research*. 61: 145–172. (Chapter 5).
- Bhattacharya SS, Garlapati VK, Banerjee R.(2011). Optimization of laccase production using response surface methodology coupled with differential evolution. *New Biotechnology*. 28(1),31-39.
- Birhanli, E., Yesilada, O. (2006). Increased production of laccase by pellets of *Funalia trogii* ATCC 200800 and *Trametes versicolor* ATCC 200801 in repeated batch mode. *Enzyme and Microbial Technology*. 39: 1286-293.
- Biermann, C. J. (1996a). *Wood and fiber fundamentals*. Handbook of Pulping and Paper making. Academic. San Diego. p 13.
- Biermann, C. J. (1996b). *Pulping fundamentals*. Handbook of Pulping and Paper making. Academic. San Diego. p 55.
- Bjorklund, M., Germgard, U., Basta, J. (2004). Formation of AOX and OCl in ECF bleaching of birch pulp. *TAPPI* 3(8), 7-12.
- Boran, F., Yeşilada, O. (2011). Enhanced production of laccase by fungi under solid substrate fermentation condition. *BioResources*. 6(4): 4404-4416.
- Boruah, P., Dowarah, P., Hazarika, R., Yadav, A., Barkakati, P., Goswami, T. (2016). Xylanase from *Penicillium meleagrimum* var. *viridiflavum* - A potential source for bamboo pulp bleaching. *Journal of Clean Production*. 116: 259-267.
- Bourbonnais, R., Paice, M. G., Freiermuth, B., Bodie, E., Borneman, S. (1997). Reactivities of various mediators and laccases with kraft pulp and lignin model compounds. *Applied and Environmental Microbiology*. 63, 4627–4632.
- Bradford M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein. Dye Binding Analytical Biochemistry. 72: 248-254.
- Brage, C., Eriksson, T., Gierer, J., Eriksson, T., Gierer, J. (1991). Reactions of chlorine dioxide with lignins in unbleached pulps part I. *Holzforschung* 45(45), 23-30.
- Brijwani, K., Rigdon, A., Vadlani, P.V. (2010). Fungal laccases: Production, function, and applications in food processing. *Enzyme Research* .10: 149-748
- Buswell, J. A., Cai, Y., Chang, S. T. (1995). Effect of nutrient nitrogen and manganese on manganese peroxidase and laccase production by *Lentinula (Lentinus) edodes*. *FEMS Microbiology Letters*. 128(1): 81-88.
- Camarero, S., David, I., Angel, T., Javier, R., Ana, G., Jose, C. (2007). Paper pulp delignification using laccase and natural mediators. *Enzyme and Microbial Technology*. 40: 1264-1271.
- Cadena, EM., Vidal, T., Torres, AL. (2010). Can the laccase mediator system affect the chemical and refining properties of the eucalyptus pulp? *Bioresource Technology* 101: 8199–8204.

- Cañas, A., and Camarero, S. (2010) Laccases and their natural mediators: Biotechnological tools for sustainable eco-friendly processes. *Biotechnology Advances*. 28(6):694-705.
- Champagne, P., Ramsay, J. A. (2007). Reactive blue 19 decolouration by laccase immobilized on silica beads. *Applied Microbiology and Biotechnology*. 77(4): 819-823.
- Changwei, Z., Guangwen, B., Shun, H., (2016). Optimization of laccase production in the white-rot fungus *Pleurotus ostreatus* (ACCC 52857) induced through yeast extract and copper. *Biotechnology and Biotechnological equipment*. 30(2): 270-276
- Chakroun, H., Mechichi, T., Martinez, M. J., Dhouib, A., Sayadi, S. (2010). Purification and characterization of a novel laccase from the ascomycete *Trichoderma atroviride*: Application on bioremediation of phenolic compounds. *Process Biochemistry*. 45(4):507-513.
- Chaparro, T.R., Botta, C.M. Pires, E.C. (2010). Toxicity and recalcitrant compound removal from bleaching pulp plant effluents by an integrated system: anaerobic packed bed bioreactor and ozone. *Water Science & Technology-WST*. 61 (1): 199-205.
- Choinowski, T., Blodig, W., Winterhalter, K. H., Piontek, K. (1999). The crystal structure of lignin peroxidase at 1.70 Å resolution reveals a hydroxy group on the C β of tryptophan 171: a novel radical site formed during the redox cycle. *Journal of Molecular Biology*. 286: 809–827
- Collins, P. J., Dobson, A. (1997). Regulation of laccase gene transcription in *Trametes versicolor*. *Applied Environment Microbiology*. 63: 3444-3450.
- Comlekcioglu, U., Tutus, A., Cicekler, M., Gunes, M., and Aygan, A. (2014). Application of recombinant xylanase from *Orpinomyces* sp. in elemental chlorinefree bleaching of kraft pulps. *Romanian Biotechnological Letters*. 19 (1): 8941-8950.
- Costa, S. M., Goncalves, A. R., Esposito, E. (2005). *Ceriporiopsis Subvermispora* used in delignification of sugarcane bagasse prior to soda/antraquinone pulping. In: Davison BH, Evans BR, Finkelstein M and McMillan JD. Twenty-Sixth Symposium on Biotechnology for Fuels and Chemicals. *Humana Press*. p. 695–706.
- Couto, S. R., Rosales, E., and Sanromán, M. A. (2006). Decolourization of synthetic dyes by *Trametes hirsuta* in expanded-bed reactors. *Chemosphere*. 62(9): 1558-1563.
- Couto, R.S., Sanroman, M.A., (2005). Application of solid-state fermentation to ligninolytic enzyme production. *Biochemical Engineering Journal*. 22:211–219.
- Couto, S., Toca-Herrera, J. (2006). Laccases in the textile industry. *Biotechnology and Molecular Biology Reviews*. 1:115-120.
- Cui, Y. Q., van der Lans, R. G. J. M., Luyben, K. C. A. M. (1997). Effect of agitation intensities on fungal morphology of submerged fermentation. *Biotechnology and Bioengineering*. 55(5): 715-726.

- Cui, Y. Q., van der Lans, R. G. J. M., & Luyben, K. C. A. M. (1998). Effects of dissolved oxygen tension and mechanical forces on fungal morphology in submerged fermentation. *Biotechnology and Bioengineering*. 57(4): 409-419.
- Da Re, V., Papinutti, L., Villalba, L., Forchiassin, F., Levin, L. (2008). Preliminary studies on the biobleaching of loblolly pine kraft pulp with *Trametes trogii* crude extracts. *Enzyme Microbiology. Technology*. 43, 164–168.
- Dai, Y., Song, X., Gao, C., He, S., Nie, S., Qin, C. (2016). Xylanase-aided chlorine dioxide bleaching of bagasse pulp to reduce AOX formation. *BioResources* 11(1), 3204-3214.
- D'Souza, T.M., Merritt, C.S., Reddy, C.A.(1999). Lignin-modifying enzymes of the white rot basidiomycete *Ganoderma lucidum*. *Applied and Environmental Microbiology* 65(12):5307-5313.
- DeSouza, C. G. M., Tychanowicz, G.K., De Souza, D. F., Peralta, R.M. (2004). Production of laccase isoforms by *Pleurotus pulmonarius* in response to presence of phenolic and aromatic compounds. *Journal of Basic Microbiology*. 44: 129-136.
- Desia, S.S and Nityanand, C.(2011) Microbial Laccases and their Applications: A Review. *Asian Journal of Biotechnology*. 3 (2) 98 – 124.
- DeJong E, De Vries FP, Field JA, Van Der Zwan RP, De Bont JAM (1992). Isolation and screening of basidiomycetes with high peroxidative activity. *Mycological Research*. 96: 1098-1104.
- Demir, A., Aytar, P., Gedikli, S., Çabuk, A., Arısoy, M. (2011). Laccase Production with Submerged and Solid State Fermentation: Benefit and Cost Analysis. Hacettepe. *Journal Biology and Chemistry*. 39 (3): 305–313
- Dekker, R. F. H., Barbosa, A. M. (2001). The effects of aeration and veratryl alcohol on the production of two laccases by the ascomycete *Botryosphaeria* sp. *Enzyme and Microbial Technology*. 28(1): 81-88
- Del Pilar Castillo, M., Ander, P., Stenstrom, J. (1997). Lignin and manganese peroxidase activity in extracts from straw solid substrate fermentations. *Biotechnology Techniques*. 11: 701–706.
- Divya, L. M., Prasanth, G. K., Sadasivan, C. (2013). Isolation of a salt tolerant laccase secreting strain of *Trichoderma* sp. NFCCI-2745 and optimization of culture conditions and assessing its effectiveness in treating saline phenolic effluents. *Journal of Environmental Science*. 12: 2411-2416.
- Dias A, Sampaio A, Bezerra R. (2007). Environmental applications of fungal and plant systems: decolourisation of textile wastewater and related dyestuffs. In: Singh S and Tripathi R. *Environmental Bioremediation Technologies*. 445–463.
- Dias, A. A., Freitas, G. S., Marques, G. S. M., Sampaio, A., Fraga, I. S., Rodrigues, M. A. M. (2010). Enzymatic saccharification of biologically pre-treated wheat straw with white-rot fungi. *Bioresource Technology*. 101:6045–50.

- Ding, Z., Chen, Y., Xu, Z., Peng, L., Xu, G., Gu, Z., Zhang, L., Shi, G., Zhang, K. (2014). Production and characterization of laccase from *Pleurotus ferulae* in submerged fermentation. *Annals of Microbiology*. 64: 121–129.
- Du, Y., Qin, C., Huang, X., Nie, S., Song, X. (2015). Enzyme and alkali-aided ECF bleaching of kraft bamboo pulp *BioResources* 10(4), 7372-7385.
- Dwivedi, U. N., Singh, P., Kumar, A. (2011). Structure–function relationship among bacterial, fungal and plant laccases. *Journal of Molecular Catalysis B: Enzymatic*. 68: 117–128.
- Elisbetta, A., Josep, F.C., Teresa, V. (2009). Application of laccase – natural mediator systems to sisal pulp: An effective approach to biobleaching or functionalizing pulp fibres? *Bioresource Technology* 100, 5911-5916.
- Elisashvili, V., Penninckx, M., Kachlishvili, E., Tsiklauri, N., Metreveli, E. K. T., Kvesitadze, G. (2008a). Lentinus edodes and Pleurotus species lignocellulolytic enzymes activity in submerged and solid-state fermentation of lignocellulosic wastes of different composition. *Bioresource Technology*. 99: 457-462.
- Elisashvili, V., Kachlishvili, E., Penninckx, M. (2008b). Effect of growth substrate, method of fermentation, and nitrogen source on lignocellulose-degrading enzymes production by white-rot basidiomycetes. *Journal India Microbiology Biotechnology* 35, 1531-1538.
- Elisashvili, V., Parlar, H., Kachlishvili, E., Chichua, D., Kvesitadze, G. (2001). Ligninolytic activity of basidiomycetes grown under submerged and solid-state fermentation on plant raw material (sawdust of grapevine cuttings). *Advances in Food Sciences*. 23: 117–123
- Elisashvili, V., Kachlishvili, E., Tsiklauri, N., Metreveli, E., Khardziani, T., Agathos, S. N. (2009). “Lignocellulose-degrading enzyme production by white-rot Basidiomycetes isolated from the forests of GeorgiaWorld. *Journal Microbiology Biotechnology*. 25: 331-339.
- Erden, E., Ucar, M.C., TekinGezer N.K. Pazarlioglu (2009). Screening for ligninolytic enzymes from autochthonous fungi and applications for decolorization of remazole marine blue. *Brazilian Journal of Microbiologyl*. 40: 346-353
- Elsayed, M.A., Hassan, M.M., Elshafei, A.M., Haroun, B.M., Othman, A.M. (2012). Optijmization of cultural and nutritional parameters for the production of laccase by *Pleurotus ostreatus* ARC280. *British Biotechnology Journal*. 2(3): 115-132.
- Emre, B., Özfer, Y. (2013). The utilization of lignocellulosic wastes for laccase production under semisolid-state and submerged fermentation conditions. *Turkish Journal of Biology*. 37: 450-456.
- Eugenio, M., Santos, S., Carbajo, J., Marti´n, J., Marti´n-Sampedro, R., Gonza lez, A., Villar, J. (2010). Kraft pulp biobleaching using an extracellular enzymatic fluid produced by *Pycnoporus sanguineus*. *Bioresources Technology*. 101(6):1866–1870

- Eugenio, M.E., Herna'ndez, M., Moya, R., Marti'n-Sampedro, R., Villar, J.C., Arias, M. E. (2011) Evaluation of a new laccase produced by *Streptomyces ipomoea* on biobleaching and ageing of kraft pulps. *Bioresources*. 6 (3): 3231–3241.
- Eudes, A., Liang, Y., Mitra, P., Loqu'ed D. (2014). Lignin bioengineering. *Current Opinion in Biotechnology*. 26:189–198.
- Ethaib, S., Rozita, O., Mustapa, K. S. M., Awang, B. D. R., S. Syafie (2016). Microwave- assisted Dilute Acid Pretreatment and Enzymatic Hydrolysis of Sago Palm Bark. *BioResources*. 11(3): 5687-5702.
- Fang, T.J., Liao, B.C. Lee, S.C. (2010). Enhanced production of xylanase by *Aspergillus carneus* M34 in solid- state fermentation with agricultural waste using statistical approach. *New Biotechnology*. 27(1): 25-32.
- Ferraroni, M, Myasoedova, N., Schmatchenko, V., Leontievsky, A., Golovleva, L., Scozzafava, A., Briganti, F. (2007) Crystal structure of a blue laccase from *Lentinus tigrinus*: evidences for intermediates in the molecular oxygen reductive splitting by multicopper oxidases. *BMC Structural Biology*. 7:60.
- Flegel, T. W., Meevootisom, V., and Kitapapan, S.(1982). Indications of ligninolysis by *Trichoderma* strains isolated from soil during simultaneous screening for fungi with cellulase and laccase activity. *J. Ferm. Technol*, 60: 473-475.
- Fillat, U., Roncero, M.B. (2010). Optimization of laccase–mediator system in producing biobleached flax pulp. *Bioresource Technology*. 101: 181–187.
- Fillat, U., Roncero, M. B. (2009). Effect of process parameters in laccase-mediator system delignification of flax pulp: Part I. Pulp properties. *Chemical Engineering Journal*. 152 (2–3): 322–329.
- Fillat, A., Gallardo, O., Vidal, T., Pastor, F. I. J., Diaz, P., Roncero, M. B. (2012). Enzymatic grafting of natural phenols to flax fibres: development of antimicrobial properties. *Carbohydrate Polymers*. 87: 146–152.
- Fonseca-Maldonado, R., Ribeiro, L. F., Furtado, G. P., Arruda, L. M., Meleiro, L. P., Alponi, J. S., Botelho-Machado, C., Vieira, D. S., Bonneil, E., Furriel, R. d. P. M., Thibault, P., Ward, R. J. (2014). Synergistic action of co-expressed xylanase/laccase mixtures against milled sugar cane bagasse. *Process Biochemistry*. 49(7), 1152-1161.
- Forough, N., Dzulkefly, K. A., Norhafizah, A., Nazila, M., Reza, Z. (2013). Biological pretreatment of Rubberwood with *Ceriporiopsis subvermispora* for enzymatic hydrolysis and bioethanol production. *BioMed Research International*. pages 9.
- Galhaup, C., Goller, S., Peterbauer, C. K., Strauss, J., Haltrich, D. (2002). Characterization of the major laccase isoenzyme from *Trametes pubescens* and regulation of its synthesis by metal ions. *Microbiology Society Journals*. 148 (7): 2159–2169.
- Galli, C., Gentili, P. (2004). Chemical messengers: mediated oxidations with the enzyme laccase. *Journal of Physical organic chemistry* 17, 973–977.

- Gangwar, A. K., Tejo Prakash, N., Prakash, R. (2014). Applicability of microbial xylanases in paper pulp bleaching: A review. *BioResources* 9(2), 3733-3754.
- Gangwar, A.K., Tejo Prakash, N., Prakash, R. (2015). Amenability of Acacia and Eucalyptus hardwood pulps to elemental chlorine-free bleaching: Application and efficacy of microbial xylanase. *BioResources* 10(4), 8405-8413.
- Gangwar, A.K., Tejo Prakash, N., Ranjana, P., (2016). An eco-friendly approach: incorporating a xylanase stage at various places in ECF and chlorine-based bleaching of Eucalyptus pulp. *BioResources* 11(2), 5381-5388.
- Gharpuray, M.M., Lee, Y.H., Fan, L.T. (1983). Structural modification of lignocellulosics by pretreatments to enhance enzymatic hydrolysis. *Biotechnology and Bioengineering*. 25, 157-172.
- Gonzalez, J. C., Medina, C. S., Rodriguez, A., Osma, F. J., Alméciga-Dáz, C. J., Sánchez, F. O. (2013). Production of *Trametes pubescens* Laccase under Submerged and Semi-Solid Culture Conditions on Agro-Industrial Wastes. *plos journal*. 8 (9): 73-721
- Giese, C. E., Dekker, F. H. R., Barbosa, M. A. (2008). Orange bagasse as substrate for the production of pectinase and laccase by *Botryosphaeria rhodina* MAMB-05 in submerged and solid state fermentation. *Bioresources*. 3: 335–345.
- George, S., Daniel, S., Gloria A., Jimenéz-Tobón, Charles, J., Gerhard, K., Michel J., Penninckx. (2015). Production of a high level of laccase by submerged fermentation at 120-L scale of *Cerrena unicolor* C- 139 grown on wheat bran. *Comptes Rendus Biologies*. 338(2): 121–125.
- Glenn, J. K., Gold, M. H. (1985). Purification and characterization of an extracellular Mn (II)-dependent peroxidase from the lignin-degrading basidiomycete, *Phanerochaete chrysosporium*. *Archives of Biochemistry and Biophysics*. 242(2): 329–341.
- Gochev, V. K., Krastanov, A. I. (2007). Isolation of laccase producing *Trichoderma* spp. *Bulgarian Journal of Agricultural Science*. 13: 171-176
- Gomes, E., Aguiar, A. P., Carvalho, C. C., Bonfá M. R., daSilva, R., Boscolo, M. (2009). Ligninases production by basidiomycetes strains on lignocellulosic agricultural residues and their application in the decolorization of synthetic dyes. *Brazilian Journal of Microbiology*. 40: 31–39.
- Gullichsen, J. (2000). *Fiber line operations*. In: Gullichsen J, Fogelholm C-J (eds) Chemical pulping – papermaking science and technology. Fapet Oy, Helsinki, p A19 (Book 6A).
- Gutierrez, A., Bocchini, P., Galletti, G. C., & Martinez, A. T. (1996). Analysis of lignin- polysaccharide complexes formed during grass lignin degradation by cultures of *Pleurotus* species. *Applied and Environmental Microbiology*. 62: 1928-1934.
- Hanana,S., Elloumi, A., Placet,V., Tounsi, H., Belghith, H., and Bradai, C.(2015) An efficient enzymatic-based process for the extraction of high-mechanical properties alfa fibres. *Industrial Crops and Products*.70:190-200.

- Hammel, K.E., Cullen, D. (2008). Role of fungal peroxidases in biological ligninolysis. *Current Opinion in Plant Biology*. 11:349–355.
- Hatakka, A. (2001). Biodegradation of Lignin. In: Hofrichter M, Steinbuechel A (eds). (ed) Biopolymers, Wiley-VCH Verlag GmbH & Co. KGaA.
- Harman, G. E., Herrera-Estrella A. H., Benjamin, A., Matteo, L. (2012). Special issue: Trichoderma – from Basic Biology to Biotechnology. *Microbiology*. 58:1-2.
- Hao, J., Song, F., Huang, F., Yang, C., Zhang, Z., Zheng, Y., Tian, X. (2007). Production of laccase by a newly isolated deuteromycete fungus *Pestalotiopsis* sp. and its decolorization of azo dye. *Journal of Industrial Microbiology and Biotechnology*. 34: 233-240.
- He, J. X., Zhang, M., Cui, S. Z., Wang, S. Y. (2009). High-quality cellulose triacetate prepared from bamboo dissolving pulp. *Journal of Applied Polymer Science*. 113: 456-465.
- He, P., Lige, P., Xinsheng, H., and Chunping, X. (2014). Optimization of culture conditions for laccase production from *Abortiporus biennis* in a pilot- scale bioreactor. *Arch. Biol. Sci., Belgrade*, 66 (4), 1567-1574.
- Hendro, R., Elis, S., Sri, H. S., Tjandra, S. (2012). Optimisation of laccase production using white rot fungi and agriculture wastes in solid state fermentation. *ITB Journal of Engineering Science*. 44(2): 93-105.
- Hendricks, A. T., Zeeman, G. (2009). Pretreatments to enhance the digestibility of lignocellulosic biomass. *Bioresource Technology*. 100:10–108.
- Himmel, M. E., Ding, S. Y., Johnson, D. K., Adney, W. S., Nimlos, M. R., Brady, J. W. (2007). Biomass recalcitrance: engineering plants and enzymes for biofuel production. *Science*. 315, 804-807.
- Hofrichter, M. (2010). Industrial Applications. The Mycota, A Comprehensive Treatise on Fungi as Experimental Systems for Basic and Applied Research, *Springer Science & Business Media*. 10: 319–340.
- Hofrichter, M. (2002). Review: lignin conversion by manganese peroxidase (MnP). *Enzyme and Microbial Technology*. 30 (4): 454–466.
- Hoigné J., Bader, H. (1994). Kinetics of reactions of chlorine dioxide (OCIO) in water—I. Rate constants for inorganic and organic compounds. *Water Research* 28(1), 45-55.
- Huiju, G., Xiang, C., Yanwen, W., Fei, Z., Kai, Z., Zhimei, M., Qingxin, L. (2013). Media optimization for laccase production by *Trichoderma harzianum* ZF-2 using response surface methodology. *Journal Microbiology Biotechnology*. 23(12): 1757-1764.
- Ibrahim, V., Mendoza, L., Mamo, G., Hatti-Kaul, R. (2011). Blue laccase from *Galerina* sp. properties and potential for Kraft lignin demethylation. *Process Biochemical*. 46:379–384.
- Iqbal, H. M. N., Asgher, M., Bhatti, H. (2011). Optimization of physical and nutritional factors for synthesis of lignin degrading enzymes by a novel strain of *Trametes versicolor*. *BioResources*. 6(2): 1273-1287.

- Irshad, M., Anwar, Z., But, H. I., Afroz, A., Ikram, N., Rashid, U. (2013). The industrial applicability of purified cellulose complex indigenously produced by *Trichoderma viride* through solid-state bio-processing of agro-industrial and municipal paper wastes. *BioResources*. 8(1): 145-157.
- Janusz, G., Kucharzyk, K. H., Pawlik, A., Staszczak, M., Paszczynski, A. J. (2013). Fungal laccase, manganese peroxidase and lignin peroxidase: Gene expression and regulation. *Enzyme and Microbial Technology*. 52(1): 1-12.
- Jeon, J. R., Chang, Y. S. (2013). Laccase-mediated oxidation of small organics: Bifunctional roles for versatile applications. *Trends Biotechnology*. 31: 335-341.
- Jenitta, E. P., and Rangunathan, R. (2014). "Production, purification, characterization, and copper induction of laccase isoenzyme in the lignolytic fungus *Pleurotus florida*. *Journal Microbiology Biotechnolgy Research*. 4(2): 10-16.
- Jersson, P., Sergio, C. (2015). Ligninolytic enzymes: a biotechnological alternative for bioethanol production. Plácido and Capareda. *Bioresources and Bioprocessing*. 2:23.
- Jesus, D.C., Carlos, C., Esperanza, T. (2015). Optimization of the production, purification and characterization of a laccase from the native fungus *Xylaria* sp. *Biocatalysis and Agricultural Biotechnology*. 4:710–716.
- Jiménez, L., Pérez, A., Rodríguez, A., de la Torre, M.J. (2006). New raw materials and pulping processes for production of pulp and paper. *Afinidad* 63 (525), 362– 369.
- Jirka, A. M., Carter, M. J. (1975). *Analytical chemistry*. Federal register. 45 (78): 26811-26812.
- Jose, L., Joel, G. J. (2014). Optimization of culture conditions form improved laccase production by *Agaricus* sp. LCJ262. *International Journal of Current Research*. 6 (11): 9517-9522.
- Kalra, K., Chauhan, R., Shavez, M., Sachdeva, S. (2013). Isolation of laccase producing *Trichoderma spp.* and effect of pH and temperature on its activity. *International Journal of ChemTech Research*. 5 (5), 2229– 2235.
- Kapoor, M., Kapoor, R., Kuhad, R. (2007). Differential and synergistic effects of xylanase and laccase mediator system (LMS) in bleaching of soda and waste pulps. *Journal of Applied Microbiology*. 103 (2): 305– 317.
- Khambhaty, Y., Ananth, S., Sreeram, K. J., Rao, J.R., Nair, B.U. (2015). Dual utility of a novel, copper enhanced laccase from *Trichoderma aureoviridae*. *International Journal of Biological Macromolecules*. 81: 69–75.
- Kumar, P., Barrett, D. M., Delwiche, M. J., Stroeve, P. (2009). Methods for pretreatment of lignocellulosic biomass for efficient hydrolysis and biofuel production. *Industrial and Engineering Chemistry Research*. 48: 3713e3729.
- Kuntal, K., Rohit, C., Mohd, S., Sarita, S. (2013). Isolation of laccase producing *Trichoderma* sp. and effect of pH and temperature on its activity. *Journal of ChemTech Research*. 5: 974-4290.

- Kyoung-M. L., Dayanand, K., Manish. K. T., Tae-Su. K., Saurabh. S. D., Jung-Kul. L., In-Won. K. (2012). Enhanced enzymatic hydrolysis of rice straw by removal of phenolic compounds using a novel laccase from yeast *Yarrowia lipolytica*. *Bioresource Technology*. 123: 636–645.
- Khushal, B., Anne, R., Praveen V, V. (2010). Fungal Laccases: Production, Function, and Applications in Food Processing. *Enzyme Research*. 10
- Kunamneni, A., Ballesteros, A., Plou, F. J. and Alcalde, M. (2007). Fungal laccase-a versatile enzyme for biotechnological applications, in Communicating Current Research and Educational. *Topics and Trends in Applied Microbiology*. A. Mendez-Vilas, Ed. 1: 233–245.
- Kusum, D., Anita, P. (2013). Laccase Production from a Temperature and pH Tolerant Fungal Strain of *Trametes hirsuta* (MTCC 11397). *Enzyme Research*. 9.
- Kim, S., Holtzaple, M. T. (2006). Effect of structural features on enzyme digestibility of corn stover. *Bioresource Technology*. 97(4): 583-591.
- Kong, W., Chen, H., Lyu, S, Ma, F, Yu, H., Zhang ,X. (2016). Characterization of a novel manganese peroxidase from white-rotfungus *Echinodontium taxodii* 2538, and its use for the degradation of lignin-related compounds. *Process Biochemistry*. 51: 1776–1783.
- Lehtimaa, T., Tarvo, V., Kuitunen, S., Jaaskelainen, A.-S., Vuorinen, T. (2010). The effect of process variables in chlorine dioxide prebleaching of birch kraft pulp. Part 2. AOX and OX formation *Journal of Wood Chemistry and Technology*. 30(1), 19-30.
- Leutbecher, H., Constantin, M. A., Mika, S., Conrad, J., Beifuss, U. (2011). A new laccase-catalyzed domino process and its application to the efficient synthesis of 2 aryl-1H-benzimidazoles. *Tetrahedron Letters*. 52(5): 605-608.
- Li, J., He, S., Li, Z., Xu, J., Mo, L. (2014). Comparison of ECF short-sequence bleaching processes for bagasse pulp. *Journal of South China University of Technology Natural Science*. 42 (2): 14-20.
- Li, S., Tang, B., Liu, Y., Chen, A., Tang, W., Wei, S. (2016). High-level production and characterization of laccase from a newly isolated fungus *Trametes* sp. LS-10C. *Biocatalysis and Agricultural Biotechnology* 8 : 278–285.
- Liu, L., Cheng, L., Huang, L., Yu, J. (2012). Enzymatic Treatment of Mechanochemical Modified Natural Bamboo Fibers. *Fibers and Polymers*. 13: 600-605.
- Liu, J., Cai, Y., Liao, X., Huang, Q., Hao, Z., Hu, M. (2013). Efficiency of laccase production in a 65-L air-lift reactor for potential green industrial and environmental application. *Journal of Cleaner Production*. 39: 154- 160.
- Lu, C., Wang, H., Luo, Y., Guo, L. (2010). An efficient system for predelignification of gramineous biofuel feedstock in vitro: application of a laccase from *Pycnoporus sanguineus* H275. *Process Biochemistry*. 45:1141–1147.
- Luterek, J., Gianfreda, L., Wojtaś-Wasilewska, M. (1997). Screening of the wood-rotting fungi for laccase production: induction by ferulic acid, partial

- purification, and immobilization of laccase from the high laccase-producing strain, *Cerrena unicolor*. *Acta Microbiologica Polonica*. 46 (3): 297–311.
- Ma, X., Huang, L., Cao, S., Chen, Y., Luo, X., Chen, L. (2012). Preparation of dissolving pulp from bamboo for textile part 2. Optimization of pulping conditions of hydrolyzed bamboo and its kinetics. *BioResources*. 7 (2): 1866-1875.
- Mahmudin,S., Mohamed, A. Z., Jasmani, L.,Shahwahid, Hj.M., Tahir,Md.P., Harun,J. Hashim,N.,Yusoff,M.N.M. Saffian,H.A.(2012).*Kenaf a potential fibre for pulp and paper manufacture*.Malaysin Timber Industry Board.page 19-20.
- Majeau, J. A., Brar, S. K., Tyagi, R. D. (2010). Laccases for removal of recalcitrant and emerging pollutants. *Bioresource Technology*. 101: 2331-2350.
- Martín-Sampedro, R., Eugenio, M. E., Carbajo, J. M., Villar, J. C. (2011). Combination of steam explosion and laccase-mediator treatments prior to *Eucalyptus globulus* kraft pulping. *Bioresource Technology*. 102:7183–7189.
- Martin-Sampedro, R.,Rodriguez, A., Ferrer, A., Garcia-Fuentevilla, L.L., Eugenio, M.E. (2012). Biobleaching of pulp from oil palm empty fruit bunches with laccase and xylanase. *Bioresource Technology*. 110: 371–378.
- Martin-Sampedro, R., Miranda, J., Garc ía-Fuentevilla, L.L., Hernández, M., Arias, ME., Diaz, MJ., Eugenio, M.E. (2015). Influence of process variables on the properties of laccase biobleached pulps. *Bioprocess Biosyst Engineering* 38(1):113-23.
- Maria, d., R., F., Amin, K., Carlos, F., Jose, M., A. (2008). Production of laccase and xylanase from *Coriolus versicolor* grown on tomato pomace and their chromatographic behaviour on immobilized metal chelates. *Process Biochemistry*. 43: 1265–1274.
- Márquez-rocha, F. J., Guillén, G. K., Sánchez, J. E., Vázquez-duhalt, R. (1999). Growth characteristics of *Pleurotus ostreatus* in bioreactors. *Biotechnology Techniques*. 13(1). 29-32.
- Mathur, G., Nigam, R., Jaiswal, A., Kumar, C. (2013). Bioprocess parameter optimization for laccase production in solid state fermentation. *Journal of Biotechnology and Bioengineering Research*. 2231-1238(4): 521-530.
- Matera I, Gullotto A, Tilli S, Ferraroni M, Scozzafava A, Briganti F (2008). Crystal structure of the blue multicopper oxidase from the white-rot fungus *Trametes trogii* complexed with p-toluate. *Inorganica Chimica Acta*. 361:4129–4137.
- Mattinen, M., Maijala, P., Nousiainen, P., Smeds, A., Kontro, J., Sipilä J., Tamminen, T., Willför, S., Viikari, L. (2011). Oxidation of lignans and lignin model compounds by laccase in aqueous solvent systems. *Journal of Molecular Catalysis B*. 72:122–129.
- Mattinen, M., Suortti, T., Gosselink, R., Dimitris, S., Argyropoulos, D., Evtugin, A.,Suurnakki, E., de Jong, T. (2008). Polymeriazion of different lignins by laccase *Bioresources*. 3 (2) 549–565.

- Mayer, A.M., Staples, R.C. (2002). Laccase: new functions for an old enzyme. *Phytochemistry*. 60 (6): 551–565.
- Mchunu, N., Permaul, K., Alam, M., Singh, S. (2013). Carbon utilization profile of a thermophilic fungus, *Thermomyces lanuginosus* using phenotypic microarray. *Advances in Bioscience and Biotechnology*. 4: 24-32.
- Merve, A., Raziye, O. U. (2013). Extracellular ligninolytic enzymes production by *Pleurotus eryngii* on agroindustrial wastes. *Preparative Biochemistry and Biotechnology*. 44: 8.
- Mikiashvili, N., Elisashvili, V., Wasser, S., Nevo, E. (2005). Carbon and nitrogen sources influence the ligninolytic enzyme activity of *Trametes versicolor*. *Biotechnology Letter*. 27: 955–959.
- Moldes, D., D áz, M., Tzanov, T., Vidal, T. (2008). Comparative study of the efficiency of synthetic and natural mediators in laccase-assisted bleaching of eucalyptus kraft pulp. *Bioresource Technology*. 99, 7959–7965.
- Moldes, D., Vidal, T. (2008) Laccase–HBT bleaching of eucalyptus kraft pulp: influence of the operating conditions. *Bioresources Technology*. 99 (18): 8565–8570.
- Moldes, D., Cadena, E.M, Vidal, T. (2010). Biobleaching of eucalypt kraft pulp with a two laccase-mediator stages sequence. *Bioresource Technology* 101, 6924–6929.
- Monteiro, M. C., De Carvalho, M. E. A. (1998). Pulp bleaching using laccase from *Trametes versicolor* under high temperature and alkaline conditions. *Applied Biochemistry and Biotechnology*. 70-72: 983.
- Montgomery, D. C.(2008). Design and analysis of experiments. New Jersey: John wiley and Sons.
- Moradbak, A., Tahir, Md, P., Mohamed, A. Z., Halis, R. (2016). Alkaline Sulfite Anthraquinone and Methanol Pulping of Bamboo (*Gigantochloa scortechinii*). *BioResources* 11(1), 235-248.
- Morozova, O.V., Shumakovich, G.P., Shleev, S.V., Yaropolov, Y.I. (2007). Laccasemediator systems and their applications: a review. *Applied Biochemistry and Microbiology*. 43: 523–535
- Mot, A.C., Silaghi-Dumitrescu, R. (2012). Laccases: Complex architectures for one-electron oxidations. *Biochemistry*. 77: 1395–1407.
- Morselli, M., Semplice, M., Villa, S., and Di Guardo, A. (2014). Evaluating the temporal variability of concentrations of POPs in a glacier-fed stream food chain using a combined modelling approach. *Science of the Total Environment* 493(0), 571-579.
- Mou, H. Y., Heikkila, E., Fardim, P. (2013). Topochemistry of alkaline, alkaline peroxide and hydrotropic pretreatments of common reed to enhance enzymatic hydrolysis efficiency *Bioresource Technology* 150(1), 36-41.
- Muhammad, N. S. (2013) Fermentation and kinetic studies on laccase production by *pycnoporus sanguineus*. University Malaysia. Master of Science.

- Mustafa, M. T., Wahab, R., Sudin, M., Khalid, I., Kamal, N. A. M. (2011). Anatomical properties and microstructures features of four cultivated bamboo *Gigantochloa* species *Journal of Asian Scientific Research*. 1 (7): 328-339.
- Neifar, M., Kamoun, A., Jaouani, A., Ellouze-Ghorbel, R., Ellouze-chaabouni, S. (2011). Application of asymmetrical and hoke designs for optimization of laccase production by white-rot fungus *fomes fomentarius* in solid –state fermentation. *Enzyme Research*. 1(1):1-12.
- Nigam, P., Gupta, N., Anthwal, A. (2009). Pre-treatment of agro-industrial residues. In: Nigam P and Pandey A. *Biotechnology for Agro-Industrial Residues Utilisation*. Springer Netherlands; p. 13–33.
- Nie, S., Liu, X., Wu, Z., Zhan, L., Yin, G., Yao, S., Song, H., Wang, S. (2014a). Kinetics study of oxidation of the lignin model compounds by chlorine dioxide. *Chemical Engineering Journal*. 241(1): 410-417.
- Nie, S., Yao, S., Qin, C., Li, K., Liu, X., Wang, L., Song, X., Wang, S. (2014b). Kinetics of AOX formation in chlorine dioxide bleaching of bagasse pulp. *BioResources*. 9 (3): 5604-5614.
- Nie, S. X., Wu, Z. M., Liu, J. C., Liu, X. L., Qin, C. R., Song, H. N., Wang, S. F. (2013). Optimization of AOX formation during the first chlorine dioxide bleaching stage (D-0) of soda AQ bagasse pulp. *Appita Journal*. 66 (4): 306-312.
- Nie, S., Wang, S., Qin, C., Yao, S., Ebonka, J. F., Song, X., Li, K. (2015). Removal of hexenuronic acid by xylanase to reduce adsorbable organic halides formation in chlorine dioxide bleaching of bagasse pulp. *Bioresource Technology*. 196, 413-7.
- Norhidayah, C.S.,(2011). Evaluation of growth performance of bamboo (*Gigantochloa* sp.) in integration with banana (*musa paradisiacal.*) and misai kucing (*orthosiphon stamineus benth*). Universiti Putra Malaysia. Master of Science.
- Nooshin, R. (2013). *Utilization of Trichoderma harzianum SNRS3 for sugar recovery from rice straw for acetone-butanol-ethanol production by Clostridium acetobutylicum ATCC824*. Universiti Putra Malaysia. Doctor of Philosophy.
- Nyanhongo, G.S., Gomesa, J., Gubitze, G.M., Zvauyab, R., Readd, J., Steinera, W. (2002). Decolorization of textile dyes by laccases from a newly isolated strain of *Trametes modesta*. *Water research*. 36: 1449-1456.
- Osma, J.F., Toca-Herrera, J.L., Rodríguez-Couto, S. (2011). Cost analysis in laccase production. *Journal of Environmental Management*. 92: 2907–2912.
- Osman, W. H., Sheikh Abdullah, S. R., Abu Bakar M., Kadhun, H. A., Rahman, A. R. (2013) Simultaneous removal of AOX and COD from real recycled paper wastewater using GAC-SBBR. *Journal of Environmental Management* 121: 80-86.
- Ofori-Boateng, C., Lee, K. T. (2013). Sustainable utilization of oil palm wastes for bioactive phytochemicals for the benefit of the oil palm and nutraceutical

- industries. *Phytochemistry Reviews*. 1-18.
- Palaez, F., Mart ínez, M. J., Mart ínez, A. T. (1995). Screening of 68 species of basidiomycetes involved in lignin degradation. *Mycological Research*. 99: 37–42.
- Palmieri, G., Cennamo, G., Faraco, V., Amoresano, A., Sannia, G., Giardina, P. A. (2003). Typical isoenzymes from copper supplemented *Pleurotus ostreatus* cultures. *Enzyme Microbiology. Technology*. 33: 220–230.
- Patel, H., Gupte, A., (2016). Optimization of different culture conditions for enhanced laccase production and its purification from *Tricholoma giganteum* AGHP. *Bioresources and Bioprocessing*. 3: 11.
- Pathan, A. K., Bond, J., Gaskin, R.E. (2008). Sample preparation for scanning electron microscopy of plant surfaces--horses for courses. *Micron*. 39 (8):1049-61.
- Paszczynski, A., Huynh, V. B., Crawford, R. (1985). Enzymatic activities of an extracellular, manganese-dependent peroxidase from *Phanerochaete chrysosporium*. *FEMS Microbiology Letters*. 29: 37–41.
- Pei, Y., Wang, S., Qin, C., S, J., Nie, S., Song, X., (2016). Optimization of Laccase-Aided Chlorine Dioxide Bleaching of Bagasse Pulp. *BioResources* 11(1): 696-712.
- Periasamy, R., (2016) Bleaching of paper pulp by laccase from *Pleurotus ostreatus* IMI 395545 with artificial mediators. *International Journal of Advanced Life Sciences (IJALS)*. 9:(3).
- Piontek, K., Glumoff, T., Winterhalter, K. (1993). Low pH crystal structure of glycosylated lignin peroxidase from *Phanerochaete chrysosporium* at 2.5 Å resolution. *FEBS Letters*. 315:119–124.
- Piontek, K., Antorini, M., Choinowski, T. (2002). Crystal structure of a laccase from the Fungus *Trametes versicolor* at 1.90-Å resolution containing a full complement of coppers. *Journal of Biological Chemistry*. 277 (40): 37663–37669.
- Pointing, S. B., Jones, E. B. G., Vrijmoed, L. L. P. (2000). Optimization of laccase production by *Pycnoporus sanguineus* in submerged liquid culture. *Mycologia* 92(1): 139 -144.
- Qiu, W., Chen, H. (2012). Enhanced the enzymatic hydrolysis efficiency of wheat straw after combined steam explosion and laccase pretreatment. *Bioresource Technology*. 118: 8–12.
- Quintana, E., Valls, C., Barneto, A. G., Vidal, T., Ariza, J., Roncero, M. B. (2015). Studying the effects of laccase treatment in a softwood dissolving pulp: Cellulose reactivity and crystallinity. *Carbohydrate Polymers*. 119: 53-61.
- Rafaella, C. B. S., Lucia, R. D., Manuela, d. S. Lara, D. S. (2010). Production of laccase, manganese peroxidase and lignin peroxidase by Brazilian marine-derived fungi. *Enzyme and Microbial Technology*. 46: 32–37.
- Rajesh, K., Jaswinder, K., Saurabh, J., Ashwani, K. (2016). Optimization of laccase production from *Aspergillus flavus* by design of experiment technique:

- Partial purification and characterization. *Journal of Genetic Engineering and Biotechnology*. 14 (1): 125-131.
- Rahman, N. A., Rahman, N. A. A., Aziz, S., Hassan, M. A. (2013). Production of Ligninolytic Enzymes by Newly Isolated Bacteria from Palm Oil Plantation Soils. *BioResources*. 8(4): 6136-6150.
- Ramirez-Cavazos L.I., Junghanns, C., Nair, R. (2014). Enhanced production of thermostable laccases from a native strain of *Pycnoporus sanguineus* using central composite design. *Journal of Zhejiang University Science B*. 15: 343-352.
- Ravalason, H., Bertaud, F., Herpoel-Gimbert, I., Meyer, V., Ruel, K., Joseleau, J. P., Grisel, S., Olive, C., Sigoillot, J. C., Petit-Conil, M. (2012). Laccase/HBT and laccase-CBM/HBT treatment of softwood kraft pulp: Impact on pulp bleachability and physical properties. *Bioresource Technology*. 121: 68-75.
- Rehan, A. M., Enas, A. H., Elshahat, M. R. (2016). Production of laccase enzyme for their potential application to decolorize fungal pigments on aging paper and parchment. *Annals of Agricultural Science*. 61(1): 145-154
- Rennie, E. A., Scheller, H. V. (2014). Xylan biosynthesis. *Current Opinion in Biotechnology*. 26: 100–107.
- Revankar, M.S., Lele, S.S., 2006. Enhanced production of laccase using a new isolate of white rot fungus WR-1. *Process Biochem*. 41, 581–588.
- Rittstieg, K., Suurnakki, A., Suortti, T., Kruus, K., Guebitz, G., Buchert, J. (2002). Investigations on the laccase-catalyzed polymerization of lignin model compounds using size-exclusion HPLC. *Enzyme Microbiology Technology*. 31 (4): 403–410.
- Rivera-Hoyos, C. M., Edwin, D. M. Á., Raúl, A. P. P., Aura, M. P. R., Refugio, R. V., Julio, M. D. B. (2013). Fungal laccases. *Fungal Biology Reviews*. 27 (3-4): 67-82.
- Riva, S. (2006). Laccases: blue enzymes for green chemistry. *Trends Biotechnology*. 24: 219–226.
- Robles, A., Lucas, R., Martínez-Cañamero, M., Omar, N.B., Pérez, R., Gálvez, A. (2002). Characterisation of laccase activity produced by the hyphomycete *Chalara* (syn. *Thielaviopsis*) *paradoxa* CH32. *Enzyme Microbiology Technology*. 31:516–522.
- Rodgers, C.J., Blanford, C.F., Giddens, S.R., Skamnioti, P., Armstrong, F.A., Gurr, S.J., 2010. Designer laccases: a vogue for high-potential fungal enzymes? *Trends Biotechnology*. 28 (2): 63–72.
- Rodríguez Fernández, D. E., Rodríguez Fernández, L., de Carvalho, J.C., Karp, S., Soccol, C. R. (2012). Process Development to Recover Pectinases Produced by Solid-State Fermentation. *Journal of Bioprocessing and Biotechniques*. 2(121).
- Ruttimann, C., Schwember, E., Salas, L., Cullen, D., Vicuna, R. (1992). Ligninolytic enzymes of the white-rot basidiomycetes *Phlebia brevispora* and *Ceriporiopsis subvermispora*. *Biotechnology and Applied Biochemistry*. 16:

- R.T. Mendonça, J.F. Jara, V. Gonzalez, J.P. Elissetche, J. Freer, J. Ind. *Microbiology*. SM Biotechnology. 35: 1323-1330.
- Sadhasivam, S., Savitha, S., Swaminathan, K., Feng-Huei, L. (2008). Production, purification and characterization of mid-redox potential laccase from a newly isolated *Trichoderma harzianum* WL1. *Process Biochemistry*. 43: 736-742.
- Salmela, M., Alén, R., Mân Vu, T. H. (2008). Description of kraft cooking and oxygen–alkali delignification of bamboo by pulp and dissolving material analysis. *Industrial Crops and Products*. 28: 47-55.
- Salvachúa, D., Prieto, A., López-Abelairas, M., Lu-Chau, T., Martínez ÁT, Martínez M. J. (2011). Fungal pretreatment: an alternative in second-generation ethanol from wheat straw. *Bioresource Technology*. 102:7500–7506.
- Sanchez, C. (2009). Lignocellulose residues: Biodegradation and bioconversion by fungi. *Biotechnology Advances*. 27: 185-194.
- Samuels, G.J., Chaverri, P., Farr, D.F. McCray, E.B. (2002a). *Trichoderma* Online. Systematic Mycology and Microbiology Laboratory, ARS, USDA; Retrieved 24, August 2014 from <http://nt.ars-grin.gov/taxadescriptions/keys/TrichodermaIndex.cfm>.
- Samuels, G. J., Chaverri, P., Farr, D. F., McCray, E. B. (2004). *Trichoderma* Online. Systematic Botany & Mycology Laboratory, ARS, USDA. Retrieved August 31, from <http://nt.ars-grin.gov/taxadescriptions/keys/TrichodermaIndex.cfm>.
- Samuels, G. J., Dodd, S. L., Gams, W., Castlebury, L. A., Petrini, O. (2002b). *Trichoderma* species associated with the green mold epidemic of commercially grown *Agaricus bisporus*. *Mycologia* . 94: 146- 170.
- Saijonkari-Pahkala, K. (2001). *Non-wood plants as raw material for pulp and paper*. (ACADEMIC DISSERTATION) Faculty of Agriculture and Forestry, University of Helsinki, for public criticism at Infokeskus Korona, Auditorium 1, on November 30.
- Savoie, J. M., Mata, G. Billette, C. (1998). Extracellular laccase production during hyphal interactions between *Trichoderma* sp. and shiitake, *Lentinula edodes*. *Applied Microbiology and Biotechnology*. 49: 589-593
- Schmidt, O. (2007). Indoor wood-decay basidiomycetes: damage, causal fungi, physiology, identification and characterization, prevention and control. *Mycological Progress*. 6: 261–279.
- Schuster, A., Schmoll, M. (2010). Biology and biotechnology of *Trichoderma*. *Applied Microbiology and Biotechnology*. 87 (3): 787–799.
- Scurlock, J.M.O., Dayton, D.C., Hames, B. (2000). Bamboo: an overlooked biomass resource? *Biomass and Bioenergy*. 19: 229-244.
- Scotti, T.C., Vergoignan, C., Feron, G. Durand, A. (2001). Glucosamine measurement as indirect method for biomass estimation of *Cunninghamella elegans* grown in solid state cultivation conditions. *Biochemical Engineering Journal*. 7:1-5.

- Segal, L., Creely, J. J., Martin, A. M. Jr., Conrad, C. M. (1959). An empirical method for estimating the degree of crystallinity of native cellulose using the X-ray diffractometer *Textile Research Journal*. 29 (10): 786-794.
- Semra, S., Nese, A., Inci, A. (2013). Studies on laccase activity in the filamentous fungus *Trichoderma reesei*. *Journal of Biology*. 72(2): 37-42.
- Sharma, A., Thakur, V. V., Shrivastava, A., Jain, R. K., Mathur, R. M., Gupta, R., Kuhad, R. C. (2014). Xylanase and laccase based enzymatic kraft pulp bleaching reduces adsorbable organic halogen (AOX) in bleach effluents: A pilot scale study. *Bioresource Technology*. 169: 96-102.
- Sharma, P., Sood, C., Singh, G., Capalash, N. (2015). An eco-friendly process for biobleaching of eucalyptus kraft pulp with xylanase producing *Bacillus halodurans*. *Journal of Cleaner Production*. 87: 966-970.
- Shashirekha, M. N., Rajarathnam, S., Bano, Z. (2002). Enhancement of bioconversion efficiency and chemistry of the mushroom, *Pleurotus sajor-caju* (Berk and Br.) Sacc. produced on spent rice straw substrate, supplemented with oil seed cakes. *Food Chemistry*. 76: 27-31.
- Shi, L., Yu, H., Dong, T., Kong, W., Ke, M., Ma, F., Zhang, X. (2014). Biochemical and molecular characterization of a novel laccase from selective lignin-degrading white-rot fungus *Echinodontium taxodii*2538. *Process Biochemistry* 49: 1097–1106.
- Shigematsu, A., Mizoue, N., Kajisa, T., Yoshida, S. (2011). Importance of rubberwood in wood export of Malaysia and Thailand. *New Forests*. 41(2): 179-189.
- Shraddha, Ravi, S., Simran, S., Mohit, K., Ajay, K. (2011). Laccase: Microbial sources, production, purification, and potential biotechnological applications. *Enzyme Research*. 217861: 11.
- Shleev, S., Persson, P., Shumakovich, G., Mazhugo, Y., Yaropolov, A., Ruzgas, T., Gorton, L. (2006). Interaction of fungal laccases and laccase-mediator systems with lignin. *Enzyme and Microbial Technology*. 39 (4): 841–847.
- Siddhartha, S., Tapobrata, P. (2015). Optimization of Laccase Fermentation and Evaluation of Kinetic and Thermodynamic Parameters of a Partially Purified Laccase Produced by *Daedalea flavida*. *Preparative Biochemistry and Biotechnology*. 45. 4.
- Silvério, S. C., Rodríguez, O., Tavares, A. P. M., Teixeira, J. A., Macedo, E. A. (2013). Journal of Molecular Catalysis B: Enzymatic Laccase recovery with aqueous two-phase systems: Enzyme partitioning and stability. *Journal of Molecular Catalysis. B, Enzymatic* 87: 37–43.
- Sindhu, R., Kuttiraja, M., Binod, P., Sukumaran, R. K., Pandey, A. (2014). Physicochemical characterization of alkali pretreated sugarcane tops and optimization of enzymatic saccharification using response surface methodology. *Renewable Energy*. 62: 362-368.
- Singh, M. M., Parkayastha, S. K., Bhola, P. P., Lal, K., Singh, S. (1976). Fibre morphology pulp sheet properties of Indian bamboos. *Indian Forester*. 102 (9): 579- 595.

- Singh, G., Ahuja, N., Batish, M., Capalash, N., Sharma, P. (2008). Biobleaching of wheat straw rich soda pulp with alkalophilic laccase from gamma-proteobacterium JB: optimization of process parameters using response surface methodology. *Bioresource Technology*. 99, 7472e7479.
- Singh, A., Kaur, A., Dua, A., Ritu, M. (2015). An efficient and improved methodology for the screening of industrially valuable xylano-pectino-cellulolytic microbes. *Enzyme Resrech* p.7.
- Singhania, R. R., Sukumaran, K. R., Patel, K. A., Larroche, C., Ashok, P. (2010). Advancement and comparative profiles in the production technologies using solid-state and submerged fermentation for microbial cellulases. *Enzyme and Microbial Technology*. 46(7): 541–549.
- Singhal, A., Choudhary, G., Shekhar .T. I. (2012). Charcterization of laccase activity produced by *Cryptococcus albidus*. *Preparative Biochemistry & Biotechnology*. 42:113–124.
- Singhania, R.R., Patel, K., Soccol, C.L., Pandey, A., (2009). Recent advances in solid-state fermentation. *Biochemical Engineering Journal*. 44:13–18 18.
- Sitompul, A., Yutaka, T., Toshihiro, W., Mitsuru, O. (2009). Screening of white rot fungi for biobleaching of Acacia oxygen-delignified kraft pulp. *World Journal of Microbiology and Biotechnology* 25(4):639-647.
- Smook, GA. (1992a). *Wood and chip handling*. Handbook for Pulp & Paper Technologists, 2nd edn. Angus Wilde Publications. Vancouver. p 20.
- Smook, G. A. (1992b). *Overview of pulping methodology*. Handbook for Pulp & Paper Technologists, 2nd edn. Angus Wilde Publications. Vancouver. p 36.
- Song, X.,Pei, Y.,Su, J.,Qin, C.,Wang,S.,Nie, S.(2016). Kinetics of adsorable oranic halides (AOX) reduction in laccae-aided chlorine dioxide bleaching of bagasse pulp. *BioResource*. 11(3), 7462-7475.
- Srebotnik, E., Messner, K., and Foisner, R. (1988). Penetrability of white rot-degraded pine wood by the lignin peroxidase of *Phanerochaete chrysosporium*. *Applied and Environmental Microbiology*. 54: 2608–2614.
- Srinivasakannan, C.; Zailani Abu Bakar, M. (2004). Production of activated carbon from rubber wood sawdust. *Biomass Bioenergy*. 27: 89–96.
- Svobodova, K., Majcherczyk, A., Novotny, C., Ku ës, U. (2008). Implication of mycelium-associated laccase from *Irpex lacteus* in the decolorization of synthetic dyes. *Bioresource Technology*. 99: 463-471.
- Tamilvendan, M., Arulmani, M., Kalaichelvan P., Thangavelua, Klaus, H., (2013). Characterization of optimized production, purification and application of laccase from *Ganoderma lucidum*. *Biochemical Engineering Journal* 70: 106– 114.
- Tamura, K., Stecher, G., Peterson, D., Filipski, A., Kumar, S.(2013) MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Molecular Biology and Evolution* 30(12): 2725-2729.
- TAPPI Test Method T 204 cm-97 (1997). “Solvent extractives of wood and pulp,” TAPPI Press, Atlanta, GA, USA.

- TAPPI T230 om-04 (2004). "Viscosity of pulp," TAPPI Press, Atlanta, GA.
- TAPPI T217 wd-77 (2004). "Brightness of pulp," TAPPI Press, Atlanta, GA.
- TAPPI T201 wd-76 (2004). "Cellulose in pulp (cross and bevan method)," TAPPI Press, Atlanta, GA.
- TAPPI T222 om-02 (2004). "Acid-insoluble lignin in wood and pulp," TAPPI Press, Atlanta, GA.
- TAPPI T260 wd-98 (2004). "Test to evaluate the aging properties of bleached chemical pulps," TAPPI Press, Atlanta, GA.
- Tavares, A., Coelho, M., Agapito, M., Coutinho, J., Xavier, A. (2006). Optimization and modeling of laccase production by *Trametes versicolor* in a bioreactor using statistical experimental design. *Applied Biochemistry and Biotechnology*. 134(3): 233-248.
- Tien M, Kirk T K (1988) Lignin peroxidase of *Phanerochaete chrysosporium*. In: Willis A. Wood S T K (ed.). *Methods in Enzymology*, Academic Press, 238–249.
- Thakur, V. V., Rakesh, K. J., Rajeev, M. M. (2012). Studies on xylanase and laccase enzymatic prebleaching to reduce chlorine-based chemicals during CEH and ECF bleaching. *BioResources*. 7(2): 2220-2235.
- Thiruchelvam, A.T., Ramsay, J.A. (2007). Growth and laccase production kinetics of *Trametes versicolor* in a stirred tank reactor. *Applied Microbiology and Biotechnology*. 74(3):547-54.
- Thurston, C. F. (1994). The structure and function of fungal laccases. *Microbiology*. 140: 19-26.
- Valls, C., Roncero, M. B. (2009). Using both xylanase and laccase enzymes for pulp bleaching. *Bioresource Technology*. 100(6): 2032-2039.
- Valls, C., Colom, J.F., Baffert, C., Gimbert, I., Roncero, M.B., Sigoillot, J. (2010). Comparing the efficiency of the laccase–NHA and laccase–HBT systems in eucalyptus pulp bleaching. *Biochemical Engineering Journal*. 49: 401–407.
- Valls, C., Quintana, E., Roncero, M. B. (2012). Assessing the environmental impact of biobleaching effects of the operational conditions. *Bioresource Technology* 104, 557- 564.
- Vasconcelos, A. F. D., Barbosa, A. M., Dekker, R. F. H., Scarminio, I. S., & Rezende, M. I. (2000). Optimization of laccase production by 161 *Botryosphaeria* sp. in the presence of veratryl alcohol by the responsesurface method. *Process Biochemistry*. 35(10):1131-1138.
- Velazquez-Cedeiio, M. A., Farnet, A. M., Ferre, E. (2004). Variations of lignocellulosic activities in dual cultures of *Pleurotus ostreatus* and *Trichoderma longibrachiatum* on unsterilized wheat straw. *Mycologia*. 96 (4): 712-719.
- Vishwanath, B., Chandra, M.S., Pallavi, H., Reddy, B.R. (2008). Screening and assessment of laccase producing fungi isolated from different environmental samples. *African Journal of Biotechnology*. 7(8): 1129-1133.

- Vivekanand, V., Dwivedi, P., Sharma, A., Sabharwal, N., Singh, R. P. (2008) Enhanced delignification of mixed wood pulp by *Aspergillus fumigatus* laccase mediator system. *World Journal of Microbiol Biotechnol* 24:2799–2804.
- Walker, G.M. (2010). *Bioethanol: science and technology of fuel alcohol*. Ventus Publishing ApS. ISBN 978-87-7681-681-0.
- Wahab, R., Mustafa, M.T., Salam, M.A., Tabert, T.A., Sulaiman, O., Sudin, M. (2012). Potential and structural variation of some selected cultivated bamboo species in peninsular Malaysia. *International Journal of Biology*. 4(3): 102.
- Wan, C., Li, Y. (2010). Microbial delignification of corn stover by *Ceriporiopsis subvermispora* for improving cellulose digestibility. *Enzyme Microbiology Technology*. 47:31–36.
- Wan, C., Li, Y. (2011). Effect of hot water extraction and liquid hot water pretreatment on the fungal degradation of biomass feedstocks. *Bioresource Technology* .102:9788–93.
- Wan Osman, H. W., Sheikh Abdullah, S. R., Abu Bakar M., Kadhum, H. A., Rahman, A. R. (2013). Simultaneous removal of AOX and COD from real recycled paper wastewater using GAC-SBBR. *Journal of Environmental Management*. 121: 80-86.
- Wang, C., Zhao, M., Li, D., Cui, D., Lu, L., Wei, X.D. (2010). Isolation and characterization of a novel *Bacillus subtilis* WD23 exhibiting laccase activity from forest soil. *African Journal of Biotechnology*. 9(34): 5496-5502.
- Wang, P., Hu, X., Cook, S., Begonia, M., Lee, K.S., Hwang, H.-M. (2008). Effect of culture conditions on the production of ligninolytic enzymes by white rot fungi *Phanerochaete chrysosporium* (ATCC 20696) and separation of its lignin peroxidase. *World Journal of Microbiology and Biotechnology*. 24: 2205–2212
- Ward, G., Hadar, Y., Bilkis, I., Dosoretz, C. G. (2003). Mechanistic features of lignin peroxidase-catalyzed oxidation of substituted phenols and 1, 2-dimethoxyarenes. *Journal of Biological Chemistry*. 278: 39726–39734.
- Wilkołazkaa, A.J, Rdesta, J.K., Yka, E.M., Wardasb, W., Leonowicza, A. (2002). Fungi and their ability to decolourize azo and anthraquinonic dyes. *Enzyme and Microbial Technology*. 30: 566-572
- Winquist, E., Moilanena, U., Mettälä, A., Leisola, M., Hatakka, A. (2008). Production of lignin modifying enzymes on industrial waste material by solid-state cultivation of fungi, *Biochemical Engineering Journal*. 42:128-132.
- Wise LE, Murphy, M., D'Addieco, AA. (1946) Chlorite holocellulose, its fractionation and bearing on summative wood analysis and on studies on the hemicelluloses. *Pap Trade Journal* 122(2):35–43.
- Wong, D. S. (2009). Structure and action mechanism of ligninolytic enzymes. *Applied Microbiology and Biotechnology*. 157: 174–209.

- Wong, L. (2016) Production of pectinase by locally-isolated fungus *Aspergillus Fumigatus* R6 in solid state fermentation for use in kenaf biorefining. Universiti Putra Malaysia. Doctor of Philosophy.
- Wu, W., Zhu, Y., Zhang, L., Yang, R., Zhou, Y. (2012). Extraction, preliminary structural characterization, and antioxidant activities of polysaccharides from *Salvia miltiorrhiza* Bunge Carbohydrate Polymers 87(2), 1348-1353.
- Xia, Z., Yoshida, T., Funaoka, M. (2003). Enzymatic degradation of highly phenolic lignin-based polymers (lignophenols). *European Polymer Journal*. 39 (5): 909–914.
- Xu, Q. H., Wang, Y. P., Qin, M. H., Fu, Y. J., Li, Z. Q., Zhang, F. S., Li, J. H. (2011). Fiber surface characterization of old newsprint pulp deinked by combining hemicellulase with laccase-mediator system *Bioresource Technology* 102 (11), 6536-40.
- Xu, G., Wang, X., Hu, J. (2013). Biobleaching of wheat straw pulp using laccase and xylanase. *BioResources* 8(3), 3181-3188.
- Yao, S., Gao, C., Zhu, H., Zhang, Y., Wang, S., Qin, C. (2016). Effects of Additives on Absorbable Organic Halide Reduction in Elemental Chlorine-Free Bleaching of Bagasse Kraft Pulp. *BioResources* 11(1), 996-1006.
- Yu, H., Guo, G., Zhang, X., Yan, K., Xu, C. (2009) .The effect of biological pretreatment with the selective white-rot fungus *Echinodontium taxodii* on enzymatic hydrolysis of softwoods and hardwoods. *Bioresource Technology*. 100: 5170–5175.
- Yoshida, H. (1883). LXIII.—Chemistry of lacquer (Urushi). Part I. Communication from the chemical society of Tokio. *Journal of the Chemical Society, Transactions*. 43: 472–486.
- Yobouet, Y.A., Adouby, K., Trokourey, A., Yao, B. (2010). Cadmium, copper, lead and zinc speciation in contaminated sBajaj, B. K., Sharma, M., Sharma, S. (2011). Alkali stable endo- β -1,4-xylanase production from a newly isolated alkali tolerant *Penicillium* sp. SS1 using agroresidues. *3 Biotech*. 1(2): 83-90.
- Zemin, F., Xiaoman, L., Liyuan, C., Yu, S., Xuecheng, Z., Wei, F., Xiaotang, W., Xiaoming, B., Yazhong, X. (2015). Identification of a laccase Glac15 from *Ganoderma lucidum* 77002 and its application in bioethanol production. *Biotechnology for Biofuels*. 8: 54.
- Zhao, G., Lai, R., He, B., Greschik, T., Li, X. (2010a). Replacement of soft wood kraft pulp with ECF-bleached bamboo kraft pulp in fine paper. *Bioresources*. 5 (3): 1733-1744.
- Zhao, D. Q., Chen, K. F., Mo, L. H., Li, J. (2010b). Chlorine dioxide bleaching reinforced by alkaline extraction and corresponding ECF bleaching sequences for wheat straw pulp. *Journal South China University of Technol.* 8,45-50.
- Zheng, Z.M., Obbard, J.P. (2001). Effect of nonionic surfactants on elimination of polycyclic aromatic hydrocarbons (PAHs) in soil slurry by *Phanerochaete chrysosporium*. *Journal of Chemical Technology and Biotechnology*. 76: 423 – 429.

Zheng, Z., Huazhong, L., Lun, L., Weilan, S. (2012). Biobleaching of wheat straw pulp with recombinant laccase from the hyperthermophilic *Thermus thermophiles*. *Biotechnology Letters* 34(3):541-7.

Zhu, H., Yao, S., Jiang, L., Wang, S., Qin, C. (2016). Kinetics of Adsorbable Organic Halogen Formation During the First Chlorine Dioxide Bleaching Stage of Eucalyptus Kraft Pulp. *BioResources* 11(4), 8820-8830.

