



**UNIVERSITI PUTRA MALAYSIA**

***ROOT COLONIZATION OF OIL PALM  
(*Elaeis guineensis* Jacq.) USING GFP-EXPRESSING *Ganoderma  
boninense*  
AND EFFECTS OF LIGNIN ON DISEASE PROGRESSION***

**NISHA A/P THOPLA GOVENDER**

**ITA 2016 4**



**ROOT COLONIZATION OF OIL PALM  
(*Elaeis guineensis* Jacq.) USING GFP-EXPRESSING *Ganoderma boninense*  
AND EFFECTS OF LIGNIN ON DISEASE PROGRESSION**

By

**NISHA A/P THOPLA GOVENDER**

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

**January 2016**



© COPYRIGHT UPM

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright© Universiti Putra Malaysia



## DEDICATION

Special dedication to:

*My late grandfather, Mr. Kandasamy Krishnan PJK.*



© COPYRIGHT UPM

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**ROOT COLONIZATION OF OIL PALM  
(*Elaeis guineensis* Jacq.) USING GFP-EXPRESSING *Ganoderma boninense* AND  
EFFECTS OF LIGNIN ON DISEASE PROGRESSION**

By

**NISHA A/P THOPLA GOVENDER**

**January 2016**

**Chair : Associate Professor Wong Mui Yun, PhD**  
**Faculty : Institute of Tropical Agriculture**

Oil palm is the world's most efficient oil-bearing tree. Major diseases impeding the oil palm productivity have been caused by fungi, particularly *Ganoderma boninense*, the causal agent of basal stem rot (BSR). Visible symptoms can only be observed nearing the plant death stage while early penetration and infection strategy remain cryptic due to the fungal hyaline nature. The underlying principles on how *Ganoderma* penetrates and infects oil palm roots are unknown. Therefore, a tagged *G. boninense* harbouring GUS-GFP fusion gene would ideally serve as a tool to unravel early pathogenesis of *G. boninense*. Lignin, a heterogeneous complex polymer is poorly understood during BSR development and thus, assessments of lignin content and composition in different planting materials and during the disease development were performed. In addition, enzyme activity and gene expression of key components in the phenylpropanoid pathway were investigated. Lignin content and composition were screened in oil palm lines with differential tolerance to BSR. Both parameters were associated to growth factors (height, weight and girth) and micronutrients depositions were measured using X-Ray Fluorescence (XRF). Efficient *Agrobacterium*-mediated transformation protocol was established via optimization of several parameters; *Agrobacterium* strain (LBA4404, GV3101, EHA101 and EHA105), explants (mycelia, spore and protoplast), *vir* gene induction period and modification of binary vector. The transformant was utilized to discern early stage colonization of BSR using the confocal microscopy. Glass-house trial on BSR development was performed to evaluate enzyme activities and gene expression of the following defence genes; phenylalanine ammonia lyase (PAL), cinnamyl alcohol dehydrogenase (CAD) and peroxidase (POD) using enzyme assay and quantitative real-time PCR respectively. Lignin content and composition were significantly different among the oil palm seedlings with different tolerance to BSR. The susceptible and intermediate lines showed significantly higher lignin content in comparison to the tolerant line, while lignin composition denoted as S/G ratio was higher in tolerant line in comparison to both susceptible and intermediate lines. Apparent lignin accumulation was supported by micronutrients deposition which comprised copper, silicon, titanium and sulphur. A successful transformation system was developed for *G. boninense* using protoplast and *Agrobacterium* strain LBA 4404. The binary vector pCAMBIA 1304, modified to harbour GPD fungal promoter from

plasmid p416 improved the expression of GUS-GFP fusion protein. Colonization pattern was initiated with active differentiation of the tagged *G. boninense* into microhyphae. The needle-like structure was able to penetrate the epidermis layer randomly and progressed longitudinally into exodermis and cortex region. Induced lignification during defense showed great participation from both PAL and CAD genes and enzymes. The S/G ratio increased significantly in the induced lignin as compared to constitutive lignin indicated alterations employed by host as part of their defense strategy during *Ganoderma* infection. Low lignin content supported growth without compromising oil palm biomass while creating an avenue for greater proportion of induced lignin which consists of S monomer during *G. boninense* infection. The findings can be adopted in oil palm breeding strategies aimed to produce resistant planting materials.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KOLONISASI AKAR POKOK KELAPA SAWIT (*Elaeis guineensis* Jacq)  
MENGUNAKAN *Ganoderma boninense* BEREKSPRES-GFP DAN KESAN  
LIGNIN SEMASA PERKEMBANGAN PENYAKIT**

Oleh

**NISHA A/P THOPLA GOVENDER**

**Januari 2016**

**Pengerusi : Profesor Madya Wong Mui Yun, PhD**  
**Fakulti : Institut Pertanian Tropika**

Kelapa sawit merupakan pokok paling efisien di dunia bagi penghasilan minyak sayuran. Kebanyakan penyakit yang membantutkan produktiviti kelapa sawit adalah berpunca dari kulat khususnya *Ganoderma boninense*. Kulat ini merupakan agen penyebab penyakit reput pangkal batang. Simptom-simptom penyakit ini hanya jelas kelihatan pada hujung peringkat kematian. Strategi penembusan kulat ini untuk melancarkan infeksi masih kurang diketahui memandangkan kulat ini bersifat halus. Oleh itu, *G. boninense* bertag yang mempunyai gen GUS-GFP mampu mengenalpasti corak patogenisiti penyakit reput pangkal batang. Lignin, polimer heterogenus kompleks, menjalankan peranan pertahanan ketika perkembangan penyakit reput pangkal batang dan mekanisma ini tidak banyak diketengahkan. Dengan itu, pemahaman lignin boleh diadaptasi sebagai faktor ketahanan terhadap penyakit reput pangkal batang. Kandungan dan komposisi lignin dari tisu akar pokok kelapa sawit yang terdiri daripada kumpulan pelbagai toleran terhadap penyakit reput pangkal batang dikenalpasti. Kedua-dua parameter ini dikaitkan dengan faktor tumbesaran (tinggi, berat dan diameter) sementara peratus mikronutrien ditentukan dengan menggunakan X-Ray Fluorescence (XRF). Sebuah protokol yang efisien berasaskan penggunaan *Agrobacterium* telah dibina bagi tujuan transformasi kulat *G. boninense*. Pengoptimuman beberapa parameter seperti berikut dilakukan: Isolat *Agrobacterium* (LBA4404, GV3103, EHA101 dan EHA105), explain (miselium, spora dan protoplas), tempoh induksi gen *vir* dan modifikasi vektor binari. Transforman digunakan bagi tujuan mengenalpasti corak kolonisasi penyakit reput pangkal batang pada peringkat awal penyakit dengan menggunakan mikroskop konfokal. Satu eksperimen rumah kaca mengkaji interaksi *Ganoderma*-perumah dijalankan untuk menentukan aktiviti enzim dan ekspresi gen (PAL, CAD dan POD) masing-masing dengan menggunakan esei enzim dan tindak balas rantaian. Kandungan lignin dan komposisi menunjukkan perbezaan yang signifikan di kalangan anak pokok kelapa sawit dari kumpulan pelbagai toleran terhadap penyakit batang pangkal reput. Kumpulan yang rendah dan sederhana toleran, masing-masing menunjukkan kandungan lignin yang lebih tinggi berbanding kumpulan toleran manakala komposisi lignin yang terdiri daripada nisbah S/G menunjukkan nisbah yang tinggi di kalangan kumpulan yang toleran berbanding kumpulan yang kurang toleran dan kumpulan tidak toleran. Pembentukan lignin



disokong bersama pemendapan mikronutrien yang terdiri daripada kuprum, silikon, titanium dan sulfur. Sistem transformasi berjaya dibangunkan untuk *G. boninense* dengan menggunakan protoplas sebagai sumber eksplan dan *Agrobacterium* strain LBA4404. Vektor binari pCAMBIA1304 diubahsuai dengan menambah promoter kulat GPD dari plasmid p416. Modifikasi ini berjaya meningkatkan lagi ekspresi gen gabungan bersama GUS-GFP. Induksi proses lignifikasi ketika interaksi *Ganoderma*-perumah disokong kuat oleh kedua-dua gen dan enzim CAD. Nisbah S/G meningkat secara signifikan di dalam lignin induksi berbanding dengan lignin konstitutif. Perubahan komposisi lignin yang ketara menjelaskan strategi pertahanan perumah, kelapa sawit ketika infeksi *Ganoderma*. Keputusan ini didapati seiring dengan kumpulan anak pokok kelapa sawit yang bertoleran tinggi terhadap penyakit reput pangkal batang, yang menunjukkan nisbah S/G yang tinggi. Kandungan lignin yang rendah boleh menyokong proses tumbesaran tanpa sebarang kesan signifikan ke atas biomasa dan menyediakan ruang untuk pembentukan lignin induksi pada kadar yang lebih tinggi. Hasil kajian ini boleh diadaptasi dalam usaha pembiakbakaan kelapa sawit yang bertoleran terhadap penyakit reput pangkal batang.

## ACKNOWLEDGEMENTS

I would like to express my heartiest appreciation to my mother, Vijayaletchery Kandasamy, who in spite of her limited education sacrificed everything to give me the most and best education possible.

Next, to my supervisory committee led by Assoc. Prof Dr. Wong Mui Yun, who deliberately trained and coached me to be a skilful young researcher. I sincerely thank you for helping in the many facets of this research and for being equally enthusiastic and vigorous on our research.

I also would like to acknowledge all staffs and friends from Department of Plant Protection, Faculty of Agriculture for your kindness and assistance towards completion of this research project. Not forgetting the FRGS grant from UPM, Malaysian Palm Oil Board (MPOB) and Applied Agricultural Resources (AAR) for financial assistance and providing materials required.

My gratitude also goes to Prof. Maziah Mahmood, Dr. Idris Abu Seman (MPOB, Selangor), Dr. Sreeramanan (Universiti Sains Malaysia, Penang) and Dr. Wong Han Ling (Universiti Tunku Abdul Rahman, Perak) for your kindness to provide selected strains and constructive advice needed to facilitate the research project. My gratitude to the following institutions for services and facilities provided to complete my study; Institute of Bioscience, Olympus confocal microscopy, Faculty of Science and Physics, Faculty of Food Technology, glasshouse 2D, Animal science and Crop science Departments, Faculty of Agriculture.

Lastly, I am indebtedful to Ministry of Higher Education (MOHE), Malaysia for their generosity to support both tuition fees and my monthly allowances (MyBrain PhD 15) throughout the entire study period which took approximately 4 years.

I certify that a Thesis Examination Committee has met on 19 January 2016 to conduct the final examination of Nisha a/p Thopla Govender on her thesis entitled "Root Colonization of Oil Palm (*Elaeis guineensis* Jacq.) using GFP-Expressing *Ganoderma boninense* and Effects of Lignin on Disease Progression" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

**Ho Chai Ling, PhD**

Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Chairman)

**Datin Siti Nor Akmar binti Abdullah, PhD**

Professor  
Institute of Tropical Agriculture  
Universiti Putra Malaysia  
(Internal Examiner)

**Jugah bin Kadir, PhD**

Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Internal Examiner)

**Robert Russell Monteith Paterson, PhD**

Senior Lecturer  
University of Minho  
Portugal  
(External Examiner)



---

**ZULKARNAIN ZAINAL, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 21 April 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Wong Mui Yun, PhD**  
Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Chairman)

**Maziah Mahmood, PhD**  
Professor  
Faculty of Biomolecular and Biotechnology Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Idris Abu Seman, PhD**  
Ganoderma and Diseases Research for Oil Palm Unit  
Malaysian Palm Oil Board (MPOB)  
Malaysia  
(External Examiner)

---

**BUJANG KIM HUAT, PhD**  
Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.


Signature: \_\_\_\_\_ Date: \_\_\_\_\_

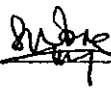
Name and Matric No.: Nisha A/P Thopla Govender, GS 30632

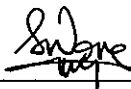
## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:   
Name of Chairman of  
Supervisory  
Committee: Wong Mui Yun, PhD

Signature:   
Name of Member of  
Supervisory  
Committee: Maziah Mahmood, PhD

Signature:   
Name of Member of  
Supervisory  
Committee: Idris Abu Seman, PhD

## TABLE OF CONTENTS

		Page
<b>ABSTRACT</b>		i
<b>ABSTRAK</b>		iii
<b>ACKNOWLEDGEMENTS</b>		v
<b>APPROVAL</b>		vi
<b>DECLARATION</b>		viii
<b>LIST OF TABLES</b>		xiv
<b>LIST OF FIGURES</b>		xv
<b>LIST OF ABBREVIATIONS</b>		xviii
<b>CHAPTER</b>		
<b>1</b>	<b>INTRODUCTION</b>	1
<b>2</b>	<b>LITERATURE REVIEW</b>	4
	2.1 Oil palm	4
	2.1.1 Origin and Botany	4
	2.1.2 Economic uses and value	4
	2.2 Basal stem rot disease	5
	2.2.1 Basal stem rot epidemiology	5
	2.2.2 <i>Ganoderma boninense</i> , the white rot fungi	6
	2.2.3 Host-pathogen interaction	7
	2.2.4 Management of basal stem rot disease	7
	2.3 Genetic transformation of Basidiomycetes	8
	2.3.1 <i>Agrobacterium</i> -mediated transformation of Basidiomycetes	8
	2.3.2 Application of tagged <i>G. boninense</i> in disease management	9
	2.4 Lignin	10
	2.4.1 Lignin biosynthesis pathway	11
	2.4.2 Shikimate pathway	12
	2.4.3 Phenylpropanoid pathway	12
	2.4.4 Lignin branch pathway	15
	2.4.5 Lignin variables: Implication to plant defense mechanism	15
	2.4.6 Lignin and its association to agronomic characters	15
	2.4.7 Lignin biodegradation	16
<b>3</b>	<b>ISOLATION AND REGENERATION OF PROTOPLAST AND BASIDIOSPORE FROM <i>G. boninense</i></b>	18
	3.1 Introduction	18
	3.2 Material and Methods	19
	3.2.1 Optimization: Protoplast preparation and regeneration	19
	3.2.2 Basidiocarp development and basidiospore Harvest	20
	3.3 Results and Discussions	20

3.3.1	Optimization for maximal protoplast regeneration	20
3.3.2	Basidiocarp development for efficient basidiospore isolation and regeneration into <i>G. boninense</i> (mycelia) single colonies	24
3.4	Conclusion	28
<b>4</b>	<b>DEVELOPMENT OF AN EFFICIENT <i>Agrobacterium</i>-MEDIATED TRANSFORMATION METHOD FOR <i>Ganoderma boninense</i></b>	<b>28</b>
4.1	Introduction	28
4.2	Material and Methods	29
4.2.1	Growth conditions for bacterial strains and fungal cultures	29
4.2.2	Antibiotic selection of <i>Ganoderma boninense</i> against Hygromycin B	30
4.2.3	Antibiotic selection of <i>Agrobacterium tumefaciens</i> strains against kanamycin and cefotaxime	30
4.2.4	Transformation of competent <i>Agrobacterium</i> strains with modified pCAMBIA 1304	31
4.2.4.1	Preparation of Freeze/Thaw-Competent <i>Agrobacterium</i> cells	31
4.2.4.2	Extraction of plasmid DNA from <i>E. coli</i>	31
4.2.4.3	Modification of pCAMBIA 1304 with GPD promoter	32
4.2.4.4	Gel extraction to harvest GPD promoter	32
4.2.4.5	Linearization and dephosphorylation of pCAMBIA 1304	32
4.2.4.6	Rapid DNA Ligation of pCAMBIA 1304 with GPD promoter	33
4.2.4.7	Transformation of <i>Agrobacterium</i> through Freeze/Thaw Method	33
4.2.5	<i>Vir</i> gene induction of the <i>Agrobacterium</i> strains	33
4.2.6	Transformation protocol	34
4.2.7	Screening of transformants	34
4.2.7.1	Integration verification using Polymerase Chain Reaction (PCR)	34
4.2.7.2	Gel electrophoresis	35
4.2.7.3	Gene copy number	35
4.2.7.4	Evaluation: GUS-GFP protein expression in transformant <i>G. boninense</i>	36
4.2.8	Artificial infection of oil palm plantlets by tagged <i>G. boninense</i>	36
4.3	Results and Discussions	37
4.3.1	Transformation procedure	37
4.3.2	Effect of co-cultivation conditions and acetosyringone supplementation	37
4.3.3	Effect of selection medium	38
4.3.4	Effect of explant on transformation efficiency	38
4.3.5	Effect of <i>Agrobacterium</i> strains over transformation efficiency	38



4.3.6	T-DNA integration and copy number in transformant <i>G. boninense</i>	39
4.3.7	Quantification of GFP distribution in transformant <i>G. boninense</i>	40
4.3.8	GFP marker in <i>G. boninense</i> transformant as a tool to discern early stage infection of BSR	43
4.3.9	Microhyphae: Penetrating structure during basal stem rots	45
4.4	Conclusion	45
<b>5</b>	<b>DETERMINATION OF LIGNIN CONTENT AND COMPOSITION OF OIL PALM SEEDLINGS WITH DIFFERENTIAL TOLERANCE TO BSR</b>	<b>47</b>
5.1	Introduction	47
5.2	Materials and Methods	48
5.2.1	Plant materials	48
5.2.2	Growth and mineral content assessment	49
5.2.3	Lignin (TGA) content determination	49
5.2.4	Nitrobenzene oxidation/HPLC	50
5.2.5	Statistical analysis	50
5.3	Results and Discussions	51
5.3.1	Lignin content and composition variability in palm genotypes	51
5.3.2	Growth parameter and association to lignin	53
5.3.3	Micronutrient absorption and lignin	55
5.3.4	Lignin changes in association to oil palm defense mechanism	56
5.4	Conclusion	57
<b>6</b>	<b>ACTIVITIES OF DEFENSE LIGNIN DURING BASAL STEM ROT PATHOGENESIS</b>	<b>58</b>
6.1	Introduction	58
6.2	Material and Methods	59
6.2.1	Planting materials and inoculum	59
6.2.2	Experimental Design: Growth and Disease assessment	60
6.2.3	Determination of lignin content and lignin composition	60
6.2.4	Determination of enzyme activities	60
6.2.5	Relative quantification of lignin biosynthetic transcripts by qRT-PCR	61
6.2.5.1	RNA extraction	61
6.2.5.2	cDNA synthesis	62
6.2.5.3	Real-time Polymerase Chain Reaction (qPCR)	62
6.2.6	Statistical Analysis	63
6.3	Results and Discussions	63
6.3.1	Disease profile of oil palm seedlings upon <i>G. boninense</i> challenge	63

6.3.2	Growth profile of oil palm seedlings upon <i>G. boninense</i> challenge	66
6.3.3	Lignin content: Changes during BSR progression	68
6.3.4	Lignin composition: Changes during BSR progression	69
6.3.5	Activities of lignin-associated enzymes during BSR progression	70
6.3.6	Gene expression patterns of lignin biosynthetic genes during early stage infection of BSR	72
6.3.7	Accumulation syringyl-rich lignin during oil palm defense response against <i>G. boninense</i> is up-regulated by PAL and CAD	73
6.3.8	CAD, key regulator in biosynthesis of S-enriched lignin	73
6.3.9	Lignin for systemic acquired resistance in oil palm	74
6.4	Conclusion	74
<b>7</b>	<b>SUMMARY, GENERAL CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	<b>76</b>
	<b>REFERENCES</b>	<b>78</b>
	<b>APPENDICES</b>	<b>104</b>
	<b>BIODATA OF STUDENT</b>	<b>136</b>
	<b>LIST OF PUBLICATIONS</b>	<b>137</b>

## LIST OF TABLES

Table		Page
3.1	Effect of incubation time of lytic enzyme (10 mg/ml) on protoplast yield/ml from <i>G. boninense</i> mycelia treated in different osmotic stabilizer and regeneration media	21
3.2	Basidiocarp diameter and number of spores regenerated from <i>Ganoderma boninense</i>	26
4.1	Primer sequences for PCR and qPCR analysis of wild and transformant <i>G. boninense</i>	36
4.2	Transformation efficiency for <i>G. boninense</i> at all possible combinations using varying <i>Agrobacterium</i> strains and fungal explants	39
5.1	The products of nitrobenzene oxidation: Syringyl to guaicyl ratios (S/G) in oil palm roots (as% of total amount of identified compounds) of differential tolerance to BSR	52
5.2	Pearson Coefficient values between growth parameters and lignin Content within oil palm lines with differential tolerance to BSR	54
6.1	Primer pairs and its description for qRT-PCR	62
6.2	Lignin composition in root tissues of oil palm at different disease severities of basal stem rot	70

## LIST OF FIGURES

Figure	Page
3.1	23
Single colonies regenerated on PDA suspended in phosphate buffer and 0.6 M sucrose from protoplast suspension after 3 h enzymatic digestion using 10 mg/ml lytic enzyme from <i>Trichoderma harzianum</i> and 0.6 M KCl as osmotic stabilizer at day 10 after inoculation	
3.2	25
Fruiting body of <i>G. boninense</i> harvested at different developmental stages. A) Sporophore; B) Budding sporophores; C) Basidiocarp; D) Matured basidiocarp; E) Senescence basidiocarp; Fi) Hymenium observed at underside of matured basidiocarp and, Fii) Hymenium of matured basidiocarp observed under light microscope at 40X	
3.3	26
Spore regeneration from <i>G. boninense</i> basidiocarp upon maturity on PDA (above) and GCM (below) in absence of antibiotics supplementation on day 9 after inoculation	
4.1	40
Gel photograph of PCR analysis of GFP (1-3) sequence and GUS-GFP fusion (4-6) from tagged <i>G. boninense</i> . Lane M; DNA size marker Lane PC; plasmid pCAMBIA 1304 Lane NC; Untransformed <i>G. boninense</i> Lane 1 and 4; Transformed <i>G. boninense</i> from spore Lane 2 and 5; Transformed <i>G. boninense</i> from protoplast Lane 1 and 6; Transformed <i>G. boninense</i> from mycelia	
4.2	40
Copy number of GFP transcripts in transformant <i>G. boninense</i> originating from different explants; protoplast, spore and mycelia. The bars represent standard error computed from three technical replicates per sample	
4.3	42
Transformant from different explants observed under the confocal laser microscopy (A-C) and overlay bright field microscopy(A'-C') at 100 X magnification. Explant is denoted as A; protoplast, B; spore and C; mycelia	
4.4	44
Artificial inoculation of oil palm plantlets with transformant <i>G. boninense</i> and corresponding colonization of BSR on root tissues observed under the confocal laser microscopy. A: First generation transformant from different explants a;protoplast b; spore c; mycelia and negative; wild. B; Ball like oil palm root medium for inoculum preparation (right) and fully colonized inoculum infected with transformant <i>G. boninense</i> (left). C; Oil palm plantlets (left) and infected oil palm plantlets using the ball inoculum pasting technique (right). D; Transformant hyphae from root surface under the bright light microscopy E; Transformant hyphae with GFP fluorescence (green signal) under the confocal laser microscopy F; Uninfected oil palm plantlet (left) and infected oil plam plantlet (right) G; Root fluorescence (yellow and red	

signal) of infected root tissue at 7 DAI H; Fungal colonization at 14 DAI I: Fungal colonization at 21 DAI. Red arrow indicates mycelium attachment (C and F) and projections of micro-hyphae (H and I)

5.1	Lignin content in root tissues of oil palm. Values were expressed as mean±standard deviation of thirty (one-year-old) and ten (five-year-old) biological replicates	51
5.2	Growth assessment in one-year-old oil palm seedlings. Values are expressed mean±standard deviation of thirty biological replicates	54
5.3	Micronutrient deposition in root tissues of one-year-old oil palm seedlings. Values are expressed as mean± standard deviation of ten biological replicates	55
6.1	Basal stem rot development on oil palm seedlings challenged against <i>G. boninense</i> inoculums under glasshouse conditions. Numbers represent (0, I, II, III and IV) months after inoculation A; uninfected seedlings (control) B: infected seedlings	64
6.2	Basal stem rot progression on 3-month old oil palm seedlings via artificial infection of <i>Ganoderma boninense</i> . A: Uninfected seedlings (control) B: Asymptomatic seedlings at week 1 after inoculation C: Seedlings at week 2 after inoculation with at least 2 yellowing leaves D: Seedlings at week 3 after inoculation with three or more browning leaves and visible fruiting bodies E: Seedlings at week 4 after inoculation showing complete necrosis F: Healthy roots of uninfected control seedlings G: Infected roots showing colonization of <i>G. boninense</i> , red arrows indicates mycelium attachment H) Healthy boles of uninfected (control) seedlings I: Infected boles of seedlings showing brown coloration. Red arrows indicated sporophore (left) and necrosis (right)	65
6.3	Basal stem rot progressive curve showing relative area under disease at different months after inoculation (0-4)	66
6.4	Growth assessment; weight (A) and height (B) of oil palm seedlings subjected artificial infection of <i>Ganoderma boninense</i> . Bars represent average mean±standard deviation of ten biological replicates. Letters with different alphabet show significant difference at $p<0.05$ .	67
6.5	Lignin content in root tissues of oil palm seedlings collected at different disease severities of basal stem rot. Bars represent average mean±standard deviation of three technical replicates from a pooled sample of ten biological replicates. Letters with different alphabet show significant difference at $p<0.05$ .	69
6.6	Enzyme activities of phenylalanine ammonia lyase (PAL), cinnamyl alcohol dehydrogenase (CAD) and peroxidase (POD) in root tissues of oil palm seedlings collected at different severities of basal stem rot.	71

Bars represent average mean±standard deviation of three technical replicates from a pooled sample of ten biological replicates. Letters with different alphabet show significant difference at  $p<0.05$ .

- 6.7 Relative quantification of phenylammonia lyase (PAL), cinnamyl alcohol dehydrogenase (CAD) and peroxidase (POD) transcripts at early stage infection of basal stem rot disease. Bars represent average mean±standard deviation of three technical replicates from a pooled sample of four biological replicates.

72



## LIST OF ABBREVIATIONS

A	Absorbance
ANOVA	Analysis of variance
AAR	Applied Agricultural Resources
AMT	Agrobacterium-mediated transformation
$\beta$ -Actin	Beta-Actin
BH	bole height
BLAST	Basic Local Alignment Search Tool
BSA	N,O-bis(trimethylsilyl)acetamide
BSR	basal stem rot
CAD	cinnamyl alcohol dehydrogenase
CCoAOMT	Caffeoyl-CoA 3-O-Methyltransferase
cDNA	complementary deoxyribonucleic acid
CRD	Completely Randomized Design
DNA	deoxyribonucleic acid
dNTPs	deoxynucleotides
DNase	deoxyribonuclease
DI	disease incidence
DTT	dithiothreitol
EDTA	Ethylenediaminetetracetic acid
FAOSTAT	Food and Agriculture
F5H	Ferulate 5-hydroxylase
GAPDH	Glyceraldehyde phosphate dehydrogenase
GCM	Ganoderma Complete Medium
GC-MS	Gas Chromatography-Mass Spectrometer
GFP	green fluorescent protein
GPD	Glyceraldehydes 3-phosphate dehydrogenase
GUS	$\beta$ -glucuronidase
HCl	hydrochloric acid
HPLC	High Performance Liquid Chromatography
IM	induction medium
LB	Luria-Bertani
M	Molar
MAI	month after inoculation
MCS	multiple cloning site
MEA	Malt Extract Agar
MES	2-(N-morpholino)-ethanesulfonic acid
MM	minimal medium
min	minute
mL	mililiter
mM	miliMolar
$\mu$ M	microMolar
$\mu$ g	microgram
MPOB	Malaysian Palm Oil Board
MPOC	Malaysian P alm Oil Council
ng	nano gram
$N_A$	Avogadro's number
NAA	Naphtaleneacetic acid

OD	optical density
OMT	O-methyltransferase
PAL	phenyl ammonia lyase
pM	picoMolar
POD	peroxidase
PCR	Polymerase Chain Reaction
qPCR	quantitative Polymerase Chain Reaction
PDA	Potato Dextrose Agar
PDB	Potato Dextrose Broth
RNA	ribonucleic acid
RSPO	Round-table for Sustainable Palm Oil
RWB	rubber wood block
STDV	standard deviation
SYBR	syber
Taq	<i>Thermus aquaticus</i>
T-DNA	transfer DNA
TFA	Trifluoroacetic acid
TGA	Thioglycolic acid
Ti	tumor inducing
UPM	Universiti Putra Malaysia
USA	United States of America
<i>Vir</i>	virulence
XRF	X-ray Fluorescence



## CHAPTER I

### INTRODUCTION

Oil palm (*Elaeis guineensis*) is the world's most efficient oil bearing tree with an average yield of 3-4 tonnes of oil per hectare per year (Basiron, 2015; Wahid et al., 2005). Palm oil input to output ratio exceeds 10 times greater than its competitors, which includes soybean, rapeseed and sunflower oils. At present, the crop is widely cultivated nearing the equatorial belts (tropical regions) of the world such as Malaysia, Indonesia, Thailand, Zaire, Nigeria, Columbia and Cameroon, which collectively accounts about 9.3-10.2% of total world's permanent agricultural land (FAOSTAT, 2013). The crop thrives well under hot and humid weather, which endeavours successful growth and subsequently results to good productivity. In Malaysia, the palm oil industry has been the nation's utmost important key economic driver since its introduction in 1917, in which Tennamaram Estate in Selangor became the first oil palm plantation (Hartley, 1988). Over the years, the industry has graduated steadily to generate more income needed to beef up the nation's development while creating employment opportunities amongst its people.

The basal stem rot (BSR) disease has been recognized as the most destructive disease infecting oil palm trees in South East Asia. The disease manifests a decrease in palm stands and yield which subsequently renders economic losses (Susanto et al., 2005; Idris, 2009). To date, the status of the disease never changed, since methods to address the disease effectively remain scant. No genetic resistance to the disease has been described while control strategies remain unsatisfactory (Soepena et al., 2000). Basal stem rot (BSR) is caused by white rot fungi, *Ganoderma boninense* Pat. May. The pathogen is a saprophytic soil inhabitant, borne with ability to initiate pathogenesis at every stage of the plant growth followed by progressive death of the host. The BSR disease utilizes reproducible infected roots for pathogenesis. Rapid progression throughout the root is initiated by biotrophic phase in root cortex and lower stem via lignin degradation followed by rapid starch depletion and simultaneous cell wall breakdown (Rees et al., 2009; Flood et al., 2011).

Reports have documented an up to 500 million USD loss per year solely attributed by fungal diseases (Ommelna et al., 2012; Arif et al., 2011) in the oil palm industry. In a study conducted at oil palm plantation in Johor (Roslan and Idris, 2012), *Ganoderma* attack was shown to reduce fresh fruit bunch (FFB) production between 0.04 tonnes to 4.34 tonnes per hectare on 10 years and 22 years of planting periods respectively. The study had also estimated a total of 400 thousand hectares of oil palm plantation area in Malaysia could be affected with *Ganoderma* by 2020.

Transgenic fungal pathogens are latest advancements in genetic engineering. Since the first successful yeast *Saccharomyces cerevisiae* (Bundock et al., 1999) transformation, genetic transfer has been widely employed to explore molecular insights of fungal

pathogenicity. The underlying principle involves transferring foreign genes into organism of interest from both related and non-related organisms. *Ganoderma boninense* has never been shown amenable to transformation system. A transgenic *G. boninense* would create an avenue of understandings during BSR development. Thus, assigning function of genes and their products relevant to pathogenicity could be deduced. Biochemical changes of fungal cell wall composition during disease development can be learnt in the quest for BSR resistance. *Agrobacterium*-mediated transformation utilizes *Agrobacterium* species as the vector for its ability to transfer a discrete segment of its DNA into the fungal cells. The transferred DNA (T-DNA) is transported to fungal cell where it is integrated into the fungal genomic DNA. This is based on the presence of a large tumor inducing (Ti) plasmid in *Agrobacterium*, which contains a set of genes (the virulence genes) that can mobilize the T-DNA. *Agrobacterium*-mediated transformation is highly preferred compared to other gene transfer method due to its feasibility in preparation and higher transformation frequencies. Different kinds of intact host cells such as conidia, mycelia, sexual spores and fruiting body tissues from fungi can be relatively used through this method (Michielse et al., 2005).

Lignin supports the vascular system for transportation of water and nutrients and first line defense mechanism through lignifications in plants (Maeda et al. 2011). *Ganoderma boninense*, degrades polysaccharides and lignin components of host tissue during development of BSR. Thus, host's degree of lignifications may explain differential tolerance to BSR. Lignin content and composition selective to *G. boninense* degradation would provide information on defense strategy employed by host through lignification. The complex biochemical grid of lignin biosynthesis results from accumulative participation of numerous enzymes, each serves as confounding unit to one another to produce the end product, lignin polymer. Lignin biosynthesis is supported by a variable degree of regulation of each individual enzymes participating in the phenylpropanoid pathway. Regulatory functions of key enzyme in the phenylpropanoid pathway during defense mechanism vary in response to growth and defense (Higuchi and Kyoto, 1990). Therefore, understanding the phenylpropanoid pathway alterations would provide insights on defense lignin accumulation during BSR development.

In this study, the fungus *G. boninense* was tagged with GUS-GFP fusion gene via *Agrobacterium*-mediated transformation to discern early stage colonization pattern of *G. boninense*. Characterization of oil palm lignin in association to growth and defense was deduced. The expression of several lignin biosynthetic genes and enzymes were evaluated to understand defense responses of oil palm seedlings during pathogenesis of BSR disease and its subsequent implications on the phenylpropanoid pathway. Thus, the objectives of this study were as followings:-

i) To determine lignin content and composition of oil palm seedlings with different tolerance to BSR and its association to growth and micronutrient content.

ii) To develop an efficient *Agrobacterium*-mediated transformation protocol for *G. boninense* harbouring GUS-GFP fusion gene.

iii) To establish early stage infection pattern of basal stem rot using transgenic *G. boninense*.

iv) To determine the lignin content, lignin composition, expression and activity of lignin biosynthetic genes and enzymes (PAL, CAD and POD) during BSR development.



## REFERENCES

- Adams, D.J. (2004). Fungal cell wall chitinases and glucanases. *Microbiology* 150:2029-2035.
- Agrios, G.N. (2004). Plant Pathology. 5<sup>th</sup> Ed. Elsevier Academic Press. San Diego, CA. p. 45-62.
- Ahmad, H. and Salmah, J. (1998). Palm oil as diesel fuel: field trial on cars with Elsbett engine. In: Proceeding of the 1998 PORIM international biofuel and lubricant conference, PORIM, Bangi, pp 165-174.
- Ainsworth, A.M. (1995). Technical Information Sheet No. 11. Isolation techniques for basidiomycetes. *World Journal of Microbiology and Biotechnology* 11:364-366.
- Ali, S. and Bakkeren, S. (2011). Introduction of large DNA inserts into the barley pathogenic fungus, *Ustilago hordei*, via recombinant binary BAC vectors and *Agrobacterium*-mediated transformation. *Current Genetics* 57:63-73. doi:10.1007/s00294-010-0324-0.
- Ali, M.B., Khatam, S., Hahn, E.J. and Paek, K.Y. (2006). Enhancement of phenylpropanoid enzymes and lignin in *Phalaenopsis* orchid and their influence on plant acclimatisation at different levels of photosynthetic photon flux. *Plant Growth Regulator*. 49:137-146. doi 10:1007/s10725-006-9003-z.
- Ali, M.B. and Mc Near, D.H. (2014). Induced transcriptional profiling of phenylpropanoid pathway genes increased flavonoid and lignin content in Arabidopsis leaves in response to microbial products. *BMC Plant Biology* 14:84.
- Alizadeh, F., Siti, N.A.A., Khodavandi, A., Faridah, A., Umi, K.Y. and Chong, P.P. (2011). Differential expression of oil palm pathology genes during interactions with *Ganoderma boninense* and *Trichoderma harzianum*. *Journal of Plant Physiology* 168:1106-1113.
- An, Z. (2005). Handbook of Industrial Mycology. CRC Press. Marcel Dekker New York. p. 515-538.
- Anna, K., Tarja, T., Tapio, L., Mervi, M., Seppanen, Mika, I., Maarit, H., Perttu, V. and Pekka S. (2014). Effect of lignin content and subunit composition on digestability in clones of Timothy (*Phleum pratense* L.). *Journal of Agricultural Food Chemistry*. 62:6091-6099.
- Ana, R.D. and Ian, A.D. (2000). Dubery Panama Disease: Cell Wall Reinforcement in Banana Roots in Response to Elicitors from *Fusarium oxysporum* f. sp. *cubense* Race Four. *Biochemistry and Cell Biology*. Vol. 90, No. 10, 1173-1180.

- Annamalai, P. and Lalithakumari, D. (1991). Isolation and regeneration of protoplasts from mycelium of *Dreschlera oryzae*. *Journal of Plant Disease Protection*. 98:197-204.
- Anterola, A.M. and Lewis, N.G. (2002). Trends in lignin modification: a comprehensive analysis of the effects of genetic manipulations/ mutations on lignification and vascular integrity. *Phytochemistry* 61:221-294.
- Arif, M.S., Roslan, A. and Idris, A.S. (2011). Proceedings of the Third MPOB-IOPRI International Seminar: Integrated Oil Palm Pests and Diseases Management. Kuala Lumpur: Convention Centre; Economics of oil palm pests and Ganoderma disease and yield losses
- Ariffin, D., Idris, A.S. and Singh, G. (2000). Status of Ganoderma in Oil Palm. CAB *International*. *Ganoderma Diseases of Perennial Crops*. (eds J. Flood, P.D. Bridge and M. Holderness).
- Asada, Y., Matsumoto, I. and Tashiro, T. (1972). The formation of phenolic acids in the root of downy mildew infected Japanese radish. *Annual Phytopathology Society of Japan* 38:405-9.
- Balasubramanian, N., Gnanam, A.J., Srikalaivani, P. and Lalithakumari, D. (2003). Release and regeneration of protoplast from the fungus *Trichothecium roseum*. *Canadian Journal of Microbiology* 49:263-268.
- Barber, M.S and Mitchell, H.J. (1997). Regulation of phenylpropanoid metabolism in relation to lignin. *International Review of Cytology* Vol. 172.
- Barriere, Y. and Argiller, O. (1993). Brown-midrib genes of maize. A review. *Agronomic* 13:865-876.
- Barrière Y., Courtial A., Soler M. and Pettenati J.G. (2015). Toward the identification of genes underlying maize QTLs for lignin content, focusing on colocalizations with lignin biosynthetic genes and their regulatory MYB and NAC transcription factors. *Molecular Breeding* 35:87 doi 10.1007/s11032-015-0275-8.
- Barros-Rios, J., Malvar, R.A., Jung, H.J.G. and Santiago, R. (2011). Cell wall composition as maize defense mechanism against corn borers. *Phytochemistry* 72:365-371.
- Basiron, Y. (2015). Oil palm wrongly targeted. Retrived 16 May 2015 from [www.thestar.com/my/oilpalm-wrongly-targeted](http://www.thestar.com/my/oilpalm-wrongly-targeted).
- Basiron, Y. (2016). The Paradox of oil palm sustainability. Proceedings 11<sup>th</sup> February, 2016 in Putra Plantation Forum Series, Universiti Putra Malaysia.
- Basiron, Y. (2007). Palm oil production through sustainable plantations. *European Journal of Lipid Science and Technology* 109:289-295.

- Basiron, Y. 1996. Palm Oil. In: Hui, Y. H. (ed) Bailey's Industrial Oil and Fat Products: Edible Oil and Fat Products; Oil and Oilseeds, Vol 2, 5<sup>th</sup> ed. John Wiley, New York, p 271-375.
- Baucher, M., Halpin, C., Petit-Conil, M. and Boerjan, W. (2003). Lignin: Genetic engineering and impact on pulping. *Critical Review in Biochemistry and Molecular Biology* 38:305-350.
- Beddington, J. (2011). The future of food and farming. Retrieved on 10 December 2015 from [www.gov.uk/government/news/the-future-of-food-farming](http://www.gov.uk/government/news/the-future-of-food-farming).
- Bhuiyan N.H., Selvaraj G., Wei Y.D. and King J. (2009). Gene expression profiling and silencing reveal that monolignol biosynthesis plays a critical role in penetration defence in wheat against powdery mildew invasion. *Journal of Experimental Botany* 60:509-521.
- Bi, C., Chen, F., Jackson, L., Gill, B.S. and Li, W. (2011). Expression of lignin biosynthetic genes in wheat during development and upon infection by fungal pathogens. *Plant Molecular Biology Reports* 29:149-161. doi 10.1007/s11105-010-0219-8.
- Blanchette, R.A. (1991). Delignification by wood-decay fungi. *Annual Review of Phytopathology* 29:381-403.
- Boerjan, W., Ralph, J. and Baucher, M. (2003). Lignin biosynthesis. *Annual Review Plant Biology* 54:519-546.
- Boudet, A.M., Lapierre, C. and Grima-Pettenati, J. (1995). Transley review No. 80. Biochemistry and molecular biology of lignifications. *New Phytopathology* 129:203-236.
- Brinkmann, K. Blaschke, L. and Polle, A. (2002). Comparisons of different methods for lignin determination as a basis for calibration of near-infrared reflectance spectroscopy and implications of lignoproteins. *Journal of Chemical Ecology* 28 (12):2483-501.
- Bruce, R.J. and West, C.A. (1989). Elicitation of lignin biosynthesis and isoperoxidase activity by pectic fragments in suspension cultures of castor bean. *Plant Physiology* 91:889-897.
- Bundock, P., Mroczek, K., Winkler, A.A., Steensma, H.Y. and Hooykaas, P.J. (1999). T-DNA from *Agrobacterium tumefaciens* as an efficient tool for gene targeting in *Kluyveromyces lactis*. *Molecular General Genetics* 261:115-121.
- Campell, A.G., Kim, W.J. and Koch, P. (1990). Chemical variation in lodgepole pine with sapwood/heartwood, stem height and variety. *Wood Fiber Science* 122:22-30.

- Campbell, C.L. and Madden, L.V. (1990). Introduction to plant disease epidemiology. John Wiley & Sons, New York, New York, USA.
- Campbell, M.M. and Sederoff, R.R. (1996). Variation in lignin content and composition, mechanisms of control and implications for the genetic improvements of plants. *Plant Physiology* 110:3-13.
- Carver, T.L.W., Zeyen, R.J., Robbins, M.P., Vance, C.P. and Boyles, D.A. (1994). Suppression of host cinnamyl alcohol dehydrogenase and phenylalanine ammonia-lyase increases oat epidermal cell susceptibility to powdery mildew penetration. *Physiology and Molecular Plant Pathology* 44:243-259.
- Chabannes, M., Barakate, A., Lapierre, C., Marita, J. M., Ralph, J., Pean, M., Danoun, S., Halpin, C., Grima-Pettenati, J. and Boudet, A.M. (2001). Strong decrease in lignin content without significant alteration of plant development is induced by simultaneous down-regulation of cinnamoyl CoA reductase (CCR) and cinnamyl alcohol dehydrogenase (CAD) in tobacco plants. *The Plant Journal* 28:257-270.
- Chanda, A., Roze, L.V., Kang, S., Artymovich, K.A., Hicks, G.R., Raikhel, N.V., Calvo, A.M. and Linz, J.E. (2009). A key role for vesicles in fungal secondary metabolism. *Proceedings of National Academy of Sciences U.S.A.* 106:19533-19538.
- Chapple, C.C., Vogt, T., Ellis, B.E. and Somerville, C.R. (1992). An Arabidopsis mutant defective in the general phenylpropanoid pathway. *Plant Cell* 4:1413-1424.
- Chen, F., Wang, M., Zheng, Y., Luo, J., Yang, X. and Wang, X. (2010). Quantitative changes of plant defense enzymes and phytohormone in biocontrol of cucumber Fusarium wilt by *Bacillus subtilis* B579. *World Journal of Microbiology Biotechnology*. 26:675–684 doi 10.1007/s11274-009-0222-0.
- Chen, X., Stone, M., Schlagnhauser, C. and Romaine, C.P. (2000). A fruiting body tissue method for efficient Agrobacterium-mediated transformation of *Agaricus bisporus*. *Applied Environmental Microbiology* 66:4510- 4513.
- Cheng, H., Li, L., Xu, F., Cheng, S., Cao, F., Wang, Y., Yuan, H., Jiang, D. and Wu, C. (2013). Expression patterns of cinnamyl alcohol dehydrogenase gene involved in lignin biosynthesis and environmental stress in *Ginkgo biloba*. *Molecular Biology Reports* 40:707-721. doi 10.1007/s11033-012-2111-0.
- Chin, C., Lee, Y.W., Tan, J.S. and Syed Alwee, S.S.R. (2012). Amplification and sequencing of partial-length disease resistance gene homologues coding for NBS–LRR proteins in oil palm (*Elaeis guineensis*). *AsPac Journal of Molecular Biology and Biotechnology* 20:25-31.

- Choi, S.H., Kim, B.K., Kim, H.W., Kwak, K.C., Choi, E.C., Kim, Y.C., Yoo, Y.B. and Park, Y.B. (1987). Studies on protoplast formation and regeneration of *Ganoderma lucidum*. *Archives of Pharmacology Research* 10(3):158-164.
- Choi, Y.W., Hyde, K.D. and Ho, W.W.H. (1999). Single spore isolation of fungi. *Fungal Diversity* 3:29-38.
- Choo, Y.M., Yung, C.L. and Ma, A.N. (2012). Chapter 17 Oil palm. In handbook of Bioenergy Crop Plants. p.433-451.
- Choo, Y.M. and Cheah, K.Y. (2000). Biofuel: In: Yusof, B., Jalanni B. S., Chan, K.W. (eds), *Advances of Oil Palm Research, Vol II*. Malaysian Palm Oil Board, Malaysia. pp 214-241.
- Choo, Y.M., Yap, S.C., Ooi, C.K., Ma, A.N., Goh, S.H. and Ong, A.S.H. (1996). Recovered oil from palm pressed fiber: a good source of natural carotenoids, vitamin E and sterols. *Journal of American Oil Chemistry Society* 73:599-602.
- Chou, T.H. and Tzean, S.S. (2015). Protoplasting, regeneration and transformation of medicinal mushroom *Ganoderma multipileum* using succinate dehydrogenase mutation gene as a selection marker. *Annals of Microbiology*. doi.10.1007/s13213-015-1087-0.
- Chiang, V.L. and Funaoka, M. (1988). The formation and quantity of diphenylmethane type structures in residual lignin during kraft delignification of Douglas-fir. *Holzforschung* 42:385-389.
- Collopy, P.D., Colot, H.V., Park, G., Ringelberg, C., Crew, C.M., Borkovich, K.A. and Dunlap, J.C. (2010). High-throughput construction of gene deletion cassettes for generation of *Neurospora crassa* knock-out strains. *Methods Molecular* 638:33-40.
- Conesa, A., Punt, P.J., van Luijk, N. and van den Hondel, C.A.M.J.J. (2001). The secretion pathway in filamentous fungi: A biotechnological view. *Fungal Genetics Biology* 33:155-171.
- Collopy, P.D., Colot, H.V., Park, G., Ringelberg, C., Crew, C. M., Borkovich, K.A. and Dunlap, J.C. (2010). High-throughput construction of gene deletion cassettes for generation of *Neurospora crassa* knock-out strains. *Methods Molecular* 638:33-40.
- Corley, R.H.V. (1983). Potential productivity of tropical perennial crops. *Experimental Agriculture* 19:217-237.
- Corley, R.H.V and Tinker, P.B. (2003). *The Oil palm*. 4<sup>th</sup> Edition. Blackwell Sciences, Ames, Iowa, USA.



- Dean, R.A., Talbot, N.J., Ebbole, D.J., Farman, M.L., Mitchell, T., Orbach, M., Thon, M., Kulkarni, R., Xu, J.R., Pan, H. et al., (2005). The genome sequence of the rice blast fungus *Magnaporthe grisea*. *Nature* 434:980–986.
- de Groot, M.J.A., Bundock, P., Hooykaas, P.J.J. and Beijersbergen, A.G.M. (1998). *Agrobacterium tumefaciens*-mediated transformation of filamentous fungi. *Nature Biotechnology* 16:839-842.
- DeWick, P.M. (1994). The biosynthesis of shikimate metabolites. *National Proceeding of Reproduction* 11:173-203.
- Dietrich, F.S., Voegeli, S., Brachat, S., Lerch, A., Gates, K., Steiner, S., Mohr, C., Pohlmann, R., Luedi, P., Choi, S. et al. (2004). The *Ashbya gossypii* genome as a tool for mapping the ancient *Saccharomyces cerevisiae* genome. *Science* 304:304–307.
- Dhar, P. and Kaur, G. (2009). Optimization of different factors for efficient protoplast release from entomopathogenic fungus *Metarhizium anisopliae*. *Annals of Microbiology* 59 (1):183-186.
- Dharmaputra, O.S., Tjitrosomo, H.S. and Abadi, A.I. (1989). Antagonistic effect of four fungal isolates to *Ganoderma boninense*, the causal agent of basal stem rot of oil palm. *Biotrophia* 3:41-49.
- Diaz, J., Bernal, A., Pomar, F. and Merino, F. (2001). Induction of shikimate dehydrogenase and peroxidase in pepper (*Capsicum annuum* L.) seedlings in response to copper stress and its relation to lignifications. *Plant Science* 161:179-188.
- Dong, Y.C., Dai, Y.N., Xu, T.Y., Cai, J., and Chen, Q.H. (2014). Biodegradation of chestnut shell and lignin-modifying enzyme production by the white rot fungi *Dichomitus squalens*, *Phlebia radiata*. *Bioprocess Biosystem Engineering* 37:75.
- Donk, M.A. (1964). A conspectus of families of Aphyllophorales. *Persoonia* 3:199-324.
- Dorak, M. T. Real-time PCR. 2007; Taylor and Francis Group /<http://www.gene-quantification.com/dorak-book-real-time-pcr-2006.pdf>.
- Dunkle L.D. In Chapter 2 Factors in Pathogenesis. Plant-Microbe Interactions. Molecular and Genetic Perspectives Vol 1. Macmillian Publishing company, London.
- Durkovic, J., Kack, F., Olck, D., Kucerova, V. and Krajnakova. (2014). Host responses and metabolic profiles of wood components in Dutch elm hybrids with a contrasting tolerance to Dutch elm disease. *Annals of Botany* 114:47-59.

- Dutse, S.W., Yusof, N.A., Ahmad, H., Hussein, M.Z. and Hushiarian, R. (2012). DNA based biosensor for detection of *Ganoderma boninense* pathogen of the oil palm utilizing a new ruthenium complex, Cl<sub>2</sub>. *International Journal of Electrochemical Science* 7:8105-8115.
- Dutse, S.W., Yusof, N.A., Ahmad, H., Hussein, M.Z. and Hushiarian, R. (2013). DNA-based biosensor for detection of *Ganoderma boninense*, an oil palm pathogen utilizing a new ruthenium complex, [Ru (dppz) <sub>2</sub> (qtpy)] <sub>2</sub> based on a PEDOT-PSS/Ag nanoparticles modified electrode. *International Journal Electrochemical Science* 8:11048-11057.
- Elissetche, J.P., Valenzuela, S., Garcia, R., Norambuena, M., Iturra, C., Rodriguez, J., Mendonea, R.T. and Balocchi, C. (2011). Transcript abundance of enzymes involved in lignin biosynthesis of Eucalyptus globules genotypes with contrasting levels of pulp yield and wood density. *Tree Genetics and Genomes* 7:697-705. Doi.10.1007/s11295-011-0367-5.
- Eudes, A., Pollet, B., Sibout, R., Do, C.T., Seguin, A., Lapierre, C. and Jouanin, L. (2006). Evidence for a role of AtCAD1 in lignifications of elongating stems of *Arabidopsis thaliana*. *Planta* 225:23-39.
- Eppendorfer, W.H. and Eggum, B.O. (1994). Effects of sulphur, nitrogen, phosphorus, potassium and water stress on dietary fibre fractions, starch, amino acids and on the biological value of potato protein. *Plant Foods for Human Nutrition* 45:299-313.
- Eriksson K.E.L., Blanchette, R.A. and Ander, P. (1990). Microbial and enzymatic degradation of wood components . Springer, Berlin-Heidelberg, New York.
- Espino, J.J., Gutiérrez-Sánchez, G., Brito, N., Shah, P., Orlando, R., González, C. (2010). The *Botrytis cinerea* early secretome. *Proteomics* 10:3020-3034.
- Eynck, C., Seguin-swartz, G., Clarke, W.E. and Parkin, I.A.P. (2012). Monolignol biosynthesis is associated with resistance to *Sclerotinia sclerotiorum* in *Camelia sativa*. *Molecular Plant Pathology* 13(8):887-899.
- Faix, O., Mozuch, M.D. and Kirk, T.K. (1985). Degradation of gymnosperm (Guaiacyl) vs. angiosperm (Syringyl/guaiacyl) lignins by *Phanerochaete chrysosporium*. *Holzforshung* 39:203-208.
- FAO (2016). FAO Crop Statistics. Retrived on 14 March 2014 from [www.fao.org/crop/statistics](http://www.fao.org/crop/statistics).
- Fang, W.C. and Kao, C.H. (2000). Enhanced peroxidase activity in rice leaves in response to excess iron, copper and zinc. *Plant Science* 158:71-76.

- Fitzgerald, A.M., Mudge, A.M., Gleave, A.P. and Plummer, K.M. (2003). *Agrobacterium* and PEG-mediated transformation of the phytopathogen *Venturia inaequalis*. *Mycological Research* 107:803-810.
- Flood, J., Cooper, R., Rees, R., Potter, U. and Hasan, Y. (2011). Some latest research and development on Ganoderma Diseases in Oil Palm. *Bulletin*.
- Flor, H.H. (1956). The complementary genetic systems in flax and flax rust. *Advances in Genetics* 8:29-54.
- Flowers, J.L. and Vaillancourt, L.J. (2005). Parameters affecting the efficiency of *Agrobacterium tumefaciens*-mediated transformation of *Colletotrichum graminicola*. *Current Genetics* 48:380-388.
- Francoz, E., Ranocha, P., Nguyen-Kim, H., Jamet, E., Burlat, V. and Dunand. (2014) Roles of cell wall peroxidases in plant development. *Phytochemistry*. doi:10.1016/j.phytochem. 2014.07.020.
- Frandsen, R.J. (2011). A guide to binary vectors and strategies for targeted genome modification in fungi using *Agrobacterium tumefaciens*-mediated transformation. *Journal of Microbiological Methods* 87(3):247-262.
- Frandsen, R.J.N. (2015). Chapter 14. *Agrobacterium tumefaciens*-mediated transformation. In Transformation Systems in Fungi, Vol 1201510.1007/978-3-319-10142-2\_14. Bergand, A.A.V.D & Maruthachalam, K. Springer International Publishing Switzerland.
- Gardner, C.J., Quinn, N.W.T. Gerpen, J.V. and Simonpietri, J. (2010). Oilseed and algal oils as biofuel feedstocks. Available at <http://www.Swcs.org/documents/resources/Chapter 8 .pdf>. (Accessed 2 July 2015)
- Geetha, N.E, Amruthesh, K.N., Sharathchandra, R.G. and Shetty, H.S. (2005). Resistance to downy mildew in pearl millet is associated with increased Phenylalanine ammonia-lyase activity. *Functural Plant Biology* 32:1-9.
- Genc, Y., Humphries, J.M., Lyons, G.H. and Graham, R.D. (2005). Exploiting genetic variation in plant nutrient accumulation to alleviate micronutrient deficiency in populations. *Journal of Trace Elements in Medicine and Biology* 18:319-324.
- Gilbertson, R.L. and Ryvarden, L. (1986). *North Americal Polypores*. Part I. Fungiflora, Oslo, Norway.
- Giordano, A, Liu, Z., Panter, S.N., Dimech, A.M., Shang, Y., Wijesinghe, H, Fulgueras, F., Ran, Y., Mouradov, A., Rochfort, F., Patron, N. J. and Spangenberg, G.C. (2014). Reduced lignin content and altered lignin composition in the warm season forage grass *Paspalum dilatatum* by a down-regulation of a Cinnamyl CoA Reductase Gene. *Transgenic Research* 25:503-517.

- Gnanam, A.J. (2013). Protoplast Fusion techniques in Fungi. In Gupta, V. et al., Eds. Laboratory Protocols in Fungal Biology; Current Methods in Fungal Biology. *Fungal Biology*, pp 483-488.
- Goffeau, A., Barrell, B.G., Bussey, H., Davis, R.W., Dujon, B., Feldmann, H., Galibert, F., Hoheisel, J.D., Jacq, C., Johnston, M. et al., (1996). Life with 6000 genes. *Science* 546:563-567.
- Grabber J.H., Jung, G.A. and Hill, R. (1991). Chemical composition of parenchyma and sclerenchyma cell-walls isolated from orchard-grass and switchgrass. *Crop Sci* 3(4):1058-1065.
- Grand, C., Boudet, A. M. and Ranjeva, R. (1982). Natural variations and controlled changes in lignification process. *Holzforschung* 36:217-223.
- Gruber, S. and Seidl-Seiboth, V. (2012). Self versus non-self fungal cell wall degradation in *Trichoderma*. *Microbiology* 158:26-34.
- Gunston, F. D. (2011). Production and trade of vegetable oils. In: Gunstone, F. D. (ed) Vegetable Oils in Food Technology: Composition, Properties and uses, 2<sup>nd</sup> Edn. Blackwell Publishing, Oxford, UK, pp. 1-24.
- Hale, M.D. and Eaton, R.A. (1985). The ultrastructure of soft rot fungi. II. Cavity-forming hyphae in wood cells. *Mycologia* 77:594-605.
- Halpin, C., Knight, M. E., Foxon, G. A., Campbell, M. M., Boudet, A. M., Boon, J. J., Chabbert, B., Toller, M. T. and Schuch, W. (1994). Manipulation of lignin quality by down-regulation of cinnamyl alcohol dehydrogenase. *Planta* 6:339-350.
- Hartley, C.W.S. (1988). The Oil palm. 3<sup>rd</sup> Edition. Longmans, London, UK.
- Hashiba, T. 1992. Isolation of fungal protoplasts. In Handbook of Applied Mycology. Vol. 4. Fungal Biotechnology (D.K. Arora, R.P. Elander & K.J. Mukerji, eds):129-149. Marcel Dekker, New York.
- Hatakka, A. and Hammel, K.E. (2010). Fungal Biodegradation of Lignocelluloses. In: Hofrichter M (ed.), Industrial Applications, 2<sup>nd</sup> edn. The Mycota X. Springer, Berlin, Heidelberg, pp. 319-340.
- Hatakka, A. (1994). Lignin modifying enzymes from selected white rot fungi production and role in lignin biodegradation. *FEMS Microbiology Review* 13:125-135.
- Highley, T.L. 1982. Influence of type and amount of lignin on decay by *Coriolus versicolor*. *Canadian Journal of Forest Research* 12:435-438.
- Higuchi, T. (1990). Lignin biochemistry: Biosynthesis and biodegradation. *Wood Science Technology* 24:23-63.

- Higuchi, T. (1985). In Biosynthesis and Biodegradation of Wood Components (Higuchi, T., ed.), p. 141. Academic Press, Orlando.
- Hoch, H.C. and Staples, R.C. (1987). Structural and chemical changes among the rust fungi during appressorium development. *Annual Review Phytopathology* 25:231-247.
- Ho, Y.W. and Nawawi, A. (1985). *Ganoderma boninense* Pat. from basal stem rot of oil palm (*Elaeis guineensis*) in Peninsular Malaysia. *Pertanika* 8:425-428.
- Hu, W.J., Harding, S.A., Lung, J., Popko, J.L., Stokke, D.D., Tsai, C.J. and Chiang, V.L. (1999). Repression of lignin biosynthesis promotes cellulose accumulation and growth in transgenic trees. *Nature Biotechnology* 17:808-812.
- Ianir, G. and Idnurm, A. (2015). *Agrobacterium tumefaciens*-Mediated transformation of pucciniomycotina red yeasts. Springer International Publishing Switzerland.
- Idnurm, A., Reedy, J.L., Nussbaum, J.C. and Heitman, J. (2004). *Cryptococcus neoformans* virulence gene discovery through insertional mutagenesis. *Eukaryot Cell* 3:420-429.
- Idris, A.S. (2009). Basal stem rot in Malaysia-Biology, epidemiology, economic importance, detection and control. Proc. Of the international Workshop on Awareness, Detection and Control of Oil Palm Devastating Diseases. p.18-62.
- Idris, A.S. and Ariffin, D. (2004). Basal Stem Rot-Biology, Detection and Control. In: International Conference on Pest and Disease of Importance to the Palm Industry, 18-19 May 2004, Kuala Lumpur, Malaysia. Paper no 9.
- Idris, A.S., Khusairi, D., Ismail, S. and Ariffin, D. (2002). Selection for partial resistance in oil palm to *Ganoderma* basal stem rot. In: The Seminar Recent Progress in the Management of Peat and *Ganoderma*, 6-7 may 2002, Bangi, 22 pp.
- Idris, A.S. and Rafidah, A.R. (2008). Polyclonal antibody for detection of *Ganoderma*. MPOB Inf Ser MPOB TT No. 405.
- Idris, A.S., Rajinder, S., Madihah, A.Z. and Mohd, B.W. (2010). Multiplex PCR-DNA kit for early detection and identification of *Ganoderma* species in oil palm. MPOB Inf Ser, MPOB TS No. 73.
- Idris, A.S., Nasyaruddin, M.N.M., Maizatul, S.M. and Zaiton, S. (2010). GanoEB1- A bacterial biocontrol agent for *Ganoderma* in oil palm. MPOB Information Series No. 500.

- Idris, AS., Yamaoka, M., Hayakawa, S., Basri, M.W., Noorhasimah, I. and Ariffin, D. (2003). PCR technique for detection of *Ganoderma*. MPOB Infer Ser MPOB Tt No. 188: MPOB TT No. 188, MPOB TT No. 188.
- Iiyama, K., Lam, T.B.T., and Stone, B.A. (1994). Covalent cross-links in the cell wall. *Plant Physiology* 104:315-320.
- Izzati, M.Z. and Abdullah, F. (2008). Disease suppression in *Ganoderma*-infected oil palm seedlings treated with *Trichoderma harzianum*. *Plant Protection Science* 44(3):101-107.
- Jameel, R.A., Yusmin, M.Y., Nurhanani, R., Jaime, J.J., Tey, C.C., Normahnani, M.N., Sarni, M.J., Rofina, Y.O. and Onn, H.H. (2014). Identification of proteins of altered abundance in oil palm infected with *Ganoderma boninense*. *International Journal of Molecular Sciences* 15:5175-5192.
- Jalani, B.S., Yusof, B., Ariffin, D., Chan, K.W. and Rajanaidu, N. (2002). Prospect of Elevating National Oil Palm Productivity: A Malaysian Perspective. MPOB, Bangi.
- Jayaraj, J., Yi, H., Liang, G.H., Muthukrishnan, S. and Velazhahan, R. (2004). Foliar application of *Bacillus subtilis* AUBS1 reduces sheath blight and triggers defense mechanisms in rice. *Journal of Plant Disease Protection* 111:115–125.
- Jin, Q., Ye, H., and Zhang, M. (2003). Relationship between the activity of PAL and resistance of corn to maize sheath blight. *Journal of Sichuan Agriculture* 2:13
- Johal, G.S., Gray, J. and Briggs, S.P. (1995). Convergent insights into mechanisms determining disease and resistance response in plant-fungal interactions. *Canadian Journal of Botany* 73(Suppl.):468-474.
- Jones, D.L. (1994). *Palms Throughout the World*. Reed Books, Australia.
- Jones, J.D., Henstrand, J.M., Handa, A.K., Herrmann, K.M. and Weller, S.C. (1995). Impaired wound induction of 3-deoxy-D-arabino-heptulosonate-7-phosphate (DAHP) synthase and altered stem development in transgenic potato plants expressing a DAHP synthase antisense construct. *Plant Physiology* 108:1413-1421.
- Jounin, L., Goujon, T., de Nadai, V., Martin, M.T., Mila, I., Vallet, C., Pollet, B., Yoshinnaga, A., Chabbert, B., Petit-Conil, M. and Lapierre, C. (2000). Lignification in transgenic Poplars with extremely reduced Caffeic Acid O-Methyltransferase activity. *Plant Physiology* 123:1363-1374.
- Jung , H.J.G., Vogel, K.P. (1992). Lignification of switchgrass (*Panicum virgatum*) and big bluestem (*Andropogon gerardii*) plant-parts during maturation and its effects on fiber degradability. *Journal of Science Food Agriculture* 59(2):169-176.

- Kamada, T. (2002).Molecular genetics of sexual development in the mushroom *Coprinus cinereus*. *Bioessay* 24:449-459.
- Kan, W. (2006). Ed Methods in Agrobacterium protocols. *Methods in Molecular Biology*.Springer.
- Kanoa, S., Kuritaa, T., Kanematsub, S. and Morinagaa, T. (2011).Agrobacteriumtumefaciens mediated Transformation of the Plant Pathogenic Fungus *Rosellinia necatrix*. ISSN 00262617, Microbiology, Vol. 80, No. 1, pp. 82-88.
- Kato, M., Hayakawa, Y., Hyodo, Y. and Yano, M. (2000). Wound-induced ethylene synthesis and expression and formation of 1-aminocyclopropane-1-carboxylate (ACC) synthase, ACC oxidase, phenylalanine ammonia-lyase and peroxidase in wounded mesocarp tissue of *Cucurbita maxima*. *Plant Cell Physiology* 41:440-447.
- Kawaoka, A., Nanto, K., Ishii, K. and Ebinuma, H. (2006). Reduction of lignin content by suppression of expression of the LIM domain transcription factor in *Eucalyptus camaldulensis*. *Silvae Genetics* 55: 269-277.
- Keen, N.T. (1992).The molecular biology of disease resistance. *Plant Molecular Biology* 19:109-122.
- Kellis, M., Patterson, N., Endrizzi, M., Birren, B. and Lander, E.S. (2003). Sequencing and comparison of yeast species to identify genes and regulatory elements.*Nature* 423:241–254.
- Kelly, F.D. and Nurse, P. (2011).de Novo growth zone formation from fission yeast spheroplasts. *PLoS One* 6:e27977.
- Kim, Y.H., Bae, J.M. and Huh, G.H. (2010).Transcriptional regulation of the cinnamyl alcohol dehydrogenase gene from sweetpotato in response to plant developmental stage and environmental stress.*Plant Cell Reports* 29:779-791. doi 10.1007/s00299-010-0864-2.
- Kitamoto Y., N. Mori, M. Yamamoto., T. Ohiwa. and Y. Ichikawa. (1988). A simple method for protoplast formation and improvement of protoplast regeneration from various fungi using an enzyme from *Trichoderma harzianum*. *Applied Microbiology and Biotechnology* 28:445-450.
- Koike, N., Hyakumachi, M., Kageyama, K., Doke, N. (2001). Induction of systemic resistance in cucumber against several diseases by plant growth-promoting fungi: lignification and superoxide generation. *European Journal of Plant Pathology* 107:523-533.
- Kosuga, K. and Nestler, P. (1984). Plant-Microbe Interactions. Molecular and Genetic Perspectives Vol 1. Macmillian Publishing company, London.

- Kues, U. (2000). Life history and developmental processes in the basidiomycete *Coprinus cinereus*. *Microbiology and Molecular Biology Reviews* Vol. 64, No. 2:316-353.
- Kuo, C.Y., Chou, S.Y. and Huang, C.T. (2004). Cloning of glyceraldehydes-3-phosphate dehydrogenase gene and use of the *gpd* promoter for transformation in *Flammulina velutipes*. *Applied Microbiol Biotechnol.* 65:593-599. doi:10.1007/s00253-004-1635-1.
- Kruske C.R., Kaysie L.B., Dante L.A., Peter C.S. Karen K.H. and Paul J.J. (1998). Small-scale DNA sample preparation method for field PCR Detection of Microbial Cells and Spores in Soil. *Applied and Environmental Microbiology*. 64:2463-2472.
- Lalithakumari, D. (1996). Protoplasts-A biotechnological tool for plant pathological studies. *Indian Phytopathology* 49:199-212.
- Leclerque, A., Wan, H., Abschutz, A., Chen, S., Mitina, G.V. and Schairer, G.Z.H. (2004). *Agrobacterium* -mediated insertional mutagenesis (AIM) of the entomopathogenic fungus *Beauveria bassiana*. *Current Genetics* 45:111-119. doi 10.1007/s00294-003-0468-2.
- Lam, T.B.T., Iliyama, K. and Stone, B.A. (2003). Hot alkali-labile linkages in the walls of the forage grass *Phalaris aquatic* and *Lolium perenne* and their relation to in vitro digestability. *Phytochemistry* 64:603-607.
- Lange, B.M., Lapiere, C. and Sanderman, H. (1995). Elicitor-induced spruce stress lignin. Structural similarity to early developmental lignin. *Plant Physiology* 108:1277-1287.
- Law K.N., Kokta, B.V. and Mao, C.B. (2001). Fibre morphology and soda sulphite pulping of switchgrass. *Bioresource Technology* 77(1):1-7.
- Leon, J., Lawton, M. A. and Raskin, L. (1995). Hydrogen peroxide stimulates salicylic acid biosynthesis in tobacco. *Plant Physiology* 108:1673-1678.
- Lewis, N.G. and Yamamoto, E. (1990). Lignin: Occurrence, biogenesis, and biodegradation. *Annual Review of Physiology and Plant Molecular Biology* 41:455-496.
- Li, S., Fang, X., Li, M., Mu, D., Ren, A., Tan, Q. and Zhao, M. (2011). Development of a simple and efficient transformation system for the basidiomycetous medicinal fungus *Ganoderma lucidum*. *World Journal of Microbiol Biotechnology* doi: 10.1007/s11274-011-0818-z.



- Li, C.Q., Shao, J.F., Wang, Y.J., Li, W.B., Guo, D.J., Yan, B., Xia, Y. and Peng, M. (2013). Analysis of banana transcriptome and global gene expression profiles in banana roots in response to infection by race 1 and tropical race 4 of *Fusarium oxysporum* f. sp. cubense. *BMC Genomics*. 14:851.doi:10.1186/1471-2164-14-851.
- Lin, C.C., Chen, L.L. and Liu, Z.H. (2005). Rapid effect of copper on lignin biosynthesis in soybean roots. *Plant Science* 168:855-861.
- Lin, S.Y. and Dence, C.W. (1992). *Methods in Lignin Chemistry*, Springer-Verlag, Berlin.
- Liu, Q., Zheng, L., He, F., Zhao, F. J., Shen, Z. and Zheng, L. (2015). Transcriptional and physiological analyses identify a regulatory role for hydrogen peroxide in the lignin biosynthesis of copper-stressed rice roots. *Plant Soil* 387:323-336.
- Livak, K.J. and Schmittgen, T.D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the  $2^{-\Delta\Delta C_T}$  Method. *Methods*. 25(4):402-8.
- Lorenz, A.J., Anex, R.P., Isci, A., Coors, J.G., Leon, N. and Weimer, P.J.X. (2009). Forage quality and composition measurements as predictors of ethanol yield from maize (*Zea mays* L.) stover. *Biotechnology in Biofuel* 2:5.
- Lundell, T.K., Makela, M .R. and Hilden, K. (2010). Lignin-modifying enzymes in filamentous basidiomycetes-ecological, functional and phylogenetic reviews. *Journal of Basic Microbiology* 50:5-20.
- Makela, M.R., Lundell, T., Hatakka, A. and Hilden, K. (2013). Effect of copper, nutrient nitrogen, and wood-supplement on the production of lignin-modifying enzymes by the white-rot fungus *Phlebia radiata*. *Fungal biology* 117:62-70.
- Malmierca M.G., Cardoza, R.E. and Gutierrez, S. (2015). *Trichoderma* Transformation Methods. In van den Berg M.A. and K. Maruthachalam (eds), Genetic Transformation Systems 41 in Fungi, Vol 1, *Fungal Biology*.
- Malonek, S. and Meinhardt, F. (2001). *Agrobacterium tumefaciens* mediated genetic transformation of the phytopathogenic ascomycete *Calonectaria morganii*. *Current Genetics* 40:152-155/doi 10.1007/s002940100236.
- Mann, D.G.J., Burris, J.N., Sykes, R.W., Gracom, K., Kline, L., Swamidoss, I.M., Davis, M, Stewart Jr., C.N. (2009). Rapid assessment of lignin content and structure in switchgrass (*Panicum virgatum* L.) grown under different environmental conditions. *Bioenergy Research* 2:246-256. doi 10.1007/s12155-009-9054-x.

- Martinez, D., Larrondo, L.F., Putnam, N., Gelpke, M.D., Huang, K., Chapman, J., Helfenbein, K.G., Ramaiya, P., Detter, J.C., Larimer, F. et al., (2004). Genome sequence of the lignocellulose degrading fungus *Phanerochaete chrysosporium* strain RP78. *Nature Biotechnology* 22:695-700.
- Masarin, F., Gurpilhares, D.B., Baffa, D.C.F., Barbosa, M.H.P., Carvalho, W., Ferraz, A. and Milagres, A.M.F. (2011). Chemical composition and enzyme digestability of sugarcane clones selected for varied lignin content. *Biotechnology for Biofuels* 4:55.
- Maeda, H., Shasany, A.K., Schnepf, J., Orlova, I. and Taguchi, G. (2010). RNAi suppression of *Arogenate Dehydratase1* reveals that phenylalanine is synthesized predominantly via the arogenate pathway in petunia petals. *Plant Cell* 22:832-849.
- Maeda, H., Yoo, H. and Dudareva, N. (2011). Prephenate aminotransferase directs plant phenylalanine biosynthesis via arogenate. *Nature Chemistry and Biology* 7:19-21.
- Menden, B., Kohlhoff, M. and Moerschbacher, B.M. (2007). Wheat cells accumulate a syringyl-rich lignin during the hypersensitive resistance response. *Phytochemistry* 68:513-520.
- Mendoza, C.G. (1992). Cell wall structure and protoplast reversion in basidiomycete. *World Journal of Microbiology and Biotechnology*. Vol 8:Supplement 1.
- Meyer, V., Mueller, D., Strowig, T. and Stahl, U. (2003). Comparison of different transformation methods for *Aspergillus giganteus*. *Current Genetics* 43:371-377.
- Mikosch, T.S.P., Lavrijssen, B., Sonnenberg, A.S.M. and van Griensven, L.J.L.D. (2001). Transformation of the cultivated mushroom *Agaricus bisporus* (Lange) using T-DNA from *Agrobacterium tumefaciens*. *Current Genetics* 39:35-39.
- Michielse, C.B., Ram, A.F., Hooykaas, P.J. and Hondel, C.A. (2004). Role of bacterial virulence proteins in *Agrobacterium*-mediated transformation of *Aspergillus awamori*. *Fungal Genetics Biology* 41:571-578.
- Michielse, C.B., Hooykaas, P.J.J., van den Hondel, C.A.M.J.J. and Ram, A.F.J. (2005). *Agrobacterium*-mediated transformation as a tool for functional genomics in fungi. *Current Genetics* 48:1-17.
- Mims, C.W. (1991). Using electron microscopy to study plant pathogenic fungi. *Mycologia* 83:1-19.
- Mitchell, A. and Walters, D. (1995). Hydrogen peroxide stimulates salicylic acid biosynthesis in tobacco. *Plant Physiology* 108:1673-1678.

- Mohammadi, M. and Kazemi, H. (2002). Changes in peroxidase and polyphenol oxidase activities in susceptible and resistant wheat heads inoculated with *Fusarium graminearum* and induced resistance. *Plant Science* 162:491-498.
- Moller, R., Steward, D., Phillips, L., Heather, F. and Wagner, A. (2005). Gene silencing of cinnamyl alcohol dehydrogenase in *Pinus radiata* callus cultures. *Plant Physiology and Biochemistry* 43:1061-1066.
- Monzon, G.C., Pinedo, M., Rienzo, J.D., Esther, N.U., Pomar, F., Lamattina, L. and Canal, L.D.L. (2014). Nitric oxide is required for determining root architecture and lignin composition in sunflower. Supporting evidence from microarray analyses. *Nitric Oxide* 39:20-28.
- Moura, J.C.M.S., Bonine, C.A.V., De Oliveira, F.V.J., Dornelas, M.C. and Mazzafera, P. (2010). Abiotic and biotic stresses and changes in the lignin content and composition in plants. *Journal of Integrative Plant Biology* 52(4):360-376.
- Moročko-Bičevska, I. and Fatehi, J. (2011). Infection and colonization of strawberry by *Gnomonia fragariae* strain expressing green fluorescent protein. *European Journal of Plant Pathology* 129:567-577.
- Moura-Sobczak, J., Souza, U. and Mazzafera, P. (2011). Drought stress and changes in the lignin content and composition in *Eucalyptus*. *BMC Proceedings* 7:103.
- Moura, J.C.M.S., Bonine, C.A.V., Viana, J.O.F., Dornelas, M.C. and Mazzafera, P. Abiotic and biotic stresses and changes in the lignin content and composition in Plants. *Journal of Integrated Plant Biology* 52(4):360-376.
- Mrinalini, C. and Lalithakumari, D. (1996). Protoplast fusion: A biotechnological tool for strain improvement of *Trichoderma sp.* *Current Trend Life Sciences* 21:133-146.
- Mullin, E.D. and Kang, S. (2001). Transformation: A tool for studying fungal pathogens of plants. *Cell and Molecular Life Sciences* 58:2043-2052.
- Murlidhar, R.V. and Panda, T. (2000). Fungal protoplast fusion: A revisit. *Bioprocess Biosystem Engineering* 22:429-431.
- Murphy, D.J. 2007. Future prospects for oil palm in the 21<sup>st</sup> century: biological and related challenges. *European Journal of Lipid Science and Technology* 109:296-306.
- Naher, L., Ho, C.L., Tan, S.G., Yusuf, U.K., Ahmad, S.H. and Abdullah, F. (2011). Cloning of transcripts encoding chitinases from *Elaeis guineensis* Jacq. and their expression profiles in response to fungal infections. *Physiology and Molecular Plant Pathology* 76:96-103.

- Nakamura, M., Kuwahara H., Onoyama, K. and Iwai, H. (2012). *Agrobacteriumtumefaciens*-Mediated Transformation for Investigating Pathogenicity Genes of the Phytopathogenic Fungus *Colletotrichum sansevieriae*. *Current Microbiology* 65:176-182 doi 10.1007/s00284-012-0140-5.
- Nakashima, J., Chen, F., Jackson, L., Shadle, G. and Dixon, R.A. (2008). Multi-site genetic modifications of monolignol biosynthesis in alfalfa (*Medicago sativa*); effects on lignin composition in specific cell types. *New Phytologist* 179:738-750.
- Narayanasamy, P. 2011. Microbial plant pathogen-detection and disease diagnosis. In: *Bacterial and Phytoplasmal Pathogens*. The Netherlands: Springer. 2:5-169.
- Neish, A.C. (1961). Formation of m- and p-coumaric acids by enzymatic deamination of the corresponding isomers of tyrosine. *Phytochemistry* 1:1-24.
- Neth, J., Niemann, G.J. and Baayen, R.P. (1990). Localization of phytoalexin accumulation and determination of changes in lignin and carbohydrate composition in carnation (*Dianthus caryophyllus L.*) xylem as a consequence of infection with *Fusarium oxysporum* f. sp. Dianthi, by pyrolysis-mass spectrometry. *Journal of Plant Pathology* 6:133-153.
- Ngah, H. (2010). *Ganoderma punca pangkal sawit* reput. Retrived 21 December 2010 from [www.bharian.com.my/bharian/articles/ganodermapuncapangkalsawitreput](http://www.bharian.com.my/bharian/articles/ganodermapuncapangkalsawitreput).
- Nicholson, R.L. (1992). Phenolic compounds and their role in disease resistance. *Annual Reviews in Phytopathology* 30:369-389.
- Nicole, M., Ruel, K. and Ouellette, B. (1994). Fine morphology of fungal structures involved in host wall alterations. In *Host wall alterations in parasitic fungi*. Pertini O. APS press St. Paul, Minnesota.
- Novaes, E., Kirst, M., Winter-Sederoff, H. and Sederoff, R. (2010). Lignin and biomass: a negative correlation for wood formation and lignin content in trees. *Plant Physiology* 154:555-561.
- Novaes, E., Osorio, L., Drost, D.R., Miles, B.L., Boaventura-Novaes, C.R., Benedict, C. et al., (2009). Quantitative genetic analysis of biomass and wood chemistry of *Populus* under different nitrogen levels. *New Phytology* 182:878-890.
- Novo, M., Gayoso, G.M., Pomar, F., Lucas, M.M., Ros Barcelo, A. and Merino, F. (2007). Sulphur accumulation after *Verticillium dahlia* infection of two pepper cultivars differing in degree of resistance. *Plant Pathology* 56:998-104.
- Nusaibah, S.A., Siti Nor Akmar, A., Mohamad Pauzi, Z. and Idris, A.S. (2011). Detection of phytosterols in *Ganoderma boninense*-infected oil palm seedlings through GC-MS analysis. *Journal of Oil Palm Research* 23:1069-1077.

- Nur Ain Izzati, M.Z. and Abdullah, F. (2008). Disease suppression in *Ganoderma*-infected oil palm seedlings treated with *Trichoderma harzianum*. *Plant Protection Science* 44:101-107
- Nyilasi, I., Acs, K., Papp, T. and Vagvolgyi, C. 2005. *Agrobacterium tumefaciens*-mediated transformation of *Mucorcircinelloides*. *Foliar Microbiology* 50:415-420.
- Oleksandr Skyba, A., Carl, J., Douglas, B., Shawn, D. and Mansfielda, P. (2013). Syringyl-rich lignin renders Poplars more resistant to degradation by wood decay fungi. *Applied and Environmental Microbiology* 3:2560-2571.
- Ommelna, B.G., Jennifer, A.N. and Chong, K.P. (2012). The potential of chitosan in suppressing *Ganoderma boninense* infection in oil-palm seedlings. *Journal of Sustainable Science Management* 7(2):186-192.
- Ong, S.S. and Wickneswari, R. (2011). Expression profile of small RNAs in *Acacia mangium* secondary xylem tissue with contrasting lignin content-potential regulatory sequences in monolignol biosynthetic pathway. *BMC Genomics* 12(Suppl 3): S13.
- Pantzaris, T.P. (2000). Pocketbook of palm oil uses, 5<sup>th</sup> Edn, MPOB, Bangi.
- Pantzaris, T.P and Ahmad, M.J. (2001). Properties and utilization of palm kernel oil. *Palm Oil Derivatives* 35:11-23.
- Pardo, A.G., Hanif, M., Raudaskoski, M. and Gorfer, M. (2002). Genetic transformation of ectomycorrhizal fungi mediated by *Agrobacterium tumefaciens*. *Mycological Research* 106(2):132-137.
- Park, G., Xue, C., Zhao, X., Kim, Y., Orbach, M. and Xu, J.R. (2006). Multiple upstream signals converge on the adaptor protein Mst50 in *Magnaporthe grisea*. *Plant Cell* 18:2822-2835.
- Paterson, R.R.M. (2007). *Ganoderma* disease of oil palm-A white rot perspective necessary for integrated control. *Crop Protection* 26:1369-1376.
- Paterson, R.R.M., Holderness, M., Kelley, J., Miller, and O'Grady, E. (2000). In vitro degradation of oil palm stem using macroscopic fungi from South East Asia: A preliminary investigation. In: Flood, J., Bridge, P.D., Holderness, M. (Eds.), *Ganoderma Diseases of Perennial Crops*. CABI Publishing, Wallingford, UK, pp. 129-138.
- Paterson, R.R.M. (2006). *Ganoderma*-A therapeutic fungal biofactory. *Phytochemistry* 67:1985-2001.
- Pegg, G.F. (1985). Life in a black hole: The micro-environment of the vascular pathogen. *British Mycological Society* 85:1-20.

- Peng, M., Lemke, P.A. and Shaw, J.J. (1993). Improved conditions for protoplast formation and transformation of *Pleurotus ostreatus*. *Applied Microbiology and Biotechnology* 40:101.
- Peberdy, J.F. (1979). Fungal protoplasts: isolation, reversion and fusion. *Annual Review of Microbiology* 33:21-39.
- Pieterse, C. M J., van Pelt, J.A., Verhagen, B.W.M., Ton, J., van Wees, S.C.M., Leon-Kloosterziel, K.M. and van Loon, L.C. (2003). Induced systemic resistance by plant growth promoting rhizobacteria. *Symbiosis* 35:39-54.
- Pilate, G., Guiney, E., Holt, K., Petit-Conil, M., Lapierre, C., Leple, J.C., Pollet, B., Mila, I., Webster, E.A. et al. (2002). Field and pulping performances of transgenic trees with altered lignifications. *Nature Biotechnology* 20:607-612.
- Pilotti, C.A. (2005). Stem rots of oil palm caused by *Ganoderma boninense*: Pathogen biology and epidemiology. *Mycopathologia* 159:129-137.
- Podile, A.R., Laxmi, V.D.V. (1998). Seed bacterization with *Bacillus subtilis* AF 1 increase phenylalanine ammonia lyase and reduces the incidence of Fusarial wilt in pigeonpea. *Journal of Phytopathology* 146:255-259.
- Prabavathy, V.R., Mathivanan, N., Sagadevan, E., Murugesan, K. and Lalithakumari, D. (2006). Self fusion of protoplast enhances chitinase production and biocontrol activity in *Trichoderma harzianum*. *Bioresource Technology* 97:2330-2334.
- Rahamah-Bivi, M., Siti Noor Farhanaa, M.D., Khairulmazmi, A., Idris, A.S., Susilawati, K. and Sariah, M. (2014). Assessment of plant secondary metabolites in oil palm seedlings after being treated with calcium, copper ions and salicylic acid. *Archives of Phytopathology and Plant Protection*. Vol. 47, No. 9:1120-1135.
- Ralph, J. (2010). Hydroxycinnamates in lignification. *Phytochemistry Reviews*, 9(1), 65-83.
- Rajasekaran, K., Cary, J.W., Cotty, P.J. and Cleveland, T.E. (2008). Development of a GFP-Expressing *Aspergillus flavus* Strain to study fungal invasion, colonization, and resistance in cottonseed. *Mycopathologia* 165:89-97.
- Rees, R.W., Flood, J., Hassan, Y., Potterd, U. and Cooper, R.M. (2009). Basal stem rot of oil palm (*Elaeis guineensis*): mode of infection and lower stem invasion by *Ganoderma boninense*. *Plant Pathology* 58:982-989. doi. 10.1111/j. 1365-3059.2009.02100x.
- Ride, J.P. (1975). Lignification in wounded wheat leaves in response to fungi and its possible role in resistance. *Physiological Plant Pathology* 5:125.

- Ride, J.P. (1983). Cell walls and other structural barriers in defense. Pages 215-236 In: Biochemical Plant Pathology. J.A. Callow, ed. Wiley Publishers, Chichester, U.K.
- Rivera, A.L., Magana-Ortiz, M. Gomez-Lim, F. Fernandez, L. and Loske, A.M. (2014). Physical methods for genetic transformation of fungi and yeast. *Physical life Review* 11(2):184-203.
- Robersten, B. and Svalheim, O. (1990). The nature of lignin-like compounds in cucumber hypocotyls induced by a-1,4-linked oligogalacturonides. *Physiologia Plantarum* 79:512.
- Robinson, H.L. and Deacon, H. (2001). Protoplast preparation and transient transformation of *Rhizoctonia solani*. *Mycological Research* 105 (11):1295-1303.
- Rogers, L. and Campbell, M.M. (2004). The genetic control of lignin deposition during plant growth and development. *New Phytology* 164:17-30.
- Roslan, A and Idris, A.S. (2011). Economic impact of Ganoderma incidence on Malaysian oil palm plantation-a case study in Johor. *Oil Palm Industry Economic Journal* Vol. 12 No. 1:24-30.
- Sambanthamurthi R., Sundram, K. and Tan, Y.A. (2000). Chemistry and biochemistry of palm oil. *Progress in Lipid Research* 39:507-558.
- San José, C., Monge, R. A., Pérez-Díaz, R., Pla, J. and Nombela, C. (1996). The mitogen-activated protein kinase homolog HOG1 gene controls glycerol accumulation in the pathogenic fungus *Candida albicans*. *Journal of Bacteriology* 178(19):5850-5852.
- Sarkanen, K.V. and Ludwig, C.H. (1971). Lignins, occurrence, formation, structure and reactions. Wiley-Interscience, New York.
- Sariah, M. and Zakaria, H. (2000). The use of soil amendments for the control of basal stem rot of oil palm seedlings. *Ganoderma disease of Perennial Crops* (Flood, J; Bridge, P.D. & Holderness, M.). CABI Publishing, UK. P. 89-100.
- Schuster, A., Bruno, K.S., Collett, J.R., Baker, S.E., Seiboth, B., Kubicek, C.P. and Schmoll, M. (2012). A versatile toolkit for high throughput functional genomics with *Trichoderma reesei*. *Biotechnology and Biofuels* 5:1754-6834.
- Schwarzott, D. and Schubler, A. (2001). A simple and reliable method for SSU rRNA gene DNA extraction, amplification and cloning from single AM fungal spores. *Mycorrhiza* 10:203-207.
- Seo, G.S. and Kirk, P.M. (2000). Ganodermataceae: Nomenclature and Classification. In CAB International. *Ganoderma Diseases of Perennial Crops* (eds Flood, J., Bridge, P.D. and Holderness, M.).

- Shamala, S. and Idris, A.S. (2009). *Trichoderma* as biocontrol agent against *Ganoderma* in oil palm. MPOB Information Series No. 463.4 pp.
- Sharma, K.K. and Kuhad, R.C. (2010). Genetic transformation of lignin degrading fungi facilitated by *Agrobacterium tumefaciens*. *BMC Biotechnology* 10:67.
- Shi, L., Fang, X., Li, M.J., Mu, D., Ren, A., Tan, Q. and Zhao, M. (2011). Development of a simple and efficient transformation system for the basidiomycetous medicinal fungus *Ganoderma lucidum*. *World Journal of Microbiology and Biotechnology* doi:10.1007/s11274-011-0818-z.
- Shin, G.C. and Seo, G.S. (1988). Formation of the non-basidiocarpous basidiospore of *Ganoderma lucidum*. *Korean Journal of Mycology* 16:230-234.
- Shukla, A.N. and Uniyal, K. (1989). Antagonistic interactions of *Ganoderma lucidum* (lyss) karst. against some soil microorganisms. *Journal Current Science* 58:265-267.
- Sibout, R., Eudes, A., Pollet, B., Goujon, T., Mila, I., Granier, F., Seguin, A., Lapiere, C. and Jouanin, L. (2003). Expression pattern of two paralogs encoding cinnamyl alcohol dehydrogenases in *Arabidopsis*. Isolation and characterization of the corresponding mutants. *Plant Physiology* 132:848-860.
- Sibout, R., Eudes, A., Mouille, G., Pollet, B., Lapiere, C., Jouanin, L. and Seguin A. (2005). Cinnamyl alcohol dehydrogenase-C and -D are the primary genes involved in lignin biosynthesis in the floral stem of *Arabidopsis*. *Plant Cell* 17:2059-2076.
- Singh, G. (1991). *Ganoderma*- the scourge of oil palm in the coastal areas. *The Planter* 67(786):421-444.
- Singh, R. (2014). An Integrative Omics Approach to Plant Research. Proceedings in International Conference on Advances in Plant Biotechnology and Biotechnology. Universiti Putra Malaysia.
- Singh, R., Ong-Abdullah, M., Low, E.T., Manaf, M.A., Rosli, R., Nookiah, R., Ooi, L.C., Ooi, S.E., Chan, K.L. and Halim, M.A. (2013). Oil palm genome sequence reveals divergence of infertile species in old and new worlds. *Nature* 500:335-339.
- Sitbon, F., Hennion, S., Little, C.H.A. and Sundberg, B. (1999). Enhanced ethylene production and peroxidase activity in IAA-overproducing transgenic tobacco plants is associated with increased lignin content and altered lignin composition. *Plant Science* 141:165-173.
- Sjostrom, E. (1993). Wood Chemistry. Fundamentals and Applications. Academic Press, San Diego, California, pp. 80-86.



- Skyba, O., Douglas, C.J. and Mansfield, S.D. (2013). Syringyl-rich lignin renders poplars more resistant to degradation by wood decay fungi. *Applied Environmental Biology* 79(8): 2560.
- Solis, S., Flores, M.E. and Huitron, C. (1996). Protoplasts from pectinolytic fungi: Isolation, regeneration and pectinolytic enzyme production. *Letters in Applied Microbiology* 23:36-42.
- Souza A.G.C., Herrero, S., Mafia, L.A. and Daub, E. (2014). Methods for *Cercospora coffeicola* protoplast isolation and genetic transformation with green fluorescent protein. *European Journal of Plant Pathology* 139:241-244.
- Soepena, H., Purba, R.Y. and Pawirosukarto, L. (2000). A control strategy for basal stem rot (Ganoderma) on Oil Palm. CABI International 2000. *Ganoderma Diseases of Perennial Crops* (eds J. Flood, Bridge, P.D. & Holderness).
- Srivastava, S., Vishwakarma, R.K., Arafat, Y.A., Gupta, S.K. and Khan, B.M. (2015). Abiotic stress induces change in Cinnamoyl CoA Reductase (CCR) protein abundance and lignin deposition in developing seedlings of *Leucaena leucocephala*. *Physiology and Molecular Biology of Plants* 21(2):197-205.
- Stewart, J.J., Akiyama, T., Chapple, C., Ralph, J. and Mansfield, S.D. (2009). The effects on lignin structure of overexpression of ferulate 5-hydroxylase in hybrid poplar. *Plant Physiology* 150:621-635.
- Syafii, W. and Yoshimoto, T. (1991). Effect of lignin structure on decay resistance of some tropical woods. *Indonesian Journal of Tropical Agriculture* 3:32-37.
- Stout, A.T., Davis, A.A., Domec, J.C., Yang, C., Shi, R. and King, J.S. (2014). Growth under field conditions affects lignin content and productivity in transgenic *Populus trichocarpa* with altered lignin biosynthesis. *Biomass and Bioenergy* 68:228-239.
- Sumathi, S., Chai, S. P. and Mohamed A.R. (2008). Utilization of oil palm as a source of renewable energy in Malaysia. *Renewable and Sustainable Energy Reviews* 12:2404-2421.
- Sundram, S., Meon, S., Seman, I.A. and Othman, R. (2014). Application of arbuscular mycorrhizal aeruginosa UPMP3 reduces the development of Ganoderma basal fungi with *Pseudomonas* stem rot disease in oil palm seedlings. *Mycorrhiza*. doi 10.1007/s00572-014-0620-5
- Susanto, A., Sudharto, P.S. and Purba, R.Y. (2005). Enhancing biological control of basal stem rot disease (*Ganoderma boninense*) in oil palm plantations. *Mycopathologia* 159:153-157.
- Tan, Y.C., Yeoh, K.A., Wong, M.Y. and Ho, C.L. (2013). Expression profiles of putative defence-related proteins in oil palm (*Elaeis guineensis*) colonised by *Ganoderma boninense*. *Journal of Plant Physiology* 170:1455-1460.

- Tarmizi, A., Yunus, R.M. and Zaitun, R. (2002). Oil palm liquid tissue culture protocol, MPOB series.
- Tee, S.S., Tan, Y.C., Faridah, A., Meilina, O.A. and Ho, C.L. (2011). Transcriptome of oil palm (*Elaeis guineensis* Jacq.) roots treated with *Ganoderma boninense*. *Tree Genetics and Genomes*. doi 10.1007/s11295-012-0559-7.
- ten Have R. and Teunissen, P.J.M. (2001). Oxidative mechanisms involved in lignin degradation by white-rot fungi. *Chemistry Review* 101:3397-3413.
- Terashima, N. and Fukushima, K. (1989). In Plant Cell Polymers. Biosynthesis and Biodegradation (Lewis, N. G. and Paice, M. G., eds), p. 160. Am. Chem. Soc., Washington, DC.
- Tiburzy, R. and Reisener, H.J. (1990). Resistance of wheat to *Puccinia graminis* f.sp.tritici: association of the hypersensitive reaction with the cellular accumulation of lignin-like material and callose. *Physiology and Molecular Plant Pathology* 36:109-120.
- Thilagavathi, R., Saravanakumar, D., Ragupathi, N. and Samiyappan, R. (2007). A combination of biocontrol agents improves the management of dry root rot (*Macrophomina phaseolina*) in greengram. *Phytopathology Mediterranean* 46:157-167.
- Thompson, A. (1931). Stem rot of the oil palm in Malaya. Bull. Depart Agric. Sci Ser.,23.
- Tomlinson, P.B., Horn, J.W. and Fisher, J.B. (2011). The Anatomy of Palms, Oxford University Press, New York, USA.
- Tomlinson, P.B. (1966). Anatomy of the monocotyledons. II. Palmae. Oxford University Press, London, UK.
- Tronchet, M., Balague, C., Kroj, T., Jouanin, L. and Roby, D. (2010). Cinnamyl alcohol dehydrogenase-C and D, key enzymes in lignin biosynthesis, play an essential role in disease resistance in Arabidopsis. *Molecular Plant Pathology* 11:83-92.
- Turner, P.D. (1981). Oil palm Diseases and Disorders. Oxford University Press. Kuala Lumpur. pp. 280.
- Turner, P.D. and Gillbanks, R.A. (2003). Oil palm cultivation and management. The Incorporated Society of Planters, Kuala Lumpur, p 633.
- Unnithan, R. (2015). Oil palm wrongly targeted. Retrived 16 May 2015 from www.thestar.com/my/oilpalm-wrongly-targeted.

- Utomo, C., Werner, S., Niepold, F. and Deising, H.B. (2005). Identification of Ganoderma, the causal agent of basal stem rot disease in oil palm using a molecular method. *Mycopathologia* 159:159-170.
- Urban, M., King, R., Hassani-Pak, K. and Hammond-Kosack, K.E. (2015). Whole-genome analysis of *Fusarium graminearum* insertional mutants identifies virulence associated genes and unmasks untagged chromosomal deletions. *BMC Genomics*.doi10.1186/s12864-015-1412-9.
- Valette-Collet, O., Cimerman, A., Reignault, P., Levis, C. and Boccara, M. (2003). Disruption of *Botrytis cinerea* pectin methylesterase gene *Bcpme1* reduces virulence on several host plants. *Molecular Plant-Microbe Interactions* 16:360-367.
- van den, M.A. and Maruthachalam, B.K. (YEAR). Genetic Transformation Systems in Fungi, Volume 1 Fungal Biology. doi.10.1007/978-3-319-10142-2\_15.
- Vanholme, R., Morreel, Kris., Raph, J. and Boerjan, W. (2008). Lignin Engineering. *Current Opinion in Plant Biology* 11:278-285.
- Voelker, S., Lachenbruch, B., Meinzer, F., Jourdes, M., Ki, C. and Patten, A. (2010). Antisense down-regulation of 4CL expression alters lignifications, tree growth, and saccharification potential of field-grown poplar. *Plant Physiology* 154:874-886.
- Wadhwa, N., Joshi, U.N. and Gandhi, S.K. (2013). Copper and manganese increase resistance of clusterbean to root rot caused by *Rhizoctonia*. *Journal of Phytopathology* 161:172-179.
- Wakefield, E.M. (1920). Diseases of the oil palm in West Africa. *Kew Bulletin*, 306-308.
- Wang, Y., DiGuistini, S., Wang, T.T., Bohlmann, J. and Breuil, C. (2010). *Agrobacterium*-mediated gene disruption using split-marker in *Grosmannia clavigera*, a mountain pine beetle associated pathogen. *Current Genetics* 56:297-307.doi. 10.1007/s00294-010-0294-2.
- Wang, J., Guo, L.Q., Zhang, K. and Lin, J.F. (2008). Highly efficient *Agrobacterium*-mediated transformation of *Volvariella volvacea*. *BioresourceTechnology* 99:8524-8527.
- Wang, M., Liu, S., Li, Y., Xu, R., Lu, C., and Shen, Y. (2010). Protoplast mutation and genome shuffling induce the endophytic fungus *tubercularia* sp. TF5 to produce new compounds. *Current Microbiology* 61:254-260.
- Wong, L.C., Bong, C.F.J. and Idris, A.S. (2012). Ganoderma Species Associated with Basal Stem Rot Disease of Oil Palm. *American Journal of Applied Sciences* 9(6):879-885.

- Xiao, X., Xie, J., Cheng, J., Li, G., Yi, X., Jiang, D. and Fu, F. (2014). Novel secretory protein Ss-Caf1 of the plant-pathogenic fungus *Sclerotinia sclerotiorum* required for host penetration and normal sclerotial development. *Molecular Plant-Microbe Interactions* 27: 40-55.
- Xie, X.M., Zhang, X.Q., Dong, Z.X. and Guo, H.R. (2011). Dynamic changes of lignin contents of MT-1 elephant grass and its closely related cultivars. *Biomass and Bioenergy* 35:1732-1738.
- Xu, W.H., Hu, Y.L., Gao, Y. and Lin, Z.P. (2001). Efficient transformation of the medicinal mushroom *Ganoderma lucidum*. *Plant Molecular Biology Reporter* 19:383a-383j.
- Yan, P.S., Jiang, J.H. and Cui, W.S. (2004). Characterization of protoplast prepared from the edible fungus, *Stropharia rugosa-annulata*. *World Journal of Microbiology and Biotechnology* 20:173-177.
- Yang, C., Xu, Z., Song, J., Conner, K., Barrena, G.V. and Wilson, Z.A. (2007). Arabidopsis MYB26/MALE STERILE35 regulates secondary thickening in the endothecium and is essential for anther dehiscence. *Plant Cell* 19:534-548.
- Yang, M., Yang, Q., Sun, K., Tian, Y. and Li, H. (2011). *Agrobacterium tumefaciens* Mediated Transformation of ChiV Gene to *Trichoderma harzianum*. *Journal of Applied Biochemistry and Biotechnology* 163:937-945 doi 10.1007/s12010-010-9097-7.
- Zaiton, S., Sariah, M. and Zainal, A.M.A. (2008). Effect of endophytic bacteria on growth and suppression of *Ganoderma* infection in oil palm. *International Journal of Agriculture and Biology* 10:127-32.
- Zasloff, M. (2002). Antimicrobial peptides of multicellular organisms. *Nature* 415(6870):389-39.
- Zaprometov, M.N., Zagorskina, V. and Elkin, V.V. (1993). Comparative study of lignins produced by the tea-plant and by tea-plant derived callus tissues. *Phytochemistry* 3:709-711.
- Zhang, K., Qian, Q., Huang, Z., Wang, Y., Li, M., Hong, L., Zeng, D., Gu, M., Chu, C. and Cheng, Z. (2006). GOLD HULL AND INTERNODE2 encodes a primarily multifunctional cinnamyl alcohol dehydrogenase in rice. *Plant Physiology* 140:972-983.
- Zhang, Z., Qin, G., Li, B., Tian, S. (2014). Knocking out *Bcsas1* in *Botrytis cinerea* impacts growth, development, and secretion of extracellular proteins, which decreases virulence. *Molecular Plant-Microbe Interactions* 27(6):590-600.
- Zhao, J.D. (1989). The Ganodermataceae in China. *Bibliotheca mycologica* 132. J. Cramer, Berlin, Stuttgart.

Zhao, J.D. and Zhang, X.Q. (1994). Importance, distribution and taxonomy of Ganodermataceae in China. In: Buchanan, P.K., hseu, R.S. and moncalvo, J.M. (eds) *Ganoderma-Systematics, Phytopathology and Pharmacology*. Proceedings of Contributed Symposia 59A, B, Fifth International mycological Congress, Vancouver, August 14-21, 1994.pp.1-2.

Zhou, Y.P., Chen, M.H., Lu, J.J., Kang, X., Chen, Q.H., Huang, X.L. and Tian, C.E. (2015). A simple and efficient genetic transformation method of *Ganoderma weberianum*. *Folia Microbiologica*.doi. 10. 1007/s12223-015-1377.

Zhu, J., Oger, P.M., Schrammeijer, B., Hooykaas, P., Farrand, S.K. and Winnans, S. (2000). The bases of crown gall tumorigenesis. *Journal of bacteriology* 182:3885-3895.

