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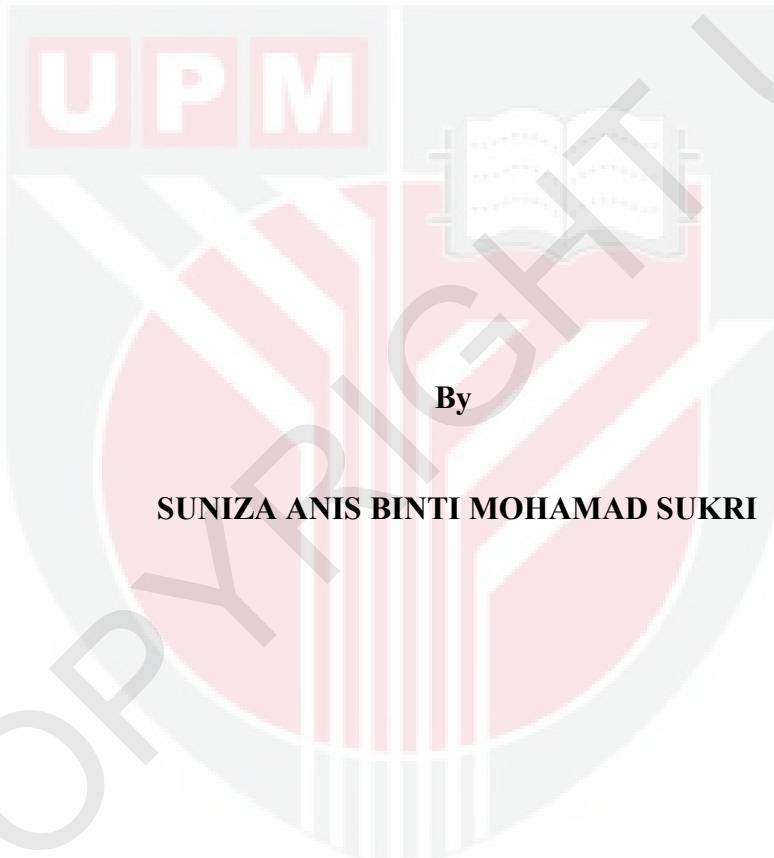
**ASSESSMENT OF SELECTED MICROALGAE (*Spirulina sp.*,
Nannochloropsis sp. AND *Chlorella sp.*) AS FEED SUPPLEMENT FOR
Macrobrachium rosenbergii (de Man) JUVENILES**

SUNIZA ANIS BINTI MOHAMAD SUKRI

FP 2017 56



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Macrobrachium rosenbergii (de Man) JUVENILES**



SUNIZA ANIS BINTI MOHAMAD SUKRI



**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

February 2017

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DEDICATION

To my mom and dad who encourage me
TUAN YAH BINTI TUAN LONG
MOHAMAD SUKRI BIN MOHD ZAIN

Beloved Husband
JALAL IKRAM BIN MOHAMED NORI

Beloved sons
AHMAD FURQAN MAHDI BIN JALAL IKRAM
AHMAD FURQAN ZUHDI BIN JALAL IKRAM
AHMAD FIRMAN ZIKRI BIN JALAL IKRAM

Siblings
SAIFUDDIN BAKHTIAR BIN MOHAMAD SUKRI
SYURIYANA BINTI MOHAMAD SUKRI
SAIFULNIZAM BOKHARI BIN MOHAMAD SUKRI
SAIFULHISAM BOKHIRI BIN MOHAMAD SUKRI
NURSOFIAH ANIS BINTI MOHAMAD SUKRI
MUHAMMAD SYAFIQ BIN MOHAMAD SUKRI

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

**ASSESSMENT OF SELECTED MICROALGAE (*Spirulina* sp.,
Nannochloropsis sp. AND *Chlorella* sp.) AS FEED SUPPLEMENT FOR
Macrobrachium rosenbergii (de Man) JUVENILES**

By

SUNIZA ANIS BINTI MOHAMAD SUKRI

February 2017

Chairman : Associate Professor Che Roos Bin Saad, PhD
Faculty : Agriculture

A series of experiment were conducted to study the potential of feed formulation for freshwater prawn *Macrobrachium rosenbergii* juvenile. Study on the potential of selected microalgae (*Spirulina* sp., *Nannochloropsis* sp. and *Chlorella* sp.) as feed for *M. rosenbergii* juvenile was conducted with amino acid and fatty acid profile. From the results, methionine and tryptophan in amino acid were not detected in *Spirulina* sp., *Nannochloropsis* sp. and *Chlorella* sp. in all four different levels. The arginine, isoleucine, leucine, alanine, aspartic acid and glutamic acid level were high with increasing level of *Spirulina* sp. *Nannochloropsis* sp. and *Chlorella* sp. showed high percentage in leucine, lysine and arginine. For fatty acid analysis, only *Spirulina* showed a small portion of γ -linolenic acid (GLA). *Chlorella* showed no detection on Eicosapentaenoic acid (EPA) when compared to *Spirulina* and *Nannochloropsis*.

For second experiment, a series of experiments were conducted to study the potential of the feed formulations for freshwater prawn *Macrobrachium rosenbergii* juvenile based on three types of microalgae (*Spirulina* sp., *Nanochloropsis* sp. and *Chlorella* sp.). An experiment in triplicate with prawn (2.5-2.6 g) were fed until an apparent satiation with 32% crude protein diet containing three types of algae with different level (Diet 1= 0 %, Diet 2=2.5 %, Diet 3= 5 % and Diet 4= 10%) in varying algae levels ranging from 0, 2.5, 5 and 10% respectively for 56 days. At the end of the feeding trial, prawn given Diet 4 containing 10 % of *Spirulina* sp. gave the optimum growth which also contributed to the best feed efficiency. There was no significant difference in FCR in all treatments ($p<0.05$). There were significant differences ($p<0.05$) in growth rate among treatments and similarly prawns fed Diet 4 in *Spirulina* sp. had the highest weight gain (WG) and specific growth rate (SGR) 5.53 g and 2.05 %/day, respectively. The percentage survival of *M. rosenbergii* ranged from 63-83 %. This study also demonstrated that the nutrient from *Spirulina* sp. is more suitable to be

supplemented in prawn diet compared with *Nannochloropsis* sp. and *Chlorella* sp. The used of Cr₂O₃ marker is reliable for digestibility study.

Spirulina sp., *Nannochloropsis* sp. and *Chlorella* sp. were analyzed for antimicrobial activity to determine the potential of the algae against the selected pathogens. *Chlorella* sp. had successfully inhibited *Aeromonas hydrophila* followed by *A. salmonicida* and *Vibrio alginolyticus*. The maximum inhibition zone by *Chlorella* sp. were 14.67 mm against *A. hydrophila*, followed by 11.33 mm against *V. alginolyticus* and 10.00 mm against *A. sobria*. Studies on the effect of *Spirulina* sp, *Nannochloropsis* sp. and *Chlorella* sp. on prawn challenged with *A. hydrophila* for 48 hours showed there are no significant difference among the different treatments. Prawn's mortality was 100 % in control group.

Spirulina sp. with 10% algae level showed a good potential to be as a feed supplement for *M. rosenbergii* juveniles. *Spirulina* sp. contained a variety of amino acids and fatty acids to support good growth performance, digestibility and disease resistance as compared to *Nannochloropsis* sp. and *Chlorella* sp.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai
memenuhi keperluan untuk ijazah Doktor Falsafah

**PENILAIAN MIKROALGAL TERPILIH (*Spirulina* sp., *Nannochloropsis* sp.
DAN *Chlorella* sp.) UNTUK MAKANAN SUPLEMEN JUVENIL
Macrobrachium rosenbergii (de Man)**

Oleh

SUNIZA ANIS BINTI MOHAMAD SUKRI

Februari 2017

Pengerusi : Profesor Madya Che Roos Bin Saad, PhD
Fakulti : Pertanian

Satu siri kajian telah dijalankan untuk mengkaji potensi makanan rumusan pilihan untuk juvenile udang galah *Macrobrachium rosenbergii*. Potensi mikroalga pilihan (*Spirulina* sp., *Nannochloropsis* sp. dan *Chlorella* sp.) telah dikaji sebagai makanan untuk juvenil *M. rosenbergii* dengan mengkaji profil asid amino dan asid lemak. Daripada keputusan, metionina dan triptofan dalam asid amino tidak dikesan dalam *Spirulina* sp., *Nannochloropsis* sp. and *Chlorella* sp. dalam keempat-empat tahap yang berbeza. Arginina, isoleusina, leusina, alanina, asid aspartik dan glutamik asid mempunyai paras yang tinggi terhadap peningkatan tahap dalam *Spirulina* sp. *Nannochloropsis* sp. dan *Chlorella* sp. menunjukkan peratusan yang tinggi dalam leusina, lisina dan arginina. Untuk analisa asid lemak, hanya *Spirulina* sp. menunjukkan sebahagian kecil asid γ -linoletik (GLA). *Chlorella* sp. tidak menunjukkan kehadiran asid eicosapentaenoik (EPA) berbanding *Spirulina* sp. dan *Nannochloropsis* sp.

Untuk kajian kedua, satu siri kajian telah dijalankan bagi mengkaji potensi makanan rumusan untuk juvenile udang galah *M. rosenbergii* berdasarkan tiga jenis mikroalga (*Spirulina* sp., *Nannochloropsis* sp. dan *Chlorella* sp.). Satu kajian dijalankan dalam tiga replikasi, dengan menggunakan udang (2.5-2.6 g) yang diberi makan sehingga kenyang dengan 32 % diet protein mentah yang mengandungi tiga jenis peringkat alga (Diet 1 = 0 %, Diet 2 = 2.5 %, Diet 3 = 5 % dan Diet 4 = 10 %) dalam pelbagai peringkat alga dari 0, 2.5, 5 dan 10 % masing-masing selama 56 hari. Pada akhir kajian, makanan udang yang diberikan Diet 4 mengandungi *Spirulina* 10% memberi pertumbuhan yang optimum yang juga menyumbang kepada kecekapan makanan terbaik. Tidak ada perbezaan yang signifikan dalam FCR dalam semua rawatan ($p < 0.05$). Terdapat perbezaan yang signifikan ($p < 0.05$) dalam kadar pertumbuhan antara rawatan dan begitu juga udang diberi makan *Spirulina* sp. Diet 4 mempunyai berat badan (WG) yang tertinggi dan kadar tertentu pertumbuhan (SGR) masing-

masing 5.53 g dan 2.05 %/hari. Peratusan kadar hidup *M. rosenbergii* adalah di antara 63-83 %. Kajian ini juga menunjukkan bahawa nutrien dari *Spirulina* sp. lebih sesuai ditambah dalam diet udang berbanding *Nannochloropsis* sp. dan *Chlorella* sp. Penanda Cr₂O₃ yang digunakan adalah teknik yang boleh dipercayai untuk kajian penghadaman.

Spirulina sp., *Nannochloropsis* sp. dan *Chlorella* sp. telah dianalisa untuk aktiviti antimikrob bagi memeriksa potensi alga terhadap patogen yang dipilih. *Chlorella* sp. telah berjaya merencatkan *Aeromonas hydrophilla* diikuti oleh *A. salmonicida* dan *Vibrio alginolyticus*. Zon perencatan maksimum *Chlorella* sp. terhadap *A. hydrophila* adalah 14.67 mm, diikuti oleh *V. alginolyticus* 11.33 mm dan *A. sobria* 10.00 mm. Kajian cabaran udang galah yang diberi makan diet yang mengandungi *Spirulina* sp., *Nannochloropsis* sp. dan *Chlorella* sp. ke atas udang dengan *A. hydrophilla* selama 48 jam tidak menunjukkan terdapat perbezaan yang signifikan dalam kesemua rawatan tersebut. Kadar kematian 100% diperhatikan dalam rawatan kawalan.

Spirulina sp. dengan peringkat alga 10% berpotensi sebagai makanan tambahan yang sesuai untuk makanan juvenil *M. rosenbergii*. *Spirulina* sp. mengandungi asid amino dan asid lemak yang sesuai untuk menyokong pertumbuhan dan daya tahan penyakit berbanding *Nannochloropsis* sp. dan *Chlorella* sp.

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I certify that a Thesis Examination Committee has met on 8 February 2017 to conduct the final examination of Suniza Anis binti Mohamad Sukri on her thesis entitled "Assessment of Selected Microalgae (*Spirulina* sp., *Nannochloropsis* sp. and *Chlorella* sp.) as Feed Supplement for *Macrobrachium rosenbergii* (de Man) Juveniles" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

AAS	Atomic Absorption Spectrophotometer
ADC	Apparent Digestibility Coefficients
ALA	Alanine
ANOVA	Analysis of Variance
ARA	Arachidonic acid
CFU	Colony-Forming Unit
DHA	Decosahexaenoic acid
DO	Dissolved Oxygen
DW	Dry Weight
EAA	Essential Amino Acid
EPA	Eicosapentaenoic acid
FAME	Fatty Acid Methyl Esters
FCR	Feed Conversion Ratio
FW	Final Weight
GC	Gas Chromatography
GLA	γ -linolenic acid
HPA	Heneicosapentaenoic acid
HPLC	High Performance Liquid Chromatography
HUFA	Highly Unsaturated Fatty Acids
IW	Initial Weight
mt	metric tonnes
%	Percentage
PUFA	Polyunsaturated Fatty Acids
SD	Standard deviation

SGR	Specific Growth Rate
TSA	Tryptic Soy Agar
WG	Weight Gain



CHAPTER 1

INTRODUCTION

1.1 Prawn nutrition

Microalgae is a single-celled algae or phytoplankton represent the natural nutritional base and primary source of nutrient in aquatic food chain and play as nutritional role to marine animals and also in aquaculture (Catarina and Davier, 2012). Microalgae are an indispensable food sources in the commercial rearing of many cultivated species, including all growth stage of bivalve molluscs (clams, oyster, scallops), larval stages of some crustacean species, gastropods (abalone, conch) and very early growth stage of some fish species such as cod, halibut and tilapia (Coutteau, 1996; Catarina and Davier, 2012; Becker, 2013). The algae size ranging from 5 to 25 microns are ideally good for early stages of aquatic animals. They can easily and rapidly grow, stable in diverse environmental conditions, and rich in good nutrient compositions. Algae such as Spirulina sp., Chlorella sp., Tetraselmis sp. and Chaetoceros sp. have been used in aquaculture mostly for nutritional purposes (Muller-Feuga, 2000). Microalgae can be given directly as feed or mixed in formulated diets for early larval stages. This leads to better results than a diet contained of single microalgae (Yamayugi, 1997; Becker, 2013).

Nutritional composition of algae is well documented and mostly contains protein, carbohydrate, lipids and trace nutrients such as vitamins, antioxidants and trace elements (Yaacob et al., 2014). Microalgae with good nutritional value also contains high PUFA, EPA and DHA content, that can provide a high quality nutritional package for different stages of aquatic animals. Combinations of suitable algal species provide a well-balanced diet, which facilitates the larval development and they are source of essential vitamins such as A, B, C, E, folic acid and pantothenic acid (Nakagawa and Montgomery, 2007; Priyadarshani and Rath, 2012; Shanmugapriya and Ramanathan, 2012). Furthermore, they are also rich source of essential amino acid (protein), minerals, carbohydrates and essential fatty acid such as linolenic acid (Quoc and Pascaud, 1996).

The production of live algae will remain a critically important aspect of successful hatchery management into the foreseeable future. Microalgae have been widely used as an aquaculture food as first feeding of fish larvae (Reitan et al., 1997; Spolaore et al., 2006; Cai et al., 2007; Catarina and Davier, 2012) and as green water for Macrobrachium rosenbergii larvae (Cohen et al., 1976). The use of microalgae during the fish feeding process, either directly or through improving the nutritional value of the rotifer can improve the nutritional conditions of the fish larvae (Reitan et al., 1997; Thipot et al., 2016). The cultivation of micro-algae is an inherent part of aquaculture and phytoplankton comprises the base of the food chain in the marine and freshwater environment and also provided energy and nutrient essential for growth and development of marine organism (Cai et al., 2007).

Fish meal is the most expensive ingredient in fish feeding. According to Tacon and Metian (2008), in year 2006, aquaculture was consumed 3.7 million tonnes of fish meal. Many researchers tried to solve the problem and tried to decrease the use of fish meal in aquafeed. They tried to find alternative protein source to replace protein source from fish meal (Grabner, 1985; Lemos et al., 2009) and tried to check the potential of oyster processing by-product to replace fish meal for tilapia fry (Serrano et al., 2016). Another study by Ganon-Naret (2014a) reported that, 10% of *Moringa oleifera* leaf meals as plant protein to replace fish meal in *Lates calcarifer* diet showed an effect on growth performance and health welfare of the fish.

The researchers also tried to find alternative protein source to replace protein from soybean. Study by Maisashvili et al. (2015), was used *Chlorella pyrenoidosa* to replace protein soybean meal in aquaculture. A study by Ganon-Naret (2014b), mung bean seed was used to replace soybean meal protein in the seabass diet. Mung bean seed contain 20 to 25% of protein and amino acid except methionine, cycteine, and tryptophan. El-Saidy and Amal (2011) reported that, soybean meal was replaced with cottonseed meal and showed good growth performance of Nile tilapia. Cottonseed meal also used to replace soybean for growth, feed utilization and blood physiological of juvenile black carp (Hu et al., 2015).

The freshwater prawn, *Macrobrachium rosenbergii* has been popular as a diet for Asia and the Pacific region since the 1960s and the farming of this species has spread throughout the world such as Bangladesh, China, India, Malaysia, Taiwan, Thailand, Vietnam (New, 2002). The freshwater prawn is in high demand in national and international markets (Soundarapandian et al., 2009). According to FAO (2009), five major countries that producing freshwater prawn such as China (56.3%), Thailand (12.5%), India (12.3%), Bangladesh (11%) and Taiwan (4.5%) in year 2006 with total production of 3.2 million tonnes of *M. rosenbergii*. The production of the freshwater prawn in Malaysia reached RM106.9 million (DoF, 2008) to RM139.3 million in 2013 (DoF, 2013).

Giant freshwater prawn can survive in a wide range of salinity between 0 to 18 ppt but this species has some inherent problem such as uneven growth and low harvest yields (FAO, 2004). This current study try to use microalgae such as *Spirulina* sp. *Nannochloropsis* sp. and *Chlorella* sp. to solve the culture problem and tried to increase production of *M. rosenbergii*.

1.2 Objectives

Objectives of this study were described as follow:

1. To determine the profile of amino acid and fatty acid of feed and *Macrobrachium rosenbergii* carcass fed with different microalgae supplemented diets.
2. To determine the growth performance and survival of *Macrobrachium rosenbergii* fed with microalgae supplemented diets.

3. To measure the digestibility of the microalgae supplemented feed in *Macrobrachium rosenbergii* juvenile.
4. To examine the antimicrobial activity and efficacy of feed containing microalgae (*Spirulina* sp., *Nannochloropsis* sp. and *Chlorella* sp.) and following *Aeromonas hydrophila* immersion challenge.

1.3 Justification

Microalga of different species are known as having very high nutritional value (high in energy, amino acid, vitamins, pigments, sterols, PUFAs). The successes of the research will provide new information in the field of fish nutrition. The use of microalgae during the fish-feeding process either directly or through improving the zooplankton nutrition can improve the nutritional conditions of the larvae and also can provide energy and nutrient essential for the growth and development. This research was conducted in order to study the amino acid as well as the fatty acid profiles in various species of algae (*Spirulina* sp., *Nannochloropsis* sp. and *Chlorella* sp.). Growth performance, digestibility, antimicrobial activity and bacterial challenge on these algae also were carried out. These algae species are believed to provide a good balance nutrient as they contain higher protein especially Essential Amino acid (EAA) and unsaturated fatty acids composite of Eicosapentaenoic acid (EPA), Decosahexaenoic acid (DHA) and Arachidonic acid (ARA) for fish growth as compared to single algae species.

Hypothesis:

H_0 : Selected microalgae (*Spirulina* sp., *Nannochloropsis* sp. and *Chlorella* sp.) did not have any effect on feed for *Macrobrachium rosenbergii* juveniles.

H_A : Selected microalgae (*Spirulina* sp., *Nannochloropsis* sp. and *Chlorella* sp.) have positive effect on feed for *Macrobrachium rosenbergii* juveniles.

REFERENCES

- Abdulkadir, S. and Tsuchiya, M. (2008). One-step method for quantitative and qualitative analysis of fatty acids in marine animal samples. *Journal of Experiment Marine Biology and Ecology*, 354:1-8.
- Abdolnabi, S., Ina-Salwany, M. Y., Daud, H. M., Mariana, N. S. and Abdulhadi, Y. M. (2015). Pathogenicity of *Aeromonas hydrophila* in giant freshwater prawn, *Macrobrachium rosenbergii*, cultured in East Malaysia. *Iranian Journal of Fisheries Sciences*, 14(1): 232-245.
- Adisukresno, S. (1980). Management of Grow-out Farm In V. Soesanto, S. Adisukresno and G. L. Escriptor (Eds.), South China Sea fisheries development and coordinating programme. Report of the trainingcourse on prawn farming for Asia and the pacific. Food and Agriculture Organisation of the United Nations, Rome.
- Ahmed, H. A. (2003). Bacterial Flora of Freshwater Prawn, *Macrobrachium rosenbergii* (de Man), Cultured in Concrete Tanks in Saudi Arabia. *Journal of Applied Aquaculture*, 14: 113-124.
- Akhtar, N., Ahmad, M. M., Sarker, N., Mahbub, K. R. and Sarker, M. A. M. (2012). Growth response of *Spirulina platensis* in papaya skin extract and antimicrobial activities of *Spirulina* extracts in different culture media. *Bangladesh Journal of Scientific and Industrial Research*, 47(2): 147-152.
- Akiyama, D. M., Coelho, S., Lawrence, A. L. and Robinson, T. H. (1989). Apparent digestibility of feedstuff by the marine shrimp *Penaeus vannamei* Bonne. *Nippon Suisan Gakkaiishi*, 55(1): 91-98.
- Akiyama, D. M., Dominy, W. G., Lawrence, A. (1992). Penaeid shrimp nutrition. In Fast, A. W., Lester, J. (Eds.), *Marine Shrimp Culture: Principles and Practices*. Elsevier, Amsterdam. Pp. 535-537.
- Aragão, C., Conceição, L. E. C., Dinis, M. T., Fyhn, H. J. (2004). Amino acid pools of rotifers and Artemia under different conditions: nutritional implications for fish larvae. *Aquaculture*, 234: 429-445.
- AOAC (1999). Official methods of analysis. Association of official analytical chemists, Washington.DC.
- AOAC (2000). Official methods of analysis. 15thed., Association of official analytical chemists, Washington.DC.
- Becker, W. (2004). Microalgae for aquaculture. The nutritional value of microalgae for aquaculture. In A. Richmond (Ed.), *Handbook of microalgal culture: Biotechnology and Applied Phycology*. Blackwell Publishing Ltd. Pp. 380-391.

Becker, E. W. (2007). Micro-algae as a source of protein. Biotechnology Advances, 25(2): 207-210.

Becker, E. W. (2014). Microalgae for Aquaculture: Nutritional Aspect. In A. Richmond and Qiang, H. (Ed.), Handbook of microalgal culture: Biotechnology and Applied Phycology. Blackwell Publishing Ltd. Pp. 380-391.

Belay, A., Ota, Y., Miyakawa, K., Shimamatsu, H. (1993). Current knowledge on potential health benefits of Spirulina. Journal of Applied Phycology, 5(2): 235-241.

Belay, A., Kato, T. and Ota, Y. (1996). Spirulina (*Arthrospira*): Potential as an animal feed supplement. Journal of Applied Phycology, 8: 303-311.

Benedito-Palos, L., Navarro, J. C., Sitj-Bobadilla, A., Bell, J. G., Kaushik, S. and Perea-Sanchez, J. (2008). High levels of vegetable oils in plant protein-rich diets fed to gilthead sea bream (*Sparus aurata* L.): growth performance, muscle fatty acid profiles and histological alterations of target tissues. British Journal of Nutrition, 100: 992-1003.

Bhavan, P. S., Devi, V. G., Shanthi, R., Radhakrishnan, S. and Poongodi, R. (2010). Basic Biochemical Constituents and Profiles of Amino Acids in the Post Larvae of *Macrobrachium rosenbergii* Fed with Spirulina and Yeast Enriched Artemia. Journal of Scientific Research, 2(3):539-547.

Bhavani, M., Hareesh, K., Suneetha, Y. and Srinivasulu Reddy, M. (2014). The effect of dietary protein on the growth potentials and nitrogen excretion in freshwater prawn *Macrobrachium rosenbergii*. International Journal of Fisheries and Aquatic Studies, 2(2): 243-248.

Bhowmik, D., Dubey, J. and Mehra, S. (2009). Probiotic Efficiency of *Spirulina platensis*-Stimulating Growth of Lactic Acid Bacteria. American-Eurasian J. Agric & Environ. Science, 6(5): 546-549

Board, N. O. S. (2008). Proposed organic aquaculture standard: fish feed and relative management issues (2008). National Organic Standard Board (NOSB).

Bragagnolo, N. and Rodrigues-Amaya, D. B. (2001). Total Lipid, Cholesterol, and Fatty Acids of Farmed Freshwater Prawn (*Macrobrachium rosenbergii*) and Wild Marine Shrimp (*Penaeus brasiliensis*, *Penaeus schmitti*, *Penaeus kroyeri*). Journal of Food Composition and Analysis, 14: 359-369.

Brinker, A. and Reiter, R. (2011). Fish meal replacement by plant protein substitution and guar gum addition in trout feed, Part I: Effect on feed utilization and fish quality. Aquaculture, 310: 350-360.

Brown, M. R. (1991). The amino-acid and sugar composition of 16 species of microalgae used in mariculture. Journal of Experimental Marine Biology and Ecology, 145: 79-99.

- Brown, M. R., Jeffrey, S. W., Volkman, J. K. and Dunstan, G. A. (1997). Nutritional properties of microalgae for mariculture. Aquaculture, 151: 315-331.**
- Brown, M. R. (2002). Nutritional value of microalgae for aquaculture. In Cru-Su-re, L. E., Ricque-Marie, D., Tapia-Salazar, M., Gaxiola-Cortés, M.G., Siimoes, N. (Eds.), Avances en Nutrición Acuícola VI. Memorias del VI Simposium Internacional de Nutrición Acuícola. 3 al 6 de septiembre del 2002. Cancún, Quintana Roo, México.**
- Brown, M. R. and Blackburn, S. I. (2013). Live Microalgae as feed in aquaculture hatcheries. In Allan and Burnell (Eds.), Advances in Aquaculture Hatchery Technology (pp. 117-158). Woodhead Publishing Limited.**
- Cai, S., Hu, C. and Du, S. (2007). Comparisons of Growth and Biochemical Composition between Mixed Culture of Algae and Yeast and Monocultures. Journal of Bioscience and Bioengineering, 104:391-397.**
- Catarina, G. A. and Cavier, M. F. (2012). Nutritional Value and Uses of Microalgae. (pp. 59-78). In M. Zainal. (Ed.). Aquaculture. InTech. Pp 59-78.**
- Cereuela, R., Guardiola, F. A., Gonçalves, P., Meseguer, J. and Esteban, M. A. (2012). Effects of dietary *Bacillus subtilis*, *Tetraselmis chuii*, and *Phaeodactylum tricornutum*, singularly or in combination, on the immune response and disease resistance of sea bream (*Sparus aurata* L.) Fish and Shellfish Immunology, 33(2): 342-349.**
- Chettri, J. K., Sahu, N. P., Pal, A. K., Reddy, A. K., Kumar, S. and Kumar, V. (2007). Comparative performance of Gamma Amino Butyric Acid (GABA) and 5-Hydroxytryptamine (5-HT) in the diet of larvae and post larvae of giant freshwater prawn, *Macrobrachium rosenbergii*: Effect of dose and route of administration on growth and survival. Aquaculture, 270: 240-248.**
- Cho, C. Y. and Slinger, S. J. (Eds) (1979). Apparent digestibility measurement in feedstuffs for rainbow trout. Proceeding from World Symposium on Finfish Nutrition and Fish Feed Technology, Hamburg.**
- Cho, S. H., Ji, S. C., Hur, S. B., Bae, J., Park, I. S. and Sons, Y. C. (2007). Optimum Temperature and Salinity Conditions for growth of green algae *Chlorella ellipsoidea* and *Nannochloris oculata*. Fish Sci, 73: 1050-1056.**
- Choubert, G., De La None, J. and Laquet, P. (1979). Continuous quantitative automatic collector for fish feces. Progressive Fish Culturist, 41: 64-67.**
- Cleber Bertoldi, F., Sant'Anna, E., Villela da Costa Braga, M. and Barcelos Oliveira, J. L. (2006). Lipids, fatty acids composition and carotenoids of *Chlorella vulgaris* cultivated in hydroponic wastewater. Grasas Y Aceites, 57(3): 270-274.**
- Cohen, D., Finkel, A. and Sussman, M. (1976). On the role of algae in larvaculture of *Macrobrachium rosenbergii*. Aquaculture, 8: 199-207.**

- Colla, L.M., Bertolin, T.E. and Costa, J.A.V. (2004). Fatty acids Profile of Spirulina platensis Grown Under Different Temperature and Nitrogen Concentrations. Z Naturforsch, 59:55-59.**
- Concei^o, L. E. C., Y^ofera, M., Makridis, P., Morais, S. and Dinis, M. T. (2009). Live feeds for early stages of fish rearing. Aquaculture Research, 41(5):613-640.**
- Cousin, M., Crillon, G., Guillaume, J. and Aquacop. (1996). Digestibility of starch in *P. vannamei*: in vivo and in vitro study on eight samples of various origin. Aquaculture, 140: 361-372.**
- Coutteau, P. (1996). Micro-algae. In P. Lavens and P. Sorgeloos (Eds.), Manual on the production and use of live food for aquaculture (pp. 7-48). Food and Agriculture Organization of the United Nations, Rome.**
- D'Abromo, L. R., Sheen, S. S. (1993). Polyunsaturated fatty acid nutrition in juvenile freshwater prawn *Macrobrachium rosenbergii*. Aquaculture, 115: 63-86.**
- Davassi, L. A. (2011). Survival and growth of the Freshwater Prawn *Macrobrachium rosenbergii* in relation to Different Nutrients Composition. Journal of Fisheries and Aquatic Science, 6(6): 649-654.**
- Davis, D.A., Samocha, T. M., Bullis, R. A., Patnaik, S., Browdy, C. L., Stokes, A. D., Atwood, H. L. (2004). Practical diets for *Litopenaeus vannamei* (Boone, 1931): Working Towards Organics and/or All Plant Production Diets. Avances en Nutriciⁿ Acu^cola VII: Memorias del Septimo Simposium Internacional de Nutriciⁿ Acu^cola. Hermosillo, Sonora.**
- Davila, J., Marcial-Martinez, L. M., Viana, M. T. and Valque-Duhalt, R. (2010). The effect of broccoli in diet on the cytochrome P450 activities of tilapia fish (*Oreochromis niloticus*) during phenol exposure. Aquaculture, 304: 58-65.**
- Dhert, P., 1996. Rotifers. In P. Lavens and P. Sorgeloos (Eds.). Manual on the production and use of live food for aquaculture (pp. 66). Food and Agriculture Organization of the United Nations, Rome.**
- Diraman, H., Koru, E., Dibeklioglu, H. (2009). Fatty Acid Profile of Spirulina platensis Used as a Food Supplement. The Israeli Journal of Aquaculture - Bamidgeh, 61(2): 134-142.**
- Divya, K. R., Isamma, A., Arunjith, T. S., Sureshkumar, S. and Krishnakumar, V. (2014). Effect of Enriched Artemia franciscana on Production, Survival, Growth and Biochemical Composition of the Freshwater Fish Catla catla (Hamilton, 1922). International Journal of Recent Biotechnology 2(3):15-24.**
- DoF. (2008). Department of Fisheries Malaysia. Perangkaan 2008. Department of Fisheries Malaysia. <http://www.dof.gov.my/en/annual-fisheries-statistics-20082>.**

DoF. (2013). Department of Fisheries Malaysia. Perangkaan 2013. Department of Fisheries Malaysia. <http://www.dof.gov.my/senarai-perangkaan-perikanan-tahunan-2013>.

D'Souza, F. M. L., Lecossois, D., Heasman, M. P., Diemar, J. A., Jackson, C. J. and Pendrey, R. C. (2000). Evaluation of centrifuged microalgae concentrates as diets for *Penaeus monodon* Fabricius larvae. *Aquaculture Research*, 31(8): 661-670.

D'Souza, F. M. L., Knuckey, R. M., Hohmann, S. and Pendrey, R.C. (2002). Flocculated microalgae concentrates as diets for larvae of the tiger prawn *Penaeus monodon* Fabricius. *Aquaculture Research*, 8(2): 113-120.

Eco Nurcahya Dewi, Ulfah, A. and Maiirwan, M. (2016). The Effect of Different Treatments to the Amino Acid Contents of Micro Algae Spirulina sp. *Aquatic Procedia* 7: 59-65.

El-Saidy, D. M. S. D. and Amal, S. S. (2011). Effect of partial and complete replacement of soybean meal with cottonseed meal on growth, feed utilization and haematological indexes for mono-sex male Nile tilapia, *Oreochromis niloticus* (L.) fingerlings. *Aquaculture Research*, 42(3): 351-359.

Equerro, J. M., García Carreño, F. L., Civera, R. and Haard, N. (1997). pH-stat method to predict protein digestibility in white shrimp (*Penaeus vannamei*) *Aquaculture*, 157: 251-262.

FAO. (2009). The State of World Fisheries and Aquaculture 2008. Food and Agricultural Organization, Rome.

Fernández-Gimenes, A. V., Cristina Dávila, A., María Velurtas, S. and Lino Fenucci, J. (2009). In vivo and In vitro Protein Digestibility of Formulated Feeds for *Artemesia longinaris* (Crustacea, Penaeidae). *Brasilian Archives of Biology and Technology*, 52(6): 1379-1386.

Folch, J., Lees, M., Stanley, S. G. H. (1957). A simple method for the isolation and pure purification of total lipides from animal tissues. *Journal of Biochemistry*, 226: 497-509.

Fowden, L. (1952). The Composition of the Bulk Protein of Chlorella. *Biochemical Journal*, 50: 355-358.

Frappaolo, P. J. and Guest, G. B. (1986). Regulating status of tetracyclines, penicillin and other antibacterial drugs in animal feeds. *Journal of Animal Science*, 62: 86-92.

Freire, I., Cortina-Burguelo, A., Grille, P., Arias, M. A., Abellán, E. Segura, M., Sousa, F. W. and Otero, A. (2016). *Nannochloropsis limnetica*: A freshwater microalgae for marine aquaculture. *Aquaculture*, 459: 124-130.

Furukawa and Tsukahara, H. (1966). On the acid digestion method for the determination of chromic oxide as an index substance in the study of digestibility of fish feed. *Bulletin of The Japanese Society of Scientific Fisheries*, 32: 502-508.

Ganon-Naret, E. S. (2014a). Utilization of *Moringa oleifera* leaf meals as plant protein sources at different inclusion levels in fish meal based diets fed to *Lates calcarifer*. *International Journal of the Bioflux Society*, 6(2): 158-167.

Ganon-Naret, E. S. (2014b). Use of raw and heat-treated mung bean seeds (*Phaseolus aureus*) as replacement for soybean meal protein in the diets for sea bass, *Lates calcarifer* fingerlings in tank: effects on growth performance, nutrient utilization and survival rate. *International Journal of the Bioflux Society*, 7(6): 458-468.

Glencross, B. D. and Smith, D. M. (1997). Comparison of triacylglycerols, esterified and free fatty acids as neutral lipid sources in the diet of the prawn *Penaeus monodon*. *Aquaculture*, 159: 67-85.

Glencross, B. D., Smith, D. M., Thomas, M. R. and Williams, K. C. (2002). Optimising the essential fatty acids in the diet for weight gain of the prawn, *Penaeus monodon*. *Aquaculture*, 204:85-99.

Grabner, M. (1985). An in vitro method for measuring protein digestibility of fish feed components. *Aquaculture*, 48: 97-110.

Guérin-Faublè, V., Delignette-Muller, M. L., Vigneulle, M. and Flandrois, J. -P. (1996). Application of a modified disc diffusion technique to antimicrobial susceptibility testing of *Vibrio anguillarum* and *Aeromonas salmonicida* clinical isolates. *Veterinary Microbiology*, 51: 137-149.

Gurure, R. Atkinson, J. and Moccia, R. D. (2007). Amino acid composition of Arctic charr, *Salvelinus alpinus* (L.) and the prediction of dietary requirement for essential amino acids. *Aquaculture Nutrition* 13: 266-272.

Habashy, M. M. (2009). Growth and body composition of juvenile freshwater prawn, *Macrobrachium rosenbergii*, Fed Different Dietary Protein / Starch Ratios. *Global Veterinaria*, 3(1): 45-50.

Habib, A., Das, N. G. and Hossain, M. B. (2014). Growth performance and survival rate of *Macrobrachium rosenbergii* (De Man 1979) larvae using different doses of probiotics. *Pakistan Journal of Biological Science*, 17(7): 920-924.

Hanel, H., Broekman, D., de Graaf, S. and Schnack, D. (2007). Partial replacement of fish meal by lyophilized powder of the microalgae *Spirulina platensis* in pacific white shrimp diets. *The Open Marine Biology Journal*, 1: 1-5.

Hanninen, M., Oivanenb, P. and Hirvelä-koskic, V. (1997). *Aeromonas* species in fish, fish-eggs, shrimp and freshwater. *International Journal of Food Microbiology*, 34(1): 17-26.

- Hanley, F. (1987). The digestibility of feedstuffs and the effects of feeding selectivity on digestibility determinations in tilapia (*Oreochromis niloticus*). *Aquaculture*, 66:163-179.
- Healy, J. M. (2001). The mollusca. In: D.T. Anderson, ed. *Invertebrate Zoology*. Oxford University Press, South Melbourne. In Gosling, E. (Ed), *Bivalve Culture, in Bivalve Molluscs* (pp. 147-155). *Biology, Ecology and Culture*, Blackwell Publishing Ltd, Oxford, UK.
- Hu, H. and Gao, K. (2006). Response of growth and fatty acid compositions of *Nannochloropsis* sp. to environmental factors under elevated CO₂ concentration. *Biotechnol Lett*, 28:987-992.
- Hu, Y., Huang, Y., Feng, F., Zhong, L., Ai, G., Tian, T. and Wen, H. (2015). Effect of soybean meal replacement by cottonseed meal on growth, feed utilization and some blood physiological/biochemical indices of juvenile black carp, *Mylopharyngodon piceus*. *Aquaculture Research*, 46(10): 2490-2500.
- Huang, M. T. F., Eble, A. F. and Hammen, C. S. (1981). Immune response of the prawn, *Macrobrachium rosenbergii*, to bacteria infection. *Journal of Invertebrate Pathology*, 38: 213-219.
- Hulatt, C. J., Wijffels, R. H., Bolla, S. and Kiron, V. (2017). Production of Fatty Acids and Protein by *Nannochloropsis* in Flat-Plate Photobioreactors. *PloS ONE*, 12(1): 1-17.
- Ide, N. F. and Fitzpatrick, S., 1999. Marine microalgae as a source of ω3 fatty acids. <http://www.irishscientist.ie/GITIF114.htm> (3 March 2011).
- Jaime-Ceballos, B. J., Hernández-Llamas, A., García-Galano, T. and Villarreal, H. (2006). Substitution of *Chaetoceros muelleri* by *Spirulina platensis* meal in diets for gricultural Research Communication Centre *Litopenaeus schmitti* larvae. *Aquaculture*, 260: 215-220.
- Jakhar, V., Sihag, R. C. and Gahlawat, S. K. (2013). Survey and Characteristic of Disease in Giant Freshwater Prawn (*Macrobrachium rosenbergii* II De Man) in Haryana. *Indian Journal of Animal Research*, 47(6): 479-485.
- Janda, J.M., Abbott, S.L. (2010). The genus *Aeromonas*: taxonomy, pathogenicity, and infection. *Clin. Microbiol. Rev.*, 23: 35-73.
- Jean, H. B. and Sung, B. H. (2011). Selection of Suitable of Chlorella, *Nannochloris* and *Nannochloropsis* in High- and Low-Temperature Season for Mass Culture of the Rotifer *Brachionus plicatilis*. *Fisheries and Aquatic Sciences*, 14(4):323-332.
- Jobling, M. 1994. Fish bioenergetics. *Fish and fishery series, volume 13*. Chapman and Hall, London, UK.

- Johnson, M. M., Swan, D.D., Surette, M, E., Stegner, J., Chilton, T., Fonteh, A. N. and Chilton, F. H. (1997). Dietary Supplementation with Linolenic Acid Alters Fatty Acid Content and Eicosanoid Production in Healthy Humans^{1,2}. Journal of Nutrition: 1435-1444.**
- Jones, P. L., and De Silva, S. S. (1997). Influence of differential movement of the marker chromic oxide and nutrients on digestibility estimations in the Australian freshwater crayfish *Cherax destructor*. Aquaculture, 154: 323-336.**
- Ju, Z. Y., Deng, D. and Dominy, W. (2012). A defatted microalgae (*Haematococcus pluvialis*) meal as a protein ingredient to partially replace fishmeal in diets of Pacific white shrimp (*Litopenaeus vannamei*, Boone, 1931). Aquaculture, 354: 50-55.**
- Kagan, M. L. and Matulka, R. A. (2015). Safety assessment of the microalgae *Nannochloropsis oculata*. Toxicology Reports, 2: 617-623.**
- Kanaawa, A. S., Teshima, S., Tokiwa, S. and Ceccaldi, H. J. (1979). Requirement of Prawn, *Penaeus japonicus* for essential fatty acids. Mem. Fac. Of Fish Kagoshima Univ., 28: 17-20.**
- Katrina, L. P., Peter, D. N. and George, D. J. (2002). Lipid and fatty acid composition of the mantle and digestive gland of four Southern Ocean squid species: implications for food-web studies. Antarctic Science, 14: 212-220.**
- Khan, M. S., Ang, K. J., Ambak, M. and Saad, C. R. (1994). Optimum dietary protein requirement of a Malaysian catfish, *Mystus numurus*. Aquaculture, 112: 227-235.**
- Khan, M., Shobhal, C. J., Mohan, I. K., Rao, N. M. U., Prayag, A. and Kutala, V. K. (2006). Spirulina attenuates cyclosporine-induced nephrotoxicity in rats. Journal of Applied Toxicology, 26(5): 444-451.**
- Khatoon, H., Banerjee, S., Yusoff, F. M. and Shariff, M. (2013). Use of microalgal-enriched *Diaphanosoma celebensis* Stingelin, 1900 for rearing *Litopenaeus vannamei* (Boone, 1931) postlarvae. Aquaculture Nutrition, 19(2): 163-171.**
- Khatoon, H., Nora'ira, A. R., Sanjoy, B., Na'urah, H., Siti, S. S., Nur, H. Z., Fathurrahman, L., Siti, H. A. H. and A'i'ah, E. (2014). Effect of different salinities and pH on the growth and proximate composition of *Nannochloropsis* sp. and *Tetraselmis* sp. isolated from South China Sea cultured under control and natural condition. International Biodeterioration & Biodegradation, 95: 11-18.**
- Keysami, M. A. and Mohammadpour, M. (2013). Effect of *Bacillus subtilis* on *Aeromonas hydrophila* infection resistance in juvenile freshwater prawn, *Macrobrachium rosenbergii* (de Man). Aquaculture International, 21(3):553-562.**

- Kiran, R. B. P., Rajendran, K. V., Jung, S. J. and Oh, M. J. (2002). Experimental susceptibility of different life-stages of the giant freshwater prawn, *Macrobrachium rosenbergii* (de Man), to white spot syndrome virus (WSSV). *Journal of Fish Diseases*, 25: 201-207.
- Krienitz, L., Hepperle, D., Stich, H. B. and Weiler, W. (2000). *Nannochloropsis limnetica* (Eustigmatophyceae), a new species of picoplankton from freshwater. *Phycologia*, 39: 219-227.
- Krienitz, L. and Wirth, W. (2006). The high content of polyunsaturated fatty acids in *Nannochloropsis limnetica* (Eustigmatophyceae) and its implication for food web interactions, freshwater aquaculture and biotechnology. *Limnologica*, 36: 204-210.
- Kaushik, P. Chauhan, A. (2008). In vitro antimicrobial activity of laboratory grown culture of *Spirulina platensis*. *Indian Journal of Microbiology* 48(3): 348-352.
- Lakshmi, B., Viswanath, B. and Gopal, D. V. R. S. (2013). Probiotic as Antiviral Agents in Shrimp Aquaculture. *Journal of Pathogen*: 1-13.
- Lemos, D., Lawrence, A. L. and Siccardi, A. J. (2009). Prediction of apparent protein digestibility of ingredients and diets by in vitro pH-stat degree of protein hydrolysis with species-specific enzymes for juvenile Pacific white shrimp *Litopenaeus vannamei*. *Aquaculture*, 295: 89-98.
- Li, R., Yokota, A., Sugiyama, J., Watanabe, M., Hiroki, M. and Watanabe, M.M. (1998). Chemotaxonomy of planktonic cyanobacteria based on non-polar and 3-hydroxy fatty acid composition. *Phycological Research*, 46(1): 21-28.
- Lijon, M. B., Khatun, M. M., Islam, A., Khatun, M. M. and Islam, M. A. (2015). Detection of multidrug resistance *Aeromonas hydrophila* in farm raised fresh water prawns. *Journal of Advanced Veterinary and Animal Research*, 2(4): 469-474.
- Lober, M. and Eng, C. (2009). Effect of microalgae concentration on larval survival, development and growth of an Australian strain of giant freshwater prawn *Macrobrachium rosenbergii*. *Aquaculture*, 289: 95-100.
- Lubelleno, E., Gibson, O., Zmora, O. and Sukenik, A. (1995). Potential advantages of green algae (*Nannochloropsis* sp.) for rotifer (*Brachionus plicatilis*) culture. *Aquaculture*, 133: 295-309.
- Macias-Sancho, J., Poersch, L. H., Bauer, W., Romano, L. A., Wasielesky, W. and Tesser, M. B. (2014). Fishmeal substitution with *Arthrospira* (*Spirulina platensis*) in practical diet for *Litopenaeus vannamei*: Effect on growth and immunological parameters. *Aquaculture*, 426: 120-125.
- Madkour, F. F., Kamil, W., Nasr, H. (2012). Production and nutritive Value of *S. platensis* in Reduced Cost Medi. National Institute of Oceanography and Fisheries. *Egyptian Journal of Aquatic Research*, 38: 51-57.

- Maisashvili, A., Henry, B., James, R., David, A., Tryon, W. and Merit, D. (2015).**
The value of whole algae and lipid extracted algae meal for aquaculture. Algae Research, 8:133-142.
- Mala, R., Sarojini, M., Saravanababu, S. and Umadevi, G. (2009).** Screening for antimicrobial activity od crude extract of *Spirulina platensis*. Journal of cell and Tissue Research, 9(3): 1951-1955.
- Maliwat, G. C., Velasque, S., Robil, J. L., Chan, M., Traifalgal, R. F., Tayamen, M. and Raga, J. A. (2017).** Growth and immune response of giant freshwater prawn *Macrobrachium rosenbergii* (De Man) postlarvae fed diets containing *Chlorella vulgaris* (Beijerinck). Aquaculture Research, 48: 1666-1676.
- Maria, C. M., Makoto, M., Jesu, V. V., Claudio, C. O. and Nilson, E. S. (2008).** Comparative analysis of eight esterification methods in the quantitative determination of vegetable oil fatty acid methyl esters (FAME). Journal of the Brazilian Chemical Society, 19(8): 1475-1483.
- Mary, A. M. A., Raquel, A. G. B., Wilsonne, A. K. T. C., Jaliyah, J. E. L., Khurt, C. S. R. and Mary, B. B. M. (2016).** Immune response of *Macrobrachium rosenbergii* immersed in aqueous extract of *Gracilaria edulis* challenged with white spot syndrome virus. AACL Bioflux, 9(2): 215-226.
- McGoogan and Reigh. (1996).** Apparent digestibility of selected ingredients in red drum (*Sciaenops ocellatus*) diets. Aquaculture, 141: 233-244.
- Meng-Umphan, K. (2009).** Growth performance, sex hormone levels and maturation ability of pla poa (*Pengasius bocourti*) fed with *Spirulina* supplementary pellet and hormone application. International Journal of Agriculture & Biology, 11:458-462.
- Metcalfe, L. D., Schmit, A. A. and Pelka, J. R. (1996).** Rapid Preparation of Fatty Acid Esters from Lipids for Gas Chromatographic Analysis. Analytical Chemistry, 38(3):514-515.
- Milena, G., Edilmar, C., Roberto, C., Denis, R. and Luis, R. M. (2013).** Apparent carbohydrate and lipid digestibility of feed for whiteleg shrimp, *Litopenaeus vannamei* (Decapoda: Penaeidae, cultivated at different salinities. Revista de Biología Tropical, 61(3): 1201-1213.
- Mitra G., Mukhopadhyay P.K. and Chattopadhyay D.N. (2005).** Nutrition and feeding in freshwater prawn (*Macrobrachium rosenbergii*) farming. Aquatic Feeds: Formulation and Beyond, 2: 17-19.
- Mostary, S., Rahman, M. S., Mandal, A. S. M. S., Hasan, K. M. M., Rehena, Z. and Basar, S. M. A. (2010).** Culture of *Brachionus plicatilis* feeding with powdered dried *Chlorella*. The Bangladesh Veterinarian, 27(2):91-98.
- Mühling, M., Belay, A. and Whitton, B. A., 2005.** Variation in fatty acid composition of *Arthrospira* (*Spirulina*) strains. J Appl Phycol, 17:137-146.

- Muller-Feuga, A.** 2000. The role of microalgae in aquaculture: situation and trends. *Journal of Applied Phycology*, 12: 527-534.
- Muller-Feuga, A., Robert, R., Cahu, C., Robin, J. and Divanach, P.** (2003a). *Uses of Microalgae in Aquaculture*. In J. G. St̄ttrup and L. A. McEvoy (Eds.), *Live Feeds in Marine Aquaculture*. Blackwell Science Ltd, Oxford, UK.
- Muller-Feuga, A., Moal, J. and Kaas, R.** (2003b). *The Microalgae of Aquaculture*, in *Live Feeds in Marine Aquaculture*. In J. G. St̄ttrup and L. A. McEvoy (Eds.), *Live Feeds in Marine Aquaculture*. Blackwell Science Ltd, Oxford, UK.
- Mustafa, Md. G. and Nakagawa, H.** (1995). A review: dietary benefits of algae as an additive in fish feed. *Israeli Journal of Aquaculture-Bamidgeh*, 47: 155-162.
- Nakagawa, H. and Gome-Dia, G.** (1995). Usefulness of Spirulina sp. Meal as feed additive for giant fresh water prawn, *Macrobrachium rosenbergii*. *Suisan-Oshoku*, 7:521-526.
- Nakagawa, H. and Montgomery, W. L.** (2007). Algae. In: Nakagawa, H., sato, M. and Gatlin III, D. M. (Ed.), *Dietary supplements for the health and quality of cultured fish*. CABI International, : 133-167.
- New, M. B., 1995.** Status of freshwater prawn farming: a review. *Aquaculture Research*, 26: 1-54.
- New, M. B. (2000).** History and Global Status of Freshwater Prawn Farming. In M. B. New, W. C. Valenti, J. H. Tidwell, L. R. D'Abramo and M. N. Kutty(ed.), *Freshwater Prawns: Biology and Farming* (pp. 1-17). Wiley-Blackwell, Oxford, UK.
- New, M.B. (2002).** Farming freshwater prawn. A manual for the culture of giant river prawn (*Macrobrachium rosenbergii*). FAO fisheries technical paper, vol 428. FAO, Rome, Italy.
- New, M. B. (2004).** Cultured Aquatic Species Information Programme. *Macrobrachium rosenbergii*. FAO Fisheries and Aquaculture Department. FAO, Rome, Italy.
- New, M. B. and Kutty, M.N. (2010).** Commercial freshwater prawn farming and enhancement around the world. In M. B. New, W. C. Velenti, J. H. Tidwell, L. R. D' Abramo and M. N. Kutty (Eds.), *Freshwater Prawns Biology and Farming* (pp: 18-34). Wiley-Blackwell Publishing.
- Ng, W. K. and Hung, S. S. O. (1994).** Amino acid composition of whole body, egg and selected tissues of white sturgeon (*Acipenser transmontanus*). *Aquaculture*, 126: 329-339.
- Norambuena, F., Hermon, K., Skrypcyk, V., Emery, J. A., Sharon, Y., Beard, A. and Turchini, G. M. (2015).** Alage in Fish Feed: Performances and Fatty Acid Metabolism in Juvenile Altantic Salmon. *PloS ONE*, 10(4): 1-17.

- les, S. and Pire, R. (2001). Fatty acid composition of Chlorella and Spirulina microalgae species. *Journal of AOAC International*, 84:1708-1714.
- O□demir, G., Karabay, N. U., Dalay, M. C. and Pa□arbasi, B. (2004). Antibacterial activity of volatile components and various extracts of *Spirulina platensis*. *Phytotherapy Research*, 18:754-757.
- Patterson, G. M. L., Larsen, L. K. and Moore, R. R. E. (1994). Bioactive natural product from blue-green algae. *Journal of Applied Phycology*, 6: 151-157.
- Piya, R. and Madhumita, M. (2013). Role of algal mixture in Food intake of *Macrobrachium rosenbergii* during larval development. *Indian Journal of Geo-Marine Sciences*, 42(5): 647-652.
- Praksh, M. and Karmagam, N. (2013). A study on bacterial flora associated with fresh water prawn, *Macrobrachium rosenbergii*. *International Journal of Current Research and Academic Review*, 1(1): 1-16.
- Priyadarshani, I. and Rath, B. (2012). Bioactive Compound from Microalgae and Cyanobacteria: Utility and Applications. *International Journal of Pharmaceutical Sciences and Research*, 3(11): 4123-4130.
- Promya, J. and Chitmanat, C. (2011). The effect of *Spirulina platensis* and *Cladophora* algae on the Growth Performance, Meat Quality and Immunity Stimulating Capacity of the African Sharptooth Catfish (*Clarias gariepinus*). *International Journal of Agriculture □ Biology*, 13:77-82.
- Quoc, K.P. and Pascaud, M. (1996). Effect of dietary gammalinolenic acid on the tissue phospholipid fatty acid composition and the synthesis of eicosanoids in rats. *Annals of Nutrition and Metabolism*, 40:99-108.
- Radhakrishnan, S., Saravana, B. P., Seenivasan, C. and Muralisankar, T. (2015). Effect of dietary replacement of fishmeal with *Chlorella vulgaris* on growth performance, energy utilization and digestive enzymes in *Macrobrachium rosenbergii* post larvae. *International Journal of Fisheries and Aquaculture*, 7(5): 62-70.
- Rania, M. A. A. and Hala, M. T. (2008). Antibacterial and antifungal activity of cyanobacteria and green microalga. Evaluation of medium component by plackett-Burman Design for antimicrobial activity of *Spirulina platensis*. *Global Journal of Biotechnology and Biochemistry*, 3(1):22-31.
- Rasool, M and Sabrina, E. P. (2009). Appraisal of immunomodulatory potential of *Spirulina fusiformis*: an in vivo and in vitro study. *J nat Med*, 63: 169-175.
- Rasoarahona, J. R. E., Barnathan, G., Bianchini, J. and Gaydou, E. M. (2005). Influence of season on the lipid content and fatty acid profiles of the three tilapia species (*Oreochromis niloticus*, *O. macrochir* and *Tilapia rendalli*) from Madagascar. *Food Chemistry*, 91: 683-694.

- Rees, M. W. (1946). The estimation of threonine and serine in protein. Biochemical Journal, 40: 632-640.**
- Reitan, K. I., Rainu□o, J. R., □ie, G. and Olsen, Y. (1997). A review of the nutritional effect of algae in marine fish larvae. Aquaculture, 155: 207-221.**
- Ridha, M. T. and Cru□, E. M. (2001). Effect of biofilter media on water quality and biological performance of the Nile tilapia *Oreochromis niloticus* L. reared in a sample recirculating system. Aquacultural Engineering, 24: 157-166.**
- Robert, R. and G□nard, A. (1999). Bivalve Hatchery Technology: The current situation for the pacific oyster *Crassostrea gigas* and the scallop *Pecten maximus* in France. Aqua Living Resource, 12:121-130.**
- Saad, A. S., Habashy, M. M. and Sharshar, K. M. (2009). Growth response of the freshwater prawn, *Macrobrachium rosenbergii* (De Man) to Diets Having Different Level of Biogen□. World Applied Sciences Journal, 6(4): 550-556.**
- Sandifer, P. A. and Joseph, J. D. (1976). Growth responses and fatty acid composition of juvenile prawns (*Macrobrachium rosenbergii*) fed a prepared ration augmented with shrimp head oil. Aquaculture, 8: 129-138.**
- Sarac, Z., Thaggard, H., Sounders, J., Gravel, M., Neill, A., Cowan, R. T. (1993). Observations on the chemical composition of some commercial prawn feeds and associated growth responses in *Penaeus monodon*. Aquaculture 115:97-110.**
- Serano Jr, A. E., Peralta, E. M. and Tumbokan, B. L. M. (2016). Use of Oyster Processing Byproduct to replace Fish meal and Minerals in the Diet of Nile Tilapia *Oreochromis niloticus* Fry. The Israeli Journal of Aquaculture-Bamidgeh, :1-6.**
- Seong, S. S. (2005). High Performance Liquid Chromatographic Determination Homocysteine and Cystathionine in biological Samples by Derivati□ation with 6-Amino-quinolyl-N-Hydroxysuccinimidyl Carbamate (AQC). Journal of Korean Chemical Society, 49(3):278-282.**
- Shanmugapriya, R. and Ramanathan, T. (2012). Antioxidant and Radial Scavenging Effect of Blue-Green Alga *Spirulina platensis*. International Journal of Pharmaceutical □ Biogical Archives, 3(5): 1086-1090.**
- Siddhanta, A.K, Mody, K.H., Ramavat, B.K., Chauhan, V.D., Garg, H.S., Goel, A.K., Jinandra, D. M., Srivastava, M.N., Patnaik, G.K. and Kamboj, V.P. (1997). Bioactivity of marine organisms: Part VIII-Screening of some marine flora of Western coast of India. Indian Journal Experimental Biology, 35:638-643.**
- Siripornadulsi, W., Thongserm, M. and Siripornadulsi, S. (2014). Pathogenicity of halophilic *Vibrio harveyi* in giant freshwater prawns (*Macrobrachium rosenbergii* De Man). Aquaculture Research, 45(12): 1979-1988.**

Sivakumar, J. and Santhanam, P. (2011). Antipathogenic activity of Spirulina Powder. Recent Research in Science and Technology, 3(4):158-161.

Skrede, A., Mydland, L. T., Ahlstrøm, Ø., Reitan, K. I., Gislerød, H. R. and Øverland, M. (2011). Evaluation of microalgae as sources of digestible nutrients for monogastric animals. Journal of Animal and Feed Science, 20: 131-142.

Smith. (1971). A method for measuring digestibility and metabolic energy of fish feeds. Progressive Fish Culturist, 33: 132-134.

Smith and Lovell. (1971). Determination of apparent protein digestibility in feeds for channel catfish. Transactions of the America Fisheries Society, 102: 831-835.

Soundarapandian, P., Prakash, K.S. and Dinakaran, G.K. (2009). Simple Technology for the Hatchery Seed Production of Giant Palaemonid Prawn *Macrobrachium rosenbergii* (De Man). International Journal of Animal and Veterinary Advances, 1: 49-53.

Spolaore, P., Joannis-Cassan, C., Duran, E. and Isambert, A. (2006). Commercial Applications of Microalgae. Journal of Bioscience and Bioengineering, 101: 87-96.

Srivastava, A., Hamre, K., Stoss, J., Chakrabarti, R. and Tonheim, S. K. (2006). Protein content and amino acid composition of live feed rotifer (*Brachionus plicatilis*): with emphasis on the water soluble fraction. Aquaculture, 254: 534-543.

Sung, H., Hwang, S. and Tasi, F. (2000). Responses of Giant Freshwater Prawn (*Macrobrachium rosenbergii*) to Challenge by Two Strains of *Aeromonas* spp. Journal of Invertebrate Pathology, 76: 278-284.

Sukenik, A. Carmeli, Y. and Berner, T. (1989). Regulation of Fatty Acid composition by Irradiance Level in the Eustigmatophyte *Nannochloropsis* sp. Journal of Phycology, 25(4): 686-692.

Syarifah, E. and Eguchi, M. (2012). Benefits of live phytoplankton, *Chlorella vulgaris*, as a biocontrol agent against fish pathogen *Vibrio anguillarum*. Fisheries Science 78(2): 367-373.

Tacon and Metian (2008). Global overview on the use of fish meal and fish oil in industrially compounded aquafeeds: Trends and future prospects. Aquaculture, 285:146-158.

Teshima, S., Koshio, S. and Ishikawa, M. (2006). Protein Requirement of the Freshwater Prawn *Macrobrachium rosenbergii* Evaluated by Factorial Method. Journal of The World Aquaculture Society, 37(2): 145-153.

Terrafierro, M., Civera-Cerecedo, R., Ibarra-Martínez, L., Goytortúa-Bores, E., Herrera-Andrade, M. and Reyes-Becerra, A. (2010). Apparent digestibility of dry matter, protein, and essential amino acid in marine feedstuffs for juvenile whiteleg shrimp *Litopenaeus vannamei*. Aquaculture, 308:166-173.

- Thibout, V., Mangott, A. and Piroddi, I. (2016). Rotifers enriched with a mixed algal diet promote survival, growth and development of barramundi larvae, *Lates calcarifer* (Bloch). *Aquaculture Report*, 3:147-158.
- Thomson, R., Macpherson, H. L., Riaño, A. and Birkbeck, T. H. (2005). *Vibrio splendidus* biotype 1 as a cause of mortalities in hatchery-reared larval turbot, *Scophthalmus maximus* (L.). *Journal of Applied Microbiology*, 99: 243-250.
- Tibbetts, S. M., Bjornsson, W. J. and McGinn, P. J. (2015). Biochemical composition and amino acid profile of *Nannochloropsis granulata* algal biomass before and after supercritical fluid CO₂ extraction at two processing temperatures. *Animal Feed Science and Technology*, 201: 62-71.
- Tietze, h. W. (2004). *Spirulina. Micro food Macro Blessing*. Fourth Edition. Harald W. Tietze Publishing, Australia.
- Uddin, M., Islam, M. S., Hoq, M. E., and Chowdhury, M. B. R. (1998). Investigation on bacterial flora of farmed freshwater prawn (*Macrobrachium rosenbergii* de Man) in Bangladesh. *Bangladesh J. Fish. res*, 2(2): 171-175.
- Uslu, L. H., Isik, sayin, Durmaç, Goksan and Gokpinar. (2009). The Effect of Temperature on Protein and Amino Acids composition of *S. platensis*. *Journal of Fisheries and Aquatic Sciences*, 26: 139-142.
- Usydus, Z., Słominder-Richert, J., Adamczyk, M. and Słatkowska, U. (2011). Marine and farmed fish in the Polish market: Comparison of the nutritional value. *Food Chemistry*, 126: 78-84.
- Vivekanandhan, G., Hatha, A. A. M. and Lakshmanaperumalsamy, P. (2005). Prevalence of *Aeromonas hydrophila* in fish and prawn from the seafood market of Coimbatore, South India. *Food Microbiology*, 22:133-137.
- Volkman, J. K., Jeffrey, S. W., Nichols, P. D., Rogers, G. W., and Garland, C. D. (1989). Fatty acid and lipid composition of 10 species of microalgae used in mariculture. *Journal of Experimental Marine Biology and Ecology*, 128: 219–240.
- Wacker, A. and von Elert, E. (2003). Food quality controls production of zebra mussel (*Dreissena-polymorpha*). *Oecologia*, 135: 332-338.
- Watanabe, T., Kitajima, C., Fujita, S. (1983). Nutritional values of live organisms used in Japan for mass propagation of fish. A review. *Aquaculture*, 34:115–143.
- Weers, P. and Gulati, R. (1997). Effect of the addition of polyunsaturated fatty acids to be the diet on the growth and fecundity of *Daphnia galeata*. *Freshwater Biology*, 38: 721-729.

□u, J., Yan, B., Teng, Y., Lou, G. and Lu, Z. (2010). Analysis of nutrient composition and fatty acid profiles of Japanese sea bass *Lateolabrax japonicus* (Cuvier) reared in seawater and freshwater. *Journal of Food Composition and Analysis*, 23: 401-405.

Yamayugi, K. (1997). Recent advances in microalgal bioscience in Japan, with special reference to utilization of biomass and metabolites: a review. *Journal of Applied Phycology*, 8: 487-502.

Yano, Y., Hamano, K., Tsutsui, I., Aue-umneoy, D., Ban, M. and Satomi, M. (2015). Occurrence, molecular characterization, and antimicrobial susceptibility of *Aeromonas* spp. in marine species of shrimps cultured at inland low salinity ponds. *Food Microbiology*, 47: 21-27.

Yaakob, Z., Ali, E., Zainal, A., Mohamad, M. and Takniff, M. S. (2014). An overview: biomolecules from microalgae for animal feed and aquaculture. *Journal of Biological Research-Thessaloniki*, 21:6-15.