



UNIVERSITI PUTRA MALAYSIA

***ENZYMATIC ESTERIFICATION OF DIHYDROCAFFEIC ACID AND
EVALUATION OF ANTIOXIDANT ACTIVITIES OF THE
SYNTHESIZED ALKYL-ESTERS***

SOMAYEH GHOLIVAND

FSTM 2016 29



**ENZYMATIC ESTERIFICATION OF DIHYDROCAFFEIC ACID AND
EVALUATION OF ANTIOXIDANT ACTIVITIES OF THE
SYNTHESIZED ALKYL-ESTERS**

By

SOMAYEH GHOLIVAND

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Master of Science**

December 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

This thesis is dedicated to my family for nursing me with affections and love and their dedicated partnership for success in my life.



© COPYRIGHT UPM

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

**ENZYMATIC ESTERIFICATION OF DIHYDROCAFFEIC ACID AND
EVALUATION OF ANTIOXIDANT ACTIVITIES OF THE
SYNTHESIZED ALKYL-ESTERS**

By

SOMAYEH GHOLIVAND

December 2016

Chairman : Associate Professor Lasekan Olusegun Olaniyi, PhD
Faculty : Food Science and Technology

Solubility limitations of phenolic acids in many lipidic environments restrict their application as effective antioxidants. Enzymatic esterification of phenolic acids in a green media such as ionic liquid improves the lipophilicity of the resultant compounds. Herein, the investigation of a biocatalysts process for the esterification of dihydrocaffeic acid with different ionic liquids in order to improve the conversion of the phenolic acid in a shorter time and increase productivity of target product was carried out. To achieve the maximum conversion yield, the lipophilization of dihydrocaffeic acid (DHCA) by its enzymatic esterification with hexanol in the selected ionic liquid (as a model) was optimized by response surface methodology (RSM) using a five-level and four independent variables including: dosage of the enzyme (in relation to the total weight of substrates), hexanol/dihydrocaffeic acid mole ratio, reaction temperature and reaction time. The obtained optimal conditions were applied for the synthesis of the other n-alkyl esters with different chain lengths. Subsequently, the synthesized esters were separated from unreacted compounds and purified via a silica gel column. The purified alkyl-esters were identified by Fourier transform infrared (FTIR) and Nuclear magnetic resonance (NMR) analysis. Finally, the effects of alkyl chain length and concentration on the anti-oxidative properties of the resultant purified esters was investigated using β -carotene bleaching (BCB) and free radical scavenging method DPPH and also compared with butylated hydroxytoluene (BHT) as reference compound. According to the attained results, among ionic liquids tested, 1-butyl-3-methylimidazolium bis (trifluoromethylsulfonyl) imide was the best solvent with the highest bioconversion. Furthermore, results of the optimization showed that hexyl dihydrocaffeate (HDHCA) conversion yield was significantly ($p < 0.05$) affected by the linear, quadratic and the interaction effects of all the variables examined. Thus, an empirical equation was developed to describe and predict the variation of the response variable. The optimization procedure showed that the optimum molar conversion yield for hexyl dihydrocaffeate (HDHCA) ($Y_1 = 84.41\%$) was obtained using 41.6%

enzyme load at 39.4°C for 77.5 h and 9.3×10^{-4} mM hexanol to DHCA ratio. According to the obtained results from synthesized alkyl esters, both esters' concentration and their chain-length had significant ($p < 0.05$) effects on their antioxidant assays. Moreover, the scavenging activity of the tested compounds was methyl ester > hexyl ester > dodecyl ester > octadecyl ester > BHT while the order of *BCB* anti-oxidative activity was BHT > octadecyl ester > dodecyl ester > hexyl ester > methyl ester. Therefore, in the emulsion system, the hydrophobicity and solubility of the tested esters affected the antioxidant activity of esters which revealed that the antioxidant with hydrophobic character had better anti-oxidative activity in an emulsion system.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

PENGESTERAN ENZIM ASID DIHIDROKAFEIK DENGAN HEXANOL DI DALAM CECAIR IONIK YANG BERBEZA DAN PENILAIAN AKTIVITI ANTIOKSIDAN DARIPADA ALKIL-ESTER YANG DISINTESIS

Oleh

SOMAYEH GHOLIVAND

Disember 2016

Pengerusi : Profesor Madya Lasekan Olusegun Olaniyi, PhD
Fakulti : Sains dan Teknologi Makanan

Keterlarutan asid fenolik yang terhad di dalam pelbagai persekitaran berlipid telah menyekat pengaplikasiannya sebagai antioksidan yang efektif. Pengesteran enzim asid fenolik di dalam media mesra alam seperti cecair ionik meningkatkan kecenderungan lipofilik bagi sebatian yang dihasilkan. Oleh itu, proses pemangkin biologi tentang pengesteran asid dengan pelbagai jenis cecair ionic dikaji untuk menyinkatkan masa penukaran asid fenolik dan meningkatkan produktiviti produk sasaran. Untuk mencapai hasil penukaran maksimum, kecenderungan lipofilik asid dihidrokafeik (DHCA) oleh pengesteran dengan heksanol di dalam cecair ionik terbaik (1-butyl-3-methylimidazolium bis (trifluorometilsulfonil) imida, dalam kehadiran *Candida antarctica* lipase B (Novozyme 435) telah dioptimumkan dengan kaedah tindak balas permukaan (RSM) menggunakan lima peringkat dan empat pemboleh ubah bebas termasuk: dos enzim (berhubung dengan jumlah berat substrat); nisbah mol heksanol/asid dihidrokafeik; suhu tindak balas dan masa tindak balas. Syarat-syarat optimum yang diperolehi telah digunakan untuk sintesis ester n-alkil yang lain dengan panjang rantai yang berbeza. Selepas itu, ester yang tersintesis telah dipisahkan daripada sebatian yang tidak bertindak balas dan dituliskan melalui kolum gel silika. Alkil-ester tulen telah dikenal pasti menggunakan analisis transformasi Fourier infra merah (FTIR) dan resonans magnetik nuklear (NMR). Akhirnya, kesan kepanjangan rantai alkil pada sifat-sifat antioksidan ester tulen disiasat menggunakan pelunturan β -karotena (BCB) dan kaedah memerangkap radikal bebas DPPH dan dibandingkan dengan butylated hidrositoluen (BHT) sebagai sebatian rujukan. Selain itu, berdasarkan keputusan yang diperolehi, 1-butyl-3-methylimidazolium bis (trifluoromethylsulfonyl) imide ialah pelarut yang terbaik dengan kadar penukaran yang tertinggi antara cecair ionic yang diuji. Hasil pengoptimuman menunjukkan hasil penukaran heksil dihidrokafeik yang bersandar secara signifikan ($p < 0.05$) terhadap linear, kuadratik dan kesan interaksi semua pemboleh ubah yang diperiksa. Oleh itu, persamaan empirikal telah dibangunkan untuk menerang dan meramalkan perubahan pemboleh ubah bergerak balas. Prosedur pengoptimuman menunjukkan bahawa hasil penukaran molar optimum

untuk heksil dihidrokafeik ($Y_1 = 84,41\%$) telah diperoleh dengan menggunakan 41.6% beban enzim pada 39.4°C 77.5 jam dan 9.3×10^{-4} mM heksanol kepada nisbah DHCA. Berdasarkan keputusan yang diperoleh daripada sintesis ester alkil, kepekatan dan rantai panjang ester mempunyai kesan signifikan ($p < 0.05$) ke atas aktiviti antioksidan. Tambahan lagi, aktiviti memerangkap oleh sebatian ujian adalah metil ester > heksil ester \geq dodesil ester > oktadesil ester > BHT manakala susunan aktiviti BCB antioksidan adalah BHT > oktadesil ester > dodesil ester > heksil ester > metil ester. Oleh itu, di dalam sistem emulsi, sifat hidrofobik dan keterlarutan ester yang diuji telah berhubung kait dengan aktiviti antioksidan ester, yang mendedahkan bahawa antioksidan dengan karakter hidrofobik mempunyai aktiviti pengantioksidan yang lebih baik di dalam sistem emulsi.



ACKNOWLEDGEMENTS

I would first like to thank my thesis advisor Associate Professor Dr. Lasekan Olusegun Olaniyi of the Food Science and Technology faculty at Universiti Putra Malaysia who gave me lots of encouragement, enhanced my knowledge and any efforts that he had put from the start until the completion of my research work. Besides, I appreciate the experts who were involved in the validation investigation for this research project: Professor Tan Chin Ping, Associate Professor Dr. Faridah Abas and Dr. Leong Sze Wei. Without their passionate participation and input, the validation survey could not have been successfully conducted.

Secondly, I would also like to show my gratitude to my family especially to my parents who strongly supported and encouraged me to continue with my study by providing financial and good environment for me to focus in my master research.

Last but not least, I want to acknowledge all the staff of Faculty of Food Science and Technology (FSTM), Universiti Putra Malaysia who always take care of us and our needs in the lab.

I certify that a Thesis Examination Committee has met on 9 December 2016 to conduct the final examination of Somayeh Gholivand on her thesis entitled "Enzymatic Esterification of Dihydrocaffeic Acid and Evaluation of Antioxidant Activities of the Synthesized Alkyl-Esters" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Chong Gun Hean, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Chairman)

Anis Shobirin binti Meor Hussin, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Internal Examiner)

Nor Azah Mohamad Ali, PhD

Senior Lecturer
Forest Research Institute Malaysia
Malaysia
(External Examiner)



NOR AINI AB. SHUKOR, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 26 January 2017

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Lasekan Olusegun Olaniyi, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Chairman)

Tan Chin Ping, PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

Faridah Abas, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____ Date: _____

Name and Matric No.: Somayeh Gholivand / GS38273

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee:

Associate Professor Dr. Lasekan Olusegun Olaniyi

Signature: _____

Name of Member
of Supervisory
Committee:

Professor Dr. Tan Chin Ping

Signature: _____

Name of Member
of Supervisory
Committee:

Associate Professor Dr. Faridah Abas

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xiv
LIST OF ABBREVIATIONS	xvi
CHAPTER	
1 INTRODUCTION	1
1.1 Background	1
2 LITERATURE REVIEW	3
2.1 Phenolic acids	3
2.2 Classification of phenolic compounds	3
2.3 Antioxidants	5
2.4 Potential application of antioxidants	5
2.4.1 Applications of antioxidant in food products	6
2.4.2 Adding antioxidants to livestock diets	10
2.5 Antioxidants from natural sources	10
2.5.1 Antioxidant properties of phenolic acids	10
2.5.2 Caffeic and dihydrocaffeic acids	11
2.5.3 Sources of caffeic and dihydrocaffeic acids	13
2.6 Esterification of phenolic acids	16
2.7 Esterification by chemical methods	16
2.8 Enzymatic esterification of phenolic acids	18
2.8.1 Lipase as a biocatalyst:	18
2.6.2 Immobilization and modification of enzymes	20
2.9 Ionic liquids as green media	25
2.10 Enzymatic esterification of dihydrocaffeic acid	26
3 ENZYMATIC ESTERIFICATION OF DIHYDROCAFFEIC ACID WITH HEXANOL IN DIFFERENT IONIC LIQUIDS	29
3.1 Introduction	29
3.2 Materials and methods	30
3.2.1 Procedure of enzymatic esterification of dihydrocaffeic acid with hexanol in different ionic liquids	30
3.2.2 Procedure of enzymatic esterification of dihydrocaffeic acid in presence of different amounts of hexanol in the selected ionic liquid	31
3.2.3 Ester purification	31
3.2.4 Characterization of purified product	31
3.3 Results and discussion	32

3.3.1	Effect of ionic liquid structure on conversion yield	32
3.3.2	Effect of hexanol concentration on the conversion yield of hexyl- dihydrocaffeate	35
3.3.3	Purification and identification of hexyldihydrocaffeate	36
3.4	Conclusion	39
4	OPTIMIZATION OF ENZYMATIC ESTERIFICATION OF DIHYDROCAFFEIC ACID BY RESPONSE SURFACE METHODOLOGY IN THE SELECTED IONIC LIQUID	40
4.1	Introduction	40
4.2	Materials and methods	41
4.2.1	Materials	41
4.2.2	Experimental design	42
4.2.3	Statistical analysis	44
4.2.4	Optimization procedure	44
4.2.5	Model verification	45
4.2.6	Enzymatic esterification of DHCA with hexanol in the selected ionic liquid	45
4.2.7	HPLC Analysis	45
4.2.8	Calculation of degree of esterification	45
4.3	Results and discussion	46
4.3.1	Fitting the response surface model	46
4.3.2	Optimization procedure	50
4.3.3	Model verification	52
4.4	Conclusion	54
5	COMPARATIVE STUDY OF THE ANTIOXIDANT ACTIVITIES OF SOME SYNTHESIZED ALKYL DIHYDROCAFFEATES IN THE SELECTED IONIC LIQUID	55
5.1	Introduction	55
5.2	Materials and methods	56
5.2.1	Materials	56
5.2.2	Procedure of enzymatic synthesis of dihydrocaffeates with different chain-length alcohols in the selected ionic liquid	56
5.2.3	Ester purification	57
5.2.4	Identification of alkyl dihydrocaffeates	57
5.2.5	Determination of antioxidant activity with the β -carotene bleaching (<i>BCB</i>) test	57
5.2.6	Free radical scavenging method DPPH	58
5.2.7	Statistical analysis	58
5.3	Results and discussion	59
5.3.1	Esterification of dihydrocaffeic acid with different alcohols in the selected ionic liquid	59
5.3.2	Anti-oxidative property of alkyl dihydrocaffeate esters using free radical scavenging DPPH assay	60
5.3.3	Anti-oxidative property of alkyl dihydrocaffeate esters using β -carotene bleaching method	65

5.3.4	Characterization of the purified products	68
5.4	Conclusion	70
6	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	71
6.1	Conclusion	71
6.2	Recommendations for future research	72
	REFERENCES	73
	APPENDICES	101
	BIODATA OF STUDENT	135
	LIST OF PUBLICATIONS	136



LIST OF TABLES

Table		Page
2.1	Examples of antioxidants and their effects in different omega-3 enriched foods	8
2.2	Physical characters and application of some synthetic antioxidants	9
2.3	Total caffeic acid (CA) content in milligram per 100 gram of fresh weight beverage (Mean \pm SD)	13
2.4	Total caffeic acid (CA) content in milligram per 100 gram of fresh weight fruit (Mean \pm SD)	14
2.5	Total caffeic acid (CA) content in milligram per 100 gram of fresh weight berry (Mean \pm SD)	15
2.6	Enzymatic esterification of phenolic compounds reported in recent years	22
4.1	Levels of independent variables established according to central composite design for enzymatic esterification of hexyl dihydrocaffeate condition level (coded and, uncoded)	42
4.2	Matrix of central composite design (CCD)	43
4.3	Regression coefficient of the polynomial response surface model along with R ² and R ² (adjusted) for response variable	47
4.4	ANOVA and regression coefficient of the first and second degree polynomial regression model	49
4.5	Central composite design: factor (X _i), response variable (Y) and Residual	53
5.1	DPPH radical scavenging activity of alkyl dihydrocaffeate esters	62
5.2	The significance of each independent variable effect shown by F-ratio and P-value	64
5.3	Total antioxidant activity of alkyl dihydrocaffeate esters using β -carotene bleaching method	66
5.4	Representative FTIR spectra (cm ⁻¹) of dihydrocaffeic acid esters	69

LIST OF FIGURES

Figure		Page
2.1	Main classes of phenolic acids and chemical structures of the cinnamic acid derivatives	4
2.2	Oxidation of phenolic compounds	11
2.3	Proposed mechanism of the Fischer esterification	17
2.4	Alkylation of acidic functional groups	18
2.5	The reaction mechanism for lipase-catalyzed synthesis of cinnamic acid esters based on the known structure of the <i>C. antarctica</i> active center and, by analogy to other lipolytic reactions	20
3.1	(Scheme 1) Proposed esterification reaction of hexyl dihydrocaffeate	30
3.2	Effect of ionic liquid structure on degree of esterification (DHCA) after five days; IL1 (1-butyl-3-methylimidazolium bis (trifluoromethylsulfonyl) imide), IL2 (1-butyl-3-methylimidazoliumhexafluorophosphate), IL3 (1-hexyl-3-methylimidazoliumhexafluorophosphate), IL4 (1-octyl-3-methylimidazoliumhexafluorophosphate)	34
3.3	IL1 (1-butyl-3-methylimidazolium bis (trifluoromethylsulfonyl) imide), IL2 (1-butyl-3-methylimidazoliumhexafluorophosphate), IL3 (1-hexyl-3-methylimidazoliumhexafluorophosphate), IL4 (1-octyl-3-methylimidazoliumhexafluorophosphate). Other conditions: DHCA/hexanol = 1.4, 35 mg enzyme and 250 rpm at 55°C	35
3.4	Esterification of DHCA in 1-3 days by varying molar ratio of dihydrocaffeic acid to hexanol: solution 1(1:2), solution 2 (1:4), solution 3 (1:8) and solution 4 (1:16) keeping all other parameters constant (at temperature 55°C and 250rpm in the selected ionic liquid	36
3.5	HPLC chromatogram of the hexyldihydrocaffeate after purification	37
3.6	Fourier transforms infrared spectra of hexyl dihydrocaffeate synthesis	38
3.7	The crystal image (a) and NMR spectrum (b) of the purified hexyl dihydrocaffeate	39

4.1	The three-dimensional (3D) response surface plots	51
4.2	Numerical response optimizer	52
5.1	Degree of esterification(%) of alkyl dihydrocaffeate esters after 3 days	59
5.2	Decoloration of DPPH solution in the presence of methyl (row 1), hexyl (row 2), dodecyl (row 3), octadecyl (row 4) dihydrocaffeates and BHT (row 5) with concentrations ranging from 5 to 75 ($\mu\text{g/mL}$)	61
5.3	DPPH radical scavenging activity of alkyl dihydrocaffeate esters	63
5.4	Total antioxidant activity of alkyl dihydrocaffeate esters using β -carotene bleaching method	67

LIST OF ABBREVIATIONS

AA	Antioxidant Activity
AILs	Aprotic ionic liquids
ANOVA	Analysis of variance
BCB	β -Carotene Bleaching
BF ₄	Tetrafluoroborate
BHA	Butylated hydroxyanisole
BHT	Butylated hydroxytoluene
CA	Caffeic acid
CCD	Central composite design
CnMe ₃ N	Alkyltrimethylammonium
Cm	Centimetre
D	Desirability
DHCA	Dihydrocaffeic acid
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
DPPH	1, 1-diphenyl-1-picrylhydrazyl
DTGS	Deuterated Triglycine Sulphate
Emim (NTf ₂)	1-butyl-3-methylimidazoliumbis(trifluoromethylsulfonyl)imide
eNOS	Endothelial Nitric Oxide Synthase
FTIR	Fourier transform infrared
G	Gram
H	Hydrogen
HCAs	Hydroxycinnamic acids

HDHCA	Hexyl Dihydrocaffeate
HPLC	High Performance Liquid Chromatography
Hydro-PCs	Hydrophilic phenolic compounds
IL 1	1-butyl-3-methylimidazoliumbis(trifluoromethylsulfonyl)imide
IL 2	1-butyl-3-methylimidazoliumhexafluorophosphate
IL 3	1-hexyl-3-methylimidazolium hexafluorophosphate
IL 4	1-octyl-3-methylimidazolium hexafluorophosphate
ILs	Ionic liquids
IM	Imidazolium
LDL	Low-Density Lipoprotein
Lipo-PCs	Lipophilic Phenolic Compounds
μL	Microliter
μm	Micrometer
Mg	Milligram
MHz	Megahertz
Min	Minute
mL	Millilitre
mM	Milimolar
Mm	Millimeter
MTOA	Methyl trioctylammonium
NMR	Nuclear Magnetic Resonance
NTf ₂	Bis (trifluoromethylsulfonyl)imide
$^{\circ}\text{C}$	Celsius
OG	Octyl gallate

%	Percent
%T	Percent of Transmittance
<i>P</i>	Probability
PF ₆	Hexafluorophosphate
Ph•	Phenoxy radical
PhH	Phenolic antioxidant
PILs	Protic ionic liquids
PUFA	Polyunsaturated fatty acids
R ²	Regression
ROO•	Peroxy radical
ROOH	Hydro peroxide
RSM	Response Surface Methodology
TBHQ	Tert-butylated hydroquinone
TLA	Trilinolein
TLNA	Trilinolenin
TLC	Thin-Layer Chromatography
TY	Trans-esterification Yield
3D	Three-dimensional
UV	Ultraviolet
v/v	Volume per volume
w/w	Weight per weigh

CHAPTER 1

INTRODUCTION

1.1 Background

The several thousand phenolic compounds naturally present in almost all plant materials, including food products of plant origin (Psomiadou & Tsimidou, 2002). Interest in food phenolic has grown greatly because of their antioxidant and free radical scavenging ability and potential health benefits as well. Attention has increased mostly in finding naturally occurring antioxidants for use in foods or medicinal materials to replace synthetic antioxidants that, in some cases, have been reported to be carcinogenic (Zheng & Wang, 2001).

Among natural antioxidants, phenolic acids as well as their derivatives are the major subgroup of phenolics broadly dispersed in the plant kingdom such as vegetables, fruits and cereals. There has been on-going research attention in phenolic derivatives acid as the common constituents of plant tissues not only due to antioxidant effects but also due to the variety of several pharmacological and biological applications such as neuroprotective, anti-carcinogenic and anti-inflammatory abilities (Castelluccio et al., 1996; Laranjinha et al., 1994; Rice-Evans et al., 1996). In addition, some of the phenolic acids have been reported to possess UV-absorbing efficiency for application in sunscreen formulation (Compton et al., 2000). Consequently, phenolic acids have the potential to be suitable components in cosmetics, health, food and pharmaceuticals.

Most of the food systems are emulsion. In general, due to the hydrophilic character of phenolic compounds, they will most probably be found in the aqueous phase (Frankel et al., 1994; Porter, 1993). One major barrier in the functional properties of phenolic acids, is their hydrophilicity character in non-polar media, emulsions and oil-based formulae which restricts their application in lipophilic formulation of food processing and cosmetic industries due to the reduction of their antioxidant capabilities in these systems (Figuroa-Espinoza & Villeneuve, 2005; Lue et al., 2005).

A practical approach to address the solubility limitation of phenolic acids is lipophilization through esterifying their carboxylic acid group with aliphatic side-chain groups (Buisman et al., 1998; Lue et al., 2005; Sabally et al., 2005b; Stamatis et al., 2001) or trans-esterification with triacylglycerols (Compton et al., 2000; Laszlo et al., 2003) so that it retains its original functional properties. Furthermore, conventional esterification such as chemical processes can be applied for synthesizing lipophilic compounds from phenolic acids; however, their disadvantages are loss of product yield due to oxidation under certain pH conditions besides the susceptibility of phenolics to heating. Thus, the various new synthesis

pathways are needed to utilize the esterification of natural antioxidants. As an alternative, enzyme-catalyzed biosynthesis of phenolic esters in ionic liquids or organic solvents is known as a suitable approach for esterification of phenolic compounds owing to its capability of product separation and recovering the used enzyme (Auerbach et al., 1998; Jain et al., 2005).

Enzymatic esterification of phenolic acid (using lipase) with short or medium chain length aliphatic alcohols has been studied (Buisman et al., 1998; Stamatis et al., 2001). A lipase-catalyzed reaction has been mostly applied in organic solvent medium in order to synthesize valuable esters under mild conditions with the purpose of their applications in food industries (Auerbach et al., 1998; Gandhi et al., 2000). The chief reason for the inefficiency of this kind of lipolization is low solubility of phenolic acids in most organic reaction solvents whereas the enzyme is active and *vice versa* (Eastoe et al., 2005).

Ionic liquids are able to give different reactivity and selectivity to various biocatalytic reactions as a medium and also can dissolve numerous compounds due to existence of some properties compared with conventional organic solvents. In fact, their structures are composed of bulky cation and a small anion. The properties of ionic liquids can be easily altered by combination of different cations and anions as well. Besides, they possess no vapor pressures and entail a wide-ranging temperature for the liquid phase. Consequently, synthesis of plentiful ionic liquids with different compositions is possible (Jain et al., 2005). Many studies have been done on enzymatic transformation of various phenolic acids in several ionic liquids as a replacement reaction media with organic solvents (Jain et al., 2005; Kurata et al., 2010; Yang et al., 2012b).

Dihydrocaffeic acid (DHCA) is a hydrophilic compound and has low solubility in oil based foods. Thus, its modification via esterification with alcohols can be used as a tool to produce derivatives with increased lipophilicity and antioxidant potency. In comparison with caffeic acid, little information is presented about the anti-oxidative activity of dihydrocaffeic acid and its derivatives. Therefore, the specific objectives of this study were:

1. To investigate the enzymatic esterification of dihydrocaffeic acid (DHCA) and hexanol in different ionic liquids;
2. To optimize the enzymatic esterification conditions of dihydrocaffeic acid and hexanol in selected ionic liquid (NTf₂) by response surface methodology (RSM); and
3. To evaluate the antioxidant activity of some synthesized alkyl dihydrocaffeic acid esters in the selected ionic liquid.

REFERENCES

- Akarpat, A., Turhan, S., & Ustun, N. (2008). Effects of hot-water extracts from myrtle, rosemary, nettle and lemon balm leaves on lipid oxidation and color of beef patties during frozen storage. *Journal of Food Processing and Preservation*, 32(1), 117-132.
- Andjelkovic, M., Van Camp, J., De Meulenaer, B., Depaemelaere, G., Socaciu, C., Verloo, M., & Verhe, R. (2006). Iron-chelation properties of phenolic acids bearing catechol and galloyl groups. *Food Chemistry*, 98(1), 23-31.
- Araujo, M. E. M. B. D., Contesini, F. J., Franco, Y. E. M., Sawaya, A. C., Alberto, T. G., Dalfre, N., & Carvalho, P. D. O. (2011). Optimized enzymatic synthesis of hesperidin fatty acid esters in a two-phase system containing ionic liquid. *Molecules*, 16(8), 7171-7182.
- Ardhaoui, M., Falcimaigne, A., Ognier, S., Engasser, J., Moussou, P., Pauly, G., & Ghoul, M. (2004). Effect of acyl donor chain length and substitutions pattern on the enzymatic acylation of flavonoids. *Journal of Biotechnology*, 110(3), 265-272.
- Aruoma, O. I., & Halliwell, B. (1991). *Free Radicals and Food Additives*: Taylor & Francis Ltd, London, 129-150.
- Auerbach, M., Chang, P., Kosmark, R., O'Neill, J., & Philips, J. (1998). Salatrim: A family of reduced-calorie structured lipids. *Structurally Modified Food Fats: Synthesis, Biochemistry, and Use*, Illinois: AOCS Press, Champaign, 89-116.
- Balasundram, N., Sundram, K., & Samman, S. (2006). Phenolic compounds in plants and agri-industrial by-products: Antioxidant activity, occurrence, and potential uses. *Food Chemistry*, 99(1), 191-203.
- Baltazar, Q. Q., Chandawalla, J., Sawyer, K., & Anderson, J. L. (2007). Interfacial and micellar properties of imidazolium-based monocationic and dicationic ionic liquids. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 302(1), 150-156.
- Barlow, S. M. (1990). Toxicological aspects of antioxidants used as food additives *Food Antioxidants*, Ed by Hudson BJB, Elsevier, New York, USA, 253-307
- Bas, D., & Boyacı, İ. H. (2007). Modeling and optimization I: Usability of response surface methodology. *Journal of Food Engineering*, 78(3), 836-845.
- Bassil, D., Makris, D. P., & Kefalas, P. (2005). Oxidation of caffeic acid in the presence of L-cysteine: isolation of 2-S-cysteinylcaffeic acid and evaluation of its antioxidant properties. *Food Research International*, 38(4), 395-402.

- Basso, A., Braiuca, P., Cantone, S., Ebert, C., Linda, P., Spizzo, P., Gardossi, L. (2007). In Silico analysis of enzyme surface and glycosylation effect as a tool for efficient covalent immobilisation of CalB and PGA on Sepabeads®. *Advanced Synthesis and Catalysis*, 349(6), 877-886.
- Bertelli, D., Plessi, M., & Miglietta, F. (2004). Effect of industrial microwave treatment on the antioxidant activity of herbs and spices. *Italian journal of Food Science*, 16(1).
- Bezbradica, D., Mijin, D., Siler-Marinkovic, S., & Knezevic, Z. (2007). The effect of substrate polarity on the lipase-catalyzed synthesis of aroma esters in solvent-free systems. *Journal of Molecular Catalysis B: Enzymatic*, 45(3), 97-101.
- Bhullar, K. S., Warnakulasuriya, S. N., & Rupasinghe, H. V. (2013). Biocatalytic synthesis, structural elucidation, antioxidant capacity and tyrosinase inhibition activity of long chain fatty acid acylated derivatives of phloridzin and isoquercitrin. *Bioorganic and Medicinal Chemistry*, 21(3), 684-692.
- Boler, D. D., Gabriel, S., Yang, H., Balsbaugh, R., Mahan, D., Brewer, M., . . . Killefer, J. (2009). Effect of different dietary levels of natural-source vitamin E in grow-finish pigs on pork quality and shelf life. *Meat Science*, 83(4), 723-730.
- Borneman, W. S., Hartley, R. D., Morrison, W. H., Akin, D. E., & Ljungdahl, L. G. (1990). Feruloyl and p-coumaroyl esterase from anaerobic fungi in relation to plant cell wall degradation. *Applied Microbiology and Biotechnology*, 33(3), 345-351.
- Box, G. E., & Wilson, K. (1951). On the experimental attainment of optimum conditions. *Journal of the Royal Statistical Society. Series B (Methodological)*, 13(1), 1-45.
- Braca, A., De Tommasi, N., Di Bari, L., Pizza, C., Politi, M., & Morelli, I. (2001). Antioxidant Principles from *Bauhinia t arapotensis*. *Journal of Natural Products*, 64(7), 892-895.
- Brannan, R. (2008). Effect of grape seed extract on physicochemical properties of ground, salted, chicken thigh meat during refrigerated storage at different relative humidity levels. *Journal of Food Science*, 73(1), 36-40.
- Bravo, L. (1998). Polyphenols: chemistry, dietary sources, metabolism, and nutritional significance. *Nutrition Reviews*, 56(11), 317-333.
- Brewer, M. (2011). Natural antioxidants: sources, compounds, mechanisms of action, and potential applications. *Comprehensive Reviews in Food Science and Food Safety*, 10(4), 221-247.
- Budryn, G., & Nebesny, E. (2005). The structure and properties of antioxidant polyphenols of cocoa beans. *Bromatology and Toxicological Chemistry*, 3 (38), 203-209.

- Buisman, G., Van Helteren, C., Kramer, G., Veldsink, J., Derksen, J., & Cuperus, F. (1998). Enzymatic esterifications of functionalized phenols for the synthesis of lipophilic antioxidants. *Biotechnology Letters*, 20(2), 131-136.
- Burke, T. R. J., Fesen, M., Mazumder, A., Yung, J., Wang, J., Carothers, A. M., Kohn, K. (1995). Hydroxylated aromatic inhibitors of HIV-1 integrase. *Journal of Medicinal Chemistry*, 38(21), 4171-4178.
- Cabrera, C., Artacho, R., & Gimenez, R. (2006). Beneficial effects of green tea: a review. *Journal of the American College of Nutrition*, 25(2), 79-99.
- Cassani, J., Luna, H., Navarro, A., & Castillo, E. (2007). Comparative esterification of phenylpropanoids versus hydrophenylpropanoids acids catalyzed by lipase in organic solvent media. *Electronic Journal of Biotechnology*, 10(4), 508-513.
- Castelluccio, C., Bolwell, G. P., Gerrish, C., & Catherine, R.-E. (1996). Differential distribution of ferulic acid to the major plasma constituents in relation to its potential as an antioxidant. *Biochemical Journal*, 316(2), 691-694.
- Celiz, G., & Daz, M. (2011). Biocatalytic preparation of alkyl esters of citrus flavanone glucoside prunin in organic media. *Process Biochemistry*, 46(1), 94-100.
- Chalas, J., Claise, C., Edeas, M., Messaoudi, C., Vergnes, L., Abella, A., & Lindenbaum, A. (2001). Effect of ethyl esterification of phenolic acids on low-density lipoprotein oxidation. *Biomedicine & Pharmacotherapy*, 55(1), 54-60.
- Chandra, K. L., Saravanan, P., Singh, R. K., & Singh, V. K. (2002). Lewis acid catalyzed acylation reactions: scope and limitations. *Tetrahedron*, 58(7), 1369-1374.
- Chen, C. P., Yokozawa, T., & Chung, H. Y. (1999). Inhibitory effect of caffeic acid analogues isolated from *Salviae Miltiorrhizae Radix* against 1, 1-diphenyl-2-picrylhydrazyl radical. *Experimental and Toxicologic Pathology*, 51(1), 59-63.
- Chen, H.-C., Twu, Y.-K., Chieh-ming, J. C., Liu, Y.-C., & Shieh, C.-J. (2010). Optimized synthesis of lipase-catalyzed octyl caffeate by Novozym® 435. *Industrial Crops and Products*, 32(3), 522-526.
- Chen, J.-H., Shao, Y., Huang, M.-T., Chin, C.-K., & Ho, C.-T. (1996). Inhibitory effect of caffeic acid phenethyl ester on human leukemia HL-60 cells. *Cancer Letters*, 108(2), 211-214.
- Chen, J. H., & Ho, C.-T. (1997). Antioxidant activities of caffeic acid and its related hydroxycinnamic acid compounds. *Journal of Agricultural and Food Chemistry*, 45(7), 2374-2378.

- Chigorimbo-Murefu, N. T., Riva, S., & Burton, S. G. (2009). Lipase-catalysed synthesis of esters of ferulic acid with natural compounds and evaluation of their antioxidant properties. *Journal of Molecular Catalysis B: Enzymatic*, 56(4), 277-282.
- Chimi, H., Cillard, J., Cillard, P., & Rahmani, M. (1991). Peroxyl and hydroxyl radical scavenging activity of some natural phenolic antioxidants. *Journal of the American Oil Chemists Society*, 68(5), 307-312.
- Choi, H. R., Choi, J. S., Han, Y. N., Bae, S. J., & Chung, H. Y. (2002). Peroxynitrite scavenging activity of herb extracts. *Phytotherapy Research*, 16(4), 364-367.
- Clifford, M. N. (1999). Chlorogenic acids and other cinnamates—nature, occurrence and dietary burden¹. *Journal of the Science of Food and Agriculture*, 79, 362-372.
- Compton, D. L., Laszlo, J. A., & Berhow, M. A. (2000). Lipase-catalyzed synthesis of ferulate esters. *Journal of the American Oil Chemists' Society*, 77(5), 513-519.
- Cornellas, A., Perez, L., Comelles, F., Ribosa, I., Manresa, A., & Garcia, M. T. (2011). Self-aggregation and antimicrobial activity of imidazolium and pyridinium based ionic liquids in aqueous solution. *Journal of Colloid and Interface Science*, 355(1), 164-171.
- Cos, P., Bruyne, T., Hermans, N., Apers, S., Berghe, D. V., & Vlietinck, A. (2004). Proanthocyanidins in health care: current and new trends. *Current Medicinal Chemistry*, 11(10), 1345-1359.
- Cos, P., Ying, L., Calomme, M., Hu, J. P., Cimanga, K., Van Poel, B., Berghe, D. V. (1998). Structure-activity relationship and classification of flavonoids as inhibitors of xanthine oxidase and superoxide scavengers. *Journal of Natural Products*, 61(1), 71-76.
- Couling, D. J., Bernot, R. J., Docherty, K. M., Dixon, J. K., & Maginn, E. J. (2006). Assessing the factors responsible for ionic liquid toxicity to aquatic organisms via quantitative structure–property relationship modeling. *Green Chemistry*, 8(1), 82-90.
- Croft, K. D. (1998). The chemistry and biological effects of flavonoids and phenolic acids. *Annals of the New York Academy of Sciences*, 854(1), 435-442.
- Cuppert, S., Schnepf, M., & Hall, C. (1997). Natural antioxidant—are they a reality? *Natural Antioxidants: Chemistry, Health Effects, and Applications*: AOCS Press, Champaign. IL, 12–24.
- Cuvelier, M.-E., Richard, H., & Berset, C. (1992). Comparison of the antioxidative activity of some acid-phenols: structure-activity relationship. *Bioscience, Biotechnology, and Biochemistry*, 56(2), 324-325.

- De Diego, T., Lozano, P., Gmouh, S., Vaultier, M., & Iborra, J. L. (2005). Understanding Structure-Stability Relationships of *Candida antarctica* Lipase B in Ionic Liquids. *Biomacromolecules*, 6(3), 1457-1464.
- Decker, E. (1998). Strategies for manipulating the prooxidative/antioxidative balance of foods to maximize oxidative stability. *Trends in Food Science & Technology*, 9(6), 241-248.
- Deprez, S., Brezillon, C., Rabot, S., Philippe, C., Mila, I., Lapierre, C., & Scalbert, A. (2000). Polymeric proanthocyanidins are catabolized by human colonic microflora into low-molecular-weight phenolic acids. *The Journal of Nutrition*, 130(11), 2733-2738.
- Dewick, P. M. (2002). *Medicinal Natural Products: a Biosynthetic Approach*: John Wiley & Sons, New York, USA.
- Eastoe, J., Gold, S., Rogers, S. E., Paul, A., Welton, T., Heenan, R. K., & Grillo, I. (2005). Ionic liquid-in-oil microemulsions. *Journal of the American Chemical Society*, 127(20), 7302-7303.
- Eckstein, M., Sesing, M., Kragl, U., & Adlercreutz, P. (2002). At low water activity α -chymotrypsin is more active in an ionic liquid than in non-ionic organic solvents. *Biotechnology Letters*, 24(11), 867-872.
- Eckstein, M., Wasserscheid, P., & Kragl, U. (2002). Enhanced enantioselectivity of lipase from *Pseudomonas* sp. at high temperatures and fixed water activity in the ionic liquid, 1-butyl-3-methylimidazolium bis [(trifluoromethyl) sulfonyl] amide. *Biotechnology Letters*, 24(10), 763-767.
- Estevez, M., Kylli, P., Puolanne, E., Kivikari, R., & Heinonen, M. (2008). Oxidation of skeletal muscle myofibrillar proteins in oil-in-water emulsions: Interaction with lipids and effect of selected phenolic compounds. *Journal of Agricultural and Food Chemistry*, 56(22), 10933-10940.
- Etzenhouser, B., Hansch, C., Kapur, S., & Selassie, C. D. (2001). Mechanism of toxicity of esters of caffeic and dihydrocaffeic acids. *Bioorganic & Medicinal Chemistry*, 9(1), 199-209.
- Fasseas, M., Mountzouris, K., Tarantilis, P., Polissiou, M., & Zervas, G. (2008). Antioxidant activity in meat treated with oregano and sage essential oils. *Food Chemistry*, 106(3), 1188-1194.
- Feddern, V., Yang, Z., Xu, X., Badiale-Furlong, E., & de Souza-Soares, L. A. (2011). Synthesis of octyl dihydrocaffeate and its transesterification with tricaprylin catalyzed by *Candida antarctica* lipase. *Industrial & Engineering Chemistry Research*, 50(12), 7183-7190.
- Figuroa-Espinoza, M.-C., & Villeneuve, P. (2005). Phenolic acids enzymatic lipophilization. *Journal of Agricultural and Food Chemistry*, 53(8), 2779-2787.

- Finkle, B. J., Lewis, J., Corse, J., & Lundin, R. (1962). Enzyme reactions with phenolic compounds: formation of hydroxystyrenes through the decarboxylation of 4-hydroxycinnamic acids by *Aerobacter*. *Journal of Biological Chemistry*, 237(9), 2926-2931.
- Fischer, E., & Speier, A. (1924). Darstellung der ester. *Untersuchungen aus Verschiedenen Gebieten*, 285-291.
- Floros, J., & Chinnan, M. (1988). Computer graphics- assisted optimization for product and process development. *Food Technology*, 42(2), 72-78.
- Forsyth, S. A., & MacFarlane, D. R. (2003). 1-Alkyl-3-methylbenzotriazolium salts: ionic solvents and electrolytes. *Journal of Materials Chemistry*, 13(10), 2451-2456.
- Frankel, E. N., Huang, S.-W., Kanner, J., & German, J. B. (1994). Interfacial phenomena in the evaluation of antioxidants: bulk oils vs emulsions. *Journal of Agricultural and Food Chemistry*, 42(5), 1054-1059.
- Frankel, E. N., & Meyer, A. S. (2000). The problems of using one-dimensional methods to evaluate multifunctional food and biological antioxidants. *Journal of the Science of Food and Agriculture*, 80(13), 1925-1941.
- Fresco, P., Borges, F., Diniz, C., & Marques, M. (2006). New insights on the anticancer properties of dietary polyphenols. *Medicinal Research Reviews*, 26(6), 747-766.
- Friedman, M. (1997). Chemistry, biochemistry, and dietary role of potato polyphenols. A review. *Journal of Agricultural and Food Chemistry*, 45(5), 1523-1540.
- Fukumoto, L., & Mazza, G. (2000). Assessing antioxidant and prooxidant activities of phenolic compounds. *Journal of Agricultural and Food Chemistry*, 48(8), 3597-3604.
- Furniss, B. S. (1989). Vogel's textbook of practical organic chemistry: Pearson Education India, 5th Edn.; Wiley, New York, USA.
- Gallagher, E., O'Brien, C., Scannell, A., & Arendt, E. (2003). Use of response surface methodology to produce functional short dough biscuits. *Journal of Food Engineering*, 56(2), 269-271.
- Galonde, N., Nott, K., Richard, G., Debuigne, A., Nicks, F., Jerome, C., & Fauconnier, M.-L. (2013). Study of the influence of pure ionic liquids on the lipase-catalyzed (trans) esterification of mannose based on their anion and cation nature. *Current Organic Chemistry*, 17(7), 763-770.
- Gandhi, N. N., Patil, N. S., Sawant, S. B., Joshi, J. B., Wangikar, P. P., & Mukesh, D. (2000). Lipase-catalyzed esterification. *Catalysis Reviews*, 42(4), 439-480.

- Gauthier, L., Verdal, M.-N., Marchegay, G., Pinson-Gadais, L., Ducos, C., Richard-Forget, F., & Atanasova-Penichon, V. (2016). Fungal biotransformation of chlorogenic and caffeic acids by *Fusarium graminearum*: New insights in the contribution of phenolic acids to resistance to deoxynivalenol accumulation in cereals. *International Journal of Food Microbiology*.
- Gayot, S., Santarelli, X., & Coulon, D. (2003). Modification of flavonoid using lipase in non-conventional media: effect of the water content. *Journal of Biotechnology*, *101*(1), 29-36.
- Giovanelli, G., & Buratti, S. (2009). Comparison of polyphenolic composition and antioxidant activity of wild Italian blueberries and some cultivated varieties. *Food Chemistry*, *112*(4), 903-908.
- Giovanni, M. (1983). Response surface methodology and product optimization. *Food Technology*, *38*, 41–101
- Giuliani, S., Piana, C., Setti, L., Hochkoeppler, A., Pifferi, P., Williamson, G., & Faulds, C. (2001). Synthesis of pentylferulate by a feruloyl esterase from *Aspergillus niger* using water-in-oil microemulsions. *Biotechnology Letters*, *23*(4), 325-330.
- Gobert, M., Gruffat, D., Habeanu, M., Parafita, E., Bauchart, D., & Durand, D. (2010). Plant extracts combined with vitamin E in PUFA-rich diets of cull cows protect processed beef against lipid oxidation. *Meat Science*, *85*(4), 676-683.
- Gocer, H., & Gülçin, İ. (2011). Caffeic acid phenethyl ester (CAPE): correlation of structure and antioxidant properties. *International Journal of Food Sciences and Nutrition*, *62*(8), 821-825.
- Goldstein, D. S., Stull, R., Markey, S. P., Marks, E. S., & Keiser, H. R. (1984). Dihydrocaffeic acid: a common contaminant in the liquid chromatographic-electrochemical measurement of plasma catecholamines in man. *Journal of Chromatography B: Biomedical Sciences and Applications*, *311*, 148-153.
- Gonthier, M.-P., Cheynier, V., Donovan, J. L., Manach, C., Morand, C., Mila, I., Scalbert, A. (2003a). Microbial aromatic acid metabolites formed in the gut account for a major fraction of the polyphenols excreted in urine of rats fed red wine polyphenols. *The Journal of Nutrition*, *133*(2), 461-467.
- Gonthier, M.-P., Donovan, J. L., Texier, O., Felgines, C., Remesy, C., & Scalbert, A. (2003b). Metabolism of dietary procyanidins in rats. *Free Radical Biology and Medicine*, *35*(8), 837-844.
- Gonthier, M.-P., Remesy, C., Scalbert, A., Cheynier, V., Souquet, J.-M., Poutanen, K., & Aura, A.-M. (2006). Microbial metabolism of caffeic acid and its esters chlorogenic and caftaric acids by human faecal microbiota in vitro. *Biomedicine & Pharmacotherapy*, *60*(9), 536-540.

- Graf, E. (1992). Antioxidant potential of ferulic acid. *Free Radical Biology and Medicine*, 13(4), 435-448.
- Greaves, T. L., & Drummond, C. J. (2008). Protic ionic liquids: properties and applications. *Chemical Reviews*, 108(1), 206-237.
- Groenewoud, G., & Hundt, H. (1986). The microbial metabolism of condensed (+)-catechins by rat-caecal microflora. *Xenobiotica*, 16(2), 99-107.
- Gryglewski, R. J., Korbut, R., Robak, J., & Swies, J. (1987). On the mechanism of antithrombotic action of flavonoids. *Biochemical Pharmacology*, 36(3), 317-322.
- Gutfinger, T. (1981). Polyphenols in olive oils. *Journal of the American Oil Chemists Society*, 58(11), 966-968.
- Guyot, B., Bosquette, B., Pina, M., & Graille, J. (1997). Esterification of phenolic acids from green coffee with an immobilized lipase from *Candida antarctica* in solvent-free medium. *Biotechnology Letters*, 19(6), 529-532.
- Ha, S. H., Van Anh, T., Lee, S. H., & Koo, Y.-M. (2012). Effect of ionic liquids on enzymatic synthesis of caffeic acid phenethyl ester. *Bioprocess and Biosystems Engineering*, 35(1-2), 235-240.
- Haerens, K., Matthijs, E., Chmielarz, A., & Van der Bruggen, B. (2009). The use of ionic liquids based on choline chloride for metal deposition: A green alternative? *Journal of Environmental Management*, 90(11), 3245-3252.
- Hakkinen, S. (2000). Flavonols and phenolic acids in berries and berry products (Doctoral dissertation, University of Kuopio) Publications Finland.
- Halliwell, B. (1990). How to characterize a biological antioxidant. *Free Radical Research Communications*, 9(1), 1-32.
- Hatzidimitriou, E., Nenadis, N., & Tsimidou, M. Z. (2007). Changes in the catechin and epicatechin content of grape seeds on storage under different water activity (a_w) conditions. *Food Chemistry*, 105(4), 1504-1511.
- Heim, K. E., Tagliaferro, A. R., & Bobilya, D. J. (2002). Flavonoid antioxidants: chemistry, metabolism and structure-activity relationships. *The Journal of Nutritional Biochemistry*, 13(10), 572-584.
- Herrman, V. (1956). Uber Kaffeesaure and chlorogensaure. *Pharmazie*, 11, 433-449.
- Herrmann, K., & Nagel, C. W. (1989). Occurrence and content of hydroxycinnamic and hydroxybenzoic acid compounds in foods. *Critical Reviews in Food Science & Nutrition*, 28(4), 315-347.

- Hettiarachchy, N., Glenn, K., Gnanasambandam, R., & Johnson, M. (1996). Natural antioxidant extract from fenugreek (*Trigonella foenumgraecum*) for ground beef patties. *Journal of Food Science*, 61(3), 516-519.
- Horning, M. G., Boucher, E. A., & Moss, A. M. (1967). The study of urinary acids and related compounds by gas phase analytical methods. *Journal of Chromatographic Science*, 5(6), 297-302.
- Hou, X. D., Li, N., & Zong, M. H. (2013). Renewable bio ionic liquids-water mixtures-mediated selective removal of lignin from rice straw: Visualization of changes in composition and cell wall structure. *Biotechnology and Bioengineering*, 110(7), 1895-1902.
- Hough, W. L., & Rogers, R. D. (2007). Ionic liquids then and now: from solvents to materials to active pharmaceutical ingredients. *Bulletin of the Chemical Society of Japan*, 80(12), 2262-2269.
- Hsieh, H.-J., Nair, G. R., & Wu, W.-T. (2006). Production of ascorbyl palmitate by surfactant-coated lipase in organic media. *Journal of Agricultural and Food Chemistry*, 54(16), 5777-5781.
- Hu, Y., Guo, Z., Lue, B.-M., & Xu, X. (2009). Enzymatic synthesis of esculin ester in ionic liquids buffered with organic solvents. *Journal of Agricultural and Food Chemistry*, 57(9), 3845-3852.
- Huang, H. M., Johanning, G. L., & O'Dell, B. L. (1986). Phenolic acid content of food plants and possible nutritional implications. *Journal of Agricultural and Food Chemistry*, 34(1), 48-51.
- Huang, J., de Paulis, T., & May, J. M. (2004). Antioxidant effects of dihydrocaffeic acid in human EA. hy926 endothelial cells. *The Journal of Nutritional Biochemistry*, 15(12), 722-729.
- Huang, W.-Y., Cai, Y.-Z., & Zhang, Y. (2009). Natural phenolic compounds from medicinal herbs and dietary plants: potential use for cancer prevention. *Nutrition and Cancer*, 62(1), 1-20.
- Hudson, B. J., & Mahgoub, S. E. (1980). Naturally-occurring antioxidants in leaf lipids. *Journal of the Science of Food and Agriculture*, 31(7), 646-650.
- Ingraham, L. L., & Corse, J. (1951). Enzymatic browning of fruits. I. Autoxidation of chlorogenic acid. *Journal of the American Chemical Society*, 73(12), 5550-5553.
- Ishihara, K., Katsube, Y., Kumazawa, N., Kuratani, M., Masuoka, N., & Nakajima, N. (2010). Enzymatic preparation of arbutin derivatives: lipase-catalyzed direct acylation without the need of vinyl ester as an acyl donor. *Journal of Bioscience and Bioengineering*, 109(6), 554-556.

- Ito, N. (1982). Induction of squamous cell carcinoma in the forestomach of F344 rats treated with butylated hydroxyanisole. *Cancer*, 73(2), 332-334.
- Itoh, T., Akasaki, E., Kudo, K., & Shirakami, S. (2001). Lipase-Catalyzed Enantioselective Acylation in the Ionic Liquid Solvent System. Reaction of Enzyme Anchored to the Solvent. *Chemistry Letters*(3), 262-263.
- Itzstein, M. (2002). Rapid, clean, and mild O-acetylation of alcohols and carbohydrates in an ionic liquid. *Chemical Communications* (7), 714-715.
- Jacobsen, C., Adler-Nissen, J., & Meyer, A. S. (1999). Effect of ascorbic acid on iron release from the emulsifier interface and on the oxidative flavor deterioration in fish oil enriched mayonnaise. *Journal of Agricultural and Food Chemistry*, 47(12), 4917-4926.
- Jacobsen, C., Hartvigsen, K., Lund, P., Thomsen, M. K., Skibsted, L. H., Hølmer, G., . Meyer, A. S. (2001). Oxidation in fish oil-enriched mayonnaise: 4. Effect of tocopherol concentration on oxidative deterioration. *European Food Research and Technology*, 212(3), 308-318.
- Jacobsen, C., Hartvigsen, K., Thomsen, M. K., Hansen, L. F., Lund, P., Skibsted, L. H., Meyer, A. S. (2001). Lipid oxidation in fish oil enriched mayonnaise: calcium disodium ethylenediaminetetraacetate, but not gallic acid, strongly inhibited oxidative deterioration. *Journal of Agricultural and Food Chemistry*, 49(2), 1009-1019.
- Jacobsen, C., & Nielsen, N. S. (2007). Optimization of oxidative stability of omega-3 enriched foods *Long-Chain Omega-3 Specialty Oils*: Oily Press.
- Jacobson, E., Newmark, H., Baptista, J., & Bruce, W. (1983). A preliminary investigation of the metabolism of dietary phenolics in humans [Urinary metabolites of caffeic and ferulic acid]. *Nutrition Reports International*, 28, 1409-1417.
- Jadhav, A. H., Lee, K., Koo, S., & Seo, J. G. (2015). Esterification of carboxylic acids with alkyl halides using imidazolium based dicationic ionic liquids containing bis-trifluoromethane sulfonimide anions at room temperature. *RSC Advances*, 5(33), 26197-26208.
- Jain, N., Kumar, A., Chauhan, S., & Chauhan, S. (2005). Chemical and biochemical transformations in ionic liquids. *Tetrahedron*, 61(5), 1015-1060.
- Jayaprakasam, B., Vanisree, M., Zhang, Y., Dewitt, D. L., & Nair, M. G. (2006). Impact of alkyl esters of caffeic and ferulic acids on tumor cell proliferation, cyclooxygenase enzyme, and lipid peroxidation. *Journal of Agricultural and Food Chemistry*, 54(15), 5375-5381.
- Jenner, A. M., Rafter, J., & Halliwell, B. (2005). Human fecal water content of phenolics: the extent of colonic exposure to aromatic compounds. *Free Radical Biology and Medicine*, 38(6), 763-772.

- Joglekar, A., & May, A. (1987). Product excellence through design of experiments. *Cereal Foods World*, 32(12), 857.
- Kaar, J. L., Jesionowski, A. M., Berberich, J. A., Moulton, R., & Russell, A. J. (2003). Impact of ionic liquid physical properties on lipase activity and stability. *Journal of the American Chemical Society*, 125(14), 4125-4131.
- Kamori, M., Hori, T., Yamashita, Y., Hirose, Y., & Naoshima, Y. (2000). Immobilization of lipase on a new inorganic ceramics support, toyonite, and the reactivity and enantioselectivity of the immobilized lipase. *Journal of Molecular Catalysis B: Enzymatic*, 9(4), 269-274.
- Kandaswami, C., & Middleton Jr, E. (1994). Free radical scavenging and antioxidant activity of plant flavonoids. *Free Radicals in Diagnostic Medicine*, 336, 351-376.
- Khani, R., & Shemirani, F. (2013). Simultaneous determination of trace amounts of cobalt and nickel in water and food samples using a combination of partial least squares method and dispersive liquid-liquid microextraction based on ionic liquid. *Food Analytical Methods*, 6(2), 386-394.
- Kapoor, M., & Gupta, M. N. (2012). Lipase promiscuity and its biochemical applications. *Process Biochemistry*, 47(4), 555-569.
- Katsoura, M., Polydera, A., Katapodis, P., Kolisis, F., & Stamatis, H. (2007). Effect of different reaction parameters on the lipase-catalyzed selective acylation of polyhydroxylated natural compounds in ionic liquids. *Process Biochemistry*, 42(9), 1326-1334.
- Katsoura, M., Polydera, A., Tsironis, L., Tselepis, A., & Stamatis, H. (2006). Use of ionic liquids as media for the biocatalytic preparation of flavonoid derivatives with antioxidant potency. *Journal of Biotechnology*, 123(4), 491-503.
- Katsoura, M. H., Polydera, A. C., Tsironis, L. D., Petraki, M. P., Rajacic, S. K., Tselepis, A. D., & Stamatis, H. (2009). Efficient enzymatic preparation of hydroxycinnamates in ionic liquids enhances their antioxidant effect on lipoproteins oxidative modification. *New Biotechnology*, 26(1), 83-91.
- Kefeli, V. I., Kalevitch, M. V., & Borsari, B. (2003). Phenolic cycle in plants and environment. *Journal of Cell and Molecular Biology*, 2(1), 13-18.
- Keng, P. S., Basri, M., Abdul Rahman, M. B., Salleh, A. B., Rahman, R. A., Zaliha, R. N., & Ariff, A. (2005). Optimization of palm-based wax esters production using statistical experimental designs. *Journal of Oleo Science*, 54(10), 519-528.
- Kermasha, S., Safari, M., & Goetghebeur, M. (1995). Interesterification of butter fat by lipase from *Rhizopus niveus* in cosurfactant-free microemulsion system. *Applied Biochemistry and Biotechnology*, 53(3), 229-244.

- Kerry, N., & Rice-Evans, C. (1998). Peroxynitrite oxidises catechols to o-quinones. *FEBS Letters*, 437(3), 167-171.
- Keumi, T., & Meidar, D. (1978). Synthetic Methods and Reactions; 511. A convenient and improved method for esterification over nafion-H2, a superacidic perfluorinated resinsulfonic acid catalyst. *Synthesis*, 1978(12), 929-930.
- Khadem, S., & Marles, R. J. (2010). Monocyclic phenolic acids; hydroxy-and polyhydroxybenzoic acids: occurrence and recent bioactivity studies. *Molecules*, 15(11), 7985-8005.
- Khuri, A. I., & Cornell, J. A. (1996). Response surfaces: designs and analyses, 2nd edn: Marcel Dekker, New York, USA.
- Kikuzaki, H., Hisamoto, M., Hirose, K., Akiyama, K., & Taniguchi, H. (2002). Antioxidant properties of ferulic acid and its related compounds. *Journal of Agricultural and Food Chemistry*, 50(7), 2161-2168.
- Kim, K.-W., Song, B., Choi, M.-Y., & Kim, M.-J. (2001). Biocatalysis in ionic liquids: markedly enhanced enantioselectivity of lipase. *Organic Letters*, 3(10), 1507-1509.
- Kim, M. J., Choi, M. Y., Lee, J. K., & Ahn, Y. (2003). Enzymatic selective acylation of glycosides in ionic liquids: significantly enhanced reactivity and regioselectivity. *Journal of Molecular Catalysis B: Enzymatic*, 26(3), 115-118.
- Kim, S. R., & Kim, Y. C. (2000). Neuroprotective phenylpropanoid esters of rhamnose isolated from roots of *Scrophularia buergeriana*. *Phytochemistry*, 54(5), 503-509.
- Kimura, Y., Okuda, H., Okuda, T., Hatano, T., Agata, I., & Arichi, S. (1985). Studies on the activities of tannins and related compounds from medicinal plants and drugs. VII. Effects of extracts of leaves of *Artemisia* species, and caffeic acid and chlorogenic acid on lipid metabolic injury in rats fed peroxidized oil. *Chemical and Pharmaceutical Bulletin*, 33(5), 2028-2034.
- King, P. J., Ma, G., Miao, W., Jia, Q., McDougall, B. R., Reinecke, M. G., Robinson, W. E. (1999). Structure-activity relationships: analogues of the dicaffeoylquinic and dicaffeoyltartaric acids as potent inhibitors of human immunodeficiency virus type 1 integrase and replication. *Journal of Medicinal Chemistry*, 42(3), 497-509.
- Knezevic Jugovic, Z., Jakovetic, S., Jugovic, B., Gvozdencovic, M., Grbavcic, S., Bezbradica, D., & Antov, M. (2012). Enzymatic Synthesis of Aliphatic Esters of Phenolic Acids and Evaluation of Their Antioxidant Properties. In *Proceedings of the 39th International Conference of Slovak Society of Chemical Engineering*, Tatranske Matliare, Slovakia, 1426-1432.

- Kobayashi, T. (2011). Lipase-catalyzed syntheses of sugar esters in non-aqueous media. *Biotechnology Letters*, 33(10), 1911-1919.
- Kroon, P. A., & Williamson, G. (1999). Hydroxycinnamates in plants and food: current and future perspectives. *Journal of the Science of Food and Agriculture*, 79(3), 355-361.
- Kurata, A., Kitamura, Y., Irie, S., Takemoto, S., Akai, Y., Hirota, Y., Kishimoto, N. (2010). Enzymatic synthesis of caffeic acid phenethyl ester analogues in ionic liquid. *Journal of Biotechnology*, 148(2), 133-138.
- Ky, C. L., Noirot, M., & Hamon, S. (1997). Comparison of five purification methods for chlorogenic acids in green coffee beans (*Coffea* sp.). *Journal of Agricultural and Food Chemistry*, 45(3), 786-790.
- La-Vecchia, C., Altieri, A., & Tavani, A. (2001). Vegetables, fruit, antioxidants and cancer: a review of Italian studies. *European Journal of Nutrition*, 40(6), 261-267.
- Lafay, S., & Gil-Izquierdo, A. (2008). Bioavailability of phenolic acids. *Phytochemistry Reviews*, 7(2), 301-311.
- Lago, S., Rodríguez, H., Arce, A., & Soto, A. (2014). Improved concentration of citrus essential oil by solvent extraction with acetate ionic liquids. *Fluid Phase Equilibria*, 361, 37-44.
- Lamoureux, G., & Agüero, C. (2009). A comparison of several modern alkylating agents. *Arkivoc*, 1, 251-264.
- Landry, T., Brooks, K., Poche, D., & Woolhiser, M. (2005). Acute toxicity profile of 1-butyl-3-methylimidazolium chloride. *Bulletin of Environmental Contamination and Toxicology*, 74(3), 559-565.
- Laranjinha, J., Vieira, O. I., Madeira, V. T., & Almeida, L. (1995). Two related phenolic antioxidants with opposite effects on vitamin E content in low density lipoproteins oxidized by ferrylmyoglobin: consumption vs regeneration. *Archives of Biochemistry and Biophysics*, 323(2), 373-381.
- Laranjinha, J. A., Almeida, L. M., & Madeira, V. M. (1994). Reactivity of dietary phenolic acids with peroxy radicals: antioxidant activity upon low density lipoprotein peroxidation. *Biochemical Pharmacology*, 48(3), 487-494.
- Lasekan, O., Juhari, N. H., & Pattiram, P. D. (2011). Headspace solid-phase microextraction analysis of the volatile flavour compounds of roasted chickpea (*Cicer arietinum* L.). *Journal Food Processing and Technology*, 2(3), 1-6.

- Laszlo, J. A., Compton, D. L., Eller, F. J., Taylor, S. L., & Isbell, T. A. (2003). Packed-bed bioreactor synthesis of feruloylated monoacyl- and diacylglycerols: clean production of a “green” sunscreen. *Green Chemistry*, 5(4), 382-386.
- Lau, R. M., Sorgedraeger, M. J., Carrea, G., van Rantwijk, F., Secundo, F., & Sheldon, R. A. (2004). Dissolution of *Candida antarctica* lipase B in ionic liquids: effects on structure and activity. *Green Chemistry*, 6(9), 483-487.
- Laura, A., Alvarez-Parrilla, E., & Gonzalez-Aguilar, G. A. (2009). Fruit and vegetable phytochemicals: Chemistry, nutritional value and stability: John Wiley & Sons, New York, USA.
- Leblanc, L. M., Paré, A. F., Jean-François, J., Hébert, M. J., Surette, M. E., & Touaibia, M. (2012). Synthesis and Antiradical/Antioxidant Activities of Caffeic Acid Phenethyl Ester and Its Related Propionic, Acetic, and Benzoic Acid Analogues. *Molecules*, 17(12), 14637-14650.
- Lee, D., & Paquette, L. (1995). Encyclopedia of reagents for organic synthesis. 2nd Edition, John Wiley & Sons, New York, USA.
- Lee, G.-S., Widjaja, A., & Ju, Y.-H. (2006). Enzymatic synthesis of cinnamic acid derivatives. *Biotechnology Letters*, 28(8), 581-585.
- Lee, J., Ye, L., Landen, W. O., & Eitenmiller, R. R. (2000). Optimization of an extraction procedure for the quantification of vitamin E in tomato and broccoli using response surface methodology. *Journal of Food Composition and Analysis*, 13(1), 45-57.
- Lee, S. H., Ha, S. H., Lee, S. B., & Koo, Y.-M. (2006). Adverse effect of chloride impurities on lipase-catalyzed transesterifications in ionic liquids. *Biotechnology Letters*, 28(17), 1335-1339.
- Ley, J. P., & Bertram, H.-J. (2001). Synthesis of polyhydroxylated aromatic mandelic acid amides and their antioxidative potential. *Tetrahedron*, 57(7), 1277-1282.
- Li, H., Wang, W., Wang, S., Kong, W., Ren, J., & Sun, R. (2014). Ionic Liquids: Promising Solvents And Catalysts For Conversion Of Lignocellulose Biomass Into Chemicals And Biomaterials. *International Journal of Condensed Matter, Advanced Materials, and Superconductivity Research*, 13(4), 343.
- Li, W.-N., Chen, B.-Q., & Tan, T.-W. (2011). Esterification synthesis of ethyl oleate in solvent-free system catalyzed by lipase membrane from fermentation broth. *Applied Biochemistry and Biotechnology*, 163(1), 102-111.
- Li, Y.-Q. (1999). Catalytic esterifications of carboxylic acids and alcohols by sodium bisulfate monohydrate. *Synthetic Communications*, 29(22), 3901-3903.

- Liebler, D. C., Baker, P. F., & Kaysen, K. L. (1990). Oxidation of vitamin E: Evidence for competing autoxidation and peroxy radical trapping reactions of the tocopheroxyl radical. *Journal of the American Chemical Society*, *112*(19), 6995-7000.
- Lin, L.-C., Kuo, Y.-C., & Chou, C.-J. (1999). Immunomodulatory principles of *Dichrocephala bicolor*. *Journal of Natural Products*, *62*(3), 405-408.
- Liu, L., Jin, C., & Zhang, Y. (2014). Lipophilic phenolic compounds (Lipo-PCs): emerging antioxidants applied in lipid systems. *RSC Advances*, *4*(6), 2879-2891.
- Liu, Q.-P., Hou, X.-D., Li, N., & Zong, M.-H. (2012). Ionic liquids from renewable biomaterials: synthesis, characterization and application in the pretreatment of biomass. *Green Chemistry*, *14*(2), 304-307.
- Long, H., Zhu, Y., Cregor, M., Tian, F., Coury, L., Kissinger, C. B., & Kissinger, P. T. (2001). Liquid chromatography with multi-channel electrochemical detection for the determination of epigallocatechin gallate in rat plasma utilizing an automated blood sampling device. *Journal of Chromatography B: Biomedical Sciences and Applications*, *763*(1), 47-51.
- Lozano, P., Bernal, J. M., & Navarro, A. (2012). A clean enzymatic process for producing flavour esters by direct esterification in switchable ionic liquid/solid phases. *Green Chemistry*, *14*(11), 3026-3033.
- Lozano, P., de Diego, T., Gmouh, S., Vaultier, M., & Iborra, J. L. (2004). Criteria to design green enzymatic processes in ionic liquid/supercritical carbon dioxide systems. *Biotechnology Progress*, *20*(3), 661-669.
- Lozano, P., De Diego, T., Carrie, D., Vaultier, M., & Iborra, J. (2001). Over-stabilization of *Candida antarctica* lipase B by ionic liquids in ester synthesis. *Biotechnology Letters*, *23*(18), 1529-1533.
- Lozano, P., de Diego, T., Guegan, J. P., Vaultier, M., & Iborra, J. L. (2001). Stabilization of α -chymotrypsin by ionic liquids in transesterification reactions. *Biotechnology and Bioengineering*, *75*(5), 563-569.
- Lu, Y., & Luthria, D. (2016). Influence of gelatinization on the extraction of phenolic acids from wheat fractions. *Food Chemistry*, *194*, 1138-1142.
- Lue, B.-M., Guo, Z., Glasius, M., & Xu, X. (2010a). Scalable preparation of high purity rutin fatty acid esters. *Journal of the American Oil Chemists' Society*, *87*(1), 55-61.
- Lue, B.-M., Nielsen, N. S., Jacobsen, C., Hellgren, L., Guo, Z., & Xu, X. (2010b). Antioxidant properties of modified rutin esters by DPPH, reducing power, iron chelation and human low density lipoprotein assays. *Food Chemistry*, *123*(2), 221-230.

- Lue, B. M., Karboune, S., Yeboah, F. K., & Kermasha, S. (2005). Lipase-catalyzed esterification of cinnamic acid and oleyl alcohol in organic solvent media. *Journal of Chemical Technology and Biotechnology*, 80(4), 462-468.
- Ma, X., Yan, R., Yu, S., Lu, Y., Li, Z., & Lu, H. (2012). Enzymatic acylation of isoorientin and isovitexin from bamboo-leaf extracts with fatty acids and antiradical activity of the acylated derivatives. *Journal of Agricultural and Food Chemistry*, 60(43), 10844-10849.
- Macfarlane, D. R., Golding, J., Forsyth, S., Forsyth, M., & Deacon, G. B. (2001). Low viscosity ionic liquids based on organic salts of the dicyanamide anion. *Chemical Communications* (16), 1430-1431.
- Macheix, J. J., & Fleuriet, A. (1990). *Fruit phenolics*: CRC Press, Boca Raton, Florida, 20- 34.
- Madeira Lau, R., Van Rantwijk, F., Seddon, K., & Sheldon, R. (2000). Lipase-catalyzed reactions in ionic liquids. *Organic Letters*, 2(26), 4189-4191.
- Magnusson A (2005) Ph. D. Thesis, Royal Institute of Technology, Stockholm, Sweden.
- Manach, C., Scalbert, A., Morand, C., Rémésy, C., & Jiménez, L. (2004). Polyphenols: food sources and bioavailability. *The American Journal of Clinical Nutrition*, 79(5), 727-747.
- Martin, K., & Appel, C. (2010). Polyphenols as dietary supplements: a double-edged sword. *Nutrition and Dietary Supplements*, 2, 1-12.
- Martins, A. B., Graebin, N. G., Lorenzoni, A. S., Fernandez-Lafuente, R., Ayub, M. A., & Rodrigues, R. C. (2011). Rapid and high yields of synthesis of butyl acetate catalyzed by Novozym 435: reaction optimization by response surface methodology. *Process Biochemistry*, 46(12), 2311-2316.
- Maruyama, T., Nagasawa, S., & Goto, M. (2002). Poly (ethylene glycol)-lipase complex that is catalytically active for alcoholysis reactions in ionic liquids. *Biotechnology Letters*, 24(16), 1341-1345.
- Maruyama, T., Yamamura, H., Kotani, T., Kamiya, N., & Goto, M. (2004). Poly (ethylene glycol)-lipase complexes that are highly active and enantioselective in ionic liquids. *Organic and Biomolecular Chemistry*, 2(8), 1239-1244.
- Mason, R. L., Gunst, R. F., & Hess, J. L. (2003). Statistical design and analysis of experiments: with applications to engineering and science. John Wiley & Sons, Inc. Hoboken, New Jersey.

- Mat-Radzi, S., Basri, M., Salleh, A. B., Ariff, A., Mohammad, R., Abdul Rahman, M. B., Zaliha, R. N. (2006). Optimisation study of large-scale enzymatic synthesis of oleyl oleate, a liquid wax ester, by response surface methodology. *Journal of Chemical Technology and Biotechnology*, 81(3), 374-380.
- Mattila, P., Hellström, J., & Törrönen, R. (2006). Phenolic acids in berries, fruits, and beverages. *Journal of Agricultural and Food Chemistry*, 54(19), 7193-7199.
- Maximo, G. J., Santos, R. J., Lopes-da-Silva, J. A., Costa, M. C., Meirelles, A. J., & Coutinho, J. o. A. (2013). Lipidic Protic Ionic Liquid Crystals. *Acs Sustainable Chemistry & Engineering*, 2(4), 672-682.
- McCarthy, T., Kerry, J., Kerry, J., Lynch, P., & Buckley, D. (2001). Evaluation of the antioxidant potential of natural food/plant extracts as compared with synthetic antioxidants and vitamin E in raw and cooked pork patties. *Meat Science*, 58(1), 45-52.
- Michaluart, P., Masferrer, J. L., Carothers, A. M., Subbaramaiah, K., Zweifel, B. S., Koboldt, C., Tanabe, T. (1999). Inhibitory effects of caffeic acid phenethyl ester on the activity and expression of cyclooxygenase-2 in human oral epithelial cells and in a rat model of inflammation. *Cancer Research*, 59(10), 2347-2352.
- Milasinovic, N., Knezevic-Jugovic, Z., Jakovljevic, Z., Filipovic, J., & Krusic, M. K. (2012). Synthesis of n-amyl isobutyrate catalyzed by *Candida rugosa* lipase immobilized into poly (N-isopropylacrylamide-co-itaconic acid) hydrogels. *Chemical Engineering Journal*, 181, 614-623.
- Mirhosseini, H., Tan, C. P., Hamid, N. S., & Yusof, S. (2008). Optimization of the contents of Arabic gum, xanthan gum and orange oil affecting turbidity, average particle size, polydispersity index and density in orange beverage emulsion. *Food Hydrocolloids*, 22(7), 1212-1223.
- Miyake, Y., Yamamoto, K., & Osawa, T. (1997). Metabolism of antioxidant in lemon fruit (*Citrus limon* BURM. f.) by human intestinal bacteria. *Journal of Agricultural and Food Chemistry*, 45(10), 3738-3742.
- Monino, I., Martínez, C., Sotomayor, J. A., Lafuente, A., & Jordan, M. J. (2008). Polyphenolic Transmission to Segureño Lamb Meat from Ewes' Diet Supplemented with the Distillate from Rosemary (*Rosmarinus officinalis*) Leaves. *Journal of Agricultural and Food Chemistry*, 56(9), 3363-3367.
- Montgomery, D. C., Runger, G. C., & Hubele, N. F. (2009). *Engineering statistics*: John Wiley & Sons, New York, USA.
- Montgomery, D. (2001). *Design and Analysis of Experiments*, 5th edn, 2001: John Wiley & Sons, Inc. New York, USA. 51-117.

- Moon, J.-H., & Terao, J. (1998). Antioxidant activity of caffeic acid and dihydrocaffeic acid in lard and human low-density lipoprotein. *Journal of Agricultural and Food Chemistry*, 46(12), 5062-5065.
- Morishita, H., & Ohnishi, M. (2001). Absorption, metabolism and biological activities of chlorogenic acids and related compounds. *Studies in Natural Products Chemistry*, 25, 919-953.
- Murota, S., & Koshihara, Y. (1984). New lipoxygenase inhibitors isolated from Chinese plants. Development of new anti-allergic drugs. *Drugs under Experimental and Clinical Research*, 11(9), 641-644.
- Myers, R. H., & Montgomery, D. H. (1995). Response surface methodology. John Wiley & Sons, New York, USA.
- Myers, R. H., Montgomery, D. C., Vining, G. G., Borror, C. M., & Kowalski, S. M. (2004). Response surface methodology: a retrospective and literature survey. *Journal of Quality Technology*, 36(1), 53.
- Nagaoka, T., Banskota, A. H., Tezuka, Y., Saiki, I., & Kadota, S. (2002). Selective antiproliferative activity of caffeic acid phenethyl ester analogues on highly liver-metastatic murine colon 26-L5 carcinoma cell line. *Bioorganic & Medicinal Chemistry*, 10(10), 3351-3359.
- Nakashima, K., Maruyama, T., Kamiya, N., & Goto, M. (2005). Comb-shaped poly (ethylene glycol)-modified subtilisin Carlsberg is soluble and highly active in ionic liquids. *Chemical Communications* (34), 4297-4299.
- Nakashima, K., Okada, J., Maruyama, T., Kamiya, N., & Goto, M. (2006). Activation of lipase in ionic liquids by modification with comb-shaped poly (ethylene glycol). *Science and Technology of Advanced Materials*, 7(7), 692-698.
- Nakatani, N. (2000). Phenolic antioxidants from herbs and spices. *Biofactors*, 13(1-4), 141-146.
- Nardini, M., D'Aquino, M., Tomassi, G., Gentili, V., Di Felice, M., & Scaccini, C. (1995). Inhibition of human low-density lipoprotein oxidation by caffeic acid and other hydroxycinnamic acid derivatives. *Free Radical Biology and Medicine*, 19(5), 541-552.
- Nardini, M., Natella, F., Gentili, V., Di Felice, M., & Scaccini, C. (1997). Effect of Caffeic Acid Dietary Supplementation on the Antioxidant Defense System in Rat: An In Vivo Study. *Archives of Biochemistry and Biophysics*, 342(1), 157-160.
- Natella, F., Nardini, M., Di Felice, M., & Scaccini, C. (1999). Benzoic and cinnamic acid derivatives as antioxidants: structure-activity relation. *Journal of Agricultural and Food Chemistry*, 47(4), 1453-1459.

- Olivier-Bourbigou, H., Magna, L., & Morvan, D. (2010). Ionic liquids and catalysis: recent progress from knowledge to applications. *Applied Catalysis A: General*, 373(1), 1-56.
- Owen, R., Haubner, R., Mier, W., Giacosa, A., Hull, W., Spiegelhalder, B., & Bartsch, H. (2003). Isolation, structure elucidation and antioxidant potential of the major phenolic and flavonoid compounds in brined olive drupes. *Food and Chemical Toxicology*, 41(5), 703-717.
- Park, S., & Kazlauskas, R. J. (2001). Improved preparation and use of room-temperature ionic liquids in lipase-catalyzed enantio- and regioselective acylations. *The Journal of Organic Chemistry*, 66(25), 8395-8401.
- Park, S., & Kazlauskas, R. J. (2003). Biocatalysis in ionic liquids—advantages beyond green technology. *Current Opinion in Biotechnology*, 14(4), 432-437.
- Parr, A. J., & Bolwell, G. P. (2000). Phenols in the plant and in man. The potential for possible nutritional enhancement of the diet by modifying the phenols content or profile. *Journal of the Science of Food and Agriculture*, 80(7), 985-1012.
- Passos, H., Freire, M. G., & Coutinho, J. A. (2014). Ionic liquid solutions as extractive solvents for value-added compounds from biomass. *Green Chemistry*, 16(12), 4786-4815.
- Paya, M., Halliwell, B., & Hoult, J. (1992). Interactions of a series of coumarins with reactive oxygen species: scavenging of superoxide, hypochlorous acid and hydroxyl radicals. *Biochemical Pharmacology*, 44(2), 205-214.
- Peleg, H., Naim, M., Rouseff, R. L., & Zehavi, U. (1991). Distribution of bound and free phenolic acids in oranges (*Citrus sinensis*) and grapefruits (*Citrus paradisi*). *Journal of the Science of Food and Agriculture*, 57(3), 417-426.
- Peppercorn, M. A., & Goldman, P. (1971). Caffeic acid metabolism by bacteria of the human gastrointestinal tract. *Journal of Bacteriology*, 108(3), 996-1000.
- Pereira, D. M., Valentão, P., Pereira, J. A., & Andrade, P. B. (2009). Phenolics: From chemistry to biology. *Molecules*, 14(6), 2202-2211.
- Peric, B., Sierra, J., Martí, E., Cruañas, R., Garau, M. A., Arning, J., . . . Stolte, S. (2013). (Eco) toxicity and biodegradability of selected protic and aprotic ionic liquids. *Journal of Hazardous Materials*, 261, 99-105.
- Persson, M., & Bornscheuer, U. T. (2003). Increased stability of an esterase from *Bacillus stearothermophilus* in ionic liquids as compared to organic solvents. *Journal of Molecular Catalysis B: Enzymatic*, 22(1), 21-27.
- Pham, M.-Q., Yoon, H.-S., Khare, V., & Ahn, S.-H. (2014). Evaluation of ionic liquids as lubricants in micro milling—process capability and sustainability. *Journal of Cleaner Production*, 76, 167-173.

- Pietta, P.-G. (2000). Flavonoids as antioxidants. *Journal of Natural Products*, 63(7), 1035-1042.
- Porter, W. L. (1993). Paradoxical behavior of antioxidants in food and biological systems. *Toxicology and Industrial Health*, 9(1-2), 93.
- Priya, K., & Chadha, A. (2003). Synthesis of hydrocinnamic esters by *Pseudomonas cepacia* lipase. *Enzyme and Microbial Technology*, 32(3), 485-490.
- Psomiadou, E., & Tsimidou, M. (2002). Stability of virgin olive oil. 1. Autoxidation studies. *Journal of Agricultural and Food Chemistry*, 50(4), 716-721.
- Plou, F. J., Barandiaran, M., Calvo, M. V., Ballesteros, A., & Pastor, E. (1996). High-yield production of mono- and di-oleylglycerol by lipase-catalyzed hydrolysis of triolein. *Enzyme and Microbial Technology*, 18(1), 66-71.
- Rababah, T. M., Ereifej, K. I., & Howard, L. (2005). Effect of ascorbic acid and dehydration on concentrations of total phenolics, antioxidant capacity, anthocyanins, and color in fruits. *Journal of Agricultural and Food Chemistry*, 53(11), 4444-4447.
- Rao, X., Zhang, J., Zheng, J., Song, Z., & Shang, S. (2014). Chiral ionic liquid crystals with a bulky rigid core from renewable camphorsulfonic acid. *RSC Advances*, 4(48), 25334-25340.
- Razzaghi-Asl, N., Garrido, J., Khazraei, H., Borges, F., & Firuzi, O. (2013). Antioxidant properties of hydroxycinnamic acids: a review of structure-activity relationships. *Current Medicinal Chemistry*, 20(36), 4436-4450.
- Reblova, Z. (2006). The effect of temperature on the antioxidant activity of tocopherols. *European Journal of Lipid Science and Technology*, 108(10), 858-863.
- Reis, I. A., Santos, S. B., Pereira, F. D., Sobral, C. R., Freire, M. G., Freitas, L. S., . . . Lima, Á. S. (2014). Extraction and recovery of rutin from acerola waste using alcohol-salt-based aqueous two-phase systems. *Separation Science and Technology*, 49(5), 656-663.
- Reische, D. W., Lillard, D. A., & Eitenmiller, R. R. (1999). Antioxidants. In C. Akoh & D. Min (Eds.), *Food Lipids: Chemistry, Nutrition, and Biotechnology* (. 409-434). UK: CRC Press.
- Ren, M.-Y., Bai, S., Zhang, D.-H., & Sun, Y. (2008). pH memory of immobilized lipase for (\pm)-menthol resolution in ionic liquid. *Journal of Agricultural and Food chemistry*, 56(7), 2388-2391.
- Rice-Evans, C. A., Miller, N. J., & Paganga, G. (1996). Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radical Biology and Medicine*, 20(7), 933-956.

- Rios, L. Y., Gonthier, M.-P., Rémésy, C., Mila, I., Lapiere, C., Lazarus, S. A., Scalbert, A. (2003). Chocolate intake increases urinary excretion of polyphenol-derived phenolic acids in healthy human subjects. *The American Journal of Clinical Nutrition*, 77(4), 912-918.
- Robards, K., Prenzler, P. D., Tucker, G., Swatsitang, P., & Glover, W. (1999). Phenolic compounds and their role in oxidative processes in fruits. *Food Chemistry*, 66(4), 401-436.
- Robbins, R. J. (2003). Phenolic acids in foods: an overview of analytical methodology. *Journal of Agricultural and Food Chemistry*, 51(10), 2866-2887.
- Roche, M., Dufour, C., Mora, N., & Dangles, O. (2005). Antioxidant activity of olive phenols: mechanistic investigation and characterization of oxidation products by mass spectrometry. *Organic and Biomolecular Chemistry*, 3(3), 423-430.
- Roosen, C., Müller, P., & Greiner, L. (2008). Ionic liquids in biotechnology: applications and perspectives for biotransformations. *Applied Microbiology and Biotechnology*, 81(4), 607-614.
- Rumbau, V., Marcilla, R., Ochoteco, E., Pomposo, J. A., & Mecerreyes, D. (2006). Ionic liquid immobilized enzyme for biocatalytic synthesis of conducting polyaniline. *Macromolecules*, 39(25), 8547-8549.
- Sabally, K., Karboune, S., St-Louis, R., & Kermasha, S. (2006a). Lipase-catalyzed transesterification of dihydrocaffeic acid with flaxseed oil for the synthesis of phenolic lipids. *Journal of Biotechnology*, 127(1), 167-176.
- Sabally, K., Karboune, S., St-Louis, R., & Kermasha, S. (2006b). Lipase-catalyzed transesterification of trilinolein or trilinolenin with selected phenolic acids. *Journal of the American Oil Chemists' Society*, 83(2), 101-107.
- Sabally, K., Karboune, S., Yeboah, F. K., & Kermasha, S. (2005a). Lipase-catalyzed esterification of selected phenolic acids with linolenyl alcohols in organic solvent media. *Applied Biochemistry and Biotechnology*, 127(1), 17-27.
- Sabally, K., Karboune, S., Yeboah, F. K., & Kermasha, S. (2005b). Enzymatic esterification of dihydrocaffeic acid with linoleyl alcohol in organic solvent media. *Biocatalysis and Biotransformation*, 23(1), 37-44.
- Sabally, K., Karboune, S., St-Louis, R., & Kermasha, S. (2007). Lipase-catalyzed synthesis of phenolic lipids from fish liver oil and dihydrocaffeic acid. *Biocatalysis and Biotransformation*, 25(2-4), 211-218.
- Salem, J. H., Humeau, C., Chevalot, I., Harscoat-Schiavo, C., Vanderesse, R., Blanchard, F., & Fick, M. (2010). Effect of acyl donor chain length on isoquercitrin acylation and biological activities of corresponding esters. *Process Biochemistry*, 45(3), 382-389.

- Salunkhe, D. K., Chaven, J. K., & Kadam, S. S. (1990). Plant phenolics: structure, classification, and biosynthesis. *Dietary Tannins: Consequences and Remedies*, CRC Press: Boca Raton, Florida. 5-28.
- Santos, L. D., Coutinho, J. A., & Ventura, S. P. (2015). From water-in-oil to oil-in-water emulsions to optimize the production of fatty acids using ionic liquids in micellar systems. *Biotechnology progress*, 31(6), 1473-1480.
- Saraji, M., & Mousavi, F. (2010). Use of hollow fibre-based liquid–liquid–liquid microextraction and high-performance liquid chromatography–diode array detection for the determination of phenolic acids in fruit juices. *Food Chemistry*, 123(4), 1310-1317.
- Sarma, A. D., Mallick, A. R., & Ghosh, A. (2010). Free radicals and their role in different clinical conditions: an overview. *International Journal of Pharma Sciences and Research*, 1(3), 185-192.
- Scholz, E., Heinrich, M., & Hunkler, D. (1994). Caffeoylquinic acids and some biological activities of *Pluchea symphytifolia*. *Planta Medica*, 60(4), 360-364.
- Setzer, W. N., Setzer, M. C., Bates, R. B., Nakkiew, P., Jackes, B. R., Chen, L., Meehan, E. J. (1999). Antibacterial hydroxycinnamic esters from *Piper caninum* from Paluma, north Queensland, Australia. The crystal and molecular structure of (+)-bornyl coumarate. *Planta Medica*, 65(8), 747-749.
- Shah, S., Solanki, K., & Gupta, M. N. (2007). Enhancement of lipase activity in non-aqueous media upon immobilization on multi-walled carbon nanotubes. *Chemistry Central Journal*, 1(1), 1.
- Shahidi, F., Naczki, M., & Griffiths, W. (1996). Food phenolics: sources, chemistry, effects, applications. *Trends in Food Science and Technology*, 7(7), 235-277.
- Shahidi, F., & Zhong, Y. (2010). Lipid oxidation and improving the oxidative stability. *Chemical Society Reviews*, 39(11), 4067-4079.
- Sheldon, R. A., Lau, R. M., Sorgedraeger, M. J., van Rantwijk, F., & Seddon, K. R. (2002). Biocatalysis in ionic liquids. *Green Chemistry*, 4(2), 147-151.
- Shelke, K. (1990). Chinese wet noodle formulation: A response surface methodology study. *Cereal Chemistry*, 67, 338-342.
- Silva, F. A., Borges, F., Guimarães, C., Lima, J. L., Matos, C., & Reis, S. (2000). Phenolic acids and derivatives: studies on the relationship among structure, radical scavenging activity, and physicochemical parameters. *Journal of Agricultural and Food Chemistry*, 48(6), 2122-2126.
- Simitzis, P., Deligeorgis, S., Bizelis, J., Dardamani, A., Theodosiou, I., & Fegeros, K. (2008). Effect of dietary oregano oil supplementation on lamb meat characteristics. *Meat Science*, 79(2), 217-223.

- Singh, A. K., & Mukhopadhyay, M. (2012). Overview of fungal lipase: a review. *Applied Biochemistry and Biotechnology*, 166(2), 486-520.
- Son, S., & Lewis, B. A. (2002). Free radical scavenging and antioxidative activity of caffeic acid amide and ester analogues: structure-activity relationship. *Journal of Agricultural and Food Chemistry*, 50(3), 468-472.
- Sorensen, A. D. M., Friel, J., Winkler-Moser, J. K., Jacobsen, C., Huidrom, D., Reddy, N., & Thiyam-Holländer, U. (2013). Impact of endogenous canola phenolics on the oxidative stability of oil-in-water emulsions. *European Journal of Lipid Science and Technology*, 115(5), 501-512.
- Sorensen, A. D. M., Nielsen, N. S., Yang, Z., Xu, X., & Jacobsen, C. (2012). Lipophilization of dihydrocaffeic acid affects its antioxidative properties in fish-oil-enriched emulsions. *European Journal of Lipid Science and Technology*, 114(2), 134-145.
- Stalikas, C. D. (2007). Extraction, separation, and detection methods for phenolic acids and flavonoids. *Journal of Separation Science*, 30(18), 3268-3295.
- Stamatis, H., Sereti, V., & Kolisis, F. (1999). Studies on the enzymatic synthesis of lipophilic derivatives of natural antioxidants. *Journal of the American Oil Chemists' Society*, 76(12), 1505-1510.
- Stamatis, H., Sereti, V., & Kolisis, F. (2001). Enzymatic synthesis of hydrophilic and hydrophobic derivatives of natural phenolic acids in organic media. *Journal of Molecular Catalysis B: Enzymatic*, 11(4), 323-328.
- Stuwe, H.-T., Bruhn, G., König, W., & Hausen, B. (1989). The synthesis of caffeic acid esters, a new group of naturally occurring contact allergens. *Naturwissenschaften*, 76(9), 426-427.
- Sudina, G., Mirzoeva, O., Pushkareva, M., Korshunova, G. A., Sumbatyan, N., & Varfolomeev, S. (1993). Caffeic acid phenethyl ester as a lipoxygenase inhibitor with antioxidant properties. *FEBS Letters*, 329(1-2), 21-24.
- Sugiura, M., Naito, Y., Yamaura, Y., Fukaya, C., & Yokoyama, K. (1989). Inhibitory Activities and Inhibition Specificities of Caffeic Acid Derivatives and Related Compounds. *Chemical and Pharmaceutical Bulletin*, 37(4), 1039-1043.
- Sun, B., Qi, L., & Wang, M. (2014). Determination of preservatives in soft drinks by capillary electrophoresis with ionic liquids as the electrolyte additives. *Journal of Separation Science*, 37(16), 2248-2252.
- Sun, S., Shan, L., Liu, Y., Jin, Q., Zhang, L., & Wang, X. (2008). Solvent-free enzymatic preparation of feruloylated monoacylglycerols optimized by response surface methodology. *Journal of Agricultural and Food Chemistry*, 56, 442-447.

- Sureshkumar, M., & Lee, C.-K. (2009). Biocatalytic reactions in hydrophobic ionic liquids. *Journal of Molecular Catalysis B: Enzymatic*, 60(1), 1-12.
- Susan, M. A., Noda, A., Mitsushima, S., & Watanabe, M. (2003). Brønsted acid–base ionic liquids and their use as new materials for anhydrous proton conductors. *Chemical Communications*(8), 938-939.
- Swatloski, R. P., Spear, S. K., Holbrey, J. D., & Rogers, R. D. (2002). Dissolution of cellulose with ionic liquids. *Journal of the American Chemical Society*, 124(18), 4974-4975.
- Tan, Z., & Shahidi, F. (2012). Optimization of enzymatic synthesis of phytosteryl caprylates using response surface methodology. *Journal of the American Oil Chemists' Society*, 89(4), 657-666.
- Thomas, R. H., Bernards, M. A., Drake, E. E., & Guglielmo, C. G. (2010). Changes in the antioxidant activities of seven herb-and spice-based marinating sauces after cooking. *Journal of Food Composition and Analysis*, 23(3), 244-252.
- Tomaino, A., Cimino, F., Zimbalatti, V., Venuti, V., Sulfaro, V., De Pasquale, A., & Saija, A. (2005). Influence of heating on antioxidant activity and the chemical composition of some spice essential oils. *Food Chemistry*, 89(4), 549-554.
- Tomas-Barberan, F. A., & Espin, J. C. (2001). Phenolic compounds and related enzymes as determinants of quality in fruits and vegetables. *Journal of the Science of Food and Agriculture*, 81(9), 853-876.
- Topakas, E., Stamatis, H., Biely, P., Kekos, D., Macris, B., & Christakopoulos, P. (2003). Purification and characterization of a feruloyl esterase from *Fusarium oxysporum* catalyzing esterification of phenolic acids in ternary water–organic solvent mixtures. *Journal of Biotechnology*, 102(1), 33-44.
- Tsevegsuren, N., Edrada, R., Lin, W., Ebel, R., Torre, C., Ortlepp, S., Proksch, P. (2007). Biologically active natural products from Mongolian medicinal plants *Scorzonera divaricata* and *Scorzonera pseudodivaricata*. *Journal of Natural Products*, 70(6), 962-967.
- Uchida, K. (2000). Role of reactive aldehyde in cardiovascular diseases. *Free Radical Biology and Medicine*, 28(12), 1685-1696.
- Uppenberg, J., Hansen, M. T., Patkar, S., & Jones, T. A. (1994). The sequence, crystal structure determination and refinement of two crystal forms of lipase B from *Candida antarctica*. *Structure*, 2(4), 293-308.
- Vafiadi, C., Topakas, E., Alissandratos, A., Faulds, C. B., & Christakopoulos, P. (2008). Enzymatic synthesis of butyl hydroxycinnamates and their inhibitory effects on LDL-oxidation. *Journal of Biotechnology*, 133(4), 497-504.

- Vaisey-Genser, M., Ylimaki, G., & Johnston, B. (1987). The selection of levels of canola oil, water, and an emulsifier system in cake formulations by response surface methodology. *Cereal Chemistry*, 64(1), 50-54.
- Valentao, P., Fernandes, E., Carvalho, F., Andrade, P., Seabra, R., & Bastos, M. (2003). Hydroxyl radical and hypochlorous acid scavenging activity of small centaury (*Centaureum erythraea*) infusion. A comparative study with green tea (*Camellia sinensis*). *Phytomedicine*, 10(6), 517-522.
- Valentao, P., Fernandes, E., Carvalho, F., Andrade, P. B., Seabra, R. M., & Bastos, M. d. L. (2002a). Antioxidant activity of *Hypericum androsaemum* infusion: scavenging activity against superoxide radical, hydroxyl radical and hypochlorous acid. *Biological and Pharmaceutical Bulletin*, 25(10), 1320-1323.
- Valentao, P., Fernandes, E., Carvalho, F., Andrade, P. B., Seabra, R. M., & Bastos, M. d. L. (2002b). Studies on the antioxidant activity of *Lippia citriodora* infusion: scavenging effect on superoxide radical, hydroxyl radical and hypochlorous acid. *Biological and Pharmaceutical Bulletin*, 25(10), 1324-1327.
- Valentao, P., Fernandes, E., Carvalho, F., Andrade, P. B., Seabra, R. M., & Bastos, M. L. (2002c). Antioxidative properties of cardoon (*Cynara cardunculus* L.) infusion against superoxide radical, hydroxyl radical, and hypochlorous acid. *Journal of Agricultural and Food Chemistry*, 50(17), 4989-4993.
- Van-Rantwijk, F., & Sheldon, R. A. (2007). Biocatalysis in ionic liquids. *Chemical Reviews*, 107(6), 2757-2785.
- Ventura, S. P., Santos, L. D., Saraiva, J. A., & Coutinho, J. A. (2012). Ionic liquids microemulsions: the key to *Candida antarctica* lipase B superactivity. *Green Chemistry*, 14(6), 1620-1625.
- Vining, G. G., & Kowalski, S. (2010). *Statistical methods for engineers*: Cengage Learning, chapter 1, Boston, USA.
- Viuda-Martos, M., Ruiz-Navajas, Y., Fernandez-Lopez, J., & Perez-Alvarez, J. A. (2010). Effect of adding citrus fibre washing water and rosemary essential oil on the quality characteristics of a bologna sausage. *LWT-Food Science and Technology*, 43(6), 958-963.
- Voisin-Chiret, A. S., Bazin, M.-A., Lancelot, J.-C., & Rault, S. (2007). Synthesis of new L-ascorbic ferulic acid hybrids. *Molecules*, 12(11), 2533-2545.
- Von Gadow, A., Joubert, E., & Hansmann, C. F. (1997). Comparison of the antioxidant activity of aspalathin with that of other plant phenols of rooibos tea (*Aspalathus linearis*), α -tocopherol, BHT, and BHA. *Journal of Agricultural and Food Chemistry*, 45(3), 632-638.

- Vosmann, K., Weitkamp, P., & Weber, N. (2006). Solvent-free lipase-catalyzed preparation of long-chain alkyl phenylpropanoates and phenylpropyl alkanoates. *Journal of Agricultural and Food Chemistry*, 54(8), 2969-2976.
- Walt, D. R., & Agayn, V. I. (1994). The chemistry of enzyme and protein immobilization with glutaraldehyde. *TrAC Trends in Analytical Chemistry*, 13(10), 425-430.
- Wanasundara, P., & Shahidi, F. (2005). Antioxidants: science, technology, and applications. *Bailey's Industrial Oil and Fat Products*, 11, 431-489.
- Watanabe, Y., Nakanishi, H., Goto, N., Otsuka, K., Kimura, T., & Adachi, S. (2010). Antioxidative properties of ascorbic acid and acyl ascorbates in ML/W emulsion. *Journal of the American Oil Chemists' Society*, 87(12), 1475-1480.
- Weitkamp, P., Vosmann, K., & Weber, N. (2006). Highly efficient preparation of lipophilic hydroxycinnamates by solvent-free lipase-catalyzed transesterification. *Journal of Agricultural and Food Chemistry*, 54(19), 7062-7068.
- Weng, W.-L., Liu, Y.-C., & Lin, C.-W. (2001). Studies on the optimum models of the dairy product Kou Woan Lao using response surface methodology. *Asian Australasian Journal of Animal Sciences*, 14(10), 1470-1476.
- Weuster-Botz, D. (2000). Experimental design for fermentation media development: statistical design or global random search? *Journal of Bioscience and Bioengineering*, 90(5), 473-483.
- Widjaja, A., Yeh, T.-H., & Ju, Y.-H. (2008). Enzymatic synthesis of caffeic acid phenethyl ester. *Journal of the Chinese Institute of Chemical Engineers*, 39(5), 413-418.
- Wills, T., DeWitt, C. M., & Sigfusson, H. (2007). Improved antioxidant activity of Vitamin E through solubilization in ethanol: A model study with ground beef. *Meat Science*, 76(2), 308-315.
- Xanthakis, E., Theodosiou, E., Magkouta, S., Stamatis, H., Loutrari, H., Roussos, C., & Kolisis, F. (2010). Enzymatic transformation of flavonoids and terpenoids: Structural and functional diversity of the novel derivatives. *Pure and Applied Chemistry*, 82(1), 1-16.
- Yadav, G. D., & Lathi, P. S. (2004). Synthesis of citronellol laurate in organic media catalyzed by immobilized lipases: kinetic studies. *Journal of Molecular Catalysis B: Enzymatic*, 27(2), 113-119.
- Yan, Y., Bornscheuer, U. T., & Schmid, R. D. (1999). Lipase-catalyzed synthesis of vitamin C fatty acid esters. *Biotechnology Letters*, 21(12), 1051-1054.

- Yang, C. S., Landau, J. M., Huang, M.-T., & Newmark, H. L. (2001). Inhibition of carcinogenesis by dietary polyphenolic compounds. *Annual Review of Nutrition*, 21(1), 381-406.
- Yang, R.-L., Li, N., Li, R.-F., Smith, T. J., & Zong, M.-H. (2010). A highly regioselective route to arbutin esters by immobilized lipase from *Penicillium expansum*. *Bioresource Technology*, 101(1), 1-5.
- Yang, Z., Feddern, V., Glasius, M., Guo, Z., & Xu, X. (2011). Improved enzymatic production of phenolated acylglycerols through alkyl phenolate intermediates. *Biotechnology Letters*, 33(4), 673-679.
- Yang, Z., Guo, Z., & Xu, X. (2012a). Enzymatic lipophilisation of phenolic acids through esterification with fatty alcohols in organic solvents. *Food Chemistry*, 132(3), 1311-1315.
- Yang, Z., Guo, Z., & Xu, X. (2012b). Ionic liquid-assisted solubilization for improved enzymatic esterification of phenolic acids. *Journal of the American Oil Chemists' Society*, 89(6), 1049-1055.
- Yang, Z., & Pan, W. (2005). Ionic liquids: Green solvents for nonaqueous biocatalysis. *Enzyme and Microbial Technology*, 37(1), 19-28.
- Yasuko, K., Tomohiro, N., Sei-Itsu, M., Ai-Na, L., Yasuo, F., & Takashi, T. (1984). Caffeic acid is a selective inhibitor for leukotriene biosynthesis. *Biochimica et Biophysica Acta (BBA)-Lipids and Lipid Metabolism*, 792(1), 92-97.
- Yu, H.-h., Liu, X.-g., Xing, R.-e., Liu, S., Guo, Z.-y., Wang, P.-b., Li, P.-c. (2006). In vitro determination of antioxidant activity of proteins from jellyfish *Rhopilema esculentum*. *Food Chemistry*, 95(1), 123-130.
- Zhang, G.-S. (1999). Fe₂(SO₄)₃·xH₂O in Synthesis: A Convenient and Efficient Catalyst for the Esterification of Aromatic Carboxylic Acids With Alcohols. *Synthetic Communications*, 29(4), 607-611.
- Zhang, Q., Zhang, S., & Deng, Y. (2011). Recent advances in ionic liquid catalysis. *Green Chemistry*, 13(10), 2619-2637.
- Zhao, D., Liao, Y., & Zhang, Z. (2007). Toxicity of ionic liquids. *Clean-Soil, Air, Water*, 35(1), 42-48.
- Zhao, H. (2010). Methods for stabilizing and activating enzymes in ionic liquids: a review. *Journal of Chemical Technology and Biotechnology*, 85(7), 891-907.
- Zhao, Z., & Moghadasian, M. H. (2008). Chemistry, natural sources, dietary intake and pharmacokinetic properties of ferulic acid: A review. *Food Chemistry*, 109(4), 691-702.

Zheng, W., & Wang, S. Y. (2001). Antioxidant activity and phenolic compounds in selected herbs. *Journal of Agricultural and Food Chemistry*, 49(11), 5165-5170.



© COPYRIGHT UPM