

# **UNIVERSITI PUTRA MALAYSIA**

# ANTRAL FOLLICULAR DEVELOPMENT AND OESTROUS RESPONSE IN OESTROUS SYNCHRONIZED AND NATURALLY CYCLING GOATS

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DOCTOR OF PHILOSOPHY UNIVERSITI PUTRA MALAYSIA

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By

# MUHAMMAD MODU BUKAR

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirement for the Degree of Doctor of Philosophy

December 2011

### DEDICATION

In the name of Allah, the Beneficient, and the Merciful. This thesis is dedicated to my precious mother, Hajja Kaltume Mustapha who nurtured me with unconditional love. The thesis is also dedicated to my father, Alhaji Bukar Madu for his unflinching support and encouragement all these years, to all my siblings for their confidence in me; to my wife, Yagana Baba Mele for her support and finally, to my children, Aisha, Kaltume, Abubakar, Idris and Maryam. Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosopy

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December 2011

Chairman: Associate Professor Rosnina Yusoff, PhD

**Faculty: Veterinary Medicine** 

At present, most of the information on the pattern of antral follicular development on ruminants were gathered from studies conducted in cattle and sheep. However, very little amount of literature exists for goats, especially the non-seasonal polyoestrous goats domesticated in the humid tropics. Thus, the focus of this thesis was to examine the effects of oestrus synchronisation with  $PGF_{2\alpha}$  and  $P_4$ , alone or in combination, with and without gonadotrophins on antral follicular development in goats. In the first experiment, the effects of oestrus synchronisation with  $PGF_{2\alpha}$  and CIDR on follicular

development, plasma IGF-1 concentration and its association with follicle population were studied.

Twenty-four Boer x Australian feral crossbred goats that were between 3-4 years of age, with a mean bodyweight of  $35.0 \pm 2.7$  kg and a median body condition score of 3 were used in this study. The goats were equally divided into 3 groups: PGF<sub>2α</sub> (A), CIDR (B) and naturally cycling (C). Group A was synchronised with two intramuscular injections of 125 µg cloprostenol, 11 days apart while Group B was synchronised with CIDR inserted in the vagina for 17 days. Group C was not oestrus synchronised. Three waves of follicular development were most frequently observed (58%), followed by 4 waves (31.6%). There were no significant differences (P>0.05) between treatment groups for the other parameters associated with follicular development. A low positive correlation (r=0.14) was observed between IGF-1 concentration and mean number of follicles (r=0.13). The low correlation between IGF-1 concentration and follicular development in goats.

The objective of the second experiment was to analyse the effects of  $PGF_{2\alpha}$  and  $P_4$  methods of oestrus synchronisation with or without eCG and FSH on oestrus response and ovulation rates in the goats. The serum cortisol concentrations were also measured to determine if stress occurred during handling for ultrasonography, in the goats raised

under a hot and humid tropical environment. There were 9 groups of goats and each group was synchronised with different protocol using  $PGF_{2\alpha}$ , FGA and their combinations and either 5 mg FSH or 300 IU eCG. All the synchronised goats that were given eCG exhibited oestrus (100%). However, the number of follicles was higher (P < 0.05) in FSH synchronised groups than the eCG synchronised groups. It was concluded that the PGF<sub>2 $\alpha$ </sub> + FGA + FSH method of oestrus synchronisation is the most promising alternative to oestrus synchronisation with eCG in goats. The serum cortisol concentrations were not significantly different (P< 0.05) between goats handled for ultrasonography and the control group. The third experiment was conducted to examine the effects of eCG on preovulatory follicle growth and time of ovulation. The number of follicles, maximum size of the ovulatory follicles and time of ovulation in a total of fifty-one FGA +  $PGF_{2\alpha}$  + eCG synchronised goats and those without eCG were determined. It was found that eCG significantly (P<0.05) increased the total follicle number, maximum follicle diameter and reduced the time to ovulation by 20 hrs.

In conclusion, this is the first report of the pattern of follicular development during natural and synchronised oestrous cycles in non-seasonal polyoestrous goats raised in the hot humid tropics. The effects of oestrus synchronisation with prostaglandin  $F_{2\alpha}$ and  $P_4$  on follicular development were similar and both hormones increased the number of follicles and IGF-1 concentration in the synchronised and subsequent oestrous cycles compared with the naturally cycling group. Of the FSH-based oestrus synchronisation protocols, the  $PGF_{2\alpha} + FGA + FSH$  resulted in higher number of follicles than any of the eCG-based oestrus synchronisation protocols evaluated and is the most promising alternative to oestrus synchronisation with eCG in goats.

**Key words:** Ultrasonography, follicular development, oestrus response, IGF-1 concentration, cortisol

Abstrak tesis ini dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

### PERKEMBANGAN FOLIKEL ANTRUM DAN GERAKBALAS OESTRUS PADA KAMBING YANG DISELARASKAN ESTRUS DAN YANG BERKITARAN SEMULA JADI

Oleh

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Pengerusi: Profesor Madya Rosnina Yusoff, PhD

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Sehingga kini banyak maklumat mengenai perkembangan folikel antrum pada haiwan ruminan diperolehi daripada kajian yang melibatkan lembu dan biri-biri, dan maklumat mengenainya pada kambing adalah sangat sedikit, terutama bagi kambing poliestrus tidak bermusim yang dibelajinak di tropik melengas panas. Sehubungan itu, fokus tesis ini adalah untuk mengkaji kesan penyelarasan estrus dengan  $PGF_{2\alpha}$  dan  $P_4$ , bersendirian atau secara kombinasi, dengan dan tanpa gonadotropin terhadap perkembangan folikel antrum. Dalam eksperimen pertama, kesan penyelarasan estrus menggunakan  $PGF_{2\alpha}$  dan CIDR terhadap perkembangan folikel, kepekatan plasma IGF-1 dan korelasi dengan populasi folikel dikaji. Dua-puluh empat ekor kambing kacukan Boer x feral Australia yang berumur antara 3 dan 4 tahun, dengan berat badan

 $35.0 \pm 2.7$  kg dan skor median 3 bagi keadaan badan digunakan untok kajian ini. Kambing tersebut dibahagi sama kepada 3 kumpulan: PGF<sub>2a</sub> (A), CIDR (B) dan berkitaran semulajadi/kawalan (C). Kumpulan A diselaraskan dengan dua kali suntikan 125µg cloprostenol melalui intraotot dengan jarak 11 hari, manakala Kumpulan B diselaraskan dengan CIDR yang diletakkan ke dalam faraj selama 17 hari. Kumpulan C tidak diselaraskan estrusnya. Perkembangan folikel 3-gelombang paling kerap dicerap (58%), diikuti oleh 4 gelombang (31.6%). Tiada perbezaan signifikan (P>0.05) di antara kumpulan yang dikaji bagi parameter yang berkaitan dengan perkembangan folikel. Suatu korelasi positif rendah (r = 0.14) dicerap antara kepekatan IGF-1 dan bilangan 3 mm folikel dan antara kepekatan IGF-1 dan purata bilangan folikel (r = 0.13). Korelasi yang rendah antara kepekatan IGF-1 dan populasi folikel mencadangkan perhubungan lemah antara kepekatan plasma IGF-1 dan perkembangan folikel pada kambing.

Objektif eksperimen kedua adalah untuk menganalisis kesan kaedah penyelarasan estrus menggunakan PGF<sub>2a</sub> dan P<sub>4</sub> dengan atau tanpa eCG dan FSH terhadap gerakbalas estrus dan kadar pengovulanan pada kambing. Kepekatan serum kortisol juga disukat untok menentukan jikalau stress berlaku semasa pengendalian untok ultrasonografi di kalangan kambing yang dibelajinak di bawah persekitaran tropik panas dan melengas. Terdapat 9 kumpulan kambing dan setiap kumpulan diselaras dengan protokol yang berbeza menggunakan PGF<sub>2a</sub>, FGA serta kombinasi mereka dan 5 mg FSH atau 300 IU eCG. Semua kambing terselaras dan diberikan eCG mempamerkan oestrus (100%). Walau bagaimanapun, bilangan folikel adalah lebih

tinggi (P<0.05) dalam kumpulan FSH daripada kumpulan eCG. Kesimpulannya ialah kaedah PGF<sub>2 $\alpha$ </sub> + FGA + FSH merupakan alternatif harapan terhadap penyelarasan estrus mengguna eCG pada kambing. Kepekatan serum kortisol tidak memberi perbezaan signifikan (P>0.05) antara kambing yang dikendalikan untok ultrasonografi dan kumpulan kawalan.

Eksperimen ketiga dijalankan untuk meneliti kesan eCG terhadap pertumbuhan folikel praovulatori dan masa pengovulanan. Bilangan folikel, saiz maksimum folikel ovulatori dan masa pengovulanan pada 51 ekor kambing yang diselaraskan dengan FGA + PGF<sub>2a</sub> + eCG dan tanpa eCG ditentukan. Didapati bahawa eCG meningkatkan dengan signifikan (P<0.05) jumlah bilangan folikel, garispusat folikel maksimum dan mengurangkan masa pengovulanan ovulasi sebanyak 20 jam.

Kesimpulannya, ini merupakan laporan pertama terhadap corak perkembangan folikel semasa kitaran estrus semulajadi dan estrus terselaras di kalangan kambing poliestrus tidak bermusim yang dibelajinak di tropik melengas panas. Kesan penyelarasan estrus menggunakan PGF<sub>2</sub> dan P<sub>4</sub> terhadap perkembangan folikel adalah sama, dan keduadua hormon telah meningkatkan bilangan folikel dan kepekatan IGF-1 pada kitaran estrus yang diselaraskan dan kitaran estrus berikutnya berbanding kumpulan kitaran semulajadi. Dari protokol penyelarasan estrus berdasarkan FSH, kumpulan PGF<sub>2</sub> + FGA + FSH menghasilkan bilangan folikel yang tinggi berbanding penyelarasan estrus berasaskan eCG dan ia merupakan alternatif harapan terhadap penyelarasan estrus menggunakan eCG pada kambing.

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# DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

# MUHAMMAD MODU BUKAR

Date: 16<sup>th</sup> December 2011

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# LIST OF ABBREVIATIONS

ACTH	adrenocorticotrophic hormone
AI	artificial insemination
ANOVA	analysis of variance
BCS	body condition score
CIDR	controlled internal drug release
CL	corpus luteum
DF	dominant follicle
eCG	equine chorionic gonadotrophin
EIA	enzyme immunoassay
ELISA	enzyme-linked immunosorbent assay
FGA	flugestone acetate
FAOSTAT	food and agriculture organization of the united nations
FSH	follicle stimulating hormone
g	gravity
GnRH	gonadotropin releasing hormone
HPA	hypothalamic-pituitary axis
<sup>125</sup> I	Iodine 125
IGFBP	insulin-like growth factor binding protein

IGF-1	insulin-like growth factor 1
IGF-2	insulin-like growth factor 2
IGFR-1	insulin-like growth factor receptor 1
IGFR-2	insulin-like growth factor receptor 2
IU	international unit
kDa	kilodalton
LH	luteinizing hormone
МАР	medroxyprogesterone acetate
MHz	megahertz
P <sub>4</sub>	progesterone
P450scc	cytochrome P450 side chain cleavage
PBS	phosphate buffer saline
PGF <sub>2a</sub>	protaglandin F2 alpha
RIA	radioimmunoassay
SEM	standard error of mean

#### **CHAPTER 1**

#### **GENERAL INTRODUCTION**

In 2009, the world goat population was estimated at 879 million (Dubeuf and Boyazoglu, 2009; FAOSTAT, 2011). Out of this total, the largest population is found in Asia (524 million), followed by Africa (298 million). When these two populations are combined, they make up 94 % of the world goat population (FAOSTAT, 2011). The world-wide appeal of goats is due to their small manageable size, prolificacy, superior adaptive characteristics and their ability to thrive in adverse conditions and limited feed resources of arid, semi arid or tropical environment (Silanikove, 2000a; Lebbie, 2004). Overall goat productivity depends on genotype, environment and husbandry factors, thus the intensive management system is used to mitigate the negative effects of these factors in order to improve the efficiency of goat reproduction (Alexandre and Mandonnet, 2005).

In tropical countries like Indonesia and Malaysia, goats breed throughout the year from January to December (Devendra and Burns, 1983; Sodiq et al., 2003). This provides an opportunity for a greater control of their reproductive cycle to exploit the naturally high prolificay of the goats. Manipulation of the oestrous cycle through oestrus synchronisation is an effective tool to improve reproduction in food animals (Greyling, 2010). Exogenous hormones are used to control the oestrous cycle so that a large number of females exhibit oestrus behaviour within a short period of time. In goats, the treatments used for oestrus synchronisation include the administration of prostaglandin  $F_{2\alpha}$  (PGF<sub>2 $\alpha$ </sub>), or progesterone (P<sub>4</sub>) and their combination (Adams et al., 1992a; Wildeus, 2000; Whitley and Jackson, 2004).

In earlier studies, oestrus synchronisation gave better results when  $PGF_{2\alpha}$  or  $P_4$  was co-treated with gonadotrophins (Oliveira et al., 2001; Husein et al., 2007). Equine chorionic gonadotrophin (eCG) and follicle stimulating hormone (FSH) are the common gonadotrophins in oestrus synchronisation of domestic animals (Wildeus, 2000). However, eCG is more commonly used than FSH despite previous reports of the development of antibodies against eCG which adversely affects subsequent fertility (Baril et al., 1996). On the other hand, there are no reports of similar effects associated with the use of FSH (Viudes De Castro et al., 2009). Thus, there is a need to evaluate the use of FSH in oestrus synchronisation in goats.

Most research on oestrus synchronisation had been conducted in the developed temperate countries such as France and USA, in seasonally polyoestrous goat breeds and in temperate climates (Baril et al., 1993; Wildeus, 2000; Whitley and Jackson, 2004). However, it is necessary to observe the oestrus response in oestrus synchronised goats under the conditions they are raised. Consequently, many previous oestrus synchronisation studies in goats have been carried out in many countries under different management and environmental conditions (Ahmed et al., 1998; Amarantidis et al., 2004; De Santiago-Miramontes et al., 2009; Zhao et al., 2010).

Further studies could also be aided by serial ultrasonography which was introduced as a safe tool to monitor the growth and regression of ovarian antral follicles, repeatedly in the same animal regardless of their depth within the ovary (Bartlewski et al., 2000a; Bartlewski et al., 2002; Filho et al., 2007). This has made ultrasonography the preferred technique for the study of follicular development in domestic animals. However, ultrasonography has not been used to monitor follicular development in naturally cycling or oestrus synchronised non-seasonal polyoestrous goats in the humid tropics. Thus, an accurate depiction of the relationship between follicle development and plasma reproductive hormones concentrations could provide valuable information which could be used as a tool to control reproductive cycles in tropical goat breeds.

Follicular dynamics is closely associated with ovarian factors, which include insulin-like growth factor-1 (IGF-1), their binding proteins, and proteases (Hwa et al., 1999; Monget and Bondy, 2000; Monget et al., 2002). The positive effects of IGF-1 on ovarian physiology and follicular development has been reported in sheep (Scaramuzzi et al., 1999). In cattle, close association was found between endocrine IGF-1 concentration and some reproductive traits such as early age at first calving and first service to conception (Yilmaz et al., 2006). These associations led to the suggestion that endocrine IGF-I levels could be used as a selection tool for reproductively high performing sheep and cattle (Roberts et al., 1990; Taylor et al., 2004). However, in this study, the focus was to determine the correlation between endocrine IGF-1 and ovarian follicular development during induced and natural oestrous cycle as a more reliable indicator of the relationship between them. Thus, a better understanding of the effects of oestrus synchronisation on endocrine IGF-1 concentration would also aid in the assessment of the actual clinical significance and usefulness of the measurement of circulating IGF-1 concentration.

It was hypothesized that oestrus synchronisation with  $PGF_{2\alpha}$  and  $P_4$  increased antral follicular development and plasma IGF-1 concentration and gonadotrophins stimulate the follicular development, increased the percentage oestrus response and the ovulation rate in goats. Thus, the aim of this study was to examine the relationship between plasma IGF-1 concentrations and antral follicular development during the oestrous cycles of  $PGF_{2\alpha}$  or  $P_4$  synchronised and naturally cycling goats. The efficiency of oestrus synchronisation with prostaglandin  $F_{2\alpha}$  and  $P_4$  with and without gonadotrophins was also evaluated.

The specific objectives were:

- 1. To determine the effects of  $PGF_{2\alpha}$  or  $P_4$  on follicular dynamics in oestrus synchronised and naturally cycling goats.
- 2. To determine the relationship between follicular development and plasma IGF-1 concentration in oestrus synchronised and naturally cycling goats.
- 3. To determine the effects of eCG on preovulatory follicle growth and time of ovulation in non-seasonal, polyoestrous goats
- 4. To determine the effects of  $PGF_{2\alpha}$  and  $P_4$  with or without eCG and FSH treatments on oestrus response and ovulation rates in non-seasonal polyoestrous goats raised under hot and humid condition.

### REFERENCES

- Abecia, J. A., Forcada, F., Zuniga, O. and Valares, J. A. 2002. The effect of progestagen treatment on sheep reproductive performance at different phases of the oestrous cycle. *Animal Research* 51: 149-155.
- Acuti, G., Todini, L., Malfatti, A., Antonini, M., Barbato, O. and Trabalza-Marinucci, M. 2009. Effects of field bean (Vicia faba L. var. minor) dietary supplementation on plasma thyroid hormones, insulin, insulin-like growth factor-1 concentrations and mohair characteristics in growing Angora goat kids. *Journal of Animal Physiology and Animal Nutrition* 93: 456-466.
- Adams, G. P., Matteri, R. L. and Ginther, O. J. 1992a. Effect of progesterone on ovarian follicles, emergence of follicular waves and circulating follicle stimulating hormone in heifers. *Journal of Reproduction and Fertility* 95: 627-640.
- Adams, G. P., Matteri, R. L., Kastelic, J. P., Ko, J. C. H. and Ginther, O. J. 1992b. Association between surges of follicle-stimulating hormone and the emergence of follicular waves in heifers. *Journal of Reproduction and Fertility* 94: 177-188.
- Adams, G. P., Jaiswal, R., Singh, J. and Malhi, P. 2008. Progress in understanding ovarian follicular dynamics in cattle. *Theriogenology* 69: 72-80.
- Ahmed, M. M. M., Makawi, S. E. and Jubara, A. S. 1998. Synchronisation of oestrus in Nubian goats. *Small Ruminant Research* 30: 113-120.
- Al-Azraqi, A. A. 2007. Effect of fasting on luteal function, leptin and steroids concentration during oestrous cycle of the goat in natural photo-status. *Animal Reproduction Science* 98: 343-349.
- Al-Merestani, M. R., Zarkawi, M. and Wardeh, M. F. 2003. Improving the reproductive efficiency, pregnancy diagnosis and monitoring the resumption of luteal activity in indigenous Damascus goats. *Reproduction in Domestic Animals* 38: 36-40.
- Alexandre, G. and Mandonnet, N. 2005. Goat meat production in harsh environments. *Small Ruminant Research* 60: 53-66.
- Allrich, R. D. 1994. Endocrine and neural control of estrus in dairy cows. *Journal* of Dairy Science 77: 2738-2744.

- Alvarez, L., Arvizu, R. R., Luna, J. A. and Zarco, L. A. 2010. Social ranking and plasma progesterone levels in goats. *Small Ruminant Research* 90: 161-164.
- Amarantidis, I., Karagianidis, A., Saratsis, P. H. and Brikas, P. 2004. Efficiency of methods used for estrous synchronisation in indigenous Greek goats. *Small Ruminant Research* 52: 247-252.
- Amer, H. A. and Hazzaa, A. B. 2009. The effect of different progesterone protocols on the reproductive efficiency of ewes during the non-breeding season. *Veterinarski Arhiv* 79: 19-30.
- Amoah, E. A., Gelaye, S., Guthrie, P. and Rexroad, C. E. 1996. Breeding season and aspects of reproduction of female goats. *Journal of Animal Science* 74: 723-728.
- Arashiro, E. K., Fonseca, J. F., Siqueira, L. G. B., Fernandes, C. A., Brandao, F. Z., Oba, E. and Viana, J. H. 2010. Assessment of luteal function in goats by ultrasonographic image attribute analysis. *Small Ruminant Research* 94: 176-179.
- Ariyaratna, H. B. S. and Gunawardana, V. K. 1997. Morphology and morphometry of ovarian follicles in the goat. *Small Ruminant Research* 26: 123-129.
- Azawi, O. I. and Al-Mola, M. K. M. A. 2010. A study on superovulation using FSH and eCG in Awassi ewes. *Tropical Animal Health and Production* 42: 799-801.
- Baby, T. E. and Bartlewski, P. M. 2011 Progesterone as the driving regulatory force behind serum FSH concentrations and antral follicular development in cycling ewes. *Reproduction, Fertility and Development* 23: 303-310.
- Badinga, L., Thatcher, W. W., Diaz, T., Drost, M. and Wolfenson, D. 1993. Effect of environmental heat stress on follicular development and steroidogenesis in lactating Holstein cows. *Theriogenology* 39: 797-810.
- Bao, B. and Garverick, H. A. 1998. Expression of steroidogenic enzyme and gonadotropin receptor genes in bovine follicles during ovarian follicular waves: a review. *Journal of Animal Science* 76: 1903-1921.
- Baril, G., Leboeuf, B. and Saumande, J. 1993. Synchronisation of estrus in goats: The relationship between time of occurrence of estrus and fertility following artificial insemination. *Theriogenology* 40: 621-628.

- Baril, G., Remy, B., Vallet, C., Beckers, J. F. and Saunmade, J. 1996. Synchronisation of estrus in goats: the relationship between eCG binding in plasma, time of occurrence of estrus and fertility following artificial insemination. *Theriogenology* 45: 1553-1559.
- Baril, G., Touze, J. L., Pignon, R. and Saumande, J. 2000. Evaluation of the efficiency of transrectal ultrasound to study ovarian function in goats. [Abstract]. *Theriogenology* 53: 370.
- Bartlewski, P. M., Beard, A. P. and Rawlings, N. C. 2000a. An ultrasound-aided study of temporal relationships between the patterns of LH/FSH secretion, development of ovulatory-sized antral follicles and formation of corpora lutea in ewes. *Theriogenology* 54: 229-245.
- Bartlewski, P. M., Vanderpol, J., Beard, A. P., Cook, S. J. and Rawlings, N. C. 2000b. Ovarian antral follicular dynamics and their associations with peripheral concentrations of gonadotropins and ovarian steroids in anoestrous Finnish Landrace ewes. *Animal Reproduction Science* 58: 273-291.
- Bartlewski, P. M., Beard, A. P., Cook, S. J. and Rawlings, N. C. 2002. Ovarian activity during sexual maturation and following introduction of the ram to ewe lambs. *Small Ruminant Research* 43: 37-44.
- Bartlewski, P. M., Duggavathi, R., Aravindakshan, J., Barrett, D. M. W., Cook, S. J. and Rawlings, N. C. 2003. Effects of a 6-day treatment with medroxyprogesterone acetate after prostaglandin F2α-induced luteolysis at midcycle on antral follicular development and ovulation rate in nonprolific Western white-faced ewes. *Biology of Reproduction* 68: 1403-1412.
- Bazer, F. W., Thatcher, W. W., Hansen. P.S., Mirando, M. A., Ott, T. L. and Plante, C. 1991. Mechanisms of pregnancy recognition in ruminants. *Journal of Reproduction and Fertility* 43: 39-47.

Bearden, H. J., Fuquay, J. W. and Willard, S. T. (ed.) 2004. Applied Animal Reproduction. 6th Edition. New Jersey: Pierson Educ. Inc. pp. 67-70.

Beg, M. A., Bergfelt, D. R., Kot, K. and Ginther, O. J. 2002. Follicle selection in cattle: dynamics of follicular fluid factors during development of follicle dominance. *Biology of Reproduction* 66: 120-126.

- Bister, J. L., Noel, B., Perrad, B., Mandiki, S. N. M., Mbayahaga, J. and Paquay, R. 1999. Control of ovarian follicles activity in the ewe. *Domestic Animal Endocrinology* 17: 315-328.
- Bley, M. A., Sarageta, P. E. and Baraao, J. L. 1997. Concerted stimulation of rat granulosa cell deoxyribonucleic acid synthesis by sex steroids and folliclestimulating hormone. *The Journal of Steroid Biochemistry and Molecular Biology* 62: 11-19.
- Bondy, C. A. and Zhou, J. 2005. Growth hormone, insulin-like growth factors and the female reproductive system. In: Varela-Nieto, I. and Chowen, J. A. (ed.), *The Growth Hormone/Insulin-Like Growth Factor Axis During Development.* Springer, New York, USA. pp. 91-115.
- Boscos, C. M., Samartzi, F. C., Dellis, S., Rogge, A., Stefanakis, A. and Krambovitis, E. 2002. Use of progestagen-gonadotrophin treatment in estrus synchronisation of sheep. *Theriogenology* 58: 1261-1272.
- Boyazoglu, J., Hatziminaoglou, I. and Morand-Fehr, P. 2005. The role of the goat in society: past, present and perspectives for the future. *Small Ruminant Research* 60: 13-23.
- Breen, K. M. and Karsch, F. J. 2004. Does cortisol inhibit pulsatile luteinizing hormone secretion at the hypothalamic or pituitary level? *Endocrinology* 145: 692-698.
- Bukar, M. M., Amin, J. D., Sivachelvan M. N. and Ribadu. A.Y. (2006). Postnatal Histological Development of the Ovaries and Uterus and the attainment of sexual maturity in kid Female Sahel Goats. *Small Ruminant Research* 65: 200-208.
- Bunter, K. L., Hermesch, S, Luxford, B. G., Graser, H. U. and Crump, R. E. 2005. Insulin-like growth factor-1 measured in juvenile pigs is genetically correlated with economically important performance traits. *Australian Journal of Experimental Agriculture* 45: 783-792.
- Burkholder, W. J. 2000. Use of body condition scores in clinical assessment of the provision of optimal nutrition. *Journal of the American Veterinary Medical Association* 217: 650-654.

- Burns, D. S., Jimenez-Krassel, F., Ireland, J. L., Knight, P. G. and Ireland, J. J. 2005. Numbers of antral follicles during follicular waves in cattle: evidence for high variation among animals, very high repeatability in individuals, and an inverse association with serum follicle-stimulating hormone concentrations. *Biology of Reproduction* 73: 54-62.
- Campbell, B. K., Dobson, H. and Scaramuzzi, R. J. 1998. Ovarian function in ewes made hypogonadal with GnRH antagonist and stimulated with FSH in the presence or absence of low amplitude LH pulses. *Journal of Endocrinology* 156: 213-222.
- Campbell, B. K., Kendall, N. R. and Baird, D. T. 2007. The effect of the presence and pattern of luteinizing hormone stimulation on ovulatory follicle development in sheep. *Biology of Reproduction* 76: 719-727.
- Casey, N. H. and Van Niekerk, W. A. 1988. The boer goat. I. Origin, adaptability, performance testing, reproduction and milk production. *Small Ruminant Research* 1: 291-302.
- Casey, N. H. and Webb, E. C. 2010. Managing goat production for meat quality. Small Ruminant Research 89: 218-224.
- Chao, L. M., Takayama, K., Nakanishi, Y., Hamana, K., Takagi, M., Kubota, C. and Kojima, T. 2008. Luteal lifespan and fertility after estrus synchronization in goats. *Journal of Veterinary Science* 9: 95-101.
- Chemineau, P., Daveau, A., Maurice, F. and Delgadillo, J. A. 1992. Seasonality of estrus and ovulation is not modified by subjecting female Alpine goats to a tropical photoperiod. *Small Ruminant Research* 8: 299-312.
- Chowdhury, S. A., Bhuiyan, M. S. A. and Faruk, S. 2002. Rearing Black Bengal goat under semi-intensive management 1: Physiologic and reproductive performances. *Asian-Australasian Journal of Animal Science* 15: 477-484.
- Christenson, K. L. and Devoto, L. 2003. Cholesterol transport and steroidogenesis by the corpus luteum. *Reproductive Biology and Endocrinology* 1: 1-9.
- Cruz, J. F., Rondina, D. and Freitas, V. J. 2005. Ovarian follicular dynamics during anoestrus in Anglo-Nubian and Saanen goats raised in tropical climate. *Tropical Animal Health and Production* 37: 395-402.

- Cueto, M., Gibbons, A., Alberio, R., Taddeo, H. and Gonzalez-Bulnes, A. 2006. Timing of emergence of ovulatory follicles in polyovulatory goats. *Animal Reproduction Science* 91: 275-284.
- Daftary, S. S. and Gore, A. C. 2005. IGF-1 in the brain as a regulator of reproductive neuroendocrine function. *Experimental Biology and Medicine* 230: 232-306.
- Davis, M. E. and Simmen, R. C. (2006). Genetic parameter estimates for serum insulin-like growth factor I concentrations, and body weight and weight gains in Angus beef cattle divergently selected for serum insulin-like growth factor I concentration. *Journal of Animal Science*. 84:2299–308.
- de Castro, T., Rubianes, E., Menchaca, A. and Rivero, A. 1999. Ovarian dynamics, serum estradiol and progesterone concentrations during the interovulatory interval in goats. *Theriogenology* 52: 399-411.
- De Santiago-Miramontes, M. A., Malpaux, B. and Delgadillo, J. A. 2009. Body condition is associated with a shorter breeding season and reduced ovulation rate in subtropical goats. *Animal Reproduction Science* 117: 175-182.
- DesCoteaux, L., Carriere, P. D. and Durocher, J. 2006. Ultrasonography of the reproductive system of the cow: Basic principles, practical uses and economic aspects of this diagnostic tool in dairy production. Proceedings of the 24<sup>th</sup> World Buiatrics Congress, Nice, France, 15-19 October, 2006, Nice, France. pp 303-314.
- Devendra, C. and Nozawa, K. 1976. Goats in South East Asia their status and production. Zeitschrift für Tierzüchtung und Züchtungsbiologie 93: 101-120.
- Devendra, C. and Burns, M. 1983. Goat production in the tropics. Commonwealth Agricultural Bureaux, Farnham Royal, Slough, U.K. pp. 182-183
- Devendra, C. and Solaiman, S. G. 2010. Perspectives on goats and global production. In: Solaiman, S. G. (ed.), *Goat Science and Production*. 1<sup>st</sup> Edition. Iowa, USA: Blackwell Publishing. pp. 3-20.
- Devendra, C. and Leng, R. A. 2011. Feed resources for animals in Asia: issues, strategies for use, intensification and integration for increased productivity. *Asian-Australasian Journal of Animal Sciences* 24: 303-321.

- Diaz, F. J., Anderson, L. E., Wu, Y. L., Rabot, A., Tsai, S. J. and Wiltbank, M. C. 2002. Regulation of progesterone and prostaglandin  $F_{2\alpha}$  production in the corpus luteum. *Molecular and Cellular Endocrinology* 191: 65-80.
- Dogan, I., Nur, Z., Gunay, U., Sagirkaya, H., Soylu, M. K. and Sonmez, C. 2005. Estrous synchronisation during the natural breeding season in Anatolian black does. *Veterinarni Medicina-Czech* 50: 6.
- Driancourt, M. A., Reynaud, K. and Smitz, J. 2001. Differences in follicular function of 3-month-old calves and mature cows. *Reproduction* 121: 463-474.
- Drummond, A. 2006. The role of steroids in follicular growth. *Reproductive Biology and Endocrinology*, 4(1), 16. Retrieved from <u>http://www.ncbi.nlm.nih.gov/pmc/articles/PMC1459164</u>/1477-7827-4-16
- Dubeuf, J.-P. and Boyazoglu, J. 2009. An international panorama of goat selection and breeds. *Livestock Science* 120: 225-231.
- Erasmus, J. A. 2000. Adaptation to various environments and resistance to disease of the improved Boer goat. *Small Ruminant Research* 36: 179-187.
- Erickson, B. H. 1966. Development and Senescence of the Postnatal Bovine Ovary. Journal of Animal Science 25: 800-805.
- Evans, A. C. O., Adams, G. P. and Rawlings, N. C. 1994. Follicular and hormonal development in prepubertal heifers from 2 to 36 weeks of age. *Journal of Reproduction and Fertility* 102: 463-470.
- Evans, A. C. O., Duffy, P., Hynes, N. and Boland, M. P. 2000. Waves of follicle development during the estrous cycle in sheep. *Theriogenology* 53: 699-715.
- Evans, A. C. O. 2003. Characteristics of ovarian follicle development in domestic animals. *Reproduction in Domestic Animals* 38: 240-246.
- FAOSTAT. 2011. World goat population. Food and Agriculture Organization of the United Nations. Rome, Italy http://faostat.fao.org. Retrieved 12-05-2011.

- Fatet, A., Pellicer-Rubio, M.-T. and Leboeuf, B. 2011. Reproductive cycle of goats. *Animal Reproduction Science* 124: 211-219.
- Fenwick, M. A., Fitzpatrick, R., Kenny, D. A., Diskin, M. G., Patton, J. and Murphy, J. J. 2008. Interrelationships between negative energy balance (NEB) and IGF regulation in liver of lactating dairy cows. *Domestic Animal Endocrinology* 34: 31-44.
- Ferin, M. 2006. Stress and the Reproductive System. In: Neill, J. D. (ed.), Knobil and Neill's Physiology of Reproduction. 3rd Edition. St Louis, Missouri: Elsevier, USA. pp. 2627-2667.
- Fernandez-Moro, D., Veiga-Lopez, A., Ariznavarreta, C., Tresguerres, J. A., Encinas, T. and Gonzalez-Bulnes, A. 2008. Preovulatory follicle development in goats following oestrous synchronisation with progestagens or prostaglandins. *Reproduction in Domestic Animals* 43: 9-14.
- Filho, T. F., Santos, M. H. B., Carrazzoni, P. G., Paula-Lopes, F. F., Neves, J. P., Bartolomeu, C. C., Limaa, P. F. and Oliveira, M. A. L. 2007. Follicular dynamics in Anglo-Nubian goats using transrectal and transvaginal ultrasound. *Small Ruminant Research* 72: 51-56.
- Fonseca, J. F., Torres, C. A. A., Santos, A. D. F., Maffili, V. V., Amorim, L. S. and Moraes, E. A. 2008. Progesterone and behavioral features when estrus is induced in Alpine goats. *Animal Reproduction Science* 103: 366-373.
- Fortune, J. E., Rivera, G. M., Evans, A. C. O. and Turzillo, A. M. 2001. Differentiation of dominant versus subordinate follicles in cattle. *Biology of Reproduction* 65: 648-654.
- Fortune, J. E. 2003. The early stages of follicular development: activation of primordial follicles and growth of preantral follicles. *Animal Reproduction Science* 78: 135-163.
- Ginther, O. J. and Kot, K. 1994. Follicular dynamics during the ovulatory season in goats. *Theriogenology* 49: 987-1001.
- Ginther, O. J., Kot, K., Kulick, L. J., Martin, S. and Wiltbank, M. C. 1996. Relationships between FSH and ovarian follicular waves during the last six months of pregnancy in cattle. *Journal of Reproduction and Fertility* 108: 271-279.

- Ginther, O. J., Bergfelt, D. R., Kulick, L. J. and Kot, K. 2000. Selection of the dominant follicle in cattle: role of estradiol. *Biology of Reproduction* 63: 383-389.
- Ginther, O. J., Beg, M. A., Donadeu, F. X. and Bergfelt, D. R. 2003. Mechanism of follicle deviation in monovular farm species. *Animal Reproduction Science* 78: 239-257.
- Gonzalez-Bulnes, A., Santiago-Moreno, J., Gomez-Brunet, A., Inskeep, E. K., Townsend, E. C. and Lopez-Sebastian, A. 1999. Follicular dynamics during the oestrous cycle in dairy goats. *Animal Science* 68: 547-554.
- Gonzalez-Bulnes, A., Santiago-Moreno, J., Cocero, M. J. and Lopez-Sebastian, A. 2000. Effects of FSH commercial preparation and follicular status on follicular growth and superovulatory response in Spanish Merino ewes. *Theriogenology* 54: 1055-1064.
- Gonzalez-Bulnes, A., Diaz-Delfa, B., Urrutia, J. A., Carrizosa, A. and Lopez-Sebastian, A. 2004. Ultrasonographic screening of the ovulatory process in goats. *Small Ruminant Research* 52: 165-168.
- Gonzalez-Bulnes, A., Diaz-Delfa, C., Garcia-Garcia, R. M., Urrutia, B., Carrizosa, J. A. and Lopez-Sebastian, A. 2005. Origin and fate of pre-ovulatory follicles after induced luteolysis at different stages of the luteal phase of the estrous cycle in goats. *Animal Reproduction Science* 86: 237-245.
- Gonzalez-Bulnes, A., Pallares, P. and Vazquez, M. I. 2010. Ultrasonographic imaging in small ruminant reproduction. *Reproduction in Domestic Animals* 45: 9-20.
- Goodwin, N., Hayssen, V., Deakin, D. W. and Flint, A. P. F. 1998. Influence of social status on ovarian function in farmed red deer (*Cervus elaphus*). *Physiology and Behavior* 65: 691-696.
- Gordon, I. (ed.) 2004. *Reproductive Technologies in Farm Animals*. Wallingford, Oxfordshire, UK: CAB International. pp. 332.
- Greyling, J. and Van Niekerk, C.H., 1990. Ovulation in the Boer goat doe. *Small Ruminant Research* 3: 457-46.
- Greyling, J. 2000. Reproduction traits in the Boer goat doe. *Small Ruminant Research* 36: 171-177.

- Greyling, J. 2010. Applied reproductive physiology. In: Solaiman, S. G. (ed.), *Goat Science and Production*. 1<sup>st</sup> Edition. Iowa, USA: Blackwell Publishing. pp. 139-155.
- Gupta, P. (ed.) 2008. Goats of the world. 1<sup>st</sup> Edition. New Delhi: Chawla Offset Printers. pp. 268.
- Gurung, N. K. and Solaiman, S. G. 2010. Goat breeds. In: Solaiman, S. G. (ed.), Goat Science and Production. 1<sup>st</sup> Edition. Iowa, USA: Blackwell Publishing. pp. 21-37.
- Haenlein, G. F. W. 2004. Goat milk in human nutrition. *Small Ruminant Research*: 155-163.
- Hashizume, T., Ohtsuki, T. and Matsumoto, N. 2000. Plasma insulin-like growth factor-I concentrations increase during the estrous phase in goats. *Domestic Animal Endocrinology* 18: 253-263.
- Husein, M. Q., Ababneh, M. M. and Abu-Ruman, D. S. 2007. The effects of short or long term FGA treatment with or without eCG on reproductive performance of ewes bred out-of-season. *American Journal of Animal and Veterinary Sciences* 2: 23-28.
- Hwa, V., Oh, Y. and Rosenfeld, R. G. 1999. The insulin-like growth factor-binding protein (IGFBP) superfamily. *Endocrinology Reviews* 20: 761-768.
- Jaisuwal, R. S., Singh, J., Marshal, L. and Adams, G. P. 2009. Repeatability of 2wave and 3-wave patterns of ovarian follicular development during the bovine estrous cycle. *Theriogenology* 72: 81-90.
- Jessen, C. and Pongratz, H. 1979. Air humidity and carotid rete function in thermoregulation of the goat. *The Journal of Physiology* 292: 469-479.
- Kawashima, C., Kida, K., Hayashi, K.-G., Amaya Montoya, C., Kaneko, E., Matsunaga, N., Shimizu, T., Matsui, M., Miyake, Y.-I., Schams, D. and Miyamoto, A. 2007. Changes in plasma metabolic hormone concentrations during the ovarian cycles of Japanese black and holstein cattle. *The Journal* of Reproduction and Development 53: 247-254.
- Kazanskaya, E., Kuznetsova, M. and Danilkin, A. 2007. Phylogenetic reconstructions in the genus *Capra* (Bovidae, Artiodactyla) based on the mitochondrial DNA analysis. *Russian Journal of Genetics* 43: 181-189.

- Kenyon, P. R., Jenkinson, C. M. C., Blair, H. T., Morel, P. C. H., Breier, B. H. and Gluckman, P. D. 2009. Reproductive performance of progesterone synchronised IGF-1 selection line ewes. *New Zealand Journal of Agricultural Research* 52: 307 - 314.
- Kosgey, I. S., Baker, R. L., Udo, H. M. J. and Van Arendonk, J. A. M. 2006. Successes and failures of small ruminant breeding programmes in the tropics: a review. *Small Ruminant Research* 61: 13-28.
- Kusina, N., Tarwirei, T., Hamidukuwanda, H., Agumba, G. and Mukwena, J. 2000. A comparison of the effects of progesterone sponges and ear implants,  $PGF_{2\alpha}$ , and their combination on efficacy of estrus synchronisation and fertility of Mashona goat does. *Theriogenology* 53: 1567-1580
- Lassala, A., Hernandez-Ceron, J., Rodriguez-Maltos, R. and Gutierrez., C. G. 2004. The influence of the corpus luteum on ovarian follicular dynamics during estrous synchronisation in goats. *Animal Reproduction Science* 84: 369-375.
- Lebbie, S. H. B. 2004. Goats under household conditions. *Small Ruminant Research* 51: 131-136.
- Leboeuf, B., Manfredi, E., Boue, P., Piacere, A., Brice, G., Baril, G., Brogua, C., Humblot, P. and Terqui, M. 1998. Artificial insemination of dairy goats in France. *Livestock Production Science* 15: 193-203.
- Lehloenya, K. C., Greyling, J. P. C. and Grobler, S. 2008. Effect of season on the superovulatory response in Boer goat does. *Small Ruminant Research* 78: 74-79.
- Letelier, C.A., Contreras-Solis, I., Garcia-Fernandez, R.A., Sanchez, M.A., Garcia-Palencia, P., Sannchez, B., Ariznavarreta, C., Tresguerres, J.A.F., Flores, J.M. and Gonzalez-Bulnes, A. 2011. Effects of oestrus induction with progestagens or prostaglandin analogues on ovarian and pituitary function in sheep. *Animal Reproduction Science* 126: 61-69.
- Leyva, V., Buckrell, B. C. and Walton, J. S. 1998. Regulation of follicular activity and ovulation in ewes by exogenous progestagen. *Theriogenology* 50: 395-416.

- Ligtvoet, C. M., Bom, N. and Gussenhoven, W. J. 1989. Technical principles of ultrasound. In: Taverne, M. A. M. and Willemse, A. H. (ed.), *Diagnostic Ultrasound and Animal Reproduction*. Dordrecht, The Netherlands: Kluwer Academic Publishers. pp. 123.
- Liu, X., Hart, E. J., Dai, Q., Rawlings, N. C., Pierson, R. A. and Bartlewski, P. M. 2007. Ultrasonographic image attributes of non-ovulatory follicles and follicles with different luteal outcomes in gonadotropin-releasing hormone (GnRH)-treated anestrous ewes. *Theriogenology* 67: 957-969.
- Logan, K. A., Juengel, J. L. and McNatty, K. P. 2002. Onset of steroidogenic enzyme gene expression during ovarian follicular development in sheep. *Biology of Reproduction* 66: 906-916.
- Lu, C. D. 1989. Effects of heat stress on goat production. *Small Ruminant Research* 2: 151-162.
- Lucy, M. C., Bilby, C. R., Kirby, C. J., Yuan, W. and Boyd, C. K. 1999. Role of growth hormone in development and maintenance of lutea. *Journal of Reproduction and Fertility* 54: 49-59.
- Lucy, M. C. 2000. Regulation of ovarian follicular growth by somatotropin and insulin-like growth factors in cattle. *Journal of Dairy Science* 83: 1635-1647.
- Magistrelli, D., Valli, A. and Rosi, F. 2005. Insulin and IGF-1 in goat milk: Influence of the diet. *Italian Journal of Animal Science* 4: 386-388.
- Magistrelli, D. and Rosi, F. 2009. Plasma insulin and IGF-1 and hepatic activity in Saanen goat kids, around weaning. *Tropical and Subtropical Agroecosystems* 11: 205-208.
- Malan, S. W. 2000. The improved Boer goat. Small Ruminant Research 36: 165-170.
- Martemucci, G. and D'Alessandro, A. G. 2011. Synchronisation of oestrus and ovulation by short time combined FGA,  $PGF_{2\alpha}$ , GnRH, eCG treatments for natural service or AI fixed-time. *Animal Reproduction Science* 123: 32-39.

- Mazerbourg, S., Zapf, J., Bar, R. S., Brigstock, D. R. and Monget, P. 2000. Insulinlike growth factor (IGF)-binding protein-4 proteolytic degradation in bovine, equine, and porcine preovulatory follicles: regulation by IGFs and heparin-binding domain-containing peptides. *Biology of Reproduction* 63: 390-400.
- Mazerbourg, S., Bondy, C., Zhou, J. and Monget, P. 2003. The insulin growth factor system: a key determinant role in the growth and selection of ovarian follicles? a comparative species study. *Reproduction in Domestic Animals* 38: 247-258.
- Menchaca, A. and Rubianes, E. 2002. Relation between progesterone concentrations during the early luteal phase and follicular dynamic in goats. *Theriogenology* 57: 1411-1419.
- Menchaca, A. and Rubianes, E. 2004. New treatments associated with timed artificial insemination in small ruminants. *Reproduction Fertility and Development* 16: 403-413.
- Menchaca, A., Miller, V., Salveraglio, E. and Rubianes, E. 2007. Endocrine, luteal and follicular responses after the use of the short-term protocol to synchronise ovulation in goats *Animal Reproduction Science* 102: 76-82.
- Mihm, M. and Austin, E. J. 2002. The final stages of dominant follicle selection in cattle. *Domestic Animal Endocrinology* 23: 155-166.
- Mihm, M. and Bleach, E. C. L. 2003. Endocrine regulation of ovarian antral follicle development in cattle. *Animal Reproduction Science* 78: 217-237.
- Minka, N. S., Ayo, J. O., Sackey, A. K. B. and Adelaiye, A. B. 2009. Assessment and scoring of stresses imposed on goats during handling, loading, road transportation and unloading, and the effect of pretreatment with ascorbic acid. *Livestock Science* 125: 275-282.
- Monget, P. and Bondy, C. 2000. Importance of the IGF system in early folliculogenesis. *Molecular and Cellular Endocrinology* 163: 89-93.
- Monget, P., Fabre, S., Mulsant, P., Lecerf, F., Elsen, J.-M., Mazerbourg, S., Pisselet, C. and Monniaux, D. 2002. Regulation of ovarian folliculogenesis by IGF and BMP system in domestic animals. *Domestic Animal Endocrinology* 23: 139-154.

- Monniaux, D., Monget, P., Besnard, N., Huet, C. and Pisselet, C. 1997. Growth factors and antral follicular development in domestic ruminants. *Theriogenology* 47: 3-12.
- Montlomelo, K. C., Greyling, J. P. C. and Schwalbach, L. M. J. 2002. Synchronisation of oestrus in goats: the use of different progestagen treatments. *Small Ruminant Research* 45: 45-49.
- Morand-Fehr, P., Boutonnet, J. P., Devendra, C., Dubeuf, J. P., Haenlein, G. F. W., Holst, P., Mowlem, L. and Capote, J. 2004. Strategy for goat farming in the 21st century. 51: 175-183.
- Mostl, E. and Palme, R. 2002. Hormones as indicators of stress. *Domestic Animal Endocrinology* 23: 67-74.
- Nozawa, K. 1991. Domestication and history of goats. In: K., M. (ed.), *Genetic resources of Pig, Sheep and Goat*. Amsterdam: Elsevier Science Publishers. pp. 391-404.
- Obese, F. Y., Humphrys, S., Macmillan, K. L. and Egan, A. R. 2008. Measuring Concentrations of Insulin-Like Growth Factor-I with an Enzyme-Linked Immunosorbent Antibody Assay in Plasma Samples from Holstein Cows. *Journal of Dairy Science* 91: 160-168.
- Oliveira, M. A. L., Guido, S. I. and Lima, P. F. 2001. Comparison of different protocols used to induce and synchronise estrus cycle of Saanen goats. *Small Ruminant Research* 40: 149-153.
- Olivier, J. J., Cloete, S. W. P., Schoeman, S. J. and Muller, C. J. C. 2005. Performance testing and recording in meat and dairy goats. *Small Ruminant Research* 60: 83-93.
- Orskov, E. R. 2011. Goat production on a global basis. *Small Ruminant Research* 98: 9-11.
- Ozawa, M., Tabayashi, D., Latief, T. A., Shimizu, T., Oshima, I. and Kanai, Y. 2005. Alterations in follicular dynamics and steroidogenic abilities induced by heat stress during follicular recruitment in goats. *Reproduction* 129: 621-630.
- Peacock, C. 2005. Goats A pathway out of poverty. *Small Ruminant Research* 60: 179-186.

- Pfeifer, L. F. M., Siqueira, L. G., Mapletoft, R. J., Kastelic, J. P., Adams, G. P., Colazo, M. G. and Singh, J. 2009. Effects of exogenous progesterone and cloprostenol on ovarian follicular development and first ovulation in prepubertal heifers. *Theriogenology* 72: 1054-1064.
- Picton, H. M., Tsonis, C. G. and McNeilly, A. S. 1990. The antagonistic effect of exogenous LH pulses on FSH-stimulated preovulatory follicle growth in ewes chronically treated with a gonadotrophin-releasing hormone agonist. *Journal of Endocrinology* 127: 273-283.
- Picton, H. M. 2001. Activation of follicle development: the primordial follicle. *Theriogenology* 55: 1193-1210.
- Pierce, B. N., Clarke, I. J., Turner, A. I., Rivalland, E. T. A. and Tilbrook, A. J. 2009. Cortisol disrupts the ability of estradiol-17 to induce the LH surge in ovariectomized ewes. *Domestic Animal Endocrinology* 36: 202-208.
- Pikuleva, I. A. 2006. Cytochrome P450s and cholesterol homeostasis. *Pharmacology and Therapeutics* 112: 761-773.
- Pineda, M. H. and Dooley, M. P. 2003. McDonald's Veterinary Endocrinology and Reproduction. 5<sup>th</sup> Edition. Iowa, USA: Iowa State Press. pp. 597.
- Ramwell, P. W., Leovey, E. M. K. and Sintetos, A. C. 1977. Regulation of the arachidonic acid cascade. *Biology of Reproduction* 16: 70-88.
- Randel, R. D., Lammoglia, M. A., Lewis, A. W., Neuendorff, D. A. and Guthrie, M. J. 1996. Exogenous  $PGF_{2\alpha}$  enhanced GnRH-induced LH release in postpartum cows. *Theriogenology* 45: 643-654.
- Regueiro, M., Perez Clariget, R., Ganzabal, A., Aba, M. and Forsberg, M. 1999. Effect of medroxyprogesterone acetate and eCG treatment on the reproductive performance of dairy goats. *Small Ruminant Research* 2: 331.
- Restall, B. J. 1992. Seasonal variation in reproductive activity in Australian goats. *Animal Reproduction Science* 27: 305-318.
- Riesenberg, S., Meinecke-Tillman, Y. and Meinecke, B. 2001. Ultrasonic survey of follicular development following superovulation with a single application of pFSH, eCG or hMG in goats. *Small Ruminant Research* 40: 83-93.

- Roberts, C. A., McCutcheon, S. N., Blair, H. T., Gluckman, P. D. and Breier, B. H. 1990. Developmental patterns of plasma insulin-like growth factor-1 concentrations in sheep. *Domestic Animal Endocrinology* 7: 457-463.
- Rosnina, Y., Jainudeen, M. R. and Nihayah, M. 1992. Superovulation and egg recovery in goats in the tropics. *Veterinary Record* 130: 97-99.
- Rubianes, E. and Menchaca, A. 2003. The pattern and manipulation of ovarian follicular growth in goats. *Animal Reproduction Science* 78: 271-287.
- Rubianes, E., Menchaca, A. and Carbajal, B. 2003. Response of the 1-5 day-aged ovine corpus luteum to prostaglandin  $F_{2\alpha}$ . Animal Reproduction Science 78: 47-55.
- Russel, A. J. F. 1989. The application of real-time ultrasonic scanning in commercial sheep, goats and cattle production enterprise. In: Taverne, M. A. M. and Willemse, A. H. (ed.), *Diagnostic Ultrasound and Animal Reproduction*. Dordrecht, The Netherlands: Kluwer academic publishers. pp. 73-87.
- Sakkinen, H., Tornbeg, J., Goddard, P. J., Eloranta, E., Ropstad, E. and Saarela, S. 2004. The effect of blood sampling method on indicators of physiological stress in reindeer (*Rangifer tarandus tarandus*). Domestic Animal Endocrinology 26: 87-98.
- Sales, J. N. S., Crepaldi, G. A., Girotto, R. W., Souza, A. H. and Baruselli, P. S. 2011. Fixed-time AI protocols replacing eCG with a single dose of FSH were less effective in stimulating follicular growth, ovulation, and fertility in suckled-anestrus Nelore beef cows. *Animal Reproduction Science* 124: 12-18.
- Sangha, G. K., Sharma, R. K. and Guraya, S. S. 2002. Biology of corpus luteum in small ruminants. *Small Ruminant Research* 43: 53-64.
- Sarath, T., Mehrotra, S., Agarwal, S. K., Varshney, V. P., Hoquec, M., Shankar, U. and Singh, S. K. 2008. Effect of insulin administration on ovarian function and estrus induction in acyclic goats. *Animal Reproduction Science* 108: 216-225.
- Savio, J. D., Thatcher, W. W., Badinga, L., de la Sota, R. L. and Wolfenson, D. 1993. Regulation of dominant follicle turnover during the oestrus cycle in cows. *Journal of Reproduction and Fertility* 97: 197-203.

- Sawada, T., Takahara, Y. and Mori, J. 1995. Secretion of progesterone during long and short days of the estrous cycle in goats that are continuous breeders. *Theriogenology* 43: 789-795.
- Scaramuzzi, R. J., Murray, J. F., Downing, J. A. and Campbell, B. K. 1999. The effects of exogenous growth hormone on follicular steroid secretion and ovulation rate in sheep. *Domestic Animal Endocrinology* 17: 269-277.
- Scherf, B. D. (ed.) 2000. World watch list of domestic animal diversity. 3<sup>rd</sup> Edition. Rome: Food and Agriculture Organization of the United Nations. pp. 726.
- Schwarz, T. and Wierzchos, E. 2000. Relationship between FSH and ovarian follicular dynamics in goats during the estrous cycle. *Theriogenology* 53: 381.
- Seekallu, S., Toosi, B., Duggavathi, R., Barrett, D., Davies, K., Waldner, C. and Rawlings, N. 2010. Ovarian antral follicular dynamics in sheep revisited: comparison among estrous cycles with three or four follicular waves. *Theriogenology* 73: 670-680.
- Sejian, V. and Srivastava, R. S. 2010. Effects of melatonin on adrenal cortical functions of Indian goats under thermal stress. *Veterinary Medicine International*. Electronic journal retrieved from: www.hindawi.com/journals/vmi. 348919.
- Shrestha, J. N. B. and Fahmy, M. H. 2005. Breeding goats for meat production: a review: 1. Genetic resources, management and breed evaluation. *Small Ruminant Research* 58: 93-106.
- Shrestha, J. N. B. and Fahmy, M. H. 2007. Breeding goats for meat production 3. Selection and breeding strategies. *Small Ruminant Research* 67: 113-125.
- Silanikove, N. 2000a. The physiological basis of adaptation in goats to harsh environments. *Small Ruminant Research* 35: 181-193.
- Silanikove, N. 2000b. Effects of heat stress on the welfare of extensively managed domestic ruminants. *Livestock Production Science* 76: 1-18.
- Silva, J. R. V., Figueiredo, J. R. and Van den Hurk, R. 2009. Involvement of growth hormone (GH) and insulin-like growth factor (IGF) system in ovarian folliculogenesis. *Theriogenology* 71: 1193-1208.

- Simoes, J., Potes, J., Azevedo, J., Almeida, J. C., Fontes, P., Baril, G. and Mascarenhas, R. 2005. Morphometry of ovarian structures by transrectal ultrasonography in Serrana goats. *Animal Reproduction Science* 85: 263-273.
- Simoes, J., Almeida, J. C., Valentim, R., Baril, G., Azevedo, J., Fontes, P. and Mascarenhas, R. 2006. Follicular dynamics in Serrana goats. *Animal Reproduction Science* 95: 16-25.
- Simoes, J., Baril, G., Almeida, J. C., Valentim, R., Azevedo, J., Fontes, P. and Mascarenhas, R. 2008. Time of ovulation in nulliparous and multiparous goats. *Animal* 2: 761-768.
- Smitz, J. and Cortvrindt, R. 2002. The earliest stages of folliculogenesis in vitro. *Reproduction* 123: 185-202.
- Sodiq, A., Adjisoedarmo, S. and Tawfik, E. S. 2003. Reproduction rate of Kacang and peranakan Etawah goats under village production systems in Indonesia. Paper presented at the International Research on Food Security, Natural Resource Management and Rural Development, Deutscher Tropentag – Göttingen, Germany 8-10, October, pp 1-7. retrieved from http://www.tropentag.de/2003/abstracts/full/143.pdf
- Sousa, F. C., Melo, C. H. S., Albuquerque Teles Filho, A. C., Avelar, S. R. G., Alencar Araripe Moura, A., Martins, J. A. M., Figueirêdo Freitas, V. J. and Teixeira, D. I. A. 2011. Ovarian follicular response to different hormonal stimulation treatments in Canindé goats. *Animal Reproduction Science* 125: 88-93.
- Spahr, I. L. (2009). Body Condition Scoring in Meat Goats. Retrieved from: http://bedford.extension.psu.edu/agriculture/goat.htm
- Spicer, L. J., Enright, W. J., Murphy, M. G. and Roche, J. F. 1991. Effect of dietary intake on concentrations of insulin-like growth factor-I in plasma and follicular fluid, and ovarian function in heifers. *Domestic Animal Endocrinology* 8: 431-437.
- Spicer, L. J., Crowe, M. A., Prendiville, D. J., Goulding, D. and Enright, W. J. 1992. Systemic but not intraovarian concentrations of insulin-like growth factor-I are affected by short-term fasting. *Biology of Reproduction* 46: 920-925.

- Spicer, L. J., Hanrahan, J. P., Zavy, M. T. and Enright, W. J. 1993. Relationship between ovulation rate and concentrations of insulin-like growth factor-1 in plasma during the oestrous cycle in various genotypes of sheep. *Journal of Reproduction and Fertility* 97: 403-409.
- Spicer, L. J. and Echternkamp, S. E. 1995. The ovarian insulin and insulin-like growth factor system with an emphasis on domestic animals. *Domestic Animal Endocrinology* 12: 223-245.
- Stevenson, J. S. 2007. Clinical reproductive physiology of the cow. In: Youngquist, R. S. and Threlfall, W. R. (ed.), *Current Therapy in Large Animal Theriogenology*. 2<sup>nd</sup> Edition. St. Louis, Missouri, USA: Saunders, Elsevier. pp. 23-47.
- Sudo, N., Shimizu, T., Kawashima, C., Kaneko, E., Tetsuka, M. and Miyamoto, A. 2007. Insulin-like growth factor-I (IGF-I) system during follicle development in the bovine ovary: Relationship among IGF-I, type 1 IGF receptor (IGFR-1) and pregnancy-associated plasma protein-A (PAPP-A). *Molecular and Cellular Endocrinology* 264: 197-203.
- Suganuma, C., Kuroiwa, T., Tanaka, T. and Kamomae, H. 2007. Changes in the ovarian dynamics and endocrine profiles in goats treated with a progesterone antagonist during the early luteal phase of the estrous cycle. *Animal Reproduction Science* 101: 285-294.
- Swali, A. and Wathes, D. C. 2006. Influence of the dam and sire on size at birth and subsequent growth, milk production and fertility in dairy heifers. *Theriogenology* 66: 1173-1184.
- Takada, T., Kikkawa, Y., Yonekawa, H., Kawakami, S. and Amano, T. 1997. Bezoar (*Capra aegagrus*) is a matriarchal candidate for ancestor of domestic goat (*Capra hircus*): Evidence from the mitochondrial DNA diversity. *Biochemical Genetics* 35: 315-326.
- Taylor, V. J., Beever, D. E., Bryant, M. J. and Wathes, D. C. 2004. First lactation ovarian function in dairy heifers in relation to prepubertal metabolic profiles. *Journal of Endocrinology* 180: 63-75.
- Torres-Júnior, J. R., Pires, M., de Sá, W. F., Ferreira, A., Viana, J. H. M., Camargo, L. S. A., Ramos, A. A., Folhadella, I. M., Polisseni, J., de Freitas, C., Clemente, C. A. A., de Sá Filho, M. F., Paula-Lopes, F. F. and Baruselli, P. S. 2008. Effect of maternal heat-stress on follicular growth and oocyte competence in Bos indicus cattle. *Theriogenology* 69: 155-166.

- Vazquez, M. I., Blanch, M. S., Alanis, G. A., Chaves, M. A. and Gonzalez-Bulnes, A. 2010. Effects of treatment with a prostaglandin analogue on developmental dynamics and functionality of induced corpora lutea in goats. *Animal Reproduction Science* 118: 42-47.
- Velazquez, M. A., Newman, M., Christie, M. F., Cripps, P. J., Crowe, M. A. and Smith, R. F. 2005. The usefulness of a single measurement of insulin-like growth factor-1 as a predictor of embryo yield and pregnancy rates in a bovine MOET program. *Theriogenology* 64: 1977-1994.
- Velazquez, M. A., Spicer, L. J. and Wathes, D. C. 2008. The role of endocrine insulin-like growth factor-I (IGF-I) in female bovine reproduction. *Domestic Animal Endocrinology* 35: 325-342.
- Viudes De Castro, M. P., Cortell, C., Mocé, E., Marco-Jiménez, F., Joly, T. and Vicente, J. S. 2009. Effect of recombinant gonadotropins on embryo quality in superovulated rabbit does and immune response after repeated treatments. *Theriogenology* 72: 655-662.
- Webb, R., Campbell, B. K., Garverick, H. A., Gong, J. G., Gutierrez, C. G. and Armstrong, D. G. 1999. Molecular mechanisms regulating follicular recruitment and selection. *Journal of Reproduction and Fertility* 54: 33-48.
- Webb, R., Woad, K. J. and Armstrong, D. G. 2002. Corpus luteum (CL) function: local control mechanisms. *Domestic Animal Endocrinology* 23: 277-285.
- Webb, R., Garnsworthy, P. C., Gong, J. G. and Armstrong, D. G. 2004. Control of follicular growth: Local interactions and nutritional influences. *Journal of Animal Science* 82: E63-74.
- Weems, C. W., Weems, Y. S. and Randel, R. D. 2006. Prostaglandins and reproduction in female farm animals. *The Veterinary Journal* 171: 206-228.
- Whitley, N. C. and Jackson, D. J. 2004. An update on estrus synchronisation in goats: A minor species. *Journal of Animal Science* 82: E270-E276.
- Wildeus, S. 2000. Current concepts in synchronisation of estrus: Sheep and goats. *Journal of Animal Science* 77: 1-14.

- Wu, M., Wang, A., Bernard, G. C., Hall, J. B., Beal, W. E., Michael, A. R., Boisclair, Y. R. and Jiang, H. M. 2008. Increased degradation of insulin-like growth factor-I in serum from feed-deprived steers. *Domestic Animal Endocrinology* 35: 343-351.
- Yilmaz, A., Davis, M. E. and Simmen, R. C. M. 2006. Analyses of female reproductive traits in Angus beef cattle divergently selected for blood serum insulin-like growth factor I concentration. *Theriogenology* 65: 1180-1190.
- Yu, Y. S., Luo, M. J., Han, Z. B., Li, W., Sui, H. S. and Tan, J. H. 2005. Serum and follicular fluid steroid levels as related to follicular development and granulosa cell apoptosis during the estrous cycle of goats. *Small Ruminant Research* 57: 57-65.
- Zarkawi, M., AI-Merestani, M. R. and Wardeh, M. F. 1999. Induction of synchronised oestrous in indigenous Damascus goats outside the breeding season. *Small Ruminant Research* 33: 193-197.
- Zeder, M. A. 2000. The initial domestication of goats (*Capra hircus*) in the Zagros mountains 10,000 years ago. *Science* 287: 2254-2257.
- Zhao, Y., Zhang, J., Wei, H., Sun, X., Mu, B., Yu M. and Wang, L. 2010. Efficiency of methods applied for goat estrous synchronisation in subtropical monsoonal climate of South-west China. *Tropical Animal Health and Production* 42: 1257-1262.
- Zohary, D., Tchernov, E. and Horwitz, L. K. 1998. The role of unconscious selection in the domestication of sheep and goats. *Journal of Zoology* 245: 129-135.