

# **UNIVERSITI PUTRA MALAYSIA**

DEVELOPMENT OF PNA ELECTROCHEMICAL BIOSENSOR BASED ON SCREEN-PRINTED ELECTRODE-MODIFIED AMINE-FUNCTIONALIZED GRAPHENE COMPOSITE FOR Mycobacterium tuberculosis DETECTION

# **MOHD HAZANI BIN MAT ZAID**

**ITMA 2018 11** 



### DEVELOPMENT OF PNA ELECTROCHEMICAL BIOSENSOR BASED ON SCREEN-PRINTED ELECTRODE-MODIFIED AMINE-FUNCTIONALIZED GRAPHENE COMPOSITE FOR *Mycobacterium tuberculosis* DETECTION



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the requirements for the Degree of Doctor of Philosophy

October 2017

### COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icon, photographs and all other artwork, is copying material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purpose from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of the thesis submitted to the senate of, Universiti Putra Malaysia, in fulfillment of the requirement for the Degree of Doctor of Philosophy

#### DEVELOPMENT OF PNA ELECTROCHEMICAL BIOSENSOR BASED ON SCREEN-PRINTED ELECTRODE-MODIFIED AMINE-FUNCTIONALIZED GRAPHENE COMPOSITE FOR *Mycobacterium tuberculosis* DETECTION

By

#### MOHD HAZANI B. MAT ZAID

October 2017

Chairman: Jaafar b. Abdullah, PhD Faculty : Institute Advance Technology

Agarose gel electrophoresis is a fundamental and essential technique for analysis of PCR products related of mycobacterium tuberculosis. However this technique imposes some sort of limitation such as unquantifiable results and low specificity. Therefore, a novel PNA electrochemical biosensor based on modified screen printed carbon electrode (SPCE) has been developed for detection of mycobacterium tuberculosis which could overcome the limitation of agarose gel electrophoresis technique. In this study, the fabricated electrochemical biosensor SPCE has been modified with two different nanomaterial consist of amine functionalized reduced graphene oxide (NH<sub>2</sub>-GO) composited with water soluble CdS Quantum dots (QDs) and NH<sub>2</sub>-GO composited with Tempo-nanocellulose (TNCC) for enhance sensitivity detection of target ssDNA sequence related to Mycobacterium tuberculosis. The prepared composite materials have been characterized by Raman, FTIR, and TEM. Meanwhile, field emission microscope (FE-SEM) and energy dispersive X-ray spectroscopy (EDX) analysis had confirmed that both nanomaterials (NH2-GO/ODs and NH2-GO/TNCC) were deposited and uniformly distributed on the surface of SPCE. Furthermore, based on impedance spectroscopy (EIS) and cyclic voltammetry characterization (CV), the modified electrode has shown an enhancement of surface active area and better conductivity compared to the unmodified electrode. Subsequently, both fabricated electrode was further explored as electrochemical biosensor platform based on immobilized PNA probe and methylene blue (MB) was used as an electrochemical indicator to evaluate the performance of PNA electrochemical biosensor. At optimum condition, PBS buffer was choose as supporting buffer, PNA probe concentration of 10 µM, ratio of EDC/NHS of 4 mM:5 mM, MB immersion time of 45 min, MB concentration of 35 µM, pH MB of 7.5, and hybridization temperatures of 27 °C for NH2-GO/QDs/SPCE biosensor. Meanwhile, for biosensor based on NH2-GO/TNCC/SPCE, The optimum detection can be reach by using borate saline buffer as supporting buffer, PNA probe concentration of 20 µM, ratio of EDC/NHS concentration of 6 mM:5 mM (v/v), MB immersion time of 30 min, MB concentration of 45 µM, pH MB of 9 and hybridization temperature at 40 °C. Moreover, reproducibility study exhibited good result with the RSD value of 4.46% for SPCE/NH<sub>2</sub>-GO/ODs and 5.96% for SPCE/NH<sub>2</sub>-GO/NCC, respectively with sufficient selectivity to discriminate between complementary, non-complimentary and one base mismatch DNA. The reduction peak current of MB after hybridization was proportional to the concentration of target MTB DNA in the range from  $1.0 \times 10^{-13}$  to  $1.0 \times 10^{-6}$  mol/L with a detection limit of 8.948  $\times 10^{-13}$  M for NH<sub>2</sub>-GO/QDs/SPCE and 3.14 x 10<sup>-14</sup> M for NH<sub>2</sub>-GO/TNCC/SPCE, respectively. In addition, the stability study has also shown that both fabricated biosensor could be stored at 4°C for 4 weeks with minimum degradation. Meanwhile, for the detection of Mycobacterium tuberculosis from the real sample, DNA amplification step is necessary to amplify genomic MTB DNA to obtain a highly specific MTB DNA fragment sequence. Six difference samples were tested on both fabricated biosensors and demonstrate good result with real sample based on polymerase chain reaction (PCR) amplification product of M. tuberculosis DNA. Both fabricated biosensor also showed successful discrimination on the positive and negative sample with a limit of detection of 2.49  $ng/\mu l$  (NH<sub>2</sub>-GO/QDs/SPCE) and 1.52 ng/µl (NH<sub>2</sub>-GO/TNCC/SPCE), respectively. Therefore, the combination of PCR amplification with electrochemical detection provides a sensitive method for specific sequence detection of mycobacterium tuberculosis (MTB).

Abstrak tesis yang dikemukan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

#### PEMBANGUNAN BIOSENSOR PNA ELEKTROKIMIA BERASASKAN ELEKTROD SKRIN BERCETAK KARBON DIUBAHSUAI BAHAN KOMPOSIT AMINA GRAFIN OKSIDA UNTUK PENGESANAN Mycobacterium tuberculosis

Oleh

#### MOHD HAZANI B. MAT ZAID

Oktober 2017

Pengerusi : Jaafar b. Abdullah, PhD Fakulti : Insititut Teknologi Maju

Elektroforesis gel agarose adalah teknik asas dan penting untuk analisis produk PCR berkaitan mycobacterium tuberculosis. Walau bagaimanapun teknik ini mempunyai beberapa jenis kelemahan seperti menghasilkan keputusan yang tak kuantitatif dan mempunyai kepekaan yang rendah. Oleh itu, satu biosensor elektrokimia PNA baharu berdasarkan elektrod karbon dicetak skrin yang diubahsuai (SPCE) telah dibangunkan untuk pengesanan mycobacterium tuberculosi yang mana boleh mengatasi kelemahan teknik elektroforesis gel agarose. Dalam kajian ini, SPCE telah diubahsuai dengan dua nanomaterial komposit yang berbeza mempunyai fungsian amina grafin oksida terturun (NH<sub>2</sub>-GO) dikompositkan dengan bahan larut air CdS titik kuantum (QDs) dan NH<sub>2</sub>-GO dikompositkan dengan nanoselulos kristal (NH2-GO/NCC) bagi meningkatkan sensitif pengesanan jujukan DNA sasaran berkaitan Mycobacterium tuberculosis. Bahan komposit yang disediakan ini telah diciri menggunakan Raman, FTIR dan TEM. Manakala, mikroskop medan pelepasan (FESEM) dan spektroskopi penyebaran tenaga sinaran-x (EDX) telah mengesahkan bahawa kedua-dua bahan-nano iaitu NH<sub>2</sub>-GO/QDs dan NH2-GO/NCC telah didepositkan dan diselerakkan secara sekata diatas permukaan SPCE. Selain itu, berdasarkan pencirian spektroskopi impeden (EIS) dan voltametri berkitar (CV), elektrod diubahsuai telah menunjukkan peningkatan luas permukaan aktif dan kekonduksian yang lebih baik berbanding elektrod yang tidak diubahsuai. Selanjutnya, kedua-dua elektrod yang difabrikasikan telah diterokai sebagai tapak biosensor elektrokimia berasaskan prob PNA pegun dan metilena biru (MB) telah digunakan sebagai bahan penunjuk elektrokimia untuk menilai prestasi kedua-dua biosensor PNA elektrokimia. Pada keadaan optimum, pemilihan PBS sebagai penampan sokongan, kepekatan prob PNA 10 µM, kadar EDC/NHS 4 mM:5 mM, masa rendaman MB 45 minit, kepekatan MB 35 µM, pH MB 7.5, dan suhu hibridisasi pada 27 °C telah digunakan untuk biosensor NH2-GO/ QDs/SPCE. Sementara itu, untuk biosensor berdasarkan NH2-GO/TNCC/SPCE menggunakan larutan penampan salin borate sebagai penampan sokongan, kepekatan probe PNA 20

uM, nisbah kepekatan EDC/NHS sebanyak 6mM: 5mM (v/v), masa rendaman MB30 minit, kepekatan MB 45 uM, pH MB pada 9 dan suhu hibridisasi 40 °C untuk mendapatkan tahap pengesana optimimum. Tambahan lagi, kajian kebolehulangan juga menunjukkan hasil yang baik dengan nilai RSD iaitu 4.46% untuk NH2-GO/ODs/SPCE dan 5.96% untuk NH<sub>2</sub>-GO/NCC/SPCE, masing-masing dengan keselektifan memadai dapat membezakan diantara DNA pelengkap, DNA bukanpelengkap dan DNA satu asas tidak sepadan. Penurunan puncak arus MB selepas penghibridan berkadar terus kepada kepekatan DNA MTB sasaran dalam julat  $1.0 \times 10^{-13}$  to  $1.0 \times 10^{-6}$  mol/L dengan had pengesanan (LOD) masing-masing iaitu 8.948 × 10-13 M untuk NH2-GO/ODs/ SPCE dan 3.14 x 10-14 M untuk NH2-GO/NCC/SPCE. Tambahan pula, kajian kestabilan juga telah menunjukkan bahawa kedua-dua biosensor yang difabrikasikan boleh disimpan pada 4°C selama 4 minggu (28 hari) dengan kemerosotan yang minimum. Sementara itu, untuk pengesanan Mycobacterium tuberculosis dari sampel sebenar, dapat dilihat bahawa peningkatan isyarat arus MB dicerap apabila penghibridan prob PNA dengan DNA genomik berlaku. Walaubagaimanapun, hasil yang ditunjukan tidak cukup jelas untuk membuktikan keupayaan kedua-dia biosensor yang telah difibrikasikan kerana isu pelekatan tidak spesifik dari DNA genomik yang panjang. Oleh itu, teknik tindakbalas rantai polimerase (PCR) telah digunakan untuk pengandaan jujukan spesifik sasaran untuk mendapatkan jujukan spesifik DNA MTB yang tinggi. Enam sampel berlainan telah diuji keatas kedua-dua biosensor yang difabrikasi dan menunjukkan hasil yang baik dengan sampel sebenar berasaskan penggandaan produk DNA M. tuberculosis melalui tindakbalas rantai polimerase (PCR). Kedua-dua biosensor juga berjaya membezakan diantara sampel positif dan negatif dengan had pengesanan (LOD) adalah 2.49 ng/ul (NH<sub>2</sub>-GO/ODs/SPCE) and 1.52 ng/ul (NH<sub>2</sub>-GO/TNCC/SPCE), masing-masing. Oleh itu, kombinasi penggandaan produk DNA M. tuberculosis mengunakan amplifikasi PCR dan pengesanan elektokimia telah meyediakan kaedah yang berkesan untuk pengesanan mycobacterium tuberculosis (MTB).

#### ACKNOWLEDGEMENTS

Alhamdulillah, this is long journey for me. All the praises and gratitudes are due to Allah S.W.T. First and foremost I would like to thanks my supervisor Dr. Jaafar bin Abdullah for his great guidance, patience, constructive comment and support during all the stages of this Ph.D. study. I would also like to greatly appreciate for my cosupervisor Prof. Nor Azah Yusof and Dr. Helmi Wasoh for their limitless help during my research study. Gratefully acknowledge to Dr. Rahizan Issa for her helps and advice during my Ph.D. program. Special thank goes to my close research partner Puzi Alttas, Fariza, and Amalina. I am also thankful to the people in the biosensor group in 103 lab and bacteriology unit staff under Institute of medical research (IMR). I am grateful and pray for your success in this life and the next to come. To the University Putra Malaysia (UPM) thank you so much for providing research facilities and Kementerian Pengajian Tinggi Malaysia (KPTM) for providing financial aids in Mybrain 15 (My Ph.D.)

Special thanks to my mother Nahyati binti Salleh and my father Mat Zaid bin Ismail for their love, support, and encouragement. Not forget to my brothers and lastly, I would acknowledge to Nur Hafiza Nasir who accompanied me throughout the entire process in pursuing my degree of doctor of philosophy. Thank you very much I certify that a Thesis Examination Committee has met on 25 October 2017 to conduct the final examination of Mohd Hazani bin Mat Zaid on his thesis entitled "Development of PNA Electrochemical Biosensor Based on Screen-Printed Electrode-Modified Amine-Functionalized Graphene Composite for *Mycobacterium tuberculosis* Detection" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

### Suraya binti Abdul Rashid, PhD

Associate Professor Faculty of Engineering Universiti Putra Malaysia (Chairman)

Mohd Nizar bin Hamidon, PhD Associate Professor Institute of Advanced Technology Universiti Putra Malaysia (Internal Examiner)

#### Zulkarnain bin Zainal, PhD

Professor Faculty of Science Universiti Putra Malaysia (Internal Examiner)

#### Bambang Kuswandi, PhD

Professor University of Jember Indonesia (External Examiner)

NOR AINI AB. SHUKOR, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 28 December 2017

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the supervisory committee are as follows:

#### Jaafar Abdullah, PhD

Senior Lecturer Faculty of Science Universiti Putra Malaysia (Chairman)

#### Nor Azah Yusof, PhD Professor Faculty of Science Universiti Putra Malaysia (Member)

### Helmi wasoh, PhD

Senior Lecturer Faculty of Biotechnoly and Biomolecule Universiti Putra Malaysia (Member)

### **ROBIAH BINTI YUNUS, PhD**

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

#### **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations, and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the university Putra Malaysia (research) rules 2012;
- written permission must be obtained from supervisor and the office of deputy vice-chancellor (research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the university Putra Malaysia (research) rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the university Putra Malaysia (graduate studies) rules 2003 (revision 2012-2013) and the university Putra Malaysia (research) rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_

Date:

Name and Matric No: Mohd Hazani B. Mat Zaid, GS 41859

#### Declaration by Members of Supervisory committee

This is to confirm that:

 $(\mathbf{C})$ 

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to :

| Signature:<br>Name of<br>Chairman of<br>Supervisory<br>Committee: | JPM |
|-------------------------------------------------------------------|-----|
| Signature:<br>Name of<br>Member of<br>Supervisory<br>Committee:   |     |
| Signature:<br>Name of<br>Member of<br>Supervisory<br>Committee:   |     |

# TABLES OF CONTENTS

|                    | Page  |
|--------------------|-------|
| ABSTRACT           | i     |
| ABSTRAK            | iii   |
| ACKNOWLEDGEMENTS   | v     |
| APPROVAL           | vi    |
| DECLARATION        | viii  |
| LIST OF TABLES     | xvii  |
| LIST OF FIGURES    | xviii |
| LIST OF SCHEMATICS | xxiii |
|                    |       |

| 1 | INT | RODUCTIO     | DN                                                |          |
|---|-----|--------------|---------------------------------------------------|----------|
|   | 1.1 | Backgrou     | ind of study                                      | 1        |
|   | 1.2 | Problem      | Statement                                         | 3        |
|   | 1.3 | Objective    | e of the study                                    | 4        |
|   | 1.4 | Novelty a    | and benefits of research                          | 5        |
|   | 1.5 | Scope an     | d limitation                                      | 5        |
|   |     |              |                                                   |          |
|   |     |              |                                                   |          |
| 2 | LIT | ERATURE      | REVIEW                                            |          |
|   | 2.1 | Status tube  | erculosis in Malaysia                             | 6        |
|   | 2.2 | Common t     | est for TB detection and analysis                 | 6        |
|   | 2.3 | Biosensor    | for TB diagnostics                                | 9        |
|   | 2.4 | Electroche   | mical biosensor                                   | 9        |
|   |     | 2.4.1        | Electrochemical DNA biosensor principle           | 10       |
|   |     | 2.4.2        | Electrochemical biosensor based on Screen         | 11       |
|   |     |              | printed electrode                                 |          |
|   | 2.5 | Peptide nu   | clide acid (PNA)                                  | 12       |
|   |     | 2.5.1        | The advantages of PNA in biosensor                | 13       |
|   |     |              | application                                       |          |
|   |     | 2.5.2        | Developed electrochemical biosensor based         | 14       |
|   |     |              | on PNA probe                                      |          |
|   | 2.6 | Indirect ele | ectrochemical DNA detection                       | 15       |
|   |     | 2.6.1        | Electrochemistry of Methylene blue as redox       | 17       |
|   |     |              | indicator                                         | 17       |
|   | 2.7 | Nanomater    | rial in development of biosensor                  | 18       |
|   | 2.8 | Properties   | of graphene oxide                                 | 19       |
|   |     | 2.8.1        | Functionalization of graphene oxide               | 21       |
|   |     | 2.8.2        | Amine functionalization graphene oxide            | 21       |
|   | 2.9 | Graphene/    | RGO/fGO nanocomposite                             | 22       |
|   |     | 2.9.1        | Properties of Quantum dots                        | 24       |
|   |     | 2.9.2        | Properties of nanocellulose                       | 25       |
|   |     | 2.9.3        | Surface modification of nanocellulose             | 26       |
|   |     | 2.9.4        | The advantages of nanocrystalline cellulose (NCC) | 23<br>27 |
|   |     | 2.9.5        | Graphene composited nanocellulose                 | 27       |
|   |     |              |                                                   |          |

| 2.10 | Oligonucleotide immobilization on solid support |                                      |    |
|------|-------------------------------------------------|--------------------------------------|----|
| 2.11 | Oligonucleotide Surface density                 |                                      |    |
| 2.12 | Electrochemical technique                       |                                      |    |
|      | 2.12.1                                          | Cyclic Voltammetry (CV) theory       | 31 |
|      | 2.12.2                                          | Laviron approach                     | 31 |
|      | 2.12.2                                          | Cyclic voltammetry theory            | 33 |
|      | 2.12.3                                          | Differential Pulse Voltammetry (DPV) | 33 |
|      | 2.12.3                                          | Chronocoulometry (CC)                | 33 |

# METHODOLOGY

| 3.1  | Nanomaterial preparation                                            |    |
|------|---------------------------------------------------------------------|----|
|      | 3.1.1 Preparation of CdS QDs                                        | 36 |
|      | 3.1.2 Functionalization of grapheme oxide with                      | 36 |
|      | ethylenediamine                                                     |    |
|      | 3.1.3 Preparation amine functionalized graphene                     | 36 |
|      | oxide /CdS QDs (NH <sub>2</sub> - GO/QDs)                           |    |
|      | 3.1.4 Preparation TEMPO-oxidize Nanocrystalline                     | 36 |
|      | cellulose                                                           |    |
|      | 3.1.5 Preparation amine functionalized graphene                     | 36 |
|      | oxide /TEMPO-nano crystalline cellulose                             |    |
|      | (NH <sub>2</sub> -GO/TNCC)                                          |    |
|      | 3.1.6 Preparation of NH <sub>2</sub> -GO/QDs modified SPCE          | 36 |
|      | 3.1.7 Preparation of NH <sub>2</sub> -GO/TNCC modified on           | 37 |
| 2 2  | Descents and solutions                                              | 27 |
| 3.2  | 2.2.1 Chamical solvent and biological reagents                      | 37 |
| 33   | Apparatus and instrumentation                                       | 30 |
| 5.5  | 3 3 1 Flectrochemical measurement                                   | 39 |
|      | 3.3.2 Instrumentation                                               | 40 |
| 34   | Peptide Nucleic Acid (PNA) probe related to                         | 42 |
| 5.1  | Mycobacterium tuberculosis and other DNA sequences                  | .2 |
| 3.5  | Other buffer solution                                               | 42 |
| 3.6  | PNA immobilization and hybridization procedure on NH <sub>2</sub> - | 43 |
|      | GO/QDs and NH <sub>2</sub> -GO/TNCC                                 |    |
| 3.7  | Characterization of modified electrode                              | 44 |
| 3.8  | Effect of supporting electrolyte                                    | 45 |
| 3.9  | Effect of EDC/NHS ratio on Surface probe PNA density                | 45 |
| 3.10 | Effect of pH.                                                       | 46 |
| 3.11 | Effect of temperature                                               | 46 |
| 3.12 | Selectivity study                                                   | 46 |
| 3.13 | Sensitivity study                                                   | 46 |
| 3.14 | Stability study                                                     | 46 |
| 3.15 | Regeneration study                                                  | 47 |
| 3.16 | Scan rate study                                                     | 47 |
| 3.17 | Preparation of Mycobacterium tuberculosis (MTB)                     | 47 |
|      | genomic sample                                                      |    |
|      | 3.17.1 Laboratory procedure                                         | 47 |
| 3.18 | MTB genomic PCR amplification                                       | 48 |
| 3.19 | Preparation an electrophoresis agarose gel                          | 49 |

3

|     | 3.19.1                                                               | Evaluation agarose gel result                                                                                                                      | 50       |
|-----|----------------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------|----------|
| 3.2 | 0 Applicatio                                                         | on with portable potentiostat                                                                                                                      |          |
| 3.2 | 1 Summary<br>electroche<br>for <i>Mycol</i><br>NH <sub>2</sub> -GO/0 | of work flow for development of<br>mical PNA biosensor<br><i>pacterium tuberculosis</i> based on SPCE modified<br>QDs and NH <sub>2</sub> -GO/TNCC | 51       |
|     |                                                                      |                                                                                                                                                    |          |
| 4 F | RESULTS AN                                                           | D DISCUSSION                                                                                                                                       |          |
| 4.  | 1 Preparatio<br>composite<br>GO/QDs)                                 | n of amine functionalized graphene oxide<br>d water soluble CdS quantum dots (NH <sub>2</sub> -                                                    | 53       |
| 4.  | 2 Characteri                                                         | zation of synthesis of CdS Quantum dots                                                                                                            | 53       |
| 4.  | .3 Functional ethylenedi                                             | lization of graphene oxide (GO) with amine (EDA)                                                                                                   | 55       |
|     | 4.3.1                                                                | Effect of ethylenediamine (EDA) on surface                                                                                                         | 55       |
| 4.  | 4 Cyclic vol                                                         | tammetry study                                                                                                                                     | 56       |
| 4.  | 5 Raman Ch                                                           | aracterization                                                                                                                                     | 57       |
| 4.  | 6 Optimizat                                                          | ion ratio between NH <sub>2</sub> -GO and CdS QDs                                                                                                  | 59       |
| 4.  | .6 4.6.1                                                             | FTIR characterization                                                                                                                              | 61       |
|     | 4.6.2                                                                | Electrodeposition of NH <sub>2</sub> -GO/QDs on screen printed electrode                                                                           | 62       |
|     | 4.6.3                                                                | FESEM-EDX Characterization                                                                                                                         | 63       |
| 4.  | 7 Preparatio                                                         | n of amine functionalized graphene oxide                                                                                                           | 64       |
|     | composite                                                            | water TEMPO nanocrystalline cellulose                                                                                                              | 61       |
|     | 4.7.1                                                                | Characterization of TEMPO-nanocrystalline                                                                                                          | 04<br>65 |
|     | 4.7.2                                                                | cellulose (TNCC)                                                                                                                                   | 05       |
|     | 4.7.3                                                                | Optimization amount of NH <sub>2</sub> -GO and TNCC                                                                                                | 66       |
|     | 4.7.4                                                                | FTIR characterization for amine                                                                                                                    | 68       |
|     |                                                                      | Nanocellulose ( $NH_2$ -GO/TNCC)                                                                                                                   |          |
|     | 4.7.5                                                                | Drop casting technique of NH <sub>2</sub> -GO/T-NCC                                                                                                | 69       |
|     | 4.7.6                                                                | FESEM-EDX Characterization of NH <sub>2</sub> -<br>GO/TNCC                                                                                         | 70       |
|     | 4.7.7                                                                | Electrochemical behavior of modified                                                                                                               | 72       |
|     |                                                                      | electrode for NH <sub>2</sub> -GO/QDs and NH <sub>2</sub> -<br>GO/TNCC on screen printed carbon<br>electrode                                       |          |
| 4.  | 8 4.8.1                                                              | Cyclic voltammetry investigation                                                                                                                   | 72       |
|     | 4.8.2                                                                | Electrochemical Impedance Spectroscopy                                                                                                             | 74       |
|     |                                                                      | (EIS) investigation                                                                                                                                |          |
| 4.  | 9 Electro ac<br>GO/TNCC                                              | tive surface area of NH <sub>2</sub> -GO/QDs and NH <sub>2</sub> -C                                                                                | 76       |
| 4.  | 1 The electr                                                         | ochemical behavior of methylene blue as redox                                                                                                      | 79       |
|     | mediator                                                             | in fabrication PNA biosensor for                                                                                                                   |          |
|     | Mycobact                                                             | erium Tuberculosis detection                                                                                                                       | 00       |
|     | 4.10.1                                                               | Preliminary Investigation                                                                                                                          | 80       |
| 4.1 | i Optimizat                                                          | ion Condition for biosensor performance                                                                                                            | 81       |

|      | 4.11.1                           | Effect of buffer solution as supporting electrolyte.                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                | 83  |
|------|----------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----|
|      | 4.11.2                           | Effect of PNA probe concentration                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                   | 83  |
|      | 4.11.3                           | Effect of probe density                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                             | 84  |
|      | 4.11.4                           | Effect of MB concentration and immersion time                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                       | 87  |
|      | 4.11.5                           | Effect of pH.                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                       | 88  |
|      | 4.11.6                           | Effect of hybridization temperature and time                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                        | 90  |
| 4.12 | Selectivity                      |                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                     | 93  |
| 4.13 | Sensitivity                      |                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                     | 95  |
| 4.14 | Reproducib                       | ility                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                               | 97  |
| 4.15 | Regeneration and Stability Study |                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                     |     |
| 4.16 | Scan rate st                     | udy for the determination rate electron transfer                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                    |     |
|      | of NH <sub>2</sub> -GO           | /QDs/SPCE and NH <sub>2</sub> -GO/TNCC/SPCE                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                         |     |
| 4.17 | Real Sampl<br>genomic            | e analysis using the amplification of MTB                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                           | 103 |
| 4.18 | Polymerase                       | chain reaction (PCR) technique for DNA                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                              | 104 |
|      | amplificatio                     | on and a second s |     |
|      | 4.18.1                           | Effect of varying the number of amplification cycles                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                | 104 |
|      | 4.18.2                           | Selectivity study on real sample                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                    | 106 |
|      | 4.18.3                           | Sensitivity                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                         | 107 |
| 4.19 | Application                      | PNA electrochemical biosensor on portable                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                           | 107 |
|      | device                           |                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                     |     |

### 5 SUMMARY, CONCLUSION, AND RECOMMODATION

| 5.1 | Research Summary                    | 108 |
|-----|-------------------------------------|-----|
| 5.2 | Conclusion                          | 113 |
| 5.3 | Recommendations for future research | 113 |

112

142

148

149

REFERENCES APPENDICES BIODATA OF STUDENT LIST OF PUBLICATIONS

xiii

# LIST OF TABLES

| Table |                                                                                                                                                          | Page |
|-------|----------------------------------------------------------------------------------------------------------------------------------------------------------|------|
| 2.1   | TB cases reported in 2015 in Malaysia (Source: World health of organization (WHO)                                                                        | 6    |
| 2.2   | Comparison of different biosensors for MTB detection technology                                                                                          | 9    |
| 2.3   | List of redox mediator used in the previous study.                                                                                                       | 16   |
| 2.4   | List of difference modified by methylene blue redox<br>indicator                                                                                         | 23   |
| 3.1   | List of chemical, solvent and buffer                                                                                                                     | 37   |
| 3.2   | List of apparatus used in this study                                                                                                                     | 40   |
| 3.3   | The sequences of oligonucleotides involved in this study                                                                                                 | 42   |
| 3.4   | Component mixture in MTB master mix                                                                                                                      | 48   |
| 4.1   | The Current and charge transfer resistance at different modified electrodes obtained in in 5 mM potassium ferrocyanide( $[Fe(CN)_6]^{3-/4-}$ / 1.0 M KCl | 75   |
| 4.2   | Comparison performance of different biosensors for the detection of M. Tuberculosis DNA reported in the literature.                                      | 96   |

C,

# LIST OF FIGURES

| Figure |                                                                                                                                                                           | Page |
|--------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------|
| 2.1    | Current tuberculosis diagnostics pipeline listing the development phases at every stages (Wang et al., 2011)                                                              | 8    |
| 2.2    | Components of a typical electrochemical biosensor.                                                                                                                        | 11   |
| 2.3    | Three electrode system screen printed been used in analytical analysis                                                                                                    | 12   |
| 2.4    | a) Differences between DNA and PNA; b)<br>Hybridization between PNA and DNA/RNA                                                                                           | 13   |
| 2.5    | The schematic of mechanism of methylene blue (MB) reduction at modified electrode                                                                                         | 18   |
| 2.6    | a) Graphene honeycomb lattice; b) Oxidation of graphite to graphene oxide and reduction to reduced graphene oxide (Source : Wang et al. (2012)                            | 21   |
| 2.7    | Proposed mechanism simultaneous reduction and functionalization of GO with ethylenediamine (Source: Kim et al. (2013)                                                     | 22   |
| 2.8    | Illustrated CdS Quantum dots capped with MPA                                                                                                                              | 25   |
| 2.9    | The chemical structure of cellulose                                                                                                                                       | 26   |
| 2.10   | Sulfo-NHS plus EDC (carbodiimide) crosslinking reaction. (Source Panchaud et al. (2008))                                                                                  | 29   |
| 2.11   | Variation the applied potential as a function of time in<br>a cyclic voltammetry experiment                                                                               | 31   |
| 2.12   | A typical cyclic voltammogram of current verses potential                                                                                                                 | 32   |
| 2.13   | (a)Potential wave from employed in<br>chronocoulometry. (b) A typical chronocoulometry<br>response                                                                        | 34   |
| 3.1    | NH <sub>2</sub> -GO sample after dried at 60 °C                                                                                                                           | 35   |
| 3.2    | The electrodeposition of NH <sub>2</sub> -GO on SPCE                                                                                                                      | 37   |
| 3.3    | a) Images of electrochemical instrumentation used in<br>this study; b) Images of SPCE used for NH <sub>2</sub> -<br>GO/ODs : c) Images of SPCE used for NH <sub>2</sub> - | 40   |

G

GO/TNCC

| 3 | 3.4 | Preparation step electrochemical PNA biosensor<br>modified electrode based on amine functionalized<br>graphene oxide composite quantum dots (NH <sub>2</sub> -<br>GO/QDs/SPCE)                                                                    | 43 |
|---|-----|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----|
| 3 | 3.5 | Preparation step of electrochemical PNA biosensor<br>modified electrode based on amine functionalized<br>graphene oxide composite nanocrystalline cellulose<br>(NH <sub>2</sub> -GO/TNCC/SPCE)                                                    | 44 |
| 3 | 3.6 | The inoculated treated sample in ogawa or LJ (Lowenstein Jensen) media; b) Smearing technique using fluorescent microscopy                                                                                                                        | 48 |
|   | 3.7 | a) Termocycler to run PCR amplification; b) Setup gel tray with the well comb for electrophoresis study.                                                                                                                                          | 49 |
|   | 3.8 | Portable Potentiostat                                                                                                                                                                                                                             | 50 |
| 3 | 3.9 | Flow chart process for development of electrochemical PNA biosensor                                                                                                                                                                               | 52 |
| 2 | 4.1 | Illustration of possible mechanism for the preparation of NH <sub>2</sub> -GO/QDs composite                                                                                                                                                       | 54 |
| 2 | 4.2 | a) CdS QDs solution images b) Absorption spectra of<br>CdS QDs sample c) TEM micrograph at 25K<br>magnification of CdS QDs nanoparticles d) Zeta<br>potential analysis                                                                            | 55 |
| 2 | 4.3 | Zeta potential result of A) GO; B) NH <sub>2</sub> -GO. C) Graph<br>Zeta potential at difference pH of a) GO; b) NH <sub>2</sub> - GO                                                                                                             | 56 |
| 2 | 4.4 | Effect of current at different EDA concentration on cyclic voltammetry in 5 mM ferro-/ferricyanide $(Fe(CN)_6^{4/3-})$ containing 0.1 M KCl; (a) 0.1M, (b) 0.2 M, (c) 0.3 M, (d) 0.4 M.                                                           | 57 |
|   | 4.5 | Raman spectra a) GO; b) NH <sub>2</sub> -GO                                                                                                                                                                                                       | 58 |
|   | 4.6 | A) CV response B) Histogram plot at difference ratio between CdS QDs and NH <sub>2</sub> -GO at a) 1:7.5 b) 1:5 c) 1:2.5 in W%, C) Images of composite material of NH <sub>2</sub> -GO, D) TEM image of composite material of NH <sub>2</sub> -GO | 60 |
|   | 4.7 | FTIR spectra and Raman of a) Graphane oxide (GO);<br>b) Amine functionalized reduced graphane oxide<br>(NH <sub>2</sub> -GO); c) NH <sub>2</sub> -GO/QDs                                                                                          | 61 |
| 2 | 4.8 | CV Voltammograms after electrodeposition of A) CdS                                                                                                                                                                                                | 62 |

|      | B) NH <sub>2</sub> -GO (C) NH <sub>2</sub> -GO/QDs in 0.1 Lithium perchlorate; (D) Histogram effect of current at difference CV cycle                                                                                                                                                                         |    |
|------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----|
| 4.9  | FESEM images-EDX analysis of (ai-aii) CdS QDs ;(bi-bii) NH <sub>2</sub> -GO; (ci-cii) NH <sub>2</sub> -GO/QDs                                                                                                                                                                                                 | 64 |
| 4.1  | Illustration of possible mechanism for preparation of (TEMPO)-nanocrystalline cellulose (TNCC) and amine functionalized graphene oxide (NH <sub>2</sub> -GO) to produce NH <sub>2</sub> -GO/TNCC composite                                                                                                    | 65 |
| 4.11 | a) Prepared TEMPO-nanocellulose (TNCC); b) UV VIS; c) Zeta potential analysis; d) TEM images                                                                                                                                                                                                                  | 66 |
| 4.12 | Ai) CV Voltommagram (Aii) Histogram effect<br>amount of TEMPO-nanocrystalline celulose (TNCC);<br>(Bi) CV Voltommagram (Bii) Histogram effect of<br>difference amount of NH <sub>2</sub> -GO (C) NH <sub>2</sub> -GO/T-NCC<br>in glass sample bottle; (D) TEM images of prepared<br>NH <sub>2</sub> -GO/TNCC. | 67 |
| 4.13 | FTIR spectra of a) Nanocrystalline cellulose (NCC);<br>b) TEMPO-nanocrystalline nanocellulose (TNCC); c)<br>NH <sub>2</sub> -GO ; d) NH <sub>2</sub> -GO/TNCC                                                                                                                                                 | 68 |
| 4.14 | Effect of NH <sub>2</sub> -GO volume on working electrode surface of SPCE                                                                                                                                                                                                                                     | 69 |
| 4.15 | FESEM images with EDX analysis of NH <sub>2</sub> -GO-(a-<br>aii); (b) TNCC (b-bii); NH <sub>2</sub> -GO/TNCC (c-cii)                                                                                                                                                                                         | 71 |
| 4.16 | Cyclic voltammetry characterization; (A); bare SPCE<br>(a), NH <sub>2</sub> -GO (b), NH <sub>2</sub> -GO/SPCE (c), NH <sub>2</sub> -GO/QDs<br>(d). (B); a) Bare SPCE, TNCC/SPCE, c) NH <sub>2</sub> -GO/SPCE, NH <sub>2</sub> -GO/TNCC (d) in 5 mM<br>([Fe(CN) <sub>6</sub> ] <sup>3-/4-)</sup> /1.0 M KCl    | 73 |
| 4.17 | Electrochemical impedance spectroscopy (EIS): (A)<br>Bare SPCE (a), CdS/SPCE (b), NH <sub>2</sub> -GO/SPCE (c)<br>NH <sub>2</sub> -GO/QDs (d); (B) Bare SPCE (a), TNCC/SPCE<br>(b), NH <sub>2</sub> -GO/SPCE (c) NH <sub>2</sub> -GO TNCC/SPCE (d)                                                            | 75 |
| 4.18 | CV response at different scan rate (50mV to 350mV)                                                                                                                                                                                                                                                            | 78 |
|      |                                                                                                                                                                                                                                                                                                               |    |

|  |      | consist of a) Bare SPCE, b) NH <sub>2</sub> -GO/QDs/SPCE, c) The plot of oxidation and reduction peak current of $(Fe(CN)_6]^{3./4}$ for bare SPCE and NH <sub>2</sub> -GO/QDs/SPCE against square roots of scan rate, d) Bare SPCE; e) NH <sub>2</sub> -GO/TNCC; f) The plot of oxidation and reduction peak current of $(Fe(CN)_6]^{3./4-}$ of bare SPCE and NH <sub>2</sub> -GO/TNCC/SPCE against square roots of scan rate, in 5mM $(Fe(CN)_6]^{3./4-}$ solution containing supporting electrolyte (0.1 M KCl) (pH 7.4) |    |
|--|------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----|
|  | 4.19 | CV diagram of A) NH <sub>2</sub> -GO/QDs/SPCE; B) NH <sub>2</sub> -GO/TNCC/SPCE with methylene blue $(35\mu M \text{ and } 45\mu M)$ respectively(a) in the absent of PNA b) Immobilization with PNA c) Hybridization with DNA target (Scan rate: 50 mV)                                                                                                                                                                                                                                                                    | 80 |
|  | 4.2  | CV for NH <sub>2</sub> -GO/QDS (A) NH <sub>2</sub> -GO/TNCC (B) for<br>effect of buffer solution on the voltammetry MB<br>signal with PNA-DNA hybridization of (a) Tris-HCl<br>buffer (b) TE buffer (c) TAE buffer (d) PBS buffer<br>with equal strength (0.1 M) with 50 $\mu$ M Methylene<br>blue at 50 mV scan rate                                                                                                                                                                                                       | 82 |
|  | 4.21 | DPV response by difference probe concentration a)<br>NH <sub>2</sub> -GO/QDs; b) NH <sub>2</sub> -GO/TNCC                                                                                                                                                                                                                                                                                                                                                                                                                   | 83 |
|  | 4.22 | Chronocoulometry response at difference ratio of EDC/NHS (a-b) NH <sub>2</sub> -GO/QDs; (c-d) NH <sub>2</sub> -GO/TNCC                                                                                                                                                                                                                                                                                                                                                                                                      | 85 |
|  | 4.23 | Chronocoulometry response at individual modified<br>SPCE a(i) CdS QDs; a(ii) NH <sub>2</sub> -GO; a(iii) NH <sub>2</sub> -<br>GO/QDs ; b(i) TNCC; b(ii) NH <sub>2</sub> -GO; b(iii) NH <sub>2</sub> -<br>GO/TNCC : DPV response of difference probe density<br>c) NH <sub>2</sub> -GO/QDs/SPCE; d) NH <sub>2</sub> -GO/TNCC/SPCE.                                                                                                                                                                                           | 87 |
|  | 4.24 | Optimization of methylene blue (MB) concentrations<br>(a) NH <sub>2</sub> -GO/QDs; b) NH <sub>2</sub> -GO/TNCC; Optimization<br>of immersion time (c) NH <sub>2</sub> -GO/QDs (d) NH <sub>2</sub> -<br>GO/TNCC                                                                                                                                                                                                                                                                                                              | 88 |
|  | 4.25 | DPV response and plot current again pH buffer solution containing Methylene blue for (a-b) NH <sub>2</sub> -GO/QDs (c-d) NH <sub>2</sub> -GO/TNCC                                                                                                                                                                                                                                                                                                                                                                           | 90 |
|  | 4.26 | Effect of hybridization temperature and time on the DPV signal current of methylene blue (a) NH <sub>2</sub> -GO/QDs/SPCE/PNA-DNA; (b) NH <sub>2</sub> GO/TNCC/SPCE/PNA-DNA                                                                                                                                                                                                                                                                                                                                                 | 92 |
|  | 4.27 | DPV response curve and bar graph of MB current at                                                                                                                                                                                                                                                                                                                                                                                                                                                                           | 94 |

xviii

various condition of target DNA; a) PNA; b) Non-Complimentary; c) two base mismatch; d) One base mismatch d) PNA-DNA of (A-B) SPCE/NH<sub>2</sub>-GO/QDs; (C-D) SPCE/NH<sub>2</sub>-GO/TNCC

| 4.28 | DPV curve at different concentrations of complementary DNA. Inset is linear relationship between DPV peak current (Ip) with the different concentrations of (A) NH <sub>2</sub> -GO/QDs/SPCE with complementary DNA (From a to g was $1 \times 10^{-6}$ M to $1 \times 10^{-11}$ M) in 0.1 M PBS buffer pH 7. (B) NH <sub>2</sub> -GO/TNCC/SPCE with complementary DNA (From a to g was $1 \times 10^{-7}$ M $\times 10^{-13}$ M in borate buffer pH 9. | 96  |
|------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----|
| 4.29 | DPV response for five consecutive electrode; inset<br>current against number of electrode (a) NH <sub>2</sub> -GO/QDs<br>(b) NH <sub>2</sub> -GO/TNCC                                                                                                                                                                                                                                                                                                   | 98  |
| 4.3  | (A) Regeneration studies of a) NH <sub>2</sub> -GO/QDs/SPCE b) NH <sub>2</sub> -GO/TNCC/SPCE Stability study for 28 days (a) NH <sub>2</sub> -GO/QDs (b) NH <sub>2</sub> -GO/TNCC                                                                                                                                                                                                                                                                       | 100 |
| 4.31 | Plotted graph log of scan rate (V) vs log of peak<br>current (Ip) at 50-350 mV/s) before and after<br>hybridization (a-b); Ln scan rate (V) vs variation<br>potential (Ep) before and after hybridization (c-d) for<br>NH <sub>2</sub> -GO/QDs/SPCE (A) NH <sub>2</sub> -GO/TNCC(B)                                                                                                                                                                     | 102 |
| 4.32 | Bar graph current reading at difference cycle of a)<br>NH <sub>2</sub> -GO/QDs/SPCE; b) NH <sub>2</sub> -GO/TNCC; c) gel<br>electrophoresis photographs of PCR products obtained<br>by difference cycle                                                                                                                                                                                                                                                 | 105 |
| 4.33 | Gel electrophoresis photographs of PCR products for<br>six difference sample                                                                                                                                                                                                                                                                                                                                                                            | 107 |
| 4.34 | DPV response of (A-B) NH <sub>2</sub> -GO/QDs/SPCE and (C-D) NH2-GO/TNCC/SPCE in real samples detection;<br>a) PNA probe, b) Negative sample, c) M227/1<br>(MOTT), d) 1914,e) 1915, f) 3316,g) 8316                                                                                                                                                                                                                                                     | 107 |
| 4.35 | DPV response and plotted graph log concentration<br>amplified DNA at difference concentration of; (a-b)<br>NH <sub>2</sub> -GO/QDs/SPCE; (c-d) NH <sub>2</sub> -GO/TNCC/SPCE                                                                                                                                                                                                                                                                            | 109 |
| 4.36 | Current response with difference types of sample by using portable potentiostat                                                                                                                                                                                                                                                                                                                                                                         | 110 |

C

# LIST OF ABBREVIATIONS

| PNA             | Peptide Nucleic Acid                   |
|-----------------|----------------------------------------|
| DNA             | Dioxyribonucleic Acid                  |
| GO              | Graphene Oxide                         |
| RGO             | Reduce Graphene Oxide                  |
| QDs             | Quantum Dots                           |
| TNCC            | TEMPO-Nanocrystalline cellulose        |
| NCC             | Nanocrystalline cellulose              |
| MTB             | Mycobacterium tuberculosis             |
| SPCE            | Screen Printed Carbon Electrode        |
| МВ              | Methylene Blue                         |
| CV              | Cyclic Voltammetry                     |
| EIS             | Electrochemical Impedance Spectroscopy |
| DPV             | Differential Pulse Voltammetry         |
| PCR             | Polymerase chain reaction              |
| Eo              | Formal Reduction Potential             |
| Ei              | Initial Potential                      |
| E <sub>pa</sub> | Anodic Peak potential                  |
| E <sub>pc</sub> | Cathodic peak potential                |
| $\Delta E_p$    | Peak potential separation              |
| i               | Current                                |
| i <sub>pa</sub> | Anodic Peak Current                    |
| i <sub>pc</sub> | Cathodic Peak Current                  |
| n               | Number of electron                     |
| Q               | Quantity of charge                     |

| POC            | Point Of Care                                                         |
|----------------|-----------------------------------------------------------------------|
| EDC/NHS        | 1-Ethyl-3-(3-dimethylaminopropyl (EDC)/<br>N-hydroxysuccinimide (NHS) |
| T <sub>m</sub> | Melting temperature                                                   |
| Thyb           | Hybridization temperature                                             |



 $\bigcirc$ 

#### **CHAPTER 1**

#### INTRODUCTION

#### 1.1 Background of study

*Mycobacterium tuberculosis* (MTB) is an obligate pathogenic bacterial species in the family of *mycobacteriaceae* and a causative agent of most cases of tuberculosis. Tuberculosis bacteria can spread through the air, either by coughing, sneezing, talking, or spitting. Only a few of these bacteria can inflict a healthy person to be infected (WHO, 2015).

In the current situation in Malaysia, tuberculosis (TB) is fast rising as a noncommunicable disease (NCD), with the death rate even higher than that of dengue where a total of 1.520 deaths from TB were recorded in 2012 compared to 34 deaths from dengue in the same year. Latest data provided by world health organization (WHO) shows that a total number of TB cases were recorded in Malaysia about 24220 TB cases with the Figures of mortality was 2400 people (WHO, 2015b). Traditionally, microbial culture-based tests were used as a diagnostic method to identify mycobacterium bacteria based on sputum sample from the patient. However, this approach is time consuming and laborious, especially for the slow-growing bacteria like MTB (Tortoli et al., 1997; Colebunders et al., 2000). As time evolved, alternative methods that have been developed for rapid detection of MTB include polymerase chain reaction (PCR) (Brisson et al., 1999, Wilson et al., 1994), Latex agglutination (Krambovitis et al., 1984), enzyme-linked immunosorbent assay (Chan et al., 2000), radiometric detection (Middlebrook et al., 1997), gen-probe amplified M. tuberculosis direct test (AMTDT) (Gamboa et al., 1997), TB rapid cultivation detection technique, such as MB/Bact system (Liu et al., 2001), BactecMGIT 960 system and flow cytometry (Qin et al., 2008). These methods are more sensitive and rapid than the traditional microbial culture-based method (Siddigi et al., 2003). However, they cannot provide the detection results in real-time, and most of these methods require complex instrumentation and highly qualified personnel. Therefore, the development of portable, real-time, sensitive, rapid, and accurate methods detection is essential to prevent tuberculosis infection effectively.

The biosensor is the best candidate to overcome this situation. Currently, various reviews on biosensors have been reported for tuberculosis detection includes electrochemical sensors (Liu et al., 2014), piezoelectric biosensors (Kaewphinit et al., 2010), optical biosensors (Prabhakar et al., 2008) and magnetoelastic biosensors (Park et al., 2012). Although the reported biosensors have gained excellent performance. However, the sensitivity is a major issue that requires further improvement.

Electrochemical DNA biosensors recently have found significant applications for onsite detection especially for clinical and medical diagnostics (Zhou et al., 2011). While offering simplicity in operation and sample manipulation, the electrochemical biosensor also yields highly sensitive and specific measurements for a broad spectrum of biomolecules (Liepold et al., 2005). Moreover, sample size requirement for performing electrochemical biosensors is small, ranging from several microliters to hundreds of nanoliters, which includes the sample pretreatment reagents (Wei et al., 2010). Additionally, the latest technology has introduced commercially available screen printed electrode (SPE) which offer disposability, lower cost, and miniaturization (Dominguez et al., 2007). However, the most important feature of electrochemical biosensors is their potential to be easily transformed from a laboratorybased instrument to the point of care (POC) device.

Compared to their DNA counterparts, Peptide Nucleic Acid (PNA) recognition layers can offer significant advantages for sequence-specific DNA biosensors. As previously reported, PNA provides higher sensitivity and specificity including greater discrimination against single-base mismatches, faster hybridization at room temperatures, and minimal dependence on ionic strength, and promising for possible employment in the fabrication of biosensors for various applications (Wang et al., 1996). As a consequence, PNA has stimulated several studies to test new capabilities and opportunities afforded by the use of PNA as surface probes.

With exceptional achievements in nanotechnology and nanoscience, nanomaterialbased electrochemical signal amplifications offer huge potential of improving both sensitivity and selectivity for electrochemical sensors and biosensors (Zhu et al., 2015). Graphene oxide (GO) is one of the advance nanomaterials can be used as an electrode modifier in electrochemical biosensor provided a biocompatible matrix to improve the performance of electrochemical biosensor (Sharma et al., 2015). More recently, the applications of being explored widely on the surface functionalization of GO by combination with long chain aliphatic group, aromatic amines, amino acids, and polymers (Kuila et al., 2012). As a result, reduced GO is also called functionalized GO or chemically reduced GO provide abundant structural defects and functional groups offer advantageous for electrochemical applications (Tang et al., 2009).

In addition, the combination of GO or reduced graphene oxide (rGO) with others nanoparticles may contribute additional advantages such as to enhance the shelf-life of the chosen bio-recognition element (Artiles et al., 2011). Over the past few years, semiconductor quantum dots (QDs) have attracted increasing attention for sensing application due to its unique properties such as high surface activity, promote electron transfer and fixing the immobilization of biomolecules (Matsuno et al., 2006). The combination of reduced graphene oxide (rGO) and QDs will be given advantages in the development of electrochemical sensors, especially as immobilization matrix. Moreover, hydrophilicity, biocompatibility, high specific surface, good electrical conductivity and be able to form a good film on electrode surface also provide extra benefits for sensor febrication. Although these types of method are well established to be used as a detection mechanism, however, the nanocomposite material based on reduced graphene oxide composited quantum dots has not been reported yet for the *M. Tuberculosis* detection.

Nanocrystalline cellulose (NCC) is another nanomaterial with high surface area that can be easily functionalized and is biocompatible and ecofriendly. It has been used stand alone and in combination with other nanomaterials to improve biosensor fabrication (Edward et al., 2013). Depend on preparation method, the obtained NCC currently has been studied due to its biocompatible, mechanical, functional, and biodegradable properties, which enable a wide dimension application in composite material (Lin et al., 2014). All these features offer the great potential of NCC in the electrochemical biosensor construction. Thus, combination NCC and GO will provide a new platform for the attachment of biomolecule without losing their bioactivity. Additionally, the designed biosensor based on the GO and NCC composite is expected to display excellent analytical performances for the detection of *mycobacterium tuberculosis* with high sensitivity, wide linearity range and lower detection limit.

#### 1.2 Problem Statement

Early detection of tuberculosis remains crucial for its effective control as it is one of the leading causes of death worldwide. It is essential for global public health protection to detect, analyze, and quantify *Mycobacterium tuberculosis* (MTB) by sensitive and cost effective methods.

Current clinical diagnostic for the detection of mycobacteria relies on molecular biology diagnostic via polymerase chain reaction (PCR) combine with agarose electrophoresis technique for separation and purification of nucleic acids. This approach is proven to be highly sensitive and can easily be detected by simple visual observation of the test and the control tubes even at the lowest concentration of the targeted sequence (Alli et al., 2011). However, the main limitation when agarose gel electrophoresis is employed for identifying genes (DNA) of mycobacteria tuberculosis by size separation of the DNA fragment (Brody et al., 2004). Firstly, gel electrophoresis relies on the observation on band density of different spots on the gel and observation of this band density does not yield unquantifiable results and has low specificity, therefore cannot differentiate the size of amplified DNA between target DNA and non-complimentary target DNA. Secondly, this method has some degree of error since the bands formed in the gels tend to be fuzzy and spread apart and samples are usually run multiple times to get clean results. Thirdly, the analysis procedure of agarose gel electrophoresis is time consuming, tedious step, and require hazardous element such as ethidium bromide for gel staining to visualize DNA in agarose gel. Thus, electrochemical PNA biosensor seems to be the best candidate to replace agarose gel electrophoresis part in PCR diagnostics method because PNA electrochemical biosensor could display a high specificity of amplified nucleic acid between specific and nonspecific DNA target. Moreover, the level of concentration in amplified nucleic acid can be quantified by monitoring the current signal. However, electrochemical detection needs to undergo electrode surface modification process to enhance the sensitivity of the electrodes. For this purpose, utilization of electrode modifier material for amplification purposes is necessary for the bare electrode.

To date reduced graphene oxide (rGO) incorporate with quantum dots (QDs) and nanocrystalline cellulose (NCC) as a novel nanomaterial platform for PNA immobilization for *Mycobacterium tuberculosis* detection has not yet been explored. Thus, suitability of these composite materials for PNA immobilization and hybridization is still unknown. Additionally, understanding the nature of PNA as a

hybridization probe on GO with QDs and GO with NCC probably produce another advantage that offers compatible microenvironment with DNA related *Mycobacterium tuberculosis*. Hence, the potential applications of both composite materials are beneficial in the designs for point of care device based biosensors. Additionally, the utilization of PNA instead of DNA as sensing probe in the fabrication of electrochemical biosensor can enhance selectivity and sensitivity of detection of *Mycobacterium tuberculosis* DNA that cannot be achievable with traditional oligonucleotides technique.

### **1.3** Objective of the study

The goal of this study is to develop a simple and sensitive PNA electrochemical biosensor as an alternative to the gel electrophoresis method for early detection of nucleic acid related *Mycobacterium tuberculosis*. This can be achived by incorporate reduced graphene oxide (rGO) with quantum dots (QDs) and reduced graphene oxide (rGO) with nanocrystalline cellulose (NCC) as a sensing material for enhance the sensitivity of detection. The following specific objectives are designed to achieve the aim of this research;

- 1) To prepare and characterize amine functionalized graphene oxide/CdS quantum dots and amine functionalized graphene oxide/Tempo-nanocellulose composite as a sensing material.
- 2) To fabricate PNA electrochemical biosensor based on amine functionalized graphene oxide/CdS quantum dots and amine functionalized graphene oxide/TEMPO-nanocellulose composite composite for the detection of *mycobacterium tuberculosis*.
- 3) To evaluate the analytical performance of the fabricated electrochemical PNA biosensor from amplified real sample related to *Mycobacterium tuberculosis*.

#### **1.4** Novelty and benefits of research

Presently, the application of electrochemical biosensor PNA probe as biorecognition element on the modified electrode based on reduced graphene oxide composited water soluble quantum dots (CdS) is the first being employed. Similarly, reduced graphene oxide composited nanocrystalline nanocelulose was first use with PNA probes for the detection of *Mycobacterium tuberculosis*. Additionally, both fabricated electrochemical PNA biosensor could be used as an alternative way to the gel electrophoresis in PCR detection method greatly reduce the cost of chemical and time detection.

### 1.5 Scope and limition of study

Agarose gel electrophoresis is common method for the detection of *M. tuberculosis* from clinical samples by PCR. However, this method relies on the band density (darkness) of the different spots on the gel which are analysis of sample is based on qualitative determination. Another limitation of this method are involve tedious preparation procedure, required longer time for analysis of sample (more than one hour) and the sample usually have to be run multiple times to obtain clean and accurate result. Therefore, PNA based electrochemical biosensor was suggested as an alternative method for the detection of *M. tuberculosis* which offers rapid detection time, simple pretreatment of sample, less expensive and possible for portable equipment. Moreover, the suggested method also offers quantifiable measurement, sensitivity and selectivity detection compared to agarose gel electrophoresis technique.

#### REFERENCES

- Abdullah, M. A. A and Salah, A, J. (2012). Synthesis, structure, and optical properties of CdS thin films nanoparticles prepared by chemical bath technique. *Journal of the Association of Arab Universities for Basic and Applied Sciences* 11(1): 27–31
- Abibi A, Protozanova E., Demidov V,V, Frank-Kamenetskii, M.D. (2004). Specific versus nonspecific binding of cationic PNAs to duplex DNA. *Biopysical Journal*. 86(5):3070-8.
- Abitol T, Cranston E. (2014) Chiral nematic self-assembly of cellulose nanocrystals in Suspension and solid films. In: In: Oksman K, Bismarck A, Rojas OJ, Sain M, editors. Handbook of green materials, Part 3: self- and direct-assembling of bionanomaterials, Chapter 4, Singapore:World Scientific Publishing. 37–56
- Agui, L., Yanez-Sedeno, P. and Pingarron, J.M. (2008). Role of carbon nanotubes in electroanalytical chemistry. A review, *Analytica Chimica Acta* 622:11–47.
- Aghdam, K., Panahi, H.A., Alaei, E., Hasani, A.E., Moniri, E. (2016). Preparation of functionalized graphene oxide and its application as a nanoadsorbent for Hg<sup>2+</sup> removal from aqueous solution. *Environmental Monitoring and Assessment* 188:223
- Akhtar, Hayat and Jean Louis Marty, (2014). Disposable Screen Printed Electrochemical Sensors: Tools for Environmental Monitoring. Sensors (Basel) 14(6): 10432–10453.
- Ali, M., Neumann, R and Ensinger, W. (2010). Sequence-specific recognition of DNA oligomer using peptide nucleic acid (PNA)-modified synthetic ion channels: PNA/DNA hybridization in nanoconfined environment ACS Nano 4(12): 7267-74
- Alli, O.A.T., Ogbolu, O.D.and Alaka, O.O. (2011). Direct mo lecular detection of Mycobacterium tuberculosis complex from clinical samples an adjunct to cultural method of laboratory diagnosis of tuberculosis. *North American Journal of Medical Sciences* 3(6): 281–288.
- Anson, F.C. (1976). Electroanalytical Chemistry, ACS Audio Course Manual, American Chemical Society 55
- Artiles, M., Rout. C and Fisher, T. (2011). Graphene based hybrid materials and devices for bio sensing. *Advanced Drug Delivery Reviews* 63: 1352–1360.
- Bard, A.J. and Faulkner, L. R. (2000). Electrochemical Methods, Fundamentals and Applications, *John Wiley & Sons, New York* 2: 115-130
- Barglik-Chory C., Buchold, D., Schmitt, M., Kiefer, W., Heske, C., Kumpf. C, Fuchs O, Weinhardt L, Stahl, A, Umbach, E, Lentze, M, Geurts, J, and Mqller, G

(2003). Synthesis, structure and spectroscopic characterization of watersoluble CdS nanoparticles. *Chemical Physics Letters* 379: 443–451

- Barry, C.E., Boshoff, H. I., Dartois, V., Dick T., Ehrt, S and Flynn, J. (2009). The spectrum of latent tuberculosis: rethinking the biology and intervention strategies. *Nature reviews Microbiology* 7: 845–55.
- Basirun, W. J., Sookhakian, M., Baradaran, S., Mahmoudian, M.R., and Ebadi, M. (2013). Solid-phase electrochemical reduction of graphene oxide films in alkalinesolution, *Nanoscale Research Letters* 8: 397-402
- Bianco, A., Kostarelos, K., and Prato, M. (2008). Opportunities and challenges of carbon-based nanomaterials for cancer therapy, *Expert Opinion on Drug Deliver* 5: 331–342.
- Biswas, S., and Drzal, L.T. (2009). A Novel Approach to Create a Highly Ordered Monolayer Film of Graphene Nanosheets at the Liquid–Liquid Interface. Nano Letters 9: 167
- Bocking, T., Kilian, K. A., Reece, P.J., Gaus, K., Galb, M and Gooding, J.J. (2012). Biofunctionalization of free-standing porous silicon films for self-assembly of photonic devices. *Soft Matter*, 8: 360-366
- Boon, E. M, Jackson, N.M., Wightman ,M.D., Kelley, S.O., Hill, M.G and Barton, J.K.(2003). Intercalative Stacking: A Critical Feature of DNA Charge-Transport Electrochemistry, *The Journal of Physical Chemistry*. B 107 (42): 11805–11812
- Bora, U.,Singh, A and Deepika. (2013). Nucleic Acid Based Biosensors for Clinical Applications, *Biosensors Journal* 2: 104

Bolognia J, Jorizzo J. and Rapini, R. (2008). Dermatology, Elsevier 2, 3<sup>rd</sup> edition. Bott, A.W and Howell, J.O. (1992). Chronocoulometry. *Current. Separations*. 11(21).

- Brandt, O and Hoheisel, J.D. (2004) Peptide nucleic acids on microarrays and other biosensors. *Trends in Biotechnology*, 22 (12).
- Brisson, N.A., Aznar, C., Chureau C, Nguyen S, Pierre C, Bartoli M, Bonete R, Pialoux G, Gicquel B, and Garrigue G. (1999). Diagnosis of tuberculosis by DNA amplification in clinical practice evaluation. *The Lancet* 338:364-366.
- Brody, J. R. and Kern, S. E. (2004). History and principles of conductive media for standard DNA electrophoresis. *Analytical biochemistry* 333 (1): 1-13
- Butun, S., Ince, F.G., Erdugan, H. and Sahiner, N. (2011). One-step fabrication of biocompatible carboxymethyl cellulose polymeric particles for drug delivery systems. *Carbohydrate Polymers* 86: 636–643.
- Cao Y.C. and Wang, J. (2004). One-pot synthesis of high-quality zinc- blende Cd Snanocrystals. *Journal of the Chemical Society* 126: 14336–14337

- Carter, M,T., Rodriguez, M and Bard, A, J. (1989). Voltammetric Studies of the Interaction of Metal Chelates with DNA. *Journal of the American Chemical Society:* 8901-8911
- Cattamanchi, A., Smith, R., Steingart, K. R., Metcalfe, J.Z., Date, A., Coleman, C., Marston, B.J, Huang, L., Hopewell, P.C., and Pai, M. (2011). Interferongamma release assays for the diagnosis of latent tuberculosis infection in HIVinfected individuals: a systematic review and meta-analysis. *Journal of acquired immune deficiency syndromes* 56(3): 230-8
- Chaubey, A and Malhotra, B. D. (2002). Mediated biosensors. *Biosensors & Bioelectronics* 17(6): 441–456.
- Chan, E.D., Heifets L. and Iseman M.D (2000). Immunologic diagnosis of tuberculosis: a review. Tuber Lung, 80:131-140 Complexes of Cobalt (111) and Iron(11) with 1,IO-Phenanthroline and 2,2'-Bipyridine. *Journal of the American Chemical Society* 111
- Chavolla, E.T and Alocilja, E.C. (2011). Nanoparticle based DNA biosensor for tuberculosis detection using thermophilic helicase-dependent isothermal amplification. *Biosensors and Bioelectronics* 26: 4614–4618
- Chen, D., Feng, H. and Li, J. (2012) Graphene Oxide: Preparation, Functionalization, and Electrochemical Applications, *Chemical Reviews*. 112: 6027–6053
- Chen, J., Zhang, R., Wang J., Liu L, Zheng, Y and Shen, Y. (2011). Interferon-gamma release assays for the diagnosis of active tuberculosis in HIV-infected patients: a systematic re-view and meta-analysis, *PLoS One* 6:e26827.
- Chen, K., Lu, G., Chang, J., Mao, S., Yu, K., Cui, S. and Chen, J. (2012). Analytical Chemistry 84: 4057–4062
- Chen, L., Tang, Y., Wang K, Liu, C. and Luo. (2011). Direct electrodeposition of reduced graphene oxide on glassy carbon electrode and its electrochemical application *Electrochemistry Communications*. 13: 133–137.
- Chen, S., Ding, R., Ma, X., Xue, X., Lin, X., Fan, X and Zhimin Luo . (2012). Preparation of highly dispersed reduced graphene Oxide modified with carboxymethyl chitosan for highly Sensitive Detection of Trace Cu(II) in Water. *polymer* 8(4): 78-81
- Chen, Y., and Rosenzweig, Z. (2002). Luminescent CdS Quantum Dots as Selective Ion Probes. *Analytical Chemistry* 74 (19): 5132–5138
- Chiu, S.K., Hsu, M., Ku, W.C., Tu, C.Y., Tseng, Y.T., Lau, W.K., Yan, R.Y., MA, J.T and Tzeng. C.M. (2003). Synergistic effects of epoxy and amine silanes on microarray DNA immobilization and hybridization. *Biochemistry* 374: 625– 632
- Christensen, L., Fitzpatrick, R., Gildea, B., Petersen, K. H., Hansen, H. F., Koch, T., Egholm, M., Buchardt, O., Nielsen, P. E., Coull, J., and Berg, R. H. (1995)

Solid-phase synthesis of peptide nucleic acids. *Journal of Peptide Science* 3: 175–183

- Chua, C. K. and Pumera, M. (2014). Chemical reduction of graphene oxide: a synthetic chemistry viewpoint. *Chemical Society Reviews* 43: 291-312
- Cirtiu, C.M, Dunlop-Briere, A.F, and Moores, A. (2011). Cellulose nanocrystallites as an efficient support for nanoparticles of palladium: application for catalytic hydrogenation and Heck coupling under mild conditions. *Green Chemistry* 13:288–91
- Cole S.verma, T., Brosch R., Parkhill J., Garnier T., Churcher, C., Harris, D., Gordon, S.V, Eiglmeier K, Gas S, Barry CE 3rd, Tekaia F, Badcock K, Basham D, Brown D, Chillingworth T, Connor R, Davies R, Devlin K, Feltwell T, Gentles S, Hamlin N, Holroyd S, Hornsby T, Jagels K, Krogh A, McLean J, Moule S, Murphy L, Oliver K, Osborne J, Quail MA, Rajandream MA, Rogers J, Rutter S, Seeger K, Skelton J, Squares, R., Squares, S., Sulston, J.E, Taylor K, Whitehead S, Barrell. (1998), BG Deciphering the biology of Mycobacterium tuberculosis from the complete genome sequence. *Nature* 393(6685): 537-44
- Colebunders, R. and Bastian, I. (2000). A review of the diagnosis and treatment of smear-negative pulmonary tuberculosis. *International Journal of Tuberculosis and Lung Disease*. *Lung* 4: 97.
- Compton, O.C. and Nguyen, S.T. (2010). Graphene Oxide, Highly Reduced Graphene Oxide and Graphene :Versatile Building Blocks for Carbon-Based Materials. Small 6: 711–723.
- Corey, D. R. (1997). Peptide nucleic acids: expanding the scope of nucleic acid recognition. *Trends in Biotechnology* 15: 224–229
- Dahoumane, S. A., Nguyen M. N., Thorel A., Boudou J-P., Chehimi M. M., and Mangeney, C. (2009). Protein-functionalized hairy diamond nanoparticles. *Langmuir* 25: 9633–9638.
- Dai, Z., Chen, J., Yan, F., Ju, H. (2005). Electrochemical sensor for immunoassay of carcinoembryonic antigen based on thionine monolayer modified gold electrode. *Cancer Detection and Prevention* 29: 233–240
- Daniele, S., Ghazzai, M.N., Hubert-Pfalzgrai, L.G., Duchamp, C., Guillard, C., and Ledoux, G. (2006). Preparation of nano particles nanocomposites and fibers of ZnO from an amide precursor: photocatalytic decomposition of (CH3)2Sn in a continuous flow reactor, *Materials Research Bulletin* 41: 2210–2218
- Das, M., Sumana, G., Nagarajan, R. and Malhotra, B.D. (2010). Zirconia based nucleic acid sensor for Mycobacterium tuberculosis detection *Applied Physics Letters* 96: 133703
- Dean, D.A. (2000). Peptide nucleic acids: versatile tools for gene therapy strategies *Advanced Drug Delivery Reviews* 15: 81–95.

- Degefa, T. H. and Kwak, J. (2008). Electrochemical impedance sensing of DNA at PNA self-assembled monolayer. *Journal of Electroanalytical Chemistry* 612: 37–41
- Deng, M., Tu, N., Bai, F., and Wang, F. (2012). Surface Functionalization of Hydrophobic Nanocrystals with One Particle per Micelle for Bioapplications *Chemistry of Materials* 24 (13): 2592–2597
- Diaz-Gonzalez, M., Gonzalez-Garcia, M.B., and Costa-Garcia, A. (2005). Immunosensor for Mycobacterium tuberculosis on screen-printed carbon electrodes. *Biosensors and Bioelectronics* 20: 2035–2043
- Ding, L., Zhou, Li, S. Q, Shi, G.Y, Zhong, T., and Wu, M. (2011). Spectroscopic Studies on the Thermodynamics of L-Cysteine Capped CdSe/CdS Quantum Dots-BSA Interactions. *Journal of Fluorescence* 21, 17–24.
- Ding, Yu, S., Wei, J. and Xiong, H. (2016). Full-Color Light-Emitting Carbon Dots with a Surface-State-Controlled Luminescence. *Mechanism ACS Nano* 10 (1): 484–491
- Dominguez, R. O., Alonso, L. M. A., and Arcos, M. M. J. (2007). Recent developments in the field of screen-printed electrodes and their related applications, *Talanta*, 73: 202-209
- Dominguez, R ,O., Ruiz-Espelt, L., García-Astorgano, N., and Arcos-Martínez, M.J. (2008). Electrochemical determination of chromium(VI) using metallic nanoparticle-modified carbon screen-printed electrodes. *Talanta* 76(4):854–858.
- Dreyer, D. R., Ruoff, R. S. & Bielawski, C. W.(2010). From conception to realization: an historical account of graphene and some perspectives for its future. Angew. *Angewandte Chemie International Edition* 49: 9336–9344
- Drummond, T,G., Hill, M, G and Barton, J, K. (2003). Electrochemical DNA sensors. *Nature biotechnology* 21:1192-1199.
- Drummond, T. G. Hill, M. G, and Barton, J. K. (2004). Electron Transfer Rates in DNA Films as a Function of Tether Length. *Journal of the American Chemical Society* 126: 15010–15011
- D'Souza, S.F. (2001).immobilization and stabilization of biomaterials for biosensor applications. *Applied Biochemistry and Biotechnology*. 96: 225-38
- Du, D., Guo, S., Tang, L., Ning, Y., Yao, Q and Zhang. G. J. (2013). Graphene modified electrode for DNA detection via PNA–DNA hybridization. *Sensors* and Actuators B 186:563–570
- Dubale, A.A., Su, W.N., Tamirat, A.G., Pan, C.J., Aragaw, B.A., Chen, H.M., Chen, C.H and Hwang, B.J. (2014). The synergetic effect of graphene on Cu2O nanowire arrays as a highly efficient hydrogen evolution photocathode in water splitting. *Journal of Materials Chemistry A* 2:18383-18397

- Ducker R. E., Montague M. T., and Leggett G. J. (2008). A comparative investigation of methods for protein immobilization on self-assembled monolayers using glutar aldehyde, carbodiimide, and anhydride reagents. *Bio interphases* 3:59–65.
- Dufresne, A., Cavaille, J.Y, and Vignon, M.R (1997). Mechanical behavior of sheets prepared from sugar beet cellulose microfibrils. *Journal of Applied Polymer Science* 64: 1185–94.
- Dutta, D., Chandra, S., Swain, A.K and Bahadur, D. (2014). SnO<sub>2</sub> Quantum Dots-Reduced Graphene Oxide Composite for Enzyme-Free Ultrasensitive Electrochemical Detection of Urea. *Analytical Chemistry*. 86: 5914–5921
- Edwards, J.V., Prevost N, French A, ConchaM, Delucca A, and Wu Q. (2013). Nanocellulose-based biosensors: design, preparation, and activity of peptidelinked cotton cellulose nanocrystals having fluorimetric and colorimetric elastase detection sensitivity. *Engineering* 5: 20–8
- Efferen, L. S. (1999). Tuberculosis: Practical Solutions to Meet the Challenge. *Journal* of Respiratory Diseases 20: 772-785.
- Egholm, M., Buchardt, O., Christensen, L., Behrens, C., Freier, S.M., Driver, D.A., Berg, R.H., Kim, S.K., Norden, B. and Nielsen, P.E. (1993). PNA hybridizes to complementary oligonucleotides obeying the Watson-Crick hydrogen-bonding rules. *Nature* 365: 566–568.
- Eichhorn, S.J. Dufresne, A., Aranguren, M., Marcovich, N.E., Capadona, J.R., Rowan,
  S.J., Weder, C., Thielemans, W., Roman, M., Renneckar, S., Gindl, W.,
  Veigel, S., Keckes, J., Yano, H., Abe, K. (2010). Review: current international research into cellulose nanofibres and nanocomposites. *Journal of Materials* Science 45(1): 1-33.
- Ellina, N., Izaanin,N., Abdullah, J., Arif,N. and Azah, N. (2015) A simple and sensitivefluorescence based biosensor for the determination of uric acid using H<sub>2</sub>O<sub>2</sub> sensitivequantum dots/dual enzymes, *Biosensor and Bioelectronic* 67: 129–133.
- Erdem, A., Kerman, K., Meric, B., Ozkan, D, Kara, P. and Ozsos, M. (2002). Detection by Using Methylene Blue and Ruthenium Complex as Electrochemical Hybridization Labels. *Turkish Journal of Chemistry*. 26: 851 – 862.
- Erdem, A., Kerman,K., Meric,B., Akarca, U, S and Ozsoz, M. (2000). Novel hybridization indicator methylene blue for the electrochemical detection of short DNA sequences related to the hepatitis B virus *Analytical Chemistry*. *Acta* 422: 139-149
- Erhan, D.E., and E, Arkan. (2017). Investigation of dilution agent effect onto interactions between methylene blue and DNA using carbon fiber based DNA biosensor. *Journal of New Materials for Electrochemical Systems*, 20(2): 65-70.

- Esmaeili, C., Abdi, M, M., Mathew, A, P, Jonoobi, M, Oksman, K and Rezayi, M. (2015). Synergy Effect of Nanocrystalline Cellulose for the Biosensing Detection of Glucose Sensors 15: 24681-24697
- Eyley, S and Thielemans. W. (2014). Surface modification of cellulose nanocrystals. *Nanoscale* 6: 7764-7779
- Fang. B, Jiao, S., Li, M., Qu, Y. and Jiang, X. (2008). Label- free electrochemical detection of DNA using ferrocene-containing cationic polythiophene and PNA probes on nanogold modified electrodes. *Biosensors and Bioelectronics* 28: 1175–1179.
- Farjami, E., Clima, L, Gothelfa, K.V and Ferapontova, E, E. (2010). DNA interactions with aMethylene Blue redox indicator depend on the DNA length and are sequence specific. *Analyst* 135:1443–1448.
- Feng, R., Hu, X., He, C., Li, X. and Luo, X. (2017). Electrochemical Determination of the p53 Tumor Suppressor Gene Using a Gold Nanoparticle-Graphene Nanocomposite Modified Glassy Carbon Electrode. Journal. Analytical Letters 50 (2): 336-349
- Fojta, M., Havran, L., Cahova, K., Kostecka, P., Jelen, F., Palecek, E. (2008). Electroactive DNA Markers and Probes Based on Osmium Tetroxide. *Analytical Chemistry*. 80 (12): 4598–4605.
- Foulds, J. and O' Brien, R. (1998). New tools for the diagnosis of tuberculosis: the perspective of developing countries. *International Journal of Tuberculosis and Lung Disease* 2:778-788
- Fujisawa, S., Saito, T., ura., S., Iwata T and Isogai, A. (2014).Comparison of mechanical rein- forcement effects of surface-modified cellulose nanofibrils and carbon nanotubes in PLLA composites. *Composites Science and Technology* 90: 96–101
- Fukuzumi, H., Saito, T., Iwamoto, S., Kumamoto, Y., Ohdaira, T., Suzuki, and Isogai, A. (2011). Pore Size Determination of TEMPO-Oxidized Cellulose Nanofibril Films by Positron Annihilation Lifetime Spectroscopy *Biomacromolecules*, 12 (11): 4057–4062.
- Fukuzumi, H., Saito T, Iwata, T., Kumamoto, Y and Isogai, A. (2009). Transparent and High Gas Barrier Films of Cellulose Nanofibers Prepared by TEMPO-Mediated Oxidation. *Biomacromolecules* 10(1), 162–165.
- Gamboa, F., Manterola, J.M., Lonca, J. (1997). Detection and identification of mycobacteria by amplification of RNA and DNA in pretreated blood and bone marrow aspirates by a simple lysismethod. *Journal of ClinicalMicrobiology* 35(8): 2124–2128
- Gao, H., Sun, M., Lin, C., and Wang, S. (2012). Electrochemical DNA Biosensor Based on Graphene and TiO2 Nanorods Composite Film for the Detection of

Transgenic Soybean Gene Sequence of MON89788. *Electroanalysis* 12: 2283–2290

- Geim, A.K., and Novoselov, K. S. (2007). The rise of graphene. *Nature Materials*. 6:183–191.
- George, J and Sabapathi, S. N. (2015). Cellulose nanocrystals: synthesis, functional properties, and applications. *Nanotechnology, Science and Applications* 8: 45– 54
- Gibbs, R. A. (1990). DNA Amplification by the Polymerase Chain Reaction. Analytical Chemistry 62 (13):1202-1214.
- Goda, T., Singi, A.B., Maeda, Y., Matsumoto, A., Torimura, M., Aoki, H. and Miyahara, Y. (2013). Label-Free Potentiometry for Detecting DNA Hybridization Using Peptide Nucleic Acid and DNA Probes. Sensors 13: 2267-2278.
- Gooding, J, J. (2002). Electrochemical DNA Hybridization Biosensors, *Electroanalysis* 14: 1149–1156.
- Gu, J.Y., Lu, X.J and Ju, H. X. (2002). DNA sensor for recognition of native yeast DNA sequence with methylene blue as an electrochemical hybridization indicator. *Electroanalysis* 14: 949-954
- Guidelli, R., Compton, R.G., Feliu, J. M., Gileadi, E., Lipkowski, J., Schmickler, W. and Trasatti, S. (2014). Defining the transfer coefficient in electrochemistry: An assessment (IUPAC Technical Report. *Pure and Applied Chemistry*, 86(2): 245–258
- Gupta, V., Chaudhary, N., Srivastava, R., Sharma, G.D., Bhardwaj, R. and Chand, S. (2011). Luminscent Graphene Quantum Dots for Organic Photovoltaic Devices. *Journal of the American Chemical Society* 133 (26), 9960–9963
- Habibi, Y., Lucia, L. A, Rojas, O. J. (2010). Cellulose nanocrystals: chemistry, selfassembly and applications. *Chemical Reviews* 110:3479–500.
- Hagan, M. F and Chakraborty, A.K. (2004). Hybridization dynamics of surface immobilized DNA. *The Journal of Chemical Physics* 120: 4958–4968.
- Hajime, F., Tomoyuki, M., Toshikazu,G.,Kazuo, S and Tokuhiro, I. (2002). Sensitivity of Acid- Fast Staining for Mycobacterium tuberculosis in Formalin-fixed Tissue, American Journal of Respiratory and Critical Care Medicine, 166(7): 994-997.
- Hakimi, M., and Alimard, P. (2012). Graphene: Synthesis and Applications in Biotechnology : A Review. *World Applied Programming* 2(6): 377-388
- Halperin, A., Buhot, A and Zhulina, E. B (2006). On the hybridization isotherms of DNA microarrays: the Langmuir model and its extensions. *Journal of Physics: Condensed Matter:* 18:S463–S490

- Hamidi-Asl, E., Raoof, J.B., Ojani, R. and Hejazib, M.S. (2013). Indigo Carmine as New Label in PNA Biosensor for Detection of Short Sequence of p53 Tumor Suppressor Gene. *Electroanalysis* 25(9): 2075 – 2083
- Han X ., Fang X ., Shi A ., Wang J ., and Zhang Y . (2013). An electrochemical DNA biosensor based on gold nanorods decorated graphene oxide sheets for sensing platform. *Analytical biochemistry* 2: 117-23
- Hansen, J.A. Wang, J., Kawde, A.N., Xiang, Y., Gothelf, K.V. and Collins, G. (2006). Quantum-dot/aptamer-based ultrasensitive multi analyte electrochemical biosensor. *Journal of the American Chemical Society*, 128(7): 2228–2229
- Harries, A.D., Maher, D. and Nunn, P. (1997). Practical and affordable measures for the protection of health care workers from tuberculosisin low-income countries. *Bull World Health Organ* 75: 477–89
- He, F., Zhao, J., Zhang, L., and Su, X. (2003). A rapid method for determining Mycobacterium tuberculosis based on a bulk acoustic wave impedance biosensor, *Talanta* 59(5): 935–941.
- He, X. Y., Li, J., Hao, J., Chen, H.B., Zhao, Y.Z., Huang, X.Y., He,K., Xiao, L., Ye, P., Qu,Y. M and Ge, L. H. (2011). Assessment of Five Antigens from Mycobacterium tuberculosis for Serodiagnosis of TuberculosisClinical and Vaccine Immunology. *Hindawi* 18(4): 565-570
- Hernandez, Y., Nicolosi, V., Lotya, M., Blighe, F.M., Sun, Z., De, S., McGovern, I.T., Holland, B., Byrne, M., Gun Ko, Y. K., Boland, J.J., Niraj, P., Duesberg, G., Krishnamurthy, S.,Goodhue, R., Hutchison, J.,Scardaci, V., Ferrari, A.C. and Coleman, J. N (2008). High-yield production of graphene by liquid-phase exfoliation of graphite. *Nature Nanotechnology* 3: 563–568.
- Hianik, T., Ostatna, V., Sonlajtnerova, M. and Grman. I. (2007). Influence of ionic strength, pH and aptamer conFigureuration for binding affinity to thrombin *Bioelectrochemistry* 70:127–133
- Holzinger, M., Le, Goff., A, Cosnier S. Front. (2014). Nanomaterials for biosensing applications: a review. *Frontier in Chemistry*. 2:63.
- Hook, F., Ray, A., Norden, B and Kasemo, B. (2001). Characterization of PNA and DNA Immobilization and Subsequent Hybridization with DNA Using Acoustic-Shear-Wave Attenuation Measurements, *Langmuir* 17: 8305–831.
- Howley, P.M, Israel, M.A, Law, M.F, Martin MA. (1979). A rapid method for detecting and mapping homology between heterologous DNAs. Evaluation of polyomavirus genomes. *Journal Biology Chemistry* 254(11):4876-83.
- Hsu, S.H., Lin , Y.Y., Lu , S.H., Tsai, I.F., Lu, Y.T., and Ho, H.T.(2014). Mycobacterium tuberculosis DNA Detection Using Surface Plasmon Resonance Modulated by Telecommunication Wavelength. *Sensors*, 14:458-467.

- Hu, R., Wu, Z.S., Zhang, S., Shen, G.L and Yu, R (2011). Robust electrochemical system for screening single nucleotide polymorphisms. *Chemical Communications* 47:1294-1296.
- Hutton, J.R. 1977. Renaturation kinetics and thermal stability of DNA in aqueous solutions of formamide and urea. *Nucleic Acids Research* 4, 3537–3555.
- Huang, J.G., Hung, C.C., Lai, H.C, Lee, C.K., Lin, S.M, Feng, P, and Lin C.W.(2004). Surface plasmon resonance biochips for tuberculosis bacillus detection. *Engineering in Medicine and Biology Society*. 4: 2553-6.
- Ignas T. K. and Emmanuel, N. (2011). Voltammetric Analysis of Pesticides, Pesticides in the Modern World Trends in Pesticides *Analysis*. *Intech* 19
- Incani, V., Danumah, C, and Boluk, Y. (2013). Nanocomposites of nanocrystalline cellulose for enzyme immobilization. *Cellulose* 20(1): 191–200.
- Isogai, A., T. Saito, and H. Fukuzumi. (2011). TEMPO-oxidized cellulose nanofibers. *Nanoscale*, 3(1): 71-85.
- Issa, R., Hamdam. N.A and Noh, M.F.M. (2010). Differential pulse voltametric determinition of DNA hybridizationusing methylene blue on screen printed carbon electrode for the detection of mycobacterium tuberculosis, *Biotechnology* 9: 304–311.
- Iyawoo, K. (2004). Tuberculosis in Malaysia: problems and prospect of treatment and control. *Tuberculosis*. 84: 4–7.
- Jang, H.D., Kim, S.K., Chang, H. (2012). A glucose biosensor based on TiO2-Graphene composite. *Biosensors & Bioelectronics* 38(1):184-188.
- Jennifer, A., Harnisch, Andrew, D., Pris, and Marc. (2001). D. Porter Attachment of Gold Nanoparticles to Glassy Carbon Electrodes via a Mercaptobenzene Film. *Journal of the American Chemical Society* 123(24):5829–5830.
- Jensen, K.K., Orum, H., Nielsen, P.E. and Norden, B. (1997). Kinetics for hybridization of peptide nucleic acids (PNA) with DNA and RNA studied with the biacore technique. *Biochemistry* 36:5072–5077.
- Jiang, Y. and Gunasekaran, S. (2013). Electrochemically reduced graphene oxide sheets for use in high performance supercapacitors. *Carbon* 51: 36-44.
- Jiao, K., Li, Q.J., Sun, W., and Wang. Z. J. (2005). Voltammetric Detection of the DNA Interaction with Toluidine Blue. *Electroanalysis* 17(11): 997–1002
- Jin, S., Hu, Y., Gu, Z., Liu, H, Wu, C. (2011). Application of Quantum Dots in Biological Imaging. *Journal of Nanomaterial* 13.
- Joshi, T., Patra, M., Spiccia, L and Gasser, G. (2013). Di-heterometalation of thiolfunctionalized peptide nucleic acids. *Artificial DNA: PNA XNA*, 4(1): 11–18.

- Joung, D and Khondaker, S.I. (2013). Structural Evolution of Reduced Graphene Oxide of Varying Carbon sp<sup>2</sup> Fractions Investigated via Coulomb Blockade Transport. *The Journal of Physical Chemistry C* 117 (50): 26776–26782
- Joucken, F., Tison, Y., Fèvre, P. L., Tejeda, A., Taleb-Ibrahimi, A., Conrad, E., Repain, V., Chacon, C., Bellec, A., Girard, Y., Rousset, S., Ghijsen, J., Sporken, R., Amara, H., Ducastelle, F and Lagoute, J. (2015). Charge transfer and electronic doping in nitrogen-doped graphene *Scientific Reports 5*
- Ju, H., Zhou, J., Cai, C and C, H. (1994). The electrochemical behaviour of methylene blue at a microcylinder carbon fiber electrode. *Electroanalysis* 7(12).
- Jung, D.H., Kim, B.H. Ko, Y.K.; Jung, M.S., Jung, S.K., Lee, S.Y., and Jung, H.T. (2004). Covalent attachment and hybridization of DNA oligonucleotides on patterned single-walled carbon nanotube films. *Langmuir* 20: 8886–8891.
- Kaewphinit, T., Santiwatanakul, S., Promptmas, C., and Chansiri, K. (2010). Detection of non-amplified Mycobacterium tuberculosis genomic DNA using piezoelectric DNA-based biosensors. *Sensors* 10: 1846–1858.
- Kalle.E., Kubista, M and Rensing, C. (2014). Multi-template polymerase chain reaction. *Biomolecular Detection and Quantification*, 2: 11-29.
- Kang, S.H., Mhin, S., Han, H., Kim, K.M., Jones, J.L., Ryu, J.H., Kang, S.S., Kim, S.H. and Shim, K.B. (2016). Ultrafast Method for Selective Design of Graphene Quantum Dots with Highly Efficient Blue Emission. *Scientific Reports*. 6: 38423.
- Kara, P., Meric, B., and Ozsoz, M. (2008). Application of Impedimetric and Voltammetric Genosensor for Detection of a Biological Warfare: Anthrax *Electroanalysis* 20: 2629–2634.
- Karuwan, C, Wisitsoraat, A., Phokharatkul, D., Sriprachuabwong, C., Lomas, T., Nacapricha, D and Tuantranont, A. (2013). A disposable screen printed graphene carbon paste electrode and its application in electrochemical sensing. *RSC Advances* 3:25792-25799.
- Keighley, S.D., Li, P., Estrela, P. and Migliorato, P. (2008). Optimization of DNA immobilization on gold electrodes for label-free detection by electrochemical impedance spectroscopy. *Biosensors and Bioelectronics* 23:1291–1297.
- Kelley, S.O and Barton, J.K. (1997) Electrochemistry of Methylene Blue Bound to a DNA-Modified Electrode Bioconjugate. *Chemistry* 8 (1): 31–37.
- Kemell, M., Pore, V., Ritala, M., Leskela, M and Linden, M. (2005). Atomic layer deposition in nanometer-layer replication of cellulosic substances and preparation of photocatalytic TiO2/cellulose composites. *Journal of the American Chemical Society* 127, 14178–14179.
- Khalil, I., Julkapli, N.M., Yehye, W.A., Basirun, W. J and Bhargava. S. K (2016). Graphene–Gold Nanoparticles Hybrid—Synthesis, Functionalization, and Application in a Electrochemical and Surface-Enhanced Raman Scattering Biosensor. *Materials*. 9:406

- Khomenko. V., Frackowiak, E., and Beguin, F. (2005). Determination of the specific capacitance of conducting polymer/nanotubes composite electrodes using different cell conFigureurations. *Electrochimica Acta*. 50(12), 2499-2506.
- Kim, J. R., Santiano, B., Kim, H., and Kan, E. (2013). Heterogeneous Oxidation of Methylene Blue with Surface-Modified Iron-Amended Activated Carbon. *American Journal of Analytical Chemistry*, 4, 115-122.
- Kim, N.H., Kuila, T.,and Lee, J. H. (2013). Simultaneous reduction, functionalization and stitching of graphene oxide with ethylenediamine for composites application, *Journal of Materials Chemistry A*. 1:1349–1358.
- Kim, S. K., Cho, H., Jeong, J., Kwon, J.N., Jung.W and Chung, B.H. (2010). Labelfree and naked eye detection of PNA/DNA hybridization using enhancement of gold nanoparticles. *Chemical Communications*. 46: 3315-3317.
- Kim, N.H.,Kuila, T., and Lee, J.H., (2013). Simultaneous reduction, functionalization and stitching of graphene oxide with ethylenediamine for composites application, *Journal of Materials Chemistry A* 1:1349.
- Klemm D, Kramer F, Moritz S, Lindström T, Ankerfors M, Gray D, (2011). Nanocelluloses: a new family of nature based materials. *Angewandte Chemie International Edition* 50: 5438–66.
- Knemeyer, J.P., Marme, N and Sauer., M. (2000). Probes for Detection of Specific DNA Sequences at the Single-Molecule Level. *Analytical Chemistry*. 72 (16): 3717–3724.
- Komeda, S., Moulaei, T., Woods, K. K., Chikuma, M., Farrell, N. P. and Williams, L. D. J. and Am, A. (2006). Third Mode of DNA Binding: Phosphate Clamps by a Polynuclear Platinum Am. *Journal of the American Chemical Society*. 128(50): 16092–16103.
- Koneswaran, M., and Narayanaswamy, R. (2009) Mercaptoacetic acid capped CdS quantum dots as fluorescence single shot probe for mercury(II). *Sensors and Actuators B: Chemical*, 139(1): 91–96.
- Konios, D., Stylianakis, M.M., Stratakis, E and Kymakis. E. (2014). Dispersion behaviour of graphene oxide and reduced graphene oxide. *Journal of Colloid and Interface Science*, 430: 108–112.
- Konopka, S, J. and Mcduffie, B. (1970) Diffusion Coefficients of Ferri and Ferrocyanide Ionsin Aqueous Media, Using Twin-Electrode Thin-layer Electrochemistry, *Analytical Chemistry*, 42: 1741–1746.
- Krambovitis, E., McIllmurray, M.B., Lock, P.E., Hendrickse, W., Holzel, H., (1984). Rapid diagnosis of tuberculosis meningitis by latex particle agglutination. *Lancet*, 2, 1229–1231.
- Kubista, M., Andrade, J.M., Bengtsson, M., Forootan, A., Jonak. J., Lind K., Sindelka R., Sjoback R., Sjogreen, B, Stromborn, L., Stahlberg, A and Zoric,

N. (2006). The real-time polymerase chain reaction. *Molecular Aspects of Medicine*. 27(2-3), 95-125.

- Kudin, K.N., Ozbas, B., Schniepp, H.C., Prudhomme, R.K., Aksay, I.A and Car, R. (2008). Raman Spectra of Graphite Oxide and Functionalized Graphene Sheets. *Nano Letters.*, 8,36–41.
- Kuila T, Bose S, Mishra A, Khanra P, Kim N. and Lee J. (2012). Chemical functionalization of graphene and its applications. *Progress in Materials Science*, 57:1061–1105.
- Kuilla, T., Bhadra,S., Yao,D., Kim, N.H., Bose, S., and Hee, J. (2010). Recent advances in graphene based polymer composites. *Progress in Polymer Science*, 12:1350–1375.
- Kumar, P., Kukkar, D., Deep, A., Sharma, S.C. and Bharadwaj, L.M. (2012). Synthesis of mercaptopropionic acid stabilized CDS quantum dots for bioimaging in breast cancer. *Advanced Materials Letters* 3(6): 471-475.
- Kumar, S., Ahlawat, W., Kumar, R and Dilbaghi, N. (2015). Graphene, carbon nanotubes, zinc oxide and gold as elite nanomaterials for fabrication of biosensors for healthcare. *Biosensors and Bioelectronics*. 70: 498-503.
- Kundu, S., Ghosh, S., Fralaide, M., Narayanan, T.N., Pillai, V.K and Talapatra, S. (2015). Fractional photo-current dependence of graphene quantum dots prepared from carbon nanotubes. *Physical Chemistry Chemical Physics* 17: 24566-24569.
- Lasseuguette E., Roux, D and Nishiyama, Y.(2008). Rheological properties of microfibrillar suspension of TEMPO-oxidized pulp. *Cellulose*, 15:425–33.
- Laschi, S., Bulukin, E., Palchetti, I., Cristea, C and M. Mascini. (2008). Disposable electrodes modified with multi-wall carbon nanotubes for biosensor applications. *IRBM* 29: 202–207.
- Lawn. S.D., Kerkhoff, A.D., Vogt, M., and Wood, R. (2012). Diagnostic accuracy of a low-cost, urine anti- gen, point-of-care screening assay for HIV-associated pulmonary tuberculosis be- fore antiretroviral therapy: a descriptive study. *Lancet Infection* 12:201–9.
- Lazcka., Campo, F. J. D. and Munoz, F. X. (2007) Pathogen detection: a perspective of traditional methods and biosensors, *Biosensors and Bioelectronics* 22, (7):1205–1217.
- Lee and Kwon, O. S., Park. C. S. (2016). Conducting Polymer Based Nanobiosensors. *Polymers* 8: 249.
- Lehninger, A. L. (1982). Principles of Biochemistry, Worth Publisher Inc, USA 27:803-825

- Li, D., Muller, M.B., Gilje, S., Kaner, R. B. and Wallace, G.G. (2008). Processable aqueous dispersions of graphene nanosheets. *Nature Nanotechnology* 3: 101.
- Li, J. and Wu, N. (2017) Biosensors Based on Nanomaterials and Nanodevices, CRC Press, 517.
- Li, X. Q., Zhang, H., and Yan, Y. J. (2001). A Super exchange-Mediated Sequential Hopping Theory for Charge Transfer in DNAJ. *The Journal of Physical Chemistry A*, 105(41), 9563–9567.
- Li, Y, Zhou, Y., Wang, H.Y., Perrett, S., Zhao, Y., Tang, Z., and Nie, G.(2011). Chirality of glutathione surface coating affects the cytotoxicity of quantum dots. Angew. Angewandte Chemie International Edition 50: 5860-5864.
- Lian, K.Y., Ji,Y, F., Li,X, F, Jin., M, X, Ding, D, J and Luo,Y. (2013). Big Bandgap in Highly Reduced Graphene Oxides. *The Journal of Physical Chemistry A* 117(12): 6049–6054.
- Liang, H. F., Smith, C.T.G., Mills, C.A., and Silva, S.R.P. (2015). The Band Structure of Graphene Oxide Examined Using Photoluminescence Spectroscopy. *Journal of Materials Chemistry*. C 3: 12484-12491.
- Liepold P, Wieder H, Hillebrandt H, Friebel A, and Hartwich G.(2005). DNA-arrays with electrical detection: A label-free low cost technology for routine use in life sciences and diagnostics. *Bioelectrochemistry* 67:143–150.
- Lim, H., Lee, J., Shin, H. (2010). Spatially resolved spontaneous reactivity of diazonium salt on edge and basal plane of graphene without surfactant and its doping effect. *Langmuir* 26(14): 12278-12284.
- Lin N, Huang J, and Dufresne A. (2012). Preparation, properties and applications of polysaccharide nanocrystals in advanced functional nanomaterials: a review. *Nanoscale* 4(32): 74–94.
- Lin, N. and.Dufresne, A. (2014). Nanocellulose in biomedicine: Current status and future prospect. *European Polymer Journal* 59:302-325.
- Lin, X., Wu, P., Chen, W., Zhang, Y., Xia, X. (2007). Electrochemical DNA biosensor for the detection of short DNA species of Chronic Myelogenous Leukemia by using methylene blue. *Talanta* 72, 468–471.
- Liu G., Wang J., Kim J., and Jan M. R. (2004). Electrochemical Coding for Multiplexed Immunoassays of Proteins, *Analytical Chemistry* 76, 7126-7130.
- Liu, C, Jiang, D, Xiang G, Liu L, Liu F, Pu, X. (2014). An electrochemical DNA biosensor for the detection of Mycobacterium tuberculosis, based on signal amplification of graphene and a gold nanoparticle-polyaniline nanocomposite. *Analyst* 139(21):5460-5.
- Liu, X., Chen, M., Hou, T., Wang, X., Liu, S., and Li, F. (2013). A novel electrochemical biosensor for label-free detection of uracil DNA glycosylase activity based on enzyme-catalyzed removal of uracil bases inducing strand release. *Electrochimica Acta* 13, 514–518.

- Liu, Z., Robinson, J. T.,, X. M. and Dai, H. J. (2008). PEGylated nanographene oxide for delivery of water-insoluble cancer drugs. *Journal of the American Chemical Society* 130 (33), 10876-10900.
- Liu, Z.H., Shi, X.D., and Wu, X.Y.(2001). The method of Mycobacterium tuberculosis rapid cultivation fluorescence detection. *Chinese Journal of Clinical Laboratory Science* 19: 347-348.
- Liu. D., Zou, X., Zhong, L., Lou,Y., Yang, B. and Yin, Y. (2014). New features of DNA damage by acid hydrolysis in MALDI-TOF mass spectrum. *International Journal of Mass Spectrometry* 374: 20-25.
- Loh K.P. and Bao Q.L. (2010). The chemistry of graphene. Journal of Materials Chemistry. 20, 2277–2289.
- Long, W and Fu, L. (2017). Hydrothermal synthesis of ZnO flower-reduced graphene oxide composite for electrochemical determination of ascorbic acid, *Fullerenes, Nanotubes and Carbon Nanostructures Journal* 25(7):404-409
- Lucarelli, F., Marrazzaa, G., Turner, A.P.F., Mascini, M. (2004). Carbon and gold electrodes as electrochemical transducers for DNA hybridization sensors *Biosensors and Bioelectronics*, 19, 515–530.
- Lucarelli, F., Palchetti, I., Marrazza, G and Mascini, M. (2002). Electrochemical DNA biosensor as a screening tool for the detection of toxicants in water and wastewater samples. *Talanta* 56(5): 949–957.
- Ma, H.H., Zhang, Y.,Hu,Q. H.,Yan, D.,Yu, Z.Z.,and Zhai, M. (2012) Chemical reduction and removal of Cr(vi) from acidic aqueous solution by ethylenediamine-reduced graphene oxide. *Journal of Materials Chemistry* 22: 5914.
- Mackay, I.M. (2004). Real-time PCR in the microbiology laboratory. *Clinical Microbiology and Infection*.10(3):190-212
- Magana-Arachchi, D., Perera , J, Gamage, S and Chandrasekharan V. (2008). Low cost in house PCR for the routine diagnosis of extra-pulmonary tuberculosis. *International Journal of Tuberculosis and Lung Disease* 12(3): 275-80.
- Mahaisavariya, P., Chaiprasert, A., Manonukul, J., Khemngern, S. and Tingtoy, N. (2005) identification of Mycobacterium species by polymerase chain reaction (PCR) from paraffin- embedded tissue compare to AFB staining in pathological sections *Journal of the Medical Association of Thailand* 88: 108-113.
- Mandal A and Chakrabarty D (2011). Isolation of nanocellulose from waste sugarcane bagasse (SCB) and its characterization, *Carbohydrate. Polymers* 86: 1291-1299,

- Marmur, J and Doty, P. (1961). Thermal renaturation of deoxyribonucleic acids. *Journal of Molecular Biology* 3: 585–594.
- Marmur, J. and Ts'o, P.O.P., (1961). Denaturation of deoxyribonucleic acid by formamide. *Biochimica et Biophysica Acta* 51:32–36.
- Matsuno, A., Itoh, J., Takekoshi, S., Nagashima, T, and Osamura, R.Y. (2006). Three-Dimensional Imaging of the Intracellular Localization of Growth Hormone and Prolactin and Their mRNA Using Nanocrystal (Quantum dot) and Confocal Laser Scanning Microscopy Techniques. *Journal of Histochemistry* & Cytochemistry 53(7): 833-838.
- Maybodi, A.S., Abbasi, F. and Akhoondi, F. (2014). Aqueous synthesis and characterization of CdS quantum dots capped with some amino acids and investigations of their photocatalytic activities. *Colloids and Surfaces A: Physicochem. Eng. Aspects* 447: 111–119.
- Mc Creery, R.L. (1991). Electroanalytical Chemistry, 17 (Dekker, New York Eds).
- McNerney, R., Wondafrash, B, A., Amena, K., Tesfaye, A., McCash, E.M., and Nicol, J. Murray. (2010). Field test of a novel detection device for Mycobacterium tuberculosis antigen in cough. *BMC Infectious Diseases* 10: 161.
- Gonzalez, M., Xu, Y., Chen, W.Z., Farhanghi, B., and Charpentier, P.A. (2011). CdS and CdTeS quantum dot decorated TiO<sub>2</sub> nanowires. Synthesis and photoefficiency. *Nanotechnology* 22: 065603.
- Merkoci A. (2007). Electrochemical biosensing with nanoparticles. *The feb journal*. 274(2): 310-316.
- Metaferia, B., Wei, J.S., Song, Y.K., Evangelista, J., Aschenbach, K., Johansson, P., Wen X, Chen Q, Lee, A., Hempel, H., Gheeya, J.S., Getty, S., Gomez, R. and Khan, J. (2013). Development of peptide nucleic acid probes for detection of the HER2 oncogene. *PLoS One* 8(4):e58870.
- Middlebrook, G., Tigertt, W. D. and Z. Reggiardo.(1977). Automat- able radiometric detection of grwoth of Mycobacterium tuberculosis in selective media. *American Review of Respiratory Disease* 115(6):1066–1069.
- Miodek, A., Mejri, N. Gomgnimbou., Sola, M., Korri Youssoufi, H. (2015). E-DNA Sensor of Mycobacterium tuberculosis Based on Electrochemical Assembly of Nanomaterials (MWCNTs/PPy/ PAMAM). *Analytical Chemistry* 87: 9257–9264.
- Moccia, M., Adamo, M,F,A and Saviano. M. (2014). Insights on chiral, backbone modified peptide nucleic acids: Properties and biological activity. Artif DNA *PNA XNA* 5(3): e1107176.
- Mohamad, N.R., Marzuki, N.H.C., Buang, N.H., Huyop, F., and Wahab R. A. (2015). An overview of technologies for immobilization of enzymes and surface analysis techniques for immobilized enzymes. *Biotechnol Biotechnol Equip* 29(2): 205–220

- Moon, R.J., Martini, A., Nairn, J., Simonsenf, J. and Youngblood, J. (2011). Cellulose nanomaterials review: structure, properties and nanocomposites. *Chemical Society, Reviews* 40(7): 3941-3994.
- Moser, J. and Bachtold, A. (2009). Fabrication of large addition energy quantum dots in graphene. *Applied Physics Letters* 95, 173506.
- Mu, X and Gray D.G (2015). Droplets of cellulose nanocrystal suspensions on drying give iridescent 3-D coffee-stain rings .*Cellulose* 22 :1103-1107.
- Mundinamani, S.P and Rabinal, M, K. (2014). Cyclic Voltammetric Studies on the Role of Electrode Electrode Surface Modification and Electrolyte Solution of an Electrochemical Cell. *Journal of Applied Chemistry* 7:45–52.
- Natarajan, A., Shalini Devi,K.S., Raja, S., and Kumar,A.S. (2017). An Elegant Analysis of White Spot Syndrome Virus Using a Graphene Oxide/Methylene Blue based Electrochemical Immunosensor Platform. *Scientific Reports* 7: 46169.
- Nicholson, R,S. (1965). Theory and application of cyclic voltametry for measurement of electrode reaction kinetic, *Analytical Chemistry* 37(1): 1351–1355.
- Nielsen, P. E., and Michael Egholm. (1999). An Introduction to Peptide Nucleic Acid. Current Issues *Journal of Molecular Biology*. 1(2): 89-104.
- Niemz A, Ferguson, T.M, and Boyle, D.S. (2011). Point-of-care nucleic acid testing for infectious dis- eases. *Trends Biotechnol* 29: 240–50.
- Nirsch, M., Reuter, F and Voros. J. (2011). Review of transducer principles for labelfree, Biomolecular Interaction. *Analysis Biosensors (Basel)* 1(3), 70–92.
- Niu, M.C. Y. (2000). Composite Airframe Structures, 2nd ed., Hong Kong Conmilit, *Press Limited* 3.
- Noel, M and Vasu, K. (1999). Cyclic Voltammetry and the frontiers of electrochemistry eds. Oxford & IBH Publishing Co.Pvt.Ltd. New Delhi Eds.
- Nogueira, J. J. and González, L. (2014). Molecular Dynamics Simulations of Binding Modes between Methylene Blue and DNA with Alternating GC and AT Sequences, *Biochemistry* 53 (14): 2391–2412.
- Noordhoek, G.T., Kaan, J, A., Mulder, S., Wilke., H. and Kolk, A.H. (1995). Routine application of the polymerase chain reaction for detection of Mycobacterium tuberculosis in clinical samples. *J Clin Pathol* 48(9): 810-814.
- Orum, H., Nielsen, P., Jorgensen, M., Larsson, C., Stanley, C., and Koch, T. (1995). Sequence specific purification of nuceic acids by PNA controlled hybrid selection. *Biotechniques* 19: 472-480.

- Ozkan, D., Erdem, A. Kara, P., Kerman, K. Gooding, J.J. Nielsen, P.E. and Ozsoz, M. (2002). Electrochemical detection of hybridization using peptide nucleic acids and methylene blue on self-assembled alkanethiol monolayer modified gold electrodes. *Electrochem. Commun* 4: 796–802.
- Palanisamy, S., Chen ,S.M., and Sarawathi , R. (2012). A novel nonenzymatic hydrogen peroxide sensor based on reduced graphene oxide/ZnO composite modified electrode. *Sensors and Actuators B: Chemical* 166: 372–377.
- Palecek, E and Bartosik, M. (2012). Electrochemistry of Nucleic Acids. *Chem. Rev* 112 (6), 3427–3481.
- Palecek, E., Tomschik, M., Stankova, V. and Havran, L. (1997). Chronopotentiometric stripping of DNA at mercury electrodes *Electroanalysis*, 9-990.
- Patolsky, F.; Weizmann, Y.; Willner, I.(2002). Redox-active nucleic-acid replica for the amplified bioelectrocatalytic detection of viral DNA,. *Journal of the American Chemical Society*124:770-772
- Pan, S., Sun, X., Lee.J. K. (2006). Stability of complementary and mismatched DNA duplexes: Comparison and contrast in gas versus solution phases. *International Journal of Mass Spectrometry* 253: 238–248.
- Pandey, D., R. Reifenberger Piner, R. (2008). Scanning probe microscopy study of exfoliated oxidized graphene sheets. *Surface Science*. 602 (9), 1607-1613.
- Pandikumar, A., Gregory T, Peik See, T., Omar, F,S., Jayaba, S., Kamali, K, Z., Yusoff, N., Jamil, Ramaraj, A, R., John, S, A., Lim. H. N., and Huang, N. M. (2014) Graphene and its nanocomposite material based electrochemical sensor platform for dopamine. *RSC Advance* 4: 63296-63323.
- Paredes, J. I., Villar-Rodil, S., Solis-Fernandez, P., Martinez-Alonso, A and Tas- con, J.M.D. (2009). Atomic force and scanning tunneling microscopy imaging of graphene nanosheets derived from graphite oxide, *Langmuir* 25, 5957-5968.
- Park, M., Chang, H., Jeong, D.H., and Hyun, J. (2013). Spatial deformation of nanocellulose hydrogel enhances SERS. *BioChip J* 7: 234–241.
- Park. M. K., Park. J.W., Wikle, H.C. and Chin, B.A. (2012). Evaluation of phage-based magnetoelastic biosensors for direct detection of Salmonella Typhimurium on spinach leaves. Sensors and Actuators B: Chemical.176: 1134-1140
- Parsons, L.M., Somoskovi, A., Gutierrez, C., Lee, E., Paramasivan C.N and Abimiku A. (2011). Labora tory diagnosis of tuberculosis in resource poor countries, challenges and opportunities. *Clinical Microbiology Reviews* 24. 314–50.
- Pattnaik, S., Swain , K., and Lin, Z. (2016). Graphene and graphene-based nanocomposites:biomedical applications and biosafety. *Journal of Materials Chemistry*. B 4: 7813-7831.
- Paul, A., Watson, R.M., Lund, P., Xing, Y., Burke, K., He, Y., Borguet, E., (2008). Achim, C and Waldeck, D.H (2008). Charge Transfer through Single-

Stranded Peptide Nucleic Acid Composed of Thymine Nucleotides. *The Journal of Physical Chemistry C* 112 (18):7233–7240.

- Peik-See, T., Alagarsamy, P., Huang Nay, M., Lim H.N and Yusran, S (2014). Simultaneous Electrochemical Detection of Dopamine and Ascorbic Acid Using an Iron Oxide/Reduced Graphene Oxide Modified Glassy Carbon Electrode. Sensors 14: 15227-15243.
- Pertile, R.A. (2012). Bacterial Cellulose: Long-Term Biocompatibility Studies. *Journal* of Biomaterials Science, Polymer Edition 23(10):1339-54.
- Peter T. K and William R. H.(1983) Cyclic voltammetry. *Journal of Chemical Education*. 60 (9): 702.
- Peterson, A.W., Heaton, R.J and Georgiadis, R.M. (2001). The effect of surface probe density on DNA hybridization. *Nucleic Acids Research* 29(24): 5163–5168.
- Pheeney, C. G., and Barton, J. K. (2012) DNA Electrochemistry with Tethered Methylene Blue. *Langmuir* 28: 7063–7070.
- Piatek, A.S., Tyagi, S., Pol, A.C., Telenti, A, Miller, L.P. and Kramer F.R. (1998). Molecular beacon se quence analysis for detecting drug resistance in Mycobacterium tuberculosis. *Nature Biotechnology* 16: 359–63
- Pohanka, M (2008) Electrochemical biosensors principles and applications Journal of Applied Biomedicine . *Journal of Applied Biomedicine* 6(2): 57–64.
- Powell, R.D., Whitworth, W. C., Bernardo, J., Moonan, P.K and Mazurek, G.H. (2011). Unusual interferon gamma measurements with Quanti FERON TB Gold and Quantferon TB Gold in-tube tests. *PLoS One* 6(6):e20061.
- Prabhakar, N., Solanki, P.R., Kaushik, A., Pandey, M.K. and Malhotra B.D. (2010). Peptide Nucleic Acid Immobilized Biocompatible Silane Nanocomposite Platform for Mycobacterium tuberculosis Detection. *Electroanalysis* 22, 2672 – 2682.
- Prabhakar, N., Singh, H. and Malhotra, B.D. (2008) Nucleic acid immobilized polypyrrole-polyvinylsulphonate film for *Mycobacterium tuberculosis* detection. *Electrochemistry Com munications* 10(6): 821–826.
- Prabhakar, N., Arora, K., Arya, S. K., Solanki, P.R., Iwamoto, M., Singh, H. and Malhotra, B.D. (2008). Nucleic acid sensor for M. tuberculosis detection based on surface plasmon resonance. *Analyst* 133: 1587–1592
- Prakash, S., Chakrabarty, T., Singh, A.K., Shahi, V.K. (2013). Polymer thin films embedded with metal nanoparticles for electrochemical biosensors applications. *Biosensors and Bioelectronics* 41, 43–53.
- Pumera, M., Castan-eda, A. T., Pividori, M. I., Eritja, R., Merkoc, A. and Salvador Alegret. (2005). Magnetically Trigged Direct Electrochemical Detection of

DNA Hybridization Using Au67 Quantum Dot as Electrical Tracer. *Langmuir* 21: 9625-9629.

- Pumera, M., Sanchez, S., Ichinose, I., and Tang, J.(2007). Electrochemical nanobiosensors. Sensors and Actuators B 123(2): 1195–1205.
- Putzbach, W. and Ronkainen, N.J. (2013). Immobilization Techniques in the Fabrication of Nanomaterial-Based Electrochemical Biosensors: A Review Sensors (Basel) 13(4): 4811–4840.
- Qi, X., Gao, H., Zhang, Y., X.Wang, Y.Chen, W. Sun. (2012). Electrochemical DNA biosensor with chitosan-Co3O4 nanorod-graphene composite for the sensitive detection of staphylococcus aureus nuc gene sequence. *Bioelectrochemistry*. 88: 42–47.
- Qin, D, L., He, X.X., Wang, K.M. and Tan, W.H. (2008). Using fluorescent nanoparticles and SYBR Green I based two- color flow cytometry to determineMycobacterium tuberculosis avoiding false positives. *Biosensors* and Bioelectronics 24(4):626–631.
- Qing, W., Wang,Y., Wang,Y., Zhao,D., Liu,X., Zhu,J. (2016). The modified nanocrystalline cellulose for hydrophobic drug delivery. *Applied Surface Science* 366: 404–409.
- Qutub, N. and Sabir, S. (2012). Optical, Thermal and Structural Properties of CdS Quantum Dots Synthesized by A Simple Chemical Route Int. J. Nanosci. Nanotechnol 8 (2): 111-120.
- Raj, A., Singh, N., Gupta, K. B., Chaudhary, D., Yadav, A., Chaudhary, A., Agarwal, K., Varma-Basil, M., Prasad, R., Khuller, G.K., and Mehta. P. K. (2016). Comparative Evaluation of Several Gene Targets for Designing a Multiplex-PCR for an Early Diagnosis of Extrapulmonary Tuberculosis. *Yonsei Medical Journal* 57(1): 88–96.
- Rao, A.R. and Grainger, D.W. (2014). Biophysical properties of nucleic acids at surface relevent to microarray performance. *Biomaterials Science*, 2(4): 436– 471.
- Rashid, J.I.A., Yusof, N.A., Abdullah, J., Hashim, and U.,Hajian, R.(2014). The utilization of SiNWs/AuNPs-modified indium tin oxide (ITO) in fabrication of electrochemical DNA sensor. *Materials Science and Engineering C* 45 :270–276.
- Rasmussen, H., Kastrup, J. S., Nielsen, J. N., Nielsen, J. M. and Nielsen, P. E. (1997) Crystal structure of a peptide nucleic acid (PNA) duplex at 1.7 Å resolution. *Nature Structural & Molecular Biology*. 4, 98–101
- Ricci, F., Lai, R.Y., Heeger ,A.J., Plaxco, K.W. and Sumner, J.J. (2007). Effect of Molecular Crowding on the Response of an Electrochemical DNA Sensor, *Langmuir* 23(12):6827–6834.

- Rivas, G.A., Rubianes, M.D. and Rodríguez, M.C. (2007). Carbon nanotubes for electrochemical biosensing, *Talanta* 74(3) 291–307
- Rodriguez-Perez, L., Herranza, M.A and Martin, N. (2013) The chemistry of pristine graphene, *Chemical Communications*. 49: 3721–3735.
- Rohs, R., and Sklenar, H. (2004). Methylene Blue Binding to DNA with Alternating AT Base Sequence: Minor Groove Binding is Favored over Intercalation. *Journal of Biomolecular Structure and Dynamics* 21(5):699–711.
- Rundi, C, Fielding K, Godfrey, F. P., Rodrigues, L.C. and Mangtani, P. (2011). Delays in seeking treatment for symptomatic tuberculosis in Sabah, East Malaysia: factors for patient delay. *International Journal of Tuberculosis and Lung Disease* 15(9): 1231–8.
- Ruzin, S. (1999). Plant Microtechnique and Microscopy.oxford University Press 14
- Saito T, Uematsu T, Kimura S, Enomae T, and Isogai A. (2011). Self-aligned integration of native cellulose nanofibrils towards producing diverse bulkmaterials. *Soft Matter* 7: 8804–8809.
- Saito, T., Kimura, S, Nishiyama, Y. and Isogai, A. (2007). Cellulose nanofibers prepared by TEMPO-mediated oxidation of native cellulose. *Biomacromolecules* 8(8): 2485-2491
- Salajkova, M., Berglund, L. A. and Zhou, Q. (2012). Hydrophobic cellulose nanocrystals modified with quaternary ammonium salts. *Journal of Materials Chemistry* 22(37):19798-19805.
- Salih, E., Rabeay, M. M., Hassan, Y.A., El-Sherbiny, M.M. (2016). Synthesis, characterization and electrochemical-sensor applications of zinc oxide/graphene oxide nanocomposite. *Journal of Nanostructure in Chemistry* 6 (2): 137–144
- Sam, S., Touahir L., Andresa, J. S., Allongue, P., Chazalviel, J. N and Gouget-Laemmel, A. C. C., de Villeneuve, C. H., Moraillon, A., Ozanam, F., Gabouze, N. and Djebbar, S. (2009) Semiquantitative study of the EDC/NHS activation of acid terminal groups at modified porous silicon surfaces. *Langmuir* 26: 809–814.
- Sato, Y., Fujimoto, K and Kawaguchi, H. (2003) Detection of a K-ras point mutation employing peptide nucleic acid at the surface of a SPR biosensor. *Colloids Surface B*, 27: 23–31.
- Schmidt, P., Nielsen, P.E and Orgel, L. E. (1996) Separation of 'uncharged' oligodeoxynucleotide analogs by anion-exchange chromatography at high pH. *Analytical Biochemistry* 235: 239–241.
- Shamsudin, S.A., Takenaka, M., and Hasegawa, H. (2014). Introducing the Safe Capsule for CdS Quantum Dots as Bio-Labeling Device. *Journal of the Japan Society of Powder and Powder Metallurgy* 61:129-132.

- Shan, C., Yang, H., Han, D., Zhang, Q., Ivaska, A., and Niu, L. (2009). Water-soluble graphene covalently functionalized by biocompatible poly-L-lysine. *Langmuir* 25: 12030–12033.
- Shao Y, Wang J, Engelhard M, Wang C, and Lin Y, (2010). Facile and controllable electrochemical reduction of graphene oxide and its applications. *Journal of Materials Chemistry* 20:743–748.
- Sharma, D., Kanchi, S., Myalowenkos, Sabela and Bisetty. K. (2015) Insight into the biosensing of graphene oxide: Present and future prospects. *Arabian Journal* of Chemistry 9(2): 238–261.
- Sharma, R., Baik, J. and Perera, C. (2010). Anomalously large reactivity of single graphene layers and edges toward electron transfer chemistries. *Nano Letters* 10(2): 398-405.
- Sharma, S., Rodríguez, J. Z., Estrela, P. and O'Kennedy, R. (2015). Point-of-Care Diagnostics in Low Resource Settings: Present Status and Future Role of Microfluidics. *Biosensors* 5: 577-601.
- Sheka, E.F. (2014). The uniqueness of physical and chemical natures of graphene: Their coherence and conflicts. *International Journal of Quantum Chemistry* 114:1079-1095.
- Shen, J., Dudik, L and Liu, C.C. (2007). An iridium nanoparticles dispersed carbon based thick film electrochemical biosensor and its application for a single use, disposable glucose biosensor, *Sensors and Actuators B* 125(1):106–113.
- Shi, Z., Philip, G.O., and Yang, G. (2013). Nanocellulose electroconductive composites. *Nanoscale* 5:3194–3201.
- Shih, W.Y. and Shih.W.H. (2007).Synthesis and Characterization of Aqueous Carboxyl-Capped CdS Quantum Dots for Bioapplications. *Industrial & Engineering Chemistry Research* 46:2013-2019.
- Shimotoyodome, A, Suzuki J, Kumamoto Y, Hase T, Isogai A. (2011). Regulation of postprandial blood metabolic variables by TEMPO-oxidized cellulose nanofibers. *Biomacromolecules* 12:3812–3818.
- Shin, H. J, Kim, K.K, Benayad, A, Yoon, SM, Park, H.K, Jung IS, Jin MH, Jeong, H.K, Kim, J.M, Choi, J.Y, Lee, Y.H. (2009). Efficient reduction of graphite oxide by sodium borohydride and its effect on electrical conductance. *Angewandte Chemie International Edition* 19(12):1987–1992.
- Shin, M.K., Lee, B., Kim, S.H., Lee, J.A., Spinks, G.M., Gambhir, S., Wallace, G.G Kozlov, M.E., Baughman, R.H and Kim, S.J. (2011). Synergistic toughening of composite fibres by self-alignment of reduced graphene oxide and carbon nanotubes. *Nature Comunications*. 3

- Shopsowitz, K.E., Hamad W.Y., MacLachlan, M.J (2011). Chiral nematic mesoporous carbon derived from nanocrystalline cellulose. Angew *Angewandte Chemie International Edition* 50:10991-10995.
- Shukla, S.K. (2014). Recent developments in biomedical applications of quantum dots. *Advance. Material. Review.* 1(1): 2-12.
- Siddiqi K, Lambert ML, Walley J. (2003). Clinical diagnosis of smear-negative pulmonary tuberculosis in low-income countries: the current evidence. *Lancet Infect* 3:288-296.
- Singh, S., Kaushal, A., Khare, S., Kumarab, P., and Kumar, A (2014). Gold mercaptopropionic acid–polyethylenimine composite based DNA sensor for early detection of rheumatic heart disease. *Analyst*, 139, 3600–3606.
- Slinker, J., Muren, N., Renfrew, S and Barton, J. K.(2011). DNA charge transport over 34 nm. *Nature Chemistry*. 3: 228–233.
- Smalley, J. F., Finklea, H.O., Chidsey, C.E.D., Linford, M.R., Creager, S.E., Ferraris, J.P., Chalfant, K., Zawodzinsk, T., Feldberg, O.S.W and Newton, M.D. (2003) Heterogeneous Electron-Transfer Kinetics for Ruthenium and Ferrocene Redox Moieties through Alkanethiol Monolayers on Gold. *Journal of the American Chemical Society*
- Sobhana, S. S., Vimala Dev M., Sastry, T. P. and Mandal. A. B. (2010) .CdS quantum dots for measurement of the size-dependent optical properties of thiol capping. *Journal of Nanoparticle Research* 3(4): 1747–1757.
- Sobhana, S.S.L., Devi, M.V.,Sastry, T.P and Mandal, A. B. (2011). CdS quantum dots for measurement of the size- dependent optical properties of thiol capping, Journal of Nanoparticle Research 13: 1747–1757.
- Soomro, S.A., Memon, N., Solangi, A.R., Sirajuddin, Qureshi, T., Behzad, A.R (2014). Disposable screen printed graphite electrode for the direct electrochemical determination of ibuprofen in surface water. *EnvironmentalNanotechnology, Monitoring & Management* 1(2): 8–13.
- Soni, B., Hassan el B., Mahmoud B. (2015). Chemical isolation and characterization of different cellulose nanofibers from cotton stalks, Carbohydrate Polymers 134(10):581-589
- Srikanta, S and Lennart, N. (1998). Molecular Dynamics of Duplex Systems Involving PNA: Structural and Dynamical Consequences of the Nucleic Acid Backbone, *Journal of the American Chemical Society* 120 (4): 619–631.
- Stankovich S, Piner, R.D., Nguyen, S.T., Ruoff, R.S.(2006). Synthesis and exfoliation of isocyanate-treated graphene oxide nanoplatelets. *Carbon* 44 (15): 3342–3347.
- Stankovich, S., Dikin, D. A., Piner, R. D., Kleinhammes, K. A., Jia, Y., Wu, Y., Nguyen, S. B. T. and Ruoff, R. S.(2007). *Carbon*, 45: 1558–1565.

- Stankovich, S.,Dikin, D. A., Dommmett, G. H. B. Kohlhaas, K. M. Zimney, E. J. Stach, E. A.Piner, R. D. Nguyen, S. T. Ruoff, R. S. (2006). Graphene based composite materials. *Nature*, 442: 282–286.
- Steel, A. B., Levicky, R. L., Herne, T. M. and Tarlov, M. J. (2000). Immobilization of nucleic acids at solid surfaces: effect of oligonucleotide length on layer assembly. *Biophysics. J.* 79: 975–981.
- Steel, A.B., Herne, T.M., and Tarlov M. J. (1999). Electrostatic Interactions of Redox Cations with Surface-Immobilized and Solution DNA Bioconjugate Chemistry 10(3):419-23.
- Steiner, D., Dirk D, Uri B, Fabio D. S, Liberato, M and Oded M. (2008) Determination of Band Offsets in Heterostructured Colloidal Nanorods Using Scanning Tunneling Spectroscopy Nano Letter 8(9): 2954–2958
- Stellwagen, N.C. (1998). DNA gel electrophoresis. Nucleic acid electrophoresis laboratory manual Verlag. *Berlin-Heidelberg- New York*.
- Su B, Tang D, Li Q, Tang J, and Chen G. (2011). Gold silver graphene hybrid nanosheets-based sensors for sensitive amperometric immunoassay of alpha-fetoprotein using nanogold-enclosed titania nanoparticles as labels. *Analytica Chimica Acta* 692(1-2): 116-24.
- Sun, B., Hou, Q., He, Z., Liu, Z., and Ni, Y. (2014). Cellulose nanocrystals (CNC) as carriers for a spirooxazine dye and its effect on photochromic efficiency, Carbohydr. *Polymer* 111:419–424.
- Sun, X. Y., He, P.G., Liu, S.H., Ye, J.N. and Fang, Y.Z. (1998) Immobilization of singlestranded deoxyribonucleic acid on gold electrode with self-assembled aminoethanethiol monolayer for DNA electrochemical sensor applications, *Talanta* 47:487–495.
- Tachibana, Y., Akiyama, H.Y, Ohtsuka, Y., Torimoto, T., and Kuwabata, S. (2010). CdS Quantum Dots Sensitized TiO2 Sandwich Type Photoelectrochemical Solar Cells. CSJ Journal 36: 88-89.
- Tan, C., Huang, X and Zhang, H. (2013). Synthesis and applications of graphene-based noble metal nanostructures. *Materials Today* 16(2): 29–36.
- Tan, S. C and Yiap, B.C. (2009). DNA, RNA, and Protein Extraction: The Past and The Present. *Journal of Biomedicine and Biotechnology* 10.
- Tan, X.X, Actor, J.K, Chen Y. (2005). Peptide nucleic acid antisense oligomer as a therapeutic strategy against bacterial infection: proof of principle using mouse intraperitoneal infection. *Antimicrobial Agents and Chemotherapy* 49(8):3203-7.
- Tanner, E.E.L., Barnes, E.O., Tickell, C.B., Goodrich, P., Hardacre, C and Compton, R.G. (2015). Application of Asymmetric Marcus–Hush Theory to

Voltammetry in Room-Temperature Ionic Liquids. *The Journal of Physical Chemistry C*, 119 (13):7360–7370

- Tang, L., Wang, Y., Li, Y., Feng H., Lu J, and Li, J.(2009) Preparation, structure, and electrochemical properties of reduced graphene sheet films. *Advanced Functional Materials* 19: 2782–2789.
- Tang, L., Zeng, G., Shen, G., Li, Y., Liu, C and Li, Z. (2009). Sensitive detection of lip genes by electrochemical DNA sensor and its application in polymerase chain reaction amplicons from Phanerochaete chrysosporium. *Biosensors and Bioelectronics* 24: 1474–9.
- Tang, S.R., Tong, P., Lu,W., Chen, J.F. and Yan, Z.M., (2014). A novel label-free electrochemical sensor for Hg2+ based on the catalytic formation of metal nanoparticle Biosensors and Bioelectronics: 59, 1–5.
- Tichoniuk, M., Ligaj, M., and Filipiak, M. (2008). Application of DNA Hybridization Biosensor as a Screening Method for the Detection of Genetically Modified Food Components. *Sensors* (Basel) 8(4): 2118–2135.
- Torati, S.T., Reddya, V., Yoon, S.S., and Kim, C. G. (2015). Electrochemical Biosensor for Mycobacterium Tuberculosis DNA Detection Based on Gold Nanotubes Array Electrode Platform. *Biosensors and Bioelectronics* 78: 483-488.
- Torres-Costa, J., Silva R., Ringshausen, F.C. and Nienhaus A. (2011). Screening for tuberculosis and prediction of disease in Portuguese healthcare workers. *Journal of Occupational Medicine and Toxicology* 6: 19.
- Tortoli, E. F., Lavinia, and Simonetti. M. T. (1997). Evaluation of a commer- cial ligase chain reaction kit (Abbott LCx) for direct detection of Mycobac- terium tuberculosis in pulmonary and extrapulmonary specimens. *Journal of Clinical Microbiology*. 35:2424–2426.
- Tsiafoulis, C.G., Florou, A.B., Trikalitis, P.N., Bakas, T., and Prodromidis, M.I. (2005) Electrochemical study of ferrocene intercalated vanadium pentoxide xerogel/polyvinyl alcohol composite films: Application in the development of amperometric biosensors. *Electrochemistry Communications* 7:781–788.
- Ulianas, A., Heng, L.Y., Hanifah, S.B., and Ling, T.L. (2012). An Electrochemical DNA Microbiosensor Based on Succinimide-Modified Acrylic Microspheres. Sensors 12:5445-5460.
- Valentini, L and Bon, S. B. (2016) Graphene Based Bionic Composites with Multifunctional and Repairing Properties ACS Appl. Mater. Interfaces, 8 (12):7607–7612.
- Voicu, R., Boukherroub, R., Bartzoka, V., Ward, T., Wojtyk, J. T. C., and Wayner, D. D. M. (2004) Formation, characterization, and chemistry of undecanoic acid-terminated silicon surfaces: Patterning and immobilization of DNA. *Langmuir* 20:11713–11720.

- Verma, D., Gope, P.C., Maheshwari, M.K. and Sharma, R.K. (2012). Bagasse Fiber Composites-A Review. J. Mater. Environ. Sci. 3 (6): 1079-1092
- Walch, N. J., Nabok, A., Davis, F. and Higson, S.P.J. (2016). Characterization of thin films of graphene surfactant composites produced through a novel semiautomated method. *Nanotechnology* 7: 209–219.
- Wang, C., Shim, M. and Guyot-Sionnest, P. (2001). Electrochromic Nanocrystal Quantum Dots. *Science* 291: 2390-2392.
- Wang, C., Yan, Q., Liu, H.B., Zhou, X.H. and Xiao, S.J. (2011) Different EDC/NHS Activation Mechanisms between PAA and PMAA Brushes and the Following Amidation Reactions *Langmuir* : 12058–12068.
- Wang, D., Li, Y., Hasin, P., Wu, Y. (2011) Preparation, characterization, and electrocatalytic performance of graphene-methylene blue thin films. *Nano research* 4(1): 124–130.
- Wang, J. (1994). Analytical Electrochemistry" T eds (VCH Publishers Inc New York Eds).
- Wang, J (2002). Review Electrochemical nucleic acid biosensors. *Analytica Chimica* Acta 469:63–71.
- Wang, J., (2006). Electrochemical biosensors: towards point-of-care cancer diagnostics. *Biosensors and Bioelectronics* 21:1887–1892.
- Wang, J., Liu, G., and Merkoçi, A. (2003). Electrochemical Coding Technology for Simultaneous Detection of Multiple DNA Targets. *Journal of the American Chemical Society*. 125:3214-3215.
- Wang, J., Palecek, E., Nielsen, P. E., Rivas, G., Cai, X., Shiraishi, H., Dontha,N.,Luo, D. and Farias, P.A.M. (1996). Peptide Nucleic Acid Probes for Sequence-Specific DNA Biosensors. *Journal of the American Chemical Society* 118 (33): 7667–7670.
- Wang J. (1998). DNA biosensors based on Peptide Nucleic Acid (PNA) recognition layers. *Biosensors and Bioelectronics* 13: 757-762.
- Wang, J., Musameh, M., and Lin, Y. (2003). Solubilization of carbon nanotubes by Nafion toward the preparation of amperometric biosensors. *Journal of the American Chemical Society* 125(9):2408–2409.
- Wang, S., Cole, I. S and Li. Q. (2016). The toxicity of graphene quantum dots. *RSC* Adv 6: 89867-89878.
- Wang, X and Son, A. (2013). Effects of pretreatment on the denaturation and fragmentation of genomic DNA for DNA hybridization, *Environ. Sci.*: *Processes Impacts* 15, 2204-2212.

- Wang, X., Lim, H.J and Son, A (2014). Characterization of denaturation and renaturation of DNA for DNA hybridization. *Environmental Health and Toxicology* 29: e2014007.
- Wang, Z., Tammela, P., Stromme, M and Nyholm, L. (2015). Nanocellulose coupled flexible polypyrrole/graphene oxide composite paper electrodes with high volumetric capacitance. *Nanoscale* 7:3418-3423.
- Wang Z. G., Wang Y., Xu H., Li G. and Xu Z. K. (2009). Carbon nanotube-filled nanofibrous membranes electrospun from poly(acrylonitrile-co-acrylic acid) for glucose biosensor. *The Journal of Physical Chemistry C* 113: 2955–2960.
- Wagberg, L., Decher, G., Norgren, M., Lindstrom, T., Ankerfors, M., and Axnas, K. (2008). The build-up of polyelectrolyte multilayers of microfibrillated cellulose and cationic polyelectrolytes. *Langmuir* 24, 784–795.
- Wei, F., Lillehoj, P.B., and Ho,C.M. (2010). DNA Diagnostics: Nanotechnologyenhanced Electrochemical Detection of Nucleic Acids. *PMC*. 67(5):458–468.
- William, T., Parameswaran, U., Lee, W. K., Yeo, T.W., Anstey, N.M. and Ralph, A. P. (2015). Pulmonary tuberculosis in outpatients in Sabah, Malaysia: advanced disease but low incidence of HIV co-infection. *BMC Infectious Diseases*. 1–9.
- Wilson.(1994). Polymerase chain reaction for assessing treatment response in patients with pulmonary tuberculosis. *Journal. Infection* 170:713–716.
- Wong, E. L. S and Gooding, J.J. (2005). Electrochemical Transduction of DNA Hybridization by Long-Range ElectronTransfer. Aust. *Journal of Chemistry*. 58: 280–287.
- Wong, Y and Nicholas A. M. (2010). An Electrostatic Model for DNA Surface Hybridization. *Biophysical Journal* 98: 2954–2963.
- Wopschall, R.H. and Shain, I. (1967). Effects of adsorption of electroactive species in stationary electrode polarography. *Analytical Chemistry*. 39 (13): 1514–1527
- World Health Organization. (2015). Global Tuberculosis Report. 20th edition. www.who.int/tb/publications/global\_report/gtbr2015\_executive\_summary.
- World Health Organization. (2016). Global Tuberculosis Report, 214 www.who.int/tb/publications/global report/en/
- Wu, P., He, Y., Wang, F., and Yan, X.P. (2010). Conjugation of Glucose Oxidase onto Mn-Doped ZnS Quantum Dots for Phosphorescent Sensing of Glucose in Biological Fluids. *Analytical Chemistry* 82 (4):1427–1433.
- Xi, C., Balberg, M., Stephen A., Raskin, B. (2003). Use of DNA and Peptide Nucleic Acid Molecular Beacons for Detection and Quantification of rRNA in Solution and in Whole Cells. *Applied and Environmental Microbiology* 69: 95673-5678.

- Xiao, F., Li, Y., Zan, X., Liao, K., Xu, R. and Duan, H. (2012). Growth of Metal–Metal Oxide Nanostructures on Freestanding Graphene Paper for Flexible *Biosensors* 22: 2487–2494.
- Xiao, Y. and Chang, M. L (2008). Nanocomposites: from fabrications to electrochemical bioapplications. *Electroanalysis*, 20(6). 648–662
- Xie, H., Yang, D., Heller, A. and Gao, Z. (2007). Electrocatalytic Oxidation of Guanine, Guanosine, and Guanosine Monophosphate. *Biophyisc. Journal*. 92(8):170–172.
- Xu, Z. B., Zhang, Y.F., Zhang, X.Z., Chen, X.L. and Tian, J. G. (2009). Porphyrin and Fullerene Covalently Functionalized Graphene Hybrid Materials with Large Nonlinear Optical Properties. *Journal of Physical Chemistry B*, 113 (29): 9681-9686.
- Xu, F., Pellino, M., Pellino, A. M., Knoll, W. (2008). Electrostatic repulsion and steric hindrance effects of surface probe density on deoxyribonucleic acid (DNA)/peptide nucleic acid (PNA) hybridization. *Thin Solid Films* 516(23): 8634-8639.
- Xue, Y., Liu, Y., Lu,F.,Qu,J., Chen, H., and Dai., L. (2012). Functionalization of Graphene Oxide with Polyhedral Oligomeric Silsesquioxane (POSS) for Multifunctional Applications. *Journal of Physical Chemistry Letters* 1: 1607-1612.
- Yakoh, A., Pinyorospathum, C.,Siangproh, W. and Orawon Chailapakul. (2015). Biomedical Probes Based on Inorganic Nanoparticles for Electrochemical and Optical Spectroscopy Applications. *Sensors* 15(9):21427-21477
- Yan, C., Wang, J., Kang, W., Cui, M., Wang, X., Foo, C. Y., Chee, k. J. and Lee. P. S. (2014). Highly Stretchable Piezoresistive Graphene Nanocellulose Nanopaper for Strain. *Sensors Advanced Materials* 26(13):2022–2027.
- Yang, Y.C., Dong , S.W., Shen , T., Jian , C.X., Chang , H.j. and Ying, Li. (2012). A Label-Free Amperometric Immunoassay for Thrombomodulin using Graphene/Silver-Silver Oxide Nanoparticles as a Immobilization Matrix. *Analytical letters* 45 (7): 724-734.
- Yang, L. (2008). Electrical impedance spectroscopy for detection of bacterial cells in suspensions using interdigitated microelectrodes. *Talanta* 74(5): 1621-1629.
- Yang, J., Heo, M., Lee, H.J., Park, S.M., Kim, J.Y and Shin, H.S. (2011). Reduced graphene oxide (rGO)-wrapped fullerene (C60) wires. ACS Nano, 5 (10): 8365–8371.
- Yesil, M., Donmez, S and Arslan, F. (2016). Development of an electrochemical DNA biosensor for detection of specific Mycobacterium tuberculosis sequence based on poly(L-glutamic acid) modified electrode. *Journal of Chemical Sciences.* 128 (11): 1823–1829.

- Yildrim, A. and Seckin, T. (2013). In Situ Preparation of Polyether Amine Functionalized MWCNT Nanofiller as Reinforcing Agents. Advances in Materials Science and Engineering, 6.
- Yin, X., Kong, J., Leon, A.D., Li, Y., Ma, Z., Wierzbinski, E., Achim, C., and David, H. Waldeck. (2014). Luminescence Quenching by Photoinduced Charge Transfer between Metal Complexes in Peptide Nucleic Acids. J. Phys. Chem. B. 118 (30): 9037–9045
- Yu, X., Liu, J., Zuo, S., Yu, Y., Cai, K., and Yang, R. (2013). Application of mercaptosuccinic acid capped CdTe quantum dots for latent fingermark development. *Forensic Science International* 10: 125-30
- Zaib, M .and Athar, M. M. (2015). Electrochemical Evaluation of Phanerocheaete Chrysosporium Based Carbon Paste Electrode with Potassium Ferricyanide Redox System. *International Journal of Electrochemical Science* 10: 6690 – 6702
- Zaitseva, G., Gushikem, Y., Ribeiro, E.S., Rosatto, S.S. (2002). Electrochemical property of methylene blue redox dye immobilizedon porous silica–zirconia–antimonia mixed oxide. *Electrochimica Acta* 47: 1469–1474
- Zhang, W., Li, X., Zou, R., Wu, H., Shi, H., Yu, S and Liu, Y. (2015). Multifunctional glucose biosensors from Fe3O4 nanoparticles modified chitosan/graphene nanocomposites Scientific Reports 5
- Zhang, C., Lou, J., Tu, W., Bao J., and Dai, Z. (2015). Ultrasensitive electrochemical biosensing for DNA using quantum dots combined with restriction endonuclease. *Analyst* 140: 506-511
- Zhang, C., Zhang, R. Z., Ma,Y. Q., Guan,W.B., Wu, X.L., Liu,X., Li, H.,Du,L., and Pan, C.P. (2015). Preparation of Cellulose/Graphene Composite and Its Applications for Triazine Pesticides Adsorption from Water. ACS Sustainable Chemistry & Engineering 3(3): 396–405
- Zhang, F., Li, Y.,Gu, Y., Wang, Z., and Wang, C.(2011). One-pot solvothermal synthesis of a Cu2O/Graphene nanocomposite and its application in an electrochemical sensor for dopamine. *Microchimica Acta* 173(1): 103–109
- Zhang, J., Cao, Y., Feng, J., and Wu, P., (2012). Graphene-Oxide-Sheet-Induced Gelation of Cellulose and Promoted Mechanical Properties of Composite Aerogels. *The Journal of Physical Chemistry C* 116 (14): 8063–8068
- Zhang, N and Appella, D.H. (2010). Advantages of Peptide Nucleic Acids as Diagnostic Platforms for Detection of Nucleic Acids in Resource-Limited Settings. *Journal of Infection* 201: 42–45.
- Zhang, Q, D, Marcha, G, Noel, V, Piroa, B, Reisberga,S, Trana L,D, Hai L,V, Abadia E, Nielsen,P,E Sola, C,M. (2012). Label-free and reagentless electrochemical detection of PCR fragments using self-assembled quinone derivative monolayer: Application to *Mycobacterium tuberculosis*. Pham *Biosensors and Bioelectronics* 32: 163–168

- Zhang, X. and Hu. H (2014). DNA molecules site-specific immobilization and their applications. *Central European Journal of Chemistry* 12(10): 977–993
- Zhao, X.H., Kong, R.M., Zhang,X,B. (2011). Graphene-DNAzyme based biosensor for amplified fluorescence "turn-On" detection of Pb2+ with a high selectivity, *Analytical Chemistry* 83 (13): 5062–5066,
- Zhao, X.H., Ma, Q.J., Wu, and X.X. Zhu, (2012). Graphene oxide based biosensor for sensitive florescence detection of DNA based on exonuclease III-aided signal amplifica tion. *Analytical Chimica Acta* 21: 67–70
- Zhiguo ,G., Shuping, Y., Zaijun , L., Xiulan, S., Guangli, W., Yinjun, F., and Junkang, L. (2011). An ultrasensitive electrochemical biosensor for glucose using CdTe-CdS core–shell quantum dot as ultrafast electron transfer relay between graphene-gold nanocomposite and gold nanoparticle. *Electrochimica Acta*, 56 (25): 9162–9167
- Zhou, L., He, X., He, D., Wang, K. and Qin, D. (2011). Biosensing Technologies for Mycobacterium tuberculosis Detection: Status and New Developments, *Hindawi*. 8.
- Zhu, C., Yang, G., Li, He., Du,D., and Lin,Y, (2015). Electrochemical Sensors and Biosensors Based on Nanomaterials and Nanostructures. *Analytical Chemistry* 87: 230–249
- Zhu, L., Zhao, R., Wang, K., Xiang, H., and Shang, Z. (2008). Electrochemical Behaviors of Methylene Blue on DNA Modified Electrode and Its Application to the Detection of PCR Product from NOS Sequence. Sensors, 8: 5649-5660.
- Zhu, Y., Mural, S., Cai, W., Li, X., Suk, J.W., Potts, J.R., and Ruoff, R.S. (2010). Graphene and graphene oxide: Synthesis, properties, and applications. *Advance Material* 22: 3906–3924