

# **UNIVERSITI PUTRA MALAYSIA**

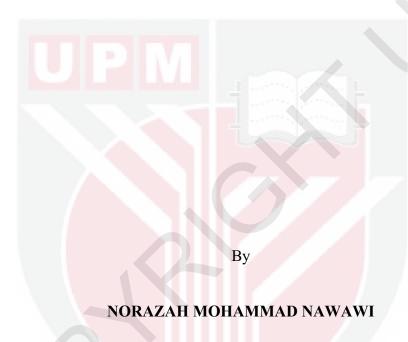
# BIODEGRADATION OF PHENOL BY A MALAYSIAN ISOLATE Rhodococcus sp. NAM 81

# **NORAZAH MOHAMMAD NAWAWI**

FBSB 2016 39



# BIODEGRADATION OF PHENOL BY A MALAYSIAN ISOLATE Rhodococcus sp. NAM 81



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of Philosophy

## **COPYRIGHT**

All materials contained within the thesis including without limitation text, logos, icons, photographs and all other artworks are copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from copyright holder. Commercial use of materials may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



## **DEDICATION**

This Thesis is dedicated to my beloved Husband,

Mr Nor Suhaime Sukirman

Daughters,

Miza Nur Wajihah & Miza Nur Wafiyah

and

My supportive daddy,

<mark>Tuan Ha</mark>ji <mark>Moh</mark>ammad Nawawi Abdul Rahman

sister and brother

as well as my late mother,

Jamilah Abdul Rahman,

my thoughts and prayers are always with you Mak

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Doctor of Philosophy

## BIODEGRADATION OF PHENOL BY A MALAYSIAN ISOLATE Rhodococcus sp. NAM 81

By

#### NORAZAH MOHAMMAD NAWAWI

#### November 2016

Chairman : Siti Aqlima Ahmad, PhD

Faculty : Biotechnology and Biomolecular Sciences

Phenol is a major threat to the environment due its toxicity effect and extensive use in various industries. Phenol can harm human and other organisms at a low dose. Phenols and phenolic compounds are organic pollutants generated from various industrial activities. Biodegradation is a technology that is currently applied for decontaminating pollutants including phenols. Biodegradation of phenols using microorganism is an eco-friendly and cost-effective approach. Bacteria are widely used in bioremediation processing. Thus, the isolation and selection of good microorganism with tolerance towards other toxicants is important to improve the performance of bioremediation. This study evaluates the potential of a local *Rhodococcus* bacteria to decompose phenol, an oxygenated hydrocarbon, one of the harmful pollutants, because of its toxic and carcinogenic effects. Ten strains of *Rhodococcus* spp. isolated from Peninsular Malaysia survived and grew in a medium that was supplemented with 0.5 gL<sup>-1</sup> of phenol. The 1500 bp phenol hydroxylase gene was amplified from the eight strains as a confirmation of phenol-degradation pathway and for future molecular marker for the presence of phenol-degrading bacteria. One strain, Rhodococcus sp. NAM 81 was selected as the most potent strains with the ability to decompose 0.6 gL<sup>-1</sup> phenols within 24 hours. The phenol biodegradation ability of *Rhodococcus* sp. NAM 81 was greatly affected by the presence of trace element in the medium. The parameters that supported the degradation and growth of *Rhodococcus* sp. NAM 81 were optimized initially by one-factor at a time approach. The strain exhibited highest cell growth and phenol degradation at the optimal incubation conditions of 30 °C and pH 7.5. Ammonium sulphate 0.4 gL<sup>-1</sup>, glycine 0.3 gL<sup>-1</sup>, and 0.1 gL<sup>-1</sup> NaCl were needed to enhance cell growth. Apart from phenol, the strain was able to utilizes 1 mg L<sup>-1</sup> of 2,4dinitrophenol, toluene, naphthalene, diesel, acetonitrile, glycerol and waste cooking oil as the sources of carbon for growth, but the phenol degradation ability was inhibited by 1 mg L<sup>-1</sup> of Ag<sup>+</sup>, Cu<sup>2+</sup>, Cr<sup>2+</sup>, Cd<sup>2+</sup>, Zn<sup>2+</sup> and Hg<sup>2+</sup>. Statistical approach by fractional factorial design (FFD) and response surface method (RSM) improved the biodegradation of phenol with pH, phenol and NaCl concentrations were found to be important parameters. The results showed good verification between theoretical and experimental data. RSM method improves the process by 1.2 fold for phenol

degradation and 1.3 fold for cell growth. Investigation on the application of immobilized cells in phenol biodegradation started with an evaluation of suitable matrices such as gellan gum, calcium alginate, agarose, agar-agar and polyacrylamide for phenol degradation using an immobilization approach. Gellan gum was found to be the most effective and suitable matrix for high phenol degradation compared to other matrices studied. Maximum phenol degradation was achieved at the gellan gum concentration of 0.75% (w/v), bead size of 3 mm diameter and bead number of 300 per 100 mL medium. Both free and immobilized bacteria exhibited similar rates of phenol degradation at the phenol concentration of 100 mgL<sup>-1</sup>, but at higher phenol concentrations, immobilized bacteria exhibited a higher degradation rate of phenol. The immobilized cells completely degrade phenol within 108, 216, and 240 h at 1100, 1500 and 1900 mgL<sup>-1</sup> phenol, respectively, whereas free cells took 240 h to completely degrade phenol at 1100 mgL<sup>-1</sup>. In overall, the rates of phenol degradation by both immobilized and free bacteria decreased gradually as the phenol concentrations were increased. It also proved that inhibition of heavy metal and respiratory inhibitors was prevented by gellan gum encapsulated cells. The immobilized cells showed no loss in phenol degrading activity after being used repeatedly for 50 repetitions of 18 h cycle and was stable after storing at 4 °C for 28 days. Study on the best disruption methods for efficient enzyme, protein and genomic DNA isolation from *Rhodococcus* cells showed that grinding for 20 minutes under liquid nitrogen was the best approach. The ortho-cleavage pathway as applied by Rhodococcus sp. NAM 81 for phenol degradation was discovered by biochemical and polymerase chain reaction. The gene for phenol hydroxylase; the enzyme that catalyzes the conversion of phenol to catechol, has been cloned, expressed and purified from *Rhodococcus* sp. NAM 81. SDS-PAGE produced a single band with a molecular weight of ~ 60 kDa. The potential of resting cells of *Rhodococcus* sp. NAM 81 as an alternative to the use of free and immobilized cells for phenol biodegradation process in liquid waste was also tested. The results from this study showed that *Rhodococcus* sp. NAM 81 has an excellent potential that can be applied in bioremediation of phenol-containing wastes especially using immobilized cells.

# BIODEGRADASI FENOL OLEH *Rhodococcus* sp. NAM 81 PENCILAN MALAYSIA

Oleh

#### NORAZAH MOHAMMAD NAWAWI

#### November 2016

Pengerusi : Siti Aqlima Ahmad, PhD

Fakulti : Bioteknologi dan Sains Biomolekul

Fenol merupakan satu bahan yang mengancam alam sekitar kerana kesan ketoksikan dan penggunaan yang meluas dalam pelbagai industri. Fenol membahayakan manusia dan organisma lain walaupun dalam dos yang minimum. Fenol dan sebatian fenolik merupakan pencemar organik yang dihasilkan dari pelbagai aktiviti industri. Biodegradasi adalah teknologi yang boleh digunakan untuk menguraikan bahan pencemar termasuk fenol. Biodegradasi fenol menggunakan mikroorganisma adalah satu pendekatan yang mesra alam dan ekonomi. Bakteria digunakan secara meluas dalam proses pemulihan alam sekitar. Maka, pemencilan dan pemilihan mikroorganisma yang bagus dengan toleransi terhadap toksik lain adalah penting untuk meningkatkan prestasi pemulihan persekitaran. Kajian ini menilai potensi bakteria Rhodococcus tempatan untuk mengurai fenol, iaitu hidrokarbon oksigen, yang merupakan salah satu daripada bahan cemar berbahaya kerana kesan toksik dan karsinogeniknya. Sepuluh strain *Rhodococcus* spp. dipencilkan dari Semenanjung Malaysia mampu tumbuh dalam medium yang mengandungi 0.5 gL<sup>-1</sup> fenol. 1500 bp gen fenol hydroxylase gen telah diamplifikasi dari lapan strain sebagai penanda molekul. Satu strain, *Rhodococcus* sp. NAM 81 yang telah dipilih sebagai strain yang paling efektif menguraikan 0.5 gL<sup>-1</sup> fenol dalam masa 24 jam. Keupayaan biodegradasi *Rhodococcus* sp. NAM 81 terjejas dengan kehadiran unsur surih dalam medium. Sebagai permulaan, parameter yang membantu biodegradasi dan pertumbuhan Rhodococcus sp. NAM 81 telah dioptimumkan menggunakan satu faktor pada satu masa. Pertumbuhan sel tertinggi dan degradasi fenol adalah optimum pada (30 °C, pH 7.5). Ammonium sulfat 0.4 gL<sup>-1</sup>, glycine 0.3 gL<sup>-1</sup> dan 0.1 gL<sup>-1</sup> NaCl diperlukan untuk meningkatkan pertumbuhan sel. Selain daripada fenol, strain ini juga mampu menggunakan 2,4-dinitrophenol, toluena, naftalena, diesel, asetonitril, gliserol dan sisa minyak memasak sebagai sumber karbon. Namun, kebolehan Rhodococcus sp. NAM 81 direncat dengan kehadiran logam berat di dalam medium yang mengandungi 1 mgL<sup>-1</sup> Ag<sup>+</sup>, Cu<sup>2+</sup>, Cr<sup>2+</sup>, Cd<sup>2+</sup>, Zn<sup>2+</sup> and Hg<sup>2+</sup>. Kaedah statistik melalui rekabentuk faktor fraksi (FFD) dan pengkaedahan tindakbalas permukaan (RSM) meningkatkan keupayaan biodegradasi fenol dengan pH, kepekatan fenol dan NaCl sebagai parameter penting yang disahkan melalui teori dan data eksperimen. RSM

telah memperbaiki proses dengan 1.2 kali ganda bagi degradasi fenol dan 1.3 kali ganda bagi pertumbuhan sel. Kajian tentang penggunaan sel sekat gerak dalam biodegradasi fenol bermula dengan penilaian matriks yang sesuai merangkumi gam gellan, kalsium alginat, agarose, agar-agar dan polyacrylamide. Gam gellan didapati matriks yang sesuai berbanding dengan matriks-matriks lain. Maksimum degradasi fenol dicapai pada kepekatan gam gellan sebanyak 0.75 %, saiz manik 3 mm dengan bilangan 300 biji dalam 100 ml media. Kedua-dua sel bebas dan sel sekat gerak mengurai pada kadar yang sama pada kepekatan fenol 100 mgL<sup>-1</sup>, namun pada kepekatan yang tinggi, sel sekat gerak sahaja mengurai pada kadar yang tinggi. Fenol terurai sepenuhnya dalam masa 108, 216, dan 240 jam dalam kepekataan fenol masing-masing 1100, 1500 dan 1900 mgL<sup>-1</sup>, manakala sel bebas mengambil masa 240 jam untuk mengurai sepenuhnya 1100 mgL<sup>-1</sup> fenol dan tidak berupaya mengurai sepenuhnya fenol pada kepekatan yang tinggi. Secara keseluruhan, kadar degradasi fenol oleh sel sekat gerak dan sel bebas menurun selari dengan peningkatan kepekatan fenol. Logam berat dan perencat respirasi memberikan kesan yang rendah pada penguraian fenol oleh sel tersekat-gerak berbanding sel bebas. Sel tersekat-gerak boleh digunakan berulang kali selama 50 kitaran 18 jam dan stabil dalam penyimpan pada 4 °C selama 28 hari. Kajian tentang teknik untuk memecahkan sel bagi mengeluarkan enzim, protein dan DNA daripada sel Rhodococcus menunjukkan pemecahan menggunakan nitrogen cecair dalam masa 20 minit adalah kaedah terbaik. Bakteria ini menggunakan tapak jalan ortho untuk menguraikan fenol yang telah ditentukan menggunakan pendekatan biokimia dan tindak balas berantai polimerase. Gen untuk fenol hydroxylase, iaitu enzim yang memangkinkan penukaran fenol kepada catechol telah diklon, dirembes serta ditulenkan daripada Rhodococcus sp. NAM 81. Gel SDS-PAGE menghasilkan jalur tunggal dengan berat molekul ~ 60 kDa. Potensi sel rehat bagi *Rhodococcus* sp. NAM 81 sebagai alternatif kepada penggunaan sel bebas dan tersekat gerak untuk bidegradasi fenol dalam pencemar cecair juga telah diuji. Keputusan daripada kajian ini menunjukkan bakteria Rhodococcus sp. NAM 81 mempunyai potensi yang baik yang boleh diaplikasikan dalam proses biopemulihan sisa mengandungi fenol terutamanya menggunakan sel tersekat-gerak.

#### **ACKNOWLEDGEMENTS**

## In the name of Allah, the Most Gracious, Most Merciful

Alhamdulillah, all praises to Allah for the strengths, guide and blessing in my study and the completion of this thesis. Special appreciation goes to my supervisor, Dr. Siti Aqlima Ahmad, who was being abundantly helpful and offered invaluable support and guidance that enables me to develop a deeper understanding on the biology of these microorganisms. I am heartily thankful to my co-supervisor, Assoc. Prof. Dr. Mohd Yunus Abd. Shukor and Prof. Dr. Mohd Arif Syed. Their invaluable constructive comments and suggestions throughout the experiments and thesis works have contributed the success of this research.

It is an honour for me to thank my mentor, Emeritus Prof. Dato' Dr. Abdul Latif Ibrahim, who initiated my doctoral programme and has made his support available in a number of ways. Not to forget, Dr. John Archer, our collaborator from University of Cambridge and currently attach to King Abdullah University of Science and Technology, Jeddah, Saudi Arabia who instigate the *Rhodococcus* research to our team in Universiti Selangor (UNISEL), Universiti Putra Malaysia (UPM) and Universiti Kebangsaan Malaysia (UKM) supported by Ministry of Science, Technology and Innovation Malaysia under Malaysia Genome Institute from Grant 07-05-16-MGI-GMB05. I would also like to acknowledge UPM and UNISEL, specifically to Institute Bio-IT Selangor for the facilities to do the experiments.

I wish to extend my warmest thanks to all of those who have helped me with my work particularly, Dr Noor Azlin for the working organism, *Rhodococcus* spp., Dr. Izuan and Dr. Nor Suhaila for RSM work and Dr. James and Dr. Julie from Faculty of Life Sciences, Universiti of Manchester, United Kingdom for molecular experiment. Deepest gratitude also goes to my friends in UNISEL, Assoc. Prof. Fatimah, Dr. Maegala, Dr. Fridelina, Nadzirah, Hasdianty, Siti Nur Marlia, Rozila, Norhatiah, Nor Azmahera, Roziah, Suhaiza Jamhor, Sashila, Jayasree and all friends from the bioremediation group in UPM, Dr. Adeela, Dr. Khalizan, Dr. Baskaran, Dr. Shamala, Norliza, Sabrina, Rahim, Shahir, Aqilah, Ain, Zira and others.

This thesis would not be possibly done without the love and support from my husband, Mr. Nor Suhaime and daughters, Miza Nur Wajihah and Miza Nur Wafiyah. Thank you for the patient, understanding, everlasting prayers as well as accompany me through sad and happy moments. To my father, brothers and sisters and all my family members, thank you for your encouragement, understanding and emotional support that have made it possible for me to finish this study.

Lastly, I offer my regards and blessings to all of those who supported me in any value during the completion of my dissertation. To those who indirectly contributed to this research, your kindness means a lot to me. Thank you very much.

Norazah Mohamad Nawawi, 2016



This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

## Siti Aqlima Ahmad, PhD

Senior Lecturer Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

### Mohd Yunus Shukor, PhD

Associate Professor
Faculty Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

## Mohd Arif Syed, PhD

Professor Faculty of Biotechnology Universiti Putra Malaysia (Member)

### Abdul Latif Ibrahim, PhD

Emeritus Professor Institute Bio-IT Selangor Universiti Selangor (Member)

## **ROBIAH BINTI YUNUS, PhD**

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

#### **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature:	Date:	

Name and Matric No: Norazah Mohammad Nawawi, GS22884

# **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature:	
Name of Chairman	
of Supervisory	
Committee:	Dr. Siti Aqlima Ahmad
Signature:	
Name of Member	
of Supervisory	
Committee:	Associate Professor Dr. Mohd Yunus Shukor
Signature:	
Name of Member	
of Supervisory	
Committee:	Professor Dr. Mohd Arif Syed
Signature:	
Name of Member	
of Supervisory	
Committee	Empritus Professor Dr. Abdul I stif Ibrahim

# TABLE OF CONTENTS

						Page
	RAK OWLI	EDGEM	ENTS			i iii V
APPRO						vii :
DECL.						ix xvii
		GURES				xix
		BREVIA	TIONS			XXII
ZIST (	, , , , , ,		110110			717111
CHAP	TER					
1	GEN	FRAL I	NTROD	UCTION		1
•	1.1		h backgr			1
	1.2		ent of pro			1
	1.3		cance of s			
	1.4				earch approach	2 2
2	LITE	ERATUR	E REVI	EW		4
_	2.1			echnology		4
	2.2			pollution		4
	2.3		-	hnologies		5
	2.5	2.3.1	Bioreme			5 5 7
	2.4	Xenobio				7
	2.5					7
		2.5.1	Sources	of phenol		8
		2.5.2		of phenol		10
		2.5.3		-	nto the environment	10
	2.6	Organic		-	st <mark>e in Mala</mark> ysia	12
	2.7	Phenol	managen	nent method	d	13
		2.7.1	Microbi	al degradat	ion of phenol	13
		2.7.2	Phenol-	degrading r	nicroorganisms	14
			2.7.2.1		grading mold	15
			2.7.2.2	Phenol-de	egrading yeast	16
			2.7.2.3	Phenol-de	egrading algae	18
			2.7.2.4	Phenol-de	grading bacteria	18
				2.7.2.4.1	Phenol-degrading bacteria from	19
					phenol and hydrocarbon-related	
				27242	industries and impacted areas	21
				2.7.2.4.2	Phenol-degrading bacteria from	21
				27242	chemical-related industries	22
				2.7.2.4.3	Phenol-degrading bacteria from	22
				27244	agricultural-related industries  Phonol degrading booterie from	22
				2.7.2.4.4	Phenol-degrading bacteria from	23
					pristine and uncontaminated	

		2.7.2.4.5 Phenol-degrading extremophilic bacteria	24
	2.8	Actinomycete in bioremediation	27
	2.0	2.8.1 Characteristics of <i>Rhodococcus</i>	28
		2.8.2 Phenol-degrading <i>Rhodococcus</i> sp.	29
	2.9	Aerobic biodegradation of phenol	30
		Phenol degradation pathway	31
		Application of immoblized cells for phenol degradation	33
	2.12		34
3	MAT	TERIALS AND METHODS	35
	3.1	Experimental design	35
	3.2	Chemicals	39
	3.3	Microorganism	39
		3.3.1 Preparation of inoculums	39
	3.4	Microbiological media	40
		3.4.1 Luria-Bertani medium	41
		3.4.2 Nutrient agar and broth	41
		3.4.3 Minimal salt medium (MSM)	41
	3.5	Oligonucleotides	41
	3.6	Isolation of genomic DNA	43
	3.7	Isolation of plasmid DNA	43
	3.8	Quantification of nucleic acids	43
	3.9	Electrophoresis	44
		Polymerase chain reaction	44
		3.10.1 Standard PCR	44
		3.10.2 Colony PCR	44
	3.11	Purification of reaction products	44
	3.12	Sequence analysis	45
		Analytical procedures	45
		Cell concentration	45
	3.15	Determination of phenol concentration	45
		Intracellular enzyme assay	46
		3.16.1 Preparation of cell extraction	46
		3.16.2 Protein determination	46
	3.17	Statistical analysis	46
4	SCRI	EENING AND CHARACTERIZATION OF PHENOL	47
	DEG	RADING Rhodococcus BACTERIA	
	4.1	Introduction	47
	4.2	Materials and methods	47
		4.2.1 Chemicals and microorganism	47
		4.2.2 Screening of phenol-degrading <i>Rhodococcus</i>	48
		4.2.3 Detection of phenol hydroxylase gene	48
		4.2.4 Effects of different medium composition on phenol	48
		degradation	
		4.2.5 Physio-chemical optimization for optimum growth	49
		and enhanced phenol degradation	
		4.2.6 Analytical procedures	50
		4.2.6.1 Cell concentration	50

			4.2.6.2 Phenol concentration	50
			4.2.6.3 Statistical analysis	50
	4.3	Result	and discussion	50
		4.3.1	Screening of phenol degrading <i>Rhodococcus</i> spp.	50
		4.3.2		54
			phenol degradation and growth of <i>Rhodococcus</i> sp.	
			NAM 81	
		4.3.3	Optimization of culture conditions for the optimum	56
			growth and phenol degradation of <i>Rhodococcus</i> sp.	
			NAM 81	
			4.3.3.1 Effects of temperature	56
			4.3.3.2 Effects of initial pH	58
			4.3.3.3 Effect of nitrogen sources and the	59
			concentration of nitrogen sources	
			4.3.3.4 Effects of salinity	61
		4.3.4	Effect of phenol concentration	62
		4.3.5		67
		4.3.6	Effects of Tween 80	69
		4.3.7	Utilisation of additional carbon source	69
	4.4	Conclu	asion	71
_	CT A	TICTIC	AL OPTIMIZATION FOR IMPROVEMENT OF	73
5				13
	5.1	Introdu	EGRADATION BY Rhodococcus sp. NAM 81	73
	5.2		als and methods	73 74
	3.2	5.2.1	Chemicals	74
		5.2.2		74 74
		5.2.3	Phenol biodegradation experiments	74 74
		5.2.4	Analytical procedures	75
		5.2.5	Experimental design and data analysis for	75
		3.2.3	optimisation of phenol degradation and cell growth	13
		5.2.6	Screening of significant physico-chemical	75
		3.2.0	determinants by FFD	13
		5.2.7	Optimization of key determinants by RSM	75
		5.2.8	Statistical analysis and modeling	76
		5.2.9	Validation of the experimental model	77
	5.3		s and discussion	77
		5.3.1	Screening of important variables using FFD	77
		5.3.2	Optimization of the key determinants by CCD for	79
			phenol degradation	
		5.3.3	Response surface plotting for the optimization of	84
			biodegradation of phenol by <i>Rhodococcus</i> sp. NAM	
			81	
		5.3.4	Response surface plotting for optimising the growth	86
			of <i>Rhodococcus</i> sp. NAM 81	
		5.3.5	Verification of experiments	89
	5.4	Conclu	•	90

6	PHE	NOL DE	EGRADATION BY IMMOBILIZED Rhodococcus	91
	sp. N	<b>IAM 81</b>		
	6.1	Introdu	ction	91
	6.2	Materia	als and methods	92
		6.2.1	Chemicals	92
		6.2.2	Microorganism and Inoculum Preparation	92
		6.2.3	Cultivation and phenol biodegradation experiments	92
		6.2.4	Preparation of immobilized cells	92
		0.2.1	6.2.4.1 Gellan gum	92
			6.2.4.2 Agarose	93
			6.2.4.3 Calcium alginate	93
			6.2.4.4 Agar	93
			6.2.4.5 Polyacylamide	93
		6.2.5	Screening of different matrices for immobilization	94
		0.2.3		94
		626	of <i>Rhodococcus</i> sp. NAM 81 for phenol degradation	0.4
		6.2.6	Optimization of immobilized condition	94
		6.2.7	Cell leakage	94
		6.2.8	Comparison of phenol degradation between free and	94
			immobilized cells	0.4
		6.2.9		94
		6.2.10	· · ·	95
			Statistical analysis	95
		6.2.12		95
	6.3		and discussion	95
		6.3.1	Phenol degradation by immobilized cells in	95
			different entrapment matrix	
		6.3.2	Optimization conditions for immobilized cells	97
			6.3.2.1 Effects of matrix component concentration	97
			6.3.2.2 Effects of crosslinker concentration	98
			6.3.2.3 Effect of bead size	99
			6.3.2.4 Effect of initial cell loading	100
			6.3.2.5 Effect of beads number	101
		6.3.3	Effect of initial phenol concentration on phenol	102
			degradation by freely suspended and immobilized	
			cells	
		6.3.4	Effect of heavy metals on phenol degradation by	104
		0.0	freely suspended and immobilized cells	10.
		6.3.5	Effect of respiratory inhibitors on phenol	106
		0.5.5	degradation by freely suspended and immobilized	100
			cells	
		6.3.6	Reusability of immobilized beads	108
		6.3.7	Storage stability of <i>Rhodococcal</i> cells and gellan	111
		0.5.7	gum encapsulated beads with phenol degradation	111
			activity	
		620	•	111
		6.3.8	Scanning electron microscopy (SEM) of immobilized cells	111
	6.1	Comple		112
	6.4	Conclu	SIOII	112

7		RACTI YMES	ERIZATI	ON OF PHENOL DEGRADING	114
	7.1	Introdu	action		114
	7.1		als and m	atha da	115
	1.2	7.2.1			115
		7.2.2		ganism and inoculum preparation	115
		7.2.3		lular quantification using different	115
				ical disruption methods	115
			7.2.3.1	Grinding under liquid nitrogen	115
			7.2.3.2	9	115
				Sonication	116
		724		Homogenisation	116
		7.2.4		lic pathway determination	116
			7.2.4.1	Metabolic pathway determination using	116
				biochemical analysis	11/
				7.2.4.1.1 Phenol hydoxylase assay	116
				7.2.4.1.2 Catechol 1,2 dioxygenase assay	116
				7.2.4.1.3 Catechol 2,3-oxygenase assay	117
				7.2.4.1.4 Test tube analysis	117
				7.2.4.1.5 Spray plate method	117
				7.2.4.1.6 Spot test for detection of	117
				catechol 2,3 dioxygenase	
				activity	
			7.2.4.2	1	117
				dioxygenase and catechol 2,3-dioxygenase	
				gene	
		7.2.5		, expression and characterization of phenol	118
			hydroxy		
			7.2.5.1		118
			7.2.5.2	Cloning and sequencing of phenol	118
				hydroxylase gene	
			7.2.5.3	Preparation of glycerol stock	120
			7.2.5.4	Expression of phenol hydroxylase in <i>E.coli</i>	120
			7.2.5.5	Purification of phenol hydroxylase	120
		7.2.6	Sodium	Dodecyl Sulphate Polyacrylamide Gel	121
			Electrop	phoresis (SDS-PAGE)	
		7.2.7	Analysis	s of protein by mass spectrometry	122
	7.3	Results	and disc	ussion	122
		7.3.1	Determi	nation of the optimum disruption methods	122
			to extra	ct intracellular metabolite from <i>Rhodococcus</i>	
			sp. NAN	M 81	
		7.3.2	Determi	ination of phenol metabolic pathway in	127
		-	Rhodoce	occus sp. NAM 81	
		7.3.3		, expression and characterization of phenol	132
			hydroxy		
			7.3.3.1	Amplification of phenol hydroxylase gene by PCR	133
			7.3.3.2	Cloning of phenol hydroxylase gene in	135
			1.3.3.4	vector pQE9	133

			7.3.3.3	Screening using PCR and restriction enzyme digestion	136
			7.3.3.4	Expression and purification of phenol hydroxylase gene	137
			7.3.3.5	Catalytic activity of His6PheA1	146
	7.4	Conclu		Catalytic activity of fillsof herri	149
8	Biolo	gical Re	moval of	Phenol From Simulated Wastewater	150
	Using	-		LS OF Rhodococcus sp. NAM 81	
	8.1	Introdu			150
	8.2		als and M		150
		8.2.1	-	ion of synthetic wastewater	150
		8.2.2	Preparat NAM 8	tion of seed culture of <i>Rhodococcus</i> sp.	150
		8.2.3		nation of growth curve	151
		8.2.4		ion of resting cells	151
		8.2.5	Effect o	f cells suspension media and cells	151
		8.2.6		nation of factor for phenol degradation by	151
			resting of		
		8.2.7		stability and reusability of resting cells	151
		8.2.8	-	ison between immobilized, growing and	152
				cells in phenol degradation at different	
		0.2.0	_	concentration	150
	8.3	8.2.9	and Disc	cal procedures	152
	0.3	8.3.1			152 152
		8.3.2		curve of <i>Rhodococcus</i> sp. NAM 81 f different medium for the production of	154
		6.3.2	resting of	-	134
		8.3.3	_	f cells suspension media and cells	155
			concent	ration on phenol degradation	
		8.3.4	Determine resting of	nation of factor for phenol degradation by	157
		8.3.5		stability and reusability of resting cells of	159
				occus sp. NAM 81	
		8.3.6		ison between immobilized, growing and	160
			_	cells in phenol degradation at different	
	0.4	G 1		concentration	1.61
	8.4	Conclu	sion		161
9				D RECOMMENDATION FOR FUTURE	162
		EARCH			1.60
	9.1	Conclu		6 6 4 1	162
	9.2	Kecom	mendatio:	n for future study	164
REFER	RENCI	ES			166
APPEN	DICE	$\mathbf{S}$			210
BIODA	TA O	F STUD	ENT		216
LIST O	F PUI	BLICAT	TIONS		217

# LIST OF TABLES

Table		Page
2.1	Sources of phenols and the other related aromatic compounds in wastewater	9
2.2	Levels of phenol in wastewater of some selected industries	11
2.3	Phenol management method	13
2.4	List of molds that are capable to degrade phenol	15
2.5	List of phenol-degrading yeast	18
2.6	List of bacteria that are capable to degrade phenol	25
2.7	Source of origin for phenol degrading Rhodococcus sp.	29
3.1	Bacterial strain used in this work	40
3.2	Oligonucleotides used this study	42
4.1	Growth of phenol-degrading <i>Rhodococcus</i> spp. on MSM containing 0.5gL <sup>-1</sup> phenol within 10 days of incubation at 30 °C	51
4.2	Effect of different phenol concentration on the phenol degradation by eight <i>Rhodococcus</i> spp. in five days incubation	53
4.3	Carbon utilization profile of <i>Rhodococcus</i> sp. NAM 81	70
5.1	Experimental range and levels of the independent variables used in fractional factorial design in terms of actual and coded units	75
5.2	Experimental range and levels of the independent variables used in RSM in terms of actual and coded units	76
5.3	Half FFD matrix for 6 independent variables for phenol degradation by <i>Rhodococcus</i> sp. strain NAM 81	78
5.4	Analysis of variance (ANOVA) for fitted second-order polynomial model as per half FFD	79
5.5	Analysis of variance (ANOVA) for phenol degradation and cell growth for response surface model	80
5.6	The predicted and experiment value of phenol degradation and cell growth of <i>Rhodococcus</i> NAM 81 in the matrix of central composite design	83

5.7	Validation of the predicted and experimental cultivation variable	89
6.1	Percentage of phenol degradation and amount of cell leakage using various immobilization matrices	96
6.2	Effect of initial phenol concentration on phenol degradation ability of immobilized and free suspended cells of <i>Rhodococcus</i> sp. NAM 81 within 12 days of cultivation period	103
6.3	Effect of storage stability on percentage of phenol degradation by immobilized and free cells	111
7.1	Components of digestion mixture	119
7.2	Components of ligation mixture	119
7.3	Analysis of total protein content and specific enzyme activity	123
7.4	Nucleic acid yield obtained by four disruption methods from Rhodococcus sp.	126
7.5	Protein expression at different temperature	141
7.6	Purification table of recombinant phenol hydroxylase using Histrap HP IMAC purification column	145
8.1	Physical factors on phenol degradation by resting cells of <i>Rhodococcus</i> sp. NAM 81	158
8.2	Thermostability of resting cells of Rhodococcus sp. NAM 81	160

# LIST OF FIGURES

Figure		Page
2.1	Schematic diagram on biological treatment from three distinct areas of biocatalyst	6
2.2	Chemical structure of phenol	8
2.3	Phenol metabolic pathway	32
3.1	Experimental design of phenol biodegradation by <i>Rhodococcus</i> sp.	36
4.1	Amplification of phenol hydroxylase gene for general detection and as the molecular marker of phenol degrading bacteria	52
4.2	Comparison of medium on the growth profile (A) and phenol degradation (B) of <i>Rhodococcus</i> sp. NAM 81	55
4.3	Effect of temperature on the growth profile and phenol degradation of <i>Rhodococcus</i> sp. NAM 81	57
4.4	Effect of initial pH of MSM containing 0.5 gL <sup>-1</sup> on the growth profile and phenol degradation of <i>Rhodococcus</i> sp. NAM 81	59
4.5	Effect of nitrogen sources on the growth profile and phenol degradation of <i>Rhodococcus</i> sp. NAM 81	60
4.6	Effect of ammonium sulphate and glycine concentration on bacterial growth and percentage of phenol degradation	61
4.7	Effect of NaCl concentration on the growth profile and phenol degradation of <i>Rhodococcus</i> sp. NAM 81	62
4.8	Effect of phenol concentration on the growth profile and phenol degradation of <i>Rhodococcus</i> sp. NAM 81	66
4.9	Effect of metal ions on the growth profile and phenol degradation of <i>Rhodococcus</i> sp. NAM 81	68
4.10	Effect of Tween 80 on growth and phenol degradation ability of <i>Rhodococcus</i> sp. NAM 81	69
5.1	Three dimensional response surface plot for phenol degradation with respect to the interactive effects of pH and phenol concentration (A); phenol concentration and NaCl concentration (B) and the interaction of phenol concetration and incubation time (C)	86

5.2	Three dimensional response surface plotted for cell growth with respect to the interactive effects of pH and phenol concentration (A); phenol concentration and NaCl concentration (B) and phenol concentration and incubation time (C)	88
6.1	Effect of different percentage of gellan gum on phenol degradation by immobilized <i>Rhodococcus</i> sp. Strain NAM 81	98
6.2	Effects of concentration of calcium chloride on phenol degradation by immobilized <i>Rhodococcus</i> sp. Strain NAM 81	99
6.3	Effects of beads size on phenol degradation by immobilized <i>Rhodococcus</i> sp. strain NAM 81	100
6.4	Effect of cell concentration on phenol degradation by immobilized <i>Rhodococcus</i> sp. strain NAM 81	101
6.5	Effect of beads number on phenol degradation by immobilized <i>Rhodococcus</i> sp. strain NAM 81	102
6.6	Effect of heavy metals on phenol degradation by immobilized (black) and free cells (white) of <i>Rhodococcus</i> sp. strain NAM 81	105
6.7	Effect of respiratory inhibitors on phenol degradation towards immobilized and free cells of <i>Rhodococcus</i> sp. strain NAM 81	107
6.8	Effect of 3 mM respiratory inhibitors on phenol degradation towards immobilized and free cell of <i>Rhodococcus</i> sp. strain NAM 81	108
6.9	Repeated usage of immobilized <i>Rhodococcus</i> sp. strain NAM 81 in the initial phenol concentration of 500 mgL <sup>-1</sup> for cycle 41-54	110
6.10	Scanning electron microscope of blank gellan gum bead (a) and cells immobilized in gellan gum bead (b) after degrading phenol 18hours under optimum condition	112
7.1	Protein extraction assessments using various mechanical disruptions	124
7.2	Analysis of phenol hydroxylase using different grinding time under liquid nitrogen	125
7.3	Metabolism of phenol-degrading pathways by <i>Rhodococcus</i> sp. strain NAM 81	128
7.4	Metabolic pathway analysis of <i>Rhodococcus</i> sp. NAM 81 using test tube method	129

7.5	spray method	130
7.6	Metabolic pathway analysis of <i>Rhodococcus</i> sp. NAM 81 using spot test method	130
7.7	The amplification of catechol 1,2 dioxygenase gene on an agarose gel	132
7.8	PCR amplification product of phenol hydroxylase gene from the genomic DNA of <i>Rhodococcus</i> sp. NAM 81 with different annealing temperatures	134
7.9	Nucleotide sequence of phenol hydroxylase gene in plasmid pQE9	138
7.10	SDS-PAGE of Escherichia coli XL 10 Gold harboring pQE9-phehyd1	139
7.11	Effect of IPTG induction on growth of E.coli XL 10 Gold	140
7.12	Time course study of protein expression induced at 37 °C	143
7.13	The purified His6Phe1NAM 81 on SDS-PAGE	145
7.14	Catalytic activity of His6Phe1Hyd1 in E. coli XL10 Gold	147
7.15	The chromoatogram of LC-MS	148
8.1	Growth curve of Rhodococcus sp. NAM 81	153
8.2	Phenol degradation by resting cells of <i>Rhodococcus</i> sp. NAM 81	155
8.3	Phenol degradation by resting cells of <i>Rhodococcus</i> sp. NAM 81 resuspend in different solution	156
8.4	Phenol degradation in synthetic wastewater containing 0.5 gL <sup>-1</sup> phenol for 24 hours at 30 °C by various cell concentration of resting cells of <i>Rhodococcus</i> sp. NAM 81	157
8.5	Percentage of phenol degradation by free, resting and immobilize <i>Rhodococcus</i> cells at diverse phenol concentration in 24 hours	161

## LIST OF ABBREVIATIONS

Abs Absorbance

CCD Central composite design

Cd Cadmium

cm Centimetre

Cr Chromium

Co Cobalt

Cu Copper

°C Degree celcius

dH<sub>2</sub>O Distilled water

Da Dalton

EDTA Ethylene diamine tetra acetic acid

FFD Fractional factorial design

G Gram

gL<sup>-1</sup> Gram per litre

> Greater than

h Hour

Fe Iron

kDa Kilo dalton

kb Kilo base

kg Kilogram

< Less than

L Liter

Hg Mercury

mL Mililiter

mgL<sup>-1</sup> Miligram per litre

mM Milimolar

μgL<sup>-1</sup> Microgram per litre

μM Micromolar

μL Microlitre

min Minute

MW Molecular weight

MSM Minimal salt medium

M Molar

μm Micrometer

h<sup>-1</sup> Per hour

L<sup>-1</sup> Per liter

% Percent

et al and friends

NA Nutrient agar

OD Optical density

PAGE Polyacrylamide gel electrophoresis

RPM Rotation per minute

RSM Response surface method

SDS Sodium dodecyl sulphate

SEM Scanning electron microscope

#### **CHAPTER 1**

#### **GENERAL INTRODUCTION**

### 1.1 Research background

Malaysia as a modern Islamic country has made a progressive living and operating environment in a stable political and consistent economic growth from various industries. Industrializations will lead to numerous environmental problems that is harmful to ecosystems and communities (Hsu *et al.*, 2014). In the Fourth Regional 3R Forum in Asia, it has been reported that about 3,281,569.21 metric tonnes of scheduled wastes have been generated in Malaysia compared to 3,087,496.84 metric tonnes (6.29%) in 2010. Besides dross, slag, clinker, ash, gypsum, mineral sludge and heavy metals sludge, about 7,904.42 metric tonnes of phenol have been also released, which contributes to about 0.49% of the total waste as the main categories of waste generated in the country (DOE, 2011).

Phenol (C<sub>6</sub>H<sub>5</sub>OH) is toxic and relatively recalcitrant to biological degradation process (Li et al., 2015; ATSDR, 2014). Phenol occurs naturally and can also originate from industrial effluents such as pharmaceutical, food industries, oil refineries and coal conversion processes (Yemendzhiev et al., 2008; Passos et al., 2010; Pradeep et al., 2014; Aida and Hanan, 2014). Phenol and phenolic compounds are also released into the wastewater from steel industries, petrochemicals, polymeric resins and dye manufacturing units. Phenol is important as raw materials and additives for industrial purposes, especially in laboratory processes, chemical industry, chemical engineering, wood and plastics processing. Phenol and phenolics are soluble in water; hence causing the carbolic odour in water. It disturbs the ecological balance since it is toxic to aquatic flora and fauna (Michalowicz and Duda, 2007; Clodio et al., 2009; Soudi and Kolahchi, 2011). It could also diffuse across a cell's membrane of an organism. Studies showed that constant administration of phenol by animals could cause pathological changes (ATSDR, 1998). Phenol discharge towards the water body will endangers aquatic life, even at concentration of between 500 and 2500 mgL<sup>-1</sup> (Yan et al., 2005).

### 1.2 Statement of problem

Based on the toxicity and risks to organism and environment, phenol removal is crucial for lowering the concentration of phenol below the regulatory limit that requires appropriate treatment of waste containing phenol before being discharged to the nature. A report prepared by the United States Government Accountability Office mentioned that their agencies have spent almost US\$30 billion from 1986 to 2008 for all environmental cleanup and restoration activities (GAO, 2010). Physical, chemical and biological methods have been widely applied to remove contaminants from the polluted area. Yet, among the techniques, biological removal offers the cheapest and safest method through bioremediation (Bijay *et al.*, 2012). It is an ideal solution for pollution abatement nowadays being a green technology that uses biological systems

for the treatment of contaminants (Desai *et al.*, 2010). Biological approach is a method of choice as it is low in cost and offers complete mineralisation compared to other methods (Shweta and Dhandayuthapani, 2013; Agarry *et al.*, 2008). Even though this recent technology is using a multidisciplinary approach, its central thrust depends on microbiology, especially with the application of bacteria in phenol biodegradation (Gami *et al.*, 2014; Aysha and Mumtaj, 2014; Shah, 2014; Krastanov *et al.*, 2013).

## 1.3 Significance of study

Realising the importance of using microorganisms as a potential remediation system for xenobiotic compounds in waste, the Malaysian Government, through the Malaysia Genome Institute (MGI) established a project on the isolation of such microbes for industrial and environmental applications. The study on the ecology of marine and terrestrial microbes around Peninsular Malaysia is in progress with a number of microorganisms from different ecological niches that have been isolated. A group of microorganisms that have been chosen for further research is the mycolic acidcontaining actinomycetes. The broad metabolic diversity of the mycolic acidcontaining actinomycetes have attracted the pharmaceutical, environmental, chemical and energy industries. The discovery of novel xenobiotics-degrading metabolic capability and bioactive compounds from this group of microbes is far from exhausted (Davies and Davies 2010; Arenskotter et al., 2004). The rhodococci is a group of aerobic, non-motile actinomycetes (Larkin et al., 2005). Rhodococci degrade an extensive array of recalcitrant pollutants that makes them promising candidates to be used in bioremediation. In addition, there are numerous other industrial applications where rhodococci, or their gene products, are actually being used or are in the pipeline. The economic significance of *Rhodococcus* species are being widely recognised. In Malaysia, informations available on the potential role of *Rhodococcus* spp. especially in phenol detoxification is slowly being discovered.

Environmental-friendly technologies are gaining increasing prominence as a safe remediation technique for phenol and phenolics pollution (Agrawal and Shahi, 2015; Dash *et al.*, 2009). In this study, the evaluation of various locally isolated *Rhodococci* will broaden the reservoir of existing phenol-degrading microbial populations that are competent in phenol degradation especially in a temperate climate.

### 1.4 Research objectives and research approach

Keeping all these points in view, the overall aim of this project was to develop an approach for phenol biodegradation performed by locally-isolated microorganism. The hypothesis for this research were; (a) the locally isolated *Rhodococcus* is able to tolerate higher concentration of phenol (b) the immobilized *Rhodococcus* sp. offer better degradation rate and (c) the enzymes and gene encoded for the enzymes could be isolated and characterized. In view of this, the following objectives are outlined:

- 1. To screen and characterize phenol-degrading *Rhodococcus*
- 2. To optimize the degradation of phenol by the selected phenol-degrading *Rhodococcus* using one factor at a time (OFAT) approach and response surface methodology (RSM) by free and immobilized cells.
- 3. To immobilize the phenol-degrading *Rhodococcus* to improve biodegradation efficiency.
- 4. To determine the mechanism of phenol degradation by characterization of the phenol-degrading enzymes
- 5. To evaluate the potential of resting cells for phenol degradation



#### REFERENCES

- Abd El-Zaher, E.H.F., Mahmoud, Y.A.G. and Aly, M.M. 2011. Effect of different concentrations of phenol on growth of some fungi isolated from contaminated soil. *African Journal of Biotechnology* 10(8): 1384-1392.
- Abd-El-Haleem, Desouky, Beshay, U., Abdu O. Hamid, A., Moawad, H. and Zaki, S. 2003. Effects of mixed nitrogen sources on biodegradation of phenol by immobilized *Acinetobacter* sp. strain W-17. *African Journal of Biotechnology* 2(1): 8 12.
- Abdel-Raof, N., Al-Homaidan, A.A. and Ibrahim, I.B.M. 2012. Microalgae and wastewater treatment. *Saudi Journal of Biological Sciences* 19(3): 257-275.
- Abdullah, P. and Nainggolan, H. 1991. Phenolic water pollutants in a Malaysian river basin. *Environmental Monitoring and Assessment* 19(1-3): 423-31.
- Abdullah-Al-Mahin, Alamgir, Z., Chowdhury, M., Alam, K., Aktar, Z. and Fakhruddin, A.N.M. 2011. Phenol biodegradation by two strains of *Pseudomonas putida* and effect of lead and zinc on the degradation process. *International Journal of Environment* 1: 27-33.
- Adepoju, T.F., Olawale, O., Ojediran, O.J. and Layokun, S.K. 2014. Application of response surface methodology (RSM) and artificial neural network (ANN) for achieving desire BA in the biotransformation of benzaldehyde using free cells of *Saccaromyces cerevisae* and the effect of B-cyclodextrin 3: 62-79.
- Adjei, M.D. and Ohta, Y. 1999. Isolation and characterization of a cyanide-utilizing Burkholderia cepacia strain. World Journal of Microbiology and Biotechnology 15: 699-704.
- Agarry, S.E. and Aremu M.O. 2012. Batch equilibrium and kinetic studies of simultaneous adsorption and bidegradation of phenol by pineapple peels immobilized *Pseudomonas aeruginosa* NCIB 950. *British Biotechnology Journal* 2(1): 24-48.
- Agarry, S.E., Durojaiye, A.O. and Solomon, B.P. 2008. Microbial degradation of phenols: A review. *International Journal Environmental Pollution* 32: 12-28.
- Agarry, S.E., Solomon, B.O. and Audu, T.O.K. 2010. Substrate utilization and inhibition kinetics: Batch degradation of phenol by indigenous monoculture of *Pseudomonas aeruginosa*. *International Journal for Biotechnology and Molecular Biology Research* 1(2): 22-30.
- Agency for Toxic Substances and Disease Registry (ATSDR), 2008. Toxicological profile for phenol. Division of toxicology and environmental medicine/applied toxicology branch US Department of health and human services. Georgia, United States 2008 1–20.

- Aggelis, G., Ehaliotis, C., Nerud, F., Stoychev, I., Lyberatos, G. and Zervakis, G.I. 2002. Evaluation of white-rot fungi for detoxification and decolorization of effluents from green olive debittering process. *Applied Microbiology and Biotechnology* 59(2-3): 353-360.
- Aghaie, E., Pazouki, M., Hosseini, M.R., Ranjbar, M. and Ghavipanjeh, F. 2009. Response surface methodology (RSM) analysis of organic acid production for Kaolin beneficiation by *Aspergillus niger*. *Chemical Engineering Journal* 147: 245-251.
- Aghalino, S.O. and Eyinla, B. 2009. Oil exploitation and marine pollution: Evidence from the Niger Delta, Nigeria. *Journal of Human Ecology* 28(3): 177-182.
- Agrawal, N. and Shahi, S.K. 2015. An environmental cleanup strategy-microbial transformation of xenobiotic compounds. *International Journal of Current Microbiology and Applied Sciences* 4(4): 429-461.
- Ahari, H., Razavilar, V., Motalebi, A., A., Akbar, A.A., Kakoolaki, S., Anvar, A.A., Shahbazadeh, D. and Mooraki, N. 2012. DNA extraction using liquid nitrogen in *Staphylococcusaureus*. *Iranian Journal of Fisheries Science* 11: 926-929.
- Ahmad, S.A., Syed, M.A., Norliza, M.A., Abdul Shulor, M.Y. and Nor Aripin, S. 2011. Isolation, identification and characterization of elevated phenol degrading *Acinetobacter* sp. strain AQ5NOL1. *Australian Journal of Basic and Applied Sciences* 5(8): 1035-1045.
- Ahmad, S.A., Shamaan, N.A., Arif, N.M., Koon, G.B., Shukor, M.Y.A. and Syed, M.A. 2012. Enhanced phenol degradation by immobilized *Acinetobacter* sp. strain AQ5NOL 1. *World Journal of Microbiology and Biotechnology* 28: 347–352.
- Ahmed, I., Zia, M.A. and Iqbal, H.M.N. 2011. Purification and kinetic parameters characterization of an alkaline protease produced from *Bacillus subtilis* through submerged fermentation technique. *World Applied Sciences Journal* 12(6): 751–757.
- Aida, M.F. and Hanan M.Abd-Elnaby. 2014. Degradation of phenol by a new-degradable marine halophilic fungus *Fennelia Flavipes* isolated from mangrove sediments. *Life Science Journal* 11(9): 1-10.
- Akcil, A., Karahan, A.G., Ciftci, H., Sagdic, O. 2003. Biological treatment of cyanide by natural isolated bacteria (*Pseudomonas* sp.). *Mineral Engineering* 16: 643–649.
- Alan, E., Tomkinson, Timothy, R.L., Howes and Nathaniel, E.W. 2013. DNA Ligase as therapeutic targets. *Translational Cancer Research* 2 (3): 1-7.
- Ali, N., Hameed, A. and Ahmed, S. 2009. Physicochemical characterization and bioremediation perspective of textile effluent, dyes and metals by indigenous bacteria. *Journal of Hazardous Materials* 164: 322-328.

- Ali, O., Namane, A. and Hellal, A. 2013. Use and recycling of Ca-alginate biocatalyst for removal of phenol from wastewater. *Journal of Industrial Engineering Chemistry* 19(4): 1384–1390.
- Ali, S., Lafuente, R.F. and Cowan, D.A. 1998. Meta-pathway degradation of phenolics by thermophilic *Bacilli*. *Enzyme Microbiology and Technology* 23: 462–468.
- Alimoradzadeh, R., Assadi, A., Nasseri, S. and Mehrasbi, M.R. 2012. Photocatalytic degradation of 4-clorophenol by UV/H<sub>2</sub>O<sub>2</sub>/NiO process in aqueous solution. *Iranian Journal of Environmental Health and Science Engineering* 9: 12.
- Aliyu, S. and Zahangir, A. 2012. Palm Oil Mill Effluent: A Waste or a Raw Material? Journal of Applied Sciences Research 8(1): 466-473.
- Alkarkhi, A.F., Ahmad, A., Ismail, N., and Easa, A.M. 2008a. Assessment of arsenic and heavy metal contents in cockles (*Anadara granosa*) using multivariate statistical techniques. *Journal of Hazardous Materials* 150: 783-789.
- Alkarkhi, A.F., Ahmad, A., Ismail, N., and Easa, A.M. 2008b. Multivariate analysis of heavy metals concentrations in river estuary. *Environmental Monitoring Assessment* 143(1-3): 179-186.
- Alkorta, I., Hernandez-Allica, J., Becerril, J.M., Amezaga, V., Albizu, I. and Garbisu, C. 2004. Recent findings on the phytoremediation of soils contaminated with environmentally toxic heavy metals and metalloids such as zinc, cadmium, lead, and arsenic. *Environmental Science and Biotechnology* 3: 71–90.
- Al-Saleh, E. and Obuekwe, C. 2005. Inhibition of hydrocarbon bioremediation by lead in a crude oil-contaminated soil. *International Biodeterioration and Biodegradation* 56: 1–7.
- Alvarez, H.M. 2010. *Biology of Rhodococcus*, Microbiology Monographs 16. DOI 10.10007/978-3-642-12937-7-1. Springer-Verlag Berlin Heidelberg.
- Alvarez, H.M., Mayer, F., Fabritius, D. and Steinbu chel, A. 1996. Formation of intracytoplasmic lipid inclusions by *Rhodococcus opacus* strain PD630. *Archieves of Microbiology* 165: 377–386.
- Alvarez, H.M., Mayer, F., Fabritius, D. and Steinbu chel, A. 2013. Metabolism of triacylglycerols in *Rhodococcus* species: insight from physiology and molecular genetics. *Journal of Molecular Biochemistry* 2: 69-78.
- Anna J. 1989. FDA safety alert: sodium azide contamination of hemodialysis water supplies. Villforth JC. 16(4): 273.
- Annadurai, G., Balan, S.M. and Murugesan, T. 2000. Design of experiments in the biodegradation of phenol using immobilized *Pseudomonas pictorium* (NICM-2077) on activated carbon. *Bioprocess Engineering* 22: 101-107.

- Annadurai, G., Ling, L.Y. and Lee, J.F. 2007. Biodegradation of phenol by *Pseudomonas pictorum* on immobilized with chitin. *African Journal of Biotechnology* 6(3): 296-303.
- Anselmo, A.M., Mateus, M., Cabral, J.M.S. and Novais, J.M. 1985. Degradation of phenol by immobilized cells of *Fusarium flocciferum*. *Biotechnology Letters* 7(12): 889-894.
- APHA (American Public Health Association). 2005. Standard Methods for the Examination of Water and Wastewater, 20th edn. Washington DC, USA, pp. 540–544.
- Aravindhan, R., Rao, J.R., and Nair, B.U. 2009. Application of a chemically modified green macro alga as a biosorbent for phenol removal. *Journal of Environment Management* 90(5): 1877-1883.
- Arenskotter, M., Broker, D. and Steinbuchel, A. 2004. Biology of the metabolically diverse genus *Gordonia*. *Applied Environmental and Microbiology* 70: 3195-3204.
- Arif, N.M., Ahmad, S.A., Syed, M.A. and Shukor, M.Y. 2012. Isolation and characterization of a phenol-degrading *Rhodococcus* sp. Strain AQ5NOL 2 KCTC 11961BP. *Journal of Basic Microbiology* 53(1): 9-19.
- Arunkumar, D., Anitha, A., Nadu, T., and Arts, N. 2014. Biodegradation of phenol and emulsification properties of native microorganisms from coal carbonization plant. *International Journal of Applied Biology and Pharmaceutical Technology* 4(5): 249–256.
- Aryal, M. and Liakopoulou-Kyriakides, M. 2015. Degradation in aqueous solutions by *Pseudomonas* sp. isolated from contaminated soil of mining industry. *Journal of Water Sustainability* 5(2): 45-57.
- Asadollahzadeh, M., Tavakoli, H. and Torab-mostaedi, M. 2014. Response surface methodology based on central composite design as a chemometric tool for optimization of dispersive-solidification liquid–liquid microextraction for speciation of inorganic arsenic in environmental water samples. *Talanta* 123: 25–31.
- ATSDR, 1998. Toxicological profile for phenol. Atlanta: US Department of Health and Human Services, Agency for Toxic Substances and Disease Registry. Available from: http://www.seagrant.umn.edu/water/report/chemicalsofconcern/phenol/phenol.pdf [Accessed March 2010].
- ATSDR, 2014. Toxicological profile for phenol. Atlanta: US Department of Health and Human Services, Agency for Toxic Substances and Disease Registry. Available from: http://www.atsdr.cdc.gov/toxprofiles/tp115.html [Accessed June 2015].

- Auffret, M., Labbe, D., Thouand, G., Charles, W., Greer and Fayoulle-Guichard, F.2009. Degradation of a mixture of hydrocarbons, gasoline, and diesel oil additives by *Rhodococcus aetherivorans* and *Rhodococcus wratislaviensis*. *Applied Environment Microbiology* 75(24): 7774-7782.
- Awan, Z.U.R., Shah, A. and Rand Amjad, M. 2013. Microbial degradation of phenol by locally isolated soil bacteria. *Global Advanced Research Journal of Microbiology* 2(4): 72-79.
- Aysha, O.S. and Mumtaj, K. 2014. Degradation of phenol by selected strains of *Bacillus* species isolated from marine water. *American Journal of Pharmacy and Health Science* 2(6): 41-56.
- Baek, S., Yin, C., Lee, S. 2001. Aerobic nitrate respiration by a newly isolated phenol degrading bacterium *Alcaligenes* P5. *Biotechnology Letter* 23: 627-630.
- Bajaj, M., Gallert, C. and Winter, J. 2009. Effect of co-substrates on aerobic phenol degradation by acclimatized and non-acclimatized enrichment cultures. *Engineering Life Sciences* 8: 125-131.
- Bak, F. and Widdel, F. 1986. Anaerobic degradation of phenol and phenol derivatives by *Desulfobacterium phenolicum* sp. *Archives of Microbiology* 146: 177–180.
- Balamurugan, P., Preetha, B. and Virithagiri, T. 2012. Study on effect of operating parameters on biodegradation of phenol by *Aspergillus fumigatus*. *International Journal of Engineering Research and Application* 2: 981–986.
- Bandhyopadhyay, K., Dasb, D., Bhattacharyya, P. and Maiti, B.R. 2012. Reaction engineering studies on biodegradation of phenol by *Pseudomonas putida* MTCC 1194 immobilized on calcium alginate. *Biochemical Engineering Journal* 8: 179–186.
- Banerjee, A. and Ghoshal, A.K. 2010. Phenol degradation by *Bacillus cereus*: Pathway and kinetic modelling. *Bioresource Technology* 101: 5501–5507.
- Baradarajan, A., Vijayaraghavan, S., Srinivasaraghavan, T., Musti, S., Kar, S. and Swaminathan, T. 1995. Biodegradation of phenol by *Arthrobacter* and modeling of the kinetics. *Bioprocess Engineering* 12: 227–229.
- Barbas, C.F., Burton, D.R., Scott, J.K. and Silverman, G.J. 2007. Quantitation of DNA and RNA. DOI:10.1101/pdb.ip47Cold Spring Harb Protoc2007.
- Barlow, Janice, and Jo Ann P. Johnson. Fact Sheet on Phenols. Breast Cancer and the Environment Research Centers. November 7, 2007. http://www.bcerc.org/COTCpubs/BCERC.FactSheet\_Phenols.pdf.
- Basak, B., Bhunia, B., Dutta, S., Chakraborty, S. and Dey, A. 2014. Kinetics of phenol biodegradation at high concentration by a metabolically versatile isolated yeast *Candida tropicalis* PHB5. *Environmental Science Pollutant Residue International* 2:1444-1454.

- Basha, K.M., Rajendran, A. and Thangavelu, V. 2010. Recent advances in the biodegradation of phenol: A review. *Asian Journal of Experimental Biological Science* 1(2): 219-234.
- Basile, L.A., and Erijman, L. 2008. Quantitative assessment of phenol hydroxylase diversity in bioreactors using functional gene analysis. *Applied Microbiology Biotechnology* (78): 863-872.
- Baskin, S.I., Kelly, J.B., MalIner, B.I., Rockwood, G.A. and Zoltani, C.K. 2006. Cyanide poisoning. In: *Medical Aspects of Chemical Warfare* pp. 371-410.
- Bayat, Z., Hassanshahian, M., and Cappello, S. 2015. Immobilization of microbes for bioremediation of crude oil polluted environments: A Mini Review. *The Open Microbiology Journal* 9: 48–54.
- BBC News, June 14, 2001. Tanker carrying toxic chemical capsizes. http://news.bbc.co.uk/2/hi/asia-pasific/1387726.stm. [Accessed on 30 October 2012].
- Bell, K.S., Philp, J.C., Aw, D.W. and Christofi, N. 1998. The genus *Rhodococcus*. *Journal of Applied Microbiology* 85: 195-210.
- Benerjee, A. and Goshal, A.K. 2010. Phenol degradation by *Bacillus cereus*: pathway and kinetic modelling. *Bioresource Technology* 101(14): 5501-5507.
- Benoit, F.M. and Lee, V.M.Y. 2000. A new link between pesticides and Parkinson's disease. *Nature Neuroscience* 3: 1227-1229.
- Benov, L. and Al-Ibraheem, J. 2002. Disrupting *Escherichia coli*: A comparison methods. *Journal of Biochemistry and Molecular Biology* 35: 428-431.
- Beshay, U., Abd-El-Haleem, D., Moawad, H. and Zaki, S. 2002. Phenol biodegradation by free and immobilized *Acinetobacter*. *Biotechnology Letters* 24: 1295–1297.
- Bettman, H. and Rehm, H.J. 1984. Degradation of phenol by polymer entrapped microorganisms. *Applied Microbiology and Biotechnology* 20 (5): 285-290.
- Bezerra, M.A., Santelli, R.E., Oliveira, E.P., Villar, L.S. and Escaleira, L.A. 2008. Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta* 75(5): 965-977.
- Bhalla, T.C and Kumar, H. 2005. *Nocardia globerula* NHB-2: a versatile nitrile-degrading organism. *Canada Journal of Microbiology* 51(8): 705-708.
- Bhattacharya, S., Das, A. and Nalin, P. 2012. *Ex situ* biodegradation of phenol by native bacterial flora isolated from industrial effluent. *Journal of Chemical, Biological and Physical Sciences* 2 (2): 1091-1101.

- Bijay, T., Kumar, A.K.C and Anish, G. 2012. A review on bioremediation of petroleum hydrocarbon contamintants in soil. *Journal of Science, Engineering and Technology* 8(1): 164-170.
- Bisht, D., Kumar, Y.S. and Darmwal, S.N. 2013. Optimization of immobilization conditions by conventional and statistical strategies for alkaline lipase production by *Pseudomonas aeruginosa* mutant cells: scale-up at bench-scale bioreactor level. *Turkish Journal of Biology* 37(4): 392–404.
- Bogan, B.W. and Sullivan, W.R. 2003. Physicochemical soil parameters affecting sequestration and mycobacterial biodegradation of polycyclic aromatic hydrocarbons in soil. *Chemosphere* 52: 1717–1726.
- Bonfá, Lindenbah, M.R., Grossman, James, M., Piubeli, Francine, Mellado, Encarnación, Durrant and Regina, L. 2013. Phenol degradation by halophilic bacteria isolated from hypersaline environments. *Biodegradation* 24: 699-709.
- Botz, M.M., Mudder, T.I. and Akcil, A.U. 2005. Cyanide treatment: Physical, chemical and biological processes. *Developments in Minéral Processing* 15:672-701.
- Boumaaza, B., Benkhelifa, M. and Belkhoudja, M. 2015. Effects of two salts compounds on mycelial growth, sporulation, and spore germination of six isolates of *Botrytis Cinerea* in the western north of Algeria. *International Journal of Microbiology* 2015: Article ID 572626
- Boyd, C., Larkin, M.J., Reid, K.A., Sharma, N.D. and Wilson, K. 1997. Metabolism of naphthalene, 1-naphthol, indene, and indole by *Rhodococcus* sp. strain NCIMB 12038. *Applied Environment Microbiology* 63: 151–155.
- Bradford, M.M. 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry* 72: 248–254.
- Brand, D., Pandey, A., Roussos, S. and Soccol, C.R. 2000. Biological detoxification of coffee husk by filamentous fungi using a solid state fermentation system. *Enzyme Microbiology and Technology* 27(1-2): 127-133.
- Brook, R.D., Franklin, B., Cascio, W., Hong, Y., Howard, G., Lipsett, M., Luepker, R., Mittleman, M., Samet, J., Smith, S.C. Jr. and Tager, I. 2004. Air pollution and cardiovascular disease: a statement for healthcare professionals from the expert panel on population and prevention science of the American heart association. *Circulation* 109(21): 2655-2671.
- Bull, S. 2007. Phenol toxicology overview. Health Protechtion Agency. http://www.hpa.org.uk/webc/HPAwebfile/HPAweb\_C/1194947332255. [Accessed on 20 September 2013].

- Busca, G., Berardinelli, S., Resini, C. and Arrighi, L. 2008. Technologies for the removal of phenol from fluid streams: A short review of recent developments. *Journal of Hazardous Materials* 160(2-3): 265–288.
- Buswell, JA. 1975. Metabolism of phenol and cresols by *Bacillus stearothermophilus*. *Journal of Bacteriology* 124(3): 1077–1083.
- Caetano, C.S., Guerreiro, L., Fonseca, I.M., Ramos, A.M., Vital, J. and Castanheiro, J.E., 2009. Esterification of fatty acid to biodiesel over polymers with sulfonic acid groups. *Applied Catalysis and Chemical* 359: 41–46.
- Cai, T., Chen, L., Ren, Q., Cai, S., and Zhang, J. 2011. The biodegradation pathway of triethylamine and its biodegradation by immobilized *Arthrobacter protophormiae* cells. *Journal of Hazardous Materials* 186: 59-66.
- Caldorera-Moore, M. and Peppas, N.A. 2009. Micro and nanotechnologies for intelligent and responsive biomaterial-based medical systems. *Advanced Drug Delivery Review* 61(15): 1391–1401.
- Camelin, I., Lacroix, C., Paquin, C., Prevost, H., Cachon, R. and Divies, C. 1993. Effects of chelants on gellan gum rheological properties and setting temperature for immobilization of living *Bifidobacteria*. *Biotechnology Progress* 9: 291–297.
- Candioti, L.V., Robles, J.C., Mantovani, V.E. and Goicoechea, H.C. 2006. Multiple response optimization applied to the development of a capillary electrophoretic method for pharmaceutical analysis. *Talanta* 69:140-147.
- Cao, J.W, Dong C.M., Cao H.B. and Shao Z.Z. 2011. Isolation of phenol-degrading bacteria from coking waswater and their degradation gene. *Environmental Science* 32(2): 560-566.
- Carls M. G. And Meador J.P. 2009. A perspective on the toxicity of petrogenic PAHs to developing fish embryos related to environmental chemistry. *Human Ecology Risk Assessment* 15: 1084–1098.
- Carp, O., C.L. Huisman and A. Reller. 2004. Photoinduced reactivity of titanium dioxide. *Progress Solid State Chemistry* 32: 33-177.
- Cassidy, M.B., Mullineers, H., Lee, H. and Trevors, J.T. 1997. Mineralization of pentachlorophenol in a contaminated soil by *Pseudomonas* sp. UG30 cells encapsulated in k-carrageenan. *Journal of Industrial Microbiology and Biotechnology* 19: 43–48.
- Castillo, I. del Castillo, P. Herna ndez, A. Lafuente, I.D. Rodri guez-Llorente, Caviedes, M.A. and Pajuel, E. 2012. Self-bioremediation of cork-processing wastewaters by (chloro) phenol-degrading bacteria immobilized onto residual cork particles *Water research* 46: 1723-1734.

- Cejkova, A., Masak, J., Jirku, V., Vesely, M., Patek, M. and Nesvera, J. 2005. Potential of *Rhodococcus erythropolis* as a bioremediation organism. *World Journal of Microbiology and Biotechnology* 21: 317–321.
- Chakraborty, B., Indra, S., Hazra, D., Betai, R., Ray, L. and Basu, S. 2013. Performance study of chromium (vi) removal in presence of phenol in a continuous packed bed reactor by *Escherichia coli* isolated from east Calcutta wetlands. *BioMed Research* 6: 1–9.
- Chakraborty, S., Bhattacharya, T., Patel, T.N. and Tiwari, K.K. 2010. Biodegradation of phenol by native microorganisms isolated from coke processing wastewater. *Journal Environmental Biology* 31: 293-296.
- Chatterjee, S., Das, S.K., Chakravarty, R., Chakrabarti, A. and Ghosh, S. 2010. Interaction of malathion, an organophosphorus pesticide with *Rhizopus oryzae* biomass. *Journal of Hazardous Materials* 174: 47–53.
- Cheetham, P.S.J., Garrett, C. and Clark, L. 1985. Isomaltulose production using immobilized cells. *Biotechnology and Bioengineering* 27:471–481.
- Chen, C., Li, W.Y., Wu, J.W. and Li, J. 2012. Screening and characterization of phenol degrading bacteria for the coking wastewater treatment. *Environmental Science* 33(5): 1652-1656.
- Chen, C.Y., Chen, S.C., Fingas, M. and Kao, C.M. 2008. Biodegradation of propionitrile by *Klebsiella oxytoca* immobilized in alginate and cellulose triacetate gel. *Journal of Hazardous Material* 177(1-3): 856-863.
- Chen, C.Y., Kao, C.M., Chen, S.C., Chien, H.Y. and Lin, C.E. 2007. Application of immobilized cells to the treatment of cyanide wastewater. *Water Science and Technology* 56(7): 99-107.
- Chen, F.Y., Jiang, D. and Tang, Y. 2012. Effects of additional stimulants on phenol degradation by a halotolerant *Rhodococcus* sp. JDD1H. *Applied Mechanical Materials* 914: 253-255.
- Chen, K.C., Lin, Y.H., Chen, E.H. and Lin, Y.C. 2002. Degradation of phenol by PAA-immobilized *Candida topicalis*. *Enzyme Microbial Technology* 31: 490-497.
- Chen, X., Shi, J., Chen, Y., Xu, X., Xu, S. and Wang, Y. 2005. Tolerance and absorption of copper and zinc by *Pseudomonas putida* CZ1 isolated from metal polluted soil. *Candian Journal of Microbiology* 52:308-316.
- Chen, X.H., Wei, G., Liu, S.Y., Sun, J.M., Wang, F.F., Li, H.Y. and Liu, Y.J. 2012. Growth kinetics and phenol degradation of highly efficient phenol-degrading *Ochrobactrum* sp. *Huan Jing Ke Xue* 33(11): 3956-3961.
- Chisti, Y. and Young, M.M. 1986. Disruption of microbial cells for intracellular products. *Enzyme Microbiology Technology* 8: 195-204.

- Cho, Y.G., Rhee, S.K. and Lee, S.T. 2000. Influence of phenol on biodegradation of *p*-nitrophenol by freely suspended and immobilized *Nocardioides* sp. NSP41. *Biodegradation* 11: 21–28.
- Chong C. S. 2011. Biodegradation of RDX in *Rhodococcus* spp. University of York.
- Chung, T.P., Tseng, H.Y., and Hang, R.S. 2003. Mass transfer effect and intermediate detection for phenol degradation in immobilized *Pseudomonas putida* systems. *Process Biochemistry* 38: 1497–1507.
- Clemmitt, R. H. and H. A. Chase .2000. Facilitated downstream processing of a histidine-tagged protein from unclarified *Escherichia coli* homogenate using immobilized metal affinity expanded-bed adsorption. *Biotechnology and Bioengineering* 67: 206-216.
- Clodio, G., Dos-Santos, Marcelo, A., Costa, Willy, A. De Morais and Vânya, M.D. Pasa. 2009. Phenolic foams from wood tar resols. *Journal of Applied Polymer Science* 115(2): 923–927.
- Cole JN, Sanderson-Smith ML, Cork AJ, Henningham A, Conlan F, Ranson M, McArthur JD, Walker MJ. Gene expression and tagging of *Streptococcal* proteins. In: Molecular biology of Streptococci. Hakenbeck R, Chhatwal GS (Ed) Horizon Bioscience 2006; pp 359-378.
- Collins, L.D. and Daugulis, A.J. 1996. Use of two phase partitioning bioreactor for the biodegradation of phenol. *Biotechnology Technique* 10: 643-648.
- Comte, A., Christen, P., Davidson, S., Pophillat, M., Lorquin, J., Auria, R., Simon, G. and Casalot, L. 2013. Biochemical, transcriptional and translational evidences of the phenol-*meta*-degradation pathway by the hyperthermophilic *Sulfolobus solfataricus*. *PLoS ONE* 8(12): e82397.
- Cordova-Rosa, S.M., Dams, R.I., Cordova-Rosa, E.V., Radetski, M.R., Corrêa, A.X.R. and Radetski, C.M. 2009. Remediation of phenol-contaminated soil by a bacterial consortium and *Acinetobacter calcoaceticus* isolated from an industrial wastewater. *Journal of Hazardous Materials* 164: 61–66.
- Cotto, M.C., Emiliano, V., Nieto, S., Duconge, J., Roque-Malherbe, R. 2009. Degradation of phenol by mechanical activation of a rutile catalyst. *Journal of Colloid and Interface Science* 339: 133-139.
- Coutinho, D.F., Sant, S., Shin, H., Oliveira, J.T., Gomes. M.E., Neves, N.M., Khademhosseini, A. and Reis, R.L. 2010. Modified gellan gum hydrogels with tunable physical and mechanical properties. *Biomaterials* 31(29): 7494–7502.
- Coves, J., Niviere, V., Eschenbrenner M, Fontecave M. 1993. NADPH-sulfite reductase from *Escherichia coli*. A flavin reductase participating in the generation of the free radical of ribonucleotide reductase. *Journal of Biology Chemistry* 268: 18604–18609.

- Crawford, D.W., Bonnevie, N.L. and Wenning, R.J. 1995. Sources of pollution and sediment contamination in Newark Bay, New Jersey. *Ecotoxilogy andEnvironmental Safety* 30: 85-100.
- Cunningham, C.J., Ivshina, I.B., Lozinsky, V.I., Kuyukina, M.S., and Philip, J.C. 2004. Bioremediation of diesel contaminated soil by microorganisms immobilized in polyvinyl alcohol. *International Biodeterioration and Biodegradation* 54: 167–174.
- Dash, R.R., Gaur, A., Balomajumder, C. and Kumar, A. 2009. Cyanide in industrial wastewater and its removal: A review on biotransformation. *Journal of Hazardous Materials* 163: 1-11.
- Davies, J. and Davies, D. 2010. Origins and evolution of antibiotic resistance. *Microbiology and Molecular Biology Reviews* 74(3): 417-433.
- De Araujo, B.S., Dec, J., Bollag, J.M. and Pletsch, M. 2006. Uptake and transformation of phenol and chlorophenols by hairy root cultures of *Daucus carota, Ipomoea batatas* and *Solanum aviculare. Hemosphere* 63(4): 642-651.
- De Oliveira-Filho E.C., Lopez R.M. and Paugmgartten F.J. 2004. Comparative study on the susceptibility of freshwater species to copper-based pesticides. *Chemosphere* 56(4): 369-374.
- Dean-Ross, D. and Rahimi, M. 1995. Toxicity of phenolic compounds to sediment bacteria. *Bulletin of Environmental Contamination and Toxicology* 55: 245-250.
- Debadatta, D., Susmita, M., and Engineering, M. 2012. Simultaneous reduction of phenol and chromium from textile industry effluent using mixed culture. Journal of Environmental Research and Development 7(2): 946–957.
- Dekant W. 2009. The role of biotransformation and bioactivation in toxicoxity. *EXS* 99: 57-86.
- Denome, S.A., Oldfield, C., Nash, L.J. and Young, K.D. 1994. Characterization of the desulfurization genes from *Rhodococcus* sp. strain IGTS8. *Journal of Bacteriology* 176: 6707–6716.
- DOE. 2011. Malaysia Environmental Quality Report 2011. Department of Environment, Ministry of Natural Resources and Environment Malaysia. Publication section Strategic Communication Division, Department of Environment Malaysia. pp 65.
- DeRito C.M. and Madsen E.L. 2009. Stable isotope probing reveals *Trichosporon* yeast to be active in situ in soil phenol metabolism. *ISME Journal* 3(4): 477-485.

- Desai, C., Pathak, H. and Madamwar, D. 2010. Advances in molecular and '-omics' technologies to gauge microbial communities and bioremediation at xenobiotic/anthropogen contaminated sites. *Bioresource Technology* 1(6): 1558-1569.
- Dey, S. and Mukherjee, S. 2010. Performance and kinetic evaluation of phenol biodegradation by mixed microbial culture in a batch bioreactor. *International Journal Water Resources Environmental Engineering* 2(3): 40–49.
- Dhatwalia, V.K. and Nanda, M. 2016. Biodegradation of phenol: Mechanisms and applications. In Rathoure, A. and Dhatwalia V.K (Eds.) *Toxicity and Waste Management Using Bioremediation*. Hershey, PA: Engineering Science Reference, pp. 198-214. DOI:10.4018/978-1-4666-9734-8.ch010
- Dikshitulu, S., Baltzis, B.C., Lewandowski, G.A. and Pavlou, S. 1993. Compitition between two microbial populations in a sequencing fed-batch reactor: theory, experimental verification, and implications for waste treatment applications. *Biotechnology Bioengineering* 42: 643-656.
- Duffner, F.M., U. Kirchner, M.P. Bauer and R. Muller, 2000. Phenol/cresol degradation by the thermophilic *Bacillus thermoglucosidasius* A7: cloning and sequence analysis of five genes involved in the pathway. *Gene* 256: 215-221.
- Duque, A.F., Hasan, S.A., Bessa, V.S., Carvalho, M.F., Samin, G., Janssen, D.B. and Castro, P.M. 2012. Isolation and characterization of a *Rhodococcus* strain able to degrade 2-florophenol. *Applied Microbiology and Biotechnology* 95(2): 511-520.
- Duràn, N. and Esposito, E. 2000. Potential application of oxidative enzymes and phenoloxidase-like compounds in wastewater and soil treatment: A review. *Applied Catalysis Environmental* 28: 89-99.
- Dursun, A.Y. and Tepe, O. 2005. Internal mass transfer effect on biodegradation of phenol by Ca-alginate immobilized *Ralstonia eutrophe*. *Journal of Hazardous Materials* 126: 105–111.
- Edalatmanesh, M., Dhib, R. and Mehrvar, M. 2008. Kinetic modelling of aqueous phenol degradation by UV/H<sub>2</sub>O<sub>2</sub> process. *International Journal of Chemical Kinetics* 40(1): 34-43.
- Elahwani, M.D. and Mabrouk, M.E. 2013. Isolation and characterization of phenol-degrading strain of *Alcaligenes* sp. AM4. *Acta Biologica Hungarica* 64(1): 106-117.
- El-Deeb, B. and Altalhi, A.D. 2009. Natural combination of genetic systems for degradation of phenol and resistance to heavy metals in phenol and cyanide assimilating bacteria. *Malaysian Journal of Microbiology* 5(2): 94–103.

- Elibol, M., and Moreira, A.R. 2003. Production of extracellular alkaline protease by immobilization of the marine bacterium *Teredinibacterturnirae*. *Process Biochem*istry 38: 1445–1450.
- El-Katatny, M.H., Ahmed, M.H., Gehan, M.S. and El-Komy, H.M. 2003. Enzyme production by alginate encapsulated *Trichoderma* sp. *Food Technology and Biotechnology* 41: 219–225.
- El-Naas, M.H., Al-Muhtaseb, S.A. and Makhlouf, S. 2009. Biodegradation of phenol by *Pseudomonas putida* immobilized in polyvinyl alcohol (PVA) gel. *Journal of Hazardous Materials* 164(2–3): 720-725.
- Essam, T., Amin, Tayeb, M.A., Bo Mattiasson, O.E. and Guieysse, B. 2010. Kinetics and metabolic versatility of highly tolerant phenol degrading *Alcaligenes* strain TW1. *Journal of Hazardous Materials* 173: 783–788.
- Evans, W.C. 1947. Oxidation of phenol and benzoic acid by some soil bacteria. Journal of Biological Chemistry 41: 373-382.
- Faizal., I., Ohba, M., Kuroda, A., Takiguchi, N., Ohtake, H., Honda, K. and Kato, J. 2007. Bioproduction of 3-methylcatechol from toluene in a two-phase (organic-aqueous) system by a genetically modified solvent-tolerant *Pseudomonas putida* strain T-57. *Journal of Environmental Biotechnology* 7(1): 39-44.
- Farliahati, M.R., Ramanan, R.N., Mohamad, R., Puspaningsih, N.N.T. and Ariff, A.B. 2010. Enhanced production of xylanase by recombinant *Escherichia coli* DH5α through optimization of medium composition using response surface methodology. *Annals of Microbiology* 60: 279–285.
- Fiaolova, A., Boschke, E. and Bley, T. 2004. Rapid monitoring of the biodegradation of phenol-like compound by the yeast *Candida maltosa* using BOD measurement. *International Biodeterioration and Biodegradation* 4: 69-76.
- Fijalkowska, S., Katarzyna, L. and Dlugonski, J. 1998. Bacterial elimination of polycyclic aromatic hidrocarbons and heavy metals. *Journal of Basic Microbiology* 38: 361-369.
- Florin, O. and Sandalescu, M. 2006. Phenol removal from wastewater using ion exchange adsorption. *Environmental Engineering and Management Journal* 5(5): 1051.
- Folsom, B.R., Chapman, P.J. and Pritchard P.H. 1990. Phenol and trichloroethylene degradation by *Pseudomonas cepacia* G4: Kinetics and interactions between substrates. *Applied Environmental Microbiology* 56 (5): 1279-1285.
- Futamata, H., Harayama, S., Watanabe, K. 2001. Group-specific monitoring of phenol hydroxylase genes for a functional assessment of phenol-stimulated trichloroethylene bioremediation. *Applied and Environmental Microbiology* 67(10): 4671-4677.

- Gaal, A. and Neujahr, H.Y. 1979. Metabolism of phenol and resorcinol in *Trichosporon cutaneum. Journal of Bacteriology* 137: 13-21.
- Galindez-Mayer, J., Ram on Gallegos, J., Ruiz-Ordaz, N., Ju'rez-Ram'irez., C., Salmer' on Alcocer, A. and Poggi-Varaldo, H.M. 2008. Phenol and 4-chrorophenol biodegradation by yeast *Candida tropicalis* in fluidized bed reactor. *Biochemical Engineering Journal* 38: 147-157.
- Gami, A.A, Shukor, M.Y., Khalil, K.A, Dahalan, F.A, Khalid, A. and Ahmad S.A. Phenol and its toxicity. *Journal of Environmental Microbiology and Toxicology* 2(1): 11-24.
- Gan, C.L., Francis, C., Cha, B.K. and Uda Hashim. 2013. Extended reliability of gold and copper ball bonds in microelectronic packaging. *Gold Bulletin* 46(2): 103-115.
- Ganguly, S. and G. Pierce. 2005. Enhanced Stabilization of Nitrile Hydratase Enzyme from *Rhodococcus* Species DAP 96253 and *Rhodococcus* rhodochrous DAP 96622. Georgia State University.
- Gani, S.A. 2011. Biodegradation of diesel by *Rhodococcus* sp. strain SeAg. Universiti Putra Malaysia.
- GAO 2010. Superfund: Interagency Agreements and Improved Project Management Needed to Achieve Cleanup Progress at Key Defense Installations, GAO-10-348 Washington, D.C. July 15, 2010.
- Garg, S.K., Tripathi, M. and Srinath, T. 2012. Strategies for chromium bioremediation of tannery effluent. *Environmental Contamination and Toxicology* 217: 75-140.
- Geciova, J., Bury, D. and Jelen, P. 2002. Methods for disruption of microbial cells for potential use in the dairy industry. *International Dairy Journal* 12: 541-533.
- Ghanem, K.M., Al-Garni, S.M. and Al-Shehri, A.N. 2009. Statistical optimization of cultural conditions by response surface methodology for phenol degradation by a novel *Aspergillus flavus* isolate. *African Journal of Biotechnology* 8(15): 3576-3583.
- Gibson, J. and Harwood, C.S. 2002. Metabolic diversity in aromatic compound utilization by anaerobic microbes. *Annual Review of Microbiology* 56: 345-369.
- Godjevargova, T., Ivanova, D., Alexieva, L., Dimova, D. 2003. Biodegradation of toxic organic components from industrial phenol production waste waters by free and immobilized *Trichosporon cutaneum* 57. *Process Biochemistry* 38: 915-920.
- Goldberg S. 2008. Mechanical/physical methods of cell disruption and tissue homogenization. *Methods molecular biology* 424: 3-22.

- González, G., Herrera, G., García, T. and Peña, M. 2001. Biodegradation of phenolic industrial wastewater in a fluidized bed bioreactor with immobilized cells of *Pseudomonas putida*. *Bioresedual Technology* 80: 137-142.
- Goswami, M., Shivaraman, N. and Singh, R.P. 2005. Microbial metabolism of 2-chlorophenol, phenol and q-cresol by *Rhodococcus erythropolis* M1 in coculture with *Pseudomonas fluorescens* P1. *Microbiological Research* 160: 101–109.
- Gotz, P. and Reuss, M. 1997. Dynamics of microbial growth: Modeling time delays by introducing polymerization reaction. *Journal of Biotechnology* 58: 101-114.
- Gracioso, H., Louise; Regina, Avanzi, Ingrid, P.G. Baltazar, Marcela; Martins-Pinheiro, Marinalva; Karolski, Bruno, Anita Mendes, Maria, Frederico Martins Menck, Carlos, Augusto Oller Nascimento, Claudio and Aquino Perpetuo, Elen. 2012. Proteome analysis of phenol-degrading *Achromobacter* sp. Strain c-1, isolated from an industrial area. *Current Proteomics* 9(4): 280-289.
- Grund, E., Denecke, B. and Eichenlaub, R. 1992. Naphthalene degradation via salicylate and gentisate by *Rhodococcus* sp. strain B4. *Applied Environment and Microbiology* 58: 1874–1877.
- Guiseley, K.B. 1989. Chemical and physical properties of algal polysaccharides used for cell immobilization. *Enzyme and Microbial Technology* 11: 706–716.
- Guosheng, L., Xiaojie, H., Peiling, D., Yingand, H. and Wenyan, Q. 2011. Identification and characteristics of a phenol-degrading bacterium UW7. *Chinese Journal of Applied Environmental Biology* 17(1): 118-120.
- Gupta, N., Balomajumder, C. and Agarwal, V.K. 2010. Enzymatic mechanism and biochemistry for cyanide degradation: A review. *Journal of Hazardous Materials* 176: 1-13.
- Gupta, R and Rajpal, T. 2012. Concise notes on biotechnology. Tata McGraw Hill Education Private Limited. New Delhi. pp 13.4-13.8.
- Gurtler, V., Mayall, B. C., and Seviour, R. 2004. Can whole genome analysis refine the taxonomy of the genus *Rhodococcus?*. *FEMS Microbiology Reviews* 28(3): 377–403.
- Gurujeyalakshmi, G. and Oriel, P. 1989. Isolation of phenol-degrading *Bacillus* stearothermophilus and partial characterisation of the phenol hydroxylase. *Applied Environmental Microbiology* 55: 500-502.
- Hagea, N., Jonathan G., Renshawb, G., Winklera, S., Gellertc, P., Stolnika, S., Franco, H.P. 2015. Improved expression and purification of the *Helicobacter pylori* adhesin BabA through the incorporation of a hexalysine tag. *Protein Expression and Purification* 106: 25-30.

- Halmi, M.I.E., Ahmad S.A., Syed, M.A., Shamaan, N.A. and Shukor, M.Y. 2014. Mathematical modelling of the molybdenum reduction kinetics in *Bacillus pumilus* strain Lbna. *Bulletin of Environmental Science and Management* 2(1): 24–29.
- Hank, D., Saidani, N., Namane, A. and Hellal, A. 2010. Batch phenol biodegradation study and application of factorial experimental design. *Journal of Engineering Science and Technology Review* 3(1): 123-127.
- Harayama, S. and Timmis, K.N. 1992. Aerobic Biodegradation of aromatic hydrocarbons by bacteria. In: Sigel, H. and Sigel, A. (Eds.) *Metal Ions in Biological Systems. Degradation of Environmental Pollutants by Microorganisms and Their Metalloenzymes*. Marcel Dekker Inc. New York. pp. 99-165.
- Harvey, P.J., Campanella, B.F., Castro, P.M.L., Harms, H., Lichtfouse, E., Schäffner, A.R., Smrcek, S. and Werck-Reichhart, D. 2002. Phytoremediation of polyaromatic hydrocarbons, anilines and phenols. *Environmental Science and Pollution Research* 9: 29-47.
- Hattab, G., Suisse, A.Y.T., Ilioaia, O., Casiraghi, M., Warnet, X.L., Miroux, B.
  Warschawski, D.E., Moncoq, K., Zoonens, M. and Miroux, B. 2014.
  Membrane Protein Production in Escherichia coli: Overview and Protocols,
  In: Mus-Veteau, I. (Ed.), Membrane Proteins Production for Structural Analysis. Springer Science Business Media, New York. pp. 87-106.
- Heinaru, E., Heinaru, A., Truu, J. Stottmesiter, U. and Heinaru, A. 2000. Three types of phenol and p-cresol catabolism in phenol- and p-cresol-degrading bacteria isolated from river water continuously polluted with phenolic compounds. *FEMS Microbiology Ecology* 31: 195-205.
- Hernandez-perez, G. Fayolle, F., vandecasteele, J-P. 2001. Biodegradation of ethyl- *t*-butyl ether (ETBE), methyl *t*-butyl ether (MTBE) and *t*-amyl ether (TAME) by *Gordonia Terue*. *Applied Microbiology Biotechnology* 55: 117-121.
- Hill, G.A and Robinson, C.W. 1975. Substrate inhibition kinetics, phenol degradation by *Pseudomonas putida*. *Biotechnology Bioengineering* 17: 1599-1615.
- Hinteregger, C.R., Leinter, M., Loidl, A. and Streichshier, F. 1992. Degradation of phenol and phenolic compounds by *Pseudomonas* EKII. *Applied Microbiology Biotechnology* 37: 252–295.
- Hoffman, A.S. 2002. Hydrogels for biomedical applications. *Advanced Drug Delivery Review* 54(1): 3-12.
- Hofmann, K.H. and Kruger, A. 1985. Induction and inactivation of phenol hydroxylase and catechol oxygenase in *Candida maltosa* L4 in dependence on the carbon source. *Journal of Basic Microbiology* 25: 373-379.

- Hori, K., Atsushi, K., Hiroshi, I. and Hajime, U. 2009. *Rhodococcus aetherivorans* IAR1, a new bacterial strain synthesizing poly(3-hydroxybutyrate-co-3-hydroxyvalerate) from toluene. *Journal of Bioscience and Bioengineering* 107(2): 145-150.
- Hossain, M.M., Jahangir, R., Raquibul Hasan, S.M., Akter, R., Ahmed, T., Imamul Islam, Md., Faruque, A. 2009. Antioxidant, analgesic and cytotoxic activity of *Micheliachampa* linn. Leaf. *Stamford Journal of Pharmaceutical Sciences* 2(2): 1-7.
- HSDB. 2013. Hazardous Substances Data Bank. *National Library of Medicine*, *National Toxicology Information Program*, Bethesda, M.D.
- Hsu, P.D., Lander, E. S. and Zhang, F. 2014. Development and Applications of CRISPR-Cas9 for Genome Engineering. *Leading Edge Review* 157: 1262-1278.
- Huang, W.Y., Zhang, H.C., Liu, W.X., Li, C.Y. 2012. Survey of antioxidant capacity and phenolic composition of blueberry, blackberry and strawberry in Nanjing. *Journal of Zhejiang University Science Biology Biomedical and Biotechnology* 13(2): 94–102. DOI: 10.1631/jzus.B1100137.
- Hupert-kocurek, K., and Guzik, U. 2012. Characterization of catechol 2,3-dioxygenase from *Planococcus*. Acta Biochimica Polonic 59(3): 345-352.
- Ingelman, M., Ramaswamy, S., Niviere, V., Fontecave, M., Eklund, H. 1999. Crystal structure of NAD(P)H:flavin oxidoreductase from *Escherichia coli*. *Biochemistry* 38: 7040–7049.
- International agency for research in cancer. IARC monograph on the evaluation of carcinogenic risks to human. Phenol. http://monographs.iarc.fr/ENG/Monographs/vol71/mono71-33.pdf.
  [Accessed on 12 October 2014].
- Iyagba, M.A., Adoki, A., and Sokari, T.G. 2008. Testing biological methods to treat rubber effluent. *African Journal of Agriculture Residue* 3(6): 448-454.
- Jabbar, M.E., Defiery, A., and Reddy, G. 2014. Influence of metal ions concentration on phenol degradation by *Rhodococcus pyridinivorans* GM3. *Mesopotamia Environmental Journal* 1(1): 30–38.
- Jacob, H.J. and Sohail, A. 2010. Isolation of two fungal strains capable of phenol biodegradation. *Journal of Biological Science* 10(2): 162-165.
- Jacobs, I.D. and Chirwa, E.M.N. 2015. Estimation of reaction parameters for phenol biodegradation using trainable artificial neural networks DOI:10.3303/CET1545208
- James, C.A and Strand, S.E. 2009. Phytoremediation of small organic contaminants using transgenic plants. *Current Opinion Biotechnology* 20(2): 237-41.

- Janice and Jo. 2007. Fact Sheet On Phenols. Bulletin Of Breast Cancer and The Environment Research Centers, pp 9-20.
- Janke, D, Al-Mofarji, T., Straube, G., Schuman, Pn. and Prauser, H. 1988. Critical steps in degradation of chloroaromatics by rhodococci. *Journal of Basic Microbiology* 28: 509-528.
- Jiang, B., Zhou, Z., Dong, Y., Wang, B., Jiang, J., Guan, X., and Sun, H. 2015. Bioremediation of petrochemical wastewater containing BTEX compounds by a new immobilized bacterium *Comamonas* sp. JB in magnetic gellan gum. *Applied Biochemistry and Biotechnology* 176(2): 572-581.
- Johnson, G., Jain, R. and Spain, J. 2000. Properties of the trihydroxytoluene oxygenase from *Burkholderia cepacia* R34: an extradiol dioxygenase from the 2,4- dinitrotoluene pathway. *Archieves Microbiology* 173: 86-90.
- Joseph, B., Ramteke, P.W. and Kumar, P.A. 2006. Studies on the enhanced production of extracellular lipase by *Staphylococcus epidermidis*. *The Journal of General and Applied Microbiology* 52: 315–320.
- Joutey, N.T., Bahafid, W., Sayel, H. and Ghachtouli, N.E. 2013. Biodegradation: Involved Microorganisms and Genetically Engineered Microorganisms. In: Biodegradation-Life Science. Chamy, R. and Rosenkranz, F. (Eds). ISBN: 978-953-51-1154-2, InTech. DOI: 10.5772/56194.
- Kalil, S.J., Maugeri, F. and Rodrigues, M.I. 2000. Response surface analysis and simulation as a tool for bioprocess design and optimization. *Journal of Process Biochemistry* 35: 539–550.
- Kamble, P.D., Raul, A.A. and Gandhi, M.B. 2011. Isolation and characterization of vegetable oil degrading bacteria. *Journal of Pure Applied Microbiology* 5: 857-890.
- Kampfer, P. 1995. An efficient method for preparation of extracts from Gram-positive bacteria for comparison of cellular protein patterns. *Journal of Microbiological Methods* 21: 55-60.
- Kao, C.M., Liu, J.K., Lou, H.R., Lin, C.S. and Chen, S.C. 2003. Biotransformation of cynide to methane and ammonia by *Klebsiella oxytoca*. *Chemospere* 50: 1055-1061.
- Karigar, C., Mahesh, A., Nagenhalli, M. and Yun, D.J. 2006. Phenol degradation by immobilized cells of *Arthrobacter citreus*. *Biodegradation* 17: 47-55.
- Kashif, N. and Ouyang, F. 2009. Effect of various additives on photocatalytic degradation of 4-nitrophenol. *E-Journal of Chemistry* 6(S1): S422-S428.
- Kavitha, V. and Palanivelu, K. 2004. The role of ferrous ion in Fenton and photo Fenton processes for the degradation of phenol. *Chemospere* 55: 1235-1243.

- Keusgen, M., Milka, P. and Krest, I. 2001. Cyanidase from bacterial sources and its potential for the construction of biosensors. In: *Proceedings of the Biosensor Symposium, Germany*.
- Khan M.A and Gouri A.M. 2011. Environmental Pollution: Its effects on life and its remedies. *Journal of Arts, Science and Commerce* 2: 276-292.
- Khan, F. and Cameotra, S.S. 2013. Aerobic degradation of 2-hexanone by a *Rhodococcus* sp. strain mb-p1 via novel pathway. *Petroleum and Environmental Biotechnology* 4: 151. DOI:10.4172/2157-7463.1000151
- Khleifat, K.M. 2006. Biodegradation of phenol by *Ewingella americana*: Effect of carbon starvation and some growth conditions. *Process Biochemistry* 41: 2010-2016.
- Khokhawala, I.M. and Gogate, P.R. 2010. Degradation of phenol using a combination of ultrasonic and UV irridiations at pilot scale operation. *Ultrasonic Sonochemistry* 17(5): 833-838.
- Kilbane, Z., Vaas, P.R., O'Cuív, P. and O'Connor, B. 2007. Cloning and heterologous expression of bovine pyroglutamyl peptidase type-1 in *Escherichia coli*: purification, biochemical and kinetic characterisation. *Molecular and Cellular Biochemistry* 297: 189-197.
- Kim, B.Y. and Hyun, H.H. 2002. Production of acrylamide using immobilized cells of *Rhodococcus rhodochrous* M33. *Biotechnology Bioprocess Enggineering* 7: 194-200.
- Kim, D., Chae, J.C., Zlystra, G.J., Kim, Y.S., Kim, S.K., Nam, M.Y., Kim, Y.M. and Kim, E. 2004. Identification of a novel dioxygenase involved in metabolism of o-xylene, toluene and ethylbenzene by *Rhodococcus* sp. strain DK17. *Applied Environment Microbiology* 70: 7086–7090.
- Kim, D., Kim, Y.S., Kim, S.K., Kim, S.W., Zylstra, G.J., Kim, Y.M. and Kim, E. 2002. Monocyclic aromatic hydrocarbon degradation by *Rhodococcus* sp. strain DK17. *Applied Environmental Microbiology* 68: 3270-3278.
- Kim, E. and Zylstra, G.J. 1995. Molecular and biochemical characterization of two meta-cleavage dioxygenases involved in biphenyl and m-xylene degradation by *Beijerinckia* sp. strain B1. *Journal Bacteriology* 177: 3095-3103.
- Kim, S.J., Choi, D.H., Sim, D.S. and Oh, Y.S. 2005. Evalution of bioremediation effectiveness on crude-oil contaminated sand. *Chemosphere* 59(6): 845-852.
- Kis, A., Laczi, K., Zsíros, S., Rakhely, G. and Perei, K. 2015. Biodegradation of animal fats and vegetable oils by *Rhodococcus erythropolis* PR4. *International Biodeterioration and Biodegradation* 105:114-119.

- Klimek-Ochab, M., Brzezińska-Rodak, M., Zymańczyk-Duda, E., Lejczak, B. 2011. Comparative study of fungal cell disruption-scope and limitations of the methods. *Folia Microbiology* 56: 469-475.
- Ko, F.W. and Hui, D.S. 2009. Outdoor air pollution: impact on chronic obstructive pulmonary disease patients. *Current Opinion Pulmanory Medicine* 15(2): 150-157.
- Kobayashi, M. and Shimizu, S. 1998 Metalloenzyme nitrile hydratase: structure, regulation and application to biotechnology. *Nature Biotechnology* 16: 733-736.
- Kocher, G. and Mishra, S. 2009. Immobilization of *Bacillus circulans* MTCC 7906 for enhanced production of alkaline protease under batch and packed bed fermentation conditions. *The Internet Journal of Microbiology* 7(2): 459–378.
- Konsoula, Z. and Kyriakides, M.L. 2006. Thermostable alpha-amylase production by *Bacillus subtilis* entrapped in calcium alginate gel capsules. *Enzyme and Microbial Technology* 39(4): 690–696.
- Kotresha, D. and Vidyasagar, G.M. 2008. Isolation and characterization of phenol-degrading *Pseudomonas aeruginosa* MTCC 4996. World Journal of Microbiology and Biotechnology 24: 541-547.
- Kovacs, A., Kende, A., Mortl, M., Volk, G., Rikker, T. and Torkos, K.J. 2008. Determination of phenolic compounds in water and urine samples using solid-phase microextraction based on sol-gel technique prior to GC-FID. *Chromatographia* 1194: 139-142.
- Kowalska, M., Bodzek, M. and Bochdziewicz, J. 1998. Biodegradation of phenols and cyanides using membranes with immobilized microorganisms. *Process Biochemistry* 33: 189-197.
- Krastanov, A., Alexieva, Z. and Yemendzhiev, H. 2013. Microbial degradation of phenol and phenolic derivatives. *Engineering in Life Sciences* 13(1): 76-87.
- Krug, M. and Straube, G. 1986. Degradation of phenolic compounds by the yeast *Candida tropicalis* HP15: Some properties of the first two enzymes of the degradation pathway. *Journal Basic Microbiology* 26: 271-281.
- Kumar, A., Kumar, S. and Kumar, S. 2005. Biodegradation kinetics of phenol and catechol using *Pseudomonas putida* MTCC 11894. *Biochemical Engineering Journal* 22: 151-159.
- Kumar, B., Smita, K. and Flores, C.L. 2014. Plant mediated detoxification of mercury and lead. *Arabian Journal of Chemistry*.DOI: org/10.1016/j.arabjc.2013.08.010

- Kumar, P., Nikakhtari, H., Nemati, M. Hill, G. and Headley, John. 2010. Oxidation of phenol in a bioremediation medium using chlorine dioxide. *Journal of Chemical Technology and Biotechnology* 85(5): 720-725.
- Kumari, A., Majumder, C.B. 2015. Phytoremediation of cyanide and phenol from wastewater by *E. crassipes* (water hyacinth). *Ijcbs Research Paper* 2(3): 42-53.
- Kumari, S., Chetty, D., Ramdhani, N. and Bux, F. 2013. Phenol degrading ability of *Rhocococcus pyrinidivorans* and *Pseudomonas aeruginosa* isolated from activated sludge plants in South Africa. *Journal of Environmental Science and Health, Part A: Toxic/Hazardous Substances and Environmental Engineering* 48(8): 947-953.
- Kumi, A., Khan, V. and Ankumah, R. 2013. Assessing the effects of solarization and sodium azide amendments on selected soil parameters, enzyme activities and microbial populations. *Journal of Environmental Protection* 4(8): 772-778.
- Kunimoto, S., Nosaka, C. and Takeuchi, T. 1999. Stimulation of cellular XTT reduction by cytochrome oxidase inhibitors. *Biological and Pharmaceutical Bulletin* 22: 660-661.
- Kuntiya, Shinji Takenaka and Phisit Seesuriyachan. 2013. High potential of thermotolerant *Candida tropicalis* no. 10 for high concentration of phenol biodegradation. *Food and Applied Bioscience Journal* 1(2): 59-68.
- Kunz, A., Reginatto, V.E., Duran, N. 2001. Combined treatment of textile effluent using the sequence *Phanerochaete chrysosporium ozone*. Chemosphere 44: 281-287.
- Kushairi, A. and Parveez, G.K.A. 2009. Development of value-added products in oil palm via genetic modification and non-genetic modification routes. In: Accelerating Commercialization in Biotechnology, Kuala Lumpur Convention Centre.
- Larkin, J.M., Kulakov, L.A and Allen, C. 2005. Biodegradation and *Rhodococcus*—masters of catabolic versatility. *Current Opinion in Biotechnology* 16: 282-290.
- Larkin, M.J., Kulakov, L.A. and Allen, C.C. 2006. Biodegradation by Members of the Genus *Rhodococcus*: Biochemistry, Physiology, and Genetic Adaptation. *Advances in Applied Microbiology* 59: 1-29.
- Lashkarian, H., JamshidRaheb, Kiana Shahzamani, Hossein Shahbani and Mehdi Shamsara. 2010. Extracellular cholesterol oxidase from *Rhodococcus* sp.: isolation and molecular characterization. *Iran Biomedical Journal* 14 (1 & 2): 49-57.
- Laxamana R.Y. and Gopalacharyulu, V.P. 2006. http://www.lce.hut.fi/teaching/S-114.2500/s2006/LaxmanAndGopal\_S114\_2500.pdf

- Lazarova, N., Valladares, A.G., Georgieva, N., and Müller, R. 2015. Phenol degradation by *Trichosporon cutaneum* r57 in the presence of copper ions. *Journal of Chemical Technology and Metallurgy*. 50(5): 613-618.
- Lee, C.M., Lu, C.J. and Chuang, M.S. 1994. Effects of immobilized cells on the biodegradation of chlorinated phenol. *Water Science and Technology* 30(9): 87–90.
- Lee, S.H., Choi, J.I., Park, I., Lee, S.Y. and Park, B.C. 2004. Display of bacterial lipase on the *Escherichia coli* cell surface by using fadl as an anchoring motif and use of the enzyme in enantioselective Biocatalysis. *Applied and Environmental Microbiology* 70(9): 5074–5080.
- Leemhuis, H., Kelly, R.M., Dijkhuizen, L. 2009. Directed evolution of enzymes: library screening strategies. *IUBMB Life* 61: 222–228.
- Leitao, A.L., Duarte, M.P. and Oliveira, J.S. 2007. Degradation of phenol by a halotolerant strain of *Penicillium chrysogenum*. *International Bioterioration Biodegradation* 59: 220-225.
- Leonard, D. and Lindley, N.D. 1998. Carbon and energy flux constraints in continuous cultures of *Alcaligenes eutrophus* grown on phenol. *Microbiology* 144(1): 241-248.
- Leonard, D., Benyouseff, C., Destruhaut, C., Lindly, N.D. and Queinnec, I. 1999. Phenol degradation by *Ralstonia eutropha*: colorimetric determination of 2-hydroxymuconate semi aldehyde accumulation to control feed strategy in fed. *Biotechnology and Bioengineering* 65: 407-415.
- Lesage-meessen, L., Navarro, D., Maunier, S., Sigoillot, J-C., Lorquin, J., Delattre, M., Simon, J-L., Asther, M. and Labat, M. 2001. Simple phenolic content in olive oil residues as a function of extraction systems. *Food Chemistry* 75(4): 501-507.
- Li, N., Jiang, J., Chen, D., Xu, Q., Li, H. and Lu, J. 2015. A reusable immobilization matrix for the biodegradation of phenol at 500 mg/L. *Scientific reports* 1: 1-5.
- Li, X.Y., Cui, Y.H., Feng, Y.J., Xie, Z.M. and Gu, J.D. 2005. Reaction pathways and mechanisms of the electrochemical degradation of phenol on different electrodes. *Water Research* 39: 1972-1981.
- Li, Y., Li, J., Wang, C. and Wang, P. 2010. Growth kinetics and phenol biodegradation of psychrotrophic *Pseudomonas putida* LY1. *Bioresource Technology* 101(17): 6740-6744.
- Li, Z., Wu, M., Jiao, Z., Bao, B. and Lu, S. 2004. Extraction of phenol from wastewater by N-octonoylpyrrolidine. *Journal of Hazardous Materials* 114(1-3): 111-114.

- Liang, Y.T., Zhang, X., Dai, D.J. and Li, G.H. 2009. Porous biocarrier-enhanced biodegradation of crude oil contaminated soil. *International Biodeterioration and Biodegradation* 63: 80–87.
- Lichtinger, T., Reiss, G., and Benz, R. 2000. Biochemical identification and biophysical characterization of a channel-forming protein from *Rhodococcus erythropolis*. *Journal of Bacteriology* 182: 764-770.
- Lika, K. and Papadakis, I.A. 2009. Modelling the biodegradation of phenolic compounds by microalgae. *Journal of Sea Research* 62:135-146.
- Liming, W., Li Y., Yu. P., ZHixiong, X., Yunbai, L. and Yanwen, Lin. 2010. Biodegradation of phenol at high concentration by a novel fungal strain *Paecilomyces variotii* JH6. *Journal of Hazardous Materials* 183: 366-371.
- Lin, W.C., Chen, Y.S. and Cheng, W.Y. 2006. Effect of metals on biodegradation kinetics for methytert-butyl ether. *Biochemical Engineering Journal* 32: 25-32.
- Liu, F.Z., Ming Z.G., Wang, J., Zhong, H., Ying D.Y. and Zhong Y.X. 2010. Effects of monorhamnolipid and Tween 80 on the degradation of phenol by *Candida tropicalis. Process Biocehmistry* 45: 805-809.
- Liu, J., Wang, Q., Yan, J., Qin, X., Li, L. and Xu, W. 2012. Isolation and Characterization of a Novel Phenol Degrading Bacterial Strain WUST-C1. *Industrial Engineering Chemical Research* 52: 258-265.
- Liu, L., Schmid, R.D., Urlacher, V.B. 2006. Cloning, expression, and characterization of a self-sufficient cytochrome P450 monooxygenase from *Rhodococcus ruber* DSM 44319. *Applied Microbiology Biotechnology* 72: 876–882.
- Liu, Y.J., Zhang, A.N. and Wang, X.C. 2009. Biodegradation of phenol by using free and immobilized cells of *Acinetobacter* sp. XA05 and *Spingomonas* sp. FG03. *Biochemical Engineering Journal* 44:187-192.
- Loh, K.C. and Wang, S.J. 1998. Enhancement of biodegradation of phenol and a non-growth substrate 4-chlorophenol by medium augmentation with conventional carbon sources. *Biodegradation* 8: 329-338.
- Loh, K.C., Chung, T.S. and Ang, W.F. 2000. Immobilized-cell membrane bioreactor for high strength phenol wastewater. *Journal of Environmental Engineering* 126: 75-79.
- Ma, H., Li, G., Fang, P., Zhang, Y. and Xu, D. 2010. Identification of phenol-degrading *Nocardia* sp. Strain C-14-1 and characterization of its ring-cleavage 2,3-dioxygenase. *International Journal of Biology* 2(1): 79-83.

- Macleod, C.T. and Daugulis, A.J. 2005. Interfacial effects in a two-phase partitioning bioreactor: Degradation of polycyclic aromatic hydrocarbons (PAHs) by a hydrophobic *Mycobacterium*. *Process Biochem*istry 40: 1799-1805.
- Maegala, M., Fridelina, S. and Latif, I. 2011. Bioremediation of cynide by optimized resting cells of *Rhodococcus* strains isolated from Peninsular Malaysia. *International Journal of Bioscience, Biochemistry and Bioinformatics* 1(2): 98–101.
- Maegala, M., Fridelina, S. and Latif, I. 2012, Cynide degradation by immobilized cells of *Rhodococcus* UKMP-5M. *Biologia* 67(5): 837-844.
- Maghsoudi, S., Vossoughi, M., Kheirolomoom, A., Tanaka, E., Katoh, S. 2001. Biodesulfurization of hydrocarbons and diesel fuels by *Rhodococcus* sp. strain P32C1. *Journal of Biochemical Engineering* 8: 151-156.
- Mahto, R.B. and Bose, K.J. 2012. Production of alkaline protease from *Bacillus subtilis* by different entrapment techniques. *Journal of Biochemical Technology* 4(1): 498–501.
- Malhautier, L., Quijano, G., Avezac, M., Rocher, J., Fanlo, J.L. 2014. Kinetic characterization of toluene biodegradation by *Rhodococcus erythropolis*: Towards a rationale for microflora enhancement in bioreactors devoted to air treatment. *Chemical Engineering Journal* 247: 199-204.
- Mamimin, C., Thongdumyu, P., Hniman, A., Prasertsanoonsuk, P., Imai, T. and Thong, S.O. 2012. Simultaneous thermophilic hydrogen production and phenol removal from palm oil mill effluent by *Thermoanaerobacterium*-rich sludge. *International Journal of Hydrogen Energy* 37: 15598-15606.
- Manahan, S.E. 2000. Environmental Science, Technology and Chemistry. In: *Environmental Chemistry* (Ed.) Boca Raton: CRC Press LLC. pp. 623.
- Mandal, S., Bhunia, B., Kumar, A., Dasgupta, D., Mandal, T., Datta, S. and Bhattacharya, P. 2013. A statistical approach for optimization of media components for phenol degradation by *Alcaligenes faecalis* using Plackett–Burman and response surface methodology. *Desalination and Water Treatment* 51(31-33): 6058-6069
- Mandani, P., Desai, K. and Highland, H. 2013. Cytotoxic effects of benzene metabolites on human sperm function: An in vitro study. *ISRN Toxicology* 2013: 397524, 6 pages. DOI:10.1155/2013/397524
- Marella, M., Seo, B.B., Nakamaru-Ogiso, E., Greenamyre, J.T., Matsuno-Yagi, A. and Yagi, T. 2008. Protection by the NDI1 gene against neurodegeneration in a rotenone rat model of Parkinson's disease. *PLoS One*. 3: e1433.
- Margesin, R. and Schinner, F. 2001. Potential of halotolerant and halophilic microorganisms for biotechnology. *Extremophiles* 5: 73-83.

- Margesin, R., Bergauer, P. and Gander, S. 2004. Degradation of phenol toxicity of phenolic compounds: a comparison of cold-tolerant *Arthrobacter* spp. and mesophilic *Pseudomonas putida*. *Extremophiles* 8: 133-141.
- Margesin, R., Fonteyne, P.A. and Redl, B. 2005. Low-temperature biodegradation of high amounts of phenol by *Rhodococcus* spp. and *Basidiomycetous* yeasts. *Residual Microbiology* 156: 68-75.
- Marimaa, M., Heinaru, E., Liivak, M., Vedler, E. and Heinaru. A. 2006. Grouping of phenol hydroxylase and catechol 2,3-dioxygenase genes among phenol- and *p*-cresol-degrading *Pseudomonas* species and biotypes. *Archives of Microbiology* 186: 287–296.
- Martínková, L., Uhnáková, B., Pátek, M., Nesvera, J. and Kren, V. 2009. Biodegradation potential of the genus *Rhodococcus*. *Environment International* 35: 62–177.
- Martins, S.C.S, Martins C.M., Larissa Maria Cidrão Guedes Fiúza1 and Sandra Tédde Santaella. 2013. Immobilization of microbial cells: A promising tool for treatment of toxic pollutants in industrial wastewater. *African Journal of Biotechnology* 12(28): 4412-4418.
- Matsumura, E., Susumu, O., Shuichiro, M., Shinji, T. and Aoki, T. 2004. Constitutive synthesis, purification, and characterization of catechol 1, 2-dioxygenase from the aniline assimilating bacterium *Rhodococcus* sp. AN-22. *Journal of Biosciences Bioengineering* 98: 71–76.
- Maza-Márquez, P., Martinez-Toledo, M.V., Fenice, L., Andrade, M., Lasse rrot, A., Gonzalez-Lopez, J. 2014. Biodegradation of olive washing wastewater pollutants by highly efficient phenol-degrading strains selected from adapted bacterial community. *International Biodeterioration and Biodegradation* 82: 192-198.
- McLeod, M.P., Warren, R.L., Hsiao, W.W.L., Araki, N., Myhre, M., Fernandes, C., Miyazawa, D., Wong, W., Lillquist, A.L., Wang, D., Dosanjh, M., Hara, H., Petrescu, A., Morin, R.D., Yang, G., Stott, J.M., Schein, J.E., Shin, H., Smailus, D., Siddiqui, A.S., Marra, M.A., Jones, S.J.M., Holt, R., Brinkman, F.S.L., Miyauchi, K., Fukuda, M., Davies, J.E., Mohn, W.W. and Eltis, L.D. 2006. The complete genome of *Rhodococcus* sp. RHA1 provides insights into a catabolic powerhouse. In: *Proceeding of National Academy Science* USA 103: 15582–15587
- McMinn, E.J., Alderson, G., Dodson, H.I., Goodfellow, M. and Ward, A.C. 2000. Genomic and phenomic differentiation of *Rhodococcus equi* and related strains. *Antonie van Leeuwenhoek* 78: 331–340.
- Meena, G.S., Kumar, N., Majumdar, G.C., Banerjee, R., Meena, P.K. and Yadav, V. 2014. Growth characteristics modeling of *Lactobacillus acidophilus* using RSM and ANN. *Brazilian Archives of Biology and Technology* 57(1): 15-22.

- Meijer, W.G. and Prescott JF. 2004. *Rhodococcus equi. Veterinar Research*. 35: 383–396
- Mendonca, E., Martins, A. and Anselmo, A. M. 2004. Biodegradation of natural phenolic compounds as single and mixed subtrates by *Fusarium flocciferum*. *Electronic Journal of Biotechnology* 7(1): 30-37.
- Michalowicz, J. and Duda, W. 2007. Phenols-Sources and Toxicity. *Polish Journal of Environmental Studies* 16: 347-362.
- Minhalma, Miguel And De Pinho, Maria N. 2001. Tannic-membrane interactions on ultrafiltration of cork processing wastewaters. *Separation and Purification Technology* 22-23(1): 479-488.
- Miroux, B. and Walker, J.E. 1996. Over-production of proteins in *Escherichia coli*: mutant hosts that allow synthesis of some membrane proteins and globular proteins at high levels. *Journal of Molecular Biology* 260: 289–298.
- Mishra, V., and Lal, R. 2014. Enhanced degradation of phenol by a new species of *Rhodococcus: R. gordoniae* through ortho-pathway. *International Journal of Basic and Applied Biology* 1(1): 23–26.
- Misson, M. and Razali, F. 2007. Immobilizatin of phenol degrader *Pseudomonas sp.* in repeated batch culture using bioceramic and sponge as support materials. *Journal of Technology* 46: 51-59.
- Mitani, Y., Meng, X., Kamagata, Y., and Tamura, T. 2005. Characterization of LtsA from *Rhodococcus erythropolis*, an enzyme with glutamine amidotransferase activity. *Journal of Bacteriology* 187(8): 2582-2591.
- Mohammadi, M., Man, H.C., Hassan, M. A. and Yee, P.L. 2010. Treatment of wastewater from rubber industry in Malaysia. *African Journal of Biotechnology* 9(38): 6233–6243.
- Mohanty, S.S. 2012. Microbial Degradation of Phenol: A Comparative Study. National Institute of Technology, Rourkela Orissa, India.
- Mohite, B.V., Pawar, S.P., and Morankar, A. 2011. Isolation, Selection and Biodegradation Profile of Phenol Degrading Bacteria from Oil Contaminated Soil. *Bulletin Environmental Contamination Toxicology* 87: 143–146.
- Mokhtar, N.A. 2011. Investigations of triacyglyceride metabolism amongst actinomycete isolates from Peninsular Malaysia. Cambridge University, United Kingdom.
- Mona, K.G. 2007. Immobilization of *Rhodococcus* sp. DG for efficient degradation of phenol. *Fresenius Environmental Bulletin* 16: 1655-1661.

- Moore, E., Arnscheidt, A., and Mau, M. 2004. Simplified protocols for the preparation of genomic DNA from bacterial cultures, 3–18. In *Molecular Microbial Ecology Manual, Second Edition* 1.01: 3–18. Kluwer Academic Publishers.
- Moslemy, P., Guiot, S.R. and Neufeld, R.J. 2002. Production of size-controlled gellan gum microbeads encapsulating gasoline-degrading bacteria. *Enzyme Microbial Technology* 30: 10-18.
- Moslemy, P., Guiot, S.R. and Neufeld, R.J. 2004. Activated sludge encapsulation in gellan gummicrobeads for gasoline biodegradation. *Bioprocess Biosystemic Engineering* 26: 197–204.
- Moslemy, P., Neufeld, R.J., Millette, D. and Guiot, S.R. 2003. Transport of gellan gum microbeads through sand: an experimental evaluation for encapsulated cell bioaugmentation. *Journal of Environmental Management* 69: 249–259.
- Mot, R.D., Nagy, I. and Baumeister, W. 1998. A self-compartmentalizing protease in Rhodococcus: the 20S proteosome. Antonie van Leeuwenhoek 74: 83-87.
- Movahedyan, H., Khorsandi, H., Salehi, R. and Nikaeen, M. 2009. Detection of phenol degrading bacteria and *Pseudomonas putida* in activated sludge by polymerase chain reaction. *Iranian Journal of Environmental*, *Health*, *Science and Engineering* 6: 115-120.
- Mrozik, A. and Piotrowska-Segete, Z. 2009. Bioaugmentation as a strategy for cleaning up of soils contaminated with aromatic compounds. *Microbiological Research* 165(5): 363-375.
- Mrozik, A., Piotrowska-Segete and Labuzek, S. 2004. Cytoplasmic bacterial Membrane Response to Environmental Perturbations. *Polish Journal of Environmental Studies* 15(5): 487-494.
- Muller, R.H. and Babel, W. 1994. Phenol and its derivatives as heterotrophic substrates for microbial growth and energetic comparison. *Applied Microbiology and Biotechnology* 42: 446–451.
- Murialdo, S.E., Fernoglio, R., Haure, P.M. and Gonzalez, J.F. 2003. Degradation of phenol and chlorophenol by mixed and pure cultures. *Water Science Academy* 29(4): 457-464.
- Muthukumarasamy, P., Allan-Wojtas, P. and Holley, R.A. 2006. Stability of *Lactobacillus reuteri* in different types of microcapsules. *Journal of Food Science* 71(1): 20–24.
- Nadaf, N.H. and Ghosh, J.S. 2011. Purification and characterization of catechol 1, 2-dioxygenase from *Rhodococcus* sp. NCIM 2891. *Residual Journal of Environment Earth Science* 3: 608-613.

- Nagamani, A. and Lowry, M. 2009. Phenol biodegradation by *Rhodococcus* coprophilus isolated from semi arid soil samples of Pali, Rajasthan. *International Journal of Applied Environmental Sciences* 4: 294-302.
- Nagarajan, J., Norazah, M.N. and Ibrahim A.L. 2014. *Rhodococcus* UKMP-5M, an endogenous lipase producing actinomycete from Peninsular Malaysia. *Biologia* 69(2): 123-132.
- Nair, C.I., Jayachandran, K. and Shashidhar, S. 2008. Biodegradation of phenol. *African Journal of Biotechnology* 7(25): 4951-4958.
- Neujahr, H.Y. and Gaal, A. 1973. Phenol hydroxylase from yeast. Purification and properties of the enzyme from *Trichosporon cutaneum*. *European Journal of Biochemistry* 35: 386-400.
- Neujahr, H.Y., Lindsjö, S. and Varga, J.M. 1974. Oxidation of phenol by cells and cell-free enzymes from *Candida tropicalis*. *Antonie van Leeuwenhoek* 40: 209-216.
- Newman, J., Peat, T.S., Richard, R. Kan, L., Swamson, P.E., Affholter, J.A., Holmes, L.H., Schindler, J.F., Unkefer, C.J. and Terwilliger, T.C. 1999. Haloalkane dehalogenase: structure of *Rhodococcus* enzyme. *Biochemistry* 38: 16105-16114.
- Nigam, P.S. 2013. Microbial enzymes with special characteristics for biotechnological applications. *Biomolecules* 3: 597-611
- Nishiuchi, Y., Tuneko, B. and Yano, I. 2000. Mycolic acids from *Rhodococcus*, Gordonia and Dietzia. Journal of Microbiology Method 40: 1-9.
- Norena-Barroso, E., Sima-Lyarez, R., Gold-Bouchot, G. and Zapata-Perez, O. 2004. Persistent organic pollutants and histological lesions in Mayan catfish *Ariopsis assimilis* from the Bay of Chetumal, Mexico. *Marine Pollution Bulletin* 48: 263-269.
- Norton, S. and Lacroix, C. 1990. Gellan gum gel as entrapped matrix for high temperature fermentation process-rheological study. *Biotechnology Techniques* 4: 351-356.
- Nunes, C., Sousa, C., Ferreira, H., Lucio, M., Lima, J.L.F.C., Travares, J., Cordeiro-da-Silva, A. and Reis, S. 2008. Substituted phenols as pollutants that affect membrane fluidity. *Journal of Environmental Biology* 29: 733-738.
- Oboirien, B.O., Amigun, B., Ojumu, T.V., Ogunkunle, O.A., Adetunji, O.A., Betiku, E. and Solomon, B.O. 2005. Substrate inhibition kinetics of phenol degradation by *Pseudomonas aeruginosa* and *Pseudomonas fluorescence*. *Biotechnology* 4(1): 56-61.
- Ogbonnaya, U. and Kirk, T. Semple. 2013. Impact of biochar on organic contaminants in soil: A tool for mitigating risk. *Agronomy* (3): 349-375.

- OhioEPA, State of Ohio Environmental Protection Agency. 2002. Pollution prevention opportunities for PBT chemicals: Phenol. Columbus, Ohio. www.epa.state.oh.us/opp. Accessed on 20 September 2013.
- Ojumu, T.V., Bello, O.O., Sonibare, J.A. and Solomon, B.O. 2005. Evaluation of microbial systems for bioremediation of petroleum refinery effluents in Nigeria. *African Journal of Biotechnology* 4(1): 31-35.
- Okungbowa, F.I., Ghosh, A.K., Chowdhury, R., Chaudhuri, P., Basu, A. and Pal, K. 2007. Mechanical lysis of *Candida* cells for crude protein and enzymatic activity estimation:vcomparison of three methods. *World Journal of Medical Sciences* 2(2): 101-104.
- Oltmanns, R.H., Muller, R., Otto, M.K. and Lingens, F. 1989. Evidence for a new pathway in the bacterial degradation of 4-fluorobenzoate. *Applied Environment Microbiology* 55: 2499-2504.
- Omokoko, B., Jantges, U.K., Zimmermann, M., Reiss, M. and Hartmeier, W. 2008. Isolation of the *phe*-operon from *G. stearothermophilus* comprising the phenol degradative meta-pathway genes and a novel transcriptional regulator. *BioMed Central Microbiology* 8: 197.
- Paca, J Jr, Kremlackova, V., Turek, M., Sucha, V., Vilimkova, L., Paca, J., Halecky, M. and Stiborova, M. 2007. Isolation and partial characterization of cytoplasmic NADPH dependent phenol hydroxylase oxidizing phenol to cathecol in *Candida tropicalis* yeast. *Enzyme Microbial Technology* 40: 919-926.
- Padmaja, P. and Soni, H. 2014. Palm shell based activated carbon for removal of bisphenol. A: An equilibrium, kinetic and thermodynamic study. *Journal of Porous Materials* 21(3): 275-284.
- Pai, L.S., Hsu, L.Y., Chong, M.N., Sheu, S.C. and Chen, H.C. 1995. Continuous degradation of phenol by *Rhodococcus* sp. immobilized on granular activated carbon and in calcium alginate. *Bioresource Technology* 51: 37-42
- Paisio, C.E., Talano, M.A., Gonzalez, P.S., Busto, V.D., Talou, J.R. and Agostini, E. 2012. Isolation and characterization of a *Rhodococcus* strain with phenol-degrading ability and its potential use for tannery effluent biotreatment. *Science and Pollution Research*. 19: 3430-3439.
- Paisio, C.E., Talano, M.A., Gonzalez, P.s., Pajuelo-Dominuez, E. and Agostini. 2013. Characterization of a phenol-degrading bacterium isolated from an industrial effluent and its potential application for bioremediation. *Environmental Technology* 34(1-4): 485-493.
- Paller, G., Hommel, R.K. and Kleber, H.P. 1995. Phenol degradation by *Acinetobacter calcoaceticus* NCIB 8250. *Journal Basic Microbiology* 35(5): 325-335.

- Pankhurst, E.S. 1965. A spot test for catechol 2,3-oxygenase in bacteria. *Journal Applied Bacteriology* 28: 309-315.
- Parales R.E., Bruce N.C., Schmid A. and Wackett L.P. 2002. Biodegradation, biotransformation, and biocatalysis (B3). *Applied Environmental Microbiology* 68: 4699–4709.
- Passos, C.T., Michelon, M., Burkert, J.F.M., Kalil, S.J. and Burkert, C.A.V. 2010.Biodegradation of phenol by free and encapsulated cells of a new *Aspergillus* sp. isolated from a contaminated site in southern Brazil. *African Journal of Biotechnology* 940: 6716-6720.
- Patel, R. and Rajkumar, S. 2009. Isolation and characterization of phenol degrading yeast. *Journal of Basic Microbiology* 49(2): 216-219.
- Pathak, A., Green, S. J., Ogram, A., and Chauhan, A. 2013. Draft genome sequence of *Rhodococcus opacus* strain M213 shows a diverse catabolic potential. *Genome Announcements* 1: e00144–e00212. DOI:10.1128/genomeA.00144-12
- Pazarlioglu, N.K. and Telefoncu, A. 2005. Biodegradation of phenol by *pseudomonas* putida immobilized on activated pumice particle. *Process Biochemistry* 40: 1807–1814.
- Peng Y., Yang G.U., Wang Q.M., Du Y.Y, Li J.R. 2013. Isolation and mutagenesis of a novel phenol degrading strain. *Advanced Materials Research* 647: 588-594.
- Peppas, N.A., Hilt, J.Z., Khademhosseini, A. and Langer, R. 2006. Hydrogels in biology and medicine: From molecular principles to bionanotechnology. *Advanced Materials* 18(11): 1345–1360.
- Perry M.B., MacLean L.L., Patrauchan M.A., Vinogradov E. 2007. The structure of the exocellular polysaccharide produced by *Rhodococcus* sp. RHA 1. *Carbohydrate Research* 342: 2223–2229.
- Philp, J.C., Atlas, R.M. and Cunningham, C.J. 2009. *Bioremediation*, Encyclopedia of Life Sciences, John Wiley and Sons, Ltd. Online posting date: 15th March 2009
- Piakong, M.T. 2006. The Performance of Phenol Biodegradation by RETL-Cr1 Using Batch and Fed-batch Fermentation Techniques. Universiti Teknologi Malaysia.
- Poh P. E. and Chong M. F. 2010. Thermophilic palm oil mill effluent (POME) treatment using a mixed culture cultivated from POME. *Chemical Engineering Transactions* 21: 811-816.
- Porebski, S., Bailey, L.G. and Baum, B.R. 1997. Modification of a CTAB DNA extraction protocol for plants containing high polysaccharide and polyphenol components. *Plant Molecular Biology* 15: 8-15.

- Pourgholam, R., Laluei, F., Saedi, A.A., Zahedi, A., Safari, R., Taghavi, M.J., Saravi, N.H. and Pourgholam, H. 2011. Distribution and molecular identification of some causative agents of *Streptococcosis* isolated from farmed rainbow trout (oncorhynchus mykiss, walbaum) in Iran. *Iranian Journal of Fisheries Science* 10: 109-122.
- Powlowski, J., Shingler, V. 1994. Genetics and biochemistry of phenol degradation by *Pseudomonas* sp. CF600. *Biodegradation* 5: 219-236.
- Prabu, C.S and Thatheyusb, A.J. 2007. Biodegradation of acrylamide employing free and immobilized cells of *Pseudomonas aeruginosa*. *International Biodeterioration and Biodegradation* 60(2): 69-73. N. V. Pradeep · S. Anupama · K. Navya · H. N. Shalini · M. Idris · U. S. Hampannava. Biological removal of phenol from wastewaters: a mini review
- Prasad, M.P. and Manjunath, K. 2011. Comparative study on biodegradation of lipid rich wastewater using lipase producing bacteria species. *Indian Journal of Biotechnology* 10: 121-124.
- Prasad, M.P., Raj, J. and Bhalla, T.C. 2009. Purification of hyperactive nitrile hydratase from resting cells of *Rhodococcus rhodochrous* PA-34. *Indian Journal of Microbiology* 49: 237-242.
- Prieto, B., Hidalgo, M.A., Rodriguez, F.C., Serra, J.L. and Llama, M.J. 2002. Biodegradation of phenol in synthetic and industrial wastewater by *Rhodococcus eryhropilis*UPV-1 immobilized in air stirred reactor with clarifier. *Applied Microbiology and Biotechnology* 58: 583–589.
- Qiu, Y. and Park, K. 2001. Environment-sensitive hydrogels for drug delivery. *Advanced Drug Delivery Review* 53(3): 321–333.
- Quek, E., Ting, Y.P. and Tan, H.M. 2006. *Rhodococcus* sp. F92 immobilized on polyurethane foam shows ability to degrade various petroleum products. *Bioresource Technology* 97: 32-38.
- Quezada, M.A., Carbaleira, J.D., García-Burgos, C.A. and Sinisterra, J.V. 2008. *Monascus kaoliang* CBS 302.78 immobilized in tailor-made agars as catalyst for reduction of ketones: On the quest for a green biocatalyst. *Process Biochemistry* 43: 1220–1226.
- Radjendirane, V., Bhat, M.A and Vaidyanathan, C.S. 1991. Affinity purification and characterization of 2,4-dichlorophneol hydroxylase from *Pseudomonas cepacia*. *Archives of Biochemistry and Biophysic* 288(1): 169-176.
- Rafigh, S.M., Yazdi, A.V., Vossoughi, M., Safekordi, A.A., and Ardjmand, M. 2014. Optimization of culture medium and modeling of curdlan production from *Paenibacillus polymyxa* by RSM and ANN. *International Journal of Biological Macromolecules* 70: 463-473.

- Rajasekharan, S., Rajasekharan, R. and Vaidyanathan, C.S. 1990. Substrate-mediated purification and characterization of a 3-hydroxybenzoic acid-6-hudroxylase from *Micrococcus*. *Archives of Biochemistry and Biophysic* 278(1): 21-25.
- Rakamthong, C. and Prasertsan, P. 2011. Decolorization and phenol removal of anaerobic palm oil mill effluent by *Phanerochaetechrysosporim* ATCC 24725. TIChE International conference.
- Ramanan, R.N., Ling, T.C. and Ariff A.B. 2008. The performance of a glass bead shaking technique for the distruption of *Escherichia coli* cells. *Biotechnology and Bioprocess Engineering* 13: 613-623.
- Ramanan, R.N., Tan, J.S., Mohd, S.M., Ling T.C., Tey, B.T. and Arbakariya, A. 2010. Optimization of osmotic shock process variables for enhancement of the release of periplasmic interferon-a2b from *Escherichia* coli using response surface method. *Process Biochemistry* 45: 196–202.
- Rani, M.J., Hemambika, B., Hemapriya, J. and Kannan, V.R. 2010. Comparative assessment of heavy metal removal by immobilized and dead bacterial cells: A biosorption approach. *African Journal of Environmental Science and Technology* 4(2): 77-83.
- Rao, C.S., Madhavendra, S.S., Rao, R.S., Hobbs, P.J. and Prakasham, R.S. 2008. Studies on improving the immobilized bead reusability and alkaline protease production by isolated immobilized Bacillus circulans (MTCC 6811) using overall evaluation criteria. *Applied Biochemistry and Biotechnology* 150: 65–83.
- Rathore, S., Desai, P.M., Liew, C.V., Chan, L.W. and Hen, P.W.S. 2013. Microencapsulation of microbial. *Journal of Food Engineering* 16: 369-381.
- Ravishankar, S., Ambady, A., Ramu, H., Mudugal, N.V., Tunduguru, R., Anbarasu, A., Sharma, U.K., Samdaburthy, V.K. and Ramaiah, S. 2015. An IPTG inducible conditional expression system for *Mycobacteria*. *PLoS ONE* 10(8): e0134562.
- Ray, S., Lalman, J.A., and Biswas, N. 2009. Using the Box-Benkhen technique to statistically model phenol photocatalytic degradation by titanium dioxide nanoparticles. *Chemical Engineering Journal* 150(1): 15–24.
- Razika, B., Abbes, B., Messaoud, C. and Soufi, K. 2010. Phenol and benzoic acid degradation by *Pseudomonas aeruginosa*. *Journal of Water Resource and Protection* 2: 788-791.
- Reardon, K., Mosteller, D., Rogers, J. and DuTeau, N. 2002. Biodegradation kinetics of aromatic hydrocarbon mixtures by pure and mixed bacterial cultures. *Environmental Health Perspectives* 110: 1005–1011.

- Reardon, K.F., Mosteller, D.C. and Rogers, J.D. 2000. Biodegradation kinetics of benzene, toluene and phenol and mixed substrates for *Pseudomonas putida* F1. *Biotechnology Bioengineering* 69 (4): 385-400.
- Reddy, L., Wee, Y., Yun, J. and Ryu, H. 2008. Optimization of alkaline protease production by batch culture of *Bacillus* sp. RKY3 through Plackett-Burmann and response surface methodolical approaches. *Bioresource Technology* 9: 2242–2249.
- Rehfuss, M. and Urban, J. 2005. *Rhodococcus phenolicus* sp. Nov., A novel bioprocessor isolated *Actinomycetes* with the ability to degrade chlorobenzene, dichlorobenzene, and phenol as sole carbon sources. *System Applied Microbiology* 28(8): 695–701.
- Ren, H.S., Wang, Y., Zhao, H.B., and Cai, B.L. 2008. Isolation and identification of phenol-degrading strains and the application in biotreatment of phenol-containing wastewater. *Huan Jing Ke Xue* 29(2): 482-487.
- Rocha, L.L., Cordeiro, R.D.A., Cavalcante, R.M., Nascimento, R.F.D., Martins, S.C.S., Santaella, S.T. and Melo, V.M.M. 2007. Isolation and characterization of phenol-degrading yeasts from an oil refinery wastewater in Brazil. *Mycopathologia* 164: 183-188.
- Rodriguez, M.J, Lebrero, J.L.A and Alvarez, E. 2009. Biotransformation of phenol to catechol by recombinant phenol hydroxylase. *Biocatalysis and Biotransformation* 17(1): 45-60.
- Rubalcaba, A., Sua', M.E., Rez-Ojeda, F., Stu", Ber, A., Fortuny, C., Bengoa, I., Metcalfe, J., Font, J., Carrera and Fabregat, A. 2012. Phenol wastewater remediation: advanced oxidation processes coupled to a biological treatment. *Water Science and Technology* 55(12): 221–227.
- Saa, L., Jaureguibeitia, A., Largo, E., Llama, M.J. and Serra. J. 2010. Cloning, purification and characterization of two components of phenol hydroxylase from *Rhodococcus erythropolis* UPV-1. *Applied Microbiology Biotechnology* 86: 201-211.
- Sabullah, M.K., Rahman, M.F., Ahmad, S.A., Sulaiman, M.R., Shukor, M.S., Shamaan, N.A. and Shukor, M.Y. 2016. Isolation and characterization of a molybdenum-reducing and glyphosate-degrading *Klebsiella oxytoca* strain Saw-5 in soils from Sarawak. *AGRIVITA* 38(1): 1-13.
- Said, M., Ahmad, A. and Wahab, A.M. 2013. Removal of phenol during ultrafiltration of Palm oil mill effluent (POME): effect of pH, ionic strength, pressure and temperature. *Der Pharma Chemica* 5(3): 190–196.
- Sally N.J. 2012. Extraction of phenol from industrial water using different solvents. *Research Journal of Chemical Sciences* 2(4): 1-12.

- Sambrook, J., Fritsch, F. and Maniatis, T. 1989. *Molecular cloning: A laboratory manual (2nd Edition)*. Cold Spring Harbor Laboratory Press. New York, pp 18.47-18.59.
- Sandhu, A., Halverson, L.J and Beattie, G.A. 2009. Identification and genetic characterization of phenol-degrading bacteria from leaf microbial communities. *Microbial Ecology* 57: 276–285.
- Santos, V.L. and Linardi, V.R. 2004. Biodegradation of phenol by a filamentous fungi isolated from industrial effluents-identification and degradation potential. *Proceeding Biochemistry*, 39: 1001-1006.
- Santos, V.L., Heilbuth, N.M., Braga, D.T., Monteiro, A.S. and Linardi, V.R. 2003. Phenol degradation by a *Graphium* sp. FIB4 isolated from industrial effluents. *Journal of Basic Microbiology* 43: 238-248.
- Santos, V.L., Monteiro, A.S., Braga, D.T. and Santoro, M.M. 2009. Phenol degradation by *Aureobasidium pullulans* FE13 isolated from industrial effluents. *Journal of Hazardous Materials* 161(2-3): 1413-1420.
- Saravanan, P., Pakshirajan, K. and Saha, P. 2008. Kinetics of phenol and m-cresol biodegradation by an indigenous mixed microbial culture isolated from a sewage treatment plant. *Journal of Environmental Sciences* 20(12): 1508–1513.
- Sarin, C. and Sarin, S. 2010. Removal of cadmium and zinc from soil using immobilized cell of biosurfactant producing bacteria. *Environment Asia* 3(2): 49-53.
- Schleinitz, H., Schmeling, S., Jehmlich, N., Von, M., Harms, B.H., Kleinsteuber, S., Vogt, C. and Fuchs, G. 2009. Phenol degradation in the strictly anaerobic iron-reducing bacterim geobacter metallreducens GS-15. Applied Environmental and Microbiology 75(12): 3912-3919.
- Schoof, Andrew. 2015. Enzymatic treatment of phenolic industrial wastewater with nitrogen management. University of Windsor.
- Schroder, M., Muller, C., Posten, C., Deckwer, W.D. and Hecht, V. 1997. Inhibition kinetics of phenol degradation from unstable steady state data. *Biotechnology and Bioengineering* 54: 567-576.
- Scott, R.I., Wills, S.J. and Bucke, C. 1989. Oxygen uptake by k-carrageenan entrapped *Streptomyces clavuligerus*. *Enzymes Microbiology Technology* 10: 258-263.
- Scragg, A.H. 2006. The effect of phenol on the growth of *Chlorella vulgaris* and *Chlorella* VT-1. *Enzyme Microbial Technology* 39: 796–799.

- Selvam, K., Vishnupriya, B. and Subhash, C.B. 2011. Screening and quantification of marine actinomycetes producing industrial enzymes amylase, cellulase and lipase from South Coast of India. *International Journal of Pharmaceutical and Biological Archieves* 2: 1481-1487.
- Shah, M.P. 2014. Microbiological removal of phenol by an application of *Pseudomonas* spp. ETL: an innovative biotechnological approach providing answers to the problems of FETP. *Journal of Applied and Environmental Microbiology* 2(1): 6-11.
- Shao, Z., Dick, W.A. and Behki, R.M. 1995. An improved *Escherichia coli-Rhodococcus* shuttle vector and plasmid transformation in *Rhodococcus* spp. using electroporation. *Letters in Applied Microbiology* 21: 261–266.
- Sharma, A., Thakur, I.S. and Dureja, P. 2009. Enrichment, isolation and characterization of pentachlorophenol degrading bacterium *Acinetobacter* sp. ISTPCP-3 from effluent discharge site. *Biodegradation* 20 (50): 643-650.
- Sharma, M. 2014. Actinomycetes: Source, identification, and their applications. 2014. *International Journal of Current Microbiology and Applied Science* 3(2): 801-832.
- Sharma, N. and Vikas, C.G. 2012. Batch biodegradation of phenol of paper and pulp effluent by *Aspergillus Niger*. *International Journal of Chemical Engineering and Applications* 3(3): 182-186.
- Sharma, S.L. and Pant, A. 2001. Biodegradation and conversion of alkanes and crude oil by a marine *Rhodococcus* sp. *Biodegradation* 11: 289-294.
- Shen, J., He, R., Wang, L., Zhang, J., Zuo, Y. Li, Y., Sun, X., Li, J., Han, W. 2009. Biodegradation kinetics of picric acid by *Rhodococcus* sp. NJUST16 in batch reactors. *Journal of Hazardous Materials* 164: 1198-1204.
- Shourian, M., Noghabi, K.A., Zahiri, H.S., Bagheri, T., Karballaei, G., Mollaei, M., Rad, I., Ahadi, S., Raheb, J. and Abbasi, H. 2009. Efficient phenol degradation by a newly characterized *Pseudomonas* sp. SA01 isolated from pharmaceutical wastewaters. *Desalination* 246: 577–594.
- Shuler, M.L. and Kargi, F. 2002. Bioprocess engineering: basic consepts. 2nd Edition. Prentice Hall, NJ. U.S.A. pp 133.
- Shumkova, E.S., Solyanikova, I.P., Plotnikova, E.G. and Golovleva, L.A. 2009. Phenol degradation by *Rhodococcus opacus* strain 1G. *Applied Biochemistry and Microbiology* 45(1): 43–49.
- Shweta and Dhandayuthapani, K. 2013. Influence of media supplements on phenol biodegradation by *Pseudomonas aeruginosa* SPD10. *International Journal of Current Microbiology and Applied Sciences* 2(6): 64-69.

- Silva, A.A.L, Pereira, M.P., Filho R.G.S, Hofer, E. 2007. Utilization of phenol in the presence of heavy metals by metal-tolerant non-fermentative gram-negative bacteria isolated from wastewater. *Revista Latinoamerca de Microbiologia* 49(3-4): 68-73.
- Silva, A.S., Jacques, R.J.S., Andreazza1, R., Bento, F.M., Roesch, L.F.W. and Camargo, F.A.O. 2013. Properties of catechol 1,2-dioxygenase in the cell free extract and immobilized extract of *Mycobacterium fortuitum*. *Brazilian Journal of Microbiology* 44: 291-297.
- Silva, C.C., Hayden, H., Sawbridge, T., Mele, P. and De Paula, S.O. 2013. Identification of genes and pathways related to phenol degradation in metagenomic libraries from petroleum refinery wastewater. *PLoS ONE* 8: e61811.
- Simpson, R.J. 2010. Homogenization of mammalian tissue. Cold Spring Harbor Protocol. 7: 1-4. DOI: 10.1101/pdb.prot5455.
- Sivasubramanian, S. and Namasivayam, S.K.R. 2013. Optimization of parameters for phenol degradation using immobilized *Candida tropicalis* SSK01 in batch reactor. *Journal of Environmental Biology* 35: 531-536.
- Sivasubramanian, S. and Namasivayam, S.K.R. 2014. Statistical optimization of physical conditions for phenol degradation using effective microorganism. *Indian Journal of Chemical Technology* 21: 14–20.
- Smit, E., Lee, H., Trevors, J.T. and van Elsas, J.D. 1996. Interaction between a genetically engineered *Pseudomonas fluorescens* and bacteriophage R2f in soil: effect of nutrients, alginate encapsulation and the wheat *Rhizospere*. *Microbial Ecology* 31: 125–140.
- Sokolovska, I., Rozenberg, R., Riez, C., Rouxhet, P.G., Aqathos, S.N. and Wattiau, P. 2003. Carbon source-induced modifications in the mycolic acid content and cell wall permeability of *Rhodococcus erythropolis* E1. *Applied and Environmental Microbiology* 69: 7019-7027.
- Solomon, B.O., Posten, C., Harder, M.P.F., Hecht, V. and Deckwer, W.D. 1994. Energetics of *Pseudomonas cepacia* growth in a chemostat with phenol limitation. *Journal of Chemical Technology and Biotechnology* 60: 275-282.
- Soudi, M.R. and Kolahchi, R.N. 2011. Bioremediation potential of a phenol degrading bacterium, *Rhodococcus erythropolis* SKO-1. *Progress in Biological Sciences* 1: 31-40.
- Spånning, Å. and Neujahr, H. Y. 1987. Growth and enzyme synthesis during continuous culture of *Trichosporon cutaneum* on phenol. *Biotechnology and Bioengineering* 29: 464-468.

- Sridevi, V., Chandana Lakshmi M.V.V., Manasa, M. and Sravani, M. 2012. Metabolic pathways for the biodegradation of phenol. *International Journal of Engineering Science and Advance Technology* 3: 695-705.
- Stackebrandt, E., Rainey, F.A., and Ward-Rainey, N.L. 1997. Proposal for a new hierarchic classification system, *Actinobacteria* classis nov. *International Journal System Bacteriology* 47: 479-491.
- Statostina, N.G., Lusta, K.A. and Fikhte, B.A. 1982. Influence of various factors in polyacrylamide gel immobilization on the viability of *Escherichia coli* B cells. *Prikladnaia Biohimmiia Mikrobiologiia* 18(2): 225-230.
- Steiert, J.G., Pignatello, J.J. and Crawford, R.L. 1987. Degradation of chlorinated phenols by a pentachlorophenol-degrading bacterium. *Applied Environmental Microbiology* 53(5): 907-10.
- Straube, G., Hensel, J., Niedan, C., Straube, E. 1990. Kinetic studies of phenol degradation by *Rhodococcus* sp. P1I. Batch cultivation. *Anton. van Leeuwenhook* 57: 29-32.
- Strnad, H., Patek, M., Fousek, J., Szokol, J., Ulbrich, P., Nesvera, J., Paces, V., Vlcek, C. 2014. Genome sequence of Rhodococcus erythropolis strain CCM2595, a phenol derivative-degrading bacterium. *Genome Announcement* 2(2):e00208-14.
- Sueoka, K., Satoh, H., Onuki, M., Mino, T. 2009. Microorganisms involved in anaerobic phenol degradation in the treatment of synthetic coke-oven wastewater detected by RNA stable-isotope probing. *FEMS Microbiology Letter* 291: 169-174.
- Suhaila, N.Y., Rosfarizan, M., Ahmad, S.A., Latif, A.I. and Ariff, A. 2013. Nutrients and culture conditions requirements for the degradation of phenol by *Rhodococcus* UKMP-5M. *Journal of Environmental Biology* 34: 635-643.
- Suhaila, N.Y., Ramanan, R.N., Rosfaziran, M., Abdul Latif, I. and Ariff, A. 2012. Optimization of parameters for improvement of phenol degradation by *Rhodococcus* UKMP-5M using response surface methodology. *Annals of Microbiology* 63(2): 513-521.
- Sun, J., Liu, Y. and Li, Z. 2011. Optimization of entrapping conditions of nitrifying bacteria and selection of entrapping agent. In *Proceedings of the 2nd International Conference on Environmental Science and Technology (IPCBEE)*, vol. 6, IACSIT Press Singapore.
- Sun, X., Ma, X., Xu, C., Bai, J. and Zhang, W. 2012. Isolation, screening and identification of phenol-degrading bacteria from coking wastewater. *Applied Mechanics and Materials* 209-211: 2027-2031.

- Sutcliffe, I.C. 2010. Characterisation of lipomannan from the mycolic acid containing *Actinomycete Dietzia maris*. *Antonie van Leewenhoek* 78: 195-201.
- Sutcliffe, I.C, Brown, A.K. and Dover, L.G. 2010. The Rhodococcal Cell Envelope: Composition, organisation and biosynthesis. H.M. Alvarez (Ed.), *Biology of Rhodococcus*, Microbiology Monographs 16, DOI 10.1007/978-3-642-12937-7\_2. Springer-Verlag Berlin Heidelberg. pp 49-71
- Szőköl, J., Rucká, L., Šimčíková, M., Halada, P., Nešvera, J. and Pátek, M. 2014. Induction and carbon catabolite repression of phenol degradation genes in *Rhodococcus erythropolis* and *Rhodococcus jostii*. *Applied Microbiology and Biotechnology* 98(19): 8267-79.
- Tabib, A., Haddadi, A., Shavandi, M., Soleimani, M. and Azizinikoo, P. 2012. Biodegradation of phenol by newly isolated phenol-degrading bacterium *Ralstonia* sp. strain PH-S1. *Journal of Petroleum Science and Technology* 2: 58-62.
- Tai, J., Adav, S.S., Sub, A. and Lee, D.J. 2010. Biological hydrogen production from phenol-containing wastewater using *Clostridium butyricum*. *International Journal of Hydrogen Energy* 35: 13345-13349.
- Takeo, M., Yasukawa, T., Abe, Y., Niihara, S., Maeda, Y., Negoro, S. 2003. Cloning and characterization of a 4-nitrophenol hydroxylase gene cluster from *Rhodococcus* sp. PN1. *Journal of Bioscience Bioengineering* 95: 139-145.
- Tan, W.S. and Adeline, S.Y.T. 2012. Efficacy and reusability of alginate-immobilized live and heat-inactivated *Trichoderma asperellum* cells for Cu (II) removal from aqueous solution. *Bioresource Technology* 123: 290-295.
- Tarighian, A., Hill, G.A and Lin, Y.H. 2001. Lag phase model for transient growth of pseudomonas putida on phenol. The Canadian Journal of Chemical Engineering 79: 732-736.
- Taskova, R.M., Zorn, H., Krings, U., Bouws, H. and Berger, R.G. 2006. A comparison of cell wall disruption techniques of intracellular metabolites from *Pleurotus* and *Lepista* sp. Z. Naturforsch. *Canadian Journal of Bioscience* 61: 347-350.
- Tauber, M.M., Paula, A.C., Robra, K.H. and Gubitz, G.M. 2000. Nitrile hydratase and amidase from *Rhodococcus rhodochrous* hydrolyze acrylic fibers and granular polyacrylnitriles. *Applied Environmental Microbiology* 66: 1634-1638.
- Tepe, O. and Dursun, A.Y. 2008. Combined effects of external mass transfer and biodegradation rates on removal of phenol by immobilized *Ralstonia eutropha* in a packed bed reactor. *Journal of Hazardous Materials* 151: 9-16.

- Tomás-Gallardo, L., Gómez-Álvarez, H., Santero, E., and Floriano, B. 2014. Combination of degradation pathways for naphthalene utilization in *Rhodococcus* sp. strain TFB. *Microbial Biotechnology* 7: 100-114.
- Tomás-Gallardo, L., Santero, E., Camafeita, E., Calvo, E., Schlömann, M. and Floriano, B. 2009. Molecular and biochemical characterization of the tetralin degradation pathway in *Rhodococcus* sp. strain TFB. *Microbial Biotechnology*. 2(2): 262-273.
- Tomás-Gallardo, L., Canosa, I., Santero, E., Camafeita, E., Calvo, E., López, J.A. and Floriano, B. 2006. Proteomic and transcriptional characterization of aromatic degradation pathways in *Rhodoccocus* sp. strain TFB. *Proteomics* 6(1): 119-132.
- Trelles, J.A., Ferna ndez, M., Lewkowicz, E.S., Iribarrena, A.M. and Sinisterra, J.V. 2003. Purine nucleoside synthesis from uridine using immobilised *Enterobacter gergoviae* CECT 875 whole cells. *Tetrahedron Letters* 44: 2605-2609.
- TRI96. 1998. Toxic Chemical Release Inventory. National Library of Medicine, National Toxicology Information Program, Bethesda, MD.
- Troyer, D.L., Reed, A., Obertz, D. and Chengapa, D.D. 1990. A rapid and simplified protocol for DNA isolation from bacteria. *Veterinary Research Communications* 14: 447-451.
- Tsai, S.C. and Li, Y.K. 2007. Purification and characterization of a catechol 1,2-dioxygenase from a phenol degrading *Candida albicans* TL3. *Archives of Microbiology* 187: 199-206.
- Tuah, P.M., Aini, N., Rashid, A., and Salleh, M. 2009. Degradation pathway of phenol through ortho-cleavage by *Candida tropicalis* RETL-Cr1. *Borneo science* 24: 1-8.
- U.S. ENVIRONMENTAL PROTECTION AGENCY (USEPA). 2010. Technology transfer air toxic website. *Phenol*. Available from: http://www.epa.gov/ttnatw01/hlthef/cresols.html.
- Uludag, H., De Vos, P. and Tresco, P.A. 2000. Technology of mammalian cell encapsulation. *Advanced Drug Delivery Reviews* 42(1–2): 29–64.
- Urai, M., Anzai, H., Ogihara, J., Iwabuchi, N., and Harayama, S. 2006. Structural analysis of an extracellular polysaccharide produced by *Rhodococcus rhodochrous* strain S-2 341: 766-775.
- Uz, I., Duan, Y.P. and Ogram, A. 2000. Characterization of the naphthalene-degrading bacterium, *Rhodococcus opacus* M213. *FEMS Microbiology Letters* 185: 231-238.

- Vaidehi, K. and Kulkarni, S. D. 2012. Microbial remediation of polycyclic aromatic hydrocarbons: An overview. *Research Journal of Chemistry and Environment* 16(4): 200-212.
- Van der Geize, R. and Dijkhuizen, L. 2004. Harnessing the catabolic diversity of rhodococci for environmental and biotechnological applications. *Current Opinion in Microbiology* 7: 255-261.
- Van Schie, P.M., and Yong, L.Y. 1998. Isolation and characterization of phenol-degrading denitrifying bacteria. *Applied Environmental Microbiology* 64: 2432-2438.
- Van Schie, P.M., Young, L.Y. 2000. Biodegradation of phenol: mechanisms and applications. *Bioremediation Journal* 4: 1-18.
- Varma, R.J. and Gaikwad, B.G. 2008. Rapid and high biodegradation of phenols catalyzed by *Candida tropicalis* NCIM 3556 cells. *Enzyme and Microbiology Technology* 43(6): 431-435.
- Veenagayathri, K. and Vasudevan, N. 2010. Effect of pH, nitrogen sources and salts on the degradation of phenol by the bacterial consortium under saline conditions. *International Journal of Biotechnology and Biochemistry* 6: 783-791.
- Veenagayathri, K., and Vasudevan, N. 2011. Ortho and meta cleavage dioxygenases detected during the degradation of phenolic compounds by a moderately halophilic bacterial consortium. *International Research Journal of Microbiology* 2(10): 406-414.
- Verschueren, K. 1983. Handbook of environmental data on organic chemicals. 2nd ed. Van Nostrand Reinhold Company, New York.
- Veselý, M., Pátek, M., Nesvera, J., Cejková, A., Masák, J. and Jirků, V. 2003. Hostvector system for phenol-degrading *Rhodococcus erythropolis* based on *Corynebacterium* plasmids. *Applied Microbiology and Biotechnology* 61: 523–527.
- Veselý, M., Knoppová, M. and Nešvera J. and Pátek M. 2007. Analysis of catRABC operon for catechol degradation from phenol-degrading *Rhodococcus erythropolis*. Applied Microbiology and Biotechnology 76: 159-168.
- Vidali, M. 2001. Bioremediation: An overview. *Pure and Applied Chemistry* 73: 1163-1172.
- Walter, M., Guthrie, J.M., Sivakumaran, S., Parker, E. and Slade, A., McNaughton, D. and Boyd-Wilson, K.S.H. 2003. Screening of New Zealand native white-rot isolates for PCP Degradation. *Bioremediation Journal* 7: 119-128.

- Wang, G., Wen, J., Li, K. Qiu, C. and Li, H. 2009. Biodegradation of phenol and m-cresol by *Candida albicans* PDY-07 under anaerobic condition. *Journal of Industrial Microbiology and Biotechnology* 36: 809-814.
- Wang, X., Xu, P., Yuan, Y., Liu, C., Zhang, D., Yang, Z., Yang, C. and Ma, C. 2006. Modeling for gellan gum production by *Sphingomonas paucimobilis* ATCC 31461 in a simplified medium. *Applied and Environmental Microbiology* 72(5): 3367-3374.
- Wang, Y., Tian, Y., Han, B., Zhao, H.B., Bi, J.N. and Cai, B.L. 2007. Biodegradation of phenol by free and immobilized *Acinetobacter* sp. strain PD12. *Journal of Environmental Science* 19(2): 222-225.
- Wang, Y.X., Wang, H.B., Zhang, Y.Q., Xu, L.H., Jiang, C.L., and Li, W.J. 2008. *Rhodococcus kunmingensis* sp. nov., an actinobacterium isolated from a rhizosphere soil. *International Journal of Systematic and Evolutionary Microbiology* 58(6): 1467-71.
- Warren, R., Hsiao, W.W.L., Kudo, H., Myhre, M., Dosanjh, M., Petrescu, A., Kobayashi, H., Shimizu, S., Miyauchi, K., Masai, E., Yang, G., Stott, J.M., Schein, J.E., Shin, H., Khattra, J., Smailus, D., Butterfield, Y.S., Siddiqui, A., Holt, R., Marra, M.A., Jones, S.J., Mohn, W.W., Brinkman, F.S., Fukuda, M., Davies, J. and Eltis, L.D. 2004. Functional characterization of a catabolic plasmid from polychlorinated-biphenyl-degrading *Rhodococcus* sp. strain RHA1. *Journal of Bacteriology* 186: 7783-7795.
- Wasi, S., Jeelani, G. and Ahmad, M. 2008. Biochemical characterization of a multiple heavy metal, pesticides and phenol resistant *Pseudomonas fluorescens* strain. *Chemosphere* 71: 1348-1355.
- Watanabe, K. 2001. Microorganisms relevant to bioremediation. Current Opinion in Biotechnology 12: 237-241.
- Weber, M. and Weber, M. 2010. In Phenolic Resins: A Century of Progress. L. Pilato (Ed.), Springer-Verlag Berlin Heidelberg. pp 9-22. DOI: 10.1007/978-3-642-04714-5 2,
- Weir, S.C., Dupuis, S.P., Providenti, M.A., Lee, H., Trevors, J.T. 1995. Nutrient enhanced survival of and phenanthrene mineralization by alginate encapsulated and free *Pseudomonas* sp. UG14Lr cells in creosotecontaminated soil slurries. *Applied Environmental Microbiology* 43:946-951.
- Whiteley A.S., and Bailey M.J. 2000. Bacterial community structure and physiological state within an industrial phenol bioremediation system. *Applied Environmental Microbiology* 66: 2400-2407.
- Whiteley C.G. & Lee D.J. 2006. Enzyme technology and biolog- ical remediation. *Enzyme Microbial and Technology* 38: 291-316.

- Whiteley, A.S., Wiles, K., Lilley, J., Philip, MJ., Babailey. 2001. Ecological and physiological analyses of *Pseudomonad species within a phenol remediation system. Journal of Microbiological Methods* 44: 79-88.
- Whitely, A.S. and Bailey, M.Y. 2000. Bacterial community structure and physiological state within an industrial phenol bioremediation system. *Applied Environmental Microbiology* 66: 2400-2407.
- WHO (World Health Organization). 1994. Phenol. Environmental Health Criteria 161. WHO, Geneva.
- Windholz, M., ed. (1983) The Merck Index, 10th ed., Rahway, NJ, Merck Co.pp. 1043.
- Wong, P.W., Nik, M.S., Nachiappa, M. and Balaraman, V. 2002. Pre-treatment and membrane ultra filtration using treated palm oil mill effluent (POME). Songklanakarin Journal of Science and Technology 24:891-898.
- Woznica, A., Dzirba, J., Manka, D., and Abuzek. 2003. Effect of electron transport inhibitors on iron reduction in *Aeromonas hydrophila* strain KB1. *Anaerobe* 9(3): 125-130.
- Wright, K.P. 2003. Effect ofinorganic and organic nitrogen sources on the growth of *Micrococcus luteus, Rhodococcus rhodochrous, Sarcina aurantiaca, and Serratia marcescen* in-vitro. Johnson C. Smith University.
- Wu, Z., Wang, Y. and Xing, Z. 2005. Study on degrading phenol by immobilized *Ralstonia metallidurans* CH34. *Microbiology* 32(4): 31-36.
- Wu, Z.L. and Li, Z.Y. 2002. Enhancement of enzyme activity and enantioselectivity via cultivation in nitrile metabolism by *Rhodococcus* sp. CGMCC 0497. *Biotechnology Applied Biochemistry* 35:61-67.
- Xi-Pu, H., Hong-Jie, L., Min, Z., Jin-tian, L., Jun-Fang, L. and Bao-shan, C. 2012. Isolation, identification of Phenol-Degrading Strain F5-2 and its Degradation Characteristics. *Journal of Chemical Engineering of Chinese Universities*. 26(6): 1001-1008.
- Yamada, T., Takahama, Y. and Yamada, Y. 2008. Biodegradation of 2,4,6-tribromophenol by *Ochrobactrum* sp. strain TB01. *Bioscience, Biotechnology and Biochemistry* 72(5): 1264-1271.
- Yan, J., Jianping, W., Hongmei, L. and Suliang, Y. 2005. The biodegradation of phenol at high initial concentration by the yeast *Candida tropicalis*. *Biochemical Engineering Journal* (3): 243-247.
- Yang, C.F. and Lee, C.M. 2007. Enrichment isolation, and characterization of phenol-degrading *Pseudomonas resinovorans* strain P-1 and *Brevibacillus* sp. strain P-6. *International Biodeterioration and Biodegradation* 59(3): 206-210.

- Yang, R.D. and Humphrey, A.E. 1975. Dynamic and steady state studies of phenol biodegradation in pure and mixed cultures. *Biotechnology and Bioengineering* 17: 1211-1235.
- Yemendzhiev, H.M., Gerginova, A., Krastanov, I., Stoilova, I. and Alexieva, Z. 2008. Growth of *Trametes versicolor* on phenol. *Journal Industrial Microbiology and Biotechnology* 35: 1309-1312.
- Yeoh, B.G. 2005. A technical and economic analysis of heat and power generation from biomethanation of palm oil mill effluent. *Electricity Supply Industry in Transition: Issues and Prospect for Asia.* pp. 20-63.
- Yin, P., Xu, Q., Qu, R., Zhao, G. and Sun, Y. 2010. Adsorption of transition metal ions from aqueous solutions onto a novel silica gel matrix inorganic-organic composite material. *Journal of Hazardous Material* 173: 710–716.
- Yoon, J.H., Shim, Y.B., Lee, B.S., Choi, S. and Won, S.M. 2012. Electrochemical degradation of phenol and 2-chlorophenol using pt/ti and boron-doped diamond electrodes. *Bulletin Korean Chemical Society* 33(7): 2274-2278.
- Yu, C., Chen, F., Zhao, X., Guo, T., Lin, R., Qu, M. and Liu, Q. 2011. Isolation, identification and degrading characteristics of phenol-degrading bacteria B3.
   In Proceeding of International Conference on Electrical and Control Engineering, ICECE 2011 art. no. 6058317. pp. 3509-3512.
- Liu, Y.G; Pan, C.; Xia, W.B., Zeng, G.M, Zhou, M., Liu, Y.Y, Ke, J., Huang, C. 2008. Simultaneous removal of Cr and phenol in consortium culture of *Bacillus* sp. and *Pseudomonas putida Migula* (CCTCC AB92019). *Transaction on Nonferous Metals Society of China* 18(4): 1014-1102.
- Yusoff, S. and Hansen, S.B. 2007. Feasibility study of performing an life cycle assessment on crude palm oil production in malaysia. *The International Journal of Life Cycle Assessment* 12(1): 50-58.
- Zaitsev, G.M. and Surovtseva, E.G. 2000. Growth of *Rhodococcus opacus* on mixtures of monohalogenated benzenes and phenols. *Microbiology* 69(4): 401-405.
- Zaki, S. 2006. Detection of *meta* and *Helicobacter pylori* ortho-cleavage dioxygenases in bacterial phenol-degraders. *Journal of Applied Sciences and Environmental Management* 10(3): 75-81.
- Zhang, J., Sun, Z., Li, Y., Peng, X., Li, W., and Yan, Y. 2008. Biodegradation of pnitrophenol by *Rhodococcus* sp. CN6 with high cell surface hydrophobicity. *Journal of Hazardous Materials* 163: 723-728.
- Zhang, Y., Meng, X., Chai, T., Sheng, W. and Bao, W.X. 2013. Characterization of phenol-degrading *Rhodococcus* sp. strain P1 from coking wastewater. *Acta Microbiologica Sinica* 53(10): 1117-1124.

- Zhang, Z.Y., Pan, L.P. and Li, H.H. 2010. Isolation, identification and characterization of soil microbes which degrade phenolics allelochemicals. *Journal of Applied Microbiology* 8: 1839-1849.
- Zhang, X., Gao, P., Chao, Q., Wang, L., Senior, E. and Zhao, L. 2004. Microdiversity of phenol hydroxylase genes among phenol-degrading isolates of *Alcaligenes* sp. from an activated sludge system. *FEMS Microbiology Letter* 237(2): 369-375.
- Zhao, Q., Han, H., Fang, F., Zhuang, H., Wang, D., and Li, K. 2015. Strategies for recovering inhibition caused by phenolic compounds in a short-cut nitrogen removal reactor treating coal gasification wastewater. *Journal of Water Reuse and Desalination* 5(4): 569-578.
- Zheng, Y.G., Chen, J., Liu, Z.Q., Wu, M.H., Xing, L.Y. and Shen, Y.C. 2007. Isolation, identification and characterization of *Bacillus subtilis* ZJB-063, a versatile nitrile-converting bacterium. *Applied Microbiology and Biotechnology* 77: 985-993.
- Zheng, H., Yin, J., Gao, Z., Huang, H., Ji, X. and Dou, C. 2011. Disruption of *Chlorella vulgaris* cells for the release of biodiesel-producing lipids: a comparison of grinding, ultrasonication, bead milling, enzymatic lysis, and microwaves. *Applied Biochemistry Biotechnology* 164(7): 1215-1224.
- Zhou, B. 2014. Kinetics of chromium (VI) reduction and phenol biodegradation by *Pseudomonas* sp. JF122. *Pakistan Journal of Pharmaceutical Sciences* 27(6): 2047-2051.
- Zhou, J., Yu, X., Ding, C., Wang, Z., Zhou, Q., Pao, H. and Cai, W. 2011. Optimization of phenol degradation by *Candida tropicalis* Z-04 using Plackett-Burman design and response surface methodology. *Journal of Environmental Sciences* 23(1): 22–30.
- Zidkove, L., Szokol, J., Rucká, L., Pátek, M., and Ne, J. 2013. Biodegradation of phenol using recombinant plasmid-carrying *Rhodococcus erythropolis* strains. *International Biodeterioration and Biodegradation* 84: 179-184.