



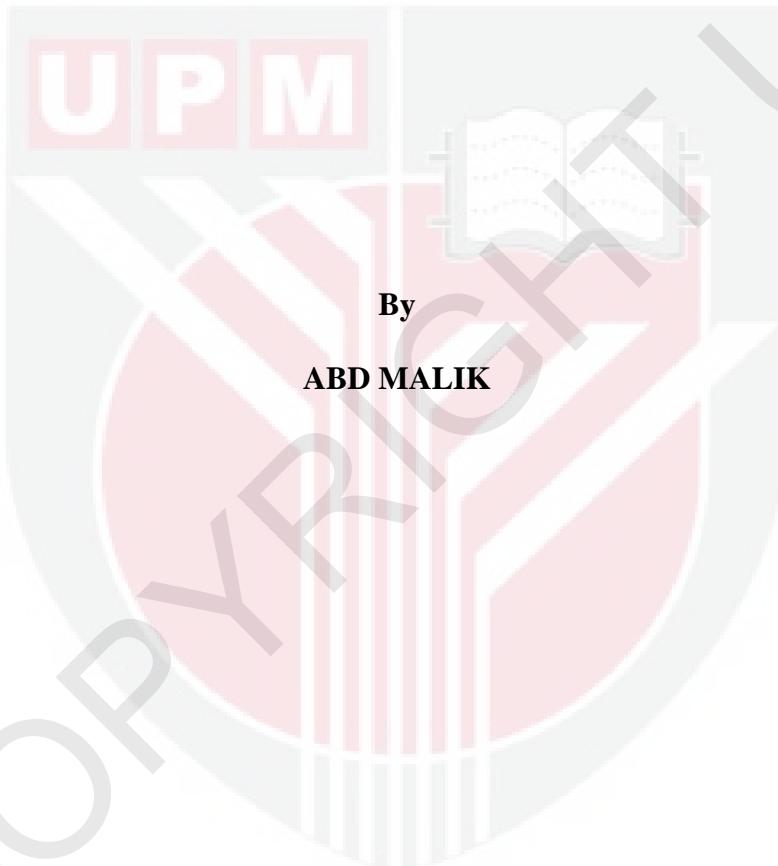
UNIVERSITI PUTRA MALAYSIA

***REPRODUCTIVE EFFICIENCY THROUGH ESTRUS
RESYNCHRONIZATION, TIMED ARTIFICIAL INSEMINATION AND
SPERMATOZOA SEPARATION IN BEEF CATTLE***

ABD MALIK

FPV 2012 1

**REPRODUCTIVE EFFICIENCY THROUGH ESTRUS RESYNCHRONIZATION,
TIMED ARTIFICIAL INSEMINATION AND SPERMATOZOA SEPARATION
IN BEEF CATTLE**



**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

Februari 2012

***Dedicated to my beloved Parents (Almarhum my father Munandar
and my mother Tasmani), my wife Anis Latifa and my sons
Mohammad Azarudin Malik and Masduki zaini Malik for their
supplication, patience and understanding***



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**REPRODUCTIVE EFFICIENCY THROUGH ESTRUS RESYNCHRONIZATION,
TIMED ARTIFICIAL INSEMINATION AND SPERMATOZOA SEPARATION
IN BEEF CATTLE**

By

ABD MALIK

February 2012

Chairman : Professor Abd Wahid Haron, PhD

Faculty : Veterinary Medicine

The objectives of this study were to improve resynchronization of oestrus, timing of artificial insemination and natural mating, verification of X- and Y-chromosomes after separation, and to analyze the motility, membrane integrity and acrosome integrity of bull spermatozoa. In the first experiment, a total 140 Brangus cows were synchronized with CIDR. Thirty to thirty five days after artificial insemination (AI), the cows were checked for pregnancy using ultrasound and those that remained open (non pregnant) were divided into two groups and resynchronized with either CIDR or two injections of PGF_{2α} further at 11 days interval. All cows were observed visually for estrus response for a period of two hours at 12 h intervals, starting immediately after CIDR removal or after the second injection of PGF_{2α}. Cows were in estrus when they were mounted at least 3 times during the period of observation. Following removal of CIDR and second injection of PGF_{2α}, cows were inseminated at 60 and 70 h later, respectively. There were no significant differences

(P>0.05) in estrus response and pregnancy rate between cows with initial synchronization and resynchronization used CIDR protocol.

In Experiment 2, a total of 185 Brangus cows were randomly divided into two experiments. First, a total of 100 cows were randomly divided into three groups. Groups 1, 2 and 3 were artificially inseminated at 48-50 h (n=30), 53-55 h (n=29) and 58-60 h (n=41) after CIDR removal. The pregnancy rates were 26.6%, 24.1% and 36.6%, respectively. The pregnancy rate was higher in the G1 (36.6%) than the G1 (26.6%) and G2 (24.1%) groups, but not significant different (P>0.05). Second, a total of 85 cows were divided into two groups. Group 1, consisted of 35 cows and further subdivided into seven sub-groups, consisting of 5 cows per group. Group 2 consisted of 50 cows which were artificially inseminated. At 58-60 h after CIDR removal, bulls were mixed in all sub-groups for one week (Group 1) and for Group 2, timed AI were conducted. The average pregnancy rate in natural mating was 28.6%. The pregnancy rate in the timed AI group was 18.0%. However, the difference in the pregnancy rate in natural mating and timed AI groups were not statistically significant (P>0.05).

In Experiment 3, verification of X- and Y-chromosome bearing spermatozoa separation by nested PCR and the motility and membrane integrity of the bull spermatozoa were evaluated. Semen samples were collected using an electro ejaculator from four Brangus bulls. Spermatozoa separation was conducted using three spermatozoa separation methods; (1) swimming speed using cervical mucus from estrus cows, (2) Percoll discontinuous gradient (45-90%), and (3) swim up using TALP medium. The nested PCR was used to verify the presence of X- and Y- chromosome. The motility and membrane integrity were

also evaluated before and after spermatozoa separation. Primers were designed using amelogenin cDNA sequence with 329 bp and 266 bp X- and Y-bearing chromosome spermatozoa, respectively. Motility was analyzed using computer assist spermatozoa analysis (CASA) system, whereas the membrane integrity was analyzed using hypo-osmotic swelling test (HOST). These results were confirmed by the absence of a single band, either for X- or Y- chromosome. The double band indicating that the spermatozoa cannot be separated was observed, after separation. However, the percentage of X-chromosome spermatozoa in the swimming speed using cervical mucus from estrus cows media (58.33%), swim-up using TALP media (50%), while that in the Percoll gradient method was 44.33%. Statistically both percentage were significantly different ($P<0.001$) compared to the theoretical ratio (50:50). However, comparison of spermatozoa motility, membrane integrity, and concentration before and after separation showed significant differences ($P<0.05$) among them.

In Experiment 4, the effect of extenders on the spermatozoa quality after cryopreservation were studied. Four extenders were used in this study namely cervical mucus collected from cows at estrus, Tris buffer, Sodium citrate and Tyrode's albumin lactate and pyruvate (TALP). All of the extender aliquots were diluted to obtained sperm concentrations of 25×10^6 sperm/mL. Computer assisted semen analyses, hypo-osmotic swelling tests and fluorescence isothiocyanate-labelled peanut agglutinin (FITC-PNA) techniques were used to determine sperm motility, plasma membrane integrity and sperm acrosome integrity, respectively, in both fresh and frozen-thawed semen. The motility ($74.9 \pm 0.60\%$), membrane integrity ($70.86 \pm 0.81\%$) and acrosome integrity ($67.31 \pm 0.94\%$) of fresh ejaculates were higher than frozen-thawed semen. Sperm motility ($47.93 \pm 0.80\%$) was significantly higher

(P<0.05) in cervical mucus extender than in the Tris buffer and TALP extenders. Furthermore, the levels of membrane ($43.09 \pm 0.83\%$) and acrosome integrity ($51.67 \pm 0.80\%$) of post-thawed spermatozoa in the TALP extender were significantly lower (P< 0.05) compared to the other three extenders. In conclusion, the cervical mucus extender resulted in sperm motility, membrane integrity and acrosome integrity comparable to that of the other tested extenders. However, acrosome integrity after thawing was higher than that observed with the TALP extender.

Abstrak thesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KECEKAPAN PEMBIAKAN DENGAN SINKRONISASI SEMULA ESTRUS,
WAKTU KAWIN DAN PENGASINGAN SPERMATOZOA DALAM LEMBU**

Oleh

ABD MALIK

Februari 2012

Pengerusi : Profesor Abd Wahid Haron, PhD

Fakulti : Perubatan Veterinar

Objektif kajian ini adalah untuk meningkatkan pensinkronian semula estrus, masa permanian beradas atau pembiakan tabii pengesahan kromosom X dan Y, dan juga untuk menganalisis motiliti, kesempurnaan selaput dan kesempurnaan akrosom menggunakan kaedah pemisahan spermatozoa pada lembu . Dalam ujikaji pertama, 140 lembu Brangus disinkroni dengan CIDR. Tiga puluh hingga tiga puluh lima hari selepas permanian beradas (AI), lembu diperiksa untuk menentukan kebuntingan menggunakan ultrabunyi dan lembu yang tidak bunting dibahagikan kepada dua kumpulan dan disinkronisasi semula dengan CIDR atau dua suntikan PGF_{2α} pada selang 11 hari. Semua lembu diperhati untuk menentukan tindakbalas estrus selama dua jam selang 12 jam, bermula selepas CIDR dikeluarkan atau selepas suntikan kedua PGF_{2α}. Lembu berada pada estrus apabila dipanjang sekurang-kurangnya 3 kali dalam tempoh pemerhatian. Selepas CIDR dikeluarkan dan suntikan kedua PGF_{2α}, lembu diinseminasi masing-masing pada 60 dan 70 jam kemudian. Tiada perbezaan yang bererti ($P > 0.05$) dalam tindakbalas estrus dan kadar kebuntingan antara pensinkronian awal dan pensinkronian semula lembu menggunakan CIDR.

Dalam Ujikaji 2, sebanyak 185 lembu Brangus diasingkan secara rawak kepada dua ujikaji. Pertama, sebanyak 100 ekor lembu telah dibahagikan secara rawak kepada tiga kumpulan. Kumpulan 1, 2 dan 3 masing-masing diinseminasi pada 48-50 jam ($n = 30$), 53-55 jam ($n = 29$) dan 58-60 jam ($n = 41$) selepas CIDR dikeluarkan. Kadar kebuntingan adalah 26.6%,

24.1% dan 36.6%. Sungguhpun kadar kebuntingan ini adalah lebih tinggi bagi kumpulan G3 (36.6%) berbanding dengan G2 (24.1%) dan kumpulan G1 (26.6%), namun tiada perbezaan bererti ($P>0.05$). Kedua, sebanyak 85 ekor lembu telah dibahagikan kepada dua kumpulan.Kumpulan 1 terdiri daripada 35 ekor lembu dibahagikan lagi kepada tujuh sub-kumpulan, terdiri daripada 5 ekor lembu bagi setiap sub-kumpulan.Sementara kumpulan 2 terdiri daripada 50 ekor lembu yang dibiakkan secara AI. Pada 58-60 jam selepas CIDRdikeluarkan, lembu jantan telah dicampur dalam setiap sub-kumpulan selama seminggu (kumpulan 1) dan bagi kumpulan 2 pula, AI telah dijalankan. Purata kadar kebuntingan dalam kumpulan 1 adalah 28.6% dan kumpulan 2 adalah 18.0%. Walau bagaimanapun, perbezaan dalam kadar kebuntingan ini adalah tidak bererti ($P> 0.05$).

Dalam Ujikaji 3, pengesahanspermatozoa berkromosom X dan Y menggunakan kaedah nested-PCR, penilaian motiliti dan kesempurnaan selaput spermatozoa juga dijalankan dengan menggunakan tiga kaedah pemisahan spermatozoa; (1) kelajuan berenang menggunakan mukus cervik daripada lembu estrus, (2) Percoll kecerunan tak selanjar (45-90%), dan (3) berenang menggunakan media TALP. Nested PCR digunakan untuk mengesahkan kehadiran kromosom X dan Y. Penilaian motiliti dan kesempurnaan selaput juga dinilai sebelum dan selepas pemisahan spermatozoa. Primers direka dengan menggunakan jujukan cDNA amelogenin dengan 329 dan 266 bp masing-masing bagi spermatozoa berkromosom X dan Y. Motiliti dianalisis menggunakan penganalisisan semen berkomputer (CASA), manakala kesempurnaan selaput dianalisis menggunakan kaedah ujian bengkak hipo-osmosis (HOST). Keputusan ini telah disahkan oleh ketiadaan jalur tunggal, sama ada untuk kromosom X atau Y. Jalur berganda menunjukkan bahawa spermatozoa tidak dapat dipisahkan juga diperhatikan, selepas pemisahan. Walau bagaimanapun, peratus

spermatozoa berkromosom X menggunakan media mukus servik daripada lembu estrus adalah 58.33%, manakala kaedah kecerunan Percoll adalah 44.33%.Statistik peratusan kedua-duanya berbeza ($P<0.001$) berbanding dengan nisbah teori (50:50).Walau bagaimanapun, perbandingan motiliti spermatozoaa, kesempurnaan selaput, dan kepekatan sebelum dan selepas pemisahan adalah mempunyai perbezaan yang bererti ($P <0.05$) di kalangan mereka.

Dalam Ujikaji 4, kesan pelarut pada motiliti, kesempurnaan selaput dan kesempurnaan akrosom spermatozoa lembu beku yang dicairkan juga dikaji. Empat pelarut digunakan dalam ujikaji ini iaitu mukus servik yang dikumpul daripada lembu estrus, penampang Tris, sodium sitrat dan media TALP.Kesemua alikuot pelarut telah dicairkan untuk menghasilkan kepekatan sebanyak 25×10^6 sperma/mL. Teknik CASA, HOST, dan florosent dilabel isothiocyanate kekacang agglutinin (FITC-PNA) digunakan untuk menentukan motili, kesempurnaan selaput dan kesempurnaan akrosom spermatozoa, masing-masing, dalam kedua-dua air mani yang segar dan beku yang dicairkan. Motiliti ($74.9 \pm 0.60\%$), kesempurnaan selaput ($70.86 \pm 0.81\%$) dan kesempurnaan akrosom spermatozoa ($67.31 \pm 0.94\%$) bagi air mani segar adalah lebih tinggi daripada air mani beku yang dicairkan.Motiliti spermatozoa ($47.93 \pm 0.80\%$) adalah berbeza ($P <0.05$) dalam mukus servik berbanding penampang Tris dan TALP. Tambahan pula, kesempurnaan selaput ($43.09 \pm 0.83\%$) dan kesempurnaan akrosom ($51.67 \pm 0.80\%$) bagi spermatozoa beku yang dicairkan dalam pengekal TALP adalah rendah ($P<0.05$) jika dibandingkan dengan pelarut lain. Kesimpulannya, pelarus mucus cervik memberikan keputusan motility spermatozoa, kesempurnaan selaput dan akrosom yang standing dengan pelarut lain. Walau bagaimanapun, kesempurnaan akrosom selepas dicairkan adalah lebih tinggi berbanding pelarut TALP.



ACKNOWLEDGEMENTS

In the name of Allah: The Most Merciful and the Most Compassionate. The praise is to Allah. Because of Allah is guide, I have completed this PhD.

I would like to deeply express my profound gratitude to my supervisor, Professor. Dr. Abd Wahid Haron, who consistently motivated and supported me with his insight, experience, and knowledge as well as for invaluable guidance and advice during the course of my PhD study, I am also highly thankful to Associate Prof Dr. Rosnina Yusoff, Associate Prof Dr. Azhar Kasim and Dr. Md Sabri Mohd Yusoff for their valuable suggestions and guidance throughout this study.

My appreciation is also directed to Directorate of Human Resources (Direktorat Ketenagaan), Directorate General of Higher Education (Dikti), and The National Education Ministry, Republic of Indonesia and government provinsi of south Kalimantan for financial support during my study. I would like thank coordinator Kopertis Wilayah XI Kalimantan, Dean Faculty of Agriculture and Rector of Islamic Kalimantan University. I wish to extend my warmest thanks to all those who have helped me with my work at the Theriogenology & Cytogenetics unit (Mr. Yap Keng Chee and Mr. Fahmi Mashuri). Department of Clinical Studies and all of staff Histopathology Laboratory, Department of Veterinary & Pathology microbiology, Faculty of Veterinary Medicine.

Finally, my deepest gratitude goes to my parents (Almarhum Munandar and Tasmani), my father in law (Almarhum Taswan) and my mother in law (Sumi) for their prayer, my entire

brother and sisters, for their motivation, and especially to my wife Anis Latifa and my sons, Mohammad Azaruddin Malik and Masduki Zaini Malik, my source of motivation and inspiration.



I certify that an Examination Committee met on 13 February 2012 to conduct the final examination of Abd Malik on his Doctor of Philosophy thesis entitled "Reproductive efficiency through estrus resynchronization, timed artificial insemination and spermatozoa separation in beef cattle" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The committee recommends that the candidate be awarded the Doctor of Philosophy.

Members of the Examination Committee are as follows:

Mohamed Ariff bin Omar, PhD

Professor

Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Mohamed Ali bin Rajion, PhD

Professor

Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Internal Examiner)

Md Zuki bin Abu Bakar @ Zakaria, PhD

Professor

Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Internal Examiner)

Patrick Lonergan, PhD

Professor

Faculty of Agriculture, Food and Veterinary Medicine
Universiti Colloge Dublin
(External Examiner)

SEOW HENG PONG, Ph D

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirements for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Abd Wahid Haron, PhD

Professor

Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Rosnina Yusoff, PhD

Associate Professor

Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Azhar Kasim, PhD

Associate Professor

Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Md Sabri Mohd Yusoff, PhD

Senior Lecturer

Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, Ph D

Professor and Dean

School of Graduate Studies
Universiti Putra Malaysia

Date:

DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institutions.

ABD MALIK

Date: 13 February 2012



TABLE OF CONTENTS

	Page
ABSTRACT	iv
ABSTRAK	viii
ACKNOWLEDGEMENTS	xi
APPROVAL	xiii
DECLARATION	xiv
TABLE OF CONTENTS	xvi
LIST OF TABLES	xix
LIST OF ABBREVIATIONS	xxii
CHAPTER	1
1 INTRODUCTION	1
2 LITERATURE REVIEW	5
2.1 Introduction	5
2.2 Bovine estrus cycle	6
2.3 Estrus synchronization protocol	10
2.4 Resynchronization of estrus cycle	11
2.5 Artificial insemination	16
2.6 Semen collection	17
2.7 Semen evaluation	18
2.7.1 Motility	19
2.7.2 Spermatozoa Concentration	21
2.7.3 Spermatozoa viability	22
2.7.4 Morphology of spermatozoa	23
2.7.5 Membrane integrity	24
2.7.6 Acrosome integrity	25
2.8 Semen extenders	26
2.9 Spermatozoa separation	28
2.9.1 Spermatozoa separation protocols	29
2.9.2 Verification of separated spermatozoa	32
3 EFFECTS OF RESYNCHRONIZATION WITH PROGESTERONE AND PROSTAGLANDIN F₂<i>a</i> ON ESTRUS RESPONSE AND PREGNANCY RATE IN BEEF CATTLE	34
3.1 Introduction	34

3.2	Materials and methods	36
3.2.1	Animal	36
3.2.2	Synchronization and resynchronization of estrus	36
3.2.3	Estrus observation	39
3.2.4	Artificial insemination	39
3.2.5	Pregnancy diagnosis	40
3.2.6	Analysis statistic	41
3.3	Results	41
3.4	Discussions	46
4	COMPARISON BETWEEN TIMED ARTIFICIAL INSEMINATION AND NATURAL MATING ON PREGNANCY RATE IN POSTPARTUMSYNCHRONIZED BEEF CATTLE	49
4.1	Introduction	49
4.2	Material and methods	51
4.2.1	Animal	51
4.2.2	Estrus synchronization	51
4.2.3	Experimental design	52
4.2.4	Pregnancy diagnosis	53
4.2.5	Statistical analysis	53
4.3	Results	53
4.4	Discussions	56
5	SPERMATOZOA SEPARATION AND VERIFICATION BY NESTED PCR AND EVALUATION OF THEIR MOTILITY AND MEMBRANE INTEGRITY	60
5.1	Introduction	60
5.2	Materials and methods	62
5.2.1	Semen collection	62
5.2.2	Assessment of motility	63
5.2.3	Semen sexing procedure	63
5.2.4	Swim up	63
5.2.5	Swimming Speed	64
5.2.6	Percoll Gradient	64
5.2.7	Assessment of spermatozoa membrane integrity	65
5.2.8	DNA extraction	66
5.2.9	Primers	67
5.2.10	PCR amplification	67
5.2.11	Detection of PCR	68
5.2.12	Statistical analysis	68

5.3	Results	68
5.4	Discussions	73
6	EFFECTS OF CERVICAL MUCUS FROM ESTRUS COWS ON THE MOTILITY, MEMBRANE AND ACROSOME INTEGRITY OF FROZEN–THAWED BULL SPERMATOZOA	77
6.1	Introduction	77
6.2	Materials and methods	79
6.2.1	Semen collection	79
6.2.2	Preparation of extenders	79
6.2.3	Semen evaluation	80
6.2.4	Assessment motility	80
6.2.5	Semen of cryopreservation	80
6.2.6	Evaluation of frozen-thawed semen	81
6.2.7	Assessment of membrane integrity	81
6.2.8	Assessment acrosome integrity	82
6.2.9	Statistical analysis	86
6.3	Results	86
6.4	Discussions	90
7	GENERAL DISCUSSION	95
8	SUMMARY GENERAL CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH	100
REFERENCES		102
APPENDICES		131
BIODATA OF STUDENT		137
LIST OF PUBLICATIONS		138

LIST OF TABLES

Tabel 2.1	Characteristic of the bovine estrus cycle from stage, day of the estrus cycle	9
Table 2.2	Hormones used for synchronization of estrus in cows	13
Table 2.3	Scoring system for wave motion of spermatozoa	20
Table 2.4	Morphology abnormality of spermatozoa	24
Table2.5	Summary of potential differences between X- and Y-chromosome spermatozoa	29
Table3.1	Percentage of estrus response and pregnancy following synchronization and resynchronization using CIDR and PGF _{2α}	43
Table 4.1	Percentage of estrus response, pregnancy rates following timed AI54	
Tabel 4.2	Pregnancy rates following natural mating and timed IA in estrus synchronized cows	55
Table 5.1	Percentages (mean ± S.E.M) of motility, membrane integrity and concentration of spermatozoa before and after separation.	72
Table6.1	Effects of different semen extenders on motility of frozen-thawed spermatozoa	87
Table 6.2	Effects of different semen extender on frozen-thawed membrane integrity spermatozoa	88
Table 6.3	Effects of different semen extenders on frozen-thawed acrosome integrity spermatozoa	89

LIST OF FIGURES

Figure3.1 Synchronization and resynchronization protocol with CIDR and PGF _{2α} administration in beef cattle	38
Figure3.2 Ultrasonographic image of a transverse section of the uterine horn (A) non pregnant 1: Endometrium 2: Myometrium 3: vascular portion of the uterus. (B) Early pregnancy 1: Amniotic fluid 2: Uterine wall 3: Fetus40	
Figure3.3 Percentage distribution of estrus response in CIDR synchronized and resynchronized cows observed at 12-hourly intervals	44
Figure3.4 Percentage distribution of estrus response in CIDR and PGF2α resynchronized cows observed at 12-hourly intervals	45
Figure 4.1 Procedure for estrus synchronization and timed AI in beef cattle	53
Figure 4.2 Pregnancy rates following in natural mating in estrus synchronized cows	55
Figure 5.1 Agarose gel results from nested PCR for X- and Y-chromosome after separation; Line M with molecular weight of 500 bp. Gel A single band of Y- Chromosome, B double band and C single band of X- chromosome	70
Figure 5.2 Percentage of X - and Y - chromosome after confirmation by nested PCR for different methods Error! Bookmark not defined.	
Figure6.1 Assessment of bulls spermatozoa membrane integrity following hypoosmotic swelling test (HOST). (▲) Normal spermatozoa are displaying coiled tails after HOST. (▽) Abnormal spermatozoa displaying coiled tails after HOST	82
Figure 6.2 Photograph of bull's spermatozoa acrosome stained with FITC-PNA. (A) Image obtained by phase contrast microscope. (B) The same field of image A, obtained by fluorescence microscope; (▼) Arrowhead indicates intact acrosome; dented arrowhead (▷) indicates partially damaged acrosome	84
Figure 6.3 Photograph of bull's spermatozoa acrosome stained with FITC-PNA. (A) Image obtained by phase contrast microscope. (B) The same field of image A, obtained by fluorescence microscope; (△) Arrowhead indicates intact acrosome; dented arrowhead (▷) indicates partially damaged acrosome; and arrow (↗) indicates lost acrosome	85

Figure 6.4 Percentage of motility, membrane integrity and acrosome integrity on each extenders in frozen-thawed spermatozoa 89



LIST OF ABBREVIATIONS

AI	Artificial insemination
ANOVA	Analysis of variance
BCS	Body condition score
CASA	Computer assisted spermatozoa analysis
CI	Calving interval
CL	Corpus luteum/Corpora lutea
CIDR	Controlled Internal Drug Release
d	Day (s)
DNA	Deoxyribonucleic acid
EB	Estradiol benzoate
eCG	Equine chorionic gonadotropine
FITC	Fluorescein isothiocyanate
FSH	Follicle stimulating hormone
FTAI	Fixed-time insemination
GnRH	Gonadotropin releasing hormone
h	Hour (s)
HEPES	N-(2-hydroxyethyl) piperazine-N'-(2-ethanesulphonic acid)
hCG	Human chorionic gonadotropine
HOST	Hypo-osmotic swelling test
i.m.	Intramuscular
IU	International unit

IVF	In-vitro fertilization
kg	Kilogram (s)
LH	Luteinizing hormone
mOsmol	Milli-osmole
MGA	Melengestrol acetate
mg	Milligram (s)
mL	Milliliter (s)
mM	Millimole (s)
PBS	Phosphate-buffered saline
PCR	Polymerase chain reaction
PGF ₂ α	Prostaglandin F ₂ α
pH	hydrogen ion concentration,-log 10 of
PNA	Peanut agglutinin
PSA	Pisum Sativum agglutinin
PRID	Progesterone releasing intra vagina device
RIA	Radioimmunoassay
RNA	Ribosome nucleate acid
ROS	Reactive oxygen species
SQA	Spermatozoa quality analyzer
S E M	Standard error of the mean
TALP	Tyrode's albumin lactate and pyruvate
Tris	Tris (hydroxymethyl) aminomethane
v/v	Volume: volume ratio

X sperm X-chromosome bearing spermatozoa

Y sperm Y-chromosome bearing spermatozoa



CHAPTER 1

INTRODUCTION

Reproductive efficiency is an essential tool that is used to evaluate beef cattle production (Dickerson, 1970). The target in cattle production is the maintenance of good reproductive performance, and it is also important to provide efficient management (Ball and Peters, 2004). Some of the reproductive technologies that can be used to increase reproductive efficiency include estrus synchronization, artificial insemination and spermatozoa separation.

Normally, estrus in cows are observed around days 21 of the estrus cycle if the cow is not pregnant. Failure of cows to become pregnant and calf losses at or immediately after birth is the main reasons for reproductive failure (Bellows and Short, 1990; Wiltbank, 1994). Synchronization of estrus can be practiced to successive early pregnancy rate, which is essential to achieve a 12 month calving interval. This also allows more time for cows to resume estrus cyclicity before the beginning of the subsequent breeding season. Synchronization of estrus also improves homogeny of a calf crop (Dziuk and Bellows, 1983; Stagg *et al.*, 1995).

Resynchronization of estrus is a new method for the improvement of calving intervals in cows. The advent of ultrasonography has allowed accurate early pregnancy diagnosis, at least 25 to 26 days after artificial insemination in cattle (Kastelic *et al.*, 1988; Fricke, 2002; Ricardo *et al.*, 2004; Bartolome *et al.*, 2005;

Bartolome *et al.*, 2009), which facilitated the resynchronization programs. Implementation of resynchronization of estrus programs was considering results of pregnancy diagnosis by ultrasonography or per rectal palpation of the uterus. If the pregnancy diagnosis shows that the cows are not pregnant, the continued estrus synchronization will reduce the calving interval in cattle (Bartolome *et al.*, 2002).

Development of reproductive biotechnology has made it possible to carry out separation of X- and Y-chromosome bearing spermatozoa. Sexing of spermatozoa, involving separation of X- from Y-chromosome bearing spermatozoa, implies its application in artificial insemination and *in vitro* fertilization (IVF). The result of sexing spermatozoa is essential for dairy and beef industry. Dairy farmers would prefer most of their calves to be female and thus potentially become replacement for the milking herd, whereas beef farmers may often prefer their cow to produce bull calves (Ball and Peters, 2004; Hamano *et al.*, 2007). For a long time there had been attempts at spermatozoa sexing of X- and Y-chromosome on the basis of the physical, biological and immunological properties (Ball and Peters, 2004).

The spermatozoa separation has been accomplished by many different procedures such as the Percoll gradient, swim-up and modified swim-up (swimming speed), Sephadex columns, albumin gradient, sex specific antibody binding, free-flow electrophoresis, PCR, and flow cytometry (Iizuka *et al.*, 1987; Khatamee *et al.*, 1999; Madrid *et al.*, 2003; Joerg *et al.*, 2004; Kobayashi *et al.*, 2004). Sexed spermatozoa could be used in the management and insemination of cows with the aim of

producing calves of the required sex, and for the fertilization of oocytes *in vitro* to produce embryo of the required sex.

Cattle production in tropical countries could be described as multipurpose, with cows being used for milking, meat, clothing, fertilizer, fuel, draft power and sometimes for status or as a form of currency (Ball and Peters, 2004). Generally, in cattle production the main problem causing decreased reproductive efficiency, especially calving interval and delayed first estrus postpartum, is due to the high incidence of postpartum anestrus (Baruselli *et al.*, 2004). The effects of a long calving interval have as a consequence, negative effects on reproductive performance.

The slow recovery of reproductive performance during postpartum is a limitation to the success of reproductive management programs that are implemented for beginning of inseminations. The synchronization, resynchronization of estrus and artificial insemination including spermatozoa separation program could be used to increase reproductive performance in beef cattle so that it will have a good economic impact for the farmer. Therefore, this study was conducted with these objectives:

1. to determine the effects of resynchronization with progesterone and prostaglandin F₂α on estrus response and pregnancy rate in beef cattle.
2. to compare between timed artificial insemination and natural mating on the pregnancy rate in postpartum estrus synchronized beef cattle.
3. to separate X- and Y-chromosome bearing spermatozoa using swim-up, swimming speed and Percoll gradient methods.

4. to verify X- and Y-chromosome bearing spermatozoa using nested PCR.
5. to determine the effects of cervical mucus from cows in estrus on the motility, membrane integrity and acrosome integrity of frozen-thawed bull spermatozoa.



REFERENCES

- Abdelhakeam, A.A., Graham, E.F., Vazquez, J.M., and Chaloner, K.M. (1991). Study of absence of glycerol in unfrozen and frozen ram semen: Development of an extender for freezing:Effects of osmotic pressure, egg yolk levels,type of sugars, and ,method of dilution. *Journal of Cryobiology*, 28: 43-49.
- Aboagla, M.E., and Terada, T. (2003). Trehalose-Enhanced fluidity of the goat spermatozoa membrane and its protection during freezing. *Journal of Biologyof Reproduction*, 69: 1245 - 1250.
- Aboagla, M.E., and Terada, T. (2004). Effects of the suplementation of trehalosa extender containing egg yolk with sodium dodecyl sulfateon the freezabilityof goat spermatozoa. *Theriogenology*, 62: 809-818.
- Acevedo, N., Galina, C.S., Pulido, A., and Orihuela, A. (2007). Dynamics in sexually active groups of Zebu cattle (*Bosindicus*) comparing two procedures for estrus induction. *Journal of Veterinary Behavior*, 2: 5-9.
- Aitken, R.J. (1995). Free radicals, lipid peroxidation and sperm function. *Journal of Reproduction Fertility and Development*, 7: 659 - 668.
- Aitken, R.J., and Krausz, C. (2001). Oxidative stress, DNA damage and the Y-chromosome. *Journal of Reprodction*, 122: 497-506.
- Aitken, R.J., Gordon, E., Harkiss, D., Twigg, J.P., Milne, P., Jennings, Z., and Irvine, D.S. (1998).Relative impact of oxidative stress on the functional competence and genomic integrity of human spermatozoa. *Journal of Biology Reproduction*, 59: 1037-1046.
- Alvarenga, M.A., and Leão, K.M. (2002). Hysteroscopic insemination of mares with low number of frozen thawed spermatozoa selected by percoll gradient. *Theriogenology*, 58: 651-653.
- Amann, R.P. (1989). Can the fertiliy potential of a seminal sample be predicted accurately? *Journal of Andrology*, 10: 89-98.
- Amirat, I., Tainturier, D., Jeanneau, L., Thorin, C., Ge'rard, O., Courtens, J., and Anton, M. (2004). Bull semen in vitro fertility after cryopreservation using egg yolk LDL: a comparison with Optidyl, a commercial egg yolk extender. *Theriogenology*, 61: 895-907.

- Andersen-Ranberg, I.M., Klemetsdal, G., Heringstad, B., and Steine, T. (2005). Heritabilities, genetic correlations, and genetic change for female fertility and protein yield in Norwegian dairy cattle. *Journal of Dairy Science*, 88: 348-355.
- Ando, T., Kamimura, S., Hamana, K., Watanabe, G., and Taya, K. (2005). GnRH treatment at CIDR insertion influences ovarian follicular dynamics in Japanese black cows. *Journal of Veterinary Medicine Science* 67:275-280.
- Ax, R.L., Dally, M.R., Didion, B.A., Lenz, R.W., Love, C.C., Varner, D.D., Hafez, B., and Bellin, M.E. (2000). Artificial insemination. In: Reproduction in Farm Animal. 7 th edition, pp 376-388, New York.Lippincot.Williams and Wilkens.
- Bader., Kojima, F.N., Schafer, D.J., Stegner, J.E., Ellersieck, M.R., Smith, M.F., and Patterson, D.J. (2005). A comparison of progestin-based protocols to synchronize ovulation and facilitate fixed-time artificial insemination in postpartum beef cows. *Journal of Animal Science*, 83: 136-143.
- Ball, P.J.H., and Peters, A.R. (2004). Reproduction in Cattle.3 ed. Hongkong, Blacwell Publising.
- Ballachey, B.E., Evenson, D.P., and Saacke, R.G. (1988). The sperm chromatin structure assay. Relationship with alternate tests of semen quality and heterospermic performance of bulls. *Journal of Andrology*, 9: 109-115.
- Ballinger, H.J. (1970). The effect of inseminations carried out early or late in oestrus on the sex ratio of calves born. *Journal of Veterinary Research*, 86: 631-640.
- Bamba, K., and Cran, D.G. (1988). Effect of rapid warming of bull and rabbit semen. *Reproduction and Fertility*, 82: 501-507.
- Barth, A.D. (2000). Bull Breeding Soundness Evaluation. Western Canadian Association of Bovine Practitioners, Saskatoon, Sask., Canada.
- Bartolome, J.A., Sozzi, A., McHale, J., Melendez, P., Arteche, A.C.M., and Silvestre, F.T. (2005). Resynchronization of ovulation and timed insemination in lactating dairy cows, II: assigning protocols according to stages of the estrus cycle, or presence of ovarian cysts or an oestrus. *Theriogenology*, 68: 1628-1642.
- Bartolome, J.A., Sheerin, P., Luznar, S., Melendez, P., Kelbert, D., Risco, C.A., and Thatcher, W.W. (2002). Conception rates in lactating dairy cows using

- ovsynch after presynchronization with Prostaglandin F2 α or Gonadotropin releasing hormone (GnRH). *The Bovine Practitioner*, 36: 35-39.
- Bartolome, J.A., Van Leeuwen, J.J., Thieme, M., Sa'filho, O.G., Melendez, P., Archbald, L.F., and Thatcher, W.W. (2009). Synchronization and resynchronization of insemination in Lactating dairy with the CIDR insert and the ovsynch protocol. *Theriogenology*, 72: 869-878.
- Baruselli, P.S., Reis, E.L., Marques, M.O., Nasser, L.F., and Bo, G.A. (2004). The use of hormonal treatments to improve reproductive performance of anestrus beef cattle in tropical climates. *Animal Reproduction Science*, 82/83: 479-486.
- Barth, A. (2000). Bull breeding Soundness evalution. The Western Canadian Assosiation of Bovine Practitioners.
- Burstein, P. and Sckenker, J.G. (1985). High long-standing fertilizing capacity of human sperm isolation for male sex preselection. *Am Jurnal of Obstet Gynecol*, 151:795-798.
- Battista, P. J., C. E. Rexroad, Jr. and W. F. Williams.(1984). Effects of progesterone administered to dairy heifers on sensitivity of corpora lutea to FGF2, and on plasmaLH concentration. *Theriogenology*, 22:47-54.
- Bearden, H.J., and Fuquay, J.W. (1980). *Applied Animal Reproductive*. Virginia, Reston Publishing Company, Inc. USA.
- Beernink, F.J., Dmowski, W.P., and Ericsson, R.J. (1993). Sex preselection through albumin separation of sperm. *Journal of Fertility and Sterility*, 59: 382-386.
- Bellows, R.A., and Short, R.E. (1990). Reproductive losses in the beef industry In In: Proc. 39th Annu. Beef Cattle Short Course, Vol.Pp. 109-133.
- Beer-Ljubic', B., Aladrović, J., Marenjak, T.S., Majic-Balic, I., Laskaj, R.(2011). Biochemical properties of bull spermatozoa separated in iodixanol density solution. *Research Veterinary Science*.doi:10.1016/j.rvsc.2011.01.011.
- Bergeron, A., and Manjunath, P. (2006). New insights towards understanding the mechanisms of sperm protection by egg yolk and milk. *Molecular Reproduction and Development*, 73: 1338-1344.
- Bernard, C., Valet, J.P., Beland, R., and Lambert, R.D. (1983). Prediction of bovine ovulation by a rapid radioimmunoassay for plasma LH. *Reproduction and Fertility*, 68: 425-430.

- Bó, G.A., Baruselli, P.S., and Martinez, M.F. (2003). Pattern and manipulation of follicular development in Bos indicus cattle. *Animal Reproduction Science*, 78: 307-326.
- Bó, G.A., Cutaia, L., Peres, L.C., Pincinato, D., Mara~na, D., and Baruselli, P.S. (2007). Technologies for fixed-time artificial insemination and their influence on reproductive performance of Bos indicus cattle. *Reproductive and Fertility (Suppl)*, 64: 223-236.
- Blockey, M.A., De.B. (1979). Observation on group mating of bulls at pasture. *Journal Applied Animal Ethol*. 5, 15-34.
- Brandiff, B.F., Gordin, L.A., Haendel, S., Singer, S., Moore, D.H., and Gledhill, L. (1986). Sex chromosome ratios determined by karyotypic analysis in albumin-isolated human sperm. *Journal of Fertility and Sterility*, 46: 678-685.
- Bridges, G. A., Helser, L. A., Grum, D. E., Mussard, M.L., Gesser, C. L. and Day, M. L. (2008). Decresing the interval between GnRH and PGF_{2α} from 7 to 5 days lengthening proestrus increases timed AI pregnancy rate in beef cows. *Theriogenology*, 69:843-851.
- Bryan, J.H., and Akruk, S.R. (1977). A Naphitol Yellow S and Erythrosin B staining procedure for use in studies of the acrosome reaction of rabbit spermatozoa. *Journal of Stain Technol*, 52: 47-51.
- Burns, B.M., Fordyce, G., and Holroyd, R.G. (2010). A review of factors that impact on the capacity of beef cattle females to conceive, maintain a pregnancy and wean a calf—Implications for reproductive efficiency in orthern Australia. *Animal Reproduction Science*, 122: 1-22.
- Busch, D.C., Wilson, D.J., Schafer, D.J., Leitman, N.R., Haden, J. K., Ellersieck, M.R., Smith, M.F., and Patterson, D.J. (2007). Comparison of progestin-based estrus synchronization protocols before fixed-time artificial insemination on pregnancy rate in beef heifers. *Journal of Animal Science*, 85: 1933-1939.
- Busch, D.C., Schafer, D.J., Wilson, D.J., Mallory, D.A., Leitman, N.R., Haden, J.K., Ellersieck, M.R., Smith, M.F., and Patterson, D.J. (2008). Timing of artificial insemination in postpartum beef cows following administration of the CO-Synch + controlled internal drug-release protocol. *Journal of Animal Science*, 86: 1519-1525.

- Cavalieri, J., and Macmillan, K.L. (2002). Synchronisation of oestrus and reproductive performance of dairy cows following administration of estradiol benzoate or GnRH during a synchronised pro-oestrus. *Australia. Veterinary*, 80: 486-493.
- Cavalieri, J., Hepworth, G., and Fitzpatrick, L.A. (2004). Comparison of two oestrus synchronization and resynchronization treatments in lactating dairy cows. *Theriogenology*, 62: 729-747.
- Cavalieri, J., Eagles, V.E., Ryan, M., and Macmillan, K.L. (2000). Patterns of onset of oestrus and reproductive performance of dairy cows enrolled in controlled breeding programs. In: Proc. Dairy Veterinary Conference in the Australia and New Zealand Combined.: 161-182.
- Cavalieri, J., V.M. Smart, G., Hepworth, M., Ryan., and Macmillan, K.L. (2008). Ovarian follicular development and hormone concentrations in inseminated dairy cows with resynchronized estrus cycles. *Theriogenology*, 70: 946-955.
- Chandler, J.E., Chenevert, R.W., Adkinson, E.B., and Moser.(1998). Sex ratio variation between ejaculates within sire evaluated by polymerase chain reaction, calving, and farrowing records. *Journal of Dairy Science*, 81: 1855-1867.
- Chebel, R., Santos, J.E.P., Cerri, R.L.A., Juchem, S., Galvao, K.N., and thatcher, W.W. (2003). Effects of resynchronization with GnRH on day 21 after artificial insemination on pregnancy rates and pregnancy loss in lactating dairy cows. *Theriogenology*, 60: 1389-1399.
- Check, J. H., and D. Katsoff. (1993). A prospective study to evaluate the efficacy of modified swim-up preparation for male sex selection. *Human.Reproduction*. 8(2):211-214.
- Check, J.H., Shanis, B.s., Cooper, S.O., and Bollendorf, A. (1989). Male sex preselection: swim-up technique and insemination of women after ovulation induction. *Journal of Arch. Androl*, 23: 165-166.
- Chenault, J.R., Boucher, J.F., Dame, K.J., Meyer, J.A., and Woods-Follis, S.L. (2003). Intra vaginal progesteron insertto synchronize return to estrus of previously inseminated Dairy cows. *Journal of Dairy Science*, 86: 2039-2049.
- Christensen, P., Brockhoff, P.B., and Lehn-Jensen, H. (2005). Discrepancies in the determination of sperm concentration using Burker-Turk, Thoma and Makler counting chambers. *Theriogenology*, 63: 992-1003.

- Claassens, O.E., Stander, F.S., Kruger, T.F., Menkveld, R., and Lombard, J.C. (1989). Does the wash-up and swim-up method of semen preparation play a role in sex selection? *Arch. Androl.*, 23 (1): 23-26.
- Claassens, O.E., Oosthuizen, J.C., Brusnick, J., Franken, D.R., and Kruger, T.f. (1995). Fluorescent in situ hybridization evaluation of human Y-bearing spermatozoa separated by albumin density gradients. *Journal of Fertility and Sterility*, 63: 417-418.
- Colazo, M.G., Kastelic, J.P., Mainar-Jaime, R.C., Gavaga, Q.A., Whittaker, P.R. and Small, J. A.(2006). Resynchronization of previously timed-inseminated beef heifers with progestins. *Theriogenology*, 65:557-72.
- Colly, A., Muhr, M., and Golovan, S.P. (2008). Single bovine sperm sex typing by amelogenin nested PCR. *Theriogenology*, 57: 1327-1346.
- Comizzoli, P., Mauget, R., and Mermilliod, P. (2001). Assessment of in vitro fertility or deer spermatozoa by heterologous IVF with zona-free bovine oocytes. *Theriogenology*, 56: 261-274.
- Convey, E.M., Tucker, H.A., and Short, R.E. (1983). Acute effect of suckling on gonadotropin, prolactin and glucocorticoid concentrations in serum of intact and ovariectomized beef cows. *Theriogenology*, 20:454-58
- Correa, J.R., and Zavos, P.M. (1994). The hypo osmotic swelling test: its employment as an assay to evaluate the functional integrity of the frozen-thawed bovine sperm membrane. *Theriogenology*, 70: 978 - 983.
- Correa, J.R., Pace, M.M., and Zavos, P.M. (1997). Relationships among frozen-thawed sperm characteristics assessed via the routine semen analysis, sperm functional tests and fertility of bulls in an artificial insemination program. *Theriogenology*, 48: 721-731.
- Courtens, J.L., Ekwall, H., Paquignon, M., and Ploen, L. (1989). Preliminary study of water and some element contents in boar spermatozoa, before, during and after freezing. *Journal of Fertility and. Sterility*, 87: 613-626.
- Cundiff, L.V., Nú~nez-Dominguez, R., Dickerson, G.E., Gregory, K.E., and Koch, R.M. (1992). Heterosis for lifetime production in Hereford, Angus, Shorthorn and crossbred cows. *Journal of Animal Science*, 70: 2397-2410.
- Cupp, A.S., Stumpf, T.T., Kojima, F.N., Werth, L.E., Wolfe, M.W., Roberson, M.S., Kittok, R.J., and Kinder, J.E. (1995). Secretion of gonadotrophins change during the luteal phase of the bovine oestrus cycle in the absence of

- corresponding changes in progesterone or 17β -oestradiol. *Animal Reproduction Science*, 37: 109-119.
- Day, M.L., Burke, C.R., Taufa V. K., and Day A. M and Macmillan K. L (2000). The strategic use of estradiol to enhance fertility and submission rates of progestin base estrus synchronization program in dairy herds. *Journal of Animal Science*, 73: 523-529.
- Dayem, A.M.H., Mahmoud, K.G.h.M., Nawito, M.F., Ayoub, M.M., and Scholkamy, T.H. (2009). Fertility evaluation in Egyptian buffalo bulls using zona pellucida binding and in vitro fertilization assays. *Livestock Science* 122: 193-193.
- de Leeuw, B., Berger, W., Sinke, R.J., Suijkerbuijk, R.F., Gilgenkrantz, S., Geraghty, M.T., Valle, D., Monaco, A.P., Lehrach, H., and Ropers, H.H. (1993). Identification of a yeast artificial chromosome (YAC) spanning the synovial sarcoma-specific t(X;18) (p11.2;q11.2) breakpoint. *Genes Chromosomes Cancer*, 6: 182-189.
- Dickerson, G. (1970). Efficiency of animal production--molding the biological components. *Journal of Animal Science*, 30: 849-859
- Dimmick, M.A., Gimenez, T., and Spitzer, J.C. (1991). Ovarian endocrine activity and development ovarian follicles during the post-partum interval in beef cows. *Animal Reproduction Science*, 24: 173-183.
- Driancourt, M.A. (2001). Regulation of ovarian follicular dynamics in farm animals: Implications for manipulation of reproduction. *Theriogenology*, 55: 1211-1239.
- Dziuk, P.J., and Bellows, R.A. (1983). Management of reproduction of beef cattle, sheep, and pigs. *Journal of Animal Science*, 57: 355-379.
- Eagles, V.E., Malmo, J., and Macmillan, K.L. (2001). Resynchronising returns to-service in anoestrus cows in Victorian dairy herds. *NZ Soc. Animal Production*, 61: 176-179.
- El-Zarkouny, S.Z., and Stevenson, J.S. (2004). Resynchronizing estrus with progesterone or progesterone plus estrogen in cows of unknown pregnancy status. *Journal of Dairy Science*, 87: 3306-3321.
- Engelmann, U., Parsch, E.M., and Schill, W.B. (1989). Modern techniques of sperm preparation--do they influence the sex of offspring. *Journal of Andrologia*, 21 (6): 523-528.

- Erickson, B.H. (1966). Development and senescence of the postnatal bovine ovary. *Journal of Animal Science*, 25: 800-805.
- Ericsson, R.J., Langevin, C.N., and Nishino, M. (1973). Isolation of fractions rich in human Y sperm. *Journal of Nature*, 246: 421-424.
- Erlich, H.A., and Arnheim, N. (1992). Genetic analysis using the polymerase chain reaction. *Annal Reolution Genetic*, 26: 479-506.
- Erlich, H.A., Gelfand, D., and Sninsky, J.J. (1991). Recent advances in the polymerase chain reaction. *Journal of Science*, 252: 1643-1651.
- Esteso, M.C., Fernández, S.M.R., Soler, A.J., and Garde, J.J. (2003). Head dimensions of cryopreserved red deer spermatozoa are affected by the thawing procedure. *Cryoletters*, 24: 261-268.
- Evans, G., and Maxwell, W.M.C. (1987). In: Maxwell, W. M. C. (Ed). Salamon's artificial insemination of Sheep and Goat. Butterworths. Sydney
- Evenson, D.P., Park, J.E., Kaproth, M.T., and Jost, L.K. (1993). Rapid determination of sperm cell concentration in bovine semen by flow cytometry. *Journal of Dairy Science*, 76: 86-94.
- Farahvash, T., Adabi, S.G., Adabi, A., Ahmadzadeh., Davoodi, A., and Ahmadzadeh and J. Davoodi. (2008). Some factors affecting sex ratio of dairy herds in East Azarbajan Iran. *Asian J. Animal Veterinary Advences*, 3: 357-362.
- Fatehi, J., and Schaeffer, L.R. (2003). Data management for the fertility project. Report to the Technical Committee of the Canadian Genetic Evaluation Board. J. Canada.
- Fazeli, A., Hage, W.J., Cheng, F.P., Voorhout, W.F., Marks, A., Bevers, M.M.(1997). *Colenbrander B.* Acosome intact boar spermatozoa initiate binding to the homologous zone pellucid in vitro. *Biology of Reproduction*, 56, 430 - 438
- Flaherty, S.P., J. Michalowska., N. J. Swann., W. P. Dmowski., and C. D. Matthews, a.R.J.A. (1997). Albumin gradients do not enrich Y-bearing human spermatozoa. *Human Reproduction*, 12: 938-942.
- Flores, R., Loper, M.L., Kreider, D.L., and Post, N.M., and Rosenkrans Jr C. F., (2006). Estrus behavior and initiation of estrus cycles in postpartum Brahman-influenced cows after treatment with progesterone and

- prostaglandin F2alpha/progesterone and prostaglandin F2alpha. *Journal of Animal Science*, 84: 1916-1925.
- Friscke, P.M., Caraviello, D.Z., and Weigel, K.A. (2003). Fertility of dairy cows after resynchronization of ovulation at three intervals following first timed insemination. *Journal of Dairy Science*, 86: 3941-3950.
- Galina, C.S., and Arthur, G.H. (1990). Review of cattle reproduction in the tropics. Part 4. Oestrus cycles. *Journal of Animal Breed. Abstr*, 58: 697-707.
- Galina, C.S., Hom, M.H., and Molina, R. (2007). Reproductive behavior in bulls raised under tropical and subtropical conditions. *Hormone Behavior*, 52: 26-31.
- Garcia, M.C., McDonnell, S. M., Kenney, R. M. (1986). Bull sexual behavior test: Stimulus cow affects performance. *Journal of Applied Animal Behavior Science*, 16. 1-10
- Garner, D.I., Gledhill, B.L., Pinkel, D., Lake, S., Sethepenshon, D., and VanDilla, M.A. (1983). Quantification of the X -and Y - chromosome bearing sperm of domestic animals by flow cytometry. *Biology of Reproduction*, 28: 312-321.
- Garner, D.L., and Johnson, L.A. (1995). Viability assessment of mammalian sperm using sybr-14 and propidium iodine. *Biology of Reproduction*, 53;544 -49.
- Garner, D.L., and Hafez, E.S.E. (2000). Spermatozoa and seminal Plasma. In: Reproduction in farm Animals. 7 th edition. (pp 96-123). New York. Lippincott Williams and Wilkins.
- Garner DL and Seidel GE (2008). History of Commercializing sexed semen for Cattle. *Theriogenology*, 68: 886-895.
- Garner, D.L., Thomas, I.L.C.A., Gravance, C.G., Marshall, C.E., DeJarnette, J.M., and Allen, C.H. (2001). Seminal plasma addition attenuates the dilution effect in bovine sperm. *Theriogenology*, 56: 31-40.
- Garverick, H.A., and Smith, M.F. (1986). Mechanisms associated with subnormal luteal function. *Journal of Animal Science*, 62(Suppl. 2): 92-105.
- Garverick, H.A., Zollers Jr, W.G., and Smith, M.F. (1992). Mechanisms associated with corpus luteum lifespan in animals having normal or subnormal luteal function. *Animal Reproduction Science*, 28: 111-124.

- Gerrard, O., and Humblot, P. (1991). Influenze of interaction between semen extender and number of spermatozoa on non return rate estimates of fertility for individual Holstein bull. *Theriogenology*, 36: 727-736.
- Ginther, O.J., Bergfelt, D.R., Kulick, L.J., and Kot, K. (2000). Selection of the dominant follicle in cattle: Role of two-way functional coupling between follicle- stimulating hormone and the follicles. *Biology of Reproduction*, 62: 920-927.
- González, S.C., Soto, E., Goicochea, J., and González, R.a.S., G.(1988). Identificación de los factores causales y control del anestro, principal problema reproductivo en la ganadería mestiza de doble propósito. Premio Agropecuario Banco Consolidado, Caracas, Venezuela.
- Gordon, I. (2005). Reproductive Technologies in Farm Animals. CABI, Oxfordshire OX10 UK.
- Graham, J.K. (2001). Assessment of sperm quality: a flow cytometric approach. *Animal Reproduction Science*, 68: 239-247.
- Graham, J.K., and Moce, E. (2005). Fertility evaluation of frozen - thawed semen. *Theriogenology*, 64: 492 - 504.
- Gupta, S.K., and Bhandari, B. (2011). Acrosome reaction: relevance of zona pellucida glycoproteins. *Asian Journal of Andrology*, 13: 97-105.
- Gutierrez-Adan, A., G. , Perez, J., Granados, J.J., Garde, M., Perez-Guzman, B., Pintado, and De La Fuente. (1999). Relationship between sex ratio and time of insemination according to both time of ovulation and maturational state of oocyte. *Journal of Zygote*, 7: 37-43.
- Hadi, P.U., and Ilham, N. (2000). Peluang pengembangan usaha pembibitan ternak sapi potong di Indonesia dalam rangka swasembada daging. In: Lap. Directur Pembibitan derectur jendral bina produksi Peternakan. Pp. 11-12. Jakarta, Indonesia.
- Hafez,, H. S. E.(2000). Preservation and Cryopreservation of Gametes and Embryos Reproductive Behavior.In: Reproduction in farm Animals. 7 ed. pp. (431-441).New York. Lippincott Williams and Wilkens.
- Hafez, B., and Hafez, E.S.E. (2000). Reproductive Behavior.In: Reproduction in farm Animals. 7 ed.pp. (293-306) New York Lippincott Williams and Wilkens,,

- Hamano, K., Li, X., Qiant, Q., Funuachi, K., Furudate, M., and Minato, Y. (2001). Sex pre selection in farm animal by flow cymetric separation of X- and Y-chromosome bearing spermatozoa *Mammalia Ova Research*, 18: 81-88.
- Hamano, K.X., Li, X., and Minato, Y. (2007). Sex preselection in bovine by Separation of X-and Y-Chromosome Bearing Spermatozoa. *Reproduction and Development*, 53: 27-38.
- Hanlon, D.W., Wichtel, J.J., Xu, Z.Z., and Burton, L.J. (2000). The reproductive performance of anoestrus dairy cows following treatment with progesterone and oestradiol prior to the start of mating. *NZ Veterinary Journal*, 48: 136-143.
- Henkel, R. R and Schill, W. B. (2003). Spermatology preparation for assisted reproduction technology (ART). *Reproductive Biology and Endocrinology*. 1:108 doi: 10.1186/2477-7827-1-108.
- Hansel, W., and Convey, E.M. (1983). Physiology of the estrus cycle. *Journal of Animal Science*, 57: 404-424.
- Harrison, R.A., and Vickers, S.E. (1990). Use of fluorescent probes to assess membrane integrity in mammalian spermatozoa. *Reproductive and Fertility*, 88: 343-352.
- Hayashi, T., and Matsukawa, T. (1979). Free amino - acid composition of bovine genital tract fluid (cervical mucus, uterine fluid). *Japan Animal Reproduction*, 25: 1-5.
- Helbig, L., Woodbury, M. R., Haigh, J. C and Marth, A. D. (2007). The onset of puberty in North American bison (*Bison bison*) bulls. *Animal Reproduction Science*. 96:12-24.
- Herschler. R. C. (1983). Estrus synchronization and conception rates in first heifers using fenprostalene in both single- and double-injection programs. *Journal of Agri-Practice*, 4:28.
- Hoflack, G., Van Soom, A., Maes, D., de Kruif , A., Opsomer, G., Duchateau, L. (2006). Breeding soundness and libido examination of Belgian Blue and Holstein Friesian artificial insemination bulls in Belgium and The Netherlands. *Theriogenology*, 66. 207–216

- Holden, C.A., Hyne, R.V., and Sathananthan, A.H. (1990). Trounson AO. Assessment of the human sperm acrosome reaction using concanavalin A lectin. *Molecular Reproduction Development*, 25: 247-257.
- Holt, W.V. (2000). Basic aspects of frozen storage of semen. *Animal Reproduction Science*, 62: 3-22.
- Hong, J.H.U., Wang, Q.L.I., Chen, Y.L., Jlang, Z.L., Jia, Y.H., Wang, L.Q., and Ou, B.B. (2009). Effects of addition of vitamin B12 to the extender on post-thaw motility, acrosome morphology, and plasma membrane integrity in bull semen. *Turk J Veterinary and Animal Science*, 35: 379 - 384.
- Hopkins, F.M., and Spitzer, J.C. (1997). The new society for theriogenology breeding soundness evaluation system. *Vet Clin North Am Food Animal Practice*, 13: 283-293.
- Hossepien de Lima ,V.F.M., Ramalho, M.D.T, Rodrigues, L.H., Malheiros, E.B, Moreira-Filho, C.A. (2000). Separation of X- and Y- bearing bovine spermatozoa by Percoll density gradient centrifugation. *Theriogenology*, 53:480-489
- Houghton, P.L., Lemenager, R.P., Hendrix, K.S., Mos, G.E., and Sterwart, T.S. (1990). Effects of body composition pre- and postpartum energy intake and stage of production of energy utilization by beef cows. *Journal of Animal Science*, 68: 1447 - 1456
- Hu, J.H., Li, Q.W., Li, G., Jiang, Z.L., Bu, S., Yang, H., and Wang, L.Q. (2009). The cryoprotective effects of trehalose supplementation on boar spermatozoa quality. *Animal Reproduction Science*, 112: 107-118.
- Hung, P.H., Miller, M.G., Mayers, S.A., and VandeVoort, C.A. (2008). Sperm mitochondrial integrity is not required for hyperactivated motility, zone binding, or acrosome reaction in the Rhesus macaque. *Biology of Reproduction*, 79: 367-375.
- Hunter, R.H. (1975). In *The Biology of spermatozoa*. (ads E.S.E. Hafez and C.F. Thiboult) p.145. Kargel, Basel.
- Iizuka, R., Kaneko, S., Aoki, R., and Kobayashi, T. (1987). Sexing of human sperm by discontinuous Percoll density gradient and its clinical application. *Human Reproduction*, 2(7): 573-575.

Jainudeen, M.R., and Hafez, E.S.E. (2000a). Cattle and Buffalo. In: Reproduction in farm Animals. 7 th edition. (pp 159-170). New York. Lippincott Williams and Wilkins

Jainudeen, M.R., and Hafez, E.S.E. (2000b). Reproductive failure in females. In: Reproduction in farm Animals. 7 th edition. (pp 261-278). New York. Lippincott Williams and Wilkins .

Jainudeen, M.R., Wahid, H., and Hafez, E.S.E. (2000). Ovulation induction, embryo,Production and Transfer. In:Reproduction in Farm Animals. 7 ed Lippincott pp. (405-430) New York, Lippincott, Williams and Wilkens,

Januskauskas, A., Johannisson, A., and Rodriguez-Martinez, H. (2001).Assessment of sperm quality through fluorometry and sperm chromatin structure assay in relation to field fertility of冻融 semen from Swedish AI bulls. *Theriogenology*, 55: 947-961.

Januskauskas, A., Johannisson, A., and Rodriguez-Martinez, H. (2003). Subtle membrane changes in cryopreserved bull semen in relation with sperm viability, chromatin structure, and field fertility. *Theriogenology*, 60: 743-758.

Januskauskas, A., Soderquist, L., Haard, M.G., Haard, M.C., Lundeheim, N., and Rodriguez-Martinez, H. (1996). Influenze of sperm number per straw on the post-thaw sperm viability and fertility of swedish redand white AI bull. *Acta Veterinary Scandaniva*, 37: 461-470.

Jasko, D.J. (1992). Evaluation of stallion semen.The Veterinary Clinics of North America.*Equine practice*, 8: 129-148.

Jeyendran, R.S., Van der Ven , H.H., Pe'rez Pela'ez , M., Crabo, B., G., , and Zaneveld, L.J.D. (1984). Development of an assay to assess the functional integrity of the human sperm membrane and its relationship to the other semen characteristics. *Reproduction and Fertility*, 70: 219-228.

Joerg, B.H., Asali, M., Graphodatskaya, D., Janett, F., and Stranzinger, G. (2004). Validating bovine sexed semen samples using quantitative PCR. *Animal Breeding Genetic*, 121: 209-215.

Johnson, L.A. (1995). Sex Preselection by Flow Cytometric separation of X- and Y-chromosome bearing sperm based on DNA difference: a review. *Reproductive Fertility Development*, 7: 893-903.

- Johnson, L.A. (2000). Sexing mammalian sperm for production of offspring: the state-of-the-art. *Animal Reproduction Science*, 60-61: 93-107.
- Jones, A. L. and Lamd, G. C. (2008). Nutrition, synchronization, and management of beef embryo transfer recipients. *Theriogenology*, 69:107-115.
- Kandukuri, S.P., Rao, B., and Masson, P.L. (1977). Study of the primary structures of the peptide core of bovine oestrus cervical mucus. *Biology of Chemical*, 252: 7788-7795.
- Kaneko, S., J. , Yamaguchi, T., Kobayashi., and Iizuka, R. (1983). Separation of human spermatozoa X- and Y-bearing sperm using percoll density gradient centrifugation. *Fertility and Sterility*, 40: 661-665.
- Kastelic, J.P., Curran, S., Pierson, R.A., and Ginther , O.J. (1988). Ultrasonic evaluation of the bovine conceptus. *Theriogenology*. 29: 39-54.
- Katayama, K.P., Stehlik, E., and Jeyendran, R.S. (1989). In vitro fertilization outcome: glass wool-filtered sperm versus swim-up sperm. *Fertility and Sterility*, 52: 670-672.
- Kawasaki, T., Sone, M., Yoshida, M., and Bamba, K. (1996). Rapid and simultaneous detection of chromosome Y- bearing porcine spermatozoa by fluorescence in situ hybridization. *Molecular Reproduction Developmant*, 43: 548-553.
- Khatamee, M.A., Horn, S., Weseley, A., Farooq, T., Jaffe, S.B., and Jewelewicz, R. (1999). A controlled study for gender selection using swim-up separation. *Gynecol Obstet Invest*, 48: 17-13.
- Kiracofe, G. H..L. E. Keay and K. G. Odde. (1985). Synchronization of estrus in cyclic beef heifers with the prostaglandin analog alfa prostol. *Theriogenology* 24:737
- Klemm, W.R., Hawkins, G.N., and Santos, D.L. (1986).Identification of compounds in bovine cervico-vaginal mucus estrus that evoke male sexual behavior. *Chemical Science*, 12: 11-18.
- Kobayashi, J., Oguro, H., Uchida, H., Koshaka, T., Hasada, H., and Sato, E. (2004). Assessment of bovine X- and Y- bearing spermatozoa infractions by discontinuous percoll gradients with rapid fluorescence in situ hybridization. *Reproduction and Developmant*, 50: 463-469

- Kojima, F.N. (2003). The estrus cycle in cattle: Physiology, endocrinology, and follicular waves. *The proof Animal Science*, 19: 83-95.
- Kommisrud, E., Graffer, T., and Steine, T. (1996). Comparison of two processing systems for bull semen with regard to post-thaw motility and nonreturn rates. *Theriogenology*, 45: 1515-1521.
- Krininger, C.E., Block, J., Al-Katanani, Y.M., Rivera, R.M., Chase C.C., and Hansen, P.J. (2003). Differences between Brahman and Holstein cows in response to estrus synchronization, superovulation and resistance of embryos to heat shock. *Animal Reproduction Science*, 78: 13-24.
- Kumi-Diaka, J. (1993). Subjecting canine semen to the hypo-osmotic test. *Theriogenology*, 39: 1289-1993.
- Kuster, C. (2005). Sperm concentration determination between hemacythometric and CASA systems: why the can different. *Theriogenology*, 64: 614-617.
- Lamb, G.C. (2010). Estrus synchronization protocols for cows. In Applied reproductive strategies in Beef Cattle. San Antonio, TX. January 28-29, pp. 113-128.
- Lamb, G.C., Stevenson, J.S., Kesler, D.J., Garverick, H.A., and Brown , D.R.a.S.B.E. (2001). Inclusion of an intravaginal progesterone insert plus GnRH and prostaglandin F2 alpha for ovulation control in postpartum suckled beef cows. *Journal of Animal Science*, 79: 2253-2259.
- Lamb, G.C., Larson, J.E., Geary, T.W., Stevenson, J.S., Johnson, K.J., Day, M.L., Ansotegui, R.P., Kesler, D.J., DeJarnette, J.M., and Landblom, D.G. (2006). Synchronization of estrus and artificial insemination in replacement beef heifers using gonadotropin-releasing hormone, prostaglandin F 2α , and progesterone. *Journal of Animal Science*, 84(3000-3009).
- Larson, J.E., Lamb, G.C., Stevenson, J.S., Johnson, S.K., Day, M.L., Geary , T.W., Kesler, D.J., Dejarnette, J.M., Schrick , F., and Costanzo A and Arseneau, J.D. (2006). Synchronization of estrus in suckled beef cows for detected estrus and artificial insemination and timed artificial insemination using gonadotropin-releasing hormone, prostaglandin F 2α , and progesterone. *Journal of Animal Science*, 84: 332-342.
- Leitmana, N.R., Buscha , D.C., Mallorya, D.A., Wilsona, D.J., Ellersieckb, M.R., Smitha , M.F., and Pattersona D. J. (2009). Comparison of long-term CIDR-based protocols to synchronize estrus in beef heifers. *Animal Reproduction. Science*, 114: 345-355.

- Loskutoff, N.M., and Crichton, E.G. (2001). Standard operating procedures for genome resource banking. The Bill and Bernice Grawcock center for conservation and research. pp1-16, Omaha's Henry Doorly Zoo.
- Lucy, M.C. (2001). Reproductive loss in high-producing dairy cattle: where will it end. *Journal of Dairy Science*, 84: 1277-1293.
- Macmillan, K.L., and Watson, J.D. (1975). Fertility differences between groups of sires relative to the stage of oestrus at the time of insemination. *Journal of Animal Science*, 21: 243-249.
- Macmillan, K.L., Taufa, V.K., Barnes, D.R., and Day, A.M. (1991). Plasma progesterone concentrations in heifers and cows treated with a new intravaginal device. *Animal Reproduction Science*, 26: 25-40.
- Macmillan, K.L., Taufa V. K., Day A. M., and Eagles, V.E. (1999). Some effects of post estrus hormonal therapies on conception rates and resubmission rates in lactating dairy cows In In: Fertility in the high- producing Dairy cows Br.Soc.Animal Sci, pp. pp.195-208.
- Macoda, M., Carvalho, J. O., Siqueira Filho, E., Caixeta, E.S., Franco, M. M., Rumpf, R and Dode, M. A. N. (2009). Effect of Percoll volume, duration and force of centrifugation, on in vitro production and sex ratio of bivine embryos. *Theriogenology*, 71:1289-1297.
- Madrid, B.N., Raul, F., Adela Jimenez, S., P. G., , Pedro, N.M., and Belen Pintado, J., F and Alfonso, G. A., (2003). Effect of ejaculate, bull, and a double swim-up sperm processing method on sperm sex ratio. *Journal of Zygote*, 11: 229-235.
- Maffeo, G.R. Ballabio, V. Olgiati and F. Guidobono. (1983). Induction of estrus in cows by a new analogue of PGF_{2α} (alfaprostol). *Prostaglandins*, 25:541.
- Mammoto, A., Masumoto, N., Tahara, M., Ikebuchi, Y., Ohmichi, M., Tasaka, K., and Miyake, A. (1996). Reactive oxygen species block sperm-egg fusion via oxidation of sperm sulphhydryl proteins in mice. *Biology of Reproduction*, 55: 1063-1068.
- Manjunath, P., Nauc, V., Bergeron, A., and Menard, M. (2002). Major proteins of bovine seminal plasma bind to the low densitylipoprotein fraction of hen's egg yolk. *Biology of Reproduction*, 67: 1250-1258.
- Maquivar, A., Verduzco, C.S.G., A., Pulido S., Rojas K, Forster, G., Van der Laan., and Arnoni., R. (2007). Relationship Among Follicular Growth, Oestrus,

- Time of Ovulation, Endogenous Estradiol 17 β and Luteinizing Hormone in Bos Indicus Cows After a Synchronization. *Reproduction Domestic Animal*, 42: 571-576.
- Martin Rillo, S., Martinez, A., Garcia-Artiga, C., and De Alba, C. (1996). Boer semen evaluation in practise. *Reproduction Domestic Animal*, 31: 519-526.
- Mazur, P. (1984). Freezing of living cells: Mechanisms and implications. *Am Journal Physiology*, 247: 125-142.
- McCracken, J.A., Custer, E.E., and Lamsa, J.C. (1999). Luteolysis: A neuroendocrine- mediated event. *Physiology Reviews*, 79: 263-324.
- McCracken, J.A., Custer, E.E., Eldering, J.E., and Robinson, A.G. (1996). The central oxytocin pulse generator: A pacemaker for the ovarian cycle. *Acta Neurobiol. Exp.*, 56: 819-832.
- McDougall, S., and Loeffler, S.H. (2004). Resynchrony of postpartum dairy cows previously treated for anestrus. *Theriogenology*, 61: 239-253.
- Mehmood, A., Anwar, M., and Saqlan Naqvi, S.M. (2008). Motility, acrosome integrity, membrane integrity and oocyte cleavage rate of sperm separation by swim-up or percoll gradient method from frozen-thawed buffalo semen. *Animal Reproduction Science*, DOI:10.1016/j.anireprosci.2008.02.011.
- Mendoza, C., Carreras, A., Moos, J., and Tesaric, J. (1992). Distinction between true acrosome reaction and degenerative loss by a one step staining method using Pisum Sativum agglutinin. *Reproduction and Fertility*, 95: 755-763.
- Meneghetti, M., Sá Filho, O.J., Peres, R., Lamb, G., and Vasconcelos, J.L.M. (2009). Fixed-time artificial insemination with estradiol and progesterone for Bos indicus cows. I. Basis for development of protocols. *Theriogenology*, 72: 179-189.
- Molina, R., Galina, C.S., Maquivar, M., Estrada, S., Chavez, A., and Diaz, G.S. (2003). Pregnancy rate in zebu cows with two different postpartum interval exposed to a two-Bull rotational system. *Veterinary Research Comm*, 27: 671-680.
- Molina, R., Bolaños, I., Galina, C. S., Pérez, E., Paniagua, G., Estrada, S. (2000). Sexual behaviour of Zebu bulls in the humid tropics of Costa Rica: single versus multiple-sire groups. *Animal Reproduction Science*.64.139–148.

- Molina, R., Galina, C.S., Martinez, J., Monge, M., (1997). Sexual behavior of Zebu bulls with females in either natural or induced estrus. *Int. Journal of Animal Science*, 12, 181–186.
- Moreire, F., Risco, C.A., Pires, M., F. A., Ambrose, J.D., Drost, M., and Thatcher, W.W. (2000). Use of bovine somatotropin in lactating dairy cows receiving timed artificial insemination. *Journal of Dairy Science*, 83: 1237-1247.
- Morton, K.M., Evans, G., and Maxwell, W.M.C. (2010). Effect of glycerol concentration, Equex STM® supplementation and liquid storage prior to freezing on the motility and acrosome integrity of frozen-thawed epididymal alpaca (*Vicugna pacos*) sperm. *Theriogenology*, 74: 311–316.
- Muino, R., Tamargo, C., Hidalgo, C, O. and Pena, A, I. (2008). Identification of sperm subpopulations with defined motility characteristics in ejaculates from Holstein bulls:Effect of cryopreservation and between-bull variation. *Animal Reproduction Science*, 109:27-39.
- Moruzzi, J.F. (1979). Selecting a mammalian species for the separation of X- and Y-chromosome bearing spermatozoa. *Reproductive and Fertility*. 57: 319-323.
- Munoz, O.V.L., Amirat-Briand, T., Diaz, L., Va'squez, E., Schmidt, S., Desherces, M., Anton, D., Bencharif, D., and Tainturier.(2009). Effect of semen dilution to low-sperm number per dose on motility and functionality of cryopreserved bovine spermatozoa using low-density lipoproteins (LDL) extender: Comparison to Triladyl and Bioxcell.*Theriogenology* 71(895-900).
- Mushayandebvu, T.I., Santoro, N.F., Lipetz, K.J., and Colon, J.M. (1995). Comparison of Percoll and swim up techniques for sperm recovery in patients with male factor infertility. In: IX Word Congress on in vitro fertilization and alternative assisted reproduction. Mondizzi Editore.Bologna, Italy: pp.577-580.
- Naing, S.W., Wahid, H., Azam, K.M., Rosnina, Y., Zuki, A.B., Kazhal, S., Bukar, M.M., Thein, M., Kyaw, T., and San, M.M. (2010). Effect of sugars on characteristics of Boer goat semen after cryopreservation. *Animal Reproduction Science*, 122(1-2): 23-28.
- Niles, D., Risco, C.A., and Thatcher, M.J. (2002). Seasonal evaluation of artificial insemination and natural service pregnancy rates in dairy herds. Compend.Contin. *Veterinary Medicine Practice Education*, 24: 544-548.

- Nur, Z., Dogan, I., Gunay, U., and Soylu, M.K. (2005). Relationship between sperm membrane integrity and others semen quality characteristics of the semen Saanen goat bucks. *Bull Veterinary Inst Pulawy*, 49: 183-187.
- Obando, H., Tamayo, T., and Alvaro, C. (1984). Evaluation of some factors affecting swine spermatozoa during freezing. In: Proceedings of the Tenth International Congress on Animal Reproduction AI, Vol 2: Urbanda, p.193.
- O'Brien, J.K., Steinman, K.J. and Robeck, T.R. (2009). Application of sperm sorting and associated reproductive technology for wildlife management and conservation. *Theriogenology*, 71: 98–107.
- Odde, K. G. (1990). A review of synchronization of estrus in postpartum cattle. *Journal of Animal Science*, 68:817-830.
- Olivera-Angel, M., and Martínez, G. (1990). Evaluation of an implant to synchronize oestrus and/or to resolve suckling anoestrus in Brahman cows In: Livestock Reproduction in Latin America, IAEA: 221-225.
- Overton, M.W., and Sischo, W.M. (2005). Comparison of reproductive performance by artificial insemination versus natural service sires in California dairies. *Theriogenology*, 64: 603-613.
- Pace, M.M., Sullivan, J.J., Elliott, P.I., Graham, E.F., and Coulter, G.H. (1981). Effects of thawing temperature, number of spermatozoa and spermatozoal quality on fertility of bovine spermatozoa packaged in 5-ml French straw. *Journal of Animal Science*, 53: 693-701.
- Paleologou, A.M. (1979). A study of the cervicovaginal secretions of cows during the different phase of the estrus cycle. *Inst.itite Animal Technicians*, 30: 83-94.
- Palmer, C.W., Barth, A.D. (2003). Comparison of the BullMateTM sperm quality analyzer with conventional means of assessing the semen quality and breeding soundness of beef bulls. *Animal Reproduction Science*, 77:173–185.
- Parrish, J.J., Kroghnaes, A., and Susko-Parrish, J.L. (1995). Effect of bovine sperm separation by either swim-up or Percoll method on success of in vitro fertilization and early embryonic development. *Theriogenology*, 44: 859-869.
- Parrish, J.J., Susko-Parrish, J., Winer, M.A., and First, N.L. (1988). Capacitation of bovine sperm by heparin. *Biology and Reproductive*, 38: 1171-1180.

- Patterson, D.J., Kojima, and F. N. and Smith. M. F. (2003). Methods to synchronize estrus cycles of postpartum beef cows with melengestrol acetate. *Journal of Animal Science*, 19: 109-115.
- Peeler, I. D., R. L. Nebel, R. E. Pearson, W. S. Swecker, and A. Garcia. (2004). Pregnancy rates after timed AI of heifers following removal of intravaginal progesterone inserts. *Journal of Dairy Science*, 87:2868–2873.
- Pe'rez-Llano, B., Sala, R., Reguera, G., and Garcí'a-Casado, P. (2009). Changes in subpopulations of boar sperm defined according to viability and plasma and acrosome membrane status observed during storage at 15 °C. *Theriogenology*, 73: 311–317.
- Pena, F.J. Johannsson, A., Wallgren, M., and Rodriguez-Martínez, H. (2003a). Assessment of fresh and frozen-thawed boar semen using an annexin-v assay: a new method of evaluating sperm membrane integrity. *Theriogenology*, 60: 677-689.
- Pena, F.J., Johannsson, A., Wallgren, M., and Rodriguez-Martínez, H. (2003b). Antioxidant supplementation in vitro improves boar sperm motility, and mitochondrial membrane potential after cryopreservation of different fractions of the ejaculate. *Animal Reproduction Science*, 78: 85-98.
- Perry, G.A., Smith, M.F., and Geary, T.W. (2004). Ability of intravaginal progesterone inserts and melengestrol acetate to induce estrus cycles in postpartum beef cows. *Journal of Anial Science*, 82: 695-704.
- Pertoft, H., Laurent, T.C., La°a°s, T., and Ka°gedal L. (1978). Density gradients prepared from colloidal silica particles coated by polyvinylpyrrolidone (Percoll). *Anal Biochem*, 88: 271-282.
- Pierson, R.A. (1984). Ultrasonography for detection of pregnancy and study of embryonic development in heifers. *Theriogenology*, 22: 225-235.
- Pierson, R.A., and Ginther, O.J. (1984). Ultrasonography of the bovine ovary. *Theriogenology*, 21: 495-504.
- Polge, C., Salamon, S., and Wilmut, I. (1970). Fertilizing capacity of frozen boar semen following surgical insemination. *Veterinary Reserach*, 87: 434-428.
- Ponthier, J., Franck, T., Detilleux, J., Mottart, E., Serteyn, D., and Deleuze, S. (2010). Association between Myeloperoxidase Concentration in Equine Frozen Semen and Post-Thawing Parameters. *Reproduction in Domestic Animals*, 45(5): 811-816.

- Prathalingan, N.S., Holt, W.V., Revel, S.G., Jones, S., and Watson, P.F. (2006). The precision and accuracy of six different methods to determine sperm concentration. *Journal of Andrology*, 27: 257-262.
- Purdy, P.H. (2006). A review on goat sperm cryopreservation. *Small Ruminant Research*, 63: 215-225.
- Pursel, V.G., and Johnson, L.A. (1975). Freezing of boar spermatozoa: fertilizing capacity with concentrated semen and a new thawing procedure. *Journal of Animal Science*, 40: 99-102.
- Quinlivan, W.L., Preciado, K., Long, T.L., and Sullivan, H. (1982). Separation of human X and Y spermatozoa by albumin gradients and Sephadex chromatography. *Fertility and Sterility*, 37: 104-107.
- Rajamahendran, R., Robinson, J., Desbottes, S. and Walton, J.S. (1989). Temporal relationships among estrus, body temperature, milk yield, progesterone and luteinizing hormone levels, and ovulation in dairy cows. *Theriogenology*, 31(6): 1173-1182.
- Randel, R.D. (1981). Effect of once-daily suckling on post-partum interval and cow-calf performance of first-calf Brahman×Hereford heifers. *Journal of Animal Science*, 53: 755-757.
- Randel, R.D. (1990). Nutrition and post-partum rebreeding in cattle. *Journal of Animal Science*, 68: 853-862.
- Rasby, R.J., Day, M.L., Johnson , S.K., Kinder , J.E., Lynch , J.M., Short , R.E., Wettemann, R.P., and Hafs, H.D. (1998). Luteal function and estrus in peripubertal beef heifers treated with an intravaginal progesterone releasing device with or without a subsequent injection of estradiol. *Theriogenology*, 50: 55-63.
- Rasul, Z., Ahmad, N., and Anzar, M. (2001). Changes in motion characteristics, plasma membrane integrity, and acosome morphology during cryopreservation of buffalo spermatozoa. *Journal of Andrology*, 22(2): 278-283.
- Rhodes, F.M., Fitzpatrick, L.E., Entwistle, K.W., and Kinder, J.E. (1995). Hormone concentrations in the caudal vena cava during the first ovarian follicular wave of the oestrus cycle in heifers. *Reproductive and Fertility*, 104: 33-39.
- Ricardo C., Chebel., José, E.P., Santos, James, P., Reynolds, Ronaldo, L.A., Cerri, Sérgio, O., Juchem, and Overton, M. (2004). Factors affecting conception

- rate after artificial insemination and pregnancy loss in lactating dairy cows. *Animal Reproduction Science*, 84: 239-255.
- Rodriguez-Martínez, H. (2003). Laboratory semen assessment and prediction of fertility. *Reproduction Domestic Animal*, 38: 312-318.
- Rodriguez-Martínez, H., Larson, J.E., and Pertoft, H. (1997). Evaluation of sperm damage and techniques for sperm clean up. *Reproductive Fertility Developmant*, 9: 297-308.
- Rosenkranz, H.S., Mednis, A., and Marks., P.A., and Rose, H . M. (2002). Metabolic effects of phenethyl alcohol on mammalian cells. *Biochemical Biophys*, 149(2).123-127.
- Rota, A., Penzo, N., Vincenti, L., and Mantovani, R. (2000). Hypo-osmotic swelling (HOS) as ascreening assay for testing in vitro fertility of bovine spermatozoa. *Theriogenology*, 53: 1415-1420.
- Roth, T.L., Weiss, R.B., Buff , J.L., Wildt, D.E., and Bush, L.M. (1998). Heterologous in vitro fertilization and sperm capacitation in an endangered African antelope the scimitar-horned oryx (*Oryx dammah*). *Biology Reproduction*, 58: 475-482.
- Rutter, L.M., and Randel, R.D. (1984). Post-partum nutrient intake and body condition: effect on pituitary function and onset of estrus in beef cattle. *Animal Reproduction Science*, 58: 265-275.
- Sá Filho, M.F., Crespilho, A.M., Santos, J.E.P., Perry, G.A., and Baruselli, P.S. (2010). Ovarian follicle diameter at timed insemination and estrus response influence likelihood of ovulation and pregnancy after estrus synchronization with progesterone or rogestin-based protocols in suckled *Bos indicus* cows. *Animal Reproduction Science*, 120: 23-30.
- Saacke, R.G., and White, J.M. (1972). Semen quality tests and their relationship to fertility. In: Proc. of the 4th NAAB Technical Conference on Artificial Insemination and Reproduction. pp, 22-27.
- Salisbury, G.W., Van Demark, N.L., and Lodge, J.R. (1978). Extender and extention of un frozen semen. In: Salisbury GW, editor. *Physiology of reproduction and artificial insemination of Cattle*. 2nd ed, San Francisco, W. H. Freeman and Company, 442-93.
- Samardzi, M., Karadjole, M., Matkovic, M., Cergolj, M., Getz, I., Dobranic, T., Tomaskovic, A., Petric, J., Surina, J., Grizelj, J., and Karadjole, T. (2006).

Comparison of bovi Pure[®] and Percoll[®] on bull sperm separation protocols for IVF. *Animal Reproduction Science*, 91: 237 - 247.

Sanchez Sarmiento, C.A., Coetzee, K., Kruger, T.F., van der Merwe, J.P., Stander, F.S.H., Henkel, R.R., and Lombard, C.J. (1996). Comparison between swim-up and glass wool column filtration of human semen in a gamete intrafallopian transfer program. *Journal of Arch. Andrology*, 36: 155-160.

Schafer, D.J., Bader, J.F., Meyer, J.P., Haden , J.K., Ellersieck, M.R., Lucy, M.C., and Smith, M.F.a.P., D. J., (2007). Comparison of progestin-based protocols to synchronize estrus and ovulation before fixed-time artificial insemination in postpartum beef cows. *Journal of Animal Science*, 85: 1940-1945.

Schallenberger, E., D., Schams, B., Bullermann., and Walters, D.L. (1984). Pulsatile secretion of gonadotrophins, ovarian steroids and ovarian oxytocin during prostaglandin-induced regression of the corpus luteum in the cow. *Reproductive Fertility*, 71: 493-501.

Schenk, J.L., Amann, R.P. and Allen, C.H. (1987). Influenze of semen collection interval and tactile stimuli on semen quality and sperm output in bulls. *Journal of Dairy Science*, 70: 1458-1464.

Seidel, G.E., Jr L.A. Herickhoff, J.L., Schenk, S.P., and Doyle and R.D. Green.(1998). Artificial insemination of heifers with cooled, unfrozen sexed semen. *Theriogenology*, 49: 365 (Abstr)

Seidel, G.E., Jr Allen, L.A., Johnson, M.D., Holland, Z., Brink, G.R., , Welch, J.K., and Graham and Cattell, M.B. (1997). Uterine horn insemination of heifers with very low numbers of non frozen and sexed spermatozoa. *Theriogenology*, 48: 1255- 1264.

Seidel, G.E., Jr D.G. Cran., , L.A. Herickhoff., J.L. Schenk, S.P., and Doyle and R.D. Green. (1999). Insemination of heifers with sexed frozen or sexed liquid semen. *Theriogenology*, 51: 400 (Abstr.).

Seidel, G.E., Jr. (2003). Sexing mammalian sperm--intertwining of commerce, technology, and biology. *Animal Reproduction Science*, 79(3-4): 145-156.

Seidel, G.E.J. (1995). Reproductive biotechnologies for profitable beef production. In: Proc. Beef Improvement Federation: Pp. 28 Sheridan, WY.

Seidel, J.R.G.E. (1988). Sexing mammalian sperm and embryos, In: Proceeding of the XIth International congress on animal reproduction and artificial insemination., 5.

Senger, P.L. (2003). Pathways to pregnancy and parturition Current Conceptions Inc., Pullman, WA. .

Sheldon, I.M., Lewis, G.S., LeBlanc, S., and Gilbert, R.O. (2006). Defining was postpartum uterine disease in cattle. *Theriogenology*, 65: 1516-1530.

Shibaya, M., Murakami, S., Tatsukawa, Y., Skarzynsky, D.J., Acosta, T.J., and Okuda, K. (2006). Bovine corpus luteum is an extrapituitary site of prolactin production. *Molecular Reproductive Developmant*, 73: 512-515.

Short, R.E., Bellows, R.A., Staigmiller, R.B., Berardinelli, J.G., and Custer, E.E. (1990). Physiological mechanisms controlling anestrus and infertility in postpartum beef cattle. *Journal of Animal Science*, 68: 799-816.

Silva, P. F. N and Gadella, B. M. (2006). Detection of demage in mammalian sperm cells. *Theriogenology*, 65:958-978.

Smith, R.D., Pomerantz, A.J., Beal, W.E., McCann, J.P., Pilbeam, T.E., and Hansel, W. (1984). Insemination of Holstein heifers at a preset time after estrus cycle synchronization using progesterone and prostaglandin. *Journal of Animal Science*, 58: 792-800.

Smith, T.T., Hosid, S., and AScott, L. (1995). Use of post separation sperm parameters to determine the methods of choice for sperm separation for assisted reproductive technology. *Fertility and Sterility*, 63: 591-597.

Soderquist, L., Janson, L., Larsson, K., and Einarsson, S. (1991). Sperm morphology and fertility in bulls. *Zentralbl Veterinarmed* 38: 534-543.

Son , D.S., Choe , C.Y., Cho, S.R., Choi, S.H., Kim, H.J., Hur, T.Y., Jung , Y.G., and Kang , H.G.a.K.I.H. (2007). A CIDR Based timed embryo transfer protocol increases the pregnancy rate of lactating repeat breeder dairy cows. *Reproduction Developmant*, 53: 1313-1318

Stagg, K., Diskin, M.G., Sreenan, J.M., and Roche, J.F. (1995). Follicular development in long-term anestrus suckled beef cows fed two levels of energy postpartum. *Animal Reproduction Science*, 38: 49-61.

Stalhammar, E.M., Janson, L., and Philipsson, J. (1994). The impact of sperm motility on non return rate in preselected dairy bulls. *Reproduction Nutrition Developmant*, 34: 37-445.

- Steeno, O., Adimoelja, A., and Steeno, J. (1975). Separation of X- and Y-bearing human spermatozoa with the sephadex gel-filtration method. *Journal of Andrology*, 7: 95-97.
- Stevenson, J.S., Kobayashi, Y., and Thompson, K.E. (1999). Reproductive performance of dairy cows in various programmed breeding systems including Ovsynch and combinations of gonadotropin-releasing hormone and prostaglandin F2 α . *Journal of Dairy Science*, 99: 506-515.
- Stevenson, J.S., Cartmill, J.A., Hensley, B.A., and El-Zarkouny, S.Z. (2003). Conception rates of dairy cows following early not-pregnant diagnosis by ultrasonography and subsequent treatments with shortened Ovsynch protocol. *Theriogenology*, 60:560.
- Stevenson, J. S., Johnson, S.K., Medina-Britos, M.A., Richardson-Adams, A.M. and Lamb, G.C. (2003). Resynchronization of estrus in cattle of unknown pregnancy status using estrogen, progesterone, or both. *Journal of Animal Science*, 81:1681–1692.
- Stumpf, T.T., Day, M.L., Wolfe, M.W., Clutter, A.C., Stotts, J.E., Wolfe, P.L., Kittok, R.J., and Kinder, J.E. (1989). Effect of estradiol on secretion of luteinizing hormone during the follicular phase of the bovine estrus cycle. *Biology Reproduction*, 41: 91-97.
- Subiharta, U., Nuschat, B., Utomo, D., Pramono, S., Prawirodikromo, T., Prasetyo, A., Musofa, , Ernawati, J., Purmijanto, , and Suharno. (2000). In: Lap. Hasil kegiatan pengkajian sistem usaha tani, Pertanian sapi potong di daerah lahan kering.Balai Pembibitan Ternak Potong (BPPT) Ungaran.Central java.Indonesia.: Page 24.
- Suzuki, K., Geshi, M., Yamauchi, N., and Nagai, T. (2003). Functional changes and motility characteristics of Japanese Black bull spermatozoa separated by Percoll. *Animal Reproduction Science*, 77: 157-172.
- Szyda, J., Liu, Z., Dflek., and Lien, S. (2000). Application of a mixed inheritance model to the detection of quantitative trait loci in swine. *Applied. Genetic*, 43 (1): 69-83.
- Talbot, P., and Chacon, R.S. (1981). A triple stain technique for evaluating normal acrosome reactions of human spermatozoa. *Exp.Zoology*, 215: 201-208.
- Tardif, S., Laforest, J.P., Cormier, N., and Bailey, J.L. (1999). The important of porcine sperm parameters on fertility in vivo. *Theriogenology*, 52: 447-459.

- Tanghe, S., Van Soom, A., Sterckx, V., Maes, D., de Kruif, A. (2002). Assesment of different sperm quality parameters to predict *in vitro* fertility of bulls. *Reproduction Domestic Animal*, 37: 127–132.
- Tauck, S.A., Wilkinson, J.R.C., Olsen, J.R., Janitell, J.N., and Berardinelli, J.G. (2007). Comparison of controlled internal drug release device and melengesterol acetate as progestin sources in an estrus synchronization protocol for beef heifers. *Theriogenology*, 68: 162-167.
- Thatcher, W.W., Bilby, T.R., Bartolome, J.A., Silvestre, F., Staples, C.R., and Santos, J.E.P. (2006). Strategies for improving fertility in the modern dairy cow. *Theriogenology*, 65: 30-44.
- Thuwanut, P., Chatdarong, K., Techakumphu, M., and Axner, E. (2008). The effects of antioxidants on motility, viability, acrosome integrity and DNA integrity of frozen-thawed epididymal cat spermatozoa. *Theriogenology*, 70: 233-240.
- Toelihere, M.R., Yusuf, T.L., and Taurin, M.B. (1980). Artificial Insemination. 6 edition. Department of Reproduction Faculty of Veterinary Medicine, Institute Pertanian Bogor. Indonesia.
- Trenkle, A., and Willham, R.L. (1997). Beef production efficiency: The efficiency of beef production can be improved by applying knowledge of nutrition and breeding. *Journal of Science*, 198: 1009-1015.
- Tucker, K.E., and Jansen, C.A.M. (2002). Sperm separation techniques: comparison and evaluation of gradient products. In: Proceedings 2nd International workshop for Embryologists. Troubleshooting Activities in the ART lab. Ed. Basuray R, Mortimer D.
- Valciircel , M.A., de las Hems, D.F., Moses, L.J., Perez, H., and Baldassme. (1996). Comparison between Sephadex G-10 and Percoll for preparation of normospermic, asthenospermic and frozen/thawed ram semen. *Animal Reproduction Science*, 41: 215-224.
- Valergakis, G.E., Arsenos, G., and Banos, G. (2007). Comparison of artificial insemination and natural service cost effectiveness in dairy cattle. *Journal of Animal*, 1: 293-300.
- Van Cleeff, J., Macmillan, K.L., Drost, M., Lucy, M.C., and Thatcher, W.W. (1996). Effects of administering progesterone at selected intervals after insemination of synchronized heifers on pregnancy rates and resynchronization of returns to service. *Theriogenology*, 46: 1117-1130.

- Van Kooij, R.J., and Van Oost, B.A. (1992). Determination of sex ratio of spermatozoa with a deoxyribonucleic acid-probe and quinacrine staining: a comparison. *Fertility and Sterility*, 58: 384-386.
- Vazquez, J.M., Martinet, E., Roca, J., Coy, P., and Pastor, L.M. (1993). Acrosome reaction of boar spermatozoa in homologous in vitro fertilization. *Molecular Reproductive Development*, 36: 84 - 88
- Verstegen, J., Igner-Ouada, M., and Onclin, K. (2002). Computer assisted semen analyzers in andrology research and veterinary practice. *Theriogenology*, 57: 149-179.
- Vishwanath, R., and Shannon, P. (2000). Storage of bovine semen in liquid and frozen state. *Animal Reproduction Science*, 62: 23-53.
- Visser, D., and Salamon, S. (1974). Effect of composition of tris-based diluent on survival of boar spermatozoa following deep-freezing. *Australia. Biology Science*, 27: 485-497.
- Voh, A.A., Oyedipe, E., Buvanendran, V., and Kumi-Diaka, J. (1987). Estrus response of indigenous Nigerian zebu cows after prostaglandin F_{2alpha} analogue treatment under continuous observations for two seasons. *Theriogenology*, Vol. 28 No. 1.
- Wall, R.J., and Foote, R.H. (1999). Fertility of bull sperm frozen and stored in clarified egg yolk, Tris, glycerol extender. *Journal of Dairy Science*, 82: 817-821.
- Walters, D.L., and Schallenberger, E.D. (1984). Pulsatile secretion of gonadotrophins, ovarian steroids and ovarian oxytocin during the periovulatory phase of the oestrus cycle in the cow. *Reproductive and Fertility*, 71: 503-512.
- Wang, H.X., Flaherty, S.P., Swann, N.J., and Matthews, C.D. (1994). Assessment of the separation of X- and Y- bearing sperm on albumin gradients using double label fluorescence in situ hybridization. *Fertility and Sterility*, 61: 720 - 726.
- Waterhouse, K. E., Gjeldnes, A., Tverdal, A., De Angelis, P. M., Farstad, W., Håård, M., Kommisrud, E. (2010). Alterations of sperm DNA integrity during cryopreservation procedure and in vitro incubation of bull semen. *Animal Reproduction Science*, DOI:10.1016/j.anireprosci.2009.04.011.
- Watson, P.F. (2000). The causes of reduced fertility with cryopreserved semen. *Animal Reproduction Science*, 60: 481-492. .

- Webb, R., Gong, J.G., Law, A.S., and Rusbridge, S.M. (1992). Control of ovarian function in cattle. *Reproductive Fertility*, 45: 141-156.
- Welch, G.R., Waldebesseer, G.C., Wall, R.J., and Johnson, L.A. (1995). Flow cytometry sorting and PCR to confirm separation of X- and Y- chromosome bearing bovine sperm. *Animal Biotechnology*, 6: 131-139.
- Wheaton, J.E., and Lamb, G.J. (2007). Induction of cyclicity in postpartum anestrus beef cows using progesterone, GnRH and estradiol cypionate (ECP). *Animal Reproduction Science*, 102: 208-216.
- Widsor, D.P., Evans, G., and White, I.G. (1993). Sex predetermination by separation of X- and Y-chromosome bearing sperm. *Reproductive Fertility Development*, 5: 155-171.
- Williams, G.L. (1990). Sucking as a regulator of postpartum rebreeding in cattle: a review. *Journal of Animal Science*, 68: 831-852.
- Wiltbank, J.N. (1990). Challenges for improving calf crop. In: Proc. 39th Annu. Beef Cattle Short Course: Pp. 1-22.
- Wiltbank, M.C. (1994). Cell types and hormonal mechanisms associated with mid-cycle corpus luteum function. *Journal of Animal Science*, 72: 1873-1883.
- Wishart, D.F. (1972). Observations on the oestrus cycle of the Friesian heifer. *Veterinary Research*, 90: 595-597.
- Woelders, H. (1991). Overview of in vitro methods for evaluation of semen quality. *Reproduction Domestic Animal, Suppl* 1: 145-164.
- Wright, P.J., and Malmo, J. (1992). Pharmacological manipulation of fertility. *Vet Clin North Am Food. Animal Practice*, 8: 57-89.
- Yan, J., Huai, L., Feng Chen, Z.J., Hu, J., , Gao, X., and Qin, Y. (2006). Influence of swim up time on the ratio of X - and Y-bearing spermatozoa. *European J Obstetric & Gynecology and Reproductive Biology*, 129: 150-154.
- Yanagimachi, R. (1994). Mammalian fertilization In:Knobil, E., Neill, J.D. (eds), the physiology of reproduction, 2nd. ed. Vol. 1,: pp 189-317, New York, Reven Press.
- Yimer, N., Rosnina, Y., Wahid, H., Saharee, A.A., and Yap, K.C.G., P (2010). Ovarian activity in beef and dairy cows with prolonged postpartum period

and heifers that fail to conceive. *Tropical Animal Health Production*, 42: 607-615.

Zelinski, M.B., Hirota, N.A., Keenan, E.J., and and Stormshak, F. (1980). Influence of exogenous estradiol- 17β on endometrial progesterone and estrogen receptors during the luteal phase of the ovine estrus cycle. *Biology and Reproductive*, 23:345-351.

