



UNIVERSITI PUTRA MALAYSIA

***METABOLITE CHARACTERIZATION OF *Neptunia oleracea* Lour. AND
ITS
CORRELATION WITH ANTIOXIDANT AND ALPHA-GLUCOSIDASE
INHIBITORY ACTIVITIES USING NMR-BASED METABOLOMICS
APPROACH***

LEE SOO YEE

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By

LEE SOO YEE

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

October 2017

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

METABOLITE CHARACTERIZATION OF *Neptunia oleracea* Lour. AND ITS CORRELATION WITH ANTIOXIDANT AND ALPHA-GLUCOSIDASE INHIBITORY ACTIVITIES USING NMR-BASED METABOLOMICS APPROACH

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October 2017

Chairman : Associate Professor Faridah Abas, PhD
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Diabetes mellitus is a metabolic disease characterized by high blood glucose levels. Due to the unpleasant side effects of currently available synthetic drugs, traditional medicinal plants have drawn the attention of researchers who are seeking natural compounds for the control of diabetes. This present study focused on the profiling and characterization of metabolites in local medicinal plants with potential antioxidant and α -glucosidase inhibitory activities. The first part of the study assessed the total phenolic contents (TPC), DPPH free radical scavenging and α -glucosidase inhibitory activities of 5 local medicinal plants. Of these plants, *Neptunia oleracea* exhibited the highest TPC and most potent DPPH free radical scavenging and α -glucosidase inhibitory activities. Subsequently, the second part of the study examined the influence of different drying methods on metabolite variations among *N. oleracea* leaves and stems using an NMR-based metabolomics approach. The results showed that freeze drying was the most suitable drying method for preserving the metabolites in *N. oleracea*; whereas the *N. oleracea* leaves possessed higher TPC and better bioactivities than its stems. Multivariate data analysis (MVDA) revealed that the identified phenolics, including catechin, caffeic and gallic acids and derivatives of apigenin, quercetin, kaempferol and myricetin were responsible for the discrimination and potent bioactivities of freeze-dried *N. oleracea* leaves. Then, the third part of the study focused on an investigation of the relationship between these phenolics and the tested bioactivities. The effects of different extraction methods and ethanol ratios were also investigated. Both partial least square (PLS) and random forests (RF) analysis identified caffeic acid and derivatives of apigenin, quercetin and kaempferol as the most effective DPPH scavengers and α -glucosidase inhibitors of *N. oleracea*. To extract these phenolics, sonication with absolute ethanol was the most suitable extraction condition. Consequently, in the next part of the study, the freeze-dried *N. oleracea* leaf extracts obtained via sonication with absolute ethanol were fractionated using solid phase extraction (SPE) to obtain hexane (HF), chloroform (CF), ethyl acetate (EF) and methanol (MF) fractions. Of

these fractions, EF and MF displayed the most significant bioactivities, and were then subjected to UPLC-MS/MS analysis, which resulted in the identification of 37 metabolites, with mostly phenolics. Through various chromatographic techniques, 5 phenolics including quercetin-3-O- β -D-xylopyranoside (**28**), quercetin-3-O- α -L-arabinopyranoside (**30**), quercetin-3-O- α -L-rhamnoside (**32**), methylgallate (**11**) and rutin (**22**) were isolated from these fractions. Individual testing of the bioactivities of these phenolics showed that methylgallate (**11**) exhibited the most potent DPPH scavenging and α -glucosidase inhibitory activities, with the IC₅₀ values of 17.25 and 50.76 μ M, respectively. In summary, this study suggested that *N. oleracea* is a prominent source of phenolics which can be potential antioxidant and α -glucosidase inhibitors for the management of diabetes. This study is the first report on the antidiabetic potentiality of *N. oleracea*, and the first application of metabolomics approach for extensive study of the phytoconstituents and their correlation with antioxidant and α -glucosidase inhibitory activities of this plant. The results of this study lay the foundation for future research regarding the antidiabetic effect of *N. oleracea*.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENCIRIAN METABOLIT *Neptunia oleracea* Lour. DENGAN KORELASI
KEPADA AKTIVITI ANTIOKSIDAN DAN PERENCATAN ALFA-
GLUKOSIDASE MENGGUNAKAN PENDEKATAN METABOLOMIK
BERASASKAN NMR**

Oleh

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Oktober 2017

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Kencing manis merupakan satu penyakit metabolik yang dicirikan oleh kandungan gula yang tinggi dalam darah. Disebabkan oleh kesan sampingan yang tidak menyenangkan daripada ubat sintetik yang sedia ada, ubatan tradisional berasaskan tumbuhan telah menarik perhatian penyelidik dalam pencarian sebatian semulajadi untuk mengawal penyakit kencing manis. Kajian ini memberi tumpuan kepada profil dan pencirian metabolit dalam tumbuhan ubatan tempatan yang berpotensi untuk aktiviti antioksidan dan perencatan α -glukosidase. Bahagian pertama kajian ini adalah menilai jumlah kandungan fenolik (TPC), aktiviti pemerangkap radikal bebas DPPH dan perencatan α -glukosidase bagi 5 tumbuhan ubatan tempatan. Dalam kalangan tumbuhan ini, *Neptunia oleracea* mempamerkan TPC yang paling tinggi dan aktiviti pemerangkapan radikal bebas DPPH dan perencatan α -glukosidase yang paling kuat. Seterusnya, bahagian kedua kajian ini adalah mengkaji kesan kaedah pengeringan yang berbeza terhadap variasi metabolit antara daun dan batang *N. oleracea* dengan menggunakan metabolomik berasaskan NMR. Keputusan menunjukkan pengeringan sejuk beku adalah kaedah pengeringan yang paling sesuai untuk mengekalkan metabolit dalam *N. oleracea*; manakala daun *N. oleracea* memiliki TPC yang lebih tinggi dan bioaktiviti lebih baik daripada batang. Analisis data multivariat (MVDA) membuktikan bahawa fenolik yang dikenalpasti, antaranya katekin, asid kafeik dan galik, dan terbitan apigenin, kuersetin, kaempferol dan miricetin bertanggungjawab terhadap diskriminasi dan bioaktiviti yang tinggi pada daun *N. oleracea* yang dikering dengan pengeringan sejuk beku. Selanjutnya, bahagian ketiga kajian ini adalah memberi tumpuan kepada kajian hubungan antara fenolik dan bioaktiviti yang diuji. Pengaruh kaedah pengekstrakan dan nisbah etanol juga dikaji. Analisa separa persegi (PLS) dan hutan rawak (RF) telah mengenalpasti asid kafeik dan terbitan apigenin, kuersetin dan kaempferol sebagai pemerangkap DPPH dan perencat α -glukosidase yang paling mempengaruhi dalam *N. oleracea*. Bagi pengekstrakan fenolik ini, sonication dengan etanol mutlak merupakan kaedah

pengekstrakan yang paling sesuai. Kesannya, ekstrak daun *N. oleracea* yang dihasilkan dari pengeringan sejuk beku dan sonication dengan etanol mutlak telah difraksionasi menggunakan pengekstrakan fasa pepejal (SPE) untuk mendapatkan fraksi heksans (HF), klorofom (CF), etil asetat (EF) dan metanol (MF). Di antara kesemua fraksi, EF dan MF mempamerkan bioaktiviti yang paling ketara, dan kemudian dijadikan subjek untuk analisis UPLC-MS/MS, yang membawa kepada pengenalpastian 37 metabolit, dengan kebanyakannya fenolik. Melalui pelbagai teknik kromatografi, 5 fenolik termasuk kuersetin-3-O- β -D-xilopiranosida (**28**), kuersetin-3-O- α -L-arabinopiranosida (**30**), kuersetin-3-O- α -L-rhamnosida (**32**), metilgallate (**11**) dan rutin (**22**) telah diasingkan dari kedua-dua fraksi ini. Kajian individu pada fenolik ini dalam bioaktiviti menunjukkan bahawa metilgallate mempamerkan aktiviti pemerangkapan radikal bebas DPPH dan perencatan α -glikosidase yang paling kuat dengan nilai IC₅₀ masing-masing sebanyak 17.25 dan 50.76 μ M. Kesimpulannya, kajian ini mencadangkan *N. oleracea* adalah sumber utama fenolik yang boleh berpotensi sebagai antioksidan dan perencat α -glikosidase bagi menangani masalah kencing manis. Kajian ini merupakan laporan pertama mengenai potensi antidiabetik *N. oleracea*, serta aplikasi pertama pendekatan metabolomik untuk analisis secara meluas kandungan fitokimia *N. oleracea*, dan korelasi dengan aktiviti pemerangkapan radikal bebas DPPH dan perencatan α -glikosidase tumbuhan ini. Hasil kajian ini meletakkan asas untuk penyelidikan masa depan mengenai aktiviti antidiabetik *N. oleracea*.

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I certify that a Thesis Examination Committee has met on 25 October 2017 to conduct the final examination of Lee Soo Yee on her thesis entitled "Metabolite Characterization of *Neptunia oleracea* Lour. and its Correlation with Antioxidant and Alpha-Glucosidase Inhibitory Activities using NMR-Based Metabolomics Approach" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

ABTS	2,2'-azinobis-(3-ethylbenzothiazoline-6-sulfonic acid)
AD	Air drying
ANOVA	Analysis of variance
APCI	Atmospheric pressure chemical ionization
API	Atmospheric ionization
BHA	Butylated hydroxyanisole
BHT	Butylated hydroxytoluene
CF	Chloroform fraction
CID	Collision-induced dissociation
d	Doublet
DAD	Diode array detector
dd	Doublet of doublet
DPPH	2,2-diphenyl-1-picrylhydrazyl free radicals
EF	Ethyl acetate fraction
ESI	Electrospray ionization
FD	Freeze drying
FRAP	Ferric reducing antioxidant power
GAE	Gallic acid equivalent
gCOSY	Gradient homonuclear correlation spectroscopy
gHMBCAD	Adiabatic gradient heteronuclear multiple bond coherence
gHSQCAD	Adiabatic gradient heteronuclear single quantum correlation
HF	Hexane fraction

HPLC	High performance liquid chromatography
HRAM	High resolution accurate mass
LC	Liquid chromatography
LC/MS	Liquid chromatography/mass spectrometry
m	Multiplet
m/z	Mass-to-charge ratio
MF	Methanol fraction
MS	Mass spectrometry
MVDA	Multivariate data analysis
NMR	Nuclear magnetic resonance
OD	Oven drying
ORAC	Oxygen radical absorption capacity
PCA	Principle component analysis
PLS	Partial least squares
PLS-DA	Partial least squares–discriminant analysis
PNP	<i>p</i> -nitrophenol
PNPG	<i>p</i> -nitrophenyl- α -D-glucopyranose
ppm	Part per million
QTOF	quadrupole-time of flight
RF	Random forests
RNS	Reactive nitrogen species
s	Singlet
SPE	Solid phase extraction
t	Triplet

TIC	Total ion chromatogram
TLC	Thin layer chromatography
TMS	Tetramethyl silane
TOF	Time of flight
TPC	Total phenolic content
TPU	Agricultural Park
UHPLC-MS/MS	Ultra-high performance liquid chromatography tandem mass spectrometry analysis
UPM	Universiti Putra Malaysia
UV	Ultraviolet
WHO	World Health Organization
δ	Chemical shift in ppm
1D	One-dimensional
^1H	Proton
2D	Two-dimensional
^{13}C	Carbon-13

CHAPTER 1

INTRODUCTION

1.1 Background

Diabetes mellitus is a metabolic complicated disease in which the patient experiences a high level of blood glucose. Over the past few decades, the occurrence of diabetes mellitus have increased drastically. Globally, the number of diabetic cases in adult population has increased from 108 million to 422 million from 1980 to 2014 (World Health Organization, 2016). Among the diabetic patient, majority are suffering from type 2 diabetes. In Malaysia, 20.8% of its adults are affected by type 2 diabetes (Hussein et al., 2015). Diabetes ultimately results in complications, such as cardiovascular disease, retinopathy, nephropathy and impaired wound healing. As a result, it is claimed to be a major factor contributing to blindness, kidney failure, heart attacks, stroke and amputation of lower limb.

Currently, there are many synthetic drugs available for the treatment of diabetes, such as acarbose, voglibose and miglitol. However, they meet few of the needs of diabetic patients and cause many undesirable side effects (Hung et al., 2012). On the other hand, accumulated evidence suggests that antioxidants and α -glucosidase inhibitors from natural sources may exert antidiabetic effects (Hung et al., 2012; Yao et al., 2010; Zhang et al., 2013). Hence, these natural compounds have drawn the attention of researchers worldwide. Phenolic compounds are one of the most important groups of secondary natural compounds. They have been reported to exhibit various biological effects, including antioxidant and carbohydrate hydrolyzing enzyme inhibition activities (Pukalskienė et al., 2015; Yao et al., 2010). Traditional medicinal plant is one of the most studied source of these compounds.

The tropical rainforest in Malaysia harbors precious medicinal plants. These plants has been traditionally used by the folks to enhance their health and as remedy to treat several ailments, including diabetes. The examples of some well-known Malaysian medicinal plant used traditionally for controlling diabetes are *Andrographis paniculata* (hempedu bumi), *Cosmos caudatus* (ulam raja), *Orthosiphon stamineus* (misai kucing), *Centella asiatica* (pegaga), *Ficus deltoidea* (mas cotek), *Piper sarmentosum* (sireh) and *Phyllanthus niruri* (dukung anak). Many studies had been performed to scientifically prove the antidiabetic efficacy of these plants and to identify the active principles (Mustaffa et al., 2011; Sekar et al., 2014). This shows that the traditional medicinal plants in Malaysia offers a great opportunity for seeking the antidiabetic components.

Neptunia oleracea is a floating aquatic plant widely distributed over tropical regions of the world. It is consumed as a vegetable, especially in Southeast Asia, and as a traditional medicinal plant. The leaf is traditionally used as antipyretic and antidote (Daduang et al.,

2011). The roots are used to treat necrosis of the nose and hard palate and as a remedy to cure fever whereas the juice from stem is squeezed into the ear to cure earache (Deb et al., 2013). Several biological activities had been reported for this plant, including antioxidant (Thalang et al., 2001), antiulcer (Bhoomannavar et al., 2011a), analgesic and anti-inflammatory (Paul et al., 2012), α -glucosidase inhibitory (Wongsa et al., 2012) as well as hepatoprotective (Bhoomannavar et al., 2011b) activities. Though the presence of flavonoids, carbohydrates, tannins and triterpenes had been reported in this plant (Vijayashree et al., 2006), the extensive metabolite profiling and correlation between metabolites with the claimed bioactivities have not yet been reported.

Metabolite composition can have significant variation at the different parts of the plant and may be altered by the post-harvest processing, such as drying and extraction. Generally, drying is applied to remove moisture from plants to retard the enzymatic degradation and hence extend their shelf life for storage. Air-, oven- and freeze drying are some of the common drying methods applied in medicinal plants research. There are various study reported regarding the effect of different drying methods on the plant metabolites (Borchani et al., 2011; Chan et al., 2009). Furthermore, extraction is a crucial step in the assessment of metabolites present in plant materials as extraction helps to release the valuable metabolites from the complex plant matrix. There is no single extraction procedure to simultaneously extract all classes of metabolites with great efficiency due to the complexity of the metabolites in their physical as well as chemical properties. Hence, depending on the plant samples, the effect of different drying as well as the extraction conditions should be evaluated and optimized.

Metabolomics is a competent approach for evaluating the metabolite variation in organism under certain conditions. The variation in the metabolite composition arise from different processing parameters can hence be measured by metabolomics approach, using advanced analytical methods coupled with multivariate statistical analysis. Nuclear magnetic resonance (NMR) spectroscopy is one of the most utilized analytical tools for applications in plant metabolomics. It provides information about the metabolites in plant. The metabolites identification is made possible based on the chemical shift and coupling constants provided by NMR spectroscopy (Kim et al., 2006). Multivariate data analysis (MVDA) such as principle component analysis (PCA) and partial least squares (PLS) are useful to manage the huge dataset generated from NMR analyses and to highlight the important phytochemical markers contributing to the studied bioactivities of the samples (Kim et al., 2010b). Recently, a machine learning method, random forests (RF) has drawn increased popularity in the application as MVDA tool in omics research as well.

Often at times, the metabolite profile of crude extracts are complicated due to the numerous amount of metabolites that could be present. Hence, fractionation of the crude extract by different polarity may facilitate the analysis and make identification of the important metabolites more effective (Grkovic et al., 2014). Fractionation can also enrich the important metabolites in certain fractions and these fractions can be subjected to different chromatographic techniques to isolate the metabolites. The isolated metabolites can be further characterized based on their physical properties and spectral

data obtained from advance spectroscopic method such as mass spectrometry (MS) and NMR to confirm their structures and identities. In addition, the isolated metabolites can be individually tested for the studied bioactivities to validate their contributions towards the bioactivities.

1.2 Problem statements

Negative effects arise from available synthetic drugs such as acarbose, voglibose and miglitol has drawn the attention of researchers to seek for antioxidants and α -glucosidase inhibitors from natural sources. The medicinal plants in Malaysia is a potential natural resources for this purpose. However, there are still a large number of these plants which have none or limited study regarding their antidiabetic potency. Little is known regarding the metabolite profile of *N. oleracea* and its relationship between the metabolites and antidiabetic-related bioactivities. There is also no information has been reported on the effects of different post-harvest processing on the metabolites in this plant. Besides, the metabolite profile of crude extract is often complicated owing to the large amount of metabolites present. This makes the analysis and identification of the important metabolites less effective.

1.3 Objectives

To address the aforementioned problems, the present study was designed and conducted, with the goal to seek for the phytochemical components to be used for the management of diabetes and its complications. To achieve these goals, several specific objectives were proposed. First part of the work aimed to screen the antioxidant and α -glucosidase inhibitory activities of 5 local traditional medicinal plants (section 4.1). The most active plant was chosen to evaluate the effect of different drying methods on the antioxidant and α -glucosidase inhibitory activities using ^1H NMR-based metabolomics approach (section 4.2). Following this, two regression models applied in MVDA were compared for evaluating relationship between phenolics and the bioactivities studied (section 4.3). The effect of different extraction conditions on the phenolics were also assessed (section 4.3). Subsequently, the plant extract obtained from the most suitable drying and extraction conditions were fractionated and profiling of the chemical constituents in the bioactive fractions were performed (section 4.4). Lastly, attempt was made to isolate the phenolics from the bioactive fractions, followed by individually assessment for their bioactivities and quantitative determination (section 4.5).

The five specific objectives are listed below:

1. To screen the leaves and stems of 5 selected traditional medicinal plants (*Mitragyna speciosa*, *Clinacanthus nutans*, *Strobilanthes crispus*, *Neptunia oleracea* and *Mentha asiatica*) for antioxidant and α -glucosidase inhibitory activities.

(From screening results, *N. oleracea* was identified as having the highest activity and was thus selected for further study.)

2. To evaluate the effect of drying methods on the phytochemical constituents of different parts of *N. oleracea* and the correlation with antioxidant and α -glucosidase inhibitory activities using ^1H NMR-based metabolomics.
3. To compare partial least squares (PLS) and random forests (RF) for evaluating relationship between phenolics and bioactivities of *N. oleracea* and to evaluate the effects of different extraction methods and ethanol ratios on the phenolics in *N. oleracea*.
4. To differentiate the fractions of *N. oleracea* leaves extract and correlate with antioxidant and α -glucosidase inhibitory activities using ^1H NMR-based metabolomics, as well as to profile the bioactive fractions using UPLC–MS/MS.
5. To isolate and characterize the phenolics in the bioactive fractions of *N. oleracea* and to quantitate their contents in their respective fractions using HPLC.

BIBLIOGRAPHY

- Abas, F., Lajis, N. H., Israf, D. A., Khozirah, S., & Umi Kalsom, Y. (2006). Antioxidant and nitric oxide inhibition activities of selected Malay traditional vegetables. *Food Chemistry*, 95(4), 566–573.
- Abdel-Farid, I. B., Hye, K. K., Young, H. C., & Verpoorte, R. (2007). Metabolic characterization of *Brassica rapa* leaves by NMR spectroscopy. *Journal of Agricultural and Food Chemistry*, 55(19), 7936–7943.
- Abdul-Hamid, N. A., Abas, F., Ismail, I. S., Shaari, K., & Lajis, N. H. (2015). Influence of different drying treatments and extraction solvents on the metabolite profile and nitric oxide inhibitory activity of Ajwa Dates. *Journal of Food Science*, 80(11), H2603–H2611.
- Abdullah, S. A., & Nakagoshi, N. (2008). Changes in agricultural landscape pattern and its spatial relationship with forestland in the State of Selangor, peninsular Malaysia. *Landscape and Urban Planning*, 87(2), 147–155.
- Abu Bakar, M. F., Mohamed, M., Rahmat, A., & Fry, J. (2009). Phytochemicals and antioxidant activity of different parts of bambangan (*Mangifera pajang*) and tarap (*Artocarpus odoratissimus*). *Food Chemistry*, 113(2), 479–483.
- Abu Bakar, M. H., Sarmidi, M. R., Cheng, K.K., Ali Khan, A., Suan, C. L., Zaman Huri, H., & Yaakob, H. (2015). Metabolomics - the complementary field in systems biology: a review on obesity and type 2 diabetes. *Molecular BioSystems*, 11(7), 1742–1774.
- Acharjee, A., Kloosterman, B., de Vos, R. C. H., Werij, J. S., Bachem, C. W. B., Visser, R. G. F., & Maliepaard, C. (2011). Data integration and network reconstruction with ~ omics data using Random Forest regression in potato. *Analytica Chimica Acta*, 705(1–2), 56–63.
- Ado, M. A., Abas, F., Ismail, I. S., Ghazali, H. M., & Shaari, K. (2015). Chemical profile and antiacetylcholinesterase, antityrosinase, antioxidant and α -glucosidase inhibitory activity of *Cynometra cauliflora* L. leaves. *Journal of the Science of Food and Agriculture*, 95(3), 635–642.
- Agnolet, S., Jaroszewski, J. W., Verpoorte, R., & Staerk, D. (2010). ¹H NMR-based metabolomics combined with HPLC-PDA-MSSPE- NMR for investigation of standardized *Ginkgo biloba* preparations. *Metabolomics*, 6(2), 292–302.
- Al-Bayati, F. A., & Al-Mola, H. F. (2008). Antibacterial and antifungal activities of different parts of *Tribulus terrestris* L. growing in Iraq. *Journal of Zhejiang University-Science B*, 9(2), 154–159.
- Alam, M. N., Bristi, N. J., & Rafiquzzaman, M. (2013). Review on in vivo and in vitro methods evaluation of antioxidant activity. *Saudi Pharmaceutical Journal*. 21(2), 143-152.

- Amari, N. O., Bouzouina, M., Berkani, A., & Lotmani, B. (2014). Phytochemical screening and antioxidant capacity of the aerial parts of *Thymelaea hirsuta* L. *Asian Pacific Journal of Tropical Disease*, 4(2), 104–109.
- Ardrey, R. E. (2003). *Liquid chromatography-mass spectrometry: an introduction*. John Wiley & Sons.
- Arslan, D., & Musa Özcan, M. (2008). Evaluation of drying methods with respect to drying kinetics, mineral content and colour characteristics of rosemary leaves. *Energy Conversion and Management*, 49(5), 1258–1264.
- Aslam, M. S., Ahmad, M. S., & Mamat, A. S. (2015). A review on phytochemical constituents and pharmacological activities of *Clinacanthus nutans*. *International Journal of Pharmacy and Pharmaceutical Sciences*, 7(2), 30–33.
- Ayouni, K., Berboucha-rahmani, M., Kim, H. K., Atmani, D., Verpoorte, R., & Choi, Y. H. (2016). Metabolomic tool to identify antioxidant compounds of *Fraxinus angustifolia* leaf and stem bark extracts. *Industrial Crops & Products*, 88, 1–13.
- Barchan, A., Bakkali, M., Arakrak, A., Pagán, R., & Laglaoui, A. (2014). The effects of solvents polarity on the phenolic contents and antioxidant activity of three *Mentha* species extracts. *International Journal of Current Microbiology and Applied Sciences*, 3(11), 399–412.
- Barros, E., Lezar, S., Anttonen, M. J., Van Dijk, J. P., Röhlig, R. M., Kok, E. J., & Engel, K. H. (2010). Comparison of two GM maize varieties with a near-isogenic non-GM variety using transcriptomics, proteomics and metabolomics. *Plant Biotechnology Journal*, 8(4), 436–451.
- Baser, K. H. C., Nuriddinov, K. R., Nigmatullaev, A. M., & Aripov, K. N. (1997). Essential Oil of *Mentha asiatica* Boriss. from Uzbekistan. *Journal of Essential Oil Research*, 9(4), 453–454.
- Beckmann, M., Enot, D. P., Overy, D. P., & Draper, J. (2007). Representation, comparison, and interpretation of metabolome fingerprint data for total composition analysis and quality trait investigation in potato cultivars. *Journal of Agricultural and Food Chemistry*, 55(9), 3444–3451.
- Benavides, A., Montoro, P., Bassarello, C., Piacente, S., & Pizza, C. (2006). Catechin derivatives in *Jatropha macrantha* stems: Characterisation and LC/ESI/MS/MS quali-quantitative analysis. *Journal of Pharmaceutical and Biomedical Analysis*, 40(3), 639–647.
- Bendary, E., Francis, R. R., Ali, H. M. G., Sarwat, M. I., & El Hady, S. (2013). Antioxidant and structure–activity relationships (SARs) of some phenolic and anilines compounds. *Annals of Agricultural Sciences*, 58(2), 173–181.
- Bennett, B. Y. R. N., & Wallsgrove, R. M. (1994). Secondary metabolites in plant defence mechanisms. *New Phytologist*, (127), 617–633.
- Bhoomannavar, V. S., Patil, V. P., Hugar, S., & Nanjappaiah, H. M. (2011a). Anti-Ulcer

activity of *Neptunia oleracea* Lour. *Pharmacologyonline*, 3, 1015–1020.

- Bhoomannavar, V. S., Shivakumar, S. I., Halliheri, C. S., & Hetapakki, B. C. (2011b). Hepatoprotective activity of leaves of *Neptunia oleracea* Lour in carbon tetrachloride induced rats. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 2(2), 309–314.
- Bihmidine, S., Baker, R. F., Hoffner, C., & Braun, D. M. (2015). Sucrose accumulation in sweet sorghum stems occurs by apoplasmic phloem unloading and does not involve differential Sucrose transporter expression. *BMC Plant Biology*, 15(1), 186.
- Boath, A. S., Stewart, D., & McDougall, G. J. (2012). Berry components inhibit α -glucosidase in vitro: Synergies between acarbose and polyphenols from black currant and rowanberry. *Food Chemistry*, 135(3), 929–936.
- Borchani, C., Besbes, S., Masmoudi, M., Blecker, C., Paquot, M., & Attia, H. (2011). Effect of drying methods on physico-chemical and antioxidant properties of date fibre concentrates. *Food Chemistry*, 125(4), 1194–1201.
- Brito, A., Ramirez, J. E., Areche, C., Sepúlveda, B., & Simirgiotis, M. J. (2014). HPLC-UV-MS profiles of phenolic compounds and antioxidant activity of fruits from three citrus species consumed in Northern Chile. *Molecules*, 19(11), 17400–17421.
- Ceriello, A., & Motz, E. (2004). Is oxidative stress the pathogenic mechanism underlying insulin resistance, diabetes, and cardiovascular disease? The common soil hypothesis revisited. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 24(5), 816–823.
- Chan, E. W. C., Lim, Y. Y., Wong, S. K., Lim, K. K., Tan, S. P., Lianto, F. S., & Yong, M. Y. (2009). Effects of different drying methods on the antioxidant properties of leaves and tea of ginger species. *Food Chemistry*, 113(1), 166–172.
- Chen, T., Cao, Y., Zhang, Y., Liu, J., Bao, Y., Wang, C., Jia, W., Zhao, A. (2013). Random forest in clinical metabolomics for phenotypic discrimination and biomarker selection. *Evidence-Based Complementary and Alternative Medicine*, 2013.
- Chew, K. K., Ng, S. Y., Thoo, Y. Y., Khoo, M. Z., Wan Aida, W. M., & Ho, C. W. (2011). Effect of ethanol concentration, extraction time and extraction temperature on the recovery of phenolic compounds and antioxidant capacity of *Centella asiatica* extracts. *International Food Research Journal*, 18, 571–578.
- Choi, H. K., Choi, Y. H., Verberne, M., Lefeber, A. W. M., Erkelens, C., & Verpoorte, R. (2004). Metabolic fingerprinting of wild type and transgenic tobacco plants by ¹H NMR and multivariate analysis technique. *Phytochemistry*, 65(7), 857–864.
- Choi, J. G., Mun, S. H., Chahar, H. S., Bharaj, P., Kang, O. H., Kim, S. G. Shin, D. W., & Kwon D. Y. (2014). Methyl gallate from *Galla rhois* successfully controls clinical isolates of *Salmonella* infection in both *in vitro* and *in vivo* systems. *PloS*

one, 9(7), e102697.

- Choi, Y. H., Kim, H. K., Linthorst, H. J. M., Hollander, J. G., Lefeber, A. W. M., Erkelens, C., Nuzillard, J., & Verpoorte, R. (2006). NMR metabolomics to revisit the tobacco mosaic virus infection in *Nicotiana tabacum* leaves. *Journal of Natural Products*, 69(5), 742–748.
- Colegate, S. M., & Molyneux, R. J. (2007). *Bioactive natural products: detection, isolation, and structural determination* (Second). CRC press.
- Contini, M., Baccelloni, S., Massantini, R., & Anelli, G. (2008). Extraction of natural antioxidants from hazelnut (*Corylus avellana* L.) shell and skin wastes by long maceration at room temperature. *Food Chemistry*, 110(3), 659–669.
- Cooper, W. T. (2006). Normal-phase liquid chromatography. In *Encyclopedia of Analytical Chemistry*. John Wiley & Sons, Ltd.
- Daduang, J., Vichitphan, S., Daduang, S., Hongprabhas, P., & Boonsiri, P. (2011). High phenolics and antioxidants of some tropical vegetables related to antibacterial and anticancer activities. *African Journal of Pharmacy and Pharmacology*, 5(5), 608–615.
- Davison, G. W., George, L., Jackson, S. K., Young, I. S., Davies, B., Bailey, D. M., Peters, J. R., & Ashton, T. (2002). Exercise, free radicals, and lipid peroxidation in type 1 diabetes mellitus. *Free Radical Biology and Medicine*, 33(11), 1543–1551.
- De Hoffmann, E., & Stroobant, V. (2007). *Mass spectrometry: principles and applications* (Third). John Wiley & Sons.
- Deb, P. K., Das, S., Bhaumik, K. N., Ghosh, R., Ghosh, T. K., & Bhakta, T. (2013). Pharmacognostic & Preliminary Phytochemical Investigations of *Neptunia prostrata* L. *Journal of Pharmacognosy and Phytochemistry*, 2(3), 5–11.
- Deutschländer, M. S., Venter, M. Van De, Roux, S., Louw, J., & Lall, N. (2009). Hypoglycaemic activity of four plant extracts traditionally used in South Africa for diabetes. *Journal of Ethnopharmacology*, 124(3), 619–624.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S., & Ju, Y. H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatica*. *Journal of Food and Drug Analysis*, 22(3), 296–302.
- Elsadig Karar, M. G., Quiet, L., Rezk, A., Jaiswal, R., Rehders, M., Ullrich, M. S., Brix, K., & Kuhnert, N. (2016). Phenolic profile and *in vitro* assessment of cytotoxicity and antibacterial activity of *Ziziphus spina-christi* leaf extracts. *Medicinal Chemistry*, 6(3), 143–156.
- Eriksson, L., Johansson, E., Kettaneh-Wold, N., Trygg, J., Wikström, C., & Wold, S. (2006). *Multi- and megavariable data analysis : Part I: Basic principles and applications*. Umetrics Inc.

- Es-Safi, N. E., & Gómez-Cordovés, C. (2014). Characterization of flavonoid glycosides from fenugreek (*Trigonella foenum-graecum*) crude seeds by HPLC-DAD-ESI/MS analysis. *International Journal of Molecular Sciences*, 15(11), 20668–20685.
- Eyles, A., Jones, W., Riedl, K., Cipollini, D., Schwartz, S., Chan, K., Herms, D.A., & Bonello, P. (2007). Comparative phloem chemistry of Manchurian (*Fraxinus mandshurica*) and two North American Ash Species (*Fraxinus americana* and *Fraxinus pennsylvanica*). *Journal of Chemical Ecology*, 33(7), 1430–1448.
- Fan, G., Luo, W.Z., Luo, S.H., Li, Y., Meng, X.L., Zhou, X.D., & Zhang, Y. (2014). Metabolic discrimination of *Swertia mussotii* and *Swertia chirayita* known as “Zangyinchen” in traditional Tibetan medicine by ¹H NMR-based metabolomics. *Journal of Pharmaceutical and Biomedical Analysis*, 98, 364–370.
- Fan, L., Li, J., Deng, K., & Ai, L. (2012). Effects of drying methods on the antioxidant activities of polysaccharides extracted from *Ganoderma lucidum*. *Carbohydrate Polymers*, 87(2), 1849–1854.
- Fernando, I. D. N. S., Abeysinghe, D. C., & Dharmadasa, R. M. (2013). Determination of phenolic contents and antioxidant capacity of different parts of *Withania somnifera* (L.) Dunal. from three different growth stages. *Industrial Crops & Products*, 50, 537–539.
- Fossen, T., Larsen, Å., Kiremire, B. T., & Andersen, Ø. M. (1999). Flavonoids from blue flowers of *Nymphaea caerulea*. *Phytochemistry*, 51(8), 1133–1137.
- Francescato, L. N., Debenedetti, S. L., Schwanz, T. G., Bassani, V. L., & Henriques, A. T. (2013). Identification of phenolic compounds in *Equisetum giganteum* by LC-ESI-MS/MS and a new approach to total flavonoid quantification. *Talanta*, 105, 192–203.
- Furusawa, M., Tanaka, T., Ito, T., Nakaya, K., Iliya, I., Ohyama, M., Linuma, M., Murata, H., Inatomi, Y., Inada, A., Nakanishi, T., Matsushita, S., Kubota, Y., Sawa R., & Takahashi, Y. (2005). Flavonol glycosides in leaves of two *Diospyros* species. *Chemical & Pharmaceutical Bulletin*, 53(5), 591–3.
- Georgiev, M. I., Ali, K., Alipieva, K., Verpoorte, R., & Choi, Y. H. (2011). Metabolic differentiations and classification of *Verbascum* species by NMR-based metabolomics. *Phytochemistry*, 72(16), 2045–2051.
- Gogna, N., Hamid, N., & Dorai, K. (2015). Metabolomic profiling of the phytomedicinal constituents of *Carica papaya* L. leaves and seeds by ¹H NMR spectroscopy and multivariate statistical analysis. *Journal of Pharmaceutical and Biomedical Analysis*, 115, 74–85.
- González-Domínguez, R., García-Barrera, T., & Gómez-Ariza, J. L. (2015). Application of a novel metabolomic approach based on atmospheric pressure photoionization mass spectrometry using flow injection analysis for the study of Alzheimer’s disease. *Talanta*, 131, 480–489.

- Grata, E., Boccard, J., Guillarme, D., Glauser, G., Carrupt, P. A., Farmer, E. E., Wolfender, J.L., & Rudaz, S. (2008). UPLC-TOF-MS for plant metabolomics: A sequential approach for wound marker analysis in *Arabidopsis thaliana*. *Journal of Chromatography B*, 871(2), 261–270.
- Grkovic, T., Pouwer, R. H., Vial, M., Gambini, L., No, A., Hooper, J. N. A., Wood, S.A., Mellick, G.D., & Quinn, R. J. (2014). NMR fingerprints of the drug-like natural-product space identify irochlorotazine A : A chemical probe to study Parkinson's disease. *Angewandte Chemie International Edition*, 53(24), 6070–6074.
- Gromski, P. S., Muhamadali, H., Ellis, D. I., Xu, Y., Correa, E., Turner, M. L., & Goodacre, R. (2015). A tutorial review: Metabolomics and partial least squares-discriminant analysis - a marriage of convenience or a shotgun wedding. *Analytica Chimica Acta*, 879, 10–23.
- Guehi, T. S., Zahouli, I. B., Ban-Koffi, L., Fae, M. A., & Nemlin, J. G. (2010). Performance of different drying methods and their effects on the chemical quality attributes of raw cocoa material. *International Journal of Food Science & Technology*, 45(8), 1564–1571.
- Hartmann, T. (1996). Diversity and variability of plant secondary metabolism: A mechanistic view. In E. Städler, M. Rowell-Rahier, & R. Bauer (Eds.), *Proceedings of the 9th International Symposium on Insect-Plant Relationships* (pp. 177–188). Dordrecht: Springer Netherlands.
- He, D., Gu, D., Huang, Y., Ayupbek, A., Yang, Y., Akber, H., & Ito, Y. (2009). Separation and purification of phenolic acids and myricetin from black currant by high speed countercurrent chromatography. *Journal of Liquid Chromatography & Related Technologies*, 32(20), 3077–3088.
- Hofmann, T., Nebhaj, E., & Albert, L. (2016). Antioxidant properties and detailed polyphenol profiling of European hornbeam (*Carpinus betulus* L.) leaves by multiple antioxidant capacity assays and high-performance liquid chromatography/multistage electrospray mass spectrometry. *Industrial Crops and Products*, 87, 340–349.
- Hossain, M. B., Barry-Ryan, C., Martin-Diana, A. B., & Brunton, N. P. (2010a). Effect of drying method on the antioxidant capacity of six Lamiaceae herbs. *Food Chemistry*, 123(1), 85–91.
- Hossain, M. B., Rai, D. K., Brunton, N. P., Martin-Diana, A. B., & Barry-Ryan, A. C. (2010b). Characterization of phenolic composition in Lamiaceae spices by LC-ESI-MS/MS. *Journal of Agricultural and Food Chemistry*, 58(19), 10576–10581.
- Hromadkova, Z., Ebringerova, A., & Valachovič, P. (2002). Ultrasound-assisted extraction of water-soluble polysaccharides from the roots of valerian (*Valeriana officinalis* L.). *Ultrasonics Sonochemistry*, 9(1), 37–44.
- Hu, X., You, J., Bao, C., Zhang, H., Meng, X., Xiao, T., Zhang, K., Wang, Y., Wang, H., Zhang, H., & Yu, A. (2008). Determination of total flavonoids in *Scutellaria*

- barbata* D. Don by dynamic ultrasonic extraction coupled with on-line spectrophotometry. *Analytica Chimica Acta*, 610(2), 217–223.
- Hung, H.Y., Qian, K., Morris-Natschke, S. L., Hsu, C.S., & Lee, K.H. (2012). Recent discovery of plant-derived anti-diabetic natural products. *Natural Product Reports*, 29(5), 580-606.
- Hussein, Z., Taher, S. W., Gilcharan Singh, H. K., & Chee, W. S. S. (2015). Diabetes care in Malaysia: Problems, new models, and solutions. *Annals of Global Health*, 81(6), 851–862.
- Ibrahim, R. M., El-Halawany, A. M., Saleh, D. O., Naggar, E. M. B. El, El-Shabrawy, A. E.-R. O., & El-Hawary, S. S. (2015). HPLC-DAD-MS/MS profiling of phenolics from *Securigera securidaca* flowers and its anti-hyperglycemic and anti-hyperlipidemic activities. *Revista Brasileira de Farmacognosia*, 25(2), 134–141.
- Ismail, M., Bagalkotkar, G., Iqbal, S., & Adamu, H. A. (2012). Anticancer properties and phenolic contents of sequentially prepared extracts from different parts of selected medicinal plants indigenous to Malaysia. *Molecules*, 17(5), 5745-5756.
- Ismail, M., Manickam, E., Danial, A. M., Rahmat, A., & Yahaya, A. (2000). Chemical composition and antioxidant activity of *Strobilanthes crispus* leaf extract. *The Journal of Nutritional Biochemistry*, 11(11–12), 536–542.
- Jaimand, K., & Rezaee, M. B. (2002). Chemical constituents of essential oils from *Mentha longifolia* (L.) Hudson var. *asiatica* (Boriss.) Rech. f. from Iran. *Journal of Essential Oil Research*, 14(2), 107–108.
- Javadi, N., Abas, F., Hamid, A. A., Simoh, S., Shaari, K., Ismail, I. S., Mediani, A., & Khatib, A. (2014). GC-MS-based metabolite profiling of *Cosmos caudatus* leaves possessing alpha-glucosidase inhibitory activity. *Journal of Food Science*, 79(6),
- Jay, D., Hitomi, H., & Griendling, K. K. (2006). Oxidative stress and diabetic cardiovascular complications. *Free Radical Biology and Medicine*, 40(2), 183-192.
- Jeong, C. H., Jeong, H. R., Choi, G. N., Kim, D. O., Lee, U., & Heo, H. J. (2011). Neuroprotective and anti-oxidant effects of caffeic acid isolated from *Erigeron annuus* leaf. *Chinese Medicine*, 6(1), 25.
- Ji, M., Li, C., & Li, Q. (2015). Rapid separation and identification of phenolics in crude red grape skin extracts by high performance liquid chromatography coupled to diode array detection and tandem mass spectrometry. *Journal of Chromatography A*, 1414, 138–146.
- Jin, J. L., Lee, Y. Y., Heo, J. E., Lee, S., Kim, J. M., & Yun-Choi, H. S. (2004). Anti-platelet pentacyclic triterpenoids from leaves of *Campsis grandiflora*. *Archives of Pharmacol Research*, 27(4), 376–380.
- Jo, S. H., Ka, E. H., Lee, H. S., Apostolidis, E., Jang, H. D., & Kwon, Y. I. (2009).

Comparison of antioxidant potential and rat intestinal α -glucosidases inhibitory activities of quercetin, rutin, and isoquercetin. *International Journal of Applied Research in Natural Products*, 2(4), 52–60.

Jorge, T. F., Rodrigues, J. A., Caldana, C., Schmidt, R., van Dongen, J. T., Thomas-Oates, J., & António, C. (2016). Mass spectrometry-based plant metabolomics: Metabolite responses to abiotic stress. *Mass Spectrometry Reviews*, 35(5), 620–649.

Jung, Y., Lee, J., Kwon, J., Lee, K. S., Ryu, D. H., & Hwang, G. S. (2010). Discrimination of the geographical origin of beef by ^1H NMR-based metabolomics. *Journal of Agricultural and Food Chemistry*, 58(19), 10458–10466.

Kalhan, S. C., Guo, L., Edmison, J., Dasarathy, S., McCullough, A. J., Hanson, R. W., & Milburn, M. (2011). Plasma metabolomic profile in nonalcoholic fatty liver disease. *Metabolism*, 60(3), 404–413.

Kamatham, S., Kumar, N., & Gudipalli, P. (2015). Isolation and characterization of gallic acid and methyl gallate from the seed coats of *Givotia rottleriformis* Griff. and their anti-proliferative effect on human epidermoid carcinoma A431 cells. *Toxicology Reports*, 2, 520–529.

Khakimov, B., Tseng, L. H., Godejohann, M., Bak, S., & Engelsens, S. B. (2016). Screening for triterpenoid saponins in plants using hyphenated analytical platforms. *Molecules*, 21(12), 1–19.

Khoo, L. T., Abdullah, J. O., Abas, F., Tohit, E. R. M., & Hamid, M. (2015a). Bioassay-guided fractionation of *Melastoma malabathricum* Linn. leaf solid phase extraction fraction and its anticoagulant activity. *Molecules*, 20(3), 3697–3715.

Khoo, L. W., Mediani, A., Zolkeflee, N. K. Z., Leong, S. W., Ismail, I. S., Khatib, A., Shaari, K., & Abas, F. (2015b). Phytochemical diversity of *Clinacanthus nutans* extracts and their bioactivity correlations elucidated by NMR based metabolomics. *Phytochemistry Letters*, 14, 123–133.

Kim, H. K., Choi, Y. H., & Verpoorte, R. (2010a). NMR-based metabolomic analysis of plants. *Nature Protocols*, 5(3), 536–549.

Kim, D.O., Lee, K. W., Lee, H. J., & Lee, C. Y. (2002). Vitamin C equivalent antioxidant capacity (VCEAC) of phenolic phytochemicals. *Journal of Agricultural and Food Chemistry*, 50(13), 3713–3717.

Kim, H. K., Choi, Y. H., & Verpoorte, R. (2006). Metabolomic analysis of *Catharanthus roseus* using NMR and principal component analysis. In *Plant Metabolomics* (pp. 261–276). Springer Berlin Heidelberg.

Kim, H. K., Choi, Y. H., & Verpoorte, R. (2011). NMR-based plant metabolomics: Where do we stand, where do we go? *Trends in Biotechnology*, 29(6), 267–275.

Kim, H. K., Saifullah, Khan, S., Wilson, E. G., Kricun, S. D. P., Meissner, A., Gorolaer,

- S., Deelder, A. M., Choi, Y. H., & Verpoorte, R. (2010b). Metabolic classification of South American *Ilex* species by NMR-based metabolomics. *Phytochemistry*, 71(7), 773–784.
- Kim, J. K., Park, S. Y., Lim, S. H., Yeo, Y., Cho, H. S., & Ha, S. H. (2013). Comparative metabolic profiling of pigmented rice (*Oryza sativa* L.) cultivars reveals primary metabolites are correlated with secondary metabolites. *Journal of Cereal Science*, 57(1), 14–20.
- Koay, Y. C., Wong, K. C., Osman, H., Ibrahim, M. S., & Asmawi, M. Z. (2013). Chemical constituents and biological activities of *Strobilanthes crispus* L. *Records of Natural Products*, 7(1), 59–64.
- Kongpichitchoke, T., Hsu, J. L., & Huang, T. C. (2015). Number of hydroxyl groups on the B-ring of flavonoids affects their antioxidant activity and interaction with phorbol ester binding site of PKC δ C1B domain: In vitro and in silico studies. *Journal of Agricultural and Food Chemistry*, 63(18), 4580–4586.
- Korus, A. (2012). Amino acid retention and protein quality in dried kale (*Brassica oleracea* L. var. *acephala*). *Journal of Food Processing and Preservation*, 38(2), 676–683.
- Kovalishyn, V., Aires-de-Sousa, J., Ventura, C., Elvas Leitão, R., & Martins, F. (2011). QSAR modeling of antitubercular activity of diverse organic compounds. *Chemometrics and Intelligent Laboratory Systems*, 107(1), 69–74.
- Kriaa, W., Fetoui, H., Makni, M., Zeghal, N., & Drira, N. E. (2012). phenolic contents and antioxidant activities of date palm (*Phoenix dactylifera* L.) leaves. *International Journal of Food Properties*, 15(6), 1220–1232.
- Kumar, S., Chandra, P., Bajpai, V., Singh, A., Srivastava, M., Mishra, D. K., & Kumar, B. (2015). Rapid qualitative and quantitative analysis of bioactive compounds from *Phyllanthus amarus* using LC/MS/MS techniques. *Industrial Crops and Products*, 69, 143–152.
- Kwon, D. J., Jeong, H. J., Moon, H., Kim, H. N., Cho, J. H., Lee, J. E., Hong, K. S., & Hong, Y. S. (2015). Assessment of green coffee bean metabolites dependent on coffee quality using a ^1H NMR-based metabolomics approach. *Food Research International*, 67, 175–182.
- Lanz, C., Patterson, A. D., Slavík, J., Krausz, K. W., Ledermann, M., Gonzalez, F. J., & Idle, J. R. (2009). Radiation metabolomics. 3. Biomarker discovery in the urine of gamma-irradiated rats using a simplified metabolomics protocol of gas chromatography-mass spectrometry combined with random forests machine learning algorithm. *Radiation Research*, 172(2), 198–212.
- Lapornik, B., Prošek, M., & Wondra, A. G. (2005). Comparison of extracts prepared from plant by-products using different solvents and extraction time. *Journal of Food Engineering*, 71(2), 214–222.
- Lasisi, D. (2014). A comparative study of effects of drying methods on quality of cocoa

- beans. *International Journal of Engineering Research & Technology*, 3(1), 991–996.
- Lee, J. E., Lee, B. J., Chung, J. O., Shin, H. J., Lee, S. J., Lee, C. H., & Hong, Y. S. (2011). ¹H NMR-based metabolomic characterization during green tea (*Camellia sinensis*) fermentation. *Food Research International*, 44(2), 597–604.
- Leiss, K. A., Choi, Y. H., Abdel-Farid, I. B., Verpoorte, R., & Klinkhamer, P. G. L. (2009). NMR metabolomics of thrips (*Frankliniella occidentalis*) resistance in *Senecio* hybrids. *Journal of Chemical Ecology*, 35(2), 219–229.
- Li, H., Pordesimo, L., & Weiss, J. (2004). High intensity ultrasound-assisted extraction of oil from soybeans. *Food Research International*, 37(7), 731–738.
- Li, H., Song, F., Xing, J., Tsao, R., Liu, Z., & Liu, S. (2009a). Screening and structural characterization of α -glucosidase inhibitors from hawthorn leaf flavonoids extract by ultrafiltration LC-DAD-MSn and SORI-CID FTICR MS. *Journal of the American Society for Mass Spectrometry*, 20(8), 1496–1503.
- Li, Y. Q., Zhou, F. C., Gao, F., Bian, J. S., & Shan, F. (2009b). Comparative evaluation of quercetin, isoquercetin and rutin as inhibitors of α -glucosidase. *Journal of Agricultural and Food Chemistry*, 57(24), 11463–11468.
- Li, Z., Meng, F., Zhang, Y., Sun, L., Yu, L., Zhang, Z., Peng, S., & Guo, J. (2016). Simultaneous quantification of hyperin, reynoutrin and guaijaverin in mice plasma by LC-MS/MS: Application to a pharmacokinetic study. *Biomedical Chromatography*, 30(7), 1124–1130.
- Liaw, A., & Wiener, M. (2002). Classification and regression by random forest. *R News*, 2(3), 18–22.
- Lima, I. C., Nora, R., de Carvalho, M. G., & Chaves, D. S. A. (2014). Distribution and chemotaxonomic significance of phenolic compounds in *Spermocoe verticillata* (L.) G. Mey. *Journal of Pharmacy & Pharmacognosy Research*, 2(1), 14–18.
- Lin, L., Lei, F., Sun, D. W., Dong, Y., Yang, B., & Zhao, M. (2012). Thermal inactivation kinetics of *Rabdosia serra* (Maxim.) Hara leaf peroxidase and polyphenol oxidase and comparative evaluation of drying methods on leaf phenolic profile and bioactivities. *Food Chemistry*, 134(4), 2021–2029.
- Liza, M. S., Abdul Rahman, R., Mandana, B., Jinap, S., Rahmat, A., Zaidul, I. S. M., & Hamid, A. (2010). Supercritical carbon dioxide extraction of bioactive flavonoid from *Strobilanthes crispus* (Pecah Kaca). *Food and Bioproducts Processing*, 88(2), 319–326.
- Llorent-Martinez, E. J., Spinola, V., Gouveia, S., & Castilho, P. C. (2015). HPLC-ESI-MSn characterization of phenolic compounds, terpenoid saponins, and other minor compounds in *Bituminaria bituminosa*. *Industrial Crops and Products*, 69, 80–90.
- Lobo, V., Patil, A., Phatak, A., & Chandra, N. (2010). Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacognosy Reviews*, 4(8), 118.

- Lu, S., Tran, B. N., Nelsen, J. L., & Aldous, K. M. (2009). Quantitative analysis of mitragynine in human urine by high performance liquid chromatography-tandem mass spectrometry. *Journal of Chromatography B*, 877(24), 2499–2505.
- Ludwig, C., & Viant, M. R. (2010). Two-dimensional J-resolved NMR spectroscopy: Review of a key methodology in the metabolomics toolbox. *Phytochemical Analysis*. John Wiley & Sons, Ltd.
- Ma, K., Han, J., Bao, L., Wei, T., & Liu, H. (2014). Two sarcoviolins with antioxidative and α -glucosidase inhibitory activity from the edible mushroom *Sarcodon leucopus* collected in Tibet. *Journal of Natural Products*, 77(4), 942–947.
- Ma, L., Chen, H., Zhu, W., & Wang, Z. (2013). Effect of different drying methods on physicochemical properties and antioxidant activities of polysaccharides extracted from mushroom *Inonotus obliquus*. *Food Research International*, 50(2), 633–640.
- Ma, Y. Q., Ye, X. Q., Fang, Z. X., Chen, J. C., Xu, G. H., & Liu, D. H. (2008a). Phenolic compounds and antioxidant activity of extracts from ultrasonic treatment of Satsuma Mandarin (*Citrus unshiu* Marc.) peels. *Journal of Agricultural and Food Chemistry*, 56(14), 5682–5690.
- Ma, Y., Ye, X., Hao, Y., Xu, G., Xu, G., & Liu, D. (2008b). Ultrasound-assisted extraction of hesperidin from Penggan (*Citrus reticulata*) peel. *Ultrasonics Sonochemistry*, 15(3), 227–232.
- Madeira, P. J. A., & Florêncio, M. H. (2012). *Applications of tandem mass spectrometry: From structural analysis to fundamental studies*. INTECH Open Access Publisher.
- Materska, M. (2014). Bioactive phenolics of fresh and freeze-dried sweet and semi-spicy pepper fruits (*Capsicum annuum* L.). *Journal of Functional Foods*, 7, 269–277.
- Matt, P., Krapp, A., Haake, V., Mock, H. P., & Stitt, M. (2002). Decreased rubisco activity leads to dramatic changes of nitrate metabolism, amino acid metabolism and the levels of phenylpropanoids and nicotine in tobacco antisense RBCS transformants. *Plant Journal*, 30(6), 663–677.
- Maulidiani, H., Khatib, A., Shaari, K., Abas, F., Shitan, M., Kneer, R., Neto, V., & Lajis, N. H. (2012). Discrimination of three *pegaga* (*Centella*) varieties and determination of growth-lighting effects on metabolites content based on the chemometry of ^1H nuclear magnetic resonance spectroscopy. *Journal of Agricultural and Food Chemistry*, 60(1), 410–417.
- Maulidiani, M., Sheikh, B. Y., Mediani, A., Wei, L. S., Ismail, I. S., Abas, F., & Lajis, N. H. (2015). Differentiation of *Nigella sativa* seeds from four different origins and their bioactivity correlations based on NMR-metabolomics approach. *Phytochemistry Letters*, 13, 308–318.
- Maulidiani, Abas, F., Khatib, A., Shaari, K., & Lajis, N. H. (2014). Chemical characterization and antioxidant activity of three medicinal Apiaceae species. *Industrial Crops and Products*, 55, 238–247.

- Maulidiani, Abas, F., Khatib, A., Shitan, M., Shaari, K., & Lajis, N. H. (2013). Comparison of Partial Least Squares and Artificial Neural Network for the prediction of antioxidant activity in extract of *Pegaga (Centella)* varieties from ^1H Nuclear Magnetic Resonance spectroscopy. *Food Research International*, 54(1), 852–860.
- Mavel, S., Nadal-Desbarats, L., Blasco, H., Bonnet-Brilhault, F., Barthélémy, C., Montigny, F., Sarda, P., Laumonnier, F., Vour, P., Andres, C. R., & Emond, P. (2013). ^1H – ^{13}C NMR-based urine metabolic profiling in autism spectrum disorders. *Talanta*, 114, 95–102.
- Mazlan, N. A., Mediani, A., Abas, F., Ahmad, S., Shaari, K., Khamis, S., & Lajis, N. H. (2013). Antioxidant, antityrosinase, anticholinesterase, and nitric oxide inhibition activities of three Malaysian *macaranga* species. *The Scientific World Journal*, 2013.
- Mediani, A., Abas, F., Khatib, A., Maulidiani, H., Shaari, K., Choi, Y. H., & Lajis, N. H. (2012). ^1H -NMR-based metabolomics approach to understanding the drying effects on the phytochemicals in *Cosmos caudatus*. *Food Research International*, 49(2), 763–770.
- Mediani, A., Abas, F., Khatib, A., Tan, C. P., Ismail, I. S., Shaari, K., Ismail, A., & Lajis, N. H. (2015a). Phytochemical and biological features of *Phyllanthus niruri* and *Phyllanthus urinaria* harvested at different growth stages revealed by ^1H NMR-based metabolomics. *Industrial Crops and Products*, 77, 602–613.
- Mediani, A., Abas, F., Khatib, A., Tan, C. P., Ismail, I. S., Shaari, K., Ismail, A., & Lajis, N. H. (2015b). Relationship between metabolites composition and biological activities of *Phyllanthus niruri* extracts prepared by different drying methods and solvents extraction. *Plant Foods for Human Nutrition*, 70(2), 184–192.
- Michalak, A. (2006). Phenolic compounds and their antioxidant activity in plants growing under heavy metal stress. *Polish Journal of Environmental Studies*, 15(4), 523–530.
- Mohd Nazrul, H. D., Mohd, L. J., Mohd, N. J., Normah, A., & Nurul Nabilah, M. F. (2011). Identification and isolation of methyl gallate as a polar chemical marker for *Labisia pumila* Benth. *Journal of Tropical Agriculture and Food Science*, 39(2), 279–284.
- Mushtaq, M. Y., Choi, Y. H., Verpoorte, R., & Wilson, E. G. (2014). Extraction for metabolomics: Access to the metabolome. *Phytochemical Analysis*, 25(4), 291–306.
- Mustafa, R. A., Hamid, A. A., Mohamed, S., & Bakar, F. A. (2010). Total phenolic compounds, flavonoids, and radical scavenging activity of 21 selected tropical plants. *Journal of Food Science*, 75(1), C28–C35.
- Mustaffa, F., Indurkar, J., Ali, N. I. M., Hanapi, A., Shah, M., Ismail, S., & Mansor, S. M. (2011). A review of Malaysian medicinal plants with potential antidiabetic

- activity. *Journal of Pharmacy Research*, 4(11), 4217–4224.
- Nacef, S., Ben, H., Abreu, P., & Mighri, Z. (2010). Phenolic constituents of *Convolvulus dorycnium* L. flowers. *Phytochemistry Letters*, 3(2), 66–69.
- Nakamura, Y., Murakami, A., Koshimizu, K., & Ohigashi, H. (1996). Identification of pheophorbide a and its related compounds as possible anti-tumor promoters in the leaves of *Neptunia oleracea*. *Bioscience, Biotechnology, and Biochemistry*, 60(6), 1028–1030.
- Nash, H., & Stroupe, S. (2003). *Complete guide to water garden plants*. Sterling Publishing Company, Inc..
- Niki, E. (2010). Assessment of antioxidant capacity *in vitro* and *in vivo*. *Free Radical Biology and Medicine*, 49(4), 503–515.
- Norfarizan-Hanoon, N. A., Asmah, R., Rokiah, M. Y., Fauziah, O., & Faridah, H. (2009). Antihyperglycemic, hypolipidemic and antioxidant enzymes effect of *Strobilanthes crispus* juice in normal and streptozotocin-induced diabetic male and female rats. *International Journal of Pharmacology*, 5(3), 200–207.
- Oboh, G., Ademiluyi, A. O., Akinyemi, A. J., Henle, T., Saliu, J. A., & Schwarzenbolz, U. (2012). Inhibitory effect of polyphenol-rich extracts of jute leaf (*Corchorus olitorius*) on key enzyme linked to type 2 diabetes (α -amylase and α -glucosidase) and hypertension (angiotensin I converting) *in vitro*. *Journal of Functional Foods*, 4(2), 450–458.
- Oshiro, T. M., Perez, P. S., & Baranauskas, J. A. (2012). How many trees in a random forest? *Lecture Notes in Computer Science (Including Subseries Lecture Notes in Artificial Intelligence and Lecture Notes in Bioinformatics)*, 7376 LNAI, 154–168.
- Oszmiański, J., Kolniak-Ostek, J., & Wojdyło, A. (2013). Characterization and content of flavonol derivatives of *Allium ursinum* L. plant. *Journal of Agricultural and Food Chemistry*, 61(1), 176–184.
- Oszmiański, J., Wojdyło, A., Nowicka, P., Teleszko, M., Cebulak, T., & Wolanin, M. (2015). Determination of phenolic compounds and antioxidant activity in leaves from wild *Rubus* L. species. *Molecules*, 20(3), 4951–4966.
- Park, B. J., Matsuta, T., Kanazawa, T., Park, C. H., Chang, K. J., & Onjo, M. (2012). Phenolic compounds from the leaves of *Psidium guajava* II. quercetin and its glycosides. *Chemistry of Natural Compounds*, 48(3), 477–479.
- Parthasarathy, S., Azizi, J. Bin, Ramanathan, S., Ismail, S., Sasidharan, S., Mohd, M. I., & Mansor, S. M. (2009). Evaluation of antioxidant and antibacterial activities of aqueous, methanolic and alkaloid extracts from *Mitragyna speciosa* (rubiacaceae family) leaves. *Molecules*, 14(10), 3964–3974.
- Paudel, L., Wyzgoski, F. J., Giusti, M. M., Johnson, J. L., Rinaldi, P. L., Scheerens, J. C., Chanon, A. M., Bomser, J. A., Miller, A. R., Hardy, J. K., & Reese, R. N. (2014). NMR-based metabolomic investigation of bioactivity of chemical

- constituents in black raspberry (*Rubus occidentalis* L.) fruit extracts. *Journal of Agricultural and Food Chemistry*, 62(8), 1989–1998.
- Paul, S. B., Choudury, S. N., & De, B. (2012). Structural elucidation of a bioactive compound from the leaves of *Neptunia prostrate*. *Asian Journal of Chemistry*, 24(4), 1469–1472.
- Perron, N. R., & Brumagim, J. L. (2009). A review of the antioxidant mechanisms of polyphenol compounds related to iron binding. *Cell Biochemistry and Biophysics*, 53(2), 75–100.
- Pinela, J., Barros, L., Carvalho, A. M., & Ferreira, I. C. F. R. (2011). Influence of the drying method in the antioxidant potential and chemical composition of four shrubby flowering plants from the tribe Genisteae (Fabaceae). *Food and Chemical Toxicology*, 49(11), 2983–2989.
- Pukalskienė, M., Venskutonis, P. R., & Pukalskas, A. (2015). Phytochemical composition and antioxidant properties of *Filipendula vulgaris* as a source of healthy functional ingredients. *Journal of Functional Foods*, 15, 233–242.
- Rao, V. S. R., Qasba, P. K., Balaji, P. V., & Chandrasekaran, R. (1998). *Conformation of carbohydrates*. Harwood Academic Publishers.
- Rattanasena, P. (2012). Antioxidant and antibacterial activities of vegetables and fruits commonly consumed in Thailand. *Pakistan Journal of Biological Sciences*, 15(18), 877–882.
- Rochfort, S. J., Ezernieks, V., Maher, A. D., Ingram, B. A., & Olsen, L. (2013). Mussel metabolomics - Species discrimination and provenance determination. *Food Research International*, 54(1), 1302–1312.
- Rodrigues, S., & Pinto, G. A. S. (2007). Ultrasound extraction of phenolic compounds from coconut (*Cocos nucifera*) shell powder. *Journal of Food Engineering*, 80(3), 869–872.
- Routray, W., & Orsat, V. (2014). MAE of phenolic compounds from blueberry leaves and comparison with other extraction methods. *Industrial Crops and Products*, 58, 36–45.
- Ruiz-Montañez, G., Ragazzo-Sánchez, J. A., Calderón-Santoyo, M., Velázquez-De La Cruz, G., Ramírez De León, J. A., & Navarro-Ocaña, A. (2014). Evaluation of extraction methods for preparative scale obtention of mangiferin and lupeol from mango peels (*Mangifera indica* L.). *Food Chemistry*, 159, 267–272.
- Saboonchian, F., Jamei, R., & Sarghein, S. H. (2014). Phenolic and flavonoid content of *Elaeagnus angustifolia* L. (leaf and flower). *Avicenna Journal of Phytomedicine*, 4(4), 231.
- Sakdarat, S., Shuyprom, A., Pientong, C., Ekalaksananan, T., & Thongchai, S. (2009). Bioactive constituents from the leaves of *Clinacanthus nutans* Lindau. *Bioorganic and Medicinal Chemistry*, 17(5), 1857–1860.

- Sanchez-Rabaneda, F., Jauregui, O., Casals, I., Andres-Lacueva, C., Izquierdo-Pulido, M., & Lamuela-Raventos, R. M. (2003). Liquid chromatographic/electrospray ionization tandem mass spectrometric study of the phenolic composition of cocoa (*Theobroma cacao*). *Journal of Mass Spectrometry*, 38(1), 35–42.
- Sato, Y., Itagaki, S., Kurokawa, T., Ogura, J., Kobayashi, M., Hirano, T., Sugawara, M., & Iseki, K. (2011). *In vitro* and *in vivo* antioxidant properties of chlorogenic acid and caffeic acid. *International Journal of Pharmaceutics*, 403(1), 136–138.
- Savic, I., Nikolic, V., Savic, I., Nikolic, L., Jovic, M., & Jovic, M. (2014). The qualitative analysis of the green tea extract using ESI-MS method. *Savremene Tehnologije*, 3(1), 30–37.
- Schievano, E., Stocchero, M., Morelato, E., Facchin, C., & Mammi, S. (2012). An NMR-based metabolomic approach to identify the botanical origin of honey. *Metabolomics*, 8(4), 679–690.
- Schripsema, J. (2010). Application of NMR in plant metabolomics: Techniques, problems and prospects. *Phytochemical Analysis*, 21(1), 14–21.
- Sekar, M., Zulhilmi, M., Hamdi, A. Y., Nabila, N., Zahida, Z., & Shafiq, M. (2014). Ten commonly available medicinal plants in Malaysia used for the treatment of diabetes-a review. *Asian Journal of Pharmaceutical and Clinical Research*, 7(1), 1–5.
- Selmar, D., & Kleinwächter, M. (2013). Influencing the product quality by deliberately applying drought stress during the cultivation of medicinal plants. *Industrial Crops and Products*, 42, 558–566.
- Shaham, O., Wei, R., Wang, T. J., Ricciardi, C., Lewis, G. D., Vasan, R. S., Carr, S. A., Thadhani, R., Gerszten, R. E., & Mootha, V. K. (2008). Metabolic profiling of the human response to a glucose challenge reveals distinct axes of insulin sensitivity. *Molecular Systems Biology*, 4(1), 214.
- Shobana, S., Sreerama, Y. N., & Malleshi, N. G. (2009). Composition and enzyme inhibitory properties of finger millet (*Eleusine coracana* L.) seed coat phenolics: Mode of inhibition of α -glucosidase and pancreatic amylase. *Food Chemistry*, 115(4), 1268–1273.
- Shou, M., Galinada, W. A., Wei, Y. C., Tang, Q., Markovich, R. J., & Rustum, A. M. (2009). Development and validation of a stability-indicating HPLC method for simultaneous determination of salicylic acid, betamethasone dipropionate and their related compounds in Diprosalic Lotion®. *Journal of Pharmaceutical and Biomedical Analysis*, 50(3), 356–361.
- Shuib, N. H., Shaari, K., Khatib, A., Maulidiani, Kneer, R., Zareen, S., Raof, S. M., Lajis, N. H., & Neto, V. (2011). Discrimination of young and mature leaves of *Melicope ptelefolia* using ^1H NMR and multivariate data analysis. *Food Chemistry*, 126(2), 640–645.
- Siddhuraju, P., & Becker, K. (2003). Antioxidant properties of various solvent extracts

- of total phenolic constituents from three different agroclimatic origins of Drumstick Tree (*Moringa oleifera* Lam.) leaves. *Journal of Agricultural and Food Chemistry*, 51(8), 2144–2155.
- Silverstein, R. M., Webster, F. X., Kiemle, D. J., & Bryce, D. L. (2014). *Spectrometric identification of organic compounds*. John Wiley & sons.
- Simirgiotis, J. M., Schmeda-Hirschmann, G., Bórquez, J., & Kennelly, J. E. (2013). The *Passiflora tripartita* (Banana Passion) fruit: A Source of bioactive flavonoid c-glycosides isolated by HSCCC and characterized by HPLC–DAD–ESI/MS/MS. *Molecules*, 18(2), 1672–1692.
- Simoh, S., Quintana, N., Kim, H. K., Choi, Y. H., & Verpoorte, R. (2009). Metabolic changes in *Agrobacterium tumefaciens*-infected *Brassica rapa*. *Journal of Plant Physiology*, 166(10), 1005–1014.
- Sintayehu, B., Asres, K., & Raghavendra, Y. (2012). Radical scavenging activities of the leaf extracts and a flavonoid glycoside isolated from *Cineraria abyssinica* Sch. Bip. Exa. Rich. *Journal of Applied Pharmaceutical Science*, 2(4), 44–49.
- Siqueira, S., Falcão-Silva, V. D. S., Agra, M. de F., Dariva, C., Siqueira-Júnior, J. P. De, & Fonseca, M. J. V. (2011). Biological activities of *Solanum paludosum* Moric. extracts obtained by maceration and supercritical fluid extraction. *Journal of Supercritical Fluids*, 58(3), 391–397.
- Siriamornpun, S., Kaisoon, O., & Meeso, N. (2012). Changes in colour, antioxidant activities and carotenoids (lycopene, β -carotene, lutein) of marigold flower (*Tagetes erecta* L.) resulting from different drying processes. *Journal of Functional Foods*, 4(4), 757–766.
- Smolinska, A., Blanchet, L., Buydens, L. M. C., & Wijmenga, S. S. (2012). NMR and pattern recognition methods in metabolomics: From data acquisition to biomarker discovery: A review. *Analytica Chimica Acta*, 750, 82–97.
- Son, H., Hwang, G., Ahn, H., Park, W., Lee, C., & Hong, Y. (2009). Characterization of wines from grape varieties through multivariate statistical analysis of ^1H NMR spectroscopic data. *Food Research International*, 42(10), 1483–1491.
- Soto, C., Caballero, E., Pérez, E., & Zúñiga, M. E. (2014). Effect of extraction conditions on total phenolic content and antioxidant capacity of pretreated wild *Peumus boldus* leaves from Chile. *Food and Bioprocess Processing*, 92(3), 328–333.
- Stoker, H. S. (2012). *General, organic, and biological chemistry* (5th ed.). Nelson Education.
- Struijs, K., Vincken, J., & Gruppen, H. (2008). Comparison of atmospheric pressure chemical ionization and electrospray ionization mass spectrometry for the detection of lignans from sesame seeds. *Rapid Communications in Mass Spectrometry*, 22(22), 3615–3623.
- Sugiwati, S., Setiasih, S., & Afifah, E. (2009). Antihyperglycemic activity of the

- Mahkota Dewa [*Phaleria macrocarpa* (Scheff.) Boerl.] leaf extracts as an α -glucosidase inhibitor. *Makara Kesehatan*, 13(2), 74–78.
- Sulaiman, S. F., Sajak, A. A. B., Ooi, K. L., Supriatno, & Seow, E. M. (2011). Effect of solvents in extracting polyphenols and antioxidants of selected raw vegetables. *Journal of Food Composition and Analysis*, 24(4–5), 506–515.
- Sultana, B., Anwar, F., & Ashraf, M. (2009). Effect of extraction solvent/technique on the antioxidant activity of selected medicinal plant extracts. *Molecules*, 14(6), 2167–2180.
- Sun, J., Liang, F., Bin, Y., Li, P., & Duan, C. (2007). Screening non-colored phenolics in red wines using liquid chromatography/ultraviolet and mass spectrometry/mass spectrometry libraries. *Molecules*, 12(3), 679–693.
- Svetnik, V., Liaw, A., Tong, C., Culberson, J. C., Sheridan, R. P., & Feuston, B. P. (2003). Random forest: A classification and regression tool for compound classification and QSAR modeling. *Journal of Chemical Information and Computer Sciences*, 43(6), 1947–1958.
- Tadera, K., Minami, Y., Takamatsu, K., & Matsuoka, T. (2006). Inhibition of α -glucosidase and α -amylase by flavonoids. *Journal of Nutritional Science and Vitaminology*, 52(2), 149–153.
- Takayama, H. (2004). Chemistry and pharmacology of analgesic indole alkaloids from the rubiaceous plant, *Mitragyna speciosa*. *Chemical & Pharmaceutical Bulletin*, 52(8), 916–928.
- Thalang, N. V., Trakoontivakorn, G., & Nakahara, K. (2001). Determination of antioxidant activity of some commonly consumed leafy vegetables in Thailand. *JIRCAS Journal for Scientific Papers (Japan)*, 9, 39–46.
- Tirzitis, G., & Bartosz, G. (2010). Determination of antiradical and antioxidant activity: basic principles and new insights. *Acta Biochimica Polonica*, 57(1), 139–142.
- Touw, W. G., Bayjanov, J. R., Overmars, L., Backus, L., Boekhorst, J., Wels, M., & Sacha van Hijum, A. F. T. (2013). Data mining in the life science with random forest: A walk in the park or lost in the jungle? *Briefings in Bioinformatics*, 14(3), 315–326.
- Uttara, B., Singh, A. V., Zamboni, P., & Mahajan, R. T. (2009). Oxidative stress and neurodegenerative diseases: a review of upstream and downstream antioxidant therapeutic options. *Current Neuropharmacology*, 7(1), 65–74.
- Valko, M., Leibfritz, D., Moncol, J., Cronin, M. T. D., Mazur, M., & Telser, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. *The International Journal of Biochemistry & Cell Biology*, 39(39), 44–84.
- Vijayashree, S. B., Suresh, H. M., Hemantkumar, V., Hatapakki, B. C., Shivakumar, S. I., Hallikeri, C. S., & Chandur, V. K. (2006). Anti-inflammatory activity of leaves of *Nepenthes oleracea*. *Advances in Pharmacology and Toxicology*, 3, 21–24.

- Waksmundzka-Hajnos, M., & Sherma, J. (2010). *High performance liquid chromatography in phytochemical analysis*. CRC Press.
- Wan-Ibrahim, W. I., Sidik, K., & Kuppasamy, U. R. (2010). A high antioxidant level in edible plants is associated with genotoxic properties. *Food Chemistry*, 122(4), 1139–1144.
- Wan, C., Yuan, T., Cirello, A. L., & Seeram, N. P. (2012). Antioxidant and α -glucosidase inhibitory phenolics isolated from highbush blueberry flowers. *Food Chemistry*, 135(3), 1929–1937.
- Wang, D. M., Pu, W. J., Wang, Y. H., Zhang, Y. J., & Wang, S. S. (2012). A new isorhamnetin glycoside and other phenolic compounds from *Callianthemum taipaicum*. *Molecules*, 17(4), 4595–4603.
- Wang, Y., Xiang, L., Wang, C., Tang, C., & He, X. (2013). Antidiabetic and antioxidant effects and phytochemicals of mulberry fruit (*Morus alba* L.) polyphenol enhanced extract. *PLOS ONE*, 8(7), e71144.
- Wansi, J. D., Lallemand, M. C., Chiozem, D. D., Toze, F. A. A., Mbaze, L. M., Naharkhan, S., Iqbal, M. C., Tillequin, F., Wandji, J., & Fomum, Z. T. (2007). α -Glucosidase inhibitory constituents from stem bark of *Terminalia superba* (Combretaceae). *Phytochemistry*, 68(15), 2096–2100.
- Welch, C. R., Wu, Q., & Simon, J. E. (2008). Recent advances in anthocyanin analysis and characterization. *Current Analytical Chemistry*, 4(2), 75–101.
- Wójciak-Kosior, M., Sowa, I., Kocjan, R., & Nowak, R. (2013). Effect of different extraction techniques on quantification of oleanolic and ursolic acid in *Lamii albi flos*. *Industrial Crops and Products*, 44, 373–377.
- Wolfender, J. L., Rudaz, S., & Young, H. C., & Hye, K. K. (2013). Plant metabolomics: From holistic data to relevant biomarkers. *Current Medicinal Chemistry*, 20(8), 1056–1090.
- Wong, K., & Cheung, P. C. (2001). Influence of drying treatment on three *Sargassum* species. *Journal of Applied Phycology*, 13(1), 43–50.
- Wongsa, P., Chaiwarit, J., & Zamaludien, A. (2012). *In vitro* screening of phenolic compounds, potential inhibition against α -amylase and α -glucosidase of culinary herbs in Thailand. *Food Chemistry*, 131(3), 964–971.
- World Health Organization. (2016). Diabetes (Fact sheet No. 312). Retrieved from <http://www.who.int/mediacentre/factsheets/fs312/en/>
- Wu, Y., Jiang, X., Zhang, S., Dai, X., Liu, Y., Tan, H., Gao, L., & Xia, T. (2016). Quantification of flavonol glycosides in *Camellia sinensis* by MRM mode of UPLC-QQQ-MS/MS. *Journal of Chromatography B*, 1017, 10–17.
- Wyrepkowski, C. C., Lu, D., Gomes, M., Sinhorin, A. P., Vilegas, W., De Grandis, R. A., Resende, F. A., Varanda, E. A., & do Santos, L. C. (2014). Characterization

- and quantification of the compounds of the ethanolic extract from *Caesalpinia ferrea* stem bark and evaluation of their mutagenic activity. *Molecules*, 19(10), 16039–16057.
- Xiao, J., Kai, G., Yamamoto, K., & Chen, X. (2013). Advance in dietary polyphenols as α -glucosidases inhibitors: A Review on structure-activity relationship aspect. *Critical Reviews in Food Science and Nutrition*, 53(8), 818–836.
- Yan, Z., Yang, X., Wu, J., Su, H., Chen, C., & Chen, Y. (2011). Qualitative and quantitative analysis of chemical constituents in traditional Chinese medicinal formula Tong-Xie-Yao-Fang by high-performance liquid chromatography/diode array detection/electrospray ionization tandem mass spectrometry. *Analytica Chimica Acta*, 691(1–2), 110–118.
- Yang, S. Y., Kim, H. K., Lefeber, A. W. M., Erkelens, C., Angelova, N., Choi, Y. H., & Verpoorte, R. (2006). Application of two-dimensional nuclear magnetic resonance spectroscopy to quality control of ginseng commercial products. *Planta Medica*, 72(4), 364–369.
- Yao, Y., Sang, W., Zhou, M., & Ren, G. (2010). Antioxidant and α -glucosidase inhibitory activity of colored grains in China. *Journal of Agricultural and Food Chemistry*, 58(2), 770–774.
- Yaya, S., Amian, K., Benjamin, B., Fanté, B., Sorho, S., Amadou, T. S., & Jean-marie, C. (2012). Flavonoids and gallic acid from leaves of *Santaloides afzelii* (Connaraceae). *Rasayan Journal of Chemistry*, 5(3), 332–337.
- Ye, M., Yang, W. Z., Liu, K. Di, Qiao, X., Li, B. J., Cheng, J., Feng, J., Guo, D. A., & Zhao, Y. Y. (2012). Characterization of flavonoids in *Millettia nitida* var. *hirsutissima* by HPLC/DAD/ESI-MSn. *Journal of Pharmaceutical Analysis*, 2(1), 35–42.
- You, Q., Chen, F., Wang, X., Jiang, Y., & Lin, S. (2012). Anti-diabetic activities of phenolic compounds in muscadine against alpha-glucosidase and pancreatic lipase. *LWT - Food Science and Technology*, 46(1), 164–168.
- Zhang, Q., Zhang, J., Shen, J., Silva, A., Dennis, D. A., & Barrow, C. J. (2006). A simple 96-well microplate method for estimation of total polyphenol content in seaweeds. In *Eighteenth International Seaweed Symposium* (pp. 219–224). Springer Netherlands.
- Zhang, Z., Luo, A., Zhong, K., Huang, Y., Gao, Y., Zhang, J., Gao, H., Xu, Z., & Gao, X. (2013). α -Glucosidase inhibitory activity by the flower buds of *Lonicera japonica* Thunb. *Journal of Functional Foods*, 5(3), 1253–1259.
- Zhao, Y., Li, X., Zeng, X., Huang, S., Hou, S., & Lai, X. (2014). Characterization of phenolic constituents in *Lithocarpus polystachyus*. *Analytical Methods*, 6(5), 1359–1363.
- Zheng, W., Zhang, M., Zhao, Y., Miao, K., Pan, S., Cao, F., & Dai, Y. (2011). Analysis of antioxidant metabolites by solvent extraction from sclerotia of *Inonotus*

obliquus (Chaga). *Phytochemical Analysis*, 22(2), 95–102.

Zhou, X., Peng, J., Fan, G., & Wu, Y. (2005). Isolation and purification of flavonoid glycosides from *Trollius ledebouri* using high-speed counter-current chromatography by stepwise increasing the flow-rate of the mobile phase. *Journal of Chromatography A*, 1092(2), 216–221.

