



UNIVERSITI PUTRA MALAYSIA

***PREVALENCE, CHARACTERIZATION AND ANTIBIOTIC RESISTANCE
OF Salmonella spp. ISOLATED FROM RAW VEGETABLES***

NAJWA BINTI MOHD SHAHRIL

FSTM 2016 12



**PREVALENCE, CHARACTERIZATION AND ANTIBIOTIC RESISTANCE
OF *Salmonella* spp. ISOLATED FROM RAW VEGETABLES**

By

NAJWA BINTI MOHD SHAHRIL

**Thesis Submitted to the School of Graduate Studies,
Universiti Putra Malaysia, in Fulfillment of the
Requirements for the Degree of Master of Science**

August 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

**Dedicated to my beloved parents, siblings and friends for their eternity love
and endless support**



© COPYRIGHT UPM

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Masters of Science

**PREVALENCE, CHARACTERIZATION AND ANTIBIOTIC RESISTANCE
OF *Salmonella* spp. ISOLATED FROM RAW VEGETABLES**

By

NAJWA BINTI MOHD SHAHRIL

August 2016

Chairman : Professor Son Radu, PhD
Faculty : Food Science and Technology

Growing consumption of fresh vegetables has led to an increase in the number of outbreaks of food-borne disease linked to fresh produce including salmonellosis. In Malaysia, some vegetables are eaten raw, known as *ulam* or salad in other countries. Therefore, this research was conducted to study the prevalence and antibiotic resistance of *Salmonella* spp., *Salmonella* Enteritidis (*S. Enteritidis*) and *Salmonella* Typhimurium (*S. Typhimurium*) in *ulam*. This study also aimed to determine the relation among different isolates of same serovar of *Salmonella* spp. by employing random amplification of polymorphic DNA-polymerase chain reaction (RAPD-PCR) method and the risk acquiring salmonellosis through consumption of *ulam*.

A total of 96 samples of *ulam* were purchased from wet markets and hypermarkets in limited geographical locations of Selangor. Most probable number (MPN) method was combined with multiplex polymerase chain reaction (PCR) and plating method for detection of *Salmonella* spp. Prevalence of *Salmonella* spp., *S. Enteritidis* and *S. Typhimurium* was higher in samples from hypermarkets than wet markets which were 100%, 64.6% and 87.5% respectively at hypermarkets and 95.8%, 43.8% and 75% respectively at wet markets. The density of *Salmonella* spp., *S. Enteritidis* and *S. Typhimurium* ranged from <3 to >2400 MPN/g. MPN-multiplex PCR detected more *Salmonella* spp., *S. Enteritidis* and *S. Typhimurium* which were 97.9%, 54.2% and 81.3% respectively, as compared to MPN-plating which were 7.3%, 6.3% and 10.4% respectively.

Six isolates of *S. Enteritidis* and 10 isolates of *S. Typhimurium* were recovered from *ulam*. Both *Salmonella* spp. serovars showed high resistance to amoxicillin/clavunic acid, cephalothin, streptomycin and ciprofloxacin with 100% resistant to ampicillin and erythromycin. All isolates showed resistance to at least three antibiotics tested. *S. Typhimurium* was more resistant compared to *S. Enteritidis* with multiple antibiotic resistances (MAR) index ranging from 0.27 to 0.82 and 0.27 to 0.55 respectively.

RAPD-PCR dendograms showed that most isolates of *S. Enteritidis* and *S. Typhimurium* tend to type together based on same sample type which 3 RAPD types (RAPD type 3 of *S. Enteritidis* and RAPD type 1 and 6 of *S. Typhimurium*) were typed together respectively on same sample type and same sampling locations. The other 3 RAPD types (RAPD type 1 of *S. Enteritidis*, RAPD type 2 and 4 of *S. Typhimurium*) were typed based on the same sample type but different sampling locations. RAPD type 3 of *S. Typhimurium* was typed on different sample types but same sampling location was observed.

The quantitative microbiological risk assessment (QMRA) simulations by single-hit concept showed that overall risk of acquiring salmonellosis due to consumption of *ulam* was higher for *S. Typhimurium* than *S. Enteritidis*. The mean risk of illness per serving per year for *S. Typhimurium* was 1 while for *S. Enteritidis*, the risk was ~1 in the exposed population.

All in all, the prevalence of *Salmonella* spp. in *ulam* is alarming and should be monitored from time to time seems prevention is better than cure.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

**PREVALENS, PENCIRIAN DAN RINTANGAN ANTIBIOTIK *Salmonella*
spp. YANG DIPENCILKAN DARI SAYUR-SAYURAN MENTAH**

Oleh

NAJWA BINTI MOHD SHAHRIL

Ogos 2016

Pengerusi : Professor Son Radu, PhD
Fakulti : Sains dan Teknologi Makanan

Peningkatan penggunaan sayur-sayuran segar telah membawa peningkatan bilangan wabak penyakit bawaan makanan berkaitan dengan produk segar termasuk salmonellosis. Di Malaysia, beberapa sayur-sayuran dimakan secara mentah, dikenali sebagai ulam atau salad di negara-negara lain. Oleh itu, penyelidikan ini dijalankan untuk mengkaji prevalens dan rintangan antibiotik *Salmonella* spp., *Salmonella* Enteritidis (*S. Enteritidis*) dan *Salmonella* Typhimurium (*S. Typhimurium*) pada ulam. Kajian ini juga bertujuan untuk menentukan kaitan antara pencilan-pencilan yang berbeza pada serovar *Salmonella* spp. yang sama dengan menggunakan kaedah profil amplikasi poliformik DNA rawak- tindak balas berantai polimerasi (RAPD-PCR) dan risiko mendapat salmonellosis melalui penggunaan ulam.

Sejumlah 96 sampel ulam dibeli dari pasar dan pasar raya di tempat-tempat secara geografi yang terhad di Selangor. Kaedah jumlah paling mungkin (MPN) telah digabungkan dengan multipleks tindak balas berantai polimerasi (PCR) dan kaedah piring untuk mengesan *Salmonella* spp. Prevalens *Salmonella* spp., *S. Enteritidis* dan *S. Typhimurium* adalah lebih tinggi di pasar raya dari pasar di mana masing-masing 100%, 64.6% dan 87.5% di pasar raya dan masing-masing 95.8%, 43.8% dan 75% di pasar. Ketumpatan *Salmonella* spp., *S. Enteritidis* dan *S. Typhimurium* berada antara <3 sehingga >2400 MPN/g. MPN-multipleks PCR boleh mengesan lebih *Salmonella* spp., *S. Enteritidis* dan *S. Typhimurium* berbanding MPN-piring. MPN-multipleks PCR mengesan masing-masing 97.9%, 54.2% dan 81.3% *Salmonella* spp., *S. Enteritidis* dan *S. Typhimurium* sementara MPN-pring mengesan masing-masing hanya 7.3%, 6.3% dan 10.4% *Salmonella* spp., *S. Enteritidis* dan *S. Typhimurium*.

Enam belas pencilan *S. Enteritidis* dan *S. Typhimurium* telah didapati daripada ulam. Kedua-dua *Salmonella* spp. serovar menunjukkan rintangan tinggi terhadap amoxicillin/asid clavunic, cephalothin, streptomycin dan ciprofloxacin dengan 100% rintangan terhadap ampicillin dan erythromycin. Semua pencilan menunjukkan rintangan pada sekurang-kurangnya tiga antibiotik yang diuji. *S. Typhimurium* adalah lebih rintangan berbanding *S. Enteritidis* dengan kerintangan antibiotik pelbagai (MAR) indeks masing-masing antara 0.27 ke 0.82 dan 0.27 ke 0.55.

Dendogram RAPD-PCR menunjukkan kebanyakan pencilan *S. Enteritidis* dan *S. Typhimurium* cenderung untuk taip bersama berdasarkan jenis sampel yang sama dimana 3 RAPD taip (RAPD taip 3 *S. Enteritidis* dan RAPD taip 1 dan 6 *S. Typhimurium*) ditaipkan bersama masing-masing pada jenis sampel dan lokasi persampelan yang sama. 3 RAPD taip yang lain (RAPD taip 1 *S. Enteritidis*, RAPD taip 2 dan 4 *S. Typhimurium*) ditaipkan berdasarkan jenis sampel yang sama tetapi berbeza lokasi persampelan. RAPD taip 3 *S. Typhimurium* telah ditaip pada jenis sampel yang berbeza tetapi lokasi persampelan yang sama diperhatikan.

Penilaian risiko kuantitatif mikrobiologi (QMRA) simulasi oleh konsep satu-hit menunjukkan keseluruhan risiko mendapat salmonellosis disebabkan pada penggunaan ulam lebih tinggi untuk *S. Typhimurium* dari *S. Enteritidis*. Purata risiko penyakit per hidangan per tahun untuk *S. Typhimurium* adalah 1 manakala untuk *S. Enteritidis*, risikonya adalah ~1 pada populasi yang terdedah.

Kesimpulannya, prevalens *Salmonella* spp. pada ulam adalah membimbangkan dan perlu dipantau dari masa ke semasa memandangkan pencegahan lebih baik daripada mengubati.

ACKNOWLEDGEMENTS

Here, I would like to thank my supervisor, Professor Dr. Son Radu for his help in the process of completing this thesis. I am really appreciated on his knowledge, expertise, advice, time and patience he has contributed.

Besides that, I am very grateful to my co-supervisor, Associate Professor Dr. Yaya Rukayadi (Faculty of Food Science and Technology, UPM) for his guidance, support and time he had been spent for helping me along I am finishing the thesis.

A million thanks to my labmates of Chang Wei San, Soopna Puspanadan, Aimi Syazana, Lye Ying Ling, Ubong Anyi, Loo Yuet Ying, Goh Sur Guat, Kuan Chee Hao, Nina Fouladynezhad, Elexson Nillian, Malcomn Tan Turk Hsern, Krishanthi Premarathne, Thung Tze Young, Sylvester William Silan and Elham Taghavi for helping me and analyze the results of experiment. I will always remember the memories of us working together. Also a big thank to Dr. Natasha Lee from Division of Postgraduate, Research and Innovation for your guide and help. Not to forget, special thanks to my friends of Raihana, Rosma, Siti Isa, Farahin, Nadia, Ada, Zira, Athirah, Siti Sarah Hani and Dinie for the continuous support and motivation.

Besides that, a big appreciation to Kak Liza, Kak Fatimah, all staff in Faculty of Food Science and Technology and School of Graduate Studies, UPM in organizing and meet the needs for all postgraduates student here.

Finally, I would like to express my deepest gratitude to my parents (Mohd Shahril bin Haji Ahmad and Sabariah binti Mat Amin), family and siblings (Mohd Israr, Siti Fatimah and Najihah) for their unfailing encouragement for me to finish up the research until the end. A sincere apology for the misbehaved, less time spent and worries I had brought to them. No words I can describe how much I appreciate and love all of you. Thank you once again for always be around me in ups and down of life.

All in all, without help from all of you, it is impossible for me to go this far. Thank you, thank you and thank you for zillion times!

I certify that a Thesis Examination Committee has met on 22 August 2016 to conduct the final examination of Najwa binti Mohd Shahril on her thesis entitled "Prevalence, Characterization and Antibiotic Resistance of *Salmonella* spp. Isolated from Raw Vegetables" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Anis Shobirin binti Meor Hussin, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Chairman)

Abdulkarim Sabo Mohammed, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Internal Examiner)

Samuel Lihan, PhD

Senior Lecturer
Universiti Malaysia Sarawak
Malaysia
(External Examiner)



ZULKARNAIN ZAINAL, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 28 September 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Son Radu, Ph.D

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Chairman)

Yaya Rukayadi, Ph.D

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

BUJANG KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Najwa Binti Mohd Shahril (GS30388)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of
Chairman of

Supervisory Committee: Prof. Dr. Son Radu

Signature: _____

Name of
Chairman of

Supervisory Committee: Assoc. Prof. Dr. Yaya Rukayadi

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xv
LIST OF ABBREVIATIONS	xvii
CHAPTER	
1 GENERAL INTRODUCTION	1
2 LITERATURE REVIEW	4
2.1 <i>Salmonella</i> spp.	4
2.1.1 Taxonomy	4
2.1.2 Distribution	6
2.1.3 <i>Salmonella</i> outbreaks	7
2.1.4 Source and Transmission	12
2.1.5 Clinical manifestation	12
2.1.5.1 Enteric fever	12
2.1.5.2 Gastroenteritis	13
2.1.5.3 Systemic infections and other complications of nontyphoidal salmonellosis	13
2.1.5.4 Chronic carrier state	13
2.1.6 Pathogenicity	13
2.2 Detection of <i>Salmonella</i>	15
2.2.1 Conventional methods	15
2.2.2 Rapid methods	15
2.2.3 Most probable number	16
2.2.4 MPN-PCR method	17
2.3 Antibiotic	17
2.3.1 Treatment of <i>Salmonella</i> infections	18
2.3.2 Antibiotic resistance	18
2.4 Typing methods	19
2.4.1 Phenotyping methods	20
2.4.2 Genotyping methods	20
2.4.2.1 Random amplification of polymorphic DNA-PCR (RAPD-PCR)	20
2.5 Risk assessment	21
2.5.1 Hazard identification	21
2.5.2 Exposure assessment	21

	2.5.3	Hazard characterization	22
	2.5.4	Risk characterization	22
3		PREVALENCE AND QUANTIFICATION OF <i>Salmonella</i> spp., <i>S. Enteritidis</i> AND <i>S. Typhimurium</i> IN ULAM	23
	3.1	Introduction	23
	3.2	Materials and Methods	23
	3.2.1	Sample collection	23
	3.2.2	Most probable number (MPN) method	24
	3.2.3	Plating method	24
	3.2.4	DNA extraction	24
	3.2.5	Multiplex PCR	24
	3.2.6	Statistical analysis	25
	3.3	Results	26
	3.4	Discussion	31
	3.5	Conclusion	34
4		ANTIBIOTIC SUSCEPTIBILITY OF <i>S. Enteritidis</i> AND <i>S. Typhimurium</i> ISOLATES	35
	4.1	Introduction	35
	4.2	Materials and Methods	35
	4.2.1	Antimicrobial susceptibility testing	35
	4.2.1.1	Bacterial inoculum preparation	35
	4.2.1.2	Standard disk diffusion method	35
	4.2.1.3	Data interpretation	36
	4.2.1.4	Multiple antibiotic resistances (MAR) indexing	36
	4.3	Results	36
	4.4	Discussion	38
	4.5	Conclusion	41
5		RANDOM AMPLIFICATION OF POLYMORPHIC DNA-PCR (RAPD-PCR) OF <i>S. Enteritidis</i> AND <i>S. Typhimurium</i> ISOLATES	42
	5.1	Introduction	42
	5.2	Materials and Methods	42
	5.2.1	Bacterial isolates and DNA extraction	42
	5.2.2	DNA primers	42
	5.2.3	RAPD-PCR	43
	5.2.4	RAPD-PCR analysis	43
	5.3	Results	44
	5.4	Discussion	50
	5.5	Conclusion	52

6	QUANTITATIVE MICROBIAL RISK ASSESSMENT (QMRA) OF RETAILED <i>ULAM</i>	53
6.1	Introduction	53
6.2	Materials and Methods	53
6.2.1	Risk characterization	53
6.2.1.1	Prevalence	57
6.2.1.2	Concentration	57
6.2.1.3	Consumption	57
6.2.1.4	Probability of illness per serving	57
6.2.1.5	Probability of illness per year	58
6.2.1.6	Expected number of cases per year	58
6.2.1.7	Simulations settings and analysis of model outputs	58
6.3	Results	58
6.4	Discussion	67
6.5	Conclusion	68
7	SUMMARY, GENERAL CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH	70
	REFERENCES	72
	APPENDICES	90
	BIODATA OF STUDENT	92
	LIST OF PUBLICATIONS	93

LIST OF TABLES

Table	Page
2.1 <i>Salmonella</i> outbreaks	8
3.1 Types of <i>ulam</i>	23
3.2 Primers used for detection of <i>Salmonella</i> spp., <i>S. Enteritidis</i> and <i>S. Typhimurium</i> in the multiplex PCR	25
3.3 Incidence of <i>Salmonella</i> spp., <i>S. Enteritidis</i> and <i>S. Typhimurium</i> in <i>ulam</i> from wet markets and hypermarkets	27
3.4 Density of <i>Salmonella</i> spp., <i>S. Enteritidis</i> and <i>S. Typhimurium</i> in <i>ulam</i> samples	29
3.5 Sensitivity of MPN-multiplex PCR and MPN-plating methods in detecting <i>Salmonella</i> spp., <i>S. Enteritidis</i> and <i>S. Typhimurium</i> in <i>ulam</i>	30
4.1 Percentage of <i>Salmonella</i> isolates resistant to antimicrobial agents	37
4.2 Antibiograms of <i>S. Enteritidis</i> and <i>S. Typhimurium</i> isolated from <i>ulam</i>	38
5.1 RAPD primers	43
5.2 RAPD type of <i>S. Enteritidis</i>	49
5.3 RAPD type of <i>S. Typhimurium</i>	50
6.1 The risk assessment model of illness by <i>S. Enteritidis</i> and <i>S. Typhimurium</i> due to consumption of <i>ulam</i>	54
6.2 Outputs of the QMRA model in the population exposed due to consumption of <i>ulam</i> contaminated with <i>Salmonella</i>	67
A1 Comparison for prevalence of <i>Salmonella</i> spp., <i>S. Enteritidis</i> and <i>S. Typhimurium</i> in <i>ulam</i> between wet markets and hypermarkets	90
A2 Comparison of prevalence of <i>Salmonella</i> spp., <i>S. Enteritidis</i> and <i>S. Typhimurium</i> between leafy vegetables and non-leafy vegetables	90

- A3 Comparison in sensitivity of detecting *Salmonella* spp., *S. Enteritidis* and *S. Typhimurium* in *ulam* between MPN-multiplex PCR method and MPN-plating method 91



LIST OF FIGURES

Figure		Page
2.1	Schematic diagram showing <i>Salmonella</i> invasion through host mucosal membrane	14
2.2	'Farm to fork' sequence	22
3.1	A representative gel of <i>Salmonella</i> spp., <i>S. Enteritidis</i> and <i>S. Typhimurium</i> amplified for <i>invA</i> gene, <i>sdf</i> gene and STM4492 gene respectively. Lane 1 and Lane 8, molecular size marker (100 bp DNA ladder); Lane 2, negative control; Lane 3, positive control (<i>S. Enteritidis</i> ATCC 13076 and <i>S. Typhimurium</i> ATCC 14028); Lane 4, <i>Salmonella</i> spp. (sample); Lane 5, <i>S. Enteritidis</i> (sample); Lane 6, <i>S. Typhimurium</i> (sample) and Lane 7, <i>S. Enteritidis</i> and <i>S. Typhimurium</i> (sample)	26
5.1	RAPD fingerprints of <i>S. Enteritidis</i> (SE) and <i>S. Typhimurium</i> (ST) isolates using GEN 1-50-08. Lane M: 1 kb DNA ladder; Lane 1: SE1; Lane 2: SE2; Lane 3: SE3; Lane 4: SE4; Lane 5: SE5; Lane 6: SE6; Lane 7: ST1; Lane 8: ST2; Lane 9: ST3	44
5.2	RAPD fingerprints of <i>S. Enteritidis</i> (SE) and <i>S. Typhimurium</i> (ST) isolates using GEN 1-50-08. Lane M: 1 kb DNA ladder; Lane 10: ST4; Lane 11: ST5; Lane 12: ST6; Lane 13: ST7; Lane 14: ST8; Lane 15: ST9; Lane 16: ST10; Lane 17: A2; Lane 18: A3	45
5.3	RAPD fingerprints of <i>S. Enteritidis</i> (SE) and <i>S. Typhimurium</i> (ST) isolates using GEN 1-50-09. Lane M: 1 kb DNA ladder; Lane 1: SE1; Lane 2: SE2; Lane 3: SE3; Lane 4: SE4; Lane 5: SE5; Lane 6: SE6; Lane 7: ST1; Lane 8: ST2; Lane 9: ST3	45
5.4	RAPD fingerprints of <i>S. Enteritidis</i> (SE) and <i>S. Typhimurium</i> (ST) isolates using GEN 1-50-09. Lane M: 1 kb DNA ladder; Lane 10: ST4; Lane 11: ST5; Lane 12: ST6; Lane 13: ST7; Lane 14: ST8; Lane 15: ST9; Lane 16: ST10; Lane 17: A2; Lane 18: A3	46
5.5	Dendogram for <i>S. Enteritidis</i> using GEN-1-50-08 and GEN-1-50-09 primers	47
5.6	Dendogram for <i>S. Typhimurium</i> using GEN-1-50-08 and GEN-1-50-09 primers	48

6.1	Probability of illness per year of <i>S. Enteritidis</i> in Asiatic pennywort	59
6.2	Expected number of cases per year of <i>S. Enteritidis</i> in Asiatic pennywort	59
6.3	Probability of illness per year of <i>S. Enteritidis</i> in water dropwort	60
6.4	Expected number of cases per year of <i>S. Enteritidis</i> in water dropwort	60
6.5	Probability of illness per year of <i>S. Enteritidis</i> in long bean	61
6.6	Expected number of cases per year of <i>S. Enteritidis</i> in long bean	61
6.7	Probability of illness per year of <i>S. Enteritidis</i> in winged bean	62
6.8	Expected number of cases per year of <i>S. Enteritidis</i> in winged bean	62
6.9	Probability of illness per year of <i>S. Typhimurium</i> in Asiatic pennywort	63
6.10	Expected number of cases per year of <i>S. Typhimurium</i> in Asiatic pennywort	63
6.11	Probability of illness per year of <i>S. Typhimurium</i> in water dropwort	64
6.12	Expected number of cases per year of <i>S. Typhimurium</i> in water dropwort	64
6.13	Probability of illness per year of <i>S. Typhimurium</i> in long bean	65
6.14	Expected number of cases per year of <i>S. Typhimurium</i> in long bean	65
6.15	Probability of illness per year of <i>S. Typhimurium</i> in winged bean	66
6.16	Expected number of cases per year of <i>S. Typhimurium</i> in winged bean	66

LIST OF ABBREVIATIONS

%eat	% of Malaysia population eating <i>ulam</i>
× g	Times gravity
16S rRNA	16 subunit ribosomal ribose nucleic acid
A2	<i>S. Typhimurium</i> ATCC 14028
A3	<i>S. Enteritidis</i> ATCC 13076
ACSSuT	Ampicillin, chloramphenicol, streptomycin, sulfonamides, tetracycline
AFLP	Amplified fragment length polymorphism
AMC	Amoxycillin/clavunic acid
Amp	Ampicillin
ARDRA	Amplified ribosomal DNA restriction analysis
ATCC	American Type Culture Collection
a_w	Water activity
BGA	Brilliant green agar
BOX-PCR	BOX-A1R-based repetitive extragenic palindromic-PCR
bp	Base pair
BPW	Buffered peptone water
C	Concentration
C	Cholaramphenicol
<i>C. jejuni</i>	<i>Campylobacter jejuni</i>
CDC	Centers for Disease Control and Prevention
Cip	Ciprofloxacin
CLSI	Clinical Laboratory Standards Institute
Cn	Gentamicin
D	Dose per serving
DNA	Deoxyribonucleic acid
dNTP	Deoxynucleoside triphosphate
DTs	Definitive phage types
E	Exposure (no. of serving/year)
E	Erythromycin
<i>E. coli</i>	<i>Escherichia coli</i>
<i>eaeA</i>	Intimin
Ec	Expected no. of cases per year
EPS	Exopolymer substance
ERIC-PCR	Enterobacterial repetitive intergenic consensus-polymerase chain reaction
Fim	Type I fimbriae
<i>fliC</i> _{h7}	Flagellar antigen
H ₂ S	Hydrogen sulfide
HE	Hektoen enteric
HIV	Human Immunodeficiency Virus
<i>hly</i> ₉₃₃	Hemolysin
ICMSF	International Commission on Microbiological Specifications for Foods

<i>inv</i>	invasion
<i>K. pneumonia</i>	<i>Klebsiella pneumonia</i>
Kf	Cephalothin
L	Long bean
<i>L. monocytogenes</i>	<i>Listeria monocytogenes</i>
LIA	Lysine iron agar
Lpf	Long polar fimbriae
LPS	Lipopolysaccharides
LT	Heat labile
MAR	Multiple antibiotic resistances
Max	Maximum
MDR	Multidrug resistance
Med	Median
MgCl ₂	Magnesium chloride
MH	Mueller Hinton
MIC	Minimum inhibitory concentration
Min	Minimum
MLCB	Mannitol lysine crystal violet brilliant green
MLST	Multilocus sequence typing
MPN	Most probable number
MPN-multiplex	Most probable number-multiplex polymerase chain reaction
PCR	Most probable number-polymerase chain reaction
MPN-PCR	Most probable number-plating
MPN-plating	Nalidixic acid
Na	National Committee on Risk Analysis
NCRA	Non-typhoidal <i>Salmonella</i>
NTS	Prevalence
P	Asian pennywort
P	Polymerase chain reaction
PCR	Population of Malaysia eating <i>ulam</i>
P _{eat}	Plasmid-encoded fimbriae
P _{ef}	Pulsed field gel electrophoresis
PFGE	Probability of illness per positive serving
P _{ill/pos}	Probability of illness per serving
P _{ill/serv}	Probability of illness per year
P _{ill/year}	Population of Malaysia
P _M	Quantitative microbial risk assessment
QMRA	Quinolone resistance-determining region
QRDR	Random amplification of polymorphic DNA-polymerase chain reaction
RAPD-PCR	Restriction fragment length polymorphism
RFLPs	Rappaport-Vassiliadis
RV	Serving size
S	Water dropwort
S	Streptomycin
S	<i>Salmonella</i> Enteritidis
S. Enteritidis	

S. Paratyphi	<i>Salmonella</i> Paratyphi
S. Typhi	<i>Salmonella</i> Typhi
S. Typhimurium	<i>Salmonella</i> Typhimurium
S. Typhimurium DT104	<i>Salmonella</i> Typhimurium definitive phage type 104
SCV	<i>Salmonella</i> containing vacuole
<i>sdf</i>	<i>Salmonella</i> difference fragment
SE	<i>S. Enteritidis</i>
SGI1	<i>Salmonella</i> genomic island 1
spp.	Species
SS	Salmonella-Shigella
SSCP	Single strand conformation polymorphism
Sspp	<i>Salmonella</i> spp.
ST	<i>S. Typhimurium</i>
STM4492	Cytoplasmic protein of <i>Salmonella</i> Typhimurium LT2
<i>stx</i> ₁ , <i>stx</i> ₂	Shiga toxin
Sxt	Trimethoprim-Sulphamethoxazole
<i>Taq</i>	<i>Thermus aquaticus</i>
TBE	Tris-Borate EDTA electrophoresis buffer
TE	Tris-EDTA buffer
Te	Tetracycline
TSI	Triple sugar iron agar
TT	Tetrathionate
U	Unit
UPGMA	Unweighted pair group linkage analysis method
USA	United States of America
<i>V. parahaemolyticus</i>	<i>Vibrio parahaemolyticus</i>
VBNC	Viable but non-culturable
VNTR	Variable number of tandem repeat
W	Winged bean
WHO	World Health Organization
XLD	Xylose lysine deoxycholate
α	Alpha
β	Beta

CHAPTER 1

GENERAL INTRODUCTION

1.1 Introduction

Consuming vegetables is gaining popularity nowadays as many people are starting to be concerned in leading a healthy lifestyle. Ministry of Health, Malaysia recommends eating plenty of vegetables and fruits every day. According to Malaysian Food Pyramid, it is recommended to take three servings of vegetables and two servings of fruits daily. Malaysian Food Pyramid is a visual tool in designing a healthy diet which consists of four levels of food groups and recommended number of servings per day. Vegetables and fruits provide important sources of vitamins, minerals, fibre and other bioactive compounds such as phytochemicals. Hence, by eating vegetables and fruits daily is able to prevent cardiovascular disease, diabetes, coronary heart disease, stroke, high blood pressure and cancers. A raw eaten vegetables known as *ulam* in Malay language is one of the healthier choices than cooked vegetables. It is because, it can prevent nutrient loss since cooking process will damage certain nutrients and phytochemicals in vegetables (MOH, 2010).

Even though the vegetables and fruits are known with the healthy and nutritional values, people also are faced with the increasing of reported foodborne pathogens contamination on fresh produce worldwide such as *Salmonella*, *Escherichia coli* O157:H7, *Vibrio parahaemolyticus*, *Campylobacter* and *Listeria monocytogenes* (Losio *et al.*, 2015; Tunung, 2012; Pui *et al.*, 2011; Gabriel *et al.*, 2007). *Salmonella* can grow on the surface of alfafa sprouts, tomatoes, other fruits and vegetables (Forsythe, 2010). Centers for Disease Control and Prevention (CDC) reported that fresh produce such as cucumbers, bean sprouts, mangoes, cantaloupe, papaya and tomatoes contaminated with *Salmonella* can lead to outbreaks. For instance, in 2012, there was multistate outbreaks regarding *Salmonella* Typhimurium (*S. Typhimurium*) and *Salmonella* Newport (*S. Newport*) linked to cantaloupe throughout 24 states of United States of America (USA). From the outbreaks, about 261 persons were infected with 94 persons were hospitalized and 3 deaths were reported in Kentucky (CDC, 2013).

A foodborne pathogen, *Salmonella* is first discovered in 1885 and it can be divided into two groups: typhoid *Salmonella* and nontyphoid *Salmonella*. Typhoid *Salmonella* causes systemic infections known as a typhoid fever while nontyphoid *Salmonella* causes a self-limiting gastroenteritis. Reservoir of typhoid fever is limited to human whereas animal is for nontyphoid *Salmonella*

(Sánchez-Vargas *et al.*, 2011; Bhunia, 2008). *Salmonella* can contaminate fresh produce during harvesting, processing, retailing and preparation at home through cross-contamination and mishandling practice.

Conventional methods and rapid methods specifically polymerase chain reaction (PCR) can be employed to detect the presence of *Salmonella* in food. Conventional methods involve enrichment, plating, biochemical test and serological confirmation provides an acceptable detection limit and specificity but they are time-consuming, laborious and not suitable for large number of samples (Lee *et al.*, 2015). Meanwhile, PCR and other PCR-based methods promise rapid detection and high degree of specificity (Saeki *et al.*, 2013). In the present study, most probable number (MPN) method was combined with PCR to detect and quantify *Salmonella* in food sample.

Characterization of *Salmonella* spp. is important for epidemiological study in order to track the source of outbreaks and to study the genetic diversity of bacterial pathogens. It involves typing methods such as antibiotic resistance patterns, molecular typing methods including random amplification of polymorphic DNA-PCR (RAPD-PCR) and pulsed field gel electrophoresis (PFGE) (Ranjbar *et al.*, 2014).

Malaysian government as well as Malaysians are more concern about the food safety issues nowadays. Hence, Food Safety and Quality Division under Ministry of Health, Malaysia had established National Committee on Risk Analysis (NCRA) in 2002 in order to examine and investigate food safety issues in the country (FAO, 2004). Information on safety of *Salmonella* spp. in *ulam* is still scarce but salmonellosis has been reported in Malaysia. To the best of author's knowledge, there is no published data on prevalence of *Salmonella* spp., *Salmonella* Enteritidis (*S. Enteritidis*) and *Salmonella* Typhimurium (*S. Typhimurium*) using most probable number-multiplex polymerase chain reaction (MPN-multiplex PCR) technique on these *ulam*. Thus, this study will provide a glance on the Malaysia scenario and alert the related health authorities on safety issues of *ulam* at the retail markets. Furthermore, it also gives us insight into the prevalence of *Salmonella* spp., *S. Enteritidis* and *S. Typhimurium* in *ulam* and risk acquiring *Salmonella* through consumption of *ulam*.

1.2 Objectives

The objectives of this study are:

1. To ascertain the prevalence and contamination level of *Salmonella* spp., *S. Enteritidis* and *S. Typhimurium* in *ulam* at retail level.
2. To examine antibiotic resistance patterns of *S. Enteritidis* and *S. Typhimurium* isolates from *ulam*.
3. To characterize *S. Enteritidis* and *S. Typhimurium* isolates using random amplification of polymorphic DNA-PCR (RAPD-PCR).
4. To estimate the risk of illness (salmonellosis) through consumption of *ulam*.



REFERENCES

- Abadias, M., Usall, J., Anguera, M., Solsona, C., & Viñas, I. (2008). Microbiological quality of fresh, minimally processed fruit and vegetables, and sprouts from retail establishments. *International Journal of Food Microbiology*, 123(1): 121-129.
- Abakpa, G.O., Umoh, V.J., Ameh, J.B., Yakubu, S.E., Kwaga, J.K.P., & Kamaruzaman, S. (2015). Diversity and antimicrobial resistance of *Salmonella enterica* isolated from fresh produce and environmental samples. *Environmental Nanotechnology, Monitoring & Management*, 3: 38-46.
- Abdul-Mutalib, N. A., Syafinaz, A. N., Sakai, K., & Shirai, Y. (2015). An overview of foodborne illness and food safety in Malaysia. *International Food Research Journal*, 22(3): 896-901.
- Adzitey, F., Rusul, G., & Huda, N. (2012). Prevalence and antibiotic resistance of *Salmonella* serovars in ducks, duck rearing and processing environments in Penang, Malaysia. *Food Research International*, 45(2): 947-952.
- Adzitey, F., Rusul, G., Huda, N., Cogan, T., & Corry, J. (2012). Prevalence, antibiotic resistance and RAPD typing of *Campylobacter* species isolated from ducks, their rearing and processing environments in Penang, Malaysia. *International Journal of Food Microbiology*, 154(3): 197-205.
- Andrews, H. L., & Baumler, A. J. (2005). *Salmonella* species. In P. M. Fratamico, A. K. Bhunia, & J. L. Smith (Eds.), *Foodborne pathogens: Microbiology and Molecular Biology* (pp. 327-339). United Kingdom: Horizon Scientific Press Ltd.
- Arumugaswamy, R. K., Rusul, G., Hamid, S. A., & Cheah, C. T. (1995). Prevalence of *Salmonella* in raw and cooked foods in Malaysia. *Food Microbiology*, 12: 3-8.
- Baharom, C. B. (2014, March 04). Death of 5-yr old boy was caused by *Salmonella* poisoning: Terengganu healthdirector. *New Straits Times*, <http://www2.nst.com.my/latest/death-of-5yr-old-boy-was-caused-by-salmonella-poisoning-terengganu-health-director-1.497963>. Assessed date: 30 January 2015.
- BAM (Bacteriological Analytical Manual) (2010). BAM Appendix 2: Most Probable Number from Serial Dilutions. U.S. Food and Drug Administration. <http://www.fda.gov/Food/FoodScienceResearch/LaboratoryMethod/ucm10656.htm>. Assessed date: 5 November 2015.

- Bauer, A. W., Kirby, W. M. M., Sherris, J. C. T., & Turck, M. (1966). Antibiotic susceptibility testing by a standardized single disk method. *American Journal of Clinical Pathology*, 45(4): 493-496.
- Bemrah, N., Bergis, H., Colmin, C., Beaufort, A., Millemann, Y., Dufour, B., Benet, J. J., Cerf, O., & Sanaa, M. (2003). Quantitative risk assessment of human salmonellosis from the consumption of a turkey product in collective catering establishments. *International Journal of Food Microbiology*, 80(1): 17-30.
- Beuchat, L. R. (2002). Ecological factors influencing survival and growth of human pathogens on raw fruits and vegetables. *Microbes and Infection*, 4(4): 413-423.
- Bhunja, A. K. (2008). *Foodborne Microbial Pathogens*. United States of America: Springer Science+Business Media, LLC.
- Bordini, M. E. B., Ristori, C. A., Jakabi, M., & Gelli, D. S. (2007). Incidence, internalization and behavior of *Salmonella* in mangoes, var. Tommy Atkins. *Food Control*, 18(8): 1002-1007.
- Borowsky, L. M., Schmidt, V., & Cardoso, M. (2007). Estimation of most probable number of *Salmonella* in minced pork samples. *Brazilian Journal of Microbiology*, 38(3): 544-546.
- Buchanan, R. L., Smith, J. L., & Long, W. (2000). Microbial risk assessment: dose response relations and risk characterization. *International Journal of Food Microbiology*, 58(3): 159-172.
- Budiati, T., Rusul, G., Wan-Abdullah, W. N., Arip, Y. M., Ahmad, R., & Thong, K. L. (2013). Prevalence, antibiotic resistance and plasmid profiling of *Salmonella* in catfish (*Clarias gariepinus*) and tilapia (*Tilapia mossambica*) obtained from wet markets and ponds in Malaysia. *Aquaculture*, 372: 127-132.
- Campioni, F., Bergamini, A. M. M., & Falcão, J. P. (2012). Genetic diversity, virulence genes and antimicrobial resistance of *Salmonella* Enteritidis isolated from food and humans over a 24-year period in Brazil. *Food Microbiology*, 32(2): 254-264.
- Carraminana, J. J., Humbert, F., Ermel, G., & Colin, P. (1997). Molecular epidemiological investigation of *Salmonella* Typhimurium strains related to an egg-borne outbreak. *Research in Microbiology*, 148(7): 633-636.

- CDC (Centers for Disease Control and Prevention) (2012). What is salmonellosis. <http://www.cdc.gov/salmonella/general/>. Assessed date: 30 January 2015.
- CDC (Centers for Disease Control and Prevention) (2013). Antibiotic Resistance Questions & Answers. <http://www.cdc.gov/getsmart/antibioticuse/antibiotic-resistance-faqs.html#>. Assessed date: 1 April 2015.
- CDC (Centers for Disease Control and Prevention) (2013). Multistate Outbreak of *Salmonella* Typhimurium and *Salmonella* Newport Infections Linked to Cantaloupe (FinalUpdate). <http://www.cdc.gov/salmonella/typhimurim-cantaloupe-0812/index.html>. Assessed date: 14 Disember 2015.
- CDC (Centers for Disease Control and Prevention) (2014). Pathogens causing US foodborne illnesses, hospitalizations, and deaths, 2000-2008. <http://www.cdc.gov/foodborne-burden/PDFs/pathogens-complete-list-01-12.pdf>. Assessed date: 30 January 2015.
- CDC (Centers for Disease Control and Prevention) (2014). Reports of Selected Salmonella Outbreaks Investigation. <http://www.cdc.gov/salmonella/outbreaks>. Assessed date: 30 January 2015.
- Cerna-Cortes, J. F., Gómez-Aldapa, C. A., Rangel-Vargas, E., Ramírez-Cruz, E., & Castro-Rosas, J. (2013). Presence of indicator bacteria, *Salmonella* and diarrheagenic *Escherichia coli* pathotypes on mung bean sprouts from public markets in Pachuca, Mexico. *Food Control*, 31(2): 280-283.
- Ceuppens, S., Hessel, C. T., de Quadros Rodrigues, R., Bartz, S., Tondo, E. C., & Uyttendaele, M. (2014). Microbiological quality and safety assessment of lettuce production in Brazil. *International Journal of Food Microbiology*, 181: 67-76.
- Chai, L. C., Robin, T., Ragavan, U. M., Gunsalam, J. W., Bakar, F. A., Ghazali, F. M., Radu, Son., & Kumar, M. P. (2007). Thermophilic *Campylobacter* spp. in salad vegetables in Malaysia. *International Journal of Food Microbiology*, 117: 106-111.
- Chen, H. M., Wang, Y., Su, L. H., & Chiu, C. H. (2013). Nontyphoid *Salmonella* Infection: Microbiology, Clinical features, and Antimicrobial Therapy. *Pediatrics and Neonatology*, 54(3): 147-152.
- Chia, T. W. R., Goulter, R. M., McMeekin, T., Dykes, G. A., & Fegan, N. (2009). Attachment of different *Salmonella* serovars to materials commonly used in a poultry processing plant. *Food Microbiology*, 26(8): 853-859.

- CLSI (Clinical and Laboratory Standards Institute) (2013). Performance standards for antimicrobial susceptibility testing; twenty-third Informational Supplement. M100 S23. http://www.wsmicrobiology.com/clsi-m100_s23_performance_standards_for_antimicrobial-susceptibility_testing_-_twenty_third_informational_supplement-2013/. Assessed date: 2 February 2014.
- Cooke, F.J., Threlfall, E.J., & Wain, J. (2007). Current Trends in the Spread and Occurrence of Human Salmonellosis: Molecular Typing and Emerging Antibiotic Resistance. In M. Rhen, D. Maskell, P. Mastroeni, & J. Threlfall (Eds.), *Salmonella: Molecular Biology and Pathogenesis* (pp. 1-30). United Kingdom: Cronwell Press Trowbridge Wiltshire.
- Cruchaga, S., Echeita, A., Aladueña, A., García-Peña, J., Frias, N., & Usera, M. A. (2001). Antimicrobial resistance in salmonellae from humans, food and animals in Spain in 1998. *Journal of Antimicrobial Chemotherapy*, 47(3): 315-321.
- DAN (Dasar Agromakanan Negara) (2015). Dasar Agromakanan Negara 2011-2020. <http://www.moa.gov.my/web/guest/dasar-agro-makanan-negara-2011-2020-dan>. Assessed date: 2 February 2015.
- De Giusti, M., Aurigemma, C., Marinelli, L., Tufi, D., De Medici, D., Di Pasquale, S., De Vito, C., & Boccia, A. (2010). The evaluation of the microbial safety of fresh ready-to-eat vegetables produced by different technologies in Italy. *Journal of Applied Microbiology*, 109(3): 996-1006.
- de Oliveira, F. A., Brandelli, A., & Tondo, E. C. (2006). Antimicrobial resistance in *Salmonella* Enteritidis from foods involved in human salmonellosis outbreaks in southern Brazil. *New Microbiologica*, 29(1): 49-54.
- de Oliveira, M. A., De Souza, V. M., Bergamini, A. M. M., & De Martinis, E. C. P. (2011). Microbiological quality of ready-to-eat minimally processed vegetables consumed in Brazil. *Food Control*, 22(8): 1400-1403.
- Deering, A. J., Mauer, L. J., & Pruitt, R. E. (2012). Internalization of *E. coli* O157:H7 and *Salmonella* spp. in plants: A review. *Food Research International*, 45(2): 567- 575.
- Dennis, S. B., Buchanan, R. L., & Miller, A. J. (2002). Microbial Risk Assessment: Achievements and Future Challenges. *In Food Safety Magazine*.
- Dewaai, C. S., & Bhuiya, F. (2007). Outbreaks by the numbers: fruits and vegetables 1990–2005. *International Association for Food Protection 94th Annual Meeting*.

- Diana, J. E., Pui, C. F., & Son, R. (2012). Enumeration of *Salmonella* spp., *Salmonella* Typhi and *Salmonella* Typhimurium in fruit juices. *International Food Research Journal*, 19(1): 51-56.
- Embun, M. (2013, October 02). 'Ayam masak merah' contaminated. *The Star*, [http://www.thestar.com.my/News/Nation/2013/10/02Ayam masak merah contaminated -Cook -who -made -the -dish -is -in shock say relatives/](http://www.thestar.com.my/News/Nation/2013/10/02Ayam_masak_merah_contaminated_Cook_who_made_the_dish_is_in_shock_say_relatives/). Assessed date: 30 January 2015.
- FAO (Food and Agriculture Organization of the United Nations) (2004). Application of risk analysis in food control; challenges and benefits risk analysis in Malaysia. <http://www.fao.org/docrep/meeting/006/ad699e.htm>. Assessed date: 14 Desember 2015.
- Fatimah, A. M. Z., Norazian, M. H., & Rashidi, O. (2012). Identification of carotenoid composition in selected 'ulam' or traditional vegetables in Malaysia. *International Food Research Journal*, 19(2): 527-530.
- FDA (U.S. Food and Drug Administration) (2014). Be *Salmonella* Safe!. [http://www.fda.gov/animal_veterinary/resources_for_you/animalhealth/teracy/ucm136197.htm](http://www.fda.gov/animal_veterinary/resources_for_you/animal_health/teracy/ucm136197.htm). Assessed date: 30 January 2015.
- Food Consumption Statistics of Malaysia. 2003. Viewed on 18th June 2014.
- Forsythe, S. J. (2010). *The Microbiology of Safe Food*. United Kingdom: Blackwell Publishing Ltd.
- Franz, E., Tromp, S. O., Rijgersberg, H., & Van Der Fels-Klerx, H. J. (2010). Quantitative microbial risk assessment for *Escherichia coli* O157: H7, *Salmonella*, and *Listeria monocytogenes* in leafy green vegetables consumed at salad bars. *Journal of Food Protection*, 73(2): 274-285.
- Fratamico, P. M., Bagi, L. K., & Pepe, T. (2000). A multiplex polymerase chain reaction assay for rapid detection and identification of *Escherichia coli* O157:H7 in foods and bovine feces. *Journal of Food Protection*, 63(8): 1032-1037.
- Gabriel, A. A., Berja, M. C., Estrada, A. M. P., Lopez, M. G. A. A., Nery, J. G. B., & Villaflor, E. J. B. (2007). Microbiology of retail mung bean sprouts vended in public markets of National Capital Region, Philippines. *Food Control*, 18(10): 1307-1313.
- Ge, C., Lee, C., & Lee, J. (2012). The impact of extreme weather events on *Salmonella* internalization in lettuce and green onion. *Food Research International*, 45(2): 1118-1122.

- Goh, S. G. (2013). *Prevalence, risk and antibiogram of Listeria monocytogenes in raw chicken meat*. (Unpublished Master Thesis). Universiti Putra Malaysia.
- Goh, S. G., Kuan, C. H., Loo, Y. Y., Chang, W. S., Lye, Y. L., Soopna, P., Tang, J. Y. H., Nakaguchi, Y., Nishibuchi, M., Afsah-Hejri, L., & Son, R. (2012). *Listeria monocytogenes* in retailed raw chicken meat in Malaysia. *Poultry Science*, 91(10): 2686-2690.
- Gonzales-Barron, U. A., Redmond, G., & Butler, F. (2012). A risk characterization model of *Salmonella* Typhimurium in Irish fresh pork sausages. *Food Research International*, 45(2): 1184-1193.
- Gordon, M. A., Graham, S. M., Walsh, A. L., Wilson, L., Phiri, A., Molyneux, E., Zijlstra, E. E., Heyderman, R. S., Hart, C. A., & Molyneux, M. E. (2008). Epidemics of Invasive *Salmonella enterica* Serovar Enteritidis and *S. enterica* serovar Typhimurium Infection Associated with Multidrug Resistance among Adults and Children in Malawi. *Clinical Infectious Diseases*, 46(7): 963- 969.
- Graham, S. M., & English, M. (2009). Non-typhoidal salmonellae: a management challenge for children with community acquired invasive disease in tropical African countries. *Lancet*, 373(9659): 267-269.
- Gray, J. T., & Fedorka-Cray, P. J. (2002). *Salmonella*. In D. O. Cliver, & H. P. Riemann (Eds.), *Foodborne diseases* (pp. 55-68). San Diego: Academic Press.
- Grimont, P. A. D., Grimont, F., & Bouvet, P. (2000). Taxonomy of the genus *Salmonella*. In C. Wray, & A. Wray (Eds.), *Salmonella in domestic animals* (pp. 1-18). United Kingdom: CABI Publishing.
- Guillier, L., Danan, C., Bergis, H., Delignette-Muller, M. L., Granier, S., Rudelle, S., Beaufort, A., & Brisabois, A. (2013). Use of quantitative microbial risk assessment when investigating foodborne illness outbreaks: The example of a monophasic *Salmonella* Typhimurium 4, 5, 12: i:- outbreak implicating beef burgers. *International Journal of Food Microbiology*, 166(3): 471-478.
- Gunel, E., Kilic, G. P., Bulut, E., Durul, B., Acar, S., Alpas, H., & Soyer, Y. (2015). *Salmonella* surveillance on fresh produce in retail in Turkey. *International Journal of Food Microbiology*, 199: 72-77.
- Gwack, J., Lee, K. C., Lee, H. J., Kwak, W., Lee, D. W., Choi, Y. H., Kim, J. S., & Kang, Y. A. (2010). Trends in water-and foodborne disease outbreaks in Korea, 2007–2009. *Osong Public Health and Research Perspectives*, 1(1): 50-54.

- Hanes, D. (2003). Nontyphoid *Salmonella*. In O. Henegariu, N. A. Heerema, S. R. Dloughy, G. H. Vance, & P. H. Vogt (Eds.), *International handbook of foodborne pathogens* (pp. 137-149). New York: Marcel Dekker, Inc.
- Hardy, A. (2004). *Salmonella*: a continuing problem. *Postgraduate Medical Journal*, 80: 541-545.
- Haryani, Y., Noorzaleha, A. S., Fatimah, A. B., Noorjahan, B. A., Patrick, G. B., Shamsinar, A. T., Laila, R. A. S., & Son, R. (2007). Incidence of *Klebsiella pneumoniae* in street foods sold in Malaysia and their characterization by antibiotic resistance, plasmid profiling, and RAPD PCR analysis. *Food Control*, 18(7): 847-853.
- Hendriksen, R. S., Vieira, A. R., Karlsmose, S., Lo Fo Wong, D. M., Jensen, A. B., Wegener, H. C., & Aarestrup, F. M. (2011). Global Monitoring of *Salmonella* Serovar Distribution from the World Health Organization Global Foodborne Infections Network Country Data Bank: Results of Quality Assured Laboratories from 2001 to 2007. *Foodborne Pathogens and Disease*, 8(8): 887- 900.
- Hoorfar, J., Ahrens, P., & Rådström, P. (2000). Automated 5' nuclease PCR assay for identification of *Salmonella enterica*. *Journal of Clinical Microbiology*, 38(9): 3429-3435.
- Jacobsen, C. S., & Bech, T. B. (2012). Soil survival of *Salmonella* and transfer to freshwater and fresh produce. *Food Research International*, 45(2): 557-566.
- Joseph, B., Otta, S. K., Karunasagar, I., & Karunasagar, I. (2001). Biofilm formation by *Salmonella* spp. on food contact surfaces and their sensitivity to sanitizers. *International Journal of Food Microbiology*, 64(3): 367-372.
- Khoodoo, M. H. R., Issack, M. I., & Jaufferally-Fakim, Y. (2002). Serotyping and RAPD profiles of *Salmonella enterica* isolates from Mauritius. *Letters in Applied Microbiology*, 35(2): 146-152.
- Kovats, R. S., Edwards, S. J., Hajat, S., Armstrong, B. G., Ebi, K. L., & Menne, B. (2004). The effect of temperature on food poisoning: a time-series analysis of salmonellosis in ten European countries. *Epidemiology and Infection*, 132(3): 443-453.
- Krumperman, P. H. (1983). Multiple antibiotic resistance indexing of *Escherichia coli* to identify high-risk sources of fecal contamination of foods. *Applied and Environmental Microbiology*, 46(1): 165-170.

- Kuan, C. H., Goh, S. G., Loo, Y. Y., Chang, W. S., Lye, Y. L., Puspanadan, S., Tang, J. Y. H., Nakaguchi, Y., Nishibuchi, M., Mahyudin, N. A., & Radu, S. (2013). Prevalence and quantification of *Listeria monocytogenes* in chicken offal at the retail level in Malaysia. *Poultry Science*, 92(6): 1664-1669.
- Kunwar, R., Singh, H., Mangla, V., & Hiremath, R. (2013). Outbreak investigation: *Salmonella* food poisoning. *Medical Journal Armed Forces India*, 69(4): 388-391.
- Lambertini, E., Danyluk, M. D., Schaffner, D. W., Winter, C. K., & Harris, L. J. (2012). Risk of salmonellosis from consumption of almonds in the North American market. *Food Research International*, 45(2): 1166-1174.
- Lammerding, A. M., & Fazil, A. (2000). Hazard identification and exposure assessment for microbial food safety risk assessment. *International Journal of Food Microbiology*, 58: 147-157.
- Lapidot, A., Romling, U., & Yaron, S. (2006). Biofilm formation and the survival of *Salmonella* Typhimurium on parsley. *International Journal of Food Microbiology*, 109(3): 229-233.
- Learn-Han, L., Yoke-Kqueen, C., Shiran, M. S., Sabrina, S., Noor Zaleha, A. S., Sim, J. H., Chai-Hoon, K., & Son, R. (2009). Molecular characterization and antimicrobial resistance profiling of *Salmonella enterica* subsp. *Enterica* isolated from 'Selom' (*Oenanthe stolonifera*). *International Food Research Journal*, 16(1): 191-202.
- Lee, H. Y., Chai, L. C., Tang, S. Y., Jinap, S., Ghazali, F. M., Nakaguchi, Y., Nishibuchi, M., & Son, R. (2009). Application of MPN-PCR in biosafety of *Bacillus cereus* s.l. for ready-to-eat cereals. *Food Control*, 20(11): 1068-1071.
- Lee, H. Y., Su, L. H., Tsai, M. H., Kim, S. W., Chang, H. H., Jung, S. I., Park, K. H., Perera, J., Carlos, C., Tan, B. H., Kumarasinghe, G., So, T., Chongthaleong, A., Hsueh, P. R., Liu, J. W., Song, J. H., & Chiu, C. H. (2009). High rate of reduced susceptibility to ciprofloxacin and ceftriaxone among nontyphoid *Salmonella* clinical isolates in Asia. *Antimicrobial Agents and Chemotherapy*, 53(6): 2696-2699.
- Lee, K. M., Runyon, M., Herrman, T. J., Phillips, R., & Hsieh, J. (2015). Review of *Salmonella* detection and identification methods: Aspects of rapid emergency response and food safety. *Food Control*, 47: 264-276.

- Lertworapreecha, M., Sutthimusik, S., & Tontikapong, K. (2013). Antimicrobial Resistance in *Salmonella enterica* Isolated From Pork, Chicken, and Vegetables in Southern Thailand. *Jundishapur Journal of Microbiology*, 6(1): 36-41.
- Li, Y., & Mustapha, A. (2004). Simultaneous detection of *Escherichia coli* O157:H7, *Salmonella*, and *Shigella* in apple cider and produce by a multiplex PCR. *Journal of Food Protection*, 67(1): 27-33.
- Li, Y., Zhuang, S., & Mustapha, A. (2005). Application of a multiplex PCR for the simultaneous detection of *Escherichia coli* O157:H7, *Salmonella* and *Shigella* in raw and ready-to-eat meat products. *Meat Science*, 71: 402-406.
- Lim, H., Lee, K. H., Hong, C. H., Bahk, G. J., & Choi, W. S. (2005). Comparison of four molecular typing methods for the differentiation of *Salmonella* spp. *International Journal of Food Microbiology*, 105(3): 411-418.
- Liu, C., Hofstra, N., & Franz, E. (2013). Impacts of climate change on the microbial safety of pre-harvest leafy green vegetables as indicated by *Escherichia coli* O157 and *Salmonella* spp. *International Journal of Food Microbiology*, 163(2): 119-128.
- Löfström, C., Knutsson, R., Axelsson, C. E., & Rådström, P. (2004). Rapid and specific detection of *Salmonella* spp. in animal feed samples by PCR after culture enrichment. *Applied and Environmental Microbiology*, 70(1): 69-75.
- Losio, M. N., Pavoni, E., Bilei, S., Bertasi, B., Bove, D., Capuano, F., Farneti, S., Blasi, G., Comin, D., Cardamone, C., Decastelli, L., De Santis, P., Di Pasquale, S., Gattuso, A., Goffredo, E., Fadda, A., Pisanu, M., & Decastelli, L. (2015). Microbiological survey of raw and ready-to-eat leafy green vegetables marketed in Italy. *International Journal of Food Microbiology*, 210: 88-91.
- Maistro, L. C., Miya, N. T. N., Sant'Ana, A. S., & Pereira, J. L. (2012). Microbiological quality and safety of minimally processed vegetables marketed in Campinas, SP-Brazil, as assessed by traditional and alternative methods. *Food Control*, 28(2): 258-264.
- Majowicz, S. E., Musto, J., Scallan, E., Angulo, F. J., Kirk, M., O'Brien, S. J., Jones, T. F., Fazil, A., & Hoekstra, R. M. (2010). The global burden of nontyphoidal *Salmonella* gastroenteritis. *Clinical Infectious Diseases*, 50(6): 882-889.

- Manfredi, R. (2009). The phenomenon of antibiotic resistance in an evolutionary perspective. In A. R. Bonilla, & K. P. Muniz (Eds.), *Antibiotic Resistance: Causes and Risk Factors, Mechanisms and Alternatives* (pp. 1-4). New York: Nova Science Publishers, Inc.
- Manijeh, M., Mohammad, J., & Roha, K. K. (2008). Biofilm Formation by *Salmonella* Enteritidis on Food Contact Surfaces. *Journal of Biological Sciences*, 8(2): 502-505.
- Marian, M. N., Aminah, S. S., Zuraini, M. I., Son, R., Maimunah, M., Lee, H. Y., Wong, W. C., & Elexson, N. (2012). MPN-PCR detection and antimicrobial resistance of *Listeria monocytogenes* isolated from raw and ready-to-eat foods in Malaysia. *Food Control*, 28(2): 309-314.
- Martin, B., Jofre, A., Garriga, M., Hugas, M., & Aymerich, T. (2004). Quantification of *Listeria monocytogenes* in fermented sausages by MPN PCR method. *Letters in Applied Microbiology*, 39(3): 290-295.
- Martinez, J. L. (2009). Environmental pollution by antibiotics and by antibiotic resistance determinants. *Environmental Pollution*, 157(11): 2893-2902.
- Maurer, J. (2006). *PCR Methods in Foods*. United States of America: Springer Science+Business Media, Inc.
- McCarthy, N., Reen, F. J., Buckley, J. F., Frye, J. G., Boyd, E. F., & Gilroy, D. (2009). Sensitive and rapid molecular detection assays for *Salmonella enterica* serovars Typhimurium and Heidelberg. *Journal of Food Protection*, 72(11): 2350-2357.
- Miko, A., Pries, K., Schroeter, A., & Helmuth, R. (2005). Molecular mechanisms of resistance in multidrug resistant serovars of *Salmonella enterica* isolated from foods in Germany. *Journal of Antimicrobial Chemotherapy*, 56(6): 1025-1033.
- MOH (Ministry of Health Malaysia) (2008). Malaysia's Health. [www.moh.gov.my/images/gallery/publications/m Malaysia%20Health%202008-2.pdf](http://www.moh.gov.my/images/gallery/publications/m%20Malaysia%20Health%202008-2.pdf). Accessed date: 12 September 2015.
- MOH (Ministry of Health) (2010). *Malaysian Dietary Guidelines*. Malaysia: Nutrition Society of Malaysia.
- Montville, T. J., & Matthews, K. R. (2008). *Food Microbiology: An Introduction*. United States of America: American Society for Microbiology Press.
- Morpeth, S. C., Ramadhani, H. O., & Crump, J. A. (2009). Invasive nonTyphi *Salmonella* disease in Africa. *Clinical Infection of Disease*, 49(4): 606-611.

- Muhammad, I. N. (2016, April 11). Roti jala dicemari tiga bakteria. myMetro, [http:// www. hmetro. com.my/ node / 129647](http://www.hmetro.com.my/node/129647). Assessed date: 30 April 2016.
- MysIDC (Malaysia Informative Data Centre, Department of Statistics Malaysia) (2014). [http:// mysidc. statistics.gov.my / index. php? lang=en#](http://mysidc.statistics.gov.my/index.php?lang=en#). Assessed date: 21 March 2014.
- Newell, D. G., Koopmans, M., Verhoef, L., Duizer, E., Aidara Kane, A., Sprong, H., Giessen, J. v. d., & Kruse, H. (2010). Food-borne diseases: The challenges of 20 years ago still persist while new ones continue to emerge. *International Journal of Food Microbiology*, 139: S3-S15.
- Ngoi, S. T., & Thong, K. L. (2013). Molecular characterization showed limited genetic diversity among *Salmonella* Enteritidis isolated from humans and animals in Malaysia. *Diagnostic Microbiology and Infectious Disease*, 77(4): 304-311.
- Nguyen, H. D. N., Yang, Y. S., & Yuk, H. G. (2014). Biofilm formation of *Salmonella* Typhimurium on stainless steel and acrylic surfaces as affected by temperature and pH level. *LWT-Food Science and Technology*, 55(1): 383-388.
- Nicholson, A. M., Gurtler, J. B., Bailey, R. B., Niemira, B. A., & Douds, D. D. (2015). Influence of mycorrhizal fungi on fate of *E. coli* O157: H7 and *Salmonella* in soil and internalization into Romaine lettuce plants. *International Journal of Food Microbiology*, 192: 95-102.
- Nicholson, F. A., Groves, S. J., & Chambers, B. J. (2005). Pathogen survival during livestock manure storage and following land application. *Bioresource Technology*, 96(2): 135-143.
- Nielsen, H., Gradel, K. O., & Schönheyder, H. C. (2006). High incidence of intravascular focus in nontyphoid *Salmonella* bacteremia in the age group above 50 years: A population - based study. *Acta Pathologica, Microbiologica Et Immunologica Scandinavica*, 114(9): 641- 645.
- NIH (National Institute of Allergy and Infectious Diseases) (2015). Salmonellosis. [http://www. niaid.nih. Gov / topics / salmonellosis / Pages Treatment .aspx](http://www.niaid.nih.gov/topics/salmonellosis/Pages/Treatment.aspx). Assessed date: 1 April 2015.
- Nik, I. N., & Sharifah, M. S. A. (2005, April 04). Typhoid hits Kelantan. *New Straits Times*, [https :/ /www. High beam. com / doc / 1P1-107602013. html](https://www.highbeam.com/doc/1P1-107602013.html). Assessed date: 30 January 2015.

- Nillian, E., Ching, C. L., Fung, P. C., Robin, T., Anyi, U., Chilek, T. Z. T., Son, R., & Nishibuchi, M. (2011). Simultaneous detection of *Salmonella* spp., *Salmonella* Enteritidis and *Salmonella* Typhimurium in raw salad vegetables and vegetarian burger patties. *Food and Nutrition Sciences*, 2: 1077-1081.
- Nipa, M. N., Mazumdar, R. M., Hasan, M. M., Fakruddin, M. D., Islam, S., Bhuiyan, H. R., & Iqbal, A. (2011). Prevalence of multi drug resistant bacteria on raw salad vegetables sold in major markets of Chittagong city, Bangladesh. *Middle-East Journal of Scientific Research*, 10(1): 70-77.
- Norhana, M. W., Poole, S. E., Deeth, H. C., & Dykes, G. A. (2010). The effects of temperature, chlorine and acids on the survival of *Listeria* and *Salmonella* strains associated with uncooked shrimp carapace and cooked shrimp flesh. *Food Microbiology*, 27(2): 250-256.
- Okafo, C. N., Umoh, V. J., & Galadima, M. (2003). Occurrence of pathogens on vegetables harvested from soils irrigated with contaminated streams. *Science of the Total Environment*, 311(1): 49-56.
- Olaimat, A. N., & Holley, R. A. (2012). Factors influencing the microbial safety of fresh produce: A review. *Food Microbiology*, 32(1): 1-19.
- O'Regan, E., McCabe, E., Burgess, C., McGuinness, S., Barry, T., Duffy, G., Whyte, P., & Fanning, S. (2008). Development of a real-time multiplex PCR assay for the detection of multiple *Salmonella* serotypes in chicken samples. *BMC Microbiology*, 8(1): 156.
- Oscar, T. P. (2004). A quantitative risk assessment model for *Salmonella* and whole chickens. *International Journal of Food Microbiology*, 93(2): 231-247.
- Parry, C. M., & Threlfall, E. J. (2008). Antimicrobial resistance in typhoidal and nontyphoidal salmonellae. *Current Opinion in Infectious Diseases*, 21(5): 531-538.
- Patel, J., & Sharma, M. (2010). Differences in attachment of *Salmonella enterica* serovars to cabbage and lettuce leaves. *International Journal of Food Microbiology*, 139(1): 41-47.
- Ponniah, J., Robin, T., Paie, M. S., Radu, S., Ghazali, F. M., Kqueen, C. Y., Nishibuchi, M., Nakaguchi, Y., & Malakar, P. K. (2010). *Listeria monocytogenes* in raw salad vegetables sold at retail level in Malaysia. *Food Control*, 21(5): 774-778.

- Pui, C. F., Wong, W. C., Chai, L. C., Lee, H. Y., Tang, J. Y. H., Ahmad, N., Farinazleen, M. G., Cheah, Y. K., & Son, R. (2011). Biofilm formation by *Salmonella* Typhi and *Salmonella* Typhimurium on plastic cutting board and its transfer to dragon fruit. *International Food Research Journal*, 18(1): 31-38.
- Pui, C. F., Wong, W. C., Chai, L. C., Nillian, E., Ghazali, F. M., Cheah, Y. K., Nakaguchi, Y., Nishibuchi, M., & Son, R. (2011). Simultaneous detection of *Salmonella* spp., *Salmonella* Typhi and *Salmonella* Typhimurium in sliced fruits using multiplex PCR. *Food Control*, 22(2): 337-342.
- Pui, C. F., Wong, W. C., Chai, L. C., Tunung, R., Jeyaletchumi, P., Noor Hidayah, M. S., Ubong, A., Farinazleen, M. G., Cheah, Y., & Radu, S. (2011). *Salmonella*: A foodborne pathogen. *International Food Research Journal*, 18(2): 465-473.
- Puspanadan, S., Afsah-Hejri, L., Loo, Y. Y., Nillian, E., Kuan, C. H., Goh, S. G., Chang, W. S., Lye, Y. L., John, Y. H. T., Rukayadi, Y., Yoshitsugu, N., Nishibuchi, M., & Son, R. (2012). Detection of *Klebsiella pneumoniae* in raw vegetables using Most Probable Number Polymerase Chain Reaction (MPN-PCR). *International Food Research Journal*, 19(4): 1757-1762.
- Quiroz-Santiago, C., Rodas-Suárez, O. R., Vázquez, Q., Carlos, R., Fernández, F. J., Quiñones-Ramírez, E. I., & Vázquez-Salinas, C. (2009). Prevalence of *Salmonella* in vegetables from Mexico. *Journal of Food Protection*, 72(6): 1279-1282.
- Radhika, M., Saugata, M., Murali, H. S., & Batra, H. V. (2014). A novel multiplex PCR for the simultaneous detection of *Salmonella enterica* and *Shigella* species. *Brazilian Journal of Microbiology*, 45(2): 667-676.
- Ranjbar, R., Karami, A., Farshad, S., Giammanco, G. M., & Mammina, C. (2014). Typing methods used in the molecular epidemiology of microbial pathogens: a how-to guide. *The New Microbiologica*, 37(1): 1-15.
- Rezende, A. C. B., de Castro, M. F. P., Porto, E., Uchima, C. A., Benato, E., & Penteado, A. L. (2009). Occurrence of *Salmonella* spp. in persimmon fruit (*Diospyrus kaki*) and growth of *Salmonella enteritidis* on the peel and in the pulp of this fruit. *Food Control*, 20(11): 1025-1029.
- Saeki, E. K., Alves, J., Bonfante, R. C., Hirooka, E. Y., & Oliveira, T. C. R. M. (2013). Multiplex PCR (mPCR) for the Detection of *Salmonella* spp. and the Differentiation of the Typhimurium and Enteritidis Serovars in Chicken Meat. *Journal of Food Safety*, 33(1): 25-29.

- Salleh, N. A., Rusul, G., Hassan, Z., Reezal, A., Isa, S. H., Nishibuchi, M., & Radu, S. (2003). Incidence of *Salmonella* spp. in raw vegetables in Selangor, Malaysia. *Food Control*, 14(7): 475-479.
- Salyers, A. A., & Whitt, D. D. (2005). *Revenge of the Microbes*. United States of America: American Society for Microbiology Press.
- Sánchez-Vargas, F. M., Abu-El-Haija, M. A., & Gómez-Duarte, O. G. (2011). *Salmonella* infections: An update on epidemiology, management, and prevention. *Travel Medicine and Infectious Disease*, 9(6): 263-277.
- Sant'Ana, A. S., Landgraf, M., Destro, M. T., & Franco, B. D. (2011). Prevalence and counts of *Salmonella* spp. in minimally processed vegetables in São Paulo, Brazil. *Food Microbiology*, 28(6): 1235-1237.
- Sant'Ana, A. S., Franco, B. D., & Schaffner, D. W. (2014). Risk of infection with *Salmonella* and *Listeria monocytogenes* due to consumption of ready-to eat leafy vegetables in Brazil. *Food Control*, 42: 1-8.
- Scaria, J., Warnick, L. D., Kaneene, J. B., May, K., Teng, C. H., & Chang, Y. F. (2010). Comparison of phenotypic and genotypic antimicrobial profiles in *Escherichia coli* and *Salmonella enterica* from the same dairy cattle farms. *Molecular and Cellular Probes*, 24(6): 325-345.
- Schönenbrücher, V., Mallinson, E. T., & Bülte, M. (2008). A comparison of standard cultural methods for the detection of foodborne *Salmonella* species including three new chromogenic plating media. *International Journal of Food Microbiology*, 123(1): 61-66.
- Singla, R., Goel, H., & Ganguli, A. (2014). Novel synergistic approach to exploit the bactericidal efficacy of commercial disinfectants on the biofilms of *Salmonella enterica* serovar Typhimurium. *Journal of Bioscience and Bioengineering*, 118(1): 34-40.
- Smith, R. J., Newton, A. T., Harwood, C. R., & Barer, M. R. (2002). Active but nonculturable cells of *Salmonella enterica* serovar Typhimurium do not infect or colonize mice. *Microbiology*, 148(9): 2717-2726.
- Soares, V. M., Pereira, J. G., Viana, C., Izidoro, T. B., dos Santos Bersot, L., & Pinto, J. P. D. A. N. (2012). Transfer of *Salmonella* Enteritidis to four types of surfaces after cleaning procedures and cross-contamination to tomatoes. *Food Microbiology*, 30(2): 453-456.
- Solomon, E. B., Niemira, B. A., Sapers, G. M., & Annous, B. A. (2005). Biofilm formation, cellulose production, and curli biosynthesis by *Salmonella* originating from produce, animal, and clinical sources. *Journal of Food Protection*, 68(5): 906-912.

- Soon, J. M., Singh, H., & Baines, R. (2011). Foodborne diseases in Malaysia: A review. *Food Control*, 22(6): 823-830.
- Steenackers, H., Hermans, K., Vanderleyden, J., & De Keersmaecker, S. C. (2012). *Salmonella* biofilms: An overview on occurrence, structure, regulation and eradication. *Food Research International*, 45(2): 502-531.
- Su, H. P., Chiu, S. I., Tsai, J. L., Lee, C. L., & Pan, T. M. (2005). Bacterial foodborne illness outbreaks in northern Taiwan, 1995-2001. *Journal of Infection and Chemotherapy*, 11(3): 146-151.
- Su, L. H., Chiu, C. H., Chu, C., & Ou, J. T. (2004). Antimicrobial resistance in nontyphoid *Salmonella* serotypes: A global challenge. *Clinical Infectious Diseases*, 39(4): 546-551.
- Sutton, S. (2010). The most probable number method and its uses in enumeration, qualification, and validation. *Journal of Validation Technology*, 16(3): 35-38.
- Tan, Y. F., Chai, L. C., Mohamad Ghazali, F., Radu, S., & Hareesh, K. K. (2008). Prevalence of *Campylobacter* spp. in retailed ready-to-eat sushi. *International Food Research Journal*, 15(3): 331-336.
- Tenover, F. C., Arbeit, R. D., & Goering, R. V. (1997). How to select and interpret molecular strain typing methods for epidemiological studies of bacterial infections: A review for healthcare epidemiologists. *Infection Control*, 18(6): 426-439.
- The Malaysian Times (2014). Two pupils discharged, five still treated for food poisoning. <http://www.themalayian.com.my/two-pupils-discharged-five-still-treated-for-food-poisoning>. Assessed date: 30 January 2015.
- The Star (2015). Typhoid on the rise in KL. <http://www.thestar.com.my/news/nation/2015/10/20/typhoid-on-the-rise-in-kl-32-cases-reported-since-august-source-of-infection-still-unknown/>. Assessed date: 8 January 2016.
- Thong, K. L., & Modarressi, S. (2011). Antimicrobial resistant genes associated with *Salmonella* from retail meats and street foods. *Food Research International*, 44(9): 2641- 2646.
- Thong, K. L., Goh, Y. L., Radu, S., Noorzaleha, S., Yasin, R., Koh, Y. T., Lim, V. K. E., Rusul, G., & Puthuchery, S. D. (2002). Genetic diversity of clinical and environmental strains of *Salmonella enterica* serotype Weltevreden isolated in Malaysia. *Journal of Clinical Microbiology*, 40(7): 2498-2503.

- Tikoo, A., Tripathi, A. K., Verma, S. C., Agrawal, N., & Nath, G. (2001). Application of PCR fingerprinting techniques for identification and discrimination of *Salmonella* isolates. *Current Science*, 80(8): 1049-1052.
- Tuan Zainazor, T. C. (2006). *Prevalence and molecular characterization of Vibrio Parahaemolyticus isolated from cultured tiger prawns (Penaeus monodon) from Malacca*. (Unpublished Master Thesis). Universiti Putra Malaysia.
- Tunung, R. (2012). *Biosurveillance of Vibrio parahaemolyticus in raw salad vegetables at pre-harvest, retail and domestic kitchen level*. (Published doctoral dissertation). Universiti Putra Malaysia.
- Tunung, R., Chai, L. C., Usha, M. R., Lesley, M. B., Cheah, Y. K., Patrick, G. B., Farinazleen, M. G., Fatimah, A. B., Malakar, P. K., & Son, R. (2007). Incidence and characterization of *Salmonella* species in street food and clinical samples. *Journal of Food Safety*, 27(4): 345-361.
- Tunung, R., Margaret, S. P., Jeyaletchumi, P., Chai, L. C., Tuan Zainazor, T. C., Ghazali, F. M., Nakaguchi, Y., Nishibuchi, M., & Son, R. (2010). Prevalence and quantification of *Vibrio parahaemolyticus* in raw salad vegetables at retail level. *Journal of Microbiology and Biotechnology*, 20(2): 391-396.
- Ubong, A. (2011). *Prevalence and detection of Vibrio cholera in fruit juices and flavored drinks*. (Unpublished Master Thesis). Universiti Putra Malaysia.
- USDA (United States Department of Agriculture) (2012). Microbial Risk Assessment Guideline: Pathogenic Microorganisms with Focus on Food and Water. http://www.fsis.usda.gov/wps/wcm/connect/d79eaa29c53a451eba1c36a76a6c6434/Microbial_Risk_Assessment_Guideline_201001.pdf?MOD=AJPERES. Assessed date: 10 July 2015.
- USDA (United States Department of Agriculture) (2014). Most Probable Number Procedure and Tables. http://www.fsis.usda.gov/wps/wcm/connect/8872ec11-d6a34fcf86df4d87e57780f5/MLG_Appendix2.pdf?MOD=AJPERES. Assessed date: 6 November 2015.
- Valeriano, C., De Oliveira, T. L. C., De Carvalho, S. M., das Graças Cardoso, M., Alves, E., & Piccoli, R. H. (2012). The sanitizing action of essential oil based solutions against *Salmonella enterica* serotype Enteritidis S64 biofilm formation on AISI 304 stainless steel. *Food Control*, 25(2): 673-677.

- van Belkum, A., Tassios, P. T., Dijkshoorn, L., Haeggman, S., Cookson, B., Fry, N. K., Fussing, V., Green, J., Feil, E., Gerner-Smidt, P., Brisse, S., & Struelens, M. (2007). Guidelines for the validation and application of typing methods for use in bacterial epidemiology. *Clinical Microbiology and Infection*, 13(s3): 1-46.
- Van Hoorebeke, S., Van Immerseel, F., Schulz, J., Hartung, J., Harisberger, M., Barco, L., Ricci, A., Theodoropoulos, G., Xylouri, E., De Vylder, J., Ducatelle, R., Haesebrouck, F., Pasmans, F., de Kruif, A., & Dewulf, J. (2010). Determination of the within and between flock prevalence and identification of risk factors for *Salmonella* infections in laying hen flocks housed in conventional and alternative systems. *Preventive Veterinary Medicine*, 94(1): 94-100.
- Viswanathan, P., & Kaur, R. (2001). Prevalence and growth of pathogens on salad vegetables, fruits and sprouts. *International Journal of Hygiene and Environmental Health*, 203(3): 205-213.
- Vugia, D. J., Samuel, M., Farley, M. M., Marcus, R., Shiferaw, B., Shallow, S., Smith, K., & Angulo, F. J. (2004). Invasive *Salmonella* infections in the United States, FoodNet, 1996-1999: Incidence, serotype distribution, and outcome. *Clinical Infectious Diseases*, 38(Supplement 3): S149-S156.
- Waddington, C. S., Darton, T. C., Woodward, W. E., Angus, B., Levine, M. M., & Pollard, A. J. (2014). Advancing the management and control of typhoid fever: A review of the historical role of human challenge studies. *Journal of Infection*, 68(5): 405-418.
- Wang, H., Zhang, X., Zhang, Q., Ye, K., Xu, X., & Zhou, G. (2014). Comparison of microbial transfer rates from *Salmonella* spp. biofilm growth on stainless steel to selected processed and raw meat. *Food Control*, 50: 574-580.
- Westrell, T., Ciampa, N., Boelaert, F., Helwich, B., Korsgaard, H., Chriél, M., Ammon, A., & Mäkelä, P. (2009). Zoonotic infections in Europe in 2007: A summary of the EFSA-ECDC annual report. *Euro surveillance*, 14(3): 785-794.
- WHO (World Health Organization) (2002). Risk assessments for *Salmonella* in eggs and broiler chickens. <ftp://ftp.fao.org/docrep/fao/y4392e/y4392e00.pdf>. Assessed date: 2 April 2014.
- WHO (World Health Organization) (2013). *Salmonella* (non typhoidal). http://www.who.int/media centre/fact sheets/fs_139en/. Assessed date: 30 January 2015.

- Williams, J. G., Kubelik, A. R., Livak, K. J., Rafalski, J. A., & Tingey, S. V. (1990). DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. *Nucleic Acids Research*, 18(22): 6531-6535.
- Wong, D. L. F., Hald, T., van der Wolf, P. J., & Swanenburg, M. (2002). Epidemiology and control measures for *Salmonella* in pigs and pork. *Livestock Production Science*, 76: 215-222.
- Yang, S. J., Park, K. Y., Kim, S. H., No, K. M., Besser, T. E., Yoo, H. S., Kim, S. H., Lee, B. K., & Park, Y. H. (2002). Antimicrobial resistance in *Salmonella enterica* serovars Enteritidis and Typhimurium isolated from animals in Korea: comparison of phenotypic and genotypic resistance characterization. *Veterinary Microbiology*, 86(4): 295-301.
- Yoke-Kqueen, C., Learn-Han, L., Noorzaleha, A. S., Son, R., Sabrina, S., Jiun Horng, S., & Chai-Hoon, K. (2008). Characterization of multiple antimicrobial-resistant *Salmonella enterica* Subsp. *enterica* isolated from indigenous vegetables and poultry in Malaysia. *Letters in Applied Microbiology*, 46(3): 318-324.
- Yousef, A. E., & Carlstrom, C. (2003). *Salmonella*. In A. E. Yousef, & C. Carlstrom (Eds.), *Food Microbiology: A laboratory manual* (pp. 167-205). New Jersey: John Wiley & Sons, Inc.