



UNIVERSITI PUTRA MALAYSIA

***ENHANCEMENT OF METHANE PRODUCTION FROM PALM OIL MILL
EFFLUENT THROUGH CO-DIGESTION AND BIO-AUGMENTATION
TREATMENT***

NURLIYANA MOHD YAAKOB

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By

NURLIYANA MOHD YAAKOB

**Thesis Submitted to the School of Graduate Studies,
Universiti Putra Malaysia, in Fulfilment of the Requirements for the
Degree of Doctor of Philosophy**

November 2016

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fulfilment of the requirement for the degree of Doctor of Philosophy

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November 2016

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Degradation process of palm oil mill effluent (POME) via anaerobic digestion produced a valuable methane gas which has recently become a potential source of renewable energy in Malaysia. As to date, only a few palm oil mills in Malaysia are involved in recovering the biogas since current methane production by conventional technologies is below expectation. The composition of POME exhibits nutrient availability for the indigenous microorganisms and will affect the direction of the degradation process. Similarly, the performance of microbial community during the treatment process plays a significant role as they are primarily the agents that accomplish the desired treatment. This research aims at assessing the effects of POME characteristics on the methane production and subsequently investigating ways of enhancing the methane productivity by improving the feedstock characteristics and microbial consortia. This study was divided into three parts. The first part evaluates the POME physicochemical and biochemical characteristics. Total carbon of POME was found to be relatively low leading to low carbon to nitrogen (C/N) ratio value which affects the production of methane gas. In addition, a low pH value of POME was recorded, influencing the optimum condition of indigenous microbes specifically the methanogens during the digestion process. The second and third part of this study were conducted to improve the feedstock characteristics as well as to increase the methane productivity and substrate degradability by conducting the co-digestion and bioaugmentation treatment.

Co-digestion treatment of POME with natural carbon substrate in the palm oil mill, specifically empty fruit bunch (EFB), was investigated by varying the C/N ratio of the substrates loading. A significant effect of different C/N ratios towards methane production and methane yield were observed. The co-digestion treatment resulted in higher methane production where C/N ratio of 45 delivers the highest result with twelve times and 61.7% improvement in methane production and methane yield, respectively. While for substrate biodegradability,

a significant reduction of organic substances was identified with different C/N ratios except for degradation of cellulose content in EFB. However, co-digestion samples were found to be ineffective in terms of substrate degradability compared to mono-digestion of POME in total reduction of total solid (TS), biological oxygen demand (BOD) and chemical oxygen demand (COD) respectively for POME substrate. While in EFB substrate degradability, a low total reduction was observed for all co-digestion samples. Cellulose content particularly indicates an increase in percentage after the digestion process.

Bioaugmentation treatment towards POME and POME co-digested with EFB was investigated by addition of different microbial loadings of cellulolytic bacteria and methanogenic archaea by single or co-augmentation technique, to improve the microbial activity in the feedstock and to enhance the methane productivity and substrate biodegradability. Treatment of single augmentation by *Bacillus subtilis* in co-digestion sample at 10% microbial loading (C-B10) resulted in the highest methane production with 14.3% increment compared to non-augmented co-digestion sample. In the matter of substrate degradability, bioaugmentation treatment of C-B10 activated the cellulose reduction in EFB with 1.95% reduction as compared to non-augmented co-digestion sample which had an increased value of 4.4%. Besides, sample C-B10 recorded better TS, total volatile solid (TVS), COD, organic matter (OM) and lignin reduction with 27.2%, 22.3%, 90.9% 29.4% and 2.8 times higher degradation, respectively compared to the non-augmented co-digestion sample.

Treatment of co-augmentation between *B. subtilis* and mix methanogens was operated to maximize the methane productivity and substrate degradability. Sample of single digestion co-augmented at 5% microbial loading (S-B5-M5) gives a significant increase in methane productivity with 9.4-29.6% and relatively 2.1 times improvement compared to single augmented sample of similar microbial loading (S-B5 and S-M5) and the non-augmented single digestion sample respectively. In terms of substrate degradability, sample of S-B5-M5 showed a notable results in TVS and COD reduction where it attained the highest removal efficiencies among all co-augmented samples with 37.3% and 55.2% improvement measure up to non-augmented single digestion sample.

Co-digestion and bioaugmentation treatment were demonstrated as effective means of enhancing the methane productivity and substrate biodegradability of the feedstock. Optimum loading of the co-substrate and microbial cultures from this research can be implemented in the field to better develop the methane production system specifically in the closed anaerobic pond. A substantial number of palm oil millers will be able to apply the results of this study and broaden the chances to recover the biogas produced into a valuable renewable energy.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**PENINGKATAN PENGHASILAN GAS METANA DARIPADA AIR
PEMROSESAN KILANG KELAPA SAWIT MELALUI RAWATAN
DEGRADASI BERSAMA DAN PENAMBAHAN MIKROORGANISMA**

Oleh

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Proses degradasi air pemprosesan kilang kelapa sawit (POME) melalui degradasi anaerobik menghasilkan gas metana berharga yang kebelakangan ini telah menjadi sumber potensi tenaga boleh diperbaharui di Malaysia. Sehingga kini, hanya beberapa kilang minyak sawit di Malaysia yang terlibat dalam mendapatkan biogas memandangkan pengeluaran semasa metana menggunakan teknologi konvensional adalah di bawah jangkaan. Komposisi POME menunjukkan ketersediaan nutrien untuk mikroorganisma sedia ada dan akan memberi kesan kepada arah tuju proses degradasi. Begitu juga prestasi komuniti mikrob semasa proses rawatan yang memainkan peranan penting, terutama kerana mereka adalah agen yang mencapai rawatan yang dikehendaki. Kajian ini bertujuan untuk menilai kesan ciri-ciri POME dalam pengeluaran metana dan seterusnya menyiasat cara untuk mempertingkatkan produktiviti metana dengan memperbaiki ciri-ciri bahan mentah dan konsortia mikrob. Kajian ini dibahagikan kepada tiga bahagian. Bahagian pertama adalah menilai ciri-ciri fisikokimia dan biokimia POME. Jumlah karbon POME secara relatif didapati rendah dan mengakibatkan nilai nisbah karbon kepada nitrogen (C/N) turut menjadi rendah dan akan menjejaskan pengeluaran gas metana. Di samping itu, POME didapati mencatatkan nilai pH yang rendah dan akan mempengaruhi keadaan optimum mikrob asli terutamanya metanogen ketika proses degradasi. Bahagian kedua dan ketiga kajian ini telah dijalankan untuk meningkatkan ciri-ciri bahan mentah serta untuk meningkatkan produktiviti metana dan keupayaan degradasi substrat dengan menjalankan rawatan untuk degradasi bersama dan penambahan mikroorganisma.

Rawatan degradasi bersama antara POME dengan bahan karbon asli daripada kilang minyak sawit, khususnya tandan kosong buah kelapa sawit (EFB), telah dikaji dengan mengubah nisbah C/N muatan substrat. Terdapat kesan yang ketara antara nisbah C/N yang berbeza ke arah pengeluaran metana dan hasil metana. Rawatan degradasi bersama telah menghasilkan peningkatan dalam pengeluaran metana di mana nisbah C/N 45 menghasilkan hasil tertinggi dengan

dua belas kali dan 61.7% peningkatan masing-masing dalam pengeluaran metana dan hasil metana. Manakala bagi keupayaan degradasi substrat, pengurangan ketara bahan organik dengan nisbah C/N yang berbeza telah dikenal pasti kecuali degradasi kandungan selulosa di dalam EFB. Walaubagaimanapun, sampel degradasi bersama didapati tidak berkesan dari segi keupayaan degradasi substrat berbanding degradasi POME secara tunggal dalam jumlah pengurangan jumlah pepejal (TS), keperluan oksigen secara biologi (BOD) dan keperluan oksigen secara kimia (COD) untuk substrat di dalam POME. Bagi keupayaan degradasi substrat di dalam EFB, jumlah pengurangan yang rendah telah dikenal pasti untuk semua sampel degradasi bersama. Kandungan selulosa khususnya menunjukkan peratusan peningkatan selepas proses degradasi.

Rawatan penambahan mikroorganisma ke dalam POME dan degradasi bersama antara POME dan EFB telah dikaji dengan penambahan muatan mikrob yang berbeza daripada bakteria selulosa dan arkea metanogen dengan teknik tunggal atau penambahan bersama, untuk meningkatkan aktiviti mikrob dalam bahan mentah dan meningkatkan produktiviti metana dan keupayaan degradasi substrat. Rawatan penambahan tunggal oleh *Bacillus subtilis* dalam sampel degradasi bersama pada 10% muatan mikrob (C-B10) menghasilkan pengeluaran metana tertinggi dengan 14.3% kenaikan berbanding sampel degradasi bersama yang tidak ditambah mikroorganisma. Dalam keupayaan degradasi substrat, rawatan penambahan mikroorganisma oleh C-B10 telah meningkatkan degradasi selulosa dalam EFB dengan jumlah pengurangan 1.95% berbanding dengan sampel degradasi bersama yang tidak ditambah mikroorganisma yang mempunyai peningkatan sebanyak 4.4%. Selain itu, sampel C-B10 mencatatkan pengurangan lebih baik dalam TS, jumlah pepejal mudah meruap (TVS), COD, bahan organik (OM) dan pengurangan lignin masing-masing dengan 27.2%, 22.3%, 90.9% 29.4% dan 2.8 kali lebih tinggi degradasi berbanding dengan sampel degradasi bersama yang tidak ditambah mikroorganisma.

Rawatan penambahan mikroorganisma bersama antara *B. subtilis* dan campuran methanogens dijalankan untuk memaksimumkan produktiviti metana dan keupayaan degradasi substrat. Sampel degradasi POME dengan penambahan mikroorganisma bersama pada 5% muatan mikrob (S-B5-M5) memberikan peningkatan yang ketara dalam produktiviti metana dengan 9.4-29.6% dan secara relatif 2.1 kali peningkatan masing-masing berbanding dengan sampel ditambah muatan mikrob yang sama secara tunggal (S-B5 dan S-M5) dan sampel degradasi POME yang tidak ditambah mikroorganisma. Dari segi keupayaan degradasi substrat, sampel S-B5-M5 menunjukkan keputusan yang ketara dalam pengurangan TVS dan COD di mana ia mencapai jumlah penyingkiran tertinggi dalam kalangan sampel yang ditambah mikroorganisma dengan 37.3% dan 55.2% peningkatan berbanding sampel degradasi POME secara tunggal yang tidak ditambah mikroorganisma.

Rawatan degradasi bersama dan rawatan penambahan mikroorganisma telah dikenal pasti sebagai cara yang berkesan untuk meningkatkan produktiviti metana dan keupayaan degradasi bahan mentah. Muatan optimum substrat degradasi bersama dan kultur mikrob daripada kajian ini dapat dilaksanakan di lapangan

untuk membangunkan sistem pengeluaran metana yang lebih baik khususnya dari kolam anaerobik tertutup. Sebilangan besar daripada pengilang minyak kelapa sawit akan dapat menggunakan hasil kajian ini dan meluaskan peluang untuk menukar biogas yang dihasilkan menjadi tenaga boleh diperbaharui.



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I certify that a Thesis Examination Committee has met on 2016 to conduct the final examination of Nurliyana Mohd Yaakob on her thesis entitled Enhancement of Methane Production from Palm Oil Mill Effluent Through Co-digestion and Bio-augmentation Treatment in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

ANOVA	Analysis of Variance
APHA	American Public Health Association
BOD	Biological Oxygen Demand
C/N	Carbon to Nitrogen
CFU	Colony Forming Unit
CMCA	Carboxymethylcellulose Agar
COD	Chemical Oxygen Demand
CRA	Congo Red Agar
DOE	Department of Environment
EFB	Empty Fruit Bunches
FFB	Fresh Fruit Bunches
GHG	Green House Gases
LEL	Lower Exposure Limit
LSD	Least Significant Difference
NA	Nutrient Agar
OM	Organic Matter
PCR	Polymerase Chain Reaction
PDA	Potato Dextrose Agar
POME	Palm Oil Mill Effluent
TA	Thioglycollate Agar
TS	Total Solids
TVS	Total Volatile Solids

CHAPTER 1

INTRODUCTION

1.1 General Background

Malaysia is a global leader in the palm oil industry that accounts for 33.1% of world palm oil production and 39.6% of world palm oil products exporter (MPOC, 2014). Currently, palm oil industry is one of the nation's important industries having the status as the largest contributor in the agricultural sector and the fourth largest contributor to the national economy as it covers the entire value chain from plantations to downstream activities. Despite a high economic return to the country, the industry is under incessant criticism for its performance from the environmental perception. The palm oil industry faces the challenge to balance between their economic and development sustainability as well as environmental preservation.

Oil palm processing generates a highly polluting wastewater commonly known as palm oil mill effluent (POME) that can cause a severe pollution to the waterways if discharged without proper treatment due to the oxygen depletion and other subsequent related effects. Likewise, a huge amount of solid wastes are produced from the oil palm extraction process predominantly empty fruit bunches (EFB), usually unsystematically dumped on the land causing pollution to the surrounding area. POME and EFB are the largest wastes produce in oil palm productions and these huge waste are nonetheless a good source of biomass due to their high organic contents and has recently become a potential source of renewable energy in Malaysia (Bakar, 2014).

Degradation process of POME via anaerobic digestion will emit greenhouse gases (GHG) primarily methane gas that is effective in trapping heat in the atmosphere, with a global warming potential over 20 times than carbon dioxide (Tabak, 2009). Wijbrans and Van Zutphen (2005) have reported that methane release from POME equivalents to 9kg/tonne fresh fruit bunch (FFB) (at 0.7 m³ POME/tonne FFB, 28 m³ biogas/m³ POME and 65% methane in the biogas). Nevertheless, the methane gas produced can be a valuable source of bioenergy due to its high calorific value.

Methane collection can be carried out via various invented technologies either in a high efficiency closed-tank anaerobic digester system or covered anaerobic pond system. Harnessed methane gas is suitable not only to substitute non-renewable energy for multiple application but also can significantly reduce the GHG emission and dependence on fossil fuels to develop a more sustainable palm oil industry. Methane gas production by the conventional method of ponding system employed by the majority of the palm oil millers for POME treatment, however, are not fully recovered for utilization. As to date, only a few

palm oil mills in Malaysia are involved in recovering the biogas to be utilized as additional sources of energy in the mills or supplying the energy to the national electricity grid. This is related to the current methane production by using the conventional technologies, which is below expectation (Chin et al., 2013).

The degradation process of POME into methane consists of four main stages which include hydrolysis to disintegrate the complex polymers, acidogenesis to further break down the soluble organic compounds, acetogenesis to degrade the resulting organic acids and lastly methanogenesis that produce methane and carbon dioxide gases (Gerardi, 2003; Poh and Chong, 2009). Energy is required for these cellular activities including growth and reproduction processes which is generally derived from the breakdown of the chemical bond mainly between carbon compounds. One of the reason for low methane production by POME might be due to its unstable nutrient availability which subsequently affected the nutrient requirements of the microorganisms. Besides, a deficiency in the desired microbial colonies to execute the degradation process could also influence the methane production (Cavaleiro et al., 2010).

Thus, effective methods should be implemented to counteract the inadequacies. EFB possessed a high carbon content that is suitable to be co-digested with POME to balance the nutrient availability and subsequently to assist in producing higher methane gas (Baharuddin et al., 2010). Since POME and EFB are comprised of cellulosic compounds, bio-augmentation by cellulolytic microorganisms might assist in the degradation of the complex compounds, as well as addition of methanogenic colony which might contribute to improve the methane production.

1.2 Problem Statement

Malaysian palm oil sector consistently seeks ways to further enhance its sustainability efforts. One of the effort implemented is via Entry Point Project No 5 (EPP 5) under National Key Economic Areas (NKEA) which has been inspired by the Government in the Tenth Malaysia Plan (2011 - 2015) that aims to develop the biogas facilities at palm oil mills to boost the renewable energy sector as well as reducing the environmental impacts. Besides, Malaysian Fifth Fuel Policy targeted to achieve 5% of national grid-connected electricity generation from renewable resources, particularly biomass.

With various biogas capturing technologies offered, covered lagoon was widely opted since ponding system was used by a majority of the palm oil millers besides it is cost-effective and easier to operate. However, low methane production was observed from the covered lagoon technologies as compared to the high-tech closed anaerobic digester tank leading the millers to merely flared the methane produced instead of utilizing it for peripheral use and lowering the chances to produce electricity for grid connection. One of the reasons might be due to the unbalanced nutrient in the feedstock since POME has been indicated

to have a low carbon to nitrogen (C/N) ratio. Carbon act as the main energy sources for the indigenous microbes in the feedstock, while nitrogen support in structural material metabolism process. Hence, the unstable C/N ratio will subsequently affect their growth, methane productivity, and degradability rate.

Co-digestion of POME with a high carbon source will stabilize the nutrient availability in the feedstock. EFB is a lignocellulosic material with a high amount of carbon which makes it a potential natural co-substrate in the digestion process. Nonetheless, the loading of the co-substrate has to be specifically quantified to avoid unbalance nutrient availability during the digestion process. Loading proportion of POME as the primary substrate and EFB as the co-substrate in facultative digestion estimated by C/N ratio content on weight basis has yet to be reported.

The addition of prepared microbial cultures to a biomass treatment system or known as bio-augmentation can increase the density of desired microbes and their enzymes. The addition of cellulolytic bacteria and methanogens archaea as bio-augmentation treatment in facultative digestion can assist in improving the substrate degradability and methane productivity respectively. Thus far, limited studies had been reported towards bio-augmentation of *Bacillus subtilis* and mix methanogens in methane production from POME. Loading of the bacterium has to be quantified to avoid excess microbial colony and severe competition with the indigenous microorganisms during the digestion process.

Degradability of the substrates throughout the digestion process of POME was also in a limited search. Depletion of substrates in the feedstock in terms of organic substances after the digestion process signified total degradation and degradability of the materials. High degradability of biomass sample implies a high potential for methane productivity. Thus, the degradation rate of biomass feedstock can practically estimate the methane production and methane yield. Subsequently, methane production can be enhanced by improving the degradation rate of the substrates.

1.3 Justification

As mentioned earlier, methane production was affected by numerous factors from physical, chemical and biological properties of the feedstock as well as its operational parameters. The composition of the feedstock exhibits the nutrient availability for the indigenous microorganisms and will affect the direction of the degradation process. Similarly, the performance of microbial community during the treatment process plays a significant role in determining the production of methane gas, as they are primarily the agents that accomplish the desired treatment. Hence, it is essential to distinguish characteristics of the feedstock and to understand the microbial participating in metabolic reaction during the degradation process, in developing the digestion system operational efficiency.

Methane production from POME can be enhanced with numerous techniques and technologies available. In this study, two methods were evaluated which are 1) adding a co-substrate and 2) adding a commercialized microbes to investigate their effects toward the production of methane gas and substrates degradability. The addition of co-substrate can improve deficiency of the main substrates and subsequently enhance the methane production. Palm oil mills generate various types of cellulosic waste that can potentially be used as a co-substrate and EFB was selected for its high carbon content and high availability. In addition, microbes were added to increase the desired microbial colony in degrading the organic substrate. Cellulolytic bacteria were selected to assist in degrading the cellulosic material of POME and EFB while mix methanogens were selected to further enhance the methanogenesis process and subsequently the production of methane. Meanwhile, integration of both microorganisms was conducted to overcome the limitations of individual degradation potential and for further maximizing the yield of methane gas.

Sample of POME added with EFB at different loadings of C/N ratio was experimented in Part 1 of the study to observe the effect towards methane productivity. Eight loadings of C/N ratio were selected from 25, 30, 35, 40, 45, 50 and 55 including control where no EFB was added at all. These C/N ratios were selected in the range of original C/N ratio of both POME and EFB. Methane productivity and substrate degradability were observed throughout the digestion process and optimum C/N ratio was selected from the sample with the highest production of cumulative methane.

Meanwhile, bio-augmentation was done in Part 2 of the study where commercialized microbes of *B. subtilis* and mix methanogens were added at different concentrations of microbial loading to further observe the effect towards degradability of the substrate and methane productivity. Microbial loading was selected at 5% and 10% (v/v) for *B. subtilis* and mix methanogens at an average concentration of 10^6 colony forming unit (CFU). Both loading concentrations of singular cultures were further combined to observe the effects of their integration toward substrate degradation and methane productivity.

Optimized loading of the co-substrate and microbial colony from this research can be implemented in the field to better develop the methane production system. Since the majority of palm oil millers opted to apply ponding system for the POME treatment, a substantial number of palm oil millers will be able to apply the result of this study and broaden the chances to recover the biogas produced into valuable renewable energy. This thesis presents a research which aims at assessing the effects of palm oil mills bio-wastes characteristics on the methane production and subsequently investigating ways of enhancing the methane productivity by improving the feedstock characteristics and microbial consortia.

1.4 Objectives

The general objective of this research is to study the methane production from POME through co-digestion treatment with EFB and bio-augmentation treatment with *B. subtilis* and mix methanogens, via facultative digestion system. This research was carried out with the following specific objectives:

- 1) To determine the physicochemical, biochemical and microbial characteristics of raw POME and EFB sample.
- 2) To study the effect of different C/N ratios in co-digestion treatment of POME and EFB, on methane productivity and substrate degradability rate.
- 3) To study the effect of microbial types (*B. subtilis* and mix methanogens) and microbial loadings in bio-augmentation treatment of single and co-digestion sample (at selected C/N ratio), on methane productivity and substrate degradability rate.
- 4) To evaluate the co-augmentation treatment of single and co-digestion sample (at selected C/N ratio) towards methane productivity and substrate degradability rate, by incorporating *B. subtilis* and mix methanogens at different loadings.

REFERENCES

- Abd El-Gawad, H. S. (2014). Oil and grease removal from industrial wastewater using new utility approach. *Advances in Environmental Chemistry*, 2014, 1–6.
- Abdullah, N., & Sulaiman, F. (2013). The oil palm wastes in Malaysia. In M. D. Matovic (Ed.), *Biomass Now – Sustainable Growth and Use* (pp. 75–100). InTech.
- Abdullah, N., Sulaiman, F., & Gerhauser, H. (2011). Characterisation of oil palm empty fruit bunches for fuel application, *Journal of Physical Sciences*, 22(1), 1–24.
- Abdurahman, N. H., Rosli, Y. M., Azhari, N. H., & Tam, S. F. (2011). Biomethanation of palm oil mill effluent (POME) by membrane anaerobic system (MAS) using POME as a substrate. *World Academy of Science, Engineering and Technology*, 75(2011), 419–424.
- Abouelenien, F., Namba, Y., Kosseva, M. R., Nishio, N., & Nakashimada, Y. (2014). Enhancement of methane production from co-digestion of chicken manure with agricultural wastes. *Bioresource Technology*, 159, 80–87.
- Ahmad, A. L., Chan, C. Y., Abd Shukor, S. R., & Mashitah, M. D. (2008). Recovery of oil and carotenes from palm oil mill effluent (POME). *Chemical Engineering Journal*, 141, 383–386.
- Ahmad, A. L., Ismail, S., & Bhatia, S. (2003). Water recycling from palm oil mill effluent (POME) using membrane technology. *Desalination*, 157(May), 87–95.
- Ahmad, A., Shah, S. M. U., Othman, M. F., & Abdullah, M. A. (2014a). Enhanced palm oil mill effluent treatment and biomethane production by co-digestion of oil palm empty fruit bunches with *Chlorella* sp. *The Canadian Journal of Chemical Engineering*, 92(9), 1636–1642.
- Ahmad, A., Shah, S. M. U., Othman, M. F., & Abdullah, M. A. (2014b). Aerobic and anaerobic co-cultivation of *Nannochloropsis oculata* with oil palm empty fruit bunch for enhanced biomethane production and palm oil mill effluent treatment. *Desalination and Water Treatment*, 1(November 2014), 37–41.
- Ahmad, M. N., Mokhtar, M. N., Baharuddin, A. S., Hock, L. S., Ali, S. R. A., Abd-Aziz, S., ... Hassan, M. A. (2011). Changes in physicochemical and microbial community during co-composting of oil palm frond with palm oil mill effluent anaerobic sludge. *Bioresources.com*, 6(4), 4762–4780.

- Ahmed Yassin, A.A. (2005). *Biohydrogen production from palm oil mill effluent by anaerobic fermentation*. Doctor of Philosophy Thesis, Universiti Putra Malaysia.
- Alam, M. Z., Kabbashi, N. A., & Hussin, S. N. I. S. (2009). Production of bioethanol by direct bioconversion of oil-palm industrial effluent in a stirred-tank bioreactor. *Journal of Industrial Microbiology & Biotechnology*, 36(6), 801–808.
- Ali, A. A. M., Zainudin, M. H. M., Idris, A., Baharuddin, A. S., Sulaiman, A., Matsui, T., ... Shirai, Y. (2012). Enhanced biogas production from palm oil mill effluent supplemented with untreated oil palm empty fruit bunch biomass with a change in the microbial community. *Japan Journal of Food Engineering*, 13(3), 37–41.
- Alves, M. M., Pereira, M. A., Sousa, D. Z., Cavaleiro, A. J., Picavet, M., Smidt, H., & Stams, A. J. M. (2009). Waste lipids to energy: How to optimize methane production from long-chain fatty acids (LCFA). *Microbial Biotechnology*, 2(5), 538–550.
- Amenaghawon, A. N., Asegame, P. A., & Obahiagbon, K. O. (2013). Potential application of urea and NPK 15:15:15 fertilizers as biostimulants in the bioremediation of domestic wastewater. *American Journal of Environmental Protection*, 1(4), 91–95.
- Anonymous. (2010). *Microbiology practical guide (A)*. Budapest, Hungary. Retrieved 2 May 2013, from http://mikrobiologia.elte.hu/?q=system/files/PRACTICAL_GUIDE-2012-A.pdf
- APHA (2005). *Standard methods for examination of water and wastewater* (21st ed.). APHA, AWWA, WPCF, Washington DC, USA.
- Arjoon, A., Olaniran, A. O., & Pillay, B. (2013). Chemosphere enhanced 1, 2-dichloroethane degradation in heavy metal co-contaminated wastewater undergoing biostimulation and bioaugmentation. *Chemosphere*, 93, 1826–1834.
- Astimar, A. A., Wan Hasamudin, W. H., Zawawi, I., Rosnah, M. S., Ropandi, M., & Anis, M. (2009). A pyro-ligno binder from oil palm empty fruit bunch. *MPOB Information Series*, (467), 1–2.
- ASTM. (2000). *Standard test methods for moisture, ash and organic matter of peat and other organic soils*. West Conshohocken, United States.

- Baharuddin, A. S., Hock, L. S., Md Yusof, M. Z., Abdul Rahman, N. A., Md Shah, U. K., Hassan, M. A., ... Shirai, Y. (2010). Effect of palm oil mill effluent (POME) anaerobic sludge from 500 m³ of closed anaerobic methane digested tank on pressed-shredded empty fruit bunch (EFB) composting process. *African Journal of Biotechnology*, 9(16), 2427–2436.
- Baharuddin, A. S., Kazunori, N., Abd-Aziz, S., Tabatabaei, M., Abdul Rahman, N. A., Hassan, M. A., ... Shirai, Y. (2009). Characteristics and microbial succession in co-composting of oil palm empty fruit bunch and partially treated palm oil mill effluent. *The Open Biotechnology Journal*, 3(1), 87–95.
- Bakar, N. A. (2014). *Country presentation on status of bioenergy development in Malaysia* (Power Point slides). Retrieved 6 February 2016 from <https://www.iea.org/media/technologyplatform/workshops/southeastasiabienergy2014/Malaysia.pdf>
- Bakar, N. K. A., Abd-Aziz, S., Hassan, M. A., & Ghazali, F. M. (2010). Isolation and selection of appropriate cellulolytic mixed microbial cultures for cellulases production from OPEFB. *Biotechnology*, 9(1), 73–78.
- Bala, J. D., Lalung, J., & Ismail, N. (2015). Studies on the reduction of organic load from palm oil mill effluent (POME) by bacterial strains. *International Journal of Recycling of Organic Waste in Agriculture*, 4, 1–10.
- Basri, M., Yacob, S., & Hassan, M. (2010). Improved biogas production from palm oil mill effluent by a scaled-down anaerobic treatment process. *World Journal of Microbiology and Biotechnology*, (26), 505–514.
- Beguin, P., & Aubert, J. P. (1994). The biological degradation of cellulose. *FEMS Microbiology Review*, 13 (1), 25-58.
- Benoliel, B., Torres, F. A. G., & de Moraes, L. M. P. (2013). A novel promising *Trichoderma harzianum* strain for the production of a cellulolytic complex using sugarcane bagasse in natura. *SpringerPlus*, 2(656), 1–7.
- Bento, F. M., Camargo, F. A. de O., Okeke, B., & Frankenberger-Junior, W. T. (2003). Bioremediation of soil contaminated by diesel oil. *Brazilian Journal of Microbiology*, 34(Suppl. 1), 65–68.
- Bhattacharyya, B.C. & Banerjee, R. (2007). Classification and characterization of wastes. In: *Environmental biotechnology*. Oxford University Press, India.
- Bhumibhamon, O., Kopraserstak, A., & Funthong, S. (2002). Biotreatment of high fat and oil wastewater by lipase producing microorganisms. *Kasetsart Journal. (Natural Science.)*, 267(36), 261–267.

- Biely, P. (1985). Microbial xylanolytic systems. *Trends in Biotechnology*, 3(11), 286-290.
- Biolog. (2008). GEN III MicroPlate. Biolog Inc.
- Bogner, J., Spokas, K., Burton, E., Sweeney, R., & Corona, V. (1995). Landfills as atmospheric methane sources and sinks. *Chemosphere*, 31(9), 4119–4130.
- Bonin, A. S., & Boone, D. R. (2006). The order *Methanobacteriales*. In M. Dworkin, S. Falkow, E. Rosenberg, K.-H. Schleifer, & E. Stackebrandt (Eds.), *The prokaryotes: Volume 3: Archaea. Bacteria: Firmicutes, Actinomycetes* (pp. 231–243). New York: Springer New York.
- Borja, R., Banks, C. J., & Sinchez, E. (1996). Anaerobic treatment of palm oil mill effluent in a two-stage up-flow anaerobic sludge blanket (UASB) system. *Journal of Biotechnology*, 45(1996), 125–135.
- Botheju, D., & Bakke, R. (2011). Oxygen effects in anaerobic digestion – A review. *The Open Waste Management*, 4, 1–19.
- Botheju, D., Samarakoon, G., Chen, C., & Bakke, R. (2010). An experimental study on the effects of oxygen in bio-gasification - Part 1. In *International Conference on Renewable Energies and Power Quality*. Granada, Spain.
- Braun, R., & Wellinger, A. (2002). *Potential of Co-digestion*. IEA Bioenergy.
- Burdass, D., Grainger, J., & Hurst, J. (Eds.). (2006). *Basic practical microbiology, a manual*. Society for General Microbiology (SGM). United Kingdom. Retrieved 6 February 2016, from http://www.microbiologyonline.org.uk/media/transfer/doc/sgm_basic_practical_microbiology_2.pdf
- Burke D.A. (2001). *Options for recovering beneficial products from dairy manure: Dairy waste anaerobic digestion handbook*. Olympia, WA: Environmental Energy Company.
- Busu, Z., Sulaiman, A., Hassan, M. A., Shirai, Y., Abd-Aziz, S., Yacob, S., & Wakisaka, M. (2010). Improved anaerobic treatment of palm oil mill effluent in a semi-commercial closed digester tank with sludge recycling and appropriate feeding strategy. *Pertanika Journal of Tropical Agricultural Science*, 33(1), 27–37.
- Buswell, A. M., & Mueller, H. F. (1952). Mechanism of methane fermentation. *Industrial and Engineering Chemistry*, 44(3), 550–552.

- Camarillo, M. K., Stringfellow, W. T., Spier, C. L., Hanlon, J. S., & Domen, J. K. (2013). Impact of co-digestion on existing salt and nutrient mass balances for a full-scale dairy energy project. *Journal of Environmental Management*, 128, 233–242.
- Casper, J. K. (2010). *Greenhouse Gases: Worldwide Impacts: Facts On File*.
- Cavaleiro, A. J., Sousa, D. Z., & Alves, M. M. (2010). Bioaugmentation of anaerobic sludge with *Syntrophomonas zehnderi* as a prospect for enhanced methane production from oleate. In *International Water Association (IWA) 12th. World Congress on Anaerobic Digestion*.
- Chai, E. W., H'ng, P. S., Peng, S. H., Wan-Azha, W. M., Chin, K. L., Chow, M. J., & Wong, W. Z. (2013). Compost feedstock characteristics and ratio modelling for organic waste materials co-composting in Malaysia. *Environmental Technology*, 34(20), 2859–2866.
- Chaisri, R., Boonsawang, P., & Prasertsan, P. (2007). Effect of organic loading rate on methane and volatile fatty acids productions from anaerobic treatment of palm oil mill effluent in UASB and UFAF reactors. *Songklanakarin J. Sci. Technol.*, 29(April 2006), 311–323.
- Chan, Y. J., Chong, M. F., & Law, C. L. (2012). An integrated anaerobic – aerobic bioreactor (IAAB) for the treatment of palm oil mill effluent (POME): Start-up and steady state performance. *Process Biochemistry*, 47, 485–495.
- Chen, B., Yen, C., & Hsueh, C. (2009). Cost-effective biostimulation strategy for wastewater decolorization using immobilized-cell systems. *Bioresource Technology*, 100, 2975–2981.
- Chen, Y., Cheng, J. J., & Creamer, K. S. (2008). Inhibition of anaerobic digestion process: A review. *Bioresource Technology*, 99(10), 4044–4064.
- Chernicharo, C. A. L. (2007). *Anaerobic Reactors: IWA Publishing*.
- Chernoglazov, V. M., Ermolova, O. V., & Klyosov, A. A. (1988). Adsorption of high-purity endo-1,4- β -glucanases from *Trichoderma reesei* on components of lignocellulosic materials: cellulose, lignin, and xylan. *Enzyme and Microbial Technology*, 10, 503–507.
- Chin, K.L., H'ng, P.S., Wong, L.J., Tey, B.T., & Paridah, M.T. (2011). Production of glucose from oil palm trunk and sawdust of rubberwood and mixed hardwood. *Applied Energy*, 88(11), 4222–4228.
- Chin, M. J., Poh, P. E., Tey, B. T., Chan, E. S., & Chin, K. L. (2013). Biogas from palm oil mill effluent (POME): Opportunities and challenges from Malaysia's perspective. *Renewable and Sustainable Energy Reviews*, 26, 717–726.

- Chin, S. X., Chia, C. H., & Zakaria, S. (2013). Production of reducing sugar from oil palm empty fruit bunch (EFB) cellulose fibres via acid hydrolysis. *Bioresources.com*, 8(1), 447–460.
- Choorit, W., & Wisarnwan, P. (2007). Effect of temperature on the anaerobic digestion of palm oil mill effluent. *Electronic Journal of Biotechnology*, 10(3), 376–385.
- Collins, C. H., & Beale, A. J. (2015). *Safety in industrial microbiology and biotechnology*. Elsevier Science.
- Csuros, M. & Csaba, C. (1999). *Microbial examination of water and wastewater*. Lewis Publisher.
- Daoud, J. I., & Alam, M. Z. (2010). Statistical optimization of fermentation conditions for cellulase production from palm oil mill effluent. *American Journal of Environmental Sciences*, 6(1), 66–70.
- Davies, P. S. (2005). *The biological basis of wastewater treatment*. Glasgow, UK: Strathkelvin Instruments Ltd.
- Delzer, G. C., & McKenzie, S. W. (2003). Five-day biochemical oxygen demand. In *U.S. geological survey techniques of water-resources investigations* (Vol. 7, pp. 1–21). Retrieved 6 February 2016, from <http://pubs.water.usgs.gov/twri9A/>
- Demirbas, A. (2008). *Biofuels: Securing the planet's future energy needs* (pp. 86-102). London: Springer.
- Demirel, B., & Scherer, P. (2008). Production of methane from sugar beet silage without manure addition by a single-stage anaerobic digestion process. *Biomass and Bioenergy*, 32(3), 203–209.
- Department of Environment, (1999). *Industrial processes & the environment (Handbook no. 3) crude palm oil industry*. Malaysia: Aslita Sdn. Bhd.
- Deublein, D., & Steinhauser, A. (2008). *Biogas from waste and renewable resources: An introduction*. Germany: Wiley-VCH Verlag GmbH & Co. KGaA.
- Duran, M., Tepe, N., Yurtsever, D., Punzi, V.L., Bruno, C., Mehta, R.J., 2006. Bioaugmenting anaerobic digestion of biosolids with selected strains of *Bacillus*, *Pseudomonas*, and *Actinomycetes* species for increased methanogenesis and odor control. *Applied Microbiology and Biotechnology*, 73, 960–966.
- Economic Planning Unit (EPU). (2000). Eighth Malaysia plan- Energy. *Eighth Malaysia Plan 2001-2005*, 303–336.

- Economy Planning Unit (EPU). (2006). Ninth Malaysia plan-Sustainable energy development. *Ninth Malaysia Plan 2006-2010*, 393–411.
- Emmyrafedziawati, A. K. R. (2013). Identification and characterisation of *Bacillus subtilis* as cellulase-producing bacteria. *Journal of Tropical Agriculture and Food Science*, 41(2), 319–327.
- Environmental Quality Act Regulations 1974 (2000). *Environmental Quality (Sewage and Industrial Effluents) (Amendment) Regulations 2000*. Percetakan Nasional Malaysia Bhd., Kuala Lumpur, Malaysia.
- Eriksen, J., Andersen, A. J., Poulsen, H. V, Adamsen, A. P. S., & Petersen, S. O. (2012). Sulfur turnover and emissions during storage of cattle slurry: Effects of acidification and sulfur addition. *Journal of Environmental Quality*, 41, 1633–1642.
- ETP Annual Report (2014). Palm Oil and Rubber.
- Faisal, M., & Unno, H. (2001). Kinetic analysis of palm oil mill wastewater treatment by a modified anaerobic baffled reactor. *Biochemical Engineering Journal*, 9(1), 25–31.
- Fang, C., O-thong, S., Boe, K., & Angelidaki, I. (2011). Comparison of UASB and EGSB reactors performance, for treatment of raw and deoiled palm oil mill effluent (POME). *Journal of Hazardous Materials*, 189, 229–234.
- Fang, H. H. P., & Yu, H. Q. (2000). Effect of HRT on mesophilic acidogenesis of dairy wastewater. *Journal of Environmental Engineering*, 126(December), 1145–1148.
- Fannin, K.F. (1987). Start-up, operation, stability, and control. In D.P. Chynoweth, & I. Ron (Eds.), *Anaerobic digestion of biomass* (pp. 171-196). England: Elsevier Applied Science.
- FAO (Food and Agriculture Organization) (n.d.). *Biogas process for sustainable development*. Retrieved 23 February 2015, from <http://www.fao.org/docrep/t0541e0c.htm>
- Fletcher, L. E. (1998). *Potential explosive hazards from hydrogen sulfide production in ship ballast and sewage tanks*. Melbourne, Australia: DSTO Aeronautical and Maritime Research Laboratory.
- Florentino, H. D. O. (2003). Mathematical tool to size rural digesters. *Scientia Agricola*, 60(1), 185–190.
- Gerardi, M. H. (2003). *The microbiology of anaerobic digesters*. Wiley-Interscience. New Jersey: John Wiley & Sons, Inc.

- Girardin, H., Monod, M., Latgé, J. P., & Ge, L. (1995). Molecular characterization of the food-borne fungus *Neosartorya fischeri* (Malloch and Cain). *Applied and Environmental Microbiology*, 61(4), 1378–1383.
- Gray, K. A., Zhao, L., and Emptage, M. (2006). *Current Opinion in Chemical Biology*, 10(2), 141-146.
- Habib, M. A. B., Yusoff, F. M., Phang, S. M., Ang, K. J., & Mohamed, S. (1997). Nutritional values of chironomid larvae grown in palm oil mill effluent and algal culture. *Aquaculture*, 158, 95–105.
- Hansen, S. B., Olsen, S. I., & Ujang, Z. (2012). Greenhouse gas reductions through enhanced use of residues in the life cycle of Malaysian palm oil derived biodiesel. *Bioresource Technology*, 104, 358–366.
- Hendricks, D., Vos, P. D. & Whitman, W. B. (2009). *The Firmicutes* (2nd ed., Vol. 3). Dordrecht: Springer.
- Herrero, M., & Stuckey, D. C. (2015). Bioaugmentation and its application in wastewater treatment : A review. *Chemosphere*, 140, 119–128.
- Hickey, R. F., & Switzenbaumt, M. S. (1990). Behavior of carbon monoxide as a trace component of anaerobic digester gases and methanogenesis from acetate. *Environ. Sci. Technol.*, 24(11), 1642–1648.
- Hock, L. S., Baharuddin, A. S., Ahmad, M. N., Shah, U. K. M., Rahman, N. A. A., Abd-Aziz, S., Hassan M. A., & Shirai, Y. (2009). Physicochemical changes in windrow co-composting process of oil palm mesocarp fiber and palm oil mill effluent anaerobic sludge. *Australian Journal of Basic and Applied Sciences*, 3(3), 2809–2816.
- Houde, A., Kademi, A., & Leblanc, D. (2004). Lipases and their industrial applications an overview. *Applied Biochemistry and Biotechnology*, 118, 155–170.
- Howard, R. L., Abotsi, E., Jansen van Rensburg, E. L., & Howard, S. (2003). Lignocellulose biotechnology: Issues of bioconversion and enzyme production. *African Journal of Biotechnology*, 2(12), 602-619.
- Husain, Z., Zainac, Z., & Abdullah, Z. (2002). Briquetting of palm fibre and shell from the processing of palm nuts to palm oil. *Biomass & Bioenergy*, 22, 505–509.
- Hwang T. K., Ong S. M., Seow C. C., & Tan H. K. (1978). Chemical composition of palm oil mill effluents. *Planter*, 54, 749–756.
- Igwe, J. C., & Onyegbado, C. C. (2007). A review of palm oil mill effluent (POME) water treatment. *Global Journal of Environmental Research*, 1(2): 54-62.

- Irvan, Trisakti, B., Wongistani, V., & Tomiuchi, Y. (2012). Methane emission from digestion of palm oil mill effluent (POME) in a thermophilic anaerobic reactor. *International Journal of Science and Engineering*, 3(April), 32–35.
- Isa, Z. (1991). Fundamental of anaerobic waste treatment technology. In Yeoh, B.G. (Ed.), *Anaerobic digestion technology in pollution control: Proceeding of the Workshop on Anaerobic Digestion Technology in Pollution Control held on 5-7 August 1991 at SIRIM*. Shah Alam, Selangor.
- Jamal, P., Alam, M., & Mohamad, A. (2007). Microbial bioconversion of palm oil mill effluent to citric acid with optimum process conditions. In F. Ibrahim, N. A. A. Osman, J. Usman, & N. A. Kadri (Eds.), *3rd Kuala Lumpur International Conference on Biomedical Engineering* (Vol. 15, pp. 483–487). Kuala Lumpur, Malaysia: Springer Science & Business Media.
- Janani, P., Sudharsana, J. & Ravikumar, R. (2013). Performance of aqueous two phase system for the extraction of lipase from rice bran. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 4(3), 754-760.
- Jearat, K., & Suksaroj, T. T. (2014). Enhancing of methane production from palm oil mill effluent by co-digestion with empty fruit bunch pressed wastewater at mesophilic condition using an anaerobic batch reactor. In *ThellIER-SCIENCE PLUS International Conference* (pp. 28–30). Singapore.
- Jeffries, T. W. (1994). Biodegradation of lignin and hemicelluloses. *Biochemistry of Microbial Degradation*, 233–277.
- Jeoung, J., Fessler, J., Goetzl, S., & Dobbek, H. (2014). Carbon monoxide. Toxic gas and fuel for anaerobes and aerobes: Carbon monoxide dehydrogenases. In P. M. H. Kroneck & M. E. S. Torres (Eds.), *The metal-driven biogeochemistry of gaseous compounds in the environment* (pp. 37–65). Berlin, Germany: Springer.
- Johansen, J. E., & Bakke, R. (2006) Enhancing hydrolysis with micro- aeration. *Water Science and Technology*, 53(8), 43-50.
- Jørgensen, P. J. (2009). Biogas: green energy. PlanEnergi and Researcher for a Day. Faculty of Agricultural Sciences, Aarhus University.
- Jung, H. G., & Vogel, K. P. (1986). Influence of lignin in digestibility of forage cell wall material. *J. Anim. Sci.*, 62, 1703-1712.
- Kalle, G. P., Nayak, K. K., & Sa, C. D. E. (1985). An approach to improve methanogenesis through the use of mixed cultures isolated from biogas digester. *Journal of Biosciences*, 9(December), 137–144.

- Kananam, W., Tachapattaworakul, T., & Suksaroj, C. (2011). Biochemical changes during oil palm (*Elaeis guineensis*) empty fruit bunches composting with decanter sludge and chicken manure. *Science Asia*, 37(2011), 17–23.
- Karigar, C. S., & Rao, S. S. (2011). Role of microbial enzymes in the bioremediation of pollutants: A review. *Enzyme Research*, 2011(Figure 1), 1–11.
- Karim, A. N. M., Hamdan, M. A. H., Alam, M. Z., & Iqbal, M. (2012). Economic scenario of cellulase enzyme production using palm oil mill effluent (POME). *Advanced Materials Research*, 576, 723–726.
- Kelly, W.J., Asmundson, R.V., Harisson, G.L. & Huang, C.M. (1995). Differentiation of dextran-producing *Leuconostoc* strains from fermented rice cake (puto) using pulsed field gel electrophoresis. *International Journal of Food Microbiology*, 26, 345-352.
- Kendall, M. M., & Boone, D. R. (2006). The order *Methanosarcinales*. In M. Dworkin, S. Falkow, E. Rosenberg, K.-H. Schleifer, & E. Stackebrandt (Eds.), *The prokaryotes: Volume 3: Archaea. Bacteria: Firmicutes, Actinomycetes* (pp. 244–256). New York: Springer New York.
- Kenealy, W., Cao, Y., & Weimer, P. (1995). Production of caproic acid by cocultures of ruminal cellulolytic bacteria and *Clostridium kluyveri* grown on cellulose and ethanol. *Applied Microbiology and Biotechnology*, 44(3-4), 507–513.
- KeTTHA. (2009). *National Green Technology Policy*. Malaysia, Kementerian Tenaga, Teknologi Hijau dan Air.
- Khemkhao, M., Nuntakumjorn, B., Techkarnjanaruk, S., & Phalakornkule, C. (2011). Microbial diversity in thermophilic adaptation on POME treatment. *KMITL Science and Technology Journal*, 11(1), 1–8.
- Khemkhao, M., Nuntakumjorn, B., Techkarnjanaruk, S., & Phalakornkule, C. (2012). UASB performance and microbial adaptation during a transition from mesophilic to thermophilic treatment of palm oil mill effluent. *Journal of Environmental Management*, 103, 74–82.
- Kiener, A. & Leisinger, T. (1983). Oxygen sensitivity of methanogenic bacteria. *Systematic and Applied Microbiology*, 4, 305-312.
- Koch, K., Plabst, M., Schmidt, A., Helmreich, B., & Drewes, J. E. (2015). Co-digestion of food waste in a municipal wastewater treatment plant: Comparison of batch tests and full-scale experiences. *Waste Management*.

- Korsten, L., & Cook, N. (1996). Optimizing culturing conditions for *Bacillus subtilis*. *South African Avocado Growers' Association Yearbook*, 19, 54–58.
- Kuhad, R., Gupta, R., & Singh, A. (2011). Microbial cellulases and their industrial applications. *Enzyme Research*, 2011, 1-11.
- Kumar, D., Kumar, L., Nagar, S., Raina, C., Parshad, R., & Gupta, V. K. (2012). Screening, isolation and production of lipase/esterase producing *Bacillus* sp. strain DVL2 and its potential evaluation in esterification and resolution reactions. *Archives of Applied Science Research*, 4(4), 1763–1770.
- Labatut, R. A, Angenent, L. T., & Scott, N. R. (2011). Biochemical methane potential and biodegradability of complex organic substrates. *Bioresource Technology*, 102(3), 2255–64.
- Lebeau, T. (2011). Bioaugmentation for in situ soil remediation: How to ensure the success of such a process. In Singh, A., Parmar, N., & Kuhad, R. C. (Eds.), *Bioaugmentation, biostimulation and biocontrol* (pp. 129-186). Springer-Verlag Berlin Heidelberg.
- Lehtomaki, A., Huttunen, S., & Rintala, J. A. (2007). Laboratory investigations on co-digestion of energy crops and crop residues with cow manure for methane production : Effect of crop to manure ratio. *Resources Conversion & Recycling*, 51, 591–609.
- Lerm, S., Kleyböcker, A., Miethling-Graff, R., Alawi, M., Kasina, M., Liebrich, M., & Würdemann, H. (2012). Archaeal community composition affects the function of anaerobic co-digesters in response to organic overload. *Waste Management*, 32(3), 389–399.
- Li, Q. X. (2009). *Bioremediation* (Power Point slides). Retrieved 21 July 2016, from http://www.ctahr.hawaii.edu/scau/downloads/2009_0728_Li2.pdf
- Li, Q., Ng, W. T., & Wu, J. C. (2014). Isolation, characterization and application of a cellulose-degrading strain *Neurospora crassa* S1 from oil palm empty fruit bunch. *Microbial Cell Factories*, 13(1), 157.
- Lindorfer H., Pérez López C., Resch C., Braun R. & Kirchmayr R. (2007). The impact of increasing energy crop addition on process performance and residual methane potential in anaerobic digestion. *Water Science and Technology*, 56, 55-63.
- Lu, K.-M., Lee, W.-J., Chen, W.-H., Liu, S.-H., & Lin, T.-C. (2012). Torrefaction and low temperature carbonization of oil palm fiber and eucalyptus in nitrogen and air atmospheres. *Bioresource Technology*, 123, 98–105.

- Lyberatos, G., & Skiadas, I. (1999). Modelling of anaerobic digestion: A review. *Global Nest International Journal*, 1(2), 63–76.
- Lynd, L. R., Weimer, P. J., van Zyl, W. H., & Pretorius, I. S. (2002). Microbial cellulose utilization: Fundamentals and biotechnology. *Microbiology and Molecular Biology Reviews*, 66(3), 506–577.
- Ma A.N., Toh T.S., & Chua N.S. (2000). Renewable energy from oil palm industry. Oil palm and the environment: A Malaysian perspective. *Malaysia Oil Palm Growers' Council*, 113–26.
- Mamun, A. A., & Idris, A. (2008). Treatment of POME by Pilot Plant Anaerobic Fluidised Bed Reactor. *IIUM Engineering Journal*, 9(1), 9–18.
- Mancera-López, M. E., Esparza-García, F., Chávez-Gómez, B., Rodríguez-Vázquez, R., Saucedo-Castañeda, G., & Barrera-Cortés, J. (2008). Bioremediation of an aged hydrocarbon-contaminated soil by a combined system of biostimulation–bioaugmentation with filamentous fungi. *International Biodeterioration & Biodegradation*, 61(2), 151–160.
- Martin-Ryals, A. D. (2012). *Evaluating the potential for improving anaerobic digestion of cellulosic waste via routine bioaugmentation and alkaline pretreatment*. Master Thesis, University of Illinois. Retrieved 8 December 2015, from https://www.ideals.illinois.edu/bitstream/handle/2142/34285/Martin-Ryals_Ana.pdf?sequence=1
- McCarty, P.L. (1964). Anaerobic waste treatment fundamentals. *Public Works*, 95 (9), 107–112.
- Merlin Christy, P., Gopinath, L. R., & Divya, D. (2014). A review on anaerobic decomposition and enhancement of biogas production through enzymes and microorganisms. *Renewable and Sustainable Energy Reviews*, 34, 167–173.
- Michaud, S., Bernet, N., Buffiere, P., Roustan, M., & Moletta, R. (2002). Methane yield as a monitoring parameter for the start-up of anaerobic fixed film reactors. *Water Research*, 36, 1385–1391.
- Montero, B., Garcia-Morales, J.L., Sales, D. & Solera, R (2008). Evolution of microorganisms in thermophilic-dry anaerobic digestion. *Bioresource Technology*, 99, 3233–3243.
- MPOB (2010). Seminar and workshop on the palm oil mill effluent tertiary treatment technology (POMET3). Kota Kinabalu, Sabah, 5-6 July 2010.
- MPOB (2014a). *National biogas implementation (EPP5), Biogas capture and CDM project implementation for palm oil mills*.

MPOB (2014b). *Oil Palm and the Environment* (updated March 2014). Retrieved 11 Jun 2015, from <http://www.mpob.gov.my/en/palm-info/environment/520.achievements> on 5/12/2014.

MPOB (2015). *Overview of the Malaysian oil palm industry 2014*.

MPOB. (n.d.). *About palm oil*. Retrieved 21 July 2016, from http://www.palmoilworld.org/about_palmoil.html on 14/7/2015.

MPOC (n.d). Palm oil and the environment. *Journal Oil Palm & Environment*. Retrieved from http://www.mpoc.org.my/Palm_Oil_and_The_Environment.aspx on 12/12/2011.

Muryanto, Sahlan, M., & Sudiyani, Y. (2012). Simultaneous saccharification and fermentation of oil palm empty fruit bunch for bioethanol production by *Rhizopus oryzae*. *International Journal of Environment and Bioenergy*, 3(2), 111–120.

Musa, H., & Adebayo-tayo, B. C. (2012). Screening of microorganisms isolated from different environmental samples for extracellular lipase production. *AU Journal of Technology*, 15(3), 179–186.

Nahrul Hayamin, Z., Astimar, A. A., Anis, M., Ibrahim, M. H., Khalil, H. P. S. A., & Ibrahim, Z. (2012). Vermicomposting of empty fruit bunch with addition of palm oil mill effluent solid. *Journal of Oil Palm Research*, 1542–1549.

Najafpour, G. D., Zinatizadeh, A. A. L., Mohamed, A. R., Hasnain Isa, M., & Nasrollahzadeh, H. (2006). High-rate anaerobic digestion of palm oil mill effluent in an upflow anaerobic sludge-fixed film bioreactor. *Process Biochemistry*, 41, 370–379.

Nakashimada, Y., Srinivasan, K., Murakami, M., & Nishio, N. (2000). Direct conversion of cellulose to methane by anaerobic fungus *Neocallimastix frontalis* and defined methanogens. *Biotechnology Letters*, 22, 223–227.

Nasir, A., Bala, K. C., Mohammed, S. N., Mohammed, A., & Umar, I. (2015). Experimental investigation on the effects of digester size on biogas production from cow dung. *American Journal of Engineering Research*, 4(1), 181–186.

Navarro, S. S., Cimpola, R., Bruant, G., & Guiot, S. R. (2014). Specific inhibitors for identifying pathways for methane production from carbon monoxide by a nonadapted anaerobic mixed culture. *Canadian Journal of Microbiology*, 60(6), 407–415.

- Nayono, S. E., Gallert, C., & Winter, J. (2010). Co-digestion of press water and food waste in a biowaste digester for improvement of biogas production. *Bioresource Technology*, 101, 6987–6993.
- Ng, F.-Y., Yew, F.-K., Basiron, Y., & Sundram, K. (2011). A renewable future driven with Malaysian palm oil-based green technology. *Journal of Oil Palm & the Environment*, 2(January), 1–7.
- Nges, I. A. (2012). *Anaerobic digestion of crop and waste biomass : Impact of feedstock characteristics on process performance*. Lund University, Sweden.
- Nieves, D. C., Karimi, K., & Horvath, I. S. (2011). Improvement of biogas production from oil palm empty fruit bunches (OPEFB). *Industrial Crops and Products*, 34(1), 1097–1101.
- Nwoko, C. O., & Ogunyemi, S. (2010). Evaluation of palm oil mill effluent to maize (*Zea mays*. L) crop: Yields, tissue nutrient content and residual soil chemical properties. *Australian Journal of Crop Sciences*, 4(1), 16–22.
- Ohimain, E. I., Daokoru-Olukole, C., Izah, S. C., Eke, R. A., & Okonkwo, A. C. (2012). Microbiology of palm oil mill effluents. *Journal of Microbiology and Biotechnology Research*, 2(6), 852–857.
- Okwute, Loretta, O., & Isu, N. (2006). Impact analysis of palm oil mill effluent on the aerobic bacterial density and ammonium oxidizers in a dumpsite in Anyigba, Kogi State, 38–41.
- Olaniran, A. O., Pillay, D., & Pillay, B. (2006). Biostimulation and bioaugmentation enhances aerobic biodegradation of dichloroethenes. *Chemosphere*, 63(4), 600–8.
- O-Thong, S., Boe, K., & Angelidaki, I. (2012). Thermophilic anaerobic co-digestion of oil palm empty fruit bunches with palm oil mill effluent for efficient biogas production. *Applied Energy*, 93, 648–654.
- Owens, J.F. & Chynoweth D. P., (1993). Biochemical methane potential of MSW components. *Water Science and Technology*, 27: 1-14.
- Park, D., Lee, D. S., Kim, Y. M., & Park, J. M. (2008). Bioaugmentation of cyanide-degrading microorganisms in a full-scale cokes wastewater treatment facility. *Bioresource Technology*, 99(6), 2092–2096.
- Peng, X., Börner, R. A., Nges, I. A., & Liu, J. (2014). Impact of bioaugmentation on biochemical methane potential for wheat straw with addition of *Clostridium cellulolyticum*. *Bioresource Technology*, 152, 567–571.

- Poh, P. E., Yong, W.-J., & Chong, M. F. (2010). Palm oil mill effluent (POME) characteristic in high crop season and the applicability of high-rate anaerobic bioreactors for the treatment of POME. *Industrial and Engineering Chemistry Research*, 49, 11732–11740.
- Poh, P.E. & Chong, M.F. (2009). Development of anaerobic digestion methods for palm oil mill effluent (POME) treatment. *Bioresource Technology*, 100, 1-9.
- Ponnambalam, A. S., Deepthi, R. S., & A.R. Ghosh. (2011). Qualitative display and measurement of enzyme activity of isolated cellulolytic bacteria. *Biotechnology, Bioinformatics and Bioengineering*, 1(1), 33–37.
- Priest, F. G., Goodfellow, M. & Todd, C. (1988). A numerical classification of the genus *Bacillus*. *Journal of General Microbiology*, 134, 1847–1882.
- Rahman, S. H. A., Choudhury, J. P., Ahmad, A. L., & Kamaruddin, A. H. (2007). Optimization studies on acid hydrolysis of oil palm empty fruit bunch fiber for production of xylose. *Bioresource Technology*, 98, 554–559.
- Rai, B. & Upadhyay, R.S. (1982). The perfect state of *Fusarium udum*. *Mycologia*, 74(2), 343-346.
- Ram, C.S.V. (1957). Studies on cellulolytic activity of *Fusaria* with references to bacterial and other cellulose substrates. *Proceedings of the National Institute of Sciences of India*, 22(4), 204-211.
- Rashid, S. S., Alam, M. Z., Abdulkarim, M. I., & Salleh, M. H. (2009). Optimization of the nutrient supplements for cellulase production with the basal medium palm oil mill effluent. *World Academy of Science, Engineering and Technology*, 60, 809–815.
- Razuan, R., Finney, K. N., Chen, Q., Sharifi, V. N., & Swithenbank, J. (2011). Pelletised fuel production from palm kernel cake. *Fuel Processing Technology*, 92, 609-615.
- Rooney, A. P., Price, N. P. J., Ehrhardt, C., Swezey, J. L., & Bannan, J. D. (2009). Phylogeny and molecular taxonomy of the *Bacillus subtilis* species complex and description of *Bacillus subtilis* subsp. *inaquosorum* subsp. nov. *International Journal of Systematic and Evolutionary Microbiology*, 59, 2429–2436.
- Ryckebosch, E., Drouillon, M., & Vervaeren, H. (2011). Techniques for transformation of biogas to biomethane. *Biomass and Bioenergy*, 35(4), 1633–1645.

- Safitri, R., Septyana, D. V., Rossiana, N., Sri, R. R., & Amien, S. (2013). Biodegradation waste oil palm empty fruit bunch (*Elaeis guineensis* Jacq.) by lignocellulolytic fungi. *AgroLife Scientific Journal*, 2(2), 54–59.
- Salihu, A., Alam, M. Z., AbdulKarim, M. I., & Salleh, H. M. (2011). Suitability of using palm oil mill effluent as a medium for lipase production. *African Journal of Biotechnology*, 10(11), 2044–2052.
- Senawi, R., Alauddin, S. M., Saleh, R. M., & Shueb, M. I. (2013). Polylactic acid/empty fruit bunch fiber biocomposite: influence of alkaline and silane treatment on the mechanical properties. *International Journal of Bioscience, Biochemistry and Bioinformatics*, 3(1), 59–61.
- Shadab, M. (2009). Scale-up production of cellulase from palm oil mill effluent (POME) using 30 liter and 300 liter bioreactors. *Biotechnology Department IIUM*.
- Shahriarinnour, M., Wahab, M. N. A., Ariff, A., & Mohamad, R. (2015). Screening, isolation and selection of cellulolytic fungi from oil palm empty fruit bunch fibre. *Biotechnology*, 10(1), 108–113.
- Silva, E., Fialho, A.M., Sa-Correia, I., Burns, R.G., & Shaw, L.J. (2004). Combined bioaugmentation and biostimulation to cleanup soil contaminated with high concentrations of atrazine. *Environmental Science and Technology*, 38, 632.
- Simon, M. A., Bonner, J. S., Page, C. A., Townsend, R. T., Mueller, D. C., Fuller, C. B., & Autenrieth, R. L. (2004). Evaluation of two commercial bioaugmentation products for enhanced removal of petroleum from a wetland. *Ecological Engineering*, 22, 263–277.
- Singh, A., Billingsley, K., & Ward, O. (2006) Composting: A potentially safe process for disposal of genetically modified organisms. *Critical Reviews in Biotechnology*, 26, 1–16
- Singh, A., Parmar, N., Kuhad, R. C., & Ward, O. P. (2011). Bioaugmentation, biostimulation, and biocontrol in soil biology. In Singh, A., Parmar, N., & Kuhad, R. C. (Eds.), *Bioaugmentation, biostimulation and biocontrol* (pp. 1–26). Springer-Verlag Berlin Heidelberg.
- Sovacool, B. K., & Drupady, I. M. (2011). Examining the small renewable energy power (SREP) program in Malaysia. *Energy Policy*, 39(11), 7244–7256.
- Sprocati, A. R., Alisi, C., Tasso, F., Marconi, P., Sciullo, A., Pinto, V., Cremisini, C. (2012). Effectiveness of a microbial formula, as a bioaugmentation agent, tailored for bioremediation of diesel oil and heavy metal co-contaminated soil. *Process Biochemistry*, 47(11), 1651–1657.

- Stichnothe, H., & Schuchardt, F. (2010). Comparison of different treatment options for palm oil production waste on a life cycle basis. *The International Journal of Life Cycle Assessment*, 15(9), 907–915.
- Subramaniam, V., Ngan, M. A., May, C. Y., & Nik Meriam, N. S. (2008). Environmental performance of the milling process of Malaysian palm oil using the life cycle assessment approach. *American Journal of Environmental Sciences*, 4(4), 310–315.
- Sulaiman, A., Busu, Z., Tabatabaei, M., Yacob, S., Abd-aziz, S., Hassan, M. A., & Shirai, Y. (2009). The effect of higher sludge recycling rate on anaerobic treatment of palm oil mill effluent in a semi-commercial closed digester for renewable energy. *American Journal of Biochemistry and Biotechnology*, 5(1), 1–6.
- Sulaiman, A., Tabatabaei, M., & Mohd Yusoff, M. Z. (2010). Accelerated start-up of a semi-commercial digester tank treating palm oil mill effluent with sludge seeding for methane production. *World Applied Sciences Journal*, 8(2), 247–258.
- Sulthana, A., Latha, K. C., Balasubramaniam, S. (2016). Anaerobic digestion of wastewater and municipal sludge to isolate potent methanogen using BMP assay. *Integrated Waste Management in India*, 65-82.
- Sun, L. (2015). *Biogas production from lignocellulosic materials*. Doctoral Thesis, Swedish University of Agricultural Sciences, Uppsala.
- Sustainable Energy Development Authority Malaysia (SEDA) (2015). *Annual Report 2014*.
- Tabatabaei, M., Zakaria, M. R., Rahim, R. A., Wright, A. G., Shirai, Y., Abdullah, N., ... Hassan, M. A. (2009). PCR-based DGGE and FISH analysis of methanogens in an anaerobic closed digester tank for treating palm oil mill effluent. *Electronic Journal of Biotechnology*, 12(3). Retrieved 24 Jun 2015, from <http://www.ejbiotechnology.info/content/vol12/issue3/full/4/index.html>
- Tong, S. L., & Jaafar, A. B. (2004). Waste to energy: Methane recovery from anaerobic digestion of POME. *Energy Smart*, 14, 4–11.
- Torkian, A., Egbali, A. & Hashemian, S.J. (2003). The effect of organic loading rate on the performance of UASB reactor treating slaughterhouse effluent. *Resources, Conservation and Recycling*, 40, 1-11.
- Triolo, J. M., Sommer, S. G., Møller, H. B., Weisbjerg, M. R., & Jiang, X. Y. (2011). A new algorithm to characterize biodegradability of biomass during anaerobic digestion: Influence of lignin concentration on methane production potential. *Bioresource Technology*, 102(20), 9395–9402.

- Tyagi, M., da Fonseca, M. M. R., & de Carvalho, C. C. C. R. (2011). Bioaugmentation and biostimulation strategies to improve the effectiveness of bioremediation processes. *Biodegradation*, 22(2), 231–241.
- Uemura, Y., Omar, W. N., Tsutsui, T., & Yusup, S. (2011). Torrefaction of oil palm wastes. *Fuel*, 90(8), 2585–2591.
- Ueno, A., Ito, Y., Yumoto, I., & Okuyama, H. (2007). Isolation and characterization of bacteria from soil contaminated with diesel oil and the possible use of these in autochthonous bioaugmentation. *World Journal of Microbiology and Biotechnology*, 23(12), 1739–1745.
- Ugwu, K., & Agbo, K. (2013). Evaluation of binders in the production of briquettes from empty fruit bunches of *Elais Guinensis*. *International Journal of Renewable and Sustainable Energy*, 2(4), 176–179.
- Vavilin, V. A., Rytov, S. V., & Lokshina, L. Y. (1996). A description of hydrolysis kinetics in anaerobic degradation of particulate organic matter. *Bioresource Technology*, 56, 229–237.
- Vega, G. C. (2012). *Environmental impacts of alternative co-substrates for biogas production*. Master Thesis, Swedish University of Agricultural Sciences, Uppsala. Retrieved 17 February 2016, from <http://stud.epsilon.slu.se/4785/>
- Vijaya, S., Chow, M. C. & Ma A. N. (2004). Energy database of the oil palm. *MPOB Palm Oil Engineering Bulletin*, 70, 15-22.
- Wang, M., Li, Z., Fang, X., Wang, L., & Qu, Y. (2012). Cellulolytic enzyme production and enzymatic hydrolysis for second-generation bioethanol production. In *Biotechnology in China III: Biofuels and bioenergy* (pp. 1–24). Springer-Verlag Berlin Heidelberg.
- Wang, Q. H., Kuninobu, M., Ogawa, H.I., & Kato, Y. (1999). Degradation of volatile fatty acids in highly efficient anaerobic digestion. *Biomass & Bioenergy*, 16, 407–416.
- Wang, X., Lu, X., Li, F., & Yang, G. (2014). Effects of temperature and carbon–nitrogen (C/N) ratio on the performance of anaerobic co-digestion of dairy manure, chicken manure and rice straw: focusing on ammonia inhibition. *PLoS One*, 9(5), e97265.
- Wenderoth, D. F., Rosenbrock, P., Abraham, W.-R., Pieper, D.H., & Hofle, M.G. (2003). Bacterial community dynamics during biostimulation and bioaugmentation experiments aiming at chlorobenzene degradation in groundwater. *Microbial Ecology*, 46, 161–176.

- Wijbrans & Van Zutphen (2005). Environmental impact study for the combustion of palm fatty acid distillate in a power plant. *Zwolle* (the Netherlands), May 2005.
- Wu, L. C., & Hang, Y. D. (2000). Acid protease production from *Neosartorya fischeri*. *Lebensmittel-Wissenschaft and Technologie*, 33(1), 44–47.
- Yacob, S., Hassan, M. A., Shirai, Y., Wakisaka, M., & Subash, S. (2005). Baseline study of methane emission from open digesting tanks of palm oil mill effluent treatment. *Chemosphere*, 59, 1575–1581.
- Yacob, S., Hassan, M. A., Shirai, Y., Wakisaka, M., & Subash, S. (2006a). Baseline study of methane emission from anaerobic ponds of palm oil mill effluent treatment. *Science of the Total Environment*, 366, 187–196.
- Yacob, S., Shirai, Y., Hassan, M. A., Wakisaka, M., & Subash, S. (2006b). Start-up operation of semi-commercial closed anaerobic digester for palm oil mill effluent treatment. *Process Biochemistry*, 41, 962–964.
- Yu, K. S. H., Wong, A. H. Y., Yau, K. W. Y., Wong, Y. S., & Tam, N. F. Y. (2005). Natural attenuation, biostimulation and bioaugmentation on biodegradation of polycyclic aromatic hydrocarbons (PAHs) in mangrove sediments. *Marine Pollution Bulletin*, 51(8-12), 1071–1077.
- Zhang, X.-Z., & Zhang, Y.-H. P. (2010). One-step production of biocommodities from lignocellulosic biomass by recombinant cellulolytic *Bacillus subtilis*: Opportunities and challenges. *Engineering in Life Sciences*, 10(5), 1–9.
- Zinatizadeh A.A.L., (2006). *Biological treatment of palm oil mill effluent (POME) using an up-flow anaerobic sludge fixed film (UASFF) bioreactor*. Doctor of Philosophy Thesis, Universiti Sains Malaysia.
- Zupančič, G. D., & Grilc, V. (2012). Anaerobic treatment and biogas production from organic waste. In S. Kumar (Ed.), *Management of organic waste* (Vol. 2, pp. 1–27). Croatia: InTech.
- Zupančič, G. D., Uranjek-Ževart, N., & Roš, M. (2008). Full-scale anaerobic co-digestion of organic waste and municipal sludge. *Biomass and Bioenergy*, 32(2), 162–167.