



UNIVERSITI PUTRA MALAYSIA

COMPLETE GENOME SEQUENCING AND ANALYSIS OF *Pasteurella multocida* STRAIN PMTB2.1 AND EXPRESSION OF SELECTED GENES IN IRON-RESTRICTED ENVIRONMENT

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By

SHAGUFTA JABEEN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

December 2017

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DEDICATION

To My Murshid Sayed Abdul Rasheed Mian Sarkar (Rehmatu-Allh Ailah)

To my respectable teachers, specially Prof. Dr. Saleem Hafiz

To my parents and my Husband, my son and to my all dear family members
I dedicate to all, my work with love and gratitude



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

COMPLETE GENOME SEQUENCING AND ANALYSIS OF *Pasteurella multocida* STRAIN *PMTB2.1* AND EXPRESSION OF SELECTED GENES IN IRON-RESTRICTED ENVIRONMENT

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Pasteurella multocida (PM) is a Gram-negative, facultative anaerobic bacterium, belonging to the family *Pasteurellaceae* that commonly found as commensal in the upper respiratory tract of mammals and birds. However, *P. multocida* is often associated with acute as well as chronic infections in avian and bovine leading to significant morbidity and mortality, such as pasteurellosis and hemorrhagic septicemia (HS) in cattle and buffaloes. *P. multocida* subspecies *multocida* strain *PMTB2.1* was first isolated from buffaloes died of septicemia. The bacterium has been characterized based on biochemical tests and molecular identification based on PCR. Interestingly, based on HS causing serogroup B-specific PCR (HSB-PCR), the isolate is not from serogroup B. Hence, an in depth genome wide analysis of *PMTB2.1* was carried out. In this study, the genome of *P. multocida* strain *PMTB2.1* was sequenced using third-generation sequencing technology, PacBio and analysed bioinformatically via *de novo* method followed by in depth characterization of the genome. In addition, expression of selected genes of *PMTB2.1* grown in iron-restricted condition was also demonstrated based on real-time PCR study.

Bioinformatics analysis based on *de novo* assembly of PacBio raw reads generated 3 supercontigs that were assembled to generate a draft genome with unresolved gaps regions. The gaps between the contigs in the assembled draft genome sequence were closed by PCR sequencing with primer walking strategy using Sanger sequencing. Start position of the circular genome of *PMTB2.1* was set based on homology to reference genome *P. multocida* strain *PM36950* and the circularity of the genome was confirmed by PCR. The complete genome sequence of *P. multocida* strain *PMTB2.1* is composed of a single circular chromosome of 2,315,138 base pairs with 40.32 % GC content and a total of 2,176 potential genes. The genome was submitted to NCBI

under the accession number, CP 007205.1. The annotated complete genome sequences of *PMTB2.1* have 2,097 protein-coding sequences, 19 rRNA genes, 56 tRNA and 4 ncRNA genes. The genome also encode for more than 41 CDS (2%) that involved in iron regulation or iron uptake, 160 virulence genes and 12 antibiotic resistance genes including the complete Tad locus. The tad locus encodes 14 gene including several previously uncharacterized genes such as *flp 2* that play important roles in the adhesion and colonization of the bacteria, biofilm formation as well as in pathogenesis of the disease.

Multi-locus sequence typing against Rural Industries Research and Development Corporation (RIRDC) scheme indicated that *PMTB2.1* matched to alleles from sequence type ST101. Comparative genome analysis showed that *PMTB2.1* is closely related with other *Pasteurella multocida* strains with genomic distance less than 0.13. However, synteny analysis showed that genome structure of *PMTB2.1* is more resembles to that of *P. multocida* serogroup A strain *PM36950* as compared to that of *P. multocida* serogroup F strain *PM70*. However, *PMTB2.1* genome lacks the Integrative Conjugative Element (ICE) of 86 kb that can only be detected in *PM36950*. Nevertheless, two intact prophage sequences of approximately 62 kb that were found in *PMTB2.1*, were absent in *PM36950* and *PM70*. One of the phages is similar to transposable Mu like phage SfMu; however, the phage regions of *PMTB2.1* were not associated with toxin-related genes, as detected in serogroup D toxigenic strain of *P. multocida*. Moreover, *PMTB2.1* complete genome is approximately 34,380 kb smaller than *PM36950* genome (2,349,518 bp), on the other hand approximately 15 kb specific region of *PMTB2.1* was absent in *PM70* genome. The capsular sequence analysis of *PMTB2.1* indicated that it is resembling the capsular sequence of *P. multocida* serogroup A with 99% sequence identity with A:1 capsular sequences. Furthermore, OrthoMCL analysis based on similarity among common genes showed that *PMTB2.1* was clustered with bovine isolates and were separated from other *P. multocida* strains that infect avian and swine.

Since *P. multocida* including *PMTB2.1* has more than 2% of the genome encode for iron-regulated genes, the expression profiling of iron uptake genes namely *fbpb*, *yfea*, *fece*, *fur* and sialidase encoded by *nana* were characterized under iron-restricted environment where *PMTB2.1* was grown in broth with and without iron chelating agent 2,2' Bipyridine. Results of this study reflect that iron-reduced conditions have significant effect on the expression profiles of iron-regulating genes ($p < 0.05$) and all of the four iron-related genes (*fbpb*, *yfea*, *fece*, *fur*) behave differently in response to iron reduction in media. The highest relative fold change (281.2 fold) of *fece* gene was observed at early, 30 minutes of treatment reveal that *P. multocida* may utilizes its periplasmic protein at early stage to acquire iron. Furthermore, down-regulation expression of *fece* from 4 to -1.5 with the elevated expression of other genes (*fbpb* and *yfea*) at later time points, 60 and 120 minutes suggest that *PMTB2.1* control their iron requirements in response to iron availability by down regulating the expression of iron proteins. Moreover, the significant increase ($p \leq 0.05$) in *fbpb* expression (25 fold) at time point 60 and in *Yfea* expression (26 fold) at early time point 30 minutes with highest expression (42 fold) at 120 minutes reflect the utilization of multiple iron

systems in *P. multocida* strain *PMTB2.1*. These results demonstrate the importance of iron in the survival of *P. multocida*.

In conclusion, this study has provided insight on the genomic structure of *PMTB2.1* in terms of potential genes that can function as virulence factors and comparative pathogenomic information of valuable importance for future study in elucidating the mechanisms behind the ability of the bacterium in causing diseases in susceptible animals.



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**JUJUKAN GENOM LENGKAP DAN ANALISIS *Pasteurella multocida*
STRAIN *PMTB2.1* DAN EKSPRESI GEN TERPILIH DALAM
PERSEKITARAN BESI TERHAD**

Oleh

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Pasteurella multocida (PM) ialah bakteria anaerob fakultatif Gram-negatif, kepunyaan famili *Pasteurellaceae* yang biasa ditemui dalam saluran pernafasan atasan mamalia dan unggas. Namun, *P. multocida* selalunya dikaitkan dengan penyakit akut serta kronik dalam unggas dan bovin yang boleh menyebabkan morbiditi dan kadar kematian yang signifikan, seperti penyakit pasteurelosis dan hawar berdarah (HS) dalam lembu dan kerbau. *PMTB2.1* mula diasingkan dari kerbau mati akibat septisemia. Bakteria tersebut telah dicirikan berdasarkan ujian biokimia dan pengenalpastian molekul berdasarkan PCR. Yang menariknya, berdasarkan PCR khusus bagi serokumpulan B penyebab HS (HSB-PCR), isolat tersebut bukan dari serokumpulan B. Maka, analisi genom menyeluruh secara mendalam ke atas *PMTB2.1* telah dilakukan. Dalam kajian ini, genom *P. multocida* strain *PMTB2.1* diunjukkan menggunakan teknologi jujukan generasi ketiga iaitu PacBio dan analisi bioinformatik dilakukan melalui keadah *de novo* dan diikuti oleh pencirian genom secara mendalam. Di samping itu, ekspresi gen terpilih *PMTB2.1* yang ditumbuhkan dalam persekitaran besi terhad juga dicirikan berdasarkan kajian PCR nyata-masa.

Analisis bioinformatik berdasarkan penggabungan data kasar PacBio secara *de novo* menjana tiga “supercontigs” yang dihimpunkan untuk menghasilkan draf genom dimana terdapat bahagian jurang yang tidak dapat dikenalpasti. Penjujukan PCR dengan strategi “primer walking” berdasarkan jujukan Sanger telah digunakan untuk mengenalpasti jurang antara kontig dalam jujukan draf genom. Posisi permulaan genom membulat *PMTB2.1* telah ditetapkan berdasarkan homologi genom rujukan *P. multocida* strain *PM36950* dan bukti genom membulat telah dikenalpasti dengan menggunakan PCR. Jujukan genom lengkap *P. multocida* strain *PMTB2.1* terdiri daripada satu kromosom membulat yang mempunyai 2,315,138 bp dan 40.32 %

jumlah GC serta 2,176 gen potensi. Jujukan genom *PMTB2.1* ini juga telah dimuat naik ke NCBI dengan nombor kemasukan *CP007205.2*. Hasil anotasi genom lengkap mendapati bahawa genom ini mempunyai 2,097 protein pengkodan, 19 gen rRNA, 56 gen tRNA dan 4 gen ncRNA. Genom *PMTB2.1* juga mengkod lebih daripada 41 CDs (2%) yang terlibat di dalam regulasi besi serta pengambilan besi, 160 gen virulen dan 12 gen rintangan antibiotik, termasuk lokus Tad lengkap yang mengkod 14 gene termasuk gene belum terciri seperti *flp2* yang memainkan peranan penting semasa pelekatan dan kolonisasi bakteria, pembentukan biofilm serta pathogenesis penyakit.

Pengelasan jujukan multilokus menggunakan skim *Rural Industries Research and Development Corporation (RIRDC)* menunjukkan *PMTB2.1* sepadan dengan alel daripada jujukan jenis ST101. Analisis komprehensif genom menunjukkan strain *PMTB2.1* adalah berkait rapat dengan *Pasteurella multocida* strain lain dengan jurang genom kurang daripada 0.13. Namun, analisis “synteny” menunjukkan struktur genom *PMTB2.1* adalah lebih bersamaan dengan *P. multocida* serokumpulan A strain *PM36950* berbanding dengan *P. multocida* serokumpulan F strain *PM70*. Tetapi, genom *PMTB2.1* tidak mempunyai “Integrative Conjugative Element” (ICE) yang bersaiz 82 kb yang hanya boleh didapati dalam *PM36950*. Walau bagaimanapun, dua jujukan profaj lengkap bersaiz kira-kira 62kb ditemui hanya di dalam *PMTB2.1*, tetapi tidak didapati dalam *PM36950* dan *PM70*. Salah satu faj didapati serupa dengan faj “transposable” Mu seperti SfMu. Namun, jujukan faj *PMTB2.1* tidak dikaitkan dengan gen toksin, yang terdapat pada *P. multocida* strain toksigenik dalam serokumpulan D. Tambahan pula, genom lengkap *PMTB2.1* adalah kira-kira 34,380 kb lebih kecil daripada genom *PM36950* (2,349,518 bp). Walau bagaimanapun, *PMTB2.1* mempunyai 15 kb bahagian khusus yang tidak hadir dalam genom *PM70*. Analisis jujukan kapsul *PMTB2.1* menunjukkan ia bersamaan dengan jujukan kapsul serokumpulan A *P. multocida* dengan 99% identiti jujukan kapsul A:1. Tambahan pula, analisis OrthoMCL berdasarkan persamaan antara gen umum menunjukkan *PMTB2.1* berkumpulan dengan isolat bovin dan adalah diasingkan dari strain *P. multocida* lain yang menjangkiti unggas dan babi.

Bakteria *P. multocida* termasuk strain *PMTB2.1* mempunyai lebih daripada 2% genom yang dikod untuk gen regulasi besi. Profil ekspresi pengambilan besi terutamanya *fbpb*, *yfea*, *fece*, *fur* dan sialidase yang terkod *nana* telah dicirikan di dalam persekitaran besi terhad di mana strain *PMTB2.1* telah ditumbuh di dalam media yang mempunyai pengikat besi, 2, 2' Bipyridine. Hasil kajian menunjukkan persekitaran besi terhad mempunyai kesan signifikan kepada profil ekspresi gen regulasi besi ($p < 0.05$) dan kesemua empat gen (*fbpb*, *yfea*, *fece*, *fur*) bertindak secara berlainan dengan pengurangan besi di dalam media. Ekspresi tertinggi (281.2 kali ganda) gene *fece* direkodkan pada pada peringkat awal, 30 minit pertama dan ini mencadangkan bahawa *P. multocida* menggunakan protein periplasmik pada peringkat awal untuk memperolehi besi. Tambahan pula, ekspresi *fece* menurun dari 4 ke -1.5 dengan peningkatan ekspresi gen lain (*fbpb* dan *yfea*) pada masa terkemudian, 60 dan 120 minit mencadangkan bahawa bakteria mengawal keperluan besi mengikut keperluan besi dengan menurunkan ekspresi protein besi. Tambahan lagi peningkatan secara signifikan ($p \leq 0.05$) pada ekspresi *fbpb* (25 kali ganda) pada

minit ke 60 dan ekspresi *yfeA* (26 kali ganda) pada masa seawal 30 minit dengan ekspresi tertinggi (42 kali ganda) pada minit ke 120 menunjukkan bahawa *P. multocida* strain *PMTB2.1* menggunakan sistem besi berbilang. Hasil kajian ini menunjukkan kepentingan besi dalam daya tahan hidup *P. multocida*.

Kesimpulannya, kajian ini telah memberi pemahaman dalam struktur genom *PMTB2.1* dari segi gen potensi yang berfungsi sebagai faktor virulen dan maklumat patogenomik komparatif yang berharga untuk kajian pada masa akan datang seperti mekanisme yang terlibat dalam keupayaan bakteria tersebut dalam menyebabkan penyakit.



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I certify that a Thesis Examination Committee has met on 13 December 2017 to conduct the final examination of Shagufta Jabeen on her thesis entitled "Complete Genome Sequencing and Analysis of *Pasteurella multocida* Strain PMTB2.1 and Expression of Selected Genes in Iron-Restricted Environment" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

ACT	Artemis Comparison Tool
APV	alum-precipitated vaccine
Bp	base pair
BLAST	Basic Local Alignment Search Tool
B2G	BLAST to Gene Ontology
BP	biological process
CC	cellular component
CDs	coding sequences
cDNA	complementary DNA
CGView	Circular Genome Viewer
DNA	deoxyribonucleic acid
°C	degree Celsius
dsDNA	double-stranded DNA
EMBL	European Molecular Bioinformatics Laboratory
GC	guanosine/cytosine
GFF	General Feature Format
GO	gene ontology
HA	hyaluronic acid
HS	hemorrhagic septicemia
Kb	kilobase
kDA	kilodalton
LPS	lipopolysaccharide
min	minute

mL	milliliter
μl	microliter
MF	molecular function
MLST	multi locus sequence typing
Ng	nanogram
nM	nanomolar
NGS	next-generation sequencing
ORF	open reading frame
OM	outer membrane
OAV	oil adjuvant vaccine
PCR	polymerase chain reaction
PM	<i>Pasteurella multocida</i>
RNA	ribonucleic acid
RIRDC	Rural Industries Research and Development Corporation
mRNA	messenger RNA
rRNA	ribosomal RNA
tRNA	transcript RNA
<i>Taq</i>	<i>Thermus aquaticus</i>
TAE	Tris-acetic-EDTA
VF _s	virulence factors
UPM	Universiti Putra Malaysia

CHAPTER 1

GENERAL INTRODUCTION

P. multocida is a Gram-negative facultative anaerobic bacterium. It belongs to family *Pasteurellaceae* (Kuhnert and Christensen, 2008) and is an important opportunistic pathogenic bacterium of human and animals. It carries different types of polysaccharides on its capsule and based on that *P. multocida* is grouped into A, B, D, E and F capsular types and further classified into 16 serotypes (1-16) based on lipopolysaccharide antigen (Carter, 1955; Heddleston *et al.*, 1972). *P. multocida* is found as a commensal in many mammals including cattle, buffaloes, domestic cats, dogs and birds in the upper respiratory tract; however, under certain circumstances it can cause opportunistic infections (Boyce *et al.*, 2010). *P. multocida* is associated with a wide range of veterinary diseases in wild and domestic animals of economic significance throughout the world (Harper *et al.*, 2006). In general, *P. multocida* is often associated with chronic as well as acute infections in animals that can lead to significant morbidity manifested as pasteurellosis, pneumonia, atrophic rhinitis, dermonecrosis cellulitis and hemorrhagic septicemia. *P. multocida* can cause pneumonia and respiratory disease in cattle, fowl cholera in avian and hemorrhagic septicemia (HS) and pasteurellosis in cattle and buffaloes.

Pasteurellosis caused by *P. multocida* is an acute septicemic disease characterized by high morbidity and is a high-impact disease in livestock, according to the World Animal Health Organization, Office International des Epizooties, (OIE, 2012). The predominant syndrome of pasteurellosis caused by *P. multocida* in cattle and buffaloes is referred to as bovine respiratory disease (BRD) which is manifested by pneumonic pasteurellosis (Anderson and Rings 2009). In addition, *P. multocida* serogroup A has been implicated in fatal pneumonia of cattle in India (Dabo *et al.*, 2007).

HS is an acute septicemic disease caused by *P. multocida* serogroups B: 2 and E: 2 in cattle and buffaloes and is the most important veterinary bacterial disease of ruminants (De Alwis, 1999; OIE, 2012). Moreover, previous study also indicated the involvement of multiple strains of *P. multocida* in a single outbreak of HS in cattle and buffalo based on molecular study (Bisvas *et al.*, 2004) although HS caused by serogroup A strain is only occasionally seen in North America (Rimler and Wilson, 1994; Taylor *et al.*, 1996). *Pasteurella* persist in crypts of the tonsils of experimentally exposed buffaloes and induced carrier animal and is hard to detect as animals remain healthy for several months (De Alwis *et al.*, 1990; Harper *et al.*, 2006). Carrier animals present in the endemic area act as reservoir for new outbreaks. The modes of transmission of the disease were through aerosol or ingestion of feed and water contaminated with infected saliva and discharges.

Over the past 15 year, major advancement has been made in genome sequencing research through the development of next-generation sequencing (NGS) technology

such as 454 Life Science, Solex Illumina, SOLiD and Ion Personal Genome Machine (Quail *et al.*, 2012). This technology has clear advantages over the Sanger sequencing method in term of their high throughput and low cost and the ability to perform multiple parallel analyses simultaneously. Furthermore, with further improvement in NGS technology the development of Pacific Biosciences (PacBio) (English *et al.*, 2012) and Oxford Nanopore Technologies (Timp *et al.*, 2010) based NGS platforms have greatly influenced the field of bacterial genome sequencing . Consequently, not only are the number of completed bacterial genomes increase, but also the coverage and the size of sequenced genomes. Currently, one hundred and twenty one (121) complete or draft *P. multocida* genomes are publicly available in Gene Bank (NCBI, National Center for Biotechnology). However, none of the characterized *P. multocida* genomes were sequenced using Pacific Biosciences except for the recent genome sequencing of *P. multocida* serogroup B: 2 strain Razi 0001(CP017961.1).

The first genomic structure of *P. multocida* was determined by the analysis of the first completely sequenced genome of avian isolate *PM70* (serogroup F) accession no. AE004439.1 (May *et al.*, 2001) which identifies a number of predicted virulence genes and iron uptake genes. At present, only a few genomes have been studied and examined in detail such as *PM36950* (serogroup A) (Michael *et al.*, 2012), PM HB01 (serogroup A) (Peng *et al.*, 2016) and genomes of *P. multocida* strains harboring *P. multocida* toxin (PMT) gene such as *PMHN06* (serogroup D) (Liu *et al.*, 2012).

Currently, the complete genome sequences of *P. multocida* capsular serogroup A, D and F have been generated and characterized and are available publicly as complete or draft genome. However, the complete genome of HS causing serogroups (B: 2 and E: 5) is not available until the recent release of complete genome of Razi 0001 (CP017961.1) a vaccine strain for pasteurellosis in January 2017. Recently, a Malaysian isolate of *P. multocida* serogroup B: 2 strain PMTB have been sequenced as draft genome (AWTD01000000) (Yap *et al.*, 2013). But none of the Malaysian *Pasteurella* strain has been sequenced as a finish high quality complete genome, nor analyzed hence there is no published complete genome sequence of *P. multocida* from Malaysia. Furthermore, *Pasteurella multocida* virulent genes and their interactions in influencing the virulence of the bacteria and pathogenicity in the infected animals are poorly characterized. One of the main reasons is that only a few complete genomes of *Pasteurella multocida* belongs to serogroups A and F has been analyzed and studied in detail and most of the complete genome produced during last decade is present as unpublished data. Also the differences in the genetic structure of the major virulence-related factors between different strains are poorly understood.

Iron is an important nutrient for nearly all life forms. It is possible that iron acquisition in *P. multocida* plays an important role in its survival and pathogenesis in the host, particularly considering that more than 2.5% (53 coding DNA sequences which was later found to be more than 60) of the *PM70* genes are predicted to encode proteins homologous to known proteins involved in iron uptake or acquisition (May *et al.*, 2001; Boyce *et al.*, 2012). Bacterial pathogens, when in a vertebrate host environment, will encounter a depletion of iron, triggering release of the transcriptional control of

ferric uptake regulator (Fur), which represses genes under its control in the presence of iron. Gene expression profiling under iron-limiting conditions based on microarray experiment have identified several iron acquisition genes at increased expression levels, and indeed, different sets of genes appear to be expressed in response to the nature of the iron source (Boyce *et al.*, 2012). In addition, *P. multocida* strains have siderophore-independent iron acquisition systems homologous to the *Actinobacillus* AfeABCD system and the periplasmic binding protein-dependent iron transport systems homologous to *E. coli* FecBCDE and Neisseria FbpABC system (Wilson and Ho, 2013). The presence of multiple iron acquisition systems in *Pasteurella* species may account for their ability to acquire iron under iron limited condition by utilizing certain set of gene under certain conditions.

The general objective of this study is to characterize the complete genome sequence of *P. multocida* strain *PMTB2.1* and to determine the functional importance of selected iron-related genes of the bacteria. Hence, the specific objectives of this study are:

1. To sequence the genome of *Pasteurella multocida* strain *PMTB2.1* based on third-generation sequencing technology using PacBio platform and to perform *de novo* assembly of the generated genome sequences ;
2. To generate and annotate the complete genome sequences of *PMTB2.1* based on a reference genome *P. multocida* strain *PM36950* ;
3. To analyze the complete genome sequences of *PMTB2.1* based on identifications of important sequence motifs and to compare the genome with other *P. multocida* complete genome sequences by using various bioinformatics tools ;
4. To determine the expression profile of selected iron-acquiring genes of *P. multocida* strain *PMTB2.1* grown under iron-restricted environment.

REFERENCES

- Abdul Aziz Bin Saharee. (1992). Studies on Hemorrhagic septicemia in cattle and buffaloes in Peninsular Malaysia. Ph.D. thesis, Universiti Putra Malaysia.
- Adhikari, P., Berish, S.A., Nowalk, A.J., Veraldi, K.L., Morse, S.A. and Mietzner, T.A. (1996). The *fbpABC* locus of *Neisseria gonorrhoeae* functions in the periplasm-to-cytosol transport of iron. *J Bacteriol*, 178, 2145-2149.
- Agersø, Y. and Petersen, A. (2007). The tetracycline resistance determinant Tet 39 and the sulphonamide resistance gene *sulII* are common among resistant *Acinetobacter* spp isolated from integrated fish farms in Thailand, (November 2006). *J Antimicrob Chemother*, 59(1), 23–27. doi.org/10.1093/jac/dkl419.
- Ali, O.S., Adamu, L., Abdullah, F.F.J., Abba, Y., Hamzah, H.B., Yusuf, A.,... Mohd Zamri-Saad. (2015) Haematological and histopathological vicissitudes following oral inoculation of graded doses of *Pasteurella multocida* type B: 2 and its lipopolysaccharide in mice. *J Vet Sci Technol*, 6, 220. doi:10.4172/2157-7579.1000220.
- Altschul, S.F., Gish, W., Miller, W., Myers, E.W. and Lipman, D.J. (1990). Basic local alignment search tool. *J Mol Biol*, 215, 403-410.
- Anderson, D.E., Rings M.D. (2009). Current Veterinary Therapy: Food Animal Practice, 5th Ed. Saunders, St. Louis MO. pp. 164-170.
- Ameen, Mahmood Jamal H, Chua K, Frederick D, Mahmood Ameen A, S. (2005). plasmid dna analysis of *Pasteurella multocida* serotype B isolated from Hemorrhagic septicemia outbreaks in Malaysia. *Malay J Microbiol*, 1(2), 35–39.
- Andrews, S. C., Robinson, A. K. and Rodríguez-Quñones, F. (2003). Bacterial iron homeostasis. *FEMS Microbiol Rev*, 27(2–3), 215–237. [https://doi.org/10.1016/S0168-6445\(03\)00055-x](https://doi.org/10.1016/S0168-6445(03)00055-x).
- Angerer, A., Gaisser, S. and Braun, V. (1990). Nucleotide sequences of the *sfuA*, *sfuB*, and *sfuC* genes of *Serratia marcescens* suggest a periplasmic-binding-protein-dependent iron transport mechanism. *J Bacteriol*, 172(2), 572–8.
- Annas, S., Zamri-Saad, M., Jesse, F. F. A. and Zunita, Z. (2014). New sites of localisation of *Pasteurella multocida* B:2 in buffalo surviving experimental hemorrhagic septicemia. *BMC Vet Res*, 10(1), 88. doi.org/10.1186/1746-6148-10-88.
- Assefa, S.1., Keane, T.M., Otto, T.D., Newbold, C., Berriman, M. (2009). ABACAS: algorithm-based automatic contiguation of assembled sequences. *Bioinformatics*, 25(15), 1968-9. doi: 10.1093/bioinformatics/btp347.

- Bauer, A. W., Kirby, W. M., Sherris, J. C., Turck, M. (1966). Antibiotic susceptibility testing by a standardized single disk method. *Am J Clin Pathol*, 45(4), 493-496.
- Bearden, S. W. and Perry, R. D. (1999). The Yfe system of *Yersinia pestis* transports iron and manganese and is required for full virulence of plague. *Mol Microbiol*, 32(2), 403–414. doi.org/mmi1360 [pii].
- Beinert, H., Holm, R., and Münck, E. (1997). Iron-sulfur clusters: nature's modular, multipurpose structures. *Science*, 277(5326), 653-659.
- Benkirane, A., and De Alwis, M. C. L. (2002). Hemorrhagic septicemia, its significance, prevention and control in Asia. *Vete Medic*, 47(8), 234–240.
- Bernard, C. S., Bordi, C., Termine, E., Filloux, A., and De Bentzmann, S. (2009). Organization and PprB-dependent control of the pseudomonas aeruginosa tad locus, involved in Flp pilus biology. *J Bacteriol* 191(6), 1961–1973. https://doi.org/10.1128/JB.01330-08.
- Besemer, J. and Borodovsky, M. (2005). GeneMark: web software for gene finding in prokaryotes, eukaryotes and viruses. *Nucleic Acids Res*, 33, 451–454. dx.doi.org/10.1093/nar/gki487.
- Biswas, A.1., Shivachandra, S. B., Saxena, M.K., Kumar, A.A., Singh, V.P., Srivastava, S. K. (2004). Molecular variability among strains of *Pasteurella multocida* isolated from an outbreak of haemorrhagic septicaemia in India. *Vet Res Commun*, 28(4), 287-298.
- Broeders, S., Huber, I., Grohamann, L., Berben, G., Tavemiers, I.... Morisset D. (2014). Guidelines for validation of qualitative real-time PCR methods. *Trends Food Sci and Technol*, 37, 115-126.
- Bronzwaer, S. L., O. Cars, U. Buchholz, S. Mölsted, W. Goettsch, I. K. Veldhuijzen, J. L. Kool, M. J. Sprenger, and J. E. Degener. (2002.) A European study on the relationship between antimicrobial use and antimicrobial resistance. *Emerg Infect Dis*, 8, 278-282.
- Bosch, M., Tarragó, R., Garrido, M. E., Campoy, S., Fernández de Henestrosa, A. R., Pérez de Rozas, A. M., ... Barbé, J. (2001). Expression of the *Pasteurella multocida* ompH gene is negatively regulated by the Fur protein. *FEMS Microbiol Lett*, 203(1), 35–40. doi.org/10.1016/S0378-1097(01)00329-9.
- Bosch, M., Garrido, M.E., Llagostera, M., de Rozas, A.M.P., Badiola, I. and Barbe, J.(2002). Characterization of the *Pasteurella multocida* hgbA gene encoding a hemoglobin- binding protein. *Infect Immun*, 70, 5955–5964.
- Boyce, J., D. and Adler, B. (2000). The capsule is a virulence determinant in pathogenesis of *Pasteurella multocida* M1404 (B: 2). *Infct.Immu*, 68, 3463-3468.

- Boyce, J., D. Chung, J., Y. and Adler, B. (2000). Genetic organisation of the capsule biosynthetic locus of *Pasteurella multocida* M1404 (B:2). *Vet Microbiol*, 72(1-2), 121-34.
- Boyce, J.D., Seemann, T., Adler, B., Harper, M. (2012). Pathogenomics of *Pasteurella multocida*. *Curr. Top. Microbiol. Immunol.* 361, 23–38. doi.org/10.1007/82_2012_203.
- Bustin, S. A., Benes, V., Nolan, T., and Pfaffl, M. W. (2005). Quantitative real-time RT-PCR - a perspective. *J Mol Endocrinol*, 34(3), 597–601. doi.org/10.1677/jme.1.01755.
- Butler, J., MacCallum, I., Kleber, M., Shlyakhter, I.A., Belmonte, M.K.,...Jaffe, D. B. (2008). ALLPATHS: de novo assembly of whole-genome shotgun microreads. *Genome Res*, 18(5), 810–820.
- Bubner, B. and Baldwin, I.T. (2004). Use of real-time PCR for determining copy number and zygosity in transgenic plants. *Plant Cell Rep*, 23(5), 263-71.
- Bustin, S. A. V., Benes, J. A., Garson, J., Hellemans, J., Huggett, M., Kubista R.... Carl, T. W. (2009). The MIQE guidelines: minimum information for publication of quantitative real-time pcr experiments. *Clin Chem*, 55, 611-622.
- Byers, B. R., and J. E. L. Arceneaux. 1998. Microbial iron transport: iron acquisition by pathogenic microorganisms. In *Metal ions in biological systems: iron transport and storage in microorganisms, plants, and animals*, ed. A. Sigel and H. Sigel, 35, p. 37-66. New York, NY: Marcel Dekker, Inc.
- Capitini C. M., Herrero, I.A., Patel, R., Ishitani, M.B. and Boyce, T., G. (2002). Wound infection with *Neisseria weaveri* and a Novel sub species of *Pasteurella multocida* in a child who sustained a tiger bite. *Clin Infec Dis*, 34(12), 74–76.
- Carver, T. J., Rutherford, K. M., Berriman, M., Rajandream, M., Barrell, B. G., and Parkhill, J. (2005). ACT: The Artemis comparison tool. *Bioinformatics*, 21(16), 3422–3423. doi.org/10.1093/bioinformatics/bti553.
- Carter, G.R. (1955). A haemagglutination test for the identification of serological types. *Am. J. Vet. Res*, (16), 481–484.
- Carter, G. R., and De Alwis, M. C. L. (1989). Hemorrhagic septicemia. In *Pasteurella and Pasteurellosis*, ed. Adlam, C. and Rutter, pp. 131–160. London: Academic Press.
- Chen, L., Zheng, D., Liu, B., Yang, J., and Jin, Q. (2015). VFDB 2016 : hierarchical and refined dataset for big data analysis — 10 years on, 44(November 2015), 694–697. https://doi.org/10.1093/nar/gkv1239.

- Chain, D. V., Grafham, R. S., Fulton, M. G., FitzGerald, J., Hostetler, D., Muzny, J. A., Birren, B., Bruce, D. C., Buhay, C., Cole, J. R., ... Tiedje, G. Weinstock, A. W. (2009). Genome project standards in a new era of sequencing. *Science*, 326(1), 236–237.
- Chung, J.Y., Zhang, Y. and Adler B. (1998). The capsule biosynthetic locus of *Pasteurella multocida* A:1. *FEMS Microbiol Lett*, 166, 289–296.
- Clock, S. A., Planet, P. J., Perez, B. A., and Figurski, D. H. (2008). Outer membrane components of the tad (tight adherence) secreton of *Aggregatibacter actinomycetemcomitans*. *J Bacteriol*, 190(3), 980–990. <https://doi.org/10.1128/JB.01347-07>.
- Corfield, T. (1992). Bacterial sialidases--roles in pathogenicity and nutrition. *Glycobiol*, 2(6), 509–21.
- Creevey, C. J., Muller, J., Doerks, T., Thompson, J. D., Arendt, D., and Bork, P. (2011). Identifying single copy orthologs in metazoa. *PLoS Computational Biol*, 7(12). doi.org/10.1371/journal.pcbi.1002269.
- Dabo SM, Confer AW, Quijano-Blas RA. (2003). Molecular and immunological characterization of *Pasteurella multocida* serotype A:3 OmpA: evidence of its role in *P. multocida* interaction with extracellular matrix molecules. *Microb. Pathog*, 35, 147–157.
- Dabo, S.M., Confer, A.W., Hartson, S.D. (2005). Adherence of *Pasteurella multocida* to fibronectin. *Vet. Microbiol*, 110, 265–275.
- Dabo, S. M., Taylor, J. D., and Confer, A., W. (2007). *Pasteurella multocida* and bovine respiratory disease. *Animal health research reviews / Conference of Research Workers in Animal Diseases*, 8(2), 129–150. doi.org/10.1017/S1466252307001399.
- Darling, A. C. E., Mau, B., Blattner, F. R., and Perna, N. T. (2004). Mauve: multiple alignment of conserved genomic sequence with rearrangements. *Genome Res*, 14(7), 1394–1403. doi.org/10.1101/gr.2289704.
- Darling, A. E., Mau, B., Perna, N. T., Batzoglou, S., and Zhong, Y. (2010). progressive Mauve: Multiple genome alignment with gene gain, loss and rearrangement. *PLoS ONE*, 5(6), e 11147. doi.org/10.1371/journal.pone.0011147.
- David H. Figurski, Daniel H. Fine, Brenda A. Perez-Cheeks, Valerie W. Grosso, Karin E. Kram, J. H., and Ke Xu and Jamila Hedhli. (2013). Targeted mutagenesis in the study of the tight adherence (tad) locus of *Aggregatibacter actinomycetemcomitans*. In Genetic Manipulation of DNA and Protein - Examples from Current Research, ed. D. Figurski, pp. 953–978. InTech publisher. doi.org/10.5772/55714.

- Deangelis, P.L. and White, C.L. (2004) Identification of a distinct, cryptic heparosan synthase from *Pasteurella multocida* types A,D, and F. *J Bacteriol*, 186, 8529–8532.
- De Angelis P.L. and Padgett-McCue A.J. (2000). Identification and molecular cloning of a chondroitin synthase from *Pasteurella multocida* type F. *J Biol Chem*, 275, 24124–24129.
- De Alwis, M. C. L. (1984). Hemorrhagic septicemia in cattle and buffaloes. *Rev Sci Tec Off Int Epiz*, 3(4), 707–730. <https://doi.org/10.20506/rst.3.4.185>.
- De Alwis, M. C. L. (1999). *Hemorrhagic Septicemia ACIAR Monograph Series*. Australia: ACIAR.
- De Alwis, M. C. L., Wijewardana, T. G., Gomis, A. I. U., and Vipulasiri, A. A. (1972). Persistence of the carrier status in hemorrhagic septicemia (*Pasteurella multocida* serotype 6:B infection) in Buffaloes. *Trop Anim Health and Prod*, 16(4), 925–36. doi.org/10.1007/BF02241015.
- Delilhas, N. (2011). Impact of small repeat sequences on bacterial genome evolution. *Ge Biol and Evol*, 3, 959–973. doi.org/10.1093/gbe/evr077.
- Delcher, A.L., Bratke, K.A., Powers, E.C., Salzberg, S.L. (2007). Identifying bacterial genes and endosymbiont DNA with Glimmer. *Bioinformatics*, 23, 673–679.
- De Sá, P. H. C. G., Miranda, F., Veras, A., de Melo, D. M., Soares, S., Pinheiro, K., ... Ramos, R. T. J. (2016). GapBlaster—A Graphical gap filler for prokaryote genomes. *PloS One*, 11(5), e0155327. doi.org/10.1371/journal.pone.0155327.
- Drzeniek, R., Scharmann, W., and Balke, E. (1972). Neuraminidase and N-acetylneuraminase pyruvate-Lyase of *Pasteurella multocida*. *J Gen Microbiol*, 72(2), 357–368. doi.org/10.1099/00221287-72-2-357.
- Edwards, D. J., and Holt, K. E. (2013). Beginner’s guide to comparative bacterial genome analysis using next-generation sequence data. *Microb Inform Exp*, 3(1), 2. <https://doi.org/10.1186/2042-5783-3-2>.
- Emily J. H., Hodgson, J. C., Lainson, F. A., & Zadoks, R. N. (2011). Multilocus sequence typing of a global collection of *Pasteurella multocida* isolates from cattle and other host species demonstrates niche association. *BMC Microbiol*, 11, 115. doi.org/10.1186/1471-2180-11-115
- English, A. C., Richards, S., Han, Y., Wang, M., Vee, V., Qu, J., ... Gibbs, R. A. (2012). Mind the gap: Upgrading genomes with Pacific Biosciences RS long-read sequencing technology. *PloS One*, 7(11), 1–12. <https://doi.org/10.1371/journal.pone.0047768>.
- Escolar, L., Pérez-Martín, J., and de Lorenzo, V. (1999). Opening the iron box: transcriptional metalloregulation by the Fur protein. *J. Bacteriol*, 181(20), 6223–9.

- Field, D., Hughes, J., and Moxon, E. R. (2004). Using the genome to understand pathogenicity. *Methods Mol Biol (Clifton, N.J.)*, 266, 261–287. <https://doi.org/10.1385/1-59259-763-7:261>.
- FAO, (1991). Proceedings of 4th International workshop on Hemorrhagic septicemia. Kandy, Sri Lanka. FAO–APHCA. Pub. No. 1991/13.
- Faez, F. J. A., Lawan, A., Abdinasir, Y. O., Zunita, Z., Rasedee, A. (2013) Clinicopathological responses of calves associated with infection of *Pasteurella multocida* type B and the bacterial lipopolysaccharide and outer membrane protein immunogens. *Int J Anim Vet Adv*, 5, 190-198.
- Fernandez, D., Henestrosa, A.R., Badiola, I., Saco, M., Perez, Rozas, D.A.M., Campoy, Frawley, E. R., and Fang, F. C. (2014). The ins and outs of bacterial iron metabolism. *Mol Microbiol*, 93(4), 609–616. doi.org/10.1111/mmi.12709.
- S., Barbe, J. (1997). Importance of the galE gene on the virulence of *Pasteurella multocida*. *FEMS Microbiol Lett*, 154, 311–316.
- Garcia-del Portillo, F., Foster, J. W., and Finlay, B. B. (1993). Role of acid tolerance response genes in *Salmonella typhimurium* virulence. *Infect Immun*, 61(10), 4489–92.
- Gautam, R., Kumar, A. A., Singh, V. P., Singh, V. P., Dutta, T. K., and Shivachandra, S. B. (2004). Specific identification of *Pasteurella multocida* serogroup-A isolates by PCR assay. *Res Vet Sci*, 76(3), 179–85. <https://doi.org/10.1016/j.rvsc.2003.10.005>.
- Gautam, R., Dutta, T.K., Kumar, A.A. (2006). Molecular typing of Indian isolates of *Pasteurella multocida* serogroup-A from different animal species. *Indian J Anim Sci*, 76, 867–872.
- Gordon D, Desmarais C, Green P. (2001). Automated finishing with autofinish. *Genome Res*, 11(4), 614-25.
- Habrun, B., Kompes, G., Cvetni, Ž., Špi, S., Beni, M., and Mitak, M. (2010). Antimicrobial sensitivity of *Escherichia coli*, *Salmonella* spp., *Pasteurella multocida*, *Streptococcus suis* and *Actinobacillus pleuropneumoniae* isolated from diagnostic samples from large pig breeding farms in Croatia. *Vet Arh*, 80(5), 571–583.
- Harper, M., Boyce, J. D., and Adler, B. (2006). *Pasteurella multocida* pathogenesis: 125 years after Pasteur. *FEMS Microbiol Lett*, 265(1), 1–10. doi.org/10.1111/j.1574-6968.2006.00442.x.
- Harper, M., Cox, A.D., St Michael, F., Wilkie I.W., Boyce, J.D., Adler, B. (2004) A heptosyltransferase mutant of *Pasteurella multocida* produces a truncated lipopolysaccharide structure and is attenuated in virulence. *Infect Immun*, 72, 3436–43.

- Harris, M. A., J. Clark., A. Ireland., J. Lomax., M. Ashburner., R. Foulger., K. Eilbeck., S. Lewis.,... R. White. (2004). The Gene Ontology (GO) database and informatics resource: Gene Ontology Consortium. *Nucleic Acids Res*, 32, 258-261.
- Heddleston, K. L., Gallagher, J. E., and Rebers, P. A. (1972). Fowl cholera: gel diffusion precipitin test for serotyping *Pasteurella multocida* from avian species. *Avian Dis*, 16(4), 925–36.
- Heddleston, K.L., Watko, L.P. and Rebers, P.A. (1964). Dissociation of a fowl cholera strain of *Pasteurella multocida*. *Avian Dis*, 8, 649–657.
- Hermann, D. and Foerzler, D. (2002). Specific amplification of difficult PCR products, from small amounts of DNA using FastStart Taq DNA polymerase *Biochemical*, 4, 25-26.
- Heid, C. A., Stevens, J., Livak, K. J., and Williams, P. M. (1996). Real-time quantitative PCR. *Genome Res*, 6(10), 986–94.
- Higgs, P. I., Larsen, R. A., and Postle, K. (2002). Quantification of known components of the *Escherichia coli* TonB energy transduction system: TonB, ExbB, ExbD and FepA. *Mol Microbiol*, 44(1), 271–281. doi.org/10.1046/j.1365-2958.2002.02880.
- Higuchi, R., Fockler, C., Dollinger, G., and Watson, R. (1993). Kinetic PCR analysis: real-time monitoring of DNA amplification reactions. *Biotech (NY)*, 11(9), 1026–30.
- Holland, P. M., Abramson, R. D., Watson, R., and Gelfand, D. H. (1991). Detection of specific polymerase chain reaction product by utilizing the 5'----3' exonuclease activity of *Thermus aquaticus* DNA polymerase. *Proc Nat Acad Sci*, 88(16), 7276–7280.
- Horsburgh, M. J., Ingham, E., and Foster, S. J. (2001). In *Staphylococcus aureus*, Fur is an interactive regulator with perr, contributes to virulence, and is necessary for oxidative stress resistance through positive regulation of catalase and iron homeostasis. *J Bacteriol*, 183(2), 468–475. doi.org/10.1128/JB.183.2.468-475.2001.
- Hodgson, J.C., Finucane, A., Dagleish, M.P., Ataei, S., Parton, R., Coote, J.G. (2005). Efficacy of vaccination of calves against hemorrhagic septicemia with a live *aroA* derivative of *Pasteurella multocida* B:2 by two different routes of administration. *Infect Immun.*, (73), 1475-1481.
- Hunt, M. L., Adler, B., and Townsend, K. M. (2000). The molecular biology of *Pasteurella multocida*. *Vet Microbiol*, 72(1–2), 3–25. doi.org/10.1016/S0378-1135(99)00183-2.

- Horadagoda, N. U., Hodgson, J.C., Moon, G.M., Wijewardana, T.G., Eckersall, P.D. (2001). Role of endotoxin in the pathogenesis of hemorrhagic septicemia in the buffalo. *Microb. Pathog*, 30, 171–178.
- Jabeen, A., Khattak, M., Munir, S., Jamal, Q., and Hussain, M. (2013). antibiotic susceptibility and molecular analysis of bacterial pathogen *pasteurella multocida* isolated from cattle, *J. App Pharma Sci*, 3(4), 106–110. doi.org/10.7324/JAPS.2013.3419.
- Janakiraman, A., and Slauch, J. M. (2000). The putative iron transport system SitABCD encoded on SPI1 is required for full virulence of *Salmonella typhimurium*. *Mol Microbio*, 35(5), 1146–1155. doi.org/10.1046/j.1365-2958.2000.01783.
- Kachlany, S. C., Planet, P. J., Desalle, R., Fine, D. H., Figurski, D. H., and Kaplan, J. B. (2001). flp-1, the first representative of a new pilin gene subfamily, is required for non-specific adherence of *Actinobacillus actinomycetemcomitans*. *Mol Microbiol*, 40(3), 542–54.
- Kehrenberg, C., Werckenthin, C., Schwarz, S. (1998). Tn5706, a transposon-like element from *Pasteurella multocida* mediating tetracycline resistance. *Antimicrob. Agents Chemother*, (42), 2116–2118.
- Kehrenberg C., Schulze-Tanzil G., Martel J.-L., Dancla E.C. and Schwarz S. (2001). Antimicrobial resistance in *Pasteurella* and *Mannheimia*: Epidemiology and genetic basis. *Vet. Res.*, 32, 323–339.
- Kim, B. J., Park, J. H., Park, T. H., Bronstein, P. A., Schneider, D. J., Cartinhour, S. W., and Shuler, M. L. (2009). Effect of iron concentration on the growth rate of *Pseudomonas syringae* and the expression of virulence factors in *hrp*-inducing minimal medium. *App Environ Microbiol*, 75(9), 2720–2726. http://doi.org/10.1128/AEM.02738-08.
- Kuhnert. P., Christensen, H. (2008). *Pasteurellaceae: Biology, Genomics and Molecular Aspects*. pp. 34-39. Caister Academic Press.
- Khadak Singh Bisht. (2006). Epidemiology of hemorrhagic septicemia in cattle and buffaloes in Peninsular Malaysia. Ph.D. thesis, University Putra, Malaysia.
- Köster, W. (2001). ABC transporter-mediated uptake of iron, siderophores, heme and vitamin B12. *Res in Microbiol*, 152(3–4), 291–301.
- Kumar, A, A., Shivachandra, S. B., Biswas, A.V., Singh, V.P., Srivastava, S. K. (2004). Prevalent serotypes of *Pasteurella multocida* isolated from different animal and avian species in India. *Vet Res Commun*, 28, 657–667.
- Lee, H., Gurtowski, J., and Yoo, S. (2014). Error correction and assembly complexity of single molecule sequencing reads. *bioRxiv*, 1–17. https://doi.org/10.1101/006395.

- Lee, M. D., Brown, J., Wooley, R. E., and Glisson, J. R. (1988). The relationship of pathogenicity to the growth of *Pasteurella multocida* serotypes 3,4 isolates in normal turkey plasma. *Avian Dis*, 32(3), 509–12.
- Leonard, J. T., Grace, M. B., Buzard, G. S., Mullen, M. J. and Barbagallo, C. B. (1998). Preparation of PCR products for DNA sequencing. *BioTech*, 24, 314–317.
- Litwin, C.M. and Calderwood, S.B. (1993). Role of iron in regulation of virulence genes. *Clin. Microbiol, Rev* (6), 137-149.
- Liu, L., Li, Y., Li, S., Hu, N., He, Y., Pong, R., ... Law, M. (2012). Comparison of next-generation sequencing systems. *J Biomed Biotech*, 2012. <https://doi.org/10.1155/2012/251364>.
- Liu, W., Yang, M., Xu, Z., Zheng, H., Liang, W., Zhou, R., ... Chen, H. (2012). Complete genome sequence of *Pasteurella multocida* HN06, a toxigenic strain of serogroup D. *Genome Announc* 194(12), 3292–3293. doi.org/10.1128/JB.00215-12.
- Liu, C.M., Aziz, M., Kachur, S., Hsueh P. R., Huang, Y. T., Keim, K., and Price L. B. (2012). Bact quant: An enhanced broad-coverage bacterial quantitative real-time PCR assay. *BMC Microbiol*, 12(56), 1-13.
- Livak, K. J., and Schmittgen, T. D. (2001). analysis of relative gene expression data using real-time quantitative PCR and the 2⁻ $\Delta\Delta$ CT method. *Methods*, 25(4), 402–408. doi.org/10.1006/meth.2001.1262.
- Lowe, T.M., Eddy, S.R. (1997). tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res*, 25, 955–964. doi.org/10.1093/nar/25.5.0955.
- Lukashin., A.V., and Borodovsky, M. (1998). GeneMark.hmm: new solutions for gene finding. *Nucleic Acids Res*. 26, 1107–1115.
- Mackiewicz, P., Zakrzewska-Czerwinska, J., Zawilak, A., Dudek, M. R., and Cebrat, S. (2004). Where does bacterial replication start? Rules for predicting the oriC region. *Nucleic Acids Res*, 32(13), 3781–3791. doi.org/10.1093/nar/gkh699.
- Marioni, J. C., Mason, C. E., Mane, S. M., Stephens, M., and Gilad, Y. (2008). Comparison with gene expression arrays RNA-seq: An assessment of technical reproducibility and comparison with gene expression arrays. *Genome Res*, 18(9) 1509–1517. doi.org/10.1101/gr.079558.108.
- Marshall, B., M. and Levy S., B. (2011). Food animals and antimicrobials: impacts on human health. *Clin. Microbiol. Rev*, 24(4), 718- 733. [doi 10.1128/CMR.00002-11](https://doi.org/10.1128/CMR.00002-11).
- May, B. J., Zhang, Q., Li, L. L., Paustian, M. L., Whittam, T. S., and Kapur, V. (2001). Complete genomic sequence of *Pasteurella multocida*, PM70. *Proc Natl*

Acad Sci, 98(6), 3460–3465doi.org/10.1073/pnas.051634598.

- Marandi, M., Mittal, K.R., (1997). Role of outer membrane protein H (OmpH) and OmpA-specific monoclonal antibodies from hybridoma tumors in protection of mice against *Pasteurella multocida*. *Infect Immun*, 65, 4502–4508.
- McArthur, A. G., Waglechner, N., Nizam, F., Yan, A., Azad, M. A., Baylay, A. J., ... Wright, G. D. (2013). The comprehensive antibiotic resistance database. *Antimicrob Agents Chemother*, 57(7), 3348–3357. doi.org/10.1128/AAC.00419-13.
- Michael, G. B., Kadlec, K., Sweeney, M. T., Brzuszkiewicz, E., Liesegang, H., Daniel, R., ... Schwarz, S. (2012). ICEPmu1, an integrative conjugative element (ICE) of *Pasteurella multocida*: Structure and transfer. *J Antimicrob Chemother*, 67(1), 91–100. doi.org/10.1093/jac/dkr411
- Miranda, C. D., Kehrenberg, C., Ulep, C., Schwarz, S., Roberts, M. C., and Al, M. E. T. (2003). Diversity of tetracycline resistance genes in bacteria from Chilean salmon farms. *Antimicrob Agents and Chemother*, 47(3), 883–888. doi.org/10.1128/AAC.47.3.883
- Moustafa, A. M., Seemann, T., Gladman, S., Adler, B., Harper, M., Boyce, J. D., and Bennett, M. D. (2015). Comparative genomic analysis of asian Hemorrhagic septicemia-associated strains of *Pasteurella multocida* identifies more than 90 hemorrhagic septicemia- specific genes. *PloS One*, 10(7)(1), 1–16. https://doi.org/10.1371/journal.pone.0130296.
- Miflin J.K. and Blackall P.J. (2001). Development of a 23S rRNA-based PCR assay for the identification of *Pasteurella multocida*. *Lett. Appl. Microbiol.*, 33, 216–221.
- Namioka, S. (1978). *Pasteurella multocida*. Biochemical characteristics and serotypes. In *Methods in Microbiology*, pp. 272-292. London, UK: Academic Press.
- Nielsen, J. P., Rosdahl, V. T. (1990). Development and epidemiological applications of a bacteriophage typing system for typing *Pasteurella multocida*. *J. Clin. Microbiol.* 28, 103-107.
- OIE, 2012. Chapter 2.4.12 Hemorrhagic Septicemia. In OIE Terrestrial Manual 2012. The World Assembly of delegates of the OIE, May 2012. <http://www.oie.int/fileadmin/Home/fr/>
- Opiyo, S.O., Pardy, R.L., Moriyama, H., Moriyama, E., N. (2010). Evolution of the Kdo2-lipid A biosynthesis in bacteria. *BMC Evol Biol*, 10,362. doi: 10.1186/1471-2148-10-362.
- Otomaru, K., Kubota, S., and Tokimori, M. (2013). Maternally and naturally acquired antibody to *Mannheimia haemolytica* in Japanese black calves. *J Vet Med Sci*, 4–6. https://doi.org/10.1292/jvms.13-0262.

- Paustian, M. L., May, B. J., and Kapur, V. (2002). Transcriptional response of *Pasteurella multocida* to nutrient limitation. *J Bacteriol*, *184*(13), 3734–3739. <https://doi.org/10.1128/JB.184.13.3734>.
- Peng, Z., Liang, W., Liu, W., Wu, B., Tang, B., Tan, C., ... Chen, H. (2016). Genomic characterization of *Pasteurella multocida* HB01, a serotype A bovine isolate from China. *Gene*, *581*(1), 85–93. <https://doi.org/10.1016/j.gene.2016.01.041>
- Petersen, A., Bisgaard, M., Townsend, K., Christensen, H. (2014). MLST typing of *Pasteurella multocida* associated with hemorrhagic septicemia and development of a real-time PCR specific for hemorrhagic septicemia associated isolates. *Vet Microbiol*, *170*(3-4), 335-41. doi: 10.1016/j.vetmic.2014.02.022.
- Planet, P., J. Kachlany, S., C. Fine, D., H. Desalle, R. and Figurski, D., H. (2003). The widespread colonization island (WCI) of *Actinobacillus actinomycetemcomitans*, *Nat. Genet*, *34*, 193-198.
- Praveena P. E., Periasamy, S., Kumar, A. A., and Singh, N. (2014). Pathology of experimental infection by *Pasteurella multocida* serotype A:1. *Vet Patho*, *51*(6), 1109-1112.
- Price, S.B., Freeman, M.D., Macewen, M.W. (1993). Molecular analysis of a cryptic plasmid isolated from avian. *Vet Microbiol*, *37*(1-2), 31-43.
- Price, N.M. and Morel, F.M.M. (1998). Biological cycling of iron in the ocean. In *Metal Ions in Biological Systems*, ed. Sigel, A. and Sigel, H. 35, pp.1-36. New York: Marcel Dekker.
- Pryor, R. J. and Wittwer, C.T. (2006). Real-time polymerase chain reaction and melting curve analysis. *Methods Mol Biol*, *336*, 19-32.
- Ponchel, F., Toomes, C., Bransfield, K., Leong, F. T., Douglas, S. H., Field, S. L., ... Markham, A. F. (2003). Real-time PCR based on SYBR-Green fluorescence: an alternative to the TaqMan assay for a relative quantification of gene rearrangements, gene amplifications and micro gene deletions. *BMC Biotechnol*, *3*(1), 18. doi.org/10.1186/1472-6750-3-18.
- Pullinger, G. D., Bevir, T., and Lax, A. J. (2004). The *Pasteurella multocida* toxin is encoded within a lysogenic bacteriophage. *Mol Microbiol*, *51*(1), 255–269. doi.org/10.1046/j.1365-2958.2003.03829.x.
- Quail, M. A., Smith, M., Coupland, P., Otto, T. D., Harris, S. R., Connor, T. R., ... Gu, Y. (2012). A tale of three next generation sequencing platforms: comparison of Ion Torrent, Pacific Biosciences and Illumina MiSeq sequencers. *BMC Genomics*, *13*(1), 341. doi.org/10.1186/1471-2164-13-341.
- Ratledge, C., and Dover, L. G. (2000). Iron Metabolism in Pathogenic Bacteria. *Annual Rev Microbiol*, *54*(1), 881–941 . doi.org/10.1146/annurev.micro.54.1.881.

- Ramdani and Adler B (1991). Oposonic monoclonal antibodies against lipopolysaccharide (LPS) antigens of *Pasteurella multocida* and the role of LPS in immunity. *Vet Microbiol*, 26, 335–347.
- Ryan, K.J., Ray, C.G. (2004). Sherris Medical Microbiology. 4th ed, 8529-85399. McGraw Hill.
- Rimler, R. B., and Wilson, M. A. (1994). Re-examination of *Pasteurella multocida* serotypes that caused hemorrhagic septicemia in North America. *Vet Rec*, 134(10), 256.
- Rimler, R., Richard, B.R. (2000). Restriction endonuclease analysis using HhaI and HpaII to discriminate among group B *Pasteurella multocida* associated with hemorrhagic septicemia. *J. Med. Microbio*, 49, 81-87.
- Roberts, M. C., Schwarz, S., and Aarts, H. J. M. (2012). Erratum : Acquired antibiotic resistance genes : an overview. *Front Microbiol*, 3(November), 1–17. doi.org/10.3389/fmicb.2012.00384.
- Ross, A. and Somssich, I., E. (2016). A DNA-based real-time PCR assay for robust growth quantification of the bacterial pathogen *Pseudomonas syringae* on *Arabidopsis thaliana*. *Plant Methods* 12(48).https://doi.org/10.1186/s13007-016-0149-z
- Rafidah, O., Zamri, S. M., Nasip, E., Shahiruddin, S. and Saharee, A.A. (2010). Analysis of hemorrhagic septicemia outbreaks in cattle and buffalo in Malaysia. *Online J Vet Res*, (14), 325-333.
- Rafidah, O., Zamri, S. M., Shahirudin, S., Nasip, E. (2012). Efficacy of intranasal vaccination of field buffaloes against haemorrhagic septicaemia with a live *gdhA* derivative *Pasteurella multocida* B:2. *Vet. Rec*, 171, pp. 175-178.
- Rashida, M., Farooq, U., Bader, N., Khanum, A., Wadood, A. (2006). Restriction endonuclease analysis of *Pasteurella multocida* field isolates by Hha-1. *Pak Vet J*, 26, 80-84.
- Raymaekers, M., Smets, R., Maes, B., and Cartuyvels R. (2009). Checklist for optimization and validation of real-time PCR Assays. *J Clin Lab Ana*, 23, 145–151.
- Reeves, G. A., Talavera, D. and Thornton, J. M. (2009). Genome and proteome annotation: organization, interpretation and integration. *J R Soc Interface*, 6(31), 129–147. doi.org/10.1098/rsif.2008.0341.
- Roehrig, S.C., Tran, H.Q., Spehr, V., Gunkel, N., Selzer, P.M., Ullrich, H.J. (2007). The response of *Mannheimia haemolytica* to iron limitation: implications for the acquisition of iron in the bovine lung. *Vet. Microbiol*, 121, 316–329. doi.org/10.1016/j.vetmic.
- Saleem, L., Munir, R., and Ferrari, G. (2014). Efficacy and cross-protectivity of live

intranasal aerosol hemorrhagic septicemia vaccine in buffalo calves, *Int J Curr Microbiol App Sci*, 3(11), 300–307.

- Sanchez, S., Mizan, S., Quist, C., Schroder, P., Juneau, M., Dawe, D., ... Lee, M. D. (2004). Serological response to *Pasteurella multocida* NanH sialidase in persistently colonized rabbits. *Clin Diag Lab Immunol*, 11(5), 825–34. doi.org/10.1128/CDLI.11.5.825-834.2004.
- Saranghi, L. N., Priyadarshini, A., Kumar, S., Thomas, P., Gupta, S. K., Nagaleekar, V. K., and Singh, V. P. (2014). Virulence genotyping of *Pasteurella multocida* isolated from multiple hosts from India. *Sci World J*, 2014, 814109–10. doi.org/10.1155/2014/814109.
- Saranghi, L. N., Thomas, P., Gupta, S. K., Kumar, S., Viswas, K. N., and Singh, V. P. (2016). Molecular epidemiology of *Pasteurella multocida* circulating in India by Multilocus sequence typing. *Trans and Emerg Dis*, 63(2), e286–e292. doi.org/10.1111/tbed.12270.
- Scharmann, W., Drzeniek, R., and Blobel, H. (1970). Neuraminidase of *Pasteurella multocida*. *Infect Immun*, 1(3), 319–20.
- Sellyei, B., Varga, Z., Szentesi-Samu, K., Kaszanyitzky, E., and Magyar, T. (2009). Antimicrobial susceptibility of *Pasteurella multocida* isolated from swine and poultry. *Acta Vet Hung*, 57(3), 357–367. doi.org/10.1556/AVet.57.2009.3.2.
- Shapiro, J. A., and von Sternberg, R. (2005). Why repetitive DNA is essential to genome function. *Biologic Rev*, 80(2), 227–250. doi.org/10.1017/S1464793104006657.
- Shivachandra, S. B., Viswas, K. N., and Kumar, A. A. (2011). A review of hemorrhagic septicemia in cattle and buffalo. *Anim Health Res Rev*, 12(1), 67–82. doi.org/10.1017/S146625231100003X.
- Singh, R., Blackall, P. J., Remington, B., Turni, C., Singh, R., Blackall, P. J., ... Turni, C. (2013). Studies on the presence and persistence of *Pasteurella multocida* serovars and genotypes in fowl cholera outbreaks. *Avian Pathol* 9457, 42(6), 581–585. doi.org/10.1080/03079457.2013.854861.
- Snipes, K.P., Ghazikhanian, G.Y., and Hirsh, D.C. (1987) Fate of *Pasteurella multocida* in the blood vascular system of turkeys following intravenous inoculation: comparison of an encapsulated, virulent strain with its avirulent, acapsular variant. *Avian Dis*, 31, 254–259.
- Song, H., Wang, P., Li, C., Han, S., Lopez-Baltazar, J., Zhang, X., and Wang, X. (2016). Identification of lipoxygenase (LOX) genes from legumes and their responses in wild type and cultivated peanut upon *Aspergillus flavus* infection. *Sci Rep*, 6 (September), 35245. doi.org/10.1038/srep35245.

- Sørum, H., Roberts, M. C., and Crosa, J. H. (1992). Identification and cloning of a tetracycline resistance gene from the fish pathogen *Vibrio salmonicida*. *Antimicrob Agents Chemother*, 36(3), 611–5.
- Subaaharan, S., Blackall, L. L., and Blackall, P. J. (2010). Development of a multi-locus sequence typing scheme for avian isolates of *Pasteurella multocida*, *Vet Microbiol*, 141, 354–361. doi.org/10.1016/j.vetmic.2010.01.017.
- Tang, X., Zhao, Z., Hu, J., Wu, B., Cai, X., He, Q., and Chen, H. (2009). Isolation, antimicrobial resistance, and virulence genes of *Pasteurella multocida* strains from swine in China *J Clin Microbiol* 47(4), 951–958. http://doi.org/10.1128/JCM.02029-08.
- Tajadini, M., Panjehpour, M., and Javanmard, S. (2014). Comparison of SYBR green and TaqMan methods in quantitative real-time polymerase chain reaction analysis of four adenosine receptor subtypes. *Adv Biomed Res*, 3(1), 85. doi.org/10.4103/2277-9175.127998.
- Tabatabaei, M. Z., Liu, A., Finucane, R., Parton, Coote, J. G. (2002). Protective immunity conferred by attenuated *aroA* derivatives of *Pasteurella multocida* B:2 strains in a mouse model of hemorrhagic septicemia *Infect Immun*, (70), 3355–3362.
- Tsai, I. J., Otto, T. D. and Berriman, M. (2010). Improving draft assemblies by iterative mapping and assembly of short reads to eliminate gaps. *Genome Biol*, 11. doi: 10.1186/gb-2010-11-4-r41.
- Tatusov, R. L., Galperin, M. Y., Natale, D. A., and Koonin, E. V. (2000). The COG database: a tool for genome-scale analysis of protein functions and evolution. *Nucleic Acids Res*, 28(1), 33–36. doi.org/10.1093/nar/28.1.33.
- Taylor, S. K., Ward, A. C. S., Hunter, D. L., Gunther, K., and Kortge, L. (1996). isolation of *Pasteurella* spp. from free-ranging american bison (bison bison). *J Wildlife Dis*, 32(2), 322–325. https://doi.org/10.7589/0090-3558-32.2.322.
- Tettelin H., Riley, D., Cattuto, C. and Medini, D. (2008). Comparative genomics: the bacterial pan genome. *Curr Opin Microbiol*, 11, 472–477.
- Timp, Winston, U. M. M. (2001). Nonspecific adherence and fibril biogenesis by *Actinobacillus actinomycetemcomitans*: TadA protein is an ATPase. *J Bacteriol*, 183(20), 5927–36. doi.org/10.1128/JB.183.20.5927-5936.2001.
- Timp, W., Mirsaidov, U. M., Wang, D., Comer, J., Aksimentiev, A., and Timp, G. (2010). Nanopore sequencing: Electrical measurements of the code of life. *IEEE Trans Nanotechno*, 9(3), 281–294 doi.org/10.1109/TNANO.2010.2044418.
- Tomich, M., Planet, P. J., and Figurski, D. H. (2007). The tad locus: postcards from the widespread colonization island. *Nat Rev Microbiol*, 5(5), 363–375. https://doi.org/10.1038/nrmicro1636.

- Touati, D. (2000). Iron and oxidative stress in bacteria. *Arch Biochem and Biophys*, 373(1), 1–6. doi.org/10.1006/abbi.1999.1518.
- Townsend, K. M., Frost, A. J., Lee, C. W., John, M., Dawkins, H. J. S., and Papadimitriou, J. M. (1998). Development of PCR assays for species- and type-specific identification of *Pasteurella multocida* isolates development of PCR assays for species- and type-specific identification of *Pasteurella multocida* isolates. *J Clin Microbiol*, 36(4), 1096–1100.
- UniProt Consortium. UniProt. (2017). The universal protein knowledgebase. *Nucleic Acids Res*, 45, 158-169. doi.org/10.1093/nar/gkw1099.
- Verma, R., and Jaiswal, T. N. (1998). Hemorrhagic septicemia vaccines. *Vaccine*, 16(11–12), 1184–92.
- Verma, S., Sharma, M., Katoch, S., Verma, L., Kumar, S., Dogra, V., ... Singh, G. (2013). Profiling of virulence associated genes of *Pasteurella multocida* isolated from cattle. *Vet Res Comm*, 37(1), 83–89. doi.org/10.1007/s11259-012-9539-5.
- Wang, D. (2011). Nanopore sequencing: Electrical measurements of the code of life. *IEEE Trans Nanotechnol*, 9(3), 281–294 doi.org/10.1109/TNANO.2010.2044418.Nanopore.
- Wang, Z., Gerstein, M., and Snyder, M. (2010). RNA-Seq: a revolutionary tool for transcriptomics. *Nat Rev Genet*, 10(1), 57–63. doi.org/10.1038/nrg2484.RNA-Seq.
- Welsh, R. D., Dye, L. B., Payton, M. E., and Confer, A. W. (2004). Isolation and antimicrobial susceptibilities of bacterial pathogens from bovine pneumonia: 1994–2002. *J. Vet Diag Invest*, 16(5), 426–431. doi.org/10.1177/104063870401600510.
- Wilson, B. A., and Ho, M. (2013). *Pasteurella multocida*: from zoonosis to cellular microbiology. *Clin Microbiol Rev*, 26(3), 631–55. doi.org/10.1128/CMR.00024-13.
- White, D. G., Zhao, S., Simjee, S., Wagner, D. D., and McDermott, P., F. (2002). Antimicrobial resistance of foodborne pathogens. *Microbes Infect*, 4, 405-412.
- Wilkie, I.W., Harper, M., Boyce, J.D., Adler, B. (2012). *Pasteurella multocida*: diseases and pathogenesis. *Curr. Top Microbiol Immunol*, 361, 1–22.
- Winnenburg, R., Baldwin, T. K., Urban, M., Rawlings, C., Kohler, J., Hammond-Kosack, K. E., ... Hammond-Kosack, K. E. (2006). PHI-base: a new database for pathogen host interactions. *Nucleic Acids Res*, 34, D459-64. doi.org/10.1093/nar/gkj047.
- Wijewardana, T. G. (1992). Hemorrhagic septicemia. *Rev. Med. Microbiol*, 3, 59–63.

- Wijewardana, T.G., Wilson, C.F., Gilmour, N.J. and Poxton I.R. (1990). Production of mouse monoclonal antibodies to *Pasteurella multocida* type A and the immunological properties of a protective anti-lipopolysaccharide antibody. *J Med Microbiol*, 33, 217–222.
- Wong, M. L., and Medrano, J. F. (2005). Real-time PCR for mRNA quantitation.pdf. *BioTech*, 39(1), 75–85. doi.org/10.2144/05391RV01.
- Wootton, M. (2013). BSAC Version 12 May 2013, 44(May).
- Woodford, N. (2004). Public databases: Retrieving and manipulating sequences for beginners. *Meth in Mol Biol*. 17-28.
- Xie, H., Wasserman, A., Levine, Z., Novik, A., Grebinskiy, V., Shoshan, A., and Mintz, L. (2002). Large-scale protein annotation through Gene Ontology. *Genome Res*, 12(5), 785–794. doi.org/10.1101/gr.86902.
- Xu, Z., Yue, M., Zhou, R., Jin, Q., Fan, Y., Bei, W., and Chen, H. (2011). Genomic characterization of *Haemophilus parasuis* SH0165, a highly virulent strain of serovar 5 prevalent in China. *Plos One*, 6(5), e19631. doi.org/10.1371/journal.pone.0019631.
- Xu, Z., Zhou, Y., Li, L., Zhou, R., Xiao, S., Wan, Y., ... Chen, H. (2008). Genome biology of *Actinobacillus pleuropneumoniae* JL03, an isolate of serotype 3 prevalent in China. *PLOS ONE*, 3(1), e1450.
- Yap, H.Y., Ghazali, K., Wan-Mohamad Nazarie, W.F., Mat Isa, M.N., Zakaria, Z., Omar, A.R. (2013). Draft genome sequence of *Pasteurella multocida* sub spp *P. multocida* strain PMTB, isolated from a buffalo. *Genome Announc*, 1(5), e00872-13. doi:10.1128/genomeA.00872-13.
- Yasmin A.R., Yeap, S.K., Hair-Bejo, M., Omar, A.R. (2016). Characterization of chicken splenic-derived dendritic cells following vaccine and very virulent strains of infectious bursal disease virus infection. *Avian Dis*, 60(4), 739-751.
- Yanni, S., Jeremy B. (2007). Designing patterns for profile HMM search. *Bioinformatics*, 23(2), e36-e43. doi: 10.1093/bioinformatics/bt1323.
- Yeo, B. K., and Mokhtar, I. (1993). Hemorrhagic septicemia of buffalo in Sabah, Malaysia. *Pasteurellosis in Production Animals*, 43, 112–115.
- Zamri-Saad, M., and Annas, S. (2015). Vaccination against hemorrhagic septicemia of bovines: A Review. *Pak Vet J*, 36, 1–5. doi.org/10.1097/QCO.0b013e3283