



**UNIVERSITI PUTRA MALAYSIA**

***BIOFUNCTIONALIZED TAPERED MULTIMODE FIBER FOR DENGUE  
VIRUS ANTIBODY DETECTION***

**MOHD AZMIR BIN MUSTAPA**

**FK 2017 98**



**BIOFUNCTIONALIZED TAPERED MULTIMODE FIBER FOR  
DENGUE VIRUS ANTIBODY DETECTION**

By

**MOHD AZMIR BIN MUSTAPA**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfillment of the Requirements for the Degree of Master of  
Science**

**June 2017**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

**BIOFUNCTIONALIZED TAPERED MULTIMODE FIBER FOR DENGUE VIRUS ANTIBODY DETECTION**

By

**MOHD AZMIR BIN MUSTAPA**

**June 2017**

**Chair: Muhammad Hafiz Bin Abu Bakar, PhD**  
**Faculty: Engineering**

The use of optical tapered fibers provides good sensitivity for the detection and measurement of biological and chemical compound especially biomolecules such as proteins, nucleic acids, antibodies, lipids, and carbohydrates. This thesis looks into the fabrication of a biofunctionalized tapered multimode fiber (TMMF) as a probe for anti-Dengue virus NS1 immunoglobulin G (anti-DENV NS1 IgG) detection. This device offers advantages by improving sensitivity, rapidness of detection, the size of device by making it more compact, and the field operability. In this research, TMMF was fabricated using a Vytran GPX-3000 glass processing system. A multimode fiber (MMF) with diameter of 125  $\mu\text{m}$  is tapered until the size reduced to 10  $\mu\text{m}$  with 10 mm of its waist length and transition length. The implemented technique is proved to induce some amount of light propagating outside the core of the tapered fiber and generates evanescent wave field (EWF) allowing high sensitivity to the changes in surrounding condition. The surface modification process onto TMMF involving three stages, namely silanization, activation with glutaraldehyde (GA), and immobilization of DENV NS1 glycoprotein on the surface of TMMF. This process is required to allow the interaction between inorganic silica TMMF and organic biomolecules. The silanated-TMMF treated with 3-amino-propyltriethoxysilane (APTES) and coated with 100  $\mu\text{g}/\text{mL}$  of DENV NS1 glycoprotein specific to anti-DENV NS1 IgG antibodies in order to detect the presence of the antibodies. The generated EWF is applicable to measure the interaction between bound and unbound biomolecules on the surface by utilizing evanescent wave absorption (EWA) in the UV region. This biosensor device has the capability to detect the presence of anti-DENV NS1 IgG antibodies in a liquid sample in less than five minutes of incubation time. The concentration of the samples varies from 100  $\text{pg}/\text{mL}$  until 1000  $\text{pg}/\text{mL}$  with interval of 100  $\text{pg}/\text{mL}$ . Results showed the sensitivity of the biofunctionalized TMMF to be  $7 \times 10^{-6}$  a.u./ $\text{pg}/\text{mL}$  with R-squared of 0.9938. The experiment done in this study demonstrates the potential of the biofunctionalized-TMMF to be developed and commercialized as a rapid and label-free sensor for dengue virus detection in the future.

Abstrak tesis dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

## **GENTIAN OPTIK MULTIMOD BERFUNGSI-BIO UNTUK MENGESAN ANTIBODI VIRUS DENGGI**

Oleh

**MOHD AZMIR BIN MUSTAPA**

**Jun 2017**

**Pengerusi: Muhammad Hafiz Bin Abu Bakar, PhD**  
**Fakulti: Kejuruteraan**

Penggunaan kaedah gentian optik bertirus memberikan kelebihan terhadap peranti ini dari segi kepekaan yang tinggi dalam pengesanan dan pengukuran sebatian biologi dan kimia terutamanya biomolekul seperti protein, asid nukleik, antibodi, lipid, dan karbohidrat. Tesis ini memberikan perhatian terhadap penghasilan gentian tirus multimod (TMMF) berfungsi-bio sebagai pengesanan antibodi virus anti-denggi protein tidak berstruktur 1 immunoglobulin G (anti-DENV NS1 IgG). Peranti ini mempunyai kelebihan dari segi peningkatan kadar kepekaan peranti pengesanan, kepantasan mengesan substrat, menjadikan saiz peranti lebih padat, dan keupayaan untuk beroperasi di lapangan. Di dalam kajian ini, TMMF dihasilkan dengan menggunakan sistem pemprosesan kaca Vytran GPX-3000. Gentian multimod (MMF) yang mempunyai ukuran saiz diameternya 125  $\mu\text{m}$  telah ditiruskan sehingga 10 $\mu\text{m}$ , sementara panjang tirus dan panjang peralihan masing-masing adalah 10 mm. Kaedah yang digunapakai ini telah membuktikan bahawa beberapa jumlah cahaya akan terdorong untuk tersebar keluar dari teras gentian tirus dan mejana medan gelombang cepat berlalu (EWF) bagi menghasilkan kepekaan yang tinggi terhadap perubahan keadaan persekitarannya. Proses pengubahsuaian permukaan TMMF melibatkan tiga peringkat, iaitu menyalut permukaan gentian dengan bahan uji silana yang dikenali sebagai 3-aminopropiltriethoxysilana (APTES), pengaktifan dengan glutaraldehid (GA), dan pemegangan glikoprotein virus denggi protein tidak berstruktur 1 (DENV NS1) di atas permukaan tirus gentian tersebut. Proses ini adalah perlu bagi memastikan permukaan TMMF yang tidak organik saling bertindak dengan biomolekul berorganik. Seterusnya, permukaan TMMF yang telah dirawat dengan APTES dan GA telah disalut dengan 100  $\mu\text{g/mL}$  glikoprotein DENV NS1 spesifik kepada antibodi anti-DENV NS1 IgG untuk mengesan kehadiran antibodi tersebut. EWF yang terhasil dapat digunakan untuk mengukur kadar saling bertindak antara biomolekul yang terikat dan tidak terikat di atas permukaan dengan menggunakan penyerapan gelombang cepat berlalu (EWA) di kawasan ultraungu (UV). Peranti pengesanan-bio ini mempunyai keupayaan untuk mengesan kehadiran antibodi anti-DENV NS1 IgG di dalam sampel cecair kurang

daripada lima menit tempoh inkubasi. Kepekatan sampel yang digunakan adalah selang 100 pg/mL bermula daripada 100 pg/mL sehingga 1000 pg/mL. Data yang diperolehi daripada eksperimen ini telah menunjukkan kepekaan TMMF berfungsi-bio ini adalah  $7 \times 10^{-6}$  a.u./pg/mL dengan pekali penentuan ( $R^2$ ) adalah 0.9938. Uji kaji yang dijalankan dalam pembelajaran ini menunjukkan TMMF berfungsi-bio ini berpotensi untuk dibangunkan dan dikomersialkan pada masa hadapan sebagai peranti yang mampu mengesan virus denggi dengan pantas dan tanpa menggunakan bahan kimia penanda.



## ACKNOWLEDGEMENTS

Alhamdulillah, I am very grateful to Allah, this thesis has been carried out up to the final stage of my research. Secondly, I would like to convey my appreciation and sincerest thankfulness to my family, especially to my lovely wife (Amirah), my parents (Mustapa & Azizah), my mother in-law (Asmah Sani), my brothers (Brother Yoe and Lan), and my family in-law for their understanding and patience in the years of my MSc study. Without thrust, I might loss my passionate to make this work completed within the timeframe given. With the full of respect, I would like to thank you very much to my supervisor, Dr Muhammad Hafiz bin Abu Bakar for his priceless guidance, encouragement and all the opportunities he gave. Throughout the process, I have learnt not only about the research that I am working on but also the values that I have to grasp to keep on success in life. I would also like to express my gratitude to my co-supervisors, Professor Dr. Mohd Adzir bin Mahdi and Dr Amir Syahir Amir Hamzah for their guidance, support and knowledge throughout my study. Thanks also to other colleagues like Hadi, Shanan, Kak Maz, Noran, Kak G-jah, Kak Zura, and other that I do not mention for all the cheers and happiness that we share in the laboratory. A million thanks to my best friend Mirul, Rambo, Aud, Pak Tam and other that I do not mention here. With their kind and good assistance, the experiments are carried out successfully. Their help towards me is really much being appreciated. Last but not least, thanks to Allah for giving me and wife a cute baby boy named Muhammad Ammar who keep on inspiring me to never give up finishing my studies.

I certify that a Thesis Examination Committee has met on **15<sup>th</sup> June 2017** to conduct the final examination of **Mohd Azmir bin Mustapa** on his thesis entitled “**Biofunctionalized Tapered Multimode Fiber for Dengue Virus Antibody Detection**” in accordance with the Universities and University Colleges Acts 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

**Siti Barirah binti Ahmad Anas, PhD**

Associate Professor  
Faculty of Engineering  
Universiti Putra Malaysia  
(Chairman)

**Ahmad Shukri bin Muhammad Noor, PhD**

Associate Professor  
Faculty of Engineering  
Universiti Putra Malaysia  
(Internal Examiner)

**Hairul Azhar Abdul Rashid, PhD**

Professor  
Faculty of Engineering  
Multimedia University  
(External Examiner)

\_\_\_\_\_  
**(Insert name of current Deputy Dean)**

**(E.g. XXXXX XXXX, PhD)**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Muhammad Hafiz Abu Bakar, PhD**

Senior Lecturer  
Faculty of Engineering  
Universiti Putra Malaysia  
(Chairman)

**Amir Syahir Amir Hamzah, PhD**

Senior Lecturer  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Member)

**Mohd. Adzir Mahdi, PhD**

Professor  
Faculty of Engineering  
Universiti Putra Malaysia  
(Member)

**(ROBIAH BINTI YUNUS, PhD)**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

### Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Mohd Azmir Bin Mustapa / GS 36999

### **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_  
Name of Chairman of  
Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory  
Committee: \_\_\_\_\_

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	ii
<b>ACKNOWLEDGEMENTS</b>	iv
<b>APPROVAL</b>	v
<b>DECLARATION</b>	vii
<b>LIST OF TABLES</b>	x
<b>LIST OF FIGURES</b>	xii
<b>LIST OF ABBREVIATIONS</b>	xiii
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
1.1. An overview of Dengue Virus Fever	1
1.2. Problem Statement	3
1.3. Motivation	3
1.4. Research objectives	3
1.5. Scope of research	3
1.6. Thesis Organization	4
<b>2 THEORY AND LITERATURE REVIEW</b>	<b>5</b>
2.1 Introduction	5
2.2 Introduction to Dengue Virus	5
2.2.1 Dengue life cycle and symptoms	5
2.3 Principle of an antibody-antigen interaction	6
2.4 Design and applications of optical fiber Biosensor	8
2.5 Theoretical study of evanescent wave absorbance (EWA) based TMMF	10
2.6 Tapered multimode fiber-based sensor	11
2.7 Immobilization of bioreceptor on the surface of silica fiber	13
2.8 Anti-dengue virus immunoglobulin G (IgG) antibody detection	14
2.9 Summary	17
<b>3 TMMF FOR ANTI-DENV NS1 IgG ANTIBODY DETECTION</b>	<b>18</b>
3.1 Introduction	18
3.2 Research methodology	19
3.3 Fabrication of TMMF	20
3.4 Optimization of TMMF	23

3.5	Immobilization of DENV NS1 glycoprotein on the TMMF surface	26
3.5.1	Silanization of the TMMF with APTES	27
3.5.2	Activation of silaned-TMMF with Glutaraldehyde	29
3.5.3	Coating the DENV NS1 glycoprotein on the TMMF surface	32
3.6	Detection of anti-DENV NS1 IgG antibody	35
3.6.1	The sensitivity of the anti-DENV NS1 IgG antibodies detection	36
3.6.2	Dynamic response time of anti-DENV NS1 IgG antibodies detection	38
3.7	Summary	40
<b>4</b>	<b>CONCLUSION AND FUTURE WORK</b>	<b>41</b>
4.1	Conclusion	41
4.2	Recommendations for Future Work	42
	<b>REFERENCES</b>	<b>43</b>
	<b>BIODATA OF STUDENT</b>	<b>51</b>
	<b>LIST OF PUBLICATIONS</b>	<b>52</b>

## LIST OF TABLES

Table		Page
2.1	The summary previous study used to detect the DENV	23
3.1	The parameter of taper fibers with different waist diameter	30
3.2	Properties of the APTES	36
3.3	Properties of the glutaraldehyde	39



## LIST OF FIGURES

Figure		Page
1.1	Genome structure of DENV.	1
1.2	Statistic on the number of DENV cases in Malaysia over the past 15 years.	2
2.1	The relationship of DENV with NS1 glycoprotein, IgM, and IgG at a different stage of the infection.	8
2.2	A basic structure of immunoglobulin (Ig) and antigen binding site.	9
2.3	The geometry of optical fiber used in sensing applications.	12
2.4	The geometry of fabricated tapered fiber.	17
2.5	The light absorption spectra based on the different waist diameters and constant 10 mm taper waist length for all the tapered fiber.	18
3.1	Flow chart of research methodology used to develop optical fiber biosensor.	19
3.2	Vytran GPX-3000 glass processing unit.	21
3.3	Dimension of taper parameter that can be set on the taper properties program according to the require parameter.	21
3.4	Power needed for tapering process where it can be set in the Filament Start (W) box and being pulled at certain velocity (mm/s).	29
3.5	Tapered fiber image observed under the microscope	30
3.6	The experimental setup used to detect the DENV NS1 IgG antibody.	31
3.7	Design of acrylate TMMF platform used for TMMF placement.	32
3.8	Transmission intensity after the TMMF was exposed to UV light for eight hours.	33
3.9	Sensitivity graphs for different taper waist diameter size.	34
3.10	Molecular structure of the APTES with the primary amino group at one end.	35

3.11	Variation of spectra response based on the different incubation time for APTES solution.	37
3.12	Relationship of spectrum absorption of APTES at 510 nm with incubation time as the absorbance is increased when incubation time is longer.	38
3.13	Molecular structure of GA molecules.	39
3.14	Absorption spectra after treated with the GA.	40
3.15	Relationship of the incubation time with the absorption spectrum.	41
3.16	The SEM image of TMMF observed under Hitachi SU510 SEM.	42
3.17	Absorption spectrum of the DENV NS1 glycoprotein during the immobilization and indicate the maximum absorption at 1120 nm wavelength.	44
3.18	Relationship between the absorbance and incubation time for the DENV NS1 glycoprotein.	44
3.19	The FESEM image of TMMF without coating, coated with APTES+GA, cross-sectional image of the tapered fiber with immobilized antigenic proteins, and with higher magnification.	45
3.20	Absorption spectrum of various concentration antibodies detection.	47
3.21	Relationship of absorbance versus time in which various sort of concentration start from 100 pg/mL until 1000 pg/mL of anti-DENV NS1 IgG antibody as a constant while time is a variable.	49
3.22	Dynamic response time of the antibodies detection with the aim of to examine the rapidness of antibodies detection.	51



## LIST OF ABBREVIATIONS

APTES	3-aminopropyltriethoxysilane
AuNPs	Gold nanoparticles
bp	Base pair
BPC	Blood platelet count
BSA	Bovine serum albumin
C=N	Imine bond
C-domain	Constant domain
CS	Chitosan
DENV	Dengue virus
dH <sub>2</sub> O	Distilled water
DHF	Dengue hemorrhagic fever
DNA	Deoxyribonucleic acid
DSS	Dengue shock syndrome
DVF	Dengue virus fever
ELISA	Enzyme-linked immunosorbent assay
EWA	Evanescent wave absorbance
EWf	Evanescent wave field
Fab	Fragment antigen binding
FBG	Fiber Bragg grating
Fc	Fragment crystallizable
FESEM	Field emission scanning electron microscope
FTIC	Fluorescein Isothiocyanate
GA	Glutaraldehyde
GUI	Graphic user interface
H-chain	Heavy chain
HF	Hydrofluoric acid
HRP	Horse reddish peroxidase
Ig	Immunoglobulin
IgG	Immunoglobulin G
IgM	Immunoglobulin M
L-chain	Light chain
LPFG	Long-period fiber grating
LSPR	Localized surface plasmon resonance
M	Molar
MAC-ELISA	M antibody-capture enzyme-linked immunosorbent
MMF	Multimode fiber
MSIPs	Micro-spot integrated pillars
Ni <sup>2+</sup>	Nickel (II)
NPs	Nanoparticles
NS	non-structural protein

OFIS	Optical fiber immunosensor
-OH	Hydroxyl
PAA	Poly acrylic acid
PBS	Phosphate buffer saline
PNA	Peptide nucleic acid
QCM	Quartz crystal microbalance
RI	Refractive index
RNA	Ribosomal nucleic acid
RT-PCR	Reverse transcription polymerase chain reaction
RT-RPA	Reverse transcription-recombinase polymerase amplification
RT-RPA	Reverse transcription-recombinase polymerase amplification
SEM	Scanning electron microscope
SiNW	Silicon nanowire
SiNx	Silicon nitride
Si-O-Si	Siloxane
SMF	Single mode fiber
SPR	Surface plasmon resonance
S-S Bond	Disulfide bond
ssRNA	Single stranded ribosomal nucleic acid
TF	Tapered fiber
TIR	Total internal reflection
TMMF	Tapered multimode fiber
UV	Ultraviolet
V-domain	Variable domain

© COPYRIGHT UPM

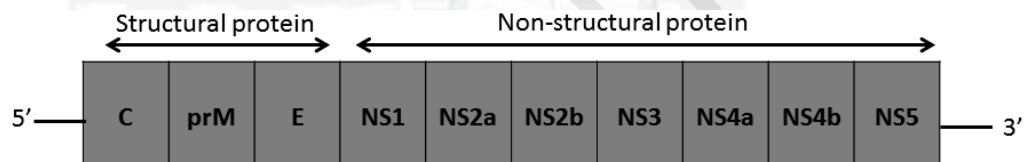


## CHAPTER 1

### INTRODUCTION

#### 1.1 An overview of DENV fever

Dengue fever is a viral disease transmitted to human through the bite of the female *Aedes aegypti* mosquito [1-5]. The virus is a single stranded ribosomal nucleic acid (ssRNA) and it is responsible in producing dengue virus (DENV) in a host cell to cause the disease [6-8]. It consists of 11000 of nucleotide base and code for ten different categories of proteins (Figure 1.1) which are three structural proteins known as Capsid protein (C), preMembrane protein (prM), and Envelop glycoprotein (E) while the other seven are non-structural proteins which are consist of NS1, NS2a, NS2b, NS3, NS4a, NS4b, and NS5 [3,9-11].



**Figure 1.1: Genome structure of DENV.**

The lipid proteins encapsulated the virus as an envelope protein. It is responsible to allow the virus attachment on the target host cell surface and subsequent replication [12]. The structure protein that surrounds the virus will help the virus to introduce their RNAs to the host cell and then the non-structural (NS) protein will play a role in producing new virus in the host cell [10,12].

Since the dengue fever shows typical symptoms such as fever, headache, muscle and joint pain thus, making it difficult to determine whether the patient has been specifically infected with dengue virus [13-15]. Example of other diseases showing similar symptoms to the DENV are normal fever, leptospirosis, and chikungunya. The latest is Zika virus, which transmitted to the human through the bite of similar type of DENV mosquitoes [16-19].

In Malaysia, dengue fever has become a very serious issue in recent years with rising number of cases [20]. Since year 2010 until 2015, the number of cases continues to increase annually with 2015 recording the highest number of cases (Figure 1.2) [20]. The DENV serotypes NS1 and NS2 are the most reported serotypes infecting human among the five strains [21]. The existence of multiple strains increases the possibility of secondary infection, which significantly alleviates the risk of contracting the potentially fatal dengue hemorrhagic fever (DHF) and dengue shock syndrome (DSS) [22].

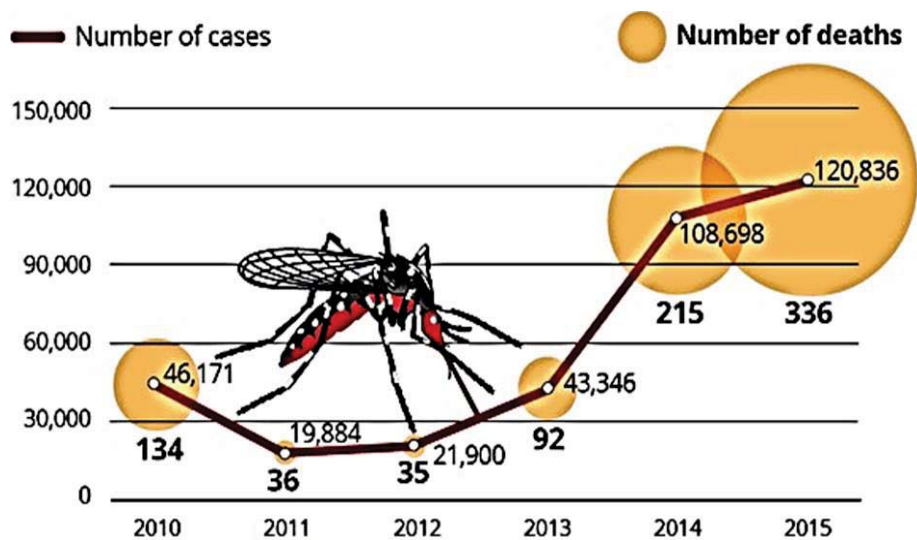


Figure 1.2: Statistic on the number of DENV cases in Malaysia over the past 15 years [23].

Therefore, early diagnostic and infection monitoring methods are crucial to prevent and control the outbreak of dengue virus. The detection of DENV IgG antibody is of high interest due to its abundance during secondary infection. Currently, the most popular technique used to detect the dengue virus is virological test and serological test. In clinical test activity, the virological test is used to detect the dengue virus infection at early stage of illness while serological test is used for further investigation to diagnose the acute phase of infection [8,24-26]. A common approach to detect Dengue virus infection in healthcare facilities is the use of rapid test kit, which utilizes the concept of immunoassay-based colorimetric on paper strips to indicate detection [27]. The most powerful technique used in clinical laboratory test for dengue detection is the enzyme-linked immunosorbent assay (ELISA), which has excellent performance in terms of specificity and sensitivity [27]. For monitoring purposes, blood platelet count (BPC) tests are performed periodically to ascertain the level of platelet in patients [5].

Nowadays, the use of optical fiber as biosensors has become increasingly attractive with multiple applications in the detection of biomolecules [28-30]. Both single mode fiber (SMF) and multimode fiber (MMF) have been explored for such purposes due to their many advantages such as high sensitivity, immunity to electromagnetic interference, and compact size [31]. MMF is of high interest as it offers added features such as coupling simplicity and good light confinement over wide wavelength range. Tapered multimode fiber (TMMF) in particular has proven itself to be a viable technology for sensing applications [32,33].

## 1.2 Problem statement

In recent years, NS1 strain has become an increasingly predominant serotype with more focus given on the detection of its IgG antibody. Although rapid test kit for dengue virus is normally sufficient to indicate infection, the approach can only produce qualitative result, which limits its deployment in disease monitoring. The test strip also lacks sensitivity and often produces false positives [34]. ELISA technique can overcome all these limitations but its detection process requires many chemical reagents, making it less economic for multiple monitoring tests and the operation restricted to trained lab personnel. While BPC test has been widely used for monitoring of patients, it is not a disease-specific approach as platelet drop can be a symptom of other viral infections as well. These two techniques involve complex preparation procedure and consume significant time.

## 1.3 Motivation

Hereby, an effective and accurate diagnostic technique is highly demanded for early, rapid, specific, and quantitative detection of the dengue virus antibody. The device should be portable for *in-situ* testing and can be operated with minimum complexity.

## 1.4 Research objectives

1. To fabricate a biofunctionalized TMMF sensing transducer by immobilizing the DENV NS1 glycoprotein on the tapered surface.
2. To investigate the sensor performance for detection of anti-DENV NS1 IgG antibody at various concentration.

## 1.5 Scope of research

There are several types of optical fiber biosensor technique used for biomolecules detection such as surface plasmon resonance, fiber Bragg grating (FBG), continuous tapered fiber and end tip tapered fiber [35-38]. It has also been employed in many applications such as in industry, manufacturing, security, biomedical, and many others [30]. In this research, it is focusing on a development of TMMF as a diagnosis tool to detect the anti-DENV NS1 IgG antibody. A chosen optimum waist diameter size of the TMMF undergoes the biofunctionalization process to allow the immobilization of DENV antigen on the surface of the TMMF and subsequently the complement antigen-antibody interaction. The goal of the research is to diagnose the dengue virus disease rapidly with the highly sensitive of biofunctionalized TMMF.

## 1.6 Thesis organization

This thesis is divided into five main chapters. Chapter 1 covers several sub-topics, which are introduction to dengue fever disease, problem statement, motivation of research, main objective of the research, and lastly is scope of research. Subsequently, continued to Chapter 2, which is the literature review. An overview of the theoretical background relevant to the work is provided in the chapter. In addition, this chapter discusses current research techniques used to diagnose dengue virus infections.

Chapter 3 elaborates the method used in this experiment starting from optimization of the TMMF, functionalize with APTES reagent, activation of it with GA, and immobilize it with DENV NS1 glycoprotein. The result for each step will be briefly explained in this chapter. This chapter also presents and reviews deliberately the results of detecting various concentration of DENV NS1 IgG antibody acquired from the detecting stage involved in the experimental work. The sensitivity and rapidness of detection will be discussed in this chapter. Lastly, Chapter 4 will conclude the overall findings of this research, which include significant contributions from this work as well as recommendations for future work.

## REFERENCES

- [1] P. Tavakolipoor, J. Schmidt-Chanasit, G. D. Burchard, and S. Jordan, "Clinical features and laboratory findings of dengue fever in German travellers: A single-centre, retrospective analysis," *Travel Med Infect Dis*, vol. 14, pp. 39-44, 2016.
- [2] M. Mustafa, V. Rasotgi, S. Jain, and V. Gupta, "Discovery of fifth serotype of dengue virus (DENV-5): A new public health dilemma in dengue control," *Medical Journal Armed Forces India*, vol. 71, pp. 67-70, 2015.
- [3] M. G. Guzman, S. B. Halstead, H. Artsob, P. Buchy, J. Farrar, D. J. Gubler, E. Hunsperger, A. Kroeger, H. S. Margolis, E. Martinez, M. B. Nathan, J. L. Pelegrino, C. Simmons, S. Yoksan, and R. W. Peeling, "Dengue: a continuing global threat," *Nat Rev Microbiol*, vol. 8, pp. S7-16, Dec 2010.
- [4] S. Chaterji, J. C. Allen, Jr., A. Chow, Y. S. Leo, and E. E. Ooi, "Evaluation of the NS1 rapid test and the WHO dengue classification schemes for use as bedside diagnosis of acute dengue fever in adults," *Am J Trop Med Hyg*, vol. 84, pp. 224-8, Feb 2011.
- [5] P. Jahanshahi, E. Zalnezhad, S. D. Sekaran, and F. R. Adikan, "Rapid immunoglobulin M-based dengue diagnostic test using surface plasmon resonance biosensor," *Sci Rep*, vol. 4, p. 3851, 2014.
- [6] A. Du Toit, "Viral infection: Dengue as mitochondrial landscapers," *Nature Reviews Microbiology*, 2016.
- [7] J. M. Zhou, Y. X. Tang, D. Y. Fang, J. J. Zhou, Y. Liang, H. Y. Guo, and L. F. Jiang, "Secreted expression and purification of dengue 2 virus full-length nonstructural glycoprotein NS1 in *Pichia pastoris*," *Virus Genes*, vol. 33, pp. 27-32, Aug 2006.
- [8] C.-L. Kao, C.-C. King, D.-Y. Chao, H.-L. Wu, and G. Chang, "Laboratory diagnosis of dengue virus infection: current and future perspectives in clinical diagnosis and public health," *J Microbiol Immunol Infect*, vol. 38, pp. 5-16, 2005.
- [9] B. Zhao, E. Mackow, A. Buckler-White, L. Markoff, R. M. Chanock, and C.-J. Lai, "Cloning full-length dengue type 4 viral DNA sequences: analysis of genes coding for structural proteins," *Virology*, vol. 155, pp. 77-88, 1986.
- [10] P. Dussart, L. Petit, B. Labeau, L. Bremand, A. Leduc, D. Moua, S. Matheus, and L. Baril, "Evaluation of two new commercial tests for the diagnosis of acute dengue virus infection using NS1 antigen detection in human serum," *PLoS Negl Trop Dis*, vol. 2, p. e280, 2008.
- [11] S. Zainah, A. H. Wahab, M. Mariam, M. K. Fauziah, A. H. Khairul, I. Roslina, A. Sairulakhma, S. S. Kadimon, M. S. Jais, and K. B. Chua, "Performance of a commercial rapid dengue NS1 antigen immunochromatography test with reference to dengue NS1 antigen-capture ELISA," *J Virol Methods*, vol. 155, pp. 157-60, Feb 2009.
- [12] W. Lucas, "Viral capsids and envelopes: Structure and function," *eLS*, 2010.
- [13] C.-C. Su, T.-Z. Wu, L.-K. Chen, H.-H. Yang, and D.-F. Tai, "Development of immunochips for the detection of dengue viral antigens," *Analytica Chimica Acta*, vol. 479, pp. 117-123, 2003.
- [14] D. Atias, Y. Liebes, V. Chalifa-Caspi, L. Bremand, L. Lobel, R. S. Marks, and P. Dussart, "Chemiluminescent optical fiber immunosensor for the detection of IgM antibody to dengue virus in humans," *Sensors and Actuators B: Chemical*, vol. 140, pp. 206-215, 2009.



- [15] A. R. Camara, P. M. Gouvêa, A. C. M. Dias, A. M. Braga, R. F. Dutra, R. E. de Araujo, and I. C. Carvalho, "Dengue immunoassay with an LSPR fiber optic sensor," *Opt Express*, vol. 21, pp. 27023-27031, 2013.
- [16] K. Aghaiypour and S. Safavieh, "Molecular detection of pathogenic *Leptospira* in Iran," *Archives of Razi institute*, vol. 62, pp. 191-197, 2016.
- [17] C. f. D. Control and Prevention, "Update: outbreak of acute febrile illness among athletes participating in Eco-Challenge-Sabah 2000---Borneo, Malaysia, 2000," 2016.
- [18] W. Dejnirattisai, P. Supasa, W. Wongwiwat, A. Rouvinski, G. Barba-Spaeth, T. Duangchinda, A. Sakuntabhai, V.-M. Cao-Lormeau, P. Malasit, and F. A. Rey, "Dengue virus sero-cross-reactivity drives antibody-dependent enhancement of infection with zika virus," *Nature immunology*, 2016.
- [19] R. Pessôa, J. V. Patriota, M. d. L. de Souza, A. C. Felix, N. Mamede, and S. S. Sanabani, "Investigation into an outbreak of dengue-like illness in Pernambuco, Brazil, revealed a cocirculation of zika, chikungunya, and dengue virus type 1," *Medicine*, vol. 95, p. e3201, 2016.
- [20] R. N. Mudin, "Dengue Incidence and Prevention and Control Program in Malaysia," *The International Medical Journal Malaysia*, vol. 14, 2015.
- [21] D. W. Vaughn, S. Green, S. Kalayanaraj, B. L. Innis, S. Nimmannitya, S. Suntayakorn, T. P. Endy, B. Raengsakulrach, A. L. Rothman, and F. A. Ennis, "Dengue viremia titer, antibody response pattern, and virus serotype correlate with disease severity," *Journal of Infectious Diseases*, vol. 181, pp. 2-9, 2000.
- [22] G. Guzman and G. Kouri, "Dengue and dengue hemorrhagic fever in the Americas: lessons and challenges," *Journal of Clinical Virology*, vol. 27, pp. 1-13, 2003.
- [23] <http://www.thestar.com.my/news/nation/2016/01/06/50-rise-in-dengue-deaths-health-ministry-upward-trend-also-observed-in-other-countries/>, "<3 aminopropyltriethoxysilane.pdf>."
- [24] M. a. G. Guzmán and G. Kourí, "Dengue diagnosis, advances and challenges," *International Journal of Infectious Diseases*, vol. 8, pp. 69-80, 2004.
- [25] R. W. Peeling, H. Artsob, J. L. Pelegrino, P. Buchy, M. J. Cardoso, S. Devi, D. A. Enria, J. Farrar, D. J. Gubler, and M. G. Guzman, "Evaluation of diagnostic tests: dengue," *Nature Reviews Microbiology*, vol. 8, pp. S30-S37, 2010.
- [26] F. S. R. R. Teles, "Biosensors and rapid diagnostic tests on the frontier between analytical and clinical chemistry for biomolecular diagnosis of dengue disease: a review," *Analytica Chimica Acta*, vol. 687, pp. 28-42, 2011.
- [27] R. W. Peeling, H. Artsob, J. L. Pelegrino, P. Buchy, M. J. Cardoso, S. Devi, D. A. Enria, J. Farrar, D. J. Gubler, M. G. Guzman, S. B. Halstead, E. Hunsperger, S. Kliks, H. S. Margolis, C. M. Nathanson, V. C. Nguyen, N. Rizzo, S. Vazquez, and S. Yoksan, "Evaluation of diagnostic tests: dengue," *Nat Rev Micro*.
- [28] H. Latifi, M. I. Zibaii, S. M. Hosseini, and P. Jorge, "Nonadiabatic tapered optical fiber for biosensor applications," *Photonic Sensors*, vol. 2, pp. 340-356, 2012.
- [29] J. Lou, Y. Wang, and L. Tong, "Microfiber optical sensors: A review," *Sensors*, vol. 14, pp. 5823-5844, 2014.
- [30] R. Bogue, "Fibre optic sensors: a review of today's applications," *Sensor Review*, vol. 31, pp. 304-309, 2011.
- [31] M. Pospisilova, G. Kuncova, and J. Trogl, "Fiber-Optic Chemical Sensors and Fiber-Optic Bio-Sensors," *Sensors (Basel)*, vol. 15, pp. 25208-59, Sep 30 2015.

- [32] S. Guo and S. Albin, "Transmission property and evanescent wave absorption of cladded multimode fiber tapers," *Opt Express*, vol. 11, pp. 215-223, 2003.
- [33] H. Qiu, S. Gao, P. Chen, Z. Li, X. Liu, C. Zhang, Y. Xu, S. Jiang, C. Yang, and Y. Huo, "Evanescent wave absorption sensor based on tapered multimode fiber coated with monolayer graphene film," *Optics Communications*, vol. 366, pp. 275-281, 2016.
- [34] V. Kumarasamy, A. H. Wahab, S. K. Chua, Z. Hassan, Y. K. Chem, M. Mohamad, and K. B. Chua, "Evaluation of a commercial dengue NS1 antigen-capture ELISA for laboratory diagnosis of acute dengue virus infection," *J Virol Methods*, vol. 140, pp. 75-9, Mar 2007.
- [35] C. Ribaut, V. Voisin, V. Malachovská, V. Dubois, P. Mégret, R. Wattiez, and C. Caucheteur, "Small biomolecule immunosensing with plasmonic optical fiber grating sensor," *Biosensors and Bioelectronics*, vol. 77, pp. 315-322, 2016.
- [36] R. Srinivasan, S. Umesh, S. Murali, S. Asokan, and S. Siva Gorthi, "Bare fiber Bragg grating immunosensor for real-time detection of Escherichia coli bacteria," *Journal of biophotonics*, 2016.
- [37] W. Yu, T. Lang, J. Bian, and W. Kong, "Label-free fiber optic biosensor based on thin-core modal interferometer," *Sensors and Actuators B: Chemical*, vol. 228, pp. 322-329, 2016.
- [38] F. Liang, Y. Zhang, W. Hong, Y. Dong, Z. Xie, and Q. Quan, "Direct tracking of amyloid and Tau dynamics in neuroblastoma cells using nanoplasmonic fiber tip probes," *Nano letters*, 2016.
- [39] M. G. Guzman and E. Harris, "Dengue," *The Lancet*, vol. 385, pp. 453-465.
- [40] S. Bhatt, P. W. Gething, O. J. Brady, J. P. Messina, A. W. Farlow, C. L. Moyes, J. M. Drake, J. S. Brownstein, A. G. Hoen, O. Sankoh, M. F. Myers, D. B. George, T. Jaenisch, G. R. Wint, C. P. Simmons, T. W. Scott, J. J. Farrar, and S. I. Hay, "The global distribution and burden of dengue," *Nature*, vol. 496, pp. 504-7, Apr 25 2013.
- [41] P.-Y. Shu and J.-H. Huang, "Current advances in dengue diagnosis," *Clinical and Diagnostic Laboratory Immunology*, vol. 11, pp. 642-650, 2004.
- [42] P. Y. Shu and J. H. Huang, "Current advances in dengue diagnosis," *Clin Diagn Lab Immunol*, vol. 11, pp. 642-50, Jul 2004.
- [43] Available: <http://www.namrata.co/structure-and-function-of-immunoglobulins/>
- [44] C. A. Janeway, P. Travers, M. Walport, and M. J. Shlomchik, *Immunobiology: the immune system in health and disease* vol. 1: Current Biology, 1997.
- [45] G. Karp, *Cell and Molecular Biology: Concepts and Experiments 5th Edition with Take Note Set*: John Wiley & Sons, Limited, 2008.
- [46] H. W. Schroeder, Jr. and L. Cavacini, "Structure and function of immunoglobulins," *J Allergy Clin Immunol*, vol. 125, pp. S41-52, Feb 2010.
- [47] E. A. Padlan, "Anatomy of the antibody molecule," *Molecular immunology*, vol. 31, pp. 169-217, 1994.
- [48] A. Leung, P. M. Shankar, and R. Mutharasan, "A review of fiber-optic biosensors," *Sensors and Actuators B: Chemical*, vol. 125, pp. 688-703, 2007.
- [49] M. D. George Y. Chen, Trevor. P. Newson and Gilberto Brambilla, "A Review of Microfiber and Nanofiber Based Optical Sensors," *The Open Optics Journal*, vol. 7, pp. 32-57, 2013.
- [50] Y.-J. He, "Novel and high-performance LSPR biochemical fiber sensor," *Sensors and Actuators B: Chemical*, vol. 206, pp. 212-219, 2015.

- [51] C. Guan, X. Tian, S. Li, X. Zhong, J. Shi, and L. Yuan, "Long period fiber grating and high sensitivity refractive index sensor based on hollow eccentric optical fiber," *Sensors and Actuators B: Chemical*, vol. 188, pp. 768-771, 2013.
- [52] P. Wang, G. Brambilla, M. Ding, Y. Semenova, Q. Wu, and G. Farrell, "High-sensitivity, evanescent field refractometric sensor based on a tapered, multimode fiber interference," *Optics Letters*, vol. 36, pp. 2233-2235, 2011/06/15 2011.
- [53] K.-T. Kim, K.-B. Hong, and J.-H. Park, "Transmission and Sensing Characteristics of the Biconically Tapered Cladded Multimode Fibers," *Journal of the Optical Society of Korea*, vol. 13, pp. 234-239, 2009.
- [54] G. Brambilla, "Optical fibre nanotaper sensors," *Optical Fiber Technology*, vol. 16, pp. 331-342, 2010.
- [55] H. Qiu, S. Gao, P. Chen, Z. Li, X. Liu, C. Zhang, Y. Xu, S. Jiang, C. Yang, Y. Huo, and W. Yue, "Evanescent wave absorption sensor based on tapered multimode fiber coated with monolayer graphene film," *Optics Communications*, vol. 366, pp. 275-281, 2016.
- [56] S. S. Chong, A. Aziz, and S. W. Harun, "Fibre optic sensors for selected wastewater characteristics," *Sensors*, vol. 13, pp. 8640-8668, 2013.
- [57] A. Leung, P. M. Shankar, and R. Mutharasan, "Model protein detection using antibody-immobilized tapered fiber optic biosensors (TFOBS) in a flow cell at 1310nm and 1550nm," *Sensors and Actuators B: Chemical*, vol. 129, pp. 716-725, 2008.
- [58] J. Villatoro, D. Luna-Moreno, and D. Monzón-Hernández, "Optical fiber hydrogen sensor for concentrations below the lower explosive limit," *Sensors and Actuators B: Chemical*, vol. 110, pp. 23-27, 2005.
- [59] M. Ahmad and L. L. Hench, "Effect of taper geometries and launch angle on evanescent wave penetration depth in optical fibers," *Biosens Bioelectron*, vol. 20, pp. 1312-9, Jan 15 2005.
- [60] J. Villatoro, D. Monzón-Hernández, and D. Luna-Moreno, "In-line optical fiber sensors based on cladded multimode tapered fibers," *Applied optics*, vol. 43, pp. 5933-5938, 2004.
- [61] "Evanescent Wave Absorption Based Fiber-Optic Sensor-Cascading of Bend and Tapered Geometry for Enhanced Sensitivity," in *Sensing Technology: Current Status and Future Trends III*, N. Punjabi, J. Satija, and S. Mukherji., Ed., ed Springer International Publishing, 2015, pp. 25-45.
- [62] J. M. Corres, I. R. Matias, J. Bravo, and F. J. Arregui, "Tapered optical fiber biosensor for the detection of anti-gliadin antibodies," *Sensors and Actuators B: Chemical*, vol. 135, pp. 166-171, 2008.
- [63] S. G. a. S. Albin, "Transmission property and evanescent wave absorption of cladded multimode fiber tapers," *Opt. Express*, vol. 11, pp. 215-223, 2003.
- [64] C. Shi, X. Wang, P. Zhou, X. Xu, and Q. Lu, "Theoretical study of mode evolution in active long tapered multimode fiber," *Opt Express*, vol. 24, pp. 19473-19490, 2016.
- [65] J. Miller, A. Castaneda, K. H. Lee, M. Sanchez, A. Ortiz, E. Almaz, Z. T. Almaz, S. Murinda, W. J. Lin, and E. Salik, "Biconically tapered fiber optic probes for rapid label-free immunoassays," *Biosensors (Basel)*, vol. 5, pp. 158-71, Jun 2015.
- [66] M. V. A. A. Díez, J.L. Cruz, "In-line fiber-optic sensors based on the excitation of surface plasma modes in metal-coated tapered fibers," *Sensors and Actuators B: Chemical*, vol. 73, pp. 95-99, 2001.

- [67] W. B. Ji, S. H. K. Yap, N. Panwar, L. L. Zhang, B. Lin, K. T. Yong, S. C. Tjin, W. J. Ng, and M. B. A. Majid, "Detection of low-concentration heavy metal ions using optical microfiber sensor," *Sensors and Actuators B: Chemical*, vol. 237, pp. 142-149, 2016.
- [68] R. Kostecki, H. Ebendorff-Heidepriem, S. Afshar V, G. McAdam, C. Davis, and T. M. Monro, "Novel polymer functionalization method for exposed-core optical fiber," *Optical Materials Express*, vol. 4, p. 1515, 2014.
- [69] R. Raghunandhan, L. H. Chen, H. Y. Long, L. L. Leam, P. L. So, X. Ning, and C. C. Chan, "Chitosan/PAA based fiber-optic interferometric sensor for heavy metal ions detection," *Sensors and Actuators B: Chemical*, vol. 233, pp. 31-38, 2016.
- [70] D. Rithesh Raj, S. Prasanth, and C. Sudarsanakumar, "Development of LSPR-Based Optical Fiber Dopamine Sensor Using L-Tyrosine-Capped Silver Nanoparticles and Its Nonlinear Optical Properties," *Plasmonics*, 2016.
- [71] D. Sun, T. Guo, Y. Ran, Y. Huang, and B. O. Guan, "In-situ DNA hybridization detection with a reflective microfiber grating biosensor," *Biosens Bioelectron*, vol. 61, pp. 541-6, Nov 15 2014.
- [72] Y. C. Tan, W. B. Ji, V. Mamidala, K. K. Chow, and S. C. Tjin, "Carbon-nanotube-deposited long period fiber grating for continuous refractive index sensor applications," *Sensors and Actuators B: Chemical*, vol. 196, pp. 260-264, 2014.
- [73] Y. Zhao, X.-g. Li, X. Zhou, and Y.-n. Zhang, "Review on the graphene based optical fiber chemical and biological sensors," *Sensors and Actuators B: Chemical*, vol. 231, pp. 324-340, 2016.
- [74] J. Zajíc, M. Bittner, T. Brányik, A. Solovyev, S. Šabata, G. Kuncová, and M. Pospíšilová, "Repetitive inductions of bioluminescence of *Pseudomonas putida* TVA8 immobilised by adsorption on optical fibre," *Chemical Papers*, vol. 70, 2016.
- [75] S. K. Chauhan, N. Punjabi, D. K. Sharma, and S. Mukherji, "A silicon nitride coated LSPR based fiber-optic probe for possible continuous monitoring of sucrose content in fruit juices," *Sensors and Actuators B: Chemical*, vol. 222, pp. 1240-1250, 2016.
- [76] S. K. Vashist, E. M. Schneider, E. Lam, S. Hrapovic, and J. H. Luong, "One-step antibody immobilization-based rapid and highly-sensitive sandwich ELISA procedure for potential in vitro diagnostics," *Sci Rep*, vol. 4, 2014.
- [77] N. S. K. Gunda, M. Singh, L. Norman, K. Kaur, and S. K. Mitra, "Optimization and characterization of biomolecule immobilization on silicon substrates using (3-aminopropyl)triethoxysilane (APTES) and glutaraldehyde linker," *Applied Surface Science*, vol. 305, pp. 522-530, 2014.
- [78] S. K. Vashist, E. Lam, S. Hrapovic, K. B. Male, and J. H. Luong, "Immobilization of antibodies and enzymes on 3-aminopropyltriethoxysilane-functionalized bioanalytical platforms for biosensors and diagnostics," *Chem Rev*, vol. 114, pp. 11083-130, Nov 12 2014.
- [79] G.-R. Han, Y.-J. Song, and C.-H. Jang, "Label-free detection of viruses on a polymeric surface using liquid crystals," *Colloids and Surfaces B: Biointerfaces*, vol. 116, pp. 147-152, 2014.
- [80] S. Teixeira, G. Burwell, A. Castaing, D. Gonzalez, R. Conlan, and O. Guy, "Epitaxial graphene immunosensor for human chorionic gonadotropin," *Sensors and Actuators B: Chemical*, vol. 190, pp. 723-729, 2014.
- [81] P. Saengdee, W. Chairiratanakul, W. Bunjongpru, W. Sripumkhai, A. Srisuwan, W. Jeamsaksiri, C. Hruanun, A. Poyai, and C. Promptmas, "Surface

- modification of silicon dioxide, silicon nitride and titanium oxynitride for lactate dehydrogenase immobilization," *Biosensors and Bioelectronics*, vol. 67, pp. 134-138, 2015.
- [82] K. Kim, C. Park, D. Kwon, D. Kim, M. Meyyappan, S. Jeon, and J.-S. Lee, "Silicon nanowire biosensors for detection of cardiac troponin I (cTnI) with high sensitivity," *Biosensors and Bioelectronics*, vol. 77, pp. 695-701, 2016.
- [83] H. Yan, N. Tang, G. A. Jairo, S. Chakravarty, D. A. Blake, and R. T. Chen, "High-sensitivity high-throughput chip based biosensor array for multiplexed detection of heavy metals," in *SPIE BiOS*, 2016, pp. 972507-972507-5.
- [84] S. Kumar, J. G. Sharma, S. Maji, and B. D. Malhotra, "Nanostructured zirconia decorated reduced graphene oxide based efficient biosensing platform for non-invasive oral cancer detection," *Biosensors and Bioelectronics*, vol. 78, pp. 497-504, 2016.
- [85] V. Mishra, P. Patel, S. Kumari, and G. Mishra, "Dengue NS1 detection used chemically modified porous silicon microcavity (PSMC)," *Silicon*, vol. 8, pp. 401-407, 2016.
- [86] R. M. Pasternack, S. Rivillon Amy, and Y. J. Chabal, "Attachment of 3-(aminopropyl) triethoxysilane on silicon oxide surfaces: dependence on solution temperature," *Langmuir*, vol. 24, pp. 12963-12971, 2008.
- [87] D. Kim and A. E. Herr, "Protein immobilization techniques for microfluidic assays," *Biomicrofluidics*, vol. 7, p. 041501, 2013.
- [88] G.-J. Zhang, L. Zhang, M. J. Huang, Z. H. H. Luo, G. K. I. Tay, E.-J. A. Lim, T. G. Kang, and Y. Chen, "Silicon nanowire biosensor for highly sensitive and rapid detection of Dengue virus," *Sensors and Actuators B: Chemical*, vol. 146, pp. 138-144, 2010.
- [89] Y. Tian, W. Wang, N. Wu, X. Zou, and X. Wang, "Tapered optical fiber sensor for label-free detection of biomolecules," *Sensors (Basel)*, vol. 11, pp. 3780-90, 2011.
- [90] C.-C. Wu, T.-M. Pan, C.-S. Wu, L.-C. Yen, C.-K. Chuang, S.-T. Pang, Y.-S. Yang, and F.-H. Ko, "Label-free detection of prostate specific antigen using a silicon nanobelt field-effect transistor," *Int. J. Electrochem. Sci*, vol. 7, pp. 4432-4442, 2012.
- [91] N. S. Gunda, M. Singh, Y. Purwar, S. L. Shah, K. Kaur, and S. K. Mitra, "Micro-spot with integrated pillars (MSIP) for detection of dengue virus NS1," *Biomed Microdevices*, vol. 15, pp. 959-71, Dec 2013.
- [92] L. Zhou, A. Zhu, X. Lou, D. Song, R. Yang, H. Shi, and F. Long, "Universal quantum dot-based sandwich-like immunoassay strategy for rapid and ultrasensitive detection of small molecules using portable and reusable optofluidic nano-biosensing platform," *Analytica Chimica Acta*, vol. 905, pp. 140-148, 2016.
- [93] R. Reverberi and L. Reverberi, "Factors affecting the antigen-antibody reaction," *Blood Transfusion*, vol. 5, p. 227, 2007.
- [94] P. W. Atkins and J. De Paula, *The elements of physical chemistry* vol. 3: Oxford University Press New York, NY, USA., 2005.
- [95] L. Xu, Y. Li, and B. Li, "Nonadiabatic fiber taper-based Mach-Zehnder interferometer for refractive index sensing," *Applied Physics Letters*, vol. 101, p. 153510, 2012.
- [96] T. Liu, Y. Zhao, Z. Zhang, P. Zhang, J. Li, R. Yang, C. Yang, and L. Zhou, "A fiber optic biosensor for specific identification of dead Escherichia coli O157:H7," *Sensors and Actuators B: Chemical*, vol. 196, pp. 161-167, 2014.

- [97] T. Q. Huy, N. T. Hanh, N. T. Thuy, P. V. Chung, P. T. Nga, and M. A. Tuan, "A novel biosensor based on serum antibody immobilization for rapid detection of viral antigens," *Talanta*, vol. 86, pp. 271-7, Oct 30 2011.
- [98] N. S. K. Gunda, M. Singh, L. Norman, K. Kaur, and S. K. Mitra, "Optimization and characterization of biomolecule immobilization on silicon substrates using (3-aminopropyl) triethoxysilane (APTES) and glutaraldehyde linker," *Applied Surface Science*, vol. 305, pp. 522-530, 2014.
- [99] O. A. Monteiro and C. Airoidi, "Some studies of crosslinking chitosan–glutaraldehyde interaction in a homogeneous system," *International Journal of Biological Macromolecules*, vol. 26, pp. 119-128, 1999.
- [100] U. Schlottmann, B. Stock, and O. H. Chemicals, "SIDS Initial Assessment Report For SIAM 17."
- [101] S. K. Vashist, E. Lam, S. Hrapovic, K. B. Male, and J. H. Luong, "Immobilization of antibodies and enzymes on 3-aminopropyltriethoxysilane-functionalized bioanalytical platforms for biosensors and diagnostics," *Chem Rev*, vol. 114, pp. 11083-11130, 2014.
- [102] J. C. Love, L. A. Estroff, J. K. Kriebel, R. G. Nuzzo, and G. M. Whitesides, "Self-assembled monolayers of thiolates on metals as a form of nanotechnology," *Chem Rev*, vol. 105, pp. 1103-1170, 2005.
- [103] G. Liu and K. Li, "Micro/nano optical fibers for label-free detection of abrin with high sensitivity," *Sensors and Actuators B: Chemical*, vol. 215, pp. 146-151, 2015.
- [104] B. Ramakrishna and V. Sai, "Evanescent wave absorbance based U-bent fiber probe for immunobiosensor with gold nanoparticle labels," *Sensors and Actuators B: Chemical*, vol. 226, pp. 184-190, 2016.
- [105] C. Ribaut, V. Voisin, V. Malachovska, V. Dubois, P. Megret, R. Wattiez, and C. Caucheteur, "Small biomolecule immunosensing with plasmonic optical fiber grating sensor," *Biosens Bioelectron*, vol. 77, pp. 315-22, Mar 15 2016.
- [106] R. Bharadwaj, V. Sai, K. Thakare, A. Dhawangale, T. Kundu, S. Titus, P. K. Verma, and S. Mukherji, "Evanescent wave absorbance based fiber optic biosensor for label-free detection of E. coli at 280nm wavelength," *Biosensors and Bioelectronics*, vol. 26, pp. 3367-3370, 2011.
- [107] V. Sai, T. Kundu, C. Deshmukh, S. Titus, P. Kumar, and S. Mukherji, "Label-free fiber optic biosensor based on evanescent wave absorbance at 280nm," *Sensors and Actuators B: Chemical*, vol. 143, pp. 724-730, 2010.
- [108] C. M. G. R. H. Garrett, and R. H. Garrett, *Student lecture notebook for Biochemistry*: Belmont, CA: Thomson/Brooks Cole, 2005.
- [109] R. J. Linhardt and H. G. Bazin, "Properties of carbohydrates," in *Glycoscience: Chemistry and Chemical Biology I-III*, ed: Springer, 2001, pp. 53-61.
- [110] I. Baianu and V. Prisecaru, "NMR, NIR, and Infrared Spectroscopy of Carbohydrate– Protein Interactions and Glycoproteins," in *NMR Spectroscopy of Polymers: Innovative Strategies for Complex Macromolecules*, ed: ACS Publications, 2011, pp. 337-352.
- [111] S. Herrmann, B. Leshem, S. Landes, B. Rager-Zisman, and R. S. Marks, "Chemiluminescent optical fiber immunosensor for the detection of anti-West Nile virus IgG," *Talanta*, vol. 66, pp. 6-14, Mar 31 2005.
- [112] A. Wilkinson and A. McNaught, "IUPAC Compendium of Chemical Terminology, (the " Gold Book")," ed: Blackwell Scientific Publications, Oxford, 1997.
- [113] G. Karp, *Cell and Molecular Biology: Concepts and Experiments*, 5th ed.: John Wiley & Sons, Limited, 2008.

- [114] J. R. C. Lima, M. Z. Rouquayrol, M. R. M. Callado, M. I. F. Guedes, and C. Pessoa, "Interpretation of the presence of IgM and IgG antibodies in a rapid test for dengue: analysis of dengue antibody prevalence in Fortaleza City in the 20th year of the epidemic," *Revista da Sociedade Brasileira de Medicina Tropical*, vol. 45, pp. 163-167, 2012.
- [115] A. D'amico, C. Di Natale, and A. Taroni, "Sensors Parameters in Sensors for domestic applications," ed: World Scientific: Singapore, 1995.
- [116] B.-T. Teoh, S.-S. Sam, K.-K. Tan, M. B. Danlami, M.-H. Shu, J. Johari, P.-S. Hooi, D. Brooks, O. Piepenburg, and O. Nentwich, "Early detection of dengue virus by use of reverse transcription-recombinase polymerase amplification," *Journal of clinical microbiology*, vol. 53, pp. 830-837, 2015.
- [117] A. Brecht and G. Gauglitz, "Optical probes and transducers," *Biosensors and Bioelectronics*, vol. 10, pp. 923-936, 1995.
- [118] B. T. Cunningham and L. G. Laing, "Advantages and application of label-free detection assays in drug screening," *Expert opinion on drug discovery*, vol. 3, pp. 891-901, 2008.