

UNIVERSITI PUTRA MALAYSIA

RUMEN FERMENTATION, FATTY ACID PROFILES, MEAT QUALITY AND ADIPOGENESIS RELATED GENE EXPRESSION OF GOAT AND SHEEP FED WITH HIGH CONCENTRATED DIET

CANDYRINE SU CHUI LEN

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By

CANDYRINE SU CHUI LEN

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December 2016

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DEDICATION

To my late grandmother (Anna Chong Kui Fah), who always tell me to 'belajar bagus- bagus di KL, jangan suka- suka ikut orang jalan sini-sana'.

May you rest in peace with the Heavenly Father (1925- 2013).



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

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Chairman: Liang Juan Boo, PhD Institute : Tropical Agriculture and Food Security

The small ruminant (goats and sheep) industry in Malaysia can be described as slow growing as compared to the demand for the products. On the other hand, ruminant products are known to contain high amount of saturated fatty acids (SFA) which are often implicated as a source of modern-day diseases. To date, limited comparative studies have been carried out to characterize differences between goats and sheep, hence the two species are often assumed to be similar and thus raised and managed similarly. To meet the local demand, there is a need to improve the productivity of the small ruminant industry, by optimizing the growth protential of each animal species; and at the same time, to produce healthier meat and meat products. It is hypothesized that the feeding of high concentrate diet supplemented with linseed oil as a source of polyunsaturated fatty acids (PUFA), could improve the growth rate of the animals and at the same time, fortifying the meat and products with health beneficial PUFA. The primary objectives of this thesis were to co-compare rumen fermentation characteristics, growth performance, meat quality attributes and fatty acids profile in tissues of goat and sheep fed high concentrate fattening diet, with and without supplementation of PUFA; and to infer the differences in fatty acids profile due to the dietary treatments and animal species (goats vs sheep) through the expression of adipogenesis related genes. Three experiments were conducted to achieve the above objectives. In the first experiment, in vitro rumen fermentation characteristics in goats and sheep with or without PUFA added to the substrate were evaluated. Results showed superior fermentation characteristics in goats with higher in vitro gas production rate (P<0.001), accompanied by higher volatile fatty acids (VFA) production and *in vitro* organic matter digestibility (P<0.05). The higher population of total bacteria and two major cellulolytic bacteria (Ruminococcus albus and Butyrivibrio fibrisolvens) in goat compared to sheep also indicates higher digestive capability in goats. The addition of 4% linseed oil to the substrate did not show any negative effect on fermentation characteristics nor the VFA production, suggesting that the level oil was appropriate.

Experiment 2 consisted of a 100-day feeding trial to determine the growth performance, digestibility and meat quality attributes of goats and sheep fed fattening diet with or without oil supplementation, in a 2 species x 2 diets factorial experiment with 6 animals (replicates) per treatment. Results of the study showed that irrespective of diet, sheep had better body weight gain (BWG) and feed efficiency (FE) compared to goats. Overall, supplementation of linseed oil at 29 g/day improved BWG by about 65 g/day (45% improvement over the control) and FE by 15% (5.01 vs 5.93). There was no species difference (except for crude protein) and no effect of dietary treatments on apparent digestibility. Goat meat had better quality attributes (P<0.001), including lower lipid oxidation and higher water holding capacity than sheep. Linseed oil supplementation did not affect meat lipid oxidation, indicating that the level of supplementation was appropriate.

The fatty acids profile in various tissue, *in vivo* rumen fermentation and expression of adipogenesis related genes from the above feeding trial are reported in Chapter 5 with the objective of determining whether differences exist in fatty acids profile in goats and sheep fed similar diets and to elucidate the possible mechanisms for any differences exist. Results of the study showed that there were significant differences in the fatty acids profile in the *longissimus dorsi* (LD) muscle, subcutaneous (SC) fat and liver tissue between species and between dietary treatments. Although no species differences were observed in the total SFA, MUFA and PUFA content in the LD muscle, several individual fatty acids were significantly different between species, including higher n-3 PUFA in LD muscle of goats. Sheep had higher total SFA content but lower UFA in the SC fat and liver tissue. Linseed oil supplementation increased the linolenic acid content by 4.5 to 6.1 folds in the tissues samples analyzed suggesting oil supplementation provides a viable approach to enrich meat and ruminant products with essential PUFA.

The result of the *in vivo* rumen fermentation characteristics explained for the differences in the fatty acids profile between species. The higher production of total VFA and acetic acid (primary precursor in fatty acid synthesis) in sheep suggest higher lipogenesis activity, and explained for the higher SFA content. The higher population of *B. fibrisolvens* in sheep than goats, which on the other hand had higher population of total methanogens and methanobacteriales, explained for the higher biohydrogenation intermediate in sheep.

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Relative expressions of adipogenesis related genes were used to elucidate the differences in the fatty acids profile between species and dietary treatment. The study found that, irrespective of diet, sheep had higher expression of fatty acid synthase gene (FASN) gene and also higher Acetyl-CoA carboxylase gene (ACACA) with oil supplementation. The higher expression of the above genes led to higher production of SFA content (especially palmitic acid) in the SC fat and liver tissue in sheep. Stearoyl-CoA desaturase (SCD) gene, which responsibles for the unsaturation of SFA, was lower in sheep than goats and was further downregulated with the addition of oil, which explains for the lower mono-unsaturated fatty acids (MUFA) content in sheep and in the oil supplemented group. On the other hand, peroxisome proliferator-activated receptor alpha (PPAR α), which serves as the transcriptional factor for many lipogenic genes, is negatively correlated with ACACA and FASN genes. Therefore the downregulation of PPAR α gene was accompanied by upregulation of ACACA and FASN genes in sheep and the opposite in goats, accounting for the different SFA production between species.

Result of this thesis suggests that high concentrate feeding supplemented with PUFA (linseed oil at 4% level) is an applicable approach to improve the growth performance of small ruminants, and to produce healthier meat with beneficial fatty acids profile, without detrimental effect on rumen fermentation or meat quality attributes. Species differences in fatty acids profile were partly influenced by the various lipogenic genes expression and therefore, dietary manipulation of adipogenesis related genes expression (nutrigenomics) can be a useful tool especially in reducing the SFA content, in order to produce healthier ruminant meat and meat products. Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENAPAIAN RUMEN, PROFIL ASID LEMAK, KUALITI DAGING DAN EKPRESI GEN BERKAITAN PENGHASILAN LEMAK PADA KAMBING DAN BEBIRI DENGAN PEMAKANAN TINGGI KONSENTRAT

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Industri ruminan kecil (kambing dan bebiri) di Malaysia boleh digambarkan dengan pertumbuhan yang perlahan berbanding dengan permintaan terhadap produk tersebut. Di samping itu, produk ruminan diketahui umum mempunyai kandungan asid lemak tepu (SFA) yang tinggi dan selalu dikaitkan sebagai sumber penyakit hari-moden. Sehingga kini, bilangan kajian yang membandingkan perbezaan antara kambing dan berbiri adalah terhad, maka kedua-dua spesis ternakan tersebut dianggap sama dan dibesarkan dan diurus dengan sama. Untuk memenuhi permintaan tempatan, maka terdapat keperluan untuk meningkatkan produktiviti industry ruminant kecil ini, dengan mengoptimumkan potensi pertumbuhan setiap spesis ternakan tersebut, dan pada masa yang sama, menghasilkan daging dan produk daging yang lebih sihat. Hipotesis kajian adalah pemakanan tinggi konsentrat, ditambah dengan minyak biji rami sebagai sumber asid lemak poli-tak tepu (PUFA), boleh meningkatkan kadar pertumbuhan binatang dan pada masa yang sama, menambahbaik daging dan produk dengan PUFA yang baik untuk kesihatan. Objektif utama tesis ini adalah untuk membandingkan ciri-ciri penapaian rumen, prestasi pertumbuhan, sifat-sifat kualiti daging dan profil asid lemak dalam tisu kambing dan bebiri di bawa pemakanan tinggi konsentrat, tanpa penambahan dan dengan penambahan PUFA; dan untuk merungkai perbezaan profil asid lemak mekanisasi yang bertanggungjawab atas perbezaan profil asid lemak yang disebabkan oleh pemakanan dan spesis binatang (kambing vs berbiri). Tiga eksperimen telah dijalankan untuk mencapai objektif-objektif tersebut. Dalam eksperimen yang pertama, ciri-ciri penapaian rumen in vitro dalam kambing dan bebiri, tanpa penambahan dan dengan penambahan PUFA pada substrat telah dikaji. Kajian menunjukan kelebihan ciri penapaian pada kambing dengan kadar penghasilan gas *in vitro* yang lebih tinggi (P<0.001), diikuti dengan kandungan asid lemak meruap (VFA) dan kebolehcernaan bahan organik in vitro yang lebih tinggi. Populasi jumlah bakteria dan juga dua bakteria utama dalam penghadaman selulosa (Ruminococcus albus dan Butyrivibrio fibrisolvens) yang lebih tinggi pada kambing berbanding bebiri juga menunjukkan keupayaan pencernaan yang lebih tinggi pada kambing. Penambahan minyak biji rami pada kadar 4% pada substrat tidak menunjukkan sebarang kesan negatif terhadap ciri penapaian atau produksi VFA, lalu mencadangkan penggunaan minyak pada kadar tersebut adalah berpatutan.

Eksperimen 2 adalah eksperimen pemakanan secara in vivo selama 100 hari untuk menguji kadar pertumbuhan, kebolehcernaan dan sifat-sifat kualiti daging pada kambing dan bebiri di bawa pemakanan pengemukkan, tanpa penambahan dan dengan penambahan minyak, dalam 2 spesis binatang x 2 jenis pemakanan, eksperimen faktorial dengan 6 binatang (replica) untuk setiap kumpulan rawatan. Hasil kajian menunjukkan bahawa tanpa mengira jenis pemakanan, bebiri mempunyai kenaikan berat badan (BWG) dan nisbah penukaran makanan (FCR) yang lebih baik berbanding kambing. Secara keseluruhan, penambahan minyak biji rami pada kadar 29 g/hari telah menambahbaik BWG sebanyak 65 g/hari (45% penambahbaikan berbanding kumpulan kawalan) dan FE sebanyak 15% (5.01 vs 5.93). Tiada perbezaan spesies (kecuali protin kasar) dan tiada kesan jenis pemakanan terhadap kebolehcernaan ketara makanan. Daging kambing mempunyai sifat-sifat kualiti daging yang lebih baik (P<0.001), termasuk kadar pengoksidaan lipid yang lebih rendah dan keupayaan menyimpan air yang lebih tinggi berbanding daging bebiri. Penambahan minyak biji rami pada makanan tidak mengganggu kadar pengoksidaan lipid, lalu mencadangkan bahawa kadar penambahan minyak tersebut adalah berpatutan.

Profil asid lemak di pelbagai tisu ciri-ciri penapaian rumen in vivo, dan ekspresi gen berkaitan penghasilan lemak daripada eksperimen pemakanan di atas telah dilaporkan di Bab 5, dengan objektif untuk menentukan sama ada wujud perbezaan pada profil asid lemak pada kambing dan bebiri di bawa pemakanan yang sama, dan menjelaskan mekanisasi yang berkemungkinan menyebabkan perbezaan tersebut. Hasil kajian menunjukkan perbezaan yang ketara pada profil asid lemak di daging longissimus dorsi (LD), lemak bawa kulit (SC) dan tisu hati antara spesis binatang dan antara jenis pemakanan. Walaupun tidak terdapat perbezaan spesis binatang pada jumlah kandungan SFA, asid lemak mono-tak tepu (MUFA) dan PUFA dalam daging LD, beberapa asid lemak adalah berbeza secara ketara antara spesis binatang, termasuklah kandungan n-3 PUFA yang lebih tinggi pada daging LD kambing. Bebiri mempunyai kandungan SFA yang lebih tinggi tetapi rendah dalam kandungan asid lemak tak tepu dalam lemak SC dan tisu hati. Penambahan minyak biji rami dalam makanan telah meningkatkan kandungan asid linolenic sebanyak 4.5 hingga 6.1 kali ganda dalam semua tisu yang dikaji lalu mencadangkan penambahan minyak tersebut merupakan jalan yang nyata untuk menambah baik daging dan produk ruminan dengan PUFA penting.

Keputusan ciri-ciri penapaian rumen *in vivo* menjelaskan perbezaan profil asid lemak antara spesis binatang. Jumlah VFA dan asid asetik (sebagai pelopor utama dalam sintesis asid lemak) yang lebih tinggi pada bebiri, mencadangkan aktivit penghasilan lemak yang lebih tinggi, dan menjelaskan kandungan SFA yang lebih tinggi. Populasi *B. fibrisolvens* yang lebih tinggi pada bebiri berbanding kambing, yang mana mempunyai populasi jumlah methanogens dan methanobacteriales yang lebih tinggi, menjelaskan kandungan hasil perantaraan biohydrogenasi pada berbiri.

Ekspresi relatif gen berkaitan penghasilan lemak digunakan untuk menjelaskan perbezaan profil asid lemak antara spesis binatang dan antara pemakanan. Kajian mendapati bahawa tanpa mengira jenis ienis pemakanan, bebiri mempunyai ekspresi gen asid lemak synthase (FASN) yang lebih tinggi, dan juga ekspresi gen Acetyl-CoA carboxylase (ACACA) yang lebih tinggi dalam pemakanan yang ditambah minyak. Ekspresi kedua gen tersebut yang lebih tinggi, mendorong kepada penghasilan SFA (terutamanya asid palmitik) yang lebih tinggi di dalam lemak SC dan tisu hati dalam bebiri. Gen Stearoyl-CoA desaturase (SCD), yang bertanggungjawab untuk menyah-tepukan SFA, adalah diekspresikan dengan lebih rendah pada bebiri dan diturun regulasi di bawa pemakanan yang ditambah dengan minyak, lalu menjawab kepada kandungan MUFA yang lebih rendah pada bebiri dan juga kumpulan pemakanan yang ditambah minyak. Di samping itu, peroxisome proliferator-activated receptor alpha (PPARa), yang merupakan factor transkripsi untuk pelbagai gen penghasilan lemak, adalah berkadar songsang dengan ACACA dan FASN gen. Maka, penuranan regulasi pada PPARα adalah diikuti dengan kenaikan regulasi gen ACACA dan FASN pada bebiri dan sebaliknya pada kambing; lalu menjawab kepada perbezaan penghasilan SFA antara spesis binatang.

Hasil kajian tesis ini mencadangkan bahawa pemakanan tinggi konsentrat ditambah dengan PUFA (minyak biji rami pada kadar 4%) adalah berguna untuk menambahbaik kadar pertumbahan ruminan kecil, dan untuk menghasilan daging yang lebih sihat dengan profil asid lemak yang bermanfaat, tanpa kesan buruk terhadap penapaian rumen ataupun sifat-sifat kualiti daging. Perbezaan spesis binatang dalam profil asid lemak adalah dipengaruhi secara separa oleh tahap ekspresi gen. Maka dengan itu, manipulasi pemakanan terhadap ekspresi gen berkaitan penghasilan lemak (nutrigenomik) boleh dijadikan alat yang berguna terutamanya dalam menurunkan kandungan SFA, untuk menghasilkan daging dan produk daging yang lebih sihat.

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I certify that a Thesis Examination Committee has met on 8 December 2016 to conduct the final examination of Candyrine Su Chui Len on his thesis entitled "Rumen Fermentation, Fatty Acid Profiles, Meat Quality and Adipogenesis Related Gene Expression of Goat and Sheep Fed with High Concentrated Diet" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

°C	Degrees centigrade
μL	Microliter
μm	Micrometer
a*	Redness
ACACA	Acetyl coenzyme A carboxylase gene
ACC	Acetyl- CoA carboxylase enzyme
ADF	Acid detergent fiber
ADG	Average daily gain
b*	Yellowness
B. fibrisolvens	Butyrivibrio fibrisolvens
BW	Body weight
BWG	Body weight gain
CLA	Conjugated linoleic acids
cm	Centimeter
CP	Crude protein
DM	Dry matter
F. succinogenes	Fibrobacter succinogenes
FAS	Fatty acid synthase enzyme
FASN	Fatty acid synthase gene
FE	Feed efficiency
g	Gram
g	Relative centrifugal force
GAPDH	Glyceraldehyde-3-Phosphate Dehydrogenase
h	Hour
IVOMD	In vitro organic matter digestibility
Kg	Kilogram
L	Liter
L*	Lightness
LD	Longissimus dorsi
m	Meter
MDA	Malondialdehyde
Messenger RNA	mRNA
Mg	Milligram
min	Minutes
MJ	Megajoules
mL	Milliliter
mm	Millimeter
mM	millimolar
mmol	Millimole
MUFA	Mono-unsaturated fatty acids
NDF	Neutral detergent fiber
OM	Organic matter
Р	Probability
PPAR	Peroxisome proliferator-activated receptor
PUFA	Polyunsaturated fatty acids
R. albus	Ruminococcus albus
R. flavefaciens	Ruminococcus flavefaciens
S	Second

SCDStearoyl coenzyme A desaturaseSDStandard deviationSEMStandard error of meanSFASaturated fatty acidsSpSpeciesTBARSThiobarbituric acid reactive substancesUFAUnsaturated fatty acidsVFAVolatile fatty acidsWHCWater holding capacity	SC	Subcutaneous
SDStandard deviationSEMStandard error of meanSFASaturated fatty acidsSpSpeciesTBARSThiobarbituric acid reactive substancesUFAUnsaturated fatty acidsVFAVolatile fatty acidsWHCWater holding capacity	SCD	Stearoyl coenzyme A desaturase
SEMStandard error of meanSFASaturated fatty acidsSpSpeciesTBARSThiobarbituric acid reactive substancesUFAUnsaturated fatty acidsVFAVolatile fatty acidsWHCWater holding capacity	SD	Standard deviation
SFASaturated fatty acidsSpSpeciesTBARSThiobarbituric acid reactive substancesUFAUnsaturated fatty acidsVFAVolatile fatty acidsWHCWater holding capacity	SEM	Standard error of mean
SpSpeciesTBARSThiobarbituric acid reactive substancesUFAUnsaturated fatty acidsVFAVolatile fatty acidsWHCWater holding capacity	SFA	Saturated fatty acids
TBARSThiobarbituric acid reactive substancesUFAUnsaturated fatty acidsVFAVolatile fatty acidsWHCWater holding capacity	Sp	Species
UFAUnsaturated fatty acidsVFAVolatile fatty acidsWHCWater holding capacity	TBARS	Thiobarbituric acid reactive substances
VFA Volatile fatty acids WHC Water holding capacity	UFA	Unsaturated fatty acids
WHC Water holding capacity	VFA	Volatile fatty acids
ö 1 y	WHC	Water holding capacity



CHAPTER 1

INTRODUCTION

Half of the world two billions goats and sheep are found in the Asia-Pacific region, making it the most populated region for goats and sheep in the world (Food and Agriculture Organization, 2014). Goats and sheep farming have been integral parts of agriculture system, contributing to the economy of many developing countries in Asia, including Malaysia. The interest in keeping these animals has increased significantly in the recent years because of their smaller body size and ability to adapt to different harsh environments such as poor quality feed and hot and humid environment compared to cattle and buffaloes. Goats and sheep also have shorter reproductive cycles compared to cattle and buffaloes, making them more reproductively efficient. The Malaysian government also intends to increase the SSL for mutton and chevon to 27.3% through driving modernization in agro-food production by year 2020 (http://www.epu.gov.my). At the moment, the self-sufficiency level (SSL) of mutton and goat meat (chevon) in Malaysia is 11.46% (Table 1.1), lagging far behind compared to poultry meat and equa which are fully self-sufficient in this country (Department of Veterinary Services, 2015), thus the above scenario provides huge opportunities for growth and development of the small ruminant industry in Malaysia.

	Buffalo	Cattle	Swine	Poultry	Goat	Sheep
Population (heads)	121,504	752,032	1,828,860	N.A.	439,667	140,049
	Beef	Pork	Poult	ry meat	Mutton/	chevon
Outputs (Metric ton)	50,493	215,760	0 1,613	.92 X10 ³	4,367.3	
Consumptions (Metric ton)	214,866	228,030	0 1,544	.78 X10 ³	38,107.	3
SSL (%)	23.5	94.62	104.4	8	11.46	

Table	1.1	1	Livestock	population,	outputs,	consumptions	and	self-			
sufficiency level (SSL, %) of livestock products in Malaysia											

N.A. = not available

(Source: Department of Veterinary Services, 2015)



The traditional goats and sheep farming system involved animals feeding and grazing on low quality forages with minimal or no feed supplementations (Khan et al., 2014). This low-input low-output system hinders growth, productivity and development of the industry which could not meet the increasing demand for meat and milk from these two small ruminant species. Thus, there is a need to increase production of animal proteins from goats and sheep in this region through various efforts including improved nutrition and feeding management (Aziz, 2010). Morand-Fehr and Sauvant (1987) suggested the incorporation of concentrate in the diets of ruminants to supply

higher energy, proteins, vitamins and minerals to improve feed utilization. Hence, the inclusion of higher proportion of concentrate in the diet can be an alternative in improving the productivity of the animals.

In the era of the 21st century, as the standard of living improves, consumers are demanding for healthy and nutritious food. Ruminant meat products (chevon and mutton) are known to contain high amount of fat, especially saturated fatty acids (SFA) which are often associated with modern-day diseases such as cardiovascular problem, obesity and cancer (Turner, 2010), making it less preferred by the consumers. However, despite containing high amount of fat, red meat is a good dietary source of protein, nutritionally dense with essential amino acids, micro-minerals and vitamins (Williamson et al., 2005; Biesalski, 2005). A recent study found that red meat is a good source of anti-inflammatory long chain n-3 polyunsaturated fatty acids (PUFA) and conjugated linoleic acid (CLA) (McAfee et al., 2010). The latter has received increasing attentions due to its potential anti-carcinogenic and anti-atherogenic properties (Lock et al., 2005, Hargrave-Barnes et al., 2008), as well as its ability to favorably modulate immune function (Tricon et al., 2004).

The use of various feed supplements in order to enhance the quality of meat products especially in improving the fatty acids profile in red meats have been studied. Alpha-linolenic acid (C18:3n-3), is an n-3 PUFA which has long been reported to be associated with reduced risk of cardiovascular diseases (Hu et al., 1999). It is also the precursor of other long chain n-3 PUFA such as eicosapentaenoic (EPA) and docosahexanoic acid (DHA). These long chain n-3 PUFA are widely known for their association with heart health, beneficial effects towards the central nervous system, retinal function and inflammatory response (Ruxton et al., 2004; Mann et al., 2006, Siddiqui et al., 2008). Linseed oil is a potential commercial source of n-3 PUFA, of which it contained more than 50% of alpha-linolenic acid (Matthews et al., 2000). The supplementation of linseed has been found to be successful in increasing the n-3 PUFA content in broiler chickens (Lopez- Ferrer et al., 2001), pigs (Fontanillas et al., 1997; Rey et al., 2001), lambs (Bessa et al., 2007; Jeronimo et al., 2009), and meat and dairy goats (Marin et al., 2012; Ebrahimi et al., 2013)..

To date, there were only handful of species comparative studies between goats and sheep although interspecies variations in rumen digestion and microbiota have been reported previously (Moon et al., 2010; Lee et al., 2012). The knowledge of putative species-specific differences in fatty acids rise only from indirect comparisons (Loor et al., 2004; Boeckaert et al., 2008a; Toral et al., 2016), due to the scarcity of available data on this subject, particularly in the caprine (Chilliard et al., 2014; Li et al., 2014), most probably because goat is deemed inferior as compared to sheep. In addition, studies on fatty acids profile in goat and sheep has been focusing more on milk and milk products and less attention has been given to meat. Tsiplakou

and Zervas (2008) reported the species differences in fatty acids profile in milk fat of goat and sheep.

Since goats and sheep are economically valuable in the developing countries across Asia including Malaysia, it is relevant that research should focus on achieving better productivity from goats and sheep. This thesis aims at achieving the above with the following hypotheses and objectives:

Hypotheses statements:

- 1. There are species differences in rumen fermentation, growth, fatty acids profiles and meat quality between goat and sheep under similar high concentrate feeding.
- 2. Linseed oil supplementation can improve the fatty acids profile in various tissue in goat and sheep especially in the n-3 PUFA content.
- 3. Differences in the expression of adipogenesis related genes and rumen fermentation characteristics leads to different fatty acids profile between species and between dietary treatments.

Objectives:

- 1. to determine the *in vitro* rumen fermentation characteristics in goat and sheep incubated with control and linseed oil added substrate;
- 2. to determine the growth performance, apparent digestibility and meat quality in goat and sheep fed similar diet;
- 3. to determine the fatty acids profile in liver, muscle and fat tissues, *in vivo* rumen fermentation and the expression of adipogenesis related genes in goat and sheep, under the same high concentrate feeding, with and without supplementation of linseed oil.

REFERENCES

- Abidi, S., Salem, H.B., Vasta, V., and Priolo, A. 2009. Supplementation with barley or spineless cactus (*Opuntia ficus indica* f. *inermis*) cladodes on digestion, growth and intramuscular fatty acid composition in sheep and goats receiving oaten hay. *Small Ruminant Research*. 87(1): 9-16.
- Aboud, A.A.O., Owen, E., Reed, J.D., Said, A.N., McAllan, A. 1993. Feeding sorghum stover to Ethiopian sheep and goats; effects of amount offered on intake, selection and performance. In *Animal Production in Developing Countries*. Eds. Gill, M., Owen, E., Pallot, G.E., Lawrence, T.L.J. pp 202-203. Occasional Publication No. 16 - British Society of Animal Production.
- Abubakr, A., Alimon, A.R., Yaakub, H., Abdullah, N. and Ivan, M. 2015. Effect of Feeding Palm Oil By-Products Based Diets on Muscle Fatty Acid Composition in Goats. *PloS One*. 10(3): p.e0119756.
- Abuelfatah, K., Zuki, A.B.Z., Goh, Y.M. and Sazili, A.Q. 2016. Effects of enriching goat meat with n-3 polyunsaturated fatty acids on meat quality and stability. *Small Ruminant Research*. 136: 36-42.
- Adeyemi, K.D., Sabow, A.B., Shittu, R.M., Karim, R. and Sazili, A.Q. 2015. Influence of dietary canola oil and palm oil blend and refrigerated storage on fatty acids, myofibrillar proteins, chemical composition, antioxidant profile and quality attributes of semimembranosus muscle in goats. *Journal of Animal Science and Biotechnology*. 6(1): 51.
- Adeyemi, K.D., Sabow, A.B., Shittu, R.M., Karim, R., Karsani, S.A. and Sazili, A.Q. 2016. Impact of chill storage on antioxidant status, lipid and protein oxidation, color, drip loss and fatty acids of semimembranosus muscle in goats. *CyTA-Journal of Food*. 14(3): 405-414.
- Aghwan, Z.A., Alimon, A.R., Goh, Y.M., Nakyinsige, K. and Sazili, A.Q. 2014. Fatty acid profiles of *supraspinatus*, *longissimus lumborum* and *semitendinosus* muscles and serum in kacang goats supplemented with inorganic selenium and iodine. *Asian-Australasian Journal of Animal Sciences*. 27(4): 543.
- Agrawal, A.R., Karim, S.A., Kumar, R., Sahoo, A. and John, P.J. 2014. Sheep and goat production: basic differences, impact on climate and molecular tools for rumen microbiome study. *International Journal of Current Microbiology and Applied Sciences*. 3(1): 684-706.
- Ahn, D.U. and Nam, K.C. 2004. Effects of ascorbic acid and antioxidants on color, lipid oxidation and volatiles of irradiated ground beef. *Radiation Physics and Chemistry*. 71(1): 151-156.

- Alam, M.B., Popp,i D.P. and Sykes, A.R. 1985. Comparative intake of digestible organic matter and water by sheep and goats. *Proceedings New Zealand Society Animal Production.* 45: 107-111.
- Alcaide, E.M., García, A.M. and Aguilera, J.F. 2000. A comparative study of nutrient digestibility, kinetics of degradation and passage and rumen fermentation pattern in goats and sheep offered good quality diets. *Livestock Production Science*. 64(2): 215-223.
- Ammar, H., López, S., Andrés, S., Ranilla, M.J., Bodas, R. and González, J.S. 2008. *In vitro* digestibility and fermentation kinetics of some browse plants using sheep or goat ruminal fluid as the source of inoculum. *Animal Feed Science and Technology*. 147(1): 90-104.
- Ammar, H., Ranilla, M.J., Tejido, M.L., Ovejero, F.J., Gonzalez, J.S. and Lopez, S. 2004. Effect of inoculum source (sheep or goat rumen fluid) on *in vitro* digestibility and gas production kinetics of the foliage of some Spanish browse plants. *Options Mediterraneennes*. Serie A, Seminaires Mediterraneenes. Centre International de Hautes Etudes Agronomiques Mediterraneens, Montpellier, France. 59: 121-126.
- AMSA (American Meat Science Association). 1991. Guidelines for meat color evaluation. *Proceedings of the 4th Reciprocal Meat Conference* Chicago, IL: National Live Stock and Meat Board. pp. 1–17.
- AOAC. 1999. Official Methods of Analysis of the Association of Official Agricultural Chemists, 16th ed. AOAC International, Arlington, VA, USA.
- Atkinson, R.L., Scholljegerdes, E.J., Lake, S.L., Nayigihugu, V., Hess, B.W. and Rule, D.C. 2006. Site and extent of digestion, duodenal flow, and intestinal disappearance of total and esterified fatty acids in sheep fed a high-concentrate diet supplemented with high-linoleate safflower oil. *Journal of Animal Science*. 84(2): 387-396.
- Aziz, M.A. 2010. Present status of the world goat populations and their productivity. *World.* 861(1078.2): 1.
- Babiker, S.A., El Khider, I.A. and Shafie, S.A. 1990. Chemical composition and quality attributes of goat meat and lamb. *Meat Science*. 28(4): 273-277.
- Bahtti, A. 2013. Feeding Behavior, Voluntary Intake and Digestibility of Various Summer Fodders in Sheep and Goats. *Pakistan Journal of Zoology*. 45(1): 53-58.
- Banskalieva, V., Sahlu, T. and Goetsch, A.L. 2000. Fatty acid composition of goat muscles and fat depots: a review. *Small Ruminant Research*. 37(3): 255-268.
- Barber, M.C. and Travers, M.T. 1995. Cloning and characterisation of multiple acetyl-CoA carboxylase transcripts in ovine adipose tissue. *Gene*. 154(2): 271-275.

- Barber, M.C., Price, N.T. and Travers, M.T. 2005. Structure and regulation of acetyl-CoA carboxylase genes of metazoa. *Biochimica et Biophysica Acta (BBA)-Molecular and Cell Biology of Lipids*. 1733(1): 1-28.
- Barber, M.C., Ward, R.J., Richards, S.E., Salter, A.M., Buttery, P.J., Vernon, R.G. and Travers, M.T. 2000. Ovine adipose tissue monounsaturated fat content is correlated to depot-specific expression of the stearoyl-CoA desaturase gene. *Journal of Animal Science*. 78(1): 62-68.
- Bas, P. and Morand-Fehr, P. 2000. Effect of nutritional factors on fatty acid composition of lamb fat deposits. *Livestock Production Science*. 64(1): 61-79.
- Bauman, D.E., Corl, B.A., Baumgard, L.H., Griinari, J.M., Garnsworthy, P.C. and Wiseman, J. 2001. Conjugated linoleic acid (CLA) and the dairy cow. In *Recent Advances in Animal Nutrition*. Eds. Garnsworthy, P.C. and Wiseman, J. pp. 221-250. Nottingham University Press, Nottingham, UK.
- Bauman, D.E., Perfield, J.W., De Veth, M.J. and Lock, A.L., 2003. New perspectives on lipid digestion and metabolism in ruminants. *Proceedings Cornell Nutrition Conference*. 65: 175-189.
- Beam, T.M., Jenkins, T.C., Moate, P.J., Kohn, R.A. and Palmquist, D.L. 2000. Effects of amount and source of fat on the rates of lipolysis and biohydrogenation of fatty acids in ruminal contents. *Journal of Dairy Science*. 83(11): 2564-2573.
- Belury, M.A. 2002. Inhibition of carcinogenesis by conjugated linoleic acid: potential mechanisms of action. *The Journal of Nutrition*. 132(10): 2995-2998.
- Berg, J.M., Tymoczko, J.L. and Stryer, L. 2002. *Biochemistry, 5th edition*. WH Freeman and Co. New York.
- Bernard, L., Bonnet, M., Leroux, C., Shingfield, K.J. and Chilliard, Y. 2009. Effect of sunflower-seed oil and linseed oil on tissue lipid metabolism, gene expression, and milk fatty acid secretion in alpine goats fed maize silage–based diets. *Journal of Dairy Science*. 92(12): 6083-6094.
- Bernard, L., Leroux, C. and Chilliard, Y. 2006. Characterisation and nutritional regulation of the main lipogenic genes in the ruminant lactating mammary gland. *Proceedings of the 10th International Symposium, Ruminant Physiology: Digestion, Metabolism and Impact of Nutrition on Gene Expression, Immunology and Stress.* August 30 – September 4, Copenhagen, Denmark. pp.295-326.
- Bernes, G., Turner, T., and Pickova, J. 2012. Sheep fed only silage or silage supplemented with concentrates: 2. Effects on lamb performance and fatty acid profile of ewe milk and lamb meat. *Small Ruminant Research*. 102(2): 114-124.

- Bessa, R.J., Alves, S.P., Jerónimo, E., Alfaia, C.M., Prates, J.A. and Santos-Silva, J. 2007. Effect of lipid supplements on ruminal biohydrogenation intermediates and muscle fatty acids in lambs. *European Journal of Lipid Science and Technology*. 109(8): 868-878.
- Bhatt, R.S., Karim, S.A., Sahoo, A. and Shinde, A.K. 2013. Growth performance of lambs fed diet supplemented with rice bran oil as such or as calcium soap. *Asian-Australasian Journal of Animal Sciences*. 26(6): 812.
- Bhatt, R.S., Soren, N.M., Tripathi, M.K. and Karim, S.A. 2011. Effects of different levels of coconut oil supplementation on performance, digestibility, rumen fermentation and carcass traits of Malpura lambs. *Animal Feed Science and Technology.* 164(1): 29-37.
- Bhatta, R., Tajima, K., Takusari, N., Higuchi, K., Enishi, O. and Kurihara, M. 2007. Comparison of *in vivo* and *in vitro* techniques for methane production from ruminant diets. *Asian Australasian Journal of Animal Sciences*. 20(7): 1049.
- Biesalski, H.K. 2005. Meat as a component of a healthy diet–are there any risks or benefits if meat is avoided in the diet?. *Meat Science*. 70(3): 509-524.
- Bionaz, M., Chen, S., Khan, M.J. and Loor, J.J. 2013. Functional role of PPARs in ruminants: Potential targets for fine-tuning metabolism during growth and lactation. *PPAR Research*. 2013: 684159.
- Blummel, M. and Ørskov, E.R. 1993. Comparison of *in vitro* gas production and nylon bag degradability of roughages in predicting feed intake in cattle. *Animal Feed Science and Technology*. 40(2): 109-119.
- Boeckaert, C., Vlaeminck, B., Dijkstra, J., Issa-Zacharia, A., Van Nespen, T., Van Straalen, W. and Fievez, V. 2008a. Effect of dietary starch or micro algae supplementation on rumen fermentation and milk fatty acid composition of dairy cows. *Journal of Dairy Science*. 91(12): 4714-4727.
- Boeckaert, C., Vlaeminck, B., Fievez, V., Maignien, L., Dijkstra, J. and Boon, N. 2008b. Accumulation of *trans* C18:1 fatty acids in the rumen after dietary algal supplementation is associated with changes in the *Butyrivibrio* community. *Applied Environmental Microbiology*. 74: 6923– 6930.
- Boles, J.A. and Pegg, R. 2010. Meat color. Montana State University and Saskatchewan Food Product Innovation, Program University of Saskatchewan. Retrieved 30 September 2015 from <u>http://safespectrum.com/pdfs/meatcolor.pdf</u>

- Bond, J.J. and Warner, R.D. 2007. Ion distribution and protein proteolysis affect water holding capacity of *Longissimus thoracis* et lumborum in meat of lamb subjected to antemortem exercise. *Meat Science*. 75(3): 406-414.
- Boots, B., Lillis, L., Clipson, N., Petrie, K., Kenny, D.A., Boland, T.M. and Doyle, E. 2013. Responses of anaerobic rumen fungal diversity (*phylum Neocallimastigomycota*) to changes in bovine diet. *Journal of Applied Microbiology*. 114(3): 626-635.
- Bouattour, M.A., Casals, R., Albanell, E., Such, X. and Caja, G. 2008. Feeding soybean oil to dairy goats increases conjugated linoleic acid in milk. *Journal of Dairy Science*. 91(6): 2399-2407.
- Brown, J.D. and Plutzky, J. 2007. Peroxisome proliferator–activated receptors as transcriptional nodal points and therapeutic targets. *Circulation*. 115(4): 518-533.
- Bu, D.P., Wang, J.Q., Dhiman, T.R. and Liu, S.J. 2007. Effectiveness of oils rich in linoleic and linolenic acids to enhance conjugated linoleic acid in milk from dairy cows. *Journal of Dairy Science*. 90(2): 998-1007.
- Burns, T., 2011. Fatty acids and lipogenesis in ruminant adipocytes. PhD thesis. Clemson University, South Carolina, USA.
- Caneque, V., Díaz, M. T., Alvarez, I., Lauzurica, S., Pérez, C., and De la Fuente, J. 2005. The influences of carcass weight and depot on the fatty acid composition of fats of suckling Manchego lambs. *Meat Science*. 70(2): 373-379.
- Cantalapiedra-Hijar, G., Yáñez-Ruiz, D.R., Martín-García, A.I. and Molina-Alcaide, E. 2009. Effects of forage: concentrate ratio and forage type on apparent digestibility, ruminal fermentation, and microbial growth in goats. *Journal of Animal Science*. 87(2): 622-631.
- Carro, M.D., Cantalapiedra-Hijar, G., Ranilla, M.J. and Molina-Alcaide, E. 2012. Urinary excretion of purine derivatives, microbial protein synthesis, nitrogen use, and ruminal fermentation in sheep and goats fed diets of different quality. *Journal of Animal Science*. 90(11): 3963-3972.
- Carro, M.D., Lopez, S., Gonzalez, J.S. and Ovejero, F.J. 1994. Comparison of laboratory methods for predicting digestibility of hay in sheep. *Small Ruminant Research*. 14(1): 9-17.
- Castro, T., Manso, T., Mantecón, A.R., Guirao, J. and Jimeno, V. 2005. Fatty acid composition and carcass characteristics of growing lambs fed diets containing palm oil supplements. *Meat Science*. 69(4): 757-764.
- Cetin, O., Bingol, E.B., Colak, H. and Hampikyan, H. 2012. Effects of electrical stimulation on meat quality of lamb and goat meat. *The Scientific World Journal*. 2012: 574202

- Chalupa, W., Galligan, D.T. and Ferguson, J.D. 1996. Animal nutrition and management in the 21st century: dairy cattle. *Animal Feed Science and Technology*. 58(1-2): 1-18.
- Cheeke, P.R. and Dierenfeld, E.S. 2010. *Comparative Animal Nutrition and Metabolism*. Cambridge University Press, Cambridge, UK.
- Chesworth, J.M., Stuchbury, T. and Scaife, J.R. 1998. An Introduction to Agricultural Biochemistry. Chapman and Hall, London, UK.
- Chikunya, S., Demirel, G., Enser, M., Wood, J.D., Wilkinson, R.G. and Sinclair, L.A. 2004. Biohydrogenation of dietary n-3 PUFA and stability of ingested vitamin E in the rumen, and their effects on microbial activity in sheep. *British Journal of Nutrition*. 91(04): 539-550.
- Chilliard, Y., Ferlay, A., Mansbridge, R.M. and Doreau, M. 2000. Ruminant milk fat plasticity: nutritional control of saturated, polyunsaturated, *trans* and conjugated fatty acids. *Annales de Zootechnie*. 49(3): 181-205.
- Chilliard, Y., Toral, P.G., Shingfield, K.J., Rouel, J., Leroux, C. and Bernard, L. 2014. Effects of diet and physiological factors on milk fat synthesis, milk fat composition and lipolysis in the goat: a short review. *Small Ruminant Research*. 122(1): 31-37.
- Christie, W. W. 1982. Lipid Analysis. Pergamon Press, New York.
- Cieslak, A., Soliva, C.R., Potkański, A., Szumacher-Strabel, M., Scheeder, M.R.L. and Machmüller, A. 2006. Effect of plant oils on methane emission and biohydrogenation *in vitro*. *International Congress Series*. 1293: 180-183.
- Clarke, S.D. and Jump, D.B. 1994. Dietary polyunsaturated fatty acid regulation of gene transcription. *Annual Review of Nutrition*. 14(1): 83-98.
- Coakley, M., Johnson, M.C., McGrath, E., Rahman, S., Ross, R.P., Fitzgerald, G.F., Devery, R. and Stanton, C. 2006. Intestinal bifidobacteria that produce *trans*-9, *trans*-11 conjugated linoleic acid: a fatty acid with antiproliferative activity against human colon SW480 and HT-29 cancer cells. *Nutrition and Cancer*. 56(1): 95-102.
- Cone, J.W., Van Gelder, A.H., Visscher, G.J. and Oudshoorn, L. 1996. Influence of rumen fluid and substrate concentration on fermentation kinetics measured with a fully automated time related gas production apparatus. *Animal Feed Science and Technology*. 61(1): 113-128.
- Costa, R.G., Santos, N.M.D., Queiroga, R.D.C.R.D., Sousa, W.H.D., Madruga, M.S. and Cartaxo, F.Q. 2015. Physicochemical characteristics and fatty acid profile of meat from lambs with different genotypes and diets. *Revista Brasileira de Zootecnia*. 44(7): 248-254.

- Crespo, N. and Esteve-Garcia, E. 2002. Dietary linseed oil produces lower abdominal fat deposition but higher de novo fatty acid synthesis in broiler chickens. *Poultry Science*. 81(10): 1555-1562.
- Czerkawski, J.W. and Breckenridge, G. 1977. Design and development of a long-term rumen simulation technique (Rusitec). *British Journal of Nutrition*. 38(03): 371-384.
- Daniel, Z.C.T.R., Wynn, R.J., Salter, A.M. and Buttery, P.J. 2004. Differing effects of forage and concentrate diets on the oleic acid and conjugated linoleic acid content of sheep tissues: The role of stearoyl-CoA desaturase. *Journal of Animal Science*. 82(3): 747-758.
- Degen, A.A., Kam, M., Pandey, S.B., Upreti, C.R., El-Meccawi, S. and Osti, N.P. 2010. *In vitro* gas production of leaves from fodder trees and shrubs from mid-hills of Nepal using cow, sheep and goat rumen liquor. *Journal of Agricultural Science*. 148(04): 445-451.
- Delerive, P., Fruchart, J.C. and Staels, B. 2001. Peroxisome proliferatoractivated receptors in inflammation control. *Journal of Endocrinology*. 169(3): 453-459.
- Delles, R.M., Xiong, Y.L., True, A.D., Ao, T. and Dawson, K.A. 2014. Dietary antioxidant supplementation enhances lipid and protein oxidative stability of chicken broiler meat through promotion of antioxidant enzyme activity. *Poultry Science*. 93(6): 1561-1570.
- Delmotte, C., Rondia, P., Raes, K., Dehareng, F. and Decruyenaere, V. 2005. Omega 3 and CLA naturally enhanced levels of animal products: Effects of grass and linseed supplementation on the fatty acid composition of lamb meat and sheep milk. In '11th Seminar of the FAO-CIHEAM sub-network on sheep and goat nutrition advanced nutrition and feeding strategies to improve sheep and goat production'. Catania. 24.
- Den Hertog-Meischke, M.J.A., Van Laack, R.J.L.M. and Smulders, F.J.M. 1997. The water-holding capacity of fresh meat. *Veterinary Quarterly*. 19(4): 175-181.
- Denke, M.A. and Grundy, S.M. 1992. Comparison of effects of lauric acid and palmitic acid on plasma lipids and lipoproteins. *The American Journal of Clinical Nutrition*. 56(5): 895-898.
- Department of Health UK. 1994. Nutritional Aspects of Cardiovascular Disease. *Report of the Health and Social Subject*. vol 46. London Department of Health of the United Kingdom. Her Majesty's Stationery Office. Retrieved 2 February 2016 from <u>http://www.actiononsalt.org.uk/salthealth/Recommendations%20on%20</u> <u>salt/42491.pdf</u>

Department of Veterinary Services, 2015. Retrieved 5 February 2016 from http://www.dvs.gov.my/index.php/pages/view/1498

http://www.dvs.gov.my/dvs/resources/user 1/DVS%20pdf/Perangkaan %202014-2015/2014 2015/kadar sara diri Muka Surat 1-15.pdf

http://www.dvs.gov.my/dvs/resources/user_1/DVS%20pdf/Perangkaan %202014-2015/2014_2015/bil_ternakan_201402015Muka_Surat_1-15.pdf

http://www.dvs.gov.my/dvs/resources/user_1/DVS%20pdf/Perangkaan %202014-2015/2014_2015/Dengeluran_besilen_ternekenMuke_Suret_1_15_pdf

2015/2014_2015/Pengeluran_hasilan_ternakanMuka_Surat_1-15.pdf

http://www.dvs.gov.my/dvs/resources/user 1/DVS%20pdf/Perangkaan %202014-2015/2014 2015/Penggunaan hasilan ternkan .Muka Surat 1-15 .pdf

- Derr, J., Kris-Etherton, P.M., Pearson, T.A. and Seligson, F.H. 1993. The role of fatty acid saturation on plasma lipids, lipoproteins, and apolipoproteins: II. The plasma total and low-density lipoprotein cholesterol response of individual fatty acids. *Metabolism*. 42(1): 130-134.
- Dervishi, E., Serrano, C., Joy, M., Serrano, M., Rodellar, C. and Calvo, J.H. 2011. The effect of feeding system in the expression of genes related with fat metabolism in semitendinous muscle in sheep. *Meat Science*. 89(1): 91-97.
- Dervishi, E., Serrano, C., Joy, M., Serrano, M., Rodellar, C. and Calvo, J.H. 2010. Effect of the feeding system on the fatty acid composition, expression of the δ 9-desaturase, peroxisome proliferator-activated receptor alpha, gamma, and sterol regulatory element binding protein 1 genes in the semitendinous muscle of light lambs of the rasa aragonesa breed. *BMC Veterinary Research*. 6(1): 1.
- Destaillats, F., Trottier, J.P., Galvez, J.G. and Angers, P. 2005. Analysis of αlinolenic acid biohydrogenation intermediates in milk fat with emphasis on conjugated linolenic acids. *Journal of Dairy Science*. 88(9): 3231-3239.
- Devendra, C. 2005. Small ruminants in Asia; Contribution to food security, poverty lleviation and opportunities for productivity enhancement. *Proceedings of International Workshop on Small Ruminant Production and Development in South East Asia.*' MEKARN, Nong Lam, HCMC, Vietnam. pp. 19-32.
- Devendra, C. and Liang, J.B. 2012. Conference summary of dairy goats in Asia: current status, multifunctional contribution to food security and potential improvements. *Small Ruminant Research.* 108: 1–11.

- Devendra, C., 2010. Small farms in Asia: revitalising agricultural production, food security and rural prosperity. *Academy of Sciences Malaysia*. Kuala Lumpur, Malaysia. pp. xiii&175.
- Devendra. C. 1978. The digestive efficiency of goats. *World Review of Animal Production*. 14: 9-22.
- Devillard, E., McIntosh, F.M., Newbold, C.J. and Wallace, R.J. 2006. Rumen ciliate protozoa contain high concentrations of conjugated linoleic acids and vaccenic acid, yet do not hydrogenate linoleic acid or desaturate stearic acid. *British Journal of Nutrition*. 96(04): 697-704.
- Diaz, M.T., Alvarez, I., De la Fuente, J., Sañudo, C., Campo, M.M., Oliver, M.A., i Furnols, M.F., Montossi, F., San Julian, R., Nute, G.R. and Caneque, V. 2005. Fatty acid composition of meat from typical lamb production systems of Spain, United Kingdom, Germany and Uruguay. *Meat Science*. 71(2): 256-263.
- Ding, X., Long, R., Zhang, Q., Huang, X., Guo, X. and Mi, J. 2012. Reducing methane emissions and the methanogen population in the rumen of Tibetan sheep by dietary supplementation with coconut oil. *Tropical Animal Health and Production*. 44(7): 1541-1545.
- Dohme, F., Fievez, V., Raes, K. and Demeyer, D.I. 2003. Increasing levels of two different fish oils lower ruminal biohydrogenation of eicosapentaenoic and docosahexaenoic acid *in vitro*. *Animal Research*. 52(4): 309-320.
- Dohme, F., Machmüller, A., Estermann, B.L., Pfister, P., Wasserfallen, A. and Kreuzer, M. 1999. The role of the rumen ciliate protozoa for methane suppression caused by coconut oil. *Letters in Applied Microbiology*. 29(3): 187-192.
- Dohme, F., Machmuller, A., Wasserfallen, A. and Kreuzer, M. 2001. Ruminal methanogenesis as influenced by individual fatty acids supplemented to complete ruminant diets. *Letters in Applied Microbiology*. 32: 47–51.
- Domingue, B.F., Dellow, D.W. and Barry, T.N., 1991. Voluntary intake and rumen digestion of a low-quality roughage by goats and sheep. *The Journal of Agricultural Science*, *117*(01), pp.111-120.
- Donkin, S.S. 2008. Glycerol from biodiesel production: the new corn for dairy cattle. *Revista Brasileira de Zootecnia*. 37: 280-286.
- Doreau, M. and Chilliard, Y. 1997. Digestion and metabolism of dietary fat in farm animals. *British Journal of Nutrition*. 78(01): 15-35.
- Doreau, M., Bauchart, D. and Chilliard, Y. 2011. Enhancing fatty acid composition of milk and meat through animal feeding. *Animal Production Science*. 51(1): 19-29.

- Dutta, T.K., Agnihotri, M.K. and Rao, S.B.N. 2008. Effect of supplemental palm oil on nutrient utilization, feeding economics and carcass characteristics in post-weaned Muzafarnagari lambs under feedlot condition. *Small Ruminant Research*. 78(1): 66-73.
- Ebrahimi, M., Rajion, M.A., Goh, Y.M. and Sazili, A.Q. 2012. Impact of different inclusion levels of oil palm (*Elaeis guineensis Jacq.*) fronds on fatty acid profiles of goat muscles. *Journal of Animal Physiology and Animal Nutrition*. 96(6): 962-969.
- Ebrahimi, M., Rajion, M.A., Goh, Y.M., Sazili, A.Q. and Schonewille, J.T. 2013. Effect of linseed oil dietary supplementation on fatty acid composition and gene expression in adipose tissue of growing goats. *BioMed Research International*. 2013: 1-11.
- Ebrahimi, M., Rajion, M.A., Meng, G.Y., Farjam, A.S., Oskoueian, E. and Jafari, S. 2015. Diet high in α-linolenic acid up-regulate PPAR-α gene expression in the liver of goats. *Electronic Journal of Biotechnology*. 18(3): 210-214.
- Eeckhoute, J., Oger, F., Staels, B. and Lefebvre, P. 2012. Coordinated Regulation of PPARγ Expression and Activity through Control of Chromatin Structure in Adipogenesis and Obesity. *PPAR Research*. 2012: 164140
- Elmore, J.S., Cooper, S.L., Enser, M., Mottram, D.S., Sinclair, L.A., Wilkinson, R.G. and Wood, J.D. 2005. Dietary manipulation of fatty acid composition in lamb meat and its effect on the volatile aroma compounds of grilled lamb. *Meat Science*. 69(2): 233-242.
- Engstrom, D.F., Goonewardene, L.A., Grimson, R.E., McKinnon, J.J., Stilborn, R.P. and Volek, R. 1994. Effects of feeding added protein and fat on feedlot performance and carcass quality in large frame steers. *Canadian Journal of Animal Science*. 74: 547–549.
- Estany, J., Ros-Freixedes, R., Tor, M. and Pena, R.N. 2014. A functional variant in the stearoyl-CoA desaturase gene promoter enhances fatty acid desaturation in pork. *PLoS One*. 9(1): p.e86177.
- Faustman, C. and Cassens, R.G. 1990. The biochemical basis for discoloration in fresh meat: a review. *Journal of Muscle Foods*. 1(3): 217-243.
- Fiorentini, G., Messana, J.D., Dian, P.H.M., Reis, R.A., Canesin, R.C., Pires, A.V. and Berchielli, T.T. 2013. Digestibility, fermentation and rumen microbiota of crossbred heifers fed diets with different soybean oil availabilities in the rumen. *Animal Feed Science and Technology*. 181(1): 26-34.
- Fliegerova, K., Kaerger, K., Kirk, P. and Voigt, K. 2015. Rumen fungi. In *Rumen Microbiology: From Evolution to Revolution*. Eds. Puniya, A.K., Singh, R. and Kamra, D.N. pp. 97-112. Springer India.

- Folch, J., Lees, M. and Sloane-Stanley, G.H. 1957. A simple method for the isolation and purification of total lipids from animal tissues. *Journal of Biological Chemistry*. 226(1): 497-509.
- Fontanillas, R., Barroeta, A., Baucells, M.D. and Codony, R. 1997. Effect of feeding highly cis-monounsaturated, *trans*, or n-3 fats on lipid composition of muscle and adipose tissue of pigs. *Journal of Agricultural and Food Chemistry*. 45(8): 3070-3075.
- Font-i-Furnols, M. and Guerrero, L. 2014. Consumer preference, behavior and perception about meat and meat products: An overview. *Meat Science*. 98(3): 361-371.
- Food and Agriculture Organization, FAO 2014. *Statistical Yearbook. Asia and the Pacific. Food and Agriculture*. Retrieved 4 January 2016 from <u>http://www.fao.org/3/a-i3590e.pdf</u>
- Galindo, J., González, N., Delgado, D., González, R., Sosa, A., Marrero, Y., Aldana, A.I., Moreira, O., Cairo, J., Torres, V. and Sarduy, L. 2016. Effect of a regulator product of the fermentation with coconut oil on the methanogenic population and other microbial of the sheep rumen Pelibuey. *Cuban Journal of Agricultural Science*. 48(4).
- Gallardo, B., Manca, M. G., Mantecón, A. R., Nudda, A., and Manso, T. 2015. Effects of linseed oil and natural or synthetic vitamin E supplementation in lactating ewes' diets on meat fatty acid profile and lipid oxidation from their milk fed lambs. *Meat Science*. 102: 79-89.
- Getachew, G., Robinson, P.H., DePeters, E.J., Taylor, S.J., Gisi, D.D., Higginbotham, G.E. and Riordan, T.J. 2005. Methane production from commercial dairy rations estimated using an *in vitro* gas technique. *Animal Feed Science and Technology*. 123: 391-402.
- Gihad, E. A. 1976. Intake, digestibility and nitrogen utilization of tropical natural grass hay by goats and sheep. *Journal of Animal Science*. 43(4): 879-883.
- Givens, D.I. 2005. The role of animal nutrition in improving the nutritive value of animal-derived foods in relation to chronic disease. *Proceedings of the Nutrition Society*. 64(03): 395-402.
- Gobert, M., Gruffat, D., Habeanu, M., Parafita, E., Bauchart, D. and Durand, D. 2010. Plant extracts combined with vitamin E in PUFA-rich diets of cull cows protect processed beef against lipid oxidation. *Meat Science*. 85(4): 676-683.
- Gomez-Cortes, P., Frutos, P., Mantecón, A.R., Juárez, M., De la Fuente, M.A. and Hervás, G. 2008. Milk production, conjugated linoleic acid content, and *in vitro* ruminal fermentation in response to high levels of soybean oil in dairy ewe diet. *Journal of Dairy Science*. 91(4): 1560-1569.

- Gordon, G.L. and Phillips, M.W. 1998. The role of anaerobic gut fungi in ruminants. *Nutrition Research Reviews*. 11(01): 133-168.
- Granit, R., Angel, S., Akiri, B., Holzer, Z., Aharoni, Y., Orlov, A. and Kanner, J. 2001. Effects of vitamin E supplementation on lipid peroxidation and color retention of salted calf muscle from a diet rich in polyunsaturated fatty acids. *Journal of Agricultural and Food Chemistry*. 49(12): 5951-5956.
- Griinari, J.M., Corl, B.A., Lacy, S.H., Chouinard, P.Y., Nurmela, K.V.V. and Bauman, D.E. 2000. Conjugated linoleic acid is synthesized endogenously in lactating dairy cows by Δ9-desaturase. *The Journal of Nutrition*. 130(9): 2285-2291.
- Griinari, J. and Bauman, D.E. 1999. Biosynthesis of conjugated linoleic acid and its incorporation into meat and milk in ruminants. In *Advances in Conjugated Linoleic Acid Research*. Eds. Yurawecz, M. P., Mossoba, M. M., Kramer, J.K.G., Pariza, M. W. and Nelson, G. J. 1: 180-200. AOCS Press Champaign, IL.
- Grummer, R.R. 1993. Etiology of lipid-related metabolic disorders in periparturient dairy cows. *Journal of Dairy Science*. 76(12): 3882-3896.
- Gupta, R., Gupta, N. and Rathi, P. 2004. Bacterial lipases: an overview of production, purification and biochemical properties. *Applied Microbiology and Biotechnology*. 64(6): 763-781.
- Gurr, M.I. and James, A.T. 1980. Lipids: what they are and how the biochemist deals with them. In *Lipid Biochemistry: An Introduction.* pp. 1-17. Springer Netherlands.
- Gurr, M.I., Harwood, J.L. and Frayn, K.N. 2002. *Lipid Biochemistry*. Blackwell Science, UK.
- Halver, J.E., 1980. Lipids and fatty acids. In *Fish Feed and Technology, Aquaculture Development and Coordination Programme. ADCP/REP/80/11, FAO/UNDP Training Course in Fish Feed Technology, College of Fisheries.* Ed. Pillay; TVR Washington, Seattle.
- Harfoot, C. G., Noble, R. C. and Moore, J. H. 1973. Factors influencing the extent of biohydrogenation of linoleic acid by rumen micro-organisms *in vitro*. *Journal of the Science of Food and Agriculture*. 24(8): 961-970.
- Harfoot, C.G. and Hazlewood, G.P. 1997. Lipid metabolism in the rumen. In *The Rumen Microbial Ecosystem*. Eds. Hobson, P.N. and Stewart, C.S. pp. 382–426. Chapman & Hall, London, UK.
- Hargrave-Barnes, K.M., Azain, M.J. and Miner, J.L. 2008. Conjugated linoleic acid-induced fat loss dependence on δ 6-desaturase or cyclooxygenase. *Obesity*. 16(10): 2245-2252.

- Heimann, E., Nyman, M. and Degerman, E. 2015. Propionic acid and butyric acid inhibit lipolysis and *de novo* lipogenesis and increase insulinstimulated glucose uptake in primary rat adipocytes. *Adipocyte*. 4(2): 81-88.
- Heinz, G. and Hautzinger, P. 2009. Meat processing technology for small to medium scale producers. FAO Regional Office for Asia and the Pacific (RAP). Bangkok. pp. 103–114.
- Hervas, G., Luna, P., Mantecón, Á.R., Castañares, N., de la Fuente, M.A., Juárez, M. and Frutos, P. 2008. Effect of diet supplementation with sunflower oil on milk production, fatty acid profile and ruminal fermentation in lactating dairy ewes. *Journal of Dairy Research*. 75(04): 399-405.
- Hess, B.W., Moss, G.E. and Rule, D.C. 2008. A decade of developments in the area of fat supplementation research with beef cattle and sheep. *Journal of Animal Science*. 86(14): 188-204.
- Ho, Y.W. and Abdullah, N. 1999. The role of rumen fungi in fibre digestion Review. *Asian-Australasian Journal of Animal Sciences*. 12(1):104-112.
- Ho, Y.W. and Barr, D.J.S. 1995. Classification of anaerobic gut fungi from herbivores with emphasis on rumen fungi from Malaysia. *Mycologia*. 655-677.
- Hodges, D.M., DeLong, J.M., Forney, C.F. and Prange, R.K. 1999. Improving the thiobarbituric acid-reactive-substances assay for estimating lipid peroxidation in plant tissues containing anthocyanin and other interfering compounds. *Planta*. 207(4): 604-611.
- Hofmann, R.R. 1989. Evolutionary steps of ecophysiological adaptation and diversification of ruminants: a comparative view of their digestive system. *Oecologia*. 78(4): 443-457.
- Honikel, K. O. 2004. Conversion of muscle to meat. In *Encyclopedia of Meat Sciences.* Eds. Jensen, W.K.C., Devine, E. and Dikeman, M. pp.314-318. Elsevier Academic Press.
- Honikel, K.O. 1998. Reference methods for the assessment of physical characteristics of meat. *Meat science*. 49(4): 447-457.
- Hook, S. E., Wright, A. D. G.and McBride, B. W. 2010. Methanogens: methane producers of the rumen and mitigation strategies. *Archaea*. 2010: 1–11.
- Horcada, A., Del Mar Campo, M., Polvillo, O., Alcalde, M.J., Cilla, I. and Sañudo, C. 2014. A comparative study of fatty acid profiles of fat in commercial Spanish suckling kids and lambs. *Spanish Journal of Agricultural Research*. 12(2): 427-435.

- Hosseini, A. and Loor, J.J. 2014, May 13. Lipid regulation of gene expression in ruminants. Retrieved 14 February 2016 from http://lipidlibrary.aocs.org/Food/content.cfm?ItemNumber=40752
- Howes, N.L., Bekhit, A.E.D.A., Burritt, D.J. and Campbell, A.W. 2015. Opportunities and implications of pasture-based lamb fattening to enhance the long-chain fatty acid composition in meat. *Comprehensive Reviews in Food Science and Food Safety*. 14(1): 22-36.
- Hristov, A.N., Kennington, L.R., McGuire, M.A. and Hunt, C.W. 2005. Effect of diets containing linoleic acid-or oleic acid-rich oils on ruminal fermentation and nutrient digestibility, and performance and fatty acid composition of adipose and muscle tissues of finishing cattle. *Journal of Animal Science*. 83(6): 1312-1321.
- Hsu, S.C. and Huang, C.J. 2006. Reduced fat mass in rats fed a high oleic acid–rich safflower oil diet is associated with changes in expression of hepatic PPARα and adipose SREBP-1c–regulated genes. *The Journal of Nutrition*. 136(7): 1779-1785.
- Hu, F.B., Stampfer, M.J., Manson, J.E., Rimm, E.B., Wolk, A., Colditz, G.A., Hennekens, C.H. and Willett, W.C. 1999. Dietary intake of α-linolenic acid and risk of fatal ischemic heart disease among women. *The American Journal of Clinical Nutrition*. 69(5): 890-897.
- Huff-Lonergan, E. and Lonergan, S.M. 2005. Mechanisms of water-holding capacity of meat: The role of postmortem biochemical and structural changes. *Meat Science*. 71(1): 194-204.
- Hungate, R.E. 1966. *The Rumen and its Microbes*. Academic Press, New York and London.
- Huss, H.H. 1995. Quality and quality changes in fresh fish. *FAO Fisheries Technical Paper. No.348.* Food and Agriculture Organization of the United Nations (FAO). Rome.
- Isac, M.D., García, M.A., Aguilera, J.F. and Alcaide, E.M. 1994. A comparative study of nutrient digestibility, kinetics of digestion and passage and rumen fermentation pattern in goats and sheep offered medium quality forages at the maintenance level of feeding. *Archives of Animal Nutrition*. 46(1): 37-50.
- Ishler, V.A., Heinrichs, A.J. and Varga, G.B. 1996. *From Feed to Milk: Understanding Rumen Function*. Vol. 422. Pennsylvania State University.
- Ismail, H.A., Lee, E.J., Ko, K.Y. and Ahn, D.U. 2008. Effects of aging time and natural antioxidants on the color, lipid oxidation and volatiles of irradiated ground beef. *Meat Science*. 80(3): 582-591.

- Ivan, M., Mir, P.S., Koenig, K.M., Rode, L.M., Neill, L., Entz, T. and Mir, Z. 2001. Effects of dietary sunflower seed oil on rumen protozoa population and tissue concentration of conjugated linoleic acid in sheep. *Small Ruminant Research*. 41(3): 215-227.
- Jenkins, T.C. 1993. Lipid metabolism in the rumen. *Journal of Dairy Science*. 76: 3851–3863.
- Jenkins, T.C., Klein, C.M. and Lee, Y.J. 2009. New insights on the pathways of lipid biohydrogenation in the rumen with possible implications on animal performance. *Proceedings of 24th Southwest Nutrition and Management Conference.* Tempe, AZ, USA. pp. 90- 105.
- Jensen, C., Flensted-Jensen, M., Skibsted, L.H. and Bertelsen, G. 1998. Effects of dietary rape seed oil, copper (II) sulphate and vitamin E on drip loss, colour and lipid oxidation of chilled pork chops packed in atmospheric air or in a high oxygen atmosphere. *Meat Science*. 50(2): 211-221.
- Jensen, R.G. 2002. The composition of bovine milk lipids: January 1995 to December 2000. *Journal of Dairy Science*. 85(2): 295-350.
- Jeronimo, E., Alves, S.P., Prates, J.A., Santos-Silva, J. and Bessa, R.J. 2009. Effect of dietary replacement of sunflower oil with linseed oil on intramuscular fatty acids of lamb meat. *Meat Science*. 83(3): 499-505.
- Jeukendrup, A.E. 2002. Regulation of fat metabolism in skeletal muscle. Annals of the New York Academy of Sciences. 967(1): 217-235.
- Jump, D.B. 2004. Fatty acid regulation of gene transcription. *Critical Reviews in Clinical Laboratory Sciences*. 41(1): 41-78.
- Jump, D.B. and Clarke, S.D. 1999. Regulation of gene expression by dietary fat. *Annual Review of Nutrition*. 19(1): 63-90.
- Jump, D.B., Clarke, S.D., MacDougald, O. and Thelen, A. 1993. Polyunsaturated fatty acids inhibit S14 gene transcription in rat liver and cultured hepatocytes. *Proceedings of the National Academy of Sciences*. 90(18): 8454-8458.
- Jump, D.B., Ren, B., Clarke, S. and Thelen, A. 1995. Effects of fatty acids on hepatic gene expression. *Prostaglandins, Leukotrienes and Essential Fatty Acids*. 52(2): 107-111.
- Kahraman, T., Ghassan, I., Bingöl, E.B., Dümen, E., Göksoy, E.Ö. and Büyükünal, S.K. 2014. Effect of rapid chilling and pelvic suspension on meat quality of *longissimus dorsi* muscle of lamb. *Kafkas Universitesi Veteriner Fakultesi Dergisi*. 20(5): 697-701.
- Kamra, D.N. 2005. Rumen microbial ecosystem. *Current Science*. 89(1): 124-135.

- Karami, M., Ponnampalam, E. N. and Hopkins, D. L. 2013. The effect of palm oil or canola oil on feedlot performance, plasma and tissue fatty acid profile and meat quality in goats. *Meat Science*. 94(2): 165-169.
- Karim, S.A. and Bhatt, R.S. 2012. Small ruminant production in India: issues and approaches. In *Trends in Small Ruminant Production: Perspectives and Prospects*. Eds. Sahoo, A., Sankhyan, S.K., Swarnkar, C.P., Shinde, A.K. and Karim, S.A. pp. 1-11. SSPH Delhi, India.
- Karim, S.A., Tripathi, M.K. and Singh, V.K. 2007. Effect of varying levels of concentrate supplementation on growth performance and carcass traits of finisher lambs. *Livestock Research for Rural Development*. 19: 173.
- Kaur, B. 2010. Consumer Preference for Goat Meat in Malaysia: Market Opportunities and Potential. *Journal of Agribusiness Marketing*. 3: 40-55.
- Kersten, S., Desvergne, B. and Wahli, W. 2000. Roles of PPARs in health and disease. *Nature*. 405(6785): 421-424.
- Keweloh, H. and Heipieper, H.J. 1996. *Trans* unsaturated fatty acids in bacteria. *Lipids*. 31(2): 129-137.
- Khan, M.F.U., Ashfaq, F. and Masood, M.A. 2014. Study on fattening potential of different sheep and goat breeds under different plan of nutrition in Pakistan. *Journal of Animal and Plant Sciences*. 24: 83-86.
- Kholif, S.M., Morsy, T.A., Matloup, O.H., Ebeid, H.M. and Kholif, A.M. 2015. Effects of crushed linseed or linseed oil supplementation on performance of dairy goats and fatty acid profile in milk. *Life Science Journal*. 12: 94-99.
- Kim, H.J., Miyazaki, M. and Ntambi, J.M. 2002. Dietary cholesterol opposes PUFA-mediated repression of the stearoyl-CoA desaturase-1 gene by SREBP-1 independent mechanism. *Journal of Lipid Research*. 43(10): 1750-1757.
- Kim, K.H. 1997. Regulation of mammalian acetyl-coenzyme A carboxylase. Annual Review of Nutrition. 17(1): 77-99.
- Kim, S.C., Adesogan, A.T., Badinga, L. and Staples, C.R. 2007. Effects of dietary n-6: n-3 fatty acid ratio on feed intake, digestibility, and fatty acid profiles of the ruminal contents, liver, and muscle of growing lambs. *Journal of Animal Science*. 85(3): 706-716.
- Kitessa, S.M., Gulati, S.K., Ashes, J.R., Fleck, E., Scott, T.W. and Nichols, P.D. 2001. Utilisation of fish oil in ruminants: I. Fish oil metabolism in sheep. *Animal Feed Science and Technology*. 89(3): 189-199.

- Koike, S. and Kobayashi,Y. 2001. Development and use of competitive PCR assays for the rumen cellulolytic bacteria: *Fibrobacter succinogenes, Ruminococcus albus* and *Ruminococcus flavefaciens*. *FEMS Microbiology Letters*. 204(2): 361–366.
- Konig, B., Rauer, C., Rosenbaum, S., Brandsch, C., Eder, K. and Stangl, G.I. 2009. Fasting Upregulates PPAR α Target Genes in Brain and Influences Pituitary Hormone Expression in a PPAR α Dependent Manner. *PPAR Research*. 2009: 801609.
- Kott, R.W., Hatfield, P.G., Bergman, J.W., Flynn, C.R., Van Wagoner, H. and Boles, J.A. 2003. Feedlot performance, carcass composition, and muscle and fat CLA concentrations of lambs fed diets supplemented with safflower seeds. *Small Ruminant Research*. 49(1): 11-17.
- Krause, D. O., McSweeney, C. S. and Forster, R. J. 1999. Molecular ecological methods to study fibrolyticruminal bacteria: phylogeny, competition and persistence. *International Symposium on Microbial Ecology*. 8: 15-19.
- Krause, D.O., Denman, S.E., Mackie, R.I., Morrison, M., Rae, A.L., Attwood, G.T. and McSweeney, C.S. 2003. Opportunities to improve fiber degradation in the rumen: microbiology, ecology, and genomics. *FEMS Microbiology Reviews*. 27(5): 663-693.
- Krishnamoorthy, U. and Moran, J. 2012. Rearing young ruminants on milk replacers and starter feeds. *FAO Animal Production and Health Manual, No. 13.* Food and Agriculture Organization of the United Nations (FAO). Rome.
- Kritchevsky, D. 2000. Antimutagenic and some other effects of conjugated linoleic acid. *British Journal of Nutrition*. 83(05): 459-465.
- Ladeira, M.M., Santarosa, L.C., Chizzotti, M.L., Ramos, E.M., Neto, O.M., Oliveira, D.M., Carvalho, J.R.R., Lopes, L.S. and Ribeiro, J.S. 2014. Fatty acid profile, color and lipid oxidation of meat from young bulls fed ground soybean or rumen protected fat with or without monensin. *Meat Science*. 96(1): 597-605.
- Laliotis, G.P., Bizelis, I. and Rogdakis, E. 2010. Comparative approach of the *de novo* fatty acid synthesis (lipogenesis) between ruminant and non ruminant mammalian species: from biochemical level to the main regulatory lipogenic genes. *Current Genomics.* 11(3): 168-183.
- Lambert, M.S., Avella, M.A., Botham, K.M. and Mayes, P.A. 1998. Comparison of short-and long-term effects of different dietary fats on the hepatic uptake and metabolism of chylomicron remnants in rats. *British Journal of Nutrition*. 79(02): 203-211.
- Lana, R.P., Russell, J.B. and Van Amburgh M.E. 1998. The role of pH in regulating ruminal methane and ammonia production. *Journal of Animal Science*. 76: 2190.–2196.

Lane, D.J. 1991. 16S/23S rRNA sequencing.nucleic acid techniques in bacterial systematics. In *Nucleic Acid Techniques in Bacterial Systematics*. Eds. Stackebrandt, E. and Goodfellow, M. pp. 114-147. John Wiley & Sons, New York, NY, USA.

Lawrie, R.A. 1991. *Meat Science. 5th edition*. Pergamon Press, Oxford.

- Lee, H.J., Jung, J.Y., Oh, Y.K., Lee, S.S., Madsen, E.L. and Jeon, C.O. 2012. Comparative survey of rumen microbial communities and metabolites across one caprine and three bovine groups, using bar-coded pyrosequencing and 1H nuclear magnetic resonance spectroscopy. *Applied and Environmental Microbiology*. 78(17): 5983-5993.
- Lee, H.J., Lee, S.C., Kim, J.D., Oh, Y.G., Kim, B.K., Kim, C.W. and Kim, K.J. 2003. Methane production potential of feed ingredients as measured by *in vitro* gas test. *Asian Australasian Journal of Animal Sciences*. 16(8): 1143-1150.
- Lee, S.S., Ha, J.K. and Cheng, K.J. 2000. Relative contributions of bacteria, protozoa, and fungi to *in vitro* degradation of orchard grass cell walls and their interactions. *Applied and Environmental Microbiology*. 66(9); 3807-3813.
- Lee, Y.J. 2009. Metabolic Tracer Studies of Linoleic and Linolenic Acids to Identify Biohydrogenation Intermediates Produced by Ruminal Microorganisms. PhD Thesis Clemson University, South Carolina, USA.
- Lee, Y.J. and Jenkins, T.C. 2011. Biohydrogenation of linolenic acid to stearic acid by the rumen microbial population yields multiple intermediate conjugated diene isomers. *The Journal of Nutrition*. 141(8): 1445-1450.
- Leonard, A.E., Pereira, S.L., Sprecher, H. and Huang, Y.S. 2004. Elongation of long-chain fatty acids. *Progress in Lipid Research*. 43(1): 36-54.
- Li, D.B.and Hou, X.Z. 2007. Effect of fungal elimination on bacteria and protozoa populations and degradation of straw dry matter in the rumen of sheep and goats. *Asian Australasian Journal of Animal Sciences*. 20(1): 70-74.
- Li, F., Yang, X.J., Cao, Y.C., Li, S.X., Yao, J.H., Li, Z.J. and Sun, F.F. 2014. Effects of dietary effective fiber to rumen degradable starch ratios on the risk of sub-acute ruminal acidosis and rumen content fatty acids composition in dairy goat. *Animal Feed Science and Technology*. 189: 54-62.
- Lillis, L., Boots, B., Kenny, D.A., Petrie, K., Boland, T.M., Clipson, N. and Doyle, E.M. 2011. The effect of dietary concentrate and soya oil inclusion on microbial diversity in the rumen of cattle. *Journal of Applied Microbiology*. 111(6): 1426-1435.

- Lock, A.L., Harvatine, K.J., Drackley, J.K. and Bauman, D.E. 2006. Concepts in fat and fatty acid digestion in ruminants. *Proceedings Intermountain Nutrition Conference*. pp. 85-100.
- Lock, A.L., Horne, C.A., Bauman, D.E. and Salter, A.M. 2005. Butter naturally enriched in conjugated linoleic acid and vaccenic acid alters tissue fatty acids and improves the plasma lipoprotein profile in cholesterol-fed hamsters. *The Journal of Nutrition*. 135(8): 1934-1939.
- Loh, T.C. 2004. Livestock production and the feed industry in Malaysia. In *Protein Sources for the Animal Feed Industry. FAO Expert Consultation and Workshop.* Bangkok, Thailand. April 29 -May 3. pp. 329-339.
- Loor, J.J., Ueda, K., Ferlay, A., Chilliard, Y. and Doreau, M. 2004. Biohydrogenation, duodenal flow, and intestinal digestibility of *trans* fatty acids and conjugated linoleic acids in response to dietary forage: concentrate ratio and linseed oil in dairy cows. *Journal of Dairy Science*. 87(8): 2472-2485.
- Lopez-Ferrer, S., Baucells, M.D., Barroeta, A.C., Galobart, J. and Grashorn, M.A. 2001. n-3 enrichment of chicken meat. 2. Use of precursors of long-chain polyunsaturated fatty acids: Linseed oil. *Poultry Science*. 80(6): 753-761.
- Lourenco, M., Ramos-Morales, E. and Wallace, R.J. 2010. The role of microbes in rumen lipolysis and biohydrogenation and their manipulation. *Animal*. 4(07): 1008-1023.
- Lynch, S.M. and Frei, B. 1993. Mechanisms of copper-and iron-dependent oxidative modification of human low density lipoprotein. *Journal of Lipid Research*. 34(10): 1745-1753.
- Madruga, M.S., Medeiros, E.J.L.D., Sousa, W.H.D., Cunha, M.D.G.G., Pereira Filho, J.M. and Queiroga, R.D.C.R.D. 2009. Chemical composition and fat profile of meat from crossbred goats reared under feedlot systems. *Revista Brasileira de Zootecnia*. 38(3): 547-552.
- Mahgoub, O., Lu, C.D. and Early, R.J. 2000. Effects of dietary energy density on feed intake, body weight gain and carcass chemical composition of Omani growing lambs. *Small Ruminant Research*. 37(1): 35-42.
- Maia, M.R., Chaudhary, L.C., Bestwick, C.S., Richardson, A.J., McKain, N., Larson, T.R., Graham, I.A. and Wallace, R.J. 2010. Toxicity of unsaturated fatty acids to the biohydrogenating ruminal bacterium, *Butyrivibrio fibrisolvens. BMC Microbiology*. 10(1): 1.
- Maia, M.R., Chaudhary, L.C., Figueres, L. and Wallace, R.J. 2007. Metabolism of polyunsaturated fatty acids and their toxicity to the microflora of the rumen. *Antonie Van Leeuwenhoek*. 91(4): 303-314.

- Man, W.C., Miyazaki, M., Chu, K. and Ntambi, J. 2006. Colocalization of SCD1 and DGAT2: implying preference for endogenous monounsaturated fatty acids in triglyceride synthesis. *Journal of Lipid Research*. 47(9): 1928-1939.
- Mancini, R.A. and Hunt, M. 2005. Current research in meat color. *Meat Science*. 71(1): 100-121.
- Mann, N., Pirotta, Y., O'Connell, S., Li, D., Kelly, F. and Sinclair, A. 2006. Fatty acid composition of habitual omnivore and vegetarian diets. *Lipids*. 41(7): 637-646.
- Mannen, H. 2012. Genes associated with fatty acid composition of beef. Food Science and Technology Research. 18(1): 1-6.
- Manso, T., Castro, T., Mantecón, A.R. and Jimeno, V. 2006. Effects of palm oil and calcium soaps of palm oil fatty acids in fattening diets on digestibility, performance and chemical body composition of lambs. *Animal Feed Science and Technology*. 127(3): 175-186.
- Mao, H.L., Wang, J.K., Lin, J. and Liu, J.X. 2012. Fatty acid profiles and stearoyl-coa desaturase gene expression in *Longissimus dorsi* muscle of growing lambs influenced by addition of tea saponins and soybean oil. *Asian-Australasian Journal of Animal Sciences*. 25(5): 648.
- Mao, H.L., Wang, J.K., Zhou, Y.Y. and Liu, J.X. 2010. Effects of addition of tea saponins and soybean oil on methane production, fermentation and microbial population in the rumen of growing lambs. *Livestock Science*. 129(1): 56-62.
- Marin, A.M., Gómez-Cortés, P., Castro, G.G., Juárez, M., Alba, L.P., Hernández, M.P. and De la Fuente, M.A. 2012. Effects of feeding increasing dietary levels of high oleic or regular sunflower or linseed oil on fatty acid profile of goat milk. *Journal of Dairy Science*. 95(4): 1942-1955.
- Martin, C., Rouel, J., Jouany, J.P., Doreau, M. and Chilliard, Y. 2008. Methane output and diet digestibility in response to feeding dairy cows crude linseed, extruded linseed, or linseed oil. *Journal of Animal Science*. 86: 2642–2650.
- Martinez Marín, A.L., Pérez Hernández, M., Alba, P., Luis, M., Carrión Pardo, D., Garzón Sigler, A.I. and Gómez Castro, G. 2013. Fat addition in the diet of dairy ruminants and its effects on productive parameters. *Revista Colombiana de Ciencias Pecuarias*. 26(2): 69-78.
- Massaro, M., Carluccio, M.A. and De Caterina, R. 1999. Direct vascular antiatherogenic effects of oleic acid: a clue to the cardioprotective effects of the Mediterranean diet. *Cardiologia*. Rome, Italy. 44(6): 507-513.

- Matsushita, M., Tazinafo, N.M., Padre, R.G., Oliveira, C.C., Souza, N.E., Visentainer, J.V., Macedo, F.A.F. and Ribas, N.P. 2007. Fatty acid profile of milk from Saanen goats fed a diet enriched with three vegetable oils. *Small Ruminant Research*. 72(2): 127-132.
- Matthews, K.R., Homer, D.B., Thies, F. and Calder, P.C. 2000. Effect of whole linseed (*Linum usitatissimum*) in the diet of finishing pigs on growth performance and on the qualityand fatty acid composition of various tissues. *British Journal of Nutrition*. 83(06): 637-643.
- Mayes, P.A. and Botham, K.M. 2003. Biosynthesis of fatty acids. In *Harper's Illustrated Biochemistry*. Eds. Murray, R.K., Granner, D.K., Mayes, P.A. and Rodwell, V.W. pp. 173-179. 26th ed. McGraw-Hill.
- McAfee, A.J., McSorley, E.M., Cuskelly, G.J., Moss, B.W., Wallace, J.M., Bonham, M.P. and Fearon, A.M. 2010. Red meat consumption: An overview of the risks and benefits. *Meat Science*. 84(1): 1-13.
- McSweeney, C.S., Blackall, L.L., Collins, E., Conlan, L.L., Webb, R.I., Denman, S.E. and Krause, D.O. 2005. Enrichment, isolation and characterisation of ruminal bacteria that degrade non-protein amino acids from the tropical legume *Acacia angustissima*. *Animal Feed Science and Technology*. 121(1): 191-204.
- Mele, M., Buccioni, A., Petacchi, F., Serra, A., Banni, S., Antongiovanni, M. and Secchiari, P. 2006. Effect of forage/concentrate ratio and soybean oil supplementation on milk yield, and composition from Sarda ewes. *Animal Research*, 55(4): 273-285.
- Menke, K.H., Raab, L., Salewski, A., Steingass, H., Fritz, D. and Schneider, W. 1979. The estimation of the digestibility and metabolizable energy content of ruminant feedingstuffs from the gas production when they are incubated with rumen liquor *in vitro*. *The Journal of Agricultural Science*. 93(01): 217-222.
- Miller, C.W. and Ntambi, J.M. 1996. Peroxisome proliferators induce mouse liver stearoyl-CoA desaturase 1 gene expression. *Proceedings of the National Academy of Sciences*. 93(18): 9443-9448.
- Min, B. and Ahn, D.U. 2005. Mechanism of lipid peroxidation in meat and meat products-A review. *Food Science and Biotechnology*. 14(1): 152-163.
- Minihane, A.M. 2009. Nutrient gene interactions in lipid metabolism. *Current Opinion in Clinical Nutrition and Metabolic Care*. 12(4): 357-363.
- Mir, Z., Rushfeldt, M.L., Mir, P.S., Paterson, L.J. and Weselake, R.J. 2000. Effect of dietary supplementation with either conjugated linoleic acid (CLA) or linoleic acid rich oil on the CLA content of lamb tissues. *Small Ruminant Research*. 36(1): 25-31.

- Mohamed, A., Jamilah, B., Abbas, K.A. and Rahman, R.A. 2008. A review on lipid oxidation of meat in active and modified atmosphere packaging and usage of some stabilizers. *Journal of Food, Agriculture and Environment*. 6(3&4): 76-81.
- Moibi, J.A., Christopherson, R.J. and Okine, E.K. 2000. Effect of environmental temperature and dietary lipid supplement on activity and protein abundance of acetyl-CoA carboxylase and fatty acid synthase in skeletal muscle, liver and adipose tissues of sheep. *Canadian Journal* of Animal Science. 80(1): 69-77.
- Moon, Y.H., Ok, J.U., Lee, S.J., Ha, J.K. and Lee, S.S. 2010. A comparative study on the rumen microbial populations, hydrolytic enzyme activities and dry matter degradability between different species of ruminant. *Animal Science Journal*. 81(6): 642-647.
- Morand-Fehr, P., and D. Sauvant. 1987. Feeding strategies in goats. *Proceedings of. 4th International Conference on Goats.* Brasilia, Brazil. Pp. 1275–1303.
- Morrissey, P.A., Sheehy, P.J.A., Galvin, K., Kerry, J.P. and Buckley, D.J. 1998. Lipid stability in meat and meat products. *Meat Science*. 49: 73-86.
- Moss, A.R., Jouany, J.P. and Newbold, J. 2000. Methane production by ruminants: its contribution to global warming. *Annales de Zootechnie.* 49(3): 231-253.
- Murthy, K.S., Reddy, M.R. and Reddy, G.V.N., 1996. Nutritive value of supplements containing poultry droppings/litter for sheep and goats. *Small Ruminant Research*, *21*(2), pp.71-75.
- Mushi, D. E., Thomassen, M. S., Kifaro, G. C. and Eik, L. O. 2010. Fatty acid composition of minced meat, *longissimus* muscle and omental fat from Small East African goats finished on different levels of concentrate supplementation. *Meat Science*. 86(2): 337-342.
- Nafikov, R.A. and Beitz, D.C. 2007. Carbohydrate and lipid metabolism in farm animals. *The Journal of Nutrition*. 137(3): 702-705.
- Najafi, M.H., Zeinoaldini, S., Ganjkhanlou, M., Mohammadi, H., Hopkins, D.L. and Ponnampalam, E.N. 2012. Performance, carcass traits, muscle fatty acid composition and meat sensory properties of male Mahabadi goat kids fed palm oil, soybean oil or fish oil. *Meat Science*. 92(4): 848-854.
- Nakamura, M.T. and Nara, T.Y. 2002. Gene regulation of mammalian desaturases. *Biochemical Society Transactions*. 30(6): 1076-1079.
- Nakamura, M.T. and Nara, T.Y. 2004. Structure, function, and dietary regulation of $\Delta 6$, $\Delta 5$, and $\Delta 9$ desaturases. *Annual Review of Nutrition*. 24: 345-376.

Nakamura, M.T., Cheon, Y., Li, Y. and Nara, T.Y. 2004. Mechanisms of regulation of gene expression by fatty acids. *Lipids*. 39(11): 1077-1083.

- Nam, I.S. and Garnsworthy, P.C. 2007. Biohydrogenation of linoleic acid by rumen fungi compared with rumen bacteria. *Journal of Applied Microbiology*. 103(3): 551-556.
- Nhan, N.T.H., Ngu, N.T., Thiet, N., Preston, T.R. and Leng, R.A. 2007. Determination of the optimum level of a soybean oil drench with respect to the rumen ecosystem, feed intake and digestibility in cattle. *Livestock Research for Rural Development*. 19(8): 117-118.
- Nicol, A.M., Poppi, D.P., Alam, M.R. and Collins, H.A. 1987. Dietary differences between goats and sheep. *Proceedings of the New Zealand Grassland Association*. 48: 199-205.
- Ntambi, J.M. 1999. Regulation of stearoyl-CoA desaturase by polyunsaturated fatty acids and cholesterol. *Journal of Lipid Research*. 40(9): 1549-1558.
- Nurnberg, K., Wegner, J. and Ender, K. 1998. Factors influencing fat composition in muscle and adipose tissue of farm animals. *Livestock Production Science*. 56(2): 145-156.
- Ogawa, J., Matsumura, K., Kishino, S., Omura, Y. and Shimizu, S. 2001. Conjugated linoleic acid accumulation via 10-hydroxy-12-octadecaenoic acid during microaerobic transformation of linoleic acid by *Lactobacillus acidophilus*. *Applied and Environmental Microbiology*. 67(3): 1246-1252.
- Oldick, B.S. and Firkins, J.L. 2000. Effects of degree of fat saturation on fiber digestion and microbial protein synthesis when diets are fed twelve times daily. *Journal of Animal Science*. 78(9): 2412-2420.
- Omar, M.A. 1998. Recent developments in animal genetics and breeding in Malaysia. *Genetic Society of Malaysia*. Retrieved 12 December 2015 from

http://www.persatuangenetikmalaysia.com/files/congress03/03Concurrent05.pdf

- Orpin, C.G. and Joblin, K.N. 1997. The rumen anaerobic fungi. In *The Rumen Microbial Ecosystem*. Eds. Hobson, P.N. and Stewart, C.S. pp. 140-195. Springer Netherlands.
- Or-Rashid, M.M., Odongo, N.E. and McBride, B.W. 2007. Fatty acid composition of ruminal bacteria and protozoa, with emphasis on conjugated linoleic acid, vaccenic acid, and odd-chain and branchedchain fatty acids. *Journal of Animal Science*. 85(5): 1228-1234.
- Orskov, E.R. and McDonald, J. 1979. The estimation of protein degradability in the rumen from incubation measurements weighed according to rate of passage. *Journal of Agricultural Science* (Cambridge) 92: 499–503.

- Osmundsen, H., Bremer, J. and Pedersen, J.I. 1991. Metabolic aspects of peroxisomal β-oxidation. *Biochimica et Biophysica Acta (BBA)-Lipids and Lipid Metabolism*. 1085(2): 141-158.
- Paillard, D., McKain, N., Chaudhary, L.C., Walker, N.D., Pizette, F., Koppova, I., McEwan, N.R., Kopečný, J., Vercoe, P.E., Louis, P. and Wallace, R.J. 2007. Relation between phylogenetic position, lipid metabolism and butyrate production by different *Butyrivibrio*-like bacteria from the rumen. *Antonie Van Leeuwenhoek*. 91(4): 417-422.
- Palmquist, D.L. 2009. Omega-3 fatty acids in metabolism, health, and nutrition and for modified animal product foods. *The Professional Animal Scientist*. 25(3): 207-249.
- Palmquist, D.L. 2010. Essential fatty acids in ruminant diets. *Proceedings of the 21st Annual Ruminant Nutrition Symposium.* pp. 2-3.
- Panchadcharam, C. 2004. Problems in the Control of Nematode Parasites of Small Ruminants in Malaysia: Resistance to Anthelmintics and the Biological Control Alternative. PhD Thesis. Swedish University of Agricultural Sciences, Uppsala, Sweden.
- Pariza, M.W., Park, Y. and Cook, M.E. 2001. The biologically active isomers of conjugated linoleic acid. *Progress in Lipid Research*. 40(4): 283-298.
- Park, Y., Albright, K.J., Liu, W., Storkson, J.M., Cook, M.E. and Pariza, M.W. 1997. Effect of conjugated linoleic acid on body composition in mice. *Lipids*. 32(8): 853-858.
- Park, Y., Storkson, J.M., Albright, K.J., Liu, W. and Pariza, M.W. 1999. Evidence that the *trans*-10, *cis*-12 isomer of conjugated linoleic acid induces body composition changes in mice. *Lipids*. 34(3): 235-241.
- Patterson, E., Wall, R., Fitzgerald, G.F., Ross, R.P. and Stanton, C. 2012. Health implications of high dietary omega-6 polyunsaturated fatty acids. *Journal of Nutrition and Metabolism*. 2012 :539426.
- Pearce, K.L., Rosenvold, K., Andersen, H.J. and Hopkins, D.L. 2011. Water distribution and mobility in meat during the conversion of muscle to meat and ageing and the impacts on fresh meat quality attributes—A review. *Meat Science*. 89(2): 111-124.
- Pell, A.N. and Schofield, P. 1993. Computerized monitoring of gas production to measure forage digestion *in vitro*. *Journal of Dairy Science*. 76(4): 1063-1073.
- Penner, G.B. and Aschenbach, J.R. 2011. Mechanisms of acid absorption in the rumen and impacts on subacute sumen acidosis. University of Saskatchewan, Canada. Retrieved 23 September 2015 from <u>http://www.diss.fuberlin.de/docs/servlets/MCRFileNodeServlet/FUDOC</u> <u>S derivate 00000002305/9penner.pdf</u>

- Perdereau, D., Narkewicz, M., Coupe, C., Ferre, P. and Girard, J. 1990. Hormonal control of specific gene expression in the rat liver during the suckling-weaning transition. *Advances in Enzyme Regulation*. 30: 91-108.
- Pereira, P.M.D.C.C. and Vicente, A.F.D.R.B. 2013. Meat nutritional composition and nutritive role in the human diet. *Meat Science*. 93(3): 586-592.
- Peters, J.M., Hollingshead, H.E. and Gonzalez, F.J. 2008. Role of peroxisome-proliferator-activated receptor β/δ (PPARβ/δ) in gastrointestinal tract function and disease. *Clinical Science*. 115(4): 107-127.
- Piperova, L.S., Teter, B.B., Bruckental, I., Sampugna, J., Mills, S.E., Yurawecz, M.P., Fritsche, J., Ku, K. and Erdman, R.A. 2000. Mammary lipogenic enzyme activity, *trans* fatty acids and conjugated linoleic acids are altered in lactating dairy cows fed a milk fat–depressing diet. *The Journal of Nutrition*. 130(10): 2568-2574.
- Ponnampalam, E.N., Sinclair, A.J., Hosking, B.J. and Egan, A.R. 2002. Effects of dietary lipid type on muscle fatty acid composition, carcass leanness, and meat toughness in lambs. *Journal of Animal Science*. 80(3): 628-636.
- Popova, T., Marinova, P., Banskalieva, V. and Vasileva, V. 2008. Content and fatty acid composition of different fat depots of lambs receiving fish oil supplemented diet. *Bulgarian Journal of Agricultural Science*. 14(1): 100-107.
- Postic, C. and Girard, J. 2008. Contribution of de novo fatty acid synthesis to hepatic steatosis and insulin resistance: lessons from genetically engineered mice. *The Journal of Clinical Investigation*. 118(3): 829-838.
- Priolo, A., Micol, D. and Agabriel, J. 2001. Effects of grass feeding systems on ruminant meat colour and flavour. A review. *Animal Research*. 50(3): 185-200.
- Racine, R.A. and Deckelbaum, R.J. 2007. Sources of the very-long-chain unsaturated omega-3 fatty acids: eicosapentaenoic acid and docosahexaenoic acid. *Current Opinion in Clinical Nutrition and Metabolic Care*. 10(2): 123-128.
- Ramirez-Retamal, J. and Morales, R. 2014. Influence of breed and feeding on the main quality characteristics of sheep carcass and meat: A review. *Chilean Journal of Agricultural Research*. 74(2): 225-233.
- Rasmussen, B.B., Holmbäck, U.C., Volpi, E., Morio-Liondore, B., Paddon-Jones, D. and Wolfe, R.R. 2002. Malonyl coenzyme A and the regulation of functional carnitine palmitoyltransferase-1 activity and fat oxidation in human skeletal muscle. *The Journal of Clinical Investigation*. 110(11): 1687-1693.

- Ratledge, C. and Kristiansen, B. 2006. *Basic Biotechnology*. Cambridge University Press.
- Ratnayake, W.N. and Galli, C. 2009. Fat and fatty acid terminology, methods of analysis and fat digestion and metabolism: a background review paper. *Annals of Nutrition and Metabolism*. 55(1-3): 8-43.
- Renerre, M. 2000. Oxidative processes and myoglobin. *Antioxidants in Muscle Foods*. 113-133.
- Rey, A.I., Kerry, J.P., Lynch, P.B., Lopez-Bote, C.J., Buckley, D.J. and Morrissey, P.A. 2001. Effect of dietary oils and alpha-tocopheryl acetate supplementation on lipid (TBARS) and cholesterol oxidation in cooked pork. *Journal of Animal Science*. 79(5): 1201-1208.
- Rezaeian, M., Beakes, G.W. and Parker, D.S. 2004. Distribution and estimation of anaerobic zoosporic fungi along the digestive tracts of sheep. *Mycological Research*. 108(10): 1227-1233.
- Rhee, K. S. 1992. Fatty acids in meats and meat products. In *Fatty Acids in Foods and their Health Implications.* Ed. Chow, C.K. pp. 65-93. Marcel Dekker, New York,
- Robbins, C.T., Spalinger, D.E. and van Hoven, W., 1995. Adaptation of ruminants to browse and grass diets: are anatomical-based browser-grazer interpretations valid?. *Oecologia*. 103(2): 208-213.
- Russell, J. B. 1998. The importance of pH in the regulation of ruminal acetate to propionate ratio and methane production *in vitro*. *Journal of Dairy Science*. 81: 3222.–3230.
- Rustan, A.C. and Drevon, C.A. 2005. Fatty acids: structures and properties. In *Encyclopedia of Life Sciences*. pp. 1-7. John Wiley & Sons, New York, NY, USA.
- Ruxton, C.H.S., Reed, S.C., Simpson, M.J.A. and Millington, K.J. 2004. The health benefits of omega-3 polyunsaturated fatty acids: a review of the evidence. *Journal of Human Nutrition and Dietetics*. 17(5): 449-459.
- Saini, J.K., Hundal, J.S., Wadhwa, M. and Bakshi, M.P.S. 2012. Effect of roughage to concentrate ratio in the diet on the rumen environment and nutrient utilization in goat and sheep. *Indian Journal of Animal Nutrition*. 4: 333-338.
- Saleem, A.Z.M., Salem, M.Z.M., El-Adawy, M.M. and Robinson, P.H. 2006. Nutritive evaluations of some browse tree foliages during the dry season: secondary compounds, feed intake and *in vivo* digestibility in sheep and goats. *Animal Feed Science and Technology*. 127(3): 251-267.

- Salim, H.M., Shahjalal, M., Tareque, A.M.M.and Akter, N. 2003. Intake and growth performance of female goats and sheep given concentrate supplement under grazing condition. *Pakistan Journal of Biological Sciences*. 6: 1304-1309.
- Sallam, S.M.A., Nasser, M.E.A., Araujo, R.C. and Abdalla, A.L. 2009. Methane emission *in vivo* by sheep consuming diet with different levels of eucalyptus essential oil. *Proceedings FAO/IAEA International Symposium on Sustainable Improvement of Animal Production and Health*. pp. 210-211.
- Santos-Silva, J., Mendes, I.A., Portugal, P.V. and Bessa, R.J.B. 2004. Effect of particle size and soybean oil supplementation on growth performance, carcass and meat quality and fatty acid composition of intramuscular lipids of lambs. *Livestock Production Science*. 90(2): 79-88.
- Santra, A. and Karim, S.A. 2000. Growth performance of faunated and defaunated Malpura weaner lambs. *Animal Feed Science and Technology*. 86: 251-260.
- Santra, A., Karim, S.A. and Chaturvedi, O.H. 2007. Rumen enzyme profile and fermentation characteristics in sheep as affected by treatment with sodium lauryl sulfate as defaunating agent and presence of ciliate protozoa. *Small Ruminant Research*. 67(2): 126-137.
- Schoonjans, K., Staels, B. and Auwerx, J. 1996. Role of the peroxisome proliferator-activated receptor (PPAR) in mediating the effects of fibrates and fatty acids on gene expression. *Journal of Lipid Research*. 37(5): 907-925.
- Schumann, B.E., Squires, E.J. and Leeson, S. 2000. Effect of dietary flaxseed, flax oil and n-3 fatty acid supplement on hepatic and plasma characteristics relevant to fatty liver haemorrhagic syndrome in laying hens. *British Poultry Science*. 41(4): 465-472.
- Scollan, N., Hocquette, J.F., Nuernberg, K., Dannenberger, D., Richardson, I. and Moloney, A. 2006. Innovations in beef production systems that enhance the nutritional and health value of beef lipids and their relationship with meat quality. *Meat Science*. 74(1): 17-33.
- Sen, A.R., Santra, A. and Karim, S.A. 2004. Carcass yield, composition and meat quality attributes of sheep and goat under semiarid conditions. *Meat Science*. 66(4): 757-763.
- Sheridan, R., Hoffman, L.C. and Ferreira, A.V. 2003. Meat quality of Boer goat kids and Mutton Merino lambs 1. Commercial yields and chemical composition. *Animal Science*. 76(01): 63-71.

- Shija, D.S., Mtenga, L.A., Kimambo, A.E., Laswai, G.H., Mushi, D.E., Mgheni, D.M., Mwilawa, A.J., Shirima, E.J. and Safari, J.G. 2013. Chemical composition and meat quality attributes of indigenous sheep and goats from traditional production system in Tanzania. *Asian-Australasian Journal of Animal Sciences*. 26(2): 295-302.
- Shingfield, K.J. and Wallace, R.J. 2014. Synthesis of conjugated linoleic acid in ruminants and humans. In *Conjugated Linoleic Acids and Conjugated Vegetable Oils.* Eds. Sels, B. and Philippaerts, A. pp. 1-65. Royal Society of Chemistry. London, UK.
- Shingfield, K.J., Ahvenjarvi, S., Toivonen, V., Arola, A., Nurmela, K.V.V., Huhtanen, P. and Griinari, J.M. 2003. Effect of dietary fish oil on biohydrogenation of fatty acids and milk fatty acid content in cows. *Animal Science*. 77 (1): 165-179.
- Shipley, L.A. 1999. Grazers and browsers: how digestive morphology affects diet selection. In *Grazing Behavior of Livestock and Wildlife*. pp.20-27. University of Idaho, Rangeland Ecology and Management.
- Siddiqui, R.A., Harvey, K.A. and Zaloga, G.P. 2008. Modulation of enzymatic activities by n-3 polyunsaturated fatty acids to support cardiovascular health. *The Journal of Nutritional Biochemistry*. 19(7): 417-437.
- Silva, T.M., Oliveira, R.L., Barbosa, L.P., Garcez Neto, A.F., Bagaldo, A.R., Lanna, D.P.D., and Jesus, I.B. 2011. Preliminary study on meat quality of goats fed levels of licury oil in the diet. *Asian Australasian Journal of Animal Sciences*. 24(8): 1112-1119.
- Simopoulos, A.P. 2008. The importance of the omega-6/omega-3 fatty acid ratio in cardiovascular disease and other chronic diseases. *Experimental Biology and Medicine*. 233(6): 674-688.
- Singh, M. 2005. Essential fatty acids, DHA and human brain. *The Indian Journal of Pediatrics*. 72(3): 239-242.
- Smith, S. 1994. The animal fatty acid synthase: one gene, one polypeptide, seven enzymes. *The FASEB Journal*. 8(15): 1248-1259.
- Smith, S. B., Gill, C. A., Lunt, D. K., and Brooks, M. A. 2009. Regulation of fat and fatty acid composition in beef cattle. *Asian-Australasian Journal of Animal Sciences*. 22(9): 1225-1233.
- Smith, S., Witkowski, A. and Joshi, A.K. 2003. Structural and functional organization of the animal fatty acid synthase. *Progress in Lipid Research*. 42(4): 289-317.
- Smith, S.B., Lunt, D.K., Chung, K.Y., Choi, C.B., Tume, R.K. and Zembayashi, M. 2006. Adiposity, fatty acid composition, and delta-9 desaturase activity during growth in beef cattle. *Animal Science Journal*. 77(5): 478-486.

- Solomon, M.B., Lynch, G.P., Paroczay, E. and Norton, S. 1991. Influence of rapeseed meal, whole rapeseed, and soybean meal on fatty acid composition and cholesterol content of muscle and adipose tissue from ram lambs. *Journal of Animal Science*. 69(10): 4055-4061.
- Soyeurt, H., Dehareng, F., Mayeres, P., Bertozzi, C. and Gengler, N. 2008. Variation of Δ 9-desaturase activity in dairy cattle. *Journal of Dairy Science*. 91(8): 3211-3224.
- Stewart, C.S., Flint, H.J. and Bryant, M.P. 1997. The rumen bacteria. In *The Rumen Microbial Ecosystem*. Eds. Hobson, P.N. and Stewart, C.S. pp. 140-195. pp. 10-72. Springer Netherlands.
- Sylvester, J.T., Karnati, S.K.R., Yu, Z., Morrison, M., Firkins, J.L. 2004. Development of an assay to quantify rumen ciliate protozoal biomass in cows using real-time PCR. *Journal of Nutrition.* 134 (12): 3378–3384.
- Tapp, W.N., Yancey, J.W.S. and Apple, J.K. 2011. How is the instrumental color of meat measured?. *Meat Science*. 89(1): 1-5.
- Theodorou, M.K., Williams, B.A., Dhanoa, M.S., McAllan, A.B. and France, J. 1994. A simple gas production method using a pressure transducer to determine the fermentation kinetics of ruminant feeds. *Animal Feed Science and Technology*. 48(3): 185-197.
- Tilley, J.M.A. and Terry, R.A. 1963. A two-stage technique for the *in vitro* digestion of forage crops. *Grass and Forage Science*. 18(2): 104-111.
- Tisserand, J.L., Hadjipanayiotou, M. and Gihad, E.A. 1991. Digestion in goats. In *Goat Nutrition*. Ed. Morand-Fehr, P. pp. 46-60. Pudoc, Wageningen.
- Toral, P.G., Bernard, L., Belenguer, A., Rouel, J., Hervás, G., Chilliard, Y. and Frutos, P. 2016. Comparison of ruminal lipid metabolism in dairy cows and goats fed diets supplemented with starch, plant oil, or fish oil. *Journal of Dairy Science*. 99(1): 301-316.
- Torok, V.A., Percy, N.J., Moate, P.J. and Ophel-Keller, K. 2014. Influence of dietary docosahexaenoic acid supplementation on the overall rumen microbiota of dairy cows and linkages with production parameters. *Canadian Journal of Microbiology*. 60(5): 267-275.
- Tricon, S., Burdge, G.C., Kew, S., Banerjee, T., Russell, J.J., Grimble, R.F., Williams, C.M., Calder, P.C. and Yaqoob, P. 2004. Effects of *cis*-9, *trans*-11 and *trans*-10, *cis*-12 conjugated linoleic acid on immune cell function in healthy humans. *The American Journal of Clinical Nutrition*. 80(6): 1626-1633.
- Tshabalala, P.A., Strydom, P.E., Webb, E.C. and De Kock, H.L. 2003. Meat quality of designated South African indigenous goat and sheep breeds. *Meat Science*. 65(1): 563-570.

- Tsiplakou, E. and Zervas, G. 2008. Comparative study between sheep and goats on rumenic acid and vaccenic acid in milk fat under the same dietary treatments. *Livestock Science*. 119(1): 87-94.
- Tsiplakou, E., Flemetakis, E., Kalloniati, C. and Zervas, G. 2011. Differences in mRNA lipogenic gene expression in the subcutaneous adipose tissue of sheep and goats under the same dietary treatments. *Small Ruminant Research*. 99(2): 110-115.
- Tsiplakou, E., Flemetakis, E., Kalloniati, C., Papadomichelakis, G., Katinakis, P. and Zervas, G. 2009. Sheep and goats differences in CLA and fatty acids milk fat content in relation with mRNA stearoyl-CoA desaturase and lipogenic genes expression in their mammary gland. *Journal of Dairy Research*. 76(04): 392-401.
- Tsiplakou, E., Flemetakis, E., Kouri, E.D., Sotirakoglou, K. and Zervas, G. 2015. The effect of long term under-and over-feeding on the expression of genes related to lipid metabolism in mammary tissue of sheep. *Journal of Dairy Research*. 82(01): 107-112.
- Turner, K. E., Belesky, D. P., Cassida, K. A., and Zerby, H. N. 2014. Carcass merit and meat quality in Suffolk lambs, Katahdin lambs, and meat-goat kids finished on a grass–legume pasture with and without supplementation. *Meat Science*. 98(2): 211-219.
- Turner, T. 2010. Influence of oilseed supplementation on ruminant meat and milk with emphasis on fatty acids. PhD Thesis. Swedish University of Agricultural Sciences, Uppsala, Sweden.
- Van Soest, P.J., Robertson, J.B.and Lewis, B.A. 1991. Methods for dietary fiber, neutral detergent fiber, and non-starch polysaccharides in relation to animal nutrition. *Journal of Dairy Science*. 74: 3583–3597.
- Veum, T.L. 2004. Feedstuffs: High-Energy Sources. In *Encyclopedia of Animal Science*. Eds. Pond, W.G. and Bell, A.W. pp. 387. Marcel Dekker, New York. USA.
- Vlaeminck, B., Fievez, V., Cabrita, A.R.J., Fonseca, A.J.M. and Dewhurst, R.J. 2006. Factors affecting odd-and branched-chain fatty acids in milk: A review. *Animal Feed Science and Technology*. 131(3): 389-417.
- Wakil, S.J. and Abu-Elheiga, L.A. 2009. Fatty acid metabolism: target for metabolic syndrome. *Journal of Lipid Research*. 50(Supplement): 138-143.
- Wallace, R.J., McKain, N., Shingfield, K.J. and Devillard, E. 2007. Isomers of conjugated linoleic acids are synthesized via different mechanisms in ruminal digesta and bacteria. *Journal of Lipid Research*. 48(10): 2247-2254.

- Wanapat, M., Petlum, A., Chanthai, S. 2005. Effects of level of urea and coconut oil on rumen ecology, milk yield and composition in lactating dairy cows fed on urea-treated rice straw. In 'Workshop on Making Better Use of Local Feed Resources'. pp. 23-25.
- Wanapat, M., Mapato, C., Pilajun, R. and Toburan, W. 2011. Effects of vegetable oil supplementation on feed intake, rumen fermentation, growth performance, and carcass characteristic of growing swamp buffaloes. *Livestock Science*. 135(1): 32-37.
- Warren, D.M., 2015. *Small Animal Care and Management*. Cengage learning, USA.
- West, D.B., Delany, J.P., Camet, P.M., Blohm, F., Truett, A.A. and Scimeca, J. 1998. Effects of conjugated linoleic acid on body fat and energy metabolism in the mouse. *American Journal of Physiology-Regulatory*, *Integrative and Comparative Physiology*. 275(3): 667-672.
- Williams, A. G. 1989. Metabolic activities of rumen protozoa. In *The Roles of Protozoa and Fungi in Ruminant Digestion*. Eds. Nolan, J.V., Leng, R.A. and Demeyer, D. I. pp.97-126. Penambul Books, Armidale, Australia.
- Williams, A.G. and Coleman, G.S. 2012. *The Rumen Protozoa*. Springer-Verlag, UK.
- Williams, A.G. and Withers, S.E. 1993. Changes in the rumen microbial population and its activities during the refaunation period after the reintroduction of ciliate protozoa into the rumen of defaunated sheep. *Canadian Journal of Microbiology*. 39(1): 61-69.
- Williams, B.A. 2000a. Cumulative gas-production techniques for forage evaluation. In *Forage Evaluation in Ruminant Nutrition*. Eds. Givens, D.I., Owen, E., Axford,R.F.E. and Omed, H.M. pp.189-213. CABI Publishing, Wallingford, UK.
- Williams, C.M. 2000b. Dietary fatty acids and human health. *Annales de Zootechnie.* 49(3): 165-180.
- Williams, P. 2007. Nutritional composition of red meat. *Nutrition and Dietetics*. 64(s4): 113-119.
- Williamson, C.S., Foster, R.K., Stanner, S.A. and Buttriss, J.L. 2005. Red meat in the diet. *Nutrition Bulletin*. 30(4): 323-355.
- Windham, W.R. and Akin, D.E. 1984. Rumen fungi and forage fiber degradation. *Applied and Environmental Microbiology*. 48(3): 473-476.
- Wong, J.W., Hashimoto, K. and Shibamoto, T. 1995. Antioxidant activities of rosemary and sage extracts and vitamin E in a model meat system. *Journal of Agricultural and Food Chemistry*. 43(10): 2707-2712.

- Wood, J.D. 1993. The relationship between the composition and quality of meat. Proceedings 39th International Congress Meat Science Technology. Calgary: pp. 37-46.
- Wood, J.D., Richardson, R.I., Nute, G.R., Fisher, A.V., Campo, M.M., Kasapidou, E., Sheard, P.R. and Enser, M. 2004. Effects of fatty acids on meat quality: a review. *Meat Science*. 66(1): 21-32.
- Woods, V.B. and Fearon, A.M. 2009. Dietary sources of unsaturated fatty acids for animals and their transfer into meat, milk and eggs: A review. *Livestock Science*. 126(1): 1-20.
- Wright, A.D.G. 2015. Rumen Protozoa. In *Rumen Microbiology: From Evolution to Revolution.* Eds. Puniya, A.K., Singh, R. and Kamra, D.N. pp. 113-120. Springer India.
- Wright, L.I., Scanga, J.A., Belk, K.E., Engle, T.E., Tatum, J.D., Person, R.C., McKenna, D.R., Griffin, D.B., McKeith, F.K., Savell, J.W. and Smith, G.C. 2005. Benchmarking value in the pork supply chain: Characterization of US pork in the retail marketplace. *Meat Science*. 71(3): 451-463.
- Yang, S.L., Bu, D.P., Wang, J.Q., Hu, Z.Y., Li, D., Wei, H.Y., Zhou, L.Y. and Loor, J.J. 2009. Soybean oil and linseed oil supplementation affect profiles of ruminal microorganisms in dairy cows. *Animal.* 3(11): 1562-1569.
- Yousefi, A. R., Kohram, H., Shahneh, A. Z., Nik-Khah, A., and Campbell, A. W. 2012. Comparison of the meat quality and fatty acid composition of traditional fat-tailed (Chall) and tailed (Zel) Iranian sheep breeds. *Meat Science*. 92(4): 417-422.
- Yu, Y., Lee, C., Kim, J., Hwang, S. 2005. Group-specific primer and probe sets to detect methanogenic communities using quantitative real-time polymerase chain reaction. *Biotechnology and Bioengineering*. 89(6): 670–679.
- Yusuf, A.M., Olafadehan, O.A., Obun, C.O., Inuwa, M., Garba, M.H. and Shagwa, S.M. 2009. Nutritional evaluation of sheabutter fat in fattening of Yankasa sheep. *Pakistan Journal of Nutrition*. 8(7): 1062-1067.
- Zeb, A. and Ullah, F. 2016. A Simple Spectrophotometric Method for the Determination of Thiobarbituric Acid Reactive Substances in Fried Fast Foods. *Journal of Analytical Methods in Chemistry*. 2016: 9412767
- Zhang, C.M., Guo, Y.Q., Yuan, Z.P., Wu, Y.M., Wang, J.K., Liu, J.X. and Zhu, W.Y. 2008. Effect of octadeca carbon fatty acids on microbial fermentation, methanogenesis and microbial flora *in vitro*. *Animal Feed Science and Technology*. 146(3): 259-269.

- Zhang, H., Zhang, X., Wang, Z., Dong, X., Tan, C., Zou, H., Peng, Q., Xue, B., Wang, L. and Dong, G. 2015. Effects of dietary energy level on lipid metabolism-related gene expression in subcutaneous adipose tissue of Yellow breed× Simmental cattle. *Animal Science Journal*. 86(4): 392-400.
- Zhou, M., Chen, Y. and Guan, L.L. 2015. Rumen bacteria. In Rumen Microbiology: From Evolution to Revolution. Eds. Puniya, A.K., Singh, R. and Kamra, D.N. pp. 79-95. Springer India.
- Zhou, M., Hernandez-Sanabria, E. and Le, L.G. 2009. Assessment of the microbial ecology of ruminal methanogens in cattle with different feed efficiencies. *Applied and Environmental Microbiology*. 75(20): 6524– 6533.

