



UNIVERSITI PUTRA MALAYSIA

***PHYSICAL-CHEMICAL-ENZYMATIC PRETREATMENT PROCESS FOR
BIOCONVERSION OF KITCHEN WASTE INTO FERMENTABLE SUGAR
AS A FEEDSTOCK FOR BIOETHANOL PRODUCTION***

HALIMATUN SAADIAH HAFID

FBSB 2017 7



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By

HALIMATUN SAADIAH HAFID

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in fulfillment of the requirement for the Degree of Doctor of Philosophy**

April 2017

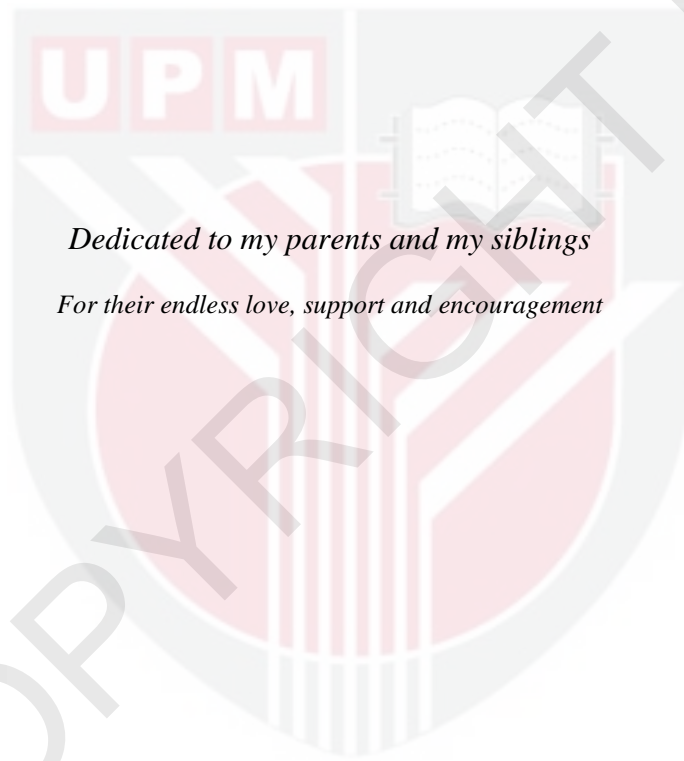


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*Dedicated to my parents and my siblings
For their endless love, support and encouragement*

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

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By

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April 2017

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Faculty : Biotechnology and Biomolecular Sciences

Sustainable conversion of municipal solid waste (MSW) into bioethanol taking into account the availability of the organic fraction of the waste which is kitchen waste that contains high carbohydrate, soluble sugar, starch, protein, lipid and other materials. It can be converted to fermentable sugar via hydrolysis and saccharification process which is still recognized as a rate-limiting step of the conversion process of kitchen waste to bioethanol. Hence, this study aimed to evaluate the physical, chemical, enzymatic and combination of pretreatment at enhancing the hydrolysis and maximizing the yield of fermentable sugar produced. This study is divided into three parts of pretreatment by (1) Hydrothermal and dilute acid; 2) Enzymatic pretreatment; 3) Combination of physical-chemical and enzymatic pretreatment, and followed by 4) Production of bioethanol using saccharified kitchen waste using locally isolated yeasts; *Saccharomyces cerevisiae*, *Candida parasilosis*, and *Lachancea fermentati*.

Initially, hydrolysis of kitchen waste by hydrothermal and dilute acid systems were carried out by varying the reaction temperatures (80, 90, 100°C) hydrolysis reactant; hot distilled water (hydrothermal), hydrochloric acid (HCl) and sulphuric acid (H₂SO₄) at different concentrations (0.5 - 2.0%) and evaluated using kinetic modelling approach by gPROMS software. A significant improvement of fermentable sugar production by 40.6% was observed at optimize condition of 1.5% HCl and 44.9% using 1.0% H₂SO₄ at 90°C as compared to hydrothermal pretreatment. The hydrolysis rate coefficient at 90°C for fermentable sugars production, $k_{r=1}$, was 0.68 gL⁻¹min⁻¹ for hydrothermal system and 1.61 and 1.88 gL⁻¹min⁻¹ in HCl and H₂SO₄ catalysed system, respectively. Enhanced hydrolysis of kitchen waste for fermentable sugar production was achieved by hydrothermal and dilute acid pretreatment.

Meanwhile, response surface methodology (RSM) technique is adopted in enzymatic pretreatment for a prediction of optimal condition of independent variables (pH, temperature, glucoamylase activity, kitchen waste loading and hydrolysis time) on fermentable sugar production and degree of saccharification. Quadratic RSM predicted maximum fermentable sugar production of 62.79 g/L and degree of saccharification (59.90%) at the optimal conditions; pH 5, temperature 60°C, glucoamylase activity of 85 U/mL and utilized 70 g/L of kitchen waste as a substrate at 10 hours hydrolysis time. The verification experiments successfully produced 65.71 ± 0.7 g/L of fermentable sugar with $55.3 \pm 0.4\%$ degree of saccharification which 31.4% higher than non-optimized condition indicating that the developed model was successfully used to predict fermentable sugar production at more than 90% accuracy.

The experiment was continued by applying single and combination pretreatments by hydrothermal, mild acid pretreatment of HCl and H₂SO₄ and with enzymatic hydrolysis by glucoamylase. The maximum total fermentable sugar produced after combination pretreatment by 1.5% HCl and glucoamylase produced 94.45 g/L of fermentable sugar consisted of 93.25 g/L glucose, 0.542 g/L sucrose, 0.348 g/L maltose, and 0.321 g/L fructose. An increase of 55.8% and 91.8% of fermentable sugar production was obtained by comparing with single glucoamylase and 1.5% HCl pretreatment, respectively. From FTIR analysis, the decrease of aliphatic absorbance bands of polysaccharides at 2851 and 2923 cm⁻¹ and the increase on structures of carbonyl absorbance bands at 1600 cm⁻¹ reflects the progress of the kitchen waste hydrolysis to fermentable sugars. For total cost and profit estimation, combination of 1.5% HCl and glucoamylase pretreatment was the most profitable process as the minimum selling price of glucose was USD 0.101/g kitchen waste. The combination pretreatment method was successfully enhance the production of fermentable sugar from kitchen waste.

Production of fermentable sugar using acid-pretreated and enzymatic hydrolysis of kitchen waste was then conducted in 2L of bioreactor. The results suggested that a significant increase in fermentable sugar production to 103.4 ± 0.04 g/L (2.04-folds) with conversion efficiency of 86.8% was observed via sequential acid-enzyme pretreatment as compared to dilute acid (42.4%) and glucoamylase enzyme (50.6%), respectively. An increased in total fermentable sugar to 150.5 ± 0.11 g/L which consist of glucose (128.47 g/L), fructose (6.24 g/L), sucrose (5.59 g/L) and maltose (10.18 g/L) was successfully recovered after downstream processing. The fermentable sugars obtained were subsequently converted to bioethanol by locally isolated yeasts; *Saccharomyces cerevisiae*, *Candida parasilosis*, and *Lachancea fermentati* produce ethanol yield ranging from 0.45 g/g to 0.5 g/g after 24 h which was equivalent to 82.06 - 98.19% of conversion efficiency based on theoretical yield that was comparable with using commercial glucose. The finding indicates that kitchen waste can be considered as a promising substrate for bioethanol production.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PROSES PRA-RAWATAN SECARA FIZIKAL-KIMIA-ENZIM BAGI
PENGHASILAN GULA TERFERMENTASI DARIPADA SISA BUANGAN
DAPUR SEBAGAI BAHAN MENTAH PENGHASILAN BIOETANOL**

Oleh

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Penukaran lestari sisa buangan pepejal bandar (MSW) kepada bioetanol mengambil kira kandungan organik iaitu sisa buangan dapur yang mengandungi karbohidrat yang tinggi, gula terlarut, kanji, protin, lipid dan bahan yang lain. Ia boleh ditukarkan kepada gula fermentasi melalui proses hidrolisis dan pensakaridaan yang masih menjadi faktor-pengehad kepada proses penukaran sisa buangan dapur kepada bioetanol. Oleh itu, kajian ini menilai pelbagai kaedah pra-rawatan fizikal, kimia, enzim dan kombinasi pra-rawatan untuk meningkatkan hidrolisis dan memaksimumkan penghasilan gula. Kajian ini terbahagi kepada tiga bahagian pra-rawatan melalui (1) Hidroterma dan pemangkin asid-cair; (2) Pra-rawatan enzim; (3) Kombinasi pra-rawatan fizikal, kimia dan enzim; dan diikuti oleh 4) Penghasilan bioetanol menggunakan hidrolisis pensakaridaan sisa buangan dapur oleh yis yang telah dipencilkan dari kawasan tempatan; *Saccharomyces cerevisiae*, *Candida parasitosis*, dan *Lachancea fermentati*.

Permulaannya, hidrolisis sisa buangan dapur melalui kaedah sistem hidroterma dan asid cair dijalankan dengan mempelbagaikan suhu tindakbalas (80, 90, 100°C), bahan tindak balas; air suling panas (hidroterma), asid hidroklorik (HCl) dan asid sulfurik (H₂SO₄) pada kepekatan berbeza (0.5-2.0%) dan dinilai menggunakan model kinetik oleh perisian gPROMS. Peningkatan ketara gula terfermentasi sebanyak 40.6% diperoleh pada keadaan optimum 1.5% HCl dan 44.9% pada keadaan 1.0% H₂SO₄ berbanding pra-rawatan hidroterma. Pekali kadar hidrolisis pada 90°C penghasilan gula terfermentasi ialah $k_{r=1}$, 0.68 gL⁻¹min⁻¹ bagi sistem hidroterma manakala 1.61 dan 1.88 gL⁻¹min⁻¹ bagi sistem HCl dan H₂SO₄. Peningkatan gula terfermentasi dalam hidrolisis sisa buangan dapur melalui pra-rawatan hidroterma dan asid cair berjaya dicapai.

Sementara itu, kaedah permukaan tindakbalas (RSM) digunakan semasa pra-rawatan enzim untuk menganggar keadaan optimum oleh faktor-faktor bebas (pH, suhu, aktiviti glukoamilase, beban sisa buangan dapur, dan masa hidrolisis) kepada penghasilan gula fermentasi dan tahap pensakaridaan. Kuadratik RSM meramalkan penghasilan gula terfermentasi tertinggi ialah 62.79 g/L dan tahap pensakaridaan (59.90%) pada keadaan optimum; pH 5, suhu 60°C, aktiviti glukoamilase 85 U/mL dan menggunakan 70 g/L sisa buangan dapur sepanjang 10 jam masa hidrolisis. Eksperimen pengesahan berjaya menghasilkan 65.71 ± 0.7 g/L gula terfermentasi dengan 55.3 ± 0.4 % tahap pensakaridaan dimana 31.4% lebih tinggi berbanding tanpa keadaan optima menunjukkan model yang direka bentuk berjaya digunakan untuk meramal penghasilan gula fermentasi dengan lebih 90% ketepatan.

Eksperimen diteruskan dengan pra-rawatan tunggal dan kombinasi pra-rawatan oleh hidroterma, pra-rawatan asid cair oleh HCl dan H₂SO₄ dan hidrolisis enzim oleh glukoamilase. Jumlah penghasilan gula terfermentasi tertinggi diperolehi melalui kombinasi pra-rawatan oleh 1.5% HCl dan glukoamilase menghasilkan 94.45 g/L gula terfermentasi dengan mengandungi 93.25 g/L glukosa, 0.542 g/L sukrosa, 0.248 g/L maltose dan 0.321 g/L fruktosa. Peningkatan sebanyak 55.8% dan 91.8% gula terfermentasi diperolehi berbanding pra-rawatan tunggal menggunakan glucoamylase dan 1.5% HCl sahaja. Dari analisa FTIR, pengurangan kuantiti penyerapan kumpulan alifatik polisakarida pada 2851 dan 2923 cm⁻¹ dan peningkatan kuantiti penyerapan pada struktur karbonil di 1600cm⁻¹ menggambarkan tindakbalas hidrolisis sisa buangan dapur kepada gula terfermentasi. Berdasarkan analisa anggaran kos dan untung, kombinasi 1.5% HCl dan glukoamilase adalah pra-rawatan yang menguntungkan dengan memberikan harga jualan minimum glukosa sebanyak USD 0.101/g sisa buangan dapur. Teknik kombinasi pra-rawatan berjaya meningkatkan penghasilan gula terfermentasi daripada sisa buangan dapur.

Penghasilan gula terfermentasi dari sisa buangan dapur terawat asid dan hidrolisis enzim kemudiannya dilakukan di dalam bioreactor berkapasiti 2 L. Keputusan menunjukkan peningkatan ketara penghasilan gula terfermentasi kepada 103.4 ± 0.04 g/L (2.04-kali ganda) dengan kecekapan penukaran 86.8% diperolehi melalui jujukan pra-rawatan asid-enzim berbanding pra-rawatan asid cair (42.4%) dan pra-rawatan enzim glukoamilase (50.6%). Peningkatan gula fermentasi kepada 150.5 ± 0.11 g/L mengandungi glukosa (128.47 g/L), fruktosa (6.24 g/L), sukrosa (5.59 g/L) dan maltose (10.18 g/L) berjaya diperolehi selepas proses hiliran dilakukan. Gula terfermentasi kemudiannya ditukarkan kepada bioetanol oleh yis yang dipencilkan; *Saccharomyces cerevisiae*, *Candida parasitosis*, dan *Lachancea fermentati* menghasilkan etanol tertinggi diantara 0.45 g/g - 0.5 g/g selepas 24 jam penapaian bersamaan dengan 82.06 - 98.19% kecekapan penukaran berdasarkan hasil teori dan setanding dengan glukosa komersil. Penemuan ini menunjukkan sisa buangan dapur dianggap substrat yang berpotensi untuk penghasilan bioetanol.

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I certify that a Thesis Examination Committee has met on 3 April 2017 to conduct the final examination of Halimatun Saadiah Hafid on her thesis entitled "Physical-Chemical-Enzymatic Pretreatment Process for Bioconversion of Kitchen Waste into Fermentable Sugar as Feedstock for Bioethanol Production" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
BOD	Biological Oxygen Demand
CCD	Central composite design
CHO	Carbohydrate
COD	Chemical oxygen demand
H ₂ SO ₄	Sulfuric acid
HAc	Acetic acid
HCl	Hydrochloric acid
HLa	Lactic acid
HPLC	High Performance Liquid Chromatography
HPr	Propionic acid
KH ₂ PO ₄ -	Potassium dihydrogen phosphate
MESP	Minimum bioethanol selling price
MgSO ₄ .7H ₂ O	Magnesium sulphate heptahydrate
MSW	Municipal Solid Waste
NaOH	Sodium hydroxide
RSM	Response Surface Methodology
TKN	Total Kjeldahl Nitrogen
TS	Total Solid
TSS	Total Suspended Solid
YPD	Yeast Peptone Dextrose

CHAPTER 1

INTRODUCTION

1.1 Research Background

The management of municipal solid waste (MSW) has been continuously be a major challenge in Malaysia attributed to the rapid urbanization and industrialization especially in urban areas. Along with the economic growth, Malaysian population has tremendously increased at the rate of 2.4% per annum, thus increase the consumption rate and accelerate the daily generation of solid wastes from 0.5-0.8 kg/person/day to 1.7 kg/person/day in major cities (Budhiarta et al., 2012). A recent study reported that Malaysians produced 33,000 tons of solid waste daily in 2012, exceeding the government's projected waste production of 30,000 tonnes daily by 2020 (Mokhtar, 2013). A typical MSW management system applied landfill as a primary choice of handling these waste, however, it displays an array of problem of crude open dumping, water pollution from the leachate generation and the breeding of flies and vermin that becoming the major environmental and public health problem due to the high concentration of organic material (71.6%) and moisture content present (Latifah et al., 2009).

The organic material of MSW is mainly referring to food or kitchen waste that is usually discharged from restaurants, kitchens, and cafeterias and leftovers from food industry. Kitchen waste is highly biodegradable comprises a rich carbon and nutrient such as carbohydrate, soluble sugar, starch, protein, lipids and other inorganic materials (Moon and Song, 2011). Pleissner and Lin (2013) reported the composition of kitchen waste consisting of approximately 60% carbohydrate, 20% proteins, 10% lipids and 10% of other materials. Current research performed is utilizing kitchen waste an alternative nutrient medium for conversion to organic acids (Omar et al., 2009; Bo et al., 2007), biogas (Ma et al., 2011; Munda et al., 2012), and bioethanol (Uncu and Cekmecelioglu, 2011; Tang et al., 2008). Various application of kitchen waste makes it a valuable raw material for the recovery of nutrient needed, however, its hydrolysis of particulates solid is still rate-limiting step in its application in many biotechnological process (Pleissner and Lin, 2013). The differences in individual biodegradability of kitchen waste compositions, such as carbohydrate, starch, protein, fat and fiber generates various degradation characteristics and may cause decreases in hydrolysis efficiency (Moon and Song, 2011; Moon et al., 2009). Hydrolysis of carbohydrate in kitchen waste may result in the breakdown of glycosidic bonds with releasing polysaccharides as monosaccharides, which are more amenable to fermentation especially for bioconversion to bioethanol (Kiran et al., 2014). Hence, the hydrolysis of kitchen waste can be performed by

physical that normally involved thermal hydrolysis, chemical by acid hydrolysis, and biological by enzymatic hydrolysis and combinations of them.

Generally, thermal hydrolysis may lead to partial degradation of sugars and other nutritional components due to Maillard reaction that make the amounts of targeted sugar and amino acids decreased (Kiran et al., 2014), whereas, chemical hydrolysis by acid may penetrate and disrupt the structure of biomass more easily thus increase the hydrolysis rate, however, the rapid degradation of sugars particularly glucose and generation of unnecessary compound make it less advantages (Li et al., 2012). In contrast, enzymatic hydrolysis by α -amylase and glucoamylase is more environmentally friendly for obtaining sugars; but, the low enzymatic accessibility due to the nature and recalcitrant of the biomass is a key problem for the kitchen waste-to-bioethanol process (Kumar et al., 2009). Moreover, most of the enzymatic processes are too slow thus limiting application at the industrial level and the excessive cost of enzymes may increase the overall processing cost (Duvernay et al., 2013; Vavouraki et al., 2012). In search of the viable alternative substrate for fermentable sugar and bioethanol production, the feasibility of the kitchen waste need to be further explored as its contain high nutritional value and organic content with a large volume of waste generated apart of its underused resources. As a pretreatment remains the most hurdles in producing technical and economically viable fermentable sugar and bioethanol production from biomass, the integrated pretreatment using a combination methods of physical and chemical by dilute acid solution together with the enzymatic saccharification is a potential efforts directed towards increasing the biodegradability of complex biomass particularly organic kitchen waste to monomeric sugars (Fred-Guelfo et al., 2011).

The integrated pretreatment has been pursued as a promising approach to break the structure of kitchen waste and expose the starch component and cellulose to enzyme action for higher efficiency of sugars production, decrease formation of inhibitory and simultaneously shortening the process time (Mood et al., 2013). The interest in utilizing integrated pretreatment is facilitate by the need to improve the efficacy of kitchen waste degradation, with a more recent focus on operating at high concentration of solid matters for fermentable sugar production. Modenbach and Nokes, (2013) reported at high solid content of biomass has been identified as the largest contributor in achieving high yields sugar in a timely manner of the kitchen waste to bioethanol conversion process, mainly because a significant portion of sugars produced translate into higher bioethanol concentration. In a study conducted by Tang et al. (2008), the combination of mechanical treatment by compact chopper and glucoamylase produced 74.1 g/L of glucose whereas about 200 g/L of glucose was successfully recovered using the same combination pretreatment (Yan et al., 2010). Vavouraki et al. (2014) reported an increase of ~300% of glucose concentrations after subjected to pretreatment with acid or alkaline in combination with enzymatic hydrolysis. Besides, combination thermo-chemical pretreatment

improve the digestibility of solid content for high rate of hydrolysis (Li et al., 2015).

In developing a technical and economically feasible pretreatment process, a lot of parameters such as type of pretreatment (physical, chemical, enzymatic), operating temperature, pH, pretreatment time, kitchen waste concentration and other unit operations involved need to be optimized to ensure higher production of fermentable sugar obtained with lower production of inhibitors and lesser environmental impacts. Bioprocess modelling and optimization has traditionally been carried out using One Variable at a Time (OVAT) and Response Surface Methodology (RSM) to evaluate the process parameters to the output products. RSM is favored by many researchers due to rapid and reliable statistical analysis and generate interactive effects of input process parameters of the complex substrate such as kitchen waste (Hafid et al., 2011). An extensive understanding of the integrated pretreatment process on the kitchen waste constituent, physical and chemical changes involved, possible generation of toxic compound is needed to evaluate the overall performance and efficiency of selected pretreatment methods. Apart from the technical aspect of fermentable sugar and bioethanol production, the effect of the pretreatment process and parameter on the economic analysis of the fermentable sugar is reflected in the cost estimation analysis. The excessive cost of pretreatments may render the overall process and economically unsustainable. In addition, the chosen pretreatment must be attractive to be applied in which it could save time, chemical used, laboratory apparatus and manpower. The most appropriate pretreatment methods takes into account the technical and economic evaluation towards the sustainable decision making process.

This study present a unique set of understanding on bioconversion of kitchen waste into fermentable sugars by integrated physical, chemical and enzymatic pretreatment. The bioethanol production was performed utilizing the fermentable sugar obtained in the kitchen waste sacharified liquid by three locally isolated yeasts; a single culture of *Saccharomyces cerevisiae*, *Lachancea fermentati* and *Candida parasilosis* in separate hydrolysis and fermentation process. The feasibility of converting kitchen waste-fermentable sugars into bioethanol is evaluated and expected to increase interest in utilizing kitchen waste as a feedstock for potential development of bioethanol production.

1.2 Problem Statements

In literature, very limited studies focus on the utilization of kitchen waste for fermentable sugar and bioethanol production as compared to lignocellulosic biomass probably due to the heterogeneity and inconsistent compositions and laborious separation process of kitchen waste. An establishment of a standard

preparation of kitchen waste need to be develop to overcome varies composition of kitchen waste before being subjected to any pretreatment for valuable products. In addition, very rare studies reported on the effect of integrated and combination pretreatment of physical and chemical prior to enzymatic hydrolysis as compared to single pretreatment in which is usually claimed to be much simpler with low-cost technology. Pretreatment strategies should be establish on a basis of the heterogeneity of the biomass to avoid selective deconstruction and fractional conversion to other by-product. A major demerit of single pretreatment is apportion of multiple products thereby reduce the concentration of main fermentable sugars. Furthermore, an appropriate pretreatment is crucial in dealing with high solid content of the biomass, particularly kitchen waste to ensure complete degradation occurs for higher yield of fermentable sugar. Despite of the many pretreatment method tested in the literature, there is still a need to establish more defined pretreatment based approach for kitchen waste conversion and evaluate the processing conditions for an effective fermentable sugar production.

1.3 Objectives

This study investigated the improvement of kitchen waste hydrolysis by single and combination pretreatments method through physical, chemical, and enzymatic approaches for higher yield of fermentable sugar production with a maximum degradation efficiency of the kitchen waste. Upon completion of the pretreatment, the hydrolysate was recovered and inoculated with ethanol-fermenting microorganisms and subjected to bioethanol production in separate hydrolysis and fermentation (SHF) process. Therefore, the specific objectives of this study were:

- 1) To investigate the environmental factors of kitchen waste hydrolysis for fermentable sugar production by hydrothermal and dilute acid pretreatment methods.
- 2) To optimize the enzymatic pretreatment method of kitchen waste for fermentable sugar production using response surface methodology.
- 3) To enhance fermentable sugar production from kitchen waste hydrolysis using combination of physical-chemical-enzymatic pretreatment method.
- 4) To produce fermentable sugar using enzymatic sequential batch of kitchen waste treated acid in 2L bioreactor for bioethanol production by locally isolated yeasts.

REFERENCES

- Abdullah, B., and Ariyanti, D. (2012). Enhancing ethanol production by fermentation using *Saccharomyces cerevisiae* under vacuum condition in batch operation. *International Journal of Renewable Energy Development* 1: 6-9.
- Abu Bakar, N. K. (2011). Microbial conversion of oil palm empty fruit bunch to fermentable sugars (polyoses) for bioethanol production. Master Thesis. Universiti Putra Malaysia.
- Adejumo, A.L., Aderibigbe, F.A., and Owolabi, R.U. (2013). Relationship between α -amylase degradation and amylose/amylopectin content of maize starches. *Advance Applied Science Research* 4(2): 315-319.
- Adnan, N. A. A. (2014). Glycerol as alternative substrate for bioethanol production using free and immobilized *Escherichia coli* SS1. Master Thesis. Universiti Putra Malaysia.
- Alvira, P., Tomas-Pejo, E., Ballesteros, M., and Negro, M. J. (2010). Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: A review. *Bioresource Technology* 101: 4851-4861.
- Anto, H., Trivedi, U.B., and Patel, K.C. (2006). Glucoamylase production by solid state fermentation using flake manufacturing waste products as substrate. *Bioresource Technology* 97: 1161-1166.
- APHA, 1985. Standard method for the examination of water and wastewater 16 th ed. American Public Health Association.
- Arapoglou, D., Varzakas, T., Vlyssides, A., and Israilidise, C. (2014). Ethanol production from potato peel waste (PPW). *Waste Management* 30: 1898-1902.
- Ariunbaatar, J., Panico, A., Esposito, G., Pirozzi, F., and Lens, P.N.L. (2014). Pretreatment methods to enhance anaerobic digestion of organic solid waste. *Applied Energy* 123: 143-156.
- Ayansina, A.D.V., and Owoseni, A.A. (2010). Studies on amylolytic enzyme synthesized by *Aspergillus flavus* associated with moldy bread. *Pakistan Journal of Nutrition* 9(5): 434-437.
- Balat, M. (2011). Production of bioethanol from lignocellulosic materials via the biochemical pathway: A review. *Energy Conversion and Management* 52: 858-875.

- Balat, M., and Balat, H. (2009). Recent trends in global production and utilization of bioethanol fuel. *Applied Energy* 86: 2273-2282.
- Ballesteros, M., Saez, F., Ballesteros, I., Manzanares, P., Negro, M.J., Martinez, J.M., Castaneda, R., and Dominguez, J.M.O. (2010). Ethanol production from the organic fraction obtained after thermal pretreatment of municipal solid waste. *Applied Biochemistry and Biotechnology* 161: 423-431.
- Bals, B., Wedding, C., Balan, V., Sendich, E., and Dale, B. (2011). Evaluating the impact of ammonia fiber expansion (AFEX) pretreatment conditions on the cost of ethanol production. *Bioresource Technology* 102: 1277-1283.
- Battista, F., Mancini, G., Ruggeri, B., and Fino, D. (2016). Selection of the best pretreatment for hydrogen and bioethanol production from olive oil waste products. *Renewable Energy* 88: 401-407.
- Behera, S., Arora, R., Nandhugural, N., and Kumar, S. (2014). Importance of chemical pretreatment for bioconversion of lignocellulosic biomass. *Renewable and Sustainable Energy Reviews* 36: 91-106.
- Bo, Z., Wei-min, C., and Pin-jing, H. (2007). Influence of lactic acid on the two phase anaerobic digestion of kitchen wastes. *Journal of Environmental Science*. 19: 244-249.
- Boelema, A. (2013). Hydrolysis as valorization route for waste streams: A case study for wastepaper. Master Programme Energy and Environmental Sciences.
- Borne, J.J.G.C., Kabel, M.A., Briens, M., Poel, A.F.B., and Hendriks, W.H. (2012). Effects of pretreatment of wheat bran on the quality of protein-rich residue for animal feeding and on monosaccharide release for ethanol production. *Bioresource Technology* 124: 446-454.
- Bougrier, C., Delgenes, J.P., and Carrere, H. (2008). Effects of thermal treatments on five different waste activated sludge samples solubilization, physical properties and anaerobic digestion. *Chemical Engineering Journal* 139: 236-244.
- Bryksa, B.C., and Yada, R.Y. (2009). *Food Biochemistry*. In *Food Science and Technology*. Ed Campbell-Platt G. pp: 57-83 United Kingdom: A John Wiley & Sons, Ltd., Publication.
- Budhiarta, I., Siwar, C., and Basri, H. (2012). Current status of municipal solid waste generation in Malaysia. *International Journal on Advanced Science Engineering Information Technology* 2: 16-21.

- Castro, E., Diaz, M.J., Cara, C., Ruiz, E., Romero, I., and Moya, M. (2011). Dilute acid pretreatment of rapeseed straw for fermentable sugar generation. *Bioresource Technology* 102: 1270-1276.
- Cekmecelioglu, D., and Uncu, O.N. (2012). Kinetic modelling of enzymatic hydrolysis of pretreated kitchen waste for enhancing bioethanol production. *Waste Management* 33(3): 735-739.
- Cesaro, A., and Belgiorno, V. (2014). Pretreatment methods to improve anaerobic biodegradability of organic municipal solid waste fractions. A review. *Chemical Engineering Journal* 240: 24-37.
- Chandel, A.K., Antunes, F.A.F., de Arruda, P.V., Milessi, T.S.S., de Silva, S.S., and Felipe, M.D.G.D.A (2012). *Dilute acid hydrolysis of Agro-residues for the depolymerisation of hemicellulose: State-of-the-art. In D-Xylitol: Fermentative production, application and commercialization.* Eds da Silva S.S., Chandel A.K. pp: 39-61 Springer-Verlag Berlin Heidelberg.
- Chen, J., Liu, D., Shi, B., Wang, H., Cheng, Y., and Zhang, W. (2013). Optimization of hydrolysis conditions for the production of glucomannooligosaccharides from konjac using -mannanase by response surface methodology. *Carbohydrate Polymers* 93: 81-88.
- Cheng, S.S., Chao, Y.C., Wong, S.C., Chen, C.C., Yang, K.H., and Yang, Y.F. (2012). Study on hydrogen production potential utilizing leachate from aerobic bio-leaching bed fed with napiergrass and kitchen waste. *Energy Procedia* 29:72-81.
- Chethana, S.H., Pratap, B., Roy, S., Jaiswal, A., Shruthi, S.D., and Vedamurthy, A.B. (2011). Bioethanol production from rice waste: A low cost motor fuel. *Pharmacologyonline* 3: 125-134.
- Choi, S.A., Choi, W.I., Lee, J.S., Kim, S.W., Lee, G.A., Yun, J., and Park, J.Y. (2015). Hydrothermal acid treatment for sugar extraction from *Golenkinia* sp. *Bioresource Technology* 190: 408-411.
- Dagnino, E.P., Chamorro, E.R., Romano, S.D., Felissia, F.E., and Area, M.C. (2013). Optimization of the acid pretreatment of rice hulls to obtain fermentable sugars for bioethanol. *Industrial Crops and Products* 42: 363-368.
- Dors, G., Mendes, A.A., Pereira, E.B., Castro, H.F.D., and Jr, A.F. (2013). Simultaneous enzymatic hydrolysis and anaerobic biodegradation of lipid-rich wastewater from poultry industry. *Applied Water Science* 3(1): 343-349.
- Dung, T.N.B., Sen, B., Chen, C.C., Kumar, G., and Lin, C.Y. (2014). Food waste to bioenergy via anaerobic processes. The 6th International Conference on Applied Energy-ICAE2014. *Energy Procedia* 61: 307-312.

- Dutta, J.R., Dutta, P.K. and Banerjee, R. (2004). Optimization of culture parameters for extracellular protease production from newly isolated *Pseudomonas* sp. using response surface and artificial neural network models. *Process Biochemistry* 39: 2193-2198.
- Duvarnay, W.H., Chinn, M.S., and Yenko, G.C. (2013). Hydrolysis and fermentation of sweet potatoes for production of fermentable sugars and ethanol. *Industrial Crops and Products* 42: 527-537.
- El-Fels, L., Zamama, M., El-Asli, A., and Hafidi, M. (2014). Assessment of biotransformation of organic matter during co-composting of sewage sludge – lignocellulosic waste chemical, FTIR analysis and phytotoxicity tests. *International Biodeterioration Biodegradation* 87: 128-137.
- El-Tayeb, T.S., Abdelhafiz, A.A., Ali, S.H., and Ramadan, E.M. (2012). Effect of acid hydrolysis and fungal biotreatment on agro-industrial wastes for obtainment of free sugars for bioethanol production. *Brazilian Journal of Microbiology*: 1523-1535.
- Fdez-Duelfo, L.A., Alvarez-Gallego, C., Sales, D., and Romero, L.I. (2011). The use of thermochemical and biological pretreatments to enhance organic matter hydrolysis and solubilization from organic fraction of municipal solid waste (OFMSW). *Chemical Engineering Journal* 168: 249-254.
- Feng, Y., Qi, X., Jian, H., Sun, R., and Jiang, J. (2012). Effects of inhibitors on enzymatic hydrolysis and simultaneous saccharification fermentation for lactic acid production from steam explosion pretreated Lespedeza stalks. *BioResources* 7(3): 3755-3766.
- Fountoulakis, M., and Lahm H.W. (1998). Hydrolysis and amino acid composition analysis of proteins. *Journal of Chromatography A* 826(2): 109-134.
- Frazier, R.A. (2009). *Food chemistry. In Food Science and Technology*. Ed Campbell-Platt G. pp 5-8. United Kingdom: A John Wiley & Sons, Ltd., Publication.
- Gabhane, J., William, S.P.M.P., Gadhe, A., Rath, R., Vaidya, A.N., and Wate, S. (2014). Pretreatment of banana agricultural waste for bioethanol production: Individual and interactive effects of acid and alkali pretreatments with autoclaving, microwave heating and ultrasonication. *Waste Management* 34: 498-503.
- Ganesan, B., Seefeldt, K., and Weimer, B.C. (2004). Fatty acids production from amino acids and α -keto acids by *Brevibacterium linens* BL2. *Applied and Environmental Microbiology* 70(11): 6385-6393.

- Gao, Y., Xu, J., Yuan, Z., Zhang, Y., Liu, Y., and Liang, C. (2014). Optimization of fed-batch enzymatic hydrolysis from alkali-pretreated sugarcane bagasse for high-concentration sugar production. *Bioresource Technology* 167: 41-45.
- Girisuta, B., Dussan, K., Haverty, D., Leahy, J.J., and Hayes, M.H.B. (2013). A kinetic study of acid catalysed hydrolysis of sugarcane bagasse to levulinic acid. *Chemical Engineering Journal* 217: 61-70.
- Grahovac, J., Dodic, J., Jokic, A., Dodic, S., and Popov, S. (2012). Optimization of ethanol production from thick juice: A response surface methodology approach. *Fuel* 93: 221-228.
- Grohmann, K., and Bothast, R.J. (1997). Saccharification of corn fibre by combined treatment with dilute sulphuric acid and enzymes. *Process Biochemistry* 32: 405-415.
- Guerro-Rodriguez, E., Portilla-Rivera, O.M., Ramirez, J.A., and Vazquez, M. (2012). Modelling of acid hydrolysis of potato (*Solanum tuberosum*) for fermentative purposes. *Biomass & Bioenergy* 42: 59-68.
- Guha, S.K., Kobayashi, H., and Fukuoka, A. (2010). *Conversion of cellulose to sugars*. In *Thermochemical conversion of biomass to liquid fuels and chemicals*. ed Crocker M. pp 344-361. United Kingdom: RSC Energy and Environment Series.
- Hafid, H.S. (2011). Optimization of microbial process for the conversion of kitchen waste into organic acids using response surface methodology. Master Thesis. Universiti Putra Malaysia.
- Hafid, H.S., Nor' Aini, A.R., Suraini, A.A., and Hassan, M.A (2011). Enhancement of organic acids production from kitchen waste using response surface methodology. *African Journal of Biotechnology* 10(65): 14507-14515.
- Hafid, H.S., Nor' Aini, A.R., Umi Kalsom, M.S., and Baharuddin, A.S. (2015). Enhanced fermentable sugar production from kitchen waste using various pretreatments. *Journal of Environmental Management* 156: 290-298.
- Hahn-Ha'gerdal, B., Galbe, M., Gorwa-Grauslund, M.F., Lide'n G., and Zacchi G. (2006). Bio-ethanol - the fuel of tomorrow from the residues of today. *TRENDS in Biotechnology* 24(12): 549-556.
- Harun, R., and Danquah, M.K. (2011). Enzymatic hydrolysis of microalgal biomass for bioethanol production. *Chemical Engineering Journal* 168:1079-1084.

- Ho, S.H., Hyang, S.W., Chen, C.Y., Hasunuma, T., Kondo, A., and Chang, J.S. (2013). Bioethanol production using carbohydrate rich microalgae biomass as feedstock. *Bioresource Technology* 135: 191-198.
- Hong, Y.S., and Yoon, H.H. (2011). Ethanol production from food residues. *Biomass and Bioenergy* 35: 3271-3275.
- Horvath, I.S., Franzen, C.J., Taherzadeh, M.J., Niklasson, C., and Liden, G. (2003). Effects of furfural on the respiratory metabolism of *Saccharomyces cerevisiae* in glucose-limited chemostats. *Applied and Environmental Microbiology* 69(7): 4076-4086.
- Hoseinpour, H., Karimi, K., Zilouei, H., and Taherzadeh, M.J. (2010). Simultaneous pretreatment of lignocellulose and hydrolysis of starch in mixtures to sugars. *Bioresources* 5(4): 2457-2469.
- Hu, C.C., Giannis, A., Chen, C.L., and Wang, J.Y. (2014). Evaluation of hydrogen producing cultures using pretreated food waste. *International Journal of Hydrogen Energy* 39(33): 19337-19342.
- Izumi, K., Okishio, Y., Nagao, N., Niwa, C., Yamamoto, S., and Toda, T. (2010). Effects of particle size on anaerobic digestion of food waste. *International Biodeterioration and Biodegradation* 64:601-608.
- Jianguo, J., Changxiu, G., Jiaming, W., Sicong, T., and Yujing, Z. (2014). Effects of ultrasound pre-treatment on the amount of dissolved organic matter extracted from food waste. *Bioresource Technology* 155: 266-271.
- Jin, Q., Zhang, H., Yan, L., Qu, L., and Huang, H. (2011). Kinetic characterisation for hemicellulose hydrolysis of corn stover in a dilute acid cycle spray. *Biomass and Bioenergy* 35(10): 4158-4164.
- Karmee, S.K., and Lin, C.S.K. (2014). Valorisation of food waste to biofuel: current trends and technological challenges. *Sustainable Chemical Processes* 2:22
- Khawla, B.J., Sameh, M., Imen, G., Donyes, F., Dhouha, G., Raoudha, E.G., and Oumema, N.E. (2014). Potato peel as feedstock for bioethanol production: A comparison of acid and enzymatic hydrolysis. *Industrial Crops and Products* 52: 144-149.
- Khraisheh, M., and Li, A. (2010). Bioethanol from municipal solid waste: The environmental impact assessment. *Proceeding of the 2nd Annual Gas Processing Symposium Qatar*, January 10-14 pp: 69-76.
- Kim, J.H., Lee, J.C., and Pak, D. (2011). Feasibility of producing ethanol from food waste. *Waste Management* 31: 2121-2125.

- Kim, S.M., Dien, B.S., and Singh, V. (2016). Promise of combine hydrothermal / chemical and mechanical refining for pretreatment of woody and herbaceous biomass. *Biotechnology for Biofuel* 9:97.
- Kim, T.H. (2013). Pretreatment of lignocellulosic biomass. In *Bioprocess technologies in biorefinery for sustainable production of fuels, chemicals, and polymers*. ed. Yang S.T., El-Enshasy H.A., and Thongchul N. pp 91-104. United States of America: John Wiley & Sons, Inc.
- Kiran, E.U., Trzcinski, A.P., and Liu, Y. (2014). Glucoamylase production from food waste by solid state fermentation and its evaluation in the hydrolysis of domestic food waste. *Biofuel Research Journal* 3:98-105.
- Kiran, E.U., Trzcinski, A.P., Ng, W.J., and Liu, Y. (2014). Bioconversion of food waste to energy: A review. *Fuel* 134: 389-399.
- Koike, Y., An, M.Z., Tang, Y.Q., Syo, T., Osaka, N., Morimura, S., and Kida, K. (2009). Production of fuel ethanol and methane from garbage by high-efficiency two-stage fermentation process. *Journal of Bioscience and Bioengineering* 108(6): 508-512.
- Kovacs, E., Wirth, R., Maroti, G., Bagi, Z., Nagy, K., Minarovits, J., Rakhely, G., and Kovacs, K.L. (2015). Augmented biogas production from protein-rich substrates and associated metagenomic changes. *Bioresource Technology* 178: 254-261.
- Kuhad, R.C., Gupta, R., Khasa, Y.P., and Singh, A. (2010). Bioethanol production from *Lantana camara* (red sage): Pretreatment, saccharification, and fermentation. *Bioresource Technology* 101: 8348-8354.
- Kumar, M., and Turner, S. (2015). Plant cellulose synthesis: CESA proteins crossing kingdoms. *Phytochemistry* 112: 91-99.
- Kumar, S., Singh, S.P., Mishra, I.M., and Adhikari, D.K. (2009). Recent advances in production of bioethanol from lignocellulosic biomass. *Chemical and Engineering Technology* 32(4): 517-526.
- Kunamneni, A., and Singh, S. (2005). Response surface optimization of enzymatic hydrolysis of maize starch for higher glucose production. *Biochemical Engineering Journal* 27: 179-190.
- Kuo, W.C., and Cheng, K.Y. (2007). Use of respirometer in evaluation of process and toxicity of thermophilic anaerobic digestion for treating kitchen waste. *Bioresource Technology* 98:1805-1811.

- Kuo, W.C., and Lai, W.L. (2010). Treatment of kitchen waste using mobile thermophilic anaerobic digestion system. *Renewable Energy* 35 (10): 2335-2339.
- Lal, B., and Sarma, P.M. (2011). *Wealth from waste: Trends and technologies*. The Energy and Resource Institute (TERI Press). Third Ed pp 60-61.
- Latifah, A.M., Abu-Samah, M.A., and Mohd-Zukki, N.I. (2009). Municipal solid waste management in Malaysia: Practices and challenges. *Waste Management* 29: 2902-2906.
- Lee, W.S., Chen, I.C., Chang, C.H., and Yang, S.S. (2012). Bioethanol production from sweet potato by co-immobilization of saccharolytic molds and *Saccharomyces cerevisiae*. *Renewable Energy* 39: 216-222.
- Li, A., Anxzar-Ladislaio, B., and Khraisheh, M. (2007). Bioconversion of municipal solid waste to glucose for bioethanol production. *Bioprocess Biosystem Engineering* 30: 189-196.
- Li, S., Zhang, X., and Andresen, J.M. (2012). Production of fermentable sugars from enzymatic hydrolysis of pretreated municipal solid waste after autoclave process. *Fuel* 92: 84-88.
- Li, Y., and Jin, Y. (2015). Effects of thermal pretreatment on acidification phase during two-phase batch anaerobic digestion of kitchen waste. *Renewable Energy* 77: 550-557.
- Limayem, A., and Ricke, S.C. (2012). Lignocellulosic biomass for bioethanol production: Current perspectives, potential issues and future prospects. *Progress in Energy and Combustion Science* 38:449-467.
- Lin, R., Cheng, J., Ding, L., Song, W., Qi, F., Zhou, J., and Cen, K. (2015). Subcritical water hydrolysis of rice straw for reducing sugar production with focus on degradation by-products and kinetic analysis. *Bioresource Technology* 186: 8-14.
- Lin, Y., Zhang, W., Li, C., Sakakibara, K., Tanaka, S., and Kong, H. (2012). Factors affecting ethanol fermentation using *Saccharomyces cerevisiae* BY4742. *Biomass and Bioenergy* 47: 395-401.
- Liu, H., Hu, H., Baktash, M.M., Jahan, M.S., Ahsan, L., and Ni, Y. (2014). Kinetics of furfural production from pre-hydrolysis liquor (PHL) of a kraft-based hardwood dissolving pulp production process. *Biomass & Bioenergy* 66: 320-327.

- Liu, X., Wang, W., Gao, X., Zhou, Y., and Shen, R. (2012). Effect of thermal pretreatment on the physical and chemical properties of municipal biomass waste. *Waste Management* 32: 249-255.
- Ma, H., Wang, Q., Qian, D., Gong, L., and Zhang, W. (2009). The utilization of acid-tolerant bacteria on ethanol production from kitchen garbage. *Renewable Energy* 34: 1466-1470.
- Ma, H., Wang, Q., Zhang, W., Xu, W., and Zou, D. (2008). Optimization of the medium and process parameters for ethanol production from kitchen garbage by *Zymomonas mobilis*. *International Journal of Green Energy* 5: 480-490.
- Ma, J., Duong, T.H., Smiths, M., Verstraete, W., and Carballa, M. (2011). Enhanced biomethanation of kitchen waste by different pretreatment. *Bioresource Technology* 102: 592-599.
- Maache-Rezzoug, Z., Pierre, G., Nouviaire, A., Maugard, T., and Rezzoug, S.A. (2011). Optimizing thermomechanical pretreatment conditions to enhance enzymatic hydrolysis of wheat straw by response surface methodology. *Biomass and Bioenergy* 35: 3129-3138.
- Malaysian Innovation Agency (2013). *National Biomass Strategy 2020: New wealth creation for Malaysia's Biomass Industry*. Retrieved on 9 May 2015 at <https://biobs.jrc.ec.europa.eu/sites/default/files/generated/files/policy/Biomass%20Strategy%202013.pdf>
- Matsakas, L., Kekos, D., Loizidou, M., and Christakopoulos, P. (2014). Utilization of household food waste for the production of ethanol at high dry material content. *Biotechnology for Biofuels* 7:4.
- Melikoglu, M. (2012). Solid-state fermentation of wheat pieces by *Aspergillus oryzae*: Effects of microwave pretreatment on enzyme production in a biorefinery. *International Journal of Green Energy* 9: 529-539.
- Meor-Hussin, A.S., Collins, S.R.A., Merali, Z., Parker, M.L., Elliston, A., Wellner, N., and Waldron, K.W. (2013). Characterisation of lignocellulosic sugars from municipal solid waste residue. *Biomass and Bioenergy* 51: 17-25.
- Miller, G.L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical Chemistry* 31: 426-428.
- Modenbach, A.A., and Nokes, S.E. (2013). Enzymatic hydrolysis of biomass at high-solids loadings - A review. *Biomass & Bioenergy* 56: 526-544.

- Modig, T., Liden, G., and Taherzadeh, M.J. (2002). Inhibition effects of furfural on alcohol dehydrogenase, aldehyde dehydrogenase and pyruvate dehydrogenase. *Biochemical Journal* 363: 769-776.
- Moh, Y.C., and Abd-Manaf, L. (2014). Overview of household solid waste recycling policy status and challenges in Malaysia. *Resources, Conservation and Recycling* 32: 50-61.
- Mokhtar, I.L. (2013, August 27). News Strait Times, "Need to act on rubbish now," <http://www.epu.gov.my/documents/10124/42a75b34-b727-4180-bc7b-6c5ffe24f1f1>
- Mokhtari-Hosseini, Z.B., Vasheghani-Farahani, E., Heidarzadeh-Vazifekhoran, A., Shojaosadati, S.A., Karimzadeh, R., and Darani, K.K. (2009). Statistical media optimization for growth and PHB production from methanol by methylothropic bacterium. *Bioresource Technology* 100:2436-2443.
- Mood, S.H., Golfeshan, A.H., Tabatabaei, M., Jouzani, G.S., Najafi, G.H., Gholami, M., and Ardjmand, M. (2013). Lignocellulosic biomass to bioethanol, a comprehensive review with a focus on pretreatment. *Renewable and Sustainable Energy Reviews* 27: 77-93.
- Moon, H.C., and Song, I.S. (2011). Enzymatic hydrolysis of food waste and methane production using UASB bioreactor. *International Journal of Green Energy* 8: 361-371.
- Moon, H.C., Song, S.I., Kim, J.C., Shirai, Y., Lee, D.H., Kim, J.K., Chung, S.O., Kim, D.H., Oh, K.K., and Cho, Y.S. (2009). Enzymatic hydrolysis of food waste and ethanol fermentation. *International Journal of Energy Research* 33: 164-172.
- Munda, U.S., Pholane, L., Kar, D.D., and Meikap, B.C. (2012). Production of bioenergy from composite waste materials made of corn waste, spent tea waste and kitchen waste co-mixed with cow dung. *International Journal of Green Energy* 9: 361-375.
- Mussatto, S.I., Dragone, G., Guimaraes, P.M.R., Silva, J.P.A., Carneiro, L.M., Roberto, I.C., Vicente, A., Domingues, L., and Teixeira, J.A. (2010). Technological trends, global market, and challenges of bioethanol production. *Bioethanol Advances* 28: 817-830.
- Natarajan, S.D., Mohamad, R., Rahim, R.A., and Nor'Aini, A.R. (2012). Potential of bioethanol production from *Nypa fruticans* sap by a newly isolated yeast *Lanchancea fermentati*. *Journal of Renewable and Sustainable Energy* 4:1-9.

- Nguyen, C.M., Choi, G.J., Choi, Y.H., Jang, K.S., and Kim, J.C. (2013). D- and L-lactic acid production from fresh sweet potato through simultaneous saccharification and fermentation. *Biochemical Engineering Journal* 81: 40-46.
- Nigam, P.S., and Singh, A. (2011). Production of liquid biofuels from renewable resources. *Progress in Energy and Combustion Science* 37: 52-68.
- Niu, Y., Zheng, D., Yao, B., Cai, Z., Zhao, Z., Wu, S., Cong, P., and Yang, D. (2017). A novel bioconversion for value-added products from food waste using *Musca domestica*. *Waste Management* 61: 455-460.
- Ohkouchi, Y., and Inoue, Y. (2007). Impact of chemical components of organic wastes on L(+)- lactic acid production. *Bioresource Technology* 98: 540-553.
- Omar, F.N., Nor' Aini, A.R., Hafid, H.S., Phang, L.Y., and Hassan, M.A. (2009). Separation and recovery of organic acids from fermented kitchen waste by an integrated process. *African Journal of Biotechnology* 8(21): 5807-5813.
- Owarnah, H.I., Dahunsi, S.O., Oranusi, U.S., and Alfa, M.I. (2014). Fertilizer and sanitary quality of digestate biofertilizer from the co-digestion of food waste and human excreta. *Waste Management* 34(4): 747-752.
- Parisutham, V., Kim, T.H., and Lee, S.K. (2014). Feasibility of consolidated bioprocessing microbes: From pretreatment to biofuel production. *Bioresource Technology* 161: 431-440.
- Peng, L., and Chen, Y. (2011). Conversion of paper sludge to ethanol by separate hydrolysis and fermentation (SHF) using *Saccharomyces cerevisiae*. *Biomass and Bioenergy* 35: 1600-1606.
- Pham, T.P.T., Kaushik, R., Parshetti, G.K., Mahmood, R., and Balasubramaniam, R. (2014). Food-waste-to-energy conversion technologies: Current status and future directions. *Waste Management* 38: 399-408.
- Pleissner, D., and Lin, C.S.K. (2013). Valorisation of food waste in biotechnological processes. A review. *Sustainable Chemical Processes* 1:21.
- Qiao, W., Yan, X., Ye, J., Sun, Y., Wang, W., and Zhang, Z. (2011). Evaluation of biogas production from different biomass wastes with/without hydrothermal pretreatment. *Renewable Energy* 36 (12): 3313-3318.
- Qin, L., Liu, Z.H., Li, B.Z., Dale, B.E., and Yuan, Y.J. (2012). Mass balance and transformation of corn stover by pretreatment with different dilute organic acids. *Bioresource Technology* 112: 319-326.
- Raghavi, S., Sindhu, R., Binod, P., Gnansnonou, E., and Pandey, A. (2016). Development of a novel sequential pretreatment strategy for the

production of bioethanol from sugarcane trash. *Bioresource Technology* 199: 202-210.

Roy, P., Tokuyasu, K., Orikasa, T., Nakamura, N., and Shiina, T. (2012). A techno-economic and environmental evaluation of the life cycle of bioethanol produced from rice straw by RT-CaCCO process. *Biomass and Bioenergy* 27: 188-195.

Ruangmee, A., and Sangwichien, C. (2013). Response surface optimization of enzymatic hydrolysis of narrow-leaf cattail for bioethanol production. *Energy Conversion and Management* 73:381-388.

Rueda, C, Fernandez-Rodriguez, J., Ruiz, G., Llano, T., and Coz, A. (2015). Monosaccharide production in an acid sulphite process: Kinetic modelling. *Carbohydrate Polymer* 116: 18-25.

Saeed, M.O., Hassan, M.N., and Mujeebu, M.A. (2009). Assessment of municipal solid waste generation and recyclable materials potential in Kuala Lumpur, Malaysia. *Waste Management* 29(7): 2209-2213.

Saha, B.C., Nichols, N.N., and Cotta, M.A. (2011). Ethanol production from wheat straw by recombinant *Escherichia coli* strain FBR5 at high solid loading. *Bioresource Technology* 102(23): 10892-10897.

Sarkar, N., Ghosh, S.K., Bannerjee, S., and Aikat, K. (2012). Bioethanol production from agricultural wastes: An overview. *Renewable Energy* 37: 19-27.

Satyarthi, J.K., Srinivas, D., and Ratnasamy, P. (2011). Hydrolysis of vegetable oils and fats to fatty acids over solid acid catalysts. *Applied Catalysis A: General* 391: 427-435.

Shafie, M., Karimi, K., and Taherzadeh, M.J. (2011). Techno-economical study of ethanol and biogas from spruce wood by NMMO-pretreatment and rapid fermentation and digestion. *Bioresource Technology* 102: 7879-7866.

Shana, A.D., Ouki, S., Asaadi, M., and Pearce, P. (2012). Influence of an intermediate thermal hydrolysis process (ITHP) on a kinetics of anaerobic digestion of sewage sludge. *Chemical Engineering Transactions* 29: 1267-1272.

Shen, F., Yuan, H., Pang, Y., Chen, S., Zhu, B., Zou, D., Liu, Y., Ma, J., Yu, L., and Li, X. (2013). Performance of anaerobic co-digestion of fruit & vegetable waste (FVW) and food waste (FW): Single-phase vs. two-phase. *Bioresource Technology* 144: 80-85.

- Shen, J., and Wyman, C.E. (2011). A novel mechanism and kinetic model to explain enhanced xylose yields from dilute sulfuric acid compared to hydrothermal pretreatment of corn stover. *Bioresource Technology* 102(19): 9111-9120.
- Shu-Hsein, T., Ching-Piao, L., and Shang-Shyng, Y. (2007). Microbial conversion of food wastes for biofertilizer production with thermophilic lipolytic microbes. *Renewable Energy* 32(6): 904-915.
- Sindhu, R., Kuttiraja, M., Binod, P., Janu, K.U., Sukumaran, R.K., and Pandey, A. (2011). Dilute acid pretreatment and enzymatic saccharification of sugarcane tops for bioethanol production. *Bioresource Technology* 102: 10915-10921.
- Singh, A., and Bishnoi, N.R. (2013). Ethanol production from pretreated wheat straw hydrolysate by *Saccharomyces cerevisiae* via sequential statistical optimization. *Industrial Crops and Products* 41: 221-226.
- Singh, A., and Bishnoi, N. (2012). Optimization of ethanol production from microwave alkali pretreated rice straw using statistical experimental designs by *Saccharomyces cerevisiae*. *Industrial Crops and Products* 37: 334-341.
- Su, W., Yu, M., Ma, H., Gao, M., and Wang, Q. (2015). Research on the recycling of distillation waste in ethanol fermentation from food waste and its influence. *International Journal of Green Energy* 12: 737-742.
- Tan, I.S., and Lee, K.T. (2014). Enzymatic hydrolysis and fermentation of seaweed solid wastes for bioethanol production: An optimization study. *Energy* 78: 53-62.
- Tang, Y.Q., Koike, Y., Liu, K., An, M.Z., Morimura, S., Wu, X.L., and Kida, K. (2008). Ethanol production from kitchen waste using the flocculating yeast *Saccharomyces cerevisiae* strain KF-7. *Biomass and Bioenergy* 32: 1037-1045.
- Tembhurkar, A.R., and Mhaisalkar, V.A. (2007). Studies on hydrolysis and acidogenesis of kitchen waste in two-phase anaerobic digestion. *Journal of IPHE* 8(2): 10-17.
- Thi, N.B.D., Kumar, G., and Lin, C.Y. (2015). An overview of food waste management in developing countries: Current status and future perspective. *Journal of Environmental Management* 157: 220-229.
- Thongdumy, P., Intrasungkha, N., and O-Thong, S. (2014). Optimization of ethanol production from food waste hydrolysate by co-culture of *Zymomonas mobilis* and *Candida shehatae* under non-sterile condition. *African Journal of Biotechnology* 13(7): 866-873.

- Tian, H., Duan, N., Lin, C., Li, X., and Zhong, M. (2015). Anaerobic co-digestion of kitchen waste and pig manure with different mixing ratios. *Journal of Bioscience and Bioengineering* 120(1): 51-57.
- Uncu, O.N., and Cekmecelioglu, D. (2011). Cost-effective approach to ethanol production and optimization by response surface methodology. *Waste Management* 31: 636-643.
- Uthumporn, U., Karim, A.A., and Fazilah, A. (2013). Defatting improves the hydrolysis of granular starch using a mixture of fungal amylolytic enzyme. *Industrial Crops and Products* 43: 441-449.
- Vani, S., Sukumaran, R.K., and Savithri, S. (2015). Prediction of sugar yields during hydrolysis of lignocellulosic biomass using artificial neural network modeling. *Bioresource Technology* 188: 128-135.
- Vavouraki, A.I., Angelis, E.M., and Kornaros, M. (2012). Optimization of thermo-chemical hydrolysis of kitchen wastes. *Waste Management* 3: 740-745.
- Vavouraki, A.I., Valiotti, V., and Kornaros, M.E. (2014). Optimization of thermo-chemical pretreatment and enzymatic hydrolysis of kitchen waste. *Waste Management* 34: 167-173.
- Vincent, M., Norrezi, N.A.M., Bujang, K., and Adeni, D.S.A. (2014). The effect of substrate concentration on enzymatic hydrolysis of selected food waste for glucose production. *Bioremediation Science and Technology Research* 2:1-4.
- Vohra, M., Manwar, J., Manmode, R., Padgilwar, S., and Patil, S. (2014). Bioethanol production: Feedstock and current technologies. *Journal of Environmental Chemical Engineering* 2(1): 573-584.
- Vucurovic, V.M., Razmovski, R.N., Miljic, U.D., Puskas, V.S., Acanski, M.M., and Pastor, K.A. (2014). Thermo-acid pretreatment of starch based kitchen waste for ethanol production. *Analecta* 1: 85-90.
- Wan, C., and Li, Y. (2011). Effect of hot water extraction and liquid hot water pretreatment on the fungal degradation of biomass feedstocks. *Bioresource Technology* 102:9788-9793.
- Wan Razali, W.A. (2014). Composting evaluation of oil palm empty fruit bunches with palm oil effluent anaerobic sludge. Master Thesis. Universiti Putra Malaysia.
- Wang, L., Shen, F., Yuan, H., Zou, D., Liu, Y., Zhu, B., and Li, X. (2014). Anaerobic co-digestion of kitchen waste and fruit/vegetable waste: Lab-scale and pilot-scale studies. *Waste Management* 34(12): 2627-2633.

- Wang, Q., Ma, H., Xu, W., Gong, L., Zhang, W., and Zou, D. (2008). Ethanol production from kitchen garbage using response surface methodology. *Biochemical Engineering Journal* 39: 604-610.
- Wang, Q., Wang, X., Wang, X., and Ma, H. (2008). Glucoamylase production from food waste by *Aspergillus niger* under submerged fermentation. *Process Biochemistry* 43: 280-286.
- Warner, K. (2008). *Chemistry of frying oil. In Food Lipids: Chemistry, nutrition, and biotechnology. Third Edition.* Ed. Akoh C.C., Min B.D. pp: 189-195 CRC Press Taylor & Francis Group, United States of America.
- Wei, W., Wu, S., and Liu, L. (2012). Enzymatic saccharification of dilute acid pretreated eucalyptus chips for fermentable sugar production. *Bioresource Technology* 110: 302-307.
- Wikandari, R., Millati, R., Syamsiyah, S., Muriana, R., and Ayuningsih, Y. (2010). Effect of furfural, hydroxymethylfurfural and acetic acid on indigeneous microbial isolate for bioethanol production. *Agricultural Journal* 5(2): 105-109.
- Woon, K.S., and Lo, I.M.C. (2016). A proposed framework of food waste collection and recycling for renewable biogas fuel production in Hong Kong. *Waste Management* 47: 3-10.
- Xiao, L., Deng, Z., Fung, K.Y., and Ng, K.M. (2013). Biohydrogen generation from anaerobic digestion of food waste. *International Journal of Hydrogen Energy* 38(32): 13907-13913.
- Xie, S., Lawlor, P.G., Frost, J.P., Wu, G., and Zhan, X. (2012). Hydrolysis and acidification of grass silage in leaching bed reactors. *Bioresource Technology* 114: 406-413.
- Xu, Q., Singh, A., and Himmel, M.E. (2009). Perspectives and new directions for the production of bioethanol using consolidated bioprocessing of lignocellulose. *Current Opinion in Biotechnology* 20: 364-371.
- Xu, Z., Wang, Q., Jiang, Z. H. Yang, X., and Ji, Y. (2007). Enzymatic hydrolysis of pretreated soybean straw. *Biomass and Bioenergy* 31: 162-167.
- Yan, S., Chen, X., Wu, J., and Wang, P. (2013). Pilot-scale production of fuel ethanol from concentrated food waste hydrolysates using *Saccharomyces cerevisiae* H058. *Bioprocess Biosystem Engineering* 36: 937-946.
- Yan, S., Li, J., Chen, X., Wu, J., Wang, P., Ye, J., and Yao, J. (2011). Enzymatic hydrolysis of food waste and ethanol production from the hydrolysate. *Renewable Energy* 36: 1259-1265.

- Yan, S., Wang, P., Zhai, Z., and Yao, J. (2010). Fuel ethanol production from concentrated food waste hydrolysates in immobilized cell reactors by *Saccharomyces cerevisiae* H058. *Journal of Chemical Technology and Biotechnology* 86: 731-738.
- Yan, S., Yao, J., Yao, L., Zhi, Z., Chen, X., and Wu, J. (2012). Fed-batch enzymatic saccharification of food waste improves the sugar concentration in the hydrolysates and eventually the ethanol fermentation by *Saccharomyces cerevisiae* H058. *Brazilian Archives of Biology and Technology* 55(2).
- Yang, F., Zhang, Q., Fan, H.X., Li, Y., and Li, G. (2013). Electrochemical control of the conversion of cellulose oligosaccharides into glucose. *Journal of Industrial and Engineering Chemistry* 20: 3487-3492.
- Yasin, N.H.M., Mumtaz, T., Hassan, M.A., and Nor' Aini, A.R (2013). Food waste and food processing waste for biohydrogen production. *Journal of Environmental Management* 130: 375-385.
- Yu, Q., Zhuang, X., Lv, S., He, M., Zhang, Y., Yuan, Z., Qi, W., Wang, Q., Wang, W., and Tan, X. (2013). Liquid hot water pretreatment of sugarcane baggase and its comparison with chemical pretreatment methods for the sugar recovery and structural changes. *Bioresource Technology* 129: 592-598.
- Zakaria, M.R., Hirata, S., and Hassan, M.A. (2014). Combined pretreatment using alkaline hydrothermal and ball milling to enhance enzymatic hydrolysis of oil palm mesocarp fiber. *Bioresource Technology* 169: 236-243.
- Zhai, N., Zhang, T., Yin, D., Yang, G., Wang, X., Ren, G., and Feng, Y. (2015). Effect of initial pH on anaerobic co-digestion of kitchen waste and cow manure. *Waste Management* 38: 126-131.
- Zhang, B., Wei-min, C., and He, P. (2007). Influence of lactic acid on the two-phase anaerobic digestion of kitchen wastes. *Journal of Environmental Sciences* 19(2): 244-249.
- Zhang, L., Lee, Y.W., and Jahng, D. (2011). Anaerobic co-digestion of food waste and piggery wastewater: Focusing on the role of trace elements. *Bioresource Technology* 102(8): 5048-5059.
- Zhang, W., Zhang, L., and Li, A. (2015). Anaerobic co-digestion of food waste with MSW incineration plant fresh leachate: process performance and synergistic effects. *Chemical Engineering Journal* 259:795-805.
- Zhang, W., Ma, H., Wang, Q., Zhao, F., and Xiao, Z. (2012). Pretreatment technology for suspended solids and oil removal in an ethanol fermentation broth from food waste separated by pervaporation process. *Desalination* 293: 112-117.

- Zhao, X., Morikawa, Y., Qi, F., Zeng, J., and Liu, D. (2014). A novel kinetic model for polysaccharide dissolution during atmospheric acetic acid pretreatment of sugarcane bagasse. *Bioresource Technology* 151: 128-136.
- Zhao, X., Zhou, Y., and Liu, D. (2012). Kinetic model for glycan hydrolysis and formation of monosaccharides during acid hydrolysis of sugarcane bagasse. *Bioresource Technology* 105: 160-168.
- Zhou, S., and Runge, T.M. (2014). Validation of lignocellulosic biomass carbohydrates determination via acid hydrolysis. *Carbohydrate Polymers* 112:179-185.





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