



UNIVERSITI PUTRA MALAYSIA

***Agrobacterium-MEDIATED TRANSFORMATION OF *Carica papaya* L.
var 'Eksotika' WITH IMPROVED RESISTANCE TO DIEBACK DISEASE***

ROSLINDA A. RAZAK

FBSB 2017 29



***Agrobacterium*-MEDIATED TRANSFORMATION OF *Carica papaya* L.
var 'Ekstotika' WITH IMPROVED RESISTANCE TO DIEBACK
DISEASE**

By

ROSLINDA A. RAZAK

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Master of Science**

April 2017

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

***Agrobacterium*-MEDIATED TRANSFORMATION OF *Carica papaya* L. var 'Eksotika' WITH IMPROVED RESISTANCE TO DIEBACK DISEASE**

By

ROSLINDA A. RAZAK

April 2017

Chair: Assoc. Prof. Janna Ong Abdullah, PhD
Faculty: Biotechnology and Biomolecular Sciences

Papaya or *Carica papaya* L. is a tropical fruit that is popular worldwide. The *Carica papaya* L. var 'Eksotika' introduced by the Malaysian Agricultural Research and Development Institute (MARDI) in 1987, is a highly sought after cultivar grown in Malaysia. Unfortunately the Malaysian papaya industry in recent years has been afflicted by many diseases. One of the major diseases identified is the papaya dieback disease caused by the bacteria *Erwinia mallotivora*. This disease was first reported in Batu Pahat, Johor in 2003. The disease can affect the whole papaya plantation via soil, airborne and environmental conditions. Current method to address this disease is by demolishing the infected area. However, this method is ineffective because the infected area will be destroyed. Therefore, this project was aimed to combat the disease by genetically engineering a papaya dieback resistance gene into the papaya plant genome via *Agrobacterium*-mediated transformation. Throughout the years, development of transgenic plants frequently relies on antibiotic as a selectable marker, which raises public concern. Hence, in this study marker-free transgenic papaya plants were created via positive selection using the *phosphomannose isomerase (pmi)* gene. Phosphomannose isomerase (PMI) is an enzyme that converts mannose-6-phosphate to fructose-6-phosphate, a glycolysis intermediate that supports the growth of plant cells. To establish a marker-free positive selection system using this PMI, the effects of mannose on the growth and development of embryogenic 'Eksotika' papaya callus was evaluated. One-month old embryogenic calli were cultured on Murashige and Skoog (MS) medium in which 60 g/L sucrose in the original recipe was replaced with different concentrations of mannose and sucrose. Mannose was supplied as the sole carbon source or in combination with sucrose at 0, 5, 10, 15, 20, 25 or 30 g/L. Embryogenic calli cultured on medium supplemented with a ratio of 0:60 g/L mannose:sucrose was used as a control. The result after six sub-cultures showed that mannose at 30 g/L mannose was effective for screening transformed embryogenic calli. Evaluation of papaya transformation efficiency using this positive selection system was pursued using one-month-old embryogenic *Agrobacterium*-transformed calli harbouring pNOV2819 carrying the *pmi* gene. Only transformed cells were capable of utilizing mannose as a carbon source to grow. After five months on mannose selection,

all 70 putative transformants obtained were PCR-positives for the *pmi* gene. Having established an antibiotic-free selection system, the next stage was targeted at combating the dieback disease. Plant disease caused by bacteria had been related to quorum sensing; whereby the bacteria monitors each other by secreting a small signal molecule known as autoinducer. It was hypothesized that interference of the signalling molecule of bacteria pathogen during infection could starve off infection. Hence, in this study, *homoserine lactone (AHL)-lactonase* gene isolated from *Bacillus* sp. strain 240B1 was used as a quorum quencher against the signalling molecule, AHL. To investigate the resistance of the 'Eksotika' papaya against *Erwinia*, the anti-pathogenic gene cassette constructed as pNOV2819:*pmi:AHL-lactonase* was transformed into the 'Eksotika' papaya embryogenic callus using *Agrobacterium tumefaciens* strain LBA4404. Mannose at 30 g/L was used during the selection process. Stable *in vitro* transgenic papaya plants were successfully obtained within 7 to 8 months on De Fossard medium during regeneration. A total of 154 transgenic plants harbouring *pmi* and *AHL-lactonase* were obtained out of 1740 transformed calli. The overall transformation frequency of pNOV2819:*pmi* was 10.8 % and pNOV2819:*pmi:AHL-lactonase* was 8.1%. PCR analysis verified to confirm that the transgene was successfully integrated into the T₀ 'Eksotika' papaya plants and the transgene was not detected in the control. In order to assess the level of resistance of T₀ plants produced against dieback, T₀ plants aged 5 to 6 months were injected with a needle at the stem area by bacteria *Erwinia mallotivora*. The result showed the control plant began to experience dieback symptoms as early as 3 days after infection. The symptoms then spread and infected other plant parts. While for transgenic plant lines, the shoots wilted and fortunately the symptoms did not spread to the other parts of the plant. The results of screening between this pathogen and *AHL-lactonase* proved that the transformation successfully disrupted the communication of the pathogen and able to reduce the spread of bacteria.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

**TRANSFORMASI *Carica papaya* L. var 'Eksotika' PENGANTARAAN-
Agrobacterium DENGAN RINTANGAN YANG BERTAMBAH BAIK
TERHADAP PENYAKIT MATI ROSOT**

Oleh

ROSLINDA A. RAZAK

April 2017

Pengerusi: Prof. Madya. Janna Ong Abdullah, PhD
Fakulti: Bioteknologi dan Sains Biomolekul

Betik atau *Carica papaya* L. adalah buah-buahan tropika yang popular di seluruh dunia. *Carica papaya* L. var 'Eksotika' yang diperkenalkan oleh Institut Penyelidikan dan Kemajuan Pertanian Malaysia (MARDI) pada tahun 1987, adalah kultivar yang dicari selepas ditanam di Malaysia. Malangnya industry betik Malaysia pada sejak kebelakangan ini telah ditimpa pelbagai penyakit. Salah satu penyakit utama yang dikenal pasti adalah penyakit betik mati rosot yang disebabkan oleh bakteria *Erwinia mallotivora*. Penyakit ini pertama kali dilaporkan di Batu Pahat, Johor pada tahun 2003. Penyakit ini boleh menjejaskan keseluruhan ladang betik melalui tanah, udara dan keadaan persekitaran. Kaedah terkini untuk menangani penyakit ini adalah dengan melupuskan kawasan yang dijangkiti. Walaubagaimanapun, kaedah ini tidak berkesan kerana kawasan yang dijangkiti akan dimusnahkan. Oleh itu, projek ini bertujuan untuk memerangi penyakit secara kejuruteraan genetik gen rintang betik mati rosot ke dalam genom tumbuhan betik melalui transformasi pengantaraan-*Agrobacterium*. Sepanjang tahun, perkembangan tumbuhan transgenik kerap bergantung kepada antibiotik sebagai penanda boleh pilih, yang menimbulkan kebimbangan orang ramai. Oleh itu, dalam kajian ini pokok betik transgenik bebas penanda telah diwujudkan melalui pemilihan positif menggunakan gen *phosphomannose isomerase (pmi)*. Phosphomannose isomerase (PMI) merupakan enzim yang menukarkan mannos-6-fosfat kepada fruktosa-6-fosfat, glikolisis perantaraan yang menyokong pertumbuhan sel-sel tumbuhan. Untuk mewujudkan satu sistem pemilihan positif penanda percuma menggunakan PMI ini, kesan mannos kepada pertumbuhan dan pembangunan embriogenik 'Eksotika' kalus betik telah dinilai. Kalus embriogenik yang berusia satu bulan telah dikulturkan pada medium Murashige dan Skoog (MS) yang mana 60 g/L sukrosa dalam resipi asal telah digantikan dengan kepekatan mannos dan sukrosa yang berbeza. Mannos telah dibekalkan sebagai sumber karbon tunggal atau dalam

kombinasi dengan sukrosa pada 0, 5, 10, 15, 20, 25 atau 30 g/L. Kalus embriogenik yang dikulturkan pada medium dengan nisbah 0:60 g/L mannosas:sukrosa telah digunakan sebagai kawalan. Hasil selepas enam kali sub-kultur menunjukkan bahawa mannosas pada 30 g/L berkesan untuk saringan berubah kalus embriogenik. Penilaian kecekapan transformasi betik menggunakan sistem pemilihan positif ini seterusnya menggunakan kalus embriogenik yang berusia satu bulan yang ditransformasikan dengan *Agrobacterium* yang mengandungi pNOV2819 pembawa gen *pmi* itu. Hanya sel-sel yang telah ditransformasi mampu menggunakan mannosas sebagai sumber karbon untuk berkembang. Selepas lima bulan pada pemilihan mannosas, ke semua 70 transformasi diperoleh adalah PCR-positif untuk gen *pmi*. Setelah menetapkan sistem pemilihan bebas antibiotik, peringkat seterusnya disasarkan untuk memerangi penyakit mati rosot ini. Penyakit tumbuhan yang disebabkan oleh bakteria telah berkaitan dengan penderiaan kuorum; dimana bakteria memantau antara satu sama lain dengan merembeskan molekul isyarat kecil yang dikenali sebagai molekul pengesan (*autoinducer*). Hipotesis telah menunjukkan bahawa gangguan isyarat molekul bakteria patogen semasa jangkitan boleh menghalang jangkitan. Oleh itu, dalam kajian ini, *homoserine lactone (AHL) -lactonase* gen diambil daripada *Bacillus* sp. strain 240B1 telah digunakan sebagai kuorum pelindapkejutan terhadap isyarat molekul, AHL. Untuk menguji kerintangan betik 'Eksotika' terhadap *Erwinia*, gen kaset anti-patogenik dibina sebagai pNOV2819:*pmi:AHL-lactonase*, telah ditransformasikan ke dalam kalus embriogenik betik 'Eksotika' dengan menggunakan *Agrobacterium tumefaciens* strain LBA4404. Mannosas pada 30 g/L digunakan semasa proses pemilihan. Tanaman *in vitro* betik transgenik telah berjaya diperolehi dalam masa 7-8 bulan keatas medium De Fossard semasa pertumbuhan semula. Sebanyak 154 tumbuhan transgenik mengandungi *pmi* dan *AHL-lactonase* diperolehi daripada 1740 kalus yang telah ditransformasikan. Kadar keseluruhan transformasi pNOV2819:*pmi* adalah 10.8% dan pNOV2819:*pmi:AHL-lactonase* adalah 8.1%. Analisis PCR telah mengesahkan bahawa transgen itu berjaya disepadukan ke dalam T₀ 'Eksotika' dan transgen itu tidak dikesan dalam kawalan. Untuk menilai tahap ketahanan tumbuhan T₀ yang dihasilkan terhadap mati rosot, pokok-pokok T₀ berumur 5 hingga 6 bulan telah disuntik dengan jarum di kawasan batang dengan bakteria *Erwinia mallotivora*. Hasilnya menunjukkan pokok kawalan mula mengalami gejala mati rosot seawal 3 hari selepas jangkitan. Tanda-tanda itu kemudiannya merebak dan telah menjangkiti ke kawasan yang lain. Manakala bagi barisan pokok transgenik, pucuk didapati layu dan bernasib baik ia tidak merebak ke bahagian-bahagian pokok yang lain. Keputusan saringan antara patogen ini dan *AHL-lactonase* membuktikan bahawa transformasi telah berjaya mengganggu komunikasi daripada patogen dan dapat mengurangkan penyebaran bakteria.

ACKNOWLEDGEMENTS

In the name of Allah, The Most Gracious, The Most Merciful

I would like to express my most sincere gratitude and deepest appreciation to my main supervisor Assoc. Prof. Janna Ong Abdullah for giving me the opportunity to further my study in the field of plant genetic engineering. Thanks a lot for your countless efforts in advising me and guiding me to finish my study. Also, special thanks to my co-supervisors Dr. Rogayah binti Sekeli and Dr Noor Azmi bin Shahrudin for their constant motivation and words of advice; they inspired me to become an independent researcher. To Dr. Rogayah, I really appreciate every single encouragement given for me to finish my laboratory work and thank you for the supply of every equipment.

Special thanks are extended also to laboratory assistants from MARDI; Puan Radnadia, Puan Noraini, Miss Siti Fatiha, Miss Arina Watikah and Miss Fadzliana for all their help and support during my study.

Finally, I wish to express my deepest appreciation to my family for their constant support during my study, especially to my mother and my family.

I certify that a Thesis Examination Committee has met on 14 March 2017 to conduct the final examination of Roslinda binti A Razak on her thesis entitled "*Agrobacterium*-Mediated Transformation of *Carica papaya* L. var 'Eksotika' with Improved Resistance to Dieback Disease" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Muhajir bin Hamid, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Christina Yong Seok Yien, PhD

Senior Lecturer
Faculty of Science
Universiti Putra Malaysia
(Internal Examiner)

Tee Chong Siang, PhD

Associate Professor
Universiti Tunku Abdul Rahman
Malaysia
(External Examiner)



NOR AINI AB. SHUKOR, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 2 June 2017

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Janna Ong Abdullah, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Noor Azmi bin Shaharuddin, PhD

Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Rogayah Sekeli, PhD

Research Officer,
Agri-Omics and Bioinformatics (BT1), Biotechnology and Nanotechnology
Research Centre
Malaysian Agricultural Research and Development Institute (MARDI)
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: _____

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman of
Supervisory
Committee: Janna Ong Abdullah

Signature: _____
Name of Member of
Supervisory
Committee: Noor Azmi bin Shahrudin

Signature: _____
Name of Member of
Supervisory
Committee: Rogayah Sekeli

TABLE OF CONTENTS

		Page
ABSTRACT		i
ABSTRAK		iii
ACKNOWLEDGEMENTS		v
APPROVAL		vi
DECLARATION		viii
LIST OF TABLES		xiv
LIST OF FIGURES		xv
LIST OF ABBREVIATIONS		xvii
CHAPTER		
1	INTRODUCTION	1
2	LITERATURE REVIEW	3
2.1	Papaya	3
2.1.1	Commercial uses of <i>Carica papaya</i> L.	3
2.1.2	General biology of <i>Carica papaya</i> L.	4
2.1.2.1	Stem	5
2.1.2.2	Fruit	5
2.1.2.3	Seed	6
2.1.3	<i>Carica papaya</i> L. Var 'Eksotika'	6
2.1.4	Genetic transformation of <i>Carica papaya</i> L.	6
2.2	Problems encountered by papaya industry	7
2.2.1	Papaya dieback disease	7
2.3	<i>Agrobacterium</i> -mediated transformations	9
2.3.1	Factors influencing the transformation efficiency	12
2.3.1.1	Explants types	12
2.3.1.2	<i>Agrobacterium</i> strains	12
2.3.1.3	Plasmolysis and co-cultivation conditions	12
2.3.2	Removal of <i>Agrobacterium tumefaciens</i> after transformation	13
2.4	Selectable-marker	13
2.4.1	Phosphomannose isomerase (PMI)	14
2.5	Genetic engineering to address the papaya dieback disease	16
2.5.1	Acyl-homoserine lactone (AHL)	16
2.5.1.1	Acyl-homoserine lactonase	19
2.5.1.2	Mechanism of AHL-lactonase in response to	19

	<i>Erwinia</i> sp. Pathogenicity	
2.6	Issues and potentials risks of transgenic plants to human and environment	20
3	MATERIALS AND METHODS / METHODOLOGY	22
3.1	Preparation of pNON2818: <i>pmi</i>	22
3.1.1	Preparation of <i>Escherichia coli</i> strain DH5 α	22
3.1.2	Transformation of pNOV2819: <i>pmi</i> into <i>Escherichia</i>	22
3.1.3	Preparation of <i>Agrobacterium tumefaciens</i> strain LBA4404 competent cells	23
3.1.4	Transformation of pNOV2819: <i>pmi</i> into <i>Agrobacterium tumefaciens</i> strain LBA4404 competent cells	23
3.1.5	Plasmid isolation of pNOV2819: <i>pmi</i> from <i>Agrobacterium tumefaciens</i>	23
3.1.6	Agarose gel electrophoresis	24
3.2	Construction of pNOV2819: <i>pmi</i> harbouring <i>Acyl-homoserine lactonase (AHL-lactonase)</i>	24
3.2.1	Transformation of pNOV2819: <i>pmi:AHL-lactonase</i> into <i>Escherichia coli</i> strain DH5 α competent cells	24
3.2.2	Transformation of pNOV2819: <i>pmi:AHL-lactonase</i> into <i>Agrobacterium tumefaciens</i> strain LBA4404 competent cells	25
3.2.3	Verification of the <i>AHL-lactonase</i> transformed in <i>Agrobacterium tumefaciens</i> strain LBA4404 cells	25
3.2.4	PCR amplification of target gene, <i>AHL-lactonase</i> gene	25
3.2.5	Restriction enzyme digestion	26
3.2.6	Glycerol stock preparation	26
3.3	Development of non-antibiotics selectable marker system	26
3.3.1	Embryogenesis callus induction of <i>Carica papaya</i> L. var 'Eksotika'	26
3.3.2	Effects of various mannose concentrations on embryogenic callus growth	26
3.4	<i>Agrobacterium</i> -mediated transformation of <i>Carica papaya</i> L. var 'Eksotika'	27
3.4.1	<i>Agrobacterium</i> -mediated transformation of embryogenic calli	27
3.4.2	Plasmolysis and co-cultivation of transformed calli	27
3.4.3	Embryogenic induction of	28

	transformed calli	
	3.4.4 Selection of transformed calli	28
	3.4.5 Maturation and regeneration of transformed calli	29
3.5	Regeneration of putative transgenic papaya T ₀ plants on De Fossard medium	29
3.6	<i>In-vitro</i> challenge of putative transgenic papaya with <i>Erwinia mallotivora</i>	29
	3.6.1 Plants materials and <i>Erwinia mallotivora</i> inoculation	30
	3.6.2 Samples treatment	30
	3.6.3 Nano drop spectrophotometer	30
3.7	Analysis of <i>pmi</i> and <i>AHL-lactonase</i> positive transgenic plants	30
	3.7.1 PCR amplification	31
4	RESULTS AND DISCUSSION	32
4.1	Development of <i>AHL-lactonase</i> gene construct	32
4.2	Confirmation of <i>AHL-lactonase</i> construct by PCR analysis	32
	4.2.1 Restriction enzyme digestion of <i>AHL-lactonase</i> gene cassette and confirmation via sequencing	33
4.3	Transformation of <i>pmi</i> and <i>AHL-lactonase</i> gene construct into <i>Agrobacterium tumefaciens</i>	34
4.4	Induction of embryogenic callus of 'Eksotika' papaya	34
4.5	Effects of mannose on growth and development of non-transformed papaya callus culture	35
4.6	Development of embryogenic callus for <i>Agrobacterium</i> -mediated transformation	37
	4.6.1 <i>Agrobacterium</i> -mediated transformation of <i>pmi</i> and <i>AHL-lactonase</i>	38
	4.6.2 <i>In-vitro</i> regeneration of putative transgenic shoots using De Fossard Medium	40
4.7	PCR analysis of putative regenerated T ₀ transgenic papaya shoots	41
4.8	<i>In-vitro</i> challenge of transgenic papaya with <i>Erwinia mallotivora</i>	43
4.9	<i>Ex-vitro</i> challenge of transgenic papaya plants with <i>Erwinia mallotivora</i>	44
5	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	50

REFERENCES	51
APPENDICES	62
BIODATA OF STUDENT	71
LIST OF PUBLICATIONS	72



LIST OF TABLES

Table		Page
3.1	Sequence of the primers used for <i>pmi</i> and <i>AHL-lactobase</i> constructs	25
3.2	Characteristics of transgenic and control plants	30
4.1	<i>Ex-vivo</i> challenge of transgenic papaya plants with <i>Erwinia mallotivora</i>	47



LIST OF FIGURES

Figure		Page
2.1	Vegetative parts of the papaya plant	4
2.2	Papaya fruits based on sex type	5
2.3	Image of <i>Erwinia mallotivora</i> under transmission electron microscope	9
2.4	Symptoms of papaya dieback disease caused by <i>Erwinia mallotivora</i>	9
2.5	Process of <i>Agrobacterium</i> -mediated transformation in a plant cell	11
2.6	Reaction catalyzed by phosphomannose isomerase (PMI)	15
2.7	Overview secretion of AHL	18
2.8	Mechanism of AHL lactonase	20
3.1	Plasmid map of pNOV2819	27
4.1	PCR analysis of AHL-lactonase construct using specific primers	33
4.2	<i>Nco</i> I digestion of pCAMBIA2301: <i>AHL-lactonase</i>	33
4.3	Morphology of embryogenic callus induced from immature zygotic embryo	35
4.4	Effects of mannose on growth and development of embryogenic papaya calli after 5 months	37
4.5	<i>Agrobacterium</i> -mediated transformation of 'Eksotika' papaya callus	40
4.6	Regeneration of putative transformants on De Fossard medium	41
4.7	PCR analysis of putative transformants harbouring <i>pmi</i> and AHL-lactonase genes	42
4.8	<i>In-vitro</i> challenges of T ₀ transgenic papaya plantlets with <i>Erwinia mallotivora</i>	44

4.9 Papaya dieback disease symptoms observed on putative transgenic T₀ plants infected with *Erwinia mallotivora*

46



LIST OF ABBREVIATIONS

2,4 D	2,4 dichlorophenoxyacetic acid
35S	cauliflower mosaic virus
AHL	Acyl-homoserine lactone
AHL-lactonase	Acyl-homoserine lactonase
BAP	6-Benzylaminopurine
BLAST	Basic Local Alignment Search Tool
bp	base pair
cm	centimeter
DMSO	dimethylsulphonyl oxide
DNA	deoxyribonucleic acid
DNase	deoxyribonuclease
<i>E.coli</i>	<i>Escherichia coli</i>
EDTA	ethylenediaminetetraacetic acid
Etbr	ethidium bromide
<i>et al.</i>	Et alia (and other)
g	gram
gusA	beta-glucuronidase
IBA	indole-3-butyric acid
LB	Luria-Bertani
M	molar
MARDI	Malaysian Agricultural Research and Development Institute
min	minute
mM	millimolar
mRNA	messenger ribonucleic acid
MS	Murashige and Skoog basal medium

NAA	α -naphthaleneacetic acid
NCBI	National Centre for Biotechnology Information
OD	optical density
PCR	Polymerase chain reaction
RNA	ribonucleic acid
RNase	ribonuclease
RT	room temperature
Sec	second
TAE	Tris boric acid EDTA
TE	Tris-EDTA
μ g	microgram
μ m	micromolar
v/v	volume per volume
w/v	weight per volume

CHAPTER 1

INTRODUCTION

Papaya or *Carica papaya* L. is a member of the dicotyledonous plant from the family Caricaceae. It consists of four genera such as *Carica*, *Jacaratia*, *Jarilla* and *Cylicomorpha*. According to Department of Statistics Malaysia, papaya is an important horticultural crop in Malaysia, with self-sufficiency ratio of about 165.4% in 2015. 'Eksotika' papaya, a cross between the 'Subang 6' and the 'Hawaiian Sunrise Solo', is a popular papaya variety grown in Malaysia. It was released by the Malaysian Agricultural Research and Development Institute (MARDI) in 1987 (Chan, 1987). The sweet taste and pleasant aroma of 'Eksotika' papaya resulted in it to be highly demanded in the local and export markets, such as China, Hong Kong, Singapore, the Europe and Middle East (Sew *et al.*, 2011).

The 'Eksotika' papaya variety is the most popular grown cultivar and one of the main varieties grown for export in Malaysia. Currently, one of the major diseases that attack the Malaysian papaya industry is the papaya dieback disease. This disease could destroy the whole papaya plant, and thereby, can result in 100% production losses. Papaya dieback was first reported in Batu Pahat, Johor in 2003. It spread to other papaya plantations in Perak, Selangor, Penang, Kedah and Pahang, affecting about 800 ha and resulting 100 % in the destruction (Roshidi, 2010). The total yield lost was estimated at 200,000 metric tons, equivalent to US\$ 58 million. The affected varieties besides 'Eksotika' are such as 'Solo', 'Hong Kong' and 'Sekaki'. The pathogen that causes the disease has been determined as *Erwinia mallotivora*, from the family *Enterobacteriaceae* (Noriha *et al.*, 2011).

Currently the only solution is by destroying the infected plant. Therefore, development of an 'Eksotika' papaya plant that harbours an *acyl-homoserine lactonase* (*AHL-lactonase*) gene conferring resistance to dieback through genetic engineering technique is seen as an alternative method to overcome and control this disease, besides upholding good management practice in the plantations. The current method for papaya transformation relies on a good antibiotic selectable marker gene for selecting the putative transformed tissues. Unfortunately, such use of antibiotic had resulted in widespread public concern, as it has potential consequences on the environment and human health (Wang *et al.*, 2000). Therefore, in order to eliminate the use of antibiotic for selection of transformed tissues, an alternative efficient selection method was developed in this project. The strategy involved using a positive selection system such as phosphomannose isomerase (PMI) to select genetically transformed tissues capable of metabolising and growing on mannose supplemented medium. It was hoped that the usage of such antibiotic-free selection can help reduce the negative perception among the public on genetically engineered products, which were created to increase crop productivity.

Therefore, the objectives for the present study were:

1. to establish a non-antibiotic selectable marker system for *Carica papaya* L. var 'Eksotika',
2. to construct a gene transformation cassette harbouring *acyl-homoserine lactonase* and *phosphomannose isomerase* for *Agrobacterium*-mediated transformation,
3. to transfer the *AHL-lactonase* and *pmi* genes into *Carica papaya* L. var 'Eksotika' callus via *Agrobacterium*-mediated transformation, and
4. to challenge the characterised T₀ transformed papaya plantlets with *Erwinia mallotivora*.



REFERENCES

- Aboshama, H. M. S. (2011). Somatic embryogenesis, proliferation, maturation and germination in *Cajanus*. *World Journal of Agricultural Sciences*, 7:86-95.
- Alcorn, J. L. (1968). Cucurbit powdery mildew on pawpaw. *Queensland Journal of Agricultural and Animal Sciences*, 2: 161–164.
- Ali, A., Ong, M.K. and Forney, C.F. (2014). Effect of ozone pre-conditioning on quality and antioxidant capacity of papaya fruit during ambient storage. *Food Chemistry*, 142:19-26.
- Asma, N., Kashif, A. and Saifullah, K. (2008). An optimized and improved method for the *in vitro* propagation of kiwifruit (*Actinidia deliciosa*) using coconut water. *Pakistan Journal of Botany*, 40:2355-2360.
- Azad, M.A. and Rabbani, M.G. (2005). Genetic transformation of *Carica papaya* by infecting mature zygotic embryo with *Agrobacterium tumefaciens* strain LBA 4404. *Biotechnology*, 4:235-237.
- Banerje, J. (2002). Tissue culture and transformation studies in Indian cultivars of papaya (*Carica papaya* L.) Ph.D. Thesis, University of Pune, India.
- Barb, A. W., Pharr, D. M., Williamson, and J. D. (2003). A *Nicotiana tabacum* cell culture selected for accelerated growth on Man as increased expression of Phosphomannose Isomerase. *Journal of Plant Science*, 165: 639-648.
- Barrios, A.F.G, Covo, V., Medina, L.M., Vives-Florez, M. and Achenie, L. (2009). Quorum quenching analysis in *Pseudomonas aeruginosa* and *Escherichia coli*: network topology and inhibition mechanism effect on the optimized inhibitor dose. *Bioprocess Biosystem Engineering*, 32:545–56.
- Beatriz W., Fabricio F., Giancarlo P., Maria H. B.Z. and Annette D. (2006). Influence of antibiotics on embryogenic tissue and *Agrobacterium tumefaciens* suppression in soybean genetic transformation. *Bragantia*, Campinas, Vol 65: 543-551. DOI: 10.1590/S0006-87052006000400002.
- Bhattacharya, J., Khuspe, S.S., Renukdas, N.N. and Rawal, S.K (2002). Somatic embryogenesis and plant regeneration from immature embryo explant of papaya (*Carica papaya* L. cv. Washington and Honey Dew). *Indian Journal of Experimental Biology*, 40:624-627.

- Bojsen, K., Donaldson, I., Haldrup, A., Joersbo, M., Kreierg, J.D., Nielsen, J., Okkels, F.T., Peterson, S. G. and Whenham, R. J. (1999). Positive Selection. United States Patent No. 5,994,629. November 30, 1999.
- Boscariol, R. L., Almeida, W. A., Derbyshire, M. T., Mourao, Filho, F. A. and Mendes, B. M. (2003). The use of the PMI/Mannose selection system to recover transgenic sweet orange plants (*Citrus sinensis* L. Osbeck). *Plant Cell Reports*, 22: 122-8.
- Boshra, V. and Tajul, A.Y. (2013). Papaya - an innovative raw material for food and pharmaceutical processing industry. *Journal of Health Environment*, 4(1):68-75.
- Burkill, I.H. (1966). *A Dictionary of the Economic Products of the Malay peninsula* (2ndEDn), Malay Ministry of Agriculture and Co-operatives, Kuala Lumpur.
- Chan, Y. K. (1987). Backcross method in improvement of papaya (*Carica papaya* L.). *Malaysian Applied Biology*.16:95-100.
- Chen G., Ye C.M., Huang J.C., Yu M., and Li B.J. (2001).Cloning of the Papaya ringspot virus (PRSV) replicase gene and generation of PRSV-resistant papayas through the introduction of the PRSV replicase gene. *Plant Cell Reports*, 20:272-277.
- Chen, M.H., Wang, P.J. and Maeda, E. (1987). Somatic embryogenesis and plant regeneration in *Carica papaya* L. tissue culture derived from root explants. *Plant Cell Report*, 6:348-351.
- Cheng Y.H., Yang J.S. and Yeh S.D. (1996).Efficient transformation of papaya by coat protein gene of papaya ringspot virus mediated by *Agrobacterium* following liquid-phase wounding of embryogenic tissue with caborundum. *Plant Cell Report*, 16:127-132.
- Chupp, C. (1953). A monograph of the fungus genus *Cercospora*. Published by the author, New York, Ithaca, pp. 667.
- Conover, R. A. (1962). Virus diseases of the papaya in Florida. *Phytopathology*, 52:6.
- Cook, A. A. and Milbrath, G. M. (1971). Virus diseases of papaya on Oahu (Hawaii) and identification of additional diagnostic host plants. *Plant Disease Report*, 55(9): 785-788.

- Czajkowski, R. and Jafra, S. (2009). Quenching of acyl-homoserine lactone-dependent quorum sensing by enzymatic disruption of signal molecules. *Acta Biochimica Polonica*, 56: 1–16.
- De Block, M., Herrera-Estrella L., Van Montagu, M., Schell, J. and Zambryski P. (1984). Expression of foreign genes in regenerated plants and in their progeny, *The EMBO Journal*, 3: 1681–9.
- De Fossard, R., Myint, A. and Lee, E. (1974). A broad spectrum tissue culture experiment with tobacco (*Nicotiana tabacum* L.) pith tissue callus. *Plant Physiology*, 31: 125-130.
- Dong, Y.H. (2001). Quenching quorum-sensing-dependent bacterial infection by an N-acyl homoserine lactonase. *Nature*, 411:813–817.
- Dong, Y.H., Xu, J.L., Li, X.Z. and Zhang, L.H. (2000). AiiA, an enzyme that inactivates the acyl-homoserine lactone quorum-sensing signal and attenuates the virulence of *Erwinia carotovora*. *Proceedings of the National Academy of Sciences U. S. A.*, 97:3526–3531.
- Eady, C.C., Weld, R. J. and Lister, C.E. (2000). *Agrobacterium tumefaciens*-mediated transformation and transgenic-plant regeneration of onion (*Allium cepa* L.). *Plant Cell Reports*, 19: 376-381.
- Enriquez-Obregon, G. A., D. L. Prieto-Samsonov, G. A. de la Riva, M. Perez, G. Selman, H. and R. I. Vazquez-Padron. (1999). *Agrobacterium*-mediated Japonica rice transformation: a procedure assisted by an anti-necrotic treatment. *Plant Cell Tissue Organ Culture*, 59:159–168.
- Fitch, M. and Manshard, T.R. (1990). Somatic embryogenesis and plant regeneration from immature zygotic embryos of papaya (*Carica papaya* L.). *Plant Cell Reports*, 9:320-324.
- Fitch, M. (1993). High-frequency somatic embryogenesis and plant regeneration from papaya hypocotyl callus. *Plant Cell, Tissue and Organ Culture*, 32:205-212.
- Fitch, M.M. (2005). *Carica papaya*. Chapter 6.1. In: RE Litz, ed. *Biotechnology of Fruit and Nut Crops*. CABI Publishing, pp. 174-207.
- Fitch, M.M., Manshardt, R.M., Gonsalves, D., Slinghtom, J.L. and Sanford, J.C. (1990). Stable transformation of papaya via microprojectile bombardment. *Plant Cell Report*, 9:189-194.

- Gadaleta, A., Giancaspro, A., and Blanco, A. (2004). Phosphomannose isomerase, PMI, as selectable marker gene in durum wheat transformation. *Proc. of XLVII Italian Society of Agricultural Genetics*.
- Gill, A. T., Snyman, S.J., Potier, B.A.M. and Hockett, B.I. (2004). Towards antibiotic resistance-free transgenic sugarcane. *African Sugarcane Technology*, 78:163-166.
- Goldsworthy, A. and Street, H.E. (1965). The carbohydrate nutrition of tomato roots VIII: The mechanism of the inhibition by D-mannose of the respiration of excised roots. *Annals of Botany*, 29: 45-58.
- Gonsalves, D., Gonsalves, C., Ferreira, S., Pitz, K., Fitch, M.M., Manshardt, R. and Slightom, J. (2004). Transgenic virus resistant papaya: from hope to reality for controlling papaya ringspot virus in Hawaii. *Phytopathology supplement: APSnet (Plant phytoogy online). ASPnet feature. American Phytopathological Society*. <http://www.apsnet.org.online.feature/ringspot>.
- Goto, M. (1976). *Erwinia mallotivora* sp. Novel, the causal organism of bacterial leaf spot of *Mallotus japonicus*. *International Journal Systematic Bacteriology*, 26: 467-473.
- Gwyne, D.C. (1984). Fire blight in perry pears and cider apples in the South West of England. *Acta Horticulturae*, 151, 41-47.
- Hentzer, M. and Givskov, M. (2003). Pharmacological inhibition of quorum sensing for the treatment of chronic bacterial infections. *Journal Clinical Investigation*, 112:1300-1307.
- Hoa, T.T. and Bong, B.B. (2002). *Agrobacterium*-mediated transformation of rice embryogenic suspension cells using phosphomannose isomerase gene, pmi, as a selectable marker. *Omonrice*, 10:1-5.
- Hoekema, A., Hirsch, P.R., Hooykaas, P.J.J. and Schilperoort, R.A. (1983). A binary plant vector strategy based on separation of vir- and T-region of the *Agrobacterium tumefaciens* Ti plasmid. *Nature*, 303:179-180.
- Horsch, R.B., Fraley, R.T., Rogers, S.G., Sanders, P.R., Lloyd, A. and Hoffmann, N. (1984). Inheritance of functional genes in plants. *Science*, 223: 496-498.
- Jaime, A., Teixeira, Da S., Zini, R., Duong, T.N., Dharin,i S., Abed, G., Manoel, T.S. and Paula, F.T. (2007). Papaya (*Carica papaya* L.) biology and biotechnology. *Tree and Forestry Science and Biotechnology*, 1:47-73.

- Jain, M., Chengalrayan, K., Abouzid, A. and Gallo, M. (2007). Prospecting the utility of a PMI/mannose selection system for the recovery of transgenic sugarcane (*Saccharum* spp. hybrid) plants. *Plant Cell Reports*, 26:581–590.
- Jime'nez, V.M., Mora-Newcomer, M. and Marco, V.M.S. (2008). *Chapter 2. Biology of the Papaya Plant*, pp. 26.
- Joersbo, M. (2001). Advances in the selection of transgenic plants using non-antibiotic marker genes. *Journal of Plant Physiology*, 111: 269-272.
- Joersbo, M., Danaldson, I., Kreiberg, J., Petersen, S.G. Brunstedt, J. and Okkels, F. T. (1998). Analysis of mannose selection used for transformation of sugar beet. *Journal of Molecular Breeding*, 4: 111-117.
- John, F.E, Bruce, D.M., David, A.M., Matthew, R.R. and Richard G.S. (2011). 2,4-Dichlorophenoxyacetic acid (2,4-D)-resistant crops and the potential for evolution of 2,4-D-resistant weeds. PNAS Early Edition www.pnas.org/cgi/doi/10.1073/pnas.1017414108.
- Jordan, M. and Velozo, J. (1996). Improvement of somatic embryogenesis in highland-papaya cell suspensions. *Plant Cell, Tissue and Organ Culture*, 44:189-194.
- Khanna, H., Becker, D., Kleidon, J. and Dale, J. (2004). Centrifugation assisted *Agrobacterium tumefaciens*-mediated transformation (CAAT) of embryogenic cell suspensions of banana (*Musa* spp. Cavendish AAA and Lady finger AAB). *Molecular Breeding*, 14: 239–252.
- Kidwell, M.G. (1993). Lateral transfer in natural populations of eukaryotes. *Annual Review of Genetics*, 27:235-256.
- Ko, W. H. (1971). Biological control of Phytophthora root rot of papaya with virgin soil. *Plant Disease*, 66:446-448.
- Kondo, T., Hasegawa, H. and Suzuki, M. (2000). Transformation and regeneration of garlic (*Allium sativum* L.) by *Agrobacterium*-mediated gene transfer. *Plant Cell Reports* 19: 989-993.
- Lai, F.M., Mei, K., Mankin, L. and Jones, T. (2007). Application of two new selectable marker genes, *dsdA* and *in* maize transformation. Proceedings of the 11th IAPTC & congress, Beijing, China. *The Netherlands: Springer*, p :141–2.
- Limanton-Grevet. and Jullien, M. (2001). *Agrobacterium*-mediated transformation of *Asparagus officinalis* L. molecular and genetic analysis of transgenic plants. *Molecular Breeding*, 7:141-150.

- Lin, Y.H., Xu, J.L., Hu, J., Wang, L.H., Ong, S.L., Leadbetter, J.R. and Zhang, L.H. (2003). Acyl-homoserine lactone acylase from *Ralstonia* strain XJ12B represents a novel and potent class of quorum-quenching enzymes. *Molecular Microbiology*, 47: 849–860.
- Litz, R.E. and Conover R.A. (1982). *In vitro* somatic embryogenesis and plant regeneration from *Carica papaya* L. ovular callus. *Plant Science Letters*, 26:153-158.
- Litz, R.E. and Gray, D.J. (1992). Organogenesis and somatic embryogenesis, in F.A. Hammerschlag and R.E. Litz (eds). *Biotechnology of perennial fruit crops. Biotechnology in agriculture. No.8*, CAB International, Wallingford, UK, pp, 3-34.
- Magdalita, P.M., Laurena A.C., Yabut-Perez, B.M., Botella, J.R., Tecson-Mendoza, E.M. and Villegas, V.N. (2002). Progress in the development of transgenic papaya: Transformation of Solo papaya using *ACC synthase* antisense construct. *Acta Horticulturae*, 575:171-176.
- Magdalita, P.M., Yabut-Perez, E.M., Mendoza, V.N., Villegas, V.N. and Botella, J.R. (2002). Towards transformation, regeneration and screening of papaya containing antisense *ACC synthase* gene. *10th International Association of Plant Tissue Culture and Biotechnology Congress, Plant Biotechnology 2002 and Beyond: A showcase, Orlando, FL*, Abstract S42.
- Mahon R.E., Bateson M.F., Chamberlain D.A., Higgins C.M., Drew R.A. and Dale J.L. (1996). Transformation of an Australian variety of *Carica papaya* using microprojectile bombardment. *Australian Journal of Plant Physiology*, 23:679-685.
- Makhtar, N.H., Kamis, S., Yusof, F.Z. and Hussain, N.H. (2008). *Erwinia papaya* causing papaya dieback in Malaysia. *New Disease Reports* 17, 4. The British Society for Plant Pathology.
- Malca, I., Endo, R.M. and Long, M.R. (1967). Mechanism of glucose counteraction of inhibition of root elongation by galactose, mannose and glucosamine. *Phytopathology*, 57:272-278.
- Manshardt, R.M. (1992). Papaya. In: Hammerschlag FA, Litz RE (Eds) *Biotechnology in Agriculture No. 8. Biotechnology of Perennial Fruit Crops*, CABI, Wallingford, pp. 489-511.

- Mathias, T.J. and Boyd, L.A. (1986). Cefotaxime stimulates callus growth, embryogenesis and regeneration in hexaploid bread wheat (*Triticumaestivum* L. EM. Thell). *Plant Science*, 46:217-233.
- Michael, J. and Bonnie, L. (2003). Interspecies communication in bacteria. *Journal of Clinical Investigation*, 112:1291-1299.
- Miller, M.B., and Bassler, B.L. (2001). Quorum sensing in bacteria. *Annual Review Microbiology*, 55:165-199.
- Morton, J. (1987). Papaya. In: Fruits of warm climates. Miami, Florida, pp. 336-346.
- Muhammad, Z.A., Iqbal, H., Aish, M., Shaukat, A.G., Muhammad, A., Sohaib R., Muhammad, A.Z. and Amir, I. (2012). Factor affecting *Agrobacterium*-mediated transformation of rice chitinase gene in *Solanum tuberosum* L. *African Journal of Biotechnology*, 11:9716-9723.
- Murashige, T. and Skoog, F. (1962). A revised medium for rapid growth and bioassay with tobacco tissue culture. *Plant Physiology*, 15:473-497.
- Murashige, T. and Skoog, F. (1962). A revised medium for rapid growth and bioassay with tobacco tissue culture. *Journal of Plant Physiology*, 15: 473-497.
- Nakano, M. and Mii, M. (1993). Antibiotics stimulate somatic embryogenesis without plant growth regulators in several *Dianthus* cultivars. *Journal of Plant Physiology*, 141: 721-725.
- Nakasone, H.Y. and Paull, R. E. (1998). Tropical Fruits. CAB International, Oxon, UK, p 443.
- Nariana, T.K. (1956). Leaf Curl of Papaya. *Indian Phytopathology*, 9:151 - 155.
- Negrotto, D., Jolley, M., Beer S., Wenck, A. and Hansen, G. (2000). The use of *phosphomannose-isomerase* as selectable marker to recover transgenic maize plants (*Zea mays* L.) via *Agrobacterium* transformation. *Plant Cell Reports*, 19:798-803.
- Noriha, M.A., Hamidun, B., Rohaiza, A.R. and Indu, B.S.J. (2011). *Erwinia mallotivora* sp., a New Pathogen of Papaya (*Carica papaya*) in Peninsular Malaysia. *International Journal Molecular Sciences*, 12:39-45.
- O'Kennedy, M. M., Burger, J. T. and Botha, F. C. (2004). Pearl millet transformation system using the positive selectable marker gene phosphomannose isomerase. *Plant Cell Reports*, 22:684-90.

- Pang, S.Z. and Sanford J.C. (1988). *Agrobacterium*-mediated gene transfer in papaya. *Journal of the American Society for Horticultural Science*, 113:287-291.
- Pantoja, A., Follett, P.A. and Villanueva-Jiménez, J.A. (2002). Pests of papaya. In: Pena J, Sharp J, Wysoki M (Eds) *Tropical Fruit Pests and Pollinators: Biology, Economic Importance, Natural Enemies and Control*, CABI Publishing, Cambridge, pp. 131-156.
- Paszkowski, J., Shillito, R.D., Saul. M., Mondfik, V., Hohn, T., Hohn, B. and Potrykus, I. (1984). Direct gene transfer to plants. *The EMBO Journal*, 3:2717-2722.
- Pego, J.V., Weisbeek, P.J. and Smeekens, S.C.M. (1999). Mannose inhibits *Arabidopsis* germination via a hexokinase-mediated step. *Plant Physiology*, 1991:1017-1023.
- Pollock, H.M., J. Holt and C. Murray. (1983). Comparison of susceptibilities of anaerobic bacteria to cefemenoxime, ceftriaxone, and other antimicrobial compounds. *Antimicrobial Agents and Chemotherapy*, 23:780-783.
- Pollock, K., Barfield, D.G., and Schields, R. (1983). The toxicity of antibiotics to plant cell cultures. *Plant Cell Reports*, (Vol 2), pp.36-39, 1983.
- Priyanga, R., Kaushalya, W.P., Sirimal Premakumara, G.A., Padmalal, G. and Saman, B. G. (2012). In vitro erythrocyte membrane stabilization properties of *Carica papaya* L. leaf extract. *Pharmacognosy Research*, 4:196-202.
- Purseglove, J.W. (1968). Caricaceae. In: *Tropical Crops. Dicotyledons* (Vol. 1), Longmans, Green and Co., Bristol, pp. 45-51.
- Qing Z., Ji J., Gang W., Jie H.W., Yuan Y. F., Han W.X. and Chun F.G. (2010). Optimization in *Agrobacterium*-mediated transformation of *Anthuriumandraeanum* using GFP as a reporter. *Electronic Journal of Biotechnology* ISSN: 0717-3458 DOI: 10.2225/vol 13-issue 5.
- Rashid, R., Moroshoshi, T, Someya, N. and Ikeda, T (2011). Degradation of N-acylhomoserine lactone quorum sensing signaling molecules by potato Root surface-associated *Chryseo bacterium* strains. *Microbes Environment*, 26:144–8.
- Razdan M.K. (2005). Introduction to plant tissue culture. Sciences Publisher, Inc. UK.
- Reifschneider, F.J.B. (1982). Lopes, C.A. Bacterial top and stalk rot of maize in Brazil. *Plant Disease*, 66, 519–520.

- Reis, F.O., Campostrini, E., Sousa, E.F. and Silva, M.G. (2006) Sap flow in papaya plants: Laboratory calibrations and relationships with gas exchanges under field conditions. *Science Horticulture*, 110(3):254–259.
- Rogayah, S., Janna, O.A., Parameswari, N., Pauziah, M. and Umi, K.A.B. (2013). Better rooting produce to enhance survival rate of field grown Malaysian Eksotika papaya transformed with 1-aminocyclopropane-1-carboxylic acid oxidase gene. *International Scholarly Research Notices: Biotechnology, Research Article Hindawi Publishing Corporation*, Vol: 2013, ID 958945.
- Roshidi, A.S. (2010). Papaya disease alert. The Star Online. Retrieved on 15 Nov. 2015 from <http://thestar.com.my/metro/story>.
- Sairam, R.V. and Prakash, C.S. (2005). OBPC Symposium: Maize 2004 and Beyond – Can agricultural biotechnology contribute to food security? *In Vitro Cellular and Developmental Biology Plant*, 41: 424 – 430.
- Sanilkumar, G., and Rathore, K.S. (2001). Transgenic cotton: Factors influencing *Agrobacterium*-mediated transformation and regeneration. *Molecular Breeding*, 8:37-52.
- Sew, Y. S., Johari, S., Maheswary, V. and Abu Bakar, U. K. (2011). Isolation of fruit ripening genes from *Carica Papaya* Var. Eksotika 1 Cdna Libraries. *Journal of Tropical Agriculture and Food Science*, 39: 203-211.
- Si-Nae, H., Poo-Reum, O., Hong-Sig, K., Hwa-Young, H., Jun, C.M., Sang-Kyu, L., Kyung-Hee, K., Yong-Weon, S. and Byung-Moo, L. (2002). Effects of antibiotics on suppression of *Agrobacterium tumefaciens* and plant regeneration from wheat embryo. *Journal Crop Science Biotech*, 10(2):92-98.
- Siritunga, D., Arias-Garcon, D., White, W. and Sayre, R. (2004). Overexpression of hydroxyl nitrile lyase in cassava roots accelerates cyanogenesis and detoxification. *Journal of Plant Biotech*, 2:37-43.
- Smith, E.F. and Townsend, C.O. (1907). A plant-tumor of bacterial origin. *Science*, 25(643):671-673.
- Songul, G., Ekrem, G., Rajvinder, K., Joshua, W., Ling, M., Han-Qi, T. and Peggy, L.G. (2009). Efficient, reproducible *Agrobacterium*-mediated transformation of sorghum using heat treatment of immature embryos. *Plant Cell Reports*, 28:429–444.

- Souza J.R., M.T. and Gonsalves D. (2005). Sequence similarity between the viral cp gene and transgene in transgenic papayas. *Brazilian Journal of Agriculture Research*, 40:479-486.
- Suksa-Ard, P., Kataoka, I., Fujime, Y. and Subhadrabandhu, S. (1999). Requirement of 2,4-D and sucrose for somatic embryogenesis of papaya. *Japanese Journal of Tropical Agriculture*, 43:1-4.
- Suzuki, S. and Nakano, M. (2002). *Agrobacterium*-mediated production of transgenic plants of *Muscariarmeniaceum*. *Plant Cell Reports*, 20:835-841.
- Toth, I. K., Bell, K.S., Holeva, M.C. and Birch, P.R.J. (2003). Soft rot *Erwiniae*: From genes to genomes. *Molecular Plant Pathology*, 4: 17–30.
- Tzfira, T., Jensen, C.S., Vainstein, A. and Altman, A. (1997). Improved rooting ability and root-system performance in transgenic plants. Biology of root formation and development, Basic Life Sciences Series. New York: *Plenum Publishing Co*, pp.181–6.
- Uroz, S. and Heinonsalo, J. (2008). Degradation of N-acyl homoserine lactone quorum sensing signal molecules by forest root-associated fungi. *FEMS Microbiology Ecology*, 65(2):271-278.
- Van Larebeke, N., Genetello, C., Schell, J., Schilperoort, R.A., Hermans, A.K., Hernalsteens, J.P. and Van Montagu, M. (1975). Acquisition of tumour inducing ability by non-oncogenic *Agrobacteria* as a result of plasmid transfer. *Nature*, 225, 742-743.
- Vijay, K. and Sriram, S. (2010). Antioxidant activity of seed extracts of *Annona squamosa* and *Carica papaya*. *Nutrition and Food Science*, 40:403-408.
- Vilasini, P., Latipah, Z. and Salasiah, A. (2000). Induction of somatic embryogenesis and plant regeneration from immature embryos of Eksotika papaya (*Carica papaya* L.). *Journal of Tropical Agriculture and Food Science*, 28:121-126.
- Visser, R.G.F., Jacobsen, E., Hesselting-Meinders, A., Schans, M.J., Withold, B. and Feenstra, W.J. (1989). Transformation of homozygous diploid potato with *Agrobacterium tumefaciens* binary vector system by adventitious shoot regeneration on leaf and stem segments. *Plant Molecular Biology*, 12: 329–337.
- Wang, A. S., Evans, R. A., Altendorf, P. R., Hanten, J. A., Doyle, M. C. and Rosichan, J. L. (2000). A mannose selection system for production of fertile transgenic maize plants from protoplasts. *Plant Cell Reports*, 19: 654-660.

- Wang, M.B. and Waterhouse, P.M. (2000). High-efficiency silencing of a beta glucuronidase gene in rice is correlated with repetitive transgene structure but is independent of DNA methylation. *Plant Molecular Biology*, 43:67-82.
- Wang, M.B., Abbott, D.C. and Waterhouse, P.M. (2000). A single copy of a virus-derived transgene encoding hairpin RNA gives immunity to barley yellow dwarf virus. *Molecular Plant Pathology*, 1:347-356.
- Woodward, B., and Puonti-Kaerlas, J. (2001). Somatic embryogenesis from floral tissue of cassava (*Manihotes culenta* Crantz). *Euphytica*, 120:1-6.
- Wright, M., Dawson, J., Dunder, E., Suttie, J., Reed, J., Kramer, R., Chang, Y. F., Novitzky, R., Wang, H. and Artim-Moore, L. (2001). Efficient biolistic transformation of maize (*Zea mays* L.) and wheat (*Triticum aestivum* L.) using the *Phosphomannose isomerase* gene, *pmi*, as the selectable marker. *Plant Cell Reports*, 20: 429-436.
- Xu J., Wang Y.Z., Yin H.A. and Liu X.J. (2009). Efficient *Agrobacterium tumefaciens*-mediated transformation of *Malus zumi* (Matsumura) Rehd using leaf explant regeneration system. *Electronic Journal of Biotechnology*, 12(1).
- Yu, T.A, Ye, S.D. and Yang, J.S. (2001). Effects of carbenicillin and cefotaxime on callus growth and somatic embryogenesis from adventitious roots of papaya. *Botanical Bulletin of Academia Sinica*, 42: 281-286.
- Yuan, D., Bassie, L., Sabalza, M. and Miralpeix, B. (2011). The potential impact of plant biotechnology on Millenium Development Goals. *Plant Cell Report*, 30: 249 – 265.
- Zai-Song, D., Ming, Z., Yu-Xiang, J., Liang-Bi, L. and Ting-Yun, K. (2006). Efficient *Agrobacterium*-mediated transformation of rice by *Phosphomannose isomerase*/mannose selection. *Plant Molecular Biology Reporter*, 24: 295-303.
- Zhang, P., Potrykus, I. and Puonti-Kaerlas, J. (2000). Efficient production of transgenic cassava using negative and positive selection. *Transgenic Research*, 9: 405-415.