



UNIVERSITI PUTRA MALAYSIA

***EFFECTS OF DRYING METHODS AND SOLVENT TYPES ON
ANTIOXIDANT PROPERTIES AND ANTI-INFLAMMATORY POTENTIAL
OF TWO GINGER VARIETIES IN MALAYSIA***

ISWAIBAH BINTI MUSTAFA

FK 2017 83



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By

ISWAIBAH BINTI MUSTAFA

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

May 2017

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Dedicated to,

My beloved father and mother, Mustafa Yusoff and Nafisah Abd. Rahman who take a big role in completing this journey. Thank you so much for your continuous love and support in many ways.

Also to all my siblings Kak, Abe, Yah, Ijad, Adam and Adik, thank you for your endless support. All of you are my priceless treasure in this duniya. I love you for ∞ (eternity)

ALHAMDULILLAH



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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By

ISWAIBAH BINTI MUSTAFA

May 2017

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Faculty: Engineering

Ginger (*Zingiber officinale*) is one of the finest medicinal herbs in the world with more than 50% of the world production coming from Asia. In Malaysia, ginger ranks among top ten potential herbs based on its remarkable medicinal properties. However, most usage of ginger is from its fresh form which causes huge postharvest loss. The objective of this study was to systematically evaluate the antioxidant properties and anti-inflammatory activity of Malaysian gingers, Halia Bentong (*Zingiber officinale* Roscoe) and Halia Bara (*Zingiber officinale* var. *Rubra*) as affected by drying methods of shade-, sun-, oven-, vacuum-, and freeze-drying with different extraction solvents of hotwater, 80% aqueous ethanol and ethanol. The changes in dried ginger extracts were analyzed for proximate analysis, phytochemical contents and antioxidant activities using FRAP, TAA by phosphomolybdenum, ability to scavenge ABTS^{•+}, DPPH[•] and OH[•] radicals. Chelation on ferrous ion was then evaluated for metal binding capacity. Anti-inflammatory activity was done using cell culture method against inhibition of NO production in LPS-stimulated RAW 264.7 cells.

The results show that drying methods have helped to reduce the moisture content of ginger whilst preserving the ginger's nutritional value, phytochemical contents as well as their antioxidant properties. Phenolic contents of dried ginger, with the highest value of 20.91 mg GAE/g extract for Halia Bentong and 20.07 mg GAE/g extract for Halia Bara were obtained in freeze-dried from ethanol extract and the lowest in fresh ginger with 8.03 mg GAE/g extract and 10.53 mg GAE/g extract, respectively. Flavonoid contents also were highest in dried gingers, which were ranged from 42.98 to 67.82 g RE/100g extract for Halia Bentong and 47.32 to 69.32 g RE/100g extract for Halia Bara when compared to fresh ginger. Meanwhile, ascorbic acid in fresh and dried gingers were less than 1.5 mg AA/g extract, which indicate that both gingers contain less amount of ascorbic acid. Shade-dried of Halia Bentong and sun-dried of Halia Bara from ethanol extracts gave the highest total antioxidant activity of ginger which were 75.23 g AA/100g extract and 81.12 g AA/100g extract, respectively. Sun-dried from ethanol extracts had

the highest antioxidant activity as exhibited by FRAP and ABTS^{•+} scavenging activity. The activity was increased by 15.35-fold and 3.5-fold for dried Halia Bentong and 3.95-fold and 2.07-fold for dried Halia Bara compared to its fresh form. However, different methods of drying did not affect the activity of inhibition of DPPH[•], OH[•] as well as the ascorbic acid contents.

Extraction solvent has a significant influence ($p < 0.05$) on the extraction of antioxidant compounds from ginger. Ethanol extracts of ginger gave the highest level of antioxidant activities as exhibited by phytochemical contents, FRAP, total antioxidant activity (TAA), ABTS^{•+} scavenging and IC₅₀ values, while hotwater extracts showed the least potent solvent extraction. However, hotwater was found as the most effective solvent in giving the strongest of ferrous ion chelating power with the ranged of 69.45 to 145.47 mg EDTA/g extract for Halia Bentong and 60.73 to 159.17 mg EDTA/g extract for Halia Bara. The extractive yield indicates that aqueous ethanol of 80% which ranged from 2.39 to 17.79% for Halia Bentong and 3.57 to 15.21% for Halia Bara, gave the maximum yield amongst the other extracts. Results suggested that not only the polarity of the solvent affected the effectiveness of extraction, the chemical structure or nature of the solvent also lead to different antioxidant capability.

The enhanced phytochemicals and antioxidant activities of dried ginger were also supported by anti-inflammatory studies against inhibition of NO production, which is known as inflammation mediator. Dried ginger from ethanol extracts were recorded as a potent inhibitor of NO at the concentrations 100 µg/mL, while fresh ginger was less potent. At the concentration of 100 µg/mL extracts, NO-inhibitory activity of dried ginger had caused inhibition with ranged from 40.85 to 50.88% for Halia Bentong and 29.16 to 45.13% for Halia Bara. While inhibitory activity of fresh ginger on NO production was 8.65% and 18.23%, respectively. Drying has demonstrated an improved post-harvest treatment of gingers' bioactivities in regard to enhanced final ginger quality.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

KESAN KAEDAH PENGERINGAN DAN JENIS PELARUT TERHADAP CIRI-CIRI ANTIOKSIDAN DAN ANTI-RADANG DARIPADA DUA JENIS HALIA MALAYSIA

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Halia adalah salah satu herba perubatan terbaik di dunia dengan lebih daripada 50% daripada pengeluaran dunia adalah dari Asia. Di Malaysia, halia berada di kedudukan sepuluh herba berpotensi berdasarkan sifat-sifat perubatannya. Walau bagaimanapun, kebanyakan penggunaan halia adalah dalam keadaan segar yang menyebabkan kerugian yang besar selepas proses menuai. Oleh itu, objektif kajian ini adalah untuk mengkaji sifat-sifat antioksidan dan aktiviti anti-radang halia Malaysia iaitu Halia Bentong dan Halia Bara yang dipengaruhi oleh kaedah pengeringan yang berbeza dengan menggunakan pelarut pengekstrakan yang berbeza. Kaedah pengeringan yang digunakan ialah teduhan, matahari, ketuhar, vakum dan beku. Manakala, pelarut pengekstrakan yang digunakan ialah air panas, 80% etanol akueus dan etanol. Perubahan dalam ekstrak halia kering telah dianalisis untuk komposisi proksimat, kandungan fitokimia dan aktiviti antioksidan dengan menggunakan FRAP, phosphomolybdenum (TAA), ABTS^{•+} kation radikal, DPPH[•] dan OH[•] radikal. Perebutan pada ion Fe²⁺ telah dinilai untuk aktiviti pengikatan logam. Aktiviti anti-radang telah dilakukan dengan menggunakan kaedah kultur sel terhadap pengurangan bagi pengeluaran NO yang dirangsang oleh LPS pada sel RAW 264.7.

Hasil kajian menunjukkan bahawa kaedah pengeringan bukan saja membantu untuk mengurangkan kandungan lembapan halia, tetapi mengekalkan nilai nutrisi, kandungan fitokimia dan sifat antioksidan. Pengeringan beku dari pelarut etanol memberi nilai fenol yang paling tinggi iaitu 20.91 mg GAE/g ekstrak untuk Halia Bentong dan 20.07 mg GAE/g ekstrak untuk Halia Bara, dan halia segar masing-masing dengan nilai yang paling rendah iaitu 8.03 mg GAE/g ekstrak dan 10.53 mg GAE/g ekstrak. Kandungan flavonoid juga lebih tinggi dalam halia kering, yang antara 42.98-67.82 g RE/ekstrak 100g untuk Halia Bentong dan 47.32-69.32 g RE/ekstrak 100g untuk Halia Bara berbanding halia segar. Sementara itu, asid askorbik dalam halia segar dan halia kering adalah kurang daripada 1.5 mg AA/g ekstrak, yang menunjukkan halia mengandungi kandungan asid askorbik yang rendah. Halia Bentong dari kaedah pengeringan teduhan

dan Halia Bara dari kaedah pengeringan matahari, yang masing-masing dari ekstrak etanol menunjukkan jumlah aktiviti antioksidan yang paling tinggi halia iaitu dengan 75.23 g AA/100g ekstrak dan 81.12 g AA/100g ekstrak. Manakala kaedah pengeringan matahari dari ekstrak etanol mempunyai aktiviti antioksidan yang paling tinggi seperti yang ditunjukkan oleh FRAP, dan ABTS⁺ kation radikal. Aktiviti ini telah meningkat sebanyak 15.35 dan 3.5 kali ganda untuk Halia Bentong kering; dan 3.95 dan 2.07 kali ganda untuk Halia Bara kering berbanding halia segar. Walaubagaimanapun, kaedah pengeringan yang berbeza tidak menunjukkan sebarang perubahan dalam aktiviti DPPH[•], OH[•] radikal dan juga kandungan asid askorbik.

Kesan pengekstrakan pelarut mempunyai pengaruh yang ketara ($p < 0.05$) terhadap antioksidan dari halia. Ekstrak halia dari pelarut etanol memberikan nilai aktiviti antioksidan yang tinggi seperti yang ditunjukkan oleh kandungan fitokimia, FRAP, TAA, ABTS⁺ dan nilai IC₅₀. Manakala pengekstrakan menggunakan air panas memberikan activity yang paling minimum. Walau bagaimanapun, air panas didapati sebagai pelarut yang paling berkesan dalam aktiviti pengikatan ion ferus dengan nilai 69.45-145.47 mg EDTA/g ekstrak untuk Halia Bentong dan 60.73-159.17 mg EDTA/g ekstrak untuk Halia Bara. Hasil ekstrak menunjukkan bahawa 80% etanol akueus memberikan hasil yang maksimum iaitu antara 2.39-17.79% untuk Halia Bentong dan 3.57-15.21% untuk Halia Bara. Kajian menunjukkan bahawa bukan sahaja keketuban pelarut mempengaruhi kesan pengekstrakan, struktur kimia atau sifat pelarut juga membawa kepada keupayaan antioksidan yang berbeza.

Kandungan fitokimia dan aktiviti antioksidan halia kering juga disokong oleh kajian anti-radang terhadap pengurangan bagi pengeluaran NO, yang juga dikenali sebagai pengantara kepada sifat keradangan. Halia kering dari ekstrak etanol menunjukkan sebagai jumlah keluaran NO yang sedikit pada 100 µg/mL, manakala halia segar menunjukkan jumlah pengeluaran NO yang banyak. Pada kepekatan ekstrak 100 µg/mL, aktiviti pengurangan NO untuk halia kering adalah antara 40.85-50.88% untuk Halia Bentong dan 29.16-45.13% untuk Halia Bara. Manakala pengurangan NO untuk halia segar masing-masing pada 8.65% dan 18.23%. Kaedah pengeringan telah menunjukkan peningkatan yang lebih baik terhadap aktiviti biologi halia.

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I certify that a Thesis Examination Committee has met on 18 May 2017 to conduct the final examination of Iswaibah binti Mustafa on her thesis entitled "Effects of Drying Methods and Solvent Types on Antioxidant Properties and Anti-Inflammatory Potential of Two Ginger Varieties in Malaysia" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

ABTS	2,2'-azinobis (3-ethyl-benzothiozoline-6-sulfonic acid) disodium salt
ANOVA	Analysis of variance
AOAC	Official methods of analysis
BHA	Butylated hydroxyanisole
DCPIP	2,6-dichlorophenolindophenol
DMEM	Dulbecco's modified eagle's medium
DPPH	1,1- diphenyl-1-picrylhydrazyl
EDTA	Ethylenediaminetetraacetic acid
FBS	Fetal bovine serum
FRAP	Ferric reducing/antioxidant power
LPS	Lipopolysaccharide
NO	Nitric oxide
PM	Phosphomolybdenum
TAA	Total antioxidant activity
TPTZ	2,4,6-tripyridyl-s-triazine
Trolox	6-hydroxy-2,5,7,8-tetramethylchromane-2-carboxylic acid

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CHAPTER 1

INTRODUCTION

1.1 Research Background

Ginger, a rhizome of the plant *Zingiber officinale* is one the most valuable medicinal herbs in the world with its innumerable uses in food and medicines. The fresh ginger has been used from ancient times as food and medicine and has gained considerable awareness for various functional benefits such as dietary supplement, food antioxidants, and as a food product for preventing diseases (Ho & Su, 2016; Yeh et al., 2013). In Malaysia, *Zingiber officinale* Roscoe and *Zingiber officinale* var. *Rubra* respectively known as Halia Bentong and Halia Bara are widely used by the locals as a major herb, health food and as an important flavouring and traditional medicine (Mojani & Ghasemzadeh, 2014). Due to its nutritive value and presence of potent bioactive compounds, the rhizome ginger are gaining importance for its health benefits and are emerging as a potential ingredients among the nutrition market. Analysis of nutritive value of ginger has been performed by several researchers (Butt & Sultan, 2011; Ghosh, 2011). The rhizome gingers are rich in minerals, vitamin, carbohydrate and other proximate compositions. Apart from being nutritious, ginger are rich sources of bioactive compounds which have shown to be effective in scavenging free radicals and to treat inflammation (Dugasani et al., 2010; Tsai, Tsai, & Ho, 2005).

However, the fresh ginger are sometimes seasonal, highly perishable and have high content of moisture. High moisture content can cause ginger to be prone to microorganism spoilage such as enzymatic reactions and other detrimental changes (Akdaş & Başlar, 2015; Roshanak, Rahimmalek, & Goli, 2016). Drying is a common traditional preservation method which significantly reduces the amount of moisture contents, and also extent the shelf-life of ginger. Despite improving the shelf life of ginger, drying processing are also recognized as one of the major factors leading to changes of natural phytochemical present in foods (Kamiloglu et al., 2016).

The health-promoting behavior of ginger is attributed to its rich natural phytochemicals, which have been widely discovered as having antioxidants and anti-inflammatory properties, protecting human from oxidative stress and inflammation related disorders. Oxidative stress is related to an imbalance between the production of free radicals and antioxidant defense system. Free radicals and their uncontrolled production will initiate and propagate the oxidation chain reactions and lead to inflammation development (Sagrin & Chong, 2013). Inflammation, which is a functionally defense response of living tissue to harmful stimuli, can be associated with uncontrolled or prolonged inflammatory responses that develop when the body's regulation of inflammation is dysfunctional (Cheung et al., 2013; Shin et al., 2013). Since a number of complex reactions take place during drying processing, the final

quality of ginger may be the results of degradation of natural phytochemicals and/or formation of new compounds of phytochemicals that further affect the therapeutic value in the dried ginger when compared to fresh ginger.

A work done on green tea leaves reported that drying processing enhanced the final product than fresh samples in terms of phytochemical contents, antioxidant activity and vitamin C (Roshanak et al., 2016). Hossain et al. (2010) observed that an increase in phytochemical contents and antioxidant activity through drying treatment of Lamiaceae herbs (Hossain, Barry-Ryan, Martin-Diana, & Brunton, 2010). Joshi et al. (2011) and Chan et al. (2013) suggested that drying treatment was able to preserve phenolic content and enhance antioxidant activity of apple and herbs, respectively (Chan, Lye, Eng, & Tan, 2013; Joshi, Rupasinghe, & Khanizadeh, 2011). In contrast, phenolic content in dried tomato (Gümüşay, Borazan, Ercal, & Demirkol, 2015) and dried mandarin (Akdaş & Başlar, 2015) were found to decrease approximately 60% and 40% from the fresh samples, respectively. The above findings have proven the drying methods may affect the natural compounds in either ways.

Therefore, this research aims to identify the photochemical contents of fresh and dried rhizome ginger as subjected to different drying treatments evaluated in terms of antioxidant and anti-inflammatory activities.

1.2 Problem Statement

Ginger has been identified as an herbal medicinal product with beneficial bioactivity effects (Yeh et al., 2013). Ginger contains a large number of active ingredients and shows a therapeutic role in the treatment of many diseases that involve inflammation and those which caused by oxidative stress (Dugasani et al., 2010). Previous study reported that some bioactive constituents present in ginger plays a crucial role in antioxidant, anti-inflammatory, anti-tumour, anti-microbial, anti-diabetic, anti-emetic, and etc. (Rahmani, Al, & Aly, 2014). In traditional practices, ginger is used to treat a wide range of ailments, including nausea and vomiting, colds and flu symptoms, stomachaches and rheumatic disorders (Kubra & Rao, 2012). Currently, two varieties of ginger rhizomes, Halia Bentong and Halia Bara are available in Malaysia and found to have potential source for natural antioxidants (Mojani & Ghasemzadeh, 2014).

Unfortunately, ginger postharvest period is relatively short due to their high moisture contents. High moisture content is highly susceptible to oxidize by external (atmospheric) and internal (enzymatic action) degradation (Roshanak et al., 2016), which in turn causes hygienic and qualitative problems. This problem also affects its phytochemicals compounds and potential bioactivity of ginger. It has been demonstrated that the health benefits of ginger against oxidative stress and inflammation diseases are mainly attributed to the occurrence of bioactive phytochemicals of phenolic and flavonoids compounds (Dugasani et al., 2010). Oxidative stress is related to an imbalance between the production of free radicals and

antioxidant defense system. Free radicals and their uncontrolled production will initiate and propagate the oxidation chain reactions and lead to inflammation development (Lu & Yen, 2015). In inflammatory disorders, nitric oxide (NO) is secreted excessively and overproduction or prolonged of NO has resulted the diseases developments. The extracts of ginger were reported to possess scavenging properties against free radicals, and exhibit anti-inflammation by inhibiting NO (Dugasani et al., 2010; Tsai et al., 2005). Phenolic and flavonoids are major contributors to its protective role against oxidative damage and its consequences such as inflammation and others, thus contributing to the overall health-protective effects of food samples.

Hence, drying is applied to prolong the shelf life of ginger and to preserve their natural phytochemical efficiency (Gümüşay et al., 2015). More recently, improvement of ginger in terms of its potential bioactivity by drying processing has been well documented (Chumroenphat, Khanprom, & Butkhup, 2011; Guo, Wu, Du, Zhang, & Yang, 2014). Drying inhibits the microbial growth and prevents the changing of certain biochemical components that might alter the antioxidant activity. Though ginger has been used by local communities for cuisines and medicine, drying processes information is still lack especially on antioxidant and anti-inflammatory activities, in particular the Malaysian ginger. Currently, there has been an increased interest to identify therapeutic potentials of natural antioxidants from plant sources which are known with their pharmacology effect with less or no side effects. As plants counteract oxidative stress caused by environmental stressor, they present a potential source of natural compounds with antioxidant activity to themselves (Azmir et al., 2013; Sudan, Bhagat, Gupta, Singh, & Koul, 2014). Antioxidant of their phytochemicals could give an additional support to the human health.

Extraction is an important step involved in the discovery of bioactive phytochemicals compounds from plant materials. A successful determination of bioactive components from plant materials most often involves type of solvent used in the extraction procedures. Many researchers on the extraction of bioactive compounds from ginger using various solvents are found despite no explanation forthcoming on their choice of selection. They include using methanol, ethanol, acetone, hexane, ethyl acetate, petrol, water, chloroform, benzene, and butanol. Among all the solvents used, ethanol, water, and their combinations (aqueous organic solvent) are the most commonly used solvent as they are considered to be GRAS solvents. However, it was well documented that different solvent will exhibit different potentiality in extracting bioactive compounds from plant materials and possess different potentiality in bioactivities (Loo, Jain, & Darah, 2008). Lapornik et al. (2005) who studied antioxidant activity of berries in ethanol and water extracts reported that ethanol extracts contain twice more bioactive phytochemicals of anthocyanins and polyphenols than water extracts. While, Benjakul et al. (2014) reported that water extraction produces higher yield of lead seed (*Leucaena leucocephala*) extracts, whereas lower yield of extracts were obtained in ethanol extracts. Additionally, water was suggested as a good solvent in metal chelating activity, as reported by Sowndhararajan et al. (2010). Metal chelating capacity is evaluated as one of the important mechanisms of antioxidant activity. Thus, the reported bioactivity in plant extracts using various extraction solvents are quite different. Despite the fact that some few recent studies have reported the extraction of bioactive compounds from ginger (Ho & Su, 2016; Li et al., 2011; Oboh, Akinyemi, & Ademiluyi, 2012), there is not any study on the extraction of bioactive compounds

from Malaysian ginger using different solvents such as water, ethanol, and their combinations (mixtures of water-ethanol). In realizing this void, this research has served as an initial exploration for extracting antioxidant compounds from Malaysian ginger with different types of solvents and evaluated in-terms of their antioxidant activity.

1.3 Objectives

The general objectives of this research are to study the effects of different drying methods and solvents extraction on extract yields, proximate compositions, phytochemicals contents, antioxidant and anti-inflammatory activities of two Malaysian ginger varieties which are Halia Bentong and Halia Bara. The specific objectives are:

1. To compare the effectiveness of different drying methods of shade-, sun-, oven-, vacuum-, and freeze-drying on proximate composition of dried ginger; and phytochemical contents and antioxidant activities of extract ginger of both Halia Bentong and Halia Bara.
2. To evaluate the antioxidant activities of dried ginger extracted from three different solvents; hotwater, aqueous ethanol and ethanol, in terms of extract yields, phytochemical contents and antioxidant activities
3. To study anti-inflammatory activity of ethanolic extracts from sun-, oven-, and freeze-dried ginger by inhibiting the nitric oxide (NO) production in LPS-stimulated RAW 264.7 cells.

1.4 Research Scope

The research covers the scope of investigation on the effects of drying methods on antioxidant and anti-inflammatory activities of two Malaysian ginger varieties, Halia Bentong and Halia Bara. Five different drying methods applied were the shade-, sun-, oven-, vacuum-, and freeze-drying. Their changes in antioxidant activities were evaluated and compared with fresh ginger. Subsequently of the drying process, the proximate analyses of ginger were analyzed. The proximate analysis involved moisture, fat, crude fibre, protein, ash and carbohydrate. Different types of solvents were also evaluated to examine their potentiality in extraction of bioactive compounds from ginger. To achieve this target, three different solvents, hotwater, aqueous ethanol (80%, w/v) and ethanol were used. The solid-liquid extraction (maceration technique) was adopted in this study during extraction process to extract bioactive compounds from the

ginger rhizome. Phytochemical content (*i.e.* phenolic, flavonoid and ascorbic acid contents), reductive ability (FRAP), total antioxidant activity by phosphomolybdenum assay (TAA), radical scavenging activity (*i.e.* ABTS⁺, DPPH[•] and OH[•]) and metal chelating activity (Fe²⁺) were carried out in this study to examine the effects of drying methods and solvent extraction. The phytochemical and antioxidant activities of dried ginger extracts were supported by anti-inflammatory studies with cell culture approach on inhibition of NO production in LPS-stimulated RAW 264.7 cells.



REFERENCES

- Akdaş, S., & Başlar, M. (2015). Dehydration and degradation kinetics of bioactive compounds for mandarin slices under vacuum and oven drying conditions. *Journal of Food Processing and Preservation*, 39(6), 1098–1107.
- Alfaro, M. J., Bélanger, J. M. R., Padilla, F. C., & Paré, J. R. J. (2003). Influence of solvent, matrix dielectric properties, and applied power on the liquid-phase microwave-assisted processes (MAP™) extraction of ginger (*Zingiber officinale*). *Food Research International*, 36(5), 499–504.
- Ali, B. H., Blunden, G., Tanira, M. O., & Nemmar, A. (2008). Some phytochemical, pharmacological and toxicological properties of ginger (*Zingiber officinale* Roscoe): A review of recent research. *Food and Chemical Toxicology*, 46(2), 409–420.
- Allothman, M., Bhat, R. and Karim, A.A. (2009). Antioxidant capacity and phenolic content of selected tropical fruits from Malaysia, extracted with different solvents *Food Chemistry* 115: 785–788.
- Amenour, M., Sendra, E., Abrini, J., Pérez-Alvarez, J. A., & Fernández-López, J. (2010). Antioxidant activity and total phenolic compounds of myrtle extracts. *CyTA - Journal of Food*, 8(2), 95–101.
- Anwar, F., Kalsoom, U., Sultana, B., Mushtaq, M., Mehmood, T., & Arshad, H. A. (2013). Effect of drying method and extraction solvent on the total phenolics and antioxidant activity of cauliflower (*Brassica oleracea* L.) extracts. *International Food Research Journal*, 20(2), 653–659.
- AOAC. (1990). Official methods of analysis (15th edition.). Washington, DC, USA: Association of Official Analytical Chemists.
- Aparadh, V. T., Aik, N. V. V., & Aradge, B. a. (2012). Antioxidative properties (TPC, DPPH, FRAP, metal chelating ability, reducing power and TAC) within some cleome species. *Annalica Di Botanica*, 2(1915), 49–56.
- Arabshahi-Delouee, S., & Urooj, A. (2007). Antioxidant properties of various solvent extracts of mulberry (*Morus indica* L.) leaves. *Food Chemistry*, 102(4), 1233–1240.
- Arunachalam, K., & Parimelazhagan, T. (2012). Evaluation of nutritional composition and antioxidant properties of underutilized *Ficus talboti* King fruit for nutraceuticals and food supplements. *Food Science and Technology*, 51(7), 1260–1268.
- Arunachalam, K., Saravanan, S., & Parimelazhagan, T. (2011). Nutritional analysis and antioxidant activity of palmyrah (*Borassus flabellifer* L.) seed embryo for potential use as food source. *Food Science and Biotechnology*, 20(1), 143–149.

- Avinash, G. R. K., Trilok, M. M., Shilpa, T., Shabnam, S., & Satish, K. B. (2012). Variation of phenols, flavonoids and antioxidant potential in various parts of *Foeniculum vulgare* on drying. *International Journal of Chemical and Pharmaceutical Sciences*, 3(1), 74–79.
- Azmir, J., Zaidul, I. S. M., Rahman, M. M., Sharif, K. M., Mohamed, et al. (2013). Techniques for extraction of bioactive compounds from plant materials: A review. *Journal of Food Engineering*, 117(4), 426–436.
- Ballard, T.S., Mallikarjunan, P., Zhou, K. and O’Keefe, S.F. (2009). Optimizing the extraction of phenolic antioxidants from peanut skins using response surface methodology. *Journal Agricultural and Food Chemistry* 57: 3064–3072.
- Barros, L., Ferreira, M. J., Queirós, B., Ferreira, I. C. F. R., & Baptista, P. (2007). Total phenols, ascorbic acid, β -carotene and lycopene in Portuguese wild edible mushrooms and their antioxidant activities. *Food Chemistry*, 103, 413–419
- Benjakul, S., & Kittiphattanabawon, P. (2014). Antioxidant activities of lead (*Leucaena leucocephala*) seed as affected by extraction solvent, prior dechlorophyllisation and drying methods. *Journal of Food Science and Technology*, 51(11), 3026–3037.
- Boeing, J. S., Barizão, É. O., e Silva, B. C., Montanher, P. F., de Cinque Almeida, V., & Visentainer, J. V. (2014). Evaluation of solvent effect on the extraction of phenolic compounds and antioxidant capacities from the berries: Application of principal component analysis. *Chemistry Central Journal*, 8(48), 1-9.
- Boligon, A. A., Machado, M. M., & Athayde, M. L. (2014). Medicinal chemistry technical evaluation of antioxidant activity. *Medicinal Chemistry*, 4(7), 517–522.
- Brand-Williams, W., Cuvelier, M. E., & Berset, C. (1995). Use of a free radical method to evaluate antioxidant activity. *LWT - Food Science and Technology*, 28(1), 25–30.
- Butt, M. S., & Sultan, M. T. (2011). Ginger and its health claims: molecular aspects. *Critical Reviews in Food Science and Nutrition*, 51(5), 383–393.
- Caceres, P. J., Penas, E., Martinez-Villaluenga, C., Amigo, L., & Frias, J. (2017). Enhancement of biologically active compounds in germinated brown rice and the effect of sun-drying. *Journal of Cereal Science*, 73, 1–9.
- Carocho, M., & Ferreira, I. C. F. R. (2013). A review on antioxidants, prooxidants and related controversy: natural and synthetic compounds, screening and analysis methodologies and future perspectives. *Food and Chemical Toxicology*, 51, 15–25.
- Chan, E. W. C., Lim, Y. Y., Wong, S. K., Lim, K. K., Tan, S. P., Lianto, F. S., & Yong, M. Y. (2009). Effects of different drying methods on the antioxidant properties of leaves and tea of ginger species. *Food Chemistry*, 113(1), 166–172.

- Chan, E. W. C., Lye, P. Y., Eng, S. Y., & Tan, Y. P. (2013). Antioxidant properties of herbs with enhancement effects of drying treatments: A synopsis. *Free Radicals and Antioxidants*, 3(1), 2–6.
- Cheok, C. Y., Chin, N. L., Yusof, Y. A., & Law, C. L. (2012). Extraction of total phenolic content from *Garcinia mangostana* Linn. hull. i. effects of solvents and UV-Vis spectrophotometer absorbance method. *Food and Bioprocess Technology*, 5(7), 2928–2933.
- Cheok, C. Y., Salman, H. A. K., & Sulaiman, R. (2014). Extraction and quantification of saponins: A review. *Food Research International*, 59, 16–40.
- Cheung, D. W. S., Koon, C. M., Wat, E., Ko, C. H., Chan, J. Y. W., Yew, D. T. W et al. (2013). A herbal formula containing roots of *Salvia miltiorrhiza* (Danshen) and *Pueraria lobata* (Gegen) inhibits inflammatory mediators in LPS-stimulated RAW 264.7 macrophagesthrough inhibition of nuclear factor κ b (NF κ B) pathway. *Journal of Ethnopharmacology*, 145, 776–783.
- Chumroenphat, T., Khanprom, I., & Butkhup, L. (2011). Stability of phytochemicals and antioxidant properties in ginger (*Zingiber officinale* Roscoe) rhizome with different drying methods. *Journal of Herbs, Spices & Medicinal Plants*, 17(4), 361–374.
- Connell, & Sutherland. (1969). A re-examination of gingerol, shogaol, and zingerone, the pungent principles of ginger (*Zingiber officinale* Roscoe). *Australian Journal of Chemistry*, 22, 1033–1043.
- Contini, M., Baccelloni, S., Massantini, R. and Anelli, G. (2008). Extraction of natural antioxidants from hazelnut (*Corylus avellana* L.) shell and skin wastes by long maceration at room temperature. *Food Chemistry* 110: 659–669.
- Dai, J., & Mumper, R. J. (2010). Plant phenolics: Extraction, analysis and their antioxidant and anticancer properties. *Molecules*, 15(10), 7313–7352.
- Deng, Y., Luo, Y., Wang, Y., & Zhao, Y. (2015). Effect of different drying methods on the myosin structure, amino acid composition, protein digestibility and volatile profile of squid fillets. *Food Chemistry*, 171, 168–76.
- Ding, S. H., An, K. J., Zhao, C. P., Li, Y., Guo, Y. H., & Wang, Z. F. (2012). Effect of drying methods on volatiles of Chinese ginger (*Zingiber officinale* Roscoe). *Food and Bioproducts Processing*, 90(3), 515–524.
- Dinis, T. C. P., Madeira, V. M. C., & Almeida, L. M. (1994). Action of phenolic derivatives (acetaminophen, salicylate, and 5-aminosalicylate) as inhibitors of membrane lipid peroxidation and as peroxy radical scavengers. *Archives of Biochemistry and Biophysics*, 315(1), 161–169.
- Diwan, R., Shinde, A., & Malpathak, N. (2012). Phytochemical composition and antioxidant potential of *Ruta graveolens* L. in vitro culture lines. *Journal of Botany*, 2012, 1–6.
- Dixon, R. A., and Paiva N. L. (1995). Stress-induced phenylpropanoid metabolism.

Plant Cell, **7**, 1085–1097.

- Dhingra, D., Michael, M., Rajput, H., & Patil, R. T. (2012). Dietary fibre in foods: A review. *Journal of Food Science and Technology*, *49*(3), 255–266.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S., & Ju, Y. H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatica*. *Journal of Food and Drug Analysis*, *22*(3), 296–302.
- Dugasani, S., Pichika, M. R., Nadarajah, V. D., Balijepalli, M. K., Tandra, S., & Korlakunta, J. N. (2010). Comparative antioxidant and anti-inflammatory effects of [6]-gingerol, [8]-gingerol, [10]-gingerol and [6]-shogaol. *Journal of Ethnopharmacology*, *127*(2), 515–520.
- El-baky, H. H. A., Farag, R. S., & Saleh, M. A. (2010). Characterization of antioxidant and antimicrobial compounds of cinnamon and ginger essential oils. *African Journal of Biochemistry Research*, *4*, 167–174.
- El-Ghorab, A. H., Nauman, M., Anjum, F. M., Hussain, S., & Nadeem, M. (2010). A Comparative study on chemical composition and antioxidant activity of ginger (*Zingiber officinale*) and cumin (*Cuminum cyminum*). *Journal of Agricultural and Food Chemistry*, *58*(14), 8231–8237.
- Esmailzadeh Kenari, R., Mohsenzadeh, F., & Amiri, Z. R. (2014). Antioxidant activity and total phenolic compounds of Dezful sesame cake extracts obtained by classical and ultrasound-assisted extraction methods. *Food Science & Nutrition*, *2*(4), 426–35.
- Eze, J., & Agbo, K. (2011). Comparative studies of sun and solar drying of peeled and unpeeled ginger. *American Journal of Scientific and Industrial Research*, *2*(2), 136–143.
- Famurewa, A. V., Emuekele, P. O., & Jaiyeoba, K. F. (2011). Effect of drying and size reduction on the chemical and volatile oil contents of ginger (*Zingiber officinale*). *Journal of Medicinal Plants Research*, *5*(14), 2941–2944.
- Flora, S. J. S., & Pachauri, V. (2010). Chelation in metal intoxication. *International Journal of Environmental Research and Public Health*, *7*, 2745–2788.
- Ghasemzadeh, A., Jaafar, H. Z. E., & Rahmat, A. (2010). Antioxidant activities, total phenolics and flavonoids content in two varieties of Malaysian young ginger (*Zingiber officinale* Roscoe). *Molecules*, *15*(6), 4324–4333.
- Ghosh, A. (2011). Pharmacognosy *Zingiber officinale*: A natural gold. *International Journal of Pharma and Bio Sciences*, *2*(1), 283–294.
- Goli, A. H., Barzegar, M., & Sahari, M. A. (2005). Antioxidant activity and total phenolic compounds of pistachio (*Pistachia vera*) hull extracts. *Food Chemistry*, *92*(3), 521–525.
- Gul, M. Z., Bhakshu, L. M., Ahmad, F., Kondapi, A. K., Qureshi, I. A., & Ghazi, I. A.

- (2011). Evaluation of *Abelmoschus moschatus* extracts for antioxidant, free radical scavenging, antimicrobial and antiproliferative activities using in vitro assays. *BMC Complementary and Alternative Medicine*, 11(64), 1-12.
- Gümüşay, O. A., Borazan, A. A., Ercal, N., & Demirkol, O. (2015). Drying effects on the antioxidant properties of tomatoes and ginger. *Food Chemistry*, 173, 156–162.
- Guo, J., Wu, H., Du, L., Zhang, W., & Yang, J. (2014). Comparative antioxidant properties of some gingerols and shogaols, and the relationship of their contents with the antioxidant potencies of fresh and dried ginger (*Zingiber officinale* Roscoe). *Journal of Agricultural Science and Technology*, 16, 1063–1072.
- Han, Y., Song, C., Koh, W., Yon, G., & Kim, Y. (2013). Anti-inflammatory effects of the *Zingiber officinale* roscoe constituent 12-Dehydrogingerdione in LPS-stimulated Raw 264.7 cells. *Phytotherapy Research*, 27, 1200–1205.
- Harbaum-Piayda, B., Palani, K., & Schwarz, K. (2016). Influence of postharvest UV-B treatment and fermentation on secondary plant compounds in white cabbage leaves. *Food Chemistry*, 197, 47–56.
- Harbourne, N., Marete, E., Jacquier, J. C., & O’Riordan, D. (2013). Stability of phytochemicals as sources of anti-inflammatory nutraceuticals in beverages - A review. *Food Research International*, 50(2), 480–486.
- Heredia, A., Castello, M. L., & Andre, A. (2014). Influence of drying method and extraction variables on the antioxidant properties of persimmon leaves. *Food Bioscience*, 6, 1–8.
- Ho, S.-C., & Su, M.-S. (2016). Optimized heat treatment enhances the anti-inflammatory capacity of ginger. *International Journal of Food Properties*, 19(8), 1884 – 1898.
- Holick, M. F. (2016). Biological effects of sunlight, ultraviolet radiation, visible light, infrared radiation and vitamin D for health. *Anticancer Res*, 36(3), 1345–1356.
- Hossain., Barry-Ryan, C., Martin-Diana, A. B., & Brunton, N. P. (2010). Effect of drying method on the antioxidant capacity of six *Lamiaceae* herbs. *Food Chemistry*, 123(1), 85–91.
- Hossain, M. A., AL-Raqmi, K. A. S., AL-Mijizy, Z. H., Weli, A. M., & Al-Riyami, Q. (2013). Study of total phenol, flavonoids contents and phytochemical screening of various leaves crude extracts of locally grown *Thymus vulgaris*. *Asian Pacific Journal of Tropical Biomedicine*, 3(9), 705–710.
- Hsu, C. (2003). Chemical composition, physical properties, and antioxidant activities of yam flours as affected by different drying methods. *Food Chemistry*, 83(1), 85–92.
- Ignat, I., Irina Volf, V., & Popa, V. I. (2011). A critical review of methods for

- characterisation of polyphenolic compounds in fruits and vegetables. *Food Chemistry*, 3(3), 1033–1036.
- Jayashree, E., Visvanathan, R., & John Zachariah, T. (2012). Quality of dry ginger (*Zingiber officinale*) by different drying methods. *Journal of Food Science and Technology*, 51(11), 3190–3198.
- Jelled, A., Fernandes, ngela, Barros, L., Chahdoura, H., Achour, L., Ferreira, I. C. F. R., & Cheikh, H. Ben. (2015). Chemical and antioxidant parameters of dried forms of ginger rhizomes. *Industrial Crops and Products*, 77, 30–35.
- Joshi, A. P. K., Rupasinghe, H. P. V, & Khanizadeh, S. (2011). Impact of drying processes on bioactive phenolics, vitamin c and antioxidant capacity of red-fleshed apple slices. *Journal of Food Processing and Preservation*, 35(4), 453–457.
- Jung, S. H., Kim, S. J., Jun, B. G., Lee, K. T., Hong, S. P., Oh, M. S., et al. (2013). α -Cyperone, isolated from the rhizomes of *Cyperus rotundus*, inhibits LPS-induced COX-2 expression and PGE₂ production through the negative regulation of NF κ B signalling in RAW 264.7 cells. *Journal of Ethnopharmacology*, 147, 208–214.
- Justo, O. R., Simioni, P. U., Gabriel, D. L., Tamashiro, W. M. S. C., Rosa, P. T. V., & Moraes, Â. M. (2015). Evaluation of in vitro anti-inflammatory effects of crude ginger and rosemary extracts obtained through supercritical CO₂ extraction on macrophage and tumor cell line: the influence of vehicle type. *BMC Complementary and Alternative Medicine*, 15(390), 1-15.
- Kabuto, H., Nishizawa, M., Tada, M., Higashio, C., Shishibori, T., & Kohno, M. (2005). Zingerone [4-(4-hydroxy-3-methoxyphenyl)-2-butanone] prevents 6-hydroxydopamine-induced dopamine depression in mouse striatum and increases superoxide scavenging activity in serum. *Neurochemical Research*, 30(3), 325–332.
- Kamiloglu, S., Toydemir, G., Boyacioglu, D., Beekwilder, J., Hall, R. D., & Capanoglu, E. (2016). A review on the effect of drying on antioxidant potential of fruits and vegetables. *Critical Reviews in Food Science and Nutrition*, 56(sup1), S110–129.
- Khangholil, S., & Rezaeinodehi, A. (2008). Effect of drying temperature on essential oil content and composition of sweet wormwood (*Artemisia annua*) growing wild in Iran. *Pakistan Journal of Biological Sciences*, 11(6), 934-937.
- Kizhakkayil, J., & Sasikumar, B. (2011). Diversity, characterization and utilization of ginger: A review. *Plant Genetic Resources*, 9(03), 464–477.
- Klein, B.P. and Perry, A.K. (1982). Ascorbic acid and vitamin A activity in selected vegetables from different geographical areas of the United States. *Journal of Food Science*, 47, 941-945.
- Klein SM, Cohen G, Cederbaum AI. (1991). Production of formaldehyde during metabolism of dimethyl sulphoxide by hydroxyl radical generating system.

Biochemistry 20, 6006-6012.

- Kratchanova, M., Denev, P., Ciz, M., Lojek, A., & Mihailov, A. (2010). Evaluation of antioxidant activity of medicinal plants containing polyphenol compounds. Comparison of two extraction systems. *Acta Biochimica Polonica*, 57(2), 229–234.
- Krishnaiah, D., Sarbatly, R., & Nithyanandam, R. (2011). A review of the antioxidant potential of medicinal plant species. *Food and Bioproducts Processing*, 89(3), 217–233.
- Kubra, I. R., & Rao, L. J. M. (2012). An impression on current developments in the technology, chemistry, and biological activities of ginger (*Zingiber officinale* Roscoe). *Critical Reviews in Food Science and Nutrition*, 52(8), 651–688.
- Kutti Gounder, D., & Lingamallu, J. (2012). Comparison of chemical composition and antioxidant potential of volatile oil from fresh, dried and cured turmeric (*Curcuma longa*) rhizomes. *Industrial Crops and Products*, 38, 124–131.
- Lantz, R. C., Chen, G. J., Sarihan, M., Solyom, A. M., Jolad, S. D., & Timmermann, B. N. (2007). The effect of extracts from ginger rhizome on inflammatory mediator production. *Phytomedicine*, 14, 123–128.
- Lapornik, B., Prošek, M., & Wondra, A. G. (2005). Comparison of extracts prepared from plant by-products using different solvents and extraction time. *Journal of Food Engineering*, 71(2), 214–222.
- Li, F., Wang, Y., Parkin, K. L., Nitteranon, V., Liang, J., Yang, W., et al. (2011). Isolation of quinone reductase (QR) inducing agents from ginger rhizome and their in vitro anti-inflammatory activity. *Food Research International*, 44(6), 1597–1603.
- Ling, A. L. M., Yasir, S., Matanjun, P., & Abu Bakar, M. F. (2014). Effect of different drying techniques on the phytochemical content and antioxidant activity of *Kappaphycus alvarezii*. *Journal of Applied Phycology*, 27, 1717–1723.
- Liu, H., Cao, J., & Jiang, W. (2015). Evaluation and comparison of vitamin C, phenolic compounds, antioxidant properties and metal chelating activity of pulp and peel from selected peach cultivars. *LWT - Food Science and Technology*, 63(2), 1042–1048.
- Loo, A. Y., Jain, K., & Darah, I. (2008). Antioxidant activity of compounds isolated from the pyrolytic acid, *Rhizophora apiculata*. *Food Chemistry*, 107(3), 1151–1160.
- Lu, C.-C., & Yen, G.-C. (2015). Antioxidative and anti-inflammatory activity of functional foods. *Current Opinion in Food Science*, 2, 1–8.
- Maroon, J. C., Bost, J. W., & Maroon, A. (2010). Natural anti-inflammatory agents for pain relief. *Surgical Neurology International*, 1(80), 1–10.
- McSweeney, M., & Seetharaman, K. (2015). State of polyphenols in the drying process

- of fruits and vegetables. *Critical Reviews in Food Science and Nutrition*, 55(5), 660–669.
- Mediani, A., Abas, F., Tan, C., & Khatib, A. (2014). Effects of different drying methods and storage time on free radical scavenging activity and total phenolic content of *Cosmos Caudatus*. *Antioxidants*, 3, 358–370.
- Mojani, M., & Ghasemzadeh, A. (2014). Assessment of bioactive compounds, nutritional composition and antioxidant activity of Malaysian young ginger (*Zingiber officinale* Roscoe). *International Food Research Journal*, 21(5), 1931–1935.
- Mphahlele, R. R., Fawole, O. A., Makunga, N. P., & Opara, U. L. (2016). Effect of drying on the bioactive compounds, antioxidant, antibacterial and antityrosinase activities of pomegranate peel. *BMC Complementary and Alternative Medicine*, 16(143), 1-12.
- Mujumdar, A. S., & Law, C. L. (2010). Drying technology: Trends and applications in postharvest processing. *Food and Bioprocess Technology*, 3(6), 843–852.
- Nagananda, G. ., Satishchandra, N., & Rajath, S. (2013). Phytochemical evaluation and in vitro free radical scavenging activity of cold and hot successive pseudobulb extracts of medicinally important orchid *Flickingeria nodosa* (Dalz.) Seidenf. *Journal of Medical Science*, 13(6), 401–409.
- Nayak, B., Liu, R. H., & Tang, J. (2013). Effect of processing on phenolic antioxidants of fruits, vegetables and grains-A review. *Critical Reviews in Food Science and Nutrition*, 1–114.
- Nicoli, M. C., Anese, M., & Parpinel, M. (1999). Influence of processing on the antioxidant properties of fruit and vegetables. *Trends in Food Science & Technology*, 10, 94–100.
- Nolle, N., Argyropoulos, D., Ambacher, S., Muller, J., & Biesalski, H. K. (2016). Vitamin D₂ enrichment in mushrooms by natural or artificial UV-light during drying. *LWT - Food Science and Technology*, [Doi: 10.1016/j.lwt.2016.11.072](https://doi.org/10.1016/j.lwt.2016.11.072).
- Nótin, B., Stéger-Máté, M., Juhász, R., Jakab, D., Monspart-Sényi, J. and Barta, J. (2011). Changes of phenolic compounds in black currant during vacuum drying process. *Acta Alimentaria* 40,120–129.
- Oboh, G., Akinyemi, A. J., & Ademiluyi, A. O. (2012). Antioxidant and inhibitory effect of red ginger (*Zingiber officinale* var. Rubra) and white ginger (*Zingiber officinale* Roscoe) on Fe²⁺ induced lipid peroxidation in rat brain in vitro. *Experimental and Toxicologic Pathology*, 64, 31–36.
- Onyeike, E. N., Anyalogbu, E. A., & Monanu, M. O. (2015). Effect of heat processing on the proximate composition and energy values of African walnut (*Plukenetiaconophora*) and African elemi (*Canariumschweinfurthii*) consumed as masticatories in Nigeria. *International Journal of Scientific and Technology Research*, 4(08), 295–301.

- Pan, M. H., Yang, J. R., Tsai, M. L., Sang, S., & Ho, C. T. (2009). Anti-inflammatory effect of *Momordica grosvenori* Swingle extract through suppressed LPS-induced upregulation of iNOS and COX-2 in murine macrophages. *Journal of Functional Foods*, 1, 145–152.
- Panda, S. K. (2012). Antioxidant Enzyme. InTech (Ed.), *Assay Guided Comparison for Enzymatic and Non-Enzymatic Antioxidant Activities with Special Reference to Medicinal Plants* (pp. 382–400). Croatia: Intech Publishers.
- Pandey, K. B., & Rizvi, S. I. (2009). Plant polyphenols as dietary antioxidants in human health and disease. *Oxidative Medicine and Cellular Longevity*, 2(5), 270–278.
- Park, Y. S., Jung, S. T., Kang, S. G., Delgado-Licon, E., Leticia Martinez Ayala, A., Tapia, M. S., et al. (2006). Drying of persimmons (*Diospyros kaki* L.) and the following changes in the studied bioactive compounds and the total radical scavenging activities. *LWT - Food Science and Technology*, 39(7), 748–755.
- Pavithra, K., & Vadivukkarasi, S. (2015). Evaluation of free radical scavenging activity of various extracts of leaves from *Kedrostis foetidissima* (Jacq.) Cogn. *Food Science and Human Wellness*, 4, 42–46.
- Pearson, D. (1976). *The chemical analysis of foods by David Pearson*. Churchill Living Stone, Edinburgh Londinum, New York: 7th Edition.
- Prieto, P., Pineda, M., & Aguilar, M. (1999). Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphomolybdenum complex: specific application to the determination of vitamin E. *Analytical Biochemistry*, 269, 337–341.
- Pruthi, J. S. (1992). Post-harvest technology of spices: pre-treatments, curing, cleaning, grading and packing. *Journal of Spices and Aromatic Crops*, 1, 1-29.
- Pulido, R., Bravo, L., & Saura-Calixto, F. (2000). Antioxidant activity of dietary polyphenols as determined by a modified ferric reducing/antioxidant power assay. *Journal of Agricultural and Food Chemistry*, 48(8), 3396–3402.
- Rahmani, A. H., Al, F. M., & Aly, S. M. (2014). Active ingredients of ginger as potential candidates in the prevention and treatment of diseases via modulation of biological activities. *International Journal of Physiology, Pathophysiology and Pharmacology*, 6(2), 125–136.
- Re, R., Pellergrini, N., Proteggente, A., Pannala, A., Yang, M., & Rice - Evans, C. (1999). Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical Biology & Medicine*, 26(98), 1231–1237.
- Roshanak, S., Rahimmalek, M., & Goli, S. A. H. (2016). Evaluation of seven different drying treatments in respect to total flavonoid, phenolic, vitamin C content, chlorophyll, antioxidant activity and color of green tea (*Camellia sinensis* or *C. assamica*) leaves. *Journal of Food Science and Technology*, 53(1), 721–729.

- Sagrin, M. S., & Chong, G. H. (2013). Effects of drying temperature on the chemical and physical properties of *Musa acuminata* Colla (AAA Group) leaves. *Industrial Crops and Products*, 45, 430–434.
- Sánchez-Moreno, C., Larrauri, J. a., & Saura-calixto, F. (1998). A procedure to measure the antiradical efficiency of polyphenols. *Journal of the Science of Food and Agriculture*, 76, 270–276.
- Schreiner, M., Mewis, I., Huyskens-Keil, S., Jansen, M. A. K., Zrenner, R., Winkler, J. B., et al. (2012). UV-B-induced secondary plant metabolites - potential benefits for plant and human health. *Critical Reviews in Plant Sciences*, 175, 449-458.
- Seidel, V. (2006). Initial and Bulk Extraction. In *Methods in Biotechnology Natural Products Isolation*, ed. S.D. Sarker, Latif, Z., and Gray, A.I. (pp. 27-46). New Jersey: Humana Press Inc.
- Shalaby, E. A., & Shanab, S. M. M. (2013). Comparison of DPPH and ABTS assays for determining antioxidant potential of water and methanol extracts of *Spirulina platensis*. *Indian Journal of Marine Sciences*, 42(5), 556–564.
- Shin, J. H., Ryu, J. H., Kang, M. J., Hwang, C. R., Han, J., & Kang, D. (2013). Short-term heating reduces the anti-inflammatory effects of fresh raw garlic extracts on the LPS-induced production of NO and pro-inflammatory cytokines by downregulating allicin activity in RAW 264.7 macrophages. *Food and Chemical Toxicology*, 58, 545–551.
- Shirin, A. P. R., & Prakash, A. (2010). Chemical composition and antioxidant properties of ginger root (*Zingiber officinale*). *Journal of Medicinal Plants Research*, 4(24), 2674–2679.
- Shirsath, S. R., Sonawane, S. H., & Gogate, P. R. (2012). Intensification of extraction of natural products using ultrasonic irradiations-A review of current status. *Chemical Engineering and Processing: Process Intensification*, 53, 10–23.
- Siddhuraju, R., Becker, K., (2003). Antioxidant properties of various solvent extracts of total phenolic constituents from three different agroclimatic origins of Drumstick tree (*Moringa olifera* Lam.) leaves. *Journal of Agricultural and Food Chemistry*, 51, 2144–2155.
- Sindhi, V., Gupta, V., Sharma, K., Bhatnagar, S., Kumari, R., & Dhaka, N. (2013). Potential applications of antioxidants - A review. *Journal of Pharmacy Research*, 7(9), 828–835.
- Singh, N., & Rajini. (2004). Free radical scavenging activity of an aqueous extract of potato peel. *Food Chemistry*, 85, 611–616.
- Sonawane, S. K., & Arya, S. S. (2014). Effect of drying and storage on bioactive components of jambhul and wood apple. *Journal of Food Science and Technology*, 52(5), 2833–2841.
- Sowndhararajan, K., Joseph, J. M., Arunachalam, K., & Manian, S. (2010). Evaluation

- of *Merremia tridentata* (L.) Hallier f. for in vitro antioxidant activity. *Food Science and Biotechnology*, 19(3), 663–669.
- Sowndhararajan, K., Siddhuraju, P., & Manian, S. (2011). Antioxidant and free radical scavenging capacity of the underutilized legume, *Vigna vexillata* (L.) A. Rich. *Journal of Food Composition and Analysis*, 24(2), 160–165.
- Sudan, R., Bhagat, M., Gupta, S., Singh, J., & Koul, A. (2014). Iron (FeII) chelation, ferric reducing antioxidant power, and immune modulating potential of *Arisaema jacquemontii* (Himalayan cobra lily). *BioMed Research International*, 2014,1-7.
- Sultan, M. T., Ahmad, A. N., Saddique, M. S., Aghazadeh, M., Imran, M., Qayyum, M. M. N., & Sibte-Abbas, M. (2014). Antioxidant and antimicrobial potential of locally sun dried garlic and ginger powder available in District Layyah, Punjab, Pakistan. *Pakistan Journal of Nutrition*, 13(11), 642–647.
- Sultana, B., Anwar, F., & Ashraf, M. (2009). Effect of extraction solvent/technique on the antioxidant activity of selected medicinal plant extracts. *Molecules*, 14, 2167–2180.
- Surveswaran, S., Cai, Y., Corke, H., & Sun, M. (2007). Systematic evaluation of natural phenolic antioxidants from 133 Indian medicinal plants. *Food Chemistry*, 102(3), 938–953.
- Tanaka, J. (2010). Anti-inflammatory properties of red ginger (*Zingiber officinale* var. *Rubra*) extract and suppression of nitric oxide production by its constituents. *Journal of Medicinal Food*, 13(1), 156–162.
- Tatiya, A. U., Tapadiya, G. G., Kotecha, S., & Surana, S. J. (2011). Effect of solvents on total phenolics, antioxidant and antimicrobial properties of *Bridelia retusa* Spreng. stem bark. *Indian Journal of Natural Products and Resources*, 2(4), 442–447.
- Tiwari, P., Kumar, B., Mandeep, K., Kaur, G., & Kaur, H. (2011). Phytochemical screening and extraction: A review. *International Pharmaceutical Science*, 1(1), 98–106.
- Tong, Y., Zhu, X., Yan, Y., Liu, R., Gong, F., Zhang, L., et al. (2015). The influence of different drying methods on constituents and antioxidant activity of saffron from China. *International Journal of Analytical Chemistry*, 2015, 1-8.
- Tsai, T. H., Tsai, P. J., & Ho, S. C. (2005). Antioxidant and anti-inflammatory activities of several commonly used compounds. *Food Chemistry and Toxicology*, 70(1), 93–97.
- Wang, T., Jónsdóttir, R., & Ólafsdóttir, G. (2009). Total phenolic compounds, radical scavenging and metal chelation of extracts from Icelandic seaweeds. *Food Chemistry*, 116(1), 240–248.

- Wohlmuth, H. (2008). *Phytochemistry and pharmacology of plants from the ginger family, Zingiberaceae*. (Unpublished doctoral dissertation). University of Lismore, Australia.
- Wu, S., Li, F., Jia, S., Ren, H., Gong, G., Wang, Y., et al. (2014). Drying effects on the antioxidant properties of polysaccharides obtained from *Agaricus blazei* Murrill. *Carbohydrate Polymers*, 103, 414–7.
- Yeh, H., Chuang, C., Chen, H., Wan, C., Chen, T., & Lin, L. (2013). Bioactive components analysis of two various gingers (*Zingiber officinale* Roscoe) and antioxidant effect of ginger extracts. *LWT - Food Science and Technology*, 1-6.
- Yoshikawa, T., & Naito, Y. (2002). What is oxidative stress? *Journal of the Japan Medical Association*, 124(11), 1549–1553.
- Zhishen, J., Mengcheng, T., & Jianming, W. (1999). The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. *Food Chemistry*, 64, 555–559.
- Zhu, K-X., Lian, C-X., Guo, X-N., Peng, W. and Zhou, H-M. (2011). Antioxidant activities and total phenolic contents of various extracts from defatted wheat germ. *Food Chemistry* 126: 1122–1126.
- Zlotek, U., Mikulska, S., Nagajek, M., & Świeca, M. (2015). The effect of different solvents and number of extraction steps on the polyphenol content and antioxidant capacity of basil leaves (*Ocimum basilicum* L.) extracts. *Saudi Journal of Biological Sciences*, 23, 628-633