



UNIVERSITI PUTRA MALAYSIA

***IDENTIFICATION AND MOLECULAR CHARACTERISATION OF
VASCULAR-RELATED DEFENSE GENES AGAINST PAPAYA DIEBACK
DISEASE***

MUHAMMAD HANAM BIN HAMID

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DISEASE**

By

MUHAMMAD HANAM BIN HAMID

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
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May 2017

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Abstract of this thesis presented to the Senate of Universiti Putra Malaysia in fulfilment
of the requirement for the degree of Master of Science

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Chairperson : Associate Professor Janna Ong Abdullah, PhD
Faculty : Biotechnology and Biomolecular Sciences

Malaysia is one of the world's main exporters of papaya with an export value worth RM120 million per year. An outbreak of papaya dieback disease in 2003 lowered the market value up to 60% from the previous year, and remains affected since. *Erwinia mallotivora*, the causal bacteria for papaya dieback disease, enters the papaya plant either through natural openings or wounds, to penetrate into the petioles and stem, and subsequently colonizes the entire vascular system. This results in the appearance of water-soak lesion at the infected region and disruption of the plant's upper meristem region, which produces papaya fruits. To date, the only solution to this problem is by demolishing all the infected papaya plants as it can easily be transmitted to other plants nearby. An alternative approach such as genetic engineering to avoid mass destruction of matured papaya plants is deemed critical. Hence, the aims of this study were to identify potential vascular-related defense genes using bioinformatics approach, isolate and characterize the genes from Eksotika papaya and subsequently to assess the functionality of the genes in transformed papaya seedlings challenged with *E. mallotivora*. This study has successfully identified vascular-related defense genes against papaya dieback disease using bioinformatics approach. After data mining of bioinformatics databases including The Arabidopsis Information Resources (TAIR), *Arabidopsis thaliana* Trans-factor and cis-element Prediction Database (ATTED), Phytozome, and National Center for Biotechnology Information (NCBI), identified five potential genes were then mapped onto the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway of Arabidopsis model plant. Orthologs of the Glycerol kinase (*NHO1*), Pathogenesis-related protein 1 (*PR1b*), Leucine-rich repeat receptor-like serine/threonine-protein kinase (*EFR*), RPM1 interacting protein 4 (*RIN4*) and Mitogen-activated protein kinase 4 (*MPK4*) genes were isolated and fully characterized from *Carica papaya* var. Eksotika I. *NHO1* gene with an estimated size of 1572 base pair (bp) was hypothesized as defense-related gene against pathogen. The similar role was predicted with *PR1b* gene (492 bp), while *EFR* gene (741 bp) was postulated to be involved in pathogen signaling. In addition, *RIN4* gene (540 bp) was suggested to be a member of resistance inducer upon pathogen invasion. *MPK4* gene (1125 bp) was estimated to work in a cascade of mediating various responses against pathogens through different signaling pathway of defense responses. These genes were then sub-

cloned, in a sense orientation, into the pEAQ virus vector for subsequent transformation into papaya seedlings via *Agrobacterium tumefaciens* strain GV3101. The constructs were designated as pEAQ.NHO1, pEAQ.PR1, pEAQ.EFR, pEAQ.RIN4 and pEAQ.MPK4. Leaves of two months old papaya seedlings were infiltrated with *A. tumefaciens* strain GV3101 ($OD_{600} = 1.5$) harbouring each respective constructs. The transformed papaya seedlings were then challenged with *E. mallotivora* ($cfu = 10^6$) and exhibited disease symptoms development as early as day two after infection. Further profiling of the transgene expression in the papaya seedlings via real-time polymerase chain reaction (qRT-PCR) analysis resulted in functional expression against the causal bacteria of this papaya dieback disease. Overexpression of *PR1b* and *NHO1* genes resulted in high expression level in infected plants compared to the uninfected plants. While overexpression of *MPK4* and *RIN4* showed high fold changes compared to control plants, expression of *MPK4* exhibited a repressed defense response via earlier development of symptoms in both infected and uninfected plants. In addition, overexpression of *EFR* showed a down-regulation expression for both infected and uninfected plants. These findings will lay the foundation for subsequent studies in developing a conceivable solution against this papaya dieback disease.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

**PENGENALPASTIAN DAN PENCIRIAN MOLEKUL GEN – GEN
PERTAHANAN BERKAITAN VASKULAR TERHADAP PENYAKIT MATI
ROSOT BETIK**

Oleh

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Mei 2017

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Malaysia merupakan salah satu daripada pengekspor utama betik di dunia dengan nilai eksport bernilai RM120 juta setiap tahun. Serangan wabak mati rosot betik pada tahun 2003 telah mengurangkan nilai pasaran betik sehingga 60% berbanding tahun sebelumnya, dan masih terkesan sehingga sekarang. *Erwinia mallotivora*, bakteria penyebab kepada penyakit mati rosot betik, memasuki pokok betik sama ada melalui bukaan semulajadi atau luka kecederaan, dengan menembusi petiol dan batang, seterusnya mengkoloni keseluruhan sistem vaskular tumbuhan. Proses ini mengakibatkan keadaan lecuh basah pada kawasan yang dijangkiti dan gangguan terhadap kawasan meristem atas tumbuhan, yang menghasilkan buah betik. Sehingga kini, satu – satunya penyelesaian terhadap masalah ini ialah dengan memusnahkan keseluruhan pokok betik yang dijangkiti disebabkan oleh penyakit ini yang senang merebak ke pokok – pokok betik berdekatan. Pendekatan alternatif seperti kejuruteraan genetik bagi mengelakkan kemusnahaan besar kepada pokok – pokok betik matang adalah keperluan yang kritikal. Objektif kajian ini adalah untuk mengenal pasti gen – gen pertahanan berkaitan vascular berpotensi melalui pendekatan bioinformatik, memencil dan mencirikan gen – gen daripada betik Eksotika dan seterusnya melalui menilai kefungsian gen – gen di dalam anak pokok betik terubah suai yang dijangkitkan dengan *E. mallotivora*. Kajian ini telah berjaya mengenal pasti gen – gen pertahanan berkaitan vaskular terhadap penyakit mati rosot betik melalui pendekatan bioinformatik. Melalui pencarian data pengkalan – pengkalan data bioinformatik seperti *The Arabidopsis Information Resources* (TAIR), *Arabidopsis thaliana Trans-factor and cis-element Prediction Database* (ATTED), *Phytozome*, dan *National Center for Biotechnology Information* (NCBI), lima gen berpotensi telah ditemui dan dipetakan pada tapakjalan *Kyoto Encyclopedia of Genes and Genomes* (KEGG) bersandarkan tumbuhan model Arabidopsis. Gen – gen ortolog *Glycerol kinase* (*NHO1*), *Pathogenesis-related protein 1* (*PR1b*), *Leucine-rich repeat receptor-like serine/threonine-protein kinase* (*EFR*), *RPM1 interacting protein 4* (*RIN4*) dan *Mitogen-activated protein kinase 4* (*MPK4*) telah dipencilkan dan dicirikan sepenuhnya daripada *Carica papaya* var. Eksotika I. Gen *NHO1* dengan saiz anggaran 1572 pasangan bes (bp) dihipotesiskan sebagai gen berkait pertahanan melawan patogen. Fungsi yang serupa diramalkan dengan gen *PR1b* (492 bp), sementara gen

EFR (741 bp) terlibat dalam fungsi pengisyaratian patogen. Gen *RIN4* (540 bp) dicadangkan sebagai sebahagian pengaruh kerintangan terhadap serangan patogen. Gen *MPK4* (1125 bp) pula dianggarkan untuk berfungsi di dalam lata pengantara kepada pelbagai gerak balas melawan patogen melalui tapakjalan pengisyaratian gerak balas pertahanan. Gen – gen ini seterusnya disub-klonkan dalam orientasi *sense* ke dalam vektor virus pEAQ untuk transformasi ke dalam anak pokok betik melalui *Agrobacterium tumefaciens* strain GV3101. Konstruk berkenaan dinamakan sebagai pEAQ.NHO1, pEAQ.PR1, pEAQ.EFR, pEAQ.RIN4 dan pEAQ.MPK4. Daun anak pokok betik berusia dua bulan disusupkan dengan *A. tumefaciens* strain GV3101 ($OD_{600} = 1.5$) yang membawa konstruk secara berasingan. Anak – anak betik yang diubah suai kemudiannya dijangkitkan dengan *E. malloivora* ($cfu = 10^6$) dan menunjukkan simptom penyakit seawal dua hari selepas jangkitan. Analisis pemprofilan tahap pengekspresan transgen dalam anak pokok betik melalui kaedah tindakbalas berantai polimerase masanya (qRTPCR) mencadangkan tahap pengekspresan transkrip gen – gen pertahanan yang dikaji adalah berkesan terhadap bakteria penyebab penyakit mati rosot betik. Pengekspresan gen – gen *PR1b* dan *NHO1* menunjukkan tahap pengekspresan yang tinggi di dalam pokok yang dijangkiti pathogen berbanding dengan pokok yang tidak dijangkiti. Sementara pengekspresan gen – gen *MPK4* dan *RIN4* menunjukkan perubahan (*fold change*) yang tinggi berbanding pokok kawalan, pengekspresan *MPK4* menunjukkan ketindasan (*repressed*) gerak balas pertahanan melalui perkembangan gejala penyakit lebih awal dalam kedua – dua pokok yang dijangkiti dan yang tidak dijangkiti patogen. Pengekspresan *EFR* pula menunjukkan pengekspresan yang menurun (*down-regulated*) bagi kedua – dua pokok yang dijangkiti dan yang tidak dijangkiti. Kajian ini memberi asas kepada penyelidikan berterusan bagi membangunkan suatu penyelesaian yang berpotensi melawan penyakit mati rosot betik ini.

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I certify that a Thesis Examination Committee has met on 23 May 2017 to conduct the final examination of Muhammad Hanam bin Hamid on his thesis entitled "Identification and Molecular Characterisation of Vascular-Related Defense Genes Against Papaya Dieback Disease" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

ASGPB	Advanced Studies in Genomics, Proteomics and Bioinformatics
ATTED	<i>Arabidopsis thaliana</i> Trans-factor and cis-element Prediction Database
ATP	Adenosine triphosphate
bp	Base pair
ca.	<i>circa</i> (around or approximately)
CDS	Coding sequence
cfu	Colony-forming unit
C _t	Threshold cycle
DEPC	Diethylpyrocarbonate or Diethyl dicarbonate
DMSO	Dimethyl sulfoxide
DTT	1,4-Dithiothreitol
EDTA	Ethylenediaminetetraacetic acid
EEO	Electroendoosmosis
EMBL	European Molecular Biology Laboratory
gDNA	Genomic deoxyribonucleic acid
HiFi	High fidelity
IPTG	Isopropyl β-D-1-thiogalactopyranoside
KEGG	Kyoto Encyclopedia of Genes and Genomes
MARDI	Malaysian Agricultural Research and Development Institute
mmHg	Millimeter mercury
OD	Optical density (absorbance unit)
PEG 8000	Polyethylene glycol 8000
RNase	Ribonuclease
rpm	Rotation per minute
RT	Reverse-transcriptase
TA	Transcript assembly
TAE	Tris-acetate-EDTA
TAIR	The <i>Arabidopsis</i> Information Resource
TIGR	The Institute for Genomic Research
Tris	Tris(hydroxymethyl)aminomethane
TSS	Transformation-Storage solution
x g	Relative centrifugal force (times gravity)
X-gal	5-bromo-4-chloro-3-indolyl-D-galactopyranoside
YEB	Yeast extract buffer

CHAPTER 1

INTRODUCTION

Papaya (*Carica papaya* L.), a member of dicotyledonous family of Caricaceae is an important tropical fruit in the international market with global production of 9.1 million metric tonnes in 2015 (FAOSTAT, 2015). Found widely in tropical and subtropical countries, papaya is also known as *papaw* or *paw paw* in Australia, and *Mamao* in Brazil. Other than being eaten fresh, papaya is also used occasionally in making jams, preserves and canned in fruit cocktails. The young fruit is eaten as a vegetable or pickled whereas, papain, an enzyme found in the white latex, is used as a meat tenderiser.

Malaysia has been reported as one of the world's main exporters of papaya, ranked at second place after Mexico, with an export value of about RM100-120 million per year (FAOSTAT, 2015). Eksotika papaya cultivar is the main variety grown for commercialization purposes and especially for export market, with a yield of about 50-70 tonnes per hectare over a two-year crop cycle (Chan, 1987; FAOSTAT, 2015).

Papaya industry faced various problems which affected its annual production including the outbreak of the papaya dieback disease in late 2003, and fruit fly quarantine restrictions from China which resulted in 60% reduction of Malaysia papaya export market (Chan and Baharuddin, 2010). The varieties affected by papaya dieback disease include the Eksotika, Solo, Hong Kong and Sekaki (Maktar *et al.*, 2008). Eksotika papaya, a variety known for its export value, dropped to 42,000 tonnes in 2007 as compared to 71,000 tonnes in 2003 upon the incident of papaya dieback disease took place. As papaya dieback disease resulted in enormous quantity loses in the papaya industry, countless resolutions are taken by researchers and farmers to reduce its implications towards papaya production.

The infection of papaya dieback disease was reported to be fast and easily transmitted within a farm from one particular infected tree to another, and this could also be further transmitted to nearby farms in a short amount of time. To date, many treatments reported are ineffective in controlling the disease, and the only advised procedure to stop the spread is by demolishing the whole papaya plantation. This, however, is a massive cost for the farmers to bear.

Hence, in this study, identification and assessment of potential vascular-related defense genes in papaya at molecular level is essential to strategize a solution against papaya dieback disease with the following objectives:

1. to identify potential vascular-related defense genes using bioinformatics approach,
2. to isolate and characterize the vascular-related defense coding sequences (CDS) from Eksotika papaya, and

3. to assess the functionality of the vascular-related defense genes in transformed Eksotika papaya seedlings challenged with *Erwinia mallotivora*.



REFERENCES

- Abdul Majid, N., and Ahmad Parveez, G. K. (2007). Evaluation of Green Fluorescence Protein (GFP) as a Selectable Marker for Oil Palm Transformation via Transient Expression. *Asia Pacific Journal of Molecular Biology and Biotechnology*, **15**(1):1–8.
- Abel, S., and Theologis, A. (1994). Transient Transformation of Arabidopsis Leaf Protoplasts: A Versatile Experimental System to Study Gene Expression. *Plant Journal*, **5**:421–27.
- Afzal, A. J., da Cunha, L., and Mackey, D. (2011). Separable Fragments and Membrane Tethering of Arabidopsis RIN4 Regulate Its Suppression of PAMP-triggered Immunity. *The Plant Cell*, **23**(10):3798–811.
- Aljabali, A. A. A., Sainsbury, F., Lomonossoff, G. P., and Evans, D. J. (2010). Cowpea Mosaic Virus Unmodified Empty Viruslike Particles Loaded with Metal and Metal Oxide. *Small*, **6**(7): 818–821.
- Al-Shanti, N., Saini, A., and Stewart, C. E. (2009). Two-Step versus One-Step RNA-to-CT™ 2-Step and One-Step RNA-to-CT™ 1-Step: Validity, Sensitivity, and Efficiency. *Journal of Biomolecular Techniques*, **20**(3):172–179.
- Aoki, Y., Okamura, Y., Tadaka, S., Kinoshita, K., and Obayashi, T. (2016). ATTED-II in 2016: A Plant Coexpression Database towards Lineage-specific Coexpression. *Plant Cell Physiology*, **57**:e5.
- Armstrong, M. R., Whisson, S. C., Pritchard, L. Bos, J. I. B., Venter, E., Avrova, A. O., Rehmany, A. P., Bohme, U., Brooks, K., Cherevach, I., Hamlin, N., White, B., Fraser, A., Lord, A., Quail, M. A., Churcher, C., Hall, N., Berriman, M., Huang, S., Kamoun, S., Beynon, J. L., and Birch, P. R. J. (2005). An Ancestral Oomycete Locus contains Late Blight Avirulence Gene *Avr3a*, Encoding a Protein that is Recognized in the Host Cytoplasm. *Proceedings of the National Academy of Science of the United State of America*, **102**(21):7766–7771.
- Axtell, M. J., and Staskawicz, B. J. (2003). Initiation of RPS2-specified Disease Resistance in Arabidopsis is coupled to the *AvrRpt2*-directed Elimination of *RIN4*. *Cell*, **112**: 369–377.
- Bechtold, N., and Pelletier, G. (1998). In Planta Agrobacterium-mediated Transformation of Adult *Arabidopsis thaliana* Plants by Vacuum Infiltration. In N. J. Clifton, (Eds), *Methods in Molecular Biology*. (pp. 259–266) Totowa, New Jersey, USA: Humana Press Incorporation.
- Bechtold, N., Ellis, J., and Pelletier, G. (1993). In planta Agrobacterium-mediated Gene Transfer by Infiltration of Adult *Arabidopsis thaliana* Plants. *Comptes Rendus de Academie des Science Paris, Life Science*, **316**:1194–1199.
- Beck, M., Komis, G., Müller, J., Menzel, D., and Samaj, J. (2010). Arabidopsis Homologs of Nucleus- and Phragmoplast-localized Kinase 2 and 3 and Mitogen-

activated Protein Kinase 4 are Essential for Microtubule Organization. *The Plant Cell*, **22**(3):755–71.

Beihaghi, M., Marashi, H., Bagheri, A., and Sankian, M. (2016). Transient Expression of CCL21 Chemokine in Tobacco via Agroinfiltration. *International Journal of Scientific and Engineering Research*, **7**(10):430–436.

Bent, A., Kunkel, B. N., Dahlbeck, D., Brown, K. L., Schmidt, R., Giraudat, J., Leung, J., and Staskawicz, B. J. (1994). *RPS2* of *Arabidopsis thaliana* - A Leucine-rich Repeat Class of Plant-disease Resistance Genes. *Science*, **265**: 1856–1860.

Bethke, G., Pecher, P., Eschen-Lippold, L., Tsuda, K., Katagiri, F., Glazebrook, J., Scheel, D., and Lee, J. (2012). Activation of the *Arabidopsis thaliana* Mitogen-activated Protein Kinase *MPK11* by the Flagellin-derived Elicitor Peptide, *flg22*. *Molecular plant-microbe interactions*, **25**(4):471–80.

Bethke, G., Scheel, D., and Lee, J. (2009). Sometimes New Results Raise New Questions, *Proceedings of the National Academy of Sciences of the United States of America*, **4**(7):672–674.

Bogs, J., and Geider, K. (1999). Migration and Gene Regulation of *Erwinia amylovora* in Host Plants Assayed with the Green Fluorescent Protein. *Proceedings of Eighth International Workshop on Fire Blight 1998 Acta Horticulture*, **489**:365–369.

Breda, C., Sallaud, C., El-Turk, J., Baffard, D., de Kozak, I., Esnault, R., and Kondorosi, A. (1996). Defense Reaction in *Medicago sativa*: A Gene Encoding a Class 10 PR Protein is Expressed in Vascular Bundles. *Molecular Plant-Microbe Interaction*, **9**(8):713–719.

Brock, A. K., Willman, R., Kolb, D., Grefen, L., Lajunen, H. M., Bethke, G., Lee, J., Nurnberger, T., and Gust, A. A. (2010). The *Arabidopsis* Mitogen-activated Protein Kinase Phosphatase *PP2C5* Affects Seed Germination, Stomatal Aperture, and Abscisic Acid-inducible Gene Expression. *Plant Physiology*, **153**(3):1098–111.

Buonauro, R. (2008). Infection and Plant Defense Responses during Plant-bacterial Interaction. In E. A. Barka and C. Clement (Eds.) *Plant-microbe Interactions*. (pp. 169-197) Kerala, India: Research Signpost.

Cai, X. Z., Xu, Q. F., Wang, C. C., and Zheng, Z. (2006). Development of a Virus-induced Gene-silencing System for Functional Analysis of the *RPS2*-dependent Resistance Signalling Pathways in *Arabidopsis*. *Plant Molecular Biology*, **62**(1-2):223–32.

Cazzonelli, C. I., and Velten, J. (2006). An In Vivo, Luciferase-based, Agrobacterium-infiltration Assay System: Implication for Post-transcriptional Gene Silencing. *Planta*, **224**:582–597.

Chan, L. K., and Teo, C. K. H. (2002). Micropropagation of Eksotika, a Malaysian Papaya Cultivar and the Field Performance of the Tissue Culture Derived Clones.

Acta Horticulture, **575**:99–105.

- Chan, Y. K. (1987). Backcross Method in Improvement of Papaya (*Carica papaya L.*). *Malaysian Applied Biology*, **16**:95–100.
- Chan, Y. K., and Baharuddin, A. G. (2010). Rejuvenating the Flagging Papaya Industry in Malaysia: The Role of MAFC. *Proceeding of International Symposium on Papaya. International Society for Horticultural Science Acta Horticulture* 851(2):37–40. Madurai, India, 9-12 December 2008.
- Chanda, B., Venugopal, S. C., Kulshreshtha, S., Navarre, D. A., Downie, B., Vaillancourt, L., Kachroo, A., and Kachroo, P. (2008). Glycerol-3-phosphate Levels are Associated with Basal Resistance to the Hemibiotrophic Fungus *Colletotrichum higginsianum* in Arabidopsis. *Plant Physiology*, **147**:2017–2029.
- Chang, S. S., Park, S. K., and Nam, H. G. (1994). Transformation of Arabidopsis by *Agrobacterium* Inoculation on Wounds. *Plant Journal*, **5**(4):551–558.
- Chen, Q., Lai, H., Hurtado, J., Stahnke, J., Leuzinger, K., and Dent, M. (2014). Agroinfiltration as an Effective and Scalable Strategy of Gene Delivery for Production of Pharmaceutical Proteins. *Advance Techniques in Biology and Medicine*, **1**(1):1–21.
- Chen, X., Equi, R., Baxter, H., Berk, K., Han, J., Agarwal, S., and Zale, J. (2010). A High-throughput Transient Gene Expression System for Switchgrass (*Panicum virgatum L.*) Seedlings. *Biotechnology for Biofuels*, **3**:9.
- Cheng, Y., Zhou, Y., Yang, Y., Chi, Y. J., Zhou, J., Chen, J. Y., Wang, F., Fan, B., Shi, K., Zhou, Y. H., Yu, J. Q., and Chen, Z. (2012). Structural and Functional Analysis of VQ Motif-containing Proteins in Arabidopsis as Interacting Proteins of WRKY Transcription Factors. *Plant Physiology*, **159**(2):810–25.
- Clough, R. C., and Vierstra, R. D. (1997). Phytochrome Degradation. *Plant, Cell and Environment*, **20**:713–721.
- Clough, S. J., and Bent, A. F. (1998). Floral Dip: a Simplified Method for *Agrobacterium*-mediated Transformation of *Arabidopsis thaliana*. *Plant Journal*, **16**:735–743.
- Coburn, B., Sekirov, I., and Finlay, B. B. (2007). Type III Secretion Systems and Disease. *Clinical Microbiology Reviews*, **20**(4): 535–49.
- Cohen, D. J., Maldera, J. A., Muñoz, M. W., Ernesto, J. I., Vasen, G., and Cuasnicu, P. S. (2011). Cysteine-Rich Secretory Proteins (CRISP) and Their Role in Mammalian Fertilization. *Biological Research*, **44**:135 – 138.
- Da Ros, V. G., Muñoz, M. W., Battistone, M. A., Brukman, N. G., Carvajal, G., Curci, L., Gómez-Elías, M. D., Cohen, D. J., and Cuasnicú, P. S. (2015). From the Epididymis to the Egg: Participation of CRISP Proteins in Mammalian Fertilization. *Asian Journal of Andrology*, **17**(5):711–715.

- Damm, B., Schmidt, R., and Willmitzer, L. (1989). Efficient Transformation of *Arabidopsis thaliana* Using Direct Gene Transfer to Protoplasts. *Molecular General Genetics*, **217**:6–12.
- Day, B., Dahlbeck, D., Huang, J., Chisholm, S. T., Li, D., and Staskawicz, B. J. (2005). Molecular Basis for the *RIN4* Negative Regulation of *RPS2* Disease Resistance. *The Plant Cell*, **17**(4):1292–1305.
- Deindl, E., Boengler, K., van Royen, N., and Schaper, W. (2002). Differential Expression of GAPDH and Beta-3-actin in Growing Collateral Arteries. *Molecular and Cellular Biochemistry*, **236**:139–146.
- Desvaux, M., Parham, N. J., Scott-Tucker, A., and Henderson, I. R. (2004). The General Secretory Pathway: a General Misnomer? *Trends in Microbiology*, **12**:306–309.
- Ding, S. W. (2010). RNA-based Antiviral Immunity. *Nature Review Immunology*, **10**:632–644.
- Doczi, R., Brader, G., Pettko-Szandtner, A., Rajh, I., Djamei, A., Pitzschke, A., Teige, M., and Hirt, H. (2007). The *Arabidopsis* Mitogen-Activated Protein Kinase Kinase *MKK3* is Upstream of Group C Mitogen-Activated Protein Kinase and Participates in Pathogen Signaling. *The Plant Cell*, **19**:3266–3279.
- Doganlar, S., Frary, A., Daunay, M. C., Lester, R. N., and Tanksley, S. D. (2002). Conservation of Gene Function in the Solanaceae as Revealed by Comparative Mapping of Domestication Traits in Eggplant. *Genetics*, **161**:1713–1726.
- Eisen, J. A. (1998). Phylogenomics: Improving Functional Predictions for Uncharacterized Genes by Evolutionary Analysis. *Genome Research*, **8**:163–167.
- Eschen-Lippold, L., Bethke, G., Palm-Forster, M. A. T., Pecher, P., Bauer, N., Glazebrook, J., Scheel, D., and Lee, J. (2012). *MPK11*—a Fourth Elicitor-responsive Mitogen-activated Protein Kinase in *Arabidopsis thaliana*. *Plant Signaling and Behavior*, **7**(9):1203–1205.
- Eskelin, K., Suntio, T., Hyvarinen, S., Hafren, A., and Makinen, K. (2010). *Renilla* Luciferase-based Quantitation of Potato Virus an Infection Initiated with *Agrobacterium* Infiltration of *N. benthamiana* Leaves. *Journal of Virological Methods*, **164**:101–110.
- Fang, G., Bhardwaj, N., Robilotto, R., and Gerstein, M. B. (2010). Getting Started in Gene Orthology and Functional Analysis. *Public Library of Sciences Computational Biology*, **6**(3):1–8.
- FAOSTAT (Food and Agriculture Organisation for the United Nations – Statistics Online Website) (1991-2011; 2003-2012). <http://faostat.fao.org> (Accessed on 25 March 2014; 25 May 2015).
- Feldmann, K. A., and Marks, M. D. (1987). *Agrobacterium*-mediated Transformation of Germinating Seeds of *Arabidopsis thaliana*: a Non-tissue Culture Approach.

Molecular and General Genetics, **208**:1–9.

- Fiil, B. K., and Petersen, M. (2011). Constitutive Expression of *MKS1* Confers Susceptibility to *Botrytis cinerea* Infection Independent of *PAD3* Expression. *Plant Signaling and Behavior*, **6**(10):1425–7.
- Fitch, W. M. (2000). Homology: a Personal View on Some of the Problems. *Trends in Genetics*, **16**:227–231.
- Fu, Z., and Dong, X. (2013). Systemic Acquired Resistance: Turning Local Infection into Global Defense. *Annual Review of Plant Biology*, **64**:839–63.
- Fullerton, R. A., Taufa, L., Vanneste, J. L., Yu J., Cornish, D. D., and Park, D. (2011). First Record of Bacterial Crown Rot of Papaya (*Carica papaya*) Caused by an *Erwinia papayae*-like Bacterium in the Kingdom of Tonga. *American Physiological Society Journal*, **95**(1):70.
- Gamborg, O. L., Miller, R. A., and Ohyama, K. (1968). Nutrient Requirements of Suspension Cultures of Soybean Root Cells. *Experimental Cell Research*, **50**:148–151.
- Gao, M., Liu, J., Bi, D., Zhang, Z., Cheng, F., Chen, S., and Zhang, Y. (2008). *MEKK1*, *MKK1/MKK2* and *MPK4* Function Together in a Mitogen-activated Protein Kinase Cascade to Regulate Innate Immunity in Plants. *Cell Research*, **18**(12):1190–8.
- Gardan, L., Christen, R., Achouak, W., and Prior, P. (2004). *Erwinia papayae* sp. nov., a Pathogen of Papaya (*Carica papaya*). *International Journal of Systematic and Evolutionary Microbiology*, **54**:107–113.
- Genoud, T., Buchala, A.J., Chua, N-H., and Metraux, J-P. (2002). Phytochrome Signalling Modulates the SA-perceptive Pathway in Arabidopsis. *The Plant Journal: For Cell and Molecular Biology*, **31**(1):87–95.
- Gibbs, G. M., Roelants, K., and O'Bryan, M. K. (2008). The CAP Superfamily: Cysteine-rich Secretory Proteins, Antigen 5, and Pathogenesis-related 1 Proteins – Roles in Reproduction, Cancer, and Immune Defense. *Endocrine Reviews*, **29**(7):865–897.
- Glare, E. M., Divjak, M., Bailey, M. J., and Walters, E. H. (2002). Beta-actin and GAPDH Housekeeping Gene Expression in Asthmatic Airways is Variable and Not Suitable for Normalising mRNA Levels. *Thorax*, **57**:765–770.
- Gorzelnik, K., Janke, J., Engeli, S., and Sharma A. M. (2001). Validation of Endogenous Controls for Gene Expression Studies in Human Adipocytes and Preadipocytes. *Hormone and Metabolic Research*, **33**:625–627.
- Halter, T., Imkampe, J., Mazzotta, S., Wierzba, M., Postel, S., Bücherl, C., Kiefer, C., Stahl, M., Chinchilla, D., Wang, X., Nürnberger, T., Zipfel, C., Clouse, S., Borst, J. W., Boeren, S., de Vries, S. C., Tax, F., and Kemmerling, B. (2014). The Leucine-rich Repeat Receptor Kinase *BIR2* is a Negative Regulator of *BAK1* in

Plant Immunity. *Current Biology*, **24**(2):134–43.

Hamalainen, H. K., Tubman, J. C., Vikman, S., Kyrola, T., Ylikoski, E., Warrington, J. A., and Lahesmaa, R. (2001). Identification and Validation of Endogenous Reference Genes for Expression Profiling of T-helper Cell Differentiation by Quantitative Real-time RT-PCR. *Analytical Biochemistry*, **299**:63–70.

Heinrich M., Baldwin I. T., and Wu J. (2011). Two Mitogen-activated Protein Kinase Kinases, *MKK1* and *MEK2*, are Involved in Wounding and Specialist Lepidopteran herbivore *Manduca Sexta*-induced Responses in *Nicotiana attenuata*. *Journal of Experimental Botany*, **62**(12):4355–4365.

Hettenhausen, C., Baldwin, I. T., and Wu, J. (2012). Silencing MPK4 in *Nicotiana attenuata* Enhances Photosynthesis and Seed Production but Compromises Abscisic Acid-induced Stomatal Closure and Guard Cell-mediated Resistance to *Pseudomonas syringae* pv tomato DC3000. *Plant Physiology*, **158**(2):759–76.

Jamir, Y., Guo, M., Oh, H. S., Petnicki-Ocwieja, T., Chen, S., Tang, X., Dickman, M. B., Collmer, A., and Alfano, J. R. (2004). Identification of *Pseudomonas syringae* type III Effectors that can Suppress Programmed Cell Death in Plants and Yeast. *The Plant Journal*, **37**(4):554–565.

Jiang, C. J., Shimono, M., Maeda, S., Inoue, H., Mori, M., Hasegawa, M., Sugano, S., and Takatsuji, H. (2009). Suppression of the Rice Fatty Acid Desaturase Gene *OsSS12* Enhances Resistance to Blast and Leaf Blight Diseases in Rice. *Molecular Plant-Microbe*, **22**:820–829.

Johansen, L. K., and Carrington, J. C. (2001). Silencing on the Spot: Induction and Suppression of RNA Silencing in the *Agrobacterium*-mediated Transient Expression System. *Plant Physiology*, **126**:930–938.

Jones, J., and Dangl, J. (2006). The Plant Immune System. *Nature reviews*, **444**:232–329.

Kanehisa, M., Sato, Y., and Morishima, K. (2016). BlastKOALA and GhostKOALA: KEGG Tools for Functional Characterization of Genome and Metagenome Sequences. *Journal of Molecular Biology*, **428**(4):726–731.

Kang, L., Li, J., Zhao, T., Xiao, F., Tang, X., Thilmony, R., He, S., and Zhou, J. M. (2003). Interplay of the Arabidopsis Nonhost Resistance Gene *NHO1* with Bacterial Virulence. *Proceedings of the National Academy of Sciences of the United States of America*, **100**(6):3519–24.

Kang, L., Yuh-Shuh, W., Srinivasa, R. U., Keri, W., Yuhong, T., Vatsala, V., Barney, J. V., Kent, D. C., Elison, B. B., and Kirankumar, S. M. (2008). Overexpression of a Fatty Acid Amide Hydrolase Compromises Innate Immunity in Arabidopsis. *Plant Journal*, **56**:336–349.

Kapila, J., Rycke, R. D., Montagu, M. V., and Angenon, G. (1997). An *Agrobacterium*-mediated Transient Gene Expression System for Intact Leaves. *Plant Science*, **122**:101–108.

- Kellis, M., Patterson, N., Endrizzi, M., Birren, B., and Lander, E. S. (2003). Sequencing and Comparison of Yeast Species to Identify Genes and Regulatory Elements. *Nature*, **423**:241–254.
- Kong, Q., Qu, N., Gao, M., Zhang, Z., Ding, X., Yang, F., Li, Y., Dong, O. X., Chen, S., Li, X., and Zhang, Y. (2012). The *MEKK1-MKK1/MKK2-MPK4* Kinase Cascade Negatively Regulates Immunity Mediated by a Mitogen-activated Protein Kinase Kinase Kinase in *Arabidopsis*. *The Plant Cell*, **24**(5):2225–36.
- Koonin, E. V. (2005). Orthologs, Paralogs, and Evolutionary Genomics. *Annual Reviews Genetics*, **39**:309–338.
- Koop, B. F., and Hood, L. (1994). Striking Sequence Similarity over Almost 100 Kilobase of Human and Mouse T-cell Receptor DNA. *Nature Genetics*, **7**:48–53.
- Kosetsu, K., Matsunaga, S., Nakagami, H., Colcombet, J., Sasabe, M., Soyano, T., Takahashi, Y., Hirt, H., and Machida, Y. (2010). The MAP Kinase *MPK4* is required for Cytokinesis in *Arabidopsis thaliana*. *The Plant Cell*, **22**(11):3778–90.
- Kunkel, B. N., and Brooks, D. M. (2002). Cross Talk between Signaling Pathways in Pathogen Defense. *Current Opinion in Plant Biology*, **5**(4):325–331.
- Kuzniar, A., van Ham, R. C., Pongor, S., and Leunissen, J. A. (2008). The Quest for Orthologs: Finding the Corresponding Gene across Genomes. *Trends in Genetics*, **24**:539–551.
- Lai, Z., Li, Y., Wang, F., Cheng, Y., Fan, B., Yu, J. Q., and Chen, Z. (2011). *Arabidopsis* Sigma Factor Binding Proteins are Activators of the *WRKY33* Transcription Factor in Plant Defense. *The Plant Cell*, **23**(10):3824–41.
- Lara-Nuñez, A., Romero-Romero, T., Ventura, J. L., Blancas, V., Anaya, A. L., and Cruz-Ortega, R. (2006). Allelochemical Stress Causes Inhibition of Growth and Oxidative Damage in *Lycopersicon esculentum* Mill. Plant. *Cell and Environment*, **29**(11):2009–16.
- Larionov, A., Krause, A., and Miller, W. (2005). A Standard Curve Based Method foliativ Real-time PCR Data Processing. *BioMed Central Bioinformatics*, **6**:62.
- Lee, M. W., and Yang, Y. (2006). Transient Expression Assay by Agroinfiltration of Leaves. In N. J. Clifton, (Eds), *Methods in Molecular Biology*. (pp. 225-229) Totowa, New Jersey, USA: Humana Press Incorporation.
- Leuzinger, K., Dent, M., Hurtado, J., Stahnke, J., Lai, H., Zhou, X., and Chen, Q. (2013). Efficient Agroinfiltration of Plants for High-level Transient Expression of Recombinant Proteins. *Journal of Visualized Experiments*. **77**:e50521.
- Li, C., Yan, J. M., Li, Y. Z., Zhang, Z. C., Wang, Q. L., and Liang, Y. (2013). Silencing the *SpMPK1*, *SpMPK2*, and *SpMPK3* Genes in Tomato Reduces Abscisic Acid—Mediated Drought Tolerance. *International Journal of Molecular Sciences*, **14**(11):21983–21996.

- Li, X. (2011). Infiltration of *Nicotiana benthamiana* Protocol for Transient Expression via *Agrobacterium*. *Bioprotocol*, **1**(14): 1-3.
- Li, X., Chanroj, S., Wu, Z., Romanowsky, S. M., Harper, J. F., and Sze, H. (2008). A Distinct Endosomal Ca²⁺/Mn²⁺ Pump Affects Root Growth Through the Secretory Process. *Plant Physiology*, **147**(4):1675–1689.
- Li, X., Lin, H., Zhang, W., Zou, Y., Zhang, J., Tang, X., and Zhou, J. M. (2005). Flagellin Induces Innate Immunity in Nonhost Interactions that is suppressed by *Pseudomonas syringae* Effectors. *Proceedings of the National Academy of Sciences of the United States of America*, **102**(36):12990–5.
- Li, X., Zhang, Y., Huang, L., Ouyang, Z., Hong, Y., Zhang, H., Li, D., and Song, F. (2014). Tomato *SLMKK2* and *SLMKK4* Contribute to Disease Resistance against *Botrytis cinerea*. *Biomedical Central Plant Biology*, **14**(1):166.
- Lindbo, J. A. (2007). TRBO: A High-efficiency Tobacco Mosaic Virus RNA-Based Overexpression Vector. *Plant Physiology*, **145**:1232–1240.
- Liu, C., and Mehdy, M. C. (2007). A Nonclassical Arabinogalactan Protein Gene Highly Expressed in Vascular Tissues, *AGP31*, is Transcriptionally Repressed in Methyl Jasmonic Acid in *Arabidopsis*. *Plant Physiology*, **145**:863-874.
- Liu, J. Z., Horstman, H. D., Braun, E., Graham, M. A., Zhang, C., Navarre, D., Qiu, W. L., Lee, Y., Nettleton, D., Hill, J. H., and Whitham, S. A. (2011). Soybean Homologs of *MPK4* Negatively Regulate Defense Responses and Positively Regulate Growth and Development. *Plant Physiology*, **157**(3):1363–78.
- Liu, W. X., Zhang, F. C., Zhang, W. Z., Song, L. F., Wu, W. H., and Chen, Y. F. (2013). Arabidopsis *Di19* Functions as a Transcription Factor and Modulates *PR1*, *PR2*, and *PR5* Expression in Response to Drought Stress. *Molecular Plant*, **6**(5):1487–1502.
- Livak, K. J., and Schmittgen, T. D. (2001). Analysis of Relative Gene Expression Data Using Real-time Quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods*, **25**(4):402–408.
- Loots, G. G., Locksley, R. M., Blankenspoor, C. M., Wang, Z. E., Miller, W., Rubin, E. M., and Frazer, K. A. (2000). Identification of a Coordinate Regulator of Interleukins 4, 13, and 5 by Cross-species Sequence Comparisons. *Science*, **288**:136–140.
- Lorenc-Kukula, K., Chaturvedi, R., Roth, M., Welti, R., and Shah, J. (2012). Biochemical and Molecular-Genetic Characterization of *SFD1*'s Involvement in Lipid Metabolism and Defense Signaling. *Frontiers in Plant Science*, **3**(2):26.
- Lorrain, S., Lin, B., Auriac, M.C., Kroj, T., Saindrenan, P., Nicole, M., Balague, C., and Roby, D. (2004). Vascular Associated Death1, a Novel GRAM Domain-containing Protein, is a Regulator of Cell Death and Defense Responses in Vascular Tissues. *The Plant Cell*, **16**:2217–2232.

- Lu, M., Tang, X., and Zhou, J. M. (2001). Arabidopsis *NHO1* is required for General Resistance against Pseudomonas Bacteria. *The Plant Cell*, **13**:437–47.
- Luo, Y., Caldwell, K. S., Wroblewski, T., Wright, M. E., and Michelmore, R. W. (2009). Proteolysis of a Negative Regulator of Innate Immunity is Dependent on Resistance Genes in Tomato and *Nicotiana benthamiana* and induced by Multiple Bacterial Effectors. *The Plant Cell*, **21**(8):2458–72.
- Lynch, M., and Conery, J. S. (2003). The Evolutionary Demography of Duplicate Genes. *Journal of Structural and Functional Genomics*, **3**:35–44.
- Maktar, N. H., Kamis, S., Mohd Yusof, F. Z., and Hussain, N. H. (2008). *Erwinia papaya* Causing Papaya Dieback in Malaysia. *Plant Pathology*, **57**:774.
- Mao, G., Meng, X., Liu, Y., Zheng, Z., Chen, Z., and Zhang, S. (2011). Phosphorylation of a WRKY Transcription Factor by Two Pathogen-responsive MAPKs Drives Phytoalexin Biosynthesis in Arabidopsis. *The Plant Cell*, **23**(4):1639–53.
- Mao, P., Duan, M., Wei, C., and Li, Y. (2007). WRKY62 Transcription Factor Acts Downstream of Cytosolic *NPR1* and Negatively Regulates Jasmonate Responsive Gene Expression. *Plant Cell Physiology*, **48**:833–842.
- Martinez-Navarro, A.C., Galvan-Gordillo, S. V., Xoconostle-Cazares, B., and Ruiz-Medrano, R. (2013). Vascular Gene Expression: a Hypothesis. *Frontiers in Plant Sciences*, **4**:261.
- Massoud, K., Barchietto, T., Le Rudulier, T., Pallandre, L., Didierlaurent, L., Garmier, M., Ambard-Bretteville, F., Seng, J. M., and Saindrenan, P. (2012). Dissecting Phosphate-induced Priming in Arabidopsis Infected with *Hyaloperonospora arabidopsidis*. *Plant Physiology*, **159**(1):286–98.
- Maxwell, D. P., Wang, Y., and McIntosh, L. (1999). The Alternative Oxidase Lowers Mitochondrial Reactive Oxygen Production in Plant Cells. *Proceedings of the National Academy of Sciences of the United States of America*, **96**(14):8271–6.
- McGuire, A. M., Hughes, J. D., and Church, G. M. (2000). Conservation of DNA Regulatory Motifs and Discovery of New Motifs in Microbial Genomes. *Genome Research*, **10**:744–757.
- Mijatovic-Rustempasic, S., Tam, K. I., Kerin, T. K., Lewis, J. M., Gautam, R., Quaye, O., Gentsch, J. R., and Bowen, M. D. (2013). Sensitive and Specific Quantitative Detection of Rotavirus A by One-Step Real-Time Reverse Transcription-PCR Assay without Antecedent Double-Stranded-RNA Denaturation. *Journal of Clinical Microbiology*, **51**(9):3047–3054.
- Millet, Y. A., Danna, C. H., Clay, N. K., Songnuan, W., Simon, M. D., Werck-Reichhart, D., and Ausubel, F. M. (2010). Innate Immune Responses Activated in Arabidopsis Roots by Microbe Associated Molecular Patterns. *The Plant Cell*, **22**:973–990.

Ming, R., Hou, S., Feng, Y., Yu, Q., Dionne-Laporte, A., Saw, J. H., Senin, P., Wang, W., Ly, B. V., Lewis, K. L., Salzberg, S. L., Feng, L., Jones, M. R., Skelton, R. L., Murray, J. E., Chen, C., Qian, W., Shen, J., Du, P., Eustice, M., Tong, E., Tang, H., Lyons, E., Paull, R. E., Michael, T. P., Wall, K., Rice, D. W., Albert, H., Wang, M. L., Zhu, Y. J., Schatz, M., Nagarajan, N., Acob, R. A., Guan, P., Blas, A., Wai, C. M., Ackerman, C. M., Ren, Y., Liu, C., Wang, J., Wang, J., Na, J. K., Shakirov, E. V., Haas, B., Thimmapuram, J., Nelson, D., Wang, X., Bowers, J. E., Gschwend, A. R., Delcher, A. L., Singh, R., Suzuki, J. Y., Tripathi, S., Neupane, K., Wei, H., Irikura, B., Paidi, M., Jiang, N., Zhang, W., Presting, G., Windsor, A., Navajas-Pérez, R., Torres, M. J., Feltus, F. A., Porter, B., Li, Y., Burroughs, A. M., Luo, M. C., Liu, L., Christopher, D. A., Mount, S. M., Moore, P. H., Sugimura, T., Jiang, J., Schuler, M. A., Friedman, V., Mitchell-Olds, T., Shippen, D. E., dePamphilis, C. W., Palmer, J. D., Freeling, M., Paterson, A. H., Gonsalves, D., Wang, L., and Alam, M. (2008). The Draft Genome of the Transgenic Tropical Fruit Tree Papaya (*Carica papaya* Linnaeus). *Nature*, **452**(7190):991–996.

Mohd Khairil, J., and Muhammad Munzir, M. (2014). Experiences in Managing Bacterial Dieback Disease of Papaya in Malaysia. *ISHS Acta Horticulturae* **1022**: III International Symposium on Papaya. **1022**: 125–132.

Mohd Waznul Adly M. Z., Aliza Z., Maheswary V., and Zuraida A. R. (2010). Validation of Housekeeping Genes for Quantitative Real-time PCR in Malaysian Papaya var. Eksotika I (*Carica papaya*) Proceeding of UMS Biotechnology Symposium IV, Kota Kinabalu, Sabah. 1 – 3 December 2010.

Moustafa, K., Abu-Qamar, S., Jarrar, M., Al-Rajab, A. J., and Tremouillaux-Guiller, J. (2014). MAPK Cascades and Major Abiotic Stresses. *Plant Cell Reports*, **33**(8):1217–25.

Murashige, T., and Skoog, F. (1962). A Revised Medium for Rapid Growth and Bioassays with Tobacco Tissue Cultures. *Physiology Plant*, **15**:473–497.

Nakagami, H., Soukupova, H., Schikora, A., Zarsky, V., and Hirt, H. (2006). A Mitogen-activated Protein Kinase Kinase Kinase Mediates Reactive Oxygen Species Homeostasis in Arabidopsis. *The Journal of Biological Chemistry*, **281**(50):387.

Nandi, A., Welti, R., and Shah, J. (2004). The *Arabidopsis thaliana* Dihydroxyacetone Phosphate Reductase Gene SUPPRESSOR OF FATTY ACID DESATURASE DEFICIENCY1 Is Required for Glycerolipid Metabolism and for the Activation of Systemic Acquired Resistance. *The Plant Cell*, **16**:465–477.

Nekrasov, V., Li, J., Batoux, M., Roux, M., Chu, Z.-H., Lacombe, S., Rougon, A., Bittel, P., Kiss-Papp, M., Chinchilla, D., van Esse, H. P., Jordà, L., Schwessinger, B., Nicaise, V., Thomma, B. P., Molina, A., Jones, J. D., and Zipfel, C. (2009). Control of the Pattern-recognition Receptor *EFR* by an ER Protein Complex in Plant Immunity. *The European Molecular Biology Organization Journal*, **28**(21):3428–38.

Nikitin, A., Egorov, S., Daraselia, N., and Mazo, I. (2003). Pathway Studio—the

- Analysis and Navigation of Molecular Networks. *Bioinformatics*, **19**(16):2155–2157.
- Nimchuk, Z., Marois, E., Kjemtrup, S., Leister, T. R., Katagiri, F., and Dangl, J. L. (2000). Eukaryotic Fatty Acylation Drives Plasma Membrane Targeting and Enhances Function of Several Type III Effector Proteins from *Pseudomonas syringae*. *Cell*, **101**:353–363.
- Norihha, M. A., Hamidun, B., Rohaiza, A. R., and Indu, B. S. J. (2011). *Erwinia mallotivora* sp., a New Pathogen of Papaya (*Carica papaya*) in Peninsular Malaysia. *International Journal Molecular Sciences*, **12**:39–45.
- Notredame, C., Higgins, D. G., and Heringa, J. (2000). T-Coffee: A Novel Method for Multiple Sequence Alignments. *Journal of Molecular Biology*, **302**:205–217.
- Novo, M., Bigey, F., Beyne, E., Galeote, V., Gavory, F., Mallet, S., Cambon, B., Legras, J. L., Wincker, P., Casaregola, S., and Dequin, S. (2009). Eukaryote-to-Eukaryote Gene Transfer Events Revealed by the Genome Sequence of the Wine Yeast *Saccharomyces cerevisiae* EC1118. *Proceedings of the National Academy of Sciences of the United States of America*, **106**:16333–16338.
- Oeltjen, J. C., Malley, T. M., Muzny, D. M., Miller, W., Gibbs, R. A., and Belmont, J. W. (1997). Large-scale Comparative Sequence Analysis of the Human and Murine Bruton's Tyrosine Kinase Loci Reveals Conserved Regulatory Domains. *Genome Research*, **7**:315–329.
- Ogata, H., Goto, S., Sato, K., Fujibuchi, W., Bono, H., and Kanehisa, M. (1999). KEGG: Kyoto Encyclopedia of Genes and Genomes. *Nucleic Acids Research*, **27**(1):29–34.
- Oh, C. S., and Beer, S. V. (2005). Molecular Genetics of *Erwinia amylovora* Involved in the Development of Fire Blight. *Federation of European Microbiological Society Microbiology Letters*, **253**:185–192.
- Oh, M. H., Wu, X., Clouse, S. D., and Huber, S. C. (2011). Functional Importance of *BAK1* Tyrosine Phosphorylation in Vivo. *Plant Signaling and Behavior*, **6**(3):400–405.
- Park, J. M., Park, C. J., Lee, S. B., Ham, B. K., Shin, R., and Paek, K. H. (2001). Overexpression of the Tobacco *Tsi1* Gene Encoding an EREBP/AP2-Type Transcription Factor Enhances Resistance against Pathogen Attack and Osmotic Stress in Tobacco. *The Plant Cell*, **13**:1035–1046.
- Pennacchio, L. A., and Rubin, E. M. (2001). Genomic Strategies to Identify Mammalian Regulatory Sequences. *Nature Review Genetics*, **2**:100–109.
- Petersen, K., Qiu, J. L., Lütje, J., Fiil, B. K., Hansen, S., Mundy, J., and Petersen, M. (2010). Arabidopsis *MKS1* is involved in Basal Immunity and Requires an Intact N-terminal Domain for Proper Function. *Public Library of Science One*, **5**(12):e14364.

- Peyret, H., and Lomonossoff, G. P. (2013). The pEAQ Vector Series: the Easy and Quick Way to Produce Recombinant Proteins in Plants. *Plant Molecular Biology*, **83**(1-2):51–8.
- Pfaffl M. W. (2001). A New Mathematical Model for Relative Quantification in Real-time RT-PCR. *Nucleic Acid Research*, **1**:29(9):e45.
- Pitzschke, A., Djamei, A., Bitton, F., and Hirt, H. (2009). A Major Role of the *MEKK1-MKK1/2-MPK4* Pathway in ROS signalling. *Molecular Plant*, **2**(1):120–37.
- Qiu, J. L., Fiil, B. K., Petersen, K., Nielsen, H. B., Botanga, C. J., Thorgrimsen, S., Palma, K., Suarez-Rodriguez, M. C., Sandbech-Clausen, S., Lichota, J., Brodersen, P., Grasser, K. D., Mattsson, O., Glazebrook, J., Mundy, J., and Petersen, M. (2008a). Arabidopsis MAP Kinase 4 Regulates Gene Expression Through Transcription Factor Release in the Nucleus. *The European Molecular Biology Organization Journal*, **27**(16):2214–21.
- Qiu, J. L., Zhou, L., Yun, B. W., Nielsen, H. B., Fiil, B. K., Petersen, K., Mackinlay, J., Loake, G. J., Mundy, J., and Morris, P. C. (2008b). Arabidopsis Mitogen-activated Protein Kinase Kinases *MKK1* and *MKK2* have Overlapping Functions in Defense Signaling Mediated by *MEKK1*, *MPK4*, and *MKS1*. *Plant Physiology*, **148**(1):212–22.
- Raaijmakers, M. H., van Emst, L., de Witte, T., Mensink, E., and Raymakers, R. A. (2002). Quantitative Assessment of Gene Expression in Highly Purified Hematopoietic Cells Using Real-time Reverse Transcriptase Polymerase Chain Reaction. *Experimental Hematology*, **30**:481–487.
- Radonic, A., Thulke, S., Mackay, I. M., Landt O., Siegert, W., and Nitsche, A. (2004). Guideline to Reference Gene Selection for Quantitative Real-time PCR. *Biochemical and Biophysical Research Communications*, **313**:858–862.
- Ramakrishna, W., Dubcovsky, J., Park, Y. J., Busso, C., Emberton, J., SanMiguel, P., and Bennetzen, J. L. (2002). Different Types and Rates of Genome Evolution Detected by Comparative Sequence Analysis of Orthologous Segments From Four Cereal Genomes. *Genetics*, **162**:1389–1400.
- Rasmussen, M. W., Roux, M., Petersen, M., and Mundy, J. (2012). MAP Kinase Cascades in Arabidopsis Innate Immunity. *Frontiers in Plant Science*, **3**(7):169.
- Rasmussen, R. (2001). Quantification on the LightCycler. In S. Meuer, C. Wittwer, K. Nakagawa (Eds.), *Rapid Cycle Real-time PCR Methods and Applications*. (pp 21-34). Berlin: Springer-Verlag Press, Heidelberg.
- Remm, M., Storm, C. E. V., and Sonnhammer, E. L. L. (2001). Automatic Clustering of Orthologs and In-paralogs from Pairwise Species Comparisons. *Journal of Molecular Biology*, **314**:1041–1052.
- Roberts, J. A., Miguel-Escalada, I., Slovik, K. J., Walsh, K. T., Hadzhiev, Y., Sanges, R., Stupka, E., Marsh, E. K., Balciuniene, J., Balciunas, D., and Muller, F.

- (2014). Targeted Transgene Integration Overcomes Variability of Position Effects in Zebrafish. *Development: The Company of Biologists*, **141**(3):715–724.
- Ronald, P. C., and Beutler, B. (2010). Plant and Animal Sensors of Conserved Microbial Signatures. *Science*, **330**:1061–1064.
- Roshidi, A.S. (2010, September 29). Papaya Disease Alert. *The Star Online*. Retrieved from <http://thestar.com.my/metro/story> (Accessed on 20 February 2015).
- Roux, M., Schwessinger, B., Albrecht, C., Chinchilla, D., Jones, A., Holton, N., Malinovsky, F. G., Tör, M., de Vries, S., and Zipfel, C. (2011). The Arabidopsis Leucine-rich Repeat Receptor-like Kinases *BAK1/SERK3* and *BKK1/SERK4* are required for Innate Immunity to Hemibiotrophic and Biotrophic Pathogens. *The Plant Cell*, **23**(6):2440–55.
- Rushton, P. J., Torres, J. T., Parniske, M., Wernert, P., Hahlbrock, K., and Somssich, I. E, (1996). Interaction of Elicitor-induced DNA Binding Proteins with Elicitor Response Elements in the Promoters of Parsley *PR1* genes. *The European Molecular Biology Organization Journal*, **15**:5690–5700.
- Sabry, A., Mohamed, A. A., and Awad, N. S. (2014) A Comprehensive in silico DNA Sequence Analysis of Sperm Surface ADAM Genes Collected from RefSeq Database. *American Journal of Bioinformatics Research*. **4**(2):23-32.
- Saijo, Y., Tintor, N., Lu, X., Rauf, P., Pajerowska-Mukhtar, K., Häweker, H., Dong, X., Robatzek, S., and Schulze-Lefert, P. (2009). Receptor Quality Control in the Endoplasmic Reticulum for Plant Innate Immunity. *The European Molecular Biology Organization Journal*, **28**(21):3439–49.
- Sainsbury, F., and Lomonossoff, G. P. (2008). Extremely High-level and Rapid Transient Protein Production in Plants without the Use of Viral Replication. *Plant Physiology*, **148**(3):1212–8.
- Sainsbury, F., Canizares, M. C., and Lomonossoff, G. P. (2007). Cowpea Mosaic Virus-based Expression Vectors. In K. Hefferon, (Eds), *Virus Expression Vectors*. (pp. 339-555) Kerala, India: Transworld Research Network.
- Sainsbury, F., Lavoie, P. O., D'Aoust, M. A., Vezina, L., P., and Lomonossoff, G. P. (2008). Expression of Multiple Proteins Using Full-Length and Deleted Versions of Cowpea Mosaic Virus RNA-2. *Plant Biotechnology Journal*, **6**:82–92.
- Sainsbury, F., Thuenemann, E. C., and Lomonossoff, G. P. (2009). pEAQ: Versatile Expression Vectors for Easy and Quick Transient Expression of Heterologous Proteins in Plants. *Plant Biotechnology Journal*, **7**:682–693.
- Sasabe, M., Kosetsu, K., Hidaka, M., Murase, A., and Machida, Y. (2011). *Arabidopsis thaliana MAP65-1* and *MAP65-2* Function Redundantly with *MAP65-3/PLEIADE* in Cytokinesis Downstream of *MPK4*. *Plant Signaling and Behavior*, **6**(5):743–747.
- Saunders, K., Sainsbury, F., and Lomonossoff, G. P. (2009). Efficient Generation of

- Cowpea Mosaic Virus Empty Virus-like Particles by the Proteolytic Processing of Precursors in Insect Cells and Plants. *Virology*, **393**(2):329–337.
- Schmittgen, T. D., and Zakrajsek, B. A (2000). Effect of Experimental Treatment on Housekeeping Gene Expression: Validation by Real-time, Quantitative RT-PCR. *Journal of Biochemical and Biophysical Methods*, **46**:69–81.
- Schob, H., Kunz, C., and Meins, F. J. (1997). Silencing of Transgenes Introduced into Leaves by Agroinfiltration: A Simple, Rapid Method for Investigation of Sequence Requirements for Gene Silencing. *Molecular Genomic Genetics*, **256**:581–585.
- Schweighofer, A., Kazanaviciute, V., Scheikl, E., Teige, M., Doczi, R., Hirt, H., Schwanninger, M., Kant, M., Schuurink, R., Mauch, F., Buchala, A., Cardinale, F., and Meskiene, I. (2007). The PP2C-type Phosphatase *AP2C1*, which Negatively Regulates *MPK4* and *MPK6*, Modulates Innate Immunity, Jasmonic Acid, and Ethylene Levels in *Arabidopsis*. *The Plant Cell*, **19**(7):2213–24.
- Schwessinger, B., and Ronald, P. (2012). Plant Innate Immunity: Perception of Conserved Microbial Signatures. *Annual Review of Plant Biology*, **63**(1):451–482.
- Schwessinger, B., Roux, M., Kadota, Y., Ntoukakis, V., Sklenar, J., Jones, A., and Zipfel, C. (2011). Phosphorylation-dependent Differential Regulation of Plant Growth, Cell Death, and Innate Immunity by the Regulatory Receptor-like Kinase *BAK1*. *Public Library of Science: Genetics*, **7**(4):e1002046.
- Seki, M., Iida, A., and Morikawa, H. (1999). Transient Expression of the Beta-glucuronidase Gene in Tissues of *Arabidopsis thaliana* by Bombardment-mediated Transformation. *Molecular Biotechnology*, **11**:251–255.
- Seki, M., Komeda, Y., Iida, A., Yamada, Y., and Morikawa, H. (1991). Transient Expression of β-glucuronidase in *Arabidopsis thaliana* Leaves and Roots and *Brassica napus* Stems Using a Pneumatic Particle Gun. *Plant Molecular Biology*, **17**:259–263.
- Selote, D., and Kachroo, A. (2010a). RIN4-like Proteins Mediate Resistance Protein-derived Soybean Defense against *Pseudomonas syringae*. *Plant Signaling and Behavior*, **5**(11):1453–6.
- Selote, D., and Kachroo, A. (2010b) RPG1-B-Derived Resistance to AvrB-Expressing *Pseudomonas syringae* Requires RIN4-Like Proteins in Soybean. *Plant Physiology*, **153**:1199–1211.
- Selvey, S., Thompson, E. W., Matthaei, K., Lea, R. A., Irving, M. G., and Griffiths, L. R. (2001). Beta-actin — An Unsuitable Internal Control for RT-PCR. *Molecular Cell Probes*, **15**:307–311.
- Seo, S., Okamoto, M., Seto, H., Ishizuka, K., Sano, H., and Ohashi, Y. (1995). Tobacco MAP Kinase: A Possible Mediator in Wound Signal Transduction Pathways. *Science*, **270**:1988–1992.

- Shan, L., He, P., Li, J., Heese, A., Peck, S. C., Nurnberger, T., Martin, G. B., and Sheen, J. (2008). Bacterial Effectors Target the Common Signaling Partner BAK1 to Disrupt Multiple MAMP Receptor-Signaling Complexes and Impede Plant Immunity. *Cell Host Microbe*, **4**(1):17–27.
- Shang, J., Xi, D. H., Xu, F., Wang, S. D., Cao, S., Xu, M. Y., Zhao, P. P., Wang, J. H., Jia, S. D., Zhang, Z. W., Yuan, S., and Lin, H. H. (2011). A Broad-spectrum, Efficient and Nontransgenic Approach to Control Plant Viruses by Application of Salicylic Acid and Jasmonic Acid. *Planta*, **233**(2):299–308.
- Shi, J., An, H. L., Zhang, L., Gao, Z., and Guo, X. Q. (2010). *GhMPK7*, a Novel Multiple Stress-responsive Cotton Group C MAPK Gene, has a Role in Broad Spectrum Disease Resistance and Plant Development. *Plant Molecular Biology*, **74**(1-2):1–17.
- Shi, J., Zhang, L., An, H., Wu, C., and Guo, X. (2011). *GhMPK16*, a Novel Stress-responsive Group D MAPK Gene from Cotton, is involved in Disease Resistance and Drought Sensitivity. *BioMed Central Molecular Biology*, **12**:22.
- Sigrist, C. J. A., De Castro, E., Cerutti, L., Cuche, B. A., Hulo, N., Bridge, A., Bougueret, L., and Xenarios, I. (2013). New and Continuing Developments at PROSITE. *Nucleic Acids Research*, **41**(D1):1–4.
- Studer, R. A., and Robinson-Rechavi, M. (2009). How Confident Can We be that Orthologs are Similar, but Paralogs Differ? *Trends in Genetics*, **25**:210–216.
- Su, S. H., Suarez-Rodriguez, M. C., and Krysan, P. (2007). Genetic Interaction and Phenotypic Analysis of the Arabidopsis MAP Kinase Pathway Mutations *mekk1* and *mpk4* Suggests Signaling Pathway Complexity. *Federation of European Biochemical Societies Letters*, **581**(17):3171–7.
- Suarez-Rodriguez, M. C., Adams-Phillips, L., Liu, Y., Wang, H., Su, S. H., Jester, P. J., Zhang, S., Bent, A. F., and Krysan, P. J. (2007). *MEKK1* is required for *flg22*-induced *MPK4* Activation in Arabidopsis Plants. *Plant Physiology*, **143**(2):661–9.
- Supian S. 2014. *Antioxidant-mediated Defense Response of Carica papaya var. Eksotika against a Compatible E. mallotivora strain BT MARDI*, Master of Science Thesis, Universiti Putra Malaysia.
- Suzuki, T., Higgins, P. J., and Crawford, D. R. (2000). Control Selection for RNA Quantitation. *Biotechniques*, **29**:332–337.
- Taj, G., Giri, P., Tasleem, M., and Kumar, A. (2014). MAPK Signalling Cascades and Transcriptional Reprogramming in Plant-Pathogen Interactions. In R. K. Gour and P. Sharma (Eds.), *Approaches to Plant Stress and their Management*, (pp. 297-298). India: Springer.
- Takahashi, Y., Soyano, T., Kosetsu, K., Sasabe, M., and Machida, Y. (2010). *HINKEL* kinesin, *ANP* MAPKKs and *MKK6/ANQ* MAPKK, which Phosphorylates and Activates *MPK4* MAPK, Constitute a Pathway that is required for Cytokinesis in *Arabidopsis thaliana*. *Plant and Cell Physiology*, **51**(10):1766–76.

- Takemoto, D., and Jones, D. A. (2005). Membrane Release and Destabilization of Arabidopsis *RIN4* Following Cleavage by *Pseudomonas syringae* AvrRpt2. *Molecular Plant-Microbe Interactions*, **18**(12):1258–68.
- Tekaia, F. (2016). Inferring Orthologs: Open Questions and Perspectives. *Genomics Insights*, **9**:17–28.
- Tekaia, F., and Dujon, B. (1999). Pervasiveness of Gene Conservation and Persistence of Duplicates in Cellular Genomes. *Journal of Molecular Evolution*, **49**(5):591–600.
- Tellmann, G., and Geulen, O. (2006). Lightcycler 480 Real-time PCR System, Innovative Solutions for Relative Quantification. *Biochemica*, **4**:16–17.
- Tenhaken, R., Doerks, T., and Bork, P. (2005). DCD - a Novel Plant Specific Domain in Proteins Involved in Development and Programmed Cell Death. *BioMed Central Bioinformatics*, **6**:169.
- Tichopad A., Dilger M., Schwarz G., and Pfaffl M.W. (2003). Standardized Determination of Real-time PCR Efficiency from a Single Reaction Set-up. *Nucleic Acids Research*, **31**(20):e122.
- Umbrasaité, J., Schweighofer, A., Kazanaviciute, V., Magyar, Z., Ayatollahi, Z., Unterwurzacher, V., Choopayak, C., Boniecka, J., Murray, J. A., Bogre, L., and Meskiene, I. (2010). MAPK Phosphatase AP2C3 Induces Ectopic Proliferation of Epidermal Cells Leading to Stomata Development in Arabidopsis. *Public Library of Science One*, **5**(12):e15357.
- Usami, S., Banno, H., Ito, Y., Nishihama, R., and Machida, Y. (1995). Cutting Activates a 46-kilodalton Protein Kinase in Plants. *Proceedings of National Academy of Science United States of America*, **92**:8660–8664.
- Van der Hoorn, J. A. L., Laurent, F., Roth, R., and de Wit, P. J. G. M. (2000). Agroinfiltration is a Versatile Tool that Facilitates Comparative Analyses of Avr9/cf-9-induced and Avr4/Cf-4-induced Necrosis. *Molecular Plant-Microbe Interaction*, **13**:439–446.
- Vandesompele, J., De Preter, K., Pattyn, F., Poppe, B., Van Roy, N., De Paepe, A., and Speleman, F. (2002). Accurate Normalization of Real-time Quantitative RT-PCR Data by Genomic Averaging of Multiple Internal Control Genes. *Genome Biology*, **3**(7):Research0034.
- Veronese, P., Chen, X., Bluhm, B., Salmeron, J., Dietrich, R., and Mengiste, T. (2004). The BOS Loci of Arabidopsis are required for Resistance to *Botrytis cinerea* infection. *The Plant Journal: Cell and Molecular Biology*, **40**(4):558–74.
- Veronese, P., Narasimhan, M. L., Stevenson, R. A., Zhu, J. K., Weller, S. C., Subbarao, K. V., and Bressan, R. A. (2003). Identification of a Locus Controlling Verticillium Disease Symptom Response in *Arabidopsis thaliana*. *The Plant Journal*, **35**(5):574–587.

- Vidhyasekaran, P. (2014). *PAMP Signals in Plant Innate Immunity*. Dordrecht: Springer Netherlands.
- Vidhyasekaran, P. (2015). *Plant Hormone Signaling Systems in Plant Innate Immunity*. Dordrecht: Springer Netherlands.
- Von Rant, A., (1931). Aceber eine Bakterienkrankheit be idem Melonenbaume (*Carica papaya* L.) auf Java. *Zentralblatt für Bakteriologie Parasitenkunde Infektionskrankheiten und Hygiene*, **84**:481–487.
- Wacker, M. J., and Godard, M. P. (2005). Analysis of One-Step and Two-Step Real-Time RT-PCR Using SuperScript III. *Journal of Biomolecular Techniques*, **16**(3):266–271.
- Wang, K. (2006). In Agrobacterium Protocols: Agro-infiltration and Co-cultivation. In J. M. Walker, (Eds), *Methods in Molecular Biology*. (pp. 213) Totowa, New Jersey, USA: Humana Press Incorporation.
- Wang, Z., Mao, H., Dong, C., Ji, R., Cai, L., Fu, H., and Liu, S. (2009). Overexpression of *Brassica napus MPK4* Enhances Resistance to *Sclerotinia sclerotiorum* in Oilseed Rape. *Molecular Plant-Microbe Interactions*, **22**(3):235–44.
- Wani, S. H., Sanghera, G. S., and Singh, N. B. (2010). Biotechnology and Plant Disease Control: Role of RNA Interference. *American Journal of Plant Science*, **1**:55–68.
- Weber, J. M., Ponti, C. G., Kappeli, O., and Reiser, J. (1992). Factors Affecting Homologous Overexpression of the *Saccharomyces cerevisiae* Lanosterol 14 α -Demethylase gene. *Yeast*, **8**(7):519–533.
- Wroblewski, T., Tomczak, A., and Michelmore, R. (2005). Optimization of Agrobacterium-Mediated Transient Expression Assays for Lettuce, Tomato and Arabidopsis. *Plant Biotechnology Journal*, **3**:259–273.
- Wu, F., Mueller, L. A., Crouzillat, D., Petiard, V., and Tanksley, S. D. (2006). Combining Bioinformatics and Phylogenetics to Identify Large Sets of Single-Copy Orthologous Genes (COSII) for Comparative, Evolutionary and Systematic Studies: A Test Case in the Euasterid Plat Clade. *Genetics*, **174**:1407–1420.
- Yadeta, K. A., and Thomma, B. P. H. J. (2013). The Xylem as Battleground for Plant Hosts and Vascular Wilt Pathogens. *Frontiers in Plant Science*, **4**:97.
- Yamaguchi-Shinozaki, K., and Shinozaki, K. (1994). A Novel *cis*-Acting Element in an Arabidopsis Gene is involved in Responsiveness to Drought, Low-Temperature or High-Salt Stress. *The Plant Cell*, **6**:251–264.
- Yasmin, A., and Debener, T. (2010). Transient Gene Expression in Rose Petals via Agrobacterium Infiltration. *Plant Cell, Tissue and Organ Culture*, **102**:245–250.
- Young, M. D., Wakefield, M. J., Smyth, G. K., and Oshlack, A. (2010). Gene Ontology Analysis for RNA-seq: Accounting for Selection Bias. *Genome Biology*,

11(2):R14.

- Younis, A., Siddique, M. I., Kim, C. K., and Lim, K. B. (2014). RNA Interference (RNAi) Induced Gene Silencing: A Promising Approach of Hi-Tech Plant Breeding. *International Journal Biology Science*, **10**(10):1150–1158.
- Zhang, J. D., Ruschhaupt, M., and Biczok, R. (2017). DdCt Method for QRT-PCR Data Analysis. www.bioconductor.org/packages/devel/bioc/vignettes/ddCt/inst/doc/rtPCR. (Accessed on 25 May 2017).
- Zhang, S., and Klessig, D. F. (2000). Pathogen-Induced MAP Kinases in Tobacco: Wounding and MAP Kinase Activation. In H. Hirt (Ed.), *MAP Kinases in Plant Signal Transduction*. (pp. 65-84) Heidelberg, Berlin, Germany: Springer-Verlag Berlin Heidelberg.
- Zhong, H., and Simons, J. W. (1999). Direct Comparison of GAPDH, Beta-actin, Cyclophilin, and 28S rRNA as Internal Standards for Quantifying RNA Levels under Hypoxia. *Biochemistry and Biophysics Research Communications*, **259**:523–526.
- Zhou, N., Tootle, T. L., Tsui, F., Klessig, D.F., and Glazebrook, J. (1998). *PAD4* Functions Upstream from Salicylic Acid to Control Defense Responses in Arabidopsis. *The Plant Cell*, **10**(6):1021–30.
- Zhu, F., Xi, D., Yuan, S., Xu, F., Zhang, D., and Lin, H. (2014). Salicylic Acid and Jasmonic Acid are Essential for Systemic Resistance against Tobacco Mosaic Virus in *Nicotiana benthamiana*. *Molecular Plant-Microbe Interactions*, **27**(6):567–577.
- Zhu, J. K. (2001). Plant Salt Tolerance. *Trends in Plant Science*, **6**:66–71.
- Zipfel, C., Kunze, G., Chinchilla, D., Caniard, A., Jones, J. D., Boller, T., and Felix, G. (2006). Perception of the Bacterial MAMP EF-Tu by the Receptor *EFR* Restricts *Agrobacterium*-mediated Transformation. *Cell*, **125**:749–760.