



UNIVERSITI PUTRA MALAYSIA

***EFFICACY OF EPIDERMAL GROWTH FACTOR TOCOTRIENOL RICH
FRACTION CREAM FORMULATION IN DEEP-PARTIAL THICKNESS
BURN IN Sprague-Dawley RATS***

ASMA BINTI AHMAD ZAINI

IB 2018 11



**EFFICACY OF EPIDERMAL GROWTH FACTOR TOCOTRIENOL RICH
FRACTION CREAM FORMULATION IN DEEP-PARTIAL THICKNESS
BURN IN *Sprague-Dawley* RATS**

By

ASMA BINTI AHMAD ZAINI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Master of Science**

January 2018

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of University Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

EFFICACY OF EPIDERMAL GROWTH FACTOR TOCOTRIENOL RICH FRACTION CREAM FORMULATION IN DEEP-PARTIAL THICKNESS BURN IN *Sprague-Dawley* RATS

By

ASMA BINTI AHMAD ZAINI

January 2018

Chairman : Huzwah Khaza'ai , PhD

Faculty : Institute of Bioscience

Silver Sulfadiazine (SSD) is primarily used as a topical burn treatment. However, it has been reported that SSD cream can cause adverse reactions like allergy and toxicity. Thus, in this study the burn healing properties of epidermal growth factor (EGF) and tocotrienol rich fraction (TRF) formulation were evaluated. This study was conducted in four stages. The first and second stages were to evaluate the efficacy of TRF (3-5%) and EGF (A-C%) separately, meanwhile the third stage was to study the synergistic effect of both compounds in burn wound healing. *Sprague-Dawley* male rats were divided into 6 groups (n=7). The deep-partial thickness burn wounds were performed on the shaved skin with the exposure to 100°C heat for 10 second. Treatment was applied topically once daily to the burned areas for 21 days. The measurable outcomes involved rate of wound contraction, clinical evaluation, H&E staining and cellular population number. The results were analyzed for statistical significance using two-way ANOVA for microscopic study and two way repeated measure for macroscopic study. Bonferonni test was performed for the significant treatment means. The optimum dosage of both ingredients obtained was further used for the formulation of EGF-TRF cream and the synergistic effects were determined. TRF at 3% concentration showed most advanced healing indicated with better cosmetic and histopathological outcome, highest percentage of wound contraction rate at day 5, 9, 13, and 17 with 43.34±2.13, 62.87±1.74, 92.38±2.48 and 100.00±0.00 respectively, lowest count of neutrophils and macrophages. The highest dose (C%) of EGF increased the healing process indicated with better cosmetic and histopathological outcome and highest percentage of wound contraction rate at day 13 and 17 with 78.82±2.40 and 100.00±0.00 respectively. Hence, for the third stage study, A-C% EGF were mixed with 3% TRF. The best formulation was further used in the fourth stage. Microscopic changes of the collagen in the dermal layer for the optimum formulation (C% EGF + 3% TRF) was monitored. Current finding

demonstrated the C% EGF + 3% TRF treatment exhibited excellent gross appearance, highest percentage of wound contraction rate in all experimental period and full histological score as early as day 14. Microscopic evaluation demonstrated that there was a significant acceleration of the epidermal and dermal repair in C% EGF + 3% TRF. Collagen staining also showed increased fibroblast proliferation and collagen synthesis in C% EGF + 3% TRF. Combination of C% EGF + 3% TRF exhibited synergistic effects with better potential than the effects of these two compounds alone in accelerating burn wound healing. In addition, combination of EGF and TRF treatments showed better healing ability as compared to SSD. In conclusion EGF-TRF formulation is capable to accelerate the burn wound healing with better cosmetic outcome in the deep-partial thickness burn on various phase of burn wound healing.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains

**EFIKASI FORMULASI EGF-TRF (FAKTOR PERTUMBUHAN
EPIDERMIS-FRAKSI KAYA TOKOTRIENOL) TERHADAP TIKUS
Sprague-Dawley YANG LUKA TERBAKAR PADA KETEBALAN SEPARA**

Oleh

ASMA BINTI AHMAD ZAINI

Januari 2018

Pengerusi : Huzwah Khaza'ai, PhD

Fakulti : Institut Biosains

Silver Sulfadiazine (SSD) digunakan sebagai ubat sapuan utama untuk luka akibat terbakar. Walau bagaimanapun, krim SSD dilaporkan boleh menyebabkan kesan buruk seperti alergi dan toksik. Oleh itu, dalam kajian ini, sifat penyembuhan luka akibat terbakar dengan formulasi faktor pertumbuhan epidermis (EGF) dan fraksi kaya tocotrienol (TRF) dinilai. Kajian ini dijalankan dalam empat peringkat. Peringkat pertama dan kedua adalah untuk menilai kesan tindak balas dos TRF dan EGF secara berasingan, manakala tahap ketiga adalah mengkaji kesan sinergistik kedua-dua sebatian dalam penyembuhan luka terbakar. Tikus jantan Sprague-Dawley dibahagikan kepada 6 kumpulan ($n = 7$). Luka terbakar ketebalan separa dilakukan ke atas kulit yang dicukur dengan pendedahan kepada suhu panas 100°C selama 10 saat. Rawatan diberikan secara sapuan sekali sehari pada kawasan yang terbakar selama 21 hari. Keputusan untuk ketiga-tiga peringkat ini ditentukan mengikut gabungan kadar pengecutan luka, penilaian klinikal, pewarnaan H & E dan bilangan populasi sel kulit. Data dianalisis menggunakan dua cara ANOVA untuk kajian mikroskopik dan langkah berulang ANOVA untuk data makroskopik. Ujian Bonferonni dilakukan untuk penentuan data yang signifikan. Dos yang optimum daripada kedua-dua ramuan ini telah digunakan untuk membuat rumusan krim EGF-TRF dan kesan sinergi telah ditentukan. 3% TRF menunjukkan kadar pengecutan luka yang paling tinggi pada hari 5, 9, 13 and 17 dengan nilai 43.34 ± 2.13 , 62.87 ± 1.74 , 92.38 ± 2.48 dan 100.00 ± 0.00 , bilangan neutrophil dan makrofaj yang terendah. Peningkatan dos yang responsif dapat diperhatikan dalam dos C% EGF ditunjukkan oleh hasil kosmetik yang baik, hasil histopatologi yang baik dan peratusan tertinggi kadar penguncupan luka pada hari 13 dan 17 dengan 78.82 ± 2.40 dan 100.00 ± 0.00 masing-masing. Oleh itu, untuk kajian peringkat ketiga, A-C% EGF telah dicampur dengan TRF 3%. Formulasi yang terbaik disiasat di peringkat keempat. Perubahan mikroskopik kolagen dalam lapisan dermal untuk formulasi terbaik telah (C% EGF + 3% TRF) dipantau. Penemuan terkini

menunjukkan rawatan C% EGF + 3% TRF menghasilkan rupa luaran luka yang baik, peratusan tertinggi untuk kadar penguncupan luka dalam semua tempoh eksperimen dan skor histologi yang penuh seawal hari 14. Penilaian mikroskopik menunjukkan bahawa terdapat akselerasi ketara pembaikan epidermis dan dermis dengan penggunaan C% EGF + 3%.TRF. Pewarnaan kolagen juga menunjukkan peningkatan proses percambahan fibroblast dan sintesis kolagen dalam kumpulan C% EGF + 3% TRF. Kombinasi C% EGF + 3% TRF menunjukkan kesan sinergi dengan potensi yang lebih baik daripada kesan kedua-dua rawatan itu secara sendirian dan rawatan lain dalam mempercepatkan penyembuhan luka terbakar ketebalan separa. Gabungan rawatan EGF dan TRF menunjukkan keupayaan penyembuhan yang lebih baik berbanding dengan SSD. Kesimpulannya, formulasi EGF-TRF mampu mempercepatkan penyembuhan luka terbakar tahap kedua di pelbagai proses tahap penyembuhan luka dan mempercepat proses penyembuhan secara langsung.

ACKNOWLEDGEMENTS

In the name of Allah, The most Gracious and Merciful.

First and foremost, I am greatly indebted to my supervisor, Dr Huzwah Khaza'ai for her endless guidance throughout the development of this study. Her comments, patience and examples have inspired me in so many ways. A highly gratitude also goes to my committee members Dr Razana Md Ali for serving as my supervisor with various suggestions and also for the help and encouragement during the research work.

I would also like to acknowledge with much appreciation to Ms. Mu'mina, Mr. Ha'iz Sokhini, and Ms. Gayathri for helping me tremendously with the animal work. Their assistances were vital for my experiment. Many thanks are also due to all staff in Histopathology laboratory, Faculty of Medicine and Health Sciences especially to Ms. Normah and Ms. Juita Chupri for their kind assistance during research. Many more persons participated in various ways to ensure my research succeed, but special mentions are due to my labmates especially to Miss Guo Hui Fang for valuable exchange of ideas and support, Miss Najwa, Miss Afifah, Miss Hazirah and Miss Sarah. I am honoured to have the opportunity to work with such a dedicated postgraduate student who taught me the meaning of teamwork and camaraderie.

My deepest gratitude to my father and mother, Mr. Ahmad Zaini bin Hussin and Mrs. Rozita Abd. Samad for their endless support. Thanks also to my siblings (Umair, Ammar and Muadz). Last but not least I am immensely thankful for the loved, advised and support of my husband Mr. Ahmad Khaliq in my pursuit for higher education and expressed understanding and consideration towards me. This thesis is dedicated to my son Faheem bin Ahmad Khaliq whose inspired me to work harder.

I certify that a Thesis Examination Committee has met on 22 January 2018 to conduct the final examination of Asma binti Ahmad Zaini on her thesis entitled "Efficacy of Epidermal Growth Factor Tocotrienol Rich Fraction Cream Formulation in Deep-Partial Thickness Burn in *Sprague-Dawley* Rats" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Suhaili binti Abu Bakar @ Jamaludin, PhD

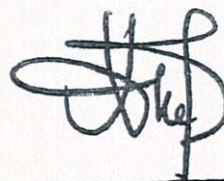
Senior Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Norhaizan binti Mohd Esa, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Internal Examiner)

Mahanem Mat Noor, PhD

Associate Professor
Universiti Kebangsaan Malaysia
Malaysia
(External Examiner)



NOR AINI AB. SHUKOR, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 28 March 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Huzwah Khaza'ai, PhD

Senior Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Razana Md Ali, PhD

Senior Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric Number: Asma binti Ahmad Zaini, GS36125

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____
Name of
Chairman of
Supervisory
Committee: Dr. Huzwah Khaza'ai

Signature: _____
Name of
Member of
Supervisory
Committee: Dr. Razana Md Ali

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xix
CHAPTER	
1 INTRODUCTION	1
1.1 Background of study	1
1.2 Problem statement	3
1.3 Research objective	3
1.3.1 General objective	3
1.3.2 Specific objectives	3
1.4 Hypothesis	4
1.5 Significance of the study	4
2 LITERATURE REVIEWS	5
2.1 Integumentary system	5
2.2 Permeability of the skin	6
2.3 Burns	7
2.3.1 Burn wound healing	8
2.3.2 Stages and mechanism of wound healing	8
2.3.2.1 Inflammatory phase	8
2.3.2.2 Proliferative phase	9
2.3.2.3 Maturation and remodeling	10
2.4 Epidermal growth factor (EGF)	11
2.4.1 Chemical structure of EGF	11
2.4.2 Sources, distribution, absorption and metabolism EGF	11
2.4.3 Properties of EGF	11
2.4.3.1 Function	11
2.4.3.2 Mechanism of action of EGF	12
2.4.3.3 EGF and wound healing	13
2.4.3.4 Pharmacological study of EGF	14
2.5 Vitamin E	15
2.5.1 Chemical structure of vitamin E	15

2.5.2	Absorption, transportation and metabolism of vitamin E	16
2.5.3	Properties of Vitamin E	17
2.5.3.1	Function and mechanism of action	17
2.5.3.2	Wound healing properties	17
2.5.3.3	Antioxidative agent	18
2.6	Redox reaction	19
2.6.1	Introduction to redox reaction	19
2.6.2	Redox reaction in wound healing	20
2.7	Antioxidants	21
2.7.1	Antioxidant enzyme	21
2.7.2	Lipid peroxidation	22
3	GENERAL MATERIALS AND METHODOLOGY	24
3.1	Materials	24
3.1.1	Experimental animal preparation	24
3.1.2	Preparation of formulation	24
3.1.3	Burn wound creation and treatment protocol	24
3.1.4	Macroscopic study	25
3.1.4.1	Gross appearance	25
3.1.4.2	Clinical evaluation	25
3.1.4.3	Wound contraction	25
3.1.5	Microscopic study	25
3.1.5.1	Hematoxylin and eosin staining (H&E staining)	25
3.1.5.2	Masson's trichrome staining	25
3.2	Methodology	26
3.2.1	Experimental design	26
3.2.2	Experimental animal	26
3.2.2.1	The effect of different concentration of TRF on burn healing	26
3.2.2.2	The effect of different concentration of EGF on burn healing	27
3.2.2.3	The effect of 3% TRF mixed with different concentration of EGF.	27
3.2.2.4	The effect of C% EGF + 3% TRF on collagen changes	27
3.2.3	Preparation of formulation	27
3.2.3.1	Preparation of base cream	27
3.2.3.2	Preparation of TRF cream	27
3.2.3.3	Preparation of EGF cream	28
3.2.3.4	Preparation of TRF-EGF cream	28
3.2.4	Preparation of the skin	28
3.2.5	The burn injury	29
3.2.5.1	Thermal source	29
3.2.5.2	Infliction of the burn wound	29
3.2.6	Treatment protocol	31
3.2.7	Macroscopic study	33

	3.2.7.1 Gross appearance of the wound	33
	3.2.7.2 Clinical evaluation	33
	3.2.7.3 Rate of wound contraction	33
3.2.8	Microscopic study	34
	3.2.8.1 Preparation of the slide	34
	3.2.8.2 Light Microscope Evaluatio	35
	3.2.8.3 Haematoxylin and Eosin staining	36
3.2.9	Statistical analysis	37
4	THE EFFECT OF DIFFERENT CONCENTRATION OF TOCOTRIENOL RICH FRACTION UPON WOUND HEALING	38
4.1	Introduction	38
4.2	Materials	39
4.3	Methodology	39
4.4	Results	39
	4.4.1 Gross appearance of the wound	39
	4.4.2 Clinical evaluation	42
	4.4.3 Rate of wound contraction	44
	4.4.4 Haematoxylin and eosin staining	45
	4.4.4.1 H&E stained section on day 1	45
	4.4.4.2 H&E stained section on day 7	47
	4.4.4.3 H&E stained section on day 14	49
	4.4.4.4 H&E stained section on day 21	51
	4.4.5 Cell populations in burn wound	54
4.5	Discussion	56
5	THE EFFECT OF DIFFERENT CONCENTRATION OF EGF ON WOUND HEALING	60
5.1	Introduction	60
5.2	Materials	61
5.3	Methods	61
5.4	Result	61
	5.4.1 Gross appearance of the wound	61
	5.4.2 Clinical evaluation	64
	5.4.3 Rate of wound contraction	66
	5.4.4 Haematoxylin and eosin staining	67
	5.4.4.1 H&E stained section on day 1	67
	5.4.4.2 H&E stained on day 7 ($\times 10$)	69
	5.4.4.3 H&E stained section on day 14	71
	5.4.4.4 H&E stained section on day 21	74
	5.4.5 Cell populations in burn wound	76
5.5	Discussion	78

6	THE SYNERGISTIC EFFECT BETWEEN 3% TRF EACH MIXES WITH A%, B% AND C% EGF UPON WOUND HEALING ACTIVITY	82
6.1	Introduction	82
6.2	Materials	82
6.3	Methods	82
6.4	Result	83
6.4.1	Gross appearance of the wound	83
6.4.2	Clinical evaluation	85
6.4.3	Estimation of wound contraction rate	86
6.4.4	Haematoxylin and eosin staining	87
6.4.4.1	H&E stained section on day 1	88
6.4.4.2	H&E stained section on day 7	89
6.4.4.3	H&E stained section on day 14	90
6.4.4.4	H&E stained section on day 21	93
6.5	Discussion	93
7	THE EFFECT OF THE OPTIMUM FORMULATON (3% TRF+ C% EGF) ON THE DERMAL COLLAGEN CHANGES	97
7.1	Introduction	97
7.2	Materials	99
7.3	Methodology	99
7.4	Results	100
7.4.1	Effects of EGF-TRF on the dermal collagen changes	100
7.4.1.1	Masson's Trichrome stained section on day 7	101
7.4.1.2	Masson's Trichrome stained section on day 14	102
7.4.1.3	Masson's Trichrome stained section on day 21	103
7.4.2	Effects of TRF-EGF on the total healing criteria	104
7.4.2.1	Masson's trichrome stained section on day 7	107
7.4.2.2	Masson's trichrome stained section on day 14	109
7.4.2.3	Masson's trichrome stained section on day 21	111
7.4.3	Fibroblasts counts	113
7.5	Discussion	114
8	GENERAL RESULTS AND DISCUSSION	118
9	CONCLUSION AND RECOMMENDATION FOR FUTURE STUDIES	122
	BIBLIOGRAPHY	123
	APPENDICES	136
	BIODATA OF STUDENT	144
	LIST OF PUBLICATION	145

LIST OF TABLES

Table	Page	
3.1	Quantitative histological scoring of H&E staining	37
4.1	Clinical evaluations of the wounds were observed for 21 days	43
4.2	Percentage of wound contraction in control groups, 3%, 4% and 5% TRF groups	44
4.3	The cell counts of control group as compared to different concentration of TRF at different time intervals	55
5.1	Clinical evaluations of the wounds were observed for 21 days	65
5.2	Percentage of wound contraction in control groups, A%, B% and C% EGF groups	66
5.3	The cell counts of control group and different concentration of EGF at different time intervals	77
6.1	Clinical evaluations of the wounds were recorded for 21 days	85
6.2	Percentage of wound contraction in EGF alone groups, 3% TRF and combination formulations (3% TRF + EGF)	86
6.3	Reference for quantitative histopathologic scoring of the wound healing parameter based on H&E staining	87
6.4	H&E score in EGF alone groups, 3% TRF and combination formulations (3% TRF + EGF)	88
7.1	Semi-quantitative histological scoring table of Masson's trichrome staining	100
7.2	Quantitative histological scoring table of Masson's trichrome staining	100
7.3	Semi quantitative histological findings in Masson's trichrome	101
7.4	Quantitative histologic findings of total dermal healing in Masson's trichrome staining	106
7.5	The cell counts of control group, 3% TRF and C% EGF + 3% TRF at different time intervals	113

LIST OF FIGURES

Figure		Page
2.1	Human skin diagram shows three layers of the skin with different cells	5
2.2	Basic skin anatomy showing the depth of injury for first, second and third degree burn	7
2.3	Epidermal growth factor (EGF) signaling pathway	12
2.4	Roles of hEGF in various applications.	15
2.5	The relation between wound healing and ROS	20
3.1	Shaved dorsal part of the rat	28
3.2	Burn wound creation apparatus	29
3.3	Infliction of the burn lesion	30
3.4	A) Standard deep partial thickness burn 5 min after the burn; B) H&E staining at 5 min post-burn ($\times 10$)	30
3.5	A) Standard deep partial thickness burn on day 7 th after the burn; B) H&E staining at day 7 th post-burn ($\times 10$)	31
3.6	Summary of the experimental design	32
3.7	A photographic method for burn wound area measurement	34
3.8	Photomicrographs of wound section showing cells population in H&E slides ($\times 40$)	36
4.1	Gross appearance of the control groups, 3%, 4% and 5% TRF	41
4.2	Microscopic view of the histological sections of control groups and different TRF groups at day 1 post-burn stained with H&E ($\times 10$)	46
4.3	Microscopic view of the histological sections of control groups and different TRF groups at day 7 th of post-burn stained with H&E ($\times 10$)	48
4.4	Microscopic view of the histological sections of control groups and different TRF groups at day 14 th of post-burn stained with H&E ($\times 10$)	50

4.5	Microscopic view of the histological sections of TRF groups at day 14 th of post-burn for granulation tissue observation and alignment of endothelial cells (×40)	51
4.6	: Microscopic view of the histological sections of control groups and different TRF groups at day 21 st of post-burn stained with H&E (×10)	53
5.1	Gross appearance of control groups, A%,B% and C% EGF	63
5.2	Microscopic view of the histological section of control groups and different EGF groups at day 1 post-burn stained with H&E (×10)	68
5.3	Microscopic view of the histological sections of control groups and different EGF groups at day 7 th post-burn stained with H&E (×10)	70
5.4	Microscopic view of the histological sections of control groups and different EGF groups at day 14 th post-burn stained with H&E (×10)	72
5.5	Microscopic view of the histological sections of EGF groups at day 14 th of post-burn for granulation tissue observation and alignment of endothelial cells (×40)	73
5.6	Microscopic view of the histological sections of control groups and different EGF groups at day 21 st post-burn stained with H&E (×10)	75
6.1	Gross appearance of wound sites in EGF alone groups, 3% TRF and combination formulations (3% TRF + EGF)	84
6.2	Photomicrographs of wound section of combination formulation; A%, B%, C% EGF each mix with 3% TRF at day 1 post burned stained with H&E (×10)	89
6.3	Microscopic view of histological section of combination formulation; A%, B%, C% EGF each mix with 3% TRF at day 7 post burned stained with H&E (×10)	90
6.4	Microscopic view of the granulation tissue of combination formulation; A%, B%, C% EGF each mix with 3% TRF at day 14 post burned stained with H&E (×10; ×40)	92
6.5	Microscopic view of the histological sections of combination formulation; A%, B%, C% EGF each mix with 3% TRF at day 21 post burned stained with H&E (×10)	93
7.1	Microscopic view of the histological sections of control groups, 3% TRF and C% EGF + 3% TRF at day 7 post-burn in Masson's trichrome staining (×10)	102

7.2	Microscopic view of the histological sections of control groups, 3% TRF and C% EGF + 3% TRF at day 14 post burned in Masson's trichrome staining (×10)	103
7.3	Microscopic view of the histological sections of control groups, 3% TRF and C% EGF + 3% TRF at day 21 post burned in Masson's trichrome staining (×10)	104
7.4	Microscopic view of histological section of total dermal healing in control groups, 3% TRF and C% EGF + 3% TRF at day 7 post burned in Masson's trichrome staining (×10)	108
7.5	Microscopic view of histological section of total dermal healing in control groups, 3% TRF and C% EGF + 3% TRF at day 14 post burned in Masson's trichrome staining (×40)	110
7.6	Microscopic view of histological section of total dermal healing in control groups, 3% TRF and C% EGF + 3% TRF at day 21 post burned in Masson's trichrome staining (×40)	112

LIST OF ABBREVIATIONS

ECM	Extracellular matrix
EGF	Epidermal growth factor
EGFR	Epidermal growth factor receptor
GPX	Glutathione peroxidase
GRB 2	Growth factor receptor-bound protein two
GTP	Guanosine-5'-triphosphate
GTPase	Guanosine-5'-triphosphatase
H&E	Hematoxylin and eosin
H ₂ O ₂	Hydrogen peroxide
HDLs	High-density lipoproteins
hEGF	Human epidermal growth factor
MAP	Mitogen-activated protein
MDA	Malondialdehyde
MMP	Matrix metalloproteinase
NADPH	Nicotinamide adenine dinucleotide phosphate
O ₂	Oxygen
PDGF	Platelet-derived growth factor
PUFA	Polyunsaturated fatty acids
ROS	Reactive oxygen species
SOD	Superoxide dismutase
SSD	Silver sulfadiazine
VLDLs	Very low density lipoprotein

α -TTP

Alpha tocopherol transport protein





© COPYRIGHT UPM

CHAPTER 1

INTRODUCTION

1.1 Background of study

Burns are the fourth most common type of trauma worldwide which results in limb deformity, large amount of expenditure in health care, and trauma in both physical and psychological status of an individual (Lazarus et al., 1994). According to the World Health Organization (WHO), 238,800 individuals died due to fire-related burn in the year 2000 (Orgill & Ogawa, 2013)(Orgill and Ogawa, 2013; Afify et al., 2012). Annually, about 2 million people are wounded, 80 000 warded, and 6,500 fatalities due to burn wound in the United States. As in Malaysia, incidence rates of burn is 31, 176 cases per year. The remodeling of damages skin initiated by edema, skin inflammation and scar formation. Poor wound management and delayed progress of wound healing will lead to keloid, hypertrophic, non-raised and contracture scar (Tavares Pereira et al., 2012). Incidence rates of hypertrophic scarring is up to 91% for burn injury (Gauglitz et al., 2011). These consequences have an impact on the psychological and social behaviour of an individual (Clouatre et al., 2013). Skin-related complications also significantly reduced the ability of an individual to move by causing joint contracture, pain from inflammatory mediators and the worst case is that it could leads to deformation from severe scarring (Orgill & Ogawa, 2013).

The damage of epithelial part of the skin exposed the wound area unprotected (Enoch & Leaper, 2005). Generally, pathophysiology and management of burn and normal wound healing is similar and divided into four different but intersecting stages: hemostasis, inflammation, proliferation and remodeling. Physiologically, wound can heal by regeneration and reparative process. Successful tissue regeneration is the ideal form of healing process giving good cosmetic and functional results. On the other hand, in reparative process specialized tissue is replaced with collagen and result in a loss of functional and cosmetic outcome. The reparative events are generally preceded by hemostatic and inflammatory phenomena which may in turn influence the final result of wound healing (Orgill and Ogawa, 2013). It was suggested by Hsu and Mustoe (2010) that a proper technique to optimize the healing process is by reducing the inflammation, increasing tissue regeneration, minimizing tissue destruction and providing a moist environment on the wound area. There are four different classes of burn namely; superficial, superficial partial thickness, deep partial thickness and full thickness burn. However, the main concern in burn unit is infection that will trigger the other complications in healing burn wound. In deep partial-thickness burn, the patients have a high risk to develop into a full-thickness burn (Chan., et al 2002). Hence, it is a critical requirement to create a better treatment for deep-partial thickness burn.

The goal of wound care is to heal wounds in the shortest time with minor pain and scarring. It is concluded that antibacterial formulation can prevent infection from bacteria, nevertheless, they can hinder healing cells from proliferating during wound healing followed by delayed in wound closure. Silver sulfadiazine (SSD) is a standard topical treatment for burn but could cause adverse side effects upon long term usage. In addition, most current therapeutic approaches for burn healing treatment can also cause scar formation and disfigurement (Aarabi et al., 2007). They are different functions of wound healing formulation in a market. The active ingredients and properties were depending on their main function; whether to clean, to protect, to keep in good condition or to change the appearance.

Vitamin E composed of eight different isoforms, four tocopherol and four tocotrienol. The difference is that tocotrienol has an unsaturated phytyl tail at carbon number 3,7 and 11 whereas tocopherol possess saturated isoprenoid side chain. These eight forms of vitamins E have different biological activity. It is well establish that vitamin E has the ability to prevent the lipid oxidation and can act as antioxidant molecules to scavenge reactive oxygen species (ROS). The action of vitamin E in accelerating wound repair is increasingly welcomed. However, much of the present work on wound healing has focused on α -tocopherol. It has been proven that tocotrienols have higher antioxidant activity than the tocopherols and it possess a few medicinal properties that are not present in tocopherol (Zingg, 2007). These characteristic are due to the presence of three double bonds on the hydrophobic side chain of tocotrienol (Sen et al., 2010). Therefore, vitamin E in the form of tocotrienol is highly welcomed and should lead to the development of strategies aimed specifically in reducing ROS produced upon burn injury.

Due to the importance of cellular proliferation during burn wound healing, any treatment that can increase the mitogenic effect of healing cells is highly desirable. Each of the phases in wound healing is controlled and regulated by cytokines called growth factors. Primary function of growth factors in wound healing is as mitogen, angiogenic and chemoattractant to command the progression of healing stages. Currently, cytokines have a limited role in clinical practice. The only growth factor currently available commercially is platelet-derived growth factor (PDGF). Wound healing managed with exogenous growth factors has been beneficial on the healing process. However, all of these studies were focused on the healing ability of vitamin E and growth factor as individual compound. By far, there are no experimental reports on the study of the healing ability of the combination of tocotrienol-rich fraction (TRF) and epidermal growth factor (EGF). Therefore it is of current interest to study on the interaction of TRF and EGF on deep partial thickness burn wound. In concert with combining antioxidant properties of TRF and mitogenic properties of EGF, increase ability of wound healing is expected. In this regards, reduced inflammation and increase cellular proliferation in wound healing could be achieved and eventually offered an alternative in topical based therapy.

1.2 Problem statement

One of the major culprit in burn injury is the presence of oxygen radicals that can form chain reaction of lipid peroxidation. Increase free radicals will result in longer inflammation which may cause the wound to lock into chronic state and resulting in delayed wound healing. Fortunately, protection against oxidative stress can be provided by radical scavenger compound such as vitamin E which can help to arrest the chain propagation. Vitamin E usually used in topical formulation, such as in wound healing and cosmetic products. Nevertheless, most vitamin E skin care products contain only alpha-tocopherol and synthetic alpha-tocopheryl acetate, which involve hydrolyzation during absorption to demonstrate its activity (Henegouwen et al., 1995). Other factor that will delayed wound healing include minimal re-epithelialization and minimal cellular proliferation. Increased cellular proliferation can be provided by cytokines and growth factor. EGF can exert a powerful mitogenic effect particularly on epithelial cells and fibroblasts. These two cells are responsible for regeneration and collagen production of the skin. Therefore in this study, the synergistic effect of both TRF and EGF in burn wound healing were evaluated.

1.3 Research objective

1.3.1 General objective

To elucidate the burn wound healing efficacy of EGF-TRF formulation in *Sprague-Dawley* rats.

1.3.2 Specific objectives

1. To evaluate the macroscopic and histopathological changes of burn wound healing treated with different concentration of TRF and EGF separately upon wound healing.
2. To investigate the synergistic effect between the optimum concentration of TRF mix with three different concentration of EGF separately upon wound healing activity via macroscopic and histopathological changes.
3. To compare and identify the best combination formulation for burn wounds.
4. To monitor the dermal collagen changes of the best treatment formulation at specific time of healing.

1.4 Hypothesis

EGF-TRF formulation is efficient in treating deep-partial thickness burn.

1.5 Significance of the study

Silver sulfadiazine (SSD) is a standard topical treatment for burn but could cause adverse side effects upon long term usage. SSD can cause allergy and toxicity. It is concluded that antibacterial formulation can prevent infection from bacteria, nevertheless, they can hinder healing cells from proliferating during wound healing followed by delayed in wound closure. So, it is important to find safer and effective treatment without any toxicity effect and aimed at promoting the stage of wound healing with better cosmeceutical outcome.

BIBLIOGRAPHY

- Aarabi, S., Longaker, M. T., and Gurtner, G. C. (2007). Burns and Trauma : New Approaches to Treatment Evolution of Patient Care, *PLoS Med* 4(9): 234.
- Afify, M. M., Mahmoud, N. F., Abd El Azzim, G. M., and El Desouky, N. A. (2012). Fatal burn injuries: A five year retrospective autopsy study in Cairo city, Egypt. *Egyptian Journal of Forensic Sciences*, 2(4), 117–122.
- Agazie, Y. M., and Hayman, M. J. (2003). Molecular mechanism for a role of SHP2 in epidermal growth factor receptor signaling. *Molecular and Cellular Biology*, 23(21), 7875–7886.
- Aggarwal, B. B., Sundaram, C., Prasad, S., and Kannappan, R. (2010). Tocotrienols, the vitamin E of the 21st century: Its potential against cancer and other chronic diseases. *Biochemical Pharmacology*, 80(11), 1613–1631.
- Alsaqr, A., Rasouly, M., and Musteata, F. M. (2015). Investigating transdermal delivery of vitamin D(3). *AAPS PharmSciTech*, 16(4), 963–972.
- Andrews, S. N., Jeong, E., and Prausnitz, M. R. (2013). Transdermal delivery of molecules is limited by full epidermis. *Pharmaceutical Research*, 30, 1099–1109.
- Arturson, G. (2000). Forty years in burns research - The postburn inflammatory response. *Burns : Journal of the International Society for Burn Injuries*, 26(7), 599–604.
- Baie, S. H., and Sheikh, K. A. (2000). The wound healing properties of *Channa striatus*-cetrimide cream-- tensile strength measurement. *Journal of Ethnopharmacology*, 71(1–2), 93–100.
- Barbosa, E., Faintuch, J., Machado Moreira, E. A., Goncalves da Silva, V. R., Lopes Pereira, M. J., Martins Fagundes, R. L., and Filho, D. W. (2009). Supplementation of vitamin E, vitamin C, and zinc attenuates oxidative stress in burned children: a randomized, double-blind, placebo-controlled pilot study. *Journal of Burn Care and Research : Official Publication of the American Burn Association*, 30(5), 859–866.
- Barrientos, S., Stojadinovic, O., Golinko, M. S., Brem, H., and Tomic-Canic, M. (2008). Growth factors and cytokines in wound healing. *Wound Repair and Regeneration : Official Publication of the Wound Healing Society and the European Tissue Repair Society*, 16(5), 585–601.
- Baum, C. L., and Arpey, C. J. (2005). Normal cutaneous wound healing: clinical correlation with cellular and molecular events. *Dermatologic Surgery : Official Publication for American Society for Dermatologic Surgery*, 31(6),

674–86.

- Baumann, L. S., and Spencer, J. (1999). The effects of topical vitamin E on the cosmetic appearance of scars. *Dermatologic Surgery : Official Publication for American Society for Dermatologic Surgery*, 25(4), 311–315.
- Bazzoni, F., Cassatella, M. A., Rossi, F., Ceska, M., Dewald, B., and Baggiolini, M. (1991). Phagocytosing neutrophils produce and release high amounts of the neutrophil-activating peptide 1/interleukin 8. *The Journal of Experimental Medicine*, 173(3), 771–774.
- Beijersbergen van Henegouwen, G. M., Junginger, H. E., & de Vries, H. (1995). Hydrolysis of RRR-alpha-tocopheryl acetate (vitamin E acetate) in the skin and its UV protecting activity (an in vivo study with the rat). *Journal of Photochemistry and Photobiology. B, Biology*, 29(1), 45–51.
- Bell, E., Ivarsson, B., and Merrill, C. (1979). Production of a tissue-like structure by contraction of collagen lattices by human fibroblasts of different proliferative potential in vitro. *Proceedings of the National Academy of Sciences of the United States of America*, 76(3), 1274–1278.
- Berlanga-acosta, J., Gabilondo-cowley, J., Pedro, L., Castro-santana, M. D., Ernesto, L., and Herrera-martinez, L. (2009). Epidermal growth factor in clinical practice – a review of its biological actions , clinical indications and safety implications, 6(5), 331–346.
- Bioscience, F., and Virginia, R. (2004). Wound healing. An overview of cute, fibrotic and delayed healing. Robert F. Diegelmann and Melissa C. Evans, (4), 283–289.
- Birben, E., Sahiner, U. M., Sackesen, C., Erzurum, S., and Kalayci, O. (2012). Oxidative Stress and Antioxidant Defense. *The World Allergy Organization Journal*, 5(1), 9–19.
- Bishop, A. (2008). Role of oxygen in wound healing. *Journal of Wound Care*, 17(9), 399–402.
- Bochaton-Piallat, M.-L., Gabbiani, G., and Hinz, B. (2016). The myofibroblast in wound healing and fibrosis. *Journal of Research*, 12 (5), 752.
- Bouayed, J., and Bohn, T. (2010). Exogenous antioxidants-double-edged swords in cellular redox state: Health beneficial effects at physiologic doses versus deleterious effects at high doses. *Oxidative Medicine and Cellular Longevity*, 3(4), 228–237.
- Boyce, D., and Thomas, D. (2008). Epidermal gorth factor - past, present and future. *The Surgeon*, 6(3), 172–177.

- Breuing, K., Andree, C., Helo, G., Slama, J., Liu, P. Y., and Eriksson, E. (1997). Growth factors in the repair of partial thickness porcine skin wounds. *Plastic and Reconstructive Surgery*, 100(3), 657–664.
- Broughton, G. 2nd, Janis, J. E., and Attinger, C. E. (2006). The basic science of wound healing. *Plastic and Reconstructive Surgery*, 117(7 Suppl), 12S–34S.
- Brown, G. L., Curtsinger, L. 3rd, Brightwell, J. R., Ackerman, D. M., Tobin, G. R., Polk, H. C. J., Schultz, G. S. (1986). Enhancement of epidermal regeneration by biosynthetic epidermal growth factor. *The Journal of Experimental Medicine*, 163(5), 1319–1324.
- Brown, G. L., Nanney, L. B., Griffen, J., Cramer, A. B., Yancey, J. M., Curtsinger, L. J. 3rd, Lynch, J. B. (1989). Enhancement of wound healing by topical treatment with epidermal growth factor. *The New England Journal of Medicine*, 321(2), 76–79.
- Buckley, A., Davidson, J. M., Kamerath, C. D., and Woodward, S. C. (1987). Epidermal growth factor increases granulation tissue formation dose dependently. *The Journal of Surgical Research*, 43(4), 322–328.
- Carpenter, G., and Cohen, S. (1990). Epidermal growth factor. *The Journal of Biological Chemistry*, 265(14), 7709–7712.
- Chan, K. Y., Mbchb, H. I., Zailani, M., Kumar, S., and Somasundaram, S. (2002). A review of burns patients admitted to the burns unit of hospital Universiti Kebangsaan Malaysia, 57(4), 418–425.
- Chang, R.-K., Raw, A., Lionberger, R., and Yu, L. (2013). Generic development of topical dermatologic products: Formulation development, process development, and testing of topical dermatologic products. *The AAPS Journal*, 15(1), 41–52.
- Chithra, P., Sajithlal, G. B., and Chandrakasan, G. (1998). Influence of Aloe vera on collagen characteristics in healing dermal wounds in rats. *Molecular and Cellular Biochemistry*, 181(1–2), 71–76.
- Church, D., Elsayed, S., Reid, O., Winston, B., and Lindsay, R. (2006). Burn wound infections. *Clinical Microbiology Reviews*, 19(2), 403–434.
- Chvapil, M., Speer, D. P., Owen, J. A., and Chvapil, T. A. (1984). Identification of the depth of burn injury by collagen stainability. *Plastic and Reconstructive Surgery*, 73(3), 438–441.
- Cohen, S., Carpenter, G., and King, L. J. (1980). Epidermal growth factor-receptor-protein kinase interactions. Co-purification of receptor and epidermal growth factor-enhanced phosphorylation activity. *The Journal of Biological Chemistry*, 255(10), 4834–4842.

- Creely, J. J., DiMari, S. J., Howe, A. M., Hyde, C. P., and Haralson, M. A. (1990). Effects of epidermal growth factor on collagen synthesis by an epithelioid cell line derived from normal rat kidney. *The American Journal of Pathology*, 136(6), 1247–1257.
- Cumming, B. D., McElwain, D. L. S., and Upton, Z. (2010). A mathematical model of wound healing and subsequent scarring. *Journal of the Royal Society Interface*, 7(42), 19–34.
- Darby, I. A., Laverdet, B., Bonté, F., and Desmoulière, A. (2014). Fibroblasts and myofibroblasts in wound healing. *Clinical, Cosmetic and Investigational Dermatology*, 7, 301–311.
- Dhall, S., Do, D. C., Garcia, M., Kim, J., Mirebrahim, S. H., Lyubovitsky, J., Martinsgreen, M. (2014). Generating and Reversing Chronic Wounds in Diabetic Mice by Manipulating Wound Redox Parameters, 25-29.
- Doillon, C. J., Dunn, M. G., Bender, E., and Silver, F. H. (1985). Collagen fiber formation in repair tissue: development of strength and toughness. *Collagen and Related Research*, 5(6), 481–492.
- Driskell, R. R., Lichtenberger, B. M., Hoste, E., Kretzschmar, K., Simons, B. D., Charalambous, M., Watt, F. M. (2013). Distinct fibroblast lineages determine dermal architecture in skin development and repair. *Nature*, 504(7479), 277–281.
- Dyson, M., Young, S., Pendle, C. L., Webster, D. F., and Lang, S. M. (1988). Comparison of the effects of moist and dry conditions on dermal repair. *The Journal of Investigative Dermatology*, 91(5), 434–439.
- Elbischger, P. J., Bischof, H., Holzzapfel, G. A., and Regitnig, P. (2005). Computer vision analysis of collagen fiber bundles in the adventitia of human blood vessels. *Studies in Health Technology and Informatics*, 113, 97–129.
- Enoch, S., and Leaper, D. J. (2005). Basic science of wound healing. *Surgery (Oxford)*, 23(2), 37–42.
- Flora, S. J. S. (2009). Structural, chemical and biological aspects of antioxidants for strategies against metal and metalloids exposure. *Oxidative Medicine and Cellular Longevity*, 2(4), 191–206.
- Foschi, D., Trabucchi, E., Musazzi, M., Castoldi, L., Di Mattia, D., Radaelli, E., Berlusconi, A. (1988). The effects of oxygen free radicals on wound healing. *International Journal of Tissue Reactions*, 10(6), 373–379.
- Franklin, J. D., and Lynch, J. B. (1979). Effects of topical applications of epidermal growth factor on wound healing. Experimental study on rabbit ears. *Plastic and Reconstructive Surgery*, 64(6), 766–770.

- Galeano, M., Torre, V., Deodato, B., Campo, G. M., Colonna, M., Sturiale, A., Altavilla, D. (2001). Raxofelast, a hydrophilic vitamin E-like antioxidant, stimulates wound healing in genetically diabetic mice. *Surgery*, 129(4), 467–477.
- Gauglitz, G. G., Korting, H. C., Pavicic, T., Ruzicka, T., and Jeschke, M. G. (2011). Hypertrophic scarring and keloids: Pathomechanisms and current and emerging Treatment Strategies, 7(12), 201-210.
- Geethalakshmi, R., Sakravarthi, C., Kritika, T., Kirubakaran, M. A., and Sarada, D. V. L. (2013). Evaluation of antioxidant and wound healing potentials of *Sphaeranthus Amaranthoides*. *Burn*, 2013.
- Getie, M., Gebre-Mariam, T., Rietz, R., and Neubert, R. H. H. (2002). Evaluation of the release profiles of flavonoids from topical formulations of the crude extract of the leaves of *Dodonaea viscosa* (Sapindaceae). *Die Pharmazie*, 57(5), 320–322.
- Girotti, A. W. (1998). Lipid hydroperoxide generation, turnover, and effector action in biological systems. *Journal of Lipid Research*, 39(8), 1529–1542.
- Grazul-Bilska, A. T., Johnson, M. L., Bilski, J. J., Redmer, D. A., Reynolds, L. P., Abdullah, A., and Abdullah, K. M. (2003). Wound healing: the role of growth factors. *Drugs of Today (Barcelona, Spain: 1998)*, 39(10), 787–800.
- Greenhalgh, D. G., Sprugel, K. H., Murray, M. J., and Ross, R. (1990). PDGF and FGF stimulate wound healing in the genetically diabetic mouse. *The American Journal of Pathology*, 136(6), 1235–1246.
- Grey, J. E., Enoch, S., and Harding, K. G. (2006). Wound assessment. *BMJ: British Medical Journal*, 332(7536), 285–288.
- Grotto, D., Maria, L. S., Valentini, J., Paniz, C., Schmitt, G., Garcia, S. C., Farina, M. (2009). Importance of the lipid peroxidation biomarkers and methodological aspects for malondialdehyde quantification. *Mica Nova*, 32, 169–174.
- Grzesik, W., and Narayanan, A. S. (2002). *Cementum and Periodontal Wound Healing and Regeneration. Critical reviews in oral biology and medicine: an official publication of the American Association of Oral Biologists* (Vol. 13).
- Guo, S., and DiPietro, L. A. (2010). Factors affecting wound healing. *Journal of Dental Research*, 89(3), 219–229.
- Hardwicke, J., Schmaljohann, D., Boyce, D., and Thomas, D. (2008). Epidermal growth factor therapy and wound healing--past, present and future perspectives. *The Surgeon: Journal of the Royal Colleges of Surgeons of Edinburgh and Ireland*, 6(3), 172–177.

- Ibrahim, M. M., Chen, L., Bond, J. E., Medina, M. A., Ren, L., Kokosis, G., ... Levinson, H. (2015). Myofibroblasts contribute to but are not necessary for wound contraction. *Laboratory Investigation*, 95, 1429. Retrieved from <http://dx.doi.org/10.1038/labinvest.2015.116>
- Iftimia, N., Ferguson, D., Mujat, M., H Patel, A., Ziyi Zhang, E., Fox, W., & Rajadhyaksha, M. (2013). Combined reflectance confocal microscopy/optical coherence tomography imaging for skin burn assessment. *Biomedical Optics Express*, 4, 680–695.
- Imanieh, M. H., Khoshneviszadeh, M., Meshksar, A., and Noorafshan, A. (2012). The Healing Effect of Arnebia Euchroma in Second Degree Burn Wounds in Rat as an Animal Model, *Journal of Dermatology*, 14(2), 70–74.
- Insights, C. N. (1997). Nutritional support for connective tissue, *Nutrition and Body*, 5 (1), 312-320.
- Johnson, K. E., and Wilgus, T. A. (2014). Vascular endothelial growth factor and angiogenesis in the regulation of cutaneous wound Repair. *Advances in Wound Care*, 3(10), 647–661.
- Jones, M. L., and Ascp, H. T. (2010). Mastering the trichrome stain, *Basic Skin Histology*, 51(2), 79–84.
- Kachiwal, A. B., Shah, M. G., Shoaib, M., Lochi, G. M., Manan, A., Haq, I., and Muhammad, F. (2014). Histological characterization of wound healing of flank verses midline ovariohysterectomy in different age groups of cats, 5(March), 6–16.
- Kalogeris, T., Baines, C. P., Krenz, M., and Korthuis, R. J. (2012). Cell biology of ischemia/reperfusion injury. *International Review of Cell and Molecular Biology*, 298, 229–317.
- Kaufman, T., Lusthaus, S. N., Sagher, U., and Wexler, M. R. (1990). Deep partial skin thickness burns: a reproducible animal model to study burn wound healing. *Burns : Journal of the International Society for Burn Injuries*, 16(1), 13–16.
- Keen, M. A., and Hassan, I. (2016). Vitamin E in dermatology. *Indian Dermatology Online Journal*, 7(4), 311–315.
- Khanna, H. D. (2009). Evaluation of wound healing activity of extracts of plantain banana in rats, 47(January), 32–40.
- Kim, H., Kong, W. H., Seong, K.-Y., Sung, D. K., Jeong, H., Kim, J. K., Hahn, S. K. (2016). Hyaluronate-epidermal growth factor conjugate for skin wound healing and regeneration. *Biomacromolecules*, 17(11), 3694–3705.
- Kim, J. S., McKinnis, V. S., Adams, K., and White, S. R. (1997). Proliferation and

repair of guinea pig tracheal epithelium after neuropeptide depletion and injury in vivo. *The American Journal of Physiology*, 273.

Kondo, T., and Ishida, Y. (2010). Molecular pathology of wound healing. *Forensic Science International*, 203(1–3), 93–98.

Kozlowski, L. P. (2017). Proteome-pI: proteome isoelectric point database. *Nucleic Acids Research*, 45, 1112–1116.

Krafts, K. P. (2010). Tissue repair: The hidden drama. *Organogenesis*, 6(4), 225–233.

Kurahashi, T., and Fujii, J. (2015). Roles of antioxidative enzymes in wound healing, 57–70.

Kwon, Y., Kim, H., Roh, D., Yoon, S., Baek, R., Kim, J., Lee, J. (2006). Veterinary science topical application of epidermal growth factor accelerates wound healing by myofibroblast proliferation and collagen synthesis in rat, 7, 105–109.

L. Catignani, G., and G. Bieri, J. (1977). Rat liver α -tocopherol binding protein. *Biochimica Et Biophysica Acta-general Subjects*. *BBA-GEN SUBJECTS* (Vol. 497).

Laato, M. (1986). Effect of epidermal growth factor (EGF) on blood flow and albumin extravasation in experimental granulation tissue. *Acta Chirurgica Scandinavica*, 152, 401–405.

Lazarus, G. S., Cooper, D. M., Knighton, D. R., Percoraro, R. E., Rodeheaver, G., and Robson, M. C. (1994). Definitions and guidelines for assessment of wounds and evaluation of healing. *Wound Repair and Regeneration: Official Publication of the Wound Healing Society and the European Tissue Repair Society*, 2(3), 165–170.

LeGrand, E. K., Burke, J. F., Costa, D. E., and Kiorpes, T. C. (1993). Dose responsive effects of PDGF-BB, PDGF-AA, EGF, and bFGF on granulation tissue in a guinea pig partial thickness skin excision model. *Growth Factors (Chur, Switzerland)*, 8(4), 307–314.

Lemmon, M. A., and Schlessinger, J. (2010). Cell signaling by receptor tyrosine kinases. *Cell*, 141(7), 1117–1134.

Lewis, M. C., MacArthur, B. D., Tare, R. S., Oreffo, R. O. C., and Please, C. P. (2016). Extracellular matrix deposition in engineered micromass cartilage pellet cultures: Measurements and modelling. *PLoS ONE*, 11(2), 302.

Li, W., Fan, J., Chen, M., Guan, S., Sawcer, D., Bokoch, G. M., and Woodley, D. T. (2004). Mechanism of human dermal fibroblast migration driven by type I collagen and platelet-derived growth factor, 15(January), 294–309.

- Lim, J. J., Ngah, W. Z. W., Mouly, V., and Abdul Karim, N. (2013). Reversal of myoblast aging by tocotrienol rich fraction posttreatment. *Oxidative Medicine and Cellular Longevity*, 2013, 101.
- Lin, T. S., Abd Latiff, A., Abd Hamid, N. A., Wan Ngah, W. Z. bt, and Mazlan, M. (2012). Evaluation of topical tocopherol cream on cutaneous wound healing in streptozotocin-induced diabetic Rats. *Evidence-Based Complementary and Alternative Medicine*, 2012, 27.
- Liu, S., Zhang, H., and Duan, E. (2013). Epidermal development in mammals : Key regulators , signals from beneath , and stem cells, 10869–10895.
- Lu, S., Xiang, J., Qing, C., Jin, S., Liao, Z., and Shi, J. (2002). Effect of necrotic tissue on progressive injury in deep partial thickness burn wounds. *Chinese Medical Journal*, 115(3), 323–325.
- Maretzky, T., Evers, A., Zhou, W., Swendeman, S. L., Wong, P.-M., Rafii, S., Blobel, C. P. (2011). Migration of growth factor-stimulated epithelial and endothelial cells depends on EGFR transactivation. *Nature Communications*, 2, 229.
- Marie, P. J., Hott, M., and Perheentupa, J. (1990). Effects of epidermal growth factor on bone formation and resorption in vivo. *The American Journal of Physiology*, 258(2 Pt 1), 81-92
- Meredith, P. A., and Elliott, H. L. (1992). Pharmacodynamis and drug action. An additive or synergistic drug interaction : Application of concentration-effect modeling.
- Monaco, J. L., and Lawrence, W. T. (2003). Acute wound healing an overview. *Clinics in Plastic Surgery*, 30(1), 1–12.
- Morimoto, S., Matsumura, Y., Ohno, Y., and Miyawaki, N. (1983). Effects of vitamin E depletion and repletion on renin release from renin granules. *Journal of Pharmacobio-Dynamics*, 6(11), 844–850.
- Muir, E. B. (1941). Gonorrheal ophthalmia and gonorrheal ophthalmia neonatorum. *American Journal of Ophthalmology*, 24(8), 879–894.
- Murphy, P. S., and Evans, G. R. D. (2012). Advances in wound healing : A review of current wound healing products, 2012.
- Musalmah, M., Fairuz, A. H., Gapor, M. T., and Zurinah, W. (2002). Effect of vitamin E on plasma malondialdehyde , antioxidant enzyme levels and the rates of wound closures, 11, 448–451.
- Musalmah, M., Nizrana, M. Y., Fairuz, A. H., NoorAini, A. H., and al, et. (2005). Comparative effects of palm vitamin E and alpha-tocopherol on healing and wound tissue antioxidant enzyme levels in diabetic rats. *Lipids*, 40(6), 575–

- Mutsaers, S. E., Laurent, G. J., Bishop, E., and Mcgrouther, G. U. S. (1997). Mechanisms of tissue repair : from wound healing to fibrosis, *29*(1), 5–17.
- Nanney, L. B. (1990). Epidermal and dermal effects of epidermal growth factor during wound repair. *The Journal of Investigative Dermatology*, *94*(5), 624–629.
- Nesaretnam, K., Meganathan, P., Veerasenan, S. D., and Selvaduray, K. R. (2012). Tocotrienols and breast cancer: the evidence to date. *Genes and Nutrition*, *7*(1), 3–9.
- Noorlander, M. L., Melis, P., Jonker, A., and Noorden, C. J. F. Van. (2002). A quantitative method to determine the orientation of collagen fibers in the dermis. *Journal of Histochemistry and Cytochemistry*, *50*(11), 1469–1474.
- Okumura, K., Kiyohara, Y., Komada, F., Iwakawa, S., Hirai, M., and Fuwa, T. (1990). Improvement in wound healing by epidermal growth factor (EGF) ointment. *Pharmaceutical Research*, *7*(12), 1289–1293.
- Olanlokun, J. O. (2008). Protective influence of vitamin E on the antioxidant defence system in the whole blood and liver of normal and alloxan-induced diabetic rats. *Indian Journal of Clinical Biochemistry*, *23*(1), 62–66.
- Orgill, D. P. M. D., and Ogawa, R. M. D. (2013). Current Methods of Burn Reconstruction. *Plastic and Reconstructive Surgery May 2013*, *131*(5).
- Palmieri, B., Gozzi, G., and Palmieri, G. (1995). Vitamin E added silicone gel sheets for treatment of hypertrophic scars and keloids. *International Journal of Dermatology*, *34*(7), 506–509.
- Panse, N., Sathe, V., Sahasrabudhe, P., and Joshi, N. (2013). Diet, wound healing and plastic surgery. *Indian Journal of Plastic Surgery*, *46*(1), 161–163.
- Pastar, I., Stojadinovic, O., Yin, N. C., Ramirez, H., Nusbaum, A. G., Sawaya, A., Tomic-Canic, M. (2014). Epithelialization in Wound Healing: A Comprehensive Review. *Advances in Wound Care*, *3*(7), 445–464.
- Ph, D., and Attinger, C. E. (2006). Wound Healing : An Overview, 1–32.
- Pierpaoli, E., Cirioni, O., Barucca, A., Orlando, F., Silvestri, C., Giacometti, A., and Provinciali, M. (2011). Vitamin E supplementation in old mice induces antimicrobial activity *The Journal of Antimicrobial Chemotherapy*. 330-351.
- Posada, J., and Cooper, J. A. (1992). Molecular signal integration. *Molecular Biology of the Cell*, *3*(6), 583–592.
- Prausnitz, M. R., and Langer, R. (2008). Transdermal drug delivery. *Nature*

Biotechnology, 26(11), 1261–1268.

Priya, K. S., Gnanamani, A., Radhakrishnan, N., and Babu, M. (2002). Healing potential of *Datura alba* on burn wounds in albino rats, 83, 193–199.

Rashid, S. A. H. N., Halim, A. S., and Muhammad, N. A. (2008). The effect of vitamin E on basic fibroblast growth factor level in human fibroblast cell culture. *The Medical Journal of Malaysia*, 69–70.

Reim, M., Busse, S., Leber, M., and Schulz, C. (1988). Effect of epidermal growth factor in severe experimental alkali burns. *Ophthalmic Research*, 20(5), 327–331.

Rittié, L. (2016). Cellular mechanisms of skin repair in humans and other mammals. *Journal of Cell Communication and Signaling*, 10(2), 103–120.

Rodero, M. P., and Khosrotehrani, K. (2010). Skin wound healing modulation by macrophages, 3(7), 643–653.

Romanovsky, A. A. (2014). Skin temperature: its role in thermoregulation. *Acta Physiologica (Oxford, England)*, 210(3), 498–507.

Rowan, M. P., Cancio, L. C., Elster, E. A., Burmeister, D. M., Rose, L. F., Natesan, S., Chung, K. K. (2015). Burn wound healing and treatment: review and advancements. *Critical Care*, 1–12.

Saito, Y., Nishio, K., Akazawa, Y. O., Yamanaka, K., Miyama, A., Yoshida, Y., Niki, E. (2010). Cytoprotective effects of vitamin E homologues against glutamate-induced cell death. *Free Radical Biology and Medicine*, 49(10), 1542–1549.

Schneider, M. R., and Yarden, Y. (2014). Structure and function of epigen, the last EGFR ligand. *Seminars in Cell and Developmental Biology*, 57–61.

Schultz, G., Rotatori, D. S., and Clark, W. (1991). EGF and TGF- α in wound healing and repair. *Journal of Cellular Biochemistry*, 45(4), 346–352.

Sen, C. K., Khanna, S., and Roy, S. (2007). Tocotrienols in health and disease. *Molecular Aspects of Medicine*, 692–728.

Sen, C. K., Rink, C., and Khanna, S. (2010). Palm oil-derived natural vitamin E α -tocotrienol in brain health and disease. *Journal of the American College of Nutrition*, 314–323.

Senthil Kumar, M., Sripriya, R., Vijaya Raghavan, H., and Kumar Sehgal, P. (2006). Wound healing potential of cassia fistula on infected albino rat model. *The Journal of surgical research*, 201-214.

Shahrim, Z., Selvaduray, K. R., and Nesaretnam, K. (n.d.). Palm tocotrienol-rich

fraction improves wound healing by modulating collagen and decreasing reactive oxygen Species, (6), 43-300.

- Shinde, A., Ganu, J., and Naik, P. (2012). Effect of free radicals and antioxidants on oxidative stress. *Journal of Dental and Allied Sciences*, 1(2), 63–66.
- Shirani, K. Z., Vaughan, G. M., Mason, A. D. J., and Pruitt, B. A. J. (1996). Update on current therapeutic approaches in burns. *Shock*, 5(1). 310-321.
- Shukla, A., Rasik, A. M., Jain, G. K., Shankar, R., Kulshrestha, D. K., and Dhawan, B. N. (1999). In vitro and in vivo wound healing activity of asiaticoside isolated from *Centella asiatica*. *Journal of Ethnopharmacology*, 65(1), 1–11.
- Singh, U., Devaraj, S., and Jialal, I. (2005). Vitamin E, oxidative Stress, and inflammation. *Annual Review of Nutrition*, 25(1), 151–174.
- Speroni, E., Govoni, P., Guizzardi, S., Renzulli, C., and Guerra, M. C. (2002). Anti-inflammatory and cicatrizing activity of *Echinacea pallida* nutt root extract. *Journal of Ethnopharmacology*, 79(2), 265–272.
- Starkey, R. H., Cohen, S., and Orth, D. N. (1975). Epidermal growth factor: identification of a new hormone in human urine. *Science*, 189(4205), 800-802.
- Stephen-haynes, J., Viability, T., Primary, W., Trusts, C., Community, N., Worcs, S., and Rosie, P. C. T. (2001). Evaluation of a honey dressing on wounds within primary care. 111-124.
- Stücker, M., Struk, A., Altmeyer, P., Herde, M., Baumgärtl, H., and Lübbers, D. W. (2002). The cutaneous uptake of atmospheric oxygen contributes significantly to the oxygen supply of human dermis and epidermis, 985–994.
- Sultana, J., Molla, M. R., and Kamal, M. (2004). Histological differences in wound healing in Maxillofacial region in patients with or without risk factors. 24(1), 20-49.
- Tan, Q., Lin, Z., Ma, W., Chen, H., Wang, L., Ning, G., and Zhou, X. (2002). Failure of Ibuprofen to prevent progressive dermal ischemia after burning in guinea pigs. *Burns : Journal of the International Society for Burn Injuries*, 28(5), 443–448.
- Tavares Pereira, D. dos S., Lima-Ribeiro, M. H. M., de Pontes-Filho, N. T., Carneiro-Leão, A. M. dos A., and Correia, M. T. dos S. (2012). Development of animal model for studying deep second-degree thermal burns. *Journal of Biomedicine and Biotechnology*, 1–7.
- Thiele, J. J., Hsieh, S. N., and Ekanayake-Mudiyanselage, S. (2005). Vitamin E: critical review of its current use in cosmetic and clinical dermatology. *Dermatologic Surgery*, 31(7 Pt 2), 805–813.

- Traber, M. G., Rallis, M., Podda, M., Weber, C., Maibach, H. I., and Packer, L. (1998). Penetration and distribution of tocotrienols applied individually onto murine skin. *Lipids*, 33(1), 87–91.
- Tripathi, R. C., Borisuth, N. S., Tripathi, B. J., and Fang, V. S. (1991). Radioimmunoassay of epidermal growth factor in human lenses. *Experimental Eye Research*, 53(6), 759–764.
- Ukong, S., Ampawong, S., and Kengkoom, K. (2008). Collagen measurement and staining pattern of wound healing comparison with fixations and stains, 37–41.
- Ulubayram, K., Cakar, A. N., Korkusuz, P., Ertan, C., and Hasirci, N. (2001). EGF containing gelatin-based wound dressings, 22, 1345–1356.
- Van Crujjsen, H., Giaccone, G., and Hoekman, K. (2005). Epidermal growth factor receptor and angiogenesis: Opportunities for combined anticancer strategies. *International Journal of Cancer*, 117(6), 883–888.
- Van der Slot-Verhoeven, A. J., van Dura, E. A., Attema, J., Blauw, B., DeGroot, J., Huizinga, T. W. J., Bank, R. A. (2005). The type of collagen cross-link determines the reversibility of experimental skin fibrosis. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*, 1740(1), 60–67.
- Vatassery, G. T., Smith, W. E., and Quach, H. T. (1989). Ascorbic acid, glutathione and synthetic antioxidants prevent the oxidation of vitamin E in platelets. *Lipids*, 1043.
- Villacorta, L., Azzi, A., and Zingg, J.M. (2007). Regulatory role of vitamins E and C on extracellular matrix components of the vascular system. *Molecular Aspects of Medicine*, 28(5–6), 507–537
- Weydert, C. J., and Cullen, J. J. (2010). Measurement of superoxide dismutase, catalase and glutathione peroxidase in cultured cells and tissue. *Nature Protocols*, 5(1), 51–66.
- Wieduwilt, M. J., and Moasser, M. M. (2008). The epidermal growth factor receptor family: Biology driving targeted therapeutics. *Cellular and Molecular Life Sciences : CMLS*, 65(10), 1566–1584.
- Winter, G. D. (2006). Some factors affecting skin and wound healing. *Journal of Tissue Viability*, 16(2), 20–23.
- Wu, S.-J., Liu, P.-L., and Ng, L.T. (2008). Tocotrienol-rich fraction of palm oil exhibits anti-inflammatory property. *Molecular Nutrition and Food Research*, 52(8), 921–929.
- Xu, C., Bentinger, M., Savu, O., Moshfegh, A., Sunkari, V., Dallner, G., Tekle, M.

- (2017). Mono-epoxy-tocotrienol enhances wound healing in diabetic mice. *Journal of Diabetes and Its Complications*, 31(1), 4–12.
- Xue, M., and Jackson, C. J. (2015). Extracellular Matrix Reorganization During Wound Healing and Its Impact on Abnormal Scarring. *Advances in Wound Care*, 4(3), 119–136.
- Zaini, A. A., Khaza, H., Ali, R. M., Mutalib, S. A., & Baharuddin, A. A. (2016). Topical Treatment of Tocotrienol- Rich Fraction (TRF) on Deep Partial-Thickness Burn Wounds in Rats. *J Dermatology Clin Res* 4(1):1063.
- Zampieri, N., Zuin, V., Burro, R., Ottolenghi, A., and Camoglio, F. S. (2010). A prospective study in children: Pre- and post-surgery use of vitamin E in surgical incisions. *Journal of Plastic, Reconstructive and Aesthetic Surgery*, 63(9), 1474–1478.
- Zimmermann, T., Yeates, R. A., Laufen, H., Scharpf, F., Leitold, M., and Wildfeuer, A. (1996). Influence of the antibiotics erythromycin and azithromycin on the pharmacokinetics and pharmacodynamics of midazolam. *Arzneimittel-Forschung*, 46(2), 213–217.
- Zingg, J.-M. (2007). Vitamin E: An overview of major research directions. *Molecular Aspects of Medicine*, 28(5–6), 400–422.
- Zohdi, R. M., Abu, Z., Zakaria, B., Yusof, N., & Sample, H. (2012). Honey Hydrogel Dressing to Treat Burn Wound in Rats - A Preliminary Report, 35(1), 67–74.



© COPYRIGHT UPM