



UNIVERSITI PUTRA MALAYSIA

***IDENTIFICATION AND FUNCTIONAL ANALYSIS OF
CYTOCHROME P450 FROM *Bacillus lehensis* G1***

ANG SWI SEE

IB 2016 10



**IDENTIFICATION AND FUNCTIONAL ANALYSIS OF CYTOCHROME P450
FROM *Bacillus lehensis* G1**

By

ANG SWI SEE

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of
Philosophy**

June 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Doctor of Philosophy

**IDENTIFICATION AND FUNCTIONAL ANALYSIS OF CYTOCHROME P450
FROM *Bacillus lehensis* G1**

By

ANG SWI SEE

June 2016

Chairman : Prof. Dato' Abu Bakar Salleh, PhD
Institute : Institute of Bioscience

Cytochrome P450s (CYPs) are a superfamily of heme monooxygenases which catalyze a wide range of biochemical reactions. The reactions involve the introduction of an oxygen atom into an inactivated carbon of a compound which is essential to produce an intermediate of a hydroxylated product. Vitamin D 25-hydroxylase catalyses the first step in vitamin D biosynthetic pathway, essential in the activation of vitamin D. Several types of CYPs had been found as potential 25-hydroxylases. However, most of them originate from eukaryotes and are membrane associated proteins. A putative gene sequence encoding a CYP, termed *CYP107CB2* was found in the genome of a new isolate *Bacillus lehensis* G1, and this gene shared sequence identity with the bacterial vitamin D hydroxylase (Vdh) from *Pseudonocardia autotrophica*. In order to deepen the understanding on the properties and biological function of CYP in *B. lehensis* G1, the objective of this study was to mine for a novel CYP from *B. lehensis* G1 with hydroxylase activity on vitamin D metabolites. Computational methods to search for the novel CYP from CYP structural databases were employed to identify the conserved pattern, functional domain and sequence properties of the uncharacterized CYP from *B. lehensis* G1. The *CYP107CB2* gene was isolated and amplified using PCR and the *CYP107CB2* protein was over-expressed in *E. coli* Rosetta-gami (DE3) followed by enzyme purification via single step affinity chromatography. The biological properties and possible functions of *CYP107CB2* were characterized through absorption spectral analysis and were assayed for vitamin D hydroxylation activity. Optimization and CYP characterization were conducted to increase the turnover of hydroxylated products with an NADPH-regenerating system. Crystallization trials on *CYP107CB2* protein were conducted via preliminary screening with Crystal Screen I and II through vapour-diffusion sitting drop method. Sequence analysis studies indicated that *CYP107CB2* contained the fingerprint heme binding sequence motif FxxGxxxCxG at amino acid position 336-345 as well as other highly conserved motifs characteristic of CYP proteins. Docking studies showed several potential substrates, including vitamin D₃, 25-hydroxyvitamin D₃ and 1 α -hydroxyvitamin D₃, were located proximally to the enzyme's heme center. The

over-expressed CYP107CB2 protein was dominantly in cytosolic and the purified fraction showed a protein band at approximately 62 kDa on SDS-PAGE, representative for CYP107CB2. Spectral analysis demonstrated that the protein was properly folded and it was in its active form. HPLC and MS analysis on the product from a reconstituted enzymatic reaction confirmed that CYP107CB2 converted vitamin D₃ and 1 α -hydroxyvitamin D₃ into 25-hydroxyvitamin D₃ and 1 α ,25-dihydroxyvitamin D₃, respectively. CYP107CB2 formed crystal in formulation No. 38 from Crystal Screen II comprising 20% (v/v) PEG 10 000 and 0.1 mM HEPES buffer pH 7.5. In conclusion, a novel CYP107CB2 was identified from *B. lehensis* G1 and these findings proved that CYP107CB2 is a biologically relevant vitamin D₃ 25-hydroxylase.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**PENGENALPASTIAN DAN ANALISIS FUNGSIAN BAGI CYTOCHROME
P450 DARIPADA *Bacillus lehensis* G1**

Oleh

ANG SWI SEE

Jun 2016

Pengerusi : Prof. Dato' Abu Bakar Salleh, PhD
Institut : Institut Biosains

Cytochrome P450 (CYP) merupakan sejenis hem monooksigenase daripada satu keluarga besar yang memangkinkan pelbagai jenis tindak balas biokimia. Tindak balas tersebut melibatkan penyatuan satu atom oksigen ke dalam komponen yang mengandungi karbon yang tidak aktif di mana tindak balas ini adalah penting untuk menghasilkan bahan perantaraan bagi produk yang dihidroksilkan. Vitamin D 25-hidroksilase memangkinkan langkah yang pertama dalam laluan bio-sintetik vitamin D yang penting dalam pengaktifan vitamin D. Beberapa jenis CYP telah dijumpai sebagai 25-hidroksilase yang berpotensi. Namun demikian, kebanyakan daripada mereka berasal daripada eukariot dan merupakan protein sekutu membran. Satu urutan gen yang mengkodkan CYP digelar sebagai *CYP102CB2* telah dijumpai dalam genom isolasi baru iaitu *Bacillus lehensis* G1, dan gen ini berkongsi struktur kesamaan dengan vitamin D hidroksilase (Vdh) dari *Pseudonocardia autotrophica*. Dalam usaha untuk memperdalam pemahaman yang berkaitan dengan sifat dan fungsi biologi CYP dalam *B. lehensis* G1, objektif kajian ini adalah untuk mengenalpasti CYP yang novel dari *B. lehensis* G1, di mana CYP tersebut dapat menghidroksilkan metabolit vitamin D. Kajian ini menggunakan kaedah pengkomputeran untuk mencari CYP yang novel dari pangkalan data struktur CYP untuk mengenalpasti corak terabadi, domain fungsi dan sifat urutan CYP yang masih belum diklasifikasikan dari *B. lehensis* G1. Gene rekombinan *CYP107CB2* kemudian diperolehi daripada strain dengan menggunakan PCR (reaksi rantai polimerase) dan protein dilebih-ekspresikan dalam *E. coli* Rosetta-gami (DE3) diikuti dengan penulenan enzim melalui afiniti kromatografi langkah tunggal. Sifat-sifat biologi dan fungsi *CYP107CB2* dicirikan melalui analisis penyerapan spektrum dan dengan itu, enzim dicerakinkan untuk aktiviti vitamin D penghidroksilan. Pengoptimuman dan pencirian CYP telah dijalankan untuk meninggikan perolehan bagi penghasilan produk hidroksil dalam sistem regenerasi-NADPH. Percubaan untuk penghabluran protein *CYP107CB2* telah dijalankan melalui penyaringan awal dengan Crystal Screen I and II melalui kaedah "vapour diffusion sitting drop". Analisis urutan menunjukkan bahawa *CYP107CB2* mengandungi urutan motif ikatan hem FxxGxxxCxG pada kedudukan 336-345

serta ciri-ciri motif terabadi yang boleh didapati dalam CYP protein. Dengan menggunakan kajian dok, beberapa substrat yang berpotensi termasuk vitamin D₃, 25-hidroksivitamin D₃ dan 1 α -hidroksivitamin D₃, terletak berhampiran di pusat hem enzim. Kebanyakan protein CYP107CB2 yang dilebih-ekspresikan berada dalam bentuk sitosol dan pecahan penulenan menunjukkan jalur protein adalah bersaiz lebih kurang 62 kDa dalam SDS-PAGE di mana jalur tersebut mewakili CYP107CB2. Analisis spektrum menunjukkan bahawa protein tersebut dilipat dengan betul dan dalam bentuk aktif. Analisis HPLC dan LCMS ke atas produk hidroksil yang dihasilkan selepas tindak balas enzim mengesahkan bahawa CYP107CB2 menukarkan vitamin D₃ dan 1 α -hidroksivitamin D₃ kepada 25-hidroksivitamin D₃ dan 1 α ,25-dua-hidroksivitamin D₃, masing-masing. CYP107CB2 membentuk hablur dalam formulasi No. 38 daripada Crystal Screen II dimana formulasi tersebut mengandungi 20% (v/v) PEG 10 000 dan 0.1 mM bufer HEPES pH 7.5. Kesimpulannya, CYP107CB2 yang novel telah dikenalpasti daripada *B. lehensis* G1 dan penemuan ini membuktikan bahawa CYP107CB2 merupakan sejenis vitamin D₃ 25-hidroksilase.

ACKNOWLEDGEMENTS

First of all, I would like to express my deepest gratitude to my supervisor, Prof. Dato' Dr. Abu Bakar Salleh for being an excellent supervisor since I pursued my Master of Science until PhD in UPM. I wish to thank him for his invaluable advices, support and intensive guidance in supervising me to complete my PhD project successfully. Thank you for the trust and offering me the opportunity to become one of the postgraduate students in Enzyme and Microbial Technology Research Centre (EMTech).

My deepest gratitude goes to my supervisory members, Dr. Normi M. Yahaya, Dr. Adam Leow, Dr. Bimo Ario Tejo and Dr. Mariam Aisha Fatima for being a great instructor who has always given me the most effective source of idea and helpful discussions. I would like to acknowledge the members of EMTech, Prof. Dr. Raja Noor Zaliha, Prof. Dr. Mohd. Basyaruddin, Dr. Mohd. Shukuri, Dr. Suriana Sabri and Dr. Siti Norbaya for their generous suggestions given towards my study.

I would also like to thank all postgraduate students from Enzyme Technology Laboratory IBS, Microbial and Enzyme Technology Laboratory, and Protein Engineering Laboratory for their friendship, sharing, tolerance and helping hands during my study here. A special thanks to Dr. Muhammad Zukhrufuz Zaman and Kok Sau Yee for solving the problems on HPLC and LC-MS analysis. Thanks for their effort, time and helpful discussions whenever I needed. Not forgotten to Dr. Ely, Randa, Farhani, Adura for sharing ideas on protein crystallography. Thank you to Ariati in helping me to operate the X-ray diffractometer for crystal diffraction.

I would like to extend my acknowledgement to Universiti Putra Malaysia for the Research University Grant (05-02-12-2185RU) for financing this project. I also gratefully acknowledge Malaysia Genome Institute (MGI) for providing the bacteria strain and its genome data. I would like to thank Ministry of Education, Malaysia for MyPhD scholarship award.

My gratitude also goes to Mr. Wahidin for allowing me to use the HPLC machine for sample analysis. I would also like to thank the staff from Institute of Bioscience, School of Graduate Study (SGS) and Faculty of Biotechnology and Biomolecular Sciences for their help and support.

I am really grateful to my friends and housemates who were always patient, encouraging and cared for me and make my time cheerful.

Most importantly, I would like to express my utmost appreciation to my family for their constant pressure in completing my study, support and unceasing love throughout the period of endeavor.

Last but not least, I would like to express my sincere thanks to anyone else whose name is not mentioned here for their invaluable help and contributions making this piece of work possible. Thank you very much.

I certify that a Thesis Examination Committee has met on 06 June 2016 to conduct the final examination of Ang Swi See on her thesis entitled "Identification and Functional Analysis of Cytochrome P450 from *Bacillus lehensis* G1" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Mohd Basyaruddin bin Abdul Rahman, PhD

Professor
Faculty of Sciences
Universiti Putra Malaysia
(Chairman)

Parameswari a/p Namasivayam, PhD

Associated Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Internal Examiner)

Rosfarizan binti Mohamad, PhD

Associated Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Internal Examiner)

Andrew W. Munro, PhD

Professor
The University of Manchester
United Kingdom
(External Examiner)



ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 26 July 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Abu Bakar Salleh, PhD

Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Leow Thean Chor, PhD

Senior Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Normi Mohd. Yahaya, PhD

Senior Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Bimo Ario Tejo, PhD

Senior Lecturer
Faculty of Life Sciences
Surya University
(Member)

Mariam Aisha Fatima, PhD

Senior Lecturer
Faculty of Health and Life Sciences
Management and Science University
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Ang Swi See GS30893

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: Professor Dr. Abu Bakar Salleh

Signature: _____
Name of Member
of Supervisory
Committee: Dr. Leow Thean Chor

Signature: _____
Name of Member
of Supervisory
Committee: Dr. Normi Mohd. Yahaya

Signature: _____
Name of Member
of Supervisory
Committee: Dr. Bimo Ario Tejo

Signature: _____
Name of Member
of Supervisory
Committee: Dr. Mariam Aisha Fatima

TABLE OF CONTENTS

		Page
ABSTRACT		i
ABSTRAK		iii
ACKNOWLEDGEMENTS		v
APPROVAL		vi
DECLARATION		viii
LIST OF TABLES		xiv
LIST OF FIGURES		xv
LIST OF APPENDICES		xix
LIST OF ABBREVIATIONS		xx
CHAPTER		
1	INTRODUCTION	1
	1.1 Hypothesis and problem statement	1
	1.2 Objectives	3
	1.3 Significance of study	4
2	LITERATURE REVIEW	5
	2.1 Cytochrome P450 (CYP)	5
	2.2 The gene and structure of CYP	6
	2.2.1 Three dimensional structures of CYP	8
	2.2.2 Catalytic mechanism of CYP	10
	2.3 The advantages and limitations of CYP	12
	2.4 Cloning and expression of CYP	13
	2.5 Applications of CYPs in biotechnology	14
	2.6 Bacterial CYP in vitamin D hydroxylation	16
	2.7 The metabolism of vitamin D in humans	17
	2.8 Vitamin D hydroxylase	20
	2.8.1 Vitamin D 25-hydroxylase	20
	2.8.2 Vitamin D 1 α -hydroxylase	24
	2.8.3 Vitamin D 24-hydroxylase	25
	2.9 Vitamin D deficiency	26
	2.9.1 The source and importance of vitamin D	28
	2.9.2 Factors causing the deficiency of vitamin D	29
	2.10 <i>Bacillus</i> sp. in biotechnology	30
3	BIOINFORMATICS ANALYSIS OF CYP SEQUENCE FROM <i>B. lehensis</i> G1	32
	3.1 Introduction	32
	3.2 Materials and methods	33
	3.2.1 Analysis of nucleotide and amino acid sequence of CYP from <i>B. lehensis</i> G1	33
	3.2.2 Multiple sequence alignment	33
	3.2.3 Homology modeling of CYP from <i>B. lehensis</i> G1	34

3.2.4	Validation of CYP model	34
3.2.5	Molecular docking with vitamin D ₃ , 25-hydroxyvitamin D ₃ and 1 α -hydroxyvitamin D ₃	34
3.3	Results and discussion	35
3.3.1	Molecular analysis of CYP gene from <i>B. lehensis</i> G1	35
3.3.2	Multiple sequence alignment of CYP proteins	38
3.3.3	Homology modelling of CYP107CB2	41
3.3.4	Validation of the CYP107CB2 model and template superimposition	43
3.3.5	Molecular docking of CYP107CB2	46
3.4	Conclusion	52
4	CLONING, EXPRESSION, PURIFICATION AND MOLECULAR WEIGHT DETERMINATION OF CYP (CYP107CB2) FROM <i>B. lehensis</i> G1	54
4.1	Introduction	54
4.2	Materials and methods	54
4.2.1	Native bacteria and strain maintenance	54
4.2.2	Genomic DNA extraction of <i>B. lehensis</i> G1	55
4.2.3	Amplification of CYP gene	55
4.2.4	Purification of PCR products	56
4.2.5	Cloning of <i>CYP107CB2</i> gene into pET102/D-TOPO vector	56
4.2.6	Preparation of <i>Escherichia coli</i> competent cells	57
4.2.7	Transformation of recombinant plasmid (pET102/D-TOPO: <i>CYP107CB2</i>) into <i>E. coli</i> TOP10	57
4.2.8	Plasmid extraction	57
4.2.9	Transformation of recombinant plasmid (pET102/D-TOPO: <i>CYP107CB2</i>) into <i>E. coli</i> Rosetta-gami (DE3)	58
4.2.10	4.2.10 Analysis of positive transformed recombinant plasmid (pET102/D-TOPO: <i>CYP107CB2</i>)	58
4.2.11	Agarose gel electrophoresis	59
4.2.12	Protein expression	59
4.2.13	Optimization of <i>CYP107CB2</i> expression	60
4.2.14	Purification of <i>CYP107CB2</i>	60
4.2.15	Protein content determination	61
4.2.16	Identification of <i>CYP107CB2</i> protein	61
4.3	Results and discussion	63
4.3.1	Genomic DNA and PCR product	63
4.3.2	Cloning of <i>CYP107CB2</i> gene into <i>E. coli</i> TOP10	65
4.3.3	Transformation of recombinant plasmid (pET102/D-TOPO: <i>CYP107CB2</i>) into <i>E. coli</i> Rosetta-gami (DE3)	66
4.3.4	Sequencing analysis	69

4.3.5	Protein expression	71
4.3.6	Purification of CYP107CB2	74
4.3.7	Western blot analysis	77
4.4	Conclusion	78
5	FUNCTIONAL CHARACTERIZATION, OPTIMIZATION AND BIOCHEMICAL ANALYSIS OF CYP107CB2 FROM <i>B. lehensis</i> G1	79
5.1	Introduction	79
5.2	Materials and methods	80
5.2.1	UV-Vis spectroscopy analysis of CYP	80
5.2.2	Measurement of CYP activity	80
5.2.3	Biochemical characterizations of purified CYP107CB2	83
5.2.4	Statistical analysis	85
5.3	Results and discussion	85
5.3.1	UV-Vis spectroscopy	85
5.3.2	Measurement of enzyme activity	86
5.3.3	Mass spectroscopy analysis	94
5.3.4	Putative hydroxylation pathway of CYP107CB2	99
5.3.5	Optimization and biochemical analysis of CYP107CB2 in an NADPH-regenerating system	99
5.4	Conclusion	112
6	STRUCTURAL STUDY AND CRYSTALLIZATION TRIALS OF CYP107CB2 PROTEIN	114
6.1	Introduction	114
6.2	Materials and methods	114
6.2.1	Circular dichroism spectroscopy	114
6.2.2	Overproduction and purification of recombinant CYP107CB2	115
6.2.3	Protein crystallization of CYP107CB2	115
6.2.4	Optimization of CYP107CB2 crystallization	116
6.2.5	Protein crystal validation	117
6.2.6	X-ray diffraction on CYP107CB2 crystal using an in-house diffractometer	117
6.3	Results and discussion	117
6.3.1	Thermal stability and secondary structure of CYP107CB2 protein	117
6.3.2	Preliminary screening of CYP107CB2 crystallization	120
6.3.3	Protein crystal validation	122
6.3.4	Optimization of CYP107CB2 crystallization using sitting drop vapour-diffusion method	124
6.3.5	X-ray diffraction analysis	129
6.4	Conclusion	131

7	SUMMARY AND RECOMMENDATION FOR FUTURE WORK	132
7.1	Summary and general conclusion	132
7.2	Recommendation for future work	134
	REFERENCES	137
	APPENDICES	155
	BIODATA OF STUDENT	162
	LIST OF PUBLICATIONS	163



LIST OF TABLES

Table		Page
2.1	List of reactions catalyzed by CYPs	6
2.2	The number of CYP genes identified in the genome sequences of some bacteria	7
2.3	Bacterial cytochrome CYPs and their functionality	7
2.4	The crystal structures of bacterial CYP and its substrate	8
2.5	The summary of the advantages and limitations of CYP application	13
2.6	CYP enzymes with 25-hydroxylation activity	23
2.7	Catalytic properties of CYP 25-hydroxylase activity	24
2.8	Risk factors for vitamin D deficiency	30
3.1	Protein sequence analysis of CYP107CB2 from <i>B. lehensis</i> G1 using NCBI protein BLAST search database against PDB	42
3.2	Binding energy and predicted contacting residues of CYP107CB2 that interact with vitamin D ₃ , 25-hydroxyvitamin D ₃ and 1 α -hydroxyvitamin D ₃	47
4.1	Thermal cycling conditions for amplification of <i>CYP107CB2</i> gene sequence	56
4.2	Composition of SDS-PAGE analysis	62
5.1	HPLC gradient elution profile for the separation of vitamin D ₃ metabolites	81
5.2	LC-MS gradient elution profile for the separation of 1 α ,25-dihydroxyvitamin D ₃	82
5.3	Summary of the hydroxylation reaction and hydroxylated product produced by CYP107CB2	88
5.4	Formation of hydroxylated product 1 α ,25-dihydroxyvitamin D ₃ by CYP107CB2 before and after optimization	111

LIST OF FIGURES

Figure		Page
2.1	General topology of CYP	9
2.2	Prosthetic of cysteinato-heme enzymes: an iron- (III) protoporphyrin-IX linked with a proximal cysteine ligand	10
2.3	Monooxygenation reaction catalyzed by CYP	11
2.4	The CYP catalytic cycle with the compound I-like ferryl species highlighted by a blue square	12
2.5	The synthesis and metabolism of vitamin D	18
2.6	The conversion of 7-dehydrocholesterol to pre-vitamin D ₃ by UV-B radiation and the formation of vitamin D ₃ induced by body temperature	19
2.7	Metabolic activation of vitamin D ₃ to its hormonal form, 1 α ,25-dihydroxyvitamin D ₃ in liver and kidney	20
2.8	Degradation of 1 α ,25-dihydroxyvitamin D ₃ into inactive excretion product, calcitric acid by CYP24	26
2.9	Nutrition form of vitamin D	28
3.1	Full length nucleotide and amino acid sequence of CYP107CB2 in <i>B. lehensis</i> G1	36
3.2	The search of the conserved domain of CYP107CB2 from <i>B. lehensis</i> G1 in NBCI CDD database	37
3.3	Theoretical isoelectric point and molecular weight of amino acid sequence of CYP107CB2 from <i>B. lehensis</i> G1	38
3.4	Multiple sequence alignment of CYP107CB2 and CYP107 superfamily of protein sequences	40
3.5	Homology model of CYP107CB2	42
3.6	The three dimensional view of heme prosthetic group and amino acid residues of CYP107CB2	43
3.7	Quality assessment of CYP107CB2 built model	44
3.8	Superimposed structure of homologue model CYP107CB2 with the structure of CYP Vdh (PDB: 3A4G)(magenta)	46

3.9	The potential substrates for <i>in silico</i> docking simulation of CYP107CB2	48
3.10	Possible binding mode of CYP107CB2 and its candidate substrates for docking analysis	49
3.11	Docking models of CYP107CB2 with the potential substrates within its active site	51
4.1	Agarose gel electrophoresis of extracted genomic DNA from <i>B. lehensis</i> G1	64
4.2	Agarose gel electrophoresis of PCR product of gene encoding CYP107CB2 from <i>B. lehensis</i> G1	64
4.3A	Schematic diagram of direct cloning of CYP107CB2 PCR product into pET102/D-TOPO vector	65
4.3B	Agarose gel electrophoresis of plasmids extraction from <i>E. coli</i> TOP10	66
4.4	<i>E. coli</i> Rosetta-gami (DE3) was grown on LB agar plate after heat shock transformation	67
4.5	Agarose gel electrophoresis of plasmids extraction from <i>E. coli</i> Rosetta-gami (DE3)	68
4.6	Agarose gel electrophoresis of PCR product generated from extracted plasmid	68
4.7	Agarose gel electrophoresis of single digested plasmids using <i>Sac</i> I restriction enzyme on empty and recombinant plasmid	69
4.8	Sequence alignment of sequencing result and putative CYP107CB2 gene sequence of <i>B. lehensis</i> G1	71
4.9	SDS-PAGE analysis of CYP107CB2 protein expression at different induction temperatures by <i>E. coli</i> Rosetta gami (DE3)	72
4.10	SDS-PAGE analysis of CYP107CB2 protein expression in <i>E. coli</i> Rosetta-gami (DE3) at different concentrations of inducer (IPTG)	74
4.11	The appearance of purified CYP107CB2 protein solution in 50 mM phosphate buffer, pH 7.5	75
4.12	SDS-PAGE analysis of cytosolic fraction of recombinant <i>E. coli</i> cells and the purified samples of CYP107CB2	76

4.13	Native-PAGE analysis of CYP107CB2 from <i>B. lehensis</i> G1	76
4.14	Western blot analysis of CYP107CB2 protein	78
5.1	Spectral analysis of CYP107CB2 protein	86
5.2	The retention time of vitamin D ₃ standards	87
5.3	HPLC analysis of the hydroxylated products of vitamin D ₃ catalyzed by CYP107CB2	89
5.4	HPLC analysis of the hydroxylated products of 1 α -hydroxyvitamin D ₃ catalyzed by CYP107CB2	91
5.5	HPLC analysis of the hydroxylated products of 25-hydroxyvitamin D ₃ catalyzed by CYP107CB2	92
5.6	HPLC chromatograms of hydroxylated product formed from 1 α -hydroxyvitamin D ₃ in different conditions of reaction mixture	94
5.7	HPLC chromatograms of hydroxylated products	95
5.8	Mass spectrum of 1 α ,25-dihydroxyvitamin D ₃	96
5.9	Mass spectrum of 25-hydroxyvitamin D ₃	98
5.10	The proposed hydroxylation pathways of A) vitamin D ₃ and B) 1 α -hydroxyvitamin D ₃ catalyzed by CYP107CB2 which acts as 25-hydroxylase	100
5.11	Effect of temperatures on the production of 1 α ,25-dihydroxyvitamin D ₃	101
5.12	Residual activity of purified CYP107CB2 towards 1 α -hydroxyvitamin D ₃	102
5.13	Effect of temperatures on the stability of CYP107CB2	103
5.14	Effect of pHs on the production of 1 α ,25-dihydroxyvitamin D ₃	104
5.15	Effect of pHs on the stability of CYP107CB2	105
5.16	Time course of 1 α ,25-dihydroxyvitamin D ₃ formation in the present and absence of NADPH regenerating system by using 1 α -hydroxyvitamin D ₃ as a substrate	108

5.17	Predicted electron flow in the catalytic system of CYP107CB2 in hydroxylating 1 α -hydroxyvitamin D ₃ into 1 α ,25-dihydroxyvitamin D ₃	109
5.18	Effect of substrate concentration on the production of 1 α ,25-dihydroxyvitamin D ₃ by CYP107CB2	110
5.19	HPLC chromatograms of hydroxylated product, 1 α ,25-dihydroxyvitamin D ₃ before and after optimization and characterization analysis	112
6.1	Crystal screening using 96 well plate	116
6.2	CD spectrum of CYP107CB2 fusion protein in 10 mM phosphate buffer pH 7.5	119
6.3	Temperature induced denaturation of CYP107CB2 fusion protein	120
6.4	Color appearance of heme-containing CYP107CB2 protein	121
6.5	The crystals of CYP107CB2 obtained from an initial screening for crystallization	122
6.6	Protein crystals stained by IZIT dye	124
6.7	The outcomes of CYP107CB2 crystallization with the variation of temperatures	126
6.8	The outcomes of CYP107CB2 crystallization with the variation of pH buffer	127
6.9	The outcomes of CYP107CB2 crystallization with the variation of PEG 10000 concentration at pH 8	128
6.10	The outcomes of CYP107CB2 crystallization with the variation of protein concentration	130
6.11	Diffraction pattern of a plate-like crystal as conducted by simple scan measurement using in house X-ray diffractometer	131

LIST OF APPENDICES

Appendix		Page
A1	The appearance of <i>B. lehensis</i> G1 after 24 h of incubation at 30 °C in Hirokoshi agar plate	156
A2	The map of pET102/D-TOPO® vector	156
A3	The feature of pET102/D-TOPO® vector	158
A4	TOPO® Cloning Site of pET102/D-TOPO® vector	159
A5	Standard curve of protein by using bovine serum albumin (BSA) as a standard	159
A6	Recipes for running buffer, sample buffer, staining and de-staining solution	160
A7	Standard curve of 25-hydroxyvitamin D ₃	160
A8	Standard curve of 1 α ,25-dihydroxyvitamin D ₃	161

LIST OF ABBREVIATIONS

α	alpha
Å	Angstrom
β	beta
BLAST	Basic Local Alignment Search Tool
BSA	bovine serum albumin
bp	base pair
CD	circular dichroism
CDD	conserved domain database
cm	centimeter
CO	carbon monoxide
CV	column volume
CYP or P450	cytochrome P450
°C	degree Celsius
δ	delta
DMSO	dimethyl sulfoxide
DNA	deoxyribonucleic acid
DIMS	direct infuse mass spectrometry
EC	enzyme classification
EDTA	ethylenediaminetetraacetic acid
ESI	electrospray ionization source
ϵ	extinction coefficient
Fe ²⁺	ferrous
FAD	flavin adenine dinucleotide
Fdr	ferredoxin NADP ⁺ reductase

Fdx	ferredoxin
g	gram
x g	gravitational force
h	hour
HPLC	high performance liquid chromatography
IPTG	isopropyl β -D-thiogalactoside
kb	kilo base pair
kDa	kilo Dalton
kV	kilovolts
L	liter
LB	Luria-Bertani
LC	liquid chromatography
LC-MS	liquid chromatography-mass spectroscopy
M	Molar
MS	mass spectroscopy
mg	miligram
min	minute
mL	milliliter
mM	milimolar
mm	millimeter
m/z	mass to charge ratio
NaCl	sodium chloride
NADPH	nicotinamide adenine dinucleotide phosphate
NaOH	sodium hydroxide
NCBI	National Center for Biotechnology Information

ng	nanogram
nm	nanometer
NMR	nuclear magnetic resonance
OD	optical density
ORF	open reading frame
PCR	polymerase chain reaction
PDB	Protein Data Bank
PEG	polyethylene glycol
ppm	part per million
psi	per square inch
Q-TOF	quadrupole time-of-flight
RMSD	root mean square deviation
RPM	revolutions per minute
RT	retention time
SDS-PAGE	sodium dodecyl sulphate-polyacrylamide gel electrophoresis
SQD	single quadrupole detector
TEMED	N, N, N, N-tetramethylenediamide
T _m	melting temperature
Tris-HCl	tris hydrochloric acid
UV-Vis	ultraviolet-visible
V	voltage
v/v	volume per volume
w/v	weight per volume
µg/mL	microgram/mililiter
µL	microliter

μm	micrometer
μM	micromolar
μmol	micromole
YASARA	Yet Another Scientific Artificial Reality Application



CHAPTER 1

INTRODUCTION

1.1 Hypothesis and problem statement

Cytochromes P450s are a superfamily of heme monooxygenases which are present in a wide variety of organisms in nature (Bernhardt, 2006; Zurek et al., 2006). These proteins are involved in monooxygenation, activates dioxygen to incorporate a single oxygen atom into a wide range of substrates (Jin et al., 2004; Zurek et al., 2006). They catalyze a broad range of biochemical reactions and play an essential role in the assimilation of carbon source. The reactions include hydroxylations, epoxidations, sulfoxidations, and so on as stated by Bernhardt, (2006). The typical reaction catalyzed by CYP is the hydroxylation of organic substrates on carbon atoms (Munro et al., 2007). Prototypical reactions catalyzed by CYPs include fatty acid hydroxylation, secondary metabolite biosynthesis and drug metabolism (Schallmeyer et al., 2011; Sono et al., 1996). They oxidize non-activated carbon at either carbon ring or lateral alkyl chain which is difficult to be achieved via chemical reactions, making CYP versatile and attractive in various fields (Schlichting et al., 2000; Lombard et al., 2011).

In biotechnology point of view, CYPs are known as interesting potential 'green' catalysts (Bernhard and Urlacher, 2014; Farinas et al., 2004) and gained much attention for the production of useful fine chemicals (Yun et al., 2007). The practical goal in CYPs research is to search for the enzymes which are able to produce chemicals that are difficult to be prepared by traditional organic synthetic (Bhattarai et al., 2012). This is due to its exclusive spectroscopic properties, catalytic diversity, and broad substrate range variety of useful chemical transformations that are essential in both biological function and chemical synthesis (Sigman et al., 1999). The enzymes produce the fine chemical in regio and stereo-selective way making CYP a versatile and powerful oxidant. Therefore, CYP had gained much attention in producing many useful fine chemicals of hydroxylated products which are essential in both biological function and chemical synthesis (Sigman et al., 1999; Urlacher and Girhard, 2012; Yun et al., 2007).

CYPs cover a wide range of applications in the production of drugs, drug metabolites and work as catalysts in various chemical (Gillam 2008; Jung et al., 2011). Despite their exclusive catalytic capabilities, only a limited number have been exploited in preparative of chemical reactions or industrial chemical processes (Julsing et al., 2008). As reported by Jung et al. (2011), many natural CYPs are insoluble, expressed at low levels, and exhibit activity insufficient for practical biocatalysis.

The biotransformation of vitamin D₃ to 1 α ,25-dihydroxyvitamin D₃ is one of the most successful applications of CYP in biotechnology. The product has been used to treat numerous diseases such as osteoporosis, chronic renal failure and hypothyroidism (Sakaki, 2012; Yasutake et al., 2009). Vitamin D₃ is a biologically inactive compound that requires one or more CYPs to catalyze the formation of the most active form of vitamin D hormone, 1 α ,25-dihydroxyvitamin D₃ or calcitriol. The major function of calcitriol is to maintain safe levels of calcium and phosphorus in the blood, by regulating the absorption of these ions in the intestine, bone and kidney. In addition, calcitriol is also recognized as an important anti-proliferative factor for dividing cells and tissues, as vitamin D deficiency is now linked to over 20 forms of cancer (Guyton et al., 2003). The chemical synthesis of 1 α ,25-dihydroxyvitamin D₃ requires complex procedures, including almost 20 reaction steps with low production yields. Therefore, development of an efficient and simplified production process for calcitriol remains an important area of investigation (Sakaki et al., 2011; Zhu and Okamura, 1995).

In the process of vitamin D₃ conversion to 1 α ,25-dihydroxyvitamin D₃, the hydroxylation of the side chain at C-25 or C-24 is a prerequisite. The enzyme-catalyzed vitamin D₃ 25-hydroxylation is an essential step for the activation of vitamin D₃ and important for understanding the entire activation process of vitamin D₃ (Aiba et al., 2006). Knowledge on vitamin D 25-hydroxylation by bacterial CYP is limited, although their involvement in several essential bioconversions has been reported. Further exploration of bacterial CYP enzymes which could metabolize vitamin D₃ is necessary.

There are several types of CYP vitamin D 25-hydroxylases and most of them are of eukaryotic origin. They are mostly present in mitochondria and microsomes of liver (Yamasaki et al., 2004). These include CYP27A1, CYP2R1, CYP3A4, CYP2C11, CYP2D25, CYP2J3 and CYP2J2. They have been found to be fairly important and critical compounds for the regulation of vitamin D metabolism as well as other hydroxylation processes in eukaryotic system. Additionally, some of these 25-hydroxylases were well identified and characterized (Aiba et al., 2006; Zhu and DeLuca, 2012).

Although eukaryotic CYPs are well studied, one of the common limitations of these proteins is that they are membrane-bound (McClean et al., 2011; Werck-Reichhart and Feyereisen, 2000). The microsomal and mitochondrial 25-hydroxylases are associated in the endoplasmic reticulum and inner membrane of mitochondria, respectively (Zhu and DeLuca, 2012). Contrastingly, bacterial CYP in particular is often soluble and targeting in cytoplasm as lacking of the N-terminal membrane anchor (Bernhardt, 2006; McClean et al., 2011; Werck-Reichhart and Feyereisen, 2000). Its CYPs are found to be useful for oxidation of various compounds including drugs, steroids and fatty acids (Hilker et al., 2008; Lewis and Wiseman, 2005). From a practical point of view, bacterial CYPs can be easily isolated and manipulated and the proteins remain in soluble active

form as compared to insoluble eukaryote CYPs (Budde et al., 2004; Urlacher and Eiben, 2006).

Since, there is an increasing industrial demand to exploit CYP as a valuable biocatalyst; considerable attempts have been devoted to search for novel enzymes with unique metabolic properties. This strategy was attempted in this study, where a novel CYP was mined from the genome of alkaliphilic bacteria, *Bacillus lehensis* G1. This bacterium dwells in soil with the capability to thrive at high pH up to 11. A survey on the complete *B. lehensis* G1 genome revealed that the strain which is usually used for the production of cyclodextrin glucanotransferase (Noor et al., 2014) contained a single candidate gene that potentially coded for a functional CYP enzyme.

A single cDNA specifying soluble CYP enzyme termed as CYP107CB2 exhibited sequence identity of 44% with vitamin D₃ hydroxylase (Vdh) from *Pseudonocardia autotrophica* which had been grouped into the CYP107 family of enzymes. CYP Vdh is a vitamin D₃ hydroxylase (protein data bank ID: 3A4G; resolution: 1.75 Å) responsible in hydroxylating vitamin D₃ to 1 α ,25-dihydroxyvitamin D₃ via 25-hydroxylation and 1 α -hydroxylation (Fujii et al., 2009; Yasutake et al., 2010). The lack of structural information for the putative CYP107CB2 from *B. lehensis* G1 prevented a more detailed characterization of its biological role. In this respect, this study seeks to address the hypothesis that an identified CYP biocatalyst from *B. lehensis* G1 is an active protein and could be used to hydroxylate vitamin D₃ metabolites.

The study was divided into four major experiments. The first was on bioinformatics analysis of CYP sequence from *B. lehensis* G1 followed by second experiment to clone, express, purify and confirm the presence of CYP107CB2 protein via molecular weight evaluation. The third experiment determined the characteristics and biological function of CYP107CB2 through UV-Vis spectroscopy analysis and hydroxylation assay. Optimization and characterization of CYP107CB2 to increase the production of hydroxylated products were also conducted. Last experiment was on crystallization trial of CYP107CB2 protein and X-ray diffraction on protein crystal.

1.2 Objectives

The main objective of this study was to mine, identify and characterize the putative CYP107CB2 from *B. lehensis* G1. The specific objectives were:

1. To conduct bioinformatics analysis on CYP sequence from the genome of *B. lehensis* G1
2. To clone, express, purify and determine the molecular weight of CYP protein
3. To characterize the spectral properties and biological function of CYP and to optimize the production of hydroxylated product

4. To crystallize the CYP protein and diffract the CYP crystal using in-house X-ray diffractometer.

1.3 Significance of study

Present study demonstrates vitamin D₃ 25-hydroxylase derived from an environmental bacilli strain in identifying the metabolic function of a new CYP107CB2. The study provides useful insights into the nature of substrate selection for CYP107CB2, which can guide future docking studies, as well as functional experiments. The discovery of a CYP107CB2 that acts as a 25-hydroxylase, has suggested the contribution of CYP family to the metabolism of vitamin D₃. This study is important as CYP107CB2 can serve as a microbial model for eukaryotic's 25-hydroxylase and provide an alternative to study the metabolism of vitamin D₃ without the technical difficulties of handling insoluble membrane proteins.

REFERENCES

- Aiba, I., Yamasaki, T., Shinki, T., Izumi, S., Yamamoto, K., Yamada, S., ... Ohyama, Y. (2006). Characterization of rat and human CYP2J enzymes as Vitamin D 25-hydroxylases. *Steroids*, 71(10), 849–856.
- Altschul, S. F., Madden, T. L., Schäffer, A. A., Zhang, J., Zhang, Z., Miller, W., & Lipman, D. J. (1997). Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Research*, 25(17), 3389–402.
- Anderson, P. H., May, B. K., & Morris, H. A. (2003). Vitamin D metabolism: new concepts and clinical implications. *The Clinical Biochemist. Reviews / Australian Association of Clinical Biochemists*, 24(1), 13–26.
- Andersson, S., & Jörnvall, H. (1986). Sex differences in cytochrome P-450-dependent 25-hydroxylation of C27-steroids and vitamin D3 in rat liver microsomes. *Journal of Biological Chemistry*, 261(36), 16932–16936.
- Andersson, S., Holmberg, I., & Wikvalls, K. (1983). 25-Hydroxylation of C27-Steroids and Vitamin D3 by a Constitutive Cytochrome P-450 from Rat Liver Microsomes *. *Journal of Biological Chemistry*, 258(11), 3–8.
- Arnold, K., Bordoli, L., Kopp, J., & Schwede, T. (2006). The SWISS-MODEL workspace: a web-based environment for protein structure homology modelling. *Bioinformatics (Oxford, England)*, 22(2), 195–201.
- Axén, E., Bergman, T., & Wikvall, K. (1994). Microsomal 25-hydroxylation of vitamin D2 and vitamin D3 in pig liver. *The Journal of Steroid Biochemistry and Molecular Biology*, 51(1-2), 97–106.
- Banik, R. M., Prakash, R., & Upadhyay, S. N. (2008). Microbial biosensor based on whole cell of *Pseudomonas* sp. for online measurement of p-Nitrophenol. *Sensors and Actuators B: Chemical*, 131(1), 295–300.
- Bateman, A. (2002). The Pfam Protein Families Database. *Nucleic Acids Research*, 30(1), 276–280.
- Battestin, V., & Macedo, G. A. (2007). Purification and Biochemical Characterization of Tannase from a Newly Isolated Strain of *Paecilomyces Variotii*. *Food Biotechnology*, 21(3), 207–216.
- Beedkar, S. D., Khobragade, C. N., Bodade, R. G., & Vinchurkar, A. S. (2012). Comparative structural modeling and docking studies of uricase: possible implication in enzyme supplementation therapy for hyperuricemic disorders. *Computers in Biology and Medicine*, 42(6), 657–666.

- Bergfors, T. M. (2009). Protein Crystallization (Second). International University Line.
- Bernhardt, R. (2006). Cytochromes P450 as versatile biocatalysts. *Journal of Biotechnology*, 124(1), 128–45.
- Bernhardt, R., & Urlacher, V. B. (2014). Cytochromes P450 as promising catalysts for biotechnological application: chances and limitations. *Applied Microbiology and Biotechnology*, 98(14), 6185–203.
- Bhattacharai, S., Liou, K., & Oh, T. J. (2012). Homology modeling and docking studies of *Streptomyces peucetius* CYP147F1 as limonene hydroxylase. *Journal of Microbiology and Biotechnology*, 22(7), 917–922.
- Bikle, D. D. (2014). Vitamin D metabolism, mechanism of action, and clinical applications. *Chemistry & Biology*, 21(3), 319–29.
- Bisswanger, H. (2014). Enzyme assays. *Perspectives in Science*, 1(1-6), 41–55.
- Bitto, N. J., Graichen, F. H. M., & Monahan, B. J. (2009). Functionality at the end of a fatty acid chain – chemical and biological routes to ω -hydroxylated fatty acids. *Lipid Technology*, 21(10), 216–219.
- Blanco, K. C., Lima, C. J. B., Monti, R., Martins, J., Bernardi, N. S., & Contiero, J. (2011). *Bacillus lehensis*—an alkali-tolerant bacterium isolated from cassava starch wastewater: optimization of parameters for cyclodextrin glycosyltransferase production. *Annals of Microbiology*, 62(1), 329–337.
- Bradford, M. (1976). A Rapid and Sensitive Method for the Quantitation of Microgram Quantities of Protein Utilizing the Principle of Protein-Dye Binding. *Analytical Biochemistry*, 72(1-2), 248–254.
- Budde, M., Maurer, S. C., Schmid, R. D., & Urlacher, V. B. (2004). Cloning, expression and characterisation of CYP102A2, a self-sufficient P450 monooxygenase from *Bacillus subtilis*. *Applied Microbiology and Biotechnology*, 66(2), 180–186.
- Cali, J. J., & Russell, D. W. (1991). Characterization of human sterol 27-hydroxylase. A mitochondrial cytochrome P-450 that catalyzes multiple oxidation reaction in bile acid biosynthesis. *The Journal of Biological Chemistry*, 266(12), 7774–8.
- Chang, Y. T., Stiffelman, O. B., Vakser, I. A., Loew, G. H., Bridges, A., & Waskell, L. (1997). Construction of a 3D model of cytochrome P450 2B4. *Protein Engineering*, 10 (2), 119–129.
- Chang, Z., Li, L., Pan, Z., & Wang, X. (2008). Crystallization and preliminary X-ray analysis of allene oxide synthase, cytochrome P450 CYP74A2, from *Parthenium argentatum*. *Acta Crystallographica Section F Structural Biology and Crystallization Communications*, 64(7), 668–670.

- Chaudhuri, R., Cheng, Y., Middaugh, C. R., & Volkin, D. B. (2014). High-throughput biophysical analysis of protein therapeutics to examine interrelationships between aggregate formation and conformational stability. *The AAPS Journal*, 16(1), 48–64.
- Chayen, N. E., & Saridakis, E. (2008). Protein crystallization: from purified protein to diffraction-quality crystal. *Nature Methods*, 5(2), 147–53.
- Cheng, J. B., Levine, M. a, Bell, N. H., Mangelsdorf, D. J., & Russell, D. W. (2004). Genetic evidence that the human CYP2R1 enzyme is a key vitamin D 25-hydroxylase. *Proceedings of the National Academy of Sciences of the United States of America*, 101(20), 7711–5.
- Cheng, J. B., Motola, D. L., Mangelsdorf, D. J., & Russell, D. W. (2003). De-orphanization of cytochrome P450 2R1: a microsomal vitamin D 25-hydroxylase. *The Journal of Biological Chemistry*, 278(39), 38084–93.
- Christakos, S., Ajibade, D. V, Dhawan, P., Fechner, A. J., & Mady, L. J. (2010). Vitamin D: metabolism. *Endocrinology and Metabolism Clinics of North America*, 39(2), 243–53, table of contents.
- Clemens, T. L., Henderson, S. L., Adams, J. S., & Holick, M. F. (1982). Increased skin pigment reduces the capacity of skin to synthesis vitamin D3. *The Lancet*, 319(8263), 74–76.
- Cryle, M. J., Stok, J. E., & De Voss, J. J. (2003). Reactions Catalyzed by Bacterial Cytochromes P450. *Australian Journal of Chemistry*, 56(8), 749.
- Cui, Y., & Rohan, T. E. (2006). Vitamin D, calcium, and breast cancer risk: a review. *Cancer Epidemiology, Biomarkers & Prevention : A Publication of the American Association for Cancer Research, Cosponsored by the American Society of Preventive Oncology*, 15(8), 1427–37.
- Cupp-Vickery, J. R., & Poulos, T. L. (1995). Structure of cytochrome P450eryF involved in erythromycin biosynthesis. *Nature Structural and Molecular Biology*, 2(2), 144–153.
- Dahlbäck, H., & Wikvall, K. (1988). 25-Hydroxylation of vitamin D3 by a cytochrome P-450 from rabbit liver mitochondria. *The Biochemical Journal*, 252(1), 207–13.
- Daniel, R., Dines, M., & Petach, H. (1996). The denaturation and degradation of stable enzymes at high temperatures. *Biochemical Journal*, 11, 1–11.
- Davis, B. J. (1964). Disc electrophoresis. II. Method and application to human serum proteins. *Annals of the New York Academy of Sciences*, 121, 404–27.

- DeLuca, H. F. (2004). Overview of general physiologic features and functions of vitamin D. *The American Journal of Clinical Nutrition*, 80(6 Suppl), 1689S–96S.
- DeLuca, H. F. (2014). History of the discovery of vitamin D and its active metabolites. *BoneKEy Reports*, 3(JANUARY), 479.
- Deng, A., Wu, J., Zhang, Y., Zhang, G., & Wen, T. (2010). Purification and characterization of a surfactant-stable high-alkaline protease from *Bacillus* sp. B001. *Bioresource Technology*, 101(18), 7100–7106.
- Denisov, I. G., Frank, D. J., & Sligar, S. G. (2009). Cooperative properties of cytochromes P450. *Pharmacology & Therapeutics*, 124(2), 151–67.
- Dietrich, M., Eiben, S., Asta, C., Do, T. A., Pleiss, J., & Urlacher, V. B. (2008). Cloning, expression and characterisation of CYP102A7, a self-sufficient P450 monooxygenase from *Bacillus licheniformis*. *Applied Microbiology and Biotechnology*, 79(6), 931–40.
- Dietrich, M., Grundmann, L., Kurr, K., Valinotto, L., Saussele, T., Schmid, R. D., & Lange, S. (2005). Recombinant production of human microsomal cytochrome P450 2D6 in the methylotrophic yeast *Pichia pastoris*. *Chembiochem: A European Journal of Chemical Biology*, 6(11), 2014–22.
- Dini, C., & Bianchi, A. (2012). The potential role of vitamin D for prevention and treatment of tuberculosis and infectious diseases. *Annali dell'Istituto Superiore Di Sanità*, 48(3), 319–327.
- Doonan, S., & Cutler, P. (2004). Protein purification protocols. *Methods Mol Biol*, 244, 85–90.
- Dunford, A. J., McLean, K. J., Sabri, M., Seward, H. E., Heyes, D. J., Scrutton, N. S., & Munro, A. W. (2007). Rapid P450 heme iron reduction by laser photoexcitation of *Mycobacterium tuberculosis* CYP121 and CYP51B1. Analysis of CO complexation reactions and reversibility of the P450/P420 equilibrium. *The Journal of Biological Chemistry*, 282(34), 24816–24.
- Durairaj, P., Malla, S., Nadarajan, S. P., Lee, P.-G., Jung, E., Park, H. H., ... Yun, H. (2015). Fungal cytochrome P450 monooxygenases of *Fusarium oxysporum* for the synthesis of ω -hydroxy fatty acids in engineered *Saccharomyces cerevisiae*. *Microbial Cell Factories*, 14(1), 45.
- Dusso, A. S., Sato, T., Arcidiacono, M. V., Alvarez-Hernandez, D., Yang, J., Gonzalez-Suarez, I., ... Slatopolsky, E. (2006). Pathogenic mechanisms for parathyroid hyperplasia. *Kidney International. Supplement*, 70(102), S8–11.

- El-Sharkawy, S. H., Yang, W., Dostal, L., & Rosazza, J. P. (1992). Microbial oxidation of oleic acid. *Applied and Environmental Microbiology*, 58(7), 2116–2122.
- Farinas, E. T., Alcalde, M., & Arnold, F. (2004). Alkene epoxidation catalyzed by cytochrome P450 BM-3 139-3. *Tetrahedron*, 60(3), 525–528.
- Friedberg, T., & Wolf, C. R. (1996). Recombinant DNA technology as an investigative tool in drug metabolism research. *Advanced Drug Delivery Reviews*, 22(1-2), 187–213.
- Fujii, Y., Kabumoto, H., Nishimura, K., Fujii, T., Yanai, S., Takeda, K., ... Tamura, T. (2009). Purification, characterization, and directed evolution study of a vitamin D3 hydroxylase from *Pseudonocardia autotrophica*. *Biochemical and Biophysical Research Communications*, 385(2), 170–5.
- Fulco, A. J. (1991). P450BM-3 and other inducible bacterial P450 cytochromes: biochemistry and regulation. *Annual Review of Pharmacology and Toxicology*, 31, 177–203.
- Furuya, T., & Kino, K. (2010). Genome mining approach for the discovery of novel cytochrome P450 biocatalysts. *Applied Microbiology and Biotechnology*, 86(4), 991–1002.
- Garland, C., Gorham, E., Mohr, S., & Garland, F. (2009). Vitamin D for cancer prevention: global perspective. *Annals of Epidemiology*, 19(7), 468–83.
- Gasteiger, E., Hoogland, C., Gattiker, A., Wilkins, M. R., Appel, R. D., & Bairoch, A. (2005). Protein identification and analysis tools on the ExPASy server. *The Proteomics Protocols Handbook*, 571–607.
- Ghosh, a, Bhardwaj, M., Satyanarayana, T., Khurana, M., Mayilraj, S., & Jain, R. K. (2007). *Bacillus lehensis* sp. nov., an alkalitolerant bacterium isolated from soil. *International Journal of Systematic and Evolutionary Microbiology*, 57(Pt 2), 238–42.
- Gillam, E. M. J. (2008). Engineering Cytochrome P450 Enzymes. *Chemical Research in Toxicology*, 21(1), 220–231.
- Giovannucci, E. (2005). The epidemiology of vitamin D and cancer incidence and mortality: a review. *Cancer Causes & Control : CCC*, 16(2), 83–95.
- Glerup, H., Mikkelsen, K., Poulsen, L., Hass, E., Overbeck, S., Andersen, H., ... Eriksen, E. F. (2000). Hypovitaminosis D myopathy without biochemical signs of osteomalacic bone involvement. *Calcified Tissue International*, 66(6), 419–24.

- Goldstein, D. A., Haldimann, B., Sherman, D., Norman, A. W., & Massry, S. G. (1981). Vitamin D metabolites and calcium metabolism in patients with nephrotic syndrome and normal renal function. *The Journal of Clinical Endocrinology and Metabolism*, 52(1), 116–21.
- Gomaa, M. S., Simons, C., & Brancale, A. (2007). Homology model of 1 α ,25-dihydroxyvitamin D₃ 24-hydroxylase cytochrome P450 24A1 (CYP24A1): active site architecture and ligand binding. *The Journal of Steroid Biochemistry and Molecular Biology*, 104(1-2), 53–60.
- Gonzalez, F. J., & Korzekwa, K. R. (1995). Cytochromes P450 expression systems. *Annual Review of Pharmacology and Toxicology*, 35, 369–90.
- Gotoh, O. (1992). Substrate recognition sites in cytochrome P450 family 2 (CYP2) proteins inferred from comparative analyses of amino acid and coding nucleotide sequences. *The Journal of Biological Chemistry*, 267(1), 83–90.
- Graham, S. E., & Peterson, J. A. (1999). How Similar Are P450s and What Can Their Differences Teach Us? *Archives of Biochemistry and Biophysics*, 369(1), 24–29.
- Graham-Lorence, S. E., & Peterson, J. A. (1996). Cytochrome P450, Part B. *Methods in Enzymology* (Vol. 272). Elsevier. [http://doi.org/10.1016/S0076-6879\(96\)72037-2](http://doi.org/10.1016/S0076-6879(96)72037-2)
- Gräslund, S., Nordlund, P., Weigelt, J., Hallberg, B. M., Bray, J., Gileadi, O., ... Gunsalus, K. C. (2008). Protein production and purification. *Nature Methods*, 5(2), 135–46.
- Green, A. J., Rivers, S. L., Cheesman, M., Reid, G. a., Quaroni, L. G., Macdonald, I. D. G., ... Munro, A. W. (2001). Expression, purification and characterization of cytochrome P450 Biol: a novel P450 involved in biotin synthesis in *Bacillus subtilis*. *JBIC Journal of Biological Inorganic Chemistry*, 6(5-6), 523–533.
- Greenfield, N. (2006). Using circular dichroism spectra to estimate protein secondary structure. *Nature Protocols*, 1(6), 2876–90. <http://doi.org/10.1038/nprot.2006.202>
- Grogan, G. (2011). Cytochromes P450: exploiting diversity and enabling application as biocatalysts. *Current Opinion in Chemical Biology*, 15(2), 241–8.
- Guengerich, F. P., Martin, M. V, Sohl, C. D., & Cheng, Q. (2009). Measurement of cytochrome P450 and NADPH-cytochrome P450 reductase. *Nature Protocols*, 4(9), 1245–51.

- Guo, Y. D., Strugnell, S., Back, D. W., & Jones, G. (1993). Transfected human liver cytochrome P-450 hydroxylates vitamin D analogs at different side-chain positions. *Proceedings of the National Academy of Sciences of the United States of America*, 90(18), 8668–72.
- Gupta, R. P., He, Y. A., Patrick, K. S., Halpert, J. R., & Bell, N. H. (2005). CYP3A4 is a vitamin D-24- and 25-hydroxylase: analysis of structure function by site-directed mutagenesis. *The Journal of Clinical Endocrinology and Metabolism*, 90(2), 1210–9.
- Gupta, R. P., Hollis, B. W., Patel, S. B., Patrick, K. S., & Bell, N. H. (2004). CYP3A4 is a human microsomal vitamin D 25-hydroxylase. *Journal of Bone and Mineral Research: The Official Journal of the American Society for Bone and Mineral Research*, 19(4), 680–8.
- Guyton, K. Z., Kensler, T. W., & Posner, G. H. (2003). Vitamin D and vitamin D analogs as cancer chemopreventive agents. *Nutrition Reviews*, 61(7), 227–238.
- Haines, D. C., Tomchick, D. R., Machius, M., & Peterson, J. A. (2001). Pivotal Role of Water in the Mechanism of P450BM-3 †. *Biochemistry*, 40(45), 13456–13465.
- Hasemann, C. A., Ravichandran, K. G., Peterson, J. A., & Deisenhofer, J. (1994). Crystal structure and refinement of cytochrome P450terp at 2.3 Å resolution. *Journal of Molecular Biology*, 236(4), 1169–1185.
- Hannemann, F., Bichet, A., Ewen, K. M., & Bernhardt, R. (2007). Cytochrome P450 systems—biological variations of electron transport chains. *Biochimica et Biophysica Acta (BBA) - General Subjects*, 1770(3), 330–344.
- Harding, S. E., & Chowdhry, B. Z. (2001). *Protein-ligand Interactions, Structure and Spectroscopy: A Practical Approach*. (B. Z. C. Stephen E. Harding, Ed.). Oxford University Press.
- Hashizume, T., Imaoka, S., Mise, M., Terauchi, Y., Fujii, T., Miyazaki, H., ... Funae, Y. (2002). Involvement of CYP2J2 and CYP4F12 in the metabolism of ebastine in human intestinal microsomes. *The Journal of Pharmacology and Experimental Therapeutics*, 300(1), 298–304.
- Hawkes, D. B., Adams, G. W., Burlingame, A. L., Montellano, P. R. O. de., & Voss, J. J. De. (2002). Cytochrome P450cin (CYP176A), Isolation, Expression, and Characterization. *Journal of Biological Chemistry*, 277(31), 27725–27732.
- Hayashi, S., Noshiro, M., & Okuda, K. (1986). Isolation of a cytochrome P-450 that catalyzes the 25-hydroxylation of vitamin D3 from rat liver microsomes. *Journal of Biochemistry*, 99(6), 1753–1763.

- Heaney, R. P., Armas, L. A. G., Shary, J. R., Bell, N. H., Binkley, N., & Hollis, B. W. (2008). 25-Hydroxylation of vitamin D3: relation to circulating vitamin D3 under various input conditions. *The American Journal of Clinical Nutrition*, 87(6), 1738–42.
- Hilker, B. L., Fukushige, H., Hou, C., & Hildebrand, D. (2008). Comparison of *Bacillus* monooxygenase genes for unique fatty acid production. *Progress in Lipid Research*, 47(1), 1–14.
- Holick, M. F. (2004). Vitamin D: importance in the prevention of cancers, type 1 diabetes, heart disease, and osteoporosis. *The American Journal of Clinical Nutrition*, 79(3), 362–371.
- Holick, M. F. (2007). Vitamin D Deficiency. *New England Journal of Medicine*, 266–281.
- Holick, M. F. (2009). Vitamin D status: measurement, interpretation, and clinical application. *Annals of Epidemiology*, 19(2), 73–8.
- Horikoshi, K. (1971). Production part of alkaline enzymes protease by alkalophilic produced microorganisms by *Bacillus* No . By Koki H0RIKOSHI The Institute of Physical and Chemical The dialyzate. *Agricultural and Biological Chemistry*, 35(9), 1407–1414.
- Horikoshi, K. (1999). Alkaliphiles: some applications of their products for biotechnology. *Microbiology and Molecular Biology Reviews*: MMBR, 63(4), 735–750, table of contents.
- Horikoshi, S., McCune, B. K., Ray, P. E., Kopp, J. B., Sporn, M. B., & Klotman, P. E. (1991). Water deprivation stimulates transforming growth factor-beta 2 accumulation in the juxtglomerular apparatus of mouse kidney. *The Journal of Clinical Investigation*, 88(6), 2117–22.
- Hosseinpour, F., & Wikvall, K. (2000). Porcine microsomal vitamin D(3) 25-hydroxylase (CYP2D25). Catalytic properties, tissue distribution, and comparison with human CYP2D6. *The Journal of Biological Chemistry*, 275(44), 34650–5.
- Jin, S., Bryson, T., & Dawson, J. (2004). Hydroperoxoferric heme intermediate as a second electrophilic oxidant in cytochrome P450-catalyzed reactions. *JBIC Journal of Biological Inorganic Chemistry*, 9(6), 644–653.
- Jion, A., Goh, L., & Oh, S. (2006). Crystallization of IgG1 by mapping its liquid–liquid phase separation curves. *Biotechnology and Bioengineering*.
- Johnston, J. B., Ouellet, H., Podust, L. M., & Ortiz De Montellano, P. R. (2011). Structural control of cytochrome P450-catalyzed ω -hydroxylation. *Archives of Biochemistry and Biophysics*, 507(1), 86–94.

- Jones, G., Strugnell, S. A., & Deluca, H. F. (1998). Current understanding of the molecular actions of vitamin D. *Physiological Review*, 78(4), 1193–1231.
- Joshi, D., Center, J., & Eisman, J. (2010). Vitamin D deficiency in adults. *Australian Prescriber*, 2–7.
- Julsing, M. K., Cornelissen, S., Bühler, B., & Schmid, A. (2008). Heme-iron oxygenases: powerful industrial biocatalysts? *Current Opinion in Chemical Biology*, 12(2), 177–86.
- Jung, S. T., Lauchli, R., & Arnold, F. H. (2011). Cytochrome P450: taming a wild type enzyme. *Current Opinion in Biotechnology*, 22(6), 809–17.
- Katsumata, T., Hasegawa, A., Fujiwara, T., Komatsu, T., Notomi, M., Abe, H., ... Kawaide, H. (2008). Arabidopsis CYP85A2 catalyzes lactonization reactions in the biosynthesis of 2-deoxy-7-oxalactone brassinosteroids. *Bioscience, Biotechnology, and Biochemistry*, 72(8), 2110–7.
- Kawauchi, H., Sasaki, J., Adachi, T., Hanada, K., Beppu, T., & Horinouchi, S. (1994). Cloning and nucleotide sequence of a bacterial cytochrome P-450VD25 gene encoding vitamin D-3 25-hydroxylase. *Biochimica et Biophysica Acta (BBA) - Gene Structure and Expression*, 1219(1), 179–183.
- Kelly, S., & Price, N. (2000). The Use of Circular Dichroism in the Investigation of Protein Structure and Function. *Current Protein and Peptide Science*, 1(4), 349–384.
- Kennel, K. a., Drake, M. T., & Hurley, D. L. (2010). Vitamin D deficiency in adults: when to test and how to treat. *Mayo Clinic Proceedings*, 85(8), 752–758.
- Khan, Q. J., & Fabian, C. J. (2010). How I treat vitamin D deficiency. *Journal of Oncology Practice*, 6(2), 97–101.
- Konagurthu, A. S., Whisstock, J. C., Stuckey, P. J., & Lesk, A. M. (2006). MUSTANG: a multiple structural alignment algorithm. *Proteins*, 64(3), 559–74.
- Krauss, I. R., Merlino, A., Vergara, A., & Sica, F. (2013). An overview of biological macromolecule crystallization. *International Journal of Molecular Sciences*, 14(6), 11643–11691.
- Krieger, E., Koraimann, G., & Vriend, G. (2002). Increasing the precision of comparative models with YASARA NOVA--a self-parameterizing force field. *Proteins*, 47(3), 393–402.
- Ku, T., Lu, P., Chan, C., Wang, T., Lai, S., Lyu, P., & Hsiao, N. (2009). Predicting melting temperature directly from protein sequences. *Computational Biology and Chemistry*, 33(6), 445–50.

- Kumar, C. G., & Takagi, H. (1999). Microbial alkaline proteases: From a bioindustrial viewpoint. *Biotechnology Advances*, 17(7), 561–594.
- Kumar, S. (2010). Engineering cytochrome P450 biocatalysts for biotechnology, medicine and bioremediation. *Expert Opinion on Drug Metabolism & Toxicology*, 6(2), 115–31.
- Kumar, V., Sharma, V. K., & Kalonia, D. S. (2009). Effect of polyols on polyethylene glycol (PEG)-induced precipitation of proteins: Impact on solubility, stability and conformation. *International Journal of Pharmaceutics*, 366(1-2), 38–43.
- Laemmli, U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 227(5259), 680–685.
- Laskowski, R. A., Rullmann, J. A., MacArthur, M. W., Kaptein, R., & Thornton, J. M. (1996). AQUA and PROCHECK-NMR: programs for checking the quality of protein structures solved by NMR. *Journal of Biomolecular NMR*, 8(4), 477–86.
- Lavie, C. J., Lee, J. H., & Milani, R. V. (2011). Vitamin D and cardiovascular disease will it live up to its hype? *Journal of the American College of Cardiology*, 58(15), 1547–56.
- Lee, D., Yamada, A., Sugimoto, H., Matsunaga, I., Ogura, H., Ichihara, K., ... Shiro, Y. (2003). Substrate recognition and molecular mechanism of fatty acid hydroxylation by cytochrome P450 from *Bacillus subtilis*. *Biochemistry*, 278(11), 9761–9767.
- Leow, T. C., Rahman, R. N. Z. R. A., Salleh, A. B., & Basri, M. (2007). High-temperature crystallization of thermostable T1 lipase. *Crystal Growth & Design*, 7(2), 406–410.
- Lewis, D. F. V., & Wiseman, A. (2005). A selective review of bacterial forms of cytochrome P450 enzymes. *Enzyme and Microbial Technology*, 36(4), 377–384.
- Leys, D., Mowat, C. G., McLean, K. J., Richmond, A., Chapman, S. K., Walkinshaw, M. D., & Munro, A. W. (2003). Atomic Structure of Mycobacterium tuberculosis CYP121 to 1.06 Å Reveals Novel Features of Cytochrome P450. *Journal of Biological Chemistry*, 278(7), 5141–5147.
- Li, H., & Poulos, T. L. (1997). The structure of the cytochrome p450BM-3 haem domain complexed with the fatty acid substrate, palmitoleic acid. *Nature Structural & Molecular Biology*, 4(2), 140–146.

- Lombard, M., Salard, I., Sari, M.-A., Mansuy, D., & Buisson, D. (2011). A new cytochrome P450 belonging to the 107L subfamily is responsible for the efficient hydroxylation of the drug terfenadine by *Streptomyces platensis*. *Archives of Biochemistry and Biophysics*, 508(1), 54–63.
- Looker, A. C., Johnson, C. L., Lacher, D. A., Pfeiffer, C. M., Schleicher, R. L., & Sempos, C. T. (2011). Vitamin D status: United States, 2001-2006. *NCHS Data Brief*, (59), 1–8.
- Luft, J., Wolfley, J., & Said, M. (2007). Efficient optimization of crystallization conditions by manipulation of drop volume ratio and temperature. *Protein Science*, (716), 715–722.
- Luscombe, N. M., Greenbaum, D., & Gerstein, M. (2001). What is Bioinformatics? A proposed definition and overview of the field. *Methods Archive*, 40(4), 346–358.
- Marchler-Bauer, A., Anderson, J. B., Chitsaz, F., Derbyshire, M. K., Deweese-Scott, C., Fong, J. H., ... Bryant, S. H. (2009). CDD: Specific functional annotation with the Conserved Domain Database. *Nucleic Acids Research*, 37(SUPPL. 1), 205–210.
- Masuda, S., & Jones, G. (2006). Promise of vitamin D analogues in the treatment of hyperproliferative conditions. *Molecular Cancer Therapeutics*, 5(4), 797–808.
- Mattson, R. H., & Gidal, B. E. (2004). Fractures, epilepsy, and antiepileptic drugs. *Epilepsy & Behavior: E&B*, 5 Suppl 2, S36–40.
- McLean, K. J., Girvan, H. M., Mason, A. E., Dunford, A. J., & Munro, A. W. (2011). *Iron-Containing Enzymes*. (S. P. de Visser & D. Kumar, Eds.). Cambridge: Royal Society of Chemistry.
- McLean, K. J., Sabri, M., Marshall, K. R., Lawson, R. J., Lewis, D. G., Clift, D., ... Munro, a. W. (2005). Biodiversity of cytochrome P450 redox systems. *Biochemical Society Transactions*, 33(4), 796.
- McLean, M., Maves, S., & Weiss, K. (1998). Characterization of a Cytochrome P450 from the Acidothermophilic Archaea *Sulfolobus solfataricus*. *Biochemical and Biophysical Research Communications*, 172(252), 166–172.
- McPherson, A. (2004). Introduction to protein crystallization. *Methods*, 34(3), 254–265.
- McPherson, A., & Cudney, B. (2014). Optimization of crystallization conditions for biological macromolecules. *Acta Crystallographica. Section F, Structural Biology Communications*, 70(Pt 11), 1445–67.

- Metzger, J. O., & Bornscheuer, U. (2006). Lipids as renewable resources: current state of chemical and biotechnological conversion and diversification. *Applied Microbiology and Biotechnology*, 71(1), 13–22.
- Meunier, B., de Visser, S. P., & Shaik, S. (2004). Mechanism of oxidation reactions catalyzed by cytochrome P450 enzymes. *Chemical Reviews*, 104(9), 3947–3980.
- Miles, C. S., Ost, T. W. ., Noble, M. A., Munro, A. W., & Chapman, S. K. (2000). Protein engineering of cytochromes P-450. *Biochimica et Biophysica Acta (BBA) - Protein Structure and Molecular Enzymology*, 1543(2), 383–407.
- Miles, J. S., & Wolf, C. R. (1989). Principles of DNA cloning. *BMJ: British Medical Journal*, 299(October), 1019–1022.
- Morris, G. M., Goodsell, D. S., Halliday, R. S., Huey, R., Hart, W. E., Belew, R. K., & Olson, A. J. (1998). Automated docking using a Lamarckian genetic algorithm and an empirical binding free energy function. *Journal of Computational Chemistry*, 19(14), 1639–1662.
- Mulchandani, P., Hangarter, C. M., Lei, Y., Chen, W., & Mulchandani, A. (2005). Amperometric microbial biosensor for p-nitrophenol using *Moraxella sp.*-modified carbon paste electrode. *Biosensors & Bioelectronics*, 21(3), 523–7.
- Munro, A. W., Girvan, H. M., & McLean, K. J. (2007). Variations on a (t)heme--novel mechanisms, redox partners and catalytic functions in the cytochrome P450 superfamily. *Natural Product Reports*, 24(3), 585–609.
- Nair, S. (2010). Vitamin d deficiency and liver disease. *Gastroenterology & Hepatology*, 6(8), 491–3.
- Nebert, D. W., & Russell, D. W. (2002). Clinical importance of the cytochromes P450. *Lancet (London, England)*, 360(9340), 1155–62.
- Nelson, D. R. (2006). Cytochrome P450 nomenclature, 2004. *Methods in Molecular Biology (Clifton, N.J.)*, 320, 1–10.
- Nelson, D. R. (2009, October 1). The Cytochrome P450 Homepage. *Human Genomics*. BioMed Central.
- Nelson, D. R., Zeldin, D. C., Hoffman, S. M. G., Maltais, L. J., Wain, H. M., & Nebert, D. W. (2004). Comparison of cytochrome P450 (CYP) genes from the mouse and human genomes, including nomenclature recommendations for genes, pseudogenes and alternative-splice variants. *Pharmacogenetics*, 14(1), 1–18.

- Noor, Y. M., Samsulrizal, N. H., Jema'on, N. A., Low, K. O., Ramli, A. N. M., Alias, N. I., ... Illias, R. M. (2014). A comparative genomic analysis of the alkalitolerant soil bacterium *Bacillus lehensis* G1. *Gene*, 545(2), 253–61.
- Norman, A. W. (1998). Sunlight, season, skin pigmentation, vitamin D, and 25-hydroxyvitamin D: integral components of the vitamin D endocrine system. *The American Journal of Clinical Nutrition*, 67(6), 1108–10.
- Oganesyan, N., Ankoudinova, I., Kim, S.-H., & Kim, R. (2007). Effect of osmotic stress and heat shock in recombinant protein overexpression and crystallization. *Protein Expression and Purification*, 52(2), 280–285.
- Ohyama, Y., & Okuda, K. (1991). Isolation and characterization of a cytochrome P-450 from rat kidney mitochondria that catalyzes the 24-hydroxylation of 25-hydroxyvitamin D₃. *The Journal of Biological Chemistry*, 266(14), 8690–5.
- O'Keefe, D. P., Romesser, J. A., & Leto, K. J. (1988). Identification of constitutive and herbicide inducible cytochromes P-450 in *Streptomyces griseolus*. *Archives of Microbiology*, 149(5), 406–412.
- Ong, R. M., Goh, K. M., Mahadi, N. M., Hassan, O., Rahman, R. N. Z. R. A., & Illias, R. M. (2008). Cloning, extracellular expression and characterization of a predominant beta-CGTase from *Bacillus* sp. G1 in *E. coli*. *Journal of Industrial Microbiology & Biotechnology*, 35(12), 1705–14.
- Pack, A. M., & Morrell, M. J. (2004). Epilepsy and bone health in adults. *Epilepsy & Behavior: E&B*, 5 Suppl 2, S24–9.
- Peterson, J. A., & Graham, S. E. (1998). A close family resemblance: the importance of structure in understanding cytochromes P450. *Structure*, 6(9), 1079–1085.
- Pitt, J. J. (2009). Principles and applications of liquid chromatography-mass spectrometry in clinical biochemistry. *The Clinical Biochemist Reviews*, 30(February), 19–34.
- Plotnikoff, G. A., & Quigley, J. M. (2003). Prevalence of severe hypovitaminosis D in patients with persistent, nonspecific musculoskeletal pain. *Mayo Clinic Proceedings*, 78(12), 1463–70.
- Podust, L. M., Kim, Y., Arase, M., Neely, B. A., Beck, B. J., Bach, H., ... Waterman, M. R. (2003). The 1.92-Å Structure of *Streptomyces coelicolor* A3(2) CYP154C1: A new monooxygenase that functionalized macrolide ring systems. *Journal of Biological Chemistry*, 278(14), 12214–12221.

- Podust, L. M., & Sherman, D. H. (2012). Diversity of P450 enzymes in the biosynthesis of natural products. *Natural Product Reports*, 29(10), 1251–66.
- Poulos, T. L., Finzel, B. C., & Howard, A. J. (1986). Crystal structure of substrate-free *Pseudomonas putida* cytochrome P-450. *Biochemistry*, 25(18), 5314–5322.
- Prescott, L.M.; Harley, J.P.; Klein, D. A. (2005). *Microbiology (Six Editio)*. New York: McGraw Hill International.
- Prior, J. E., Shokati, T., Christians, U., & Gill, R. T. (2010). Identification and characterization of a bacterial cytochrome P450 for the metabolism of diclofenac. *Applied Microbiology and Biotechnology*, 85(3), 625–33.
- Raghunathan, K., Harris, P. T., & Arvidson, D. N. (2010). Trial by fire: Are the crystals macromolecules? *Acta Crystallographica Section F: Structural Biology and Crystallization Communications*, 66, 615–620.
- Rahmaniyan, M., Patrick, K., & Bell, N. H. (2005). Characterization of recombinant CYP2C11: a vitamin D 25-hydroxylase and 24-hydroxylase. *American Journal of Physiology. Endocrinology and Metabolism*, 288(4), E753–60.
- Raussens, V., Ruyschaert, J.-M., & Goormaghtigh, E. (2003). Protein concentration is not an absolute prerequisite for the determination of secondary structure from circular dichroism spectra: a new scaling method. *Analytical Biochemistry*, 319(1), 114–121.
- Ravichandran, K. G., Boddupalli, S. S., Hasermann, C. A., Peterson, J. A., & Deisenhofer, J. (1993). Crystal structure of hemoprotein domain of P450BM-3, a prototype for microsomal P450's. *Science (New York, N.Y.)*, 261(5122), 731–6.
- Reddick, J. J., Antolak, S. a, & Raner, G. M. (2007). PksS from *Bacillus subtilis* is a cytochrome P450 involved in bacillaene metabolism. *Biochemical and Biophysical Research Communications*, 358(1), 363–7.
- Roberts, G. A., Çelik, A., Hunter, D. J. B., Ost, T. W. B., White, J. H., Chapman, S. K., ... Flitsch, S. L. (2003). A self-sufficient cytochrome P450 with a primary structural organization that includes a flavin domain and a [2Fe-2S] redox center. *Journal of Biological Chemistry*, 278 (49), 48914–48920.
- Sakaki, T. (2012). Practical application of cytochrome P450. *Biological & Pharmaceutical Bulletin*, 35(6), 844–9.
- Sakaki, T., Sugimoto, H., Hayashi, K., Yasuda, K., Munetsuna, E., Kamakura, M., ... Shiro, Y. (2011). Bioconversion of vitamin D to its active form by bacterial or mammalian cytochrome P450. *Biochimica et Biophysica Acta*, 1814(1), 249–56.

- Salazar, O., Cirino, P. C., & Arnold, F. H. (2003). Thermostabilization of a cytochrome p450 peroxygenase. *Chembiochem : A European Journal of Chemical Biology*, 4(9), 891–3.
- Sarethy, I. P., Saxena, Y., Kapoor, A., Sharma, M., Sharma, S. K., Gupta, V., & Gupta, S. (2011). Alkaliphilic bacteria: applications in industrial biotechnology. *Journal of Industrial Microbiology & Biotechnology*, 38(7), 769–790.
- Sasaki, J., & Mikami, A. (1991). Transformation of 25-and 1 alpha-hydroxyvitamin D₃ to 1 alpha, 25-dihydroxyvitamin D₃ by using *Streptomyces* sp. strains. *Applied and Environmental Microbiology*, 57(10), 2841–2846.
- Sasaki, J., Miyazaki, A., Saito, M., Adachi, T., Mizoue, K., Hanada, K., & Omura, S. (1992). Transformation of vitamin D₃ to 1 α ,25-dihydroxyvitamin D₃ via 25-hydroxyvitamin D₃ using *Amycolata* sp. strains. *Applied Microbiology and Biotechnology*, 38(2), 152–157.
- Sasaki, M., Akahira, A., Oshiman, K., Tsuchido, T., & Matsumura, Y. (2005). Purification of cytochrome P450 and ferredoxin, involved in bisphenol A degradation, from *Sphingomonas* sp. strain AO1. *Applied and Environmental Microbiology*, 71(12), 8024–30.
- Sawada, N., Sakaki, T., Ohta, M., & Inouye, K. (2000). Metabolism of Vitamin D₃ by Human CYP27A1. *Biochemical and Biophysical Research Communications*, 273(3), 977–984.
- Sawada, N., Sakaki, T., Yoneda, S., Kusudo, T., Shinkyo, R., Ohta, M., & Inouye, K. (2004). Conversion of vitamin D₃ to 1 α ,25-dihydroxyvitamin D₃ by *Streptomyces griseolus* cytochrome P450SU-1. *Biochemical and Biophysical Research Communications*, 320(1), 156–164.
- Schaller, A., & Stintzi, A. (2009). Enzymes in jasmonate biosynthesis - structure, function, regulation. *Phytochemistry*, 70(13-14), 1532–8.
- Schallmeyer, A., den Besten, G., Teune, I. G. P., Kembaren, R. F., & Janssen, D. B. (2011). Characterization of cytochrome P450 monooxygenase CYP154H1 from the thermophilic soil bacterium *Thermobifida fusca*. *Applied Microbiology and Biotechnology*, 89(5), 1475–85.
- Schlichting, I., Berendzen, J., Chu, K., Stock, A. M., Maves, S. A., Benson, D. E., ... Sligar, S. G. (2000). The catalytic pathway of cytochrome p450cam at atomic resolution. *Science (New York, N.Y.)*, 287(5458), 1615–22.
- Schuster, I. (2011). Cytochromes P450 are essential players in the vitamin D signaling system. *Biochimica et Biophysica Acta*, 1814(1), 186–99.
- Sherman, D. H. (2006). The structural basis for substrate anchoring, active site selectivity, and product formation by P450 PikC from *Streptomyces*

- venezuelae*. The Journal of Biological Chemistry, 281(36), 26289–26297.
- Shinkyo, R., Sakaki, T., Kamakura, M., Ohta, M., & Inouye, K. (2004). Metabolism of vitamin D by human microsomal CYP2R1. *Biochemical and Biophysical Research Communications*, 324(1), 451–457.
- Sigman, J. A., Pond, A. E., Dawson, J. H., & Lu, Y. (1999). Engineering cytochrome c peroxidase into cytochrome P450: a proximal effect on heme–thiolate ligation. *Biochemistry*, 38(34), 11122–11129.
- Sintupachee, S., Ngamrojanavanich, N., Sitthithaworn, W., & De-Eknamkul, W. (2014). Molecular cloning, bacterial expression and functional characterisation of cytochrome P450 monooxygenase, CYP97C27, and NADPH-cytochrome P450 reductase, CPR I, from *Croton stellatopilosus* Ohba. *Plant Science: An International Journal of Experimental Plant Biology*, 229, 131–41.
- Sirim, D., Widmann, M., Wagner, F., & Pleiss, J. (2010). Prediction and analysis of the modular structure of cytochrome P450 monooxygenases. *BMC Structural Biology*, 10(1), 34.
- Sono, M., Roach, M. P., Coulter, E. D., & Dawson, J. H. (1996). Heme-containing oxygenases. *Chemical Reviews*, 96(7), 2841–2888.
- Strushkevich, N., Usanov, S. a, Plotnikov, A. N., Jones, G., & Park, H.-W. (2008). Structural analysis of CYP2R1 in complex with vitamin D3. *Journal of Molecular Biology*, 380(1), 95–106.
- Sugimoto, H., Shinkyo, R., Hayashi, K., Yoneda, S., Yamada, M., Kamakura, M., ... Sakaki, T. (2008). Crystal structure of CYP105A1 (P450SU-1) in complex with 1 α ,25-dihydroxyvitamin D3. *Biochemistry*, 47(13), 4017–27.
- Sulistyaningdyah, W. T., Ogawa, J., Li, Q.-S., Maeda, C., Yano, Y., Schmid, R. D., & Shimizu, S. (2005). Hydroxylation activity of P450 BM-3 mutant F87V towards aromatic compounds and its application to the synthesis of hydroquinone derivatives from phenolic compounds. *Applied Microbiology and Biotechnology*, 67(4), 556–62.
- Sweet, C. R. (2003). Expression of recombinant proteins from lac promoters. *Methods in Molecular Biology (Clifton, N.J.)*, 235, 277–88.
- Syed, K., Shale, K., Nazir, K. H. M. N. H., Krasevec, N., Mashele, S. S., & Pagadala, N. S. (2014). Genome-wide identification, annotation and characterization of novel thermostable cytochrome P450 monooxygenases from the thermophilic biomass-degrading fungi *Thielavia terrestris* and *Myceliophthora thermophila*. *Genes & Genomics*, 36(3), 321–333.

- Takami, H., & Horikoshi, K. (2000). Analysis of the genome of an alkaliphilic *Bacillus* strain from an industrial point of view. *Extremophiles: Life under Extreme Conditions*, 4(2), 99–108.
- Takeda, K., Asou, T., Matsuda, A., Kimura, K., Okamura, K., Okamoto, R., ... Omura, S. (1994). Application of cyclodextrin to microbial transformation of vitamin D3 to 25-hydroxyvitamin D3 and 1 α ,25-dihydroxyvitamin D3. *Journal of Fermentation and Bioengineering*, 78(5), 380–382.
- Thompson, J. D., Higgins, D. G., & Gibson, T. J. (1994). CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research*, 22(22), 4673–80.
- Timur, S., Anik, U., Odaci, D., & Gorton, L. (2007). Development of a microbial biosensor based on carbon nanotube (CNT) modified electrodes. *Electrochemistry Communications*, 9(7), 1810–1815.
- Tsuneo Omura; Ryo Sato. (1964). The Carbon of Liver Microsomes. *The Journal of Biological Chemistry*, 239(7), 2370–2378.
- Urlacher, V. B., & Eiben, S. (2006). Cytochrome P450 monooxygenases: perspectives for synthetic application. *Trends in Biotechnology*, 24(7), 324–30.
- Urlacher, V. B., & Girhard, M. (2012). Cytochrome P450 monooxygenases: an update on perspectives for synthetic application. *Trends in Biotechnology*, 30(1), 26–36.
- Van Bogaert, I. N. A., Groeneboer, S., Saerens, K., & Soetaert, W. (2011). The role of cytochrome P450 monooxygenases in microbial fatty acid metabolism. *FEBS Journal*, 278(2), 206–221.
- Vera, A., González-Montalbán, N., Arís, A., & Villaverde, A. (2007). The conformational quality of insoluble recombinant proteins is enhanced at low growth temperatures. *Biotechnology and Bioengineering*, 96(6), 1101–6.
- Vinod, K. K. K. (2004). Total genomic DNA extraction, quality check and quantitation. *Proceedings of the Training Programme on Classical and Modern Plant Breeding Techniques—a Hands on Training*, 109–121.
- Voet, D., & Voet, J. G. (2004). *Biochemistry (Third Edit)*. John Wiley & Sons.
- Weber, J. M., Leung, J. O., Swanson, S. J., Idler, K. B., & McAlpine, J. B. (1991). An erythromycin derivative produced by targeted gene disruption in *Saccharopolyspora erythraea*. *Science*, 252(5002), 114–7.
- Werck-Reichhart, D., & Feyereisen, R. (2000). Cytochromes P450: a success story. *Genome Biology*, 1(6), REVIEWS3003.

- Wiederstein, M., & Sippl, M. J. (2007). ProSA-web: interactive web service for the recognition of errors in three-dimensional structures of proteins. *Nucleic Acids Research*, 35(Web Server issue), W407–10.
- Wolf, R., & Friedberg, T. (1996). Recombinant DNA technology as an investigative metabolism research. *Advanced Drug Delivery Reviews*, (96), 187–213.
- Wolpowitz, D., & Gilchrist, B. A. (2006). The vitamin D questions: how much do you need and how should you get it? *Journal of the American Academy of Dermatology*, 54(2), 301–17.
- Wortsman, J., Matsuoka, L. Y., Chen, T. C., Lu, Z., & Holick, M. F. (2000). Decreased bioavailability of vitamin D in obesity. *The American Journal of Clinical Nutrition*, 72(3), 690–3.
- Yamasaki, T., Izumi, S., Ide, H., & Ohyama, Y. (2004). Identification of a novel rat microsomal vitamin D3 25-hydroxylase. *The Journal of Biological Chemistry*, 279(22), 22848–56.
- Yano, J. K., Blasco, F., Li, H., Schmid, R. D., Henne, A., & Poulos, T. L. (2003). Preliminary characterization and crystal structure of a thermostable cytochrome P450 from *Thermus thermophilus*. *The Journal of Biological Chemistry*, 278(1), 608–616.
- Yasutake, Y., Fujii, Y., Cheon, W. K., Arisawa, A., & Tamura, T. (2009). Crystallization and preliminary X-ray diffraction studies of vitamin D3 hydroxylase, a novel cytochrome P450 isolated from *Pseudonocardia autotrophica*. *Acta Crystallographica. Section F, Structural Biology and Crystallization Communications*, 65(Pt 4), 372–5.
- Yasutake, Y., Fujii, Y., Nishioka, T., Cheon, W.-K., Arisawa, A., & Tamura, T. (2010). Structural evidence for enhancement of sequential vitamin D3 hydroxylation activities by directed evolution of cytochrome P450 vitamin D3 hydroxylase. *The Journal of Biological Chemistry*, 285(41), 31193–201.
- Yun, C.-H., Kim, K.-H., Kim, D.-H., Jung, H.-C., & Pan, J.-G. (2007). The bacterial P450 BM3: a prototype for a biocatalyst with human P450 activities. *Trends in Biotechnology*, 25(7), 289–98.
- Zdobnov, E. M., & Apweiler, R. (2001). InterProScan – an integration platform for the signature-recognition methods in InterPro. *Bioinformatics*, 17(9), 847–848.
- Zerbe, K., Pylypenko, O., Vitali, F., Zhang, W., Rousset, S., Heck, M., ... Schlichting, I. (2002). Crystal structure of OxyB, a cytochrome P450 implicated in an oxidative phenol coupling reaction during vancomycin biosynthesis. *Journal of Biological Chemistry*, 277(49), 47476–47485.

- Zhang, R., & Naughton, D. P. (2010). Vitamin D in health and disease: current perspectives. *Nutrition Journal*, 9, 65.
- Zhu, G.-D., & Okamura, W. H. (1995). Synthesis of Vitamin D (Calciferol). *Chemical Reviews*, 95(6), 1877–1952.
- Zhu, J., & Deluca, H. F. (2012). Vitamin D 25-hydroxylase - Four decades of searching, are we there yet? *Archives of Biochemistry and Biophysics*, 523(1), 30–36.
- Zurek, J., Foloppe, N., Harvey, J. N., & Mulholland, A. J. (2006). Mechanisms of reaction in cytochrome P450: Hydroxylation of camphor in P450cam. *Organic and Biomolecular Chemistry*, 4(21), 3931–7.



© COPYRIGHT