



UNIVERSITI PUTRA MALAYSIA

***ROOT CULTURES AND ELICITATION OF SECONDARY METABOLITES
FROM *Labisia pumila* Benth & Hook. f. var. *alata****

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By

TAN SIN LI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Science**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
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Chairman : Associate Professor Norihan Mohd Saleh, PhD
Faculty : Biotechnology and Biomolecular Sciences

Labisia pumila var *alata* (Kacip Fatimah) is currently one of the eighteen main herbs under the EPP 1 of Agriculture cluster of the Economic Transformation Programme, Malaysia. Due to the slow and non-uniform growth of this plant, destructive harvesting and the lack of plant material to sustain the demand of the bio-pharmaceutical industry, an alternative approach is crucially needed to ensure sustainable supply of the herbal raw material. The development of organ cultures such as hairy roots and adventitious roots is a suitable approach to curb the problem of sustaining raw material for the production of pharmaceuticals. Furthermore, the secondary metabolites produced in *L. pumila* has not been well studied. In this study, hairy roots of *L. pumila* were successfully established from leaf with petiole explants using three strains of *Agrobacterium rhizogenes* (A4, NCPPB2629 and NCPPB1855) which produced 16.67%, 70% and 40% transformation efficiencies, respectively. Strain A4 showed a rapid growth rate compared to strain NCPPB2629 and NCPPB1855 with a doubling time of 14 days. Subsequently, liquid cultures were established using strain A4. The presence of *rol C* gene (strain A4) and *rol B* (strain NCPPB2629 and NCPPB1855) genes were determined using PCR analysis. Adventitious roots were also successfully initiated using a combination of auxins indole-3-butyric acid (IBA) and α -naphthalene acetic acid (NAA), both supplemented in 2.69 mM concentration in GM1 medium. A total of 100% frequency of adventitious root induction with 9.93 ± 1.18 mean number of roots formed per explant were produced after 14 days of culture. Tandem mass spectrometry platforms [time of flight (TOF) based and quadruple ion-trap based (QTRAP)] were used to detect and compare secondary metabolites produced in different tissues of *L. pumila* using Principle Component Analysis (PCA). The time-of-flight based tandem mass spectrometer managed to tentatively identify three compounds (FG1, FG2 and AP1) which were produced abundantly in the aqueous extracts of tissue cultured leaves. The production of triterpene saponin, TSrA was also noted in the aqueous extracts of the natural stem (51.33 mg/g), natural roots (132.48 mg/g), tissue cultured

leaves (135.65 mg/g), tissue cultured roots (250.75 mg/g) and hairy roots (361.26 mg/g). Methanol extracts of elicited hairy roots in shake flasks using Methyl jasmonate elicitor using different two stage elicitation procedures produced higher concentrations of TSrA (11.8-fold increase in GM3-PM1 media) and AP1 (2.8-fold increase in GM3-PM3 media). Another triterpene saponin, TSrB was also obtained from methanol extracts of hairy roots elicited in GM3-PM2 medium (0.52 mg/g). This compound has never been reported found in any other tissues of *L. pumila*. Production of hairy root biomass was upscaled further in laboratory scale bioreactors. In terms of biomass production, stirred tank bioreactor with elephant ear impeller, bubbled flask and balloon type bubble bioreactor (BTBB A) produced 6.94-fold, 4.0-fold and 8.42-fold of increase in biomass respectively. The production of hairy root biomass was proven to be effective in BTBB A. Hence, further optimization were made to enhance the production of secondary metabolites. A total of 13 compounds (TSrA, TSrB, TScA, TScB, TScD, TSCH, TSmE, TSmH, TSrp, TSd xp, AP1, AP2 and FG1) were detected from various tissues of *L. pumila*. Among these compounds, TSrB, TScA, TScB, TScD, TSCH, TSrp and TSd xp productions were enhanced in the BTBB D bioreactor. This bioreactor produced a maximum biomass accumulation of 15.04 fold. The findings in this study concluded that the supply of herbal raw material can be enhanced through the usage of organ cultures, elicitation and upscaling in bioreactor platforms.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Sarjana Sains

**KULTUR AKAR DAN ELISITASI METABOLIT SEKUNDER DARI
Labisia pumila Benth & Hook. f. var. *alata***

Oleh

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Disember 2016

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Labisia pumila var *alata* (Kacip Fatimah) merupakan salah satu daripada lapan belas herba utama di bawah EPP1 Kluster Pertanian di bawah Program Transformasi Ekonomi, Malaysia. Ekoran daripada itu, kelambatan dan ketidakseragaman tumbesaran tumbuhan ini, penuaian yang berleluasa dan kekurangan bahan mentah untuk memenuhi keperluan pasaran dalam industri farmaseutikal, pendekatan yang alternatif amatlah diperlukan bagi memastikan kesinambungan bekalan bahan mentah herba. Penghasilan kultur organ seperti akar rerambut dan akar adventisius adalah sesuai untuk menyelesaikan untuk memastikan bekalan bahan mentah yang berterusan untuk pengeluaran ubat farmasiutikal. Tambahan pula, penghasilan metabolit sekunder di dalam *L. pumila* masih belum dikaji dengan teliti. Dalam kajian ini, akar rerambut *L. pumila* telah berjaya dihasilkan daripada eksplan daun dengan petiole hasil daripada penggunaan tiga strain *Agrobacterium rhizogenes* (A4, NCPPB2629, NCPPB1855) di mana 16.67%, 70% dan 40% kadar transformasi diperolehi. Di antara tiga strain *A. rhizogenes* yang digunakan, akar rerambut yang dihasilkan daripada strain A4 mempunyai kadar pertumbuhan yang lebih baik berbanding dengan strain NCPPB2629 dan NCPPB1855. Biomas akar rerambut digandakan dua kali ganda dalam masa 14 hari. Justeru itu, kultur akar rerambut di dalam media cecair dihasilkan daripada akar rerambut yang dihasilkan daripada strain A4. Kehadiran gen *rol C* (strain A4) dan gen *rol B* (strain NCPPB2629 dan NCPPB1855) ditentukan oleh analisis PCR. Akar adventisius juga telah berjaya dihasilkan menggunakan kombinasi auksin indole-3-butyric acid (IBA) dan α -naphthalene acetic acid (NAA) di mana kepekatan 2.69 mM untuk setiap auksin dibekalkan bersama-sama dengan medium GM1. Frekuensi pengaruh akar adventisius adalah 100% manakala purata bilangan akar yang dihasilkan pada setiap eksplan adalah 9.93 ± 1.18 , 14 hari selepas kultur. Platform Tandem Mass Spectrometry [time-of-flight (TOF) dan quadruple ion trap (QTRAP)] digunakan untuk mengesan dan membuat perbandingan metabolit sekunder dengan analisis komponen principal (Principle Component Analysis) (PCA). Analisis yang menggunakan

time-of flight berjaya membuat pengenalan tentatif kepada tiga jenis sebatian iaitu FG1, FG2 dan AP1 di mana tiga jenis sebatian ini dihasilkan dengan kuantiti yang banyak di dalam ekstrak akuas daun dari kultur tis. Triterpene saponin, TSrA dihasilkan di dalam ekstrak akuas batang semulajadi (51.33 mg/g), diikuti oleh akar semulajadi (132.48 mg/g), daun kultur tis (135.65 mg/g), akar kultur tis (250.75 mg/g) dan akar rerambut (361.26 mg/g). Ekstrak methanol untuk akar rerambut yang dielisitasi di dalam kelalang menggunakan elisitor methyl jasmonate serta menggunakan prosedur elisitasi yang berlainan berjaya menghasilkan nilai kepekatan yang lebih tinggi untuk TSrA (11.8 kali ganda peningkatan di dalam media GM3-PM1) dan Fatimahol (2.8 kali ganda kepekatan di dalam media GM3-PM3). Satu lagi triterpene saponin, TSrB dijumpai di dalam ekstrak methanol yang dielisitasi di dalam media GM3-PM2 (0.52 mg/g). Sebatian ini masih belum pernah dijumpai di dalam tis-tis *L. pumila* yang lain. Penghasilan biomas akar rerambut diskalakan dengan lebih besar di dalam bioreaktor skala makmal. Penghasilan biomas akar rerambut di dalam bioreaktor tangki kacau dengan pendesak telinga gajah, bioreaktor kelalang gelombang dan bioreaktor belon kolum gelombang (BTBB A) adalah masing-masing 6.94 kali ganda, 4.0 kali ganda dan 8.42 kali ganda. Penghasilan biomas akar rerambut adalah efektif di dalam BTBB A. Oleh itu, optimasi dibuat untuk meningkatkan penghasilan metabolit sekunder di dalam bioreaktor ini. Sebanyak 13 sebatian semulajadi (TSrA, TSrB, TScA, TScB, TScD, TScH, TSmE, TSmH, TSrp, TSd xp, AP1, AP2 dan FG1) dikesan dari pelbagai tis *L. pumila*. Di antara sebatian-sebatian ini, penghasilan TSrB, TScA, TScB, TScD, TScH, TSrp dan TSd xp ditingkatkan di dalam bioreaktor BTBB D. Bioreaktor ini menghasilkan biomas maksimum sebanyak 15.04 kali ganda. Kesimpulannya, hasil daripada kajian ini boleh menambahbaik bekalan bahan mentah herba melalui kultur organ, elisitasi dan penskalaan besar dalam platform bioreaktor.

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I certify that a Thesis Examination Committee has met on 23 December 2016 to conduct the final examination of Tan Sin Li on her thesis entitled "Root Cultures and Elicitation of Secondary Metabolites from *Labisia pumila* Benth & Hook.f. var. *alata*" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

AP	Alkylated phenolic compounds (<i>compound names are kept confidential at this stage due to patent filing in progress</i>)
<i>aux</i>	Auxin biosynthesis gene
BAP	6-Benzylaminopurine
BTBB	Balloon Type Bubble Bioreactor
EPP	Entry Point Project
ETP	Economic Transformation Programme
FG	Flavonoid glycoside compounds (<i>compound names are kept confidential at this stage due to patent filing in progress</i>)
GM	Growth Medium (<i>Media formulations and names are kept confidential at this stage due to patent filing in progress</i>)
IBA	Indole-3-butyric acid
LC	Liquid Chromatography
MJ	Methyl Jasmonate
MS	Murashige & Skoog
NAA	α -naphthalene acetic acid
NCPPB	National Collection of Plant Pathogenic Bacteria
NKEA	National Key Economic Areas
PCA	Principle Component Analysis
PM	Production Medium (<i>Media formulations and names are kept confidential at this stage due to patent filing in progress</i>)
QTRAP	Quadruple-ion-trap
<i>rol</i>	Root loci
TOF	Time-of-flight

TS

Triterpene saponin compounds (*compound names are kept confidential at this stage due to patent filing in progress*)

vir

Virulence gene



CHAPTER 1

INTRODUCTION

1.1 Background Information

Traditional medicinal plants from the tropical green forest of Malaysia have gained popularity due to the presence of bioactive compounds that are much needed by local biopharmaceutical industry. In fact, the herbal industry in the world is expected to reach an income of USD115 billion by the year 2020 (Global Industry Analyst Inc, 2015).

Labisia pumila Benth & Hook. f. var. *alata*. (Myrsinaceae), locally known as Kacip Fatimah can be found in the shady part of lowland primary forest and humus-rich secondary forest in Malaysia. Scientists and several publications have shown that the plant may have phytoestrogenic properties, traditionally that have been used as a post-partum medicine, to facilitate child birth, regulate menstrual, treat dysentery, flatulence and dysmenorrhoea (Burkhill 1966, Rozihawati et al., 2003). According to Sunarno (2005), seven varieties of *L. pumila* can be found in Malaysia and Indonesia region namely *L. pumila* var *alata*, *L. pumila* var *pumila*, and *L. pumila* var *lanceolata*. *L. pumila* var *discoplacenta*, *L. pumila* var *gladiata*, *L. pumila* var *nerrifolia*, *L. pumila* var *malintangensis* and *L. pumila* var *sessilifolia*. Currently, Kacip Fatimah (*Labisia pumila*) is listed as one of the five main herbs in the Entry Point Project (EPP1) under the 12 agriculture National Key Economic Areas (NKEA) under Malaysia's Economic Transformation Programme (ETP). The other four herbs are Tongkat Ali, Misai Kucing, Hempedu Bumi and Dukung Anak. In 2014, the number of medicinal plants under the EPP1 NKEA under ETP programmed have been increased to 18 plants which included the addition of Pegaga, Mengkudu, Roselle, Mas Cotek, Belalai Gajah, Halia, Peria Katak, Gelenggang, Lempoyang, Sambung Nyawa, Sireh/Kadok, Senduduk and Merunggai (Borang NRGs—A1(R), Herbal Development Office, Ministry of Agriculture & Agro-Based Industries Malaysia). The focus of Agriculture NKEA is to transform small scale herbal production sectors into large scale biobusiness sectors.

Under the Entry Point Project 1 (EPP1), the production of the five herbs including *Labisia pumila* are ensured in order to provide sufficient supply of raw plant material for the purpose of research and development and clinical trials. High value commercialization of herbal based products will take place subsequently. This EPP1 has the aim of producing an income of RM2,213.90 mil by the year 2020 and to project a number of 1822 jobs in Malaysia (Economic Transformation Programme, 2010).

The production of ginseng raw material by field cultivation and bioreactor grown adventitious root were compared previously by Murthy et al (2014). At a slightly higher cost of production of USD47/kg in the bioreactor compared to USD35/kg for field cultivation, the production of plant material was enhanced 57.36-fold with significant reduction of harvesting time from 5 to 7 years in the field compared to a one year in the bioreactor platform.

1.2 Justification

Under the EPP1 project for commercialization of high value herbal products in the Economic Transformation Programme, the cultivation of *Labisia pumila* was produced by cultivation in the field. Currently, the cultivation practices were highly affected by the weather, geographical factors, slow and non-uniform growth. *L. pumila* originated from seeds took 16 weeks to germinate and grow before they could be transferred to growth medium (Ahmad Fauzi, 2013). According to Aminah and Farah Fazwa (2012), due to the slow growth of the plant, the optimum harvesting age for *L. pumila* is 7 to 9 months old which was considered long and not suitable for large scale planting. Furthermore, *L. pumila* whole plants are commonly used in traditional herbal preparations. This leads to the lack of plant material to sustain the demand of the biopharmaceutical industry. Moreover, the secondary metabolites produced from the field-grown plants are present in minute quantities (less than 1%). The differences may be due to the morphological differentiation of field grown plants and tissue cultured plants. Furthermore, hairy root cultures has the ability to synthesize higher amounts of secondary metabolites (Niżyński et al., 2014, Kolewe et al., 2008 and Sauerwin et al., 1992).

Therefore, an alternative approach to planting should be considered to obtain sustainable production for raw plant material of *Labisia pumila* with relatively high content of bioactive secondary metabolites. Plant tissue culture is a clonal propagation method in controlled environment which can provide sustainable supply of *L. pumila* plant material. The utilization of *in vitro* organ cultures such as adventitious root and hairy root cultures can be more beneficial in the long term as these differentiated cultures are genetically stable and can be grown in controlled environment in a bioreactor. Production of specific and desired natural compounds in *L. pumila* can be manipulated in organ cultures to enhance quantity or amount of bioactive content in the herbal plant material grown under standardized conditions. This will indirectly accelerate the product to market time for *L. pumila* herbal products in Malaysia.

Organ cultures developed in a close environment such as bioreactors are needed to ensure sustainable supply of raw materials by the pharmaceutical industry. The physical and biochemical environment can be controlled and manipulated for production of specific compounds of interest.

However, the cultivation of hairy roots in bioreactors needs special considerations as hairy root cultures are sensitive to high shear stress and has tendency to form clumps which hinders the efficient delivery of oxygen to the roots. Different physical and biochemical parameters need to be assessed in order to determine suitable physical and biochemical conditions to cultivate *L. pumila* hairy roots in bioreactor. The biomass of hairy roots and highly specific secondary metabolites can be manipulated under clean and controlled conditions of a bioreactor platform.

Various strategies can be applied such as elicitation, precursor feeding, permeabilization, immobilization, selective absorption, biotransformation and nutrient replenishment to enhance the production of secondary metabolites (Murthy et al., 2014). In this way, continuous production of organ culture biomass and secondary metabolites can be performed at the same time to ensure continuing supply of plant material and phytochemical products to the bio-pharmaceutical industry.

The availability of highly sophisticated and sensitive instruments for the discovery of secondary metabolites in recent years such as Tandem Mass Spectrometry with various detectors such as triple quadrupole, quadrupole ion trap and quadrupole time-of-flight facilitated the discovery and detection of new secondary metabolites in medicinal plants. The process of identification and quantification of secondary metabolites become relatively easier, more accurate and faster.

Since *L. pumila* is an important local herb, and has a high demand in the bio-pharmaceutical industry, a sustainable production of high quality herbal raw material with relatively high amount of bioactive compounds is needed. In this study, sustainable organ cultures (hairy roots and adventitious roots) of *L. pumila* were initiated from leaf with petiole explants. Secondary metabolites' production of various tissues of *L. pumila* were investigated using high end instrumentation. Following that, scaling up of biomass and secondary metabolites production will be performed at lab scale bioreactor platforms.

1.3 Objectives

Therefore, the objectives of this study were

- 1) to establish hairy root and adventitious root cultures of *Labisia pumila* var *alata*,
- 2) to compare the production of secondary metabolites in various tissues of *Labisia pumila* var *alata*, and
- 3) to assess the secondary metabolites produced in the hairy root cultures of *Labisia pumila* var *alata* using bioreactors.

REFERENCES

- Agrofood Statistics 2014. Information Management and Statistics Section, Policy and Strategic Planning Division, Ministry of Agriculture Malaysia : 56
- Ahmad Fauzi MS (2013) Maintenance of Kacip Fatimah at nursery stage. Course of vegetative propagation of Kacip Fatimah, 21-22 March 2013, Mata Ayer, Perlis.
- Ahmad F, Zaidi MAS, Sulaiman N and Abdul Majid FA (2015a) Issues and Challenges in the Development of the Herbal Industry in Malaysia. *Prosiding Perkem* 10 : 227-238.
- Ahmad N, Fatima N, Ahmad I and Anis M (2015b) Effect of PGRs in adventitious root culture *in vitro* : present scenario and future prospects. *Rendiconti Lincei* 26 : 307-321.
- Aird ELH Hamill JD and Rhodes MJC (1988) Cytogenetic analysis of hairy root cultures from a number of plant species transformed by *Agrobacterium rhizogenes* . *Plant Cell Tissue And Organ Culture*. 15 : 47-57.
- Akramian M Tabatabaei MFT and Mirmasoumi M (2008) Virulence of different strains of *Agrobacterium rhizogenes* on Genetic Transformation of Four *Hyascyamus* Species. *American-Eurasian Journal of Agriculture & Environmental Science* 3(5): 759-763.
- Ali M, Abbasi BH and Ali GS (2015) Elicitation of antioxidant secondary metabolites with jasmonates and gibberellic acid in cell suspension cultures of *Artemisia absinthium* L. *Plant Cell, Tissue and Organ Culture* 120 : 1099-1106.
- Ali Z and Khan IA (2011) Alkyl phenols and saponins from the roots of *Labisia pumila* (Kacip Fatimah). *Phytochemistry* 72:2075-2080.
- Allan EJ Eswara JP Jarvis AP Mordue (Luntz) AJ Morgan ED and Stuchbury T (2002) Induction of hairy root cultures of *Azadirachta indica* A. Juss. and their production of azadirachtin and other important insect bioactive metabolites. *Plant Cell Reports*. 21 : 374 – 379.
- Al-Mekhlafi NA, Shaari K, Abas F, Kneer R, Jeyaraj EJ, Stanslas J, Yamamoto N, Honda T and Lajis NH. (2012) Alkenylresorcinols and cytotoxic activity of the constituents isolated from *Labisia pumila*. *Phytochemistry* 80:42-49.
- Aminah H, Naimah CL, Mohd Zaki A and Lokmal N (2008) Rooted leaf cuttings of *Labisia pumila* . *Journal of Tropical Medicinal Plants* 9(1):593-599
- Aminah H and Farah Fazwa MA (2012) Cultivation and growth performance of some selected medicinal plants. Proceedings of 1st Regional Conference

- Al-Qudah TS, Shibli RA and Alali FQ (2011) *In vitro* propagation and secondary metabolites production in wild germander (*Teucrium polium* L.) *In Vitro, Cell and Developmental Biology* 47 : 496-505
- Atansov AG, Waltenberger B, Pferschy-Wenzig E, Linder T, Wawrosch C, Urin P, Temml V, Wang L, Schwaiger S, Heiss EH, Rollinger JM, Schuster D, Breuss JM, Bochkov V, Mihovilovic MD, Kopp B, Bauer R, Dirsch VM and Stuppner H, (2015) Discover and resupply of pharmacologically active plant-derived natural products: A review *Biotechnology Advances* 33: 1582-1614
- Avula B, Wang YH, Ali Z, Smillie TJ and Khan IA (2011) Quantitative Determination of Triterpene Saponins and Alkenated –Phenolics from *Labisia pumila* Using an LC-UV/ELSD Method and Confirmation by LC-ESI-TOF. *Planta Med* 77:1742-1748.
- Bais HP, Venkatesh RT, Chandrashekar A and Ravishankar GA (2001a) *Agrobacterium rhizogenes*-mediated transformation of Witloof chicory-*In vitro* shoot regeneration and induction of flowering. *Current Science* 80(1) : 83-87
- Bais HP, Loyola Vargas VM, Flores HE, Vivanco JM (2001b) Root specific metabolism: the biology and biochemistry of underground organs. *In Vitro Cell and Developmental Biology Plant* 37: 730-741.
- Baíza AM Quiroz-Moreno A Ruíz JA Loyola-Vargas VM (1999) Genetic stability of hairy root cultures of *Datura stramonium*. *Plant Cell, Tissue and Organ Culture* 59(1) : 9-17.
- Banerjee-Chattopadhyay S Schewemmin AM and Schewemmin DJ (1985) A study of karyotypes and their alterations in cultured and *Agrobacterium* transformed roots of *Lycopersicon peruvianum* Mill. *Theoretical and Applied Genetics*. 71(2):258-262.
- Baque MA, Hahn EJ and Paek KY (2010) Growth, secondary metabolite production and antioxidant response of *Morinda citrifolia* adventitious root as affected by auxin and cytokinin. *Plant Biotechnology Report*. 4 : 109-116.
- Baque MA, Moh SH, Lee EJ, Zhong JJ and Paek KY (2011) Production of biomass and useful compounds from adventitious roots of high value added medicinal plants using bioreactors *Biotechnology Advances* 30(2012) : 1255-1267
- Baque MA, Shiragi MHK, Moh SH, Lee EJ and Paek KY (2013) Production of biomass and bioactive compounds by adventitious root suspension cultures of *Morinda citrifolia* (L.) in liquid-phase airlift balloon-type bioreactor. *In Vitro Cell and Developmental Biology* 49 : 737-749.

- Betsui F, Nishikawa NT and Shimomura K (2004) Anthocyanin production in adventitious root cultures of *Raphanus sativus* L. cv. Peking Koushin. *Plant Biotechnology* 21(5):387-391.
- Bloor SJ and Qi L (1994) Cytotoxic saponins from New Zealand *Mrysiine* species. *Journal of Natural Products*. 57 (10) : 1354-1360.
- Blume CL (1823) Catalogous van eenige der Merkwaardigste zoo in – als uitheemsche gewassen , te vinden in's Lands Plantentuin te Buitenzorg, Batavia.
- Blume CL (1826) Bijragen tot de flora van Nederlandsch Indië 13, Lands Drukkerij, Batavia.
- Bolton GW, Nester EW and Gordon MP (1986) Plant phenolic compounds induce expression of the *Agrobacterium tumefaciens* loci needed for virulence. *Science* 232 : 983-985.
- Bonfill M, Malik S, Mirjalili MH, Goleniowski M, Cusido M, Palazón J. (2013) Production and genetic engineering of terpenoids production in plant cell and organ cultures In: *Natural Products: Phytochemistry, Botany and Metabolism of Alkaloids, Phenolics and Terpenes*", eds. Ramawat KG and Mérillon JM, pp. 2761-2796, Springer, Berlin.
- Bonhomme V, Laurain-Mattar D, Fliniaux MA (2000) Effects of the *rolC* gene on hairy roots: induction development and tropane alkaloid production by *Atropa belladonna*. *Journal of Natural Products* 63 : 1249-1252.
- Bordonaro JL and Curtis WR (2000) Inhibitory role of root hairs on transport within root culture bioreactors. *Biotechnology Bioengineering*. 70 : 176-186.
- Bouchez D and Tournier J (1991) Organization of the agropine synthesis region of the *T-DNA* of the *Ri* plasmid from *Agrobacterium rhizogenes*. *Plasmid* 25 : 27-39.
- Bulgakov VP (2008) Functions of *rol* genes in plant secondary metabolism. *Biotechnology Advances*. 26 : 318 – 324.
- Burkhill IH (1966) *A dictionary of the economic products of the Malay peninsula*, 2nd ed. Vol 1, A-H. Government of Malaysia and Singapore
- Bustamante MCC, Cerri MO and Badino AC (2013) Comparison between average stress shear rates in conventional bioreactor with Rushton and Elephant Ear Impeller. *Chemical Engineering Science* 90 : 92-100.
- Camilleri C and Jouanin L (1991) The TR-DNA region carrying the auxin synthesis genes of the *Agrobacterium rhizogenes* agropine-type plasmid pRiA4: nucleotide sequence analysis and introduction into tobacco plants. *Molecular Plant Microbe Interactions* 4:155-162.

- Cardarelli M, Marriotti D, Pomponi M, Spano L, Capone L, and Costantino P. (1987) *Agrobacterium rhizogenes* T-DNA genes capable of inducing hairy root phenotype. *Mol Gen Genet* 209: 475-480.
- Cardozo T and Pagano M (2004) The SCF ubiquitin ligase: insights into a molecular machine. *Nature Reviews Molecular Cell Biology* 5 : 739-751.
- Catapano G, Czermark P, Eibl R, Eibl D and Pförtner D (2009) Equipment design considerations for large scale tissue culture. In *Cell and Tissue Reaction Engineering*. ed. Eibl R, Eibl D, Pförtner D, Catapano G and Czermark P , pp. 21-23. Berlin,Heidelberg: Springer-Verlag.
- Chan LK Ang SW Bhatt A (2010) Elicitation effect on cell biomass and production of alkaloids in cell suspension culture of the tropical tree *Eurycoma longifolia*. *Cuadernos de Investigación UNED* 2 (2) : 239-244.
- Chandra S (2012) Natural plant genetic engineer *Agrobacterium rhizogenes*: role of T-DNA in plant secondary metabolism. *Biotechnology Letters*. 34 :407-415.
- Chandra S and Chandra R (2011) Engineering secondary metabolites production in hairy roots. *Phytochem Rev* 10: 371-395.
- Chaudhury A and Pal M (2010) Induction of Shikonin production in hairy root cultures of *Anerbia hispidissima* via *Agrobacterium rhizogenes*-mediated genetic transformation. *Journal of Crop Science and Biotechnology* 13 (2) : 99-106.
- Chen H Jones AD Gregg AH (2006) Constitutive activation of the jasmonate signaling pathway enhances the production of secondary metabolites in tomato. *FEBS Letters* 580 : 2540-2546.
- Chen CC, Chang HC, Kuo CL, Agrawal DC, Wu CR and Tsay HS (2014) *In vitro* propagation and analysis of secondary metabolites in *Glossogyne tenuifolia* (Hsiang Ju)- a medicinal plant native to Taiwan *Botanical Studies* 55 : 45
- Cheruvathur MK and Thomas TD (2014) Effect of plant growth regulators and elicitors on rhinacanthin accumulation in hairy root cultures of *Rhinacanthus nasutus* (L.) Kurz. *Plant Cell, Tissue and Organ Culture* 118 : 169-177.
- Choi HK, Kim DH, Kim JW, Ngadiran S, Sarmidi MR and Park CS (2010) *Labisia pumila* extract protects skin cells from photoaging caused by UV-B irradiation. *Journal of Bioscience and Bioengineering* 109(3) : 291-296.

- Choi YE, Kim YS and Paek KY (2006) Types and designs of bioreactors for hairy root cultures. In *Plant Tissue Engineering*, ed. Gupta SD and Ibaraki Y, pp. 161-172. Neatherlands: Springer.
- Chua LS, Abdul Latiff N, Lee SY, Lee CT, Sarmidi MR, Abdul Aziz R (2011) Flavonoids and phenolic acids from *Labisia pumila* (Kacip Fatimah). *Food Chemistry* : 127 : 1186-1192.
- Comtrade Database (2015) <http://www.comtrade.un.org/>
- Danial M Chan LK Syed Alwee SSR and Subramaniam S (2012) Hairy root induction from difficult-to-transform pharmacologically important plant *Eurycoma longifolia* using wild strains of *Agrobacterium rhizogenes*. *Journal of Medicinal Plants Research* 6(3) : 479-487.
- Danphitsanuparn P, Boonsnongcheep T, Boriboonkaset T, Chintapakorn Y and Prathanturarug S (2012) Effects of *Agrobacterium rhizogenes* strains and other parameters on production on isoflavonoids in hairy roots of *Pueraria candollei* Grah ex Benth. var. *candollei*. *Plant Cell, Tissue and Organ Culture* 109 (2012) : 491-500.
- De Klerk GJ, Brugge JT and Marinova S (1997) Effectiveness of indole acetic acid, indole butyric acid and naphthaleneacetic acid during adventitious root formation *in vitro* in *Mallus* 'Jork 9'. *Plant Cell, Tissue and Organ Culture* 49 (1) : 39-44.
- Deshpande HA and Bhalsing SR (2014) Isolation and characterization of diosgenin from *in vitro* cultured tissues of *Helicteres isora* L *Physiology and Molecular Biology of Plants* 20(1) : 89-94
- Ditt RF, Kerr KF, De Figueiredo P, Delrow J, Comai L and Nester EW (2006) The *Arabidopsis thaliana* transcriptome in response to *Agrobacterium tumefaciens* . *Molecular Plant Microbe Interactions* 19 (6) : 665-681.
- Dong Y, Gao WY, Man S, Zuo B, Wang J, Huang L and Xiao P (2013) Effect of bioreactor angle and aeration rate on growth and hydromechanics parameters in bioreactor culture of ginseng suspension cells *Acta Physiologiae Plantarum* 35 : 1497-1501.
- Dörnenburg H and Knorr D (1995) Strategies for the improvement of secondary metabolite production in plant cell cultures *Enzyme Microbial Technology*. 17 : 674-684.
- Dougall DK (1980) Nutrition and metabolism In: *Plant Tissue Cultures as a Source of Biochemicals* eds. Staba EJ. Chemical Huibber Company Press, Boca Raton, FL, USA.
- Dupré P, Lacoux J, Neutelings G, Mattar-Laurain D, Fliniaux M-A, David A and Jacquin-Dubreuil A (2000) Genetic transformation of *Ginkgo biloba* by *Agrobacterium tumefaciens* *Physiologia Plantarum* 108 (4) : 413-419.

Economic Transformation Programme (ETP) -
etp.pemandu.gov.my/Agriculture-@-Agriculture_-_EPP_1;High-
Value_Herbal_Products.aspx

Eibl, R and Eibl D (2008) Design of bioreactors suitable for plant cell and tissue cultures. *Phytochemistry Reviews* 7 : 593-598.

Eilert U (1987) Elicitation: methodology and aspects of application. In: Constable F, Vasil IK eds. *Cell Culture and Somatic Cell Genetics Of Plants* Vol. 4 . New York: Academic Press . 153-196.

Estruch JJ, Parets-Soler A, Schmölling T and Spena A (1991) Cytosolic localization in transgenic plants of the *rol C* peptide from *Agrobacterium rhizogenes*. *Plant Molecular Biology* 17 : 547-550.

FAO (2005) Trade in medicinal plants. Raw Materials, Tropical and Horticultural Products Service and Commodities and Trade Division, Economic and Social Development, Food and Agriculture Organization, United Nations.

Farag MA Porzel A Schmidt J and Wessjohann LA (2012) Metabolite profiling and fingerprinting of commercial cultivars of *Humulus lupulus* L. (hop) : comparison of MS and NMR methods in metabolomics. *Metabolomics* 8 : 492 – 507.

Farah Fazwa MA, Ab Rasip AG , Maideen H and Mohamad O(2012) Selection among two varieties of *Labisia pumila* that yield high phenolic contents for establishing plant stock and further cultivation. *Journal of Tropical Medicinal Plants* : 13 (1) : 17-21

Farah Fazwa MA, Norhayati S, Syafiqah Nabilah SB and Mohd Adi Faiz AF (2013) Evaluation of rooting ability of 5 superior genotypes of *Labisia pumila* var *alata* on sand media. *Proceedings of Soils Science Conference of Malaysia 2013, 10-12 April 2012; Renaissance Hotel, Kota Bahru, Kelantan* : 329-333.

Farah Fazwa MA, Norhayati S, Syafiqah Nabilah SB, Mohamad Zaki A and Mohd Asri L (2015) Ensuring sustainability of Kacip Fatimah (*Labisia pumila*) Through Ex-Situ Conservation. *Journal of Tropical Resources Sustainability. Sci* 3 : 43-47.

Fathilah SN, Abdullah S, Mohamed N and Nazrun AS (2012b) *Labisia pumila* prevents complications of osteoporosis by increasing bone strength in a rat model of postmenopausal osteoporosis. *Evidence-Based Complementary and Alternative Medicine*. vol 2012, Article ID 948080, 7 pages.

Fathilah SN, Nazrun AS, Mohamed N, Muhammad N, Soelaiman IN (2012a) *Labisia pumila* protects the bone of estrogen-deficient rat model: A

- histomorphometric study. *Journal of Ethnopharmacology* 142 : 294-299.
- Fattahi M, Nazeri V, Torras-Claveria L, Sefidkon F, Cusido RM, Zamani Z and Palazon J (2013) A new biotechnological source of rosmarinic acid and surface flavonoids: Hairy root cultures of *Dracocephalum kotschy* Boiss. *Industrial Crops and Products* 50 : 256-263.
- Fazliana M, WanNazaimoon W, Gu HF and Östenson CG (2009) *Labisia pumila* extracts regulates body weight and adipokines in ovariectomized rats. *Maturitas* 62 : 91-97.
- Fazliana M, Gu HF, Östenson CG, Mashitah MY and WanNazaimoon WM (2012) *Labisia pumila* extract down-regulates hydroxysteroid (11-beta) dehydrogenase expression and corticosterone levels in ovariectomized rats. *Journal of Natural Medicine* 66 : 257-264.
- Fernandez SP, Nguyen M, Yow TT, Chu C, Johnston GAR, Hanrahan JR and Chebib M (2009) The flavonoid glycosides, myricitrin, gossipin and naringin exert anxiolytic action in mice" *Neurochemistry Research* 34: 1867-1875
- Foubert K, Theunis MHL, Apers S, Vlietinck A and Pieters L (2012) Chemistry, distribution, and biological activities of 13,28, epoxy-oleanane saponins from the plant families Myrsinaceae and Primulaceae *Current Organic Chemistry* 12(2008) : 629-642
- Fu CX, Zhao DX, Xue, XF, Jin, ZP and Ma, FS (2005) Transformation of *Saussurea involucreata* by *Agrobacterium rhizogenes*: Hairy root induction and syringin production. *Process Biochemistry* 40:3789-3794.
- Furmanowa M and Sykloska-Baranek K (2000) Hairy root cultures of *Taxus × media* var. *Hicksii* Rehd. as a new source of paclitaxel and 10-deacetylbaicatin III. *Biotechnology Letters*. 22 : 683 – 686.
- Gai QY, Jiao J, Luo M, Wang W, Ma W, Zu YG and Fu YJ (2014) Establishment of high-productive *Isatis tinctoria* L. hairy root cultures: A promising approach for efficient production of bioactive alkaloids. *Biochemical Engineering Journal* 95(2015) : 37-47
- Gangopadhyay M, Sircar D, Mitra A and Bhattacharya S (2008) Hairy root culture of *Plumbago indica* as a potential source for plumbagin. *Biologia Plantarum* 52 (3): 533-537.
- Gangopadhyay M, Dewanjee S and Bhattacharya S (2011) Enhanced plumbagin production in elicited *Plumbago indica* hairy root cultures. *Journal of Bioscience and Bioengineering*. 111(6):707-210.
- Gelvin SB (2000) *Agrobacterium* and plant genes involved in T-DNA transfer and integration. *Annual Reviews of Plant Physiology and Plant Molecular Biology*. 51:223-256.

- Gelvin SB (2010) Plant Proteins Involved in *Agrobacterium*-Mediated Genetic Transformation. *Annual Review of Phytopathology*. 48 : 45-68.
- Georgiev MI, Pavlov AI and Bley T (2007) Hairy root type plant in vitro systems as sources of bioactive substances. *Applied Microbial Biotechnology* 74(6):1175–1185
- Georgiev MI, Eibl R and Zhong JJ (2013) Hosting the plant cells *in vitro* : recent trends in bioreactors. *Applied Microbial Biotechnology* 97 : 3787-3800.
- Global Industry Analysts Inc, USA - Herbal Supplements and Remedies Market Trends(http://www.strategyr.com/MarketResearch/Herbal_Supplements_and_Remedies_Market_Trends.asp)
- Gromski PS, Muhamadali H, Ellis DI, Xu Y, Correa E, Turner M and Goodacre R (2015) A tutorial review: Metabolomics and partial least squares-discriminant analysis- a marriage of convenience or a shotgun wedding. *Analytica Chimica Acta* 879 : 10-23.
- Guo X, Yue Y, Tang F, Wang J, Yao X and Sun J (2013a) A comparison of C-glycosidic flavonoid isomers by electrospray ionization quadrupole time-of-flight tandem mass spectrometry in negative and positive ion mode. *International Journal of Mass Spectrometry* 333 : 59-66.
- Guo N, Ablajan K, Fan B, Yan H, Yu Y and Dou D (2013b) Simultaneous determination of seven ginsenosides in Du Shen Tang decoction by rapid resolution liquid chromatography (RRLC) coupled with tandem mass spectrometry. *Food Chemistry* 141 (4) : 4046-4050.
- Gupta R, Pandey P, Singh S, Singh DK, Saxena A, Luqman S, Bawankule DU and Banerjee S (2015) Advances in *Boerhaavia diffusa* hairy root technology: a valuable pursuit for identifying strain sensitivity and up-scaling factors to refine metabolite yield and bioactivity potentials. *Protoplasma* 253(4) : 1145-1158.
- Han B, Linden JC, Gujarathi NP and Wickramshinghe SR (2004) Population balance approach to modelling hairy root growth. *Biotechnology Progress* 20 : 872-879.
- Hartinie M and Jualang GA (2007) *In vitro* germination and plantlet establishment of *Labisia pumila* (B1.)F. Vill. *Scientia Horticulturae*. 115: 91-97.
- Herbal Development Office, Ministry of Agriculture & Agro-Based Industries, Malaysia. <http://www.moa.gov.my/en/industri-herba>
- Hisham MN , Mohd Lip J, Mohd Noh J , Normah A and Nurul Nabilah MF (2011) Identification and isolation of methyl galate as a polar chemical marker for *Labisia pumila* Benth. *Journal of Tropical Agriculture and Food Sciences* 39(2) : 279-284.

- Ho CS Lam CWK Chan MHM Cheung RCK Law LK Lit LCW Ng KF Suen MWM and Tai HL (2003) Electrospray Ionisation Mass Spectrometry: Principles and Clinical Applications. *Clin Biochem Rev* (24) : 1-10
- Hong S, Peebles C, Shanks, JV, San KY and Gibson SI (2006) Terpenoid indole alkaloid production by *Catharanthus roseus* hairy roots induced by *Agrobacterium tumefaciens* harboring *rolABC* genes. *Biotechnology Bioengineering*. 93 : 386-390.
- Hu, ZB and Du M (2006) Hairy root and its application in plant genetic engineering. *Journal of Integrative Plant Biology*. 48(2):121-127.
- Husniza H (2002) Estrogenic and Androgenic Activities of Kacip Fatimah (*Labisia pumila*). Institute of Medical Research, Ministry of Health Malaysia, Kuala Lumpur (Abstracts of Research Projects, pp. 8).
- Hussien S, Ling APK, Ng TZ, Ibrahim R and Paek KY (2012) Adventitious roots induction of recalcitrant tropical woody plant, *Eurycoma longifolia*. *Romanian Biotechnological Letters* 17 (1) : 7026-7035.
- Ibrahim MH and Jaafar HZE (2011) Photosynthetic capacity, photochemical efficiency and chlorophyll content of three varieties of *Labisia pumila* Benth. exposed to open field and greenhouse growing conditions. *Acta Physiol Plant* 33 : 2179-2185.
- Ibrahim MH and Jaafar HZE (2012) Reduced photoinhibition under low irradiance enhanced Kacip Fatimah (*Labisia pumila* Benth) secondary metabolites, phenyl alanine lyase and antioxidant activity *International Journal of Molecular Sciences* 13: 5290-5306
- Inoguchi M, Ogawa S, Furukawa S and Kondo H (2003) Production of an allelopathic polyacetylene in hairy root cultures of goldenrod (*Solidago altissima* L.). *Bioscience Biotechnology Biochemistry* 67:(4):863-868.
- Jamia, AJ, Houghton, JP and Milligan, RS (1998). Testing of *Labisia pumila* for oestrogenic properties using a recombinant yeast screen. *Journal of Pharmacy and Pharmacology* 50 : 79.
- Jamia, AJ, Houghton, JP, Milligan, RS., Jantan, I (2003). The oestrogenic and cytotoxic effects of the extracts of *Labisia pumila* var. *alata* and *Labisia pumila* var. *pumila* in vitro. *Jurnal Sains Kesihatan* 1, 53–60.
- Jansakul C, Baumann H, Kenne L and Samuelsson G (1987) Ardisicrispin A and B, two utero-contracting saponins from *Ardisia crispa*. *Planta Medica* 53 : 405-409.
- Jantan I (2004) Medicinal Plant Research in Malaysia: Scientific Interests and Advances. *Jurnal Sains Kesihatan Malaysia* 2 (2) : 24-26
- Jaremicz Z, Luczkiewicz M, Kokotkiewicz A, Krocicka A and Sowinski P (2014) Production of tropane alkaloids in *Hyoscyamus niger* (black henbane)

- hairy roots grown in bubble-column and spray bioreactors. *Biotechnology Letters* 36 : 843-853.
- Jenifer U, Francina Cecilia K and Ravindhran R (2012) *In vitro* adventitious root and hairy root cultures in *Boerhaavia diffusa* L. *International Journal of Current Research*. 4(1) : 65-67.
- Jeong GT and Park DH (2006) Characteristics of transformed *Panax ginseng* C.A. Meyer hairy roots: growth and nutrient profile. *Biotechnology and Bioprocess Engineering* 11 : 43-47.
- Jeong JA, Wu CH, Murthy HN, Hahn EJ and Paek KY (2009) Application of an airlift bioreactor system for the production of adventitious root biomass and caffeic acid derivatives of *Echinacea purpurea*. *Biotechnology and Bioprocess Engineering* 14 : 91-98.
- Jia Z, Koike K, Ohmoto T and Ni M (1994) Triterpenoid saponins from *Ardisia crenata*. *Phytochemistry* 37 (5) : 1389-1396.
- Kajani AA Moghim S Mofid MR (2012) Optimization of the basal medium for improving production and secretion of taxanes from suspension cell culture of *Taxus baccata* L. *DARU Journal of Pharmaceutical Sciences* 20 : 54.
- Kanokwaree K and Doran PM (1997) Effect of inoculum size on growth of *Atropa belladonna* hairy roots in shake flasks *Journal of Fermentation and Bioengineering* 84 : 378-381
- Karimi E and Jaafar HZE (2011) HPLC and GC-MS Determination of Bioactive Compounds in Microwave Obtained Extracts of Three Varieties of *Labisia pumila* Benth. *Molecules* 16 : 6791-6805.
- Karimi E, Jaafar HZE and Ahmad S (2011) Phytochemical analysis and antimicrobial activities of methanolic extracts of leaf, stem and root from different varieties of *Labisia pumila* Benth. *Molecules* 16 : 4438-4450.
- Kastell A Smetanska I Ulrichs C Cai Z and Mewis I (2013a) Effects of Phytohormones and Jasmonic Acid on Glucosinolate Content in Hairy Root Cultures of *Sinapsis alba* and *Brassica rapa* . *Appl Biochem Biotechnol*. 169 : 624 – 635.
- Kastell A, Smetanska I, Schreiner M and Mewis I (2013b) Hairy roots, callus, and mature plants of *Arabidopsis thaliana* exhibit distinct glucosinolate and gene expression profiles *Plant Cell, Tissue and Organ Culture* 115 : 45-54
- Kavitha G, Taghipour F and Huyop F (2010) Investigation of factors optimizing *Agrobacterium*-mediated gene transfer in *Citrullus lanatus* cv. Round Dragon. *Journal of Biological Sciences* 2010 : 1-8.

- Kevers C Jacques P, Thonart P, Gaspar T (1999) *In vitro* root cultures of *Panax ginseng* and *P. quinquefolium*. *Plant Growth Regulation*. 27: 173-178.
- Kim Y (2001) Assessment of bioreactors for transformed root cultures. PhD Thesis, Worcester Polytechnic Institute, Worcester MA.
- Kim Y, Wyslouzil BE and Weathers PJ (2001) A comparative study of mist and bubble column reactors in the *in vitro* production of artemisinin. *Plant Cell Reports*. 20 : 451-455.
- Kim YJ, Weathers PJ and Wyslouzil BE (2002a) Growth of *Artemisia annua* hairy roots in liquid and gas phase reactors. *Biotechnology Bioengineering*. 80 : 454-464.
- Kim YJ, Wyslouzil BE and Weather PJ (2002b) Invited Review: Secondary metabolism of hairy root cultures in bioreactors. *In Vitro Cell and Developmental Biology-Plant* 38 : 1-10.
- Kim OT, Manickavasagam M, Kim YJ, Jin MR, Kim KS, Seong NS and Hwang B (2005) Genetic transformation of *Ajuga multiflora* Bunge with *Agrobacterium rhizogenes* and 20-Hydroxycyclopentanone production in hairy roots. *Journal of Plant Biology* 48(2):258-262
- Kim OT, Bang KH, Shin YS, Lee MJ, Jung SJ, Hyun DY, Kim YC, Seong NS, Cha Sw, Hwang B (2007) Enhanced production of asiaticoside from hairy root cultures of *Centella asiatica* (L.) Urban elicited by methyl jasmonate. *Plant Cell Rep* 26:1941-1949.
- Kim OT, Yoo NH, Kim GS, Kim YC, Bang KH, Hung, DY, Kim SH and Kim MY (2013) Stimulation of Rg3 ginsenoside biosynthesis in ginseng hairy roots elicited by methyl jasmonate, *Plant Cell, Tissue and Organ Culture*. 112: 87-93.
- Kobayashi K, Ihara S, Kobota A, Itoh K, Kusunoki N and Yoshizaki F (2008) Inhibitory effect of *Myrica* bark on lipase activity in mouse plasma and gastrointestinal tract. *Journal of Medicinal Food* 11 : 289-293
- Kolewe ME, Gaurav V and Roberts SC (2008) Pharmaceutically active natural product synthesis and supply via plant cell culture technology *Molecular Pharmaceutics* 5 : 243-256
- Kumar V, Sharma A, Parsad BCN, Gururaj HB and Ravishankar GA (2006) *Agrobacterium rhizogenes*-mediated genetic transformation resulting in hairy root formation is enhanced by ultrasonication and acetosyringone treatment. *Electronic Journal of Biotechnology* 9 (4) : 349-357.
- Lacroix B and Citovsky V (2013) The roles of bacterial and host plant factors in *Agrobacterium*-mediated genetic transformation. *International Journal of Developmental Biology* 57: 467-481.

- Le Goff L and Beljanski M (1985) The *in vitro* effects of opines and other compounds on DNAs originating from bacteria, and from healthy and tumorous plant tissues. *Pathobiology*. 53 (6) : 335-350.
- Lee SY Xu H Kim YK and Park SU (2008) Rosmarinic acid production in hairy root cultures of *Agastache rugosa* Kuntze. *World J Microbiol Biotechnol* 24 : 969 – 972.
- Lei Z Huhman DV and Sumner LW (2011) Mass spectrometry strategies in metabolomics. *The Journal of Biological Chemistry*. 286 (29) : 25435-25442
- Li SW, Xue L, Xu S, Feng H, An L (2009) Mediators, genes and signalling in adventitious rooting. *The Botanical Reviews* 75 : 230-247.
- Li Y, Shao CH, Park SY, Piao XC and Lian ML (2014) Production of salidroside and polysaccharide in *Rhodiola sachalinensis* using airlift bioreactor systems. *Acta Physiologiae Plantarum* 36 : 2975-2983.
- Li J, Wang J, Li J, Liu D, Li H, Gao W, Li J and Liu S (2015) *Aspegillus niger* enhance bioactive compounds biosynthesis as well as expression of functional genes in adventitious roots of *Glycyrrhiza uralensis* Fisch. *Applied Biochemistry and Biotechnology* 178 (3) : 576-593.
- Libbenga KR and Mennes AM (1995) Hormone binding and signal transduction. In *Plant Hormones* , Kluwer Academic Publishers, Dordrecht, pp. 272-297.
- Ling APK, Tan KP. and Hussein S (2013) Comparative effects of plant growth regulators on leaf and stem explants of *Labisia pumila* var. *alata*. *Journal of Zhejiang University-SCIENCE B (Biomedicine & Biotechnology)*. 14(7): 621-631.
- Ma L, Li W, Wang H, Kuang X, Li Q, Wang Y, Xie P and Koike (2014) A simple and rapid method to identify and quantitatively analyse triterpenoid saponins in *Ardisia crenata* using electrospray ionization quadruple mass spectrometry *Journal of Pharmaceutical and Biomedical Analysis* 102(2015) : 400-408
- Madeira and Florêncio. Applications of tandem mass spectrometry: from structural studies to fundamental Studies. In *Tandem Mass Spectrometry – Application and Principles*. 2012. In Tech. ,pp 5-33.
- Mamun NHA, Egertsdotter U and Aidun CK (2015) Bioreactor technology for clonal propagation of plants and metabolite production. *Frontiers in Biology* doi 10.1007/s11515-015-1355-1.
- Mannerås L, Fazliana M, WanNazaimoon W, Lonn M, Gu MF, Ostenson CG and Stener-Victorin E (2010) Beneficial metabolic effects of the Malaysian herb *Labisia pumila* var *alata* in rat model of polycystic ovary syndrome. *Journal of Ethnopharmacology* 127 (2) : 346-351.

- Manuhara YSW, Kristanti AN, Utami ESW and Yachya A (2015) Effect of sucrose and potassium nitrate on biomass and saponin content of *Talinum paniculatum* Gaertn. hairy root in balloon-type bubble bioreactor. *Asian Pacific Journal of Tropical Biomedicine* 5 (12) : 1027-1032.
- Matkowski A (2008) Plant *in vitro* culture for the production of antioxidants- A review *Biotechnology Advances* 26 : 548-560.
- Matsuda H, Higashino M, Chen W, Tosa H, Iinuma M and Kubo M (1995) Studies of cuticle drugs from natural sources. III. Inhibitory effect of *Myrica rubra* on melanin biosynthesis. *Biological and Pharmaceutical Bulletin* 18 (8) :1148-1150
- Matveeva TV, Sokornova SV and Lutova LA (2015) Influence of *Agrobacterium* oncogenes on secondary metabolism in plants. *Phytochemistry Reviews* 14 : 541-554.
- Maurizio T, Bruno M, Francisco L and Paolo C (2001) The plant oncogene *rolD* encodes a functional ornithine cyclodeaminase. *Plant Biology*. 98 (23) : 13449-13453.
- Mehrotra S, Kukreja AK, Khanuja SPS and Mishra BN (2008) Genetic transformation studies and scale up in hairy root culture of *Glycyrrhiza glabra* in bioreactor. *Electronic Journal of Biotechnology* 11 (2), available online <http://www.ejbiotechnology.info/content/vol11/issue2/full/6/>
- Meijer JJ, Hoopen HJG and Libbenga KR (1993) Effects of hydrodynamic stress on cultured plant cell : a literature survey, *Enzyme and Microbial Technology* 15 : 234-238.
- Milman B (2015) General principles of identification by mass spectrometry. *Trends in Analytical Chemistry*. 69: 24-33.
- Mishra BN and Ranjan R (2008) Growth of hairy-root cultures in various bioreactors for the production of secondary metabolites. *Biotechnology and Applied Biochemistry*. 49 : 1-10.
- Mishra J, Bhandari H, Singh M, Rawat S, Agnihotri RK, Mishra S and Purohit S (2011) Hairy root cultures of *Picrorhiza kurroa* Royle ex Benth: a promising approach for the production of picrotin and picrotoxinin. 33 : 1841 – 1846.
- Mishra S, Sangwan RS, Bansal S and Sangwan NS (2012) Efficient genetic transformation of *Withania coagulans* (Stocks) Dunal mediated by *Agrobacterium tumefaciens* from leaf explants of *in vitro* multiple shoot culture. *Protoplasma* doi. 10.1007/s00709-012-0428-0.
- Murthy HN, Lee EJ and Paek KY (2014) Production of secondary metabolites from cell and organ cultures: strategies and approaches for biomass

- improvement and metabolite accumulation. *Plant Cell Tissue and Organ Culture*. 118 : 1-16.
- Murthy HN, Dandin VS and Paek KY (2016) Tools for biotechnological production of useful phytochemicals from adventitious root cultures. *Phytochemistry Reviews* 15 (1) : 129-145.
- Musharraf SG Goher M Ali A Adhikari A Choudhary MI and Rahman A (2012) Rapid characterization and identification of steroidal alkaloids in *Sarcococca coriacea* using liquid chromatography coupled with electrospray ionization quadropole time-of-flight mass spectrometry. *Steroids* 77 : 138-148.
- Nagella P, Thiruvengadam M, Jung SJ, Murthy HN and Chung IM (2013) Establishment of *Gymnema sylvestre* hairy root cultures for the production of gymnemic acid, *Acta Physiol Plant* 35 : 3067-3073.
- Nandagopal S, & Ranjitha KBD (2007) Effectiveness of auxin induced *in vitro* root culture in Chicory. *Journal of Central European Agriculture* 8(1) : 73-80.
- National Pharmaceutical Control Bureau, Malaysia.
- Nazrun AS, Leong LP, Muhammad N, Mohamed N and Soelaiman IN (2011) The effects of *Labisia pumila* var. *alata* on bone markers and bone calcium in a rat model of post-menopausal osteoporosis. *Journal of Ethnopharmacology* 133 : 538-542.
- Neumann S and Böcker S (2010) Computational mass spectrometry for metabolomics: Identification of metabolites and small molecules. *Anal Bioanal Chem* 398 : 2279-2288.
- Nguyen C., Bourgaud F., Forlot P., and Guckert A. (1992) Establishment of hairy root cultures of *Psoralea* species. *Plant Cell Reports*. 11 : 424-427
- Nguyen THY, Trinh AV, Phan VK, Chau VM, Nguyen XN, Pham QL, Luu TA, Kim N, Park SJ, Kim SH (2015) Chemical components from the leaves of *Ardisia insularis* and their cytotoxic activity *Archives of Pharmacol Research* 38: 1926-1931
- Niżyński B, Alsoufi ASM, Pączkowski C, Długosz M and Szakiel A (2014) The content of free and esterified triterpenoids of the native marigold (*Calendula officinalis*) plant and its modification in *in vitro* cultures *Phytochemistry Letters* 11(2015) : 410-417
- Ofringa IA, Melchers LS, Regensburg-Tuink AJG, Costantino P, Schilperoort RA and Hooykaas PJJ (1986) Complementation of *Agrobacterium tumefaciens* tumor-inducing *aux* mutants by genes from the TR-region of the *Ri* plasmid of *Agrobacterium rhizogenes*. *Proceedings of The National Academy of Sciences, USA* 83 : 6935-6939

- Okršlar V Štrukelj B Kreft S Bohanec B and Jana Z (2002) Micropropagation and hairy root culture of *Solanum laginiatum* Ait. In Vitro Cell Dev Biol – Plant . 38 : 352-357.
- Onrubia M Moyano E Bonfill M Cusidó RM Goossens A Palazón J (2013) Coronatine , a more powerful elicitor for inducing taxane biosynthesis in *Taxus media* cell cultures than methyl jasmonate. Journal of Plant Physiology 170 : 211 – 219.
- Paek KY, Murthy HN, Hahn EJ and Zhong JJ (2009) Large scale culture of ginseng adventitious roots for production of ginsenosides. *Advances of Biochemistry Engineering Biotechnology* 113 : 151-176
- Pal A, Swain SS, Mukherjee AK and Chand PK (2013) *Agrobacterium pRi TL-DNA rolB* and *TR-DNA* opine genes transferred to spiny amaranth (*Amaranthus spinosus* L), a nutraceutical crop. *Food Technology Biotechnology* 51 (1) : 26-35.
- Palazón J Cusidó RM Roig C and Piñol MT.(1997) Effect of *rol* genes from *Agrobacterium rhizogenes* TL-DNA on nicotine production in tobacco root culture. *Plant Physiol Biochem.* 35 : 155-162.
- Palazón J, Cusidó RM, Roig M and Piñol T (1998) Expression of the *rol C* gene and nicotine production in transgenic roots and their regenerated plants. *Plant Cell Reports* 17 (3) : 84-90.
- Palazón J, Cusidó RM, Bonfill M, Mallol A, Moyano E, Morales C and Pinol MT (2003) Elicitation of different *Panax ginseng* transformed root phenotypes for an improved ginsenoside production. *Plant Physiology and Biochemistry* 41: 1019-1025.
- Pandey P, Kaur R, Singh S, Chattopadhyay SK, Srivastava SK and Banerjee S (2014) Long term stability in biomass and production of triterpene indole alkaloids by hairy root culture of *Rauvolfia serpentina* and cost approximation to endorse commercial realism. *Biotechnology Letters* 38 (7) : 1523-1528
- Pandey V , Srivastava R, Akhtar N, Mishra J, Mishra P and Verma PC (2015) Expression of *Withania somnifera* steroidal glucosyltransferase gene enhances withanolide content in hairy roots. *Plant Molecular Biology Report* doi 10.1007/s11105-015-0955-x
- Pandolfini T, Storlazzi A, Calabria E, Defez R and Spena A (2000) The spliceosomal intron of the *rolA* gene of *Agrobacterium rhizogenes* is a prokaryotic promoter. *Molecular Microbiology* 35 : 1326-1334.
- Park NI, Park JH, Lee CY, Lee SY and Park SU (2010) *Agrobacterium rhizogenes*-mediated transformation of β -glucuronidase reporter gene in hairy roots of *Angelica gigas* Nakai, *Plant Omics Journal* 3(4) : 115-120.

- Park YJ, Thwe AA, Li X, Kim YJ, Kim JK, Mariadhas VA, Al-Dhabi NA and Park SU (2015) Triterpene and flavonoid biosynthesis and metabolic profiling of hairy roots, adventitious roots, and seedling roots of *Astragalus membranaceus*. *Journal of Agricultural and Food Chemistry* 63(40) : 8862-8869.
- Patra N and Srivastava AK (2014) Enhanced production of Artemisinin by hairy root cultivation of *Artemisia annua* in a modified stirred tank reactor. *Applied Biochemistry and Biotechnology* 174 : 2209-2222.
- Patra N and Srivastava AK (2015) Artemisinin production by plant hairy root cultures in gas- and liquid-phase bioreactors. *Plant Cell Reports*.
- Perez JAS, Porcel EMR, Lopez JLC, Sevilla JMF, Chisti Y (2006) Shear rate in stirred-tank and bubble column bioreactors. *Chemical Engineering Journal* 24 : 1-5
- Pillai DB, Jose B, Satheeshkumar K and Krishnan PN (2015) Optimization of inoculum density in hairy root culture of *Plumbago rosea* L. for enhanced growth of plumbagin production towards scaling-up in bioreactor. *Indian Journal of Biotechnology* 14 : 264-269.
- Poh SWM, Navaratnam V and Chia YY (2013) Estrogenic assessment of *Labisia pumila* extracts using a human endometrial cell line. *International Journal of Pharmacy and Pharmaceutical Sciences*. 5 (2) : 448-452.
- Praveen N and Murthy HN (2010) Production of withanolide-A from adventitious root cultures of *Withania somnifera*. *Acta Physiologiae Plantarum* 32 : 1017-1022.
- Ramakrishna A and Ravishankar GA (2011) Influence of abiotic stress signals on secondary metabolites in plants *Plant Signalling and Behaviour* vol. 6, pp. 1720-1731.
- Rao SR and Ravishankar GA (2002) Plant cell cultures: chemical factories of secondary metabolites" *Biotechnology Advances* 20 : 101-153
- Roberts SC and Shuler ML (1998) Strategies for Bioproduct Optimization in Plant Cell Tissue Cultures, In: *BioHydrogen* eds. Zaborsky OR, Benemann JR, Matsunaga T, Miyake J and SanPietro A, Springer US, New York
- Rozihawati Z, Aminah H, Lokman N (2003) Preliminary trial on rooting ability of *Labisia pumila* cuttings. In: Malaysian Science & Technology Congress. Agriculture Science, Cititel, Midvalley, Kuala Lumpur. 2003.
- Rozihawati Z and Aminah H (2005) Vegetative propagation of *Labisia pumila* through stem cuttings. *Institut Penyelidikan Perhutanan FRIM* : 368-373.

- Sauerwein M Yamazaki T Shimomura K (1991) Hernandulcin in hairy root cultures of *Lippia dulcis*. *Plant Cell Reports*. 9 : 579-581.
- Sauerwein M, Yoshimatsu K and Shimomura K (1992) Further approaches in the production of secondary metabolites by plant tissue cultures *Plant Tissue Culture Letters* 9 : 1-9
- Saleh NM and Thuc LV (2009) Assessment of hairy roots induction in *Solenostemon scutellarioides* leaves by different strains of *Agrobacterium rhizogenes*. *African Journal of Biotechnology*. 8(15):3519-3523.
- Saxena G, Banerjee S, Rahman L, Verma PC, Mallavarapu GR, Kumar (2007) Rose-scented geranium (*Pelargonium* sp.) generated by *Agrobacterium rhizogenes* mediated Ri-insertion for improved essential oil quality. *Plant Cell Tissue and Organ Culture* 90:214-223.
- Sharafi A, Sohi HH, Mousavi A, Azadi P, Razavi K and Ntui VO (2013) A reliable and efficient protocol for inducing hairy roots in *Papaver bracteatum*. *Plant Cell Tissue and Organ Culture* 113:1-9.
- Sharafi A, Sohi HH, Azadi P and Sharafi AA (2014) Hairy root induction and plant regeneration of medicinal plant *Dracocephalum kotschy*. *Physiology and Molecular Biology of Plants* 20:(2) : 257-262.
- Sharma C, Kumari T, Pant G, Bajpai V, Srivastava M, Mitra K, Kumar B and Arya KR (2015) Plantlet formation via somatic embryogenesis and LC ESI Q-TOF MS determination of secondary metabolites in *Butea monosperma* (Lam.) Kuntze” *Acta Physiologiae* 37 : 239
- Sharma P Padh H and Shrivastava N (2013) Hairy root cultures: A suitable biological system for studying secondary metabolic pathways in plants. *Eng. Life Sci* 13(1) : 62-75.
- Shiao TL and Doran PM (2000) Root hairiness: effect on fluid flow and oxygen transfer in hairy root cultures. *Journal of Biotechnology* 83 : 199-210.
- Shilpha J, Satish L, Kavikkuil M, Largia MJV and Ramesh M (2015) Methyl jasmonate elicits the solasodine production and antioxidant activity in hairy root cultures of *Solanum trilobatum* L. *Industrial Crops and Products* 71 (2015) : 54-64.
- Shimosaki S, Tsurunaga Y, Itamura H and Nakamura M (2011) Anti-allergic effects of the flavonoid myricitrin from *Myrica rubra* leaf extracts *in vitro* and *in vivo*” *Natural Product Research* 25 (4) : 374-380
- Shimura S, Tsuzuki W, Kobayashi S and Suzuki T (1992) Inhibitory effect on lipase activity of extracts from medicinal herbs. *Bioscience, Biotechnology and Biochemistry* 56 : 1478-1479.

- Shin KS, Murthy HN, Ko Jy and Paek Ky (2002) Growth and betacyanin production by hairy roots of *Beta vulgaris* in airlift bioreactors. *Biotechnology Letters* 24 : 2067-2069.
- Shkryl YN, Veremeichik GN, Bulgakov VP, Tchernoded GK, Mischenko NP, Fedoreyev SA and Zhuravlev YN (2008) Individual and combined effects of the *rol A*, *B* and *C* genes on anthraquinone production in *Rubia cordifolia* transformed calli. *Biotechnology Bioengineering* 100 : 118-125.
- Shohael AM, Murthy HN and Hahn EJ (2007) Methyl jasmonate induced overproduction of eluetherosides in somatic embryos of *Eleutherococcus senticosus* cultured in bioreactors" *Electronic Journal of Biotechnology* 10(4) : 634-637
- Siahsar B, Rahimi M, Tavassoli A and Raissi A (2011) Application of biotechnology in production of medicinal plants *American-Eurasian Journal of Agriculture and Environmental Sciences*. 11 : 439-444.
- Simões C, Albarello N, Callado CH, de Castro TC and Mansur E (2009) New approaches for shoot production and establishment of *in vitro* root cultures of *Cleome rosea* Vahl. *Plant Cell, Tissue and Organ Culture* 98: 79-86.
- Singh G and Curtis WR (1994) Reactor design for plant root culture. In: *Biotechnological applications plant cultures. CRC series of current topics in plant molecular biology*. ed. Shargool PD and Ngo TT, pp 185-206, FL: CRC Press : Boca Raton
- Sivanadhan G Dev GK Jeyaraj M Rajesh M Arjunan A Muthuselvam M Macnickavasagam M Selvaraj N and Ganapathi A (2013) Increased production of withanolide A, withanone, and withaferin A in hairy root culutres of *Withania somnifera* (L.) Dunal elicited with methyl jasmonate and salicylic acid. *Plant Cell Tissue and Organ Culture* 114 : 121 – 129.
- Skala E, Kicel A, Olszewska MA, Kiss AK and Wysokińska H (2015) Establishment of hairy root cultures of *Rhaponticum carthomoides* (Willd.) for the production of biomass and caffeic acid derivatives. *BioMed Research International* 2015 : Article ID 181098, 11 pages
- Spena A, Schmülling T, Koncz C and Schell JS (1987) Independent and synergistic activity of *rol A*, *B* and *C* loci stimulating abnormal growth in plants. *The EMBO Journal* 6(13) :3891-3899
- Srivastava S and Srivastava AK (2012) Azadirachtin production by hairy root cultivation of *Azadirachta indica* in a modified stirred tank reactor. *Bioprocess and Biosystems Engineering*.

- Srivastava S and Srivastava AK (2013) Production of biopesticide azadirachtin by hairy root cultivation of *Azadirachta indica* in liquid-phase bioreactors. *Applied Biochemistry and Biotechnology*. 171 : 1351-1361.
- Srivastava V Kaur R Chattopadhyay SK Banerjee S (2013) Production of industrially important cosmaceutical and pharmaceutical derivatives of betuligenol by *Atropa belladonna* hairy root mediated biotransformation. *Industrial Crops and Products* 44 : 171 – 175.
- Stachel SE, Messens E, VanMontagu M and Zambryski IP (1985) Identification of the signal molecules produced by the wounded plant cells that activate T-DNA transfer in *Agrobacterium tumefaciens*. *Nature* 318 : 624-629.
- Sudha CG, Sherina TV, Anand VPA, Reji JV, Padmesh P and Soniya EV (2013) *Agrobacterium rhizogenes* mediated transformation of the medicinal plant *Decalepis arayalpathra* and production of 2-hydroxy-4-methoxy benzyldehyde. *Plant Cell Tissue and Organ Culture* 112 : 217-226.
- Sujatha G Zdravković-Korać S Čalić D Flamini G Ranjitha Kumari BD (2013) High efficiency *Agrobacterium rhizogenes*-mediated genetic transformation in *Artemisia vulgaris*: Hairy root production and essential oil analysis. *Industrial Crops and Products* 44 : 643-652.
- Sun C, Huang H, Xu C, Xian L and Chen K (2013) Biological activities of extracts from chinese bayberry (*Myrica rubra* Sieb. et Zucc.) : A Review *Plant Foods For Human Nutrition* 68 : 97-106.
- Sunarno B (2005) Revision of the genus *Labisia* (Mrysinaceae) *Blumea* 50 : 579-597.
- Sung LS, Huang SY (2006) Lateral root bridging as a strategy to enhance L-DOPA production in *Stizolobium hassjoo* hairy root cultures by using a mesh hindrance mist trickling bioreactor. *Biotechnology Bioengineering* 94(3):441–447.
- Syafiqah Nabilah SB, Farah Fazwa MA, Norhayati S, Mohd Zaki A and Mohamad O(2014) Propagation of five high yielding clones of *Labisia pumila* var *alata* and evaluation of their growth performances at nursery stage. *International Journal of Agricultural Science Research* 3(7):121-125.
- Taguri T, Tanaka T and Kuono I (2006)Antibacterial spectrum of plant polyphenols and extracts depending upon hydroxyphenyl structure *Biological and Pharmaceutical Bulletin* 29 (11) : 2226-2235
- Tao L, Park SY, Ibrahim R and Paek KY (2015) Production of biomass and bioactive compounds from adventitious roots by optimization of culturing conditions of *Eurycoma longifolia* in balloon-type bubble

- bioreactor system. *Journal of Bioscience and Bioengineering*. 119 (6) : 712-717.
- Taticek RA, Moo-Young M and Legge RL (1990) Effect of bioreactor configuration on substrate uptake by cell suspension cultures of plant *Eschscholtzia californica*. *Applied Microbial Biotechnology* 33 : 280-286.
- Tawfike AF, Viegelmann C and Edrada-Ebel R (2013) Metabolomics and dereplication strategies in natural products *Methods of Molecular Biology* 1055 : 227-244, 2013
- Tepfer D (1990) Genetic transformation using *Agrobacterium rhizogenes*. *Physiologia Plantarum*. 79(1):140-146.
- Thakore D, Srivastava AK and Sinha AK (2015) Model based fed batch cultivation and elicitation for the overproduction of ajmalicine from hairy roots of *Catharanthus roseus* *Biochemical Engineering* 97 (2015) 73-80
- Tiwari RK, Trivedi M, Guang ZC, Guo GQ, and Zheng, GC (2008) *Agrobacterium rhizogenes* mediated transformation of *Scutellaria baicalensis* and production of flavonoids in hairy roots. *Biologia Plantarum* 52(1) : 26-35.
- Towler MJ, Kim YJ, Wyslouzil BE, Correl MJ and Weathers PJ (2006) Design, development and applications of mist bioreactors for micropropagation and hairy root cultures. In *Plant Tissue Culture Engineering* ed. Dutta Gupta S and Ibaraki Y, pp. 119-134. Neatherlands : Springer.
- Vásquez-Rivera A, Chicaiza-Finley D, Hoyos RA and Orozco-Sánchez F (2015) Production of limonoids with insect antifeedant activity in a two-stage bioreactor process with cell suspension culture of *Azadirachta indica*. *Applied Biochemistry Biotechnology* 177 : 334-345
- Veena V and Taylor CG (2007) *Agrobacterium rhizogenes* : recent developments and promising applications. In *Vitro Cellular & Developmental Biology*. 43(5):383-403.
- Vendruscolo F, Rossi MJ, Schmidell W and Ninow JL (2012) Determination of oxygen solubility in liquid media *ISRN Chemical Engineering* Article ID 601458 (5 pages).
- Verma P, Khan SA, Mathur AK, Shanker K and Lal RK (2014) Regulation of vincamine biosynthesis and associated growth promoting effects through abiotic elicitation , cyclooxygenase inhibition and precursor feeding of bioreactor grown *Vinca minor* hairy roots. *Applied Biochemistry and Biotechnology* 173: 663-672.
- Verpoorte R and Alfermann AW (2000) *Metabolic Engineering of Plant Secondary Metabolism*. Kluwer Academic Publishers, London, United Kingdom.

- Wang B, Zhang G, Zhu L, Chen L and Zhang Y (2006) Genetic transformation of *Echinacea purpurea* with *Agrobacterium rhizogenes* and bioactive ingredient analysis in transformed cultures. *Colloids and Surfaces B: Biointerfaces* 53:101-104.
- Wang JW Fang XK Ren XT (2002) Improved artemisinin accumulation in hairy root cultures of *Artemisia annua* by (22S, 23S)-homobrassinolide. *Biotechnology Letters*. 24 : 1573-1577.
- Wang Q, Wang J, Li J, Man S and Gao W (2015a) Use of partial least-square discriminant analysis to study the effects of gradual scale-up of culture, and optimization of bioreactor angle and aeration volume on culture of *Panax quinquefolium* L. adventitious roots in a 5-L balloon-type bubble bioreactor. *Research of Chemical Intermediates* 41 : 6707-6720.
- Wang Q, Wang J, Chai H, Li J, Man S and Gao W (2015b) Optimization of balloon-type bubble bioreactor angle and methyl jasmonate concentration to enhance metabolite production in adventitious roots of *Pseudostellaria heterophylla*. *Research on Chemical Intermediates* 41 : 5555-5563.
- Weathers PJ, Wyslouzil BE, Wobbe KK, Kim YJ and Yigit E (1999) The biological response of hairy roots to O₂ levels in bioreactors. *In Vitro Cell and Developmental Biology-Plant* 35 : 286-289.
- Weathers PJ, Towler MJ and Xu J (2010) Bench to batch: advances in plant cell culture for producing useful products. *Applied Microbiology and Biotechnology*. 85 : 1339-1351.
- Wen P, Zhang XM, Yang Z, Wang NL and Yao XS (2008) Four new triterpenoid saponins from *Ardisia gigantifolia* Stapf. and their cytotoxic activity. *Journal of Asian Natural Products Research*. 10 (9) : 873-880.
- Williams CRC and Doran PM (2000) Hairy root culture in a liquid dispersed bioreactor : characterization of spatial heterogeneity. *Biotechnology Progress* 16 : 391-401.
- Wilson PDG. (1997) The pilot-scale cultivation of transformed roots. In *Hairy roots: culture and applications*. ed. Doran PM, pp. 179-190 Amsterdam : Harwood Academic.
- Winans SC (1990) Transcriptional induction of an *Agrobacterium* regulatory gene at tandem promoters by plant-released phenolic compounds, phosphate starvation, and acidic growth medium. *Journal of Bacteriology* 172:(5):2433-2438.
- Wolfender JL, Rudaz S, Choi YH and Kim HK (2013) Plant metabolomics: from holistic data to relevant biomarkers. *Current Medical Chemistry* 20 : 1056-1090.

- Wolfender JL, Marti G, Thomas A and Bertand S (2015) Current approaches and challenges for the metabolite profiling of complex natural extracts *Journal of Chromatography A*. 1382 : 136-164
- Wu H Guo J Chen S Liu X Zhou Y Zhang X Xu X (2013) Recent developments in qualitative and quantitative analysis of phytochemical constituents and their metabolites using liquid chromatography-mass spectrometry. *Journal of Pharmaceutical and Biomedical Analysis*. 72 : 267-291.
- Zakaria MH (2015) Review of policies and issues in the Malaysian Herbal Industry. FFTC Agriculture Policy Articles.
- Zhang GH Liang YR Jin J Lu JL Borthakur D Dong JJ and Zhen XQ (2007) Induction of hairy roots by *Agrobacterium rhizogenes* in relation to L-theanine production in *Camellia sinensis*. *Journal of Horticultural Science & Biotechnology* 82(4):636-640.
- Zhang HL, Xue SH, Pu F, Tiwari RK and Wang XY (2010) Establishment of hairy root lines and analysis of gentipicroside in the medicinal plant *Gentiana macrophylla*. *Russian Journal of Plant Physiology* 57 (1) : 110-117.
- Zhao J Davis L and Verpoorte R (2005) Elicitor signal transduction leading to production of secondary metabolites. *Biotechnological Advances* 23 : 283-333.
- Zid SA and Orihara Y (2005) Polyacetylenes accumulation in *Ambrosia maritima* hairy root and cell cultures after elicitation with methyl jasmonate. *Plant Cell Tissue and Organ Culture*. 81 : 65 – 75.