

UNIVERSITI PUTRA MALAYSIA

COMPARISON AND DETERMINATION OF PROPERTIES OF OIL EXTRACTED USING SOXHLET AND SUPERCRITICAL FLUID FROM NILE TILAPIA (Oreochromis niloticus Linnaeus) PROCESSING WASTE

MASOUMEH KHAZAALI

FSTM 2016 14



COMPARISON AND DETERMINATION OF PROPERTIES OF OIL EXTRACTED USING SOXHLET AND SUPERCRITICAL FLUID FROM NILE TILAPIA (*Oreochromis niloticus* Linnaeus) PROCESSING WASTE



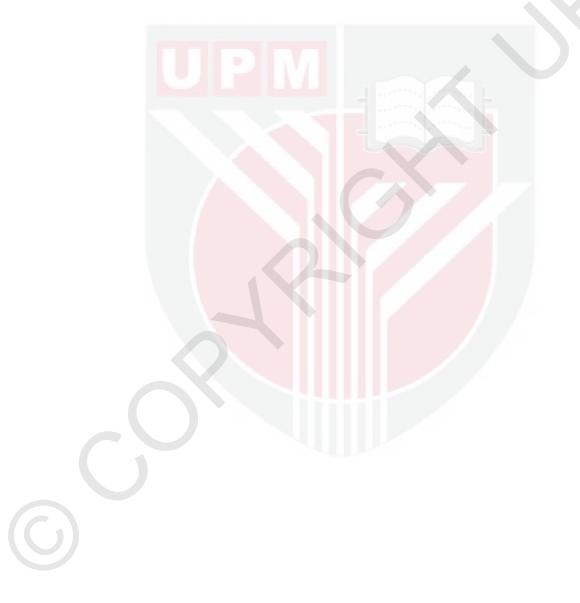
Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirement for the Degree of Master of Science

June 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

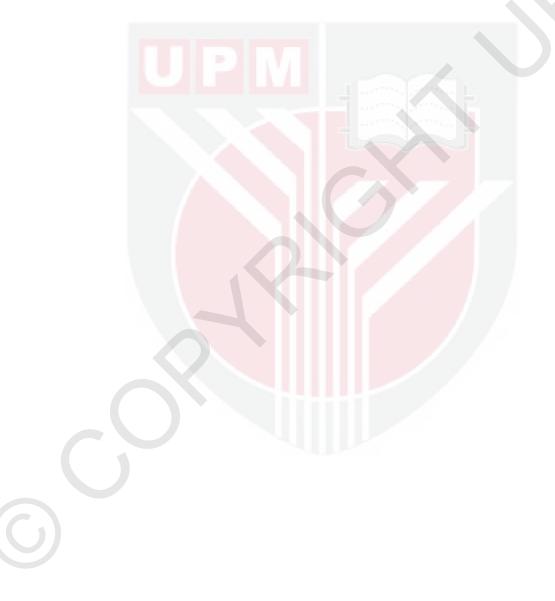
Copyright© Universiti Putra Malaysia



DEDICATION

In the name of Allah, Most Gracious, Most Merciful This thesis is dedicated to:

My caring and devoted parents who are always giving me unlimited courage, passion and support and My sister and brothers for their moral support and love



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

COMPARISON AND DETERMINATION OF PROPERTIES OF OIL EXTRACTED USING SOXHLET AND SUPERCRITICAL FLUID FROM NILE TILAPIA (*Oreochromis niloticus* Linnaeus) PROCESSING WASTE

By

MASOUMEH KHAZAALI

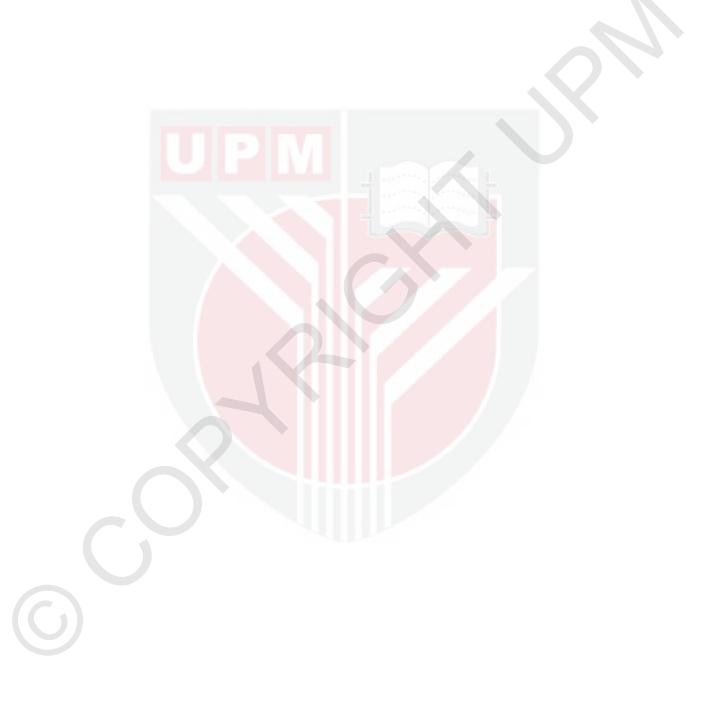
June 2016

Chairman: Professor Jamilah binti Bakar, PhDFaculty: Food Science and Technology

Fish oil contains the important omega-3 polyunsaturated fatty acids (PUFAs), Eicosapentaenoic acid (EPA) and Docosahexaenoic acid (DHA). EPA and DHA extracted from fish oil are reported to have beneficial effects to human health which resulted in a world-wide demand. There are already several methods available for extracting fish oil, with varying degrees of yield and quality. Supercritical fluid carbon dioxide extraction (SFE-CO₂) is an emerging process for the extraction of edible oils containing labile components like PUFAs. Therefore, the objectives of this study were (i) to extract and compare the distribution of oil and physicochemical properties of tilapia oil extracted from the viscera, head, skin and flesh using the soxhlet and supercritical fluid methods; (ii) to optimize the yield and extracted EPA and DHA from the identified part using supercritical fluid extraction (SFE-CO₂) technique. The results from the soxhlet method revealed that the distribution of oil was significantly (p < 0.05) different among the head, viscera, skin and flesh which is as expected. The highest oil yield was obtained from the viscera (51.81%) followed by the head (30.12%), skin (16.88%) and flesh (6.78%). The chemical properties of head, viscera, flesh and skin oil were 4.37-9.87 meg O₂/kg, 116.94-125.65 g I₂/100 g, 4.49-9.86 meq/kg and 13.23-25.08 meq/kg for PV, IV, p-AnV and TOTOX value, respectively. Although the average fatty acids composition of the viscera, head, skin and flesh samples were similar, the highest amounts of SFAs (49.74 g/100 g oil), MUFAs (24.40 g/100 g oil) and PUFAs (22.14 g/100 g oil) were found in the flesh, viscera and head, respectively. The optimization of oil yield, extraction of EPA and DHA based on central composite design with pressure (20-40 MPa), temperature (35-75 °C), flow rate (2.5-4.5 mL.min⁻¹) and extraction time (2-4 h) from the viscera was determined. The highest oil yield (40.02% on dry weight basis), EPA (0.46 g/100 g oil) and DHA (1.53 g/100g oil) was obtained at 40 MPa, 57.22 °C, 3.4 mL.min⁻¹ and 3.23 h with non-significant (p > 0.05) lack of fit and high R^2 (p > 0.90). The viscera oil from supercritical fluid extraction (SFE-CO₂) had lower total oxidation value (15.3 meq/kg) and higher amount of PUFAs composition (28.71 g/100 g oil) than oil obtained from soxhlet method (25.08 meq/kg and 17.32



g/100 g oil for TOTOX value and PUFAs, respectively). Hence, tilapia visceral oil is better extracted using supercritical fluid extraction method.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PERBANDINGAN DAN PENENTUAN CIRI-CIRI MINYAK EKSTRAKSI DARIPADA SISA IKAN NILE TILAPIA (*Oreochromis niloticus linnaeus*) MELALUI KAEDAH SOXHLET DAN FLUIDA SUPERKRITIK

Oleh

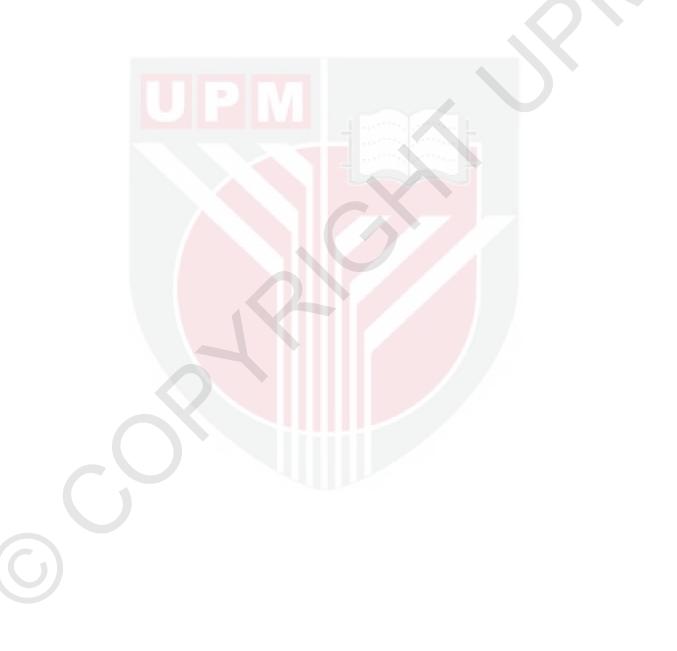
MASOUMEH KHAZAALI

Jun 2016

Pengerusi:Professor Jamilah binti Bakar, PhDFakulti:Sains dan Teknology Makanan

Minyak ikan mengandungi asid lemak omega-3 politaktepu (PUFAs), asid eikosapentaenoik (EPA) dan asid dokosaheksaenoik (DHA) yang penting. EPA dan DHA yang diekstrak daripada minyak ikan dilaporkan bermanfaat kepada kesihatan manusia. Oleh sebab demikian, ia mendapat permintaan yang tinggi dari serata dunia. Terdapat beberapa kaedah yang telah digunakan untuk mengekstrak minyak ikan, dengan kadar pengeluaran dan mutu yang berlainan. Ekstraksi fluida superkritik CO₂ (SFE-CO₂) adalah satu proses yang baru muncul untuk pengekstrakan minyak makan yang mengandungi komponen labil seperti PUFA. Tujuan kajian ini adalah untuk (i) mengekstrak dan membandingkan taburan minyak dan sifat fisiokimia minyak tilapia yang diekstrak daripada viscera, kepala, kulit dan isi, menggunakan kaedah soxhlet dan kaedah SFE-CO₂; (ii) mengoptimumkan pengeluaran minyak dan pengekstrakan EPA dan DHA daripada bahagian ikan terpilih menggunakan kaedah SFE-CO₂. Seperti yang dijangkakan, hasil kajinan kaedah soxhlet menunjukkan bahawa taburan minyak berbeza dengan ketara (p<0.05) antara kepala, viscera, kulit dan isi ikan. Kandungan minyak paling banyak didapati daripada viscera (51.81%), diikuti dengan kepala (30.12%), kulit (16.88%) dan isi ikan (6.78%). Sifat-sifat kimia minyak daripada kepala, viscera, isi dan kulit ikan adalah 4.37-9.87 meq O₂/kg (nilai peroksida), 116.94-125.65 g I₂/100 g (nilai iodine), 4.49-9.86 meq/kg (p-anisidine) and 13.23-25.08 meq/kg (nilai TOTOX) masing-masing. Walaupun purata kandungan asid lemak daripada viscera, kepala, kulit dan isi ikan adalah hampir sama, kandungan tertinggi SFA (49.74 g/100 g minyak), MUFA (24.40 g/100 g minyak) dan PUFA (22.14 g/100 g minyak) diperoleh daripada bahagian kulit, viscera dan kepala ikan. Pengoptimuman hasil minyak, pengekstrakan EPA dan DHA daripada viscera ikan telah ditentukan dengan tekanan (20-40 MPa), suhu (35-75 °C), kadar aliran (2.5-4.5 mL.min⁻¹) dan masa pengekstrakan (2-4 jam) berdasarkan reka bentuk komposit pusat. Hasil tertinggi minyak (40.02% berasaskan berat kering), EPA (0.46 g /100 g minyak) dan DHA (1.53 g /100g minyak) telah diperolehi pada 40 MPa, 57.22 °C, 3.4 mL.min-1 dan 3.23 jam dengan "lack of fit" yang tidak signifikan (p>0.05) dan R² yang tinggi (p>0.90). Minyak viscera daripada pengekstrakan fluida superkritik SFE-CO₂

mempunyai jumlah nilai pengoksidaan yang lebih rendah (15.3 meq/kg) dan komposisi PUFA yang lebih tinggi (28.71 g/100 g minyak) berbanding minyak yang diperolehi daripada kaedah soxhlet (25.08 meq/kg dan 17.32 g/100 g minyak untuk nilai TOTOX dan PUFA masing-masing). Oleh itu, minyak visceral tilapia adalah lebih baik diekstrak menggunakan kaedah pengekstrakan fluida.



ACKNOWLEDGEMENTS

First of all, all thanks belong to the most gracious Allah for granting me the good health, patient, and steadfastness accomplish my academic journey. I am honored and grateful to have the full support and commitment from many wonderful people during my study.

I would like to extend my sincere thanks and gratitude to Prof. Dr. Jamilah Bakar for her kindness, invaluable guidance, and unflinching support throughout my study. Her scrutiny, suggestions and wonderful comments indeed made me a better research student. In addition, I also wish to extend my sincere gratitude to Prof. Jinap Selamat for her role as my co-supervisor; her wisdom, knowledge, and commitment towards the achievements I made in my time working with her.

My special thanks also go to Assoc. Prof. Dzulkifly Mat hashim for his invaluable assistance and suggestions during my study. I am extremely thankful that he genuinely cared not only about my academic and professional success, but also my well-being as a person.

I would like to express my gratitude to the technical staff of the Faculty of Food Science and Technology for their support, sincere help and warm welcome throughout my study period.

Last but not least, it is my pleasure to extend my enthusiastic thankful expression to my beloved family: my father, mother, sister and brothers, without their moral support to my research journey would not have been successful. I certify that a Thesis Examination Committee has met on 20 June 2016 to conduct the final examination of Masoumeh Khazaali on her thesis entitled "Comparison and Determination of Properties of Oil Extracted using Soxhlet and Supercritical Fluid from Nile Tilapia (*Oreochromis niloticus* Linnaeus) Processing Waste" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Tan Chin Ping, PhD

Professor Faculty of Food Science and Technology Universiti Putra Malaysia (Chairman)

Abdulkarim Sabo Mohammed, PhD

Associate Professor Faculty of Food Science and Technology Universiti Putra Malaysia (Internal Examiner)

Mohamed Elwathig Saeed Mirghani, PhD

Associate Professor International Islamic University Malaysia Malaysia (External Examiner)



ZULKARNAIN ZAINAL, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 28 September 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Jamilah Bt. Bakar, PhD

Professor Faculty of Food Science and Technology Universiti Putra Malaysia (Chairman)

Jinap Selamat, PhD

Professor Faculty of Food Science and Technology Universiti Putra Malaysia (Member)

BUJANG KIM HUAT, PhD

Professor and Dean Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice- Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Date:

Name and Matric No: Masoumeh Khazaali, GS32696

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

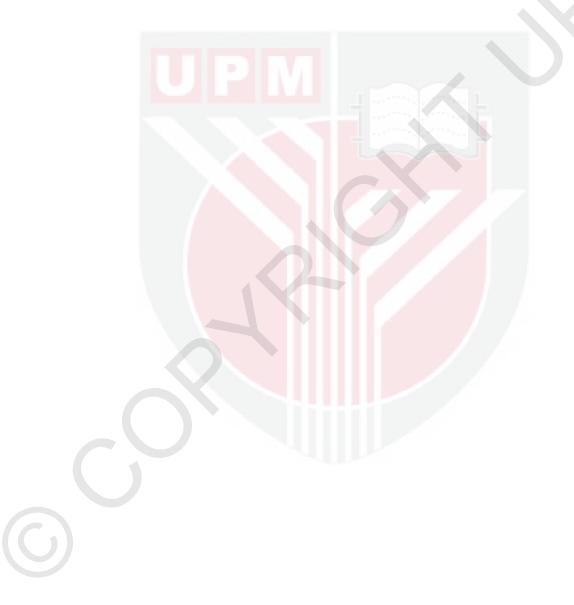
Signature: Name of Chairman of Supervisory	PM
Committee:	Professor Dr.Jamilah Bt. Bakar
Signature:	
Name of	
Member of	
Supervisory	
Committee:	Professor Dr. Jinap Selamat

TABLE OF CONTENTS

			Page
ABST ACKI		EDGMENTS	i iii v vi
	LARAT		viii
LIST	OF TA	BLES	xiii
LIST	OF FI	GURES	xiv
		BREVIATIONS	XV
LIST	OF AP	PENDICES	xvii
CHAI	PTER		
1	INTR	ODUCTION	1
	1.1	Background	1
	1.2	Problem Statement and Hypothesis	2 2
	1.3	Objectives	2
2	LITE	RATURE REVIEW	3
-	2.1	Fish Oil	3 3
		2.1.1 Source of Fish Oil	5
		2.1.2 Essential Fatty Acids	6
		2.1.3 Omega-3	6
		2.1.4 EPA and DHA	8
		2.1.5 EPA/DHA Ratio	9
	2.2	Methods of Extraction of Fish Oil	10
		2.2.1 Conventional Methods	12
		2.2.1.1 Solvent Extraction	12 13
		2.2.1.2 Rendering 2.2.1.3 Enzymatic Method	13
		2.2.2 Supercritical Fluid Extraction SFE-CO ₂	14
		2.2.2 Supercritical Fluid Extraction St E-CO ₂ 2.2.2.1 Fundamental Properties of Supercritical Fluid	14
		2.2.2.2 SFE-CO ₂ Application to Oil Extraction	15
	2.3	Quality Evaluation of Fish Oil	16
	2.4	Nile tilapia (Oreochromis niloticus)	17
		2.4.1 Economic Contribution to Global Aquaculture	18
		2.4.2 Processing of Tilapia	19
3	PHYS	SICO-CHEMICAL PROPERTIES OF SOXHLET	
		RACTED OIL FROM VARIOUS PARTS OF TILAPIA	
		chromis niloticus)	20
	3.1	Introduction	20
	3.2	Materials and Methods	20
		3.2.1 Preparation of Sample3.2.2 Proximate Analysis	20 21
		3.2.2 Proximate Analysis 3.2.2.1 Determination of Moisture	21
			<u> </u>

			3.2.2.2 Determination of Protein	21
			3.2.2.3 Determination of Oil	22
			3.2.2.3.1 Soxhlet Extraction	22
			3.2.2.4 Determination of Ash	22
			3.2.2.5 Determination of Carbohydrate Content	23
		3.2.3	Identification of EPA and DHA	23
		3.2.4	Determination of Physicochemical Properties	23
			3.2.4.1 Color 23	
			3.2.4.2 Peroxide Value	24
			3.2.4.3 Anisidine Value	24
			3.2.4.4 Iodine Value	25
			3.2.4.5 Total Oxidation Value (TOTOX value)	25
		3.2.5	· · · · · · · · · · · · · · · · · · ·	-
			Spectrometry	26
		3.2.6		26
	3.3		s and Discussion	26
	0.0		Weight Distribution and Nutritional Composition of	
		0.0.1	Tilapia Body Parts	26
		3.3.2	Exhibited Physicochemical Characterisation	27
		0.0.2	3.3.2.1 Peroxide Value, Iodine Value, Anisidine Value,	21
			Totox Value and Color	27
		3.3.3		29
			Fatty Acid Composition	29
	3.4	Conclu		32
4	FRO	M TILA	TION OF EXTRACTION OF EPA AND DHA APIA VISCERA USING SUPERCRITICAL FLUID ON (SFE-CO ₂)	
				22
				33
	4.1	Introd	uction	33
		Introd Materi	uction ials and Method	33 33
	4.1	Introd Materi 4.2.1	uction ials and Method Preparation of Raw Material	33 33 33
	4.1	Introd Mater 4.2.1 4.2.2	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction	33 33 33 33
	4.1	Introd Materi 4.2.1 4.2.2 4.2.3	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield	33 33 33 33 34
	4.1	Introd Materi 4.2.1 4.2.2 4.2.3 4.2.3 4.2.4	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA	33 33 33 33 34 34
	4.1	Introd Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM	33 33 33 33 34 34 34
	4.1	Introd Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5 4.2.6	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM Physicochemical Properties and Fatty Acids Profile	33 33 33 33 34 34 34 34 37
	4.1 4.2	Introd Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5 4.2.6 4.2.7	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM Physicochemical Properties and Fatty Acids Profile Statistical Analysis	33 33 33 34 34 34 34 37 37
	4.1	Introd Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5 4.2.5 4.2.6 4.2.7 Result	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM Physicochemical Properties and Fatty Acids Profile Statistical Analysis s and Discussion	33 33 33 33 34 34 34 34 37
	4.1 4.2	Introd Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5 4.2.6 4.2.7	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM Physicochemical Properties and Fatty Acids Profile Statistical Analysis s and Discussion Fitting the Response Surface Model for Independent	 33 33 33 34 34 34 34 37 37 37
	4.1 4.2	Introd Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5 4.2.6 4.2.7 Result 4.3.1	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM Physicochemical Properties and Fatty Acids Profile Statistical Analysis s and Discussion Fitting the Response Surface Model for Independent Variables	 33 33 33 34 34 34 37 37 37 37
	4.1 4.2	Introd Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5 4.2.6 4.2.7 Result 4.3.1 4.3.2	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM Physicochemical Properties and Fatty Acids Profile Statistical Analysis and Discussion Fitting the Response Surface Model for Independent Variables Analysis for Optimization of the Responses	 33 33 33 34 34 34 34 37 37 37
	4.1 4.2	Introd Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5 4.2.6 4.2.7 Result 4.3.1	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM Physicochemical Properties and Fatty Acids Profile Statistical Analysis s and Discussion Fitting the Response Surface Model for Independent Variables Analysis for Optimization of the Responses Effects of Temperature, Pressure, Flow Rate and Time	 33 33 33 34 34 34 34 37 37 37 41
	4.1 4.2	Introd Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5 4.2.6 4.2.7 Result 4.3.1 4.3.2 4.3.3	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM Physicochemical Properties and Fatty Acids Profile Statistical Analysis s and Discussion Fitting the Response Surface Model for Independent Variables Analysis for Optimization of the Responses Effects of Temperature, Pressure, Flow Rate and Time on the Extraction Yield	 33 33 33 34 34 34 37 37 37 37
	4.1 4.2	Introd Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5 4.2.6 4.2.7 Result 4.3.1 4.3.2	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM Physicochemical Properties and Fatty Acids Profile Statistical Analysis and Discussion Fitting the Response Surface Model for Independent Variables Analysis for Optimization of the Responses Effects of Temperature, Pressure, Flow Rate and Time on the Extraction Yield Effects of SFE Parameters on the Extraction of EPA	 33 33 33 33 34 34 34 34 37 37 37 41 42
	4.1 4.2	Intrody Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5 4.2.6 4.2.7 Result 4.3.1 4.3.2 4.3.3 4.3.4	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM Physicochemical Properties and Fatty Acids Profile Statistical Analysis s and Discussion Fitting the Response Surface Model for Independent Variables Analysis for Optimization of the Responses Effects of Temperature, Pressure, Flow Rate and Time on the Extraction Yield Effects of SFE Parameters on the Extraction of EPA and DHA	 33 33 33 34 34 34 34 37 37 37 41
	4.1 4.2	Introd Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5 4.2.6 4.2.7 Result 4.3.1 4.3.2 4.3.3	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM Physicochemical Properties and Fatty Acids Profile Statistical Analysis s and Discussion Fitting the Response Surface Model for Independent Variables Analysis for Optimization of the Responses Effects of Temperature, Pressure, Flow Rate and Time on the Extraction Yield Effects of SFE Parameters on the Extraction of EPA and DHA Comparison of Fatty Acid Profile and Physicochemical	 33 33 33 33 34 34 34 34 37 37 37 41 42
	4.1 4.2	Intrody Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5 4.2.6 4.2.7 Result 4.3.1 4.3.2 4.3.3 4.3.4	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM Physicochemical Properties and Fatty Acids Profile Statistical Analysis and Discussion Fitting the Response Surface Model for Independent Variables Analysis for Optimization of the Responses Effects of Temperature, Pressure, Flow Rate and Time on the Extraction Yield Effects of SFE Parameters on the Extraction of EPA and DHA Comparison of Fatty Acid Profile and Physicochemical Characterization of Viscera Oil from SFE and Soxhlet	 33 33 33 34 34 34 34 37 37 37 41 42 45
	4.1 4.2	Intrody Materi 4.2.1 4.2.2 4.2.3 4.2.4 4.2.5 4.2.6 4.2.7 Result 4.3.1 4.3.2 4.3.3 4.3.4	uction ials and Method Preparation of Raw Material Supercritical Fluid Extraction Determination of Crude Oil Yield Determination of EPA and DHA Experimental Design for RSM Physicochemical Properties and Fatty Acids Profile Statistical Analysis s and Discussion Fitting the Response Surface Model for Independent Variables Analysis for Optimization of the Responses Effects of Temperature, Pressure, Flow Rate and Time on the Extraction Yield Effects of SFE Parameters on the Extraction of EPA and DHA Comparison of Fatty Acid Profile and Physicochemical Characterization of Viscera Oil from SFE and Soxhlet Methods	 33 33 33 33 34 34 34 34 37 37 37 41 42

5	SUM	IMARY, CONCLUSION AND RECOMMENDATIONS	51
	5.1	Summary and Conclusion	51
	5.2	Recommendations and Future Work	52
REI	FEREN	CES	53
API	PENDIC	ES	71
BIC	DATA	OF STUDENT	75



LIST OF TABLES

	Page
EPA and DHA Content in Various Fish	9
Composition of Different Fish Species and Their Oil Content	11
Quality Properties of Crude Fish Oil	17
Proximate Analysis of Fresh Tilapia Body Parts	27
Physicochemical Properties of Different Parts of Tilapia	28
Fatty Acid Content of Oil from Different Parts of Tilapia	31
Experimental Ranges of the Independent Variables Used in the Central Composite Design (CCD) for the Oil Yield from Viscera of Tilapia	35
The Matrix of Central Composite Design (CCD)	36
The Experimental and Predicted Values for Responses	38
Analysis of Variance (ANOVA) of the Reduced Model for Yield, EPA and DHA of Viscera Oil	39
Regression Coefficients and F-Ratio of the Predicted Second- Order Model for the Response Variables, Yield, EPA and DHA	40
Optimization Parameters of Viscera Oil by SFE	42
Physicochemical Properties of Viscera Oil as Obtained from Supercritical Fluid Method	48
Fatty Acid Content of Viscera Oil	49
	 Composition of Different Fish Species and Their Oil Content Quality Properties of Crude Fish Oil Proximate Analysis of Fresh Tilapia Body Parts Physicochemical Properties of Different Parts of Tilapia Fatty Acid Content of Oil from Different Parts of Tilapia Experimental Ranges of the Independent Variables Used in the Central Composite Design (CCD) for the Oil Yield from Viscera of Tilapia The Matrix of Central Composite Design (CCD) The Experimental and Predicted Values for Responses Analysis of Variance (ANOVA) of the Reduced Model for Yield, EPA and DHA of Viscera Oil Regression Coefficients and F-Ratio of the Predicted Second- Order Model for the Response Variables, Yield, EPA and DHA Optimization Parameters of Viscera Oil by SFE Physicochemical Properties of Viscera Oil as Obtained from Supercritical Fluid Method

LIST OF FIGURES

Figure		Page
2.1	PUFA Formation Pathway	4
2.2	The Uses of Different Parts of Fish	5
2.3	Global Tilapia Production	19
4.1	3D Surface Plot of the % Oil Yield as a Function of Pressure and Temperature at Fixed Flow Rate of 3.5 mL/min and Extraction Time 3h	43
4.2	3D Surface Plot of the % Oil Yield as a Function of Pressure and Flow Rate at Fixed Temperature of 55 °C and Extraction Time of 3h	44
4.3	3D Surface Plot of the % Oil Yield as a Function of Flow Rate and Temperature at Fixed Extraction Time of 3 h and Pressure of 30 MPa	45
4.4	3D Surface Plot of the Absorption of EPA as a Function of Flow Rate and Extraction Time at Fixed Pressure 30 MPa and Temperature 55 °C	46
4.5	3D Surface Plot of the Absorption of DHA as a Function of Flow Rate and Extraction Time at Fixed Pressure 30 MPa and Temperature 55 $^{\circ}$ C	47

LIST OF ABBREVIATIONS

AA	Arachidonic Acid
AIs	Adequate Intakes
ALA	Alpha Linolenic Acid
ANOVA	Analysis of Variance
AnV	Anisidine Value
CCD	Central Composite Design
CCL	Carbon Chain Length
CHD	Coronary Heart Disease
CSE	Conventional Soxhlet Extraction
CVD	Cardiovascular Diseases
DF	Degrees of Freedom
DHA	Docosahexaenoic Acid
DOE	Design of Experiment
EFAs	Essential Fatty Acids
EP	European Pharmacopeia
EPA	Eicosapentaenoic Acid
FA	Fatty Acid
FAME	Fatty Acid Methyl Ester
FFA	Free Fatty Acids
FPH	Fish Protein Hydrolysate
GC	Gas Chromatography
GC-FID	Gas Chromatography Flame Ionization Detector
GC-MS	Gas Chromatography Mass Spectrometry
GIFT	Genetically Improved Farmed Tilapia
GOED	Global Organization for EPA and DHA
HCl	Hydrochloric Acid
HDL	High Density Lipoprotein
IOM	Institute Of Medicine
ISSFAL	International Society for the study of Fatty Acids and Lipids

IV	Iodine value
KI	Potassium Iodide
LA	Linoleic Acid
LCPUFA	Long Chain Polyunsaturated Fatty Acid
LDL	Low Density Lipoprotein
MI	Myocardium Infarction
MUFA	Monounsaturated Fatty Acid
PUFA	Polyunsaturated Fatty Acids
PV	Peroxide value
RSM	Response Surface Methodology
SC-CO ₂	Supercritical Carbon Dioxide
SCFAs	Short Chain Fatty Acids
SD	Standard Deviation
SFA	Saturated Fatty Acid
SFE	Supercritical Fluid Extraction
SS	Sum of Squared
тотох	Total Oxidation
US FDA	United States Food and Drug Administration
VLDL	Very low Density Lipoprotein

C

LIST OF APPENDICES

Appendix		Page
1	Combined Optimum Conditions of Yield (%), EPA and DHA (g/100 g oil) Ratios of SC-CO ₂ Extracted Tilapia Oil	71
2	Standards Calibration Curve; (a) Eicosapentaenoic acid (EPA); (b) Docosahexaenoic acid (DHA)	72
3	Chromatograph Standard Peaks of the Fatty Acid Composition	73
4	The GC-FID Standard Mixture of 50 (µL/L) of Eicosapentaenoic acid and Docosahexaenoic acid	74

CHAPTER 1

INTRODUCTION

1.1 Background

Fish and fishery products are excellent sources of essential fatty acids (EFAs), protein, vitamins and minerals; with all of these being crucial to a healthy diet (Gokoglu et al., 2004). Extensive worldwide farming of fish and shellfish has increased significantly and the number of fish harvested through aquaculture is actually the same as those harvested from the wild (FAO, 2010). The main species groups in aquaculture are freshwater and marine fishes, molluscs and crustaceans. The supply of marine fish is predicted to show low growth and the prospects of increasing the production of these fishes are on the pessimistic side (Vannuccini, 2005). However, the potential for enhanced supply is exceedingly good for freshwater fish because of the contribution of aquaculture production (Mohamad et al., 2010). In recent years, the production of freshwater fish has been on the rise in Malaysia. The most common species of freshwater fish being cultivated are catfish (Clarias nieuhofii), tilapia (Oreochromis niloticus) and patin (Pangasianodon hypophthalmus) (Silva & Turchini, 2009). The Nile Tilapia (Oreochromis niloticus) is one of the most significant and popular choices of fish species in fish farming (Santos et al., 2013). The popularity of this variety of fish in aquaculture is due to its rapid growth, high resistance to very poor water quality and diseases. In fact, tilapia is now being produced in over 100 countries (Yones et al., 2013). Among all the continents, Asia has seen the most rapid increase in tilapia aquaculture production.

During fish processing, a large amount of waste which include viscera, head, blood, skin and scale are usually discarded (Potaros et al., 2009). These by-products could be used as the raw materials for fish oil extraction. Unfortunately, this is not being practiced at present. Several studies have shown that fatty acids (FAs) from fish oils, particularly two of the constituent FAs, which are eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA), may have a beneficial role in promoting good human health (Russo, 2009; Domingo et al., 2007; Kris-Etherton, 2003). These ω -3 FAs are necessary for normal growth and development in the human body. Furthermore, the ω -3 FAs are also associated with reduced risks of cardiovascular diseases, diabetes mellitus, arthritis, hypertension, cancer and a wide range of inflammatory and autoimmune disorders (Lopez-Huertas, 2010). The EPA and DHA can be obtained directly from their natural sources or extracted and purified to form fish oil supplement. At present, fish oil is commercially extracted from marine species.

In the area of food technology, an extraction process is used for the following: 1) to recover essential and important compounds, 2) to isolate desired components, such as antioxidants, from food sources and 3) to remove any contaminant and undesirable compounds, such as cholesterol and heavy metals (mercury or copper). In recent times, supercritical fluid technology has emerged as an extraction method for heat-labile valuable compounds like EPA and DHA. It reduces the use of toxic

organic solvents and does not require the application of high temperatures, thus preventing the oxidation of valuable compounds such as omega-3 FAs (Moret et al., 2000). The supercritical fluid technology, which usually utilizes carbon dioxide (CO_2) as solvent, has many advantages such as low critical pressure and temperature for extraction. In addition, the final extracts are usually free of contaminants and impurities, and the CO_2 used can be easily removed from the extracts (Sarker et al., 2012).

1.2 Problem Statement and Hypothesis

The main concern of a fish factory is the management of a large amount of fish wastes during the processing of fish products (Potaros et al., 2009). In reality, these by-products can be used as the raw materials for fish oil extraction. In extracting fish oils, several methods have been reported with varying extract yields and qualities (Maqsood et al., 2012). Hitherto, the general focus has been on oil extraction technologies which can achieve maximum oil yield. However, in recent times, there has been an emergence of new technologies which can achieve maximum oil yield and at the same time, minimize the presence of undesirable impurities in the final extract (Amusan, 2008). The conventional methods for fish oil extraction such as rendering and Soxhlet extraction have disadvantages such as requiring high temperatures and long processing times, which can lead to the decomposition or degradation of the thermally-labile compounds, and the rapid oxidation of these compounds may have negative health effects on human (Lee et al., 2012). On the other hand, the use of supercritical fluid CO₂ extraction (SFE-CO₂) has attracted substantial attention because it is a safe and environmental-friendly method. The quality of oil extracted by using SFE-CO₂ is also better due to the milder operation conditions involved.

1.3 Objectives

The overall objectives of this research were to determine and compare the quality of viscera oil extracted by using SFE-CO₂ and conventional methods, and to optimize the extraction of EPA and DHA using SFE-CO₂. The specific objectives of this study were:

1) To extract and compare the distribution of oil and physicochemical properties of tilapia oil extracted from the viscera, head, skin and flesh using the soxhlet and supercritical fluid methods

2) To optimize the yield and extracted EPA and DHA from the identified part using supercritical fluid extraction (SFE-CO₂) technique.

REFERENCES

- Abbas, K. A., Abdulamir, A. S., & HA, A. (2008). A review on supercritical fluid extraction as new analytical method. *American Journal of Biochemistry and Biotechnology*, *4*(4), 345-353.
- Ackman, R. (1988). Concerns for utilization of marine lipids and oils. *Food Technology*, 42(5), 151–155.
- Aidos, I., van der Padt, A., Luten, J. B., & Boom, R. M. (2002). Seasonal changes in crude and lipid composition of herring fillets, by-products, and respective produced oils. *Journal of agricultural and food chemistry*, 50(16), 4589-4599.
- Akanda, M. J. H., Sarker, M. Z. I., Ferdosh, S., Manap, M. Y. A., Ab Rahman, N. N. N., & Ab Kadir, M. O. (2012). Applications of supercritical fluid extraction (SFE) of palm oil and oil from natural sources. *Molecules*, 17(2), 1764–94.
- Akoh, C. C., & Min, D. B. (2008). Food lipids: chemistry, nutrition, and biotechnology. CRC press.
- Akoh, Casimir C & Min, D. B. (2008). Food lipids: chemistry, nutrition, and biotechnology. CRC Press.
- Akpinar, M. A., Görgün, S., & Akpinar, A. E. (2009). A comparative analysis of the fatty acid profiles in the liver and muscles of male and female Salmo trutta macrostigma. *Food Chemistry*, 112(1), 6–8.
- Alfaro, A. D. T., Fonseca, G. G., Balbinot, E., Machado, A., & Prentice, C. (2013). Physical and chemical properties of wami tilapia skin gelatin. *Food Science and Technology*, 33(3), 592-595.
- Alkio, M., Gonzalez, C., Jäntti, M., & Aaltonen, O. (2000). Purification of polyunsaturated fatty acid esters from tuna oil with supercritical fluid chromatography. *Journal of the American Oil Chemists' Society*, 77(3), 315-321.
- Al-Souti, A., Al-Sabahi, J., Soussi, B., & Goddard, S. (2012). The effects of fish oil enriched diets on growth, feed conversion and fatty acid content of red hybrid tilapia, *Oreochromis sp. Food Chemistry*, 133(3), 723–727.
- Amusan, A. R. (2008). Optimization of procedures for oil extraction from animal tissue (master thesis). University of McGill,Canada.
- Anderson, M. J., & Whitcomb, P. J. (2000). *Design of experiments*. John Wiley & Sons, Inc.
- AOAC. (1990). Official Methods of Analysis of the Association of Analytical Chemists, 15th Ed., Association of Official Analytical Chemists, Washington, DC.
- AOAC. (1993). AOAC Official Method. Iodine value of fats and oils, Wijs (cyclohexane-acetic acid solvent) method. American Oil Chemists Society, Champaign, Illinois, USA.
- AOCS Official Method cd 8-53. (2003). AOCS Official Method Cd 8-53. Peroxides in fats and oils. American Oil Chemists Society, Champaign, Illinois, USA.

- AOCS. (2003). Official methods and recommended practices of the American Oil Chemist's Society. In Firestone (Ed.) 5th edition. Champaign. IL.
- Arab-Tehrany, E., Jacquot, M., Gaiani, C., Imran, M., Desobry, S., & Linder, M. (2012). Beneficial effects and oxidative stability of omega-3 long-chain polyunsaturated fatty acids. *Trends in Food Science & Technology*, 25(1), 24-33.
- Aro, H., Järvenpää, E., Könkö, K., Huopalahti, R., & Hietaniemi, V. (2007). The characterisation of oat lipids produced by supercritical fluid technologies. *Journal of Cereal Science*, 45(1), 116–119.
- Askin, R., & Ötles, S. (2005). Supercritical fluids. Acta Scientiarum Polonorum. *Technologia Alimentaria*, 4(1), 3-16.
- Babalola, T., & Apata, D. (2011). Chemical and quality evaluation of some alternative lipid sources for aqua feed production. *Agriculture and Biology Journal of North America*, 2(6), 935–943.
- Bandarra, N. M., Batista, I., Nunes, M. L., & Empis, J. M. (2001). Seasonal variation in the chemical composition of mackerel (*Trachurus trachurus*). *European Food Research and Technology*, 212(5), 535-539.
- Baş, D., & Boyacı, İ. H. (2007). Modeling and optimization: usability of response surface methodology. *Journal of Food Engineering*, 78(3), 836–845.
- Benyounis, KY & Olabi, Abdul-Ghani and Hashmi, M. (2008). Multi-response optimization of CO₂ laser welding process of austenitic stainless steel. *Optics & Laser Technology*, 40(1), 76–87.
- Bhaskar, N., Benila, T., Radha, C., & Lalitha, R. G. (2008). Optimization of enzymatic hydrolysis of visceral waste proteins of Catla (*Catla catla*) for preparing protein hydrolysate using a commercial protease. *Bioresource Technology*, *99*(2), 335–343.
- Bimbo, A. P. (1998). Guidelines for characterizing food-grade fish oil. *International* news on fats, oils and related materials, 9(5), 473-483.
- Blanco, M., Sotelo, C. G., & Chapela, M. J. (2007). Towards sustainable and efficient use of fishery resources : present and future trends. *Trends in Food Science & Technology*, 18, 29–36.
- Boelen, P., van Dijk, R., Damst é, J. S. S., Rijpstra, W. I. C., & Buma, A. G. (2013). On the potential application of polar and temperate marine microalgae for EPA and DHA production. *AMB Express*, *3*(1), 1.
- Bourre, J. M. (2007). Dietary omega-3 fatty acids for women. *Biomedicine & pharmacotherapy*, 61(2), 105-112.
- Bourre, J. M., Piciotti, M., Dumont, O., Pascal, G., & Durand, G. (1990). Dietary linoleic acid and polyunsaturated fatty acids in rat brain and other organs. Minimal requirements of linoleic acid. *Lipids*, 25(8), 465–472.
- Bravi, M., Bubbico, R., Manna, F., & Verdone, N. (2002). Process optimisation in sun fower oil extraction by supercritical CO₂. *Chemical Engineering Science*, 57, 2753–2764.

- Brooks MS, R. V. (2013). Fish Processing Wastes as a Potential Source of Proteins, Amino Acids and Oils: A Critical Review. *Journal of Microbial & Biochemical Technology*, 5(04), 107–129.
- Calder, P. C. (2010). Omega-3 fatty acids and inflammatory processes. *Nutrients*, 2(3), 355–374.
- Campos, H., Baylin, a., & Willett, W. C. (2008). Linolenic Acid and Risk of Nonfatal Acute Myocardial Infarction. *Circulation*, 118(4), 339–345.
- Capuzzo, A., Maffei, M. E., & Occhipinti, A. (2013). Supercritical Fluid Extraction of Plant Flavors and Fragrances. *Molecules*, *18*(6), 7194–7238.
- Catchpole, O. J., Tallon, S. J., Eltringham, W. E., Grey, J. B., Fenton, K. A., Vagi, E. M., & Zhu, Y. (2009). The extraction and fractionation of specialty lipids using near critical fluids. *The Journal of Supercritical Fluids*, 47(3), 591-597.
- Catchpole, O., Tallon, S., Dyer, P., Montanes, F., Moreno, T., Vagi, E., & Billakanti, J. (2012). Integrated Supercritical Fluid Extraction and Bioprocessing. *American Journal of Biochemistry and Biotechnology*, 8(4), 263–287.
- Çelik, M., Diler, A., & Küçükgülmez, A. (2005). A comparison of the proximate compositions and fatty acid profiles of zander (*Sander lucioperca*) from two different regions and climatic conditions. *Food Chemistry*, 92(4), 637-641.
- Chaiklahana, R., Chirasuwana, N., Lohab, V., & Bunnag, B. (2008). Lipid and fatty acids extraction from the cyanobacterium Spirulina. *Science Asia*, 34, 299-305.
- Chang, C. Y., Ke, D. S., & Chen, J. Y. (2009). Essential fatty acids and human brain. Acta neurologica Taiwanica, 18(4), 231-41.
- Chang, M. I., Puder, M., & Gura, K. M. (2012). The use of fish oil lipid emulsion in the treatment of intestinal failure associated liver disease (IFALD). *Nutrients*, 4(12), 1828-1850.
- Chantachum, S., Benjakul, S., & Sriwirat, N. (2000). Separation and quality of fish oil from precooked and non-precooked tuna heads. *Food Chemistry*, 69(3), 289-294.
- Chapman, F. A. (2000). Culture of hybrid tilapia: A reference profile. University of Florida Cooperative Extension Service, Institute of Food and Agriculture Sciences, EDIS.
- Choe, E., & Min. (2006). Mechanisms and factors for edible oil oxidation. *Comprehensive Reviews in Food Science and Food Safety*, 5(4), 169–186.
- Christie, W. W. (1993). Preparation of ester derivatives of fatty acids for chromatographic analysis. *Advances in Lipid Methodology*, 2(69), e111.
- Chung, S. T., & Shing, K. S. (1992). Multiphase behavior of binary and ternary systems of heavy aromatic hydrocarbons with supercritical carbon dioxide. *Fluid Phase Equilibria*, 81, 321–341.
- Clement, S., & Lovell, R. T. (1994). Comparison of processing yield and nutrient composition of cultured Nile tilapia (*Oreochromis niloticus*) and channel catfish (*Ictalurus punctatus*). *Aquaculture*, 119(2), 299–310.

- Connor, W. E. (2000). Importance of n-3 fatty acids in health and disease. *The American Journal of Clinical Nutrition*, 71(1), 171S–5S.
- Curtis, C. L., Harwood, J. L., Dent, C. M., & Caterson, B. (2004). Biological basis for the benefit of nutraceutical supplementation in arthritis levels to explain their mechanisms of action. *Drug Discovery Today*, *9*(4), 165–172.
- Das, S., & Bera, D. (2013). Mathematical model study on solvent extraction of carotene from carrot. *International Journal of Research in Engineering and Technology*, 2(9), 343–349.
- Das, S., Paul, B. N., Sengupta, J., & Datta, A. K. (2009). Beneficial effects of fish oil to human health : A Review. *Agricultural Reviews*, *30*(3), 199–205.
- Deepika, D., Vegneshwaran, V. R., Julia, P., Sukhinder, K. C., Sheila, T., Heather, M., & Wade, M. (2015). Investigation on Oil Extraction Methods and its Influence on Omega-3 Content from Cultured Salmon. *Journal of Food Processing & Technology*, 2014.
- Dejoye Tanzi, C., Abert Vian, M., Ginies, C., Elmaataoui, M., & Chemat, F. (2012). Terpenes as green solvents for extraction of oil from microalgae. *Molecules*, *17*(12), 8196–8205.
- Dempson, J. B., Schwarz, C. J., Shears, M., & Furey, G. (2004). Comparative proximate body composition of Atlantic salmon with emphasis on parr from fluvial and lacustrine habitats. *Journal of Fish Biology*, 64(5), 1257–1271.
- Division, P. B. (2002). Response surface optimization of the critical medium components for the production of alkaline protease by a newly isolated *Bacillus sp. Journal of Pharmacy and Pharmaceutical Sciences*, 5(3), 272–278.
- Döker, O., Salgin, U., Yildiz, N., Aydoğmuş, M., & Çalimli, A. (2010). Extraction of sesame seed oil using supercritical CO₂ and mathematical modeling. *Journal of Food Engineering*, 97(3), 360–366.
- Domingo, L., Bocio, A., Falc, G., & Llobet, J. M. (2007). Benefits and risks of fish consumption Part I . A quantitative analysis of the intake of omega-3 fatty acids and chemical contaminants. *Toxicology*, 230(2), 219–226.
- Drazen, J. C. (2007). Depth related trends in proximate composition of demersal fishes in the eastern North Pacific. *Deep-Sea Research Part I: Oceanographic Research Papers*, 54(2), 203–219.
- Dsikowitzky, L., Mengesha, M., Dadebo, E., de Carvalho, C. E. V., & Sindern, S. (2013). Assessment of heavy metals in water samples and tissues of edible fish species from Awassa and Koka Rift Valley Lakes, Ethiopia. *Environmental Monitoring and Assessment*, 185(4), 3117–3131.
- Dyerberg, J., & Sinclair, H. M. (1980). The composition of the Eskimo western food in north. *The American Journal of Clinical Nutrition*, *33*(12), 2657–2661.
- Egna, Hillary S & Boyd, C. E. (1997). Dynamics of pond aquaculture. CRC press.
- Eko, H. (1992). Fish oil: refining, stability and its use in canned fish for the Indonesian markek (Doctoral dissertation). Massey University, New Zealand.

- Elavarasan, K., Naveen Kumar, V., & Shamasundar, B. a. (2014). Antioxidant and Functional Properties of Fish Protein Hydrolysates from Fresh Water Carp (*Catla catla*) as Influenced by the Nature of Enzyme. *Journal of Food Processing and Preservation*, 38(3), 1207–1214.
- EL-Hawarry, W. N. (2012). Growth Performance, Proximate Muscle Composition and Dress-Out Percentage of Nile Tilapia (*Oreochromis niloticus*), Blue Tilapia (*Oreochromis aureus*) and their Interspecific Hybrid (♂ O. aureus and ♀ O. niloticus) Cultured in Semi-Intensive Culture System. World's Veterinary Journal, 2(2), 17–22.
- Eller, F. J., & King, J. W. (1996). Determination of fat content in foods by analytical SFE. In Seminars in food analysis (Vol. 1, No. 2, pp. 145-162).
- Endo, Yasushi and Tagiri-Endo, Misako & Kimura, K. (2005). Rapid Determination of Iodine Value and Saponification Value of Fish Oils by Near-Infrared Spectroscopy. *Journal of Food Science*, 70(30), 127–C131.
- Estiasih, T., Ahmadi, K., & Nisa, F. C. (2013). Optimizing Conditions for the Purification of Omega-3 Fatty Acids from the By-product of Tuna Canning Processing. *Journal of Food Science and Technology*, 5(5), 522–529.
- Estiasih, T., Ahmadi, K., Ginting, E., & Albab, A. U. (2013). Optimization of High EPA Structured Phospholipids Synthesis from ω -3 Fatty Acid Enriched Oil and Soy Lecithin. *Journal of Food Science and Engineering*, *3*, 25–32.
- Fallah, M., Bahram, S., & Javadian, S. R. (2015). Fish peptone development using enzymatic hydrolysis of silver carp by-products as a nitrogen source in *Staphylococcus aureus* media. *Food Science & Nutrition*, 3(2), 153–157.
- FAO. (2010). The state of world fisheries and aquaculture. Food and Agriculture Organization of the United Nations.
- Farag, R. S., & El-Anany, A. M. (2006). Improving the quality of fried oils by using different filter aids. *Journal of the Science of Food and Agriculture*, 86(13), 2228-2240.
- Ferdosh, S., Sarker, M. Z. I., Rahman, N. N. N. A., Akand, M. J. H., Ghafoor, K., Awang, M. Bin, & Kadir, M. O. A. (2013). Supercritical carbon dioxide extraction of oil from *Thunnus tonggol* head by optimization of process parameters using response surface methodology. *Korean Journal of Chemical Engineering*, 30(7), 1466–1472.
- Ferdosh, S., Sarker, Z. I., Norulaini, N., Oliveira, A., Yunus, K., Chowdury, A. J., & Omar, M. (2015). Quality of Tuna Fish Oils Extracted from Processing the By-Products of Three Species of Neritic Tuna Using Supercritical Carbon Dioxide. *Journal of Food Processing and Preservation*, 39(4), 432-441.
- Ferreira de França, L., Reber, G., Meireles, M. A. a, Machado, N. T., & Brunner, G. (1999). Supercritical extraction of carotenoids and lipids from buriti (*Mauritia flexuosa*), a fruit from the Amazon region. *The Journal of Supercritical Fluids*, 14(3), 247–256.
- Fiori, L., Solana, M., Tosi, P., Manfrini, M., Strim, C., & Guella, G. (2012). Lipid profiles of oil from trout (*Oncorhynchus mykiss*) heads, spines and viscera:

Trout by-products as a possible source of omega-3 lipids. *Food Chemistry*, 134(2), 1088–1095.

- Foh, MBK and Kamara, MT and Amadou, I and Foh, BM & Wenshui, X. (2011). Chemical and physicochemical properties of tilapia (*Oreochromis niloticus*) fish protein hydrolysate and concentrate. *International Journal of Biological Chemistry*, 5(1), 21–36.
- Food and Drug Administration. (2004). Mercury levels in commercial fish and shellfish. *Retrieved from httpy/wwwfda. gov/Food/FoodSafety/Product-SpecificInformation/Seafood/FoodbornePathogensContaminants/Methylmerc ury/ucml*, 15644.
- Gedi, M. A., Bakar, J., & Mariod, A. A. (2015). Optimization of supercritical carbon dioxide extraction of sardine (*Sardinella lemuru Bleeker*) oil using response surface methodology (RSM). *Intrenational journal of fats and oils*, 66(2), e074.
- Gelmez, N., Kincal, N. S., & Yener, M. E. (2009). Optimization of supercritical carbon dioxide extraction of antioxidants from roasted wheat germ based on yield, total phenolic and tocopherol contents, and antioxidant activities of the extracts. *Journal of Supercritical Fluids*, 48(3), 217–224.
- Gerbi, a, Zérouga, M., Debray, M., Durand, G., Chanez, C., & Bourre, J. M. (1994). Effect of fish oil diet on fatty acid composition of phospholipids of brain membranes and on kinetic properties of Na+,K(+)-ATPase isoenzymes of weaned and adult rats. *Journal of Neurochemistry*, 62(4), 1560–9.
- Ghaly, A. E., Ramakrishnan, V. V., Brooks, M. S., Budge, S. M., & Dave, D. (2013). Fish processing wastes as a potential source of proteins, amino acids and oils: a critical review. *Journal of Microbial & Biochemical Technology*, 05(04), 107–129.
- Ghazy, M. M. E., Habashy, M. M., Kossa, F. I., & Mohammady, E. Y. (2009). Effects of Salinity on Survival, Growth and Reproduction of the Water Flea, Daphnia magna. *Nature and Science*, 7(11), 28–42.
- Gokoglu, N., Yerlikaya, P., & Cengiz, E. (2004). Effects of cooking methods on the proximate composition and mineral contents of rainbow trout (*Oncorhynchus mykiss*). *Food Chemistry*, 84(1), 19–22.
- Goldburg, R., & Naylor, R. (2005). Future seascapes, fishing, and fish farming. *Frontiers in Ecology and the Environment*, 3(1), 21-28.
- Gracia, I., Garc n, M. T., Rodr guez, J. F., Fern andez, M. P., & de Lucas, a. (2009). Modelling of the phase behaviour for vegetable oils at supercritical conditions. *The Journal of Supercritical Fluids*, 48(3), 189–194.
- Gruger, E. H. (1967). Fatty Acid Composition of Fish Oils. United States. Bureau of Commercial Fisheries. Circular 276, 267.
- Guimarães, R., & Macedo, M. (2013). Sesame and flaxseed oil: nutritional quality and effects on serum lipids and glucose in rats. *Food Science and Technology*, 33(1), 209–217.
- Hajjar, T., Meng, G. Y., Rajion, M. a, Vidyadaran, S., Othman, F., Farjam, A. S., & Ebrahimi, M. (2012). Omega 3 polyunsaturated fatty acid improves spatial

learning and hippocampal Peroxisome Proliferator Activated Receptors (PPAR α and PPAR γ) gene expression in rats. *BMC Neuroscience*, 13(1), 109.

- Hall, S. J., & Mainprize, B. M. (2005). Managing by-catch and discards: how much progress are we making and how can we do better? *Fish and Fisheries*, 6(2), 134–155.
- Hao, L. P., Cao, X. J., & Hur, B. K. (2008). Separation of single component of EPA and DHA from fish oil using silver ion modified molecular sieve 13X under supercritical condition. *Journal of Industrial and Engineering Chemistry*, 14(5), 639-643.
- Hashim, N. I. K., Mustapha, N. I. K., Aziz, A. A. B. D., Mohd, N. I. K., & Hashim, H. (2013). Technical efficiency in aquaculture industry using data envelopment analysis (DEA) window : evidences from malaysia. *Journal of Sustainability Science and Management*, 8(2), 137–149.
- Hauthal, W. H. (2001). Advances with supercritical Fuids. *Chemosphere*, 43(1), 123–135.
- Herrero, M., Mendiola, J. a, Cifuentes, A., & Ib áñez, E. (2010). Supercritical fluid extraction: Recent advances and applications. *Journal of Chromatography*. *A*, *1217*(16), 2495–511.
- Heu, M. S., Kim, H. R., & Pyeun, J. H. (1995). Comparison of trypsin and chymotrypsin from the viscera of anchovy, *Engraulis japonica*. Comparative Biochemistry and Physiology. Part B, Biochemistry & Molecular Biology, 112(3), 557-567.
- Holub, B. J. (2009). Docosahexaenoic acid (DHA) and cardiovascular disease risk factors. *Prostaglandins, Leukotrienes, and Essential Fatty Acids*, 81(2), 199–204.
- Holub, D. J., & Holub, B. J. (2004). Omega-3 fatty acids from fish oils and cardiovascular disease. *Molecular and Cellular Biochemistry*, 263(1), 217–25.
- Homayooni, B., Sahari, M. A., & Barzegar, M. (2014). Concentrations of omega-3 fatty acids from rainbow sardine fish oil by various methods.*International Food Research Journal*, 21(2), 743-748.
- Horrocks, L. a, & Yeo, Y. K. (1999). Health benefits of docosahexaenoic acid (DHA). *Pharmacological Research : The Official Journal of the Italian Pharmacological Society*, 40(3), 211–225.
- Hossain, M. A. (2012). Fish as Source of n-3 Polyunsaturated Fatty Acids (PUFAs), Which One is Better Farmed or Wild. *Advance Journal of Food Science and Technology*, *3*(6), 455–466.
- Huang, J., & Sathivel, S. (2010). Purifying salmon oil using adsorption, neutralization, and a combined neutralization and adsorption process. *Journal* of Food Engineering, 96(1), 51-58.
- Huynh, M. D., & Kitts, D. D. (2009). Evaluating nutritional quality of pacific fish species from fatty acid signatures. *Food Chemistry*, 114(3), 912–918.

- İbrahim Haliloğlu, H., Bayır, A., Necdet Sirkecioğlu, a., Mevlüt Aras, N., & Atamanalp, M. (2004). Comparison of fatty acid composition in some tissues of rainbow trout (*Oncorhynchus mykiss*) living in seawater and freshwater. *Food Chemistry*, 86(1), 55–59.
- Innis, S. M., & Elias, S. L. (2003). Intakes of essential n- 6 and n- 3 polyunsaturated fatty acids among pregnant Canadian women. *The American journal of clinical nutrition*, 77(2), 473-478.
- Irwandi, J., Faridayanti, S., Mohamed, E. S. M., Hamzah, M. S., Torla, H. H., & Che Man, Y. B. (2009). Extraction and characterization of gelatin from different marine fish species in Malaysia. *International Food Research Journal*, 16(3), 381–389.
- IUPAC Official Method. (1987). Standard Method, Preparation of fatty acid methyl ester, In: Standard methods for the analysis of oils, fats and derivatives. Oxford, Blackwell.
- Jabeen, F., & Shakoor, A. (2011). Chemical compositions and fatty acid profiles of three freshwater fish species. *Food Chemistry*, 125(3), 991–996.
- Jayathilakan, K., Sultana, K., Radhakrishna, K., & Bawa, A. S. (2012). Utilization of byproducts and waste materials from meat, poultry and fish processing industries: a review. *Journal of food science and technology*, 49(3), 278-293.
- Jeppesen, P. B., Høy, C. E., & Mortensen, P. B. (2000). Deficiencies of essential fatty acids, vitamin A and E and changes in plasma lipoproteins in patients with reduced fat absorption or intestinal failure. *European Journal of Clinical Nutrition*, 54(8), 632–642.
- Justi, K. C., Hayashi, C., Visentainer, J. V., De Souza, N. E., & Matsushita, M. (2003). The influence of feed supply time on the fatty acid profile of Nile tilapia (*Oreochromis niloticus*) fed on a diet enriched with n-3 fatty acids. *Food Chemistry*, 80(4), 489–493.
- Karapanagiotidis, I. T., Bell, M. V. Little, D. C., Yakupitiyage, A., & Rakshit, S. K. (2006). Polyunsaturated fatty acid content of wild and farmed tilapias in Thailand: effect of aquaculture practices and implications for human nutrition. *Journal of Agricultural and Food Chemistry*, 54(12), 4304–10.
- Karlsson, Jens OM & Toner, M. (1996). Long-term storage of tissues by cryopreservation: critical issues. *Biomaterials*, 17(3), 243–256.
- Kaufmann, B., & Christen, P. (2002). Recent extraction techniques for natural products: microwave-assisted extraction and pressurised solvent extraction. *Phytochemical analysis*, *13*(2), 105-113.
- Khan, S., Misra, A. K., Tripathi, C. K. M., Mishra, B. N., & Bihari, V. (2006). Response surface optimization of effective medium constituents for the production of alkaline protease from a newly isolated strain of Pseudomonas aeruginosa. *Indian Journal of Experimental Biology*, 44(2), 151–156.
- Khoddami, A., Ariffin, A. A., Bakar, J., & Ghazali, H. M. (2009). Fatty acid profile of the oil extracted from fish waste (head, intestine and liver) (*Sardinella lemuru*). *World Applied Sciences Journal*, 7(1), 127-131.

- Kiessling, A., Pickova, J., Eales, J. G., Dosanjh, B., & Higgs, D. (2005). Age, ration level, and exercise affect the fatty acid profile of chinook salmon (*Oncorhynchus tshawytscha*) muscle differently. *Aquaculture*, 243(1-4), 345–356.
- Kim, H. J., Lee, S. B., Park, K. A., & Hong, I. K. (1999). Characterization of extraction and separation of rice bran oil rich in EFA using SFE process. *Separation and Purification Technology*, 15(1), 1–8.
- Kochman, K., & Czauderna, M. (2010). The necessity of adequate nutrition with diets containing omega-3 and omega-6 fatty acids for proper brain development, function and delayed aging: Review. Ournal of Animal and Feed Sciences, 19(4), 511–524.
- Kolakowska, A., & Sikorski, Z. E. (Eds.). (2003). Chemical and functional properties of food lipids (pp. 133-166). New York: CRC Press.
- Kris-Etherton, P. M. (2003). Fish Consumption, Fish Oil, Omega-3 Fatty Acids, and Cardiovascular Disease. Arteriosclerosis, Thrombosis, and Vascular Biology, 23(2), 20e–30.
- Kris-Etherton, P. M., Grieger, J. A., & Etherton, T. D. (2009). Dietary reference intakes for DHA and EPA. *Prostaglandins, Leukotrienes and Essential Fatty Acids*, 81(2), 99-104.
- Krishnan, VCA., Kuriakose, S., & Rawson, A. (2015). Ultrasound Assisted Extraction of Oil from Rice Bran: A Response Surface Methodology Approach. Journal of Food Processing & Technology & Technology.
- Kristinsson, H. G., & Rasco, B. a. (2000). Biochemical and functional properties of Atlantic salmon (*Salmo salar*) muscle proteins hydrolyzed with various alkaline proteases. *Journal of Agricultural and Food Chemistry*, 48(3), 657– 666.
- Kroes, R., Schaefer, E. J., Squire, R. a., & Williams, G. M. (2003). A review of the safety of DHA45-oil. *Food and Chemical Toxicology*, *41*(11), 1433–1446.
- Kromhout, D., Yasuda, S., Geleijnse, J. M., & Shimokawa, H. (2012). Fish oil and omega-3 fatty acids in cardiovascular disease: do they really work?. *European Heart Journal*, 33(4), 436–43.
- Kumar, R. V. (2014). Supercritical Fluid Chromatography: an Overview. International Journal of Pharmaceutical Research & Analysis, 4(7), 408–414.
- Lang, Q., & Wai, C. M. (2001). Supercritical fluid extraction in herbal and natural product studies: a practical review. *Talanta*, *53*(4), 771–782.
- Lee, J., Asaduzzaman, A. K. M., Yun, J., Yun, J., & Chun, B. (2012). Characterization of the yellow croaker larimichthys polyactis muscle oil extracted with supercritical carbon dioxide and an organic solvent. *Fisheries* and Aquatic Sciences, 15(4), 275–281.
- Lenihan-Geels, G., Bishop, K. S., & Ferguson, L. R. (2013). Alternative sources of omega-3 fats: can we find a sustainable substitute for fish?. *Nutrients*, 5(4), 1301–15.

- Létisse, M., Rozieres, M., Hiol, A., Sergent, M., & Comeau, L. (2006). Enrichment of EPA and DHA from sardine by supercritical fluid extraction without organic modifier: I. Optimization of extraction conditions. *The Journal of supercritical fluids*, 38(1), 27-36.
- L étisse, M., Rozi ères, M., Hiol, A., Sergent, M., & Comeau, L. (2006). Enrichment of EPA and DHA from sardine by supercritical fluid extraction without organic modifier. *The Journal of Supercritical Fluids*, 38(1), 27–36.
- Li, Q., Brendemuhl, J. H., Jeong, K. C., & Badinga, L. (2014). Effects of dietary omega -3 polyunsaturated fatty acids on growth and immune response of weanling pigs. *Ournal of Animal Science and Technology*, 56(1), 1–7.
- Lim, H.K., Tan, C.P., Bakar, J., & Ng, S.P. (2011). Effects of Different Wall Materials on the Physicochemical Properties and Oxidative Stability of Spray-Dried Microencapsulated Red-Fleshed Pitaya (*Hylocereus polyrhizus*) Seed Oil. *Food and Bioprocess Technology*, 5(4), 1220–1227.
- Linder, M., Fanni, J., & Parmentier, M. (2005). Proteolytic extraction of salmon oil and PUFA concentration by lipases. *Marine Biotechnology*, 7(1), 70-76.
- Liu, S., Zhang, C., Hong, P., & Ji, H. (2006). Concentration of docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA) of tuna oil by urea complexation: optimization of process parameters. *Journal of food engineering*, 73(3), 203-209.
- Lopes, C., Antelo, L. T., Franco-Ur á, A., Alonso, A. A., & Pérez-Mart n, R. (2015). Valorisation of fish by-products against waste management treatments: Comparison of environmental impacts. *Waste Management*, 46(2), 103–112.
- Łuczyńska, J., Paszczyk, B., & Łuczyński, M. J. (2014). Fatty acid profiles in marine and freshwater fish from fish markets in northeastern Poland. Archives of Polish Fisheries, 22(3), 181–188.
- Luque de Castro, M. D., & Jim énez-Carmona, M. M. (2000). Where is supercritical fluid extraction going?. *Trends in Analytical Chemistry*, 19(4), 223–228.
- Madamba, P. S. (2002). The response surface methodology: An application to optimize dehydration operations of selected agricultural crops. *Food Science and Technology*, *35*(7), 584–592.
- Maheshwari, P., Nikolov, Z. L., White, T. M., & Hartel, R. (1992). Solubility of Fatty Acids in Supercritical Carbon Dioxide. *Journal of the American Oil Chemists' Society*, 69(11), 1069–1076.
- Manresa, A., & Bastida, J. (2013). Application of Experimental Design and Canonical Analysis of Response Surfaces to the Optimization of Poly (3hydroxyalkanoates) Production by Pseudomonas aeruginosa 42A2. *Chemical and Biochemical Engineering Quarterly*, 27(4), 457–465.
- Maqsood, S., Benjakul, S., & Kamal-Eldin, A. (2012). Extraction, Processing, and Stabilization of Health-Promoting Fish Oils. *Recent Patents on Food*, *Nutrition & Agriculturee*, 4(2), 141–147.
- Marrone, C., Poletto, M., Reverchon, E., & Stassi, a. (1998). Almond oil extraction by supercritical CO₂: experiments and modelling. *Chemical Engineering Science*, *53*(21), 3711–3718.

- Mart nez, M. L., Mattea, M. a., & Maestri, D. M. (2008). Pressing and supercritical carbon dioxide extraction of walnut oil. *Journal of Food Engineering*, 88(3), 399–404.
- Mead, J. F. (1984). The non-eicosanoid functions of the essential fatty acids. *Journal* of Lipid Research, 25(13), 1517–1521.
- Melho Filho, A. A., Oliveira, H. H., & Santos, R. C. (2013). Omega-6/Omega-3 and PUFA/SFA in Colossoma macropomum grown in Roraima, Brazil. *Journal* of Chemistry, 5(1), 30-34.
- Memon, N. N., Talpur, F. N., Sherazi, S. T. H., & Bhanger, M. I. (2010). Impact of refrigerated storage on quality of oil from freshwater Jarko (*Wallago attu*) fish. *Pakistan Journal of Analytical Environmental Chemistry*, 11(2), 37–43.
- Meyer, B. J., Mann, N. J., Lewis, J. L., Milligan, G. C., Sinclair, A. J., & Howe, P. R. C. (2003). Dietary intakes and food sources of omega-6 and omega-3 polyunsaturated fatty acids. *Lipids*, 38(4), 391–8.
- Mizuno, T., Goto, M., Kodama, A., & Hirose, T. (2000). Supercritical water oxidation of a model municipal solid waste. *Industrial & engineering chemistry research*, 39(8), 2807-2810.
- Mohamad, R., Abidin, Z. Z., & Rusli, R. (2010). Consumer preference towards fresh water fish product developed by MARDI. *Economic and Technology Management Review*, 5(571), 71–77.
- Mohamed, R. S., & Mansoori, G. A. (2002). The Use of Supercritical Fluid Extraction Technology in Food Processing. *Food Technology Magazine*, 20(7), 134–139.
- Moret, S., & Conte, L. S. (2000). Polycyclic aromatic hydrocarbons in edible fats and oils: Occurrence and analytical methods. *Journal of Chromatography A*, 882, 245–253.
- Nagai, T., & Suzuki, N. (2000). Isolation of collagen from fish waste material skin, bone and fins. *Food Chemistry*, 68(3), 277–281.
- Najafabadi, H. J., Moghaddam, H. N., Pourreza, J., Shahroudi, F. E., & Golian, A. (2007). Determination of chemical composition, mineral contents, and protein quality of poultry by-product meal. *International Journal of Poultry Science*, 6(12), 875–882.
- Nakamura, Y.-N., Ando, M., Seoka, M., Kawasaki, K., & Tsukamasa, Y. (2007). Changes of proximate and fatty acid compositions of the dorsal and ventral ordinary muscles of the full-cycle cultured Pacific bluefin tuna *Thunnus orientalis* with the growth. *Food Chemistry*, 103(1), 234–241.
- Naylor, R. L., Goldburg, R. J., Primavera, J. H., Kautsky, N., Beveridge, M. C. M., Clay, J., & Troell, M. (2000). Effect of aquaculture on world fish supplies. *Nature*, 405(6790), 1017–1024.
- Ng, W. K., Chong, C. Y., Wang, Y., & Romano, N. (2013). Effects of dietary fish and vegetable oils on the growth, tissue fatty acid composition, oxidative stability and vitamin E content of red hybrid tilapia and efficacy of using fish oil finishing diets. Aquaculture, 372, 97-110.

- Nyaradi, A., Li, J., Hickling, S., Foster, J., & Oddy, W. H. (2013). Prenatal and Childhood Nutrition: Evaluating the Neurocognitive Connections. CRC Press
- Ogawa, M., Portier, R. J., Moody, M. W., Bell, J., Schexnayder, M. a., & Losso, J. N. (2004). Biochemical properties of bone and scale collagens isolated from the subtropical fish black drum (*Pogonia cromis*) and sheepshead seabream (*Archosargus probatocephalus*). *Food Chemistry*, 88(4), 495–501.
- Oku, T., Sugawara, A., Choudhury, M., Komatsu, M., Yamada, S., & Ando, S. (2009). Lipid and fatty acid compositions differentiate between wild and cultured Japanese eel (*Anguilla japonica*). Food Chemistry, 115(2), 436–440.
- Oladipo, I. C., & Bankole, S. O. (2013). Nutritional and microbial quality of fresh and dried *Clarias gariepinus* and *Oreochromis niloticus*. *International Journal of Applied Microbiology and Biotechnology Research*, 1, 1–6.
- Oliveira, L. E., Barboza, J. C. S., & Da Silva, M. L. C. P. (2013). Production of ethylic biodiesel from Tilapia visceral oil. In Proceedings of the International Conference on Renewable Energies and Power Quality (ICREPQ'13) (pp. 20-22).
- Özkal, S. G., Yener, M. E., & Bayindirli, L. (2006). The solubility of apricot kernel oil in supercritical carbon dioxide. *International Journal of Food Science and Technology*, *41*(4), 399–404.
- Özogul, Y., Özogul, F., & Alagoz, S. (2007). Fatty acid profiles and fat contents of commercially important seawater and freshwater fish species of Turkey: A comparative study. *Food Chemistry*, 103(1), 217–223.
- Ozogul, Y., Simşek, A., Balikçi, E., & Kenar, M. (2012). The effects of extraction methods on the contents of fatty acids, especially EPA and DHA in marine lipids. *International Journal of Food Sciences and Nutrition*, 63(3), 326–31.
- Perrut, M. (2000). Supercritical fluid applications: industrial developments and economic issues. *Industrial & Engineering Chemistry Research*, 39(12), 4531–4535.
- Petenuci, M. E., Stevanato, F. B., Visentainer, J. E. L., Matsushita, M., Garcia, E. E., de Souza, N. E., & Visentainer, J. V. (2008). Fatty acid concentration, proximate composition, and mineral composition in fishbone flour of Nile Tilapia. Archivos Latinoamericanos de Nutrición, 58(1), 87–90.
- Pigott, G. M., & Tucker, B. (1990). Seafood: effects of technology on nutrition (Vol. 39). CRC press.
- Popma, T. J., & Lovshin, L. L. (1996). Worldwide prospects for commercial production of tilapia. International Center for Aquaculture and Aquatic Environments.
- Popma, T. J., & Lovshin, L. L. (1996). Worldwide prospects for commercial production of tilapia. International Center for Aquaculture and Aquatic Environments.
- Potaros, T., Raksakulthai, N., & Runglerdkreangkrai, J. (2009). Characteristics of Collagen from Nile Tilapia (*Oreochromis niloticus*) Skin Isolated by Two Different Methods. *Kasetsart Journal of Natural Science*, 593(43), 584–593.

- Pourmortazavi, S. M., & Hajimirsadeghi, S. S. (2007). Supercritical fluid extraction in plant essential and volatile oil analysis. *Journal of chromatography*, *1163*(1), 2-24.
- Ramezani-Fard, E., Kamarudin, M. S., Harmin, S. A., & Saad, C. R. (2012). Dietary saturated and omega-3 fatty acids affect growth and fatty acid profiles of Malaysian mahseer. *European Journal of Lipid Science and Technology*, 114(2), 185–193.
- Rasoarahona, J. R. E., Barnathan, G., Bianchini, J.-P., & Gaydou, E. M. (2005). Influence of season on the lipid content and fatty acid profiles of three tilapia species (*Oreochromis niloticus, O. macrochir* and Tilapia rendalli) from Madagascar. *Food Chemistry*, 91(4), 683–694.
- Reverchon, E., & De Marco, I. (2006). Supercritical fluid extraction and fractionation of natural matter. *The Journal of Supercritical Fluids*, *38*(2), 146–166.
- Rezaei, K. a., & Temelli, F. (2000). Using supercritical fluid chromatography to determine diffusion coefficients of lipids in supercritical CO₂. *The Journal of Supercritical Fluids*, 17(1), 35–44.
- Rubio-Rodr guez, N., de Diego, S. M., Beltr án, S., Jaime, I., Sanz, M. T., & Rovira, J. (2008). Supercritical fluid extraction of the omega-3 rich oil contained in hake (*Merluccius capensis–Merluccius paradoxus*) by-products: Study of the influence of process parameters on the extraction yield and oil quality. *The Journal of Supercritical Fluids*, 47(2), 215–226.
- Rubio-Rodr guez, N., de Diego, S. M., Beltr án, S., Jaime, I., Sanz, M. T., & Rovira, J. (2012). Supercritical fluid extraction of fish oil from fish by-products: A comparison with other extraction methods. *Journal of Food Engineering*, 109(2), 238–248.
- Russo, G. L. (2009). Dietary n-6 and n-3 polyunsaturated fatty acids: from biochemistry to clinical implications in cardiovascular prevention. *Biochemical Pharmacology*, 77(6), 937–46.
- Rustan, A. C., & Drevon, C. A. (2005). Fatty acids: structures and properties. *The* Official Journal of Encyclopedia of Life Sciences, 16(2), 662-41.
- Ruxton, C. H. S., Reed, S. C., Simpson, M. J. a, & Millington, K. J. (2004). The health benefits of omega-3 polyunsaturated fatty acids: a review of the evidence. *Journal of Human Nutrition and Dietetics : The Official Journal of the British Dietetic Association*, 17(5), 449–59.
- Sahena, F., Zaidul, I. S. M., Jinap, S., Yazid, A. M., Khatib, A., & Norulaini, N. A. N. (2010). Fatty acid compositions of fish oil extracted from different parts of Indian mackerel (*Rastrelliger kanagurta*) using various techniques of supercritical CO₂ extraction. *Food Chemistry*, 120(3), 879-885.
- Saify, Z. S., & Akhtar, S. (2003). A Study on the Fatty Acid Composition of Fish Liver Oil from Two Marine Fish , *Eusphyra blochii* and *Carcharhinus bleekeri*. *Turkish Journal of Chemistry*, 27(2), 251–258.
- Saldaña, M. I., Gómez-álvarez, R., Mar á, C., Álvarez-Sol á, J. D., Pat-fern ández, J. M., & Carlos, F. (2014). The influence of organic fertilizers on the chemical

properties of soil and the production of Alpinia purpurata. *Ciencia E Investigacion AGRARIA*, 41(2), 215–224.

- Sánchez-Vicente, Y., Cabañas, A., Renuncio, J. A., & Pando, C. (2009). Supercritical fluid extraction of peach (*Prunus persica*) seed oil using carbon dioxide and ethanol. *The Journal of Supercritical Fluids*, 49(2), 167-173.
- Sándor, Z., Papp, Z. G., Csengeri, I., & Jeney, Z. (2011). Fish meat quality and safety. *Tehnologija Mesa*, 52(1), 97–105.
- Santos, V. B., Mareco, E. A., & Silva, M. D. P. (2013). Growth curves of Nile tilapia (*Oreochromis niloticus*) strains cultivated at different temperatures. *Animal Sciences*, 35(3), 235–242.
- Sapkale, G. N., Patil, S. M., Surwase, U. S., & Bhatbhage, P. K. (2010). A review supercritical fluid extraction. *Journal of Chemical Sciences*, 8(2), 729–743.
- Sarker, M. Z. I., Selamat, J., Habib, A. S. M. A., Ferdosh, S., Akanda, M. J. H., & Jaffri, J. M. (2012). Optimization of Supercritical CO₂ Extraction of Fish Oil from Viscera of African Catfish (*Clarias gariepinus*). *International Journal* of Molecular Sciences, 13(9), 11312–22.
- Sathivel, S., Prinyawiwatkul, W., Grimm, C. C., King, J. M., & Lloyd, S. (2002). FA composition of crude oil recovered from catfish viscera. *Journal of the American Oil Chemists' Society*, *79*(10), 989–992.
- Sathivel, S., Prinyawiwatkul, W., King, J. M., Grimm, C. C., & Lloyd, S. (2003). Oil production from catfish viscera. *Journal of the American Oil Chemists' Society*, 80(4), 377–382.
- Schakel, S. F., Buzzard, I. M., & Gebhardt, S. E. (1997). Procedures for estimating nutrient values for food composition databases. *Journal of Food Composition and Analysis*, *10*(2), 102-114.
- Shahidi, F., & Wanasundara, N. (1998). Omega-3 fatty acid concentrates: nutritional aspects and production technologies. *Trends in Food Science & Technology*, 9(6), 230–240.
- Sheibani, A., & Ghaziaskar, H. S. (2008). Pressurized fluid extraction of pistachio oil using a modified supercritical fluid extractor and factorial design for optimization. *Food Science and Technology*, *41*(8), 1472–1477.
- Shirai, N., Suzuki, H., Tokairin, S., Ehara, H., & Wada, S. (2002). Dietary and seasonal effects on the dorsal meat lipid composition of Japanese (*Silurus asotus*) and Thai catfish (*Clarias macrocephalus and hybrid Clarias macrocephalus and Clarias galipinus*). Comparative Biochemistry and Physiology Part A: *Molecular & Integrative Physiology*, 132(3), 609-619.
- Shirai, N., Terayama, M., & Takeda, H. (2002). Effect of season on the fatty acid composition and free amino acid content of the sardine Sardinops melanostictus. Comparative Biochemistry and Physiology. *Biochemistry and Molecular Biology*, 131(3), 387-393.
- Silva, S. S. De, & Turchini, G. M. (2009). Use of wild fish and other aquatic organisms as feed in aquaculture – a review of practices and implications in the Asia-Pacific. FAO Fisheries and aquaculture technical paper, 51(8), 63– 127.

- Simopoulos, A. P. (1991). Omega-3 fatty acids in health and disease and in growth and development. *The American Journal of Clinical Nutrition*, 54(3), 438–63.
- Simopoulos, A. P. (2001). Evolutionary aspects of diet, essential fatty acids and cardiovascular disease. *European Heart Journal-Supplements*, 3(8), 8–21.
- Simopoulos, A. P. (2002). Omega-3 Fatty Acids in Inflammation and Autoimmune Diseases. *Journal of the American College of Nutrition*, 21(6), 495–505.
- Simopoulos, A. P. (2009). Omega-6/omega-3 essential fatty acids: biological effects. *World Review of Nutrition and Dietetics*, *99*(2), 1–16.
- Sinclair, A. J., Hons, D. B., Mathai, M., & Weisinger, R. S. (2007). Omega 3 fatty acids and the brain : review of studies in depression. *Asia Pacific Journal of Clinical Nutrition*, 16(S1), 391–397.
- Singh, A. K., Verma, P., Srivastava, S. C., & Tripathi, M. (2014). Invasion, Biology and impact of feral population of Nile tilapia (*Oreochromis niloticus Linnaeus*) in the Ganga River (India). Asia Pacific Journal of Research Vol: I Issue XIV.
- Singh, K. P., Gupta, S., Singh, A. K., & Sinha, S. (2011). Optimizing adsorption of crystal violet dye from water by magnetic nanocomposite using response surface modeling approach. *Journal of Hazardous Materials*, 186(2-3), 1462–73.
- Singh, M. (2005). Essential fatty acids, DHA and human brain. *The Indian Journal* of Pediatrics, 72(3), 239–242.
- Siti Sarah, J., Ariff, O. M., & Goh, Y. M. (2013). Fatty acid profile of Nile and Red hybrid tilapias reared in intensive and extensive systems. *Malaysian Journal of Animal Science*, *16*(1), 13-21.
- Stevanato, F. B., Almeida, V. V., Matsushita, M., Oliveira, C. C., Souza, N. E., & Visentainer, J. V. (2008). Fatty acids and nutrients in the flour made from tilapia (*Oreochromis niloticus*) heads. *Food Science and Technology* (*Campinas*), 28(2), 440-443.
- Straccia, M. C., Siano, F., Coppola, R., Cara, L., & Volpe, G. (2012). Extraction and Characterization of Vegetable Oils from Cherry Seed by Different Extraction Processes. *Chemical engineering transactions*, 27, 391–396.
- Sullivan, J. C., & Budge, S. M. (2010). Monitoring fish oil volatiles to assess the quality of fish oil. *Lipid Technology*, 22(10), 230–232.
- Suloma, A., Ogata, H. Y., Garibay, E. S., Chavez, D. R., & El-Haroun, E. R. (2008). Fatty acid composition of Nile tilapia *Oreochromis niloticusmuscles*: a comparative study with commercially important tropical freshwater fish in Philippines. In 8th International Symposium on Tilapia in Aquaculture. Cairo: Egypt (pp. 921-932).
- Sun, M., & Temelli, F. (2006). Supercritical carbon dioxide extraction of carotenoids from carrot using canola oil as a continuous co-solvent. *Journal of Supercritical Fluids*, 37(3), 397–408.

- Suseno, S. H., Yang, T. A., & Nadiah, W. (2015). Physicochemical Characteristics and Quality Parameters of Alkali-Refined Lemuru Oil from Banyuwangi, Indonesia. *Pakistan Journal of Nutrition*, 14(2), 107–111.
- Tacon, A. G. J., & Metian, M. (2008). Global overview on the use of fi sh meal and fi sh oil in industrially compounded aquafeeds : Trends and future prospects. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 285(1-4), 146– 158.
- Tan, X. Y., Luo, Z., Xie, P., & Liu, X. J. (2009). Effect of dietary linolenic acid/linoleic acid ratio on growth performance, hepatic fatty acid profiles and intermediary metabolism of juvenile yellow catfish *Pelteobagrus fulvidraco*. *Aquaculture*, 296(1-2), 96–101.
- Tawari, C. C., Island, W., State, B., & Production, L. (2011). Some Basic Principles of Fish processing in Nigeria. Asian Journal of Agricultural Sciences, 3(6), 437–452.
- Teoh, C. Y., Turchini, G. M., & Ng, W. K. (2011). Genetically improved farmed Nile tilapia and red hybrid tilapia showed differences in fatty acid metabolism when fed diets with added fish oil or a vegetable oil blend. *Aquaculture*, 316(1-4), 144–154.
- Thammapat, P., Raviyan, P., & Siriamornpun, S. (2010). Proximate and fatty acids composition of the muscles and viscera of Asian catfish (*Pangasius bocourti*). Food Chemistry, 122(1), 223–227.
- Thana, P., Machmudah, S., Goto, M., Sasaki, M., Pavasant, P., & Shotipruk, A. (2008). Response surface methodology to supercritical carbon dioxide extraction of astaxanthin from *Haematococcus pluvialis*. *Bioresource Technology*, 99(8), 3110–3115.
- Tocher, D. R., Fonseca-Madrigal, J., Dick, J. R., Ng, W. K., Bell, J. G., & Campbell, P. J. (2004). Effects of water temperature and diets containing palm oil on fatty acid desaturation and oxidation in hepatocytes and intestinal enterocytes of rainbow trout (*Oncorhynchus mykiss*). *Biochemistry and Molecular Biology*, 137(1), 49-63.
- Topare, N. S., Raut, S. J., Renge, V. C., Khedkar, S. V., Chavanand, Y. P., & Bhagat, S. L. (2011). Extraction of oil from algae by solvent extraction and oil expeller method. *International Journal of Chemical Sciences*, 9(4), 1746-1750.
- Ugoala, C., Ndukwe, G. I., & Audu, T. O. (2008). Comparison of fatty acids profile of some freshwater and marine fishes. *Internet Journal of Food Safety*, *10*, 9-17.
- Vannuccini, S. (2005). Implication for FAO fish consumption statistics of trash fish utilization in the Asia-Pacific region. *Paper Presented at Regional Workshop on Low Value and Trash Fish in the Asia-Pacific Region*, 248.
- Vasapollo, G., Longo, L., Rescio, L., & Ciurlia, L. (2004). Innovative supercritical CO₂ extraction of lycopene from tomato in the presence of vegetable oil as co-solvent. *Journal of Supercritical Fluids*, 29(1-2), 87–96.

- Vegneshwaran VR, D. D. (2014). Investigation on Oil Extraction Methods and its Influence on Omega-3 Content from Cultured Salmon. *Journal of Food Processing & Technology*, 5(12).
- Wang, L., & Weller, C. L. (2006). Recent advances in extraction of nutraceuticals from plants. *Trends in Food Science & Technology*, 17(6), 300–312.
- Wasowicz, E., Gramza, A., Hes, M., Jelen, H. H., Korczak, J., Malecka, M. (2004). Oxidation of lipids in food. *Journal of Food and Nutrition Sciences*, 13(61), 87–100.
- Wei, Z. J., Liao, A. M., Zhang, H. X., Liu, J., & Jiang, S. T. (2009). Optimization of supercritical carbon dioxide extraction of silkworm pupal oil applying the response surface methodology. *Bioresource Technology*, 100(18), 4214-4219.
- Wenli, Y., Yaping, Z., Jingjing, C., & Bo, S. (2004). Comparison of two kinds of pumpkin seed oils obtained by supercritical CO₂ extraction. *European Journal of Lipid Science and Technology*, 106(6), 355–358.
- William W, C. (1993). Preparation of lipid extracts from tissues. *Advances in Lipid Methodology*, 2, 195–213.
- Wu, D., Sun, D. W., & He, Y. (2012). Application of long-wave near infrared hyperspectral imaging for measurement of color distribution in salmon fillet. *Innovative Food Science and Emerging Technologies*, 16, 361–372.
- Wu, T. H., & Bechtel, P. J. (2008). Salmon by-product storage and oil extraction. *Food Chemistry*, 111(4), 868-871.
- Xu, L., Zhan, X., Zeng, Z., Chen, R., Li, H., Xie, T., & Wang, S. (2011). Recent advances on supercritical fluid extraction of essential *Pharmacy and Pharmacology*, 5(9), 1196-1211.
- Yahyaee, R., Ghobadian, B., & Najafi, G. (2013). Waste fish oil biodiesel as a source of renewable fuel in Iran. *Renewable and Sustainable Energy Reviews*, 17(6), 312–319.
- Yones, A. M., El-Saidy, D. M. S. D., & Abdel-Hakim, N. F. (2013). Effects of fish oil substitution with vegetable oils in diets of juvenile Nile tilapia, *Oreochromis niloticus* (L.) on growth performance, nutrients utilization and muscle fatty acids contents. *Merit Research Journal of Food Science and Technology*, 1(1), 9-18.
- Yoshida, H., Terashima, M., & Takahashi, Y. (1999). Production of organic acids and amino acids from fish meat by subcritical water hydrolysis. *Biotechnology Progress*, 15(6), 1090–4.
- Young, K. (2009). Omega-6 (n-6) and omega-3 (n-3) fatty acids in tilapia and human health: a review. *International journal of food sciences and nutrition*, 60(5), 203-211.
- Yu, Z., Singh, B., Rizvi, S., & Zollweg, J. (1994). Solubilities of fatty acids, fatty acid esters, triglycerides, and fats and oils in supercritical carbon dioxide. *The Journal of Supercritical Fluids*, 7(1), 51–59.

- Yubin, J. I., Miao, Y., Bing, W., & Yao, Z. (2014). The extraction, separation and purification of alkaloids in the natural medicine. *Journal of Chemical and Pharmaceutical Research*, 6(1), 338–345.
- Zaidul, M., Sarker, I., Selamat, J., Sayem, A., & Habib, A. (2012). Optimization of supercritical CO₂ extraction of fish oil from viscera of african catfish (*Clarias* gariepinus). International Journal of Molecular Sciences, 13(9), 11312– 11322.
- Zenebe, T., Ahlgren, G., Gustafsson, I. B., & Boberg, M. (1998). Fatty acid and lipid content of *Oreochromis niloticus L*. in Ethiopian lakes dietary effects of phytoplankton. *Ecology of Freshwater Fish*, 7(3), 146–158.
- Zhao, S., & Zhang, D. (2013). Supercritical fluid extraction and characterisation of Moringa oleifera leaves oil. Separation and Purification Technology, 118, 497-502.
- Zuta, C. P., Simpson, B. K., Chan, H. M., & Phillips, L. (2003). Concentrating PUFA from mackerel processing waste. *Journal of the American Oil Chemists' Society*, 80(9), 933–936.