

UNIVERSITI PUTRA MALAYSIA

DEVELOPMENT OF MONOCLONAL ANTIBODIES AGAINST PORCINE BLOOD FOR DETECTION IN FISH-BASED PRODUCTS

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By

RAJA MOHD HAFIDZ BIN RAJA NHARI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

June 2017

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

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June 2017

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Animal blood mainly from porcine and bovine have been used in human food as a binder, gelling agent, emulsifier, coloring agent and iron's supplementary in most meat and fish-based products. However, certain communities including Muslims and Jews are strictly prohibited to consume animal blood in food due to prohibition assigned by each religious and related to health and personal matter. The addition of porcine plasma in fish surimi has been highlighted by the local authority in Malaysia. To date, a specific method to detect porcine plasma in food is unavailable. In this study, we have developed monoclonal antibodies (mAbs) against heated soluble proteins (HSPs) of porcine blood using fusion technology for detection of porcine plasma in fish surimi. Specificity of mAbs against blood, non-blood (meat and nonmeat) and commercial animal plasma proteins from different species were determined using indirect non-competitive ELISA. Antigenic components of porcine blood were determined using Western blot. The sensitivity of ELISA has been determined to analyze fish surimi that spiked with porcine plasma. After several screening of hybridoma was made, 27 hybridomas produced mAbs were selected. Based on that, fifteen mAbs are specific to raw and heated porcine blood, one mAb only specific to raw porcine blood and another 11 mAbs are cross-reacted with other animal blood. The fifteen mAbs specific to porcine blood are also not cross-reacted with meat and non-meat proteins. Based on specificity to animal plasma, twelve mAbs from 15 mAbs are specific to porcine plasma while other 3 mAbs are cross-reacted to chicken plasma. Western blot showed that 2 mAbs bind 60 kDa, 8 mAbs bind 85 kDa and 2 mAbs bind 250 kDa of the antigenic protein of porcine blood. The mAb labeled as B4E1 was selected to be used for detection of porcine plasma. The developed ELISA has a limit of detection (LOD) and limit of quantification (LOQ) of 0.2 μ g/g and 1 μ g/g, respectively for a standard solution of porcine plasma. The intra- and inter-assays of ELISA with coefficients of variation (CVs) less than 20% were able to detect at least 0.25% (w/w) porcine plasma in fish surimi. In conclusion, this study has successfully obtained the hybridoma-producing mAbs that are specific to porcine blood and porcine plasma. This study also has developed indirect non-competitive ELISA for

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detection of porcine plasma in laboratory model fish ball and fish surimi products using mAb B4E1 with LOD and LOQ of 0.2 μ g/g and 1 μ g/g, respectively with the sensitivity of the ELISA is 0.25% (w/w) porcine plasma in fish surimi.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

PENGHASILAN ANTIBODI MONOKLON TERHADAP DARAH KHINZIR BAGI PENGESANANNYA DI DALAM PRODUK BERASASKAN IKAN

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Darah haiwan terutamanya daripada khinzir dan lembu telah digunakan dalam makanan manusia sebagai pengikat, agen gel, pengemulsi, pewarna dan penambah zat besi di dalam kebanyakan produk berasaskan daging dan ikan. Namun begitu, sesetengah masyarakat termasuk orang Islam dan Yahudi adalah dilarang sama sekali menggunakan darah haiwan dalam makanan kerana larangan yang ditetapkan dalam agama masing-masing dan berkaitan dengan hal kesihatan dan peribadi. Penambahan plasma khinzir dalam surimi ikan telah diketengahkan oleh pihak berkuasa tempatan di Malaysia. Setakat ini, kaedah khusus bagi mengesan plasma khinzir dalam makanan masih belum diwujudkan. Dalam kajian ini, kami telah membangunkan antibodi monoklon (mAb) terhadap protein darah khinzir terlarut yang dipanaskan (HSPs) menggunakan teknologi lakuran untuk mengesan plasma khinzir dalam surimi ikan. Kespesifikan mAb terhadap protein darah, protein bukan darah (daging dan bukan daging) dan protein plasma haiwan komersil dari spesies yang berbeza telah ditentukan dengan menggunakan ELISA bukan kompetitif tidak langsung. Komponen antigen darah khinzir telah ditentukan menggunakan Western blot. Kepekaan ELISA telah ditentukan untuk menganalisis surimi ikan yang ditambahkan dengan plasma khinzir. Selepas beberapa saringan hibridoma dibuat, 27 hibridoma yang menghasilkan mAb telah dipilih. Berdasarkan itu, lima belas mAbs spesifik untuk darah khinzir mentah dan dipanaskan, satu mAb hanya spesifik untuk darah khinzir mentah dan 11 mAb yang lain bersilang tindak balas dengan darah haiwan lain. Lima belas mAb yang spesifik untuk darah khinzir didapati tidak bersilang tindak balas dengan protein daging dan protein bukan daging. Berdasarkan kespesifikan kepada plasma haiwan, dua belas mAb daripada 15 mAb spesifik untuk plasma khinzir sementara 3 mAb yang lain bersilang tindak balas kepada plasma ayam. Western blot menunjukkan bahawa 2 mAb mengikat 60 kDa, 8 mAb mengikat 85 kDa dan 2 mAb mengikat 250 kDa protein antigen darah khinzir. MAb berlabel B4E1 telah dipilih untuk digunakan bagi mengesan plasma khinzir. ELISA yang dibangunkan mempunyai had pengesanan (LOD) dan had kuantifikasi (LOQ) masing-masing 0.2 µg/g dan 1 µg/g untuk larutan piawai plasma khinzir. Asai intra dan inter ELISA

dengan pekali variasi (CV) adalah kurang daripada 20% dan ia boleh mengesan plasma khinzir sekurang-kurangnya 0.25% (w/w) plasma khinzir dalam surimi ikan. Kesimpulannya, kajian ini telah berjaya meperoleh hibridoma yang menghasilkan yang spesifik kepada darah dan plasma khinzir. Kajian ini juga telah dapat membangunkan ELISA bukan kompetitif tidak langsung untuk pengesanan plasma khinzir dalam model laboratori bebola ikan dan produk surimi ikan menggunakan mAb B4E1 dengan had pengesanan (LOD) dan had kuantifikasi (LOQ) masingmasing 0.2 μ g/g dan 1 μ g/g dengan sensitiviti ELISA iaitu 0.25% (w/w) plasma khinzir dalam surimi ikan.



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LIST OF ABBREVIATIONS

| ABTS | 2,2-azino-bis-3-ethylbenzthiazoline- 6-sulfonic acid |
|----------------------------------|--|
| AP | Autoclaved porcine blood |
| APP | Autoclaved porcine plasma |
| BP | Boiled porcine blood |
| BPP | Boiled porcine plasma |
| C_{H1} , C_{H2} and C_{H3} | Constant domains (heavy chain) |
| <u>OIE</u> | |
| CIF | Complex Initiation Factor |
| CPL | ClonePix FL |
| CL | Constant domain (light chain) |
| | Coefficient of variation |
| | Carbon dioxide |
| DMSO | |
| ELISA | Enzyme-linked immunosorbent assay |
| FBS | Fetal bovine serum |
| FCA | Freund's complete adjuvant |
| FIA | Freund's incomplete adjuvant |
| FIIC | Fluorescein isothiocyanate |
| FU | Fluorescence units |
| g | Gravitational force |
| h | Hour |
| HAT | Hypoxanthine-aminopterin-thymidine |
| HPRT | Hypoxanthine-phosphoribosyl-transferase |
| HGPRT | Hypoxanthine-guanine-phosphoribosyl-transferase |
| HRP | Horseradish peroxidase |
| HSPs | Heated soluble proteins |
| HT | Hypoxanthine thymidine |
| IgA | Immunoglobulin A |
| IgD | Immunoglobulin D |
| IgE | Immunoglobulin E |
| IgG | Immunoglobulin G |
| IgM | Immunoglobulin M |
| JAKIM | Jabatan Kemajuan Islam Malaysia |
| kDa | Kilo Dalton |
| LOD | Limit of detection |
| LOQ | Limit of quantification |
| LC-MS/MS | Liquid chromatography tandem mass spectrometry |
| mAb | Monoclonal antibody |
| min | Minute |
| mL | Milliliter |
| mM | Millimolar |
| MSG | Monosodium glutamate |
| MWCO | Molecular weight cut-off |
| OD _{405nm} | Optical reading at 405 nm |
| PBS | Phosphate buffered saline |
| PEG | Polyethylene glycol |
| | |

C

| RNA | Ribonucleic acid |
|----------------|---|
| RP | Raw porcine blood |
| rpm | Revolution per minute |
| RPMI | Roswell Park Memorial Institute |
| RPP | Raw porcine plasma |
| RT | Room temperature |
| SD | Standard deviation |
| SDS-PAGE | Sodium-dodecyl sulfate-polyacrylamide gel |
| | electrophoresis |
| TBS | Tris buffer saline |
| TEMED | <i>N</i> , <i>N</i> , <i>N</i> , <i>N</i> -tetra-methyl-ethylenediamine |
| TSE | Transmissible spongiform encephalopathy |
| TTBS | Tris buffer saline with Tween-20 |
| V _H | Variable domain (heavy chain) |
| VL | Variable domain (light chain) |
| v/v | Volume per volume |
| w/w | Weight per weight |
| μm | Micrometer – |
| μg | Microgram |
| | |

C

CHAPTER 1

INTRODUCTION

Animal blood has been utilized in human food, animal feed and in the field of the laboratory, medical, industrial and fertilizer since several years ago. Currently, advance technology in animal blood collection and processing techniques have led to a production of numerous blood protein ingredients that could give benefits to the environment, nutrition and economic derived from the maximal utilization of animal blood. The blood can be separated into two parts, a liquid and cellular portion. Plasma, the liquid portion of blood is mostly utilized in the food industry because it is tasteless and contains no dark color associated with the red blood cells. Meanwhile, the cellular portion of blood is not extensively used as compared to the plasma due to its heme component, which possesses an undesirable color, odor and gives a metallic taste to the end-product.

The production of blood products involved the processing of blood into different types of products which can deliver various functional properties. For instance, spray dried porcine plasma can be used as proteinase inhibitor to avoid irreversible proteolytic degradation of myofibrillar proteins in surimi caused by endogenous proteinases that will contribute to gel weakening of surimi and surimi-based foods (Ofori and Hsieh, 2012). Bekhit et al. (2014) reviewed several studies about the ability of porcine plasma proteins as proteinase inhibitor in the manufacturing of surimi. In addition, the porcine plasma can be used as an egg replacer in bakery products as foaming, jellification and solubility agent. The plasma and albumin are used as solubility and jellification agent in yogurt (Álvarez et al., 2012). Besides, fibrinogen and plasma from porcine blood can be added as an emulsifier in cakes and pastries. Stabilized haemoglobin is commonly used in meat products as binding and colouring agents. Decoloured globin obtained from haemoglobin can be used as solubility agent in biscuits while globulins can be used as emulsifier, solubility and jellification agent in meat products (Álvarez et al., 2012). Thus, the increasing demand of porcine plasma will occur as much as the current requirement of bovine plasma in the food industry.

However, the Muslims and Jewish are prohibited to eat food containing blood as a result of dietary restrictions obligatory of their religion (Mukhtar, A., and Mohsin Butt, 2012) and because of the commands preserved in the Halal and Kosher dietary laws, respectively. There are also people that need to avoid blood-containing food as a matter of preference (Ofori and Hsieh, 2014). Vegetarians avoid consuming products of animal origin for their ethical and religious reasons.

The prion proteins or infectious proteins are extremely resistant to heat, ultraviolet light, ionizing radiation, normal sterilization processes and common disinfectants that normally destroy viruses and bacteria (Huong et al., 2014). Thus, humans are potentially exposed to these infectious agents (abnormal prion proteins) through the various consumptions of animal blood added intentionally (adulteration) or

unintentionally in food because the process involved in the manufacturing of these blood-containing materials does not inactivate the infectious agent as a result of their resistant nature. Even though no evidences of natural case of transmissible spongiform encephalopathy (TSE) in non-ruminants including porcine (Greenlee et al., 2016), development of scientific method to detect porcine blood derivative in food is imperative because although the religious law to ban non-Halal or non-Kosher blood has been stated, the addition of porcine blood in food still occurred. This is due to the lacking in existing methods to help to enforce the laws and the manufacturer may adulterate the expensive meats with a high amount of blood proteins to fraud the protein content in their food products.

Until now, there are no scientific methods that have been developed to determine the presence of porcine blood especially porcine plasma in food. Thus, there is a need to develop a new scientific method which is able to detect porcine plasma in food especially in a processed form that undergoes the heat treatment. In addition, the method should be based on the detection of heated porcine blood as well as plasma and be able to discriminate between prohibited tissue (blood) of porcine species and allowed tissue (halal meats), as well as discriminate between blood proteins and proteins from other sources (soy protein, whey, potato extracts, egg albumin and vice versa) that commonly present in food.

The general objective of this study is to produce hybridoma that secretes monoclonal antibody (mAb) against porcine blood; where the mAb(s) will react to the heated protein of porcine blood for qualitative detection of porcine plasma in fish-based products especially surimi. The use of mAb will offer an extra advantage since the mAb itself is species-specific and could differentiate specific animal blood from other blood species, other tissues and common food proteins. The specific objectives of this study comprised of:

- a) to produce hybridoma through the fusion of immunized splenocytes and myeloma cells (P3X63Ag8.653, ATCC[®] CRL-1580[™])
- b) to characterize the mAb(s) that are specific to porcine blood especially porcine plasma
- c) to develop indirect non-competitive ELISA for detection of porcine plasma in a laboratory model of fish ball and commercial fish surimi

REFERENCES

- Álvarez, C., García, V., Rendueles, M., and Díaz, M. (2012). Functional properties of isolated porcine blood proteins modified by Maillard's reaction. *Food Hydrocolloids*. 28(2): 267–274.
- Arfat, Y. and Benjakul, S. (2012). Gelling characteristics of surimi from yellow stripe trevally (*Selaroides leptolepis*). *International Aquatic Research*. 4(1): 1-13
- Bah, C. S. F., Bekhit, A. E. D. A., Carne, A., and Mcconnell, M. A. (2013). Slaughterhouse blood: An emerging source of bioactive compounds. *Comprehensive Reviews in Food Science and Food Safety*. 12(3): 314-331
- Bekhit, A. A., Hopkins, D. L., Geesink, G., Bekhit, A. A. and Franks, P. (2014). Exogenous Proteases for Meat Tenderization. *Critical Reviews in Food Science* and Nutrition. 54(8): 1012–1031.
- Cai, Q. F., Wang, X. C., Liu, G. M., Zhang, L., Ruan, M. M., Liu, Y. and Cao, M. J. (2013). Development of a monoclonal antibody-based competitive enzyme linked-immunosorbent assay (c-ELISA) for quantification of silver carp parvalbumin. *Food Control*. 29(1): 241–247.
- Carbonetti, S., Oliver, B. G., Vigdorovich, V., Dambrauskas, N., Sack, B., Bergl, E. and Noah Sather, D. (2017). A method for the isolation and characterization of functional murine monoclonal antibodies by single B cell cloning. *Journal of Immunological Methods*. http://doi.org/10.1016/j.jim.2017.05.010
- Carrera, E., Terni, M., Montero, A., García, T., González, I. and Martín, R. (2013). ELISA-based detection of mislabeled albacore (*Thunnus alalunga*) fresh and frozen fish fillets. *Food and Agricultural Immunology*. 25(4): 569–577.
- Chanarat, S., Benjakul, S. and H-Kittikun, A. (2012). Comparative study on protein cross-linking and gel enhancing effect of microbial transglutaminase on surimi from different fish. *Journal of the Science of Food and Agriculture*. 92(4):844–852.
- Costa, J., Mafra, I., Carrapatoso, I. and Oliveira, M. B. P. P. (2016). Hazelnut Allergens: Molecular Characterization, Detection, and Clinical Relevance. *Critical Reviews in Food Science and Nutrition*. 56(15): 2579–2605.
- Cwykiel, J. and Siemionow, M. Z. (2015). Cellular Therapy Models: Ex Vivo Chimera Model by Cell Fusion. In Plastic and Reconstructive Surgery (pp. 593-603). London: Springer-Verlag London.
- Davis, J. M., Pennington, J. E., Kubler, A.-M. and Conscience, J.-F. (1982). A simple, single-step technique for selecting and cloning hybridomas for the production of monoclonal antibodies. *Journal of Immunological Methods*. 50(2): 161-171.
- de Silva, R., Dasanayake, W. M. D. K., Wickramasinhe, G. D., Karunatilake, C., Weerasinghe, N., Gunasekera, P., & Malavige, G. N. (2017). Sensitization to bovine serum albumin as a possible cause of allergic reactions to vaccines. *Vaccine*. 35(11): 1494–1500.
- Dharshanan, S., Chong, H., Cheah, S. H. and Zamrod, Z. (2014). Stable expression of H1C2 monoclonal antibody in NS0 and CHO cells using pFUSE and UCOE expression system. *Cytotechnology*. 66(4): 625–633.
- Forsthoefel, D. J., Waters, F. A., & Newmark, P. A. (2014). Generation of cell typespecific monoclonal antibodies for the planarian and optimization of sample processing for immunolabeling. *BMC Developmental Biology*. 14(1): 1-22.
- García-Bermejo, I. and de Ory, F. (2017). Rapid diagnosis in serology. *Enfermedades Infecciosas Y Microbiologia Clinica (English Ed.).* 35(4): 246–254.

- Ghanvat, M. D. (2013). Monoclonal Antibodies as Targeted Drug Delivery System: A Review. International Journal of Chemical and Life Sciences. 2(2): 1086– 1090.
- Greenlee, J. J., Kunkle, R. A., Smith, J. D. and Greenlee, M. H. W. (2016). Scrapie in Swine: a Diagnostic Challenge. *Food Safety*. 4(4): 110–114.
- Gurevich, V., Kotharu, P., McCann, K. and Bertolini, J. (2017): Improvement of ELISA procedures through simultaneous addition of antigen and detection antibody and elimination of washing steps. *Journal of Immunoassay and Immunochemistry. http://dx.doi.org/10.1080/15321819.2017.1331171*
- Hofgaard, P. O., Jodal, H. C., Bommert, K., Huard, B., Caers, J., Carlsen, H. and Bogen, B. (2012). A Novel Mouse Model for Multiple Myeloma (MOPC315.BM) That Allows Noninvasive Spatiotemporal Detection of Osteolytic Disease. *PLoS ONE*, 7(12): e51892
- Hornbeck, P., Fleisher, T. A., and Papadopoulos, N. M. (2017). Isotype Determination of Antibodies. *Current Protocols in Immunology*, 2.2.1-2.2.7.
- Hsieh, Y. H. P., Ofori, J. A., Rao, Q. and Bridgeman, C. R. (2007). Monoclonal antibodies specific to thermostable proteins in animal blood. *Journal of Agricultural and Food Chemistry*. 55(16): 6720-6725.
- Huong, V. T. L., Hoa, N. T., Horby, P., Bryant, J. E., Kinh, N. Van, Toan, T. K., and Wertheim, H. F. L. (2014). Raw pig blood consumption and potential risk for *Streptococcus suis* infection, Vietnam. *Emerging Infectious Diseases*. 20(11): 1895-1898.
- Jayathilakan, K., Sultana, K., Radhakrishna, K., and Bawa, A. S. (2012). Utilization of byproducts and waste materials from meat, poultry and fish processing industries. A review. *Journal of Food Science and Technology*. 49(3): 278-293.
- Jeong, J. Y., Nam, J. S., Kim, J. M., Jeong, H. J., Kim, K. W. and Lee, H. J. (2013). Comparison of plasma proteome expression between the young and mature adult pigs. *Reproductive & Developmental Biology*. 37(4): 247-253.
- Keiji, F. (2015). Food safety in globalized world. Bulletin of the World Health Organization. 93:212.
- Kim, S., Jin, S., and Choi, J. (2017). Effects of the addition of blood plasma proteins on physico-chemical properties of emulsion-type pork sausage during cold storage. *Journal of the Science of Food and Agriculture*. http://doi.org/10.1002/jsfa.8315
- Klinman, N. R. (1972). The mechanism of antigenic stimulation of primary and secondary clonal precursor cells. *The Journal of Experimental Medicine*. 136(2): 241-260.
- Köhler, G. and Milstein, C. (1975). Continuous cultures of fused cells secreting antibody of predefined specificity. *Nature*. 256: 495-497.
- Kreuz, G., Zagon, J., Broll, H., Bernhardt, C., Linke, B. and Lampen, A. (2012). Immunological detection of osteocalcin in meat and bone meal: a novel heat stable marker for the investigation of illegal feed adulteration. *Food Additives & Contaminant: Part A.* 29(5): 716-726.
- Li, J., Hu, F., Chen, S., Luo, P., He, Z., Wang, W. and Li, C. (2017). Characterization of novel Omp31 antigenic epitopes of Brucella melitensis by monoclonal antibodies. *BMC Microbiology*. 17(1): 1-8.
- Liu, B., Teng, D., Wang, X., Yang, Y. and Wang, J. (2012). Expression of the soybean allergenic protein P34 in Escherichia coli and its indirect ELISA detection method. *Applied Microbiology and Biotechnology*, 94(5), 1337-1345.

- Liu, J. K. H. (2014). The history of monoclonal antibody development Progress, remaining challenges and future innovations. *Annals of Medicine and Surgery*. 3(4): 113-116.
- Liu, W., Stevenson, C. D., and Lanier, T. C. (2013). Rapid heating of Alaska pollock and chicken breast myofibrillar proteins as affecting gel rheological properties. *Journal of Food Science*. 78(7): 971-977.
- Mahmuda, A., Bande, F., Jameel, K., Al-Zihiry, K., Abdulhaleem, N., Majid, R. A. Unyah, Z. (2017). Monoclonal antibodies: A review of therapeutic applications and future prospects. *Pharm Res Tropical Journal of Pharmaceutical Research Journal Citation Reports Science Edition*, 16(163): 713–713.
- Moreno, H. M., Herranz, B., Pérez-Mateos, M., Sánchez-Alonso, I. and Borderías, J. A. (2016). New Alternatives in Seafood Restructured Products. *Critical Reviews* in Food Science and Nutrition. 56(2): 237–248.
- Mukhtar, A., and Mohsin Butt, M. (2012). Intention to choose *Halal* products: the role of religiosity. *Journal of Islamic Marketing*. 3(2): 108–120.
- Nakyinsige, K., Man, Y. B. C. and Sazili, A. Q. (2012). Halal authenticity issues in meat and meat products. *Meat Science*. 91(3): 207–214.
- Ofori, J. A. and Hsieh, Y.-H. P. (2012). The use of blood and derived products as food additives. In El-Samragy, Y. *Food Additive* (pp. 229-256). Rijeka: InTech.
- Ofori, J. A. and Hsieh, Y.-H. P. (2014). Issues Related to the Use of Blood in Food and Animal Feed. *Critical Reviews in Food Science and Nutrition*. 54(5): 687-697.
- Ofori, J. A. and Hsieh, Y. H. P. (2015). Characterization of a 60-kDa Thermally Stable Antigenic Protein as a Marker for the Immunodetection of Bovine Plasma-Derived Food Ingredients. *Journal of Food Science*. 80(8): C1654–C1660.
- Olsen, R. L., Toppe, J. and Karunasagar, I. (2014). Challenges and realistic opportunities in the use of by-products from processing of fish and shellfish. *Trends in Food Science and Technology*. 36(2): 144–151.
- Ortea, I., O'Connor, G. and Maquet, A. (2016). Review on proteomics for food authentication. *Journal of Proteomics*. 147: 212–225
- Peng, J., Meng, X., Deng, X., Zhu, J., Kuang, H. and Xu, C. (2012). Development of a monoclonal antibody-based sandwich ELISA for the detection of ovalbumin in foods. *Food and Agricultural Immunology*. 25(1): 1-8.
- Rosa, E. A., Lanza, S. R., Zanetti, C. R. and Pinto, A. R. (2012). Immunophenotyping of classic murine myeloma cell lines used for monoclonal antibody production. *Hybridoma*. 31(1): 1-6.
- Soares, S., Amaral, J. S., Oliveira, M. B. P. P. and Mafra, I. (2014). Quantitative detection of soybean in meat products by a TaqMan real-time PCR assay. *Meat Science*. 98(1): 41–46.
- Toldrá, F., Aristoy, M. C., Mora, L., and Reig, M. (2012). Innovations in valueaddition of edible meat by-products. *Meat Science*. 92(3): 290-296.
- Tukiran, N. A., Ismail, A., Mustafa, S. and Hamid, M. (2015). Enzyme immunoassay for the detection of porcine gelatine in edible bird's nests. *Food Additives & Contaminants: Part A*. 32(7): 1023-1028.
- Uchigashima, M., Watanabe, E., Ito, S., Iwasa, S. and Miyake, S. (2012). Development of immunoassay based on monoclonal antibody reacted with the neonicotinoid insecticides clothianidin and dinotefuran. *Sensors*. 12(11): 15858-15872.

- Wang, H., Li, G., Wu, Y., Yuan, F. and Chen, Y. (2014). Development of an indirect competitive immunoassay for walnut protein component in food. *Food Chemistry*. 147: 106–110.
- Wang, Z., Jiyuan, Y., Su, C., Xinyuan, Q., Lijie, T., & Yijing, L. (2015). Development of an antigen capture enzyme-linked immunosorbent assay for virus detection based on porcine epidemic diarrhea virus monoclonal antibodies. *Viral Immunol*. 28(3): 184-189.
- Xi, J., Shi, Q. Q. and Lu, Q. Y. (2016). Development of an Indirect Competitive ELISA Kit for the Rapid Detection of Benzopyrene Residues. *Food Analytical Methods*. 9(4): 966–973.
- Xue, H., Xing, Y., Yin, Y., Zhang, T., Zhang, B., Zhang, Y., Song, P., Tian, X., Xu, Y., Wang, P., Meng, M. and Xi, R. (2012). Application of an enzyme immunoassay for the quantitative determination of azo dye (Orange II) in food products. *Food Additives & Contaminants: Part A*. 29(12): 1840-1848.
- Yagami, H., Kato, H., Tsumoto, K. and Tomita, M. (2013). Monoclonal antibodies based on hybridoma technology. *Pharmaceutical Patent Analyst*. 2(2): 249–263.
- Yan, X., Zhao, Y., Zhang, Y. and Qu, H. (2017). Monoclonal antibodies and immunoassay for medical plant-derived natural products: A review. *Molecules*, 22(3). http://doi.org/10.3390/molecules22030355
- Yu, T. Y., Morton, J. D., Clerens, S. and Dyer, J. M. (2017). Cooking-Induced Protein Modifications in Meat. *Comprehensive Reviews in Food Science and Food Safety*. 16(1): 141–159.
- Zhang, C. (2012). Hybridoma Technology for the Generation of Monoclonal Antibodies. In *Methods in molecular biology* (pp. 117-135). New York: Humana Press.
- Zhang, M., Liu, S., Zhuang, H. and Hu, Y. (2012). Determination of dimethyl phthalate in environment water samples by a highly sensitive indirect competitive ELISA. *Applied Biochemistry and Biotechnology*. 166(2): 436-445.
- Zhou, C., Wang, H., Chen, Y. and Chen, C. (2012). Effect of L-cysteine and lactose on color stability of porcine red blood cell during freeze-drying and powder storage. *Food Science and Biotechnology*. 21(3): 669–674.
- Zhou, J., Zhao, S., Zhang, J., Zhang, L., Cai, Y., & Zhou, L. (2013). An indirect competitive enzyme-linked immunosorbent assay for bisphenol-A based on the synthesis of a poly-L-lysine-hapten conjugate as a coating antigen. *Analytical Methods*. 5(6): 1570-1576