



**UNIVERSITI PUTRA MALAYSIA**

***ENHANCEMENT OF KEY CHEMICAL CONSTITUENTS IN  
Aquilaria malaccensis LAMARCK (KARAS) THROUGH In Vitro  
POLYPLOIDIZATION***

**SITI SUHAILA BINTI A. RAHMAN**

**FBSB 2017 16**



**ENHANCEMENT OF KEY CHEMICAL CONSTITUENTS IN  
*Aquilaria malaccensis* LAMARCK (KARAS) THROUGH *In Vitro*  
POLYPLOIDIZATION**

**By**

**SITI SUHAILA BINTI A. RAHMAN**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

**March 2017**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright© Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirements for the degree of Doctor of Philosophy

**ENHANCEMENT OF KEY CHEMICAL CONSTITUENTS IN  
*Aquilaria malaccensis* LAMARCK (KARAS) THROUGH *In Vitro*  
POLYPLOIDIZATION**

By

**SITI SUHAILA BINTI A. RAHMAN**

**March 2017**

**Chairman : Associate Professor Norihan binti Mohd. Saleh, PhD**  
**Faculty : Biotechnology and Biomolecular Sciences**

*Aquilaria malaccensis* is a highly valued timber species in Malaysia for its ability to produce fragrant resin known as agarwood. The agarwood consists of volatile chemical (sesquiterpenoid groups and phenyl ethyl chromones derivatives), reported to be triggered as a defence mechanism due to attacks by microorganisms at the wounded areas. In the natural forest, only up to 10% of the wild *A. malaccensis* were found to be able to produce agarwood. Malaysia is currently facing threats of *A. malaccensis* extinction due to illegal harvesting of the species which have caused this species to be listed in CITES (Appendix II). In view of this situation, there is a need to develop *A. malaccensis* clones with enhanced chemical constituents for commercial plantation. The *in vitro* polyploidization has been proven to improve desired characteristics in plant such as fast growing, increased in bioactive compounds and secondary metabolites. Therefore, the objectives of this study were to develop *in vitro* polyploidization protocol in *A. malaccensis*; to determine the morphological characteristics difference between the polyploids to its diploid counterparts and; to evaluate and compare the chemical constituents in the different ploidy levels of *A. malaccensis*. The *A. malaccensis in vitro* polyploidization was carried out using antimetabolic agents (colchicine and trifluralin) at different concentrations and exposure times on two types of explants (shoot tip and nodal segment). The ploidy levels of the *in vitro* induced polyploid plantlets were determined using flow cytometer, chromosome count and stomata size measurement. Diploid plantlets were used as reference. The plant height, leaf length and leaf width were also measured in tetraploids and diploids plantlets. The chemical profiling of the *in vitro* induced polyploids and diploids plantlets were evaluated using the Headspace-Solid Phase Microextraction (HS-SPME) and hydro distillation, HD (for essential oil extraction, if any); both coupled with gas chromatography-mass spectrometry (GC/MS). The experiment showed that *A. malaccensis* polyploids can be induced with tetraploids obtained using nodal segments treated with 0.1 mM trifluralin for 120 hours. However,

other treatments were only able to induce mixoploids in shoot tip (treated with 1 mM colchicine at 24 and 120 hours exposure time; 2 mM colchicine at 48 hours exposure time; while 0.05 mM and 0.1 mM trifluralin both at 120 hours exposure time) and nodal segment (0.05 mM trifluralin at 120 hours exposure time). The DNA content and genome sizes were quantified as 1.84 pg  $2C^{-1}$  and 899 Mbp in the diploid and, 3.86 pg  $2C^{-1}$  and 1887 Mbp in the tetraploids. *A. malaccensis* chromosome number was determined to be  $2x=14$  (diploids), and  $4x=28$  (tetraploids). The tetraploids showed larger stomata guard cell sizes ( $33.3\pm 0.6\ \mu\text{m}$ ) in leaves as compared to diploids ( $23.1\pm 0.5\ \mu\text{m}$ ). Plant height of 24 months old tetraploids were  $49\pm 0.05$  cm, more than double the height of diploids, stem diameter of tetraploids were  $0.7\pm 0.02$  cm compared to  $0.5\pm 0.02$  cm in diploids, the leaf area also doubled in size compared to diploids, with  $24.07\pm 0.04\ \text{cm}^2$  in tetraploids leaf sample. Chemical profiling of four different sources were compared: seedling grown under normal conditions, seedling grown *in vitro*, *in vitro* diploids and *in vitro* tetraploids. Through HS-SPME/GCMS, highest amount of important sesquiterpenes (volatile chemical constituents in agarwood oil from mature trees) such as  $\alpha$ -eudesmol and  $\alpha$ -guaiene, was found in stem and root samples of *in vitro* tetraploids. The HD-GC/MS method showed all samples did not contain essential oil. However, 60% of the hydrosol water from leaves and 49.1% of the residual water from root samples of *in vitro* tetraploids consists of important sesquiterpenes. Further evaluation in *A. malaccensis* diploid and tetraploid plants at 24 months old showed fewer amounts of chemical constituents than the 4 months old in all plant parts. The tetraploids root however, contains important sesquiterpene,  $\alpha$ -guaiene (2.92%), which was not detected in diploids. These results demonstrated that *A. malaccensis* polyploid plantlets can be induced for plant improvement. The *A. malaccensis* tetraploids contained higher chemical constituents which can be promoted as high quality *A. malaccensis* clones for commercial plantation.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**PENINGKATAN KONSTITUEN KIMIA MELALUI PENGHASILAN POLIPLIODI *Aquilaria malaccensis* Lamarck (KARAS) SECARA *In Vitro***

Oleh

**SITI SUHAILA BINTI A. RAHMAN**

Mac 2017

**Pengerusi : Profesor Madya Norihan binti Mohd. Saleh, PhD**  
**Fakulti : Bioteknologi dan Sains Biomolekul**

*Aquilaria malaccensis* adalah spesies kayu bernilai tinggi di Malaysia kerana kebolehpayaannya menghasilkan resin wangi dikenali sebagai gaharu, yang digunakan secara tradisional dan industri terapeutik/aromatik moden. Gaharu ini terdiri daripada bahan kimia meruap (kumpulan sesquiterpenoid dan olahan kromon phenyl etil), dilaporkan akan terhasil sebagai mekanisme pertahanan terhadap serangan oleh mikroorganisma di kawasan cedera pada batang pokok. Di dalam hutan semula jadi, hanya lebih kurang 10% daripada *A. malaccensis* liar mampu menghasilkan gaharu. Malaysia kini menghadapi ancaman kepupusan *A. malaccensis* ekoran daripada pengambilan gaharu secara haram yang telah mengakibatkan spesies ini disenaraikan dalam CITES (Lampiran II). Keadaan ini telah mewujudkan keperluan untuk menghasilkan klon *A. malaccensis* dengan konstituen kimia yang dipertingkatkan untuk tujuan perladangan komersial. Poliploidi telah terbukti mampu meningkatkan ciri-ciri yang dikehendaki dalam tumbuhan seperti tumbesaran yang lebih cepat, peningkatan sebatian bioaktif dan metabolit sekunder. Oleh itu, objektif kajian ini adalah untuk membangunkan protokol menghasilkan *A. malaccensis* poliploidi secara *in vitro*; untuk menentukan perbezaan ciri-ciri morfologi antara anak pokok poliploid dengan diploid dan; untuk menilai dan membandingkan konstituen kimia dari peringkat ploidi *A. malaccensis* yang berbeza. Protokol poliploidi secara *in vitro* telah dibangunkan menggunakan ejen antimitotik (colchicine dan trifluralin) pada kepekatan dan masa pendedahan yang berbeza ke atas dua jenis eksplan (pucuk dan segmen nod). Tahap ploidi anak pokok poliploid *in vitro* ditentukan dengan menggunakan kaedah aliran sitometri, bilangan kromosom dan pengukuran saiz stomata. Anak pokok diploid digunakan sebagai rujukan. Ketinggian anak pokok, panjang daun dan lebar daun diukur dalam anak pokok tetraploid dan diploid. Pemprofilan kimia terhadap anak pokok tetraploid dan diploid dinilai menggunakan kaedah *Headspace-Solid Phase Microextraction (HS-SPME)* dan hidro penyulingan, HD (untuk minyak gaharu, jika ada); kedua-duanya dinilai dengan kaedah gas kromatografi-spektrometri jisim (GC/MS). Eksperimen menunjukkan bahawa *A. malaccensis* poliploidi boleh

diaruhkan dengan terhasilnya tetraploid melalui rawatan segmen nod dengan 0.1 mM trifluralin selama 120 jam. Beberapa rawatan yang lainnya juga menghasilkan poliploidi pada peringkat mixoploidi pada eksplan pucuk (dirawat dengan 1 mM colchicine pada 24 and 120 jam masa pendedahan; 2 mM colchicine pada 48 hours pendedahan masa; 0.05 mM dan 0.1 mM trifluralin kedua-duanya pada 120 masa pendedahan) dan segmen nod (0.05 mM trifluralin pada 120 jam masa pendedahan). Kandungan DNA dan saiz genom adalah 1.84 pg  $2C^{-1}$  dan 899 Mbp dalam diploid dan, 3.86 pg  $2C^{-1}$  dan 1887 Mbp dalam tetraploid. *A. malaccensis* mempunyai bilangan kromosom  $2x = 14$  (diploid), dan  $4x = 28$  (tetraploid). Daun tetraploid menunjukkan saiz pengawal sel stomata lebih besar ( $33.3 \pm 0.6 \mu\text{m}$ ) berbanding diploid ( $23.1 \pm 0.5 \mu\text{m}$ ). Ketinggian pokok tetraploid berusia 24 bulan adalah  $49.0 \pm 0.05 \text{ cm}$ , lebih daripada dua kali ganda ketinggian pokok diploid, diameter batang tetraploid adalah  $0.7 \pm 0.02 \text{ cm}$ , berbanding  $0.5 \pm 0.02 \text{ cm}$  diploid. Keluasan daun juga adalah dua kali ganda berbanding saiz pada diploid, di mana sampel daun tetraploid adalah  $24.07 \pm 0.04 \text{ cm}^2$ . Perbandingan profil kimia dilaksanakan terhadap empat sumber yang berbeza: anak benih ditanam di bawah keadaan biasa, anak benih ditanam secara *in vitro*, diploid *in vitro* dan tetraploid *in vitro*. Melalui kaedah *HS-SPME/GCMS*, jumlah sesquiterpene penting ditemui paling tinggi di dalam batang dan akar sampel tetraploid *in vitro*. Kaedah *HD-GC/MS* menunjukkan semua sampel tidak mengandungi minyak pati. Walau bagaimanapun, 60% daripada air hydrosol dari daun dan 49.1% daripada air sisa daripada sampel akar dalam tetraploid *in vitro* terdiri daripada sesquiterpene penting. Penilaian lanjut ke atas *A. malaccensis* diploid dan tetraploid pada umur 4 bulan menunjukkan jumlah konstituen kimia yang lebih sedikit daripada pada umur 4 bulan di semua bahagian pokok. Walau bagaimanapun, akar tetraploid mengandungi sesquiterpene penting,  $\alpha$ -guaiene (2.92%), yang tidak dikesan dari akar diploid *in vitro*. Hasil kajian ini menunjukkan bahawa anak pokok *A. malaccensis* polyploid boleh diaruh untuk penambahbaikan kandungan konstituen kimia yang lebih tinggi yang boleh dipromosikan sebagai klon *A. malaccensis* berkualiti tinggi untuk perladangan komersial.

## ACKNOWLEDGEMENTS

BISMILLAH HIRRAHMAN NIRRAHEEM...

Alhamdulillah Praise to Allah SWT for the strength and courage upon me to be steadfast in this journey. This has been one of the most highlight moments in my life and I would like to say thank you to people that help me along the way. In Shaa Allah May Allah SWT granted and bless your life and hereafter for only Allah knows...

Firstly thank you to my wonderful Student Committee members for your guidance and kindness throughout this journey. I would like to express my very great appreciation to my main supervisor, Assoc. Prof. Dr. Norihan Mohd. Saleh for her useful critiques of this research work and always there to help me to better myself in research and as a person; I am particularly grateful to Dr. Norwati Muhammad for being the best second eye in over viewing the project in order to strengthen the research; I also wish to acknowledge the help by Dr. Kodi Isparan Kandasamy which has been there from the start and I appreciate all of the comments and ideas to ensure quality work; I would like to express my deepest gratitude to Prof. Mahani Mansor Clyde for being very generous to share her decades of knowledge and kind advices; also I would like to offer my special thanks to Assoc. Prof. Dr. Parameswari Namasivayam for her constructive suggestions to the project. I could not ask for a better group of intelligent people to be with in this journey.

I would also like to thank Andrew C. Chambers and Noor Nina Camille Chambers, for their love, support and encouragements throughout my study. To all my family members, this is for all of you. My special thanks are extended to all staff and friends; in spite our crazy and hectic lives, thank you for finding time making me laugh and cheerful every day. Keep those friendship and team spirits high!



I certify that a Thesis Examination Committee has met on 24 March 2017 to conduct the final examination of Siti Suhaila bt. A. Rahman on her thesis entitled "Enhancement of Key Chemical Constituents in *Aquilaria malaccensis* Lamarck (Karas) through *In Vitro* Polyploidization" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

**Noorjahan Banu binti Mohammed Alitheen, PhD**  
Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Chairman)

**Janna Ong binti Abdullah, PhD**  
Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Mohd. Puad bin Abdullah, PhD**  
Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Naglaa Abdelmoneim Abdallah, PhD**  
Professor  
Cairo University  
Egypt  
(External Examiner)



---

**NOR AINI AB. SHUKOR, PhD**  
Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 6 July 2017

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Norihan binti Mohd. Saleh, PhD**

Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Chairperson)

**Parameswari Namasivayam, PhD**

Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Member)

**Norwati binti Muhammad, PhD**

Biotechnology Programme  
Forestry Biotechnology,  
Forest Research Institute Malaysia, FRIM  
(Member)

**Mahani Mansor Clyde, PhD**

Professor Dato'  
Faculty of Science and Technology  
Universiti Kebangsaan Malaysia (retired 2016)  
(Member)

**Kodi Isparan Kandasamy, PhD**

Senior Vice President  
Malaysian Biotechnology Corporation Sdn. Bhd.  
Kuala Lumpur  
(Member)

---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice- Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Siti Suhaila Binti A. Rahman, GS26310

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: \_\_\_\_\_  
Name of Chairman of  
Supervisory Committee : \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory Committee: \_\_\_\_\_

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiii
<b>LIST OF FIGURES</b>	xvi
<b>LIST OF ABBREVIATIONS</b>	xxiv
 <b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
<b>2 LITERATURE REVIEW</b>	<b>3</b>
2.1 <i>Aquilariamalaccensis</i>	3
2.1.1 Distribution and botanical description	3
2.1.2 The history and traditional uses of <i>A. malaccensis</i> agarwood	3
2.1.3 The global economic importance of agarwood	9
2.2 The traditional and modern method of agarwood production	11
2.3 Important sesquiterpenes in <i>A. malaccensis</i> soil	12
2.4 Polyploidy in plant speciation and evolution	12
2.4.1 Classification of Polyploidy	14
2.4.2 The use of antimetabolic agents in polyploidization	14
2.4.3 Cytology study and flow cytometer analyses	16
2.4.4 Guard cell sizes and chromosomal count	16
2.4.5 Photosynthetic rates at different ploidy level	17
2.4.6 Plant growth assessment and ploidy level of timber trees	17
2.4.7 Enhancement of secondary metabolite production through polyploidization	18
<b>3 MATERIALS AND METHODS</b>	<b>19</b>
3.1 The <i>in vitro</i> polyploidization of <i>A. malaccensis</i>	19
3.1.1 Plant materials	19
3.1.2 Antimetabolic agents stock preparations	19
3.1.2.1 Colchicine stock solution	19
3.1.2.2 Trifluralin stock solution	19
3.1.3 Treatments of <i>A. malaccensis</i> explants with antimetabolic agents	20
3.2 <i>In vitro</i> regeneration of treated samples	20
3.2.1 Murashige & Skoog (1962) basal medium	20
3.2.2 Plant growth regulators (PGRs) stock solution	20
3.2.3 Shoot regeneration medium	22
3.2.4 Root induction medium	22

3.3	Plant DNA flow cytometry (FCM) analysis of <i>A. malaccensis</i>	22
3.3.1	Instrument settings and software	22
3.3.2	Nuclei isolation buffer (General Purpose Buffer)	23
3.3.3	DNA staining solution (propidium iodide) and RNase	23
3.3.4	Sample preparation and storage	23
3.3.5	Confirmation of ploidy level – DNA content analyses	23
3.3.6	Statistical analyses	24
3.4	Cytogenetic analyses	25
3.4.1	Colchicine and Carnoy I stock solution	25
3.4.2	Fuelgen staining reagent	25
3.4.3	Determination of <i>A. malaccensis</i> chromosome number	25
3.5	Stomata size assessment	25
3.6	Chemical content profiling in <i>A. malaccensis</i>	26
3.6.1	Headspace - Solid Phase Microextraction (Headspace - SPME)	26
3.6.2	Hydro distillation process (HD)	26
3.6.3	Gas Chromatography (GC)	27
3.6.4	Gas Chromatography-Mass Spectrometry (GCMS)	27
3.6.5	Statistical analysis	27
3.7	Plant growth assessment	29
3.7.1	Plant growth rates in <i>A. malaccensis</i> diploid and polyploid plantlets	29
3.7.2	Statistical Analysis	29
3.7.3	Photosynthetic rates	29
3.7.4	Statistical analysis	30
<b>4</b>	<b>RESULTS AND DISCUSSION</b>	<b>31</b>
4.1	<i>A. malaccensis</i> DNA content and genome size	31
4.2	2C DNA content comparison between six <i>Aquilaria</i> species	35
4.3	The effects of antimetabolic agent on <i>A. malaccensis</i> explants	38
4.3.1	Survival and recovery of treated shoot tip explants of <i>A. malaccensis</i>	38
4.3.2	Survival and recovery of treated nodal segment explants of <i>A. malaccensis</i>	39
4.3.3	The effects of antimetabolic agents versus exposure time to explants' recovery	42
4.3.4	<i>In vitro</i> shoots and root development in treated and untreated <i>A. malaccensis</i>	48
4.4	The effects of antimetabolic agents and exposure time on ploidy level changes	51
4.4.1	Ploidy changes in <i>A. malaccensis</i> using shoot tips explants	51
4.4.2	Ploidy changes in <i>A. malaccensis</i> using nodal segment explants	53

4.5	Differences in plant traits and chromosomal number in <i>A. malaccensis</i> diploid and tetraploid	56
4.5.1	Plant morphology	56
4.5.2	Leaf anatomy	57
4.5.3	Chromosome number	59
4.6	Acclimatization and photosynthetic rates of <i>A. malaccensis</i> plantlets	61
4.6.1	Acclimatization	61
4.6.2	Determination of photosynthetic rates in <i>A. malaccensis</i> diploids and tetraploids	61
4.7	Determination of volatile chemical constituents in diploid and tetraploid <i>A. malaccensis</i> plantlets	66
4.7.1	Volatile chemical constituents from fresh samples of 4 months old diploids and tetraploids <i>A. malaccensis</i> using Headspace - Solid Phase Microextraction (HS-SPME) and Gas Chromatography-Mass Spectrometry (GC-MS)	67
4.7.2	Volatile chemical constituents obtained from post-hydro distillation in 4 months old diploids and tetraploids <i>A. malaccensis</i> using HS-SPME/GC-MS	74
4.7.3	Chemical constituents from hydrosol in 4 month old diploids and tetraploids <i>A. malaccensis</i> using HS-SPME/GC-MS	78
4.7.4	Chemical constituents from sample-boiled water in 4 month old diploids and tetraploids <i>A. malaccensis</i> using HS-SPME/GC-MS	86
4.7.5	Volatile chemical constituents in 24 months old diploids and tetraploids <i>A. malaccensis</i> using HS-SPME/GC-MS	90
<b>5</b>	<b>CONCLUSION</b>	<b>95</b>
<b>6</b>	<b>RECOMMENDATIONS</b>	<b>96</b>
	<b>REFERENCES</b>	<b>97</b>
	<b>APPENDICES</b>	<b>119</b>
	<b>BIODATA OF STUDENT</b>	<b>160</b>
	<b>LIST OF PUBLICATIONS</b>	<b>161</b>

## LIST OF TABLES

Table		Page
1	Sources of <i>Aquilaria</i> species from around the world.	7
2	The traditional and modern use of agarwood.	7
3	Price of agarwood oil and woodchip in the global market.	10
4	Summary of CyFlow Space (Partec <sup>R</sup> ) instrument settings used in this study.	23
5	Background of the <i>in vitro</i> clones and field grown trees of <i>A. malaccensis</i> used in this study.	24
6	The DNA content from five field grown trees of <i>A. malaccensis</i> (Am461, Am462, Am463, Am464 and Am465) collected from field with Duncan's group analysis.	33
7	The DNA contents of six <i>in vitro</i> clones of <i>Aquilaria malaccensis</i> (Am <sub>KG1</sub> , Am <sub>F15</sub> , Am <sub>KG7</sub> , Am <sub>KG17</sub> , Am <sub>KG23</sub> and Am <sub>KG25</sub> ).	33
8	2C nuclear DNA contents and genome sizes of six <i>Aquilaria</i> species, determined by flow cytometry.	37
9	DNA contents and genome sizes of <i>A. malaccensis</i> , diploid and tetraploid determined by flow cytometry (FCM).	52
10	Response of <i>A. malaccensis</i> shoot tip explants expose to different types of antimetabolic agents at different duration after 60 days in cultures on MS medium supplemented with 0.1 mg/L BAP.	54
11	Response of <i>A. malaccensis</i> nodal segment explants expose to different types of antimetabolic agents at different duration after 60 days in cultures on MS medium supplemented with 0.1 mg/L BAP.	55
12	Variation in stomata size in control and tetraploid plants of <i>A. malaccensis</i> .	58
13	Plant height, stem diameter, leaf area, and root length of 6-months old <i>A. malaccensis</i> , diploids and tetraploids.	63
14	Plant height, stem diameter, leaf area, and root length of 24-months old <i>A. malaccensis</i> , diploids and tetraploids.	63



15	Photosynthetic rates of 24 months old <i>A. malaccensis</i> , diploids and tetraploids.	66
16	Volatile chemical constituents detected in 4 months old <i>A. malaccensis</i> <u>fresh leaf</u> samples using HS-SPME/GCMS.	68
17	Volatile chemical constituents detected in 4 months old <i>A. malaccensis</i> <u>fresh stem</u> samples using HS-SPME/GCMS.	69
18	Volatile chemical constituents detected in 4 months old <i>A. malaccensis</i> <u>fresh roots</u> samples using HS-SPME/GCMS.	72
19	Chemical constituents in <u>wax</u> obtained after hydro distillation of 4 months old <i>A. malaccensis</i> <u>leaf</u> samples.	76
20	Chemical constituents in <u>wax</u> obtained after hydro distillation of 4 months old <i>A. malaccensis</i> <u>leaf</u> samples.	77
21	Chemical constituents in <u>hydrosol</u> obtained after hydro distillation of 4 months old <i>A. malaccensis</i> <u>leaf</u> samples.	80
22	Chemical constituents in <u>hydrosol</u> obtained after hydro distillation of 4 months old <i>A. malaccensis</i> <u>stem</u> samples.	82
23	Chemical constituents in <u>hydrosol</u> obtained after hydro distillation of 4 months old <i>A. malaccensis</i> <u>root</u> samples.	84
24	Chemical constituents in <u>residual water</u> obtained after hydro distillation of 4 months old <i>A. malaccensis</i> <u>leaf</u> samples.	87
25	Chemical constituents in <u>residual water</u> obtained after hydro distillation of 4 months old <i>A. malaccensis</i> <u>stem</u> samples.	88
26	Chemical constituents detected in <u>residual water</u> from distillation of 4 months old <i>A. malaccensis</i> <u>root</u> samples.	89
27	Volatile chemical constituents detected in 24 months old <i>A. malaccensis</i> <u>fresh leaf samples</u> using HS-SPME/GCMS.	91
28	Volatile chemical constituents detected in 24 months old <i>A. malaccensis</i> <u>fresh stems samples</u> using HS-SPME/GCMS.	92
29	Volatile chemical constituents detected in 24 months old <i>A. malaccensis</i> <u>fresh roots samples</u> using HS-SPME/GCMS.	93
A1	Perfumes and colognes with agarwood.	119
A2	Important chemical compounds in high quality agarwood.	123

A3	Diagram of biosynthetic pathway of secondary metabolites (terpenoids, phenolic compounds and alkaloids).	123
B1	Stock of Murasige&Skoog (1962) basal medium preparation for <i>in vitro</i> plant growth and development.	124
B2	Medium formulation for <i>in vitro</i> shoot development.	125
B3	Medium formulation for <i>in vitro</i> root development.	125
C1-1	List of volatile chemical constituents obtained from <i>A. malaccensis</i> fresh samples (Mean $\pm$ Standard Deviation, SD of Area %).	132
C2-1	List of chemical constituents obtained from <i>A. malaccensis</i> wax after hydro distillation (Mean $\pm$ Standard Deviation, SD of Area %).	137
C3-1	List of chemical constituents obtained from <i>A. malaccensis</i> hydrosol after hydro distillation (Mean $\pm$ Standard Deviation, SD of Area %).	144
C4-1	List of chemical constituents obtained from <i>A. malaccensis</i> residual water after hydro distillation (Mean $\pm$ Standard Deviation, SD of Area %).	153
C5-1	List of volatile chemical constituents obtained from 24 month old <i>A. malaccensis</i> fresh samples (Mean $\pm$ Standard Deviation, SD of Area %).	158

## LIST OF FIGURES

Figure		Page
1	<i>Aquilaria malaccensis</i> morphology description. (A) <i>A. malaccensis</i> trees (>7 years old), (B) cross section of <i>A. malaccensis</i> trunk (>20 years old), (C) flowers of <i>A. malaccensis</i> , (D) fruits and leaves of <i>A. malaccensis</i> (E) longitudinal sections of fruits exposing the seeds. Source: (A) <a href="http://jotarofootsteps.blogspot.my">jotarofootsteps.blogspot.my</a> , (B) <a href="http://forestpathology.cfans.umn.edu">forestpathology.cfans.umn.edu</a> . (C) <a href="mailto:sitisuhaila@frim.gov.my">sitisuhaila@frim.gov.my</a> , (D,E) Seed Technology Lab, FRIM.	4
2	<i>Aquilaria malaccensis</i> botanical description. Illustration of <i>A. malaccensis</i> plant parts (Source: PROSEA/FRIM).	5
3	<i>A. malaccensis</i> products: (A) Agarwood (red arrow) formed in the trunk of <i>A. malaccensis</i> , (B) Agarwood oil being sold at Jalan Bukit Bintang, Kuala Lumpur, Malaysia, (C) traditional used of agarwood – house fumigations for special occasions, joss-sticks and oils/pellet/woodchips (D) agarwood as an ingredient in high-end perfumes. Sources: (A) <a href="http://researchgate.net/publication/286762450">researchgate.net/publication/286762450</a> , (B,D) <i>TheStar</i> , (C) <a href="http://flicks.com">flicks.com</a> .	8
4	Major pathways of producing polyploids <i>Geoffrey Meru</i> ; Department of Horticulture, University of Georgia(2013).	15
5	<i>In vitro</i> polyploidization protocol of <i>A. malaccensis</i> : (A) <i>in vitro</i> cultures (8 weeks old), (B) pre-treatment of shoot tip and nodal segments on modified MS hormone-free medium (9 days), (C) explants immersed in antimetabolic agents solution (the concentration and exposure time accordingly used), (D) explants aseptically rinsed, (E) explants blot dried, and ready to be transferred onto modified MS-hormone free medium, (F) shoots development from treated explants in modified MS hormone-free medium (20 days).	21
6	Extraction of chemical constituents from <i>A. malaccensis</i> samples. Plant part was sampled and measured for moisture content. The remaining fresh samples were evaluated using HS-SPME/GCMS and the rest was boiled (hydro distillation) prior to SPME/GCMS evaluation.	28

- 7 The histogram peaks of DNA content in *A. malaccensis* seedlings and *in vitro* sample. (A) Histograms peak of seedlings *A. malaccensis* sample, relative to internal standard, R; and (B) histograms peak of *in vitro* samples relative to internal standard, R. Vertical and horizontal axes represent the number of nuclei counted ( $10^3$ ) and the intensity of the fluorescence channel (FL2). Coefficient variations (CV%) represents the percentages of coefficient of variation in each sample. 32
- 8 The histogram peaks of DNA content in 6 different *Aquilaria* species grown *in vitro*. (A) *A. malaccensis*, (B) *A. hirta*, (C) *A. crassna*, (D) *A. sinensis*, (E) *A. subintegra* and (F) *A. beccariana* together with *R. sativus* (internal standard, R), showing DNA content values. Vertical and horizontal axes represent the number of nuclei counted and the intensity of the fluorescence channel (FL2), respectively. CV (%) represented the mean percentages of coefficient variation in each sample. 36
- 9 The response of *A. malaccensis* shoot tip explants to different types of antimitotic agent at different ages. Plant development from shoot tips explants after 14 days in culture: (A) Plant developed from untreated (control) shoot tips, (B) colchicine-treated shoot tips, (C) trifluralin-treated shoot tips. Plant development from shoot tips explants after 20 days in culture: (D) Plant developed from untreated shoot tips, (E) colchicine-treated shoot tips, (F) trifluralin-treated shoot tips. Some of the shoot tip explants show low tolerance exhibited (G) hyperhydricity and (H) cell death. 40
- 10 The response of *A. malaccensis* nodal segment explants to different types of antimitotic agent at different ages. Plant development from nodal segment explants after 14 days in culture: (A) Plant developed from untreated (control) shoot tips, (B) colchicine-treated nodal segment, (C) trifluralin-treated nodal segment. Plant development from nodal segment explants after 20 days in culture: (D) Plant developed from untreated nodal segment, (E) colchicine-treated nodal segment, (F) trifluralin-treated nodal segment. Some of the nodal segment explants show low tolerance exhibited (G) hyperhydricity and (H) callus proliferation (red arrow) and cell death. 41
- 11 Effects of antimitotic treatments on shoot-tip and nodal segment explants. Number of explants survived and ploidy changes after treatment with (A) colchicine and (B) trifluralin at 6 hours of exposure time ( $P0.05$ ). 43

12	Effects of antimitotic treatments on shoot-tip and nodal segment explants. Number of explants survived and ploidy changes after treatment with (A) colchicine and (B) trifluralin at 12 hours of exposure time (P0.05).	44
13	Effects of antimitotic treatments on shoot-tip and nodal segment explants. Number of explants survived and ploidy changes after treatment with (A) colchicine and (B) trifluralin at <u>24 hours</u> of exposure time (P0.05).	45
14	Effects of antimitotic treatments on shoot-tip and nodal segment explants. Number of explants survived and ploidy changes after treatment with (A) colchicine and (B) trifluralin at <u>48 hours</u> of exposure time (P0.05).	46
15	Effects of antimitotic treatments on shoot-tip and nodal segment explants. Number of explants survived and ploidy changes after treatment with (A) colchicine and (B) trifluralin at <u>120 hours</u> of exposure time (P0.05).	47
16	Morphological characteristics of <i>A. malaccensis</i> explants after <i>in vitro</i> polyploidization and cultured on MS medium supplemented with 0.1 mg/L BAP after 60 days. (A) Plantlets developed from untreated explant, (B) Explant exposed to antimitotic agents (colchicine and trifluralin, (C) excessive callus proliferation from explants treated with antimitotic agent (colchicine and trifluralin).	49
17	Induction of root from different ploidy levels of <i>A. malaccensis</i> on MS medium supplemented with 1.0 mg/L IBA at different day. (A) diploid plant at 0 day, (B) tetraploid plant 0 day, (C) diploid plant (D)tetraploid plant at 21 days in culture.	50
18	Determination of <i>A. malaccensis</i> ploidy levels using flow cytometer. (A) Determination of Amd ( <i>A. malaccensis</i> diploid) position relative to internal standard <i>R. sativus</i> (R), (B) Appearance of mixoploid profile with Amd and AmT ( <i>A. malaccensis</i> tetraploids) relative to R, (C) Appearance of true tetraploids, AmT profile. Vertical and horizontal axes represent the number of nuclei counted and the intensity of the fluorescence channel (FL2), respectively. The co-efficient variations percentage (CV%) represented the mean value of coefficient of variation percentage.	52
19	Distribution and size of stomata comparison in different ploidy levels of <i>A. malaccensis</i> (A) diploid (control) and (B) tetraploid. Bar - 20 $\mu$ m size.	58
20	Occurrence of different stages of mitosis in <i>A. malaccensis</i> : (A) interphase, (B) prophase, (C) metaphase, $2n=2x=14$ , (E) anaphase, (F) telophase.	60

21	Appearance of <i>A. malaccensis</i> chromosomes. (A) diploid $2n=2x=14$ , and (B) tetraploid $2n=4x=28$ .	60
22	Acclimatization and plant development of <i>A. malaccensis</i> plantlets. (A) <i>A. malaccensis</i> plantlets acclimatization in glass chambers with high humidity for 1 month; (B) <i>A. malaccensis</i> plantlets at greenhouse after 3 months transferred from glass chambers: (i) diploid plant and (ii) tetraploid plant. (Red arrows showing roots growing out from the potting medium, Jiffy-7 from tetraploid plantlets).	62
23	Morphological comparison of <i>A. malaccensis</i> diploid and tetraploid plant parts: (A) diploid leaf; (B) tetraploid leaf; (C) diploid stem (D) tetraploid stem; (E) diploid roots; (F) tetraploid roots.	63
24	Morphology of 24 month old (A) diploid and (B) tetraploid <i>A. malaccensis</i> grown in greenhouse.	64
25	Observation on 24 month old <i>A. malaccensis</i> diploid and tetraploid plants in the greenhouse: Leaf sizes of (A) diploid and (B) tetraploid; (B) stem diameter of (C) diploid and (D) tetraploid; (C) roots of (E) diploid and (F) tetraploid.	65
C1a	Volatile chemical constituents detected using HS-SPME/GCMS on <u>fresh leaf</u> samples of <i>A. malaccensis</i> <u>seedlings</u> (4 months old). The red labels are those important sesquiterpenes of agarwood.	126
C1b	Volatile chemical constituents detected using HS-SPME/GCMS on fresh leaf samples of <u>in vitro grown seedlings</u> <i>A. malaccensis</i> (4 months old). No important sesquiterpenes were detected.	126
C1c	Volatile chemical constituents detected using HS-SPME/GCMS on <u>fresh leaf</u> samples of <i>A. malaccensis</i> <u>in vitro diploids</u> (4 months old). No important sesquiterpenes were detected.	127
C1d	Volatile chemical constituents detected using HS-SPME/GCMS on <u>fresh leaf</u> samples of <i>A. malaccensis</i> <u>in vitro tetraploids</u> (4 months old). No important sesquiterpenes were detected.	127
C1e	Volatile chemical constituents detected using HS-SPME/GCMS on <u>fresh stem</u> samples of <u>seedlings</u> <i>A. malaccensis</i> (4 months old). Red labels are those important sesquiterpenes in agarwood.	128
C1f	Volatile chemical constituents detected using HS-SPME/GCMS on <u>fresh stem</u> samples of <i>A. malaccensis</i> <u>in vitro grown seedlings</u> (4 months old). No important sesquiterpenes were detected.	128
C1g	Volatile chemical constituents detected using HS-SPME/GCMS on	129

	<u>fresh stem</u> samples of <i>A. malaccensis in vitro</i> <u>diploids</u> (4 months old). Red labels are those important sesquiterpenes in agarwood.	
C1h	Volatile chemical constituents detected using HS-SPME/GCMS on <u>fresh stem</u> samples of <i>A. malaccensis in vitro</i> <u>tetraploids</u> (4 months old). Red labels are those important sesquiterpenes in agarwood.	129
C1i	Volatile chemical constituents detected using HS-SPME/GCMS on <u>fresh roots</u> samples of <i>A. malaccensis</i> <u>seedlings</u> (4 months old). Red labels are those important sesquiterpenes in agarwood.	130
C1j	Volatile chemical constituents detected using HS-SPME/GCMS on <u>fresh roots</u> samples of <i>A. malaccensis in vitro</i> <u>grown seedlings</u> (4 months old). Red labels are those important sesquiterpenes in agarwood.	130
C1k	Volatile chemical constituents detected using HS-SPME/GCMS on <u>fresh roots</u> samples of <i>A. malaccensis in vitro</i> <u>diploids</u> (4 months old). Red labels are those important sesquiterpenes in agarwood.	131
C1l	Volatile chemical constituents detected using HS-SPME/GCMS on <u>fresh roots</u> samples of <i>A. malaccensis in vitro</i> <u>tetraploids</u> (4 months old). Red labels are those important sesquiterpenes in agarwood.	131
C2a	Peaks of chemical constituents detected using HS-SPME/GCMS on <u>wax</u> samples extracted from <u>leaf</u> of <i>A. malaccensis in vitro</i> <u>tetraploids</u> (4 months old). No important sesquiterpenes were detected.	135
C2b	Peak of chemical constituents detected in <u>wax</u> from <u>stem</u> samples of <i>in vitro</i> <u>diploids</u> <i>A. malaccensis</i> (4 months old). No important sesquiterpenes were detected.	135
C2c	Peak of chemical constituents detected in <u>wax</u> from <u>stem</u> samples of <i>in vitro</i> <u>tetraploid</u> <i>A. malaccensis</i> (4 months old). No important sesquiterpenes were detected.	136
C3a	Peaks of chemical constituents detected in <u>hydrosol</u> from <u>leaf</u> samples of <u>seedlings</u> <i>A. malaccensis</i> (4 months old). Red labels are those important sesquiterpenes in agarwood.	138
C3b	Peaks of chemical constituents detected in <u>hydrosol</u> from <u>leaf</u> samples of <i>in vitro</i> <u>grown seedlings</u> of <i>A. malaccensis</i> (4 months old). No important sesquiterpenes were detected.	138
C3c	Peaks of chemical constituents detected in <u>hydrosol</u> from <u>leaf</u> samples of <i>A. malaccensis in vitro</i> <u>diploids</u> (4 months old). Red labels are those important sesquiterpenes in agarwood.	139

C3d	Peaks of chemical constituents detected in <u>hydrosol</u> from <u>leaf</u> samples of <i>A. malaccensis in vitro tetraploids</i> (4 months old). Red labels are those important sesquiterpenes in agarwood.	139
C3e	Peaks of chemical constituents detected in <u>hydrosol</u> from <u>stem</u> samples of <i>A. malaccensis seedlings</i> (4 months old). Red labels are those important sesquiterpenes in agarwood.	140
C3f	Peaks of chemical constituents detected in <u>hydrosol</u> from <u>stem</u> samples of <i>A. malaccensis in vitro grown seedlings</i> (4 months old). No important sesquiterpenes were detected.	140
C3g	Peaks of chemical constituents detected in <u>hydrosol</u> from <u>stem</u> samples of <i>A. malaccensis in vitro diploids</i> (4 months old). Red labels are those important sesquiterpenes in agarwood.	141
C3h	Peaks of chemical constituents detected in <u>hydrosol</u> from <u>stem</u> samples of <i>A. malaccensis in vitro tetraploids</i> (4 months old). Red labels are those important sesquiterpenes in agarwood.	141
C3i	Peaks of chemical constituents in <u>hydrosol</u> from <u>root</u> samples of <i>A. malaccensis seedlings</i> (4 months old). Red labels are those important sesquiterpenes in agarwood.	142
C3j	Peaks of chemical constituents in <u>hydrosol</u> from <u>root</u> samples of <i>in vitro grown seedlings</i> of <i>A. malaccensis</i> (4 months old). Red labels are those important sesquiterpenes in agarwood.	142
C3k	Peaks of chemical constituents in <u>hydrosol</u> from <u>root</u> samples of <i>A. malaccensis in vitro diploids</i> (4 months old). Red labels are those important sesquiterpenes in agarwood.	143
C3l	Peaks of chemical constituents in <u>hydrosol</u> from <u>root</u> samples of <i>A. malaccensis in vitro tetraploids</i> (4 months old). No important sesquiterpenes were detected.	143
C4a	Peaks of chemical constituents detected in <u>residual water</u> from <u>leaf</u> samples of <i>A. malaccensis seedlings</i> (4 months old). No important sesquiterpenes were detected.	147
C4b	Peaks of chemical constituents detected in <u>residual water</u> from <u>leaf</u> samples of <i>A. malaccensis in vitro grown seedling</i> (4 months old). Red labels are those important sesquiterpenes in agarwood.	147
C4c	Peaks of chemical constituents detected in <u>residual water</u> from <u>leaf</u> samples of <i>A. malaccensis in vitro diploids</i> (4 months old). Red labels are those important sesquiterpenes in agarwood.	148
C4d	Peaks of chemical constituents detected in <u>residual water</u> from <u>leaf</u>	148



	samples of <i>A. malaccensis in vitro</i> <u>tetraploids</u> (4 months old). Red labels are those important sesquiterpenes in agarwood.	
C4e	Peaks of chemical constituents detected in <u>residual water</u> from <u>stem</u> samples of <i>A. malaccensis</i> <u>seedlings</u> (4 months old). No important sesquiterpenes were detected.	149
C4f	Peaks of chemical constituents detected in <u>residual water</u> from <u>stem</u> samples of <i>A. malaccensis in vitro</i> <u>grown seedlings</u> (4 months old). Red labels are those important sesquiterpenes in agarwood.	149
C4g	Peaks of chemical constituents detected in <u>residual water</u> from <u>stem</u> samples of <i>A. malaccensis in vitro</i> <u>diploids</u> (4 months old). No important sesquiterpenes were detected.	150
C4h	Peaks of chemical constituents detected in <u>residual water</u> from <u>stem</u> samples of <i>A. malaccensis in vitro</i> <u>tetraploid</u> (4 months old). No important sesquiterpenes were detected.	150
C4i	Peaks of chemical constituents detected in <u>residual water</u> from <u>root</u> samples of <i>A. malaccensis</i> <u>seedlings</u> (4 months old). No important sesquiterpenes were detected.	151
C4j	Peaks of chemical constituents detected in <u>residual water</u> from <u>root</u> samples of <i>A. malaccensis in vitro</i> <u>grown seedlings</u> (4 months old). No important sesquiterpenes were detected.	151
C4k	Peaks of chemical constituents detected in <u>residual water</u> from <u>root</u> samples of <i>A. malaccensis in vitro</i> <u>diploids</u> (4 months old). No important sesquiterpenes were detected.	152
C4l	Peaks of chemical constituents detected in <u>residual water</u> from <u>root</u> samples of <i>A. malaccensis in vitro</i> <u>grown tetraploids</u> (4 months old). Red labels are those important sesquiterpenes in agarwood.	152
C5a	Peaks of chemical constituents detected in <u>residual water</u> from <u>leaf</u> samples of <i>A. malaccensis in vitro</i> <u>diploids</u> (24 months old). Red labels are those important sesquiterpenes in agarwood.	155
C5b	Peaks of chemical constituents detected in <u>residual water</u> from <u>leaf</u> samples of <i>A. malaccensis in vitro</i> <u>tetraploids</u> (24 months old). No important sesquiterpenes were detected.	155
C5c	Peaks of chemical constituents detected in <u>residual water</u> from <u>stem</u> samples of <i>A. malaccensis in vitro</i> <u>diploids</u> (24 months old). No important sesquiterpenes were detected.	156
C5d	Peaks of chemical constituents detected in <u>residual water</u> from <u>stem</u> samples of <i>A. malaccensis in vitro</i> <u>tetraploids</u> (24 months old). No	156

important sesquiterpenes were detected.

- C5e Peaks of chemical constituents detected in residual water from root samples of *in vitro* diploid *A. malaccensis* seedling (24 months old). Red labels are those important sesquiterpenes in agarwood. 157
- C5f Peaks of chemical constituents detected in residual water from root samples of *in vitro* tetraploid *A. malaccensis* (24 months old). Red labels are those important sesquiterpenes in agarwood. 157



## LIST OF ABBREVIATIONS

°C	temperature in degree Celsius
µl	microliter
BAP	6-benzylaminopurine
CITES	Conference on International Trades of Endangered Species
cm	centimeter
cm <sup>2</sup>	centimeter square
DNA	dinucleic acid
g/L	gram per liter
GC/MS	Gas Chromatography-Mass Spectrometry
GPB	general purpose buffer
HD	hydro distillation
HS-SPME/GCMS	Headspace-Solid Phase Microextraction
IBA	indo-3-butyric acid
kg	kilogram(s)
kPa	kilo Pascal pressure
m	meter
mg	milli gram(s)
mg/ml	milligram per milliliter
mm	millimeter
mM	milli Molar
MTIB	Malaysian Timber Industry Board
N <sup>ns</sup>	Normal seed grown under natural condition
pg 2C <sup>-1</sup>	DNA content in picogram per pair of chromosome
PGR	plant growth regulator
PI	propidium iodide
RM	Malaysian currency (Ringgits)
RNAse	ribonuclease enzyme
UAE	United Arab Emeritus
US	United States of America
V <sup>ns</sup>	Normal seeds grown under <i>in vitro</i> condition

## CHAPTER 1

### INTRODUCTION

*Aquilaria malaccensis* is one of the agarwood-producing species which belongs to the Thymelaeaceae family, distributed throughout Peninsular Malaysia, Indonesia, India, Vietnam and Thailand (Md. Salleh, 2010; Chang et al., 2002). *Aquilaria* species is best known for its distinguished and highly-valued fragrant resin (agarwood) used in incense making, in traditional medicines and perfumeries for hundreds of years, throughout the world (Nor Azah et al., 2013; Taha et al., 2010). The agarwood however, is not naturally produced. Its formation must be induced by wounding and/or infection by microbial attack (Dahlan, 2010; Lok and Yahya, 2010). Only up to 10% of the trees are able to produce agarwood in the wild, either through natural (wounded by nature and infected) or mechanical (artificial inoculation) methods (Chang et al., 2002; Ng et al., 1997). The agarwood contain mainly volatile sesquiterpenes and methyl ethyl groups (Tajuddin and Yusoff, 2010; Nor Azah et al., 2008).

Among the *Aquilaria* genus, *A. malaccensis* received significant attention in the past few decades and is favored by consumers especially from the Middle East and European countries, where it is worth more than 5 million US dollars (Mohd. Yusoff, 2014). However, the increase in demand had led to diminishing supplies and uncontrollable market pricing which are driving some of these species into extinction, if proper regulations are not enforced (Hashim et al., 2010). Malaysia and other agarwood-producing countries such as India, Indonesia, Vietnam and Thailand are seriously considering agarwood plantation programs (Lok and Yahya, 2010). Commercial plantation of *A. malaccensis* seems to be a practical alternative towards increasing the volume of agarwood production. At the same time, illegal harvesting can be inhibited through these plantation programmes where more manageable operations are in place, providing sustainable supply of agarwood. A sustainable commercial plantation practices coupled with proven inoculation formulation and technique seems to work in reducing harvesting of *A. malaccensis* trees from the wild. However, the metabolisms and pathways involved in the formation of agarwood remains a challenge for researchers as they discover agarwood is not a uniform product as in other commodity crops. For example, the chemical constituents were found to be highly variable between species and intraspecies, even when treated with the same inoculants. The understanding of the chemical constituents synthesized at genomic level is still at preliminary stage and genes that are involved in the agarwood regulation are still unknown. Several attempts were made to synthetically manufacture the agarwood essential oil chemical structures and/or induce the chemical compounds of agarwood using elicitors (Okudera and Ito, 2009). Although there are some initial successes, the operations and scale-up may not be economically feasible for industrial scale, at this stage. Therefore, improving the planting materials and selection of elite clones for commercial plantation programs need to be addressed to fulfill the growing demand of the agarwood and agarwood-based products.

Polyploidization (whole-genome duplication) has played a pervasive role in the evolution of eukaryotic cells (Adams and Wendel, 2005; Abbot and Lowe, 2004). Plants as well as fungi and animals undergo polyploidization as part of the natural phenomenon of speciation and evolution processes (Tate et al., 2005; Soltis, 2005). It is an important mechanism with a profound impact on biodiversity dynamics, ecosystem functioning, as well as health and socio-economic aspects (Adams and Wendel, 2005). Polyploids are frequently superior compared to their parent plants which over a long period of evolutionary time, increased the morphological complexity in plants and able to reduce the risk of species extinction (Soltis et al., 2004). The induction of artificial polyploidy has been proven to be useful, not only to enhance the morphological aspects (such as plant size, color intensity on flower petals and biomass) but also increasing the production of secondary metabolites and bioactive compounds, in important medicinal plants (Lavania et al., 2012; Gonzalez and Wheathers; 2003; Lavania, 1988). Even though it has been proven to be effective in producing superior polyploid lines in some cases, the increase in chromosome number through artificial polyploidization could inhibit or slow down the initial plant growth and development, but later showed better growth performances in comparison to the diploids (Oiyama and Okudai, 1986). There are many factors that play a role in polyploid induction, for example woody plant groups are harder to induce than herbaceous plants (Gamage et al., 2011), the response of explants towards antimetabolic agents and concentrations used (Dhooghe et al., 2011; Allum et al., 2007). Therefore, factors such as plant groups, explants types, types and concentrations of the antimetabolic agents, and exposure time, needed to be carefully assessed to determine the degree of success in a polyploidization induction study (Dhooghe et al., 2011).

In this study, the use of *in vitro* polyploidization for *A. malaccensis* for the improvement in morphological characteristics as well as enhancement of the amount of secondary metabolites produced are studied. The challenges with *A. malaccensis* being a woody plant, indicates considerably difficult to induce, and coupled with lack of information on the morphological and cytological aspects of *A. malaccensis*. Following the above considerations, this study has four major objectives; i.e. to:

1. Develop an *in vitro* polyploidization protocol for *A. malaccensis*
2. Compare the morphological characteristics in polyploids to their diploid counterparts
3. Conduct cytological study on *A. malaccensis* diploids and polyploids
4. Evaluate the key chemical constituents in *A. malaccensis* polyploids compared to the diploid counterparts

## REFERENCES

- Abbot, R.J. and Lowe, A.J. (2004). Origins, establishment and evolution of new polyploid species: *Senecio cambrensis* and *S. eboracensis* in the British Isles. *Biological Journal of the Linnean Society*. 8:467-474.
- Achakzai, A.K.K., Achakzai, P., Masood, A., Kayani, S.A. and Tareen, R.B. (2009). Response of plant parts and age on the distribution of secondary metabolites on plants found in Quetta. *Pak. J. Bot.* 41:2129-2135.
- Acquaah, G. (2007). Principles of plant genetics and breeding. *Wiley-Blackwell, Malden*.
- Adam, K.L. and Wendel, J.F. (2005). Polyploidy and genome evolution in plants. *Curr. Opin. Plant Biol.* 8:135-141.
- Ahokas, H. (1999). Spontaneous tetraploidy in strawberry (*Fragaria* spp.) *Nordic Journal of Botany*. 19:227-234.
- Ainouche, M. and Jencsewski, E. (2010). Focus on polyploidy. *New Phytologist*. 186:1-4.
- Allum, J.F., Bringle, D.H. and Roberts, A.V. (2007). Chromosome doubling in a *Rosa rugosa* Thunb. Hybrid by exposure of *in vitro* nodes to oryzalin: the effects of node length, oryzalin concentration and exposure time. *Plant Cell Reports*. 6: 1977-1984.
- Albuzio, A., Spettoli, S. and Cacco, G. (2006). Changes in gene expression from diploid to autotetraploid status of *Lycopersicon esculentum*. *Physiologia Plantarum*. 44:77-80.
- Amiri, S., Kazemitabaar, S.K., Ranjbar, G. and Azadbakht, M. (2010). The Effect of Trifluralin and Colchicine Treatments on Morphological Characteristics of Jimsonweed (*Datura stramonium* L.) *Trakia Journal of Sciences*. Vol. 8: 47-61.
- Ammal, J. and Khosla, S.N. (1969). Breaking the barrier to polyploidy in the genus *Eucalyptus*. *Regional Research Laboratory, Jammu, India*.
- Anon (2007). Kerajaan Aceh. Zaman Sultan Iskandar Muda (1607-1636). *Denys Lombard. KPG, Jakarta*. 96-97.
- Antoine-Michard, S. and Becket, M. (1997). Spontaneous versus colchicines-induced chromosome doubling in maize anther culture. *Plant Cell Tissue and Organ Culture*. 48: 203-207.
- Anssour, S., Krugell, T., Sharbel, T.F., Saluz, H.P., Bonaventure, G. and Baldwin, I.T. (2009). Phenotypic, genetic and genomic consequences of natural and

- synthetic polyploidization of *Nicotiana obtusifolia*. *Annals of Botany*. 103:1207-1217.
- Antonopoulou, M., Compton, J., Perry, L.S. and Al-Mubarak, R. (2010). The Trade and Use of Agarwood (oudh) in the United Arab Emirates. TRAFFIC Southeast Asia, *The CITES Secretariat, Petaling Jaya, Selangor, Malaysia*.
- Arumuganathan, K. and Earle, E.D. (1991). Estimation of nuclear DNA content of plants by Flow Cytometry. *Plant Molecular Biology Reporter*. 9:229-233.
- Asif, M.J., Mak, C. and Othman, R.Y. (2001). Characterization of indigenous *Musa* species based on flow cytometric analysis of ploidy and nuclear DNA content. *Caryologia*. 54:161-168.
- Banerjee (1968). Secondary metabolites. In: The Role of Chromosomal Change in Plant Evolution. Pp 140. *Oxford University Press: New York*.
- Baranyi, M. and Greilhuber, J. (1995). Flow cytometric analysis of genome size variation in cultivated and wild *Pisum sativum* (Fabaceae). *Plant Systematics and Evolution*. 194:231-239.
- Baranyi, M. and Greilhuber, J. (1996). Flow cytometric and Fuelgen densitometric analysis of genome size variation in *Pisum*. *Theoretical and Applied Genetics*. 92:297-307.
- Barden, A., Awang, A., Mulliken, T. and Song, M. (2002). Heart of the matter: Agarwood use and trade and CITES Implementation for *Aquilaria malaccensis*. *Traffic Network Report*.
- Barnabás B, Pfahkler PL and Kovacs G. (1991). Direct effect of colchicine on the microspore embryogenesis to produce dihaploid plants in wheat (*Triticum aestivum* L.) *Theoretical Applied Genetics*. 81: 675–678.
- Barnabás, B., Obert, B. and Kovacs, G. (1999). Colchicine, an efficient genome-doubling agent for maize (*Zea mays* L.) microspores cultured in anther. *Plant Cell Reports*. 18: 858–862.
- Barre P., Noiro M., Louarn J., Duperray C. and Hamon S. (1996). Reliable flow cytometric estimation of nuclear DNA content in coffee trees. *Cytometry*. 24:32-38.
- Barrett, H.C. (1974). Colchicine-induced polyploidy in Citrus. *Bot. Gaz.* 135:29-34.
- Barrett, H.C. (1992). An autotetraploid of the 'Key Lime', Citrus aurantifolia. *Fruit Varieties Journal*. 46:166-170.
- Baser, K.H. (2010). Handbook of Essential Oils: Science, Technology and Applications/K. *Hisn ü Can Baser, Gerhard Buchbauer*. ISBN 978-1-4200-6315-8. *Universitat Wien, Austria*.

- Basu, S.K., Datta, M., Sharma, M. and Kumar, A. (2011). Haploid production technology in wheat and some selected higher plants. *Australian Journal of Crop Science*. 5(9):1087-1093.
- Beck, S.L., Dunlop, R.W. and Fossey, A. (2003a). Stomatal length and frequency as a measure of ploidy level in black wattle, *Acacia mearnsii* (de Wild). *Botanical Journal of the Linnean Society*. 141:177-181.
- Beck, S.L., Visser, G. and Dunlop, R.W. (2005). A comparison of direct (flow cytometry) and other indirect (stomatal length and chloroplast numbers within stoma techniques as a measure of ploidy level in black wattle, *Acacia mearnsii* (De Wild). *South African Journal of Botany*. 71:359-363.
- Beniwal, B.S. (1989). Silvical characteristics of *Aquilaria agallocha* Roxb. *Indian Forester*. 115:17-21.
- Blanchette, R.A. and Van Beek, H.H. (2002). Cultivated agarwood. Eu Patent No. WO02094002 pub. Date 2001-11-28. Abstract: The present invention provides agarwood and/or agarwood resin from cultivated trees, and methods of generating agarwood and/or agarwood resin in cultivated trees.
- Blakeslee, A.F. and Avery, B.T. (1919). Mutation in the Jimson Weed. *Journal of Heredity*. 10:111-120.
- Blakesley, D., Allen, A., Pellny, T.K. and Roberts, A.V. (2002). Natural and induced polyploidy in *Acacia dealbata* Link. and *Acacia mangium* Wild. *Annals of Botany*. 90:391-398.
- Borodich, F.M., Gorb, E.V. and Gorb, S. N. (2010). Fracture behaviour of plant epicuticular wax crystals and its role in preventing insect attachment: a theoretical approach. *Rapid Communication*. 1:63-71
- Bose, R. and Choudhury, J. (1962) A comparative study of the cytotaxonomy, palynology, physiology of diploid plants from *Ocimum kilimandscharicum* Guerke and their yield of raw material and volatile contents. *Caryologia*. 15:435-453.
- Bowers, J.E., Chapman, B.A., Rong, J.K. and Paterson, A.H. (2003). Unravelling angiosperm genome evolution by phylogenetic analysis of chromosomal duplication events. *Nature*. 422:433-438.
- Bretagnolle, F. and Thompson, J.D. (1995). Gametes with the somatic chromosome number mechanisms of their formation and role in the evolution of autopolyploid plants. *New Phytol*. 129:1-22.
- Brummer, E.C., Cazcarro, P.M. and Luth, D. (1999). Ploidy determination of alfalfa germplasm accessions using flow cytometry. *Crop Science*. 39:1202-1207.



- Burkill, I. (1966). A Dictionary of economic products of the Malay Peninsula, I. Government of Malaysia and Singapore. *The Ministry of Agricultural and Cooperatives. Kuala Lumpur.*
- Buttler, L.G. (1989). Effects of condensed tannins on animal nutrition. In: Chemistry and significance of condensed tannins. (Eds.): R.W. Hemingway and J.J. Karchesy. *Plenum Press, New York.* pp. 391-402.
- Byrne, M.C., Nelson, C.J. and Randall, D.D. (1981). Ploidy effects on anatomy and gas exchange of tall fescue leaves. *Plant Physiology.* 68:891-893.
- Chamorro, E.R., Zambón, N.S., Morales, W.G., Sequeira, A.F. and Velasco, G.E. (2008). Study of the Chemical Composition of Essential Oils by Gas Chromatography National Technological University, Regional Faculty Resistance, UTN QUIMOBÍ Group, Argentina. *Gas Chromatography in Plant Science, 308 Wine Technology, Toxicology and Some Specific Applications.* pp.308-324.
- Chakrabarty, K., Kuner, A. and Manon, V. (1994). Trade in Agarwood. *WWF-India Traffic India, New Delhi.*
- Chang, Y.S., Nor Azah, M.A., Abu Said, A., Lok, E.H., Reader, S. and Spier, A. (2002). Gaharu. *FRIM Technical Information.* 69:7.
- Chen, H.Q., Wei, J.H., Yang, J.S., Zhang, Z., Yang, Y., Gao, Z.H., Sui, C. and Gong, B. (2012). Chemical Constituents of Agarwood Originating from the Endemic Genus *Aquilaria* Plants. *Chemistry & Biodiversity.* 9:236-250.
- Chen, Z.J. (2007). Genetic and epigenetic mechanisms for gene expression and phenotypic variation in plant polyploids. *Ann. Rev. Plant Biol.* 58:377-406.
- Cheng, Z.M. and Korban, S.S. (2011). *In vitro* ploidy manipulation in the genomic era. *Plant Cell Tiss Organ Cult.* 104:281-282.
- Chong, Z., Meiling, Q., Qinglong, S., Shan, Z. and Ruonong, F. (2007). Analysis of the volatile compounds in *Ligusticum chuanxiong* Hort. using HS-SPME-GC-MS. *Journal of Pharmaceutical and Biomedical Analysis.* 44(2):464-470.
- Compton, M., Gray, D. and Elmstrom, G. (1996). Identification of tetraploid regenerants from cotyledons of diploid watermelon cultured *in vitro*. *Euphytica.* 87:165-172.
- Cordell, G.A. (1981). Introduction to alkaloid, A Biogenetic Approach. *John Wiley & Sons, Inc. Canada.* 7-21.
- Cordeiro, G.M., Taylor, G.O and Henry, R.J. (2000). Characterization of microsatellite markers from sugarcane (*Sacharum* sp.) a highly polyploidy species. *Plant Science.* 155:161-168.

- Corner, E.J.H (1988). Wayside tree of Malaya Vol 2. *Malayan Nature Society, Kuala Lumpur*.
- Cox, A.V., Abdelnour, G.J., Bennett, M.D. and Leith, I.J. (1998). Genome size and karyotype evolution in the slipper orchids (Cypripedioideae: *Orchidaceae*). *American Journal of Botany*. 85:681-687.
- Crissman, H.A. and Steinkamp, J. (1990). Cytochemical techniques for multivariate analysis of DNA and other cellular constituents. In: Melamed, M.R., Lindmo, T. & Mendelsohn, M.L. (eds.). 'Flow cytometry and String' 2<sup>nd</sup> edition, Wiley – Liss. New York. pp227-247.
- Dahlan, T. (2010). Peluang dan cabaran industry gaharu di Malaysia. Presented at Seminar Kebangsaan dan Pameran Gaharu 2012, , 22-23 Mac. *Universiti Putra Malaysia, Serdang*.
- Dawend, J., Make, J., Philip, L., Tan, S., Roland, K. and Franklin, R.K. (2005). System Approach for Sustainable Gaharu Conservation in Sarawak: An Overview International Forestry Seminar. Synergistic approach to appropriate forestry technology for sustaining rainforest ecosystems, 2-9 March 2005, Bintulu Sarawak, Malaysia.
- De Latt, A.M.M., Gohde, W. and Vogelzang, M.J.D. (1987). Determination of ploidy of single plants and plant populations by flow cytometry. *Plant Breeding*. 99:303-307.
- de Wet, J.M.J. (1980). Origins of polyploids. pp 3-16. In: W.H. Lewis (Ed). *Polyploidy-Biological Relevance*. Plenum Press, New York.
- Decroocq, V., Hagen, L., Fave, M., Eyquard, J. and Pierronnet, A. (2004). Microsatellite markers in the hexaploid *Prunus domestica* species and parentage lineage of three European plum cultivars using nuclear and chloroplast simple-sequence repeat. *Molecular Breeding*. 13:135-142.
- Dhawan, O.P. and Lavania U.C. (1996). Enhancing the productivity of secondary metabolites *via* induced polyploidy: a review. *Euphytica* 87:81-89.
- Dhooghe, E., Van Laere, K., Eeckhaut, T., Leus, L. and Van Huylenbroeck, J. (2011). Mitotic chromosome doubling of plant tissues *in vitro*. *Plant Cell Tiss Organ Cult*. 104:359-373.
- Dijkstra and Speckmann, (1980). Secondary metabolites. In: *The Role of Chromosomal Change in Plant Evolution*. pp 140. *Oxford University Press: New York*.
- Dnyansagar, V.R. and Sudhakaran, I.V. (1970). Induced tetraploidy in *Vinca rosea* L. *Cytologia* 35: 227-241.

- Doležel, J. (1991). Flow cytometric analysis of nuclear DNA content in higher plants. *Phytochem. Anal.* 2:143-154.
- Doležel J. and Bartos, J. (2005). Plant DNA Flow Cytometry and Estimation of Nuclear Genome Size. *Annals of Botany.* 95:99-110.
- Doležel, J., Binarova, P. and Lucretti, S. (1989). Analysis of nuclear DNA content in plant cells by flow cytometry. *Biol. Plant.* 31:113-120.
- Doležel, J., Doleželová, M. & Novák, F.J. (1994). Flow cytometric estimation of nuclear DNA amount in diploid banana (*Musa acuminata* and *M. balbisiana*). *Biol. Plant* 36:351-357.
- Doležel, J., Greilhuber, J., Lucretti, S., Meister, A., Lysák, M.A., Nardi, L. & Obermayer, R. (1998). Plant genome size estimation by flow cytometry: Inter-laboratory comparison. *Ann. Bot.* 82:17-26.
- Doležel, J., Greilhuber, J. and Suda, J. (2007). Estimation of nuclear DNA content in plants using flow cytometry. *Nature Protocols.* 2:2233-2244.
- Doležel, J., Sgorbati, S. and Lucretti, S. (1992). Comparison of three DNA fluorochromes for flow cytometric estimation of nuclear DNA content in plants. *Physiol. Plant.* 85:625-631.
- Dowrick, G.J. and El Bayoumi, A.S. (1969). Nucleic acid content and Chromosome morphology in *Chrysanthemu*. *Genetical Research.* 13:241-250.
- Doyle, J.J., Flagel, L.E., Paterson, A.H., Rapp, R.A., Soltis, D.E., Soltis, P.S. and Wendel, J.F. (2008). Evolutionary genetics of genome merger and doubling in plants. *Annual Review of Genetics.* 42:443-461.
- Eeckhaut, T.G.R., Werbrouck, S.P.O., Leus, L.W.H., Van Bockstaele, E.J. and Debergh P.C. (2004). Chemically induced polyploidization in *Spathiphyllum wallisii* Regel through somatic embryogenesis. *Plant Cell, Tissue and Organ Culture.* 78:241-246.
- Eikenberry, E. (1994). Chromosome doubling of microspore-derived canola using trifluralin. *Cruciferae. Newsletter.* 16:51-52.
- Ellul P., Boscaiu M., Vincente O., Moreno V. and Rosello J.A. (2002). Intra- and interspecific variation in DNA content in *Cistus* (Cistaceae). *Annals of Botany.* 90:345-351.
- Escandon, A.S., Hagiwara, J.C. and Alderete, L.M. (2006). A new variety of *Bacopa monnieri* obtained by *in vitro* polyploidization. *Electronic Journal of Biotechnology* doi: 10.2225/vol9-issue3-fulltext-8.
- Evans (1989). Secondary metabolites. In *The Role of Chromosomal Change in Plant Evolution*. Pp. 140. *Oxford University Press: New York*.

- Fan, G.Q., Cao, Y.C., Zhao, Z.L. and Yang, Z.Q. (2007). Induction of autotetraploid of *Pawlonia fortune*. *Scientia Silvae Sinicae*. doi:CNKI:ISSN:1001-7488.0.2007-04-004.
- Feng, J. and Yang, X. (2012). Constituents from the leaves of *Aquilaria sinensis*. *Zhongguo Zhong Yao Za Zhi*. 37:230-4.
- Galbraith, D.W. (1990). Flow cytometric analysis of plant genomes. *Method Cell Biol.* 33:549-561.
- Galbraith D.W., Harkins K.R., Maddox, J.M. Ayres N.M., Sharma D.P. and Firoozabady E. (1983). Rapid flow cytometric analysis of the cell cycle in intact plant tissues. *Science*. 220:1049-1051.
- Gamage, H.K., Prentis, P., Lowe, A., Lamont, M. and Schmidt, S. (2011). Comparison of genetic and physiological traits of diploid progenitors and modified polyploidy lines of tree species. <http://www.polygenomx.com/wp-content/uploads/2011/05/Comparison-of-genetic-and-physiological-traits-of-diploid-1-progenitors-and-modified.pdf>.
- Gamage, H.K., Lamont, M., Quinn, C., Schenk, P.M. and Schmidt, S. (2007). Screening for newly generated polyploidy in trees using chromosome counts and leaf traits. <http://www.polygenomx.com/wp-content/uploads/2011/05/Gamage-et-al-for-special-issue-of-Ecofizz-2007-13-Nov.pdf>.
- Gao, S.L., Zhu, D.N., Cai, Z.H. and Xu, D.R. (1996). Autotetraploid plants from colchicines-treated bud culture of *Salvia miltiorrhiza* Bge. *Plant Cell, Tissue and Organ Culture*. 47:73-77.
- Germana, M.A. (2006). Doubled haploid production in fruit crops. *Plant Cell, Tissue and Organ Culture*. 86:131-146.
- Gibson, I.A.S. (1977). The role of fungi in the origin of oleoresin deposits (Agaru) on the wood of *Aquilaria agallocha* (Roxb.) *Bano Biggyn Patrika*. 6:16-26.
- Gil, A., Ghersa, C.M. and Leicach, S. (2000). Essential oil yield and composition of *Targetes minuta* accessions from Argentina. *Biochem. Syst. Ecol.* 28:261-274.
- Gilani, S. and Gilani, B.S.O. (2008). Competitive Strategy analysis of the Arabian perfume market: case in focus: Al Haramain perfumes. *International Journal of Business Strategy*. 8:3.
- Gonzalez, D.J. and Weathers, P.J. (2003). Tetraploid *Artemisia annua* hairy roots produce more artemisinin than diploids. *Plant Cell Reports*. 21:809-813.
- Graham M.J., Nickell, C.D., Rayburn A.L. (1994). Relationship between genome size and maturity group in soybean. *Theoretical and Applied Genetics*. 88:429-432.

- Grant, V. (1981). *Plant Speciation*. 2<sup>nd</sup> edition. New York: *Columbia University Press*.
- Grassi, P., Novak, J., Steinlesberger, H. and Franz, C. (2004). A direct liquid, non-equilibrium solid-phase microextraction application for analysing chemical variation of single peltate trichomes on leaves of *Salvia officinalis*. *Phytochem. Anal.* 15:198-203.
- Greve, B.G., Valet, A., Humpe, T. Tonn and Cassens, U. (2004). Flow cytometry in transfusion medicine: development, strategies and applications. *Transfusion Medicine and Hemotherapy*. 31:152-161.
- Griesbach, R.J. and Kamo, K.K. (1998). The effect of induced polyploidy on the flavonol of *Petunia* 'Mitchell'. *Growth and Metabolism*. 42:361-363.
- Greilhuber, J. (1988). "Self-tanning" – a new and important source of stoichiometric error in cytophotometric determination of nuclear DNA content in plants. *Plant Systematics and Evolution*. 158:87-96.
- Greilhuber, J. (1998). Intraspecific variation in genome size: a critical reassessment. *Annals of Botany*. 82:27-35.
- Gupta, P.P. (1981). Suppression of multivalent formation by B chromosomes in natural and artificial autopolyploids of scurvy-grass (*Cochlearia* L.). *Theor. Appl. Genet.* 59:221-223.
- Hamill S, Smith, M. and Dodd, W. (1992). *In vitro* induction of banana autotetraploids by colchicine treatment of micropropagated diploids. *Australian Journal of Botany*. 40: 887–896.
- Hashim, N., Yahya, A. and Hamid, Z. (2010). Polisi dan kawalan perdagangan gaharu. Paper presented at Seminar Kebangsaan dan Pameran Gaharu 2012, 22-23 Mac. *Universiti Putra Malaysia, Serdang*.
- Hansen, E. (2000). The hidden history of a scented wood. *Saudi Aramco World XX*, 2-13.
- Hansjakob, A., Bischof, S., Bringmann, G., Riederer, M. and Hilderbrandt, U. (2010). Very-long-chain aldehydes promote *in vitro* prepenetration processes of *Blumeria graminis* in a dose- and chain length-dependent manner. *New Phytol.* 188:1039-1054.
- Hansjakob, A., Riederer, M. and Hilderbrandt, U. (2011). Wax matters: absence of very-longchain aldehydes from the leaf cuticular wax of the *glossy11* mutant of maize compromises the prepenetration processes of *Blumeria graminis*. *Plant Pathol.* 60:1151-1161.

- Heping H., Shanlin G., Lanlan C. and Xiaoke, J. (2008). *In vitro* induction and identification of autotetraploids of *Dioscorea zingiberensis*. *In Vitro Cellular & Developmental Biology-Plant*. 44:448-455.
- Hopkins, W.G. and Hüner, N.P.A (2004). Introduction to Plant Physiology. 3rd Edi. John Wiley & Sons, Inc. 111 River Street, Hoboken, NJ 07030.
- <http://www.fragrantica.com/notes/Agarwood-oud-114.html>. Accessed date 5 May 2016.
- [http://www.homegrownconcept.com/?page\\_id=40](http://www.homegrownconcept.com/?page_id=40). Accessed date 23 August 2015.
- <https://islamgreatreligion.wordpress.com/tag/washing-of-kaaba/>. Accessed date 28 April 2016.
- Huang, H.P., Gao, S.L., Chen, L.L. and Wei, K.H. (2010). *In vitro* tetraploid induction and generation of tetraploids from mixoploids in *Dioscorea zingiberensis*. *Pharmacogn Mag.* 6:51-56.
- Huda, A.W.N., Munira, M.A.S.A., Fitrya, S.D. and Salmah, M. (2009). Antioxidant activity of *Aquilaria malaccensis* (Thymelaeaceae) leaves. *Phcog. Net.* 1:270-273.
- Hyder, P.W., Fredrickson, E.L., Estell, R.E., Tallez, M. and Gibbens, R.P. (2002). Distribution and connection of total phenolics, condensed tannins, and nordihydroguaiaretic acid (NDGA) I creosotebush (*larrea tridentate*). *Biochem. Syst. Ecol.* 30:905-912.
- Isaacson, T., Kosma, D.K., Matas, A.J., Buda, G.J., He, Y., Yu, B., Pravitasari, A., Batteas, J.D., Stark, R.E., Jenks, M.A., et al. (2009). Cutin deficiency in the tomato fruit cuticle consistently affects resistance to microbial infection and biomechanical properties, but not transpirational water loss. *Plant J.* 60: 363–377.
- Ishihara, M., Tsuneya, T. and Uneyama, K. (1991). Guaiene sesquiterpenes from agarwood. *Phytochemistry*. 30:3343-3347.
- Ishihara, M., Tsuneya, T. and Uneyama, K. (1993). Fragrant sesquiterpenes from agarwood. *Phytochemistry*. 33:1147-1155.
- Ismail, N., Mohd. Ali, N.A., Jamil, M., Mohd. Hezri, F.R., Tajuddin, S.N. and Mohd. Nasir, T. (2013). Analysis of high quality agarwood oil chemical compounds by means of SPME/GCMS and Z-Score Technique. *Malaysian Journal of Analytical Sciences.* 17:403-413.
- Iwata, H., Kato, T., and Ohno, S. (2000). Triparental origin of Damask roses. *Gene*: 259: 53-59.

- Janaki Ammal, E.K. and Sobti, S.N. (1962). The origin of the Jammu mint. *Current Science*. 31:387-388.
- Jelinek, L., Doleckova, M., Karabin, M., Hudcova, T., Kotlikova, B. and D., P. (2012). Influence in growing area, plant age and virus infection on the content of hop secondary metabolites. *Czech J. Food Sci.* 6:541-547.
- Jenks, M.A. and Ashworth, E.N. (1999). Plant epicuticular waxes: function, production and genetics. *Hortic Rev.* 23:1-68.
- Jetter, R, Kunst, L. and Samuels, A.L. (2006). Composition of plant cuticular waxes. In M Riederer, C Müller, eds, *Biology of the Plant Cuticle*. Blackwell, Oxford. pp145-181.
- Ježilová, E, Nožková-Hlaváčková, V and Duchoslav, M. (2014). Photosynthetic characteristics of three ploidy levels of *Allium oleraceum* L. (Amaryllidaceae) differing in ecological amplitude. doi: 10.1111/1442-1984.12053.
- Jing, S.Y., Jiao, X.W. and Zhao, S.J. (2009). Chromosomal studies on populations of *Aquilaria sinensis*. *Guangxi Zhiwu / Guihaia*. 29:192-197.
- Johnson, C.B., Kazantzis, A., Skoula, M., Mitteregger, U. and Novak, J. (2004). Seasonal, populational and ontogenic variation in the volatile oil content and composition of individuals of *Origanum vulgare* subsp. hirtum, assessed by GC headspace analysis and by SPME sampling of individual oil glands. *Phytochem. Anal.* 15:286-292.
- Jones, J.R., Ranney T.G. and Eaker T.A. (2008). A novel method for inducing polyploidy in *Rhododendron* seedlings. *J. Amer. Rhododendron Soc.* 62:130-135.
- Jorgensen, G.A. (1928). The experimental formation of heteroploid plants in the genus *Solanum*. *Journal of Genetics*. 19:133.
- Joseph, M.C., Randall, D.D. and Nelson, C.J. (1981). Photosynthesis in polyploidy tall fescue. *Plant Physiology*. 68:894-898.
- Jusoh, M.Z., Darus, M. and Abd. Azim, A.A. (2010). Anatomi Kayu Karas Dan Kehadiran Gaharu Dalam Sel Kayu. Paper presented at Seminar Kebangsaan dan Pameran Gaharu 2012, 22-23 Mac. *Universiti Putra Malaysia, Serdang*.
- Kandasamy, K.I. (2004). A chambers so cool, it's hot. *FRIM in Focus*. October.November. December Issue, pp.8,10,15.
- Kassan, I. (2013). 15 Angkubah Penentu – jaya & gagal pembentukan dammar karas. Paper presented at Quo Vadis Inductri gaharu Malaysia, 28-29 September. *Hotel Mandarin Court, Kuala Lumpur*.

- Katakoo, H., Lord, H.L. and Pawliszyn, J. (2000). Applications of solid-phase microextraction in food analysis. *Journal of Chromatography A*. 880:35-62.
- Kato, A. and Birchler, J.A. (2006). Induction of tetraploid derivatives of maize inbred lines by nitrous oxide gas treatment. *Journal of Heredity*. 97:39-44.
- Kermani MJ, Sarasan V, Roberts AV, Yokoya K, Wentworth J & Sieber VK. (2003). Oryzalin-induced chromosome doubling in *Rosa* and its effect on plant morphology and pollen viability. *Theoretical and Applied Genetics* 107:1195-1200.
- Kerr, A. (2001). Tetraploidy conversions: An easy and effective method of colchicines method. [http://members.trip\[ods.com/~h\\_syriacus/tetraploidy.html](http://members.trip[ods.com/~h_syriacus/tetraploidy.html).
- Khosravi, P., Kermani, M.J., Nematzadeh, G.A., Bihamta, M.R. and Yokoya, K. (2008). Role of mitotic inhibitors and genotype on chromosome doubling of *Rosa*. *Euphytica*. 160:267-275.
- King, D., Fan, M.Z., Ejata, G., Asem, E.K. and Adeola, O. (2000). The effects of tannins and nutrient utilization in white pekin duck. *British Poult. Sci.* 41:630-639.
- Klima, M., Vyvadilova, M. and Kucera, V. (2008). Chromosome doubling effects of selected antimetabolic agents in *Brassica napus* microspore culture. *Czech J. Genet. Plant Breed.* 44(1):30-36.
- Knight, C.A. and Beaulieu, J.M. (2008). Genome size scaling through phenotype space. *Annals of Botany*. 101: 759-766.
- Koornneef, M., Hanhart, C.J. and Thiel, F. (1989). A genetic and phenotypic description of *eceriferum* (*cer*) mutants in *Arabidopsis thaliana*. *J. Hered.* 80: 118-122.
- Kosma, D.K., Bourdenx, B., Bernard, A., Parsons, E.P., Lü S., Joubès, J. and Jenks, M.A. (2009). The impact of water deficiency on leaf cuticle lipids of *Arabidopsis*. *Plant Physiol.* 151: 1918-1929.
- Kosma, D.K., Nemacheck, J.A., Jenks, M.A. and Williams, C.E. (2010). Changes in properties of wheat leaf cuticle during interactions with Hessian fly. *Plant J.* 63: 31-43.
- Kowalski, R. and Wolski, T. (2006). Evaluation of phenolic acid content in *Silphium perfoliatum* L., leaves, inflorescences and rhizomes. *Electronic J. Polish Agric. Univ.* 6 (issue 1).
- Kron, P., Suda, J. and Husband, B.C. (2007). Applications of flow cytometry to evolutionary and population biology. *Annual Review of Ecology, Evolution and Systematics*. 38:847-876.



- Kumar, A., Altabella, T., Taylor, M.A. and Tiburcio, A.F. (1997). Recent advances in polyamine research. *Trends Plant Sci.* 2:124-130.
- Kumeta., Y. and Ito, M. (2010). Characterization of  $\delta$ -guaiene synthases from cultured cells of *Aquilaria*, responsible for the formation of the sesquiterpenes in agarwood. *Plant Physiology*. 154:1998-2007.
- Langkjer, R.B., Casaregola1, S., Ussery, D. W., Gaillardin1 C. and Piskur, J. (2003). Sequence analysis of three mitochondrial DNA molecules reveals interesting differences among *Saccharomyces* yeasts. *Nucleic Acids Research*. 31:3081-3091.
- Lavania, U.C. (1988). Enhanced productivity of the essential oil in the artificial autopolyploid of vetiver (*Vetiveria zizanioides* L. Nash). *Euphytica*. 38:271-276.
- Lavania, U.C., Srivastava, S., Lavania, S., Basu, S. Misra, N.K. and Mukai, Y. (2012). Autopolyploidy differentially influences body size in plants, but facilitates enhanced accumulation of secondary metabolites, causing increased cytosine methylation. *The Plant Journal*. 71:539-549.
- Lavania, U.C. and Srivastava, S. (1991). Enhanced productivity of tropane alkaloids and fertility in artificial autotetraploids of *Hyoscyamus niger* L. *Euphytica*. 52: 73-77.
- Levin D. (1983). Polyploidy and novelty in flowering plants. *American Naturalist*. 122:1-25.
- Levin, D. (2002). The Role of Chromosomal Change in Plant Evolution. *Oxford University Press: New York*.
- Lewis, W.H. (1980). Polyploidy in angiosperms: dicotyledons. pp. 241-268. In: W.H. Lewis (Ed). Polyploidy-Biological Relevance. *Plenum Press, New York*.
- Liao, Y.C., Wei, J.H., Xu, Y.H. and Zhang, Z. (2015). Cloning, expression and characterization of *COII* gene (*AsCOII*) from *Aquilaria sinensis* (Lour.). *Gilg Acta Pharm Sin B*. 5: 473-481.
- Lin, H., Jian, M., Liang, L.Y., pei, w.J., Liu, X.Z. and Zhang, H.Y. (2010). Production of polyploids from cultured shoot tips of *Eucalyptus globules* Labill by treatment with colchicines. *African Journal of Biotechnology*. 9:2252-2255.
- Liu, Z., Carpenter, S.B., Bourgeois, W.J., Yu, Y., Constantin, R.J., Falcon, M.J. and Adams, J.C. (1998). Variations in the secondary metabolite camptothecin in relation to tissue age and season in *Camptotheca acuminata*. *Tree Physiology*. 18:265-270.

- Liu Z. and Gao S. (2007). Micropropagation and induction of autotetraploid plants of *Chrysanthemum cinerariifolium* (Trev.) Vis. *In Vitro Cellular & Developmental Biology-Plant*. 43:404-408.
- Lok, E.H. and Yahya, A.Z. (1996). The growth and performance of plantation grown *Aquilaria malaccensis* in Peninsula Malaya. *J. Trp. For. Sci.* 8:573-575.
- Lok, E.H., Chang, Y. and Aziah, M.Y. (1999). Early survival and growth in field trials of *Aquilaria malaccensis* (karas) and *Azadirachta excelsa* (sentang). *J. Trp. For. Sci.* 11:852-854.
- Lok, E.H. and Yahya, A.Z. (2010). Teknik Penanaman dan Pengurusan lading pokok Karas. Seminar Kebangsaan dan Pameran gaharu. 22-23 Mac, Auditorium Kejuruteraan. *Universiti Putra Malaysia, Serdang, Selangor*.
- Loureiro, J., Ridriguez, E., Dolezel J. and Santos, C. (2007). Two New Nuclear Isolation Buffers for Plant DNA Flow Cytometry: A test with 7 species. *Annals of Botany*. 100:875-888.
- Love, S.L., Rhodes, B.B. and Nugent, P.E. (1986). Controlled pollination transfer of a nuclear male sterile gene from a diploid to a tetraploid watermelon line. *Euphytica*. 35:633-635.
- Lysák, M.A. and Doležel, J. (1998). Estimation of nuclear DNA content in Sesleria (*Poaceae*). *Caryologia*. 52:123-132.
- Marie, D. and Brown, S.C. (1993). A cytometric exercise in plant DNA histograms, with 2C values for 70 species. *Biology of the Cell*. 78:41-51.
- Martens, M. and Reish, B. (1988). An improved technique for counting chromosome in grapes. *HortScience*. 23:896-899.
- Mak, C. and Lim, V. (1996). *In vitro* polyploidy induction in a diploid banana. Proceeding of Second National Congress on Genetics 13-15 November 1996. *Genetics Society of Malaysia*. pp 351-354.
- Mamat, M.F., Yacob, M.R., Lim, H.F. and Rdam, A. (2010). Cost and Benefits Analysis of *Aquilaria* Species on Plantation for Agarwood Production in Malaysia. *International Journal of Business and Social Science*. 162-174.
- Martinez, E.L., Agüero, C.B., Lopez, M.E. and Galmarini, C.R. (2000). Improvement of *in vitro* gynogenesis induction in onion (*Allium cepa* L.) using polyamines. *Plant Sci* 36:17-21.
- Masterson, J. (1994). Stomata size in fossil plants: evidence for polyploidy in majority of angiosperms. *Science*. 264:421-424.
- McGregor, C.E., Lambert, C.A., Greyling, M.M., Louw, J.H. and Warnich, L. (2000). A comparative assessment of DNA fingerprinting techniques (RAPD, ISSR,

- AFLP and SSR) in tetraploid potato (*Solanum tuberosum* L.) germplasm. *Euphytica*. 113:135-144.
- Md. Salleh, M. (2010). Taburan Pokok Karas dan Sumber gaharu dalam hutan asli di Semenanjung Malaysia. Presented at Seminar Kebangsaan dan Pameran Gaharu 2012, , 22-23 Mac. *Universiti Putra Malaysia, Serdang*.
- Mehta, R.K. and Swaminathan, M.S. (1957). Studies on induced polyploidy in forage crops. *Indian Journal of Genetics and Plant Breeding*. 17:27-57.
- Michealson, M.J., Price, H.J., Johnston, J.S. and Ellison, J.R. (1999). Variation of nuclear DNA content in *Helianthus annuus* (Asteraceae). *American Journal of Botany* 78:1238-1243.
- Mishra, B.K., Pathak, S., Sharma, A., Trivedi, P.K. and Shukla, S. (2010). Modulated gene expression in newly synthesized auto-tetraploid of *Papaver somniferum* L. *South African Journal of Botany* 76:447-452.
- Moghaddam, M., Omidbiagi, R. and Sefidkon, F. (2007). Changes in Content and Chemical Composition of *Tagetes minuta* Oil at Various Harvest Times. *J. Essent. Oil Res.* 19:18-20.
- Mohd Yusoff, M.N. (2014). R&D&C for Sustainable Agarwood Production. Presented at Agarwood Technology Talk 2015 on 2-3 Dec 2014 at FRIM's Auditorium, Kepong.
- Mole, S. and Waterman, P.G. (1987). Tannins as antifeedants to mammalian herbivores: Still an open questions? In: *Allelochemicals: Role in agriculture and Forestry* (Eds.): G.R. Waller. pp. 572-587. American Chemical Society, Washington DC.
- Mole, S., Ross, J.A.M. and Waterman, P.G. (1988). Light induced variation in phenolic levels in foliage of rain-forest plants. I. *Chemical Changes. J. Chem. Ecol.* 14:1-21.
- Moscone, E.A., Baranyi, M., Ebert, I., Greilhuber, J., Dorfer F.E. and Hunziker, A.T. (2003). Analysis of nuclear DNA content in *Capsicum* (Solanaceae) by flow cytometry and Fielgen densitometry. *Annals of Botany* 92:21-29.
- Muhammad, I., Alias, A. and Jusoh, M.Z. (2010). Produk Nilai Tambah Komponen Pokok Karas.
- Murashige, T. and Skoog, F. (1962). A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiologia Plantarum*. 15: 473-497.
- Müntzing, A. (1936). The Evolutionary Significance of Autopolyploidy. *Hereditas*. 21:363-378.

- Nakanishi, T., Yamagata, E., Yoneda, K., Nagashima, T., Kawasaki, I., Yoshida, T., Mori, H. and Miura, I. (1984). Three fragrant sesquiterpenes of agarwood. *Phytochemistry*. 23:2066-2067.
- Ng L.T., Chang, Y.S. and Azizol, A.K. (1997). A review on agar (gaharu) producing *Aquilaria* species. *Journal of Tropical Forest Products*. 202:272-285.
- Nie, C., Song, Y., Chen, D., Xue, P., Tu, P., Wang, K. and Chen, J. (2009). Studies on chemical constituents of leaves of *Aquilaria sinensis*. *Zhongguo Zhong Yao Za Zhi*. 34:858-60.
- Nikolova, M., Petrova, M., Zayona, E., Vitkova, A. and Evstatieva, L. (2013). Comparative study of *in vitro*, seedlings and *in vivo* grown plants of *Arnica montana* – polyphenols and free radical scavenging activity. *Acta Bot. Croat.* 72:13-22.
- Nobel, P.S. (1976). Photosynthetic rates of sun versus shades leaves of *Hyptis emoryi* Torr. *Plant Physiology*. 58:218-223.
- Nobuchi, T. and Siripacanadilok, S. (1991). Preliminary observation of *Aquilaria crassna* wood associated with the formation of aloewood. *Bult. Kyoto Univ. Forests*. 63:226-235.
- Noirot, M., Barre, P., Louarn, J., Duperray, C. and Hamon, S. (2002). Consequences of stoichiometric error on nuclear DNA content evaluation in *Coffea liberica* var. dewevrei using DAPI and propidium iodide. *Annals of Botany*. 89:385-389.
- Noirot, M., Barre, P., Louarn, J., Duperray, C. and Hamon, S. (2000). Nucleus-Cytosol Interactions-A Source of Stoichiometric Error in Flow Cytometric Estimation of Nuclear DNA Content in Plants. *Annals of Botany*. 86:309-316.
- Nor Asmah, H. (2000). Mikroperambatan *Shorea leprosula* Miq. dan *Aquilaria malaccensis* Lamk. Master thesis.
- Nor Azah, M.A., Chang, Y.S., Mailina, J., Abu Said, A., Abd. Majid, J., Saidatul Husni, S., Nor Hasnida, H. and Nik Yasmin, Y. (2008). Comparison of chemical profiles of selected gaharu oils from Peninsular Malaysia. *The Malaysia Journal of Analytical Sciences*. 12:338-340.
- Nor Azah, M.A., Majid, J.A., Mailina, J., Said, A.A., Husni, S.S., Hasnida, H.N., Arip, M.N. and Chang, Y.S. (2009). Profiles of selected supreme Agarwood oils from Malaysia. Proceedings of the Seminar on Medicinal & Aromatic Plants, Legend Hotel, Kuala Lumpur, pp 393-398.
- Nor Azah, M.A., Nurlaila, I., Mailina, J., Mohd. Nasir, T. and Mohd. Hezri, F.R. (2013). Analysis of aroma compounds from gaharu oil by Head-Space Solid Phase Microextraction and gas chromatography-mass spectrometry.

Proceedings of the Seminar on Medicinal & Aromatic Plants, Legend Hotel, Kuala Lumpur, 25-26 September, pp 180-185.

Notsuka, K., Tsuru, T. and Shiraishi, M. (2000). Induced polyploidy grapes via in vitro chromosome doubling. *Journal of the Japanese Society of Horticultural Science*. 16:543-551.[doi.org/10.2503/jjshs.69.543](https://doi.org/10.2503/jjshs.69.543).

NIST05 – Scientific Instrument Services. NIST05 MS Library and MS Search Program v.2.0d ([www.sisweb.com/manuals/nist05manual.pdf](http://www.sisweb.com/manuals/nist05manual.pdf)).

Okudera, Y. and Ito, M. (2009). Production of agarwood fragrant constituents in *Aquilaria calli* and cell suspension cultures. *Plant Biotechnology*. 26:307-315.

Otto, S.P. and Whitton, J. (2000). Polyploid incidence and evolution. *Annual Review of Genetics*. 34:401-437.

Oiyama, I. and Okudai, N. (1986). Production of colchicines-induced autotetraploid plants through micrografting in monoembryonic citrus cultivars. *Japan J. Breed.* Rahman, M. (2006). Introduction to flow cytometry. Serotec Ltd. Endeavour House. Oxford, UK.36:371-376.

Osborn, T.C., Pires, J.C., Birchler, J.A, Auger, D.L., Chen, J.Z., Lee, H.S, Madlung, A, Doerge, R.W., Colot, V. and Martienssen, R.A. (2003). Understanding mechanisms of novel gene expression in polyploids. *Trends in Genetics*. 19:141-147.

Otto, S.P. and Whitton, J. (2000). Polyploid incidence and evolution. *Annual Review of Genetics*. 34:401-437.

Owen, H. and Miller, A (1993). A comparison of staining techniques for somatic chromosomes of strawberry. *HortScience*. 28:155-156.

Petit, T.J. and Callaway, D.J. (2000). Breeding daylilies (*Heremocallis*) In: Callaway, D. and Callaway, M.B. eds. Breeding Ornamental Plants. Timber Press, Portland, Oregon USA. pp. 49-73.

Ponchet, M., Martin-Tanguy, J., Marais, A. and Martin, C. (1982). Hydroxycinnamoyl acid amides and aromatic amines in the inflorescence of some Araceae species. *Phytochem*. 21:2865-2869.

Portugal J and Waring, J. (1988). Assignment of DNA binding sites for 4', 6'-diamidino-2-2-phenylindole and bisbenzimidazole (Hoechst 33258). A comparative footprinting study. *Biochim. Biophys. Acta*. 949:158-168.

Pramod, K., Ansari, S.H. and Ali, J. (2010). Eugenol: a natural compound with versatile pharmacological actions. *Nat Prod Commun*. 5:1999–2006.

- Pudil, F., Uvira, R. and Janda, V. (2014). Volatile compounds in stinkhorn (*Phallus impudicus* L. ex. Pers.) at different stages of growth. *European Scientific Journal*. 10:163-171.
- Puite, K.J. and Ten Broeke, W.R.R. (1983). DNA staining of fixed and non-fixed plant protoplasts for flow cytometry with Hoechst 33342. *Plant Science Letters*. 32:79-88.
- Price H.J., Hodnet G. and Johnston, J.S. (2000). Sunflower (*Helianthus annuus*) leaves contain compounds that reduce nuclear propidium iodide fluorescence. *Annals of Botany*. 86:929-934.
- Rabinovitch, P.S. (1994). DNA content histogram and cell-cycle analysis. In: Darzynkiewicz, Z, Robinson, J.P, Crissman, H.A, eds. *Methods in cell biology: flow cytometry* Vol 41. San Diego Academic Press 263-296.
- Raffaele, S., Vaillau, F., L'éger, A., Joubès, J., Miersch, O., Huard, C., Blé, E., Mongrand, S., Domergue, F. and Roby, D. (2008). A MYB transcription factor regulates very-long-chain fatty acid biosynthesis for activation of the hypersensitive cell death response in *Arabidopsis*. *Plant Cell*. 20: 752-767.
- Ramsey, J. and Schemske, D.W. (1998). Pathways, mechanisms, and rates of polyploidy formation in flowering plants. *Annual Review of Ecology and Systematics*. 29:467-501.
- Ramsey, J. and Schemske, D.W. (2002). Neoploidy in flowering plants. *Annual Review of Ecology and Systematics*. 33:589-639.
- Ranney, T.G. (2006). Polyploidy: From Evolution to New Plant Development. [http://www.Ncsu.fletcher/staff/tranney/ranney\\_pages\\_06.pdf](http://www.Ncsu.fletcher/staff/tranney/ranney_pages_06.pdf).
- Rayburn, A.L., Biradar, D.P., Bullock, D.G., Nelson, R.L., Gourmet, C. and Wetzel, J.B. (1997). Nuclear DNA content diversity in Chinese soybean introductions. *Annals of Botany*. 80:321-325.
- Ricroch, A. and Brown, S.C. (1997). DNA base composition of allium genomes with different chromosome numbers. *Gene*. 205:255-260.
- Riedel, M., Eichne, A., Meimberg, H. and Jetter, R. (2007). Chemical composition of epicuticular wax crystals on the slippery zone in pitchers of five *Nepenthes* species and hybrids. *Planta*. 225: 1517-1534.
- Ringelmann, A., Riedel, M., Riederer, M. and Hilderbrandt, U. (2009). Two sides of a leaf blade: *Blumeria graminis* needs chemical cues in cuticular waxes of *Lolium perenne* for germination and differentiation. *Planta*. 230:95-105.
- Roux, N., Dolezel, J., Swennen, R. and Zapata-Arias, F.J. (2001). Effectiveness of three micropropagation techniques to dissociate cytochimeras in *Musa* spp. *Plant Cell, Tissue and Organ Culture*. 66:189-197.

- Roux, N., Toloza, A., Radecki, Z., Zapata-Arias, F.J. and Dolezel, J. (2003). Rapid detection of aneuploidy in *Musa* using flow cytometry. *Plant Cell Reports*. 21:483-490.
- Rudolf, K., Bohanec, B. and Hansen, M. (1999). Microspore culture of white cabbage, *Brassica oleracea* var. *capitata* L.: Genetic improvement of non-responsive cultivars and effect of genome doubling agents. *Plant Breeding*. 118:237-241.
- Sadgopal (1960). Explanatory studies in the development of essential oils and their constituents in aromatic plant. Part 1: Oil of agarwood. *SPC*. 33:41-46.
- Sadgopal and Varma, B.S. (1952). Agar oil from the wood of *Aquilaria agallocha* W Roxburgh. *SPC*. 25:169-174.
- Saeed, R., Khan, I.A. and Khan, F.A. (2006). Colchicine-induced tetraploidy and changes in allele frequencies in colchicines-treated populations of diploids assessed with RAPD Markers in *Gossypium arboreum* L. *Turkish Journal of Biology* 30:93-100.
- Sajjad, Y., Jaskani, M.J. and Asim, M. (2013). Effect Of Colchicine On In Vitro Polyploidy Induction In African Marigold (*Tagetes Erecta*). *Pak. J. Bot.* 45: 1255-1258.
- Sanwal, S.K., Rai, N., Singh, J. and Buragohain, J. (2010). Antioxidant phytochemicals and gingerol content in diploid and tetraploid clones of ginger (*Zingiber officinale* Roscoe). *Scientia Horticulturae*. 124:280-285.
- Schmiderer, C., Grassi, P., Novak, J., Weber, M. and Franz, C. (2008). Diversity of essential oil glands of clary sage (*Salvia sclarea* L., Lamiaceae). *Plant Biol*. 10:433-440.
- Segura, S., Scheinvar, L., Olalde, G., Leblanc, O., Filardo, S., Muratalla, G., Gallegos, C. and Flores, C. (2007). Genome sizes and ploidy levels in Mexican cactus pear species *Opuntia* (Tourn.) Mill. Series *Streptacanthae* Britton et Rose, *Leucotrichae* DC, *Heliabravoanae* Scheinvar and *Robustae* Britton et Rose. *Genet Resour Crop Evol*. 54:1033-1041.
- Shapiro, H.M. (2005). Front matter: in Practical Flow Cytometry, 4<sup>th</sup> edn. New York: John Wiley & Sons, Inc., Hoboken, NY, USA. doi:1002/0471722131.fmatter.
- Siti Suhaila, A.R., Norwati, M., Brian Yap, J.W., Mahani M. C., Kandasamy, K.I., Faridah Q.Z., Fadelah A.A., Nor Hasnida, H., Nazirah, A., Haliza, I., Muhd. Fuad, Y., Normah, B. (2013). The potential of *P. callosum* polyploids - towards the development of improved Malaysian slipper orchid. Proceeding at Conference on Forestry and Forest Product 2013, 11-12 November, Sunway Putra Place, Kuala Lumpur, Malaysia, pp 200-204.
- Soltis, P.S. (2005). Ancient and recent polyploidy in angiosperms. *New Phytologist*. 166:5-8.

- Soltis, D.E., Soltis, P.S. and Tate J.A. (2004). Advances in the study of polyploidy since plant speciation. *New Phytologist*. 161:173-191.
- Springer, T.L., McGraw, R.L. and Aiken, G.E. (2002). Variation of condensed tennins in roundhead *Lespedeza* germplasm. *Crop. Sci.* 42:2157-2160.
- Sun, Q.R., Sun, H.S., Li, L.G. and Bell, R.L. (2009). In vitro colchicines-induced polyploidy plantlet production and regeneration from leaf explants of the diploid pear (*Pyrus communis* L.) cultivar ‘Fertility’. *J. Hortic Sci Biotech.* 84:548-552.
- Stamp, N.E. and Bowers, M.D. (1994). Effects of cages, plant age and mechanical clipping on plantain chemistry. *Oecologia*. 99:66-71.
- Stavys, V., Weckman, A., Staiene, G. and Duchovskis, P. (2005). In vitro induction of polyploidy in japanese quince (*Chaenomeles japonica*). *Plant Cell, Tissue and Organ Culture*. 84:263-268.
- Stebbins, G.L. (1980). Polyploidy in plants: unsolved problems and prospects. In polyploidy: Biological Relevance, ed. WH Lewis. New York: Plenum.
- Stebbin, G.L. (1985). Polyploidy, hybridization, and the invasion of new habitats. *Ann. Missouri Bot Gard* .72:824-832.
- Stupar, R.M., Pudota, B.B., Brian, S.Y., Willem, A.R., Amy, L.H., Shu, O., Richard, E.V., James, S.B., Robert, J.E., Buell, C.R. and Jiming, J. (2007). Phenotypic and transcriptomic changes associated with potato autopolyploidization. *Genetics* 176:2055-2067.
- Svehlikova, V. and Repcak, M. (2000). Variation of apigenin quantity in diploid and tetraploid *Chamomilla rectita* (L.) Rauschert. *Plant Biol*. 2:403-407.
- Taha, D. (2010). Peluang dan cabaran industri gaharu di Malaysia. Paper presented at Seminar Kebangsaan dan Pameran Gaharu 2012, Universiti Putra Malaysia, Serdang, 22-23 Mac.
- Tajuddin, S.N. and Yusoff, M.M. (2010). Chemical composition of volatile oils of *Aquilaria malaccensis* (Thymeleaceae) from Malaysia. *Natural Product Communications*. 5: 1965-1968.
- Taiz, L. and Zeiger, E. (1991). Plant Physiology. The Benjamin/Cumming Pub. Co., Inc. 390 Bridge Parkway, Redwood City, California 90465. USA.
- Tao, R., Gao, M., Esumi, T., Kitamura, Y. and Yamada, A. (2009). High frequency ploidy variation observed in seedlings of a hexaploid persimmons cultivar ‘Fujiwaragosho’. *Acta Horticulture*. 833:631-638.
- Tate, J.A., Soltis, D.E and Soltis, P.S. (2005). Polyploidy in Plants. In The Evolution of the Genome, Elsevier. pg. 371-426.



- Thiem, B. and Sliwinska, E. (2003). Flow cytometric analysis of nuclear DNA content in cloudberry (*Rubus chamaemorus* L.) *in vitro* cultures. *Plant Science*. 164:129-134.
- Thiersch, T.R., Chandler, R.W., Wachtel, S.S. and Elias, S. (1989). Reference standards for flow cytometry and application in comparative studies of nuclear DNA content. *Cytometry*. 10:706-710.
- Tsuba, M., Katagiri, C., Takeuchi, Y. and Yamaoka, N. (2002). Chemical factors of the leaf surface involved in the morphogenesis of *Blumeria graminis*. *Physiol Mol Plant Pathol*. 60:51-57.
- Ulrich, S. (2000). Solid-phase microextraction in biomedical analysis. *Journal of Chromatography A*. 902:167-194.
- Väinölä, A. (2000). Polyploidisation and early screening of *Rhododendron* hybrids. *Euphytica*. 112: 239-244.
- van Dyke, M.W. and Dervan, P.B. (1983). Chromomycin, mithramycin, and olivomycin binding sites on heterogenous deoxyribonucleic acid. Footprinting with (methidiumpropyl-EDTA) iron (II). *Biochemistry*. 22:2373-2377.
- Vermes, I., Haanen, C. and Reutelingsperger, C. (2000). Flow cytometry of apoptotic cell death. *Journal of Immunological Methods*. 243:167-190.
- Wakana, A., Hanada, N., Park, S.M., Fukudome, I. and Kajiwara, K. (2005). Production of tetraploid forms of acid citrus cultivars by top grafting of shoots with sprouting axillary buds treated with colchicines. *J. Fac. Agr., Kyushu Univ* 50:93-102.
- Wakana, A., Sarikhani, H. Hanada, N., Fukudome, I., Kajiwara, K., Yasukochi, K., Hiramatsu, M. and Sakai, K. (2007). Characteristics of seedless berries of triploid hybrid grapes (*Vitis vinifera* Complex) derived from eighteen crosses. *J. Fac. Agr.* 52:337-344.
- Warner, D.A. and Edwards, G.E. (1989). Effects of polyploidy on photosynthetic rates, photosynthetic enzymes, contents of DNA, chlorophyll and sizes and number of photosynthetic cells in the C<sub>4</sub> dicot *Atriplex confertifolia*. *Plant Physiology*. 91:1143-1151.
- Warner, D.A., Ku, M.S.B. and Edwards, G.E. (1987). Photosynthesis, leaf anatomy, and cellular constituents in the polyploid C<sub>4</sub> grass *Panicum virgatum*. *Plant Physiology*. 84:461-466.
- Weaver, D.K., Wells, C.D., Dankel, F.V., Bertsch, W., Sing, S.E. and Sirharan, S. (1994). Insecticidal activity of floral, foliar and root extracts of *Tagetes minuta* (Asterales: Asteraceae) against adult Mexican bean weavils (Coleoptera: Bruchidae). *J. Econ. Entomol.* 87:1718-1725.

- Weber, S., Unker, F. and Friedt, W. (2005). Improved doubled haploid production protocol for *Brassica napus* using microspore colchicines treatment *in vitro* and ploidy determination by flow cytometry. *Plant Breeding*. 124:511-513.
- Wehner, T.C. (2008). Watermelon. Springer Science, New York.
- Whitmore, T.C. (1973). Tree Flora of Malaysia. (ed). Longman Group, London.
- Whitmore, T.C. (1972). Tree Flora of Malaya. (ed). A manual for foresters. Vol 2. Longman, Malaysia.
- Wilkes, J.G., Conte, E.D., Yongkyoung, K., Holcomb, M., Sutherland, J.B. and Miller, D.W. (2000). Sample preparation for the analysis of flavors and off-flavors in foods. *Journal of Chromatography A*. 880:3-33.
- Wu, Q., Deng, Ch., Shen, S., Song, G., Hu, Y., Fu, Du., Chen, J. and Zhang, X. (2004). Solid-Phase Microextraction followed by Gas Chromatography-Mass Spectrometry analysis of the volatile components of *Chrysanthemi indicis* in different growing areas. *Chromatographia*. 59:763-767.
- Xu, Y.H., Zhang, Z., Wang, M.X., Wei, J.H., Chen, H.J., Gao, Z.H., Sui, C., Luo, H.M., Zhang, X.L., Yang, Y., Meng H. and Li, W.L. (2013). Identification of genes related to agarwood formation: transcriptome analysis of healthy and wounded tissues of *Aquilaria sinensis*. *BMC Genomics*. 14:227. doi: 10.1186/1471-2164-14-227.
- Yagura, T., Ito, M., Kiuchi, F., Honda, G. and Shimada, Y. (2003). Four new 2-(2-phenylethyl) chromone derivatives from the withered wood of *Aquilaria sinensis*. *Chem Pharm Bull*. 51:560-564.
- Yan, G. (2001). Chromosome doubling of wax flower. Plant regenerated *in vitro*. The Proceeding of Wax Flower, pp. 11-20.
- Yang, X., Ye, C.Y., Cheng, Z.M., Tschaplinski, T.J., Wullschleger, S.D., Yin, W., Xia, Z. and Tuskan, G.A. (2011). Genomic aspects of research involving polyploidy plants. *Plant Cell Tiss. Organ Cult*. 104:387-397.
- Yokoya, K., Roberts, A.V. Mottley, J., Lewis, R. and Brandham, P.E. (2000). Nuclear DNA amounts in roses. *Annals of Botany*. 85:557-561.
- Zaki M and Dickinson H. (1995). Modification of cell development *in vitro*: the effect of colchicines on anther and isolated microspore culture in *Brassica napus*. *Plant Cell Tissue Organ Culture*. 40:255-270.
- Zderewicz (1971). Secondary metabolites. In: The Role of Chromosomal Change in Plant Evolution. Pp 140. *Oxford University Press: New York*.
- Zhang J, Zhang M and Deng X. (2007). Obtaining autotetraploids *in vitro* at a high frequency in *Citrus sinensis*. *Plant Cell Tissue Organ Culture*. 89: 211-216.

- Zhao, J. and Simmonds, D.H. (1995). Application of trifluralin to embryogenic microspore culture to generate doubled haploid plants in *Brassica napus*. *Physiologia Plantarum*. 95:304-309.
- Zheng, Y., Kristina, H., Verena, E.S., Alfons, G. and Ramon, A.T.R. (2009). A large number of tetraploid *Arabidopsis thaliana* lines, generated by a rapid strategy, reveal high stability of neo-tetraploid during consecutive generations. *Theoretical and Applied Genetics*. 118:1107-1119.
- Zlesak, D., Thill, C. and Anderson, N. (2005). Trifluralin-mediated polyploidisation of *Rosa chinensis minima* (Sims) Voss. *Euphytica*. 141: 281-290.
- Zoldoš, V., Papeš, D. Brown, S.C., Panaud, O. and Šiljak-Yakovlev, S. (1998). Genome size and base composition of seven *Quercus* species: inter- and intrapopulation variation. *Genome*. 41:162-168.
- Zygadlo, J.A., Maestri, D.M. and Ariza, E.L. (1993). The volatile oil *Tagetes* Argentina Cabrera. *J. Essent. Oil Res.* 5:85-86.