

UNIVERSITI PUTRA MALAYSIA

USE OF REPRESENTATIONAL DIFFERENCE ANALYSIS TO REVEAL DIFFERENCES BETWEEN TRUNCATED LEAF SYNDROME AND NORMAL OIL PALM RAMET

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Ву

KANAGAMALAR SILVARAJOO

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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By

KANAGAMALAR SILVARAJOO

November 2015

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Truncated Leaf Syndrome (TLS) is a commonly found abnormality amongst tissue cultured plantlets of oil palm (*Elaies guineensis* Jacq.) which, if severe, will eventually lead to the death of the ramets. It was hypothesized that this phenotype could be due to genetic or epigenetic variability. The first part of this study was aimed to identify the genetic variability via Genomic-Representational Difference Analysis (G-RDA), a technique whereby the differences between two closely related genomes can be identified. In this part, 2 clones of oil palm ramets (1181; severe vs normal and 2751; mild vs normal) were used as starting material with 4 rounds of forward and reverse G-RDA were performed. A total of 18 unique sequences from G-RDA were successfully obtained. Primers were designed and verification of forward G-RDA products through PCR analyses and sequence comparison was carried out using 12 clones of TLS and normal oil palm ramets. Two out of 18 set of primers [F4(6)-1181Bgl and F4(10)-1181Bgl] were identified as potential markers and further verified by PCR and Southern analyses. The primer set F4(6)-1181Bgl was only able to distinguish between TLS and normal ramets of only one genotype (Yangambi) with the presence of expected band in TLS but was absent in normal ramets. The primer set F4(10)-1181Bgl showed the presence of multiple banding pattern in the genotype of La Me and Yangambi. Analysis of the multiple bands sequences revealed that those sequences represent multiple regions within the same genome, hence it is potentially a polymorphic marker. The 2 primer sets mentioned above could be classified as potential genotype specific primers as it is only functional in selected genotype. The second part of this study was aimed to identify the epigenetic variability via Methylation Sensitive-Representational Difference Analysis (MS-RDA). In this part, 2 rounds of forward and reverse MS-RDA were carried out and a total of 4 differentially methylated sequences with matches to known gene were successfully obtained. Primers were designed based on the 4 target genes namely protein ycf68, 3-ketoacyl-CoA synthase 12, Scarecrow-like protein 9 and Nucleotide-diphosphosugar transferases superfamily protein and verification process was carried out by Quantitative-PCR to identify the respected expression level in normal and TLS ramets. The relative expression level of uncharacterized protein ycf68 is up-regulated, while 3ketoacyl-CoA synthase 12, Scarecrow-like protein 9 and Nucleotide-diphospho-sugar transferases superfamily protein were down-regulated in TLS ramets as compared to the normal ramets. Further verification with extensive number of samples is needed to elucidate the potential of the above 2 primer sets from G-RDA and 4 primer sets from MS-RDA to be used as markers across all genotypes of oil palm.



PENGGUNAAN ANALISIS PERBEZAAN PERWAKILAN UNTUK MENGENALPASTI PERBEZAAN ANTARA ANAK POKOK KELAPA SAWIT YANG MENGALAMI SINDROM DAUN TERBANTUT DAN NORMAL

Oleh

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November 2015

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Sindrom daun terpangkas (TLS) adalah ketidaknormalan yang biasa ditemui di kalangan anak pokok tisu kultur daripada kelapa sawit (Elaies guineensis Jacq.) yang mana, jika teruk, akhirnya akan membawa kepada kematian anak pokok tersebut. Ia telah dihipotesiskan bahawa fenotip ini mungkin disebabkan oleh perubahan genetik atau epigenetik. Bahagian pertama kajian ini bertujuan untuk mengenal pasti perubahan genetik melalui teknik genomik-analisis perbezaan perwakilan (G-RDA), teknik di mana perbezaan antara dua genom berkait rapat dapat dikenal pasti. Dalam bahagian ini 2 jenis klon anak pokok kelapa sawit (1181; parah vs normal dan 2751; kurang parah vs normal) telah digunakan sebagai bahan permulaan dengan 4 pusingan G-RDA ke hadapan dan ke belakang telah dijalankan. Sebanyak 18 jujukan unik daripada G-RDA telah berjaya diperolehi. Pencetus telah direka dan verifikasi produk G-RDA melalui analisis PCR dan perbandingan jujukan telah dilaksanakan dengan menggunakan 12 klon kelapa sawit yang TLS dan normal. Dua daripada 18 set pencetus [F4 (6) -1181Bgl dan F4 (10) -1181BgI] telah dikenal pasti sebagai penanda molekular berpotensi dan seterusnya disahkan dengan PCR dan analisis Southern. Set primer F4 (6) -1181Bgl hanya dapat membezakan satu genotip (Yangambi) ramets TLS dan normal dengan kehadiran jalur di ramet TLS tetapi tidak di ramet normal. Set primer F4 (10) -1181Bgl menunjukkan kehadiran beberapa corak jalur dalam genotip La Me dan Yangambi. Analisis kehadiran beberapa corak jalur tersebut mendedahkan bahawa jalur-jalur tersebut mewakili pelbagai kawasan di dalam genom yang sama yang boleh dikatakan penanda berpotensi polimorfik. Dua set pencetus yang dinyatakan di atas boleh diklasifikasikan sebagai pencetus genotip berpotensi tertentu kerana ia hanya berfungsi dalam genotip tertentu sahaja. Bahagian kedua kajian ini bertujuan untuk mengenal pasti kepelbagaian epigenetik melalui teknik metilasi sensitif-analisis perbezaan perwakilan (MS-RDA). Dalam bahagian ini, 2 pusingan MS-RDA ke hadapan dan ke belakang telah dijalankan dan sebanyak 4 jujukan bermetil yang menunjukkan padanan dengan gen sasaran telah berjaya diperolehi. Empat set primer telah direka berdasarkan 4 sasaran gen iaitu "uncharacterized proteiun ycf68" adalah diekspresikan sedikit, manakala "3-ketoacyl-CoA synthase 12", "Scarecrow-like protein 9" dan "Nucleotide-diphospho-sugar

transferases superfamily protein" dan pengesahan telah dilakukan dengan menggunakan kaedah kuantitatif-PCR untuk mengenal pasti tahap ekspresi jujukan-jujukan tersebut dalam klon TLS dan normal. Tahap ungkapan relatif "uncharacterized proteiun ycf68" adalah diekspresikan sedikit, manakala "3-ketoacyl-CoA synthase 12", "Scarecrow-like protein 9" dan "Nucleotide-diphospho-sugar transferases superfamily protein" adalah diekspresikan banyak di klon TLS berbandingan dengan klon normal. Pengesahan lanjutan dengan jumlah samel yang banyak diperlukan untuk menjelaskan potensi dua set primer peroleh daripada G-RDA dan 4 set primer yang diperoleh daripada MS-RDA untuk digunakan sebagai marker di semua genotip kelapa sawit.



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I certify that a Thesis Examination Committee has met on 20 November 2015 to conduct the final examination of Kanagamalar Silvarajoo on her thesis entitled "Use of Representational Difference Analysis to Reveal Differences between Truncated Leaf Syndrome and Normal Oil Palm Ramet" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the (insert the name of relevant degree).

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LIST OF ABBREVIATIONS

 $\begin{tabular}{lll} \% & percentage \\ ^{\circ}C & degree Celsius \\ \mu g & microgram \\ \mu l & microliter \\ \mu M & micromolar \\ & & female \\ & & & \\ \end{tabular}$

2,4-D 2,4-dichlorophenoxyacetic acid

A adenosine

AFLP amplified fragment length polymorphism

ATP adenosine triphosphate

BLAST basic local alignment search tool

bp base pair

BSA bovine serum albumin

C cytidine

cDNA complementary DNA

CTAB cetyl trimethylammonium bromide dATP 2'-deoxy-adenosine-5'-triphosphates dCTP 2'-deoxy-cytidine-5'-triphosphate diethylpyrocarbonate treated water dGTP 2'-deoxy-guanisine-5'-triphosphate

DNA deoxyribonucleic acid

dNTPs deoxynucleotides triphosphates

dsDNA double stranded DNA

DTT dithiothreitol

dTTP 2'-deoxy-thymidine-5'-triphosphate dUTP 2'-deoxyuridine, 5'-triphosphate

EDTA ethylene diaminetetra acetic acid disodium salt

EEPS N-(2-hydroxyethyl) piperazine-N-3-ropanesulfonic acid

E-value expect value

FELCRA federal land consolidation and rehabilitation authority

FELDA federal land development authority agriculture services sdn. bhd.

g gram

g gravitational acceleration

G guanosine

GC content guanine-cytosine content

G-RDA genomic representational difference analysis IPTG isopropyl β-D-1-thiogalactopyranoside

ISSR inter-simple sequence repeat

Jacq. Jacquin
kb kilo base pair
KCl potassium chloride
LB Luria Bertani

M molar mg milligram

MgCl₂ magnesium chloride MgSO₄ magnesium sulfate

ml millilitre mM milimolar MOPS 3-morpholinopropane-1-sulfonic acid

MPOB malaysian palm oil board

MS-RDA methylation sensitive representational difference analysis

N normality

 Na_2HPO_4 disodium phosphate NAA α -napthalene acetic acid

NaCl sodium chloride

NaH₂PO₄ monosodium phosphate

NaOAc sodium acetate NaOH sodium hydroxide

NCBI national centre for biotechnology information

ng nanogram nm nanometer

ORF open reading frame
PCR polymerase chain reaction
PVPP polyvinylpolypyrrolidone

Q-PCR quantitative PCR

RAPD randomly amplified polymorphic DNA

RE restriction enzyme

RFLP restriction fragment length polymorphism

RNA ribonucleic acid RNase ribonuclease

rpm revolutions per minute rRNA ribosomal RNA

SDS sodium dodecyl sulphate

SNPs single nucleotide polymorphisms
SSC sodium chloride sodium citrate buffer

ssDNA single stranded DNA SSR simple sequence repeat

T thymidine

TAE tris-acetate-EDTA

TE tris-EDTA

TLS truncated leaf syndrome
Tm melting temperature

Tris-Cl tris-chloride

Tris-HCl tris-hydrochloric acid

tRNA transfer RNA

U unit UV Ultraviolet

v/v volume per volume w/v weight per volume

X-gal 5-bromo-4-chloro-3-indolyl-β-D-galactopyranoside

β beta

CHAPTER 1

INTRODUCTION

Elaies guineensis, known as African oil palm or macaw-fat native to west and southeast Africa, is a monocotyledonous plant belonging to the Aracaceace family. It is the most important and valuable palm species which is mainly used in commercial agriculture for the production of palm oil and palm kernel oil. It is also known as an important food that serves as a major source of vegetable oils and fats/lipids. The palm oil and palm kernel oil is derived from fleshy mesocarp and hard kernel of oil palm fruit respectively. The palm oil is normally utilized in food industries whereas palm kernel oil is utilized in oleochemical industries for soaps, detergents and toiletry products making. The African palm is mainly propagated in Asia, particularly in Malaysia and Indonesia.

In Malaysia, it was reported that throughout the year of 2013 the production of palm oil is 19.21 million tonnes per hectare and the exports reached 18.15 million tonnes with the earning of RM 45, 269.23 million has been recorded (MPOB, 2014). In the year of 2014, the production of palm oil is 19.67 million tonnes per hectare with an increase of 2.29 % compared to the previous year. The export of palm oil in 2014 reached 17.31 million tonnes earning RM 44, 498.45 million (MPOB, 2015). As for the year of 2015, the production of palm oil is 7.28 million tonnes per hectare and the exports reached 6.17 million tonnes with an earning of RM 14, 623.40 from January – May 2015 (MPOB, 2015). The demand for palm oil as a major source of vegetable oil increases every year and is assumed to reach up to 240 million tonnes by the year of 2050 (Corley, 2009). To maintain as one of the most important producer and exporter of palm oil, Malaysia tries to meet the future demands by producing clonal palms through biotechnology approaches.

The propagation by tissue culture was first described in the 1970's (Jones, 1974). Since then, the commercial advantage of tissue culture planting materials over conventional seedlings of oil palm has been well established (Sogeke, 1998). The whole process that involves the production of oil palm via tissue culture from initial level to the development of the matured oil palm tree and also including the testing part takes about 10 years. Tissue culture method enables the replication of individual high yielding oil palm in large scale in shorter time compared to the conventional method. Tissue culture usually produces offsprings identical to that of the original palm and desired trait such as higher oil or bunch is highly heritable and transferable to the next generation with slow vertical growth and disease resistance (Mutert et al, 1999). It has been proven that clonal palms are able to increase the oil yield compared to the commercial palms (Sharifah and Abu, 2007; Khaw and Ng, 1997; Donough and Lee, 1993).

Unfortunately, the tissue culture process sometimes generates somaclonal variants or abnormality in oil palm known as Truncated Leaf Syndrome (TLS). The mean frequency of this abnormality occurrence in a clonal palm is approximately 20 %. This abnormality is only obvious after few weeks of the plantlets are transferred from the

culture media to the nursery. The TLS ramets will show different characteristics compared to the normal ramets. The TLS ramets had stunted growth as well as produced undeveloped roots and the leaves looked like grasshopper damage which will eventually lead to the death of the TLS ramets. It is not known why only some ramets in an oil palm clone become TLS while others produce normal ramets. It is hypothesized that the presence of such phenotype could be due to genetic changes at the genomic level or epigenetic variability namely altered methylation level.

As such, the specific objectives of this study were to compare and identify possible genomic differences between TLS and normal oil palm ramets using the Genomic Representational Difference Analysis (G-RDA) method and to compare and identify possible methylation differences between TLS and normal oil palm ramets using the Methylation Sensitive Representational Difference Analysis (MS-RDA).

This study was specifically carried out to identify genetic and methylation differences associated to the TLS ramets. The identification of such differences could serve as a marker to screen for the TLS ramets at the early stage of the oil palm tissue culture process. For this study, two different approaches were carried out to identify the differences between TLS and normal ramets. The subtraction of the TLS genome with the genome of normal ramet was performed to identify the differences in the genomic fragments using the G-RDA and MS-RDA approach. Selected oil palm sequences from G-RDA were subjected to Polymerase Chain Reaction (PCR) analysis, sequencing and southern analysis while those sequences selected from MS-RDA were investigated on its expression profile using Real-Time PCR analysis.

The genomic fragments isolated from G-RDA and MS-RDA approach may correspond to the genetic or epigenetic changes that might cause the differences between the genome of TLS and normal ramets. Therefore, these fragments have the potential to be used as molecular markers for differentiating the TLS ramets from normal ramets in the early stage of oil palm tissue culture process. The detection of genetic or epigenetic differences between TLS and normal oil palm ramets could improve the oil palm tissue culture efficiency which will reduce cost, labour and time. Once a promising molecular marker has been established, oil palm tissue culture becomes efficient in producing economically valuable clonal palm that eventually increases the yield of palm oil.

REFERENCES

- Abdullah, R., Zainal, A., Yew Heng, W., Chui Li, L., Chee Beng, Y., Mei Phing, L., ... & Azma Jusoh, S. (2005). Immature embryo: A useful tool for oil palm (Elaeis guineensis Jacq.) genetic transformation studies. *Electronic Journal of Biotechnology*, 8(1), 24-34.
- Allen, N. L., Hilton, A. C., Betts, R., & Penn, C. W. (2001). Use of representational difference analysis to identify Escherichia coli O157-specific DNA sequences. *FEMS microbiology letters*, 197(2), 195-201.
- Ayyanathan, K., Francis, V. S. N. K., Datta, S., & Padmanaban, G. (1995). Development of specific DNA probes and their usage in the detection of Plasmodium vivax infection in blood. *Molecular and cellular probes*, 9(4), 239-246.
- Bairu, M. W., Aremu, A. O., & Van Staden, J. (2011). Somaclonal variation in plants: causes and detection methods. *Plant Growth Regulation*, 63(2), 147-173.
- Bairu, M. W., Fennell, C. W., & van Staden, J. (2006). The effect of plant growth regulators on somaclonal variation in Cavendish banana (Musa AAA cv. 'Zelig'). *Scientia horticulturae*, 108(4), 347-351.
- Balzon, T. A., Luis, Z. G., & Scherwinski-Pereira, J. E. (2013). New approaches to improve the efficiency of somatic embryogenesis in oil palm (Elaeis guineensis Jacq.) from mature zygotic embryos. *In Vitro Cellular & Developmental Biology-Plant*, 49(1), 41-50.
- Baránek, M., Křižan, B., Ondrušíková, E., & Pidra, M. (2010). DNA-methylation changes in grapevine somaclones following in vitro culture and thermotherapy. *Plant Cell, Tissue and Organ Culture (PCTOC), 101*(1), 11-22.
- Bart, A., Dankert, J., & van der Ende, A. (2000). Representational difference analysis of Neisseria meningitidis identifies sequences that are specific for the hyper-virulent lineage III clone. *FEMS microbiology letters*, 188(2), 111-114.
- Basiron, Y. (2007). Palm oil production through sustainable plantations. *European Journal of Lipid Science and Technology*, 109(4), 289-295.
- Behjati, S., & Tarpey, P. S. (2013). What is next generation sequencing? *ADC Education & Practice Edition*, 98(6), 236-238.
- Beisson, F., Li, Y., Bonaventure, G., Pollard, M., & Ohlrogge, J. B. (2007). The acyltransferase GPAT5 is required for the synthesis of suberin in seed coat and root of Arabidopsis. The Plant Cell Online, 19(1), 351-368.
- Beló, A., Zheng, P., Luck, S., Shen, B., Meyer, D. J., Li, B., ... & Rafalski, A. (2008). Whole genome scan detects an allelic variant of fad2 associated with increased oleic acid levels in maize. *Molecular Genetics and Genomics*, 279(1), 1-10.

- Benfey, P. N., Linstead, P. J., Roberts, K., Schiefelbein, J. W., Hauser, M. T., & Aeschbacher, R. A. (1993). Root development in Arabidopsis: four mutants with dramatically altered root morphogenesis. Development, 119(1), 57-70.
- Besse, I., Verdeil, J. L., Duval, Y., Sotta, B., Maldiney, R., & Miginiac, E. (1992). Oil palm (*Elaeis guineensis* Jacq.) clonal fidelity: endogenous cytokinins and indoleacetic acid in embryogenic calluscultures. *Journal of Experimental Botany*, 43(7), 983-989.
- Bhojwani, S. S., & Dantu, P. K. (2013). Somaclonal variation. In *Plant Tissue Culture: An Introductory Text* (pp. 141-154). Springer India.
- Birkenmeyer, L. G., Leary, T. P., Muerhoff, A. S., Dawson, G. J., Mushahwar, I. K., & Desai, S. M. (2003). Selectively primed adaptive driver RDA (SPAD-RDA): An improved method for subtractive hybridization. *Journal of medical virology*, 71(1), 150-159.
- Blanc-Potard, A. B., Tinsley, C., Scaletsky, I., Le Bouguenec, C., Guignot, J., Servin, A. L., ... Bernet-Camard, M. F. (2002). Representational difference analysis between Afa/Dr diffusely adhering *Escherichia coli* and nonpathogenic E. coli K-12. *Infection and immunity*, 70(10), 5503-5511.
- Bossdorf, O., Richards, C. L., & Pigliucci, M. (2008). Epigenetics for ecologists. *Ecology letters*, 11(2), 106-115.
- Bowler, L. D., Hubank, M., & Spratt, B. G. (1999). Representational difference analysis of cDNA for the detection of differential gene expression in bacteria: development using a model of iron-regulated gene expression in Neisseria meningitidis. *Microbiology*, 145(12), 3529-3537.
- Brower, V. (2011). Biomarkers: Portents of malignancy. *Nature*, 471(7339), S19-S21.
- Brown, P. T. H., Göbel, E., & Lörz, H. (1991). RFLP analysis of Zea mays callus cultures and their regenerated plants. *Theoretical and applied genetics*, 81(2), 227-232.
- Brown, P. T. H., Kyozuka, J., Sukekiyo, Y., Kimura, Y., Shimamoto, K., & Lörz, H. (1990). Molecular changes in protoplast-derived rice plants. *Molecular and General Genetics MGG*, 223(2), 324-328.
- Buzdin, A. A. (2007). Nucleic Acids Hybridization: Potentials and Limitations. In *Nucleic Acids Hybridization Modern Applications* (pp. 1-28). Springer Netherlands.
- Calia, K. E., Waldor, M. K., & Calderwood, S. B. (1998). Use of representational difference analysis to identify genomic differences between pathogenic strains of Vibrio cholerae. *Infection and immunity*, 66(2), 849-852.
- Cecchini, E., Natali, L., Cavallini, A., & Durante, M. (1992). DNA variations in regenerated plants of pea (Pisum sativum L.). *Theoretical and Applied Genetics*, 84(7-8), 874-879.

- Chang, Y., Cesarman, E., Pessin, M. S., Lee, F., Culpepper, J., Knowles, D. M., & Moore, P. S. (1994). Identification of herpesvirus-like DNA sequences in AIDSassociated Kaposi's sarcoma. *Science*, 266(5192), 1865-1869.
- Chen, Z. J., Phillips, R. L., & Rines, H. W. (1998). Maize DNA enrichment by representational difference analysis in a maize chromosome addition line of oat. *Theoretical and applied genetics*, *97*(3), 337-344.
- Cho, T. J., & Park, S. S. (1998). A simulation of subtractive hybridization. *Nucleic Acids Research*, 26(6), 1440-1448.
- Choi, C. S., & Sano, H. (2007). Abiotic-stress induces demethylation and transcriptional activation of a gene encoding a glycerophosphodiesterase-like protein in tobacco plants. *Molecular Genetics and Genomics*, 277(5), 589-600.
- Chuang, S. J., Chen, C. L., Chen, J. J., Chou, W. Y., & Sung, J. M. (2009). Detection of somaclonal variation in micro-propagated Echinacea purpurea using AFLP marker. *Scientia Horticulturae*, 120(1), 121-126.
- Cochard, B., Durand-Gasselin, T., Amblard, P., Konan, E. K., & Gogor, S. (2000). Performance of adult oil palm clones. In *Emerging technologies and opportunities in the next millennium*. Agriculture conference: proceedings of 1999 PORIM International Palm Oil Congress (PIPOC 1999), Kuala Lumpur, Malaysia (pp. 53-64).
- Collins, F. S. (1992). Positional cloning: let's not call it reverse anymore. *Nature Genetics*, 1(1), 3-6.
- Corley, R. H. V. (2009). How much palm oil do we need? *Environmental Science & Policy*, 12(2), 134-139.
- Corley, R. H. V., & Teo, C. (1976). Disbudding of mature oil palms as a method of controlling yield fluctuation. *MARDI Research Bulletin*, 4(1), 1-6.
- Corley, R. H. V., & Tinker, P. B. (2003). *The classification and morphology of the oil palm*. The Oil Palm 4th Edn. BS Ltd. Blackwell Publishing, (pp. 27-51).
- Corley, R. H. V., & Tinker, P. B. (2003). Vegetative propagation and biotechnology. *The oil palm*, *4*, 201-215.
- Corley, R. H. V., Barrett, J. N., & Jones, L. H. (1977). Vegetative propagation of oil palm. In *International Developments in Oil Palm; Proceedings of the Malaysian International Agricultural Oil Palm Conference*. Inc. Soc. of Planters, Kuala Lumpur, (pp. 1-8).
- Corley, R. H. V., Lee, C. H., Law, L. H., & Wong, C. Y. (1986). Abnormal flower development in oil palm clones. *Planter (KualaLumpur)* 62, 233-240.
- Cornelius, J. A. (1977). Palm oil and palm kernel oil. *Progress in the Chemistry of Fats and other Lipids*, 15(1), 5-27.

- Corniquel, B., & Mercier, L. (1994). Date palm (Phoenix dactylifera L.) cultivar identification by RFLP and RAPD. *Plant Science*, *101*(2), 163-172.
- Cullis CA, Cullis MA, Ong Abdullah M (2007) Development of markers for the mantled phenotype in oil palm. In: PIPOC 2007. Proceedings agriculture, biotechnology and sustainability conference. Kuala Lumpur, (pp 299–312).
- Cullis, C. A., & Kunert, K. (1999). Isolation of tissue culture-induced polymorphisms in bananas by representational difference analysis. In *International Symposium on Methods and Markers for Quality Assurance in Micropropagation* 530 (pp. 421-428).
- Dahl, C., & Guldberg, P. (2003). DNA methylation analysis techniques. *Biogerontology*, 4(4), 233-250.
- Damasco, O. P., Graham, G. C., Henry, R. J., Adkins, S. W., Smiths, M. K., & Godwin, I. D. (1996). Random amplified polymorphic DNA (RAPD) detection of dwarf off-types in micropropagated Cavendish (Musa spp. AAA) bananas. *Plant Cell Reports*, 16(1-2), 118-123.
- d'Amato, F. (1977). Cytogenetics of differentiation in tissue and cell cultures. *Applied and fundamental aspects of plant cell, tissue and organ culture*, 343-357.
- De Touchet, B., Duval, Y., & Pannetier, C. (1991). Plant regeneration from embryogenic suspension cultures of oil palm (Elaeis guineensis Jacq.). *Plant Cell Reports*, 10(10), 529-532.
- Dellaporta, S. L. (1983). A plant DNA minipreparation: Version II. *Plant Molecular Biology Reporter*, 19-21.
- Dhar, M. S., Pethe, V. V., Gupta, V. S., & Ranjekar, P. K. (1990). Predominance and tissue specificity of adenine methylation in rice. *Theoretical and applied genetics*, 80(3), 402-408.
- Donnelly, P. (2008). Progress and challenges in genome-wide association studies in humans. *Nature*, 456(7223), 728-731.
- Donnison, I. S., Siroky, J., Vyskot, B., Saedler, H., & Grant, S. R. (1996). Isolation of Y chromosome-specific sequences from Silene latifolia and mapping of male sex-determining genes using representational difference analysis. *Genetics*, 144(4), 1893-1901.
- Donough, C. R., & Lee, C. H. (1993). Longer term results from clone trials at PAMOL Plantations and Golden Hope Plantations. In *Recent Developments in Oil Palm Tissue Culture and Biotechnology, Eds. V. Rao, IE Henson and N. Rajanaidu, Palm Oil Res. Inst. Malaysia, Kuala Lumpur* (pp. 16-133).
- Duncan, R. R. (1997). Tissue culture-induced variation and crop improvement. *Advances in agronomy*, 58, 201-240.

- Dunwell, J. M. (2010). Haploids in flowering plants: origins and exploitation. *Plant Biotechnology Journal*, 8(4), 377-424.
- Eeuwens, C. J. (1976). Mineral requirements for growth and callus initiation of tissue explants excised from mature coconut palms (Cocos nucifera) and cultured in vitro. *Physiologia Plantarum*, *36*(1), 23-28.
- Eeuwens, C. J., & Blake, J. (1977). Culture of coconut and date palm tissue with a view to vegetative propagation. In *Symposium on Tissue Culture for Horticultural Purposes* 78 (pp. 277-286).
- Eeuwens, C. J., Lord, S., Donough, C. R., Rao, V., Vallejo, G., & Nelson, S. (2002). Effects of tissue culture conditions during embryoid multiplication on the incidence of "mantled" flowering in clonally propagated oil palm. *Plant Cell, Tissue and Organ Culture*, 70(3), 311-323.
- Evans, D. A. (1989). Somaclonal variation-genetic basis and breeding applications. *Trends in genetics*, 5, 46-50.
- Finnegan, E. J., Brettell, R. I. S., & Dennis, E. S. (1993). The role of DNA methylation in the regulation of plant gene expression. In DNA Methylation (pp. 218-261). Birkhäuser Basel.
- Forero, D. C., Hormaza, P., & Romero, H. M. (2012). Phenological growth stages of African oil palm (*Elaeis guineensis*). *Annals of Applied Biology*, 160(1), 56-65.
- Fraga, M. F., Ballestar, E., Paz, M. F., Ropero, S., Setien, F., Ballestar, M. L., ... & Esteller, M. (2005). Epigenetic differences arise during the lifetime of monozygotic twins. *Proceedings of the National Academy of Sciences of the United States of America*, 102(30), 10604-10609.
- Fukaki, H., Fujisawa, H., & Tasaka, M. (1996). SGR1, SGR2, and SGR3: novel genetic loci involved in shoot gravitropism in Arabidopsis thaliana. Plant Physiology, 110(3), 945-955.
- Gerritsma, W., & Soebagyo, F. X. (1999). An analysis of the growth of leaf area of oil palms in Indonesia. *Experimental agriculture*, *35*(03), 293-308.
- Giménez, C., de García, E., de Enrech, N. X., & Blanca, I. (2001). Somaclonal variation in banana: cytogenetic and molecular characterization of the somaclonal variant CIEN BTA-03. *In Vitro Cellular & Developmental Biology-Plant*, *37*(2), 217-222.
- Giorgetti, L., Vergara, M. R., Evangelista, M., Schiavo, F. L., Terzi, M., & Ronchi, V. N. (1995). On the occurrence of somatic meiosis in embryogenic carrot cell cultures. *Molecular and General Genetics MGG*, 246(6), 657-662.
- Giulietti, A., Overbergh, L., Valckx, D., Decallonne, B., Bouillon, R., & Mathieu, C. (2001). An overview of real-time quantitative PCR: applications to quantify cytokine gene expression. *Methods*, 25(4), 386-401.

- Grad, Y. H., Lipsitch, M., Feldgarden, M., Arachchi, H. M., Cerqueira, G. C., FitzGerald, M., ... & Hanage, W. P. (2012). Genomic epidemiology of the Escherichia coli O104: H4 outbreaks in Europe, 2011. Proceedings of the National Academy of Sciences, 109(8), 3065-3070.
- Grafi, G., Chalifa-Caspi, V., Nagar, T., Plaschkes, I., Barak, S., & Ransbotyn, V. (2011). Plant response to stress meets dedifferentiation. *Planta*, *233*(3), 433-438.
- Habib, S. H., Ooi, S. E., Novák, O., Tarkowská, D., Rolčík, J., Doležal, K., ... & Namasivayam, P. (2012). Comparative mineral and hormonal analyses of wild type and TLS somaclonal variant derived from oil palm (Elaeis guineensis Jacq. var. tenera) tissue culture. *Plant Growth Regulation*, 68(2), 313-317.
- Habib, S. H., Syed-Alwee, S. S. R., Ho, C. L., Ong-Abdullah, M., Sinniah, U. R., & Namasivayam, P. (2012). Morpho-histological characterization of truncated leaf syndrome seedlings: an oil palm (E. guineensis Jacq.) somaclonal variant. *Acta Physiologiae Plantarum*, 34(1), 17-28.
- Hänsch, R., & Mendel, R. R. (2009). Physiological functions of mineral micronutrients (Cu, Zn, Mn, Fe, Ni, Mo, B, Cl). *Current opinion in plant biology*, 12(3), 259-266.
- Harding, K. (1994). The methylation status of DNA derived from potato plants recovered from slow growth. *Plant cell, tissue and organ culture, 37*(1), 31-38.
- Hartley, C. W. S. (1988). The oil palm. Tropical Agriculture Series.
- Hartley, C. W. S. (2000). *The Oil Palm*. 3rd Edn., Longman, London and New York, pages: 806.
- Hartly, C. W. S. (1988). *The Oil Palm (Elaeis guineensis* Jacq.) 3rd eds, pp. 47-94. Longman Scientific and Technical, London.
- Helgadottir, A., Thorleifsson, G., Manolescu, A., Gretarsdottir, S., Blondal, T., Jonasdottir, A., ... & Stefansson, K. (2007). A common variant on chromosome 9p21 affects the risk of myocardial infarction. *Science*, *316*(5830), 1491-1493.
- Henderson, I. R., & Dean, C. (2004). Control of Arabidopsis flowering: the chill before the bloom. *Development*, 131(16), 3829-3838.
- Henderson, J., & Osborne, D. J. (2000). The oil palm in all our lives: how this came about. *Endeavour*, 24(2), 63-68.
- Hirschhorn, J. N., & Daly, M. J. (2005). Genome-wide association studies for common diseases and complex traits. *Nature Reviews Genetics*, 6(2), 95-108.
- Hubank, M., & Schatz, D. G. (1994). Identifying differences in mRNA expression by representational difference analysis of cDNA. *Nucleic acids research*, 22(25), 5640-5648.
- Ithnin, M., Singh, R., & Din, A. K. (2011). Elaeis. In *Wild Crop Relatives: Genomic and Breeding Resources* (pp. 113-124). Springer Berlin Heidelberg.

- Itoh, J. I., Kitano, H., Matsuoka, M., & Nagato, Y. (2000). SHOOT ORGANIZATION genes regulate shoot apical meristem organization and the pattern of leaf primordium initiation in rice. *The Plant Cell*, *12*(11), 2161-2174.
- Jablonka, E., & Raz, G. (2009). Transgenerational epigenetic inheritance: prevalence, mechanisms, and implications for the study of heredity and evolution. *The Quarterly review of biology*, 84(2), 131-176.
- Jaligot, E., Rival, A., Beule, T., Dussert, S., & Verdeil, J. L. (2000). Somaclonal variation in oil palm (Elaeis guineensis Jacq.): the DNA methylation hypothesis. *Plant Cell Reports*, 19(7), 684-690.
- Johannes, F., Porcher, E., Teixeira, F. K., Saliba-Colombani, V., Simon, M., Agier, N., ... & Colot, V. (2009). Assessing the impact of transgenerational epigenetic variation on complex traits. *PLoS Genetics*, *5*(6), e1000530-e1000530.
- Jones, L. H. (1974). Propagation of clonal oil palms by tissue culture. Oil Palm News, (17), 1-8.
- Jones, L. H., 1974. Propagation of clonal oil palms by tissue culture. *Oil Palm News* 17, 1-8.
- Joubès, J., Raffaele, S., Bourdenx, B., Garcia, C., Laroche-Traineau, J., Moreau, P., ... Lessire, R. (2008). The VLCFA elongase gene family in Arabidopsis thaliana: phylogenetic analysis, 3D modelling and expression profiling. Plant molecular biology, 67(5), 547-566.
- Kaeppler, S. M., & Phillips, R. L. (1993). Tissue culture-induced DNA methylation variation in maize. *Proceedings of the National Academy of Sciences*, 90(19), 8773-8776.
- Kaeppler, S. M., Kaeppler, H. F., & Rhee, Y. (2000). Epigenetic aspects of somaclonal variation in plants. *Plant molecular biology*, 43(2-3), 179-188.
- Kaeppler, S.M. 1992. Molecular and genetic studies of tissue-culture induced variation in maize. PhD thesis. St.Paul University, St. Paul, Minnesota.
- Kakutani, T., Jeddeloh, J. A., Flowers, S. K., Munakata, K., & Richards, E. J. (1996). Developmental abnormalities and epimutations associated with DNA hypomethylation mutations. *Proceedings of the National Academy of Sciences*, 93(22), 12406-12411.
- Kanchanapoom, K., & Domyoas, P. (1999). The origin and development of embryoids in oil palm (*Elaeis guineensis* Jacq) embryo culture. *ScienceAsia*, 25(4).
- Karp, A. (1994). Origins, causes and uses of variation in plant tissue cultures. In *Plant cell and tissue culture* (pp. 139-151). Springer Netherlands.
- Kawiak, A., & Lojkowska, E. (2004). Application of RAPD in the determination of genetic fidelity in micropropagated Drosera plantlets. *In Vitro Cellular & Developmental Biology-Plant*, 40(6), 592-595.

- Kelsey, G., Bodle, D., Miller, H. J., Beechey, C. V., Coombes, C., Peters, J., & Williamson, C. M. (1999). Identification of imprinted loci by methylation-sensitive representational difference analysis: application to mouse distal chromosome 2. *Genomics*, 62(2), 129-138.
- Khaw, C. H., & Ng, S. K. (1997). Performance of commercial scale clonal oil palm (*Elaeis guineensis* Jacq.) plantings in Malaysia. In *International Symposium on Biotechnology of Tropical and Subtropical Species Part 2 461* (pp. 251-258).
- Koornneef, M., Cone, J. W., Karssen, C. M., Kendrick, R. E., Van der Veen, J. H., & Zeevaart, J. A. D. (1985). Plant hormone and photoreceptor mutants in arabicopsis and tomato. In UCLA symposia on molecular and cellular biology (USA).
- Kovar, A., Koukalova, B., Bezde, M., & Opatrn, Z. (1997). Hypermethylation of tobacco heterochromatic loci in response to osmotic stress. *Theoretical and Applied Genetics*, 95(1-2), 301-306.
- Kubis, S. E., Castilho, A. M., Vershinin, A. V., & Heslop-Harrison, J. S. P. (2003). Retroelements, transposons and methylation status in the genome of oil palm (Elaeis guineensis) and the relationship to somaclonal variation. *Plant molecular biology*, 52(1), 69-79.
- Kushairi, A., Rajanaidu, N., Jalani, B. S., & Isa, Z. A. (1999). PORIM Series 1-PORIM elite oil palm planting materials. *PORIM Information Series*, (100).
- Kwong, J. C., McCallum, N., Sintchenko, V., & Howden, B. P. (2015). Whole genome sequencing in clinical and public health microbiology. *Pathology*, 47(3), 199.
- Lammens, E., Ceyssens, P. J., Voet, M., Hertveldt, K., Lavigne, R., & Volckaert, G. (2009). Representational Difference Analysis (RDA) of bacteriophage genomes. *Journal of microbiological methods*, 77(2), 207-213.
- Larkin, P. J., & Scowcroft, W. R. (1981). Somaclonal variation—a novel source of variability from cell cultures for plant improvement. *Theoretical and Applied Genetics*, 60(4), 197-214.
- Le, V. T. (2009). Molecular Characterization and Functional Analysis of Selected Expressed Sequence Tags from Oil Palm Cell Suspension Culture (Doctoral dissertation, Universiti Putra Malaysia).
- Lee, M., & Phillips, R. L. (1987). Genetic variants in progeny of regenerated maize plants. *Genome*, 29(6), 834-838.
- Lee, M., & Phillips, R. L. (1988). The chromosomal basis of somaclonal variation. *Annual Review of Plant Physiology and Plant Molecular Biology*, 39(1), 413-437.
- Lee, S. H., Shon, Y. G., Kim, C. Y., Chun, H. J., Cheong, Y. H., Kim, Z. H., ... & Cho, M. J. (1999). Variations in the morphology of rice plants regenerated from protoplasts using different culture procedures. *Plant cell, tissue and organ culture*, 57(3), 179-187.

- Lei, C. P., Jiun, K. S., Choo, C. S., & Singh, R. (2006). Analysis of tissue culturederived regenerants using methylation sensitive AFLP. Asia Pacific Journal of Molecular Biology and Biotechnology, 14, 47-55.
- Leljak-Levanić, D., Bauer, N., Mihaljević, S., & Jelaska, S. (2004). Somatic embryogenesis in pumpkin (Cucurbita pepo L.): control of somatic embryo development by nitrogen compounds. *Journal of plant physiology*, 161(2), 229-236.
- Leva, A. R., Rinaldi, L. M. R., & Petruccelli, R. (2012). Somaclonal variation in tissue culture: a case study with olive. INTECH Open Access Publisher.
- Li, H. M., Chen, H., Yang, Z. N., & Gong, J. M. (2012). Cdi gene is required for pollen germination and tube growth in Arabidopsis. FEBS letters, 586(7), 1027-1031.
- Li, X., Xu, M., & Korban, S. S. (2002). DNA methylation profiles differ between field-andin vitro-grown leaves of apple. *Journal of Plant Physiology*, 159(11), 1229-1234.
- Li, Y., Beisson, F., Pollard, M., & Ohlrogge, J. (2006). Oil content of Arabidopsis seeds: the influence of seed anatomy, light and plant-to-plant variation. Phytochemistry, 67(9), 904-915.
- Lim, C. C., Teo, K. W., Rao, V., & Chia, C. C. (2003). Performances of some pisiferas of Binga, Ekona, URT and Angolan origins: Part 1-Breeding background and fruit bunch traits. *Journal of Oil Palm Research*, 15(1), 21-31.
- Lippman, Z., Gendrel, A. V., Black, M., Vaughn, M. W., Dedhia, N., McCombie, W. R., ... Martienssen, R. (2004). Role of transposable elements in heterochromatin and epigenetic control. *Nature*, 430(6998), 471-476.
- Lisitsyn, N. A. (1995). Representational difference analysis: finding the differences between genomes. *Trends in Genetics*, 11(8), 303-307.
- Lisitsyn, N., & Wigler, M. (1993). Cloning the differences between two complex genomes. *Science*, 259(5097), 946-951.
- Lisitsyn, N., & Wigler, M. (1995). Representational difference analysis in detection of genetic lesions in cancer. *Methods in enzymology*, 254, 291-304.
- Lord, E. M., & Russell, S. D. (2002). The mechanisms of pollination and fertilization in plants. Annual Review of Cell and Developmental Biology, 18(1), 81-105.
- LoSchiavo, F., Pitto, L., Giuliano, G., Torti, G., Nuti-Ronchi, V., Marazziti, D., ... Terzi, M. (1989). DNA methylation of embryogenic carrot cell cultures and its variations as caused by mutation, differentiation, hormones and hypomethylating drugs. *Theoretical and Applied Genetics*, 77(3), 325-331.
- Low, E. T. L., Alias, H., Boon, S. H., Shariff, E. M., Tan, C. Y. A., Ooi, L. C., ... & Singh, R. (2008). Oil palm (Elaeis guineensis Jacq.) tissue culture ESTs: identifying genes associated with callogenesis and embryogenesis. *BMC Plant Biology*, 8(1), 62.

- Lund, G., Ciceri, P., & Viotti, A. (1995). Maternal-specific demethylation and expression of specific alleles of zein genes in the endosperm of Zea mays L. *The Plant Journal*, 8(4), 571-581.
- Magyar-Tábori, K., Dobránszki, J., da Silva, J. A. T., Bulley, S. M., & Hudák, I. (2010). The role of cytokinins in shoot organogenesis in apple. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 101(3), 251-267.
- Manolio, T. A., Brooks, L. D., & Collins, F. S. (2008). A HapMap harvest of insights into the genetics of common disease. *The Journal of Clinical Investigation*, 118(5), 1590.
- Marsh, I. B., & Whittington, R. J. (2005). Deletion of an mmpL gene and multiple associated genes from the genome of the S strain of Mycobacterium avium subsp. paratuberculosis identified by representational difference analysis and in silico analysis. *Molecular and cellular probes*, 19(6), 371-384.
- Martin, K. P., Pachathundikandi, S. K., Zhang, C. L., Slater, A., & Madassery, J. (2006). RAPD analysis of a variant of banana (Musa sp.) cv. grande naine and its propagation via shoot tip culture. *In Vitro Cellular & Developmental Biology-Plant*, 42(2), 188-192.
- Martinelli, L. U. C. I. A., Zambanini, J. E. S. S. I. C. A., & Grando, M. S. (2004). Genotype assessment of grape regenerants from floral explants. *VITIS-GEILWEILERHOF-*, 43(3), 119-122.
- Maziah, M., Zuraida, A. R., Halimi, M. S., Zulkifli, H. S., & Sreeramanan, S. (2010). Influence of boron on the growth and biochemical changes in plant growth promoting rhizobacteria (PGPR) inoculated banana plantlets. *World Journal of Microbiology and Biotechnology*, 26(5), 933-944.
- McClintock, B. (1950). The origin and behavior of mutable loci in maize. *Proceedings* of the National Academy of Sciences, 36(6), 344-355.
- Mengel, K., Kosegarten, H., Kirkby, E. A., & Appel, T. (Eds.). (2001). Principles of plant nutrition. Springer Science & Business Media.
- Messeguer, R., Ganal, M. W., Steffens, J. C., & Tanksley, S. D. (1991). Characterization of the level, target sites and inheritance of cytosine methylation in tomato nuclear DNA. *Plant molecular biology*, *16*(5), 753-770.
- Meyer, K., Leube, M. P., & Grill, E. (1994). A protein phosphatase 2C involved in ABA signal transduction in Arabidopsis thaliana. *Science*, 264(5164), 1452-1455.
- Mgbeze, G. C., & Iserhienrhien, A. (2014). Somaclonal variation associated with oil palm (Elaeis guineensis Jacq.) clonal propagation: a review. *African Journal of Biotechnology*, 13(9), 989-997.
- Michiels, L., Van Leuven, F., Van den Oord, J. J., De Wolf-Peeters, C., & Delabie, J. (1998). Representational ifference analysis using minute quantities of DNA. *Nucleic acids research*, 26(15), 3608-3610.

- Middendorf, B., & Gross, R. (1999). Representational difference analysis identifies a strain-specific LPS biosynthesis locus in Bordetella spp. *Molecular and General Genetics MGG*, 262(1), 189-198.
- Milner, J. J., Cecchini, E., & Dominy, P. J. (1995). A kinetic model for subtractive hybridization. *Nucleic acids research*, 23(1), 176-187.
- Morcillo, F., Gagneur, C., Adam, H., Richaud, F., Singh, R., Cheah, S. C., ... Tregear, J. W. (2006). Somaclonal variation in micropropagated oil palm. Characterization of two novel genes with enhanced expression in epigenetically abnormal cell lines and in response to auxin. *Tree physiology*, 26(5), 585-594.
- MPOB (2014). *Economics and Industry Development Division*. Retrieved on December 12, 2014 from bepi.mpob.gov.my
- MPOB (2015). *Economics and Industry Development Division*. Retrieved on June 22, 2015 from bepi.mpob.gov.my
- Muerhoff, A. S., Leary, T. P., Desai, S. M., & Mushahwar, I. K. (1997). Amplification and subtraction methods and their application to the discovery of novel human viruses. *Journal of medical virology*, 53(1), 96-103.
- Mujib, A., Banerjee, S., & Dev Ghosh, P. (2007). Callus induction, somatic embryogenesis and chromosomal instability in tissue culture-raised hippeastrum (Hippeastrum hybridum cv. United Nations). *Propag Ornam Plants*, 7, 169-174.
- Murphy, D. J. (2007). Future prospects for oil palm in the 21st century: Biological and related challenges. *European Journal of Lipid Science and Technology*, 109(4), 296-306.
- Mutert, E., & Fairhurst, T. H. (1999). Oil palm clones: Productivity enhancement for the future. *Better Crops International*, 13(1), 45.
- Neelakandan, A. K., & Wang, K. (2012). Recent progress in the understanding of tissue culture-induced genome level changes in plants and potential applications. *Plant Cell Reports*, 31(4), 597-620.
- Nehra, N. S., Kartha, K. K., Stushnott, C., & Giles, K. L. (1992). The influence of plant growth regulator concentrations and callus age on somaclonal variation in callus culture regenerants of strawberry. *Plant cell, tissue and organ culture,* 29(3), 257-268.
- Niizeki, M. (1974). Studies on Plant Cell and Tissue Culture: V. Effect of different kinds of media on the variation of chromosome numbers in tobacco callus and regenerated plant. *Journal of the Faculty of Agriculture, Hokkaido University*, 57(4), 357-367.

- Nur Fatihah, M. Y., Sharifah Shahrul, R. S. A., Abdullah, M. O., ChaiLing, H., & Namasivayam, P. (2012). A time course anatomical analysis of callogenesis from young leaf explants of oil palm (Elaeis guineensis Jacq.). *Journal of Oil Palm Research*, 24, 1330-1341.
- Nwankwo, B. A., & Krikorian, A. D. (1983). Morphogenetic potential of embryo-and seedling-derived callus of Elaeis guineensis Jacq. var. pisifera Becc. *Annals of Botany*, *51*(1), 65-76.
- Oh, T. J., Cullis, M. A., Kunert, K., Engelborghs, I., Swennen, R., & Cullis, C. A. (2007). Genomic changes associated with somaclonal variation in banana (Musa spp.). *Physiologia plantarum*, 129(4), 766-774.
- O'hanlon, P. C., Peakall, R., & Briese, D. T. (2000). A review of new PCR-based genetic markers and their utility to weed ecology. WEED RESEARCH-OXFORD-, 40(3), 239-254.
- Panaud, O., Vitte, C., Hivert, J., Muzlak, S., Talag, J., Brar, D., & Sarr, A. (2002). Characterization of transposable elements in the genome of rice (Oryza sativa L.) using representational difference analysis (RDA). *Molecular Genetics and Genomics*, 268(1), 113-121.
- Paranjothy, K., & Rohani, O. (1982). In vitro propagation of oil palm. *Plant tissue culture*, 747-748.
- Paranjothy, K., Saxena, S., Banerjee, M., Jaiswal, V. S., & Bhojwani, S. S. (1990). Clonal multiplication of woody perennials. *Developments in crop science*, 19, 190-219.
- Pastorian, K., Hawel, L., & Byus, C. V. (2000). Optimization of cDNA representational difference analysis for the identification of differentially expressed mRNAs. *Analytical biochemistry*, 283(1), 89-98.
- Pathak, H., & Dhawan, V. (2012). ISSR assay for ascertaining genetic fidelity of micropropagated plants of apple rootstock Merton 793. *In Vitro Cellular & Developmental Biology-Plant*, 48(1), 137-143.
- Peraza-Echeverria, S., Herrera-Valencia, V. A., & Kay, A. J. (2001). Detection of DNA methylation changes in micropropagated banana plants using methylation-sensitive amplification polymorphism (MSAP). *Plant Science*, *161*(2), 359-367.
- Peschke, V. M., & Phillips, R. L. (1991). Activation of the maize transposable element Suppressor-mutator (Spm) in tissue culture. *Theoretical and applied genetics*, 81(1), 90-97.
- Phillips, R. L., Kaeppler, S. M., & Olhoft, P. (1994). Genetic instability of plant tissue cultures: breakdown of normal controls. *Proceedings of the National Academy of Sciences*, 91(12), 5222-5226.

- Pilipenko, T. I., Solov'eva, N. S. (1979). Accumulation of zinc in tissues of Phaseolus vulgaris plants supplied with different boron rates. *Vestnik Khar' kovskogo Universiteta*, 189, 71-74
- Pluhar, S. A., Erickson, L., & Pauls, K. P. (2001). Effects of tissue culture on a highly repetitive DNA sequence (E180 satellite) in Medicago sativa. *Plant cell, tissue and organ culture*, 67(2), 195-199.
- Prado, M. J., Gonzalez, M. V., Romo, S., & Herrera, M. T. (2007). Adventitious plant regeneration on leaf explants from adult male kiwifruit and AFLP analysis of genetic variation. *Plant cell, tissue and organ culture,* 88(1), 1-10.
- Purand-Gasselin, T., Konan, K. E., Bondonin, L., & Noiret, J. M. (1999). In Performance of DXP. *Interspecific Hybrids and Clones*, 151-170.
- Pysh, L. D., Wysocka-Diller, J. W., Camilleri, C., Bouchez, D., & Benfey, P. N. (1999). The GRAS gene family in Arabidopsis: Sequence characterization and basic expression analysis of the SCARECROW-LIKE genes. The Plant Journal, 18(1), 111-119.
- Rajanaidu, N., & Rao, V. (1988). Oil palm genetic collections: their performance and use to the industry. In *Proceedings of the 1987 International Oil Palm Conference Agriculture*. Palm Oil Res. Inst, Malaysia, Kuala Lumpur, (pp. 59-85).
- Rajanaidu, N., Rohani, O., & Jalani, B. S. (1997). Oil palm clones: current status and prospects for commercial production. *Planter*, 73(853), 163-184.
- Reynolds, J. F., & Murashige, T. (1979). Asexual embryogenesis in callus cultures of palms. *In vitro*, 15(5), 383-387.
- Richards, E. J. (1997). DNA methylation and plant development. *Trends in Genetics*, 13(8), 319-323.
- Rochester, D. E., Winer, J. A., & Shah, D. M. (1986). The structure and expression of maize genes encoding the major heat shock protein, hsp70. The EMBO journal, 5(3), 451.
- Rohani, O., Sharifah, S. A., Rafii, M. Y., Ong, M., Tarmizi, A. H., & Zamzuri, I. (2000). Tissue culture of oil palm. *Advances in oil palm research*, 1, 238-283.
- Ronemus, M. J., Galbiati, M., Ticknor, C., Chen, J., & Dellaporta, S. L. (1996). Demethylation-induced developmental pleiotropy in Arabidopsis. *Science*, 273(5275), 654-657.
- Rossi, V., Motto, M., & Pellegrini, L. (1997). Analysis of the methylation pattern of the maize opaque-2 (O2) promoter and in vitro binding studies indicate that the O2 B-Zip protein and other endosperm factors can bind to methylated target sequences. *Journal of Biological Chemistry*, 272(21), 13758-13765.

- Roy, J. K., Smith, K. P., Muehlbauer, G. J., Chao, S., Close, T. J., & Steffenson, B. J. (2010). Association mapping of spot blotch resistance in wild barley. *Molecular Breeding*, 26(2), 243-256.
- Sato, T., & Mishina, M. (2003). Representational difference analysis, high-resolution physical mapping, and transcript identification of the zebrafish genomic region for a motor behavior. *Genomics*, 82(2), 218-229.
- Schellenbaum, P., Mohler, V., Wenzel, G., & Walter, B. (2008). Variation in DNA methylation patterns of grapevine somaclones (Vitis vinifera L.). *BMC plant biology*, 8(1), 78.
- Scheres, B., Di Laurenzio, L., Willemsen, V., Hauser, M. T., Janmaat, K., Weisbeek, P., & Benfey, P. N. (1995). Mutations affecting the radial organisation of the Arabidopsis root display specific defects throughout the embryonic axis. Development, 121(1), 53-62.
- Sedra, M. H., Lashermes, P., Trouslot, P., & Combes, M. C. (1998). Identification and genetic diversity analysis of date palm (Phoenix dactylifera L.) varieties from Morocco using RAPD markers. *Euphytica*, 103(1), 75-82.
- Shah, F. H., & Parveez, A. (1995). DNA variation in abnormal tissue culture regenerants of oil palm, Elaeis guineensis. *Asia-Pacific Journal of Molecular Biology and Biotechnology*, 3(1), 49-53.
- Sharifah, S. R. S. A., & Abu, Z. O. (2007). Achieving vision 35:25 through clonal planting. In *Proceeding of Agriculture, Biotechnology and Sustainability Conference* (International Palm Oil Congress), Kuala Lumpur Convention Centre. Malaysian Palm Oil Board, Kuala Lumpur (pp. 219-227).
- Sharma, V. K., Hänsch, R., Mendel, R. R., & Schulze, J. (2007). Node-derived cultures with high-morphogenic competence in barley and wheat. *Plant cell, tissue and organ culture*, 88(1), 21-33.
- Shchukin, A., Ben-Bassat, D., Israeli, Y. (1997) Plant regeneration via somatic embryogenesis in Grand Naine banana and its effect on somaclonal variation. *Acta Hort.* 447, 317-318.
- Shchukin, A., Ben-Bassat, D., Israeli, Y. (1998) Somaclonal variation and horticultural performance of 'Grand Naine' bananas multiplied via somatic embryogenesis or shoot-tip culture. Plant biotechnology and in vitro biology in the 21st century. International Association for Plant Tissue Culture, Jerusalem, pp. 14-19.
- Shiba, H., Kakizaki, T., Iwano, M., Tarutani, Y., Watanabe, M., Isogai, A., & Takayama, S. (2006). Dominance relationships between self-incompatibility alleles controlled by DNA methylation. *Nature genetics*, *38*(3), 297-299.
- Singh, R., Low, E. T. L., Ooi, L. C. L., Ong-Abdullah, M., Ting, N. C., Nagappan, J., ... Martienssen, R. A. (2013). The oil palm SHELL gene controls oil yield and encodes a homologue of SEEDSTICK. Nature, 500(7462), 340-344.

- Sivanesan, I. (2007). Shoot regeneration and somaclonal variation from leaf callus cultures of Plumbago zevlanica Linn. *Asian J. Plant Sci*, *6*, 83-86.
- Skirvin, R. M., McPheeters, K. D., & Norton, M. (1994). Sources and frequency of somaclonal variation. *HortScience*, 29(11), 1232-1237.
- Skirvin, R. M., Norton, M., & McPheeters, K. D. (1992). Somaclonal variation: has it proved useful for plant improvement?. In *II International Symposium on In Vitro Culture and Horticultural Breeding* 336 (pp. 333-340).
- Smith, R. J., & Kelsey, G. (2001). Identification of imprinted loci by methylation: use of methylation-sensitive representational difference analysis (Me-RDA). *Methods in molecular biology (Clifton, NJ)*, 181, 113.
- Smith, T. S. (2013). DNA Methylation and Transgenerational Stress Memories in Arabidopsis thaliana (Doctoral dissertation, University of York).
- Smulders, M. J. M., & De Klerk, G. J. (2011). Epigenetics in plant tissue culture. *Plant Growth Regulation*, 63(2), 137-146.
- Smulders, M. J. M., Rus-Kortekaas, W., & Vosman, B. (1995). Tissue culture-induced DNA methylation polymorphisms in repetitive DNA of tomato calli and regenerated plants. *Theoretical and Applied Genetics*, *91*(8), 1257-1264.
- Sogeke, A. K. (1998). Stages in the vegetative propagation of oil palm, Elaeis guineensis Jacq. through tissue culture. *Journal of Oil Palm Research*, 10(2), 1-9.
- Sparnaaij, L. D. (1969). *Oil palm (Elaeis guineensis* Jacquin). Miscellaneous Papers. Landbouwhogeschool Wageningen, (pp. 339-87).
- Staritsky, G. (1970). Tissue culture of the oil palm (*Elaeis guineensis* Jacq.) as a tool for its vegetative propagation. *Euphytica*, 19(3), 288-292.
- Swartz, H. J. (1991). Post culture behavior: genetic and epigenetic effects and related problems. In *Micropropagation* (pp. 95-121). Springer Netherlands.
- Takada, S., & Tasaka, M. (2002). Embryonic shoot apical meristem formation in higher plants. *Journal of Plant Research*, 115(6), 411-417.
- Tan, C. C., Wong, G., & Sohl, A. C. (1999). Acclimatization and handling of oil palm tissue cultured plantlets for large scale commercial production. In *PORIM Intl. Palm Oil Congress* (pp. 1-6).
- Tanaka, M., & Fujiwara, T. (2008). Physiological roles and transport mechanisms of boron: perspectives from plants. *Pflügers Archiv-European Journal of Physiology*, 456(4), 671-677.
- Teixeira, J. B., Söndahl, M. R., & Kirby, E. G. (1993). Somatic embryogenesis from immature zygotic embryos of oil palm. *Plant Cell, Tissue and Organ Culture*, 34(3), 227-233.

- Teixeira, J. B., Söndahl, M. R., & Kirby, E. G. (1994). Somatic embryogenesis from immature inflorescences of oil palm. *Plant Cell Reports*, *13*(5), 247-250.
- Teixeira, J. B., Söndahl, M. R., Nakamura, T., & Kirby, E. G. (1995). Establishment of oil palm cell suspensions and plant regeneration. *Plant Cell, Tissue and Organ Culture*, 40(2), 105-111.
- Thuzar, M., Vanavichit, A., Tragoonrung, S., & Jantasuriyarat, C. (2011). Efficient and rapid plant regeneration of oil palm zygotic embryos cv. 'Tenera'through somatic embryogenesis. *Acta physiologiae plantarum*, *33*(1), 123-128.
- Tinker, P. B. (1976). Soil requirements of the oil palm. In R. H. V. Corley, J. J. Hardon, & B. J. Wood, *Developments in Crop Science* (pp. 165-174). Elsevier, Amsterdam.
- Tinsley, C. R., & Nassif, X. (1996). Analysis of the genetic differences between Neisseria meningitidis and Neisseria gonorrhoeae: two closely related bacteria expressing two different pathogenicities. *Proceedings of the National Academy of Sciences*, 93(20), 11109-11114.
- Treutlein, J., & Rietschel, M. (2011). Genome-wide association studies of alcohol dependence and substance use disorders. *Current Psychiatry Reports*, 13(2), 147-155.
- Ushijima, T., & Yamashita, S. (2009). Methylation-sensitive representational difference analysis (MS-RDA). In DNA Methylation (pp. 117-130). Humana Press.
- Van der Velden, V. H. J., Cazzaniga, G., Schrauder, A., Hancock, J., Bader, P., Panzer-Grumayer, E. R., ... & Van Dongen, J. J. M. (2007). Analysis of minimal residual disease by Ig/TCR gene rearrangements: guidelines for interpretation of real-time quantitative PCR data. *Leukemia*, 21(4), 604-611.
- van Haaren, M. J., & Ow, D. W. (1993). Prospects of applying a combination of DNA transposition and site-specific recombination in plants: a strategy for gene identification and cloning. *Plant molecular biology*, 23(3), 525-533.
- Vandesompele, J., De Preter, K., Pattyn, F., Poppe, B., Van Roy, N., De Paepe, A., & Speleman, F. (2002). Accurate normalization of real-time quantitative Q-PCR data by geometric averaging of multiple internal control genes. *Genome biology*, *3*(7), research0034.
- Vaucheret, H., & Fagard, M. (2001). Transcriptional gene silencing in plants: targets, inducers and regulators. *Trends in Genetics*, 17(1), 29-35.
- Verhoeven, K. J., Jansen, J. J., van Dijk, P. J., & Biere, A. (2010). Stress-induced DNA methylation changes and their heritability in asexual dandelions. *New Phytologist*, 185(4), 1108-1118.

- Vician, L., Basconcillo, R., & Herschman, H. R. (1997). Identification of genes preferentially induced by nerve growth factor versus epidermal growth factor in PC12 pheochromocytoma cells by means of representational difference analysis. *Journal of neuroscience research*, 50(1), 32-43.
- Vidal, M. D. C., & De García, E. (2000). Analysis of aMusa spp. somaclonal variant resistant to yellow Sigatoka. *Plant Molecular Biology Reporter*, 18(1), 23-31.
- Vorster, B., Kunert, K., & Cullis, C. (2002). Use of representational difference analysis for the characterization of sequence differences between date palm varieties. *Plant Cell Reports*, 21(3), 271-275.
- Vos, P., Hogers, R., Bleeker, M., Reijans, M., Van de Lee, T., Hornes, M., ... & Zabeau, M. (1995). AFLP: a new technique for DNA fingerprinting. *Nucleic acids research*, 23(21), 4407-4414.
- Wahid, M. B., Abdullah, S. N. A., & Henson, I. E. (2005). Oil palm-achievements and potential. *Plant Production Science*, 8(3), 288-297.
- Wang, W. S., Pan, Y. J., Zhao, X. Q., Dwivedi, D., Zhu, L. H., Ali, J., ... & Li, Z. K. (2011). Drought-induced site-specific DNA methylation and its association with drought tolerance in rice (Oryza sativa L.). *Journal of Experimental Botany*, 62(6), 1951-1960.
- Wang, W. Y., Barratt, B. J., Clayton, D. G., & Todd, J. A. (2005). Genome-wide association studies: theoretical and practical concerns. *Nature Reviews Genetics*, 6(2), 109-118.
- Warington, K. (2008). The Growth and Anatomical Structure of The Carrot (Davcus Carota) As Affected By Boron Deficiency. Annals of Applied Biology, 27(2), 176-183.
- Welsh, J., & McClelland, M. (1990). Fingerprinting genomes using PCR with arbitrary primers. *Nucleic acids research*, 18(24), 7213-7218.
- Williams, C. N., & Thomas, R. L. (1970). Observations on sex differentiation in the oil palm, Elaeis guineensis L. *Annals of Botany*, *34*(4), 937-963.
- Williams, J. G., Kubelik, A. R., Livak, K. J., Rafalski, J. A., & Tingey, S. V. (1990). DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. *Nucleic acids research*, 18(22), 6531-6535.
- Wonkyi-Appiah, J. B. (1987). Genetic control of fertility in the oil palm (Elaeis guineensis Jacq.). *Euphytica*, 36(2), 505-511.
- Wooi, K. C. (1990). Oil palm (Elaeis guineensis Jacq.): tissue culture and micropropagation. In Legumes and Oilseed Crops I (pp. 569-592). Springer Berlin Heidelberg.

- Wooi, K. C. (1995). Oil palm tissue culture-current practice and constraints.Recent developments in oil palm tissue culture and biotechnology. *Malaysia, Malaysian Palm Oil Board*, 21-32.
- Wooi, K. C., Wong, C. Y., & Corley, R. H. V. (1982). Tissue culture of palms--a review. In *Tissue culture of economically important plants: proceedings of the International Symposium held at the Botany Department*, National University of Singapore, Singapore, 28-30 April 1981/edited by AN Rao.
- Zhang, Y. Y., Fischer, M., Colot, V., & Bossdorf, O. (2013). Epigenetic variation creates potential for evolution of plant phenotypic plasticity. *New Phytologist*, 197(1), 314-322.
- Zilberman, D., Gehring, M., Tran, R. K., Ballinger, T., & Henikoff, S. (2007). Genome-wide analysis of Arabidopsis thaliana DNA methylation uncovers an interdependence between methylation and transcription. *Nature genetics*, 39(1), 61-69.
- Ziogas, D. (2011). Standard and research for ovarian cancer: Emphasis on interactome-based tests for screening or treatment including HIPEC. *Gastric and Breast Cancer*, 10(3), 146-150.
- Zoldos, V., Siljak-Yakovlev, S., Papes, D., Sarr, A., & Panaud, O. (2001). Representational difference analysis reveals genomic differences between Q. robur and Q. suber: implications for the study of genome evolution in the genus Quercus. *Molecular Genetics and Genomics*, 265(2), 234-241.