

# **UNIVERSITI PUTRA MALAYSIA**

DEVELOPMENT AND VALIDATION OF AMINO ACID ANALYSIS METHODS IN GELATIN AND GELATIN-BASED PRODUCTS USING HIGH-PERFORMANCE LIQUID CHROMATOGRAPHY

**AZILAWATI MOHD ISMAIL** 

**IPPH 2016 9** 



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By

**AZILAWATI MOHD ISMAIL** 

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of Philosophy

November 2016

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## DEDICATIONS

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Doctor of Philosophy

### DEVELOPMENT AND VALIDATION OF AMINO ACID ANALYSIS METHODS IN GELATIN AND GELATIN-BASED PRODUCTS USING HIGH-PERFORMANCE LIQUID CHROMATOGRAPHY

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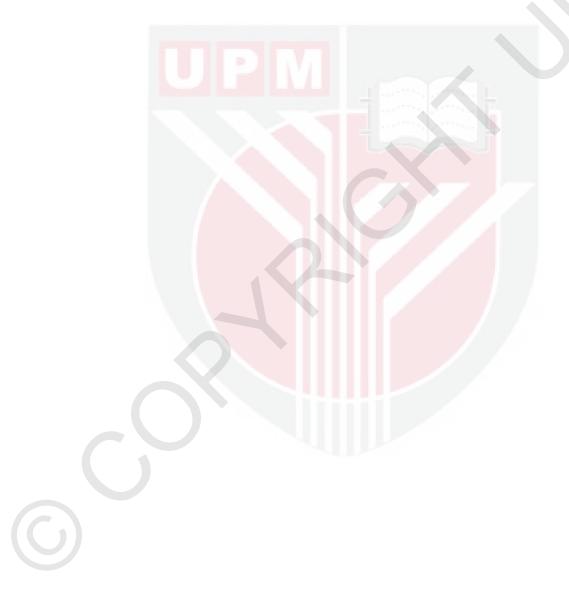
November 2016

### Chairman : Professor Amin B. Ismail, PhD, FNSM Institute : Halal Products Research Institute

The similarity of physicochemical properties among gelatin especially the porcine and bovine gelatin has sparked skepticism among Muslim consumers towards gelatinbased commercial products. This study was aimed to develop and validate a reversephase high performance liquid chromatography (RP-HPLC) method of amino acid analysis in gelatin and differentiate the bovine, porcine and fish gelatin as an ingredient and in gelatin-based commercial products using the principal component analysis (PCA). The analytical method used was amino acid analysis that using 6aminoquinolyl-N-hydroxysuccinimidyl carbamate as derivatization reagent and the chromatographic separation was determined by RP-HPLC coupled with a fluorescence detector. Method development was conducted according to ISO 17025 guidelines. In-house method validation revealed that the method was selectively performing a good chromatographic separation for 18 amino acids; the detection and quantitation limit were ranged from 5.68–12.62 and 36.0–39.0 pmol/µl, respectively; no matrix effect was observed and the linearity range was 37.5-1000 pmol/µl. Method precision revealed by HorRat values was significantly less than 2 and the method recoveries had a range of 80-115%. The uncertainty evaluation was estimated on the basis of the method validation data. The uncertainty of method precision,  $\mu(P)$ , method recovery,  $\mu(R)$  and measurement of standard,  $\mu(Std)$  for overall amino acids are within a concentration range of 0.024 to 0.113 pmol/µl, 0.006 to 0.10 pmol/µl and 0.010 to 0.0297 pmol/µl, respectively. PCA has assisted the process of distinguishing the bovine, porcine and fish gelatin. Data pre-treatments such as centering and area normalization were performed to reduce the variances of variables in dataset. Database for three gelatins were established through this work and were verified by samples from gelatin-based commercial products. Data analysis demonstrated that the fish gelatin was correlated to threonine, serine and methionine on the positive side of principal component (PC) 1; bovine gelatin was correlated to the non-polar side chains amino acids that were proline, hydroxyproline, leucine, isoleucine and valine on the negative side of PC1 and porcine gelatin was correlated to the polar side chains amino acids that were aspartate, glutamic acid, lysine and tyrosine on the negative side of



PC2. The lowest detection value for adulteration of porcine gelatin in bovine gelatin was determined at 0.05% (w/w) of porcine gelatin. The extraction of gelatin from gelatin-based commercial products was successful by samples clean-up using acetone solvent and modification on several parameters of amino acid analysis method specifically the sample digestion process. Gelatin used in the commercial products was verified using gelatin database and the results revealed that products made of porcine gelatin can be differentiated from the products that were made of bovine gelatin. This quantitative method was very useful as an alternative method for halal products authentication via laboratory testing.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

## PEMBANGUNAN DAN VALIDASI KAEDAH ANALISIS ASID AMINO DALAM GELATIN DAN PRODUK BERASASKAN GELATIN MENGGUNAKAN KROMATOGRAFI CECAIR PRESTASI TINGGI

Oleh

#### AZILAWATI MOHD ISMAIL

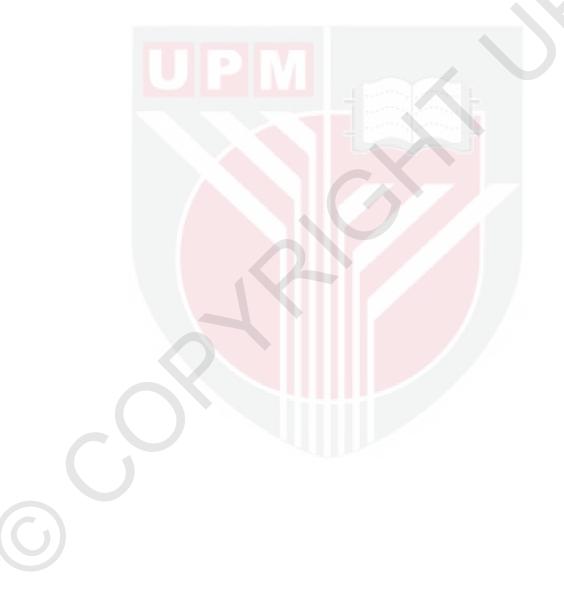
November 2016

### Pengerusi : Profesor Amin B. Ismail, PhD, FNSM Institut : Penyelidikan Produk Halal

Persamaan sifat fizikokimia di antara gelatin terutamanya gelatin porsin dan gelatin bovin telah mencetuskan keraguan di kalangan pengguna Muslim terhadap produk komersil berasaskan gelatin. Kajian ini bertujuan untuk membangunkan dan mengesahsahihan kaedah fasa-terbalik kromatografi cecair berprestasi tinggi (RP-HPLC) analisis asid amino dalam gelatin dan membezakan gelatin bovin, porsin dan ikan sebagai bahan ramuan dan di dalam produk komersil berasaskan gelatin menggunakan analisis komponen utama (PCA). Kaedah analitikal yang digunakan adalah analisis asid amino menggunakan 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate sebagai reagen derivatization dan pemisahan kromatografi ditentukan oleh RP-HPLC dengan pengesan berpendarfluor. Pembangunan kaedah dijalankan mengikut garispanduan ISO 17025. Pengesahsahihan kaedah secara dalaman menunjukkan bahawa kaedah ini adalah selektif dengan pemisahan kromatografi yang baik bagi 18 asid amino; had pengesanan dan had kuantitatif adalah dalam julat 5.68 -12.62 dan 36.0 - 39.0 pmol/µl, masing-masing; tiada pemerhatian berkaitan kesan matriks dan julat kelinearan adalah 37.5 - 1000 pmol/µl. Ketepatan kaedah didedahkan oleh nilai HorRat yang nyata sekali kurang daripada 2 dan peratusan perolehan semula bagi kaedah adalah di antara 80-115%. Penilaian ketidakpastian telah dianggarkan berdasarkan data kaedah validasi. Ketidakpastian bagi ketepatan kaedah,  $\mu(P)$ , perolehan semula kaedah,  $\mu(R)$  dan pengukuran standard,  $\mu(Std)$  untuk keseluruhan asid amino adalah dalam julat kepekatan 0.024 - 0.113 pmol/µl, 0.006 - 0.10 pmol/µl dan 0.010 - 0.0297 pmol/µl, masing-masing. PCA telah membantu proses membezakan di antara gelatin bovin, porsin dan ikan. Pra-perlakuan (treatment) data seperti pemusatan dan normalisasi kawasan puncak dilakukan untuk mengurangkan varians di antara pembolehubah dalam set data. Pangkalan data untuk tiga gelatin telah dibangunkan dan ditentusahkan oleh sampel-sampel dari produk komersial yang berasaskan gelatin. Analisis data menunjukkan bahawa gelatin ikan berkorelasi dengan treonina, serina dan metionina pada posisi positif komponen utama (PC) 1; gelatin bovin berkorelasi kepada asid amino dengan rantai sisi tidak polar iaitu prolina, hidroksiprolina, leusina, isoleusina dan valina di posisi negatif PC1 dan gelatin porsin



berkorelasi kepada asid amino dengan rantai sisi polar iaitu aspartik, asid glutamik, lisin dan tirosina di posisi negatif PC2. Nilai pengesanan terendah untuk pencemaran gelatin porsin dalam gelatin bovin ditentukan pada 0.05% (w/w) gelatin porsin. Pengekstrakan gelatin daripada produk komersial berasaskan gelatin berjaya dilakukan melalui pembersihan sampel menggunakan larutan aseton dan pengubahsuaian beberapa parameter dalam kaedah analisis asid amino khususnya bagi proses pencernaan sampel. Gelatin daripada produk komersial telah ditentusahkan menggunakan pangkalan data gelatin dan keputusan menunjukkan bahawa produk yang diperbuat daripada gelatin porsin boleh dibezakan daripada produk yang diperbuat daripada gelatin bovin. Kaedah kuantitatif ini adalah sangat berguna sebagai satu kaedah alternatif bagi pengesahan produk halal melalui ujian makmal.



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I certify that a Thesis Examination Committee has met on 7 November 2016 to conduct the final examination of Azilawati Mohd Ismail on her thesis entitled "Development and Validation of Amino Acid Analysis Methods in Gelatin and Gelatin-Based Products using High-Performance Liquid Chromatography" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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# LIST OF ABBREVIATIONS

AABA	L-Aminobutyric acid
AOAC	Association of Analytical Communities
AMQ	6-aminoquinoline
AQC	6-aminoquinolyl-N-hydroxysuccinimidyl carbamate
BSE	Bovine spongiform encephalopathy
CV	Coefficient of variation
CRM	Certified reference material
DA	Discriminant analysis
DCSI-MSMS	Doubly charged selected ion coupled with MS/MS fragments monitoring
DNA	Deoxyribonucleic acid
DSC	Disuccinimidyl carbonate
EDA	Exploratory data analysis
EMA	Economically motivated adulteration
FA	Factor analysis
FDA	Food and Drug Administration
FMOC-CL	Dansyl chloride and fluorenylmethyl chloroformate
FTIR	Fourier transform infrared spectroscopy
GC	Gas chromatography
GMIA	Gelatin Manufacturers Institute of America
GUM	Guide to the expression of uncertainty in measurement
HCL	Hydrochloric acid
HPLC-MS/MS	High-performance liquid chromatography/tandem mass spectrometry

	ICH	The International Council for Harmonisation of Technical Requirements for Pharmaceuticals for Human Use
	IR	Infrared spectroscopy
	ISO	International Organization for Standardization
	ISO/IEC 17025	General requirements for the competence of testing and calibration laboratories
	IUPAC	International Union of Pure and Applied Chemistry
	IRMS	Isotope ratio with mass spectrometry
	JAIN	State Islamic Religious Department
	JAKIM	Department of Islamic Development Malaysia
	JCGM	Joint Committee for Guides in Metrology
	K-NN	K-nearest neighbor classification
	LOD	Limit of detection
	LOQ	Limit of quantitation
	MAIN	State Islamic Religious Council
	MS	Mass spectrometry
	MALDI-TOF-MS	Matrix-assisted laser desorption/ionization time-of flight mass spectrometry
	MCE	2-mercaptoethanol
	MPA	3-mercaptopropionic acid
	NBD-Cl	4-chloro-7-nitrobenzofurazane
	NHS	N-hydroxysuccinimide
	NATA	National Association of Testing Authorities
	NaOH	Sodium hydroxide
	nanoUPLC-ESI-Q- TOF-MS	Ultra-performance liquid chromatography and electrospray ionization quadrupole time-of-flight mass spectrometry
	OPA	Orthophthalaldehyde

PC	Principal component
PCR	Polymerase chain reaction
PCA	Principal component analysis
PEG	Polyethylene glycol
PLS-DA	Partial least squares-discriminant analysis
PITC	Phenylisothiocyanate
RP-HPLC	Reversed-phase high performance liquid chromatography
RSD	Relative standard deviation
SDS-PAGE	Sodium dodecyl sulphate-polyacrylamide gel electrophoresis
SEC	Size exclusion chromatography
SIMCA	Soft independent modeling class analogy
SLV	Single laboratory validation
VAM	Valid Analytical Measurement

C

#### **CHAPTER 1**

### **INTRODUCTION**

Halal is an Arabic word which means permissible or lawful. The underlying principle for a Muslim's diet is that food intake not only has to be halal (permissible – Syariah compliant), but also toyyiban which means wholesomeness (healthy, safe, nutritious, quality). Allah has commanded in the Holy Quran for Muslims and all of mankind to eat only the halal things - "O mankind! Eat of that which is lawful and wholesome, and follow not the footsteps of the devil. Lo! he is an avowed enemy of you" (2:168). Halal food and drinks can be described as anything that man can eat or drink and there is no legal evidence prohibiting it and its constituents are free from any unlawful or impure elements. It shall be good and pure and its consumption brings no harm (Mahiah et al., 2014).

Some Islamic countries have established strict regulations for producers and importers to stamp their products with a halal certificate to distinguish them from non-halal products. Malaysia is one of the countries who implemented a halal certification to certify the products or services as pronounced by the shariah law and complying to the Halal Standard MS 1500:2009 (Halal Food - Production, Preparation, Handling and Storage - General Guidelines). The Malaysian halal certification and logo are issued at the federal level by Department of Islamic Development Malaysia (JAKIM) to certify the products for the domestic and international market and at the state level by the State Islamic Religious Department (Jabatan Agama Islam Negeri - JAIN) and State Islamic Religious Council (Majlis Agama Islam Negeri - MAIN) to issue the halal certificate for domestic market. Currently, the Halal certification is executed through the auditing of documents and at the field with regular monitoring to the manufacturers of certified halal products. However, the halal authentication based on halal laboratory analysis is not fully implemented due to lack of capabilities in terms of methods and facilities. At present, the Department of Chemistry Malaysia is appointed as the authority for performing the halal analysis specifically related to the enforcement tasks. In most cases, the halal authentication by laboratory analysis is confirmed by the DNA analysis. In this regard, development of reliable and sensitive analytical methods for evaluating halal authenticity is critically in demand.

The issue of gelatin is alarming and sometimes controversial due to commercial gelatins were offered from various sources in the global market including the porcine gelatin. The situation has prompted consumers to be more cautious, especially when it comes to religious restrictions, health hazards and safety in the intake. In the mid 1980s, the world was shaken by the emergence of bovine spongiform encephalopathy ("mad cow disease") epidemic that swept the European countries. Since then, there has been much concern about using the gelatin from the infected animals (Karim & Bhat, 2008). Besides that, almost all Hindus abstain from cow in their dietary intake. They do not worship the cow but are deeply respecting the cow that became essential to their spiritual life (Grabenstein, 2013). In addition, Islam and Judaism have forbidden the consumption of porcine derived gelatin in food materials. It was stated

in the Holy Quran (Surat Al-'An`am, 6:145) that Muslims are prohibited from taking the flesh of swine and its derivatives as indeed it is impure. For Judaism, the Torah (Lev. 11:3; Deut. 14:6) has specifies that the pig is not kosher because it does not has cloven hooves and not chews the cud. Therefore, the religious and socio-cultural factors have influenced the need for other alternative sources such as fish and poultry to fulfill halal and kosher markets.

The similarity of the physicochemical properties of different sources gelatin made them difficult to be differentiated. Several methods have been reported based on different approaches including chemisorptions, chromatographic, immunochemical, mass spectrometric, spectroscopic and molecular techniques (Yilmaz et al., 2013). A distinct identification was offered by each approach to distinguish gelatin from different sources. For example, the formation of calcium phosphate precipitation using a pH drop method (Hidaka & Liu, 2003), a deformation of N-H bonds within the range 3290-3280 and 1660-1200 cm of infrared spectra (Hashim et al., 2010), identification of marker peptides in digested gelatin using high performance liquid chromatography/tandem mass spectrometry (HPLC-MS/MS) (Zhang et al., 2009) and ultra-performance liquid chromatography and electrospray ionization quadrupole time-of-flight mass spectrometry (nanoUPLC-ESI-q-TOF-MS) (Yilmaz et al., 2013) had scientifically showed the ability of analytical methods to distinguish gelatin from various sources. However, these methods have to be confirmed and repeatability under laboratory collaborative study is recommended to assure the accuracy and quality of the reproducible results. Besides that, certain applications such using mass spectrometric is very costly, require highly skilled personnel to run the analysis and took a longer time for the profiling of peptides through chromatographic separation.

Apart from using direct methods to distinguish the gelatin, an additional of descriptive and predictive method through chemometric techniques (multivariate data analysis) was applied to facilitate discrimination among gelatins. Principal component analysis (PCA) has been the most commonly used as it is simple and revealing the internal structures of the data in a way that best explains the variance in the data. The technique has been used with analysis involving the fourier transform infrared (FTIR) spectroscopy (Hashim et al., 2010), a reversed-phase (RP) - high performance liquid chromatography (HPLC) (Nemati et al., 2004) and ultra-performance liquid chromatography-quadrupole-time of flight mass spectrometry (UPLC/Q-TOF-MS) (Cheng et al., 2012) and sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS)-PAGE (Nur Azira et al., 2014) in which differentiation among gelatins were performed. The researches indicated that bovine and porcine gelatins have large similarities in chemical/electrophoretic profiles, spectrum chromatography structure and having a very high homology between the collagen sequences at higher concentrations that make the ability to differentiate them by visual observation is impossible.

## 1.1 Objectives

The general objective of this study was to develop an RP-HPLC method of amino acid analysis that incorporated with PCA to differentiate the bovine, porcine and fish gelatins as an ingredient and in the gelatin-based commercial products.

The study encompassed of several important topics that related to a method development and uncertainty estimation for a quantitative analysis, a development of gelatin database, identification of adulteration between bovine and porcine gelatins and modification on method of amino acid analysis in order to improves the extraction of gelatin from gelatin-based commercial products. Therefore, the specific objectives of this study were:

- To perform a method validation for determination of amino acid compositions in gelatin by application of 6-aminoquinolyl-*N*-hydroxysuccinimidyl carbamate (AQC) as derivatization reagent using an RP- HPLC and estimating the uncertainty of measurement using Valid Analytical Measurement (VAM) Project 3.2.1 protocol that based on information from the validation data.
- 2) To determine the amino acid compositions in bovine, porcine and fish gelatins using amino acid analysis and differentiate the gelatins using a chemometric method specifically the principal component analysis (PCA) technique.
- 3) To determine the minimum detection value for adulteration of porcine gelatin in the bovine gelatin.
- 4) To develop a method that can enhance the gelatin extraction from gelatinbased commercial products through modification of several parameters in amino acid analysis method.

#### REFERENCES

- Adams, T.M. (2002). Guide for estimation of measurement uncertainty in testing. G104-A2LA. http://www.a2la.org/guidance/est\_mu\_testing.pdf. Retrieved 06 March 2013.
- Analytical Methods Committee. (1995). Uncertainty of measurement: Implications of its use in analytical science. Analyst 120: 2303-2308.
- AOAC Official Methods of Analysis. Guidelines for Collaborative Study Procedures to Validate Characteristics of A Method of Analysis. Interlaboratory Collaborative study, appendix D. AOAC INTERNATIONAL. Gaithersburg, MD. (2002).
- AOAC Official Methods of Analysis. Guidelines for Single Laboratory Validation of Chemical Methods for Dietary Supplements and Botanicals. AOAC INTERNATIONAL. Gaithersburg, MD. (2002).
- AOAC Official Methods of Analysis. Guidelines for Dietary Supplements and Botanicals, appendix K. AOAC INTERNATIONAL, Gaithersburg, MD. (2012).
- Agnieszka, S., Lionel, B., Lutgarde, M.C.B. and Sybren, S.W. (2012). NMR and pattern recognition methods in metabolomics: From data acquisition to biomarker discovery. Analytica Chimica Acta 750: 82-97.
- Avena-Bustillos, R.J., Olsen, C.W., Chiou, B., Yee, E., Bechtel, P.J. and McHugh, T.H. (2006). Water vapor permeability of mammalian and fish gelatin films. Journal of Food Science 71: E202–E207.
- Banerjee, S. and Bhattacharya, S. (2012). Food gels: Gelling process and new applications. Critical Reviews in Food Science and Nutrition 52(4): 334-346.
- Barbul, A. (2008). Proline precursors to sustain mammalian collagen synthesis. Journal of Nutrition 138(10): 2021S-2024S.
- Belitz, H.D., Grosch, W. and Schieberle, P. (2009). Amino acids, peptides, proteins. In Food Chemistry, pp. 8-90. Berlin Heidelberg: Springer-Verlag.
- Bartolomeo, M. P. and Maisano, F. (2006). Validation of a reversed-phase HPLC method for quantitative amino acid analysis. Journal of Biomolecular Techniques 17: 131–137.
- Barwick, V.J and Ellison, S.L.R. (2000). VAM Project 3.2.1. Part (d) : Protocol for uncertainty evaluation from validation data. In Development and harmonisation of measurement uncertainty principles. Teddington: LGC/VAM/1998/088.

- Barwick, V. J. (2012). The 'top down' approach to uncertainty estimation. In Evaluating measurement uncertainty in clinical chemistry. UK: LGC/R/2010/17.
- Bhatt, B. and Agrawal, S.S. (2007). Capsules. Pharmaceutical Technology, pp. 1-26. New Delhi, India.
- Blackburn, S. (1978). Sample preparation and hydrolytic methods. In Amino Acid Determination: Methods and Techniques, ed. S. Blackburn, 2nd Ed, pp.7-34. New York: Marcel Dekker.
- Brereton, R.G. (2003). Pattern Recognition. In Chemometrics: Data analysis for the laboratory and chemical plant, ed. R.G. Brereton, pp. 183-255. Chichester, UK: John Wiley & Sons, Ltd.
- Brereton, R.G. (2007). Applied Chemometrics for Scientists. Chichester, UK: John Wiley & Sons Ltd.
- Brian, R. (2007). Paintball. Chemistry hits its mark. ChemMatter. http://chemistry.org/education/chemmatters.html. Retrieved 23 May 2014.
- Bro, R. and Smilde, A.K. (2003). Centering and scaling in component analysis. Journal of Chemometrics 17: 16-33.
- Bro, R. and Smilde, A.K. (2014). Principal component analysis. Analytical Methods 6: 2812–2831.
- Bryant, C. M. and McClements, D.J. (2000). Influence of sucrose on NaCl-induced gelation of heat denaturated whey protein solutions. Food Research International 33: 649-653.
- Burey, P., Bhandari, B.R., Rutgers, R.P.G., Halley, P.J. and Torley, P.J. (2009). Confectionery gels: A review on formulation, rheological and structural aspects. International Journal of Food Properties 12(1): 176-210.
- CAMO Software AS. (2006). The Unscrambler Methods. The Unscrambler user manual. http://www.camo.com. Retrieved 01 January 2013.
- CAMO Process AS. (2006). The Unscrambler Tutorials. http://www.camo.com. Retrieved 01 January 2013.
- Cheng, X.L., Lin, R.C., Wei, F., Xiao, X.Y., Zhao, Y.Y. and Shi, Y. (2012). Identification of five gelatins by ultra performance liquid chromatography/time-of-flight mass spectrometry (UPLC/Q-TOF-MS) using principal component analysis. Journal of Pharmaceutical and Biomedical Analysis 62: 191–195.

- Cheng, X.L., Wei, F., Chen, J., Li, M., Zhang, L., Zhao, Y., Xiao, X., Ma, S. and Lin, R. (2014). Using the doubly charged selected ion coupled with MS/MS fragments monitoring (DCSI-MS/MS) mode for the identification of gelatin species. Journal of Analytical Methods in Chemistry 764397: 1-7.
- Choi, Y. H., Lim, S. T. and Byoungseung, Y. (2004). Measurement of dynamic rheology during ageing of gelatin-sugar composites. International Journal of Food Science and Technology 39: 935-945.
- Cohen, S.A. and Strydom, D.J. (1988). Review. Amino acid analysis utilizing phenylisothiocyanate derivatives. Analytical Biochemistry 174: 1-16.
- Cohen, S.A. and Michaud, D.P. (1993). Synthesis of a fluorescent derivatizing reagent, 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate and its application for the analysis of hydrolysate amino acids via high performance liquid chromatography. Analytical Biochemistry 211: 279–287.
- Cohen, S.A. (2005). Quantitation of amino acids as 6-aminoquinolyl-Nhydroxysuccinimidyl carbamate derivatives. In Quantitation of amino acids and amines by chromatography. Methods and protocols, ed. I. Molnar-Perl, pp. 242–267. Netherlands: Elsevier.
- Coppola, M., Djabourov, M. and Ferrand, M. (2012). Unified phase diagram of gelatin films plasticized by hydrogen bonded liquids. Polymer 53: 1483-1493.
- Cuesta, S. F., Lewi, P.I. and Massart, D.I. (1994). Effect of different preprocessing methods for principal component analysis applied to the composition of mixtures: detection of impurities in HPLC-DAD. Chemometrics and Intelligent Laboratory Systems 25: 157-177.
- Digenis, G. A., Gold, T. B. and Shah, V. P. (1994). Cross-linking of gelatin capsules and its relevance to their in vitro-in vivo performance. Journal of Pharmaceutical Sciences 83: 915-921.
- Duconseille, A., Astruc, T., Quitana, N., Meersman, F. and Sante-Lhoutellier, V. (2015). Gelatin structure and composition linked to hard capsule dissolution: A review. Food Hydrocolloids 43: 360-376.
- Djagny, K.B., Wang, Z. and Xu, S. (2010). Gelatin: A valuable protein for food and pharmaceutical industries: Review. Critical Reviews in Food Science and Nutrition 41(6): 481-492.
- Eastoe, J.E. (1955). The amino acid composition of mammalian collagen and gelatin. Biochemical Journal. 61(4): 589-600.
- Elharfaoui, N., Djabourov, M. and Babel, W. (2007). Molecular weight influence on gelatin gels: Structure, enthalpy and rheology. Macromol. Symp. 256: 149-157.

- Ellison, S. L. and Barwick, V. J. (1998). Using Validation data for ISO measurement uncertainty estimation. Part 1. Principles of an approach using cause and effect analysis. Analyst 123: 1387-1392.
- Ellison, S.L.R. and Williams, A. (2012). Quantifying Uncertainty In Analytical Measurement. In EURACHEM/CITAC Guide CG 4., 3rd ed. UK: Laboratory of Government Chemist. http://www.eurachem.org. Retrieved 20 February 2013.
- Eriksson, L., Johansson, E., Kettaneh-Wold, N., Trygg, J., Wikstrom, C. and Wold, S. (2006). Multi- and Megavariate Data Analysis. Part 1. Basic Principles and Applications. 2nd ed. Sweden: Umetrics AB.
- Esbensen, K.H, Guyot, D., Westad, F. and Houmoller, L.P. (2002). Multivariate Data Analysis-In Practice. In An Introduction to Multivariate Data Analysis and Experimental Design, ed. K.H. Esbensen, 5th ed., pp. 75-104. Norway: CAMO.
- Eurachem Working Group. The Fitness for Purpose of Analytical Methods. A Laboratory Guide to Method Validation and Related Topics. In Eurachem Guide, 2nd ed. Teddington, UK. http://www.eurachem.org. Retrieved 02 February 2012.
- Eurolab.Measurement uncertainty in testing. A short introduction on how to characterise accuracy and reliability of results including a list of useful references. Technical Report No. 1/2002. (2002).
- Everstine, K., Spink, J. and Kennedy, S. (2013). Economically motivated adulteration (EMA) of food: Common characteristics of EMA incidents. Journal of Food Protection 76(4): 723-735.
- Fennema, O.R. (1985). Amino acids, peptide and proteins. In Food Chemistry, 2nd ed., pp. 255-380. New York: Marcel Dekker, Inc.
- Food and Agriculture Organization of the United Nations. Validation of Analytical Methods for Food Control. A report of a Joint FAO/IAEA expert consultation. FAO Food and Nutrition paper: Vienna, Austria. (1997).
- Food and drug administration. Guidance for Industry. Bioanalytical Method Validation. FDA:Rockville, MD. (2001). http://www.fda.gov/cder/guidance/index.htm. Retrieved 10 May 2013.
- Food and drug administration foods program science and research steering committee. Guidelines for the Validation of Chemical Methods for the FDA Foods Program. FDA: US. (2012).
- Fountoulakis, M. and Lahm, H.W. (1998). Journal of Chromatography A 826: 109-134.

- Gelatin market by raw material (pig skin, bovine hides), by application (food & beverage, neutraceuticals, pharmaceuticals) expected to reach USD 3.18 billion by 2020. (2014). http://www.grandviewresearch.com/industry-analysis/gelatin-market-analysis. Retrieved 10 April 2014.
- Gekko, K., Li., X. and Makino, S. (1992). Effects of polyols and sugars on the sol-gel transition of gelatin. Bioscience, Biotechnology and Biochemistry 56: 1280-1284.
- Gelatin Manufacture Institute of America (GMIA). (2012). Gelatin handbook. http: //www.gelatin-gmia.com/images/GMIA\_Gelatin\_Manual. Retrieved 24 July 2013.
- George, N., Jose, J. and Zynudheen, A. A. (2014). A comparative study on the physical, chemical and functional properties of carp skin and mammalian gelatins. J Food Sci Technol 51(9): 2085-2091.
- Gorres, K.L. and Raines, R.T. (2010). Prolyl 4-hydroxylase. Critical Reviews in Biochemistry and Molecular Biology 45(2): 106-124.
- Gomez-Guillen, M.C., Gimenez, B., Lopez-Caballero, M.E. and Montero, M.P. (2011). Functional and bioactive properties of collagen and gelatin from alternative sources: A review. Food Hydrocolloids 25: 1813–1827.
- Gonzalez, A.G., Herrador, M.A. and Asuero, A.G. (2005). Practical digest for evaluating the uncertainty of analytical assays from validation data according to the LGC/VAM protocol. Talanta 65:1022-1030.
- Grabenstein, J. D. (2013). What the world's religions teach, applied to vaccines and immune globulins. Vaccine 31: 2011–2023.
- Grundy, H. H., Reece, P., Buckley, M., Solazzo, C.M., Dowle, A.A., Ashford, D., Charlton, A.J., Wadsley, M.K and Collins, M. J. (2016). A mass spectrometry method for the determination of the species of origin of gelatin in foods and pharmaceutical products. Food Chemistry 190: 276-284.
- Hashim, D.M., Che Man, Y.B., Norakasha, R., Shuhaimi, M. and Salmah, Y. (2010). Potential uses of fourier transform infrared spectroscopy for differentiation of bovine and porcine gelatines. Food Chemistry 118: 856–860.
- Haug, I.J. and Draget, K.I. (2011). Gelatin. In Handbook of food proteins, ed. G.O. Phillips, and P.A. Williams, pp. 92-115. Norway: Woodhead Publishing Limited.
- Hessam, S., Mehrangiz, M., Seyed, M. M., Ehsan, A. D., Tara, S., Mahdi, K., Maryam, R., Hossein, R. and Mahmoud, A. (2015). Halal authenticity of gelatin using species-specific PCR. Food Chemistry 184: 203-206.

- Hibbert, D.B. (2007). Systematic errors in analytical measurement results. Journal of Chromatography A 1158: 25-32.
- Hidaka, S. and Liu, S.Y. (2003). Effects of gelatins on calcium phosphate precipitation: A possible application for distinguishing bovine bone gelatin from porcine skin gelatin. Journal of Food Composition and Analysis 16: 477– 483.
- Huber, L. (2007). Validation and Qualification in Analytical Laboratories. New York, USA: Informa Healthcare.
- Huber, L. (2010). Validation of analytical methods. Germany: Agilent Technologies.
- Huda, H.N. and Adzitey, F. (2012). Fish bone and scale as a potential source of halal gelatin. Journal of Fisheries and Aquatic Sciences 6(4): 379-389.
- Hund, E., Massart, D.L and Smeyers-Verbeke, J. (2000). Inter-laboratory studies in analytical chemistry. Analytica Chimica Acta 423: 145-165.
- Horwitz, W. and Albert, R. (1985). Performance of methods of analysis used for regulatory purposes. Analytical Chemistry 68: 191-198.
- Ikhwan, I. Campuran lembu dan porcine dalam Imodium capsule Ubat gelatin babi di tarik balik. In Kosmo (http://www.kosmo.com.my). Retrieved 15 December 2015.
- International Conference on Harmonisation of Technical Requirements for Registration of Pharmaceuticals for Human Use. Guidance for Industry. Q2B Validation of Analytical Procedures: Methodology; ICH: Rockville, MD. (1996).
- International Organization for Standardization. Guide for the Expression of Uncertainty in Measurements (GUM); ISO: Geneva. (1995).
- Irwandi, J., Faridayanti, S., Mohamed, E.S.M., Hamzah, M.S. and Torla, H.H. (2009). Extraction and characterization of gelatin from different marine fish species in Malaysia. International Food Research Journal 16: 381-389.
- ISO/TS International Standard 21748. Guidance for the Use of Repeatability, Reproducibility and Trueness Estimates in Measurement Uncertainty Estimation; ISO: Geneva. (2004).
- Jeffrey, R. Analytical detection limit guidance and laboratory guide for determining method detection limits. Wisconsin Department of Natural Resources, April1996, http://www.dnr.state.wi.us. Retrieved 8 April 2012.
- Jia, Z., Xiaoshuan, Z., Lorena, D. and Cristea, V. (2011). Review of the current application of fingerprinting allowing detection of food adulteration and fraud in China. Food Control 22: 1126-1135.

- Joint committee for guides in metrology. International Vocabulary of Metrology-Basic and General Concepts and Associated Terms (VIM); JCGM 200: 2008, International Organization for Standardization: Geneva.
- Jolliffe, I. T. (1986). Principal component analysis. New York: Springer-Verlag Inc.
- Julicher, B., Gowik, P. and Uhlig, S. (1999). A top-down in-house validation based approach for the investigation of the measurement uncertainty using fractional factorial experiments. The Analyst 124: 537-545.
- Juan, M.B., Luis, C.R., Cristina, R.S and Paulina, A. (2012). Combining chromatography and chemometrics for the characterization and authentication of fats and oils from triacylglycerol compositional data. A review. Analytica Chimica Acta 724: 1-11.
- Kamal, M. and Karoui, R. (2015). Analytical methods coupled with chemometric tools for determining the authenticity and detecting the adulteration of dairy products: A review. Trends in Food Science & Technology 46: 27-48.
- Karim, A.A. and Bhat, R. (2008). Gelatin alternatives for the food industry: Recent developments, challenges and prospects. Trends in Food Science & Technology 19: 644–656.
- Karim, A.A. and Bhat, R. (2009). Fish gelatin: Properties, challenges, and prospects as an alternative to mammalian gelatins. Food Hydrocolloids 23: 563–576.
- Kittiphattanabawon, P., Benjakul, S., and Bisessanguan, W. (2005) Characterisation of acid-soluble collagen from skin and bone of bigeye snapper (Priacanthus tayenus). Food Chemistry 89: 363-372.
- Kozlov, P.V. and Burdygina, G.I. (1983). The structure and properties of solid gelatin and the principles of their modification. Polymer 24: 651-666.
- Kramer, R. (1998). Centering and scaling. In Chemometric techniques for quantitative analysis, pp. 179-180. US: Library of congress.
- Krane, S.M. (2008). The importance of proline residues in the structure, stability and susceptibility to proteolytic degradation of collagens. Amino acids 35: 703-710.
- Kurt, V. and Peter, F. (2009). Multivariate data. In Introduction to multivariate statistical analysis in chemometrics, chapter 2. US: CRC Press, Taylor & Francis.
- Leuenberger, B.H. (1991). Investigation of viscosity and gelation properties of different mammalian and fish gelatins. Food Hydrocolloids 5(4): 353-361.

- Lilli, S., Eva-Ingrid, R., Agness, K., Ivari, K. and Ivo, L. (2006). Uncertainty sources in UV-Vis spectrophotometric measurement. Accreditation and Quality Assurance 11: 246-255.
- Liu, H.J.J. (1994). Determination of amino acids by pre-column derivatization with 6aminoquinolyl-N-hydroxysuccinimidyl carbamate and high-performance with ultraviolet detection. Journal of Chromatography A 670: 59-66.
- Lourdes, B., Amparo, A. and Rosaura, F. (2006). Application of the 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate (AQC) reagent to the RP-HPLC determination of amino acids in infant foods. Journal of Chromatograhy B 831:176-183.
- Lucia, M. and Radu, M. (2008). Mining in chemometrics. Review. Analytica Chimica Acta 612:1-18.
- Mahiah, S., Faridah, H., Rosidah, M. and Rahana, N.A. (2014). Assessing consumers' perception, knowledge and religiosity on Malaysia Halal food products. Procedia-Social and Behavioral Sciences 130: 120-128.
- McNaught, A. D. and Wilkinson, A. (1997). IUPAC. Compendium of Chemical Terminology. Gold Book. 2nd ed. Oxford: Blackwell Scientific Publications. http://www.iupac.org/goldbook. Retrieved 15 Jun 2014.
- Marcus, A.L.T., Alessandro, M.S., Fabiane, L.F. and Fellipe, R.L. (2014). Measurement uncertainty in pharmaceutical analysis and its application. Journal of Pharmaceutical Analysis 4 (1): 1-5.
- Mark, H. (2003). Application of an improved procedure for testing the linearity of analytical methods to pharmaceutical analysis. Journal of Pharmaceutical and Biomedical Analysis 33: 7-20.
- Masuda, A. and Dohmae, N. (2011). Amino acid analysis of sub-picomolar amounts of proteins by pre-column fluorescence derivatization with 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate. Bioscience Trends 5(6): 231–238.
- Massart, D. L., Vandeginste, B. G. M., Buydens, L. M. C., De Jong, S., Lewi, P. J. and Smeyers, V. J. (1997). Handbook of Chemometrics and Qualimetrics Part A, Amsterdam: Elsevier.
- Matthias, O. (2007). What is chemometrics. In Chemometrics, pg. 1-11. KGaA, Weinheim: Wiley-VCH Verlag GmbH & Co.
- Meyer, V.R. (2007). Review. Measurement uncertainty. Journal of Chromatoraphy A 1158: 15 24.
- Mita, A. R., Kuwat, T., Triwahyudi and Rohman, A. (2014). Differentiation of bovine and porcine gelatins in soft candy based on amino acid profiles and chemometrics. J. Food Pharm. Sci. 2: 1-6.

- Moore, J. C., Spink, J. and Lipp, M. (2012). Development and application of a database of food ingredient fraud and economically motivated adulteration from 1980 to 2010. Journal of Food Science 77(4): R118- R126.
- Murugaiah, C., Mustakim, M., Mohd Noor, Z., & Radu, S. (2010). Identification of the species origin of commercially available processed food products by mitochondrial DNA analysis. International Food Research Journal, 17(4), 867–876.
- National Association of Testing Authorities. Guidelines for the validation and verification of quantitative and qualitative test methods; NATA, Technical Note 17: Australia. (2012).
- Nemati, M., Oveisi, M. R., Abdollahi, H. and Sabzevari, O. (2004). Differentiation of bovine and porcine gelatins using principal component analysis. Journal of Pharmaceutical and Biomedical Analysis 34: 485–492.
- Norizah, M.S., Farah, B. and Howell, N.K. (2013). Preparation and characterization of chicken skin gelatin as an alternative to mammalian gelatin. Food Hydrocolloids 30: 143-151.
- Nur Azira, T., Che Man, Y.B., Raja Mohd Hafidz, R.N., Aina, M.A. and Amin, I. (2014). Use of principal component analysis for differentiation of gelatine sources based on polypeptide molecular weights. Food Chemistry 151: 286– 292.
- Ofner, C. M., Zhang, Y. E., Jobeck, V. C. and Bowman, B. J. (2001). Crosslinking studies in gelatin capsules treated with formaldehyde and in capsules exposed to elevated temperature and humidity. Journal of Pharmaceutical Sciences 90: 79-88.
- Paul, G. (2006). Practical guide to chemometrics. US: CRC Press, Taylor & Francis.
- Parente, A. and Sutherland, J.C. (2013). Principal component analysis of turbulent combustion data: Data pre-processing and manifold sensitivity. Combustion and Flame 160: 340-350.
- Poliana, F. A. and Suzana, C.S.L. (2013). Extraction and physicochemical characterization of gelatin from chicken by-product. Journal of Food Process Engineering 36: 824-833.
- Poppe. J. (1997). Gelatin. In Thickening and gelling agents for food, ed. A. Imeson, pp. 98-123. UK: Blackie Academic and Professional.
- Rozet, E., Marini, R.D., Ziemons, E., Boulanger, B. and Hubert, P. (2011). Advances in validation, risk and uncertainty assessment of bioanalytical methods. Journal of Pharmaceutical and Biomedical Analysis 55: 848-858.

- Rousselot International. Gelatin Manufacturer. http://www.rousselot.com. Retrieved 12 Jun 2014.
- Scheilla, V. C. S., Carlos, T. P. and Roberto, G. J. (2007). In-house method validation: Application in arsenic analysis. Journal of Food Composition and Analysis 20: 241-247.
- Schrieber, R. and Gareis, H. (2007). Gelatine handbook. Theory and industrial practice. Weinheim, Germany: Wiley-VCH.
- Scheilla, V.C.S and Roberto, G.J. (2005). A procedure to assess linearity by ordinary least squares method. Analytica Chimica Acta 552: 25-35.
- Sina, A. and Cloyce, S. (2005). Case letters: Paintball purpura. J Am Acad Dermatol 901-902.
- Smolinska, A., Blanchet, L., Buydens, L.M.C. and Wijmenga, S.S. (2012). NMR and pattern recognition methods in metabolomics: From data acquisition to biomarker discovery. A Review. Analytica Chimica Acta 750: 82-97.
- Spink, J. and Moyer, D.C. (2011). Defining the public health threat of food fraud. Journal of Food Science 76(9): R157-R163.
- Strydom, D. J. and Cohen, S. A. (1994). Comparison of amino acids analyses by phenylisothiocyanate and 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate precolumn derivatization. Analytical Biochemistry 222: 19–28.
- Taverniers, I.B., Bockstaele, E.B. and Loose, M. (2004). Trends in quality in the analytical laboratory. I. Traceability and measurement uncertainty of analytical results. Trends in Analytical Chemistry 23:480 - 490.
- Thompson, M., Ellison, S.L.R. and Wood, R. (2002). Harmonized guidelines for single-laboratory validation of methods of analysis. International Union of Pure and Applied Chemistry (IUPAC) Technical Report. Journal of Pure and Applied Chemistry 74(5): 835-855.
- Ti Nijenhuis, K. (1997). Thermoreversible network: Viscoelastic properties and structure of gels. Advances Polymer Science 130: 1-267.
- Timothy, M. D. E., Lindon, J.C. and Coen, M. (2011). Processing and Modeling of Nuclear Magnetic Resonance (NMR) Metabolic Profiles. In Metabolic Profiling. Methods in Molecular Biology, ed. T.O. Metz, pp. 365-388. Springer Science: Business Media.
- Townsend, A.A. and Nakai, S. (1983). Relationships between hydrophobicity and foaming characteristics of food proteins. Journal of Food Science 48: 588–594.

- Tromp, R. H., Ten Grotenhuis, E. and Olieman, C. (2001). Self-aggregation of gelatin above the gelling temperature analysed by SEC-MALLS. Food Hydrocolloids 16: 235-239.
- van den Berg, R.A., Hoefsloot, H.C.J., Westerhuis, J.A., Smilde, A.K. and van der Werf, M.J. (2006). Centering, scaling and transformations: Improving the biological information content of metabolomics data. BMC Genomics 7: 142.
- Vessman, J. (1996). Analytical survey. Selectivity or specificity? Validation of analytical methods from the perspective of an analytical chemist in the pharmaceutical industry. Journal of Pharmaceutical and Biomedical Analysis 14: 867-869.
- Vickie, A.V. and Elizabeth, W.C. (2014). Proteins in Food: An introduction. In Essentials of food science, part 3, pg. 117-131. New York: Food Science Text Series. Springer.
- Walfish, S., Pharmaceutical Technology. A Review of Statistical Outlier Methods, 2006, http://www.pharmtech.com. Retrieved 10 Disember 2014.
- Watanabe, S., Hiraoka, Y., Endo, S., Tanimoto, Y., Tozawa, Y and Watanabe, Y. (2015). An enzymatic method to estimate the content of L-hydroxyproline. Journal of Biotechnology 199: 9-16.
- Werteker, M., Huber, S., Kuchling, S., Rossmann, B., and Schreiner, M. (2016). Differentiation of milk by fatty acid spectra and principal component analysis.Measurement.http://dx.doi.org/10.1016/j. measurement.2016.10.059
- Widyaninggar, A., Triwahyudi and Rohman, A.K. (2012). Differentiation between porcine and bovine gelatin in commercial capsule shells based on amino acid profiles and principal component analysis. Indonesian Journal of Pharmacy 23(2): 96–101.
- Wolf F.A. (2003). Collagen and gelatin. Progress in Biotechnology, vol. 23, chapter 5. Elsevier Science B.V.
- Wu, G., Bazer, F.W., Burghardt, R.C., Johnson, G.A., Kim, S.W., Knabe, D.A., Li, P., Li, X., McKnight, J.R., Satterfield, M.C. and Spencer, T.E. (2011). Proline and hydroxyproline metabolism: implications for animal and human nutrition. Amino acids 40: 1053-1063.
- Yilmaz, M. T., Kesmen, Z., Baykal, B., Sagdic, O., Kulen, O. and Kacar, O. (2013). A novel method to differentiate bovine and porcine gelatines in food products: NanoUPLC-ESI-Q-TOF-MSE based data independent acquisition technique to detect marker peptides in gelatin. Food Chemistry 141: 2450–2458.
- Zhang, G., Liu, T., Wang, Q., Chen, L., Lei, J. and Luo, J. (2009). Mass spectrometric detection of marker peptides in tryptic digest of gelatine: A new method to differentiate between bovine and porcine gelatine. Food Hydrocolloids 23: 2001–2007.

Zhang, J., Zhang, X., Dediu, L. and Victor, C. (2011). Review of the current application of fingerprinting allowing detection of food adulteration and fraud in China. Food Control 22: 1126-1135.

