



UNIVERSITI PUTRA MALAYSIA

***NISP AS A POTENTIAL SURFACE ANCHOR FOR PANDEMIC H1N1
2009 HAEMAGGLUTININ (HA1) VIRAL EPITOPE IN Lactococcus lactis***

STELLA SIAW XIU JOAN

FBSB 2014 39



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By

STELLA SIAW XIU JOAN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the Degree of Master of Science

December 2014

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of
the requirement for the degree of Master of Science

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December 2014

Chairman: Professor Raha Abdul Rahim, PhD

Faculty: Biotechnology and Biomolecular Sciences

The rapid worldwide spread of the pandemic H1N1 2009 influenza A virus in the early 2009 has contributed to high morbidity and mortality especially in children, the elderly and immuno-compromised individuals. Current available influenza vaccines are effective in terms of providing immunity protection but there are several limitations and disadvantages such as allergic reactions, biosafety issues and limited production capacity. This study uses *Lactococcus lactis* as a vehicle for cloning of the haemagglutinin (HA1) epitope of the pandemic H1N1 2009 influenza virus. The HA1 epitope was amplified and cloned into a *L. lactis* vector together with the usp45 signal peptide and nisP anchor protein (nisP-anc) in order to assemble a surface displayed HA1 recombinant *L. lactis*. The recombinant *L. lactis* were able to express HA1 epitope protein when induced with nisin and was further confirmed with Matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS) analysis. The recombinant host cells were further examined by using whole cell ELISA, fluorescence microscopy and flow cytometry to detect the surface displayed HA1 epitope protein on their surfaces. Despite it could not be confirmed that HA1 epitope has been successfully surface displayed, the recombinant cells were fed to BALB/c mice host to observe for its capability to induce immunogenic response. From the analysis of antigen-specific serum antibody by sandwich ELISA, significant anti-HA1 sIgA antibody was produced in the mice fecal suspension of mice group NZ9000 (pNZ:HN) when compared to the negative control NZ9000 (pNZ8048) suggesting that high specific anti-HA1 sIgA antibody was produced in the mice rectum. This study was conducted to gain more insights of the potential of *L. lactis* as a vaccine vehicle and future optimization can be adopted to further enhance its capability in vaccine delivery.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai
memenuhi keperluan untuk ijazah Master Sains

**NISP SEBAGAI POTENSI PAPARAN PERMUKAAN EPITOP VIRUS
PANDEMIK H1N1 2009 HEMAGGLUTININ PADA REKOMBINAN *Lactococcus
lactis***

Oleh

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Virus influenza A Pandemik H1N1 2009 yang muncul pada awal tahun 2009 telah merebak dengan cepat di seluruh dunia dan mengakibatkan morbiditi dan mortaliti terutama sekali di kalangan kanak-kanak, warga emas dan individu yang mempunyai sistem imun yang lemah. Vaksin influenza yang kini didapati adalah efektif dari segi perlindungan keimunan tetapi terdapat beberapa batasan dan keburukan seperti reaksi alergi, isu-isu keselamatan biologi dan keupayaan pengeluaran yang terhad. Kajian ini menggunakan *Lactococcus lactis* sebagai pembawa untuk pengklonan epitop hemagglutinin (HA1) dari virus influenza A Pandemik H1N1 2009. Epitop HA1 telah diamplifikasi dan diklonkan ke dalam vektor *L. lactis* bersama dengan peptida isyarat usp45 dan protein permukaan bersauh nisP (nisP-anc) untuk membentuk rekombinan *L. lactis* yang mempamerkan epitop HA1 dipermukaannya. Rekombinan *L. lactis* ini berkeupayaan untuk mengekspres protein epitop HA1 yang diaruh oleh nisin dan seterusnya disahkan dengan menggunakan analisa “Matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS)”. Sel-sel rekombinan ini seterusnya disahkan dengan menggunakan ELISA seluruh sel, mikroskopi fluoresen, dan “flow cytometry” untuk mengesan epitop HA1 yang dipamerkan di permukaan hos sel *L. lactis*. Walaupun tiada pengesahan yang epitop HA1 telah dengan jayanya dipaparkan di permukaan *L. lactis*, sel-sel rekombinan ini telah diuji dalam hos tikus BALB/c untuk mengesan keupayaannya untuk menghasilkan gerak balas imunogenik. Daripada analisa serum antibodi spesifik-antigen dengan menggunakan ELISA sandwic, antibodi sIgA anti-HA1 yang tinggi dalam suspensi tinja tikus dapat dikesan dalam kumpulan tikus NZ9000 (pNZ:HN) berbanding dengan kumpulan tikus kawalan negatif NZ9000 (pNZ8048) mencadangkan bahawa antibodi sIgA anti-HA1 yang tinggi telah dihasilkan di dubur tikus-tikus tersebut. Kajian ini dijalankan untuk mendapat pengetahuan dan menentukan potensi *L. lactis* sebagai pembawa vaksin dan pengoptimuman di masa akan datang seterusnya berupaya meninggikan keupayaannya dalam penyampaian vaksin.

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I certify that a Thesis Examination Committee has met on 1 December 2014 to conduct the final examination of Stella Siaw Xiu Joan on her thesis entitled “*NISP* as a Potential Surface Anchor for Pandemic H1N1 2009 Haemagglutinin (HA1) Viral Epitope in *Lactococcus lactis*” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A)106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

A260	-	absorbance at 260 nm
A450	-	absorbance at 450 nm
aa	-	amino acid
AcmA	-	N-acetylmuraminidase
BLAST	-	basic local alignment search tool
Bp	-	base pair
BSA	-	bovine serum albumin
CaCl ₂	-	calcium chloride
cDNA	-	complimentary DNA
cfu	-	colony forming unit
CO ₂	-	carbon dioxide
C-terminal	-	the carboxyl-terminal (-COOH) of a polypeptide
dH ₂ O	-	distilled water
DNA	-	deoxyribonucleic acid
dNTPs	-	deoxyribonucleotide triphosphate
EDTA	-	ethylenediamine tetraacetate
ELISA	-	enzyme-linked immunosorbant assay
EtBr	-	ethidium bromide
GMO	-	genetically modified organisms
GRAS	-	generally regarded as safe
H	-	hour
HA1	-	haemagglutinin 1
HCl	-	hydrochloric acid
His	-	histidine
HRP	-	horseradish peroxide
i.p	-	intraperitoneally
i.v	-	intravenously
IPTG	-	isopropyl-D-galactopyranoside
kb	-	kilo base pair
kDa	-	kilo Daltons
LAB	-	lactic acid bacteria
LB	-	Luria Bertani
M	-	molarity
MCS	-	multiple cloning site
MgCl ₂	-	magnesium chloride
min	-	minute
ml	-	milliliter
mM	-	milliMolar
mRNA	-	messenger RNA
NaCl	-	sodium chloride
NAG	-	N-acetylglucosamine
NAM	-	N-acetylmuramic acid
ng	-	nanogram
Ni ²⁺	-	nickel sulphate ion
NisP	-	serine protease
nisP-anc	-	NisP anchor protein
N-terminal	-	the amino-terminal (NH ₂) of a polypeptide
°C	-	degrees centigrade

OD	-	optical density
ORF	-	open reading frame
PBS	-	phosphate buffered saline
PCR	-	polymerase chain reaction
PEG	-	Polyethylene glycol
<i>Pfu</i>	-	<i>pyrococcus furiosus</i>
pH	-	isoelectric point
POD	-	peroxidase
PVDF	-	immobilon-polyvinylidene difluoride
RBS	-	ribosomal binding site
RE	-	restriction enzyme
RNA	-	ribonucleic acid
RNase	-	ribonuclease
rpm	-	revolution per minute
rRNA	-	ribosomal RNA
RT	-	room temperature
s	-	second
SDS-PAGE	-	sodium dodecyl sulphate-polyacrylamide gel electrophoresis
SOE	-	Splicing by overlap extension
sP.	-	species
subsp.	-	subspecies
TAE	-	tris-acetate-EDTA
Taq	-	<i>Thermus aquaticus</i>
TCA	-	tri-carboxylic acid
TE	-	tris EDTA
TEMED	-	tetramethyl-ethylene diamine
TGS	-	tris-glycine-SDS
TMB	-	3,3',5,5'-Tetramethylbenzidine
TTFC	-	tetanus toxin fragment C
U	-	unit
UV	-	ultraviolet
v/v	-	volume per volume
w/v	-	weight per volume
%	-	percent/percentage
µl	-	microliter
µg	-	microgram

CHAPTER 1

INTRODUCTION

1.1 Introduction

Pandemic H1N1 2009 influenza A virus which emerged in early 2009 has spread rapidly across the globe and contributed to morbidity and mortality especially in children, elderly and immune-compromised individuals. Although antiviral agents are able to treat influenza infections, vaccination is still the best method for controlling influenza. Current available influenza vaccines are live attenuated viruses that are propagated in embryonated chicken eggs. It is effective in terms of providing immunogenic protection but there are several limitations and disadvantages of this method such as allergic reactions to the egg protein, biosafety issues and limited production capacity.

The use of *Lactococcus lactis*, a lactic acid bacteria (LAB), in the production of recombinant vaccine is still less exploited compared to attenuated pathogens such as *Salmonella* (Levine *et al.*, 1996; Branger *et al.*, 2007). Their generally recognized as safe status and the availability of genetic tools for recombinant gene expression make them attractive to be used as potential vaccine vehicles. It can also overcome the disadvantages of egg-based vaccines and reverions. Several antigen specific immune responses have been successfully obtained using *L. lactis* as the vaccine carrier (Mozzi *et al.*, 2010). This study is a novel step towards the use of nisP-anc and LAB, *L. lactis* as a vehicle for the production of pandemic H1N1 2009 influenza virus vaccine.

1.2 Hypotheses

The hypotheses for this study are:

1. The NisP anchor protein (nisP-anc) will be able to surface display the HA1 pandemic H1N1 2009 viral epitope on *L. lactis* host cells.
2. The surface displayed recombinant *L. lactis* harbouring the HA1 viral epitope will stimulate both the cell-mediated and humoral immune responses in the BALB/c mouse model.

1.3 Problem statements

1. The production of current licensed pandemic H1N1 2009 vaccine cannot be scaled up easily.
2. Viruses propagated in the chicken eggs may be slightly modified during incubation period.
3. Inconvenient vaccination procedure such as intramuscular injection.
4. Recombinant vaccines produced with pathogenic microbes may pose threat to vaccinated person's health.

1.4 Objectives

The main objectives of this work include:

1. To construct an expression vector for *L. lactis* constituting of HA1 pandemic H1N1 2009 viral epitope together with usp45 signal peptide and nisP-anc.
2. To optimize expression conditions of the recombinant *L. lactis* constructs to improve gene expression.
3. To analyse surface display of the HA1 viral epitope on the *L. lactis* cell surface.
4. To investigate the immunogenic effect of recombinant *L. lactis* harbouring HA1 viral epitope when fed orally to BALB/c mice host.

REFERENCES

- Anuradha, K., Foo, H. L., Mariana, N. S., Loh, T. C., Yusoff, K., Hassan, M. D., Sasan, H., Raha, A. R. (2010). Live recombinant *Lactococcus lactis* vaccine expressing aerolysin genes D1 and D4 for protection against *Aeromonas hydrophila* in tilapia (*Oreochromis niloticus*). *Journal of Applied Microbiology*, 109(5), 1632-1642.
- Åvall-Jääskeläinen, S., Lindholm, A., & Palva, A. (2003). Surface display of the receptor-binding region of the *lactobacillus brevis* S-layer protein in *Lactococcus lactis* provides nonadhesive lactococci with the ability to adhere to intestinal epithelial cells. *Applied and Environmental Microbiology*, 69(4), 2230-2236.
- Bauer, G. (2000). Simplicity through complexity: Immunoblot with recombinant antigens as the new gold standard in epstein-barr virus serology. *Clinical Laboratory*, 47(5-6), 223-230.
- Belshe, R. B., Edwards, K. M., Vesikari, T., Black, S. V., Walker, R. E., Hultquist, M., Connor, E. M. (2007). Live attenuated versus inactivated influenza vaccine in infants and young children. *New England Journal of Medicine*, 356(7), 685-696.
- Bermúdez-Humarán, L. G., Cortes-Perez, N. G., Lefèvre, F., Guimarães, V., Rabot, S., Alcocer-Gonzalez, J. M., Corthier, G. (2005). A novel mucosal vaccine based on live lactococci expressing E7 antigen and IL-12 induces systemic and mucosal immune responses and protects mice against human papillomavirus type 16-induced tumors. *The Journal of Immunology*, 175(11), 7297-7302.
- Bermúdez-Humarán, L. G., Kharrat, P., Chatel, J. M., & Langella, P. (2011). Lactococci and lactobacilli as mucosal delivery vectors for therapeutic proteins and DNA vaccines. *Microb Cell Fact*, 10(Suppl 1), S4.
- Bober, J. A., & Demirci, A. (2004). Nisin Fermentation by *Lactococcus lactis* subsp. *lactis* Using Plastic Composite Supports in Biofilm Reactors. *Agricultural Engineering International: CIGR EJournal*.
- Bolotin, A., Wincker, P., Mauger, S., Jaillon, O., Malarme, K., Weissenbach, J., Sorokin, A. (2001). The complete genome sequence of the lactic acid bacterium *Lactococcus lactis* ssp. *lactis* IL1403. *Genome Research*, 11(5), 731-753.
- Branger, C. G., Curtiss III, R., Perry, R. D., & Fetherston, J. D. (2007). Oral vaccination with different antigens from *Yersinia pestis* KIM delivered by live attenuated *Salmonella typhimurium* elicits a protective immune response against plague. In *The Genus Yersinia* (pp. 387-399). Springer New York.
- Brodeur, B., Tsang, P., & Larose, Y. (1984). Parameters affecting ascites tumour formation in mice and monoclonal antibody production. *Journal of Immunological Methods*, 71(2), 265-272.

Carr, F. J., Chill, D., & Maida, N. (2002). The lactic acid bacteria: A literature survey. *Critical Reviews in Microbiology*, 28(4), 281-370.

Chan,M.

- World now at the start of 2009 influenza pandemic. Retrieved June/11, 2009, from
http://www.who.int/mediacentre/news/statements/2009/h1n1_pandemic_phase6_20090611/en/index.html
- De Dea Lindner, J. (2008). *Traditional and Innovative Approaches to Evaluate Microbial Contribution in Long Ripened Fermented Foods: The Case of Parmigiano Reggiano Cheese.*
- De Vuyst, L., & Vandamme, E. J. (1994). *Bacteriocins of lactic acid bacteria: Microbiology, genetics, and applications* Blackie Academic & Professional.
- Detmer, A., & Glenting, J. (2006). Live bacterial vaccines: A review and identification of potential hazards. *Microbial Cell Factories*, 5, 23.
- Engelke, G., Gutowski-Eckel, Z., Hammelmann, M., & Entian, K. (1992). Biosynthesis of the lantibiotic nisin: Genomic organization and membrane localization of the NisB protein. *Applied and Environmental Microbiology*, 58(11), 3730-3743.
- Fischetti, V., Pancholi, V., & Schneewind, O. (1990). Conservation of a hexapeptide sequence in the anchor region of surface proteins from gram-positive cocci. *Molecular Microbiology*, 4(9), 1603-1605.
- Garten, R. J., Davis, C. T., Russell, C. A., Shu, B., Lindstrom, S., Balish, A., Cox, N. J. (2009). Antigenic and genetic characteristics of swine-origin 2009 A(H1N1) influenza viruses circulating in humans. *Science*, 325(5937), 197-201.
- Gasson, M. J. (1983). Plasmid complements of *Streptococcus lactis* NCDO 712 and other lactic streptococci after protoplast-induced curing. *Journal of Bacteriology*, 154(1), 1-9.
- Genzel, Y., Behrendt, I., Konig, S., Sann, H., & Reichl, U. (2004). Metabolism of MDCK cells during cell growth and influenza virus production in large-scale microcarrier culture. *Vaccine*, 22(17-18), 2202-2208.
- Gupta, R. K., & Siber, G. R. (1995). Adjuvants for human vaccines - current status, problems and future prospects. *Vaccine*, 13(14), 1263-1276.
- Holmgren, J., Czerkinsky, C. (2005) Mucosal immunity and vaccines. *Nature Medicine*, 11, 545-553
- Holo, H., & Nes, I. F. (1995). Transformation of lactococcus by electroporation. *Methods Molecular Biology*, 47, 195-199.

- Hortin, G. L. (2006). The MALDI-TOF mass spectrometric view of the plasma proteome and peptidome. *Clinical Chemistry*, 52(7), 1223-1237.
- Kandler, O. (1983). Carbohydrate metabolism in lactic acid bacteria. *Antonie Van Leeuwenhoek*, 49(3), 209-224.
- Kimoto, H., Nomura, M., Kobayashi, M., Mizumachi, K., & Okamoto, T. (2003). Survival of lactococci during passage through mouse digestive tract. *Canadian Journal of Microbiology*, 49(11), 707-711.
- Kuipers, O. P., de Ruyter, P. G., Kleerebezem, M., & de Vos, W. M. (1998). Quorum sensing-controlled gene expression in lactic acid bacteria. *Journal of Biotechnology*, 64(1), 15-21.
- Kuske, C. R., Banton, K. L., Adorada, D. L., Stark, P. C., Hill, K. K., & Jackson, P. J. (1998). Small-scale DNA sample preparation method for field PCR detection of microbial cells and spores in soil. *Applied and Environmental Microbiology*, 64(7), 2463-2472.
- Laemmli, U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 227(5259), 680-685.
- Le Loir, Y., Azevedo, V., Oliveira, S. C., Freitas, D. A., Miyoshi, A., Bermúdez-Humarán, L. G., Gabriel, J. E. (2005). Protein secretion in *Lactococcus lactis*: An efficient way to increase the overall heterologous protein production. *Microbial Cell Factories*, 4(1), 2.
- Lee, S. Y., Choi, J. H., & Xu, Z. (2003). Microbial cell-surface display. *Trends in Biotechnology*, 21(1), 45-52.
- Leenhouts, K., Buist, G., & Kok, J. (1999). Anchoring of proteins to lactic acid bacteria. *Antonie Van Leeuwenhoek*, 76(1-4), 367-376.
- Leenhouts, K., & Venema, G. (1993). Lactococcal plasmid vectors. *Plasmids, a Practical Approach*. Oxford University Press, Oxford, United Kingdom, 65-94.
- Levine, M. M., Galen, J., Barry, E., Noriega, F., Chatfield, S., Sztein, M., & Tacket, C. (1996). Attenuated *Salmonella* as live oral vaccines against typhoid fever and as live vectors. *Journal of biotechnology*, 44(1), 193-196.
- Lim, S., Jahanshiri, F., Abdul Rahim, R., Sekawi, Z., & Yusoff, K. (2010). Surface display of respiratory syncytial virus glycoproteins in *Lactococcus lactis* NZ9000. *Letters in Applied Microbiology*, 51(6), 658-664.
- Makarova, K., Slesarev, A., Wolf, Y., Sorokin, A., Mirkin, B., Koonin, E., Polouchine, N. (2006). Comparative genomics of the lactic acid bacteria. *Proceedings of the National Academy of Sciences*, 103(42), 15611-15616.

- Mannam, P., Jones, K. F., & Geller, B. L. (2004). Mucosal vaccine made from live, recombinant *Lactococcus lactis* protects mice against pharyngeal infection with streptococcus pyogenes. *Infection and Immunity*, 72(6), 3444-3450.
- Mazmanian, S. K., Liu, G., Ton-That, H., & Schneewind, O. (1999). *Staphylococcus aureus* sortase, an enzyme that anchors surface proteins to the cell wall. *Science*, 285(5428), 760-763.
- Medina, E., & Guzmán, C. A. (2001). Use of live bacterial vaccine vectors for antigen delivery: Potential and limitations. *Vaccine*, 19(13-14), 1573-1580.
- Moeini, H., Rahim, R. A., Omar, A. R., Shafee, N., & Yusoff, K. (2011). *Lactobacillus acidophilus* as a live vehicle for oral immunization against chicken anemia virus. *Applied microbiology and biotechnology*, 90(1), 77-88.
- Morishita, T., Nobusawa, E., Nakajima, K., & Nakajima, S. (1996). Studies on the molecular basis for loss of the ability of recent influenza A (H1N1) virus strains to agglutinate chicken erythrocytes. *Journal of general virology*, 77(10), 2499-2506.
- Mozzi, F., Raya, R. R., & Vignolo, G. M. (2010). *Biotechnology of lactic acid bacteria: Novel applications* Wiley.com.
- Navarre, W. W., & Schneewind, O. (1994). Proteolytic cleavage and cell wall anchoring at the LPXTG motif of surface proteins in Gram-positive bacteria. *Molecular Microbiology*, 14(1), 115-121.
- Neumann, G., Noda, T., & Kawaoka, Y. (2009). Emergence and pandemic potential of swine-origin H1N1 influenza virus. *Nature*, 459(7249), 931-939.
- Odell, I. D., & Cook, D. (2013). Immunofluorescence techniques. *Journal of Investigative Dermatology*, 133(1), e4.
- Partridge, J., & Kieny, M. P. (2010). Global production of seasonal and pandemic (H1N1) influenza vaccines in 2009–2010 and comparison with previous estimates and global action plan targets. *Vaccine*, 28(30), 4709-4712. doi:DOI: 10.1016/j.vaccine.2010.04.083
- Perez-Padilla, R., de la Rosa-Zamboni, D., Ponce de Leon, S., Hernandez, M., Quiñones-Falconi, F., Bautista, E., Corrales, A. (2009). Pneumonia and respiratory failure from swine-origin influenza A (H1N1) in mexico. *New England Journal of Medicine*, 361(7), 680-689.
- Piard, J. C., Jimenez-Diaz, R., Fischetti, V. A., Ehrlich, S. D., & Gruss, A. (1997). The M6 protein of *Streptococcus pyogenes* and its potential as a tool to anchor biologically active molecules at the surface of lactic acid bacteria. *Advances in Experimental Medicine and Biology*, 418, 545-550.

- Pica, N., Hai, R., Krammer, F., Wang, T. T., Maamary, J., Eggink, D., Stein, C. R. (2012). Hemagglutinin stalk antibodies elicited by the 2009 pandemic influenza virus as a mechanism for the extinction of seasonal H1N1 viruses. *Proceedings of the National Academy of Sciences*, 109(7), 2573-2578.
- Pozzi, G., & Wells, J. M. (1997). *Gram-positive bacteria: Vaccine vehicles for mucosal immunization*. Springer.
- Raha, A. R., Varma, N. R. S., Yusoff, K., Ross, E., & Foo, H. L. (2005). Cell surface display system for *Lactococcus lactis*: a novel development for oral vaccine. *Applied microbiology and biotechnology*, 68(1), 75-81.
- Reisinger, K. S., Block, S. L., Izu, A., Groth, N., & Holmes, S. J. (2009). Subunit influenza vaccines produced from cell culture or in embryonated chicken eggs: Comparison of safety, reactogenicity, and immunogenicity. *Journal of Infectious Diseases*, 200(6), 849.
- Robertson, J. S., Cook, P., Attwell, A. M., & Williams, S. P. (1995). Replicative advantage in tissue culture of egg-adapted influenza virus over tissue-culture derived virus: Implications for vaccine manufacture. *Vaccine*, 13(16), 1583-1588.
- Robinson, K., Chamberlain, L. M., Schofield, K. M., Wells, J. M., & Le Page, R. W. F. (1997). Oral vaccination of mice against tetanus with recombinant *Lactococcus lactis*. *Nature Biotechnology*, 15(7), 653-657.
- Russell-Jones, G. (2000). Oral vaccine delivery. *Journal of Controlled Release*, 65(1), 49-54.
- Ryan, E. J., Daly, L. M., & Mills, K. H. (2001). Immunomodulators and delivery systems for vaccination by mucosal routes. *Trends in Biotechnology*, 19(8), 293-304.
- Sambrook, J., Fritsch, E. F., & Maniatis, T. (1989). *Molecular cloning: A laboratory manual (2nd edition)* (Cold Spring Harbor Laboratory Press edition). New York: Cold Spring Harbor.
- Savage, D. C. (1977). Microbial ecology of the gastrointestinal tract. *Annual Reviews in Microbiology*, 31(1), 107-133.
- Schleifer, K., Kraus, J., Dvorak, C., Kilpper-Bälz, R., Collins, M., & Fischer, W. (1985). Transfer of *Streptococcus lactis* and related streptococci to the genus *Lactococcus* gen. nov. *Systematic and Applied Microbiology*, 6(2), 183-195.
- Seegers, J. F. (2002). Lactobacilli as live vaccine delivery vectors: Progress and prospects. *Trends in Biotechnology*, 20(12), 508-515.

- Simonen, M., & Palva, I. (1993). Protein secretion in bacillus species. *Microbiological Reviews*, 57(1), 109-137.
- Smith, G. J. D., Vijaykrishna, D., Bahl, J., Lycett, S. J., Worobey, M., Pybus, O. G., Bhatt, S. (2009). Origins and evolutionary genomics of the 2009 swine-origin H1N1 influenza A epidemic. *Nature*, 459(7250), 1122-1125.
- Steen, M. T., Chung, Y. J., & Hansen, J. N. (1991). Characterization of the nisin gene as part of a polycistronic operon in the chromosome of *Lactococcus lactis* ATCC 11454. *Applied and Environmental Microbiology*, 57(4), 1181-1188.
- Steidler, L., Hans, W., Schotte, L., Neirynck, S., Obermeier, F., Falk, W., Remaut, E. (2000). Treatment of murine colitis by *Lactococcus lactis* secreting interleukin-10. *Science*, 289(5483), 1352-1355.
- Steidler, L., Viaene, J., Fiers, W., & Remaut, E. (1998). Functional display of a heterologous protein on the surface of *Lactococcus lactis* by means of the cell wall anchor of *Staphylococcus aureus* protein A. *Applied and Environmental Microbiology*, 64(1), 342-345.
- Tannock, G. W. (1999). Analysis of the intestinal microflora: A renaissance. *Antonie Van Leeuwenhoek*, 76(1-4), 265-278.
- Ton-That, H., Liu, G., Mazmanian, S. K., Faull, K. F., & Schneewind, O. (1999). Purification and characterization of sortase, the transpeptidase that cleaves surface proteins of *Staphylococcus aureus* at the LPXTG motif. *Proceedings of the National Academy of Sciences*, 96(22), 12424-12429.
- Van der Meer, J ROELOF, Polman, J., Beethuyzen, M. M., Siezen, R. J., Kuipers, O. P., & De Vos, W. (1993). Characterization of the *Lactococcus lactis* nisin A operon genes nisP, encoding a subtilisin-like serine protease involved in precursor processing, and nisR, encoding a regulatory protein involved in nisin biosynthesis. *Journal of Bacteriology*, 175(9), 2578-2588.
- Varma, S., & Raw, N. (2006). *Development of an Anchoring System for Protein Display on the Cell Wall Surface of Lactococcus Lactis MG1363*. Doctoral dissertation, Universiti Putra Malaysia, Selangor, Malaysia.
- Villena J., Medina M., Racedo S., Alvarez. (2010). Resistance of Young Mice to Pneumococcal Infection can be Improved by Oral Vaccination with Recombinant *Lactococcus lactis*. *Journal of Microbiology, Immunology and Infection*, 43(1), 1-10.
- Wells, J. M., & Mercenier, A. (2008). Mucosal delivery of therapeutic and prophylactic molecules using lactic acid bacteria. *Nature Reviews Microbiology*, 6(5), 349-362.

WorldHealthOrganization.

Pandemic (H1N1) 2009 - update 112. Retrieved August/6, 2010, from
http://www.who.int/csr/don/2010_08_06/en/index.html



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