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TURFGRASS VARIETAL IMPROVEMENT FOR SHADE AND DROUGHT TOLERANCE USING GAMMA RAY IRRADIATION

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MOHD ABDUL HALIM BIN BAHARUN AZAHAR

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Master of Science

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in partial fulfilment of the requirement for the Master of Science

TURFGRASS VARIETAL IMPROVEMENT FOR SHADE AND DROUGHT TOLERANCE USING GAMMA RAY IRRADIATION

By

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Turfgrass breeding aims to improve the characteristics of plants so that they become more desirable agronomically and economically. Alternative methods' using mutagenic treatment is a relatively quick method for improvement of turfgrass. Gamma ray irradiation can be used to improve turfgrass phenotype and enhance tolerance to environmental stress. A series of experiments were conducted to examine the response of turfgrass species to gamma ray irradiation, either in their phenotypic and genotypic characteristics, and to study turfgrass mutant lines under different shade and drought stress conditions. The mutant lines selected for evaluation in these studies were based on desirable characteristics for performance under stress.

Eight gamma ray dosages (0, 20, 40, 60, 80, 100, 150 and 200 Gy) were applied to *Axonopus compressus, Zoysia japonica* and *Cynodon dactylon* at the Gamma Cell Laboratory, Malaysian Institute of Nuclear Technology Research (MINT), Bangi, Selangor to identify the optimum dosage for turfgrass mutation. Optimum dosage was needed induce maximum mutation and to increase mutation rate. The optimum dosage was calculated based on 50% radiosensitivity tests on survival rate and plant height. The values 50% of radiosensitivity tests (LD₅₀) were determined to be 52, 76 and 90 Gy for *A.compressus, Z.japonica and C. dactylon*, respectively. The turfgrasses were radiated using the optimum dosage of gamma ray to produce numerous mutants. A total of 1500 stolons of each species were radiated and planted in biodegradable seed tray. In order to ensure the inheritance of these characteristics, all mutants were isolated using the cutting back technique.

Most of the mutants had dwarf and semi-dwarf characters. Gamma ray irradiation significantly altered the morphological parameters of turfgrass. The results showed that 2.4%, 2.6% and 1.5% rate of mutation occurred for *A. compressus*, *Z. japonica* and *C. dactylon*, respectively after exposing to the LD⁵⁰ dosages. Thirty six lines from *A. compressus* were recorded as mutants with five (A26-4-1, A61-1-1, A46-2-1, A91-3-5, A13-2-5) showing high potential for further study. Thirty nine lines from *Z. japonica* were recorded as mutant with five (Z131-3-1, Z36-3-1, Z13-1-2, Z12-2-



1, Z2-2-1) of them showing high potential for further study. Twenty two lines from *C. dactylon* were recorded as mutants with five (C43-4-1, C85-1-2, C59-2-2, C41-4-1, C5-3-1) showing high potential for further study, while six (C43-4-1, C42-4-1, C37-5-1, C83-3-2, C95-2-2, C13-3-3) of them were reselected for the shade tolerance study.

In the drought tolerance study, six most tolerant mutant lines (A48-3-5, A64-2-2, A62-3-1, A84-1-1, A26-4-1, A46-2-1) and A0 were subjected to five field capacity treatments of -20, -30, -33 (control), -40 and -50 J/kg and were assessed for visual quality and growth parameters. Shoot and root dry weights were also determined. *A. compressus* showed low quality performance under extreme drought conditions and many had died. A84-1-1 performed the best under drought conditions as it could withstand up to -50 J/kg field capacity, and this was followed by A26-4-1 and A64-2-2. In the shade tolerance study treatments were applied by exposing the grass to three, six, nine or twelve hour's of full sunlight per day. Generally, turfgrass showed slow growth and low quality when exposed to less than 3 hours of sunlight. The quality of *C. dactylon* was much better under long duration of full sunlight. Mutant line C43-4-1 performed the best under shade with its outstanding quality in terms of colour, density and uniformity. Durations with a minimum of at least 6 hours sunlight showed good responses. Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PEMBIAKBAKAAN RUMPUT TURF TERHADAP KETAHANAN TEDUHAN DAN KEMARAU DENGAN MENGGUNAKAN SINARAN GAMMA

Oleh

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Pembiakbakaan rumput bertujuan untuk menambahbaikkan sifat-sifatnya supaya lebih bernilai dari segi agronomi dan ekonomi. Cara alternatif ialah dengan menggunakan kaedah mutagenik yang mana lebih cepat untuk menambahbaikkan rumput. Sinaran gamma digunakan untuk menambahbaikkan fenotip dan ketahanan rumput terhadap tekanan persekitaran. Satu eksperimen bersiri telah dijalankan untuk menguji tindakan rumput terhadap sinaran gamma, samada terhadap sifat-sifat fenotip dan untuk menilai rumput mutan di bawah tekanan naungan dan kemarau. Mutan dipilih berdasarkan sifat-sifat yang diingini dan prestasinya di bawah tekanan.

Lapan dos sinaran gamma (0, 20, 40, 60, 80, 100, 150 dan 200 Gy) telah didedahkan kepada *Axonopus compressus, Zoysia japonica* dan *Cynodon dactylon* di Gamma Cell Laboratory, Malaysian Institute of Nuclear Technology Research (MINT), Bangi, Selangor untuk mengenalpasti dos terbaik mutasi rumput. Dos terbaik perlu digunakan supaya memaksimumumkan mutasi dan meningkatkn kadar mutasi. Dos terbaik dikira dengan berdasarkan 50% ujian radiosensitif ke atas kadar hidup dan ketinggian rumput. Nilai dos terbaik iaitu LD50 untuk *Axonopus compressus, Zoysia japonica* dan *Cynodon dactylon* ialah 52, 76 dan 90 Gy masing-masing. Rumput didedahkan dengan dos sinaran gamma terbaik untuk menghasilkan pelbagai mutan. Sejumlah 1500 stolon untuk setiap spesis diradiasikan dan ditanam dalam bekas semaian. Untuk memastikan sifat-sifat yang diingini, mutan diasingkan dengan menggunakan teknik cutting back.

Kebanyakkan mutan mempunyai sifat-sifat kerdil dan separuh-kerdil. Sinaran gamma secara signifikan telah mengubah sifat-sifat morfologi rumput. Keputusan menunjukkan sebanyak 2.4%, 2.6% and 1.5% kadar mutasi untuk *A. compressus, Z. japonica* dan *C. dactylon* masing-masing selepas didedahkan pada dos LD50. Sejumlah 36 mutan dari *A. compressus* direkodkan dan lima (A26-4-1, A61-1-1, A46-2-1, A91-3-5, A13-2-5) menunjukan potensi yang besar untuk kajian seterusnya. Sejumlah 39 mutan dari *Z. japonica* telah direkodkan dan lima (Z131-3-1, Z36-3-1, Z13-1-2, Z12-2-1, Z2-2-1) menunjukan potensi yang besar untuk kajian seterusnya. Manakala 22 mutan dari *C. dactylon* telah direkodkan dan lima (C43-4-

1, C85-1-2, C59-2-2, C41-4-1, C5-3-1) daripadanya telah menunjukan potensi yang besar untuk kajian seterusnya, sementara enam (C43-4-1, C42-4-1, C37-5-1, C83-3-2, C95-2-2, C13-3-3) daripadanya dipilih untuk kajian naungan.

Dalam kajian kesan kemarau, enam mutan (A48-3-5, A64-2-2, A62-3-1, A84-1-1, A26-4-1, A46-2-1) yang mempunyai ketahanan paling tinggi dan A0 telah dipilih dan dirawat dengan lima kapasiti lapangan iaitu -20, -30, -33 (sebagai kawalan), -40 and -50 J/kg dan dinilai dari segi kualiti visual dan pertumbuhan. Berat kering pucuk dan akar juga dinilai. *A. compressus* (A0) telah menunjukkan kualiti yang rendah di bawah tahap kemarau yang tinggi dan mengalami kematian. A84-1-1 telah menunjukkan prestasi yang terbaik di bawah keadaan kemarau dan boleh bertahan sehingga -50 J/kg kapasiti lapangan, diikuti oleh A26-4-1 dan A64-2-2. Di samping itu, kajian kesan naungan telah dilakukan dengan mendedahkan rumput kepada tiga, enam, sembilan dan dua belas jam cahaya matahari untuk setiap hari. Umumnya, pertumbuhan rumput menjadi perlahan dan kualiti juga berkurang apabila didedahkan pada 3 jam cahaya matahari. Mutan C43-4-1 mempunyai prestasi terbaik di bawah naungan dengan menunjukkan warna, kepadatan dan kesamaan yang baik.

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I certify that a Thesis Examination Committee has met on August 29, 2014 to conduct the final examination of Mohd Abdul Halim Bin Baharun Azahar on his thesis entitled "**Turfgrass varietal improvement for shade and drought tolerance using gamma ray irradiation**" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

AsA	Ascorbic Acid
APX	Ascorbate Peroxidase
ANOVA	Analysis of Variance
CIRP	Christmas Island Rock Phosphate
CAT	Catalase
CRD	Completely Randomized Design
Gy	Gray
GSH	Glutathione
GR	Glutathione Reductase
GMO	Genetically Modified Organism.
HSD	Tukey's Studentized Range
LD	Lethality dosage
Mm	Milimeter
MINT	Malaysian Institute of Nuclear Technology Research
M_1V_1	First Cutting Back
M 1 V 3	Third Cutting Back
ROS	Reactive Oxygen Species
SAS	Statistical Analyses System
SOD	Superoxide Dismutase

CHAPTER 1

INTRODUCTION

Turfgrass is a vegetative ground cover composed of closed cut thickly growing, uniform, inter-twined stems and leaves of plants which form a kind of mat, sward or sod (Beard, 1973). Turfgrass is a monocot plant that belongs to family Poaceae and only 50 grasses have been considered as turfgrasses (Christians, 2007). Each species of turfgrass has different characteristics such as shade tolerance, leaf width and colour, fertility requirements, disease resistance, growth rate, close mowing tolerance, cold hardiness, heat and drought tolerance, uniformity and ability to tolerate traffic and establishment rate (Emmons, 1984). There are about 17 species of turf grasses being used in Malaysia. However, only carpet grass (*Axonopus compressus*), bermudagrass (*Cynodon dactylon*), zoysiagrass (*Zoysia japonica*) and seashore paspalum (*Paspalum vaginatum*) are commercialized in Malaysia. These turfgrasses still have weakness like coarse texture and poor tolerance to environmental stresses.

Issues of enhanced turfgrass improvement depend primarily on increasing new cultivars and their maintenance. In Malaysia, little research has been done so far on turfgrasses. Turfgrass requires genetic variation in useful traits for turfgrass improvement and breeding. However, the desired variation is lacking especially for warm season grasses. There are several potential turfgrasses in Malaysia but due to lack of certain characteristics, these turfgrasses are not top performers. Given this challenge, existing and appropriate new technologies need to be integrated into turfgrass research, in order to focus on problems related to improving turfgrass breeding and development of new cultivars.

Turfgrass industry is still in the process of development in Malaysia. Indeed, there is a lack of turfgrass varieties and cultivars. Currently, there are several ways to obtain genetic variation. One of them is hybridization. However, hybridization is difficult in turfgrass since the florets are very small for hand emasculation and pollination, which is a limiting factor of many research institutes. Besides that, another way to obtain genetic variation is through mutation breeding. Mutations can occur either spontaneously or can be induced.

Induced mutation is a relatively quicker method for improvement of turfgrass and has been used in recent years as a valuable supplement to breeding and development of better crop cultivars (Awan, 1991). Induced mutation plays a vital role in creating additional genetic variations and gamma rays irradiation is one of the mutagenic agents available. Normally large plant populations are required to raise a segregating population. A better way would involve efficient management of first and second generations that could give the greatest possibility for selection of different mutants. For the improvement of a crop, the extent of genetic variability is more important than the total variability. The inheritance of important economic traits such as yield, quality, adaptation, pest and stress resistance, upon which much of the future of plant improvement depends, can be understood through the analysis of a wide range of induced mutations.

There are also important lessons to be learned from the attempts of hybridists and mutation breeders to introduce abiotic and biotic stress resistance into plants (Casler and Van Santen, 2010). The effects of physical and chemical mutagens are well characterized and are very similar to the spontaneous mutation arising *in vitro* or somaclonal variation. Somaclonal variation has contributed to the development of abiotic and biotic stress resistant varieties in major crops. Biotic factor due to the environment are one of the limitations to turfgrass potential. For examples *A. compressus* cannot tolerate drought conditions while *C. dactylon* can poorly tolerate shade conditions. Thus, these situations became limiting factors for turfgrass and reduces turfgrass potential.

In view of the above limitations, several studies were conducted to achieve the following objectives:

- I. Determine the optimum dose of gamma rays irradiation for the development of morphological variations in common *C. dactylon, Z. japonica* and *A. compressus*.
- II. Asses the morphological variations obtained through gamma rays irradiation.
- III. Screen the *C. dactylon* and *A. compressus* mutants for shade and drought tolerance.

REFERENCES

- Ahloowalia, B.S. and M. Maluszynski, 2001. Induced mutations A new paradigm in plant. Euphytica, 118 (2): 167-173
- Ahloowalia, B. S., 1990. In vitro radiation induced mutagenesis in potato. In: The impact of biotechnology in agriculture. Sangwan RS, SangwanNoreel BS (eds). Klumer Academic publisher, Dordrecht, Netherlands.39-46
- Ahloowalia, B. S., 1997. In vitro selection of mutants. In: Somaclonal variation and mutagenesis in plant improvement and in vitro selection of mutants. 15th IAEA/FAO Interregional Training Course on Advances in Technologies for induced mutation in crops.Siebersdorf, Vienna, Austria. 1-6
- Ahloowalia, B. S., M. Malusznyski, and K. Nichterlin, 2004. Global impact of mutation derived varieties. Euphytica.135: 187-204
- Akıncı, S. 1997 physiological responses to water stress by Cucumis sativus L. and related species. Ph. D. Thesis, University of Sheffield. U. K.
- Alshammary, S.F., Qian, Y.L. and Walner, S.J. 2004. Growth responses of four turfgrass species to salinity. *Agil. Water Manage*. 66: 97–111.
- Alshammary, S.F., Qian, Y.L. and Walner, S.J. 2004. Growth responses of four turfgrass species to stress. *Agil. Water Manage*. 66: 97–111.
- Angelis, N.G. and Angelis, B.L.D. 1999. Ornamentals: Landscaping of the other applications. *R. Bras Onramp.* 5: 12–19.
- Anon. 2002. The Bio Space Glossary. BIOSPACE. http://biospace.com/gls Accessed on 12 Feb 2013.
- Anonymous. 1986. Plant Domestication by Induced Mutations. Proceedings of an Advisory Group Meeting, Vienna, 17-21 November 1986. IAEA-STI/ PUB/793 FAO/IAEA, Vienna
- Anonymous. 1998. Soil Fertility Handbook. Ontario Ministry of Agriculture and Food. p. 72-74
- Anonymous. 2011. Golf courses in Malaysia. Retrieved 23rd February 2011 from http://www.malaysiagolfholiday.com/golf-courses.html.
- Ashraf, M. 2007. Improving plant abiotic stress resistance by ecogenous application of osmoprotectants of glycinebetine and praline. *Environ. Expt. Bot.* 59: 206–216.
- Awan, M. A. 1991. Use of induced mutations for crop improvement in Pakistan Int. symposium on the contribution of plant mutation breeding to crop improvement. Vienna Austria.IAEA. 553:67-72.

- Barnes, D. E. and Chan, L.G. 1990. Common weeds of Malaysia and their control. Ancom Berhad, Persiaran, Sellangor, 40000 Shah Alam, Malaysia, June, pp. 1–349.
- Barrios, E. P., Sundstrum, F. J., Babcock, D., and Leger, L. 1986. Quality and yield response of four warm-season lawngrasses to shade conditions. Agron. J. 78:270-273.
- Bauman, F. and Crane, P.L. 1992. Hybrid corn History, development and selection considerations. National Corn Handbook. Purdue University, US.
- Baldwin, C.M., H. Liu, L.B. McCarty, W.L. Bauerle, and J.E. Toler. 2006. Response of six bermudagrass cultivars to different irrigation intervals. HortTechnology 16:466–470.
- Beard, J. B., 1973. Turfgrass: Science And Culture. Rnglewood Cliffs, New Jersey: Prentice-Hall, Inc. A Simon & Schuster Company.
- Beard, J. B., & Green, R. L. (1994). The role of turfgrasses in environmental protection and their benefits to humans. Journal of Environmental Quality, 23 (3), 1-16. Retrieved (July 21, 2010).
- Bell, G.E., Danneberger, T.K., and McMahon, M.J. 2000. Spectral irradiance available for turfgrass growth in sun and shade. *Crop Sci.* 40:189-195.
- Blixt S, Bhrenberg L, Gelin O. 1963. Studies on induced mutations in peas VII. Mutation spectrum and mutation rate of different mutagenic agents. Agri Hortique Genetica 22, 286-94.
- Boardman, N. K. 1977. Comparative photosynthesis of sun and shade plants. Annu. Rev. Plant Physiol., 28: 355-377.
- Bohnert, H.J., D.E. Nelsen, and R.J. Jenson. 1995. Adaptations to environmental stresses. Plant Cell 7:1099–1111.
- Bottner, P., Couteaux, M. M. and Vallejo, V. R. 1995 Soil organic matter in mediterranean-type ecosystems and global climatic changes: A case studythe soils of the mediterranean basin. Global change and Mediterranean-type ecosytems. Ecological studies, Vol. 117. (ed. by Jose M. Moreno, Walter C. Oechel),. Springer-Verlag, New York. pp. 306-325
- Bowler, C., M. Van Montagu, and D. Inze. 1992. Superoxide dismutase and stress tolerance. Annu. Rev. Plant Physiol. Plant Mol. Biol. 43:83–116.
- Bottino, P. J., 1965. Radio sensitivity studies on an interspecific grass hybrid. J. Hered. 56:225-258.
- Broertjes, C., 1975. The development of (new) in vivo and in vitro techniques of significance for mutation breeding of vegetatively propagated crops. In :

Improvement of vegetatively propagated plants through induced mutations . Proc. of Res . Coordinating Meeting. Tokai, Japan. IAEA, Vienna, pp. 23-31.

- Bruneau, A. H. 2010. Morphological mutants of St. Augustinegrass induced by gamma ray irradiation Plant Breeding 129, 412–416.
- Bunnell, B.T., L.B. McCarty, J. Faust, W.C. Bridges, Jr., and N.C. Rajapakse. 2005. Quantifying a daily light integral requirement of TifEagle bermudagrass golf green. *Crop Sci.* 45:569-574.
- Bunnell, B.T., L.B. McCarty, and W.C. Bridgers, Jr. 2005. Evaluation of three bermudagrass cultivars and Meyer Japanese zoysiagrass grown in shade. *Int. Turfgrass Soc. Res. J.* 10:826-833.
- Burton, G., and W. Hanna. 1985. Bermudagrass. In M. Heath, R. Barnes, and D. Metcalfeed. Forages the science of grassland agriculture. Iowa StateUniversity Press, Ames, Iowa.643 pp.
- Burton, G. W. 1991. A history of turf research at Tifton. USGA Green Section Record, 29.
- Burton, G. W. 1974. Radiation breeding or warm season for age and turfgrasses. p. 35-39. *In* Polyploidy and induced mutations in plant breeding. IAEA, Vienna
- Busey, P., 1977. Turfgrasses for the 1980's. Proc Fla. State Hort. Soc. 90:111-114
- Busey, P., 1980. Gamma ray dosage and mutation breeding in St .Augustinegrass . Crop Sci. 20: 181-184
- Carrow, R.N. 1996. Drought resistance aspects of turfgrasses in the southeast: Rootshoot responses. *Crop Sci.* 36:687–694.
- Carrow, R.N. and Duncan, R.R. 1998. Salt-affected turfgrass sites: assessment and management. Wiley, Hoboken, NJ.
- Casler, M.D. and G.A. Pederson. 1996. Host resistance and tolerance and its deployment. pp. 475–507. In S. Chakraborty et al., Eds. Pasture and Forage Crop Pathology. American Society of Agronomy, Madison, WI.
- Casler M, and Van Santen E. 2010. Breeding objectives in forages. In: Boller B, Posselt U, Veronesi F. eds. Handbook of plant breeding: fodder crops and amenity grasses. New York: Springer Science and Business Media, 115–160.
- Chapman, G.P. and Peat, W.E. 1992. An introduction to the grasses. CAB International, Wallingford.
- Chase, A. 1948. The meek that inherit the earth. In *Grass Yearbook of Agriculture* pp. 8-15. USA: U.S. Govt. Printing Office.

- Christians Nick E. 2007. Fundamentals of turfgrass management, In: John Willey & Sons Warm season grasses, 3rdedn. Hoboken, New Jersey, 61-63
- Conger AD, Sparrow AH, Schwemmer SS, Klug EE, 1982. Relation of Nuclear volume and radiosensitivity to ploidy level in higher plants and yeast. Environmental and Experimental Botany 22, 55-74
- Da Costa, M. and B. Huang. 2007. Changes in antioxidant enzyme activities and lipid peroxidation for bentgrass species in response to drought stress. J. Amer. Soc. Hort. Sci. 132:319–326.
- Danida. 2002. Assessment of potentials and constraints for development and use of plant biotechnology in relation to plant breeding and crop production in developing countries. Working paper. Ministry of Foreign Affairs, Denmark.
- Daniel, W. H. 1969. The evaluation of a new turfgrass.p.57-64 in R.R. Davis (ed.) Proc. First Int. Turfgrass Res. Conf. Horragate, England. July 1969. Sport Turf Res. Ins., Bingley, England.
- Darwin, C. 1875. The Variation of Animals and Plants under Domestication. 2nd ed. Murray, London.
- Das, A., S. S. Gossal, J. S. Sindhu, and H. S. Dhaliwal, 2000. Induced mutations for heat tolerance in potato by using in vitro culture and radiation. Euphytica. 114: 205-209
- Dickens, R., W.J. Johnston, and R.L. Haaland. 1981. Variability observed in centipedegrass grown from 60Co-irradiated seed. Agron. J. 73:674–676.
- Dudeck, A.E. and Peacock, C.H. 1985. Effects of salinity on seashore paspalum turfgrasses. *Agron. J.* 77: 47–50.
- Dudeck, A.E. and Peacock, C.H. 1993. Salinity effects on growth and nutrient uptake of selected warm-season turfgrasses. *Int. Turfgrass Soc. Res. J.* 7: 680–686.
- Dudeck, A.E. and C.H. Peacock. 1992. Shade and turfgrass culture. In *Turfgrass, Agronomy Monograph* 32, Waddingtion D.V., Carrow R.N., and Shearma R.C. Eds., ASA-CSSA-SSSA, Madison, WI. pp. 269-284.
- Duble, R. 2003. Recreational Turf Class Notes. Ch. 1-10. Retrieved (July 7, 2010). From http://scsc302.tamu.edu/scsc302-700+/rduble/index.html
- Duncan, R.R. and Carrow, R.N. 1999. Turfgrass molecular genetic improvement for biotic/edaphic stress resistance. *Adv. Agron.* 67, 233–305.
- Emmons R. D., 1984. Turfgrass Science And Management. Albany, New York:Delmar Publishers Inc

- Erickson, P.I., M.B. Kirkham, and G.B. Adjei. 1979. Water relations, growth and yield of tall and short wheat cultivars irradiated with X-rays. Environ. Expt. Bot. 19:349–356.
- FAO/IAEA database, 2006: http://www-infocris.iaea.org/MVD/.
- Finney, D. J., 1971: Probit Analysis, 3rd edn. Cambridge University Press, Cambridge, UK
- Fly, J. and B. Huang.2004. Turf Management in the transition zone. John Wiley & Sons, Inc
- Flowers, T.J. Troke, P.F. and Yeo, A.R. 1977. The mechanism of salt tolerance in halophytes. *Ann. Rev.Plant Physiol.* 28: 89–119.
- Flowers, T.J., Flowers, S.A., Hajibagheri, M.A. and Yeo, A.R. 1990. Salt tolerance in the halophytic wild rice, *Porteresia coantata. New Phytol.* 114: 675–684.
- Gaul, H. 1964. Mutation in plant breeding. Radiat bot. 4 No 3, pp 155-232.
- Ghazali, H., A. R. Harun and S. Samsudin, 2003. Study on mutagenesis of signal grass By gamma irradiation. Malaysian Institute for Nuclear Technology Research (MINT).
- Gepts, P. 2002. A Comparison between Crop Domestication, Classical Plant Breeding, and Genetic Engineering. *Crop Sci.* 42:1780–1790
- Glenn, E.P. 1987. Relationship between cations accumulation and water content of salttolerant grasses and a sedge. *Plant Cell Environ*. 10: 205–212.
- Glenn, E.P., Watson, M.C., O'Leary, J.W. and Axelson, R.D. 1992. Comparison of salt tolerance and osmotic adjustment of low-sodium and high-sodium subspecies of the C₄ halophytes, *Atriplex canescens. Plant Cell Environ.* 15: 711–718.
- Gould, F. 1951. Grasses of southwestern United States. University of Arizona, Tucson, Arizona. 343 pp
- Hahn, E. D., and R. Soyer, 2008. Probit and logic model. Differences in a multivariate realm.
- Harlan, J., and J. de Wet. 1969. Sources of variation in Cynodondactylon. Crop Science 9:774-778
- Hanna, W., and E. Elsner, 1999: Registration of TifEagle bermudagrass. Crop Sci. 39,1258.
- Hanna, W., J. Dobson, R. R. Duncan, and D. Thompson, 1997: Registration of TifBlair centipedegrass. Crop Sci. 37, 1017.

- Harivandi, A. 2011. Purple gold: a contemporary view of recycled water irrigation USGA Green Section Record. 49 (45): 1-10.
- Haynes, R. H., and F. Eckardt, 1979. Analysis of dose-response patterns in mutation research. Can. J. Genet. Cytol. 21:277:302
- Hester, M.W., Mendelssohn, I.A. and McKee, K.L. 2001. Species and population variation to salinity stress in *Panicum hemito-mon*, *Spartina patens*, and *Spartina alternifora*: Morphological and physiological constraints. *Environ. Exp. Bot.* 46: 277–297.
- Holmes, M.G. and Smith, H. 1977. The fuction of phythochrome in the natural environment. I. Characterisation of daylight for studies in photomorgenesis and photoperiod, *Photochem. Photbiol.* 25:533-538.
- Holm, L. G., P. Donald, J. V.Pancho, and J. P. Herberger. 1977. The World's Worst Weeds: Distribution and Biology. The University Press of Hawaii, Honolulu, Hawaii. 609pp.
- Hong, Z.L., K. Lakkineni, Z.M. Zhang, and D.P.S. Verma. 2000. Removal of feedback inhibition of D1-pyrroline-5-carboxylate synthetase results in increased proline accumulation and protection of plants from osmotic stress. Plant Physiol. 122:1129–1136.
- Jain S. M., 2006. Mutation-assisted breeding for improving ornamental plants. Acta Horticulturae. 714, 85-98
- Jander G, Baerson SR, Hudak JA, Gonzalez KA, Gruys K, Last RL, 2003. Ethyl methanesulfonate saturation mutagenesis in Arabidopsis to determine frequency of herbicide resistance. Plant Physiology 131,139-146
- Jiang, Y. and R.N. Carrow. 2007. Broadband spectral reflectance models of turfgrass species and cultivars to drought stress. Crop Sci. 47:1611–1618.
- Jiang Y., R.R. Duncan, and R.N. Carrow. 2004. Assessment of low light tolerance of seashore paspalum and bermudagrass. *Crop Sci.* 44:587-594.
- Johnson, W. J., R. Dickens, R. Haaland, and J. R. Cooper. 1977. Irradiation-induced variation in centipedegrass. Agron. Abstr. P. 60.
- John, C.S and David, S.G. 2007. Shade stress and Management. In: Handbook of Turfgrass Management and Physiology. Mohammad Pessarakli, ed. pp. 447-467. Tucson. CRC Press.

Juraimi A. S. 2001. Turfgrass: types, uses and maintenance. Garden Asia. 8:40-43.

Katerjii, N., Hoorn, J.W., Hamdy, A., Mastrorilli, M., 2000. Salt tolerance classification of crops to soil salinity and to water stress day index. *Agric. Water Manage.* 43: 99-109.

- Kenicer, C. 1997. Asia's five best golf courses. *Fortune Magazine*, November 10, 1997.
- Kjelgren, R., Rupp, L. and Kilgren, D. 2000. Water conservation in urban landscapes. *Hort Sci.* 35: 1037-1043.
- Kramer, P. J. 1980 Drought, stress, and the origin of adaptations. Adaptations of plants to water and high temperature stress. (ed. by Neil C. Turner, Paul J. Kramer) pp. 7-20. John-Wiley & Sons, New York.
- Kramer, P. J. and Boyer, J. S. (1995) Water relations of plants and soils. Academic Press. San Diego.
- Lee, I.S., D. S. Kim, D.Y. Hyun, S. J. Lee, H. S. Song, Y. P. Lim, and Y. I. Lee, 2003. Isolation of gamma-inuced rice mutants with increased tolerance to salt by anther culture. J. Plant Biotechnology.5 (1): 51-57
- Lee, D.W. 1985. Duplicating foliage shade for research on plan development. *Hort Science*. 20:116-118.
- Lineberger, D. R. 1996. Origin, development and propagation of chimeras.http:/aggie-horticulture.tamu.edu/tissult/chimeralec/chimeras
- Liu L, van Zanten L, Shu QY, Maluszynski M .2004. Officially released mutant varieties in China. Mutation Breeding Reviews 14, 1-61.
- Long, J. A. 1972.Developing superior turfgrass varieties. P. 53-56. In V. B. Youngner and C. M. Mckell (eds.). The biology and utilization of grasses.Physiology ecology series. Academy Press Inc., New York.
- Lorenzi, H. and Souza, H.M. 2001. Ornamental plants in Brazil: Shrub, grass and creepers. 3. ed. *New Plantarum*. p. 558.
- Lu, S., Z. Guo, and X. Peng. 2003. Effects of ABA and S-3307 on drought resistance and antioxidative enzyme activity of turfgrass. J. Hort. Sci. Biotechnol. 78:663–666.
- Lu, S., Z. Wang, X. Peng, Z. Guo, G. Zhang, and L. Han. 2006. An efficient callus suspension culture system for triploid bermudagrass (Cynodon transvaalensis ·C. dactylon)
- Lu, S., C. Chen, Z. Wang, Y. Niu, Z. Guo, and B. Huang. 2008 Antioxidant responses of radiation-induced dwarf mutants of bermudagrass to drought stress. J. Amer. Soc. Hort. Sci. 133:360–366.
- Madison, J.H. 1971. Practical turfgrass management. pp. 18-20. New York: Van Nostrand Reinhold Co. Floradwarf and Tifdwarf bermudagrass exposed to various light regimes. *Int. Soc. Res. J.* 10:879-884.

- Marcum, K.B. 1999. Salinity tolerance mechanisms of grasses in the subfamily *Chloridodeae*. *Crop Sci.* 39: 1153–1160.
- Marcum, K.B. and Murdoch, C.L. 1990. Growth responses, ion relations, and osmotic adaptation of eleven C₄ turfgrasses to salinity. *Agron. J.* 82: 892–896.
- Marcum, K.B., Anderson, S.J. and Engelke, M.C. 1998. Salt gland ion secretion: A salinity tolerance mechanism among five zoysiagrass species. *Crop Sci.* 38: 806–810.
- Marschner, H. 1995. Adaptation of plants to adverse chemical soil conditions. *In* Mineral nutrition of higher plants. 2nd edition. Academic Press, London. pp. 596-680.
- Massoud, M., Scrimshaw M. and Lester J. 2003. Qualitative assessment of the effectiveness of the Mediterranean action plan: wastewater management in the Mediterranean region. *Ocean Coastal Manag.* 46:875-899.
- Mattioni, C., N.G. Lacerenza, A. Troccoli, A.M. De Leonardis, and N. Di Fonzo. 1997. Water and salt stress-induced alterations in proline metabolism of Triticum durum seedlings. Physiol. Plant. 101:787–792.
- McBee, G. G., & Holt, E. 1966. Shade tolerance studies on bermudagrass and other turfgrasses. *Agronomy Journal*, 58(5), 523-525.
- McBee, G.G. 1969. Association of certain variation in light quality with the performance of selected turfgrasses. *Crop Sci.* 9:14-17.
- McCullough, P., McCarty, B., Baird, V., Li, H., & Whitwell, T. (2004). Ultradwarf Bermudagrasses Exhibit Easy Mutation Tendencies. *Bermudagrass Management*, 74-78.
- Micke, A., 1976. Bibliography. Examples of literature related to the use of induced in crossbreeding mutations. In : Induced mutations in cross-breeding . IAEA, Vienna, pp .233-252
- Micke A and Donini B (1993) Induced mutations.*In* Hayward MD, Bosemark NO and Romagosa I (Eds.) Plant Breeding Principles and Prospects. Chapman and Hall, London, pp 52-62.
- Miller, G.L., Edenfield, J. T., and Nagata, R.T. 2005.Growth parameters of McCullough, P., McCarty, B., Baird, V., Li, H., & Whitwell, T. (2004). Ultradwarf Bermudagrasses Exhibit Easy Mutation Tendencies. Bermudagrass Management, 74-78.
- MINT, 1998. Mutation Breeding Manual. The Use of Induced Mutation for Plant improvement in Malaysia, National Committee on the Use of Induced Mutation in Plant Breeding. Malaysian Institute for Nuclear Technology Research (MINT), Bangi.

- Mittler, R. 2002. Oxidative stress, antioxidants and stress tolerance. Trends Plant Sci. 7:405–410.
- Michelle, M. S., R. D. Thompson and C. S. Weil, 1994. Computer programme for calculation of median effective dose (LD50) using the method of moving average interpolation. Arch Tovicol 68: 332-337.
- Munshaw, G.C., Zhang, X. and Ervin, E.H. 2004. Pass the salt. GCM J. 81–92.
- Neville P.A., Nabaya N.B., Tanguay M., 1998. Mutagenic effects of acute gamma irradiation on miniature roses: target theory approach. *Hort Sci.* 33: 127-129.
- Noctor, G. and C.H. Foyer. 1998. Ascorbate and glutathione: Keeping active oxygen under control. Annu. Rev. Plant Physiol. Plant Mol. Biol. 49:249–279.
- Pathirana, R., W. A. Wijithawanra, K. Jagoda, and A. L. Ranawaka, Selection of rice iron toxicity tolerance through irradiation caryopsis culture. Plant Cell Tissue and Organ Culture. 70: 83-90, 2002
- Patton, A. 2009. Growing Turfgrass in Shade.Agriculture and Natural Resource.Arkansas Cooperative Extension Service,University of Arkansas.
- Peacock, C.H. and Dudec, A.E. 1985. Physiological and growth responses of seashore pasplaum to salinity. *Hort. Sci.* 20: 111–112.
- Peacock, C. H. and Dudeck, A. E. 1981. Effects of shade on morphological and physiological parameters on St. Augustinegrass cultivars. Proc. 4th Int. Turfgrass Res. Conf. 493-500.
- Pereira, J. S. and Chaves, M. M. 1993. Plant water deficits in Mediterranean ecosystems. Water Deficits plant responses from cell to community. (ed. by J. A. . Smith, H. Griffiths). pp. 237-251. BIOS Sci. Ltd. Oxford.
- Pereira, J. S. and Chaves, M. M. 1995. Plant responses to drought under climate change in mediterranean-type ecosystems. Global change and Mediterraneantype ecosytems. Ecological studies, Vol. 117. (ed. by Jose M. Moreno, Walter C. Oechel), pp. 140-160. Springer-Verlag, New York.

Piperno, D.R. and Sues, H.D. 2005. Dinosaurs Dined on Grass. Sci. 310: 1126–1128.

- Philley, H. W., & Krans, J. V 1998. Turf performance of seeded bermudagrass cultivars. Golf Course Management, 11, 62 - 66. Retrieved (July 16, 2010).
- Powell, J.B., 1974. Induced mutations in turfgrasses as a source of variation for improving cultivars. P.3-8. In E.C. Roberts (eds.) Proc. Second Int. Turfgrass Res. Conf., Blacksburg, Va. June 1973. Am. Soc. Agron. And Crop Science. Soc. Am. , Madison, Wis.

- Powell, J.B., 1976. Induced mutations in highly heterozygous vegetatively propagated grasses In : Induced mutations in cross-breeding. IAEA, Vienna, pp. 219-224.
- Powell, J. B., G. W. Burton & J. R. Young, 1974. Mutations induced in vegetatively propagated turf bermudagrasses by gamma radiation. Crop Sci.14: 327-330.
- Qian, Y. L. and Engelke, M. C. 1999. Performance of five turfgrasses under linear gradient irrigation. *Hort. Sci.* 34:893–896.
- Qian, Y.L., Wilhelm, S.J. and Marcum K.B. 2007. Comparative responses of two Kentucky bluegrass cultivars to salinity stress. *Crop Sci.* 41: 1895–1900.
- Rambal, S. and Debussche, G. (1995) Water balance of Mediterranean ecosystems under a changing climate. Global change and Mediterranean-type ecosytems. Ecological studies, Vol. 117. (ed. by Jose M. Moreno, Walter C. Oechel), pp. 386-407. Springer-Verlag, New York.
- Raven, P.H., Evert, R.F. and Eichhoron, S.E. 2001. Plant Biology. Translation Antonio Salatino *et al.*, 6th ed. Janeiro : Guanabara Koogan. p.237.
- Reynolds, W. C., R. Li, K. de Silva, A. H. Bruneau, and R. Qu, 2009: Field performance of mutant and somaclonal variation lines of St. Augustinegrass [Stenotaphrum secundatum (Walt.) Kuntze]. Intl. Turfgrass Soc. Res. J. 11, 573–582.
- Rhodes, D., Nadolska-Orczyk, A. and Rich, P.J. 2002. Salinity, osmolytes and compatible solutes. In: Lauchli, A., Luttge, U.(Eds.), Salinity: Environment-Plants-Molecules. Kluwer Academic Publ., Boston, MA.
- Richards, L.A. and Weaver, L.R. 1944. "Moisture retention by some irrigated soils as related to soil moisture tension". *Journal of Agricultural Research* 69: 215–235.SAS Institute. 2009. SAS/STAT user's guide. release. Release 9.2. 4th ed. Statistical Analysis Institute, Cary, NC.
- Richard, L.D. 2001. Turfgrasses: their management and use in the southern zone (2nd Edition), pp. 4-10. Texas: A&M University Press.
- Samudio, S.-H. and A.D. Brede. 2002. Registration of 'Sundevil II' bermudagrass. Crop Sci. 42:670–671.
- Schmidt, R. E. 1966. Influence Of Light And Temperature On Turf (Both Warm And Cool Season Grasses). Dept. of Agronomy, Virginia Polytechnic Institute, Blaksburg.
- Schum A., 2003. Mutation breeding in ornamentals and efficient breeding method.Acta Hort.,612: 47-60.
- Sharp, R. E. and Davies, W. J. (1989) Regulation of growth and development of plants growing with a restricted supply of water. Plants under stress.

Biochemistry, physiology and ecology and their application to plant improvement. (ed. by Hamyln, G. Jones, T. J. Flowers, M. B. Jones). pp. 71-93. Cambridge University Press, Cambridge.

- Shatanawi, M., Hamdy, A. and Smadi, H. 2003. Urban wastewater: problem, risks and its potential use for irrigation. In *The use of non conventional water resources*, ed. Hamdy A., pp.15-44. Bari : CIHEAM-IAMB / EU-DG Research.
- Song HS, Kang SY., 2003. Application of Natural Variation and Induced Mutation in Breeding and Functional Genomics: Papers for International Symposium; Current Status and Future of Plant Mutation Breeding.Korean J. Breed. Sci., 35(1): 24-34
- Stanford, R.L., R.H. White, J.P. Krausz, J.C. Thomas, P. Colbaugh, and S.D. Abernathy. 2005. Temperature, nitrogen and light effects on hybrid bermudagrass growth and development. *Crop Sci.* 45:2491-2496.
- Tae-Woong Bae., 2009. Production of unbolting lines through gamma-ray irradiation mutagenesis in genetically modified herbicide-tolerant Zoysia japonica. Subtropical Horticulture Research Institute, Cheju National University, Jeju, 690-756, Korea
- Tan, Z.G and Qian, Y.L. 2003. Light intensity affects gibberellic acid content in Kentucky bluegrass, *Hort Sci.*, 38: 113-116.
- Taiz, L. and Zieger, E. 2006. Plant Physiology.4th ed. Sinauer Associates, Inc, Sunderland, MA.
- Toler, R. W., J. B. Beard, M. P. Grisham, and R. L. Crocker, 1985: Registration of TXSA 8202 and TXSA 8218. St. Augustinegrass germplasm resistant to Panicum mosaic virus S. Augustine decline strain. Crop Sci. 25, 371.
- Turgeon, A.J. 2008. Turfgrass management (8th edition). Upper saddle River, New Jersey: Prentice Hall Publishing Co.
- Uddin, M. K., Juraimi, A.S., Begum, M., Ismail, M.R., Radziah O. and.Rahim A.A 2009. Floristic Composition of Weed Community in Turfgrass Area of West Peninsular Malaysia. *International Journal of Agriculture and Biology*. 11: 13-20.
- Uddin, M.K., Juraimi, A.S., Begum, M., Ismail, M.R., Rahim, A.A., and Radziah, O. 2009. Floristic composition of weed community in turfgrass area of west Peninsular Malaysia. *Inter. J. Agr. Biol.* 11(1): 13-20.
- United Nation Human Development Report 2006. Beyond scarcity: Power, poverty and the global water crisis.
- Waddington, D.V., Carrow, R.N. and Shearman, R.C. 1992. Turfgrass. Madison USA: American Society of Agronomy, Inc., Crop Science Society of America, Inc. and Soil Science Society, Inc.

- Watson, J.R. 1985. Water resources in the United States. In *Turfgrass water* conservation, ed. Gibeault, V.A. and Cockerharn, S.T. pp. 19-36. Riverside: Univ. of California, Div. of Agric. and Natural Resources, Publ.
- Watson, L. and Dallwitz, M.J. 1992. The grass genera of the world, pp. 223-986. UK: CAB Publications.
- Wherley, B.G., Gardner, D.S., and Metzger, J.D. 2005. Tall fescue photomorhenesis as influenced by changes in spectral composition and light intensity, *Crop Sci*.45:562-568.
- Wilkinson, J.F. and J.B. Beard. 1975. Anatomical responses of Merion Kentucky bluegrass and Pennlawn red fescue at reduced light intensities. *Crop Sci.* 15:189-194.
- Winstead, C.W., and Ward, C.Y. 1974. Persistence of southern turfgrasses in a shade environment. in Proc. 2nd Int. Turfgrass Res Conf. Robert, E.C, Ed., ASA and CSSA, Madison, WI. pp 221-230
- Wood, G.M. 1969. Evaluating turfgrass for shade tolerance. Agron. J. 69:347-352
- Zhang, J. and M.B. Kirkham. 1994. Drought-stress-induced changes in activities of superoxide dismutase, catalase, and peroxidase in wheat species. Plant Cell Physiol. 35:785–791.