



UNIVERSITI PUTRA MALAYSIA

***ELUCIDATION OF THE ANTI-INFLAMMATORY COMPOUND PRESENT
IN *Jatropha curcas* LINN. ROOT AND ITS MODE OF ACTION***

AHMAD RAZI OTHMAN

IB 2016 6



**ELUCIDATION OF THE ANTI-INFLAMMATORY COMPOUND PRESENT IN
Jatropha curcas LINN. ROOT AND ITS MODE OF ACTION**

AHMAD RAZI OTHMAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

June 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfillment of the requirements for the Degree of Doctor of Philosophy

**ELUCIDATION OF THE ANTI-INFLAMMATORY COMPOUND PRESENT IN
Jatropha curcas LINN. ROOT AND ITS MODE OF ACTION**

By

AHMAD RAZI BIN OTHMAN

June 2016

Chairman : Professor Norhani Abdullah, PhD
Institute : Bioscience

Jatropha curcas Linn. (family *Euphorbiaceae*) is a drought resistant shrub which is widely grown in Central and South America, South-east Asia, India and Africa. The plant has been considered a traditional herb in many parts of the world. In inflammatory treatment, it has been widely accepted that non-steroidal anti-inflammatory drugs (NSAIDs) can effectively prevent inflammation. However, several studies have also revealed side effects resulting from prolonged use of NSAIDs, which include the possibility of several chronic diseases such as gastrointestinal ulcers, adverse cardiovascular side-effects, and Alzheimer's disease. Hence, alternative medicine based on natural herbs should be considered in inflammation treatment. Furthermore, the use of herbal remedies is gaining acceptance in various pharmaceutical applications. Although several studies have shown different parts of *J. curcas* possessed anti-inflammatory activity, but the nature of the compounds involved and the mode of action are not well understood. Hence, before the herbal products can be made available, detail information regarding the nature of bioactive compounds and the mode of action have to be understood. Thus, the main objective of this study was to elucidate the anti-inflammatory compounds from *J. curcas* plant and to determine the mode of action. The experiments conducted include screening of different parts of plant extracts for the anti-inflammatory activity by using Murine monocytic macrophage RAW 264.7 cells line, purifying and elucidating the structure of the anti-inflammatory compounds, determining the mode of action of the purified compounds on the inflammation pathway and the inflammatory enzymes affected. Anti-inflammatory activity was assayed by determining the inhibition of nitric oxide production in the RAW 264.7 cells, while cytotoxicity activity was determined by the (3-(4, 5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) MTT assay. In the initial study, it was observed that *Jatropha* root methanolic extract showed anti-inflammatory property higher than other plant parts (leaves, fruits and stem bark), but with high cytotoxicity towards Murine monocytic macrophage RAW 264.7 cells line. Subsequently, the root extract was fractionated into fractions with different solvents (hexane, chloroform and ethyl acetate). The hexane fraction possessed high anti-inflammatory property, but with high cytotoxicity towards cell growth. Analysis of the compounds present in the hexane fraction by gas chromatography mass spectrometry (GC-MS) showed the presence of many compounds belonging to the

terpene group which probably caused the cytotoxicity. Further purification was conducted by using an open column system to isolate and purify the compounds with anti-inflammatory activity without cytotoxicity. Five spots (labeled H-1, H-2,3, H-4 and H-5) from the hexane fraction were obtained. The anti-inflammatory assay showed the compounds present in spot H-4 and H-5 possessed high anti-inflammatory without cytotoxicity activity. Analysis of compounds present in these two spots by GC-MS showed the presence of hexadecanoic and octadecanoic acid groups in both spots. The high performance liquid chromatography (HPLC) analysis of spot H-4 showed two peaks (A and B), with eluent A (peak A) displaying better anti-inflammatory activity than eluent B (peak B), without being toxic to the cell growth. Identification of the compound present in eluent A by liquid chromatography-tandem mass spectrometry (LC-MS/MS) indicated that the compound belonged to the octadecanoic acid group. Further analysis conducted by using NuclearMagnetic Resonance (NMR) showed that this active compound was a long chain of hydrocarbons with a carboxylic group attached at the end, thus confirming that the active compound belonged to octadecanoic acid group. To determine the mode of action in the anti-inflammatory activity, the fluorescence staining assay was conducted to observe the translocation of the NF- κ B subunit (involved in inflammatory signaling pathway) from the cytoplasm into the nuclei of the RAW 264.7 cells. The results showed that octadecanoic acid did not inhibit the translocation of p65 subunit, thus could not inhibit expression of inflammatory genes. Similarly, in the gene expression study by qualitative Reverse Transcriptase PCR, genes for phospholipase A₂ (PLA₂), cyclooxygenase-1 (COX-1), cyclooxygenase-2 (COX-2), 5-lipoxygenase (5-LOX) and inducible nitric oxide synthase (iNOS) involved in inflammatory signaling pathway were not affected by octadecanoic acid added to the cells at various concentrations (0.125 mg/mL – 1.0 mg/mL). In the inflammatory enzyme assays, only PLA₂ activity, but not COX-1, COX-2, and 5-LOX were inhibited. An IC₅₀ analysis showed that at concentrations of 0.24 mg/mL, octadecanoic acid inhibited 50% of PLA₂ activity. As a conclusion, the present study showed that *J.curcas* plant possessed anti-inflammatory activity, especially the roots and several compounds belonging to the terpene group present in the root, might contributed to the cytotoxicity of the root extract. The anti-inflammatory compound was identified to be octadecanoic acid and was found to inhibit PLA₂ enzyme activity (as the possible mode of action) by competing with the enzyme substrate as indicated by the IC₅₀ analysis, where PLA₂ activity was only inhibited at high concentrations of octadecanoic acid.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai
memenuhi keperluan untuk Ijazah Doktor Falsafah

**PENJELASAN TENTANG SEBATIAN ANTI-RADANG DI DALAM AKAR
Jatropha curcas LINN. DAN MOD TINDAKANNYA**

Oleh

AHMAD RAZI BIN OTHMAN

Jun 2016

Pengerusi : Professor Norhani Abdullah, PhD
Institut : Biosains

Jatropha curcas Linn. (famili *Euphorbiaceae*) merupakan pokok renek yang mampu bertahan dalam keadaan kontang dan banyak ditanam di kawasan Amerika Tengah, Amerika Selatan, Asia Tenggara, India dan Afrika. Pokok ini digunakan sebagai ubat tradisi bagi kebanyakan masyarakat di dunia. Di dalam bidang rawatan sakit radang, non-steroidal anti-inflammatory drugs (NSAIDs) telah di buktikan keberkesanannya dalam mengawal sakit radang. Walaubagaimanapun, beberapa kajian mendedahkan kesan sampingan daripada penggunaan ubat NSAIDs yang berpanjangan yang menyebabkan kebarangkalian menghadapi penyakit kronik seperti ulcer usus, sakit jantung yang kronik dan penyakit Alzheimer's. Oleh itu, perubatan alternatif berdasarkan tumbuhan perlu dipertimbangkan dalam merawat sakit radang. Tambahan pula, penggunaan ubatan herba kini semakin diterima dalam pelbagai bidang farmasutikal. Walaupun beberapa kajian telah menunjukkan bahagian-bahagian *J. curcas* memiliki keupayaan sebagai anti-radang, tetapi bahan aktif dan mekanisma tindakannya masih belum difahami sepenuhnya. Oleh yang demikian, sebelum sesuatu produk herba boleh digunakan, maklumat terperinci berkaitan dengan bahan bioaktifnya dan mekanisma tindakannya perlulah difahami terlebih dahulu. Dengan objektif utama kajian ini adalah untuk menentukan bahan aktif yang bersifat anti-radang daripada pokok *J.curcas* dan juga mekanisma tindakannya. Kajian yang dijalankan meliputi pemeriksaan terhadap ekstrak bahagian-bahagian pokok *Jatropha* yang memiliki sifat anti-radang dengan menggunakan sel Murine monocytic macrophage RAW 264.7, penulenan dan penjelasan tentang struktur bahan aktif tersebut, pembuktian tentang mekanisma tindakan oleh bahan aktif dan enzim-enzim yang terlibat didalamnya. Aktiviti anti-radang ditentukan dengan cara perencutan penghasilan nitrik oksida di dalam sel RAW 264.7, manakala aktiviti sitotoksik ditentukan dengan (3-(4, 5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) MTT assay. Di peringkat awal kajian, dapat diperhatikan bahawa ekstrak bahagian akar *Jatropha* menggunakan methanol memiliki aktiviti sebagai anti-radang yang lebih tinggi berbanding bahagian-bahagian yang lain (daun, buah dan batang), tetapi toksik terhadap sel Murine monocytic macrophage RAW 264.7. Seterusnya, extrak akar pecahkan kepada beberapa pecahan mengikut perbezaan polariti pelarut (hexane, chloroform dan ethyl acetate). Pecahan dari pelarut hexane memiliki aktiviti anti-radang yang tinggi, tetapi masih menunjukkan kadar toksik yang tinggi

terhadap pertumbuhan sel. Analisa terhadap kandungan yang terdapat di dalam pecahan hexane menggunakan teknik gas chromatography mass spectrometry (GC-MS) menunjukkan kehadiran kompaun-kompaun dari kumpulan terpene yang berkemungkinan punca kepada sifat toksik kompaun tersebut. Teknik penulenan seterusnya menggunakan sistem kolumn terbuka bagi mengasingkan kompaun yang memiliki sifat anti-radang tanpa bersifat toksik terhadap sel. Lima kompaun dapat diasingkan dan dilabel sebagai H-1, H-2,3, H-4 dan H-5 dari pecahan hexane. Assay anti-radang menunjukkan kompaun yang terdapat pada H-4 dan H-5 memiliki sifat anti-radang yang sangat tinggi tanpa menunjukkan sifat toksik. Analisa terhadap kompaun yang terdapat didalam H-4 dan H-5 menggunakan kaedah GC-MS menunjukkan kehadiran kumpulan asid lemak (heksadekanoik dan oktadekanoik) didalam kedua-dua kompaun H-4 dan H-5. Penulenan H-4 menggunakan kaedah “high performance liquid chromatography” (HPLC) menunjukkan kehadiran dua kompaun (A dan B) dengan eluen A memaparkan aktiviti anti-radang yang lebih bagus berbanding eluen B tanpa memiliki sifat toksik terhadap pertumbuhan sel. Pengenalpastian kompaun yang terdapat eluen A menggunakan “liquid chromatography-tandem mass spectrometry” (LC-MS/MS) menunjukkan kompaun tersebut tergolong di dalam kumpulan asid lemak oktadekanoik. Analisa selanjutnya menggunakan teknik “NuclearMagnetic Resonance” (NMR) membuktikan bahawa kompaun aktif adalah terdiri daripada rantai panjang hidrokarbon dengan hujungnya memiliki kumpulan karboksilik, lalu mengesahkan bahawa kompaun aktif tersebut adalah dari kumpulan asid lemak oktadekanoik. Bagi menentukan mekanisma tindakan di dalam aktiviti anti-radang, assay menggunakan pewarna berpendafluor dijalankan bagi memerhatikan proses translokasi oleh subunit NK- κ B (terlibat di dalam proses radang) dari sitoplasma ke dalam nucleus sel RAW 264.7. Hasil kajian menunjukkan asid oktadekanoik tidak menghalang proses translokasi subunit p65, seterusnya gagal menghalang proses penyalinan gen yang terlibat dengan proses radang. Bersamaan dengan kajian ekspresi gen menggunakan kaedah PCR Transkriptase berbalik terhadap gen phospholipase A₂ (PLA₂), cyclooxygenase-1 (COX-1), cyclooxygenase-2 (COX-2), 5-lipoxygenase (5-LOX) dan inducible nitric oxide synthase (iNOS) yang terlibat di dalam proses radang, asid oktadekanoik tidak menunjukkan sebarang kesan pada kepekatan yang berbeza (0.125 mg/mL – 1.0 mg/mL). Di dalam kajian terhadap aktiviti enzim, hanya enzim PLA₂ yang direncat, tetapi tidak COX-1, COX-2 dan 5-LOX. Analisis terhadap IC₅₀ menunjukkan pada kepekatan 0.24 mg/mL, asid oktadekanoik merencat 50% aktiviti PLA₂. Sebagai kesimpulan, kajian ini membuktikan bahawa pokok *J. curcas* memiliki kebolehan sebagai anti-radang, terutamanya pada bahagian akar, tetapi beberapa kompaun daripada kumpulan terpene berkemungkinan menyumbang kepada sifat toksik ekstrak akar berkenaan. Kompaun anti-radang yang dikenalpasti sebagai asid oktadekanoik mampu merencat aktiviti enzim PLA₂ dengan cara bersaing terhadap substrat seperti yang ditunjukkan di dalam analisis IC₅₀ di mana aktiviti PLA₂ hanya direncat apabila asid oktadekanoik berkepekatan tinggi.

ACKNOWLEDGEMENTS

Assalamualaikum.

My grateful thanks to Allah S.W.T. for allowing me and granted my pray to complete my PhD research. Also for those people around me along this journey that continuously support me and encourage me to complete my study.

First and foremost, I would like to address my gratitude to my supervisor, Professor Dr. Norhani Abdullah for her precious advice, constant encouragement and guidance throughout this research.

Secondly, I would like to thanks my committee members, Dr. Syahida Ahmad and Dr. Intan Safinar Ismail for their support and thoughtful regarding my PhD research thoroughly. Without their help, I could not imagine completing this thesis.

To Natural Product Laboratory in IBS and Animal Cell Culture Lab in Biotech 2 lab members, I would like to thank all of you for helping and supporting me throughout my study.

Last but not least, my deepest thank to my family, my mother Nekmah Mat Yusof, my father Othman Sanusi, my wife Nurlizah Abu Bakar, my kids Ummar Rifqi Ahmad Razi and Sarah Sofea Ahmad Razi, my siblings and my in laws. Their endless support and encourage could not be paid by anything. Forever, I will in debt with them.

I certify that a Thesis Examination Committee has met on 10 June 2016 to conduct the final examination of Ahmad Razi bin Othman on his thesis entitled "Elucidation of The Anti-Inflammatory Compound Present in *Jatropha curcas* Linn Root and Its Mode of Action" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Muhajir bin Hamid, PhD

Associated Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Hawa binti Jaafar, PhD

Associated Professor

Faculty of Agriculture

Universiti Putra Malaysia

(Internal Examiner)

Md Zuki bin Abu Bakar @ Zakari, PhD

Professor

Institute of Bioscience

Universiti Putra Malaysia

(Internal Examiner)

Hiroshi Morita, PhD

Professor

Hoshi University

Japan

(External Examiner)



ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 26 July 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follow:

Norhani Abdullah, PhD

Professor

Faculty Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Syahida Ahmad, PhD

Senior Lecturer

Faculty Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

Intan Safinar Ismail, PhD

Associate Professor

Faculty of Science

Universiti Putra Malaysia

(Member)

BUJANG KIM HUAT, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustrations and citations have been duly referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No: Ahmad Razi Bin Othman, GS33714

Declaration by Members of Supervisory Committee

This is to confirm that:

- The research conducted and the writing of this thesis was under our supervision;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:

Name of Chairman
of Supervisory
Committee:

Professor Dr. Norhani Abdullah

Signature:

Name of Member
of Supervisory
Committee:

Dr. Syahida Ahmad

Signature:

Name of Member
of Supervisory
Committee:

Associate Professor Dr. Intan Safinar Ismail

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xiv
LIST OF ABBREVIATIONS	xvii
 CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	3
2.1 Inflammation	3
2.1.1 Acute Inflammation	3
2.1.2 Chronic Inflammation	4
2.2 Mechanism of inflammation	4
2.2.1 Receptors	4
2.2.2 Arachidonic Acid pathway	5
2.2.2.1 Phospholipase A ₂	5
2.2.2.2 Cyclooxygenase 1 and 2	6
2.2.2.3 5-Lipoxygenase	8
2.2.3 Nitric Oxide Pathway	9
2.2.3.1 Inducible Nitric Oxide Synthase (iNOS)	9
2.3 Anti-inflammation	11
2.3.1 Non-steroidal Anti-Inflammatory Drugs (NSAIDs)	11
2.4 Herbs as anti-inflammatory remedies	14
2.4.1 Medicinal herbs and inflammation	14
2.4.2 Medicinal Plants	15
2.5 Jatropha curcas Linn	16
2.5.1 Jatropha species in medicinal application	17
2.5.2 Jatropha species as a source of bioactive compounds	19
3 ANTI-INFLAMMATORY PROPERTY OF DIFFERENT PARTS OF JATROPHA CURCAS LINN.	20
3.1 Introduction	20
3.2 Materials and Methods	21
3.2.1 Sample collection	21
3.2.1.1 Leaves	21
3.2.1.2 Fruits	21
3.2.1.3 Stem bark	21
3.2.1.4 Roots	22
3.2.2 Preparation of plant extracts	22
3.2.3 Liquid – liquid fractionation	22
3.2.4 Phenolic content	23
3.2.5 Anti-inflammatory assay	23

3.2.5.1	Nitric oxide determination	24
3.2.5.2	Cytotoxicity assay	24
3.2.6	Gas chromatography mass spectra (GC-MS)	25
3.2.7	Statistical analysis	25
3.3	Results and Discussion	25
3.3.1	Freeze dried samples	25
3.3.2	Phenolic content	26
3.3.3	Effect of inducer on nitric oxide production	27
3.3.4	Anti-inflammatory activity of different parts of Jatropha curcas plant	27
3.3.5	Jatropha curcas roots fraction	29
3.3.5.1	Phenolic content for J. curcas roots fractions	30
3.3.5.2	Anti-inflammatory activity of J. curcas roots fractions	30
3.3.6	Identification of compounds in hexane fraction using GC-MS	32
3.4	Conclusion	34
4	ELUCIDATION OF ANTI-INFLAMMATORY COMPOUNDS IN HEXANE FRACTION	35
4.1	Introduction	35
4.2	Materials and Methods	36
4.2.1	Chromatographic Methods	36
4.2.1.1	Mini Open Column and Thin Layer Chromatography (TLC)	36
4.2.2	High Performance Liquid Chromatography (HPLC)	36
4.2.3	Gas Chromatography Mass Spectrometry (GC-MS)	37
4.2.4	Liquid Chromatography/ Tandem Mass Spectrometry (LC-MS/MS)	37
4.2.5	Nuclear Magnetic Resonance (NMR)	37
4.2.6	Inhibitive concentration (IC_{50})	37
4.2.7	Statistical analysis	38
4.3	Results and Discussion	38
4.3.1	Open Column and Thin Layer Chromatography (TLC)	38
4.3.2	Gas Chromatography-Mass Spectrophotometer (GC-MS)	40
4.3.3	Separation of compounds by HPLC	42
4.3.4	Identification of active compound using LC-MS/MS	43
4.3.5	Nuclear magnetic resonance (NMR) analysis	46
4.3.6	Inhibitive concentration of anti-inflammatory compound	49
4.4	Conclusion	50
5	EFFECT OF OCTADECANOIC ACID ON INFLAMMATORY SIGNALLING PATHWAY AND ENZYMES IN MURINE MACROPHAGE RAW 264.7 CELLS	51
5.1	Introduction	51
5.2	Materials and Methods	52
5.2.1	Translocation of NF- κ B cell	52
5.2.2	Expression of PLA ₂ , COX-1, COX-2, 5-LOX and iNOS genes by using Reverse Transcriptase PCR	52

5.2.2.1	Total RNA extraction	52
5.2.2.2	Reverse Transcriptase-Polymerase Chain Reaction (RT-PCR)	52
5.2.2.2.1	Phospholipase A ₂ (PLA ₂) gene expression	53
5.2.2.2.2	Cyclooxygenase 1 (COX-1) gene expression	53
5.2.2.2.3	Cyclooxygenase 2 (COX-2) gene expression	53
5.2.2.2.4	Inducible Nitric Oxide Synthase (iNOS)	54
5.2.2.2.5	5-Lipoxygenase gene (5-Lox) expression	54
5.3	Results and Discussion	54
5.3.1	Translocation of NF-κB cell	54
5.3.2	Inflammatory genes expression	55
5.3.2.1	Phospholipase A ₂ (PLA ₂) expression	55
5.3.2.2	Cyclooxygenase 1 gene expression	56
5.3.2.3	Cyclooxygenase 2 gene expression	57
5.3.2.4	5-Lipoxygenase gene expression	58
5.3.2.5	Inducible Nitric Oxide Synthase (iNOS)	59
5.4	Conclusion	60
6	EFFECT OF OCTADECANOIC ACID ON INFLAMMATORY ENZYMES	61
6.1	Introduction	61
6.2	Materials and Methods	61
6.2.1	Development of Inhibitive Concentration (IC ₅₀) of octadecanoic acid	61
6.2.2	Cyclooxygenase 1 (COX-1) and Cyclooxygenase 2 (COX-2) enzymes activities	62
6.2.3	5-lipoxygenase (5-LOX) enzyme activity	62
6.2.4	Phospholipase A ₂ enzyme activity	62
6.2.5	Statistical analysis	63
6.3	Results and Discussion	63
6.3.1	Cyclooxygenase-1 (COX-1) activity	63
6.3.2	Cyclooxygenase-2 (COX-2) activity	64
6.3.3	5-Lipoxygenase (5-LOX) activity	65
6.3.4	Phospholipase A2 (PLA2) activity	65
6.4	Conclusion	67
7	GENERAL DISCUSSION, CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE RESEARCH	68
7.1	Conclusions	71
7.2	Recommendations For Future Research	72
REFERENCES		73
APPENDICES		96
BIODATA OF STUDENT		102
PUBLICATION		103

LIST OF TABLES

Table	Page
2.1 Summary of comparison between COX-1 and COX-2 characteristics	6
2.2 List of non-selective NSAIDs and COX-2 selective NSAIDs	12
2.3 Jatropha species used in traditional medicines	18
3.1 Phenolic contents of different parts of <i>J. curcas</i>	26
3.2 Phenolic content of <i>J.curcas</i> root in different solvent fractions	30
3.3 Compounds present in hexane partition by GC-MS analysis	33
4.1 Compounds identified by LC-MS/MS	43
4.2 2D-NMR spectral data for metabolite A	47
5.1 Forward and reverse primers for genes involved in inflammation activity	53

LIST OF FIGURES

Figure		Page
2.1	Diagram showing the binding site of arachidonic acid to COX-2 enzyme on its active site at amino acid Tyr385	7
2.2	Diagram explaining the addition of 2 oxygen molecules to arachidonic acid, thereby converting it to PGG ₂ . The unstable PGG ₂ form is then rapidly converted to PGH ₂	8
2.3	Nitric oxide (NO) synthesis from L-arginine. In the presence NADPH and O ₂ , L-arginine is converted to NO and L-citrulline	11
2.4	<i>Jatropha curcas</i> Linn. plant at Field 2, Universiti Putra Malaysia.	17
3.1	A flow diagram of liquid-liquid fractionation method to fractionate the crude root sample.	23
3.2	Daily changes in weight of <i>Jatropha curcas</i> samples dried by freeze drying.	26
3.3	Nitric oxide (NO) production in RAW 264.7 macrophage cells at various concentrations of lipopolysaccharides (LPS) and interferon-gamma (IFN- γ) applied singly or in combination.	27
3.4	Anti-inflammatory activity of methanolic extract in RAW 264.7 macrophage cells.	28
3.5	Cytotoxic activity of methanolic extract of different parts of <i>J. curcas</i> plant using MTT assay.	29
3.6	A flowchart of a liquid-liquid partition step of a crude extract from <i>J. curcas</i> root.	30
3.7	Anti-inflammatory activity of different solvent fractions.	31
3.8	Cytotoxic activity of different solvent fractions of root by MTT assay.	32
3.9	GC-MS spectrum of hexane partition.	33
4.1	Anti-inflammatory activity of different spots isolated from hexane fraction by open column chromatography.	39
4.2	Cell viability of different spots isolated from hexane partition by open column chromatography using MTT assay.	40

4.3	GC-MS spectrum for H-4 spot from hexane fraction.	41
4.4	GC-MS spectrum for H-5 spot from hexane fraction.	41
4.5	HPLC profile of compounds in spot H-4	42
4.6	Anti-inflammatory assay for metabolites A and B.	42
4.7	A cytotoxicity assay for metabolites A and B.	43
4.8	LC-MS/MS spectrum aligned with database showed that peak at 8.04 minutes was identified as 12-oxooctadecanoic acid.	44
4.9	LC-MS/MS spectrum aligned with database showed that peak between 8.04 and 8.57 minutes was identified as 15, 16-dihydroxy-9Z, 12Z-octadecadienoic acid.	44
4.10	LC-MS/MS spectrum aligned with database showed that peak at 8.57 minutes was identified as octadecadienoic acid isomer.	45
4.11	A LC-MS/MS spectrum aligned with database showed that peak between 8.57 and 9.11 minutes was identified as 15S, 16S-dihydroxy-9Z, 12Z-octadecadienoic acid.	45
4.12	LC-MS/MS spectrum aligned with database showed that peak at 9.11 minutes was identified as octadecanoic acid isomer.	46
4.13	LC-MS/MS spectrum aligned with database showed that peak at 9.49 minutes was identified as cholenic acid sulfate.	46
4.14	Structure of octadecanoic acid	47
4.15	^1H -NMR spectrum of metabolite A (CDCl_3 ; 500MHz)	47
4.16	^{13}C -NMR spectrum of metabolite A (CDCl_3 ; 125MHz).	48
4.17	An IC_{50} analysis for anti-inflammatory property of octadecanoic acid	49
5.1	A fluorescent microscopy image of control and treated RAW 264.7 cells.	55
5.2	Expression of PLA ₂ gene in RAW 264.7 cells	56
5.3	Expression of COX-1 gene in RAW 264.7 cells	57
5.4	Expression of COX-2 gene in RAW 264.7 cells	58
5.5	Expression of 5-LOX gene in RAW 264.7 cells	59

5.6	Expression of iNOS gene in RAW 264.7 cells	60
6.1	Percentage activity of COX-1 enzyme at various concentrations of octadecanoic acid	63
6.2	Percentage activity of COX-2 enzyme at various concentrations of octadecanoic acid	64
6.3	Percentage activity of 5-LOX enzyme enzyme at various concentrations of octadecanoic acid	65
6.4	Percentage activity of PLA ₂ enzyme at various concentrations of octadecanoic acid	66
6.5	IC ₅₀ plot for PLA ₂ activity	67

LIST OF ABBREVIATIONS

δ	Chemical shift in ppm
$^{\circ}\text{C}$	Degree in Celsius
^{13}C	Carbon-13
DEPT	Distortionless Enhancement by Polarization Transfer
DMSO	Dimethylsulfoxide
EtOAc	Ethyl acetate
GC-MS	Gas Chromatography-Mass Spectrometry
^1H	Proton
gHMBC	Gradient Heteronuclear Multiple Bond Correlation
gHSQC	Gradient Heteronuclear Single-Quantum Coherence
gCOSY	Gradient Correlation Spectroscopy
J	Coupling in Hz
m	Multiplet
m/z	Mass per charge
MeOH	Methanol
MS	Mass Spectrum/ Mass Spectrometry
NMR	Nuclear Magnetic Resonance
s	Singlet
TLC	Thin Layer Chromatography
UV	Ultraviolet
IC	Inhibition concentration
PLA ₂	Phospholipase A ₂
COX-1	Cylooxygenase-1

COX-2	Cyclooxygenase-2
iNOS	Inducible Nitric Oxide Synthase
5-LOX	5-Lipoxygenase
FLAP	Five Lipoxygenase Activated Protein
CRP	C-Reactive Protein
NF-κB	Nuclear Factor Kappa B
I-κB	Inhibitor Kappa B
DTNB	5,5'-dithiobis-(2-nitrobenzoic acid)
AA	Arachidonic Acid
NO	Nitric Oxide
DMEM	Dulbecco's Modified Eagle Medium
LPS	Lipopolysaccharide
IFN-γ	Interferon gamma
ROS	Reactive Oxygen Species
NSAID	Non-steroidal Anti-Inflammatory Drug
ILs	Interleukins
LTs	Leukotriene
PGs	Prostaglandins
TNF-α	Tumor Necrosis Factor Alpha
TLR	Toll Like Receptor
DNA	Deoxyribonucleic Acid
mRNA	Messenger Ribonucleic Acid
RT-PCR	Reverse Transcriptase Polymerase Chain Reaction
PIC	Pro-Inflammatory Cytokines
MAPK	Mitogen Activated Protein Kinase

CDCl ₃	Deuterated chloroform
HPLC	High Performance Liquid Chromatography
LC-MS	Liquid Chromatography-Mass Spectrometry
µg	microgram
mg	milligram
mM	millimolar
µl	microliter
ml	milliliter
U/mg	Unit per milligram
U/ml	Unit per milliliter
g	gram
MTT	3-(4, 5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide

CHAPTER 1

INTRODUCTION

Inflammation is defined as the biological process which occurs when the immune system responds to pathogenic infections or damaged cells (Hansel et al., 2010). Inflammation normally causes several symptoms such as swelling, redness, pain, increase in temperature, and numbness. These common symptoms are due to the increased blood flow and capillary permeability (Hansel et al., 2010). Inflammation is also apparent in several joint illnesses such as osteoarthritis (OA), rheumatoid arthritis and gout (Siebuhr et al., 2014, de Lange-Brokaar et al., 2012, Punzi et al., 2012) which occur as a result of prolonged or chronic inflammation.

Inflammation is a condition brought about by mediators commonly known as cytokines. These are glycoproteins that are synthesized in various types of cells under stress conditions. There are numerous types of cytokines, such as interleukin-6 (IL-6), interferon (IFN) and tumor necrosis factor (TNFs) (Hansel et al., 2010). TNF- α is synthesized by macrophage and B-lymphocytes upon stimulation (O'Connor et al., 2009), which also acts as proinflammatory cytokine that triggers the release of other cytokines (IL-4 and IL-6) and C-reactive protein (CRP) in inflammation signaling pathway (Inoue et al., 2009, Ingle and Patel, 2011).

Enzymes also play a major role in regulating inflammation pathway. These enzymes are Phospholipase A₂ (PLA₂), Cyclooxygenase-1 (COX-1), Cyclooxygenase-2 (COX-2) and 5-Lipoxygenase (5-LOX). These enzymes are induced by the activation of Nuclear Factor- κ B (NF- κ B) cells in cytoplasm. Activation of NF- κ B exposes the nuclear localization sequence, thus leads to the translocation of NF- κ B from cytosol to nucleus and expresses specific genes for inflammation process (Delfino and Walker, 1999). In general, PLA₂ synthesizes arachidonic acid from *sn*-2 position of phospholipids in membrane (Murakami et al., 2010). Cyclooxygenase enzymes (COX-1 and COX-2) convert arachidonic acid to prostaglandins (PGs) and thromboxane A₂ (Marnett, 2000). 5-LOX converts arachidonic acid to leukotriene that is involved in regulation of inflammation in lung tissues (Steinhilber, 1994, Liu and Yokomizo, 2015). There have been numerous studies related to regulation of inflammatory enzymes especially COX enzymes (van Esch et al., 2013, Meskell and Ettarh, 2011). Drugs developed specifically to inhibit COX activity in human cells are named as non-steroidal anti-inflammatory drugs (NSAIDs).

Research has been conducted for decades to find a cure for inflammation and its related symptoms. Now it is widely accepted that non-steroidal anti-inflammatory drugs (NSAIDs) can effectively prevent inflammation (Shen et al., 2011, Raz, 2002). However, several studies have also revealed side effects resulting from prolonged use of NSAIDs, which include the possibility of several chronic diseases such as gastrointestinal (GI) ulcers, adverse cardiovascular side-effects, and Alzheimer's disease (Sostres et al., 2010, Niranjan et al., 2011, McGeer and McGeer, 2007). Because of the

side effects related to NSAID use, many studies have been conducted to find alternative anti-inflammatory drugs based on natural products, as these remedies may have no or less side effects. One of the plants that has been associated with anti-inflammatory properties is *Coptis chinesis*. This plant is widely used by the Chinese in treating inflammation traditionally. Scientific research proves that the presence of alkaloids group (berberine) in the plant *Berberis koreana* was the major contributor for its anti-inflammatory effect (Kim et al., 2010). *Sophora subprostrata* is another example of plant that has been used traditionally by the Chinese in treating inflammation. This plant showed an inhibition towards *in vitro* COX activity and also acted as antioxidant due the presence of quinolizidine alkaloids matrine and oxymatrine (Souto et al., 2011). *Ceanothus thyrsiflorus* was used by the American tribes to treat various types of diseases related to inflammation. The tribes used plant roots as drinks (tea) to treat sore throats and bronchitis. The presence of alkaloids group and saponins were the major compounds that contribute to anti-inflammatory effect in this plant (Darshan and Doreswamy, 2004). *Jatropha curcas* L. is another plant that has been used as a traditional medicine in treating inflammation. *Jatropha curcas* was used by tribes in South America, particularly in Brazil, to treat inflammation (Villegas et al., 1997).

Jatropha curcas is a drought tolerant, versatile perennial plant in the family of Euphorbiaceae. It originated from South America, but now abundant in South and Central America, Africa and Asia (Mandpe et al., 2005). The plant can grow in harsh conditions with low or high rainfall. This plant is considered to have enormous potentials, not only as a source of oil seed but also as a source of bioactive compounds for medicinal purposes. Different parts of the plant have been shown to possess biological activities which have been associated to the presence of phenolics, terpenoids and flavonoids (Oskoueian et al., 2011b).

However, the nature of the active compound has not been completely elucidated, and the mode of action in the anti-inflammatory activity remained unclear. Thus, the general objective of the present study was to elucidate the chemical nature of anti-inflammatory compounds present in *J. curcas* plant and to determine its effects on the inflammatory signaling pathway and enzymes.

The specific objectives were:

1. To screen the anti-inflammatory activity of extracts from different parts of *J. curcas* in Murine monocytic macrophage RAW 264.7 cell line.
2. To purify and elucidate the structure of the anti-inflammatory compounds extracted from *J. curcas* plant.
3. To determine the effects of anti-inflammatory compound on the inflammatory signaling pathway in Murine monocytic macrophage RAW 264.7 cell line.
4. To determine the effect of anti-inflammatory compound on enzymes involved in the inflammation signaling pathway.

REFERENCES

- ABDELGADIR, H. A. & VAN STADEN, J. 2013. Ethnobotany, ethnopharmacology and toxicity of *Jatropha curcas* L. (Euphorbiaceae): A review. *South African Journal of Botany*, 88, 204-218.
- ABDULLA, R., CHAN, E. S. & RAVINDRA, P. 2011. Biodiesel production from *Jatropha curcas*: a critical review. *Critical Reviews in Biotechnology*, 31, 53-64.
- ADOLF, W., OPPERKUCH, H. J. & HECKER, E. 1984. Irritant phorbol derivatives from four *Jatropha* species. *Phytochemistry*, 23, 129-132.
- AIDAH, N., ABDULLAH, N., OSKOUEIAN, E., SIEO, C. C. & SAAD, W. Z. 2014. Membrane-active antibacterial compounds in methanolic extracts of *Jatropha curcas* and their mode of action against *Staphylococcus aureus* S1434 and *Escherichia coli* E216. *International Journal Agriculture Biogyl*, 16, 723-730.
- AIYELAAGBE, O. O. 2001. Antibacterial activity of *Jatropha multifida* roots. *Fitoterapia*, 72, 544-546.
- AKTAN, F. 2004. iNOS-mediated nitric oxide production and its regulation. *Life Sciences*, 75, 639-653.
- AKULA, U. S. & ODHAV, B. 2013. In vitro 5-lipoxygenase inhibition of polyphenolic antioxidants from undomesticated plants of South Africa. *Journal of Medicinal plants research*, 2, 207-212.
- ALHAKMANI, F., KUMAR, S. & KHAN, S. A. 2013. Estimation of total phenolic content, in-vitro antioxidant and anti-inflammatory activity of flowers of *Moringa oleifera*. *Asian Pacific Journal of Tropical Biomedicine*, 3, 623-627.
- AMID, B. T. & MIRHOSSEINI, H. 2012. Influence of different purification and drying methods on rheological properties and viscoelastic behaviour of durian seed gum. *Carbohydrate Polymers*, 90, 452-461.
- APARNA, V., DILEEP, K. V., MANDAL, P. K., KARTHE, P., SADASIVAN, C. & HARIDAS, M. 2012. Anti-Inflammatory Property of n-Hexadecanoic Acid: Structural Evidence and Kinetic Assessment. *Chemical Biology & Drug Design*, 80, 434-439.
- ASASE, A. & KADERA, M. L. 2014. Herbal medicines for child healthcare from Ghana. *Journal of Herbal Medicine*, 4, 24-36.
- ASASE, A., OTENG-YEBOAH, A. A., ODAMTTEN, G. T. & SIMMONDS, M. S. J. 2005. Ethnobotanical study of some Ghanaian anti-malarial plants. *Journal of Ethnopharmacology*, 99, 273-279.

- ATANASSOVA, M., GEORGIEVA, S. & IVANCHEVA, K. 2011. Total phenolic and total flavonoid contents, antioxidant capacity and biological contaminants in medicinal herbs. *Journal of the University of Chemical Technology and Metallurgy*, 46, 81-88.
- BAE, H., KIM, R., KIM, Y., LEE, E., JIN KIM, H., PYO JANG, Y., JUNG, S.-K. & KIM, J. 2012. Effects of *Schisandra chinensis* Baillon (Schizandraceae) on lipopolysaccharide induced lung inflammation in mice. *Journal of Ethnopharmacology*, 142, 41-47.
- BAE, I.-K., MIN, H.-Y., HAN, A.-R., SEO, E.-K. & LEE, S. K. 2005. Suppression of lipopolysaccharide-induced expression of inducible nitric oxide synthase by brazilin in RAW 264.7 macrophage cells. *European Journal of Pharmacology*, 513, 237-242.
- BAEUEERLE, P. A. & BALTIMORE, D. 1988. I kappa B: a specific inhibitor of the NF-kappa B transcription factor. *Science*, 242, 540-546.
- BALBOA, M. A. A., VARELA-NIETO, I., KILLERMANN LUCAS, K. & DENNIS, E. A. 2002. Expression and function of phospholipase A₂ in brain. *Federation of European Biochemical Societies Letters*, 531, 12-17.
- BALDWIN JR, A. S. 1996. The NF-κB and IκB proteins: new discoveries and insights. *Annual review of immunology*, 14, 649-681.
- BALLOU, L. R. & CHEUNG, W. Y. 1985. Inhibition of human platelet phospholipase A₂ activity by unsaturated fatty acids. *Proceedings of the National Academy of Sciences*, 82, 371-375.
- BERENGUER, B., SÁNCHEZ, L. M., QUÍEZ, A., LÓPEZ-BARREIRO, M., DE HARO, O., GÁLVEZ, J. & MARTÍN, M. J. 2006. Protective and antioxidant effects of *Rhizophora mangle* L. against NSAID-induced gastric ulcers. *Journal of Ethnopharmacology*, 103, 194-200.
- BERNARD, M. P., BANCOS, S., SIME, P. J. & PHIPPS, R. P. 2008. Targeting Cyclooxygenase-2 in Hematological Malignancies: Rationale and Promise. *Current pharmaceutical design*, 14, 2051-2060.
- BINDU, S., MAZUMDER, S., DEY, S., PAL, C., GOYAL, M., ALAM, A., IQBAL, M. S., SARKAR, S., AZHAR SIDDIQUI, A., BANERJEE, C. & BANDYOPADHYAY, U. 2013. Nonsteroidal anti-inflammatory drug induces proinflammatory damage in gastric mucosa through NF-κB activation and neutrophil infiltration: Anti-inflammatory role of heme oxygenase-1 against nonsteroidal anti-inflammatory drug. *Free Radical Biology and Medicine*, 65, 456-467.

BIONAS. Retrieved January 2016 from <http://www.bionas.com.my/17dec2015.html>

- BISHOP-BAILEY, D., MITCHELL, J. A. & WARNER, T. D. 2006. COX-2 in Cardiovascular Disease. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 26, 956-958.
- BIZZINTINO, J. A., KHOO, S.-K., ZHANG, G., MARTIN, A. C., RUETER, K., GEELHOED, G. C., GOLDBLATT, J., LAING, I. A., LE SOUËF, P. N. & HAYDEN, C. M. 2009. Leukotriene pathway polymorphisms are associated with altered cysteinyl leukotriene production in children with acute asthma. *Prostaglandins, Leukotrienes and Essential Fatty Acids*, 81, 9-15.
- BOMBARDIER, C., LAINE, L., REICIN, A., SHAPIRO, D., BURGOS-VARGAS, R., DAVIS, B., DAY, R., FERRAZ, M. B., HAWKEY, C. J. & HOCHBERG, M. C. 2000. Comparison of upper gastrointestinal toxicity of rofecoxib and naproxen in patients with rheumatoid arthritis. *New England Journal of Medicine*, 343, 1520-1528.
- BUAPOOL, D., MONGKOL, N., CHANTIMAL, J., ROYTRAKUL, S., SRISOOK, E. & SRISOOK, K. 2013. Molecular mechanism of anti-inflammatory activity of *Pluchea indica* leaves in macrophages RAW 264.7 and its action in animal models of inflammation. *Journal of Ethnopharmacology*, 146, 495-504.
- CAI, H. & HARRISON, D. G. 2000. Endothelial Dysfunction in Cardiovascular Diseases: The Role of Oxidant Stress. *Circulation Research*, 87, 840-844.
- CALDER, P. C. 2004. n-3 Fatty acids and cardiovascular disease: evidence explained and mechanisms explored. *Clinical Science.*, 107, 1-11.
- CALDER, P. C. 2006. n-3 polyunsaturated fatty acids, inflammation, and inflammatory diseases. *The American journal of clinical nutrition*, 83, S1505-1519S.
- CALVELLO, R., PANARO, M. A., CARBONE, M. L., CIANCIULLI, A., PERRONE, M. G., VITALE, P., MALERBA, P. & SCILIMATI, A. 2012. Novel selective COX-1 inhibitors suppress neuroinflammatory mediators in LPS-stimulated N13 microglial cells. *Pharmacological Research*, 65, 137-148.
- CANALI, R., COMITATO, R., SCHONLAU, F. & VIRGILI, F. 2009. The anti-inflammatory pharmacology of Pycnogenol® in humans involves COX-2 and 5-LOX mRNA expression in leukocytes. *International Immunopharmacology*, 9, 1145-1149.
- CARMONA, F. & PEREIRA, A. M. S. 2013. Herbal medicines: old and new concepts, truths and misunderstandings. *Revista Brasileira de Farmacognosia*, 23, 379-385.
- CERELLA, C., SOBOLEWSKI, C., DICATO, M. & DIEDERICH, M. 2010. Targeting COX-2 expression by natural compounds: A promising alternative strategy to synthetic COX-2 inhibitors for cancer chemoprevention and therapy. *Biochemical Pharmacology*, 80, 1801-1815.

- CHAN, E. W. C., LIM, Y. Y., WONG, S. K., LIM, K. K., TAN, S. P., LIANTO, F. S. & YONG, M. Y. 2009. Effects of different drying methods on the antioxidant properties of leaves and tea of ginger species. *Food Chemistry*, 113, 166-172.
- CHAN, M. M.-Y., FONG, D., HO, C.-T. & HUANG, H.-I. 1997. Inhibition of Inducible Nitric Oxide Synthase Gene Expression and Enzyme Activity by Epigallocatechin Gallate, a Natural Product from Green Tea. *Biochemical Pharmacology*, 54, 1281-1286.
- CHANG, C.-C., CHUANG, C.-L., LEE, W.-S., WANG, S.-S., LEE, F.-Y., LIN, H.-C., HUANG, H.-C. & LEE, S.-D. 2013. Selective cyclooxygenase-1 inhibition improves collateral vascular reactivity in biliary cirrhotic rats. *Journal of the Chinese Medical Association*, 76, 557-563.
- CHEN, C.-H., CHAN, H.-C., CHU, Y.-T., HO, H.-Y., CHEN, P.-Y., LEE, T.-H. & LEE, C.-K. 2009. Antioxidant activity of some plant extracts towards xanthine oxidase, lipoxygenase and tyrosinase. *Molecules*, 14, 2947-2958.
- CHEN, F. E., HUANG, D.-B., CHEN, Y.-Q. & GHOSH, G. 1998. Crystal structure of p50/p65 heterodimer of transcription factor NF-κB bound to DNA. *Nature*, 391, 410-413.
- COMERFORD, S. 1996. Medicinal plants of two Mayan Healers from San Andres, Peten, Guatemala. *Economic Botany*, 50, 327-336.
- CUZICK, J., OTTO, F., BARON, J. A., BROWN, P. H., BURN, J., GREENWALD, P., JANKOWSKI, J., LA VECCHIA, C., MEYSKENS, F., SENN, H. J. & THUN, M. 2009. Aspirin and non-steroidal anti-inflammatory drugs for cancer prevention: an international consensus statement. *The Lancet Oncology*, 10, 501-507.
- DAMON, M., CHAVIS, C., CRASTES DE PAULET, A., MICHEL, F. B. & GODARD, P. 1987. Arachidonic acid metabolism in alveolar macrophages. A comparison of cells from healthy subjects, allergic asthmatics, and chronic bronchitis patients. *Prostaglandins*, 34, 291-309.
- DAN, P., ROSENBLAT, G. & YEDGAR, S. 2012. Phospholipase A₂ activities in skin physiology and pathology. *European Journal of Pharmacology*, 691, 1-8.
- DARSHAN, S. & DORESWAMY, R. 2004. Patented antiinflammatory plant drug development from traditional medicine. *Phytotherapy research*, 18, 343-357.
- DE LANGE-BROKAAR, B. J. E., IOAN-FACSIMAY, A., VAN OSCH, G. J. V. M., ZUURMOND, A. M., SCHOOONES, J., TOES, R. E. M., HUIZINGA, T. W. J. & KLOPPENBURG, M. 2012. Synovial inflammation, immune cells and their cytokines in osteoarthritis: a review. *Osteoarthritis and Cartilage*, 20, 1484-1499.
- DELFINO, F. & WALKER, W. H. 1999. Hormonal regulation of the NF-κB signaling pathway. *Molecular and cellular endocrinology*, 157, 1-9.

- DEMARIA, A. N. & WEIR, M. R. 2003. Coxibs—Beyond the GI Tract: Renal and Cardiovascular Issues. *Journal of Pain and Symptom Management*, 25, 41-49.
- DEVAPPA, R., MAKKAR, H. S. & BECKER, K. 2011. Jatropha Diterpenes: a Review. *Journal of the American Oil Chemists' Society*, 88, 301-322.
- DOS SANTOS, A. F. & SANT'ANA, A. E. G. 1999. Molluscicidal activity of the diterpenoids jatrophe and jatropholones A and B isolated from *Jatropha elliptica* (Pohl) Muell. Arg. *Phytotherapy Research*, 13, 660-664.
- DUGASANI, S., PICHKA, M. R., NADARAJAH, V. D., BALIJEPELLI, M. K., TANDRA, S. & KORLAKUNTA, J. N. 2010. Comparative antioxidant and anti-inflammatory effects of [6]-gingerol, [8]-gingerol, [10]-gingerol and [6]-shogaol. *Journal of Ethnopharmacology*, 127, 515-520.
- DUGOWSON, C. E. & GNANASHANMUGAM, P. 2006. Nonsteroidal anti-inflammatory drugs. *Physical Medicine and Rehabilitation Clinics of North America*, 17, 347-354.
- EISENBERG, D. M., DAVIS, R. B., ETTNER, S. L. & ET AL. 1998. Trends in alternative medicine use in the united states, 1990-1997: Results of a follow-up national survey. *The Journal of the American Medical Association (JAMA)*, 280, 1569-1575.
- EL-SHITANY, N. A., EL-BASTAWISSY, E. A. & EL-DESOKY, K. 2014. Ellagic acid protects against carrageenan-induced acute inflammation through inhibition of nuclear factor kappa B, inducible cyclooxygenase and proinflammatory cytokines and enhancement of interleukin-10 via an antioxidant mechanism. *International Immunopharmacology*, 19, 290-299.
- ELFAHMI, WOERDENBAG, H. J. & KAYSER, O. 2014. Jamu: Indonesian traditional herbal medicine towards rational phytopharmacological use. *Journal of Herbal Medicine*, 4, 51-73.
- EVANS, J. F., FERGUSON, A. D., MOSLEY, R. T. & HUTCHINSON, J. H. 2008. What's all the FLAP about?: 5-lipoxygenase-activating protein inhibitors for inflammatory diseases. *Trends in Pharmacological Sciences*, 29, 72-78.
- FALODUN, A., IGBE, I., ERHARUYI, O. & AGBANYIM, O. J. 2013. Chemical Characterization, Anti inflammatory and Analgesic Properties of *Jatropha Multifida* Root Bark. *Journal of Applied Sciences and Environmental Management*, 17, 357-362.
- FERGUSON, J. F., HINKLE, C. C., MEHTA, N. N., BAGHERI, R., DEROHANESSIAN, S. L., SHAH, R., MUCKSAVAGE, M. I., BRADFIELD, J. P., HAKONARSON, H., WANG, X., MASTER, S. R., RADER, D. J., LI, M. & REILLY, M. P. 2012. Translational Studies of Lipoprotein-Associated Phospholipase A₂ in Inflammation and Atherosclerosis. *Journal of the American College of Cardiology*, 59, 764-772.

- FERNANDES, E. D. S., RODRIGUES, F. A., TÓFOLI, D., IMAMURA, P. M., CARVALHO, J. E. D., RUIZ, A. L. T. G., FOGLIO, M. A., MINGUZZI, S. & SILVA, R. C. L. 2013. Isolation, structural identification and cytotoxic activity of hexanic extract, cyperenoic acid, and jatropheone terpenes from *Jatropha ribifolia* roots. *Revista Brasileira de Farmacognosia*, 23, 441-446.
- FERRERO-MILIANI, L., NIELSEN, O. H., ANDERSEN, P. S. & GIRARDIN, S. E. 2007. Chronic inflammation: importance of NOD2 and NALP3 in interleukin-1 β generation. *Clinical and Experimental Immunology*, 147, 227-235.
- FRIES, S. & GROSSER, T. 2005. The Cardiovascular Pharmacology of COX-2 Inhibition. *ASH Education Program Book*, 2005, 445-451.
- FUJIWARA, N. & KOBAYASHI, K. 2005. Macrophages in inflammation. *Current Drug Targets-Inflammation and Allergy*, 4, 281-286.
- GAJOS, G., ZALEWSKI, J., MOSTOWIK, M., KONDURACKA, E., NESSLER, J. & UNDAS, A. 2014. Polyunsaturated omega-3 fatty acids reduce lipoprotein-associated phospholipase A(2) in patients with stable angina. *Nutrition, Metabolism, and Cardiovascular Diseases : NMCD*, 24, 434-439.
- GARDNER, D. M. 2002. Evidence-based decisions about herbal products for treating mental disorders. *Journal of Psychiatry and Neuroscience: JPN*, 27, 324.
- GHISLETTI, S., MEDA, C., MAGGI, A. & VEGETO, E. 2005. 17 β -estradiol inhibits inflammatory gene expression by controlling NF- κ B intracellular localization. *Molecular and Cellular Biology*, 25, 2957-2968.
- GHOSH, G., DUYNE, G. V., GHOSH, S. & SIGLER, P. B. 1995. Structure of NF- κ B p50 homodimer bound to a kB site. *Nature*, 373, 303-310.
- GILBERT-DIAMOND, D., BAYLIN, A., MORA-PLAZAS, M. & VILLAMOR, E. 2012. Chronic inflammation is associated with overweight in Colombian school children. *Nutrition, Metabolism and Cardiovascular Diseases*, 22, 244-251.
- GOONASEKERA, M. M., GUNAWARDANA, V. K., JAYASENA, K., MOHAMMED, S. G. & BALASUBRAMANIAM, S. 1995. Pregnancy terminating effect of *Jatropha curcas* in rats. *Journal of Ethnopharmacology*, 47, 117-123.
- GRAHAM, D. J., CAMPEN, D., HUI, R., SPENCE, M., CHEETHAM, C., LEVY, G., SHOOR, S. & RAY, W. A. 2005. Risk of acute myocardial infarction and sudden cardiac death in patients treated with cyclo-oxygenase 2 selective and non-selective non-steroidal anti-inflammatory drugs: nested case-control study. *The Lancet*, 365, 475-481.
- GRIFFITHS, M. & SUNDARAM, H. 2011. Drug design and testing: profiling of antiproliferative agents for cancer therapy using a cell-based methyl-[3H]-thymidine incorporation assay. *Methods in Molecular Biology*, 731, 451-465.

- GRIMMER, K., KUMAR, S., GILBERT, A. & MILANESE, S. 2002. Non-steroidal anti-inflammatory drugs (NSAIDs): Physiotherapists' use, knowledge and attitudes. *Australian Journal of Physiotherapy*, 48, 82-92.
- GRIVENNIKOV, S. I., GRETEN, F. R. & KARIN, M. 2010. Immunity, Inflammation, and Cancer. *Cell*, 140, 883-899.
- GULLIKSSON, M., BRUNNSTRÖM, Å., JOHANNESSON, M., BACKMAN, L., NILSSON, G., HARVIMA, I., DAHLÉN, B., KUMLIN, M. & CLAESSEN, H.-E. 2007. Expression of 15-lipoxygenase type-1 in human mast cells. *Biochimica et Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids*, 1771, 1156-1165.
- HÄMÄLÄINEN, M., KORHONEN, R. & MOILANEN, E. 2009. Calcineurin inhibitors down-regulate iNOS expression by destabilising mRNA. *International Immunopharmacology*, 9, 159-167.
- HANH, T. T. H., HANG, D. T. T., VAN MINH, C. & DAT, N. T. 2011. Anti-inflammatory effects of fatty acids isolated from Chromolaena odorata. *Asian Pacific Journal of Tropical Medicine*, 4, 760-763.
- HANSEL, A., HONG, S., CAMARA, R. J. & VON KANEL, R. 2010. Inflammation as a psychophysiological biomarker in chronic psychosocial stress. *Neuroscience and Biobehavioral Reviews*, 35, 115-21.
- HEDI, H. & NORBERT, G. 2004. 5-Lipoxygenase pathway, dendritic cells, and adaptive immunity. *BioMed Research International*, 2004, 99-105.
- HENDRA, R., AHMAD, S., OSKOUIEIAN, E., SUKARI, A. & SHUKOR, M. Y. 2011. Antioxidant, Anti-inflammatory and Cytotoxicity of *Phaleria macrocarpa* (Boerl.) Scheff Fruit. *BMC Complementary and Alternative Medicine*, 11, 110-110.
- HEO, S.-J., YOON, W.-J., KIM, K.-N., OH, C., CHOI, Y.-U., YOON, K.-T., KANG, D.-H., QIAN, Z.-J., CHOI, I.-W. & JUNG, W.-K. 2012. Anti-inflammatory effect of fucoxanthin derivatives isolated from *Sargassum siliquastrum* in lipopolysaccharide-stimulated RAW 264.7 macrophage. *Food and Chemical Toxicology*, 50, 3336-3342.
- HOWES, L. G. 2007. Selective COX-2 inhibitors, NSAIDs and cardiovascular events-is celecoxib the safest choice? *Therapeutics and clinical risk management*, 3, 831.
- HUANG, R.-Y. & CHEN, G. G. 2011. Cigarette smoking, cyclooxygenase-2 pathway and cancer. *Biochimica et Biophysica Acta (BBA) - Reviews on Cancer*, 1815, 158-169.

- HUH, J.-E., SEO, B.-K., PARK, Y.-C., KIM, J.-I., LEE, J.-D., CHOI, D.-Y., BAEK, Y.-H. & PARK, D.-S. 2012. WIN-34B, a new herbal medicine, inhibits the inflammatory response by inactivating I κ B- α phosphorylation and mitogen activated protein kinase pathways in fibroblast-like synoviocytes. *Journal of Ethnopharmacology*, 143, 779-786.
- ILIC, S., DRMIC, D., FRANJIC, S., KOLENC, D., CORIC, M., BRCIC, L., KLICEK, R., RADIC, B., SEVER, M., DJUZEL, V., FILIPOVIC, M., DJAKOVIC, Z., STAMBOLIJA, V., BLAGAIC, A. B., ZORICIC, I., GJURASIN, M., STUPNISEK, M., ROMIC, Z., ZARKOVIC, K., DZIDIC, S., SEIWERTH, S. & SIKIRIC, P. 2011. Pentadecapeptide BPC 157 and its effects on a NSAID toxicity model: Diclofenac-induced gastrointestinal, liver, and encephalopathy lesions. *Life Sciences*, 88, 535-542.
- IMAI, Y., KOLB, H. & BURKART, V. 1993. Nitric Oxide Production from Macrophages Is Regulated by Arachidonic Acid Metabolites. *Biochemical and Biophysical Research Communications*, 197, 105-109.
- IMPELLIZZERI, D., ESPOSITO, E., ATTLEY, J. & CUZZOCREA, S. 2014. Targeting inflammation: New therapeutic approaches in chronic kidney disease (CKD). *Pharmacological Research*, 81, 91-102.
- INGLE, P. V. & PATEL, D. M. 2011. C-reactive protein in various disease condition—an overview. *Asian Journal of Pharmaceutical and Clinical Research*, 4, 9-13.
- INOUE, N., WATANABE, M., NANBA, T., WADA, M., AKAMIZU, T. & IWATANI, Y. 2009. Involvement of functional polymorphisms in the TNFA gene in the pathogenesis of autoimmune thyroid diseases and production of anti-thyrotropin receptor antibody. *Clinical and Experimental Immunology*, 156, 199-204.
- IRAVANI, M. M., KASHEFI, K., MANDER, P., ROSE, S. & JENNER, P. 2002. Involvement of inducible nitric oxide synthase in inflammation-induced dopaminergic neurodegeneration. *Neuroscience*, 110, 49-58.
- ISLAM, M. N., CHOI, R. J., JIN, S. E., KIM, Y. S., AHN, B. R., ZHAO, D., JUNG, H. A. & CHOI, J. S. 2012. Mechanism of anti-inflammatory activity of umbelliferone 6-carboxylic acid isolated from *Angelica decursiva*. *Journal of Ethnopharmacology*, 144, 175-181.
- J. R. VANE, BAKHLE, Y. S. & BOTTING, R. M. 1998. Cyclooxygenases 1 and 2. *Annual Review of Pharmacology and Toxicology*, 38, 97-120.
- JIA, W., GAO, W.-Y., CUI, N.-Q. & XIAO, P.-G. 2003. Anti-inflammatory effects of an herbal medicine (Xuan-Ju agent) on carrageenan- and adjuvant-induced paw edema in rats. *Journal of Ethnopharmacology*, 89, 139-141.

- JIN, M., SUH, S.-J., YANG, J. H., LU, Y., KIM, S. J., KWON, S., JO, T. H., KIM, J. W., PARK, Y. I., AHN, G. W., LEE, C.-K., KIM, C.-H., SON, J.-K., SON, K. H. & CHANG, H. W. 2010. Anti-inflammatory activity of bark of *Dioscorea batatas* DECNE through the inhibition of iNOS and COX-2 expressions in RAW264.7 cells via NF-κB and ERK1/2 inactivation. *Food and Chemical Toxicology*, 48, 3073-3079.
- JOSHI, Y. B. & PRATICÒ, D. 2014. The 5-lipoxygenase pathway: oxidative and inflammatory contributions to the Alzheimer's disease phenotype. *Frontiers in Cellular Neuroscience*, 8, 436.
- JUNG, K. H., HA, E., KIM, M. J., WON, H.-J., ZHENG, L. T., KIM, H. K., HONG, S. J., CHUNG, J. H. & YIM, S.-V. 2007. Suppressive effects of nitric oxide (NO) production and inducible nitric oxide synthase (iNOS) expression by *Citrus reticulata* extract in RAW 264.7 macrophage cells. *Food and Chemical Toxicology*, 45, 1545-1550.
- KALUPAHANA, N. S., CLAYCOMBE, K. J. & MOUSTAID-MOUSSA, N. 2011. (n-3) Fatty acids alleviate adipose tissue inflammation and insulin resistance: mechanistic insights. *Advances in Nutrition: An International Review Journal*, 2, 304-316.
- KAUR, R., ARORA, S. & SINGH, B. 2008. Antioxidant activity of the phenol rich fractions of leaves of *Chukrasia tabularis* A. Juss. *Bioresource Technology*, 99, 7692-7698.
- KAZŁOWSKA, K., HSU, T., HOU, C. C., YANG, W. C. & TSAI, G. J. 2010. Anti-inflammatory properties of phenolic compounds and crude extract from *Porphyra dentata*. *Journal of Ethnopharmacology*, 128, 123-130.
- KIM, B. H., LEE, J., SHEN, T., KIM, J. D. & CHO, J. Y. 2010. Anti-inflammatory activity of hot water extract of *Berberis koreana* in lipopolysaccharide-induced macrophage-like cells. *Journal of Medicinal Plants Research*, 4, 745-752.
- KIM, S. S., OH, O. J., MIN, H.-Y., PARK, E.-J., KIM, Y., PARK, H. J., NAM HAN, Y. & LEE, S. K. 2003. Eugenol suppresses cyclooxygenase-2 expression in lipopolysaccharide-stimulated mouse macrophage RAW264.7 cells. *Life Sciences*, 73, 337-348.
- KOSASI, S., T HART, L. A., VAN DIJK, H. & LABADIE, R. P. 1989. Inhibitory activity of *Jatropha multifida* latex on classical complement pathway activity in human serum mediated by a calcium-binding proanthocyanidin. *Journal of Ethnopharmacology*, 27, 81-89.
- KULMACZ, R. J. 2005. Regulation of cyclooxygenase catalysis by hydroperoxides. *Biochemical and Biophysical Research Communications*, 338, 25-33.
- KUMAR, A. & SHARMA, S. 2008. An evaluation of multipurpose oil seed crop for industrial uses (*Jatropha curcas* L.): A review. *Industrial Crops and Products*, 28, 1-10.

- KUMAR, K. S., VIJAYAN, V., BHASKAR, S., KRISHNAN, K., SHALINI, V. & HELEN, A. 2012. Anti-inflammatory potential of an ethyl acetate fraction isolated from *Justicia gendarussa* roots through inhibition of iNOS and COX-2 expression via NF-κB pathway. *Cellular Immunology*, 272, 283-289.
- KURUMBAIL, R. G., KIEFER, J. R. & MARNETT, L. J. 2001. Cyclooxygenase enzymes: catalysis and inhibition. *Current Opinion in Structural Biology*, 11, 752-760.
- LAINE, L. 2002. The gastrointestinal effects of nonselective NSAIDs and COX-2-selective inhibitors. *Seminars in Arthritis and Rheumatism*, 32, 25-32.
- LAINE, L. 2003. Gastrointestinal Effects of NSAIDs and Coxibs. *Journal of Pain and Symptom Management*, 25, 32-40.
- LAINE, L., WHITE, W. B., ROSTOM, A. & HOCHBERG, M. 2008. COX-2 Selective Inhibitors in the Treatment of Osteoarthritis. *Seminars in Arthritis and Rheumatism*, 38, 165-187.
- LATIMER, N., LORD, J., GRANT, R. L., O'MAHONY, R., DICKSON, J. & CONAGHAN, P. G. 2009. Cost effectiveness of COX 2 selective inhibitors and traditional NSAIDs alone or in combination with a proton pump inhibitor for people with osteoarthritis. *British Medical Journal*, 339.
- LAWRENCE, T. 2009. The nuclear factor NF-κB pathway in inflammation. *Cold Spring Harbor Perspectives in Biology*, 1-10.
- LECHNER, M., LIRK, P. & RIEDER, J. 2005. Inducible nitric oxide synthase (iNOS) in tumor biology: The two sides of the same coin. *Seminars in Cancer Biology*, 15, 277-289.
- LEE, H.-S., BILEHAL, D., LEE, G.-S., RYU, D.-S., KIM, H.-K., SUK, D.-H. & LEE, D.-S. 2013. Anti-inflammatory effect of the hexane fraction from *Orostachys japonicus* in RAW 264.7 cells by suppression of NF-κB and PI3K-Akt signaling. *Journal of Functional Foods*, 5, 1217-1225.
- LEE, H.-S., RYU, D.-S., LEE, G.-S. & LEE, D.-S. 2012a. Anti-inflammatory effects of dichloromethane fraction from *Orostachys japonicus* in RAW 264.7 cells: Suppression of NF-κB activation and MAPK signaling. *Journal of Ethnopharmacology*, 140, 271-276.
- LEE, K.-H., AB. AZIZ, F. H., SYAHIDA, A., ABAS, F., SHAARI, K., ISRAF, D. A. & LAJIS, N. H. 2009. Synthesis and biological evaluation of curcumin-like diarylpentanoid analogues for anti-inflammatory, antioxidant and anti-tyrosinase activities. *European Journal of Medicinal Chemistry*, 44, 3195-3200.

- LEE, K.-H., CHOW, Y.-L., SHARMILI, V., ABAS, F., ALITHEEN, N. B. M., SHAARI, K., ISRAF, D. A., LAJIS, N. H. & SYAHIDA, A. 2012b. BDMC33, a curcumin derivative suppresses inflammatory responses in macrophage-like cellular system: Role of inhibition in NF- κ B and MAPK signaling pathways. *International Journal of Molecular Sciences*, 13, 2985-3008.
- LEIFELD, L., FIELENBACH, M., DUMOULIN, F.-L., SPEIDEL, N., SAUERBRUCH, T. & SPENGLER, U. 2002. Inducible nitric oxide synthase (iNOS) and endothelial nitric oxide synthase (eNOS) expression in fulminant hepatic failure. *Journal of Hepatology*, 37, 613-619.
- LI, Y., ZHANG, J. & MA, H. 2014. Chronic inflammation and gallbladder cancer. *Cancer Letters*, 345, 242-248.
- LIN, T.-H., TAMAKI, Y., PAJARINEN, J., WATERS, H. A., WOO, D. K., YAO, Z. & GOODMAN, S. B. 2014. Chronic inflammation in biomaterial-induced periprosthetic osteolysis: NF- κ B as a therapeutic target. *Acta Biomaterialia*, 10, 1-10.
- LINTON, M. F. & FAZIO, S. 2004. Cyclooxygenase-2 and inflammation in atherosclerosis. *Current Opinion in Pharmacology*, 4, 116-123.
- LIU, M. & YOKOMIZO, T. 2015. The role of leukotrienes in allergic diseases. *Allergology International*, 64, 17-26.
- LONG, L., SOEKEN, K. & ERNST, E. 2001. Herbal medicines for the treatment of osteoarthritis: a systematic review. *Rheumatology*, 40, 779-793.
- MÄKI-PETÄJÄ, K. M., CHERIYAN, J., BOOTH, A. D., HALL, F. C., BROWN, J., WALLACE, S. M. L., ASHBY, M. J., MCENIERY, C. M. & WILKINSON, I. B. 2008. Inducible nitric oxide synthase activity is increased in patients with rheumatoid arthritis and contributes to endothelial dysfunction. *International Journal of Cardiology*, 129, 399-405.
- MAKKAR, H. P. S., BECKER, K. & SCHMOOK, B. 1998. Edible provenances of *Jatropha curcas* from Quintana Roo state of Mexico and effect of roasting on antinutrient and toxic factors in seeds. *Plant Foods for Human Nutrition*, 52, 31-36.
- MAKARENKO, S.P., KONENKINA, T. A. & KHOTIMCHENKO, S. V. 2007. Fatty acid composition of lipids from the vacuolar membranes of the roots of root vegetables. *Russian Journal of Plant Physiology*, 54(2), 196-201.
- MALAVIYA, R., ANSELL, J., HALL, L., FAHMY, M., ARGENTIERI, R. L., OLINI JR, G. C., PEREIRA, D. W., SUR, R. & CAVENDER, D. 2006. Targeting cytosolic phospholipase A₂ by arachidonyl trifluoromethyl ketone prevents chronic inflammation in mice. *European Journal of Pharmacology*, 539, 195-204.

- MAMDANI, M., JUURLINK, D. N., LEE, D. S., ROCHON, P. A., KOPP, A., NAGLIE, G., AUSTIN, P. C., LAUPACIS, A. & STUKEL, T. A. 2004. Cyclooxygenase-2 inhibitors versus non-selective non-steroidal anti-inflammatory drugs and congestive heart failure outcomes in elderly patients: a population-based cohort study. *The Lancet*, 363, 1751-1756.
- MANDPE, S., KADLASKAR, S., DEGEN, W. & KEPPELER, S. 2005. On road testing of advanced common rail diesel vehicles with biodiesel from the *Jatropha curcas* plant. SAE Technical Paper.
- MARIAPPAN, N., SOORAPPAN, R. N., HAQUE, M., SRIRAMULA, S. & FRANCIS, J. 2007. TNF- α -induced mitochondrial oxidative stress and cardiac dysfunction: restoration by superoxide dismutase mimetic Tempol. *American Journal of Physiology - Heart and Circulatory Physiology*, 293, H2726-H2737.
- MARINOVA, D., RIBAROVA, F. & ATANASSOVA, M. 2005. Total phenolics and total flavonoids in Bulgarian fruits and vegetables. *Journal of the University of Chemical Technology and Metallurgy*, 40, 255-260.
- MARNETT, L. J. 2000. Cyclooxygenase mechanisms. *Current Opinion in Chemical Biology*, 4, 545-552.
- MATSUSE, I. T., LIM, Y. A., HATTORI, M., CORREA, M. & GUPTA, M. P. 1998. A search for anti-viral properties in Panamanian medicinal plants.: The effects on HIV and its essential enzymes. *Journal of Ethnopharmacology*, 64, 15-22.
- MAYER, L. & BHIKHA, R. 2013. The Challenging Response of Physis to Inflammation.
- MCGEER, P. L. & MCGEER, E. G. 2007. NSAIDs and Alzheimer disease: Epidemiological, animal model and clinical studies. *Neurobiology of Aging*, 28, 639-647.
- MCGILL, K. A. & BUSSE, W. W. 1996. Zileuton. *The Lancet*, 348, 519-524.
- MCMILLAN, K., ADLER, M., AULD, D. S., BALDWIN, J. J., BLASKO, E., BROWNE, L. J., CHELSKY, D., DAVEY, D., DOLLE, R. E., EAGEN, K. A., ERICKSON, S., FELDMAN, R. I., GLASER, C. B., MALLARI, C., MORRISSEY, M. M., OHLMEYER, M. H. J., PAN, G., PARKINSON, J. F., PHILLIPS, G. B., POLOKOFF, M. A., SIGAL, N. H., VERGONA, R., WHITLOW, M., YOUNG, T. A. & DEVLIN, J. J. 2000. Allosteric inhibitors of inducible nitric oxide synthase dimerization discovered via combinatorial chemistry. *Proceedings of the National Academy of Sciences*, 97, 1506-1511.
- MEDEIRO, R., FIGUEIREDO, C. P., PASSOS, G. F. & CALIXTO, J. B. 2009. Reduced skin inflammatory response in mice lacking inducible nitric oxide synthase. *Biochemical Pharmacology*, 78, 390-395.

- MEDIANI, A., ABAS, F., KHATIB, A., MAULIDIANI, H., SHAARI, K., CHOI, Y. H. & LAJIS, N. H. 2012. *1H-NMR-based metabolomics approach to understanding the drying effects on the phytochemicals in Cosmos caudatus*. *Food Research International*, 49, 763-770.
- MENSCHIKOWSKI, M., HAGELGANS, A. & SIEGERT, G. 2006. Secretory phospholipase A₂ of group IIA: Is it an offensive or a defensive player during atherosclerosis and other inflammatory diseases? *Prostaglandins and Other Lipid Mediators*, 79, 1-33.
- MENZIES, D., NAIR, A., MELDRUM, K. T., HOPKINSON, P. & LIPWORTH, B. J. 2008. Effect of aspirin on airway inflammation and pulmonary function in patients with persistent asthma. *Journal of Allergy and Clinical Immunology*, 121, 1184-1189.e4.
- MESKELL, M. & ETTARH, R. 2011. Immunohistochemical localisation of renal cyclooxygenase-1 expression in non-steroidal anti-inflammatory drug-treated mice. *Experimental and Toxicologic Pathology*, 63, 39-42.
- MILES, E. A., ZOUBOULI, P. & CALDER, P. C. 2005. Differential anti-inflammatory effects of phenolic compounds from extra virgin olive oil identified in human whole blood cultures. *Nutrition*, 21, 389-394.
- MITA, H., YUI, Y., TANIGUCHI, N., YASUEDA, H. & SHIDA, T. 1985. Increased activity of 5-lipoxygenase in polymorphonuclear leukocytes from asthmatic patients. *Life Sciences*, 37, 907-914.
- MORITA, I. 2002. Distinct functions of COX-1 and COX-2. *Prostaglandins & Other Lipid Mediators*, 68-69, 165-175.
- MUJUMDAR, A. M. & MISAR, A. V. 2004. Anti-inflammatory activity of *Jatropha curcas* roots in mice and rats. *Journal of Ethnopharmacology*, 90, 11-15.
- MURAKAMI, M. & LAMBEAU, G. 2013. Emerging roles of secreted phospholipase A₂ enzymes: An update. *Biochimie*, 95, 43-50.
- MURAKAMI, M., TAKETOMI, Y., GIRARD, C., YAMAMOTO, K. & LAMBEAU, G. 2010. Emerging roles of secreted phospholipase A₂ enzymes: Lessons from transgenic and knockout mice. *Biochimie*, 92, 561-582.
- NAGAHARIKA, Y., KALYANI, V., RASHEED, S. & RAMADOSSKARTHIKEYAN 2013. Anti-inflammatory activity of leaves of *Jatropha gossypifolia* L. by hrbc membrane stabilization method. *Journal of Acute Disease*, 2, 156-158.
- NAHAR, P. P., DRISCOLL, M. V., LI, L., SLITT, A. L. & SEERAM, N. P. 2014. Phenolic mediated anti-inflammatory properties of a maple syrup extract in RAW 264.7 murine macrophages. *Journal of Functional Foods*, 6, 126-136.

- NAIR, A. R., MASSON, G. S., EBENEZER, P. J., DEL PIERO, F. & FRANCIS, J. 2014. Role of TLR4 in lipopolysaccharide-induced acute kidney injury: Protection by blueberry. *Free Radical Biology and Medicine*, 71, 16-25.
- NAMSA, N. D., TAG, H., MANDAL, M., KALITA, P. & DAS, A. K. 2009. An ethnobotanical study of traditional anti-inflammatory plants used by the Lohit community of Arunachal Pradesh, India. *Journal of Ethnopharmacology*, 125, 234-245.
- NAMULI, A., ABDULLAH, N., SIEO, C., ZUHAINIS, S. & OSKOUETIAN, E. 2011. Phytochemical compounds and antibacterial activity of *Jatropha curcas* Linn. extracts.
- NAYAK, B. & PATEL, K. 2010. Anti-Inflammatory screening of *Jatropha curcas* root, stem and leaf in albino rats.
- NERI, M., CERRETANI, D., FIASCHI, A. I., LAGHI, P. F., LAZZERINI, P. E., MAFFIONE, A. B., MICHELI, L., BRUNI, G., NENCINI, C., GIORGI, G., D'ERRICO, S., FIORE, C., POMARA, C., RIEZZO, I., TURILLAZZI, E. & FINESCHI, V. 2007. Correlation between cardiac oxidative stress and myocardial pathology due to acute and chronic norepinephrine administration in rats. *Journal of Cellular and Molecular Medicine*, 11, 156-170.
- NIRANJAN, R., MANIK, R., SRIVASTAVA, A. K., PALIT, G. & NATU, S. M. 2011b. Cardiovascular Side Effect Remotely Related to NSAIDs: A Comparative Experimental Study on Albino Rats. *Journal of Anatomical Society of India*, 60, 155-159.
- NOBRE, M. E. P., CORREIA, A. O., BORGES, M. D. B., SAMPAIO, T. M. A., CHAKRABORTY, S. A., GONÇALVES, D. D. O., BRITO, G. A. D. C., LEAL, L. K. A. M., FELIPE, C. F. B., LUCETTI, D. L., ARIDA, R. M. & VIANA, G. S. D. B. 2013. Eicosapentaenoic acid and docosahexaenoic acid exert anti-inflammatory and antinociceptive effects in rodents at low doses. *Nutrition Research*, 33, 422-433.
- NUNKOO, D. H. & MAHOMOODALLY, M. F. 2012. Ethnopharmacological survey of native remedies commonly used against infectious diseases in the tropical island of Mauritius. *Journal of Ethnopharmacology*, 143, 548-564.
- O'CONNOR, J. C., ANDRÉ, C., WANG, Y., LAWSON, M. A., SZEGEDI, S. S., LESTAGE, J., CASTANON, N., KELLEY, K. W. & DANTZER, R. 2009. Interferon- γ and tumor necrosis factor- α mediate the upregulation of indoleamine 2, 3-dioxygenase and the induction of depressive-like behavior in mice in response to bacillus Calmette-Guérin. *The Journal of Neuroscience*, 29, 4200-4209.
- OECKINGHAUS, A. & GHOSH, S. 2009. The NF- κ B family of transcription factors and its regulation. *Cold Spring Harbor Perspectives in Biology*, 1-14.

- OJANO-DIRAIN, C., TOYOMIZU, M., WING, T., COOPER, M. & BOTTJE, W. G. 2007. Gene expression in breast muscle and duodenum from low and high feed efficient broilers. *Poultry Science*, 86, 372-381.
- ONG, H. C. & NORDIANA, M. 1999. Malay ethno-medico botany in Machang, Kelantan, Malaysia. *Fitoterapia*, 70, 502-513.
- ORGANIZATION, W. H. 2013. WHO Expert Consultation on Rabies. Second report. *World Health Organization technical report series*, 1.
- OSKOUIEIAN, E., ABDULLAH, N., AHMAD, S., SAAD, W. Z., OMAR, A. R. & HO, Y. W. 2011a. Bioactive Compounds and Biological Activities of *Jatropha curcas* L. Kernel Meal Extract. *International Journal of Molecular Sciences*, 12, 5955-5970.
- OSKOUIEIAN, E., ABDULLAH, N., SAAD, W. Z., OMAR, A. R., AHMAD, S., KUAN, W. B., ZOLKIFLI, N. A., HENDRA, R. & HO, Y. W. 2011b. Antioxidant, anti-inflammatory and anticancer activities of methanolic extracts from *Jatropha curcas* Linn. *Journal of Medicinal Plants Research*, 5, 49-57.
- PAN, M.-H., YANG, J.-R., TSAI, M.-L., SANG, S. & HO, C.-T. 2009. Anti-inflammatory effect of *Momordica grosvenori* Swingle extract through suppressed LPS-induced upregulation of iNOS and COX-2 in murine macrophages. *Journal of Functional Foods*, 1, 145-152.
- PANDEY, V. C., SINGH, K., SINGH, J. S., KUMAR, A., SINGH, B. & SINGH, R. P. 2012. *Jatropha curcas*: A potential biofuel plant for sustainable environmental development. *Renewable and Sustainable Energy Reviews*, 16, 2870-2883.
- PARK, J.-H., JEONG, Y.-J., WON, H. K., CHOI, S.-Y., PARK, J.-H. & OH, S.-M. 2014. Activation of TOPK by lipopolysaccharide promotes induction of inducible nitric oxide synthase through NF- κ B activity in leukemia cells. *Cellular Signalling*, 26, 849-856.
- PAUL, A. G., CHANDRAN, B. & SHARMA-WALIA, N. 2013. Cyclooxygenase-2-prostaglandin E2-eicosanoid receptor inflammatory axis: a key player in Kaposi's sarcoma-associated herpes virus associated malignancies. *Translational Research*, 162, 77-92.
- PAVANELLI, W. R., GUTIERREZ, F. R. S., MARIANO, F. S., PRADO, C. M., FERREIRA, B. R., TEIXEIRA, M. M., CANETTI, C., ROSSI, M. A., CUNHA, F. Q. & SILVA, J. S. 2010. 5-Lipoxygenase is a key determinant of acute myocardial inflammation and mortality during *Trypanosoma cruzi* infection. *Microbes and Infection*, 12, 587-597.
- PAWELEC, G., GOLDECK, D. & DERHOVANESSIAN, E. 2014. Inflammation, ageing and chronic disease. *Current Opinion in Immunology*, 29, 23-28.

- PERGOLA, C., JAZZAR, B., ROSSI, A., NORTHOFF, H., HAMBURGER, M., SAUTEBIN, L. & WERZ, O. 2012. On the inhibition of 5-lipoxygenase product formation by tryptanthrin: mechanistic studies and efficacy in vivo. *British Journal of Pharmacology*, 165, 765-776.
- PERTINO, M., SCHMEDA-HIRSCHMANN, G., RODRÍGUEZ, J. A. & THEODULOUZ, C. 2007. Gastroprotective effect and cytotoxicity of terpenes from the Paraguayan crude drug "yagua rova" (*Jatropha isabelli*). *Journal of Ethnopharmacology*, 111, 553-559.
- PETERS-GOLDEN, M. & BROCK, T. G. 2003. 5-Lipoxygenase and FLAP. *Prostaglandins, Leukotrienes and Essential Fatty Acids*, 69, 99-109.
- PRASAD, S., SUNG, B. & AGGARWAL, B. B. 2012. Age-associated chronic diseases require age-old medicine: Role of chronic inflammation. *Preventive Medicine*, 54, Supplement, S29-S37.
- PUNZI, L., SCANU, A., RAMONDA, R. & OLIVIERO, F. 2012. Gout as autoinflammatory disease: New mechanisms for more appropriated treatment targets. *Autoimmunity Reviews*, 12, 66-71.
- QIU, H., GABRIELSEN, A., AGARDH, H. E., WAN, M., WETTERHOLM, A., WONG, C.-H., HEDIN, U., SWEDENBORG, J., HANSSON, G. K. & SAMUELSSON, B. 2006. Expression of 5-lipoxygenase and leukotriene A4 hydrolase in human atherosclerotic lesions correlates with symptoms of plaque instability. *Proceedings of the National Academy of Sciences*, 103, 8161-8166.
- RAGHUPATHI, R. & FRANSON, R. C. 1992. Inhibition of phospholipase A₂ by c/s-unsaturated fatty acids: evidence for the binding of fatty acid to enzyme. *Biochimica et Biophysica Acta*, 1126, 206-214.
- RAJESWAR, G., MURUGAN, M. & MOHAN, V. 2013. GC-MS analysis of bioactive components of *Hugonia mystax* L.(Linaceae). *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 3, 301-308.
- RAZ, A. 2002. Is inhibition of cyclooxygenase required for the anti-tumorigenic effects of nonsteroidal, anti-inflammatory drugs (NSAIDs)?: In vitro versus in vivo results and the relevance for the prevention and treatment of cancer. *Biochemical Pharmacology*, 63, 343-347.
- REDDY, T. R. P., RAO, R. S. V., SWAMY, A., REDDANNA, P., REDDY, G. P. & REDDY, D. R. 2003. Exploring the Anti-inflammatory and Anti-cancer compounds from the leaves of *Acalypha indica*. *Biophysical Research Communications*, 304, 385-392.
- RICCIOTTI, E. & FITZGERALD, G. A. 2011. Prostaglandins and Inflammation. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 31, 986-1000.

- RIEDL, K., KRYSAN, K., PÖLD, M., DALWADI, H., HEUZE-VOURC'H, N., DOHADWALA, M., LIU, M., CUI, X., FIGLIN, R., MAO, J. T., STRIETER, R., SHARMA, S. & DUBINETT, S. M. 2004. Multifaceted roles of cyclooxygenase-2 in lung cancer. *Drug Resistance Updates*, 7, 169-184.
- RIZZO, M. T. 2011. Cyclooxygenase-2 in oncogenesis. *Clinica Chimica Acta*, 412, 671-687.
- ROBINSON, M. A., BAUMGARDNER, J. E. & OTTO, C. M. 2011. Oxygen-dependent regulation of nitric oxide production by inducible nitric oxide synthase. *Free Radical Biology and Medicine*, 51, 1952-1965.
- RUBIN, P. & MOLLISON, K. W. 2007. Pharmacotherapy of diseases mediated by 5-lipoxygenase pathway eicosanoids. *Prostaglandins & Other Lipid Mediators*, 83, 188-197.
- SABANDAR, C. W., AHMAT, N., JAAFAR, F. M. & SAHIDIN, I. 2013. Medicinal property, phytochemistry and pharmacology of several *Jatropha* species (Euphorbiaceae): A review. *Phytochemistry*, 85, 7-29.
- SANCHEZ-MATIENZO, D., ARANA, A., CASTELLSAGUE, J. & PEREZ-GUTTHANN, S. 2006. Hepatic disorders in patients treated with COX-2 selective inhibitors or nonselective NSAIDs: A case/noncase analysis of spontaneous reports. *Clinical Therapeutics*, 28, 1123-1132.
- SEGEV, G. & KATZ, R. J. 2004. Selective COX-2 inhibitors and risk of cardiovascular events. *Hospital Physician*, 40, 39-46.
- SEOW, L.-J., BEH, H.-K., UMAR, M. I., SADIKUN, A. & ASMAWI, M. Z. 2014. Anti-inflammatory and antioxidant activities of the methanol extract of *Gynura segetum* leaf. *International Immunopharmacology*, 23, 186-191.
- SERGENT, T., PIRONT, N., MEURICE, J., TOUSSAINT, O. & SCHNEIDER, Y.-J. 2010. Anti-inflammatory effects of dietary phenolic compounds in an in vitro model of inflamed human intestinal epithelium. *Chemico-Biological Interactions*, 188, 659-667.
- SHARMA, S., DHAMIJA, H. K. & PARASHAR, B. 2012. *Jatropha curcas*: a review. *Asian Journal of Research in Pharmaceutical Science*, 2, 107-111.
- SHEN, J., GAN, L., ZHU, C., ZHANG, X., DONG, Y., JIANG, M., ZHU, J. & GAN, Y. 2011. Novel NSAIDs ophthalmic formulation: Flurbiprofen axetil emulsion with low irritancy and improved anti-inflammation effect. *International Journal of Pharmaceutics*, 412, 115-122.
- SHUKLA, S., KUMAR, A., BAHADUR, L. & PAL, M. 2015. Fatty acid composition of *Sonchus arvensis* L. roots. *Indian Journal of Natural Products and Resources*, 6(1), 62-64.

- SIEBUHR, A. S., PETERSEN, K. K., ARENDT-NIELSEN, L., EGSGAARD, L. L., ESKEHAVE, T., CHRISTIANSEN, C., SIMONSEN, O., HOECK, H. C., KARSDAL, M. A. & BAY-JENSEN, A. C. 2014. Identification and characterisation of osteoarthritis patients with inflammation derived tissue turnover. *Osteoarthritis and Cartilage*, 22, 44-50.
- SILVERSTEIN, F. E., FAICH, G., GOLDSTEIN, J. L., SIMON, L. S., PINCUS, T., WHELTON, A., MAKUCH, R., EISEN, G., AGRAWAL, N. M. & STENSON, W. F. 2000. Gastrointestinal toxicity with celecoxib vs nonsteroidal anti-inflammatory drugs for osteoarthritis and rheumatoid arthritis: the CLASS study: a randomized controlled trial. *Journal of the American Medical Association (JAMA)*, 284, 1247-1255.
- SINATRA, R. 2002. Role of COX-2 Inhibitors in the Evolution of Acute Pain Management. *Journal of Pain and Symptom Management*, 24, S18-S27.
- SIROOS, B., BALOOD, M., ZAHEDNASAB, H., MESBAH-NAMIN, S. A., POURGHOLY, F. & HARIRCHIAN, M. H. 2013. Secretory Phospholipase A₂ activity in serum and cerebrospinal fluid of patients with relapsing-remitting multiple sclerosis. *Journal of Neuroimmunology*, 262, 125-127.
- SLINKARD, K. & SINGLETON, V. L. 1977. Total phenol analysis: automation and comparison with manual methods. *American Journal of Enology and Viticulture*, 28, 49-55.
- SMITH, C. J., ZHANG, Y., KOBOLDT, C. M., MUHAMMAD, J., ZWEIFEL, B. S., SHAFFER, A., TALLEY, J. J., MASFERRE, J. L., SEIBERT, K. & ISAKSON, P. C. 1998. Pharmacological analysis of cyclooxygenase-1 in inflammation. *Proceedings of the National Academy of Sciences*, 95, 13313-13318.
- SOSTRES, C., GARGALLO, C. J., ARROYO, M. T. & LANAS, A. 2010. Adverse effects of non-steroidal anti-inflammatory drugs (NSAIDs, aspirin and coxibs) on upper gastrointestinal tract. *Best Practice and Research Clinical Gastroenterology*, 24, 121-132.
- SOUZA, L. P., ALESSANDRI, A. L., PINHO, V. & TEIXEIRA, M. M. 2013. Pharmacological strategies to resolve acute inflammation. *Current Opinion in Pharmacology*, 13, 625-631.
- SOUTO, A. L., TAVARES, J. F., DA SILVA, M. S., DINIZ, M. D. F. F. M., DE ATHAYDE-FILHO, P. F. & BARBOSA FILHO, J. M. 2011. Anti-inflammatory activity of alkaloids: An update from 2000 to 2010. *Molecules*, 16, 8515-8534.

- SPANBROEK, R., GRÄBNER, R., LÖTZER, K., HILDNER, M., URBACH, A., RÜHLING, K., MOOS, M. P. W., KAISER, B., COHNERT, T. U., WAHLERS, T., ZIESKE, A., PLENZ, G., ROBENEK, H., SALBACH, P., KÜHN, H., RÅDMARK, O., SAMUELSSON, B. & HABENICHT, A. J. R. 2003. Expanding expression of the 5-lipoxygenase pathway within the arterial wall during human atherogenesis. *Proceedings of the National Academy of Sciences*, 100, 1238-1243.
- STACHOWSKA, E., DOLEGOWSKA, B., DZIEDZIEJKO, V., RYBICKA, M., KACZMARCZYK, M., BOBER, J., RAC, M., MACHALINSKI, B. & CHLUBEK, D. 2009. Prostaglandin E₂ (PGE₂) and thromboxane A₂ (TXA₂) synthesis is regulated by conjugated linoleic acids (CLA) in human macrophages. *Acta physiologica Polonica*, 60, 77.
- STAUBMANN, R., FOIDL, G., FOIDL, N., GÜBITZ, G., LAFFERTY, R., ARBIZU, V. V. & STEINER, W. 1997. Biogas Production from Jatropha curcas Press-Cake. In: DAVISON, B., WYMAN, C. & FINKELSTEIN, M. (eds.) *Biotechnology for Fuels and Chemicals*. Humana Press.
- STEINHILBER, D. 1994. 5-lipoxygenase: enzyme expression and regulation of activity. *Pharmaceutica Acta Helvetiae*, 69, 3-14.
- SUBBARAYAN, V., XU, X.-C., KIM, J., YANG, P., HOQUE, A., SABICHI, A. L., LLANSA, N., MENDOZA, G., LOGOTHETIS, C. J., NEWMAN, R. A., LIPPMAN, S. M. & MENTER, D. G. 2005c. Inverse Relationship between 15-Lipoxygenase-2 and PPAR- γ Gene Expression in Normal Epithelia Compared with Tumor Epithelia. *Neoplasia*, 7, 280-293.
- SUN, Y., DUAN, Y., EISENSTEIN, A. S., HU, W., QUINTANA, A., LAM, W. K., WANG, Y., WU, Z., RAVID, K. & HUANG, P. 2012. A novel mechanism of control of NF κ B activation and inflammation involving A2B adenosine receptors. *Journal of cell science*, 125, 4507-4517.
- SUNIL, V. R., SHEN, J., PATEL-VAYAS, K., GOW, A. J., LASKIN, J. D. & LASKIN, D. L. 2012. Role of reactive nitrogen species generated via inducible nitric oxide synthase in vesicant-induced lung injury, inflammation and altered lung functioning. *Toxicology and Applied Pharmacology*, 261, 22-30.
- SYAHIDA, A., ISRAF, D. A., LAJIS, N. H., KHOZIRAH, S., HABSAH, M., JASRIL, PERMANA, D. & NORHADIANI, I. 2006. Effect of Compounds Isolated from Natural Products on IFN- γ /LPS-Induced Nitric Oxide Production in RAW 264.7 Macrophages. *Pharmaceutical Biology*, 44, 50-59.
- SYAM, S., BUSTAMAM, A., ABDULLAH, R., SUKARI, M. A., HASHIM, N. M., MOHAN, S., LOOI, C. Y., WONG, W. F., YAHDYU, M. A. & ABDELWAHAB, S. I. 2014. β Mangostin suppress LPS-induced inflammatory response in RAW 264.7 macrophages in vitro and carrageenan-induced peritonitis in vivo. *Journal of Ethnopharmacology*, 153, 435-445.

- TABART, J., KEVERS, C., SIPEL, A., PINCEMAIL, J., DEFRAIGNE, J.-O. & DOMMES, J. 2007. Optimisation of extraction of phenolics and antioxidants from black currant leaves and buds and of stability during storage. *Food Chemistry*, 105, 1268-1275.
- TAN, A. C., HOU, D.-X., KONCZAK, I., TANIGAWA, S., RAMZAN, I. & SZE, D. M. Y. 2011. Native Australian fruit polyphenols inhibit COX-2 and iNOS expression in LPS-activated murine macrophages. *Food Research International*, 44, 2362-2367.
- TANAKA, K.-I., SUEMASU, S., ISHIHARA, T., TASAKA, Y., ARAI, Y. & MIZUSHIMA, T. 2009. Inhibition of both COX-1 and COX-2 and resulting decrease in the level of prostaglandins E₂ is responsible for non-steroidal anti-inflammatory drug (NSAID)-dependent exacerbation of colitis. *European Journal of Pharmacology*, 603, 120-132.
- THIEBLEMONT, C., BERTONI, F., COPIE-BERGMAN, C., FERRERI, A. J. M. & PONZONI, M. 2014. Chronic inflammation and extra-nodal marginal-zone lymphomas of MALT-type. *Seminars in Cancer Biology*, 24, 33-42.
- TÓTH, L., MUSZBEK, L. & KOMÁROMI, I. 2013. Mechanism of the irreversible inhibition of human cyclooxygenase-1 by aspirin as predicted by QM/MM calculations. *Journal of Molecular Graphics and Modelling*, 40, 99-109.
- TRASK, O. J. 2004. Nuclear Factor Kappa B (NF-κB) Translocation Assay Development and Validation for High Content Screening. *Assay Guidance Manual*. Eli Lilly & Company and the National Center for Advancing Translational Sciences, Bethesda (MD).
- TSAI, C.-Y., YU, C.-L., WU, T.-H., HSIEH, S.-C. & TSAI, Y.-Y. 2004. Proinflammatory cytokines enhance COX-1 gene expression in cultured rat glomerular mesangial cells. *International Immunopharmacology*, 4, 47-56.
- UTAR, Z., MAJID, M. I. A., ADENAN, M. I., JAMIL, M. F. A. & LAN, T. M. 2011. Mitragynine inhibits the COX-2 mRNA expression and prostaglandin E₂ production induced by lipopolysaccharide in RAW264.7 macrophage cells. *Journal of Ethnopharmacology*, 136, 75-82.
- VAN DE WEERT-VAN LEEUWEN, P. B., HULZEBOS, H. J., WERKMAN, M. S., MICHEL, S., VIJFTIGSCHILD, L. A. W., VAN MEEGEN, M. A., VAN DER ENT, C. K., BEEKMAN, J. M. & ARETS, H. G. M. 2014. Chronic inflammation and infection associate with a lower exercise training response in cystic fibrosis adolescents. *Respiratory Medicine*, 108, 445-452.
- VAN DEN BERG, A. J. J., HORSTEN, S. F. A. J., KETTENES-VAN DEN BOSCH, J. J., KROES, B. H., BEUKELMAN, C. J., LEEFLANG, B. R. & LABADIE, R. P. 1995. Curcacycline A — a novel cyclic octapeptide isolated from the latex of *Jatropha curcas* L. *Federation of European Biochemical Societies (FEBS) Letters*, 358, 215-218.

- VAN ESCH, R. W., KOOL, M. M. & VAN AS, S. 2013. NSAIDs can have adverse effects on bone healing. *Medical Hypotheses*, 81, 343-346.
- VENKATESHA, S. H., BERMAN, B. M. & MOUDGIL, K. D. 2011. Herbal medicinal products target defined biochemical and molecular mediators of inflammatory autoimmune arthritis. *Bioorganic and Medicinal Chemistry*, 19, 21-29.
- VERBURG-VAN KEMENADE, B. M. L., VAN DER AA, L. M. & CHADZINSKA, M. 2013. Neuroendocrine-immune interaction: Regulation of inflammation via G-protein coupled receptors. *General and Comparative Endocrinology*, 188, 94-101.
- VICTORIEN, D. T., JEAN ROBERT, K., JACQUES, D. T., JULIEN, S., JEAN-MARC, A., ALÉODJRODO, E. P., OLUFUNKÈ, S., FERDINAND, D., CARLOS, D., FRÉDÉRIC, L. & KARIM, D. 2012. Hemostatic activity screening and skin toxicity of sap of *Jatropha multifida* L. (Euphorbiaceae) used in traditional medicine (Benin). *Asian Pacific Journal of Tropical Disease*, 2, Supplement 2, S927-S932.
- VILLEGRAS, L. F., FERNÁNDEZ, I. D., MALDONADO, H., TORRES, R., ZAVALETÀ, A., VAISBERG, A. J. & HAMMOND, G. B. 1997. Evaluation of the wound-healing activity of selected traditional medicinal plants from Perú. *Journal of Ethnopharmacology*, 55, 193-200.
- VISWANATHAN, M. B. G., JEYA ANANTHI, J. D. & SATHISH KUMAR, P. 2012. Antimicrobial activity of bioactive compounds and leaf extracts in *Jatropha tanjorensis*. *Fitoterapia*, 83, 1153-1159.
- VOGL, S., PICKER, P., MIHALY-BISON, J., FAKHRUDIN, N., ATANASOV, A. G., HEISS, E. H., WAWROSCH, C., REZNICEK, G., DIRSCH, V. M., SAUKEL, J. & KOPP, B. 2013. Ethnopharmacological in vitro studies on Austria's folk medicine—An unexplored lore in vitro anti-inflammatory activities of 71 Austrian traditional herbal drugs. *Journal of Ethnopharmacology*, 149, 750-771.
- VONKEMAN, H. E. & VAN DE LAAR, M. A. F. J. 2010. Nonsteroidal Anti-Inflammatory Drugs: Adverse Effects and Their Prevention. *Seminars in Arthritis and Rheumatism*, 39, 294-312.
- WALSH, L. S., OLLENDORFF, A. & MERSHON, J. L. 2003. Estrogen increases inducible nitric oxide synthase gene expression. *American Journal of Obstetrics and Gynecology*, 188, 1208-1210.
- WANG, D. & DUBOIS, R. N. 2008. Pro-inflammatory prostaglandins and progression of colorectal cancer. *Cancer Letters*, 267, 197-203.
- WANG, Q., KUANG, H., SU, Y., SUN, Y., FENG, J., GUO, R. & CHAN, K. 2013. Naturally derived anti-inflammatory compounds from Chinese medicinal plants. *Journal of Ethnopharmacology*, 146, 9-39.

- WANG, T. & XIA, Y. 2012. Inducible nitric oxide synthase aggresome formation is mediated by nitric oxide. *Biochemical and Biophysical Research Communications*, 426, 386-389.
- WEIR, M. 2002. Renal effects of nonselective NSAIDs and coxibs. *Cleveland Clinic Journal of Medicine*, 69, SI53.
- WONG, Y. F., ZHOU, H., WANG, J. R., XIE, Y., XU, H. X. & LIU, L. 2008. Anti-inflammatory and analgesic effects and molecular mechanisms of JCICM-6, a purified extract derived from an anti-arthritis Chinese herbal formula. *Phytomedicine*, 15, 416-426.
- WU, K. K. 1996. Cyclooxygenase 2 induction: Molecular mechanism and pathophysiologic roles. *Journal of Laboratory and Clinical Medicine*, 128, 242-245.
- WU, T. 2005. Cyclooxygenase-2 and prostaglandin signaling in cholangiocarcinoma. *Biochimica et Biophysica Acta (BBA) - Reviews on Cancer*, 1755, 135-150.
- WU, Y., ANTONY, S., MEITZLER, J. L. & DOROSHOW, J. H. 2014. Molecular mechanisms underlying chronic inflammation-associated cancers. *Cancer Letters*, 345, 164-173.
- YAM, M. F., ANG, L. F., AMEER, O. Z., SALMAN, I. M., AZIZ, H. A. & ASMAWI, M. Z. 2009. Anti-inflammatory and Analgesic Effects of *Elephantopus tomentosus* Ethanolic Extract. *Journal of Acupuncture and Meridian Studies*, 2, 280-287.
- YATES, C. M., CALDER, P. C. & ED RANGER, G. 2014. Pharmacology and therapeutics of omega-3 polyunsaturated fatty acids in chronic inflammatory disease. *Pharmacology and Therapeutics*, 141, 272-282.
- YAYLI, N., KIRAN, Z., SEYMEN, H., GENÇ, H. 2001. Characterization of Lipids and Fatty Acid Methyl Ester Contents in Leaves and Roots of *Crocus vallicola*. *Turkish Journal of Chemistry*, 25, 391-395.
- YEN, S. S. & MORRIS, H. G. 1981. An imbalance of arachidonic acid metabolism in asthma. *Biochemical and Biophysical Research Communications*, 103, 774-779.
- ZEIDLER, P. C., MILLECCHIA, L. M. & CASTRANOVA, V. 2004. Role of inducible nitric oxide synthase-derived nitric oxide in lipopolysaccharide plus interferon- γ -induced pulmonary inflammation. *Toxicology and Applied Pharmacology*, 195, 45-54.
- ZHANG, C., YU, H., NI, X., SHEN, S. & DAS, U. N. 2015. Growth Inhibitory Effect of Polyunsaturated Fatty Acids (PUFAs) on Colon Cancer Cells via Their Growth Inhibitory Metabolites and Fatty Acid Composition Changes. *PLoS ONE*, 1-18.

- ZHAO, G., ETHERTON, T. D., MARTIN, K. R., VANDEN HEUVEL, J. P., GILLIES, P. J., WEST, S. G. & KRIS-ETHERTON, P. M. 2005. Anti-inflammatory effects of polyunsaturated fatty acids in THP-1 cells. *Biochemical and Biophysical Research Communications*, 336, 909-917.
- ZHONG, H.-M., DING, Q.-H., CHEN, W.-P. & LUO, R.-B. 2013. Vorinostat, a HDAC inhibitor, showed anti-osteoarthritic activities through inhibition of iNOS and MMP expression, p38 and ERK phosphorylation and blocking NF- κ B nuclear translocation. *International Immunopharmacology*, 17, 329-335.