



**UNIVERSITI PUTRA MALAYSIA**

***IN VIVO AND IN VITRO EFFECTS OF PHALERIA MACROCARPA  
(SCHEFF.) BOERL ON LOW DENSITY LIPOPROTEIN AND PCSK9  
EXPRESSION***

**CHONG SOO CHING**

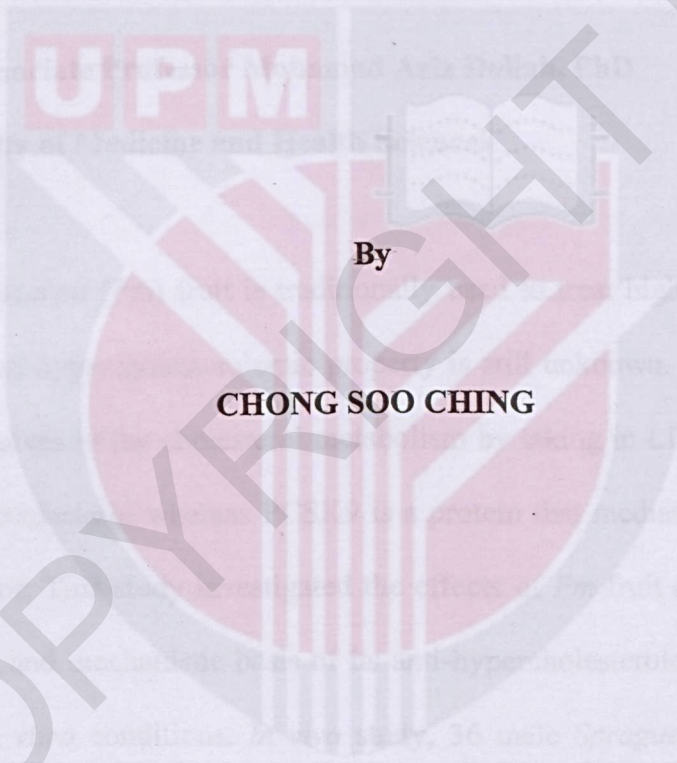
**FPSK(m) 2011 57**

**IN VIVO AND IN VITRO EFFECTS OF *PHALERIA MACROCARPA*  
(SCHEFF.) BOERL ON LOW DENSITY LIPOPROTEIN AND PCSK9  
EXPRESSION**

By

CHONG SOO CHING

December 2011



By

CHONG SOO CHING

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfillment of the Requirements for the Degree of Master of Science**

**December 2011**

Abstract of thesis presented to the Senate of University Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

***IN VIVO* AND *IN VITRO* EFFECTS OF *PHALERIA MACROCARPA* (SCHEFF.) BOERL ON LOW DENSITY LIPOPROTEIN AND PCSK9 EXPRESSION**

By

**CHONG SOO CHING**

**December 2011**

**Chairman: Associate Professor Mohamad Aziz Dollah, PhD**

**Faculty: Faculty of Medicine and Health Sciences**

*Phaleria macrocarpa* (*Pm*) fruit is traditionally used to treat high cholesterol level. However its anti-hypercholesterolemic property is still unknown. LDL receptor is a ligand that involves in the cholesterol metabolism by taking in LDL which has high proportion of cholesterol whereas PCSK9 is a protein that mediates the degradation of LDL receptor. This study investigated the effects of *Pm* fruit aqueous extract on weight control and mechanistic basis of its anti-hypercholesterolemic effect in both *in vivo* and *in vitro* conditions. *In vivo* study, 36 male *Sprague dawley* rats were randomized to 6 groups. 5 groups were given 3% (v/v) cholesterol enriched-diet for 52 days to induced to become hypercholesterolemia, followed by *Pm* fruit aqueous extract (0, 20, 30 and 40 mg extract/kg bw) or simvastatin (40 mg/kg) treatment for 84 days. The sixth group was used as a negative control. The effects of *Pm* fruit aqueous extract treatment were determined on the following parameters: 1) body weight, 2) liver weight 3) liver weight-to-body weight ratio 4) blood lipid profiles (TC, TG, HDL and LDL) and 5) expression level of hepatic LDL receptor (160 kDa

and 120 kDa) and PCSK9 proteins. *Pm* fruit aqueous extract significantly ( $P<0.05$ ) reduced body weight gain but tends to reduce liver weight and liver weight-to-body weight ratio. As for the blood lipid profiles, 20 mg extract/ kg bw of *Pm* significantly ( $P<0.05$ ) reduced TC (1.54 mmol/L), TG (0.38 mmol/L), HDL (0.68 mmol/L) and LDL (0.94 mmol/L) whereas 30 mg extract/ kg bw of *Pm* significantly ( $P<0.05$ ) reduced TC (1.55 mmol/L), TG (0.33 mmol/L) and LDL (0.93 mmol/L) as compared to the untreated hypercholesterolemic group [TC (2.4 mmol/L), TG (1.13 mmol/L), HDL (0.94 mmol/L) and LDL (1.51 mmol/L)]. 40 mg extract/ kg bw of *Pm* significantly ( $P<0.05$ ) reduced TC (1.85 mmol/L) and LDL (1.03 mmol/L). On the other hand, 20 mg extract/ kg bw of *Pm* significantly ( $P<0.05$ ) increased LDL receptor and PCSK9 proteins by 1-fold whereas 30 and 40 mg extract/ kg bw of *Pm* had no effect on LDL receptor and PCSK9. Effect of *Pm* fruit aqueous extract in *in vivo* model was then further analyzed in *in vitro* study. *In vitro* study, HepG2 cells were cultured in serum-free RPMI 1640, supplemented with 0.2% BSA with or without LDL (200  $\mu$ M) and in the presence of *Pm* fruit aqueous extract (0, 0.1, 2, 40 and 1000  $\mu$ g/ml) or simvastatin (10  $\mu$ M) for 24 hours. The abundance of both LDL receptor (160 kDa) and PCSK9 proteins and mRNA were then investigated. Similar to the *in vivo* study, *Pm* fruit aqueous extract was found to have increased LDL receptor and PCSK9 proteins by 1-fold in HepG2 cells with significant increment ( $P<0.05$ ) at the concentration of 0.1  $\mu$ g/ml. Besides that, *Pm* fruit aqueous extract at the concentration of 0.1  $\mu$ g/ml also significantly ( $P<0.05$ ) increased both LDL receptor and PCSK9 mRNA transcripts, comparable to simvastatin treated group. These study indicated that *Pm* fruit aqueous extract reduces body weight gain, liver weight, liver weight-to-body weight ratio and exhibited anti-hypercholesterolemic

effect by reducing blood lipid profile of hypercholesterolemic rats and upregulating LDL receptor and PCSK9 at both protein and mRNA level.

IN VIVO DAN IN VITRO KESAN BUAH PADI (Pennisetum purpureum L. SP. HYBRIDUM) (SCHEFF.) BOERL. PADA LIPOPROTEIN BERKECENDERUNGAN RENDAH DAN PLASMA

CHONG SHIH JIAH

December 2011

Program: Program Sarjana Sains (Sarjana Sains)

Fakulti: Fakulti Sains (Fakulti Sains)

Buah *Pennisetum purpureum* L. SP. HYBRIDUM (Padi) adalah salah satu jenis anti-hiperkolesterol tetapi mekanisme bagaimana ia bertindak terhadap kolesterol masih tidak diketahui. Receptor LDL membantu mengikat LDL untuk dipecahkan oleh enzim lipoprotein lipase manakala PCSK9 mengawal jumlah LDL. Kajian *in vivo* dan *in vitro* telah dijalankan untuk mengkaji mekanisme anti-hiperkolesterolisme buah Padi. Buah Padi 36 sampel yang berbeza telah dibahagikan kepada 6 kumpulan dan semua di beri makanan yang mengandungi tambahan 3% kolesterol selama 4 minggu untuk dalam menjadi hiperkolesterolik. 5 kumpulan akan diberi ekstrak ekstrak buah Padi (0, 20, 30 dan 40 mg ekstrak/kg) atau air masak secara oral selama 14 hari. Kumpulan ekstrak buah Padi kemudian dibandingkan dengan mengikut beberapa parameter seperti berikut: 1) berat badan, 2) berat hati, 3) berat berat hati kepada berat badan, 4) profil lipid (TC, TG, HDL, LDL) dan 5) kandungan receptor LDL (130 kDa dan 120 kDa) dan PCSK9 hati. Kajian ekstrak buah Padi menunjukkan berat badan, lemak (P<0.05), berat hati dan

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

***IN VIVO* DAN *IN VITRO* KESAN BUAH *PHALERIA MACROCARPA* (SCHEFF.) BOERL PADA LIPOPROTEIN BERKETUMPATAN RENDAH DAN PCSK9**

Oleh

**CHONG SOO CHING**

**Disember 2011**

**Pengerusi: Profesor Madya Mohamad Aziz Dollah, PhD**

**Fakulti: Fakulti Perubatan dan Sains Kesihatan**

Buah *Phaleria macrocarpa* (*Pm*) adalah ubat tradisional untuk anti-hiperkolesterol tetapi mekanisme tindakan anti-hiperkolesterolemianya masih tidak diketahui. Reseptor LDL membantu pengambilan LDL oleh sel hepar untuk didegradasi manakala PCSK9 mengdegradasi reseptor LDL. Kajian *in vivo* dan *in vitro* telah dijalankan untuk menentukan mekanisme anti-hiperkolesterolemia akues ekstrak buah *Pm*. 36 ekor *Sprague dawley* tikus jantan dibahagikan kepada 6 kumpulan dimana 5 kumpulan di beri makanan yang mengandungi tambahan 3% kolesterol selama 52 hari untuk diaruh menjadi hiperkolesterolemia. 5 kumpulan tikus itu kemudian diberi akues ekstrak buah *Pm* (0, 20, 30 dan 40 mg ekstrak/kg) atau simvastatin secara oral selama 84 hari. Kesan akues ekstrak buah *Pm* kemudian ditentukan dengan mengukur beberapa parameter seperti berikut: 1) berat badan, 2) berat hepar, 3) nisbah berat hepar kepada berat badan, 4) profil lipid (TC, TG, HDL LDL) dan 5) konsentrasi reseptor LDL (160 kDa dan 120 kDa) dan PCSK9 hepar. Akues ekstrak buah *Pm* menurunkan berat badan tikus ( $P < 0.05$ ), berat hepar dan

nisbah berat hepar kepada berat badan. Selain daripada itu, 20 mg ekstrak/ kg buah *Pm* menurunkan ( $P<0.05$ ) profil lipid TC tikus hiperkolesterolemia kepada 1.54 mmol/L, TG kepada 0.38 mmol/L, HDL kepada 0.68 mmol/L dan LDL kepada 0.94 mmol/L berbanding dengan tikus hiperkolesterolemik yang tidak diberi ekstrak buah *Pm* [TC (2.4 mmol/L), TG (1.13 mmol/L), HDL (0.94 mmol/L) and LDL (1.51 mmol/L)]. 30 mg ekstrak/ kg buah *Pm* pula menurunkan ( $P<0.05$ ) profil lipid TC tikus hiperkolesterolemia kepada 1.55 mmol/L, TG kepada 0.33 mmol/L and LDL kepada 0.93 mmol/L manakala 40 mg ekstrak/ kg buah *Pm* pula menurunkan ( $P<0.05$ ) profil lipid TC tikus hiperkolesterolemia kepada 1.85 mmol/L dan LDL kepada 1.03 mmol/L. Selain itu, 20 mg ekstrak/ kg buah *Pm* meningkatkan paras reseptor LDL (160 kDa dan 120 kDa) dan PCSK9 ( $P<0.05$ ) sebanyak satu kali ganda manakala 30 mg ekstrak/ kg dan 40 mg ekstrak/kg buah *Pm* pula tidak mempunyai apa-apa kesan terhadap reseptor LDL dan PCSK9. Kesan buah *Pm* akuaes ekstrak terhadap reseptor LDL dan PCSK9 di *in vivo* kemudian dikaji dalam eksperimen *in vitro* dengan menggunakan Hepatocellularcarcinoma (HepG2) sel. HepG2 sel diinkubasi dalam RPMI yang ditambahkan dengan 0.2% sera albumin lembu, LDL dan akues ekstrak buah *Pm* (0, 0.1, 2, 40 dan 1000  $\mu\text{g/ml}$ ) atau simvastatin (10  $\mu\text{M}$ ) selama 24 jam. Paras reseptor LDL (160kDa) dan PCSK9 protein dan mRNA kemudian ditentukan. Akues ekstrak buah *Pm* meningkatkan kepekatan protein LDL receptor (160 kDa) dan PCSK9 ( $P< 0.05$ ) di HepG2 sel sebanyak satu kali ganda walaupun pada dos serendah 0.1  $\mu\text{g/ml}$ . Selain itu, akues ekstrak buah *Pm* pada dos 0.1  $\mu\text{g/ml}$  juga meningkatkan paras reseptor LDL (160 kDa) dan PCSK9 mRNA ( $P<0.05$ ), standing dengan sel yang diinkubasi dengan simvastatin. Kesimpulannya, akues ekstrak buah *Pm* menurunkan berat badan, berat hepar dan nisbah berat hepar kepada berat badan. Selain daripada itu, akues ekstrak buah *Pm* juga menunjukkan kesan anti-

hiperkolesterolemia dengan menurunkan paras profil lipid tikus hiperkolesterolemia dan meningkatkan paras reseptor LDL dan PCSK9 pada tahap protein dan mRNA.

First and foremost, I would like to express my appreciation and gratitude to my project supervisor, Assoc. Prof. Dr. Mohamad Aziz Dollah, for his help, advice, suggestion and encouragement throughout my postgraduate study. Besides that, I would also like to extend my gratitude to my co-supervisor, Assoc. Prof. Dr. Pei Pei and Dr. Mally Abdullah for their guidance, advice and support throughout this experimental project. Without them, this experiment will not be a successful one.

My deepest appreciation goes to the administrative and non-academic staff of Faculty Medicine and Health Sciences for their support and assistance towards the completion of this project.

In addition, I would like to express my appreciation to Biomedical Lab and Pharmacology lab's staffs, Ms. Nurul Huda, Ms. Farhana and Ms. Thani Chai Ling for their constant support and assistance throughout the experimental period.

Last but not least, I would also like to thank my family members for their support, encouragement and glorious care all this time.

## ACKNOWLEDGEMENTS

First and foremost, I would like to express my appreciation and gratitude to my project supervisor, Assoc. Prof. Dr. Mohamad Aziz Dollah, for his help, advice, suggestion and encouragement throughout my postgraduate study. Besides that, I would also like to extend my gratitude to my co-supervisor, Assoc. Prof. Dr. Chong Pei Pei and Dr. Maha Abdullah for their guidance, advice and support throughout this experimental project. Without them, this experiment will not be a successful one.

My deepest appreciation also goes to all the academic and non-academic staff of Faculty Medicine and Health Sciences for their technical support and assistant towards the completion of this project.

In addition, I would also like to express my appreciation to Biomedical Lab and Immunology lab's students, Ms Shaneeta, Mr. Peishan and Ms. Tham Chau ling for their constant advice and support throughout the experimental period.

Last but not least, I would also like to thank my family members for their support, encouragement and glorious care all this time.

I certify that a Thesis Examination Committee has met on 15<sup>th</sup> December 2011 to conduct the final examination of Chong Soo Ching on her thesis entitled “*In Vivo* and *In Vitro* Effects of *Phaleria macrocarpa* (Scheff.) Boerl on Low Density Lipoprotein and PCSK9 Expression” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1988. The Committee recommends that the student be awarded the degree of Master of Science.

Members of the Thesis Examination Committee were as follows:

**Sabrina Sukardi, PhD**

Associate Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Zuraini Ahmad, PhD**

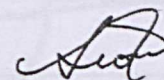
Associate Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Cheah Yoke Kqueen, PhD**

Associate Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Siti Balkis Budin, PhD**

Associate Professor  
Faculty of Health Sciences  
Universiti Kebangsaan Malaysia  
Malaysia  
(External Examiner)



---

**SEOW HENG FONG, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 2<sup>nd</sup> March 2012

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Mohamad Aziz Dollah, PhD**


Associate Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Chairman)

**Chong Pei Pei, PhD**

Associate Professor  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Member)

**Maha Abdullah, PhD**

Lecturer  
Faculty of Medicine and Health Sciences  
Universiti Putra Malaysia  
(Member)




---

**BUJANG BIN KIM HUAT, PhD**  
Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 19 MAR 2012

## DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.



**CHONG SOO CHING**  
Date: 15<sup>th</sup> December 2011

## TABLES OF CONTENTS

	<b>ABSTRACT</b>	<b>Page</b>
	<b>ABSTRAK</b>	ii
	<b>ACKNOWLEDGEMENTS</b>	v
	<b>APPROVAL</b>	viii
	<b>DECLARATION</b>	ix
	<b>LIST OF TABLES</b>	xi
	<b>LIST OF FIGURES</b>	xv
	<b>LIST OF ABBREVIATIONS</b>	xvii
	<b>LIST OF ANNOTATIONS</b>	xix
		xxii
 	<b>CHAPTER</b>	
 	<b>1 INTRODUCTION</b>	<b>1</b>
	1.1 Objectives	5
	1.1.1 General Objectives	5
	1.1.2 Specific Objectives	5
	1.2 Research Hypothesis	6
 	<b>2 LITERATURE REVIEW</b>	<b>7</b>
	2.1 Cholesterol	7
	2.1.1 Cholesterol transportation in the blood via lipoprotein	8
	2.1.2 Intracellular cholesterol metabolism	11
	2.1.3 Cholesterol metabolism in animal model	17
	2.2 LDL receptor	18
	2.3 PCSK9	19
	2.4 Hypercholesterolemia	20
	2.4.1 Non-Alcoholic Fatty Liver Disease (NAFLD)	22
	2.5 Treatment for Hypercholesterolemia	22
	2.5.1 Synthetic drugs	23
	2.5.2 Natural herbs	25
 	<b>3 MATERIALS AND METHODS</b>	<b>30</b>
	<i>In Vivo</i> study: Effect of <i>Pm</i> fruit aqueous extract on the hypercholesterolemic diet-induced rats	30
	3.1 Materials	30
	3.2 Instrument	30
	3.3 Limitation	31
	3.4 Methods	31
	3.4.1 Preparation of <i>Pm</i> fruit aqueous extract stock solution	31
	3.4.2 Preparation of Simvastatin stock solution	31
	3.4.3 Preparation of 3% cholesterol-enriched diet	32
	3.4.4 Experimental Animals	32
	3.4.5 Experimental design	33
	3.4.6 Bodyweight measurement	35
	3.4.7 Sample collection	35
	3.4.8 Sample analysis	36

3.5 Research flow chart for <i>In Vivo</i> study	44
3.6 Statistical analysis	45
<i>In Vitro</i> Study: Effect of <i>Pm</i> fruit aqueous extract on the LDL receptor and PCSK9 protein and mRNA transcript expression in Hepatocellularcarcinoma (HepG2) cells.	46
3.7 Materials	46
3.8 Instrument	46
3.9 Methods	47
3.9.1 Preparation of HepG2 media	47
3.9.2 Preparation of <i>Pm</i> fruit aqueous extract	47
3.9.3 Simvastatin preparation	47
3.9.4 Phosphate buffer saline (PBS) preparation	48
3.9.5 HepG2 cell maintenance	48
3.9.6 HepG2 growth curve	48
3.9.7 Total cell count	49
3.9.8 Cell viability using MTT assay	49
3.9.9 <i>Pm</i> fruit aqueous extract treatment	50
3.9.10 Sample analysis	52
3.10 Statistical analyses	56
<b>4 RESULTS AND DISCUSSION</b>	58
<i>In Vivo</i> study: Effect of <i>Pm</i> fruit aqueous extract on diet induced hypercholesterolemic rats	58
4.1 Effect of <i>Pm</i> fruit aqueous extract on bodyweight.	58
4.2 Effect of <i>Pm</i> fruit aqueous extract on liver weight and liver weight-to-body weight ratio	64
4.3 Effect of <i>Pm</i> fruit aqueous extract on the blood lipid profile (TC, TG, HDL and LDL)	65
4.3.1 Effect of <i>Pm</i> fruit aqueous extract on TC level	66
4.3.2 Effect of <i>Pm</i> fruit aqueous extract on TG level	69
4.3.3 Effect of <i>Pm</i> fruit aqueous extract on HDL level	72
4.3.4 Effect of <i>Pm</i> fruit aqueous extract on LDL level	75
4.4 Effect of <i>Pm</i> fruit aqueous extract on hepatic LDL receptor and PCSK9 proteins expression	78
4.4.1 Effect of <i>Pm</i> fruit aqueous extract on hepatic LDL receptor (160 kDa and 120 kDa) proteins expression.	78
4.4.2 Effect of <i>Pm</i> fruit aqueous extract on hepatic PCSK9 (63 kDa) protein expression	80
<i>In Vitro</i> study: Effect of <i>Pm</i> fruit aqueous extract on LDL receptor and PCSK9 proteins and mRNA transcripts in Hepatocellularcarcinoma (HepG2) cells.	82
4.5 HepG2 cell growth curve	82
4.6 Cell viability assay	83
4.7 Effect of <i>Pm</i> fruit aqueous extract on LDL receptor and PCSK9 protein expression in HepG2 cells	84

4.7.1 Effects of <i>Pm</i> fruit aqueous extract on LDL receptor (160 kDa) protein expression	84
4.7.2 Effect of <i>Pm</i> fruit aqueous extract on PCSK9 (63 kDa) protein expression	85
4.8 Effect of <i>Pm</i> fruit aqueous extract on LDL receptor and PCSK9 mRNA transcripts in HepG2 cells.	87
4.8.1 Effect of <i>Pm</i> fruit aqueous extract on LDL receptor mRNA transcript	87
4.8.2 Effect of <i>Pm</i> fruit aqueous extract on PCSK9 mRNA transcript	88
Discussion	90
<b>5 CONCLUSION</b>	101
<b>REFERENCES</b>	102
<b>APPENDICES</b>	112
<b>BIODATA OF STUDENT</b>	147
<b>LIST OF PUBLICATIONS</b>	148

## LIST OF TABLES

Table	Page
2.1. Cholesterol content in foods	8
3.1. Experimental design used for <i>in vivo</i> study.	34
3.2. Experimental design used for <i>in vitro</i> study	51
3.3. Primer sequences used for PCR amplification	56
4.1. Mean ( $\pm$ standard deviation) change in body weight of rats at 2 weeks interval for 20 weeks	60
4.2. Average daily gain of rats throughout the experimental period	63
4.3. Mean ( $\pm$ standard deviation) of TC level of rats fed with normal diet or 3% cholesterol enriched-diet, treated with <i>Pm</i> and simvastatin	68
4.4. Mean ( $\pm$ standard deviation) of triglyceride level of rats fed with normal diet or 3% cholesterol enriched-diet, treated with <i>Pm</i> and simvastatin	71
4.5. Mean ( $\pm$ standard deviation) of HDL level of rats fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin	74
4.6. Mean ( $\pm$ standard deviation) of LDL level of rats fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> and simvastatin	77
A.1. Data for the LDL receptor after PCR amplification	133
A.2: Data for the PCSK9 product after PCR amplification	134
A.3. ANOVA table for mean ( $\pm$ standard deviation) change in body weight of rats at 2 weeks interval for 20 weeks	143
A.4. Anova table for mean ( $\pm$ standard deviation) changes in the bodyweight of rats fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin throughout the experimental period	143
A.5. ANOVA table for average daily gain of rats throughout the experimental period	143
A.6. ANOVA table for mean ( $\pm$ standard deviation) weight of rat's liver fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin	143
A.7. ANOVA table for mean ( $\pm$ standard deviation) of liver weight-to-body weight ratio of rats fed with normal feeding or 3% cholesterol feeding, treated <i>Pm</i> or simvastatin	143

A.8. ANOVA table for mean ( $\pm$ standard deviation) of TC in rats fed with normal diet or 3% cholesterol enriched-diet, treated with <i>Pm</i> or simvastatin	144
A.9. ANOVA table for mean ( $\pm$ standard deviation) of triglyceride in rats fed with normal diet or 3% cholesterol enriched-diet, treated with <i>Pm</i> or simvastatin	144
A.10. ANOVA table for mean ( $\pm$ standard deviation) levels of HDL in rats fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin.	144
A.11. ANOVA table for mean ( $\pm$ standard deviation) levels of LDL in rats fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin	144
A.12. ANOVA table of mean ( $\pm$ standard deviation) concentration of hepatic LDL receptor (160kDa) of rats fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin	145
A.13. ANOVA table of mean ( $\pm$ standard deviation) concentration of hepatic LDL receptor (120kDa) of rats fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin	145
A.14. ANOVA table of mean ( $\pm$ standard deviation) concentration of hepatic PCSK9 of rats fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin	145
A.15. ANOVA table of mean ( $\pm$ standard deviation) concentration of LDL receptor in of HepG2 cells, treated with <i>Pm</i> or simvastatin	145
A.16. ANOVA table of mean ( $\pm$ standard deviation) concentration of PCSK9 in HepG2 cells, treated with <i>Pm</i> or simvastatin	145
A.17. ANOVA table of mean ( $\pm$ standard deviation) concentration of LDL receptor mRNA in HepG2 cells, treated with <i>Pm</i> or simvastatin	146
A.18. ANOVA table of mean ( $\pm$ standard deviation) concentration of PCSK9 mRNA in HepG2 cells, treated with <i>Pm</i> or simvastatin	146

## LIST OF FIGURE

Figure	Page
2.1. Cholesterol structure	7
2.2. Simplified flow chart of cholesterol biosynthesis. Three molecules of acetyl-CoA initiate the intracellular cholesterol synthesis pathway with HMG-CoA reductase as the rate limiting enzyme of this mechanism	13
2.3. LDL receptor pathway in cells.	14
2.4. <i>Phaleria macrocarpa</i> fruits	27
3.1. Research flow chart for in vivo study	44
4.1. Mean ( $\pm$ standard deviation) change in body weight of rats at 2 weeks interval for 20 weeks.	59
4.2. Mean ( $\pm$ standard deviation) weight of rat's liver fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin	64
4.3. Mean ( $\pm$ standard deviation) of liver weight-to-body weight ratio of rats fed with normal feeding or 3% cholesterol feeding, treated <i>Pm</i> or simvastatin.	65
4.4. Mean ( $\pm$ standard deviation) of TC in rats fed with normal diet or 3% cholesterol enriched-diet, treated with <i>Pm</i> or simvastatin	67
4.5. Mean ( $\pm$ standard deviation) of TG in rats fed with normal diet or 3% cholesterol enriched-diet, treated with <i>Pm</i> or simvastatin	70
4.6. Mean ( $\pm$ standard deviation) levels of HDL in rats fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin	73
4.7. Mean ( $\pm$ standard deviation) levels of LDL in rats fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin	76
4.8. Mean ( $\pm$ standard deviation) concentration of hepatic LDL receptor (160kDa) of rats fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin.	79
4.9. Mean ( $\pm$ standard deviation) concentration of hepatic LDL receptor (120kDa) of rats fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin	80
4.10. Mean ( $\pm$ standard deviation) concentration of hepatic PCSK9 of rats fed with normal diet or 3% cholesterol enriched diet, treated with <i>Pm</i> or simvastatin	81

4.11. Mean ( $\pm$ standard deviation) of growth curve of HepG2 cells incubated in RPMI supplemented with 10% Foetal bovine serum and 1% penicillin-streptomycin.	82
4.12. Mean ( $\pm$ standard deviation) of HepG2 growth curve of cells incubated in RPMI supplemented with 10% FBS and 1% penicillin-streptomycin for 24 hours followed by 48 hours of incubation in serum-free RPMI supplemented with 0.2% BSA and 1% penicillin-streptomycin	83
4.13. Cell viability assay for HepG2 cells, treated with or simvastatin	84
4.14. Mean ( $\pm$ standard deviation) concentration of LDL receptor in HepG2 cells, treated with <i>Pm</i> or simvastatin.	85
4.15. Mean ( $\pm$ standard deviation) concentration of PCSK9 in HepG2 cells, treated with <i>Pm</i> or simvastatin	86
4.16. Mean ( $\pm$ standard deviation) concentration of LDL receptor mRNA in HepG2 cells, treated with <i>Pm</i> or simvastatin	88
4.17. Mean ( $\pm$ standard deviation) concentration of PCSK9 mRNA in HepG2 cells, treated with <i>Pm</i> or simvastatin	89
5.1. Possible mechanism of action of anti-hypercholesterolemic effect of <i>Pm</i> fruit aqueous extract	100
A.1. ACUC approval from	114
A.2. Bloods withdrawn from the rat's eye via retro-orbital bleeding for blood lipid profile analysis	115
A.3. Livers of normal diet (control) and diet-induced hypercholesterolemic <i>Sprague dawley</i> rats	119
A.4. Standard curve for Bradford protein assay with bovine serum albumin (BSA) as standard for <i>in vivo</i> study	120
A.5. HepG2 cells (Adherent cells)	130
A.6. Standard curve for Bradford protein assay with bovine serum albumin (BSA) as standard for <i>in vitro</i> study	131
A.7. Formaldehyde agarose gel electrophoresis of RNA products after RNA extraction and DNase treatment of RNA.	132

## LIST OF ABBREVIATIONS

ACAT	acyl-CoA:cholesterol acyltransferase
ACUC	Animal Care and Use Committee
Apo	apolipoprotein
ATP	adenosine triphosphate
BMI	body mass index
BSA	bovine serum albumin
bw	by weight
cCEH	Cytosolic cholesteryl ester hydrolase
CEH	cholesteryl ester hydrolase
Cho	3% cholesterol enriched diet
CVD	cardiovascular disease
DEPC	diethyl dicarbonate
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
dNTPs	Deoxyribonucleotide triphosphate
ECL	Enhanced Chemiluminescence
EDTA	Ethylenediaminetetraacetic acid
EGF	epidermal growth-factor
ERK	extracellular signal regulated kinase
FA gel	formaldehyde agarose gel
FBS	fetal bovine serum
GAPDH	Glyceraldehyde 3-phosphate dehydrogenase
HDL	high density lipoprotein

HepG2 cell	Heptocellularcarcinoma cell
HMG-CoA	3-hydroxy-3-methylglutaryl-coenzyme A
HSDA	sodium N-(2-hydroxyl-3-sulfupropyl)-3,5-dimetoxyaniline
IDL	intermediate density lipoprotein
ISO	isoproterenol
LCAT	lecithin-cholesterol acyltransferase
LDL	low density lipoprotein
LPL	lipoprotein lipase
mRNA	messenger ribonucleic acid
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide
NADPH	nicotinamide adenine dinucleotide phosphate-oxidase
NTC	non-template control
OD	optical density
PBS	phosphate buffer saline
PCSK9	proprotein convertase subtilisin / kexin type 9
PCR	polymerase chain reaction
PEG-cholesterol esterase	polyethylene glycol- cholesterol esterase
<i>Pm</i>	<i>Phaleria macrocarpa</i>
PVDF	polyvinyl difluoride
RPMI 1640	Roswell Park Memorial Institute
RT-PCR	reverse transcriptase- polymerase chain reaction
SCAP	SREBP cleavage-activating protein

SDS	sodium dodecyl sulfate
SDS-PAGE	sodium dodecyl sulfate polyacrylamide gel electrophoresis
SR-B1	scavenger receptor B class 1
SREBP	Sterol regulatory element binding proteins
SREBP-2	Sterol regulatory element binding proteins-2
TAE	Tris-Acetate-EDTA
TBST	Tris-Buffered Saline Tween-20
TC	total cholesterol
TG	triglyceride
VLDL	very low density lipoprotein
WHO	World Health Organization

## LIST OF ANNOTATIONS

A	ampere
bp	base pair
g	gravity
kDa	kilo dalton
kg	kilogram
mg	milligram
mg/dl	milligram per deciliter
mg extract/ kg bw	milligram extract per kilogram by weight
mg/kg	milligram per kilogram
mg/ml	milligram per milliliter
ml	milliliter
nm	nanometer
rpm	revolutions per minute
U	units
µg	microgram
µg/ml	microgram per milliliter
µl	microliter
µM	micromolar
V	Volts
%	percent
°C	degree celcius
±	plus and/ or minus

## INTRODUCTION

Cholesterol, a waxy, oil-like substance is an essential metabolite that enables the normal physiological functions of the body in all animals including human. Cholesterol serves as a precursor for the synthesis of steroid hormones such as vitamin D, glucocorticoids, mineral corticoids and sex hormones. Cholesterol is also the precursor for the synthesis of bile acids that are responsible for fat absorption in the small intestine into the circulatory system via the lymphatic system. Cholesterol is found in every cell in the body. It is transported in the blood through the water-soluble carrier molecules called lipoproteins. There are five types of lipoproteins which are classified according to their density, namely chylomicron, very low density lipoprotein (VLDL), intermediate density lipoprotein (IDL), low density lipoprotein (LDL) and high density lipoprotein (HDL). Through the lipoprotein carrier, cholesterol is transported to other cells in the body such as steroidogenic cells, adrenal cortex and gonads (Owen & Farrell, 1992). Apart from dietary cholesterol, cells in the body synthesise their own cholesterol through 3-hydroxy-3-methylglutaryl-CoA reductase (HMG-CoA reductase) activity which is regulated by many regulatory elements such as SREBP-2 (Ahmed, 1994; Chale-Bozale & Abel, 1996; Burger, Gump, & Fahrenholz, 2000).

There are two types of cholesterol, bad cholesterol and good cholesterol. HDL is good cholesterol as HDL transports excess cholesterol from cells to liver where it is broken down. LDL is considered bad cholesterol as it transports cholesterol from liver to cells (Chale-Bozale & Abel, 1996). Elevated level of cholesterol is known as hypercholesterolemia and it is usually associated with an increased risk of coronary

## CHAPTER 1

### INTRODUCTION

Cholesterol, a waxy, fat-like substance is an essential metabolite that enables the normal physiological functions of the body in all animals including human. Cholesterol serves as a precursor for the synthesis of steroid hormones such as vitamin D, glucocorticoids, mineral corticoids and sex hormones. Apart from that, cholesterol is also the precursor for the synthesis of bile acids that are responsible for fat absorption in the small intestine into the circulatory system via the lymphatic system. Cholesterol is insoluble in the blood. Thus, it is transported in the blood through the water-soluble carrier molecules known as lipoprotein. There are five types of lipoprotein which can be categorized depending on their density, namely chylomicron, very low density lipoprotein (VLDL), intermediate density lipoprotein (IDL), low density lipoprotein (LDL) and high density lipoprotein (HDL). Through the lipoprotein carrier, cholesterol is transported to other cells in the body such as steroidogenic tissues, muscles and adipocytes (Grummer & Carroll, 1988). Apart from dietary cholesterol, cells in the body synthesize their own cholesterol through 3-hydroxy-3-methyl-glutaryl-CoA reductase (HMG-CoA reductase) activity which is regulated by sterol regulatory element binding protein-2 (SREBP-2) (Ahmad, 1994; Basile-Borgia & Abel, 1996; Burger, Gimpl, & Fahrenholz, 2000).

There are two types of cholesterol: bad cholesterol and good cholesterol. HDL is good cholesterol as HDL transports excess cholesterol from cells to liver where as LDL is considered bad cholesterol as it transports cholesterol from liver to cells (Basile-Borgia & Abel, 1996). Elevated level of cholesterol is known as hypercholesterolemia and it is usually associated with an increased risk of coronary

heart disease such as atherosclerosis, angina, heart attack and stroke. Instead of transporting cholesterol to the cells in the body, excess LDL deposits cholesterol into the arteries which cause the formation of the thick and hard plaque called atheroma which finally leads to atherosclerosis. On the other hand, HDL lipoprotein transports the cholesterol from the atheroma plaque back to the liver to be metabolized, thus reducing the atheroma plaque. So, it is important to maintain an ideal level of HDL (>60mg/dl) and LDL (<100mg/dl) (Stapleton, Goodwill, James, Brock & Frisbee, 2010)

Cholesterol metabolism occurs mostly in the liver via LDL receptor. LDL receptor, a cell surface membrane protein, recognizes apolipoprotein E (apoE) and apolipoprotein B100 (apoB100) in the lipoprotein. VLDL, IDL and LDL contain apoB100 where as apoE is found on most of the lipoprotein especially VLDL and HDL. Even so, the primary ligand for the LDL receptor is low-density lipoprotein, followed by beta-migrating form of VLDL ( $\beta$ -VLDL) (Lagor & Millar, 2009). After binding of the LDL to LDL receptor via apo B100, LDL is taken into the cells through receptor mediated endocytosis pathway. Then the LDL receptor is recycled back to the cell surface. Like HMG-CoA reductase, synthesis of LDL receptor is regulated by intracellular cholesterol through the SREBP-2 pathway. Mutation of LDL receptor is related to the familial hypercholesterolemia where by the LDL metabolism is disrupted, causing a high level of cholesterol in the blood (Goldstein & Brown, 2009; Goldstein, Brown, Anderson, Russell, & Schneider, 1985)

Apart from regulating LDL receptor and HMG-CoA, SREBP-2 also regulates the synthesis of proprotein convertase subtilisin-like kexin type 9 (PCSK9). PCSK9 is

identified as a new member of proprotein convertase family and has a role in the degradation of the LDL receptor. Since liver is the major organ that regulates cholesterol metabolism due to high level of LDL receptor, thus it is natural that PCSK9 is also highly expressed in liver. Synthesized as an inactive zymogen, PCSK9 undergoes autocatalytic cleavage at the Golgi apparatus. However, even though it is cleaved, PCSK9 prodomain remain associated to the mature PCSK9, assisting in escorting PCSK9 into the secretory pathway. Once outside the cells, the prodomain dissociates from the mature PCSK9. PCSK9 does not degrade the LDL receptor directly but binds to the LDL receptor on the cell surface, forming PCSK9-LDL receptor complex. This complex is then taken into the cells through receptor mediated endocytosis pathway. However, once inside the cells, the PCSK9 pump the LDL receptor into the lysosome for degradation. Like LDL receptor, PCSK9 mutation is also related to hypercholesterolemia. Gain-of-the function mutation shows a decrease in the LDL receptor where as loss-of-the function mutation shows an increase in the LDL receptor abundance in the liver (Kwon, Lagace, McNutt, Horton, & Deisenhofer, 2008; Lagor & Millar, 2009; Peterson, Fong, & Young, 2008).

Studies have shown that high cholesterol diet can leads to the formation of non-alcoholic fatty liver disease (NAFLD). An increase in dietary cholesterol is suggested to have increased the *de novo* synthesis of fatty acids in hepatocytes via Liver X Receptor Alpha (LXR $\alpha$ )-SREBP-1c pathway. Synthesized fatty acids in hepatocytes can form triglycerides, thus causing accumulation of triglyceride in liver, forming NAFLD. NAFLD is closely associated with metabolic syndrome, which is the risk factor for cardiovascular disease. Patients with NAFLD usually have an

increment in liver weight, proatherogenic lipid profile with elevated LDL, reduced HDL and postprandial hypertriglyceridemia (Cohen, Horton, & Hobbs, 2011; Smith & Adams, 2011; Zelber-Sagi, Ratzin, & Oren, 2011).

Hypercholesterolemia is defined as cholesterol level higher than the recommended level ( $>200\text{mg/dl}$ ). Hypercholesterolemia will lead to chronic heart disease. Thus, hypercholesterolemic condition must be treated. Currently there are synthetic drugs that are available in the market that are used to treat the hypercholesterolemic condition such as statins, cholesterol absorption inhibitors, resins, fibrates and niacin. Simvastatin, a statin drug used in this experiment as a positive control is a HMG-CoA reductase inhibitor. HMG-CoA is a rate-limiting enzyme for cholesterol biosynthesis. Apart from inhibiting HMG-CoA reductase, simvastatin has also been reported to be able to upregulate hepatic LDL receptor and PCSK9. However, like all synthetic drugs, simvastatin can cause side effects. Thus, herbs and plants may have potential as alternatives in medicine to control hypercholesterolemia as they have lesser side effects. One of the herbs currently used is *Phaleria macrocarpa* (*Pm*) fruit. *Pm* which originates from Papua, Indonesia is a traditional herb that has been used for a few decades to treat diabetes mellitus, hypertension, hypercholesterolemia and others (Sugiwati, Leonardus, & Bintang, 2006). Even though *Pm* fruit had been proven to be able to reduce cholesterol level (Adnyana, Yulinah, Sigit, & Fitriani, 2005; Armenia, Ermilda, Widya, & Rusdi, 2006) but the mechanism is still unknown. Thus, this study was conducted to determine the possible mechanism of anti-hypercholesterolemic property of *Pm* fruit aqueous extract.

## 1.1 Objectives

### 1.1.1 General Objectives

To assess the anti-hypercholesterolemic property of *Pm* fruit aqueous extract *in vivo* and *in vitro*.

### 1.1.2 Specific Objectives

#### 1.1.2.1 *In vivo* study

1. To determine the effects of *Pm* fruit aqueous extract on the body weight, liver weight and liver weight-to-body weight ratio in hypercholesterolemic rats.
2. To determine the effects of *Pm* fruit aqueous extract on the blood lipid profiles [Total Cholesterol (TC), Triglycerides (TG), HDL and LDL]
3. To determine the effects of *Pm* fruit aqueous extract on the expression of hepatic LDL receptor and PCSK9 proteins

#### 1.1.2.2 *In vitro* study

1. To determine the effects of *Pm* fruit aqueous extract on the expression of LDL receptor and PCSK9 proteins in HepG2 cells.
2. To determine the effects of *Pm* fruit aqueous extract on the expression of LDL receptor and PCSK9mRNA transcript in HepG2 cells.

## 1.2 Research Hypothesis

*Pm* fruit aqueous extract posses anti-hypercholesterolemic property by reducing blood TC, TG and LDL level but increases HDL of hypercholesterolemic diet-induced rats by upregulating LDL receptor and PCSK9 protein and mRNA expression. Apart from that, *Pm* fruit also reduces body weight gain, liver weight and liver weight-to-body weight ratio of diet induced hypercholesterolemic rats.

Figure 2.1. Cholesterol structure  
(Ahmad, 1994)

Cholesterol is an organic compound with molecular formula  $C_{27}H_{46}O$  (Figure 2.1). In the pure state, cholesterol is a white crystalline substance that is odorless and tasteless. Being essential to life, cholesterol serves many functions. It acts as a precursor of intermediate of the synthesis of steroid hormones, bile acids and fat soluble vitamins (vitamins A, D, E and K). Cholesterol is also essential to maintain the fluidity of the cell membrane and thereby needed for different synaptic responses (Pillay, 2001). Our body obtains cholesterol from two different sources: dietary cholesterol and synthesis in our body. Even though body stores dietary cholesterol accounts for only 20-23%, dietary cholesterol intake can affect the blood cholesterol level. Dietary cholesterol derived from meat, poultry, fish and dairy products contain higher amount of cholesterol compared to food derived from plants as shown in Table 2.1 (Ahmad, 1994; Burger et al., 2000).

## REFERENCES

- Adnyana, K., Yulinah, E., Sigit, J. I., & Fitriani, D. (2005). Anticholesterol Activity Test of Aqueous Extract of *Allium sativum* L. Bulbs, *Eugenia polyantha* Wight Leaves and *Phaleria macrocarpa* Fruit in Primary Culture. *Acta Pharmaceutica Indonesia*, 30, 43-47.
- Ahmad, J. I. (1994). Cholesterol: A current Perspective. *Nutrition & Food Science*, 1, 4.
- Aisyah, S. A. G. (2007). *Hypocholesterolemic effects of Phaleria macrocarpa (mahkota dewa) fruit aqueous extract in cholesterol diet fed-rats*. (Unpublished Bachelor Degree disertation). University Putra Malaysia, Malaysia..
- Al Rayyes, O., & Floren, C. H. (1998). Troglitazone upregulates LDL receptor activity in HepG2 cells. *Diabetes*, 47(8), 1193-1198.
- Almahdy, A., Febrianti, R., & Djamal, R. (2008). Efek Fetotoksisitas Ekstrak Biji Mahkota Dewa (*Phaleria macrocarpa* (Scheff.) Boerl) Pada Mencit *Jurnal Sains dan Teknologi Farmasi*, 13(2), 86-88.
- Animal Resource Center. (2002, March 8th). Template for rodent retro orbital bleeding. Retrieved from [http://casemed.case.edu/ora/arc/templates\\_retro\\_orbital\\_bleeding.html](http://casemed.case.edu/ora/arc/templates_retro_orbital_bleeding.html)
- Armenia, Ermilda, F., Widya, R. M., & Rusdi, D. J. (2006, 13-18 November). *Anti-Atherosclerotic Effect and Liver Toxicity of Ethanolic Extract of Phaleria macrocarpa (Scheff.) Boerl Fruit on Japanese Quail*. Paper presented at the Asian Symposium on Medicinal Plants, Spices and Other Natural Products XII, Padang, Indonesia.
- Aroor, A. R., & Shukla, S. D. (2011). Binge ethanol intake in chronically exposed rat liver decreases LDL-receptor and increases angiotensinogen gene expression. *World J Hepatol*, 3(9), 250-255.
- Asma. (2009, January 20th). Teh mahkota dewa. Retrieved from <http://shopatasma.blogspot.com/2009/08/mahkota-dewa.html>
- Attie, A. D., & Seidah, N. G. (2005). Dual regulation of the LDL receptor--some clarity and new questions. *Cell Metab*, 1(5), 290-292.
- Basile-Borgia, A., & Abel, J. H. (1996). Lipoproteins in heart disease. *Perfusion*, 11(4), 338-345.
- Bose, C., Bhuvaneswaran, C., & Udupa, K. B. (2005). Age-related alteration in hepatic acyl-CoA: cholesterol acyltransferase and its relation to LDL receptor and MAPK. *Mech Ageing Dev*, 126(6-7), 740-751.

- Brown, M. S., & Goldstein, J. L. (1997). The SREBP pathway: regulation of cholesterol metabolism by proteolysis of a membrane-bound transcription factor. *Cell*, 89(3), 331-340.
- Bundy, R., Walker, A. F., Middleton, R. W., Wallis, C., & Simpson, H. C. (2008). Artichoke leaf extract (*Cynara scolymus*) reduces plasma cholesterol in otherwise healthy hypercholesterolemic adults: a randomized, double blind placebo controlled trial. *Phytomedicine*, 15(9), 668-675.
- Burger, K., Gimpl, G., & Fahrenholz, F. (2000). Regulation of receptor function by cholesterol. *Cell Mol Life Sci*, 57(11), 1577-1592.
- Bursill, C., Roach, P. D., Bottema, C. D., & Pal, S. (2001). Green tea upregulates the low-density lipoprotein receptor through the sterol-regulated element binding Protein in HepG2 liver cells. *J Agric Food Chem*, 49(11), 5639-5645.
- Cameron, J., Ranheim, T., Kulseth, M. A., Leren, T. P., & Berge, K. E. (2008). Berberine decreases PCSK9 expression in HepG2 cells. *Atherosclerosis*, 201(2), 266-273.
- Chang, C., Dong, R., Miyazaki, A., Sakashita, N., Zhang, Y., Liu, J., Chang, T. Y. (2006). Human acyl-CoA:cholesterol acyltransferase (ACAT) and its potential as a target for pharmaceutical intervention against atherosclerosis. *Acta Biochim Biophys Sin (Shanghai)*, 38(3), 151-156.
- Chen, Y. X., Ruan, X. Z., Huang, A. L., Li, Q., Moorhead, J. F., & Varghese, Z. (2007). Mechanisms of dysregulation of low-density lipoprotein receptor expression in HepG2 cells induced by inflammatory cytokines. *Chin Med J (Engl)*, 120(24), 2185-2190.
- Chong, S.C. (2008). *Anti-hypercholesterolemic property of Phaleria macrocarpa fruit aqueous extract in hypercholesterolemic-induced rats*. (Unpublished Bachelor Degree dissertation). University Putra Malaysia, Malaysia
- Cohen, J. C., Horton, J. D., & Hobbs, H. H. (2009). PCSK9: a convertase that coordinates LDL metabolism. *J. lipid Res*, 50, S172-S177.
- Cohen, J. C., Horton, J. D., & Hobbs, H. H. (2011). Human fatty liver disease: old questions and new insights. *Science*, 332(6037), 1519-1523.
- Costet, P., Krempf, M., & Cariou, B. (2008). PCSK9 and LDL cholesterol: unravelling the target to design the bullet. *Trends Biochem Sci*, 33(9), 426-434.
- Dorfman, S. E., Wang, S., Vega-Lopez, S., Jauhiainen, M., & Lichtenstein, A. H. (2005). Dietary fatty acids and cholesterol differentially modulate HDL cholesterol metabolism in Golden-Syrian hamsters. *J Nutr*, 135(3), 492-498.

- Dubuc, G., Chamberland, A., Wassef, H., Davignon, J., Seidah, N. G., Bernier, L., & Prat, A. (2004). Statins upregulate PCSK9, the gene encoding the proprotein convertase neural apoptosis-regulated convertase-1 implicated in familial hypercholesterolemia. *Arterioscler Thromb Vasc Biol*, 24(8), 1454-1459.
- Dzien, A., Breier, C., Lisch, H. J., & Braunsteiner, H. (1986). Stimulating effect of intermediate-density lipoproteins (IDL) from hyperlipemic plasma on hepatic lipase. *Klin Wochenschr*, 64(11), 530-533.
- Eberle, D., Hegarty, B., Bossard, P., Ferre, P., & Foulfelle, F. (2004). SREBP transcription factors: master regulators of lipid homeostasis. *Biochimie*, 86(11), 839-848.
- Ehrlich, S. D. (2008, January 16th). Hypercholesterolemia. Retrieved from <http://www.umm.edu/altmed/articles/hypercholesterolemia-000084.htm>
- Erdogru, O., & Azirak, S. (2004). Review of the studies on the red yeast rice (*Monascus purpureus*). *Turkish Electronic Journal of Biotechnology*, 2, 37-49
- Fariad, A., Kurnia, D., Fariad, L. S., Usman, N., Miyazaki, T., Kato, H., & Kuwano, H. (2007). Anticancer effects of gallic acid isolated from Indonesian herbal medicine, *Phaleria macrocarpa* (Scheff.) Boerl, on human cancer cell lines. *Int J Oncol*, 30(3), 605-613.
- Food and Drug Administration. (2005). Guidance for industry estimating the maximum safe starting dose in initial clinical trials for therapeutics in adult healthy volunteers: United States of America: U.S. Department of Health and Human Services
- Ganazz. (2008, January 20th). Khasiat mahkota dewa. Retrieved from <http://www.cakap.net/index.php?showtopic=1058&mode=threaded&pid=5004>
- Gillotte, K. L., Horkko, S., Witztum, J. L., & Steinberg, D. (2000). Oxidized phospholipids, linked to apolipoprotein B of oxidized LDL, are ligands for macrophage scavenger receptors. *J Lipid Res*, 41(5), 824-833.
- Goldstein, J. L., & Brown, M. S. (2009). The LDL receptor. *Arterioscler Thromb Vasc Biol*, 29(4), 431-438.
- Goldstein, J. L., Brown, M. S., Anderson, R. G., Russell, D. W., & Schneider, W. J. (1985). Receptor-mediated endocytosis: concepts emerging from the LDL receptor system. *Annu Rev Cell Biol*, 1, 1-39.
- Goto, D., Okimoto, T., Ono, M., Shimotsu, H., Abe, K., Tsujita, Y., & Kuwano, M. (1997). Upregulation of low density lipoprotein receptor by gemfibrozil, a hypolipemic agent, in human hepatoma cells through stabilization of mRNA transcripts. *Arterioscler Thromb Vasc Biol*, 17(11), 2707-2712.

- Grummer, R. R., & Carroll, D. J. (1988). A review of lipoprotein cholesterol metabolism: importance to ovarian function. *J Anim Sci*, 66(12), 3160-3173.
- Havekes, L., van Hinsbergh, V., Kempen, H. J., & Emeis, J. (1983). The metabolism in vitro of human low-density lipoprotein by the human hepatoma cell line Hep G2. *Biochem J*, 214(3), 951-958.
- Hendra, R., Ahmad, S., Oskoueian, E., Sukari, A., & Shukor, M. Y. (2011). Antioxidant, Anti-inflammatory and Cytotoxicity of *Phaleria macrocarpa* (Boerl.) Scheff Fruit. *BMC Complement Altern Med*, 11, 110-119.
- Ikewaki, K., Tohyama, J., Nakata, Y., Wakikawa, T., Kido, T., & Mochizuki, S. (2004). Fenofibrate effectively reduces remnants, and small dense LDL, and increases HDL particle number in hypertriglyceridemic men - a nuclear magnetic resonance study. *J Atheroscler Thromb*, 11(5), 278-285.]
- Jakobsen, M. U., Berentzen, T., Sorensen, T. I., & Overvad, K. (2007). Abdominal obesity and fatty liver. *Epidemiol Rev*, 29, 77-87.
- Jang, A., Srinivasan, P., Lee, N. Y., Song, H. P., Lee, J. W., Lee, M., & Jo, C. (2008). Comparison of hypolipidemic activity of synthetic gallic acid-linoleic acid ester with mixture of gallic acid and linoleic acid, gallic acid, and linoleic acid on high-fat diet induced obesity in C57BL/6 Cr Slc mice. *Chem Biol Interact*, 174(2), 109-117.
- Javitt, N. B. (1990). HepG2 cells as a resource for metabolic studies: lipoprotein, cholesterol, and bile acids. *The FASEB journal*, 4, 161-168.
- Kanasaki, K., & Koya, D. (2011). Biology of obesity: lessons from animal models of obesity. *J Biomed Biotechnol*, 2011, 197636.
- Kim, J. L., Chae, I. S., Kang, Y. H., & Kang, J. S. (2008). Effect of onion and beet on plasma and liver lipids, platelet aggregation, and erythrocyte Na efflux in simvastatin treated hypercholesterolemic rats. *Nutr Res Pract*, 2(4), 211-217.
- Kingsley, D. M., & Krieger, M. (1984). Receptor-mediated endocytosis of low density lipoprotein: somatic cell mutants define multiple genes required for expression of surface-receptor activity. *Proc Natl Acad Sci U S A*, 81(17), 5454-5458.
- Krieger, M. (1999). Charting the fate of the "good cholesterol": identification and characterization of the high-density lipoprotein receptor SR-BI. *Annu Rev Biochem*, 68, 523-558.
- Krieger, M., Kingsley, D. M., Sege, R., Hobbie, L., & Kozarsky, K. (1985). Genetic analysis of receptor-mediated endocytosis. *Trends in Biochemical Sciences*, 10(11), 447-452.

- Kwon, H. J., Lagace, T. A., McNutt, M. C., Horton, J. D., & Deisenhofer, J. (2008). Molecular basis for LDL receptor recognition by PCSK9. *Proc Natl Acad Sci U S A*, 105(6), 1820-1825.
- Lagor, W. R., & Millar, J. S. (2009). Overview of the LDL receptor: Relevance to cholesterol metabolism and future approaches for the treatment of coronary heart disease. *Journal of Receptor, Ligand and Channel Research*, 3, 1-14.
- Landschulz, K. T., Pathak, R. K., Rigotti, A., Krieger, M., & Hobbs, H. H. (1996). Regulation of scavenger receptor, class B, type I, a high density lipoprotein receptor, in liver and steroidogenic tissues of the rat. *J Clin Invest*, 98(4), 984-995.
- Lavappa, K. S. (1978). Survey of ATCC stocks of human cell lines for HeLa contamination. *In Vitro*, 14(5), 469-475.
- Lee, J. Y., & Carr, T. P. (2004). Dietary fatty acids regulate acyl-CoA:cholesterol acyltransferase and cytosolic cholesteryl ester hydrolase in hamsters. *J Nutr*, 134(12), 3239-3244.
- Liang, K., Kim, C. H., & Vaziri, N. D. (2005). HMG-CoA reductase inhibition reverses LCAT and LDL receptor deficiencies and improves HDL in rats with chronic renal failure. *Am J Physiol Renal Physiol*, 288(3), F539-544.
- Libra, A., Ferneti, C., Lorusso, V., Visigalli, M., Anelli, P. L., Staud, F., Pascolo, L. (2006). Molecular determinants in the transport of a bile acid-derived diagnostic agent in tumoral and nontumoral cell lines of human liver. *J Pharmacol Exp Ther*, 319(2), 809-817.
- Liscum, L., & Dahl, N.K. (1992). Intracellular cholesterol transport. *J. lipid Res*, 33, 1239-1254.
- Lopez, D. (2008). PCSK9: an enigmatic protease. *Biochim Biophys Acta*, 1781(4), 184-191.
- Lu, C., & Hwang, L. S. (2008). Polyphenol contents of Pu-Erh teas and their abilities to inhibit cholesterol biosynthesis in Hep G2 cell line. *Food Chemistry*, 111(1), 67-71.
- Masyitah, I. M. M. (2008). *Effect of nigella sativa on lipid profile of cholesterol-enriched diet rats*. (Unpublished Bachelor Degree dissertation). University Putra Malaysia, Malaysia
- Mayne, J., Dewpura, T., Raymond, A., Cousins, M., Chaplin, A., Lahey, K. A., Chretien, M. (2008). Plasma PCSK9 levels are significantly modified by statins and fibrates in humans. *Lipids Health Dis*, 7, 22-30.
- McKeage, K., & Keating, G. M. (2011). Fenofibrate: a review of its use in dyslipidaemia. *Drugs*, 71(14), 1917-1946.

- Miller, N. E. (1990). HDL metabolism and its role in lipid transport. *Eur Heart J*, 11 Suppl H, 1-3.
- Miller, M., Stone, N.J., Ballantyne, C., Bittner, V., Criqui, M.H., Ginsberg, H.N., Goldberg, A.C., Howard, W.J., Jacobson, M.S., Kris-Etherton, P.M., Lennie, T.A., Levi, M., Mazzone, T., & Pennathur, S. (2011). Triglycerides and cardiovascular disease: a scientific statement from the American Heart Association. *Circulation*, 123, 2292-2333.
- Nazifi, S., Saeb, M., Rowghani, E., Hasankhani, M., Hasanshahi, F., & Ghafari, N. (2007). Studies on the Physiological Relationship Between Thyroid Hormones, Serum Lipid Profile and Erythrocyte Antioxidant Enzymes in Clinically Healthy Iranian Fat-Tailed Sheep. *Bulgarina Journal of Veterinary Medicine*, 10(3), 161-167.
- Nurul, K. A. (2005). *Effect of ficus deltoidea on cholesterol level and its antioxidant properties on lipid peroxidation in cholesterol-fed rats*. (Unpublished Bachelor Degree disertation). University Putra Malaysia, Malaysia..
- Oshimi, S., Zaima, K., Matsuno, Y., Hirasawa, Y., Iizuka, T., Studiawan, H., Morita, H. (2008). Studies on the constituents from the fruits of *Phaleria macrocarpa*. *Journal of Natural Medicine*, 62(2), 207-210.
- Ow, Y. Y., & Stupans, I. (2003). Gallic acid and gallic acid derivatives: effects on drug metabolizing enzymes. *Curr Drug Metab*, 4(3), 241-248.
- Pagler, T. A., Rhode, S., Neuhofer, A., Laggner, H., Strobl, W., Hinterndorfer, C., Stangl, H. (2006). SR-BI-mediated high density lipoprotein (HDL) endocytosis leads to HDL resecretion facilitating cholesterol efflux. *J Biol Chem*, 281(16), 11193-11204.
- Pal, S., Ho, N., Santos, C., Dubois, P., Mamo, J., Croft, K., & Allister, E. (2003). Red wine polyphenolics increase LDL receptor expression and activity and suppress the secretion of ApoB100 from human HepG2 cells. *J Nutr*, 133(3), 700-706.
- Parini, P., Jiang, Z. Y., Einarsson, C., Eggertsen, G., Zhang, S. D., Rudel, L. L., Eriksson, M. (2009). ACAT2 and human hepatic cholesterol metabolism: identification of important gender-related differences in normolipidemic, non-obese Chinese patients. *Atherosclerosis*, 207(1), 266-271.
- Park, S. W., Moon, Y. A., & Horton, J. D. (2004). Post-transcriptional regulation of low density lipoprotein receptor protein by proprotein convertase subtilisin/kexin type 9a in mouse liver. *J Biol Chem*, 279(48), 50630-50638.
- Peterson, A. S., Fong, L. G., & Young, S. G. (2008). PCSK9 function and physiology. *J Lipid Res*, 49(6), 1152-1156.

- Pfriege, F. W. (2003). Cholesterol homeostasis and function in neurons of the central nervous system. *Cell Mol Life Sci*, 60(6), 1158-1171.
- Pinzani, M. (2011). Pathophysiology of non-alcoholic steatohepatitis and basis for treatment. *Dig Dis*, 29(2), 243-248.
- Ponnian Stanely Mainzen Prince2, D. (2010). Gallic acid pretreatment ameliorates lipids, lipoproteins and HMG-CoA reductase in normal and isoproterenol induced myocardial infarction in male wistar rats. *Journal of Pharmacy Research*, 3(5), 960-964.
- Prasetya, R. K. (2009). *Pengaruh infusa daging buah mahkota dewa (phaleria macrocarpa (scheff.) boerl) terhadap penurunan glukosa darah kelinci jantan New Zealand yang dibebani glukosa oral*. (Unpublished dissertation) Universitas Muhammadiyah Surakarta, Surakarta.
- Redgrave, T. G. (2004). Chylomicron metabolism. *Biochem Soc Trans*, 32(Pt 1), 79-82.
- Ringseis, R., Konig, B., Leuner, B., Schubert, S., Nass, N., Stangl, G., & Eder, K. (2006). LDL receptor gene transcription is selectively induced by t10c12-CLA but not by c9t11-CLA in the human hepatoma cell line HepG2. *Biochim Biophys Acta*, 1761(10), 1235-1243.
- Robinson, B. (2011). Retro-orbital bleeding procedure in mice and rats. United States of America: Thomas Jefferson University.
- Roche. (2003). Triglyceride Indianapolis: Roche Diagnostics GmbH.
- Roche. (2004). Cholesterol. Indianapolis: Roche Diagnostics GmbH.
- Roche. (2005a). HDL-C plus 2nd generation. Indianapolis: Roche Diagnostics GmbH.
- Roche. (2005b). LDL-C plus 2nd generation. Indianapolis: Roche Diagnostics GmbH.
- Ruiz, J., Kouivaskaia, D., Migliorini, M., Robinson, S., Saenko, E. L., Gorlatova, N., Strickland, D. K. (2005). The apoE isoform binding properties of the VLDL receptor reveal marked differences from LRP and the LDL receptor. *J Lipid Res*, 46(8), 1721-1731.
- Sato, R., & Takano, T. (1995). Regulation of intracellular cholesterol metabolism. *Cell Structure and Function*, 20, 421-427.
- Saufi, A. (2007). *Lignans in phaleria macrocarpa (scheff.) boerl. and in linum flavum var. compactum L.* (Unpublished Doctoral Dissertation). Heinrich-Heine-Universität Düsseldorf, Germany.

- Scandinavian Simvastatin Survival Study Group. (1994). Randomised trial of cholesterol lowering in 4444 patients with coronary heart disease: the Scandinavian Simvastatin Survival Study (4S). *The Lancet*, 344(8934), 1383-1389.
- Silver, D. L., Wang, N., Xiao, X., & Tall, A. R. (2001). High density lipoprotein (HDL) particle uptake mediated by scavenger receptor class B type 1 results in selective sorting of HDL cholesterol from protein and polarized cholesterol secretion. *J Biol Chem*, 276(27), 25287-25293.
- Smith, B. W., & Adams, L. A. (2011). Non-alcoholic fatty liver disease. *Crit Rev Clin Lab Sci*, 48(3), 97-113.
- Sorci-Thomas, M., Hendricks, C. L., & Kearns, M. W. (1992). HepG2 cell LDL receptor activity and the accumulation of apolipoprotein B and E in response to docosahexaenoic acid and cholesterol. *J Lipid Res*, 33(8), 1147-1156.
- Soutar, A.K., & Naoumova, R.P. (2006). Mechanism of disease: genetic causes of familial hypercholesterolemia. *Nat Clin Pract Cardiovasc Med*, 4(4), 214-225
- Spady, D. K., Kearney, D. M., & Hobbs, H. H. (1999). Polyunsaturated fatty acids up-regulate hepatic scavenger receptor B1 (SR-BI) expression and HDL cholesteryl ester uptake in the hamster. *J Lipid Res*, 40(8), 1384-1394.
- Stancu, C., & Sima, A. (2001). Statins: mechanism of action and effects. *J Cell Mol Med*, 5(4), 378-387.
- Stange, E. F., Osenbrugge, M., Rustan, M., Reimann, F., Schneider, A., Ditschuneit, H. H., & Ditschuneit, H. (1991). Inhibition of HMG-CoA reductase in mononuclear cells during gemfibrozil treatment. *Atherosclerosis*, 91(3), 257-265.
- Stapleton, P.A., Goodwill, A.G., James, M.E., Brock, R.W., & Frisbee, J.C. (2010). Hypercholesterolemia and microvascular dysfunction: interventional strategies. *Journal of Inflammation*, 7, 54-63.
- Stein, E. A., Lane, M., & Laskarzewski, P. (1998). Comparison of statins in hypertriglyceridemia. *Am J Cardiol*, 81(4A), 66B-69B.
- Suarez, Y., Fernandez, C., Gomez-Coronado, D., Ferruelo, A. J., Davalos, A., Martinez-Botas, J., & Lasuncion, M. A. (2004). Synergistic upregulation of low-density lipoprotein receptor activity by tamoxifen and lovastatin. *Cardiovasc Res*, 64(2), 346-355.
- Sugiwati, S., Leonardus, B. S. K., & Bintang, M. (2006).  $\alpha$ -Glucosidase inhibitory activity and hypoglycemic effect of *Phaleria macrocarpa* fruit pericarp extracts by oral administration to rats. *Journal of Applied Sciences*, 6, 2312-2316.

- Tholstrup, T., Vessby, B., & Sandstrom, B. (2003). Difference in effect of myristic and stearic acid on plasma HDL cholesterol within 24 h in young men. *Eur J Clin Nutr*, 57(6), 735-742.
- Triastuti, A., & Jong, W. C. (2008). Protective effects of ethyl acetate fraction of *Phaleria macrocarpa* (scheff.) boerl. on oxidative stress associated with alloxan-induced diabetic rats. *Jurnal Ilmiah Farmasi*, 5(1), 9-17.
- Trigatti, B. L., Krieger, M., & Rigotti, A. (2003). Influence of the HDL receptor SR-BI on lipoprotein metabolism and atherosclerosis. *Arterioscler Thromb Vasc Biol*, 23(10), 1732-1738.
- Troutt, J. S., Alborn, W. E., Cao, G., & Konrad, R. J. (2010). Fenofibrate treatment increases human serum proprotein convertase subtilisin kexin type 9 levels. *J Lipid Res*, 51(2), 345-351.
- Tsutsumi, K. (2003). Lipoprotein lipase and atherosclerosis. *Curr Vasc Pharmacol*, 1(1), 11-17.
- UCSF Medical Center. (2011, February 19th). Cholesterol content of foods. Retrieved from [http://www.ucsfhealth.org/education/cholesterol\\_content\\_of\\_foods/](http://www.ucsfhealth.org/education/cholesterol_content_of_foods/)
- Williams, C. M., Bateman, P. A., Jackson, K. G., & Yaqoob, P. (2004). Dietary fatty acids and chylomicron synthesis and secretion. *Biochem Soc Trans*, 32(Pt 1), 55-58.
- Wong, J., Quinn, C. M., & Brown, A. J. (2006). SREBP-2 positively regulates transcription of the cholesterol efflux gene, ABCA1, by generating oxysterol ligands for LXR. *Biochem J*, 400(3), 485-491.
- Wong, W. Y. (2005). *Effect of orthosiphon slamineas (misai kucing) extract on blood glucose and lipid profile in streptozotocin-induced diabetic rats*. (Unpublished Bachelor Degree dissertation). University Putra Malaysia, Malaysia.
- Xiangdong, L., Yuanwu, L., Hua, Z., Liming, R., Qiuyan, L., & Ning, L. (2011). Animal models for the atherosclerosis research: a review. *Protein Cell*, 2(3), 189-201.
- Yanni, A. E. (2004). The laboratory rabbit: an animal model of atherosclerosis research. *Lab Anim*, 38(3), 246-256.
- Zainal Ariffin, S. H., Wan Omar, W. H., Zainal Ariffin, Z., Safian, M. F., Senafi, S., & Megat Abdul Wahab, R. (2009). Intrinsic anticarcinogenic effects of *Piper sarmentosum* ethanolic extract on a human hepatoma cell line. *Cancer Cell Int*, 9, 6-14.

Zelber-Sagi, S., Ratzlu, V., & Oren, R. (2011). Nutrition and physical activity in NAFLD: an overview of the epidemiological evidence. *World J Gastroenterol*, 17(29), 3377-3389.

Zhu, B., Shen, H., Zhou, J., Lin, F., & Hu, Y. (2005). Effects of simvastatin on oxidative stress in streptozotocin-induced diabetic rats: a role for glomeruli protection. *Nephron Exp Nephrol*, 101(1), e1-8.