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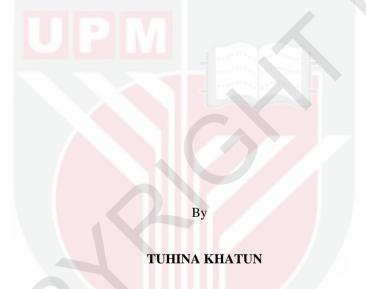
MORPHO-PHYSIOLOGICAL AND MOLECULAR CHARACTERIZATION OF UPLAND RICE GENOTYPES FOR BLAST DISEASE RESISTANCE

TUHINA KHATUN

ITA 2016 1



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Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of Philosophy

August 2016

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DEDICATION

In the name of Allah STW, the most gracious, the most merciful

To the departed soul of my beloved father, who always encouraged and wished for my higher education

To my loving mother, who makes sacrifices and does a lot to make me happy

To my younger brothers Tarek and Muaz, and my younger sister Tania who always make me delighted and cheerful



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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Doctor of Philosophy

MORPHO-PHYSIOLOGICAL AND MOLECULAR CHARACTERIZATION OF UPLAND RICE GENOTYPES FOR BLAST DISEASE RESISTANCE

By

TUHINA KHATUN

August 2016

Chairman: Professor Mohamed Hanafi Musa, PhDInstitute: Tropical Agriculture

Upland rice is a valuable base population for protecting against genetic erosion and broadening the gene pool of rice germplasm. Blast caused by the fungus Magnaporthe oryzae is the most impactful rice disease, and it can be severe in upland/dry land cultures. This study aimed to identify and characterize upland rice germplasm for blast disease resistance. Initially, 50 upland rice genotypes, including one resistant check, Pongsu Seribu-1, and one susceptible check, MR219, were evaluated to identify new sources of resistance and assess diversity based on reactions to M. oryzae. Resistant reactions were observed in the genotypes Biaw Bood Pae, Blau Noc, Chirikata 2, IPPA, IR 5533-50-1-10, IR 5533-55-1-11, Ja Hau, Ja No Nag, BR26, BRRI dhan42 and BRRI The 50 genotypes were grouped into five clusters based on the greater dhan43. similarities of their reactions to the blast fungus. The morphological, physiological and yield trait characteristics of 27 globally diverse blast-resistant genotypes were further evaluated. Additionally, the genotypes were analyzed to determine the correlations between and diversity among these characteristics. The BRRI dhan43, C, Choke Tang and Chirikata 2 were identified as early-maturing genotypes that provided the best yields. The largest heritability coupled with a high genetic advancement was recorded for a number of filled grains/panicle and yield/plant, demonstrating that these traits can be successfully transferred to offspring if selection of these characteristics is performed via a hybridization program. The 27 genotypes were grouped into six clusters based on the contributing morphological, physiological and yield characteristics. The blastresistant, early-maturing, high-yield genotype BRRI dhan43 was then finally characterized to determine the molecular mechanism underlying the plant-microbe interaction using next generation sequencing (NGS). The reference-based alignment produced total reads of 66.9 and 66.2 million in the uninoculated and inoculated plant samples, respectively. This study showed a large number of SNPs with the value of more than 95 and 88 thousand for healthy and diseased samples, respectively. A total of 2,733 differentially expressed genes (DEGs) were identified from Oryza sativa cv BRRI dhan43 and M. oryzae race P7.2 interactions, of which 43% up-regulated and 57% down-regulated, About 9.9% of genes involved in incompatible interaction between BRRI dhan43 and fungus race P7.2 pathosystem. The resistant cultivar, BRRI dhan43

responded upon colonization of virulent fungus P7.2, inducing both pathogen-associated molecular pattern (PAMP)-triggered immunity (PTI) and effector-triggered immunity (ETI). Although the functions yet to be known, based on the results of transcriptome analysis, two novel race-specific elicitors AVR4 and AVR9 in virulent race P7.2, and three resistant proteins MLA10, L6 and RPS in rice cultivar BRRI dhan43 were reported for the first time during host-pathogen interaction.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

PENCIRIAN MORFO-FISIOLOGI DAN MOLEKULAR GENOTIP PADI HUMA UNTUK RESISTEN PENYAKIT KARAH

Oleh

TUHINA KHATUN

Ogos 2016

Pengerusi: Profesor Mohamed Hanafi Musa, PhDInstitut: Pertanian Tropika

Padi huma ialah satu populasi asas berharga untuk dipelihara daripada kepupusan genetik dan memperluaskan kumpulan gen germplasms padi. Penyakit karah disebabkan oleh kulat Magnaporthe oryzae adalah penyakit padi yang paling ketara, dan ia boleh menjadi teruk dalam keadaan tanah kering/tanah tinggi. Kajian ini bertujuan untuk mengenal pasti dan mencirikan germplasms padi huma bagi resisten kepada penyakit karah. Pada mulanya, 50 genotip padi huma, termasuk satu cek resisten, 'Pongsu Seribu-1' dan satu cek mudah peka, 'MR219', telah dinilai untuk mengenal pasti sumber-sumber baharu yang resisten dan menilai kepelbagaian berdasarkan reaksi kepada M. oryzae. Reaksi resisten diperhatikan dalam genotip 'Biaw Bood Pae', 'Blau Noc', 'Chirikata 2', 'IPPA', 'IR 5533-50-1-10', 'IR 5533-55-1- 11', 'Ja Hau', 'Ja No Na', 'BR 26', 'BRRI dhan42' dan 'BRRI Dhan43'. Lima puluh genotip telah dikumpulkan ke dalam lima kelompok berdasarkan persamaan daripada tindak balas mereka kepada kulat karah. Morfologi, ciri-ciri sifat fisiologi dan hasil 27 genotip rentan karah pelbagai peringkat global telah dinilai. Selain itu, genotip dianalisis untuk menentukan hubungan dan kepelbagaian di kalangan ciri-ciri ini. 'BRRI dhan43', 'C', 'Choke Tang' dan 'Chirikata 2' telah dikenal pasti sebagai genotip awal-matang yang memberikan hasil yang terbaik. Kombinasi warisan dan kemajuan genetik yang tinggi dicatatkan bagi ciri-ciri bijirin berisi/panikel dan hasil/pokok, menunjukkan bahawa sifat-sifat ini boleh berjaya dipindahkan kepada anak-anak jika pemilihan ciri-ciri ini dilakukan melalui program penghibridan. Sebanyak 27 genotip telah dikumpulkan kepada enam kelompok berdasarkan ciri-ciri morfologi, fisiologi dan hasil. Rentan karah, awal-matang, penghasilan tinggi genotip 'BRRI dhan43' kemudiannya dicirikan untuk menentukan mekanisme molekul yang menentukan interaksi tumbuhan mikrob dengan menggunakan penjujukan generasi akan datang (NGS). Hasil jajaran mencatatkan jumlah bacaan sebanyak masing-masing 66.9 dan 66.2 juta untuk kawalan dan rawatan tumbuh-tumbuhan. Kajian ini menunjukkan bilangan SNPs yang besar, dengan nilai masng-masing 95 dan 88 ribu bagi kawalan dan sampel dirawat. Sebanyak 2,733 gen terzahir (DEGs) telah dikenal pasti daripada interaksi 'BRRI dhan43'patogen, yang mana 43% gen kawal selia huluan dan 57% gen ke bawah. Sebanyak 9.9% gen terlibat dalam interaksi yang tidak serasi antara BRRI dhan43 dan bangsa kulat P7.2 pathosystem. Hasil penjajaran getir interaksi kulat P7.2 dengan kultivar resisten, BRRI dhan43 memberikan kedua-dua corak molekul patogen-berkaitan (PAMP) kekebalan -triggered (PTI) dan imuniti effector-dicetuskan (ETI). Walaupun fungsi belum diketahui, berdasarkan keputusan analisis transcriptome, dua elisitors novel perlumbaan khusus *AVR4* dan *AVR9* dalam getir perlumbaan P7.2, dan tiga proteins tahan MLA10, L6 dan RPS dalam kultivar padi BRRI dhan43 dilaporkan buat kali pertama semasa interaksi host-pathogen.



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Mst. Tuhina Khatun UPM Serdang, Malaysia August 2016



I certify that a Thesis Examination Committee has met on 10 August 2016 to conduct the final examination of Tuhina Khatun on her thesis entitled "Morpho-Physiological and Molecular Characterization of Upland Rice Genotypes for Blast Disease Resistance" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
AS	Alternative splicing
BGI	Beijing genomics institute
BLAST	Basic local alignment search tool
BLT	Blast lesion type
bp	Base pair
BRRI	Bangladesh Rice Research Institute
CALM	Calmodulin
CDPK/CPK	Calcium-dependent protein kinase
cDNA	Complementary DNA
CC67	Chlorophyll content at 67 days after transplanting
CC97	Chlorophyll content at 97 days after transplanting
CPC	Coding potential calculator
DAI	Days after inoculation
DAT	Days after transplantation
DEGs	Differentially expressed genes
DF	Days to flowering
Df	Degrees of freedom
DGE	Digital gene expression profiling
DLA	Diseased leaf area
DM	Days to maturity
DNA	Deoxyribonucleic acid
DOA	Department of Agriculture
ds	Double stranded

	EM	Expectation-maximization
	ESTs	Expressed sequence tags
	ET	Number of effective tillers/plant
	ETI	Effector-triggered immunity
	EtOH	Ethyl alcohol
	FG	Number of filled grains/panicle
	FL	Flag leaf length
	GA	Genetic advance
	GATK	Genome analysis toolkit
	GB	Grain breadth
	GCV	Genotypic coefficient of variance
	GL	Grain length
	GO	Gene ontology
	h ² _B	Heritability in broad sense
	HCI	Hydrochloric acid
	н	Harvest index
	HR	Hypersensitive response
	HYV	High yielding variety
	Indel	Small insertions and deletions
	IRGSP	International rice genome sequencing project
	IRRI	International Rice Research Institute
	JA	Jasmonic acid
	LAI	Leaf area index
	L/B	Grain length/breadth ratio
	LSD	Least significant difference

	KADA	Kemubu Agricultural and Development Authority
	Kb	Kilobase
	KEGG	Kyoto encyclopedia of genes and genomes
	МАРК	Mitogen-activated protein kinase
	MARDI	Malaysian Agricultural Research and Development Institute
	Mb	Megabase
	MR	Moderately resistant
	MS	Moderately susceptible
	mRNA	Messenger RNA
	MSE	Mean square of error
	MSG	Mean square of accessions
	NCBI	National center for biotechnology information
	ng	Nanogram
	NGS	Next generation sequencing
	РАМР	Pathogen-associated molecular pattern
	PCA	Principal component analysis
	PCR	Polymerase chain reaction
	PCV	Phenotypic coefficient of variance
	PDA	Potato dextrose agar
	PE	Paired-end
	РН	Plant height
	PL	Panicle length
	PR	Pathogenesis related
	PRRs	Pathogen-recognition receptors
	PS-1	Pongsu seribu-1

PTI	PAMP-triggered immunity
QC	Quality control
RBOH	Respiratory burst oxidase
RCBD	Randomized complete block design
RNA	Ribonucleic acid
ROS	Reactive oxygen species
RSEM	RNA seq by expectation maximization
SA-PMPs	Streptavidin magnesphere® paramagnetic particles
SES	Standard evaluation system
SNARE	Soluble N-ethylmaleimide-sensitive factor attachment protein receptor
SNP	Single nucleotide polymorphism
TMRI	Torrey Mesa Research Institute
TT	Total number of tillers/plant
UFG	Number of unfilled grains/panicle
UPGMA	Unpaired group method of arithmetic mean
UPM	Universiti Putra Malaysia
Y	Yield/plant
C	Degree centigrade
%DLA	Percentage of diseased leaf area
$\sigma^2 e$	Mean squares of error
$\sigma^2 g$	Genotypic variance
$\sigma^2 p$	Phenotypic variance
100 GW	100-grain weight

CHAPTER 1

INTRODUCTION

1.1 Background

Rice (*Oryza sativa* L.) is the most widely consumed grain in the world, playing a vital role in combating global hunger, aiding food security, alleviating poverty and promoting socioeconomic development in most Asian countries (Pranab, 2008). Global rice consumption remains strong, as it is driven by both population and economic growth, especially in Asian and African countries (Ricepedia, 2015). In most of the developing world, rice availability is associated with food security and political stability (Ricepedia, 2015). Rice is the third most significant crop in Malaysia in terms of production, following oil palm and rubber (Karim et al., 2004). Malaysian rice production currently accounts for 72% of domestic consumption, with a goal of reaching 90% over the next 10 years (Isiaka et al., 2015). Rice in Malaysia is generally cultivated as wet paddy (Peninsular Malaysia, 503,184 ha) or upland rice (Sabah and Sarawak, 165,888 ha) per the DOA (2005).

Global agricultural production increased dramatically as a result of the "Green Revolution", which was introduced in Mexico by Norman E. Borlaug between 1950 and 1970 due to technological innovations. These advancements include producing highyielding crop varieties, as well as using chemical fertilizers, synthetic pesticides and herbicides to control weeds, kill insect pests and prevent diseases. The Green Revolution involved a concerted effort to address the lack of food self-sufficiency (David, 1997). However, the Green Revolution exaggerated both agricultural biodiversity and wild biodiversity (Kilusang, 2007), which extensively affected the environment. In addition, the movement involved the breakdown of both pathogen and insect resistant genes, which cannot be controlled by artificial pesticides. Valuable genetic resources were also permanently lost. The utilization of underground water for irrigating HYV's resulted in water shortages and droughts. In these cases, upland rice provides the best alternative for cultivation because it relies on rain-fed conditions without the use of irrigation water or chemical inputs.

The world will be faced with global rice production challenges in the coming years. Upland rice cultivation offers a promising production technique with minimal negative environmental effects and highly predictable yields (Dzido et al., 2004). Upland rice refers to rice cultures that are grown on undulated land areas with well-drained soils. The rice completely depends on rainfall for its water requirements (Fageria et al., 2011). Approximately 100 million people now depend on upland rice as their daily staple food. Upland rice comprises 11% of the total global rice production and is cultivated on approximately 14 million ha (Sohrabi et al., 2012). In Malaysia, 98,000 ha are used for upland rice cultivation, accounting for approximately 12% of the total domestic rice cultivation area (Ferdous, 2012). In underprivileged areas, the utilization of upland rice varieties will undoubtedly help farmers to combat food security (Dzido et al., 2004).

Upland rice is subject to more pressure from bio-aggressors than is rain-fed lowland rice (Sester et al., 2008). These bio-aggressors include weed infestations and diseases specially blast disease, which is caused by the fungus *Magnaporthe oryzae* (Sester et al., 2008). Blast is one of the most devastating rice diseases, posing a significant threat to global food security (Galhano and Talbot, 2011). The blast fungus causes global rice yield losses of 60-100% annually (Kihoro, 2013), leading to severe epidemics in rice growing regions around the world and increasing rice cultivation costs (Galhano and Talbot, 2011). In such a context, new strategies must be created and evaluated to control blast epidemics and ensure sustainable upland-rice production (Sester et al., 2008).

Next generation sequencing (NGS) is the latest biotechnological approach, providing a complete picture of genomes and genes. Advancements in gene sequencing and subsequent DNA sequence analyses have led to the identification of many novel genes and improved the understanding of various qualitative and quantitative genetic traits in crop plants (Ganapathy et al., 2013). These tremendous advances have created new opportunities for identifying the molecular mechanisms behind host-microbe interactions in plant species. The interaction between the blast fungus *M. oryzae* and rice has been the focus of extensive rice disease resistance and fungal infection mechanism studies (Ribot et al., 2008). A better understanding of the mechanisms associated with *M. oryzae* infection and rice plant damage may lead to new invasive disease control methods (Ribot et al., 2008).

The screening or evaluation of upland rice germplasms is essential for determining new sources of resistance and developing durable blast resistant varieties *via* hybridization programs in blast endemic upland rice areas. Further morphological characterization of blast resistance genotypes is crucial for identifying high-yielding cultivars, which are of significant value to upland farmers. The identification of resistant genes from blast resistant upland rice varieties is essential for rice breeding programme. These genes can then be used to develop durable blast resistant upland rice cultivars in future.

1.2 Problem statement

The global upland rice yield is relatively low (Fageria et al., 2013), and rice blast disease represents the major constraint for a higher yield (Hanafi et al., 2009). Few studies have focused on improving upland rice yields and disease management strategies at the national and international scales. Up to now study on molecular dissection of blast resistant high yielding varieties through NGS are scarce in rice data bases. Currently, almost no high-yielding blast resistant varieties exist for cultivation in upland rice areas. Such varieties would help to meet the future food demands of subsistence farmers.

1.3 Objectives

The general objective of this project was to morphologically characterize upland rice germplasm to identify blast resistant genotypes with high yield potentials and to understand the precise molecular infection mechanisms of host-pathogen interaction that can be used to facilitate the design of novel control strategies. The research was initiated with the following specific objectives:

- i) to assess the genetic diversity among upland rice genotypes against virulent race of blast fungus P7.2;
- ii) to characterize the blast resistant upland rice genotypes based on morphophysiological and yield attributes;
- iii) to perform transcriptome analysis of blast resistant cultivar BRRI dhan43 through next generation sequencing;
- iv) to analyze differentially expressed genes (DEG) in incompatible interaction between BRRI dhan43 and blast fungus P7.2.



REFERENCES

- Abarshahr, M., Rabiei, B. and Lahigi, H.S. (2011). Assessing genetic diversity of rice varieties under drought stress conditions. *Notulae Scientia Biologicae*. 3: 114– 123.
- Ahmad, F. (2014). Genetic diversity analysis of coloured upland rice germplasms using SSR markers, quantitative traits, and nutritional quality. MS thesis, Institute of Tropical Agriculture, Universiti Putra Malaysia, Malaysia.
- Ahmadikhah, A., Nasrollanejad, S. and Alishah, O. (2008). Quantitative studies for investigating variation and its effect on heterosis of rice. *International Journal* of Plant Production. 2: 297–308.
- Akinbile, C.O., El-Latif, K.M.A, Abdullah, R. and Yusoff, M.S. (2011). Rice production and water use efficiency for self-sufficiency in Malaysia: a review. *Trends in Applied Sciences Research*. 6(10): 1127-1140.
- Akinwale, M., Gregorio, G., Nwilene, F., Akinyele, B., Ogunbayo, S.A. and Odiyi, A.C. (2011). Heritability and correlation coefficient analysis for yield and its components in rice (*Oryza sativa* L.). African Journal of Plant Science. 5: 207-212.
- Alexandratos, N. and Bruinsma, J. (2012). World agriculture towards 2030/2050: the 2012 revision. Food and Agriculture Organization of the United Nations. ESA working paper. 12: 03.
- Andersen, M.R., Salazar, M.P., Schaap, P.J., Van de Vondervoort, P.J.I., Culley, D. Thykaer, J., Frisvad, J.C., Nielsen, K.F., Albang, R., Albermann, K., Berka, R.M., Braus, G.H., Braus-Stromeyer, S.A., Corrochano, L.M., Dai, Z., Dijck, P.W.M., Hofmann, G., Lasure, L.L., Magnuson, J.K., Menke, H., Meijer, M., Meijer, S.L., Nielsen, J.B., Nielsen, M.L., Ooyen, A.J.J., Pel, H.J., Poulsen, L., Samson, R.A., Stam, H., Tsang, A., Brink, J.M., Atkins, A., Aerts, A., Shapiro, H., Pangilinan, J., Salamov, A., Lou, Y., Lindquist, E., Lucas, S., Grimwood, J.I., Grigoriev, V. Kubicek, C.P., Martinez, D., Peij, N.N.M.E., Roubos, J.A., Nielsen, J. and Baker S.E. (2011). Comparative genomics of citric-acid-producing Aspergillus niger ATCC 1015 versus enzyme-producing CBS 513.88. Genome Research. 21(6): 885–897.

Andrews, S. (2010). Fastqc. http://www.bioinformatics.bbsrc.ac.uk/projects/fastqc/.

- Anwar, M.P., Juraimi, A.S., Man, A., Puteh, A., Selamat, A. and Begum, M. (2010). Weed suppressive ability of rice (*Oryza sativa* L.) germplasm under aerobic soil conditions. *Australian Journal of Crop Science*. 4(9): 706-717.
- Anwar, M.P. (2012). *Strategies for weed suppression in aerobic rice cultivation*. PhD thesis, Institute of Tropical Agriculture, Universiti Putra Malaysia, Malaysia.

- Arras, G., Hallewin, D.G., Molinu, M.G., Dore, A., Venditti, T., Fois, M., Lima, G. and Aqabbio, M. (2006). Induction of phytoalexins biosynthesis in orange fruit by the biocontrol yeast *Rhodotorula glutinis*. *Communications in Agriculture and Applied Biological Science*. 71: 915–921.
- Arshad, H.M.I., Khan, J.A. and Jamil, F.F. (2008). Screening of rice germplasms against blast and brown spot disease. *Pakistan Journal of Pytopathology*. 20(1): 525-527.
- Ashifield, M., Hammond-Kosack, K.E, Harrison, K. and Jones, J.D.G. (1994). Cf gene dependent induction of b1-3 glucanase promoter in tomato plants infected with *Cladosporium fulvum. Molecular Plant Microbe Interaction*. 7: 645–657.
- Ashkani, S. (2011). Molecular dissection and QTL mapping of rice blast disease resistance using simple sequence repeat markers. PhD thesis, Universiti Putra Malaysia, Malaysia.
- Ashkani, S., Yusop, M.R., Shabanimofrad, M., Harun, A.R., Sahebi, M. and Latif, M.A. (2014). Genetic analysis of resistance to rice blast: a study on the inheritance of resistance to the blast disease pathogen in an F₃ population of rice. *Journal of Phytopathology*. doi: 10.1111/jph.12323.
- Atwell, S., Huang, Y.S., Vilhjálmsson, B.J., Willems, G., Horton, M., Li, Y., Meng, D., Platt, A., Tarone, A.M., Hu, T.T., Jiang, R., Muliyati, N.W., Zhang, X., Amer, M.A., Baxter, I., Brachi, B., Chory, J., Dean, C., Debieu, M., Meaux, J.D., Ecker, J.R., Faure, N., Kniskern, J.M., Jones, J.D.G., Michael, T., Nemri, A., Roux, F., Salt, D.E., Tang, C., Todesco, M., Traw, M.B., Weigel, D., Marjoram, P., Borevitz, J.O., Bergelson, J. and Nordborg, M. (2010). Genomewide association study of 107 phenotypes in *Arabidopsis thaliana* inbred lines. *Nature*. 465(7298): 627–631.
- Awodera, V.A. and Esuruoso, O.F. (1975). Reduction in grain yield of two rice varieties infected by rice blast disease in Nigeria. *Nigerian Agricultural Journal*. 11: 170-3.
- Azizi, P., Rafii, M.Y., Abdullah, S.N., Nejat, A.N., Maziah, M., Hanafi, M.M., Latif, M.A. and Sahebi, M. (2014). Toward understanding of rice innate immunity against *Magnaporthe oryzae*. *Critical Review of Biotechnology*. doi: 10.3109/07388551.2014.946883.
- Barbazuk, W.B., Emrich, S.J., Chen, H.D., Li, L. and Schnable, P.S. (2007). SNP discovery via 454 transcriptome sequencing. *Plant Journal*. 51(5): 910–918.
- Batley, J., Barker, G., O'Sullivan, H., Edwards, K.J. and Edwards, D. (2003). Mining for single nucleotide polymorphisms and insertions/ deletions in maize expressed sequence tag data. *Plant Physiology*. 132: 84–91.
- Benjamini, Y. and Yekutieli, D. (2001). The control of the false discovery rate in multiple testing under dependency. *The Annals of Statistics*. 29: 1165-1188.

- Bennetzen, J. (2002). The rice genome-opening the door to comparative plant biology. *Science*. 296: 5.
- Bentley, D.R. (2006). Whole-genome re-sequencing. *Current Opinion in Genetics and Development.* 16: 545–552.
- Bhuiyan, M.A.R., Narimah, M.K., Rahim, H.A., Abdullah, M.Z. and Wickneswari, R. (2011). Transgressive variants for red pericarp grain with high yield potential derived from *Oryza rufipogon* × *Oryza sativa*: field evaluation, screening for blast disease, QTL validation and background marker analysis for agronomic traits. *Field Crops Research*. 121: 232–239.
- Bilgrami, K.S. and Dube, H.C. (1982). A textbook of modern plant pathology. 5th edn. New Delhi, Vikas publisher.
- Bishwajit, G., Sarker, S., Kpoghomou, M.A., Gao, H., Jun, L., Yin, D. and Ghosh, S. (2013). Self-sufficiency in rice and food security: a South Asian perspective. *Agriculture & Food Security*. 2: 10.
- Borell, A., Garside, A. and Shu, F.K. (1997). Improving efficiency of water use for irrigated rice in a semi-arid tropical environment. *Field Crops Research*. 52: 231-248.
- Bradley, D., Kjellbom, P. and Lamb, C.J. (1992). Elicitor- and wound-induced oxidative cross-linking of a proline-rich plant cell wall protein: a novel, rapid defense response. *Cell*. 70: 21–30.
- BRRI (Bangladesh Rice Research Institute). (2011). *Cultivation of modern rice*. 16th edn. Dhaka, MRI publisher, Bangladesh.
- Burton, G.W. (1952). *Quantitative inheritance in grasses*. Proceedings of the International Grassland Congress. 1: 277–283.
- Burton, G.W. and DeVane, E.H. (1953). Estimating heritability in tall fescue (*Festuca arundinacea*) from replicated clonal material. *Agronomy Journal*. 45: 478-481.
- Castano, J.B., Amril, B., Syahril, D. and Zaini, Z. (1990). Upland rice genotypes resistant to blast (B1) disease in west Sumatra. *International Rice Research Newsletter*. 15(4): 11-12.
- Chandra, R., Pradhan, S.K., Singh, S., Bose, L.K. and Singh, O.N. (2007). Multivariate analysis in upland rice genotypes. World Journal of Agricultural Sciences. 3: 295–300.
- Chang, T.T. and Vergara, B.S. (1975). Varietal diversity and morpho-agronomic characteristics of upland rice. In: *Major research in upland rice*. International Rice Research Institute, eds. (pp.72–90). Los Banos, Philippines.

- Chang, T.T., Loresto, G.C. and Tagumpay, O. (1972). *Rice breeding, Agronomic and growth characteristics of upland and lowland rice varieties*. IRRI, Los Baños, Phillippines.
- Chaudhary, B., Shrestha, S.M. and Sharma, R.C. (2005). Resistance in rice breeding lines to the blast fungus in Nepal. *Nepal Agricultural Research Journal*. 6: 49-56.
- Chen, C., Lian, B., Hu, J., Zhai, H., Wang, X., Venu, R.C., Liu, E., Wang, Z., Chen, M., Wang, B., Wang, G.L., Wang, Z. and Mitchell, T.K. (2013). Genome comparison of two *Magnaporthe oryzae* field isolates reveals genome variations and potential virulence effectors. *BMC Genomics*. 14: 887.
- Chen, H. (2001). Population structure of Pyricularia grisea from central and Southern China and comparative mapping of QTL for blast-and bacterial blightresistance in rice and barley. PhD thesis, Huazhong Agriculture University, Wuhan, China.
- Chen, W., Provart, N.J., Glazebrook, J., Katagiri, F., Chang, H.S., Eulgem, T., Mauch, F., Luan, S., Zou, G., Whitham. S.A., Budworth, P.R., Tao, Y., Xie, Z., Chen, X., Lam, S., Kreps, J.A., Harper, J.F., Si-Ammour, A., Mauch-Mani, B., Heinlein, M., Kobayashi, K., Hohn, T., Dang, J.L., Wang, X. and Zhu, T. (2002). Expression profile matrix of Arabidopsis transcription factor genes suggests their putative functions in response to environmental stresses. *Plant Cell*. 14: 559–574.
- Chen, K., Fan, B., Du, L. and Chen, Z. (2004). Activation of hypersensitive cell death by pathogen-induced receptor-like protein kinase from Arabidopsis. *Plant Molecular Biology*. 56: 271–283.
- Cheung, F., Haas, B.J., Goldberg, S.M.D., May, G.D., Xiao, Y. and Town, C.D. (2006). Sequencing *Medicago truncatula* expressed sequenced tags using 454 life sciences technology. *BMC Genomics*. 7: 272–282.
- Ching, A., Caldwell, K.S., Jung, M., Dolan, M., Smith, O.S.H., Tingey, S., Morgante, M. and Rafalski, A.J. (2002). SNP frequency, haplotype structure and linkage disequilibrium in elitemaize inbred lines. *BMC Genetics*. 3: 19.
- Christine, H.F. and Graham, N. (2003). Redox sensing and signalling associated with reactive oxygen in chloroplasts, peroxisomes and mitochondria. *Physiology of Plantarum*. 119: 355–364.
- Close, T.J., Bhat, P.R., Lonardi, S., Wu, Y., Rostoks, N., Ramsay, L., Druka, A., Stein, N., Svensson, J.T., Wanamaker, S., Bozdag, S., Roose, M.L Moscou, M.J., Chao, S., Varshney, R.K., Szűcs, P., Sato, K., Hayes, P.M., Matthews, D.E., Kleinhofs, A., Muehlbauer, G.J., Young, J.D., Marshal, D.F., Madishetty, K., Fenton, R.D., Condamine, P., Graner, A. and Waugh, R. (2009). Development and implementation of high-throughput SNP genotyping in barley. *BMC Genomics*. 10: 582.

- Couch, B.C. and Kohn, L.M. (2002). A multilocus gene genealogy concordant with host preference indicates segregation of a new species, *Magnaporthe oryzae*, from *M. grisea. Mycologia*. 94(4): 683-693.
- Correa-Victoria, F.J. and Zeigler, R.S. (1993). Pathogenic variability in *Pyricularia grisea* at a rice blast "hot spot" breeding site in Eastern Colombia. *Plant Disease*. 7: 1029-1035.
- Datta, S.K.D. (1981). *Principles and practices of rice production*. A wiley-Inter-science publication, USA.
- David, B. (1997). Food production, consumption, and policy. Chicago: Fitzroy Dearborn. *Encyclopedia of Mexico*. 1: 494.
- Dean, R.A., Talbot, N.J., Ebbole, D.J., Farman, M.L., Mitchell, T.K., Orbach, M.J., Thon, M., Kulkarni, R., Xu, J.R., Pan, H., Read, N.D., Lee, Y.H., Carbone, I., Brown, D., Oh, Y. Y., Donofrio, N., Jeong, J. S., Soanes, D.M., Djonovic, S., Kolomiets, E., Rehmeyer, C., Li, W., Harding, M., Kim, S., Lebrun, M.H., Bohnert, H., Coughlan, S., Butler, J., Calvo, S., Ma, L.J., Nicol, R., Purcell, S., Nusbaum, C. and Galagan J.E. (2005). The genome sequence of the rice blast fungus *Magnaporthe grisea*. *Nature*. 434. www.nature.com/nature.
- Deschamps, S., Rota, M-la., Ratashak, J.P., Biddle, P., Thureen, D., Farmer, A., Luck, S., Beatty, M., Nagasawa, N., Michael, L., Llaca, V., Sakai, H., May, G., Lightner, J. and Campbell, M.A. (2010). Rapid genome-wide single nucleotide polymorphism discovery in soybean and rice via deep resequencing of reduced representation libraries with the illumina genome analyzer. *The Plant Genome*. 3: 53–68. doi: 10.3835/plantgenome2009.09.0026.
- Dixit, R., Bhargava, A., Dalal, V., Plaha, P., Singh, N.K. and Sharma, T.R. (2009). Accumulation of defence response-related and unique expressed sequence tags during the incompatible interaction in the Oryza sativa–Magnaporthe oryzae pathosystem. Journal of Phytopathology. doi: 10.1111/j.1439-0434.2008.01520.x.
- DOA (Department of Agriculture). (2005). *Agricultural Statistical Handbook*. Ministry of agriculture and agro-based industry, Malaysia.
- Dong, Y., Li, Y., Zhao, M., Jing, M., Liu, X. and Liu, M., Guo, X., Zhang, X., Chen, Y., Liu, Y., Liu, Y., Ye, W., Zhang, H., Wang, Y., Zheng, X., Wang, P. and Zhang, Z. (2015). Global genome and transcriptome analyses of *Magnaporthe* oryzae epidemic isolate 98-06 uncover novel effectors and pathogenicityrelated genes, revealing gene gain and lose dynamics in genome evolution. *PLoS Pathology*. 11(4): e1004801. doi:10.1371/journal.ppat.1004801.
- Dou, X., Wang, Q., Qi, Z., Song, W., Wang, W., Guo, M., Zhang, H., Zhang, Z., Wang, P. and Zheng, X. (2011). MoVam7, a conserved SNARE involved in vacuole assembly, is required for growth, endocytosis, ROS accumulation, and pathogenesis of *Magnaporthe oryzae*. *PLoS One*. 6: e16439. doi: 10.1371/journal.pone.0016439 PMID: 21283626.

- Duistermaat, H. (1987). A revision of *Oryza* (Gramineae) in Malaysia and Australia. *Blumea*. 32: 177.
- Duran, C., Appleby, N., Clark, T., Wood, D., Imelfort, M., Batley, J. and Edwards, D. (2009a). AutoSNPdb: an annotated single nucleotide polymorphism database for crop plants. *Nucleic Acids Research*. 37: 951–953.
- Duran, C., Appleby, N., Edwards, D. and Batley, J. (2009b). Molecular genetic markers: discovery, applications, data storage and visualisation. *Current Bioinformation*. 4: 16–27.
- Duran, C., Appleby, N., Vardy, M., Imelfort, M., Edwards, D. and Batley, J. (2009). Single nucleotide polymorphism discovery in barley using autoSNPdb. *Plant Biotechnology Journal*. 7: 326–333.
- Duran, C., Eales, D., Marshall, D., Imelfort, M., Stiller, J., Berkman, P.J., Clark, T., McKenzie, M., Appleby, N., Batley, J., Basford, K. and Edwards, D. (2010). Future tools for association mapping in crop plants. *Genome*. 53: 1017–1023.
- Dzido J.L., Valès, M., Rakotoaris, J., Chabanne, A. and Ahmadi, N. (2004). Upland rice for high lands: New varieties and sustainable cropping systems for food security, promising prospects for the global challenges of rice production. FAO rice conference held on 12-13 February, 2004, Rome, Italy. p.11.
- Ebbole, D.J., Jin, Y., Thon, M., Pan, H., Bhattarai, E., Thomas, T. and Dean, R. (2004). Gene discovery and gene expression in the rice blast fungus, *Magnaporthe grisea*: Analysis of expressed sequence tags. *Molecular Plant Microbe Interaction*. 17: 1337–1347.
- Ebbole, D.J. (2007). *Magnaporthe* as a model for understanding host-pathogen interactions. *Annual Review of Phytopathology*. 45: 437–456. PMID: 17489691.
- Egan, M.J., Wang, Z.Y., Jones, M.A., Smirnoff, N. and Talbot, N.J. (2007). Generation of reactive oxygen species by fungal NADPH oxidases is required for rice blast disease. *Proceedings of the National Academy of Sciences of the United States of America*. 104(28): 11772-11777.
- Eisen, M.B., Spellman, P.T., Brown, P.O. and Botstein, D. (2001). Cluster analysis and display of genome-wide expression patterns. *Proceedings of the National Academy of Sciences of the United States of America*. 95(25): 14863-8. 2001.29: 1165-1188.
- Fageria, N.K., Barbosa Filho, M.P. and Carvalho, J.R.P. (1982). Response of upland rice to phosphorus fertilization on an Oxisol. *Agronomy Journal*. 74: 51–56.
- Fageria, N.K., Baligar, V.C. and Jones, C.A. (1997). *Growth and mineral nutrition of field crops*. 2nd edn. New York: Marcel Dekker, Inc.

- Fageria, N.K. (2007). Yield Physiology of Rice. *Journal of Plant Nutrition*. 30(6): 843-879. doi: 10.1080/15226510701374831.
- Fageria, N.K., Baligar, V.C. and Jones, C.A. (2011). Growth and mineral nutrition of field crops. 3rd edn. Boca Raton, Florida, CRC Press.
- Fageria, N.K., Moraes, O.P. and Vasconcelos, M.J. (2013). Upland rice genotypes evaluation for phosphorus use efficiency. *Journal of Plant Nutrition*. 36(12): 1868-880. doi: 10.1080/01904167.2013.818153.
- Fageria, N.K., Carvalho, M.C.S. and dos Santos, F.C. (2014). Response of upland rice genotypes to nitrogen fertilization. *Communications in Soil Science and Plant Analysis*. doi:10.1080/00103624.2014.911301.
- Fahmi, Z., Samah, B.A. and Abdullah, H. (2013). Paddy industry and paddy farmers well-being: a success recipe for agriculture industry in Malaysia. Asian Social Science. 9(3): 177-181.
- Fan, X.D., Wang, J.Q., Yang, N., Dong, Y.Y., Liu, L., Wang, F.W., Wang, N., Chen, H., Liu, W.C., Sun, Y.P., Wu, J.Y. and Li, H.Y. (2013). Gene expression profiling of soybean leaves and roots under salt, saline–alkali and drought stress by high-throughput Illumina sequencing. *Gene.* 512: 392–402.
- FAOSTAT (Food and Agriculture Organization Statistics). (2015). FAO database 2015 for rice production. FAO, Rome. http://faostat3.fao.org/browse/Q/QC/E. Accessed on 10 August 2015.
- Fedoroff, N.V., Battisti, D.S., Beachy, R.N., Cooper, P.J.M., Fischhoff, D.A., Hodges, C.N., Knauf, V.C., Lobell, D., Mazur, B.J., Molden, D., Reynolds, M.P., Ronald, P.C., Rosegrant, M.W., Sanchez, P.A., Vonshak, A. and Zhu J.K. (2010). Radically rethinking agriculture for the 21st century. *Science*. 327: 833-834.
- Ferdous J. (2012). Morphological, nutritional and molecular characterization of selected genotypes of upland improvement through marker assisted backcross breeding. PhD thesis, Institute of Tropical Agriculture, Universiti Putra Malaysia, Malaysia.
- Filichkin, S.A., Priest, H.D., Givan, S.A., Shen, R., Bryant, D.W., Fox, S.E., Wong,
 W.K. and Mockler, T.C. (2010). Genome-wide mapping of alternative splicing in *Arabidopsis thaliana. Genome Research.* 20: 45-58.
- Fukui, K. and Lijima, K. (1991). Somatic chromosome map of rice by imaging methods. *Theoretical & Applied Genetics*. 81: 589-596.
- Galhano, R. and Talbot, N.J. (2011). The biology of blast: Understanding how *Magnaporthe oryzae* invades rice plants. *Fungal Biology Reviews*. 25: 61-67.

- Ganapathy, K.N., Gomaseh, S. and Rakshit, S. (2013). Application of next generation sequencing in crop improvement. *Biotechnology Articles*. http://www.biotecharticles.com/2974.html.
- Getachew, G., Alemu, T. and Tesfaye, K. (2013). Evaluation of disease incidence and severity and yield loss of finger millet varieties and mycelial growth inhibition of *Pyricularia grisea* isolates using biological antagonists and fungicides in vitro condition. *Journal of Applied Bioscience*. 73: 5883–5901.
- Ghazanfar, M.U., Habib, A. and Sahi, S.T. (2009). Screening of rice germplasm against *Pyricularia oryzae* the cause of rice blast disease. *Pakistan Journal of Phytopathology*. 21(1): 41-44.
- Gianessi, L.P. (2014). Importance of pesticides for growing rice in South and South East Asia. International Pesticide Benefit Case Study 108 by CropLife Foundation. Accessed on 12 August 2016.
- Goff, S.A., Ricke, D., Lan, T.H., Presting, G., Wang, R.L., Dunn, M., Glazebrook, J., Sessions, A., Oeller, P., Varma, H., Hadley, D., Hutchinson, D., Martin, C., Katagiri, F., Lange, B.M., Moughamer, T., Xia, Y., Budworth, P., Zhong, J.P., Miguel, T., Paszkowski, U., Zhang, S.P., Colbert, M., Sun, W.L., Chen, L.L., and *et al.* (2002). A draft sequence of the rice genome (*Oryza sativa* L. ssp *japonica*). *Science*. 296: 92-100.
- Gordon, A. (2009). Fastx-toolkit. http://hannonlab.cshl.edu/fastx_toolkit/.
- Goto, K. (1955). History of the blast disease and changes in methods of control. *Agricultural Improvement Bureau*. Ministry of Agriculture and Forestry, Japan. 5: 1-2.
- Govindaraj, M., Vetriventhan, M. and Srinivasan, M. (2015). Importance of genetic diversity assessment in crop plants and its recent advances: an overview of its analytical perspectives. *Genetics Research International*. http://dx.doi.org/10.1155/2015/431487.
- Greer, C.A., Webster, R.K. (2001). Occurrence, distribution, epidemiology, cultivar reaction, and management of rice blast disease in California. *Plant Disease*. 85(10): 1096-1102.
- Grigoryevn and Yevgeniy A. (2011). Post-transcriptional mechanisms of gene regulation and information control in immunity. *The Scripps Research Institute*. http://gradworks.umi.com/34/88/3488985.html.
- Grist, D.H. (1959). *Rice.* 3rd edn. Longmans, Green and Co Ltd. Western printing services Ltd Bristol, Great Britain.
- Guimaraes, E.P. (2009). Rice breeding. In M.J. Carena, Ed. Cereals (pp. 1–28). Springer, New York, USA.

- Habib, S.H., Bashar, M.K., Khalequzzaman, M., Ahmed, M.S. and Rashid, E.S.M.H. (2005). Genetic analysis and morpho-physiological selection criteria for traditional Biroin Bangladesh rice germplasms. *Journal of Bioscience*. 5: 315-318.
- Hanafi, M.M., Hartinie, A., Shukor, J. and Mahmud, T.M.M. (2009). Upland rice varieties in Malaysia: agronomic and soil physico-chemical characteristics. *Pertanika Journal of Tropical Agricultural Sciences*. 32: 225-246.
- Haq, I.M., Fadnan, M., Jamil, F.F. and Rehman, A. (2002). Screening of rice germplasm against *Pyricularia oryzae* and evaluation of various fungitoxicants for control of disease. *Pakistan Journal of Phytopathology*. 14(1): 32-35.
- Hartinee, A. (2006). Increasing the yield of upland rice in idle land through nutrient and weed management. MS thesis. Faculty of Agriculture, Universiti Putra Malaysia, Malaysia.
- Hasegawa, H. (2003). High yielding rice cultivars perform best even at reduced nitrogen fertilizer rate. *Crop Science*. 43: 921–926.
- Hien, N.L., Sarhadi, W.A, Hirata, Y. and Oikawa, Y. (2007). Genetic diversity of morphological responses and the relationships among Asia aromatic rice (*Oryza sativa* L.) cultivars. *Tropics*. 16: 343-355.
- Ho, C.L., Tan, Y.C., Yeoh, K.A., Ghazali, A.K., Yee, W.Y. and Hoh, C.C. (2016). De novo transcriptome analyses of host fungal interactions in oil palm (*Elaeis guineensis* Jacq.). *BMC Genomics*. 17: 66. doi: 10.1186/s12864-016-2368-0.
- Hooker, S. (1984). The flora of British India. L. Reeve and Co. Ashford, Kent.
- Howard, R.J. and Valent, B. (1999). Breaking and entering: host penetration by the fungal rice blast pathogen *Magnaporthe grisea*. Annual Review of Microbiology. 50: 491–512.
- Hu, J., Chen, C., Peever, T., Dang, H., Lawrence, C. and Mitchell, T. (2012). Genomic characterization of the conditionally dispensable chromosome in *Alternaria arborescens* provides evidence for horizontal gene transfer. *BMC Genomics*. 13: 171.
- Hudson, M. (2008). Sequencing breakthroughs for genomic ecology and evolutionary biology. *Molecular Ecology and Resources*. 8: 3–17.
- Idowu, O.O., Salami, A.O., Ajayi, S.A., Akinwale, R.O. and Sere, Y. (2013). Varietal resistance of rice to blast fungus *Magnaporthe oryzae* at two sites in southwestern Nigeria. *African Journal of Biotechnology*. 12(33): 5174-5182. doi: 10.5897/AJB2012.2959.
- IWMI (International Water Management Institute). (2000). IWMI global water scarcity study. International Water Management Institute. Colombo, Sri Lanka.

- IRGSP (International Rice Genome Sequencing Project). (2005). The map-based sequence of the rice genome. *Nature*. 436: 793-800.
- IRRI (International Rice Research Institute). (1975). *Major research in upland rice*. Diseases of upland rice and their control through varietal resistance. Los Banos, Philippines.
- IRRI (International Rice Research Institute). (1996). *Standard evaluation system for rice*. 4th edn. Manila, Philippines.
- IRRI (International Rice Research Institute). (2011). Major research in upland rice. The international rice research institute Los Banos, Laguna, Manila, Philippines. Available at: http://en.wikipedia.org/wiki/Upland_rice#cite_note-0(Accessed 22 august 2011).
- Isiaka, K., Ismail, M.R. and Adam, P. Influence of seed priming on germination traits, gas exchange, biomass production and grain yield of rice. Proceedings of the 2nd rice research colloquium, 22 April, Universiti Putra Malaysia, Malaysia, April 2015.
- Jacquot, M. and Courtois, B. (1987). The rice plant. In *Upland Rice*. London, Macmillan Publishers.
- Jagadeb, P.N. and Samal, K.M. (1991). Multivariate analysis in niger (Guizotica abyssinicacass). Indian Journal of Genetics. 30(1): 212-221.
- Jannink, J.L., Lorenz, A.J. and Iwata, H. (2010). Genomic selection in plant breeding: from theory to practice. *Brief Functional Genomics*. 9: 166–177.
- Jantasuriyarat, C., Gowda, M., Haller, K., Hatfield, J. and Lu, G., Stahlberg, E., Zhou, B., Li, H., Kim, H.R., Yu, Y., Dean, R.A., Wing, R.A., Soderlund, C. and Wang, G.L. (2005). Large-scale identification of expressed sequence tags involved in rice and rice blast fungus interaction. *Plant physiology*. 138: 105. PMID: 15888683.
- Jhala, V.M., Mandaliya, V.B. and Thaker, V.S. (2015). Simple and efficient protocol for RNA and DNA extraction from rice (*Oryza sativa* L.) for downstream applications. *International Research Journal of Biological Sciences*. 4(2): 62-67.
- Jia, Y., Adams, S.A.M., Bryan, G.T., Hershay, H.P. and Valent, B. (2000). Direct interaction of resistance genes products confer rice blast resistance. *Embryology Journal*. 19: 4004-4014.
- Johnson, H.W., Robinson, H. and Comstock, R. (1955). Estimates of genetic and environmental variability in soybeans. *Agronomy Journal*. 47: 314–318.

- Kaku, H., Nishizawa, Y., Ishii-Minami, N., Tomiyama, C.A., Dohmae, N., Takio, K., Minami, E. and Shibuya, N. (2006). Plant cells recognize chitin fragments for defense signaling through a plasma membrane receptor. *Proceedings of the National Academy of Sciences of the United States of America*. 103: 11086–91.
- Kanehisa, M., Araki, M., Goto, S., Hattori, M., Hirakawa, M., Itoh, M., Katayama, T., Kawashima, S., Okuda, S., Tokimatsu, T. and Yamanishi, Y. (2008). KEGG for linking genomes to life and the environment. *Nucleic Acids Research*. 36 (Database issue): D480-4.
- Karim, S.M.R., Man, A.B. and Sahid, I.B. (2004). Weed problems and their management in rice fields of Malaysia: an overview. Weed Biology and Management. 4: 177-186.
- Khan, J.A., Jamil, F.F., Cheema, A.A. and Gill, M.A. Screening of rice germplasm against blast disease caused by Pyricularia oryza. In: Proceedings of National Conference of Plant Pathology, held at NARC, Islamabad, 2001.
- Khush, G.S. (2005). What it will take to feed 5.0 billion rice consumers in 2030. *Plant Molecular Biology*. 59 (1): 1–6.
- Khush, G.S. and Jena, K. (2009). Current status and future prospects for research on blast resistance in rice (*Oryza sativa* L.). In X. Wang, B. Valent (Eds.). *Advances in genetics, genomics and control of rice blast disease* (pp. 1–10). Springer, Dordrecht.
- Kiedrowski, S., Kawalleck, P., Hahlbrock, K., Somssich, I.E. and Dangl J.L. (1992). Rapid activation of a novel plant defense gene is strictly dependent on the Arabidopsis RPMI1 disease resistance locus. *European Molecular Biological* Organization (EMBO) Journal. 11: 4677–4684.
- Kihoro, J., Bosco, N.J., Murage, H., Ateka, E. and Makihara, D. (2013). Investigating the impact of rice blast disease on the livelihood of the local farmers in greater Mwea region of Kenya. *SpringerPlus*. 2:308.
- Kilusang, M.P. Layosa, P.M., Keya, A., Quijano, R.F., Adapon, S.Q., Pelegrina, W.R. and Cruz, P.A.Z. (2007). *The great rice robbery: A handbook on the impact of IRRI in Asia*. Pesticide Action Network Asia and the Pacific (PAN AP) publisher, Penang, Malaysia. ISBN 978-983-9381-35-1.
- Kim, S., Park, J., Park, S.Y., Mitchel, T.K. and Lee, Y.H. (2010). Identification and analysis of in planta expressed genes of *Magnaporthe oryzae*. *BMC Genomics*. 11:104. doi: 10.1186/1471-2164-11-104 PMID: 20146797.
- King, B.C., Waxman, K.D., Nenni, N.V., Walker, L.P., Bergstrom, G.C. and Gibson, D.M. (2011). Arsenal of plant cell wall degrading enzymes reflects host preference among plant pathogenic fungi. *Biotechnology Biofuels*. 4: 1–14.

- Koutroubas, S.D., Katsantonis, D., Ntanos, D.A. and Lupotto, E. (2009). Blast disease influence on agronomic and quality traits of rice varieties under Mediterranean conditions. *Turkish Journal of Agro-forestry*. 33: 487-494.
- Lai, K., Duran, C., Paul, J., Michał, B., Lorenc, T., Stiller, J., Manoli, S., Matthew, J., Hayden, Kerrie, L., Forrest, Fleury, D., Baumann, U., Zander, M., Annaliese S.M., Batley, J. and Edwards, D. (2012). Single nucleotide polymorphism discovery from wheat next-generation sequence data. *Plant Biotechnology Journal*. 10: 743–749.
- Lasalita-Zapico, F.C., Namocatcat, J.A. and Carino-Turner, J.L. (2010). Genetic diversity analysis of traditional upland rice cultivars in Kihan, Malapatan, Sarangani Province, Philippines using morphometric markers. *Philippines Journal of Science*. 139: 177–180.
- Levine, A., Tenhaken, R., Dixon, R. and Lamb, C.J. (1994). H₂O₂ from the oxidative burst orchestrates the plant hypersensitive disease resistance response. *Cell*. 79: 583–593.
- Liang, D., Liu, M., Hu, Q., He, M., Qi, X., Xu, Q., Zhou, F. and Chen, X. (2014). Identification of differentially expressed genes related to aphid resistance in cucumber (*Cucumis sativus* L.). Scientific Reports. 5: 9645. doi:10.1038/srep09645.
- Li, H. and Durbin, R. (2009). Fast and accurate short read alignment with Burrows– Wheeler transform. *Bioinformatics*. 25(14):1754–1760.
- Li, L., Wang, X., Stolc, V., Li, X., Zhang, D., Su, N., Tongprasit, W., Li, S., Cheng, Z., Wang, J. and Deng, X.W. (2006). Genome-wide transcription analyses in rice using tiling microarrays. *Nature Genetics*. 38: 124-129.
- Lin, S., Dittert, K., Tao, H.B., Kreye, C., Xu, Y.C., Shen, Q.R., Fan, X.L. and Sattelmacher, B. The Ground Cover Rice production System (GCRPS)-a successful new approach to save water and increase nitrogen fertilizer efficiency. In: Bouman BAM et al (eds) Water- wise Rice Production. Proceedings of the International Workshop on Water- wise Rice Production, Los Banos, Philippines, 8-11 April 2002.
- Liu, L., Li, Y., Li, S., Hu, N., He, Y., Pong, R., Lin, D., Lu, L. and Law, M. (2012). Comparison of next-generation sequencing systems. *Journal of Biomedicine* and Biotechnology. doi:10.1155/2012/251364.
- Liu, Y., Liu, M. Li, X., Cao, B. and Ma, X. (2014). Identification of differentially expressed genes in leaf of *Reaumuria soongorica* under PEG-induced drought stress by digital gene expression profiling. *PLoS One*. 9(4): e94277.
- Lu, T., Lu, G., Fan, D., Zhu, C., Li, W., Zhao Q., Feng, Q., Zhao, Y., Guo, Y., Li, W., Huang, X. and Han, B. (2010). Function annotation of the rice transcriptome at single-nucleotide resolution by RNA-seq. *Genome Research*. 20(9): 1238-49. doi: 10.1101/gr.106120.110.

- Ma, L.J., Does, V.D.H.C., Borkovich, K.A., Coleman, J.J., Daboussi, M.J., Di, P.A., Dufresne, M., Freitag, M., Grabherr, M. and Henrissat, B. Houterman, P.M., Kang, S., Shim, W.B., Woloshuk, C., Xie, X., Xu, J.R., Antoniw, J., Baker, S.E., Bluhm, B.H., Breakspear, A., Brown, D.W., Butchko, R.A.E., Chapman, S., Coulson, R., Coutinho, P.M., Danchin, E.G.J., Diener, A., Gale, L.R., Gardiner, D.M., Goff, S., Hammond-Kosack, K.E., Hilburn, K., Hua-Van, A., Jonkers, W., Kazan, K., Kodira, C.D., Koehrsen, M., Kumar, L., Lee, Y.H., Li, L., Manners, J.M., Miranda-Saavedra, D., Mukherjee, M., Park, G., Park, J., Park, S.Y., Proctor, R.H., Regev, A., Ruiz-Roldan, M.C., Sain, D., Sakthikumar, S., Sykes, S., Schwartz, D.C., Turgeon, B.G., Wapinski, I., Yoder, O., Young, S., Zeng, Q., Zhou, S., Galagan, J., Cuomo, C.A., Kistler, H.C. and Rep, M. (2010). Comparative genomics reveals mobile pathogenicity chromosomes in *Fusarium. Nature.* 464(7287): 367–373.
- Mackay, I. and Powell, W. (2007). Methods for linkage disequilibrium mapping in crops. *Trends in Plant Science*. 12: 57–63.
- Mae, T. (1997). Physiological nitrogen efficiency in rice: Nitrogen utilization, photosynthesis, and yield potential. *Plant and Soil*. 196: 201–210.
- Mahmood, T., Kakishima, M. and Komatsu, S. (2007). Proteomic analysis of jasmonic acid-regulated proteins in rice leaf blades. *Protein Peptide Letters*. 14: 311–319.
- Mardis, E.R. (2014). DNA sequencing technology: A Perspective from Dr. Elaine Mardis. http://genome.wustl.edu/articles/detail/dna-sequencing-technology-aperspective-from-dr-elaine-mardis/.
- Mathure, S., Shaikh, A., Renuka, N., Wakte, K., Jawali, N. and Thengane, N.A. (2011). Characterization of aromatic rice (*Oryza sativa* L.) germplasm and correlation between their agronomic and quality traits. *Euphytica*. 179: 237-246.
- Mazid, M.S. (2013). Genetic divergence of rice genotypes resistant to bacterial blight revealed by quantitative traits and molecular markers. MS thesis. Institute of Tropical Agriculture, Universiti Putra Malaysia, Malaysia.
- Mazid, M.S., Rafii, M.Y., Hanafi, M.M., Rahim, H.A. and Latif, M.A. (2013). Genetic variation, heritability, divergence and biomass accumulation of rice genotypes resistance to bacterial blight revealed by quantitative traits and ISSR markers. *Physiologia Plantarum.* 149: 432-447.
- Meena, B.S. (2005). Morphological and molecular variability of rice blast pathogen Pyricularia grisea (Cooke) Sacc. MS thesis. Dharwad University of Agricultural Sciences, Dharwad, India.
- Metcalf, H. (1906). A preliminary report on blast of rice, with notes on other rice disease. Bulletin of the South Carolina Agricultural Experiment Station. 121: 43.

Metcalf, H. (1907). The pathology of rice plant. Science. 25: 264 – 265.

- Meyer, N.W. (2012). No simple thing: how rice will reshape the world. http://www.internationalpolicydigest.org/2012/06/13/no-simple-thing-how-rice-will-reshape-the-world/. Access on 8 August 2015.
- Miah, G., Rafii, M.Y., Ismail, M.R., Puteh, A.B., Rahim, M.A. and Latif, M.A. (2015). Recurrent parent genome recovery analysis in a marker-assisted backcrossing program of rice (*Oryza sativa* L.). *Comptes Rendus Biologies*. 338: 83–94.
- Mohanta, B.K., Aslam, M.R., Kabir, M.E., Anam, M.K., Alam, M.K. and Habib, M.A. (2003). Performance of different genotypes/cultivars to blast disease of rice in Boro and T. Aman crop in Bangladesh. Asian Journal of Plant Science. 2(7): 575-577.
- Morishita, K. (1984). Wild plant and domestication. In *Biology of rice*, S. Tsunoda and N. Takahashi, edn. (pp. 3–30). Amsterdam: Japan Scientific Societies Press, Elsevier.
- Morozova, O., Hirst, M. and Marra M.A. (2009). Applications of new sequencing technologies for transcriptome analysis. *Annual Review of Genomics and Human Genetics*. 10:135–51. doi:10.1146/annurev-genom-082908-145957.
- Mosquera, G., Giraldo, M.C., Khang, C.H., Coughlan, S. and Valent, B. (2009). Interaction transcriptome analysis identifies *Magnaporthe oryzae* BAS1-4 as biotrophy-associated secreted proteins in rice blast disease. *Plant Cell*. 21: 1273. doi: 10.1105/tpc.107.055228 PMID: 19357089.
- Mousanejad, S., Alizadeh, A. and Safaie, N. (2010). Assessment of yield loss due to rice blast disease in Iran. *Journal of Agricultural Science and Technology*. 12: 357-364.
- Naher, U.A. (2009). Effects of root exudates on specific diazotroph-rice genotype association. PhD thesis. Faculty of Agriculture, Universiti Putra Malaysia, Malaysia.
- Nerson, H. (1980). Effects of population density and number of ears on wheat nitrogen fertilizer rate. *Crop Science*. 43: 921–926.
- Nicaise, V., Roux, M. and Zipfel, C. (2009). Recent advances in PAMP triggered immunity against bacteria: pattern recognition receptors watch over and raise the alarm. *Plant Physiology*. 150: 1638–1647.

Normile, D. (1997). Archaeology: Yangtze seen as earliest rice site. Science. 275-309.

- Novaes, E., Drost, D.R., Farmerie, W.G., Pappas, G.J.J., Grattapaglia, D., Sederoff, R.R. and Kirst, M. (2008). High-throughput gene and SNP discovery in *Eucalyptus grandis*, an uncharacterized genome. *BMC Genomics*. 9: 312.
- Nurnberger, T. and Kemmerling, B. (2009). Pathogen-associated molecular patterns (PAMP) and PAMP-triggered immunity. *Annual Plant Reviews*. 34: 16–47.

- Oghenekaro, A.O., Raffaello, T., Kovalchuk, A. and Asiegbu, F.O. (2016). De novo transcriptomic assembly and profiling of *Rigidoporus microporus* during saprotrophic growth on rubber wood. *BMC Genomics*. 17: 234. doi: 10.1186/s12864-016-2574-9.
- Oritani, T. (1995). Mechanisms of aging and senescence. In Science of the rice plant: Physiology, Vol. 2, T. Matsuo, K. Kumazawa, R. Ishii, K. Ishihara and H. Hirata, edn. (pp. 164–178). Tokyo: Food and Agriculture Policy Research Center.
- Osman, K.A., Mustafa, A.M., Ali, F., Yonglain, Z. and Fazhan, Q. (2012). Genetic variability for yield and related attributes of upland rice genotypes in semi arid zone (Sudan). *African Journal of Agricultural Research*. 7(3): 4613-4619. doi: 10.5897/AJAR12.529.
- Ou, S.H., Ling, K.C., Kauffman, H.E. and Khush, G.S. (1975). Diseases of upland rice and their control through varietal resistance. *Major research in upland rice* (pp. 126-135). Los Banos, Philippines.
- Ou, S.H. (1985). *Rice Diseases*. 2nd Edition. Commonwealth Mycological Institute, Kew, UK.
- Padmanabhan, S.Y. (1965a). Estimating losses from rice blast in India. In *The rice blast disease: Proceedings of Symposium. At IRRI, July*, 1963: (pp. 203-221). Baltimore, Maryland, Johns Hopkins.
- Padmanabhan, S.Y. (1965b). Breeding for blast resistance in India.In *The rice blast disease: Proceedings of Symposium. At IRRI, July*, 1963: (pp. 343-359). Baltimore, Maryland, Johns Hopkins Press.
- Padmavathi, G., Ram, T., Satyanarayana, K. and Mishra, B. (2005). Identification of blast (*Magnaporthe grisea*) resistance genes in rice. *Current Science*. 88: 628– 630.
- Pandey, P., Anurag, P.J., Tiwari, D., Yadav, S. and Kumar, B. (2009). Genetic variability, diversity and association of quantitative traits with grain yield in rice (*Oryza sativa* L.). *Journal of Bioscience*. 17: 77-82.
- Pandey, P. and Anurag, P.J. (2010). Estimation of genetic parameters in indigenous rice. *AAB Bioflux* 2 (1) http://www.aab.bioflux.com.ro (Accessed on 3 July 2012).
- Park, N.K. (1993). Classification of rice cultivars by the rice blast reaction and its utilization for breeding. PhD thesis, Chungnam National University, Taejon, Korea.
- Parker, D., Beckmann, M., Enot, P., Overy, D.P., Rios, Z.C., Gilbert, M. and Draper, J. (2008). Rice blast infection of *Brachypodium distachyon* as a model system to study dynamic host/pathogen interactions. *Nature Protocol.* 3(3): 435–445.

- Parkinson, J. and Blaxter, M. (2009). Expressed sequence tags: an overview. Methods in Molecular Biology. 533: 1-12.
- Pasha, A., Babaeian-Jelodar, N., Bagheri, N., Nematzadeh, G. and Khosravi, V. (2013). A field evaluation of resistance to *Pyricularia oryzae* in rice genotypes. *International Journal of Agriculture and Crop Science*. 5(4): 390-394.
- Power, J.F. and Alessi, J. (1978). Tiller development and yield of standard and semi dwarf spring wheat varieties as affected by nitrogen fertilizer. *Journal of Agricultural Science*. 90: 97–108.
- Prabhu, A.S. and Morais, O.P. (1986). Blast disease management in upland rice in Brazil. In progress in upland Rice Research. In Proceedings of the 1985 Jakarta Conference.
- Pranab, K.S. (2008). Compilation of rice production training materials. Bangladesh Rice Research Institute, Gazipur, Bangladesh.
- Puteh, A.B., Mondal, M.M., Ismail, M.R. and Latif, M.A. (2014). Grain sterility in relation to dry mass production and distribution in rice (*Oryza sativa* L.). *BioMed Research International*. http://dx.doi.org/10.1155/2014/302179.
- Rafalski, A. (2002). Applications of single nucleotide polymorphisms in crop genetics. *Current Opinion in Plant Biology*, 5: 94–100.
- Reuber, T.L. and Ausubel, F.M. (1996). Isolation of Arabidopsis genes that differentiate between resistance responses mediated by the RPS2 and RPM1 disease resistance genes. *Plant Cell*. 8: 241–249.
- Ribot, C., Hirsch, J., Balzergue, S., Tharreau, D., Notteghem, J.L., Lebrun, M.H. and Morel, J.B. (2008). Susceptibility of rice to the blast fungus, *Magnaporthe* grisea. Journal of Plant Physiology. 165: 114-124.
- Ricepedia. (2015). The online authority on rice. Ricepedia.org/challenges/food-security. Access on 25 September, 2015.
- Sah, S.K., Kaur, G. and Kaur, A. (2014). Rapid and reliable method of high-quality RNA extraction from diverse plants. *American Journal of Plant Sciences*. 5: 3129-3139.
- Sahebi, M., Hanafi, M.M., Abdullah, S.N.A., Rafii, M.Y., Azizi, P., Nejat, N. and Idris, A.S. (2014). Isolation and expression analysis of novel silicon absorption gene from roots of mangrove (*Rhizophora apiculata*) via suppression subtractive hybridization. *BioMed Research International*. http://dx.doi.org/10.1155/2014/971985.
- Saito, K., Linquist, B., Atlin, G.N., Phanthaboon, K., Shiraiwa, T. and Horie, T. (2006). Response of traditional and improved upland rice cultivars to N and P fertilizer in northern Laos. *Field Crops Research*. 96: 216-223.

- Sakata, K., Nagamura, Y., Numa, H., Antonio, B.A., Nagasaki, H., Idonuma, A., Watanabe, W., Shimizu, Y., Horiuchi, I., Matsumoto, T., Sasaki, T. and Higo, K. (2002). Rice GAAS: an automated annotation system and database for rice genome sequence. *Nucleic Acids Research*. 30: 98-102.
- Santos, A.B., Fageria, N.K. and Prabhu, A.S. (2003). Rice ratooning management practices for higher yields. *Communications in Soil Science and Plant Analysis*. 34: 881–918.
- Saravanan, R. and Senthil, N. (1997). Genotypic and phenotypic variability, heritability and genetic advance in some important traits in rice. *Madras Agricultural Journal*. 84: 276-277.
- Sariam, O., Khanif, Y.M. and Zahrah, T., (2002). Rice growth and nitrogen uptake as influenced by water management. *Malaysian Journal of Soil Science*. 6: 1-11.
- Sasaki, T.A. and Sedoroff, R. (2003). Genome studies and molecular genetics: The rice genome and comparative genomics of higher plants. *Current Opinion in Plant Biology*. 6: 97-100.
- Schirawski, J., Mannhaupt, G., Munch, K., Brefort, T., Schipper, K., Doehlemann, G., Di Stasio, M., Rossel, N., Mendoza-Mendoza A. and Pester, D., Müller, O., Winterberg, B., Meyer, E., Ghareeb, H., Wollenberg, T., Münsterkötter, M., Wong, P., Walter, M., Stukenbrock, E., Güldener, U. and Kahmann, R. (2010). Pathogenicity determinants in smut fungi revealed by genome comparison. *Science*. 330(6010): 1546–1548.
- Sester, M., Raboin, L.M., Ramanantsoanirina, A. and Tharreau, D. Endure International Conference Diversifying crop protection, 12-15 October, La Grande-Motte, France, 2008.
- Shan, X., Li, Y., Jiang, Y., Jiang, Z., Hao, W. and Yuan, Y. (2013). Transcriptome profile analysis of maize seedlings in response to high-salinity, drought and cold stresses by deep sequencing. *Plant Molecular Biology Reporters*. 31: 1485–1491.
- Sharma, T.R., Madhav, M.S., Singh, B.K., Shanker, P., Jana, T.K., Dalal, V., Pandit, A., Singh, A., Gaikwad, K., Upreti, H.C. and Singh N.K. (2005). High resolution mapping, cloning and molecular characterization of the PiKh gene of rice, which confers resistance to *M. grisea. Molecular Genetics and Genomics.* 274: 569–578.
- Singh, R.K., Singh, U.S. and Khush, G.S. (2000). *Aromatic Rices*. Oxford and IBH Publishing Co. Ltd. New Dellhi.
- Skamnioti, P. and Gurr, S.J. (2009). Against the grain: safeguarding rice from rice blast disease. *Trends in Biotechnology*. 27: 141e150.

- Soanes, D.M., Chakrabarti, A., Paszkiewicz, K.H., Dawe, A.L. and Talbot, N.J. (2012). Genomewide transcriptional profiling of appressorium development by the rice blast fungus *Magnaporthe oryzae*. *PLoS Pathogen*. 8(2): e1002514. doi:10.1371/journal.ppat.1002514.
- Sohrabi, M., Rafii, M.Y., Hanafi, M.M., Siti, N.A. and Latif, M.A. (2012). Genetic diversity of upland rice germplasm in Malaysia based on quantitative traits. *The Scientific World Journal*. doi:10.1100/2012/416291.
- Sorenson, J., Pradhan, A., Vijaychander, S., Sangari, B., Fang, S., Nguyen, T., Jones, B., Guo, D. and Doan, Q. (2004). Accurate detection and classification of heterozygous indels by direct sequencing. *Applied Biosystems*.
- Statista. (2015). The statistics portal. http://www.statista.com/search/?statistics=1. Access on 8 August 2015.
- Stoop, W., Uphoff, N. and Kassam, A. (2002). A review of agricultural research issues raised by system of rice intensification (SRI) from Madagascar: opportunities for improving farming systems for resource-poor farmers. Agricultural System. 71: 249-274.
- Strickler, S.R., Bombarely, A. and Mueller, L.A. (2012). Designing a transcriptome next-generation sequencing project for a nonmodel plant species. *American Journal of Botany*. 99(2): 257–266.
- Tabbal, D.F., Bouman, B.A.M., Bhuiyan, S.I., Sibayan, E.B. and Sattar, M.A. (2002). On-farm strategies for reducing water input in irrigated rice: case studies in the Philippines. Agricultural Water Management. 56: 93-112.
- Takahashi, N. (1984). Differentiation of ecotypes in *Oryza sativa* L. In S. Tsunoda and N. Takahashi. *Biology of rice* (pp. 31–67). Amsterdam: Japan Scientific Societies Press, Elsevier.
- Takano, Y., Choi, W.B., Mitchell, T.K., Okuno, T. and Dean, R.A. (2003). Large scale parallel analysis of gene expression during infection-related morphogenesis of *Magnaporthe grisea*. *Molecular Plant Pathology*. 4: 337–346.
- Talbot, N.J. (2003). On the trail of a cereal killer: exploring the biology of *Magnaporthe* grisea. Annual Review of Microbiology. 57: 177-202.
- Tena, G., Boudsocq, M. and Sheen, J. (2011). Protein kinase signaling networks in plant innate immunity. *Current Opinion of Plant Biology*. 14: 519–29.
- Torres, M.A., Jones, J.D. and Dang, J.L. (2005). Pathogen-induced, NADPH oxidasederived reactive oxygen intermediates suppress spread of cell death in *Arabidopsis thaliana. Nature Genetics.* 37: 1130–1134.
- Trapnell, C., Pachter, L. and Salzberg, S.L. (2009). Top Hat: discovering splice junctions with RNA-Seq. *Bioinformatics*. 25(9): 1105–1111.

- Tuhina-Khatun, M., Newaz, M.A. and Bari, M.A.A. (2007). Combining ability and heritability estimates in F2 diallel population of spring wheat under interacting environments. *Bangladesh Journal of Agricultural Science*. 34: 75-82.
- Tuong, T.P. and Bouman, B.A.M. *Rice production in water-scarce environments*. In: Proceedings of the Water Productivity Workshop, Colombo, Sri Lanka, 12-14 November 2001.
- Tyagi, A.K., Khurana, J.P., Khurana, P., Raghuvanshi. S., Gaur, A., Kapur. A., Gupta, V., Kumar, D., Ravi, V., Vij, S., Khurana, P. and Sharma, S. (2004). Structural and functional analysis of rice genome. *Journal of Genetics*. 83(1): 79–99.
- Ullah, M.Z., Bashar, M.K., Bhuiya, M.S.R., Khalequzzaman, M. and Hasan, M.J. (2011). Interrelationship and cause-effect analysis among morphophysiological traits in Biroin rice of Bangladesh. *International Journal of Plant Breeding and Genetics.* 5: 246-254.
- Vaghefi, N., Shamsudin, M.N., Makmom, A. and Bagheri, M. (2011). The economic impact of climate change on the rice production in Malaysia. *International Journal of Agricultural Researches*. 6: 67-74.
- Valent, B., Farrall, L. and Chumley, F.G. (1991). Magnaporthe grisea genes for pathogenicity and virulence identified through a series of backcrosses. *Genetics*. 127: 87-101.
- Vange, T. (2009). Biometrical studies on genetic diversity of some upland rice (*Oryza Sativa* L.) accessions. *Nature* and *Science*. 7(1): ISSN 1545-0740, http://www.sciencepub.net, naturesciencej@gmail.com.
- Varshney, R.K., Nayak, S.N., May, G.D. and Jackson S.A. (2009). Next-generation sequencing technologies and their implications for crop genetics and breeding. *Trends in Biotechnology*. 27(9): 522-530. doi:10.1016/j.tibtech.2009.05.006.
- Vergara, B.S. and Chang, T.T. (1976). The flowering response of the rice plant to photoperiod: a review of literature. *International Rice Research Institute Technology Bulletin.* 8: 75.
- Voisey, C.R. and Slusarenko, A.J. (1989). Chitinase mRNA and enzyme activity in *Phaseolus vulgaris* (L) increases more rapidly in response to avirulent than to virulent cells of *Pseudomonas syringae* pv *phaseoicola*. *Physiological and Molecular Plant Pathology*. 35: 403–412.
- Weber, A.P.M., Weber, K.L., Carr, K., Wilkerson, C. and Ohlrogge, J.B. (2007). Sampling the Arabidopsis transcriptome with massively parallel pyrosequencing. *Plant Physiology*. 144: 32–42.
- Wicker, T., Schlagenhauf, E., Graner, A., Close, T.J., Keller, B. and Stein, N. (2006). 454 sequencing put to the test using the complex genome of barley. *BMC Genomics*. 7: 275.

- Wilson, R.A. and Talbot, N.J. (2009). Under pressure: investigating the biology of plant infection by *Magnaporthe oryzae*. *Nature Reviews Microbiology*. 7: 185-195. doi:10.1038/nrmicro2032.
- Worede, F., Srecwongchai, T., Phumichai, C. and Srippichitt, P. (2014). Multivariate analysis of genetic diversity among some rice genotypes using morpho-agronomic traits. *Journal of Plant Science*. 9(1): 14-24.
- Wrather, A. (2009). Rice blast control. http://extension.missouri.edu/p/MP645.
- Wu, J., Kou, Y., Bao, J., Li, Y., Tang, M., Zhu, X., Ponaya, A., Xiao, G., Li, J., Li, C., Song, M.Y., Cumagun, C.J.R., Deng, Q., Lu, G., Jeon, J.S., Naqvi, N. and Zhou, B. (2015). Comparative genomics identifies the *Magnaporthe oryzae* avirulence effector AvrPi9 that triggers Pi9-mediated blast resistance in rice. *New Phytologist.* doi: 10.1111/nph.13310.
- Wu, S.C., Halley, J.E., Luttig, C., Fernekes, L.M., Gutie'rrez-Sanchez, G., Darvill, A.G. and Albersheim, P. (2006). Identification of an endo-b-1, 4-D-xylanase from *Magnaporthe grisea* by gene knockout analysis, purification, and heterologous expression. *Applied and Environmental Microbiology*, 72: 986–93.
- Wubben, J.P., Lawrence, C.B. and de Wit P.J.G.M. (1996). Differential induction of chitinase and 1, 3-b glucanase gene expression in tomato by *Cladosporium fulvum* and its race specific elicitors. *Physiological and Molecular Plant Pathology*. 48: 105–116.
- Xiao, M., Zhang, Y., Chen, X., Lee, Eun-Jeong., Barber, C.J.S., Chakrabarty, R., Desgagné-Penix, I., Haslam, T.M., Kim, Yeon-Bok., Liu, E., MacNevin, G., Masada-Atsum, S., Reed, D.W. Stout, J. M., Zerbe, P., Zhang, Y., Bohlmann, J., Covello, P.S., Luca, V.D., Page, J.E., Ro, Dae-Kyun., Martin, V.J.J., Facchini, P.J. and Sensena C.W. (2013). Transcriptome analysis based on nextgeneration sequencing of non-model plants producing specialized metabolites of biotechnological interest. *Journal of Biotechnology*. 166: 122–134.
- Xu, J., Li, Y., Ma, X., Ding, J., Wang, K., Wang, S., Tian, Y., Zhang, H. and Zhu, X.G. (2013). Whole transcriptome analysis using next-generation sequencing of model species *Setaria viridis* to support C4 photosynthesis research. *Plant Molecular Biology*. doi: 10.1007/s11103-013-0025-4.
- Xu, X., Liu, X., Ge, S., Jensen, J.D. Hu, F., Li, X., Dong, Y., Gutenkunst, R.N., Fang, L., Huang, L., Li, J., He, W., Zhang, G., Zheng, X., Zhang, F., Li, Y., Yu, C., Kristiansen, K., Zhang, X., Wang, J., Wright, M., McCouch, S., Nielsen, R., Wang, J. and Wen, W. (2012). Resequencing 50 accessions of cultivated and wild rice yields markers for identifying agronomically important genes. *Nature Biotechnology*. 30(1): 105–111.

- Xue, M., Yang, J., Li, Z., Hu, S. and Yao, N., Dean, R.A., Zhao, W., Shen, M., Zhang, H., Li, C., Liu, L., Cao, L., Xu, X., Xing, Y., Hsiang, T., Zhang, Z., Xu, J.R. and Peng Y.L. (2012). Comparative analysis of the genomes of two field isolates of the rice blast fungus *Magnaporthe oryzae*. *PLoS Genetics*. 8: e1002869. doi: 10.1371/journal.pgen.1002869 PMID: 22876203.
- Yockteng, R., Almeida, A.M.R., Yee, S., Andre, T., Hill, C. and Specht, C.D. (2013). A method for extracting high-quality RNA from diverse plants for next-generation sequencing and gene expression analyses. *Applications in Plant Sciences*. 1(12): 1300070.
- Yoshida, S. (1972). Physiological aspects of grain yield. Annual Review of Plant Physiology. 23: 437–464.
- Yoshida, S. (1981). *Fundamentals of rice crop science*. International Rice Research Institute. Los Banos, Philippines.
- Yoshida, K., Saitoh, H., Fujisawa, S., Kanzaki, H., Matsumura, H., Yoshida, K., Tosa, Y., Chuma, I., Takano, Y., Win, J. and Kamoun, S. and Terauchia, R. (2009). Association genetics reveals three novel avirulence genes from the rice blast fungal pathogen *Magnaporthe oryzae*. *Plant Cell*. 21(5): 1573–1591.
- Yu, C., Chen, H., Tian, F., Leach, J.E. and He, C. (2014). Differentially-expressed genes in rice infected by *Xanthomonas oryzae* pv. *oryzae* relative to aflagellin deficient mutant reveal potential functions of flagellin in host-pathogen interactions. *Rice*. 7: 20. http://www.thericejournal.com/content/7/1/20.
- Yu, J., Hu, S., Wang, J., Wong, G.K.S., Li, S., Liu, B., Deng, Y., Dai, L., Zhou, Y., Zhang, X., Cao, M., Liu, J., Sun, J., Tang, J., Chen, Y., Huang, X., Lin, W., Ye, C., Tong, W., Cong, L., Geng, J., Han, Y., Li, L., Li, W., Hu, G., Huang, X., Li, W., Li, J., Liu, Z., Li, L., Liu, J., Qi, Q., Liu, J., Li, L., Li, T., Wang, X., Lu, H., Wu, T., Zhu, M., Ni, P., Han, H., Dong, W., Ren, X., Feng, X., Cui, P., Li, X., Wang, H., Xu, X., Zhai, W., Xu, Z., Zhang, J., He, S., Zhang, J., Xu, J., Zhang, K., Zheng, X., Dong, J., Zeng, W., Tao, L., Ye, J., Tan, J., Ren, X., Chen, X., He, J., Liu, D., Tian, W., Tian, C., Xia, H., Bao, Q., Li, G., Gao, H., Cao, T., Wang, J., Zhao, W., Li, P., Chen, W., Wang, X., Zhang, Y., Hu, J., Wang, J., Liu, S., Yang, J., Zhang, G., Xiong, Y., Li, Z., Mao, L., Zhou, C., Zhu, Z., Chen, R., Hao, B., Zheng, W., Chen, S., Guo, W., Li, G., Liu, S., Tao, M., Wang, J., Zhu, L., Yuan, L., Yang, H. (2002). A draft sequence of the rice genome (*Oryza sativa* L. ssp *indica*). *Science*. 296: 79-92.
- Yun, K.Y., Park, M.R., Mohanty, B., Herath, V., Xu, F., Mauleon, R., Wijaya, E., Bajic, V.B., Bruskiewich, R. and Reyes, B.G. (2010). Transcriptional regulatory network triggered by oxidative signals configures the early response mechanisms of japonica rice to chilling stress. *BMC Plant Biology*. 10: 16.
- Zeigler, R.S., Leong, S.A. and Teng, P.S. (1994). *Rice blast disease*. International Rice Research Institute, UK: CAB International, Wallingford.

- Zhang, G., Liu, X., Quan, Z., Cheng, S., Xu, X., Pan, S., Xie, M., Zeng, P., Yue, Z., Wang, W., Tao, Y., Bian, C., Han, C., Xia, Q., Peng, X., Cao, R., Yang, X., Zhan, D., Hu, J., Zhang, Y., Li, H., Li, H., Li, N., Wang, J., Wang, C., Wang, R., Guo, T., Cai, Y., Liu, C., Xiang, H., Shi, Q., Huang, P., Chen, Q., Li, Y., Wang, J., Zhao, Z. and Wang, J. (2012). Genome sequence of foxtail millet (*Setaria italica*) provides insights into grass evolution and biofuel potential. *Nature Biotechnology*. 30(6): 549–554.
- Zhou, J., Tang, X. and Martin, G.B. (1997). The Pto kinase conferring resistance to tomato bacterial speck disease interacts with protein that bind a cis-element of pathogen related genes. *European Molecular Biology Organization (EMBO) Journal*. 16: 3207–3218.
- Zipfel, C. (2008). Pattern-recognition receptors in plant innate immunity. *Current Opinion in Immunology*. 20: 10–16.