

UNIVERSITI PUTRA MALAYSIA

ASSESSMENT OF Bambusa vulgaris Schrad. ex J.C.Wendl. AS POTENTIAL PHYTOREMEDIATION AGENT FOR HEAVY METALS

WAN RAFIEKAL BINTI WAN ABDUL RAHIM

IPTPH 2016 2



ASSESSMENT OF *Bambusa vulgaris* Schrad. *ex* J.C.Wendl. AS POTENTIAL PHYTOREMEDIATION AGENT FOR HEAVY METALS

By

WAN RAFIEKAL BINTI WAN ABDUL RAHIM

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Science

March 2016

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

ASSESSMENT OF *Bambusa vulgaris* Schrad. *ex* J.C.Wendl. AS POTENTIAL PHYTOREMEDIATION AGENT FOR HEAVY METALS

By

WAN RAFIEKAL BINTI WAN ABDUL RAHIM

March 2016

Chairman: Hazandy Abdul Hamid, PhD

Faculty: Institute of Tropical Forestry and Forest Products

Heavy metal pollution had becoming one of the most serious environmental problems today. These pollutants have highly affected agricultural productivity, human health as well as disrupting the world's ecological biodiversity system. Experimental research conducted had undergone the phytoremediation process which is bio-based low cost alternative technology, environmental friendly and represents the efficiency of using plants for treatment of heavy metal. Bamboos are potentially good plants for phytoremediation due to their widespread distribution, simple and well-known propagation mode, the full range of species and their potential as an additional use for raw material. They are fast-growing, grow more rapidly than other timber species and have various actual and possible applications for income generation and environmental conservation. This study aims in evaluate the potential of B. vulgaris as phytoremediation agent for heavy metal. The plants were grown up uniformly in lysimeter pot for three months in the presence of heavy metal copper, zinc and iron with different level of concentration to quantify the toxic effects of these metals on morphological and physiological attributes of B. vulgaris. Leachates from lysimeter were being collected periodically and undergoing screening process to determine the contents of remaining heavy metal in samples. The close positive relationship of heavy metal analysis that exhibits low concentration of metal before and after treated or infiltrated by B. vulgaris indicates that it can be used as bioaccumulator plants of heavy metal pollution by absorbing excess heavy metal from the polluted soils and water. From the study, B. vulgaris most efficient in uptake iron compared to zinc and copper. The findings indicated that heavy metals copper zinc and iron are considered to be important for plant growth due to the positive response in morphological and physiological attributes of *B. vulgaris* at low concentration however brings phytotoxicity effects at high level of concentration and time. *B.vulgaris* performance was increased at early weeks and in low concentration metal level indicated it's tolerance to the heavy metals, making it a prospective phytoremediator species. Thus, we can conclude that *B. vulgaris* has potential to be a phytoremediator plant and can be grown up on moderately heavy metal contaminated soil.



Abstrak tesis yang dikemukakan kepada senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PENILAIAN KEATAS *Bambusa vulgaris* Schrad. *ex* J.C.Wendl. YANG BERPOTENSI SEBAGAI AGEN FITOPEMULIHAN UNTUK LOGAM BERAT

Oleh

WAN RAFIEKAL BINTI WAN ABDUL RAHIM

Mac 2016

Pengerusi: Hazandy Abdul Hamid, PhD

Fakulti: Institut Perhutanan Tropika dan Produk Hutan

Pencemaran logam berat telah menjadi salah satu masalah alam sekitar yang paling serius pada hari ini. Pencemaran ini telah mempengaruhi produktiviti pertanian, kesihatan manusia dan juga mengganggu sistem biodiversiti ekologi di dunia. Kajian dijalankan mengunakan proses fitopemulihan yang merupakan teknologi alternatif yang melibatkan kos rendah berasaskan biologi, mesra alam sekitar menggunakan tumbuh-tumbuhan untuk rawatan logam berat. Buluh berpotensi untuk digunakan dalam fitopemulihan kerana ianya tumbuh secara meluas, pembiakannya yang mudah dan terkenal, mempunyai banyak spesies dan sebagai penggunaan tambahan untuk bahan mentah. Ianya tumbuh lebih cepat daripada spesies kayu lain dan mempunyai pelbagai kelebihan untuk menjana pendapatan dan pemuliharaan alam sekitar. Kajian ini bertujuan menilai potensi B. vulgaris sebagai ejen fitopemulihan untuk logam berat. Pokok buluh ditanam secara seragam di dalam lysimeter selama tiga bulan dengan kehadiran logam berat tembaga, zink dan besi dengan tahap yang berbeza untuk mengukur kesan toksik logam ini ke atas sifat-sifat morfologi dan fisiologi *B. vulgaris*. Air larut resap dari lysimeter dikumpul secara berkala dan menjalani proses saringan untuk menentukan kandungan baki logam berat dalam sampel. Hubungan positif analisis logam berat yang mempamerkan kepekatan rendah logam sebelum dan selepas dirawat oleh B. vulgaris menunjukkan bahawa ia boleh digunakan sebagai tumbuhan biopengumpul pencemaran logam berat dengan menyerap logam berat yang berlebihan dari tanah tercemar dan air. Dari kajian ini, B. vulgaris paling berkesan dalam pengambilan besi berbanding zink dan tembaga. Dapatan kajian menunjukkan bahawa logam berat tembaga, zink dan besi dianggap penting untuk pertumbuhan tumbuhan berikutan keputusan yang positif dalam sifat-sifat morfologi dan fisiologi *B. vulgaris* pada kepekatan yang rendah tetapi membawa kesan fitotosik di kepekatan yang tinggi dalam tempoh masa yang lama. Pertumbuhan *B. vulgaris* telah meningkat pada minggu-minggu awal dan pada tahap kepekatan logam rendah menunjukkan toleransi kepada logam berat, menjadikannya satu spesies yang boleh digunakan untuk fitopemulihan. Oleh itu, kesimpulan boleh dibuat bahawa *B. vulgaris* mempunyai potensi untuk fitopemulihan dan boleh ditanam di atas tanah yang sederhana tercemar dengan logam berat.



ACKNOWLEDGEMENTS

Bismillahirrahmanirrahim, first and foremost, all praise to Allah S.W.T, the most Gracious and the most Merciful, for His blessing and for giving me mental and physical strengths to accomplish this project.

I would like to express my special gratitude and respect to my supervisor, Associate Professor Dr. Hazandy Abdul Hamid for the valuable advices, guidance, suggestions, encouragement and support during my project including the preparations of this thesis. I am also grateful and sincerely thankful to other committee member Dr Azmy Mohamed and Associate Professor Dr Arifin Abdu who gave me a guide and constructive comment for the project implementation.

I wish to thank plant physiology lab members in Faculty of Forestry, UPM, assistant science officer Mr Kamil Ismail and Ms Yap Mei Ling also Mr Alagan for their cooperation in providing me equipments, their guidance and continuous assistance throughout this study. I am also greatly indebted to Mr Jamil, for his assistance in guiding me during laboratory analysis works at Faculty of Agriculture, UPM. My deepest appreciation to be extended as well to late Mr. Zakaria and staff from Forest Research Institute Malaysia Nursery for their kindness and assistance in provide the materials.

To all my friends, Noor Haslidawati, Nurul Rusmida, Asmaa' Widad, Nurul Hijrah, Kiasatina and all staffs of Institute Tropical Forestry & Forest Product especially Mrs Shuhada, thank you very much for being helpful, gives me courage and technical support during this study.

Finally, a very sincere gratitude to my family especially my dearest father, mother and siblings for their love, prayers and unconditional support that I always needed and makes me focused throughout this endeavor. Without the invaluable assistance of the people mentioned above, this study would have never reach completion. Thank you very much and all supports were deeply appreciated. May Allah S.W.T bless and repay all your kindness.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Hazandy Abdul Hamid, PhD

Associate Professor Institute Tropical Forestry and Forest Products Universiti Putra Malaysia (Chairman)

Arifin Abdu, PhD

Associate Professor Faculty of Forestry Universiti Putra Malaysia (Member)

BUJANG BIN KIM HUAT, PhD Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by Graduate Student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fullyowned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:
Name and Matric No.:	

Declaration by Members of Supervisory Committee

This is to confirm that:

5

 the research conducted and the writing of this thesis was under our supervision; supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	
Signature: Name of Member of Supervisory Committee:	

TABLE OF CONTENTS

				Page
ABSTR	ACT			i
ABSTR	AK			iii
ACKNO	V			
APPRO	vi			
DECLA		viii		
LIST O				x
LIST O				xv
LIST O	F ABB	REVIAT	IONS	xvii
СНАРТ	ED			
1		ODUCTI	ON	
1	1.1	Backgro		1
	1.1	-	n Statement	2
	1.2	Objectiv		4
	1.5	Objectiv		Т.
2	LITER	ATURE	REVIEW	
	2.1	Bamboo	J	5
		2.1.1	Bambusa vulgaris	7
	2.2	Impa <mark>ct</mark>	of Heavy Metals on Plants	
		2.2.1	Copper	9
		2.2.2	Zinc	10
		2.2.3	Iron	11
	2.3	Phytore	emediation	
		2.3.1	Advantages and Limitations of Phytoremediation	14
3	мате	RTALS A	AND METHODOLOGY	
-	3.1	Materia		
		3.1.1	Bambusa vulgaris	16
		3.1.2	Lysimeter	17
		3.1.3	Heavy Metal Stock Solution	18
	3.2	Method	dology	
		3.2.1	Heavy Metal Analysis	19
		3.2.2	Morphological Attributes	20
		3.2.3	Physiological Attributes	20
			3.2.3.1 Chlorophyll Content	20
			3.2.3.2 Chlorophyll Fluorescence	21
			3.2.3.3 Gas Exchange	22
	3.3 Sta	atistical A	Analysis	23

4

 \bigcirc

RESULTS AND DISCUSSIONS

	4.1 Heavy Metal Analysis				
		4.1.1	Copper		24
		4.1.2	Zinc		26
		4.1.3	Iron		28
	4.2	Morpholo	gical Attribu	ites	
		4.2.1	Height of C	Culm	32
		4.2.2	Diameter o	f Culm	34
		4.2.3	Number of	Branches	36
		4.2.4	Number of	Leaves	38
		4.2.5	Number of	New Culms	41
		4.2.6	Dry Weight	of Root	43
	4.3		jical Attribut		
		4.3.1	Chlorophyll	Content	45
		4.3.2		Fluorescence	
			4.3.2.1	Minimal Florurescene, (F ₀)	49
			4.3.2.2	Maximal Fluorescence, (F _m)	51
			4.3.2.3	Variable Flourescence,	53
			4.3.2.4	$F_{v}(F_{m}-F_{0})$	55
			4.3.2.4	Maximal Quantum Yield of PS II, (F_v/F_m)	55
		4.3.3	Gas Excha	nge	
			4.3.3.1	Net Photosynthesis Rate (A _{net})	58
			4.3.3.2	Stomatal Conductance (G_s)	60
			4.3.3.3	Transpiration Rate (E)	61
			4.3.3.4	Intercellular $CO_2(C_i)$	63
			4.3.3.5	Air Vapor Pressure Deficit (VpdL)	65
5	SUMM/	ARY CON	CLUSION /	AND RECOMMENDATIONS	68
REFERENCES 70				70	
				83	
BIODATA OF STUDENT 85					

LIST OF TABLES

Table		Page
2.1	Applications of phytoremediation	13
2.2	Advantages and limitations of phytoremediation	15
4.1	Mean (± standard deviation) leachate analysis for copper metal	25
4.2	Summary of paired sample t-test between top and bottom leachate analysis for copper metal	26
4.3	Mean (± standard deviation) leachate analysis for zinc metal	27
4.4	Summary of paired sample t-test between top and bottom leachate analysis for zinc metal	28
4.5	Mean (± standard deviation) leachate analysis for iron metal	29
4.6	Summary of paired sample t-test between top and bottom leachate analysis for iron metal	30
4.7	Mean (± standard deviation) height of <i>B. vulgaris</i> culm in different treatment	33
4.8	Summary of repeated measures two ways ANOVA within time and treatment on the height of <i>B. vulgaris</i> culm	34
4.9	Mean (± standard deviation) diameter of <i>B. vulgaris</i> culm in different treatment	35
4.10	Summary of repeated measures two ways ANOVA within time and treatment on the diameter of <i>B. vulgaris</i> culm	36
4.11	Mean (\pm standard deviation) number of <i>B. vulgaris</i> branches in different treatment	37
4.12	Summary of repeated measures two ways ANOVA within time and treatment on number of <i>B. vulgaris</i> branches	38
4.13	Mean (± standard deviation) number of <i>B. vulgaris</i> leaves in different treatment	39
4.14	Summary of repeated measures two ways ANOVA within time and treatment on the number of <i>B. vulgaris</i> leaves	40

 \bigcirc

	4.15	Number of <i>B. vulgaris</i> new culms emerged in different treatment	42
	4.16	Summary of repeated measures two ways ANOVA within time and treatment on number of new culms of <i>B. vulgaris</i>	43
	4.17	Mean value (±standard deviation) chlorophyll content of <i>B. vulgaris</i> between treatments	46
	4.18	Summary of repeated measures two ways ANOVA within time and treatment on chlorophyll content of <i>B. vulgaris</i>	46
	4.19	Response patterns of CF in one second measurement between treatment in three months	49
	4.20	Mean value (±standard deviation) of minimal fluorescence between treatments	50
	4.21	Summary of repeated measures two ways ANOVA within time and treatment on minimal fluorescence of <i>B. vulgaris</i>	51
	4.22	Mean value (±standard deviation) of maximal fluorescence between treatments	52
	4.23	Summary of repeated measures two ways ANOVA within time and treatment on maximal fluorescence of <i>B. vulgaris</i>	53
	4.24	Mean value (±standard deviation) of variable fluorescence between treatments	54
	4.25	Summary of repeated measures two ways ANOVA within time and treatment on F _v , variable fluorescence of <i>B. vulgaris</i>	55
	4.26	Mean value (±standard deviation) of maximum yield of photosystem II between treatment	56
	4.27	Summary of repeated measures two ways ANOVA within time and treatment on maximum yield of photosystem II	57
	4.28	Mean value (±standard deviation) of net photosynthesis rate between treatments	58
	4.29	Summary of repeated measures two ways ANOVA within time and treatment on photosynthesis rate of <i>B. vulgaris</i>	58
Θ	4.30	Mean value (±standard deviation) of stomatal conductance between treatments	61
	4.31	Summary of repeated measures two ways ANOVA within time and treatment on stomatal conductance of <i>B. vulgaris</i>	61
4			

4.32	Mean value (±standard deviation) of transpiration rate
	between treatments

- 4.33 Summary of repeated measures two ways ANOVA within time and treatment on transpiration rate of *B. vulgaris*
- 4.34 Mean value (±standard deviation) of intercellular CO₂ between treatments
- 4.35 Summary of repeated measures two ways ANOVA within time and treatment on intercellular CO₂ of B. vulgaris
- 4.36 Mean value (±standard deviation) of air vapor pressure deficit between treatments
- 4.37 Summary of repeated measures two ways ANOVA within time and treatment on vapor pressure deficit of B. vulgaris

66

62

63

64

65

66

LIST OF FIGURES

Table		Page
3.1	B. vulgaris sapling before being transplanted	17
3.2	Ilustration of plant in lysimeter (cross section view)	17
3.3	Solar Thermo Atomic Absorption Spectrometer (AAS)	19
3.4	Water leachate samples undergo filtration process to remove seuspended solids	20
3.5	Chlorophyll Meter, SPAD 502	21
3.6	(i) Chlorophyll Fluorimeter, HandyPEA (Hansatech CF Model) and (ii) Leafclips	22
3.7	Portable Photosynthesis System, LiCor 6400	23
4.1	Concentration of copper in leachate before and after infiltrate by <i>B. vulgaris</i>	25
4.2	Concentration of zinc in leachate before and after infiltrate by <i>B. vulgaris</i>	27
4.3	The concentration of iron in locating before and after infiltrating by <i>B. vulgaris</i>	29
4.4	Height of <i>B. vulgaris</i> culm within treatment periods	33
4.5	Diameter of <i>B. vulgaris</i> culm within treatments period	35
4.6	Number of <i>B. vulgaris</i> branches within treatments period	37
4.7	Number of <i>B. vulgaris</i> leaves within treatments period	39
4.8	Leaf colour of <i>B. vulgaris</i> changes after 4th week of heavy metal treatment	41
4.9	Number of <i>B. vulgaris</i> new culms emerged within treatments period	42
4.10	Root dry weight of B. vulgaris root between treatments	43
4.11	<i>B. vulgaris</i> root condition differences between treatment after harvested	44
4.12	Chlorophyll content of <i>B. vulgaris</i> within treatments period	45

6

4.13	Chlorophyll fluorescence on 1^{st} month (11^{th} June 2014)		
4.14	Chlorophyll fluorescence on 2 nd month (11 th July 2014)		
4.15	Chlorophyll fluorescence on 3 rd month (12 th Aug 2014)	48	
4.16	Minimal fluorescence of <i>B. vulgaris</i> within treatments period 5		
4.17	Maximal fluorescence of <i>B. vulgaris</i> within treatments period 5		
4.18	Variable fluorescence of <i>B. vulgaris</i> within treatments period 54		
4.19	Maximum quantum yield photosystem II of <i>B. vulgaris</i> within treatments period	56	
4.20	Net photosynthesis rate of <i>B. vulgaris</i> within treatments period	58	
4.21	Stomatal conductance of <i>B. vulgaris</i> within treatments period	60	
4.22	Transpiration rate of <i>B. vulgaris</i> within treatments period 62		
4.23	Intercellular CO ₂ of <i>B. vulgaris</i> within treatments period	64	
4.24	Air vapor pressure deficit of <i>B. vulgaris</i> within treatments period	65	

(G)

LIST OF ABBREVIATIONS

	PPM	Part per million
	ANOVA	Analysis of variance
	AAS	Atomic absorption spectrometer
	SPAD	Soil plant analysis development
	ATP	Adenosine triphosphate
	DNA	Deoxyribonucleic acid
	VOC	Volatile organic compounds
	FRIM	Forest Research Institute Malaysia
	CRD	Completely randomized design
	SPSS	Statistical package social science
	HSD	Honest significant difference
	ZnSO₄	Zinc (II) sulphate
	CuSO₄	Copper (II) sulphate
	FeSO₄	Fe (II) sulphate
	CO ₂	Carbon dioxide
	F _o	Constant minimal fluorescence in dark-adapted plant
	F _m	Maximum fluorescence in dark-adapted plant
	F _v	Variable fluorescence for a plant in dark-adapted
	F,/Fm	Maximum yield of photosystem II
\bigcirc	PSII	Photosystems II
	A _{net}	Net photosynthesis rate
	G _s	Stomatal conductance
	E	Transpiration rate

xvii

- C_i Intercellular carbon dioxide
- VpdL Air vapor pressure deficit
- Eh Oxidation potential
- pH Potential of Hydrogen
- RNA Ribonucleic acid

G

ROS Reactive oxygen species



CHAPTER 1

INTRODUCTION

This chapter highlighted the background of the study, problem statement and objectives of the study. The background of the study discusses generally about the environmental pollution by heavy metals and the role or functions of bamboo to environment. The problem statement discusses the importance of the study and why it was conducted.

1.1 Background

Environment defined as the communities we are living in. It also can be described as total circumstances surrounding a living thing or group of living things, the mix of external physical conditions which influence the growth, development and survival of them (Gore, 1997). Pollution is an accidental or deliberate contamination of the environment by waste contains pollutant due to the human activities. These pollutant substances mostly exist over a set or tolerance limit. Therefore, environmental pollution occurs with the presence of a pollutant at poisonous level in land, water and air likely causes objectionable impacts, lessening the quality of life and harmful to the other organisms in those polluted environment.

Our environment has been contaminated with organic and inorganic pollutants, because they are easily discharged into the surroundings through wide range of ways. Surface water is easily polluted by domestic wastewater effluent, runoff, and rainwater. Due to the phenomenal growth of industrialization, urbanization, and population, many surface water systems become highly contaminated (Qian *et al.*, 2007).

Chehregani (2009) states that environmental contamination by heavy metals is a worldwide catastrophe that happen because of human activities like mining, energy and fuel production, power transmission, melting operations, sludge dumping, and intensive agriculture. High concentrations of heavy metals that bring harmful toxic effects to the environment are known as environmental pollutants. Copper (Cu), zinc (Zn), cadmium (Cd), lead (Pb), nickel (Ni) and chromium (Cr) metals normally known to be severe environmental pollutants.

Heavy metal contamination to soil and water contributes a serious environmental and health issues. Metals accumulate in soil and water due to anthropogenic contamination through fertilizer, organic manure applications, irrigation, industrial and municipal wastes, and wet and/or dry deposits (Doelschet *et al.*, 2010). They are essential as micronutrient to plants and normally exist in the background of soils at different levels however they become toxic at higher concentrations. Plant roots excessively absorbed heavy metal ion which present at high level in the environment and trans located to other part of plants and lead to impaired metabolism and growth reduction of the plant. Besides that, contaminated soils with excess metal also resulted in the reduction of soil fertility, microbial activity, and yield.

Kleinhenz (2001) stated that plants have a unique capability to concentrate essential and non-essential elements from the environment into their tissues. Green plants normally use sunlight, the most important source of energy in photosynthesis to speed up the concentration process. Terrestrial plants have an extensive root system and an advanced uptake mechanism in their continuous search for water and mineral resources. Bamboo for example, a terrestrial plant that is known for its tolerance to a broad range of stressors, high biomass production and growth development rate as well as large and extended root system which probably shows good abilities for water filtration and nutrient absorption.

1.2 Problem Statement

Over the past several decades, health and pollution of the environment have becoming more apparent throughout the world. Industrialization, urbanization and rapid growth in population are the sources for the increasing of pollution rate. Metal pollution had becoming one of the most serious environmental problems today. Pollution of biosphere by toxic metals had accelerated dramatically since the beginning of the industrial revolution because of human activities such as mining and smelting of metals, energy and fuel production, electroplating, sewage, pesticide application, fertilizer, municipal waste generation and others (Srivastava S., 2007).

These organic and inorganic pollutants have highly affected agricultural productivity, human health as well as disrupting the world's ecological biodiversity system (Bridge, 2004). Contaminants can lead to land or groundwater and the toxic elements end up entered into the food chain. Due to the high public awareness regarding to this, environmental regulations have been developed to prevent pollution and also to remediate areas where contamination has occurred (Devos J., 2001).

The contamination of the water resource is a critical environmental issue. In many parts of the developing country, uncontrolled and inappropriate removal of wastewater causes severe pollution issues. However, by reusing wastewater, it can help to improve global water shortages mainly in developing country where there are no facilities for safe disposal of wastewater. In this way, phytoremediation was one of the potential outcomes on which effort were focused on, combining land cover and soil stabilisation, the utilization of the growing crops, and their possibilities on cleaning of the wastewater.

Experimental research conducted had undergone the phytoremediation process which is bio-based low cost alternative technology, environmental friendly and represents the efficiency of using plants for treatment of environmental contaminant. About 700 species of plants have been reported in accumulation of different contaminants (Xi et al., 2010). Metal accumulators are found in a large number of plant families, but most are the Brassicaceae family (Verbruggen et al., 2009) and are associated with slow plant growth and low biomass yields (Epelde et al., 2008), so there is an urgent need for identification of other plant species having fast growth and greater biomass production.

Bamboos are potentially good plants for phytoremediation due to their widespread distribution, simple and well-known propagation mode, the full range of species and their potential as an additional use for raw material. They are fast-growing, grow more rapidly than other timber species and have various actual and possible applications for income generation and environmental conservation. Because of these attributes, bamboo could have many offers in slum areas which it is usually being used to clean up waste water.

Bamboo species selected was *B. vulgaris* which is known as a highly useful plant, providing poles and timber for scaffolding, fencing, roofs, furniture, and many craft items but plays a very valuable environmental role. It is water loving plant and its extensive root system probably can take up nutrients such as nitrogen, phosphorous or dangerous contaminants such as heavy metals which some of them are locked in the root system. Hence, it suitable for disposal of effluents and reduce of waste water pollution. Planted close by waterways, ditches and creeks, *B. vulgaris* can catch the overabundance nutrients in the runoff water keeping from entering adjacent streams. Waste water from agricultural area, animal farming, manufacturing and sewerage treatment plants can be utilized to irrigate crops, consequently change it into valuable biomass. In perspective of these features, *B. vulgaris* was used as main materials for the study of the potential utilisation in plants to improve water and nutrient supplies, towards providing an environmentally compatible method for wastewater disposal.

1.3 Objectives

The main objective of this study was to evaluate the performance of *B.vulgaris* mechanism of pollutant removal and its potential for the elimination of metal contaminated through phytoremediation.

The specific objectives of this study were:

- i. To identify the ability of *B. vulgaris* to take up heavy metals contaminants
- ii. To evaluate the phytotoxicity effects of *B. vulgaris* on heavy metals towards their growth and physiological attributes

REFERENCES

- Abdu, A., Aderis, N., Abdul Hamid, H., Majid, N. A., Muhamad, N., Jusop, S. & Ahmad, K. (2011). Using orthosiphon stamineus B. for Phytoremediation of heavy metals in soils amended with sewage sludge. *American Journal of Applied Sciences*, 8(4): 323-331.
- Andrade, L. R., Farina, M., & Amado Filho, G. M. (2004). Effects of copper on Enteromorpha flexuosa (Chlorophyta) in vitro. *Ecotoxicology and environmental safety*, 58(1): 117-125.
- Arfi, V., Bagoudou, D., Korboulewsky, N., & Bois, G. (2009). Initial efficiency of a bamboo grove–based treatment system for winery wastewater. *Desalination*, 246(1): 69-77.
- Babatunde, A. I., Abiola, O. K. I., Osideko, O. A., & Oyelola, O. T. (2009). Kinetic and equilibrium studies on the adsorption of Cu²⁺ and Zn²⁺ ions from aqueous solutions by bamboo root biomassa. *African Journal of Biotechnology*,8(14).
- Becker, M., & Asch, F. (2005). Iron toxicity in rice—conditions and management concepts. *Journal of Plant Nutrition and Soil Science*, 168(4): 558-573.
- Bridge, G. (2004). Contested terrain: mining and the environment. *Annual Review Environmental Resource*, 29: 205-259.
- Brune, A., Urbach, W., & Dietz, K. J. (1994). Compartmentation and transport of zinc in barley primary leaves as basic mechanisms involved in zinc tolerance. *Plant, Cell & Environment*, 17(2): 153-162.
- Cakmak, I., Marshner, H. (1993). Effect of zinc nutritional status on superoxide radical and hydrogen peroxide scavenging enzymes in bean leaves. *In: Barrow NJ (ed) Plant nutrition-from genetic engineering field practice*. 133–137.
- Chaney, R. L., Malik, M., Li, Y. M., Brown, S. L., Brewer, E. P., Angle, J. S., & Baker, A. J. (1997). Phytoremediation of soil metals. *Current opinion in Biotechnology*, 8(3): 279-284.

- Chehregani, A., Noori, M., & Yazdi, H. L. (2009). Phytoremediation of heavymetal-polluted soils: screening for new accumulator plants in Angouran mine (Iran) and evaluation of removal ability. *Ecotoxicology and environmental safety*, 72(5): 1349-1353.
- Chen, J. R. (2014). Effect of copper toxicity on root morphology, ultrastructure, and copper accumulation in moso bamboo (*Phyllostachys pubescens*) Z Naturforsch C 69, 399–406.
- Clijsters, H., Cuypers A., Vangronsveld J., (1999). Physiological responses to heavy metals in higher plants; Defence against oxidative stress. *Z. Naturforsch.* 54: 730-734.
- Cobbett, C. S. (2000). Phytochelatin biosynthesis and function in heavy-metal detoxification. *Current opinion in plant biology*, 3(3): 211-216.
- Collin B., Doelsch E., Keller C., Panfili F. & Meunier J. D. (2012). Distribution and variability of silicon, copper and zinc in different bamboo species. *Plant soil*, 351: 377–387.
- Cooke, J. A., & Johnson, M. S. (2002). Ecological restoration of land with particular reference to the mining of metals and industrial minerals: A review of theory and practice. *Environmental Reviews*, 10(1): 41-71.
- Dahmani-Muller, H., Van Oort, F., Gelie, B., & Balabane, M. (2000). Strategies of heavy metal uptake by three plant species growing near a metal smelter. *Environmental pollution*, 109(2): 231-238.
- De Dorlodot, S., Lutts, S., & Bertin, P. (2005). Effects of ferrous iron toxicity on the growth and mineral composition of an interspecific rice. *Journal of plant nutrition*, 28(1): 1-20.
- De Vas, C.H.R., Schat, H., De Waal, M.A.M., Voojs, R. & Ernst, W.H.O. (1991). Increased ressistance to copper-induced damage of root cells plasmalemma in copper tolerant *Silence cucubalus. Physiologia Plantarum*, 82: 523-528.
- De Vries, W., Lofts, S., Tipping, E., Meili, M., Groenenberg, J. E., & Schütze, G. (2007). Impact of soil properties on critical concentrations of cadmium,

lead, copper, zinc, and mercury in soil and soil solution in view of ecotoxicological effects. In *Reviews of Environmental Contamination and Toxicology* (pp. 47-89). Springer New York.

- Demirevska-Kepova, K., Simova-Stoilova, L., Stoyanova, Z., Hölzer, R., & Feller, U. (2004). Biochemical changes in barley plants after excessive supply of copper and manganese. *Environmental and Experimental Botany*, 52(3): 253-266.
- Devi, T. S., & Singh, T. B. (2010) A Study on the Environmental Role and Economic Potential of Arundinaria callosa. Munro: *VOLUME 3 Bamboo and the Environment*, 2.
- Devos, J., Gielis, J., (2001). *Bamboo for Europe*. International Network for Bamboo and Rattan Newsletter (in the press).
- Dinelli, E., & Lombini, A. (1996). Metal distributions in plants growing on copper mine spoils in Northern Apennines, Italy: the evaluation of seasonal variations. *Applied Geochemistry*, 11(1): 375-385.
- Doelsch, E., Masion, A., Moussard, G., Chevassus-Rosset, C., & Wojciechowicz, O. (2010). Impact of pig slurry and green waste compost application on heavy metal exchangeable fractions in tropical soils. *Geoderma*,155(3): 390-400.
- Downton, W. J. S., Grant, W. J. R., & Robinson, S. P. (1985). Photosynthetic and stomatal responses of spinach leaves to salt stress. *Plant Physiology*,78(1): 85-88.
- Ebbs, S. D., & Kochian, L. V. (1997). Toxicity of zinc and copper to Brassica species: implications for phytoremediation. *Journal of Environmental Quality*, 26(3): 776-781.
- Embaye, K., Weih, M., Ledin, S., & Christersson, L. (2005). Biomass and nutrient distribution in a highland bamboo forest in southwest Ethiopia: implications for management. *Forest Ecology and Management*, 204(2): 159-169.

- Ernst, W. H. O., Verkleij, J. A. C., & Schat, H. (1992). Metal tolerance in plants. *Acta Botanica Neerlandica*, 41(3): 229-248.
- Farquhar, G. D., Ehleringer, J. R., & Hubick, K. T. (1989). Carbon isotope discrimination and photosynthesis. *Annual review of plant biology*, 40(1): 503-537.
- Gratani, L., Crescente, M. F., Varone, L., Fabrini, G., & Digiulio, E. (2008). Growth pattern and photosynthetic activity of different bamboo species growing in the Botanical Garden of Rome. *Flora-Morphology*, *Distribution, Functional Ecology of Plants*, 203(1): 77-84.
- Gang, A., Vyas, A., & Vyas, H. (2013). Toxic effect of heavy metals on germination and seedling growth of wheat. *Journal of Environmental Research And Development*, 8(2).
- Garbisu, C., Allica, J. H., Barrutia, O., Alkorta, I., & Becerril, J. M. (2002). Phytoremediation: a technology using green plants to remove contaminants from polluted areas. *Reviews on environmental health*, 17(3): 173-188.
- Gardea-Torresdey, J. L., Peralta-Videa, J. R., Montes, M., De la Rosa, G., & Corral-Diaz, B. (2004). Bioaccumulation of cadmium, chromium and copper by Convolvulus arvensis L.: impact on plant growth and uptake of nutritional elements. *Bioresource Technology*, 92(3): 229-235.
- Ghosh, M., & Singh, S. P. (2005). A review on phytoremediation of heavy metals and utilization of it's by products. *Asian Journal Energy Environment*, 6(4): 18.
- Gore, S., Aseltine, R. H. Jr., Colten, M. E. (1992). Social structure, life stress and depressive symptoms in a high school-aged population. *Journal Health Social Behaviour.* 33: 97–113.
- Gratão, P. L., Prasad, M. N. V., Cardoso, P. F., Lea, P. J., & Azevedo, R. A. (2005). Phytoremediation: green technology for the cleanup of toxic metals in the environment. *Brazilian Journal of Plant Physiology*, 17(1): 53-64.

Greipsson, S. (2011). Phytoremediation. Nature Education Knowledge, 3(10):7.

- Guo, D. P., Guo, Y. P., Zhao, J. P., Liu, H., Peng, Y., Wang, Q. M., & Rao, G. Z. (2005). Photosynthetic rate and chlorophyll fluorescence in leaves of stem mustard (Brassica juncea var. tsatsai) after turnip mosaic virus infection. *Plant Science*, 168(1): 57-63.
- Hall, J.L., (2002) Cellular mechanisms for heavy metal detoxification and tolerance. *J Exp Bot* 53: 1–11
- Hegedus, A., Erdei, S. & Horvath, G. (2001). Comparative studies of H₂O₂ detoxifying enzymes in green and greening barley seedings under cadmium stress. *Plant Science*. 160: 1085–1093.
- Hagemeyer, J. (2004). Ecophysiology of plant growth under heavy metal stress. *Heavy metal stress in plants* (pp. 201-222). Springer Berlin Heidelberg.
- Hermle, S., Madeleine S., Günthardt-Goerg, and Rainer S. (2006). Effects of metal-contaminated soil on the performance of young trees growing in model ecosystems under field conditions. *Environmental Pollution* 144(2): 703-714.
- Jadia, C.D., Fulekar, M.H. (2009). Phytoremediation of heavy metals: Recent techniques. *Afr. J. Biotechnol.* 8(6):921-928.
- Jeranyama, P., & DeMoranville, C. J. (2008). Gas exchange and chlorophyll content of cranberry under salt stress. In *IX International Vaccinium Symposium 810* (pp. 753-758).
- John, R., Ahmad, P., Gadgil, K., & Sharma, S. (2009). Heavy metal toxicity: effect on plant growth, biochemical parameters and metal accumulation by Brassica juncea L. *International Journal Plant Production*, 3(3): 65-76.
- Juneau, P., & Popovic, R. (1999). Evidence for the rapid phytotoxicity and environmental stress evaluation using the PAM fluorometric method: importance and future application. *Ecotoxicology*, 8(6): 449-455.

- Kabata-Pendias, A., & Mukherjee, A. B. (2007). Trace elements from soil to human. *Springer Science & Business Media*.
- Keller C. (2005) Efficiency and limitations of phytoextraction by high biomass plants. In *Trace elements in the environment* (pp. 611-630) CRC Press.
- Keller, C., Hammer, D., Kayser, A., Richner, W., Brodbeck, M., & Sennhauser, M. (2003). Root development and heavy metal phytoextraction efficiency: comparison of different plant species in the field. *Plant* and Soil, 249(1): 67-81.
- Khan, S., Ahmad, I., Shah, M. T., Rehman, S., & Khaliq, A. (2009). Use of constructed wetland for the removal of heavy metals from industrial wastewater. Journal of environmental management, 90(11): 3451-3457.
- Kleinhenz, V., & Midmore, D. J. (2001). Aspects of bamboo agronomy. Advances in agronomy, 74: 99-153.
- Krause, G. H., & Weis, E. (1991). Chlorophyll fluorescence and photosynthesis: the basics. *Annual review of plant biology*, 42(1): 313-349.
- Langer, I., Syafruddin, S., Steinkellner, S., Puschenreiter, M., & Wenzel, W. W. (2010). Plant growth and root morphology of Phaseolus vulgaris L. grown in a split-root system is affected by heterogeneity of crude oil pollution and mycorrhizal colonization. *Plant and soil*, 332(1-2): 339-355.
- Lee, C. W., Choi, J. M., & Pak, C. H. (1996). Micronutrient toxicity in seed geranium (Pelargonium× hortorum Bailey). *Journal of the american society for horticultural science*, 121(1): 77-82.
- Leegood R.C., (2002). C4 photosynthesis : Principles of CO2 concentration and prospects for its introduction into C3 plants. *Journal of Experimental Botany*, 53 (369): 581-590.
- Lewis, A. C. (2006). *Assessment and comparison of two phytoremediation systems treating slow-moving groundwater plumes of TCE*, Doctoral dissertation, Ohio University.

- Lewis, S., Donkin, M. E., & Depledge, M. H. (2001). Hsp70 expression in Enteromorpha intestinalis (Chlorophyta) exposed to environmental stressors. *Aquatic Toxicology*, 51(3): 277-291.
- Lelie, D. V. D., Schwitzguébel, J. P., Glass, D. J., Vangronsveld, J., & Baker, A. (2001). Peer reviewed: Assessing phytoremediation's progress in the United States and Europe. *Environmental science and technology*, 35(21): 446-452.
- Lovelock, C. E., & Ball, M. C. (2002). Influence of salinity on photosynthesis of halophytes. In *Salinity: environment-plants-molecules* (pp. 315-339). Springer Netherlands.
- Majid, N.M., Islam, M.M., Veronica, J., Arifin, A., & Parisa, A. (2011). Evaluation of heavy metal uptake and translocation by Acacia mangium as a phytoremediator of copper contaminated soil. *African Journal Biotechnology* 10: 8373-8379.
- Malik, N. J., Chamon, A. S., Mondol, M. N., Elahi, S. F., & Faiz, S. M. A. (2011). Effects of different levels of zinc on growth and yield of red amaranth (Amaranthus sp.) and rice (Oryza sativa), Variety-BR49). Journal of the Bangladesh Association of Young Researchers, 1(1): 79-91.
- Mangkoedihardjo, S. (2008). *Jatropha curcas L*. for phytoremediation of lead and cadmium polluted soil. *World Applied Sciences Journal*, 4(4): 519-522.
- Marschner, H. (1995). Functions of mineral nutrients: macronutrients. Mineral nutrition of higher plants, 2: 379-396.

McCutcheon, S. C., & Schnoor, J. L. (2003). Overview of phytotransformation and control of wastes. Phytoremediation: Transformation and control of contaminants, 358.

Meagher, R. B. (2000). Phytoremediation of toxic elemental and organic pollutants. *Current opinion in plant biology*, 3(2): 153-162.

Migliaccio, K. W., Li, Y. C., Trafford, H., & Evans, E. (2006). A simple lysimeter for soil water sampling in south Florida. *ABE361, Agricultural and* Biological Engineering Department, FL Cooperative Extension Service, IFAS, UF. URL: http://edis. ifas. ufl. edu/AE387.

- Mijovilovich, A., Leitenmaier, B., Meyer-Klaucke, W., Kroneck, P. M., Götz, B., & Küpper, H. (2009). Complexation and toxicity of copper in higher plants. II. Different mechanisms for copper versus cadmium detoxification in the copper-sensitive cadmium/zinc hyperaccumulator Thlaspi caerulescens (Ganges ecotype). *Plant Physiology*, 151(2): 715-731
- Mildvan, A.S. (1970). Metal in enzymes catalysis. In, *Boyer DD (ed) The enzymes*, (pp. 445–536) *London: Academic Press.*
- Moreno-Caselles, J., Moral, R., Pera-Espinosa, A. & Marcia, M.D. (2000). Cadmium accumulation and distribution in cucumber plants. *J. Plant. Nutri*, 23: 243–250.
- Morikawa, H., & Erkin, Ö. C. (2003). Basic processes in phytoremediation and some applications to air pollution control. Chemosphere, 52(9): 1553-1558.
- Morsomme, P., & Boutry, M. (2000). The plant plasma membrane H⁺⁻ATPase: structure, function and regulation. *Biochimica et Biophysica Acta* (*BBA*)-*Biomembranes*, 1465(1): 1-16.
- Nagajyoti, P. C., Lee, K. D., & Sreekanth, T. V. M. (2010). Heavy metals, occurrence and toxicity for plants: a review. *Environmental Chemistry Letters*, 8(3): 199-216.
- Ndimele, P. E. (2010). A review on the phytoremediation of petroleum hydrocarbon. *Pakistan Journal of Biological Sciences*, 13(15): 715.
- Ndzana, J. E., & Otterpohl, R. A Contribution to Flood Management in European Cities through Bamboo Plantations. *VOLUME 3 Bamboo and the Environment*, 10.
- Neelima, P. & Reddy, K.J. (2002). Interaction of copper and cadmium with seedlings growth and biochemical responses in Solanum melongena. *Env. Pollu. Technol.* 1:285–29

- Netto, A. T., Campostrini, E., de Oliveira, J. G., & Bressan-Smith, R. E. (2005). Photosynthetic pigments, nitrogen, chlorophyll a fluorescence and SPAD-502 readings in coffee leaves. *Scientia Horticulturae*, 104(2): 199-209.
- Nilson J., Bamboo Land PTY LTD. N.d. Retrieved 6 March 2015 http://www.bambooland.com.au/information/uses-of-bamboo
- Odjegba, V. J., & Fasidi, I. O. (2004). Accumulation of trace elements by Pistia stratiotes: implications for phytoremediation. Ecotoxicology, 13(7): 637-646
- Ouzounidou, G. (1994). Change in chlorophyll fluorescence as a result of copper treatment: dose response relations in Silene and Thlaspi. *Photosynthetica*, 29:455–462.
- Ow, D. W., & Cunningham, S. D. (1996). Promises and prospects of phytoremediation. *Plant physiology*, 110(3): 715.
- Palmroth, M. R., Pichtel, J., & Puhakka, J. A. (2002). Phytoremediation of subarctic soil contaminated with diesel fuel. *Bioresource Technology*, 84(3): 221-228.
- Panwar, B. S., Ahmed, K. S., & Mittal, S. B. (2002). Phytoremediation of nickelcontaminated soils by Brassica species. *Environment, Development and Sustainability*, 4(1): 1-6.
- Parisa, A., M.N. Azmi, A. Abdu, A.H. Hazandy and K.S. Daljit *et al.*, (2010). Uptake of heavy metals byJatropha curcas L. Planted in soils containing sewage sludge. *American Journal of Applied Sciences*, 7(10): 1291-1299.
- Pitsikki, E., Kairavuo, M., Šeršen, F., Aro, E. M., & Tyystjärvi, E. (2002). Excess copper predisposes photosystem II to photoinhibition in vivo by outcompeting iron and causing decrease in leaf chlorophyll. *Plant Physiology*, 129(3): 1359-1367.

- Pence, N. S., Larsen, P. B., Ebbs, S. D., Letham, D. L., Lasat, M. M., Garvin, D.F., & Kochian, L. V. (2000). The molecular physiology of heavy metal transport in the Zn/Cd hyperaccumulator Thlaspi caerulescens. *Proceedings of the National Academy of Sciences*, 97(9): 4956-4960.
- Perk, M.V.D. (2013). Soil and Water Contamination: From Molecular to Catchment Scale. 2nd Edn., *Taylor and Francis Group, New York, ISBN-10: 0415893437*, 450.
- Pivertz. E., & Bruce. (2001). Phytoremediation of contaminated soil and ground water at hazardous waste sites. *Environmenatal Protection Agency*, 540: 01-500.
- Prasad, M.N.V, Hagmeyer, J. (1999). Heavy metal stress in plants. *Springer, Berlin,* 16–20.
- Prasad, M. N. V., Greger, M., & Landberg, T. (2001). Acacia nilotica L. bark removes toxic elements from solution: corroboration from toxicity bioassay using Salixviminalis L. in hydroponic system. *International Journal of Phytoremediation*, 3(3): 289-300.
- Prasad, M.N.V., & de-Oliveira-Freitas, H. M. (2003). Metal hyperaccumulation in plants: Biodiversity prospecting for phytoremediation technology. *Electronic Journal of Biotechnology*, 6(3): 285-321.

Plant use project bambusa vulgaris (PROTA) assessed on 27 June 2016 http://uses.plantnet-project.org/en/Bambusa_vulgaris_(PROTA)

- Purakayastha, T. J., & Chhonkar, P. K. (2010). Phytoremediation of heavy metal contaminated soils. In *Soil Heavy Metals* (pp. 389-429). Springer Berlin Heidelberg
- Qian, Y., Wen, X.H., & Huang, X. (2007). Development and application of some renovated technologies for municipal wastewater bioremediation in China. *Frontiers Environmental Science & Engineering China* 1: 1 12.
- Ribeiro, R. V., Lyra, G. B., Santiago, A. V., Pereira, A. R., Machado, E. C., & Oliveira, R. F. (2006). Diurnal and seasonal patterns of leaf gas

exchange in bahiagrass (Paspalum notatum Flügge) growing in a subtropical climate. *Grass and Forage Science*, 61(3): 293-303.

- Romero-Puertas, M. C., Rodriguez-Serrano, M., Corpas, F. J., Gomez, M. D., Del Rio, L. A., & Sandalio, L. M. (2004). Cadmium-induced subcellular accumulation of O²⁻⁻ and H₂O₂ in pea leaves. *Plant, Cell and Environment*, 27(9): 1122-1134.
- Rout, G.R. and Das, P. (2003) Effect of metal toxicity on plant growth and metabolism: I. Zinc. *Agronomie*, 23: 3-11.
- Salt, D. E., Blaylock, M., Kumar, N. P., Dushenkov, V., Ensley, B. D., Chet, I., & Raskin, I. (1995). Phytoremediation: a novel strategy for the removal of toxic metals from the environment using plants. *Nature Biotechnology*, 13(5): 468-474.
- Sara, J.,(2013) Bamboo May Be Useful In Treating Gray Water http://www.wateronline.com/doc/bamboo-may-be-useful-in-treatinggray-water-0001 (assessed 28 June 2016)
- Schnoor, J.L. (2002). Phytoremediation of Soil and Groundwater. *Technology Evaluation Report,* GWRTAC Series TE-02-01.
- Schreiber, U., Bilger, W., & Neubauer, C. (1995). Chlorophyll fluorescence as a nonintrusive indicator for rapid assessment of in vivo photosynthesis. In *Ecophysiology of photosynthesis*. Springer Berlin Heidelberg. 49-70.
- Shahid, M., Pourrut, B., Dumat, C., Nadeem, M., Aslam, M. (2014). Heavymetal-induced reactive oxygen species: phytotoxicity and physicochemical changes in plants. *Rev Environ Contam Toxicol*. 232:1-44.
- Singh, P.K. & Tewari, S.K. (2003). Cadmium toxicity induced changes in plant water relations and oxidative metabolism of Brassica juncea L. plants. *J. Env. Biol.* 24:107–117.
- Shibata, S., Ikeda, K., Lulmuanpuia, C., Saito, T., Hasegawa, H., Nishiwaki, A., Makita, A. (2008). Mautam - Melocanna baccifera flowering - Ecological characteristics and influence to the Juhm culture. *International*

Conference on Improvement of Bamboo Productivity and Marketing for Sustainable Livelihood. New Delhi, India, 155-163.

Shkolnik, M.J. & Leningrad. (1974). Microelements in Plant Life. Nauka: Izd.

- Sinha, S., Gupta, M., & Chandra, P. (1997). Oxidative Stress Induced by Iron in Hydrilla verticillata (If) Royle: Response of Antioxidants. *Ecotoxicology and Environmental Safety*, 38(3): 286-291.
- Srivastava, S. (2007). Phytoremediation of heavy metal contaminated soils. *Journal Departmrnt Apply Science Human*, 6: 95-97.
- Tóth, S. Z., Schansker, G., & Strasser, R. J. (2005). In intact leaves, the maximum fluorescence level (F_m) is independent of the redox state of the plastoquinone pool: a DCMU-inhibition study. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, 1708(2): 275-282.
- Truyens, S., Jambon, I., Croes, S., Janssen, J., Weyens, N., Mench, M., & Vangronsveld, J. (2014). The effect of long-term Cd and Ni exposure on seed endophytes of Agrostis capillaris and their potential application in phytoremediation of metal-contaminated soils. *International journal of phytoremediation*, 16(7-8): 643-659.
- TYRER, L. (2006) Plants to the rescue. Mining Weekly.
- Van, A. F., Clijsters, H. (1990). Effects of metals on enzyme activity in plants. *Plant Cell Environ* 13:195–206.
- Verbruggen, N., Hermans, C., Schat, H. (2009). Molecular mechanisms of metal hyperaccumulation in plants. *New Phytol*, 181(4):759–776
- Vernay, P., Gauthier-Moussard, C., & Hitmi, A. (2007). Interaction of bioaccumulation of heavy metal chromium with water relation, mineral nutrition and photosynthesis in developed leaves of Lolium perenne L. *Chemosphere*,68(8): 1563-1575.
- Warne, M. S. J., Heemsbergen, D., Stevens, D., McLaughlin, M., Cozens, G., Whatmuff, M., & Penney, N. (2008). Modeling the toxicity of copper

and zinc salts to wheat in 14 soils. *Environmental Toxicology and Chemistry*, *27*(4): 786-792.

- Watmough, S. A., & Dickinson, N. M. (1995). Dispersal and mobility of heavy metals in relation to tree survival in an aerially contaminated woodland soil.*Environmental pollution*, 90(2): 135-142.
- Whiting, S. N., Leake, J. R., McGrath, S. P., & Baker, A. J. (2000). Positive responses to Zn and Cd by roots of the Zn and Cd hyperaccumulator Thlaspi caerulescens. *New Phytologist*, 145(2): 199-210.
- Xi, X.Y., Liu, M.Y., Huang, Y., (2010). Response of flue-cured tobacco plants to different concentration of lead or cadmium. 2010 4th International Conference on Bioinformatics and Biomedical Engineering (iCBBE): 1– 4.
- Yadav, S. K., Juwarkar, A. A., Kumar, G. P., Thawale, P. R., Singh, S. K., & Chakrabarti, T. (2009). Bioaccumulation and phyto-translocation of arsenic, chromium and zinc by Jatropha curcas L.: impact of dairy sludge and biofertilizer. *Bioresource technology*, 100(20): 4616-4622.
- Yamamoto, F., & Kozlowski, T. T. (1987). Effects of flooding, tilting of stems, and ethrel application on growth, stem anatomy, and ethylene production of Acer platanoides seedlings. *Scandinavian Journal of Forest Research*, 2(1-4): 141-156.
- Yiping, L., Yanxia, L., Buckingham, K., Henley, G., & Guomo, Z. (2010). Bamboo and Climate Change Mitigation: a comparative analysis of carbon sequestration. *International Network Bamboo and Rattan*.

Youssef, T. (2007). Stomatal, biochemical and morphological factors limiting photosynthetic gas exchange in the mangrove associate Hibiscus tiliaceus under saline and arid environment. *Aquatic Botany*, 87(4): 292-298.