



**UNIVERSITI PUTRA MALAYSIA**

***IDENTIFICATION OF *Trichoderma harzianum* T3.13  
AND CHARACTERIZATION OF ITS INTERACTION WITH *Neoscytalidium  
dimidiatum* U1, A PATHOGENIC FUNGUS ISOLATED FROM DRAGON  
FRUIT (*Hylocereus polyrhizus* (F.A.C. Weber) Britton & Rose)***

**WAN RUSMARINI BINTI WAN ZULKIFELI**

**FBSB 2016 34**



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By

**WAN RUSMARINI BINTI WAN ZULKIFELI**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirement for the degree of Master of Science**

**July 2016**

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Abstract of this thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

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By

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**July 2016**

**Chairman: Associate Professor Umi Kalsom Md Shah, PhD  
Faculty: Biotechnology and Biomolecular Sciences**

Endophytes are potential biological control agents. They produce enzymes which facilitate their initial colonisation of plant tissues and direct interactions with microbial pathogens. *Trichoderma* species have been broadly used in the effort to defeat soil-borne pathogens. The objectives of this study were to identify isolated endophytic fungi and pathogen of dragon fruit, and measure enzyme activity of chitinase from *Trichoderma harzianum* T3.13 grown in different types of medium. This work was also done to profile genes encoding chitinase,  $\beta$ -glucanase and N-acetylglucosamine from *Trichoderma harzianum* T3.13 against pathogen *Neoscytalidium dimidiatum* U1 and control pathogen *Colletotrichum gloeosporioides* by using reverse transcription-polymerase chain reaction (RT-PCR). In this study, endophytic fungi from the stem of healthy dragon fruit (*Hylocereus* spp.) was successfully identified as *Trichoderma harzianum* T3.13. *T. harzianum* T3.13 was shown to have the ability to produce antifungal activity against *N. dimidiatum* U1, a pathogen fungus from the stem of unhealthy dragon fruit. Mycoparasitic interactions between the two fungi were observed by scanning electron microscopy. *T. harzianum* T3.13 hyphae tangled with the hyphae of *N. dimidiatum* U1. The chitinolytic activities of *T. harzianum* T3.13 were 0.194 U/ml and 0.1 U/ml in a medium containing 3% (w/v) of colloidal chitin and 0.5 % (w/v) dried cell wall of *N. dimidiatum* U1 as sole carbon source, respectively. This study profiled the expression of genes encoding chitinase (*chit42*),  $\beta$ -glucanase (*bgn13.1*) and N-acetylglucosamine (*exc1*) from *T. harzianum* T3.13. Semi-quantitative RT-PCR was used to quantify the expression patterns of the genes during the interaction of *T. harzianum* T3.13 with pathogen *N. dimidiatum* U1 and control pathogen *C. gloeosporioides*, respectively. The expression patterns of these genes were profiled before and after the interactions occurred. The expression of the *exc1* and *chit42* genes were observed to be present before and after the interaction occurred in the presence of *N. dimidiatum* U1. However, the expression of the *bgn13.1* gene increased after 24 hours up to 96 hours of interaction

in the presence of *N. dimidiatum* U1. In the presence of *C. gloeosporioides*, the expression of *bgn13.1* and *chit42* gradually decreased during the interaction although the expression of the *exc1* gene did not change. The results suggested that the endophytic fungus *T. harzianum* T3.13 has the potential as a good biological control agent against *N. dimidiatum* U1 and *C. gloeosporioides*. Thus, the study provided an insight into cellular and molecular interactions between *T. harzianum* T3.13 and pathogenic fungi.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**Pengenalan *Trichoderma harzianum* T3.13 dan Pencirian Interaksinya dengan *Neoscytalidium dimidiatum* U1, Kulit Patogenik Diasingkan Daripada Dragon Fruit (*Hylocereus polyrhizus* (F.A.C. Weber) Britton & Rose)**

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Endofitik berpotensi sebagai ejen kawalan biologi. Ia menghasilkan enzim yang memudahkan pengkolonian awal mereka terhadap tisu tumbuhan dan berinteraksi secara langsung dengan mikroba patogen. Spesies *Trichoderma* telah digunakan secara meluas dalam usaha untuk menghapuskan patogen tanah. Objektif kajian ini adalah untuk mengenalpasti kulit endofitik dan patogen pada buah naga yang telah diasingkan dan mengenalpasti aktiviti enzim kitinase dari *Trichoderma harzianum* T3.13 yang hidup dalam media yang berbeza. Kajian ini juga dijalankan untuk memprofil gen kitin,  $\beta$ -glukan dan N-acetylglukosamine dari *Trichoderma harzianum* T3.13 apabila bertindakbalas dengan patogen *Neoscytalidium dimidiatum* U1 dan patogen kawalan *Colletotrichum gloeosporioides* dengan menggunakan tindakbalas rantaian polimerase-transkripsi berbalik. Dalam kajian ini, kulit endofitik dari batang pokok buah naga (*Hylocereus* spp) yang sihat telah berjaya dikenal pasti sebagai *Trichoderma harzianum* T3.13. *T. harzianum* T3.13 telah terbukti mempunyai keupayaan untuk menghasilkan aktiviti antikulat terhadap *N. dimidiatum* U1, iaitu kulit patogen dari batang pokok buah naga yang tidak sihat. Interaksi mikoparasitik antara kedua-dua kulit diperhatikan dengan menggunakan pengimbas mikroskop elektron. Hifa *T. harzianum* T3.13 didapati berpaut dengan hifa *N. dimidiatum*. Aktiviti kitinolitik *T. harzianum* T3.13 adalah 0.194 U/ml dan 0.1 U/ml, masing-masing dalam medium yang mengandungi 3% (w/v) kitin koloid dan 0.5% (w/v) dinding sel kering daripada *N. dimidiatum* sebagai sumber karbon tunggal. Kajian ini memprofilkan ekspresi gen kitin (*chit42*),  $\beta$ -glukan (*bgn13.1*) dan N-acetylglukosamine (*exc1*) daripada *T. harzianum* T3.13. Semi-kuantitatif RT-PCR digunakan untuk mengukur ekspresi dari gen yang berbeza apabila *T. harzianum* T3.13 berinteraksi dengan patogen *N. dimidiatum* U1 dan patogen kawalan *C. gloeosporioides*. Ekspresi gen ini telah diprofil sebelum dan selepas interaksi berlaku. Ekspresi gen *exc1* dan *chit42* wujud sebelum dan selepas interaksi berlaku dalam kehadiran *N. dimidiatum* U1. Namun, ekspresi bagi gen *bgn13.1* mula meningkat selepas

24 jam interaksi sehingga 96 jam interaksi dalam kehadiran *N. dimidiatum* U1. Dalam kehadiran *C. gloeosporioides*, ekspresi gen *bgn13.1* dan *chit42* berkurang secara beransur-ansur semasa interaksi tetapi ekspresi gen *excl* tidak berubah. Hasil kajian ini mencadangkan bahawa endofitik kulat *T. harzianum* T3.13 mempunyai potensi untuk menjadi agen kawalan biologi terhadap patogen *N. dimidiatum* U1 dan *C. gloeosporioides*. Oleh itu, kajian ini memberi pengetahuan tentang interaksi selular dan molekular antara *T. harzianum* T3.13 dan kulat patogenik.



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Finally, the greatest accomplishment is not in never falling, but in rising again after you fall. Fall forward.

*Barokallahufikum.*

Thank you.



I certify that a Thesis Examination Committee has met on 26 July 2016 to conduct the final examination of Wan Rusmarini binti Wan Zulkifeli on her thesis entitled "Identification of *Trichoderma harzianum* T3.13 and Characterization of its Interaction with *Neoscytalidium dimidiatum* U1, a Pathogenic Fungus Isolated from Dragon Fruit (*Hylocereus polyrhizus* (F.A.C.Weber) Britton & Rose)" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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## LIST OF ABBREVIATIONS

%	Percentage
µg	Microgram
BLAST	Basic Local Alignment Search Tool
BLASTN	Nucleotide-nucleotide BLAST
bp	base pair
BSA	Bovine serum albumin
cDNA	complementary deoxyribonucleic acid
DEPC	Diethyl pyrocarbonate
dH <sub>2</sub> O	distilled water
DNA	deoxyribonucleic acid
DNS	3,5-dinitrosalicylic acid
dNTPs	Deoxynucleotides
E value	Expect value
EtBr	ethidium bromide
g	gram
g	gravity
HCl	hydrochloric acid
Kg	kilogram
M	molarity
mL	Milliliter
mM	Millimolar
mRNA	messenger ribonucleic acid
MW	molecular weight
NAG	N-acetyl-D-glucosamine
NaOH	sodium hydroxide
NCBI	National Center for Biotechnology Information
ng	Nanogram
nM	Nanomolar
OD	optical density
PCI	Phenol: chloroform: isoamylalcohol
PCR	polymerase chain reaction
PDA	Potato dextrose agar
RNA	ribonucleic acid
RNase	Ribonuclease
rpm	Rotations per minute
rRNA	ribosomal RNA
RT-PCR	Reverse transcription polymerase chain reaction
SEM	scanning electron microscopy
T <sub>a</sub>	annealing temperature
TAE	Tris acetate EDTA
T <sub>m</sub>	melting temperature
U	Unit
U/mL	Unit per mL
UV	ultraviolet
V	voltage
v/v	Volume per volume
w/v	weight per volume

## CHAPTER 1

### INTRODUCTION

Dragon fruit is a cactus fruit crop that's have a high demand at national and international levels. This fruit has been planted in over 10 countries worldwide (Valencia-Botín *et al.*, 2013). Since dragon fruit starts to be farmed in commercial farms, some symptoms of rotting and spots in stems and fruits were being identified (Valencia-Botín *et al.*, 2013). Studies on stem soft rot diseases were begun at 1990s in Mexico because of the diseases caused by two types of Enterobacteria. While fungi such as *Botryosphaeria dothidea* Ces. & de Not also identified as causing agent of the spots on stems diseases (Valencia-Botín *et al.*, 2013). One more fungal disease become violent as it affects fruits and stems is anthracnose disease. Researcher in United State of America (USA) and Japan revealed their report in etiological study found that the causing agents of anthracnose disease are fungus *Colletotrichum gloesporioides* Penz (Poonpolgul and Kumphai, 2007). In Peninsular Malaysia, red-fleshed dragon fruit (*Hylocereus polyrhizus*) is very popular and widely cultivated (Masyahit *et al.*, 2009). However, anthracnose symptoms start to be detected on the stems of dragon fruit in the states of Kedah and Penang in August 2010 and January 2011, respectively (Iskandar Vijaya *et al.*, 2015).

The anthracnose disease was found in commercial plantations in the Malaysia. Prevention could be initiated for controlling plant disease through earlier detection and accurate identification of plant pathogens. It is important to prevent the introduction and distribution of new pathogens in a growing area where has not present yet. Therefore, the availability of fast, sensitive and accurate methods for detection and identification of fungal pathogens is required to improve disease management measures (Capote *et al.*, 2012).

Day now, the old-style agricultural practices have many problems such as disease, drought, and low soil fertility as a result of use of pesticides, harmful chemical, pest, pollution and global warming (Pratibha, 2011). Therefore, there is a requirement for eco-friendly biological control agents that can help in undertaking the above problems. There are many types of bacteria and fungi that's involves in biological control activities. Today, *Trichoderma* was identified has a high potential in controlling the plant diseases. *Trichoderma* species can produce different kinds of enzymes which stand as an important role in biological control activities, like cell wall degradation, hypha growth, biotic and abiotic stress tolerance and antagonistic activity against plant pathogens (Hasan, 2014). They can colonize aboveground and belowground plant organs or grow between living cells (endophytes). They can live in soil organic matter as saprophytes and could be appear in plant litter and mammalian tissues (Rai and Mehra, 2015).

*Trichoderma* species are known for their assembly of cell wall degrading enzymes such as chitinase. Chitinase enzyme has been established massive attention because of their potential use in biological control for phytopathogenic fungus and other phytopathogenic organisms that containing chitin such as insects (Souza *et al.*, 2003). In these respect, the fungi that's produce chitinase enzyme has intensively studied as biological control agents.

Genes play a major role in the biological control process by regulating signals and leads the secretion of some enzymes that help in the degradation of the pathogens. Increased in the expression of the genes such as chitinase and glucanase gene, helps in enhanced the biological control activity, promoting the plant growth and prevents the plant from pathogen attack (Massart and Jijakli, 2007). Therefore, gene such as chitinase gene can be cloned and produced in a large amount for commercial applications (Pratibha, 2011).

In the preliminary study, the endophytic *Trichoderma* from healthy stem dragon fruit and pathogen *Neoscytalidium* from unhealthy stem dragon fruit are isolated. They are shows an antagonistic activities between each other. Therefore this study has been done to initiate the identification for the genus level for *Trichoderma* and *Neoscytalidium*. The production of chitinase enzyme by *Trichoderma harzianum* T3.13 has been evaluated too in order to obtain substantial chitinase enzyme activities. Meanwhile, the profile expressions of *T. harzianum* T3.13 genes such as chitinase gene,  $\beta$ -glucanase gene and N-acetylglucosamine gene are also profiled during antagonistic activity against pathogens *Neoscytalidium dimidiatum* U1 and *Colletotrichum gloeosporioides*.

**The objectives of this study are:**

1. To identify isolated endophytic fungi and pathogen of dragon fruit plants.
2. To measure the enzyme activity of chitinase from *Trichoderma harzianum* T3.13 grown in different types of medium
3. To profile genes encoding chitinase,  $\beta$ -glucanase and N-acetylglucosamine from *Trichoderma harzianum* T3.13 against pathogen *Neoscytalidium dimidiatum* U1 and control pathogen *Colletotrichum gloeosporioides* by using reverse transcription-polymerase chain reaction (RT-PCR).

## REFERENCES

- Adams, D. J. (2004). Fungal cell wall chitinases and glucanases. *Microbiology* 150: 2029–2035.
- Ali, Asgar (2013). “Effectiveness of Submicron Chitosan Dispersions in Controlling Anthracnose and Maintaining Quality of Dragon Fruit.” *Postharvest Biology and Technology* 86: 147–53.
- Anees, M., A. Tronsmo, V. Edel-Hermann, L.G. Hjeljord, C. Heraud and C. Steinberg (2010). Characterization of field isolates of *Trichoderma* antagonistic against *Rhizoctonia solani*. *Fungal Biology* 114: 691-701.
- Aida, Wan (2010). Determination of Pitaya Seeds as a Natural Antioxidant and Source of Essential Fatty Acids 1010: 1003–9.
- Ali, A., Mahmud, T.M.M., Sijam, K., Siddiqui, Y. (2010). Potential of chitosan coating in delaying the postharvest anthracnose (*Colletotrichum gloeosporioides* Penz.) of Eksotika II papaya. *International Journal of Food Science and Technology* 45: 2134–2140.
- Ariffin, A. A. , Bakar, J. , Tan, C. P. , Rahman, R. A. , Karim, R., and Loi, C. C. (2009). Essential fatty acids of pitaya (dragon fruit) seed oil. *Food Chemistry* 114(2): 561-564.
- Azevedo, J., Maccheroni Jr., W., Pereira, J., and de Araújo, W. (2000). Endophytic microorganisms: a review on insect control and recent advances on tropical plants. *Electronic Journal of Biotechnology* 3:(1).
- Arnold, A.E., Mejía, L.C., Kylo, D., Rojas, E.I., Maynard, Z., Robbins, N. and Herre, E.A (2003). Fungal endophytes limit pathogen damage in a tropical tree. *Proc. Natl. Acad. Sci.* 100: 15649–15654
- Azevedo, J.L. (1998). *Microorganismos endofíticos*. In: Ecologia Microbiana. Melo, I.S. and Azevedo, J.L. (eds.). Editora EMBRAPA, Jaguariuna, São Paulo, Brazil. 117-137.
- Ali A., Mahmud T.M.M., Sijam K. and Siddiqui Y. (2011) Effect of chitosan coatings on the physiochemical characteristics of Eksotika II papaya (*Carica papaya* L.) fruit during cold storage. *Food Chemistry* 124: 620–626
- Andrews, K.L., J.W. Bentley, and R.D. Cave. (1992). Enhancing biological control's contributions to integrated pest management through appropriate levels of farmer participation. *Florida Entomologist* 75: 429-439.
- Agrios GN. (2005). *Plant Pathology*, 5th edn. San Diego, CA, USA, Academic press
- Boonratkwang, C., Chamswang, C., Intanoo, W. and Juntharasri, V. (2007). Effect of Secondary Metabolites from *Trichoderma harzianum* Strain Pm9 on Growth Inhibition of *Colletotrichum Gloeosporioides* and Chilli Anthracnose Control. Proceeding of the 8th National Plant Protection Conference. Naresuan University, Phisanulok, Thailand, 323-336.
- Bhattacharya, D., Nagpure, A. and Gupta, R.K. (2007) Bacterial chitinases: properties and potential. *Crit Rev Biotechnol* 27:(1):21–28.
- Bowman, Shaun M. and Stephen J. Free. (2006). “The Structure and Synthesis of the Fungal Cell Wall.” *BioEssays* 28: 799–808.
- Bailey B.A., Stream M.D. and Wood, D. (2009). *Trichoderma* species form endophytic associations within *Theobroma cacao* trichomes, *Mycol. Res.* 113: 1365–1376
- Bailey, B.A, Bae H, Strem M.D., Crozier J., Thomas S.E, Samuels G.J, Vinyard B.T and Holmes K.A. (2008). Antibiosis, mycoparasitism and colonization success for

- endo-phytic *Trichoderma* isolates with biocontrol potential in *Theobroma cacao*. *Biological Control* 46:24–35
- Baek, J. M., Howell, C. R. and Kenerley, C. M. (1999). The role of an extracellular chitinase from *Trichoderma virens* Gv29-8 in the biocontrol of *Rhizoctonia solani*. *Curr Genet* 35: 41–50.
- Carsolio, C., Benhamou, N., Haran, S., Cortés, C., Gutiérrez, A., Chet, I. and Herrera-Estrella, A. (1999). Role of the *Trichoderma harzianum* endochitinase gene, *ech42*, in mycoparasitism. *Appl Environ Microbiol* 65: 929–935.
- Carsolio, C., Gutierrez, A. and Jimenez, B., (1994). Characterization of *ech-42*, a *Trichoderma harzianum* endochitinase gene expressed during mycoparasitism. *Proceedings of National Academy of Science*, 9; (23): 10903–10907
- Cargnello M. and Roux, P.P. (2011). Activation and function of the MAPKs and their substrates, the MAPK-activated protein kinases. *Microbiol Mol Biol Rev* 75 (1): 50–83.
- Cao, Ronghua. (2009). “Mycoparasitism of Endophytic Fungi Isolated from Reed on Soilborne Phytopathogenic Fungi and Production of Cell Wall-Degrading Enzymes in Vitro.” *Current microbiology* 59(6): 584–92.
- Cheah, L.S. and W.M. Zulkarnain, (2008). *Status of pitaya cultivation in Malaysia*. Paper presented at Seminar on Pitaya Production, Market and Export Challenges and Prospects, Oct. 20, Putrajaya, Malaysia
- Chet, I. (1987). *Trichoderma*-application, mode of action, and potential as a biocontrol agent of soil borne plant pathogenic fungi. Wiley & Sons, New York, N.Y
- Crous, PW, Slippers B, Wingfield MJ, Rheeder J, Marasas WFO, Philips AJL, Alves A, Burgess T, Barber P. and Groenewald JZ. (2006). Phylogenetic lineages in the Botryosphaeriaceae. *Stud Mycol* 55:235–253.
- Chovanec, P., Kaliňák, M., Liptaj, T., Pronayová, N., Jakubík, T., Hudecová L' and Varečka, D. (2005). Study of *Trichoderma viride* metabolism under conditions of the restriction of oxidative processes. *Can J Microbiology* 51:853–862
- Castle, A., D. Speranzini, N. Rghei, G. Alm, D. Rinker and J. Bisset (1998). Morphological and molecular identification of *Trichoderma* isolates on North American mushroom farms. *Applied Environ. Microbiol.*, 64: 133-137.
- Cook, R.J. (1993) Making greater use of introduced microorganisms for biological control of plant pathogens. *Annu Rev Phytopathol* 31: 53-80
- Cohen-Kupiec, R., Broglie, K.E., Friesem, D., Broglie, R.M. and Chet, I., (1999). Molecular characterization of a novel beta-1, 3-exoglucanase related to mycoparasitism of *Trichoderma harzianum*. *Gene* 226: 147–154.
- Chuang, M.F, Ni HF, Yang HR, Hsu SL, Lai SY and Jiang YL. (2012) First report of stem canker disease of pitaya (*Hylocereus undatus*, *H. polyrhizus*) caused by *Neoscytalidium dimidiatum* in Taiwan. *Plant Dis* 96:906.
- Carsolio, C., Benhamou, N., Haran, S., Cortés, C., Gutiérrez, A., Chet, I. and Herrera-Estrella, A. (1999). Role of the *Trichoderma harzianum* endochitinase Gene *ech2* in Mycoparasitism. *Applied and Environmental Microbiology* 65:929-935.
- Chuang, M.F., Ni, H.F., Yang HR, Hsu SL, Lai SY, Jiang YL. (2012) First report of stem canker disease of pitaya (*Hylocereus undatus*, *H. polyrhizus*) caused by *Neoscytalidium dimidiatum* in Taiwan. *Plant Dis* 96:906.
- Calistru, C., McLean, M. and Berjak, P. (1997). In vitro studies on the potential for biological control of *Aspergillus flavus* and *Fusarium moniliforme* by *Trichoderma* species: a study of the production of extracellular metabolites by *Trichoderma* species. *Mycopathologia* 137:115-124



- Capote, N., Aguado, A., Pastrana, A. M., and Sánchez-Torres, P. (2012). Molecular tools for detection of plant pathogenic fungi and fungicide resistance (pp. 151-202). INTECH Open Access Publisher.
- Dennis Kunkel, Microscopy. (2007). Soil fungus conidiophore and conidia (*Trichoderma* sp.) Copyright: Created on September 9th, 2007. Last Modified on November 4th, 2009
- Deane, E.E., Whipps, J.M., Lynch, J.M. and Peberdy, J.F. (1998) The purification and characterization of a *Trichoderma harzianum* exochitinase. *Biochim Biophys Acta* 1383:101–110
- Dodd, J. C., Estrada, A., and Jeger, M. (1992). Epidemiology of *C. gloeosporioides* in the tropics. In J. A. Bailey and M. J. Jeger (Eds.), *Colletotrichum: Biology, pathology and control* (pp. 308–325). Wallingford: CAB International
- Dragon, White, and Fruit *Hylocereus*. (2012). “Phytochemical Screening And Analysis Polyphenolic Antioxidant Activity Of Methanolic Extract Of.” 23(1): 60–64.
- Dennis, C. and J. Webster, (1971). Antagonistic properties of species-groups of *Trichoderma*. Hyphal interaction. *Trans. Br. Mycol. Soc.* 57:363-369.
- Dahiya, N., Tewari R. and Hoondal GS. (2006) Biotechnological aspects of chitinolytic enzymes: a review. *Appl Microbiol Biotechnol.*71(6):773-82.
- Du, M., Schardl, C.L., Nucle, E.M. and Vaillancourt, L.J. (2005). Using mating-type gene sequences for improved phylogenetic resolution of *Colletotrichum* species complexes. *Mycologia* 97: 641e658.
- Delpin, M.W. and Goodman A.E. (2009). Nitrogen regulates chitinase gene expression in a marine bacterium. *ISME* 3:1064–9
- Druzhinina, I.S., Seidl-Seiboth V., Herrera-Estrella A., Horwitz B.A., Kenerley C. M., Monte E., Mukherjee P. K., Zeilinger S., Grigoriev I.V. and Kubicek C.P., (2011) *Trichoderma* the genomics of opportunistic success, *Nat Rev Microbiol* 9:749–759
- Druzhinina, I.S., Kopchinskiy A.G., Komon M., Bissett J., Szakacs G. and Kubicek C.P. (2005). An oligonucleotide barcode for species identification in *Trichoderma* and *Hypocrea*. *Fungal Genet Biol* 42:813–828.
- Djonovic, S., Pozo M.J. and Kenerley C.M. (2006) *Tvbgn3*, a beta-1,6- glucanase from the biocontrol fungus *Trichoderma virens*, is involved in mycoparasitism and control of *Pythium ultimum*. *Appl. Environ. Microbiol.*, 72:7661–7670
- De La Cruz, J., Rey, M., Lora, J.M., Hidalgo-Gallego, A., Dominguez, F., Pintor-Toro, J.A., Llobell, A. and Benitez, T. (1992). Isolation and characterization of three chitinases from *Trichoderma harzianum*. *Eur J Biochem*; 206: 859– 867
- De la Cruz, J., Pintor-Toro, J.A., Benítez, T., Llobell, A. and Roero, L.A., (1995). Novel endo-b-1,3-glucanase, *BGN13.1*, involved in the mycoparasitism of *Trichoderma harzianum*. *Journal of Bacteriology* 177: 6937–6945
- de Capdeville, G., Wilson, C.L., Beer, S.V. and Aist, J.R. (2002). Alternative disease control agents induce resistance to blue mold in harvested 'red delicious' apple fruit. *Phytopathology* 92: 900-908.
- de Jonge, R. and Thomma, B. P. (2009). Fungal LysM effectors: extinguishers of host immunity. *Trends Microbiol* 17:151–157
- Das, S.N., Neeraja, Ch., Sarma, P.V.S.R.N., Madhu Prakash J, Purushotham P, Kaur M, Dutta S. and Podile A.R. (2012). *Microbial chitinases for chitin waste management*. In: Microorganisms in Environmental Management. *Springer*, Netherlands, p 135–150
- da Silva Aires, R., Steindorff A.S., Ramada M.H.S., de Siqueira S.J.L. and Ulhoa C.J. (2012). Biochemical characterization of a 27 kDa 1,3-β-D-glucanase from *Trichoderma*

- asperellum* induced by cell wall of *Rhizoctonia solani*. *Carbohydr. Polym.* 8(7): 1219–1223
- El-Katatny, MH., Somitsch, W., Robra, KH., El-Katatny, MS., Gübitz, GM. (2000). Production of Chitinase and  $\beta$ -1,3 glucanase by *Trichoderma harzianum* for Control of the Phytopathogenic Fungus *Sclerotium rolfsii*. *Food Technol Biotechnology* 38:173–180
- Edward, A., Evans, Jordan and Huntley. (2011). Economics of establishing and producing pitaya in southern florida: A stochastic budget analysis. *Horticulture Technology* 21:(2) 246-251
- Eijsink, V. G., Vaaje-Kolstad, G., Varum, K. M. and Horn, S. J. (2008). Towards new enzymes for biofuels: lessons from chitinase research. *Trends Biotechnol* 26: 228–235.
- Evans, HC., Holmes, KA. and Thomas, SE. (2003). Endophytes and mycoparasites associated with an indigenous forest tree, *Theobroma gileri*, in Ecuador and a preliminary assessment of their potential as biocontrol agents of cocoa diseases. *Mycol Prog* 2:149–160,
- Ebrahim, S., Usha, K. and Singh, B. (2011). Pathogenesis-related (PR)-proteins: chitinase and  $\beta$ -1,3- glucanase in defense mechanism against malformation in mango (*Mangifera indica L.*). *Sci Hortic* 130:847–52
- Edgar, RC. ( 2004a). MUSCLE: a multiple sequence alignment method with reduced time and space complexity. *BMC Bioinformatics* 5:113.
- Edgar, RC. (2004b). MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.* 32:1792–1797
- Felsenstein, J. (1985). Confidence limits on phylogenies: An approach using the bootstrap. *Evolution* 39:783-791.
- Fialho, Mauricio Batista., Luiz Fernando, and Romanholo Ferreira. (2011). “Biocontrol Science and Technology Antimicrobial Volatile Organic Compounds Affect Morphogenesis- Related Enzymes in Guignardia Citricarpa , Causal Agent of Citrus Black Spot.” (May 2012): 37–41.
- Faramarzi, MA., Fazeli, M., Yazdi, MT., Adrangi, S., Al Ahmadi, KJ. and Tasharrofi, N. (2009). Optimization of cultural conditions for production of chitinase by a soil isolate of *Massilia timonae*. *Biotechnology* 8:93–9
- Freeman, S., Minz, D., Maymon, M. and Zveibil, A. (2001). Genetic diversity within *Colletotrichum acutatum* sensu Simmonds. *Phytopathology* 91(6):586-592.
- Freeman, S., Katan, T. and Shabi, E. (1998). Characterization of *Colletotrichum* species responsible for anthracnose diseases of various fruits. *Plant Disease* 82(6):596-605.
- Guo, Li We. (2014). “First Report of Dragon Fruit ( *Hylocereus Undatus* ) Anthracnose Caused by *Colletotrichum Truncatum* in China.” *Journal of Phytopathology* 162: 272–75.
- Gruber, S. and Zeilinger S. (2014). The Transcription Factor Ste12 Mediates the Regulatory Role of the Tmk1 MAP Kinase in Mycoparasitism and Vegetative Hyphal Fusion in the Filamentous Fungus *Trichoderma atroviride*. *PLoS ONE* 9(10): e111636.
- Gruber, S. and Seidl-Seiboth, V. (2012). Self versus non-self: fungal cell wall degradation in *Trichoderma*. *Microbiology* 158(1):26-34.
- Gruber, S., Vaaje-Kolstad G, Matarese F, Lopez-Mondejar R, Kubicek CP. and Seidl-Seiboth, V. (2011). Analysis of subgroup C of fungal chitinases containing chitin-binding and LysM modules in themycoparasite *Trichoderma atroviride*. *Glycobiology* 21:122–33



- Gams, W. and Bissett, J. (1998). *Morphology and identification of Trichoderma*. In: Harmann GE, Kubicek CP (eds) *Trichoderma and Gliocladium*. Taylor and Francis, London, pp 3–34
- Gomez Ramirez, M., L. I. Rojas Avelizapa, N. G. Rojas Avelizapa, and R. Cruz Camarillo. (2004). Colloidal chitin stained with Remazol Brilliant Blue R, a useful substrate to select chitinolytic microorganisms and to evaluate chitinases. *Journal of Microbiological Methods* 56:213-219.
- Gokul, B., Lee, J.H., Song, K.B., Rhee, S.K., Kim, C.H., Panda, T. (2000). Characterization and applications of chitinases from *Trichoderma harzianum* – a review. *Bioprocess Engineering* 23: 691–694
- Guerard, F., Sumaya-Martinez, MT., Laroque, D., Chabeaud, A. and Dufosse, L. (2007). Optimization of free radical scavenging activity by response surface methodology in the hydrolysis of shrimp processing discards. *Process Biochem* 42:1486–1491
- Gildberg, A. and Stenberg, EA. (2001). Utilisation of male Arctic capelin and Atlantic cod intestines for fish sauce production evaluation of fermentation conditions. *Process Biochem.*, 36(8-9):809-812.
- Ganley, RJ., Sniezk, RA. and Newcombe, G. (2008). Endophyte-mediated resistance against white pine blister rust in *Pinus monticola*. *Forest Ecology and Management* 255: 2751e2760
- Gams, W. and Bissett, J., (2002). Morphology and identification of *Trichoderma*. In: Kubicek, C. P. and Harman, G. E. (eds.). *Trichoderma and Gliocladium: Basic biology, taxonomy and genetics*. Taylor & Francis Ltd, pp. 3-31.
- Gooday, GW. (1999). Aggressive and defensive roles of chitinases. *Exs* 87:157–169
- Gazis, R. and Chaverri, P. (2010). *Diversity of fungal endophytes in leaves and stems of rubber trees (Hevea brasiliensis) in Tambopata, Peru*. Fungal Ecol (In press).
- Gaskell, GJ., Carter, DA., Britton, WJ., Tovey, ER., Benyon, FHL. and Lovborg, Y. (1997). Analysis of the internal transcribed spacer regions of ribosomal DNA in common airborne allergenic fungi. *Electrophoresis* 18: 1567–1569.
- Henrissat, B. (1991). A classification of glycosyl hydrolases based on amino acid sequence similarities. *Biochem. J* 280: 309–316
- Henrissat, B. (1999). *Classification of chitinase modules*. In: Jolles P, Muzzarelli RAA, eds. *Chitin and Chitinases*, Burkhauser Basel Switzerland.: 137–156
- Heydari, A., Fattahi, H., Zamanizadeh, HR., Hassanzadeh, N. and Naraghi, L. (2004). Investigation on the possibility of using bacterial antagonists for biological control of cotton seedling damping-off in green house. *Appl Entomol Phytopathol* 72: 51-68
- Heydari, A. and Misaghi, IJ. (2003). The role of rhizosphere bacteria in herbicide mediated increase in *Rhizoctonia solani* induced cotton seedling damping off. *Plant Soil* 257: 391-396.
- Howell, C. R. (2006). Understanding the mechanisms employed by *Trichoderma virens* to effect biological control of cotton diseases. *Phytopathology* 96:178–180.
- Henry, T., Iwen, PC. and Hinrichs, SH. (2000). Identification of *Aspergillus* species using internal transcribed spacer regions 1 and 2. *J Clin Microbiol*; 38: 1510–1515.
- Harman, GE. and Kubicek, CP. (1998). *Trichoderma and Gliocladium*, vol. 2: enzymes, biological control and commercial application. London: Taylor and Francis; pp. 131–151
- Hamid, R., Khan, M. A., Ahmad, M., Ahmad, M. M., Abdin, M. Z., Musarrat, J., and Javed, S. (2013). Chitinases: An update. *Journal of Pharmacy & Bioallied Sciences*, 5(1): 21–29.

- Hassan, MM. (2014). Influence of protoplast fusion between two *Trichoderma* spp. on extracellular enzymes production and antagonistic activity. *Biotechnol. Equip.* 28:1014-1023.
- Howell, CR. (2003). Mechanisms employed by *Trichoderma* species in the biological control of plant diseases: the history and evolution of current concepts. *Plant Dis.* 87:4-10.
- Haran, S., Schickler, H., Oppencheim, A., and I. Chet, (1996). Differential expression of *Trichoderma harzianum* chitinases during mycoparasitism. *Phytopathology*, 86: 980-985
- Hjeljord, L. and Tronsmo, A. (1998). *Trichoderma* and *Gliocladium* in biological control: an overview. In: Harman, G., Kubicek, C.P. (Eds.), *Trichoderma and Gliocladium*. Taylor and Francis Inc., London, pp. 185–204
- Hille, A., Neu, TR., Hempel, DC., Horn, H. (2009). Effective diffusivities and mass fluxes in fungal biopellets. *Biotechnol Bioeng.* 103:1202–13.
- Iranzo, M., Aguado, C., Pallotti, C., Canizares, JT. and Mormeneo, S. (2002). The use of trypsin to solubilize wall proteins from *Candida albicans* led to the identification of chitinase 2 as an enzyme covalently linked to the yeast wall structure. *Res Microbiol* 153: 227–232
- Ilias, G.N.M. ( 1999). *Trichoderma* and its efficacy as a bio-control agent of basal stem rot of oil palms, PhD, University of Putra, Malaysia.
- Isaac, S. (1992). *Fungal plant interactions*. London: Chapman and Hall
- Isabelle Mouyna, Lukas Hartl and Jean-Paul Latgé. (2013). Beta-1,3-glucan modifying enzymes in *Aspergillus fumigatus*. *Front. Microbiol.* 1664-302
- Ihrmark, K., Asmail, N., Ubhayasekera, W., Melin, P., Stenlid, J., & Karlsson, M. (2010). Comparative Molecular Evolution of *Trichoderma* Chitinases in Response to Mycoparasitic Interactions. *Evolutionary Bioinformatics Online* 6: 1–26.
- Intanoo, W. and Chamswarn, C. (2007). *Effect of Antagonistic Bacterial Formulations for Control of Anthracnose on Chilli Fruits*. Proceeding of the 8th National Plant Protection Conference. Naresuan University, Phisanulok, Thailand, p.309-322.
- International Seed Testing Association. (1996). International rules for seed testing. *Seed Sci. Technol.* 4: 3-49.
- Islam, MT., Hashidoko, Y., Deora, A., Ito, T. and Tahara, S. (2005). Suppression of damping off disease in host plants by the rhizoplane bacterium *Lysobacter* sp. strain SB-K88 is linked to plant colonization and antibiosis against soilborne Peronosporomycetes. *Appl Environ Microbiol* 71: 3786-3796.
- Iskandar Vijaya, S., Mohd Anuar, I. S. and Zakaria, L. (2015). Characterization and Pathogenicity of *Colletotrichum truncatum* Causing Stem Anthracnose of Red-Fleshed Dragon Fruit (*Hylocereus polyrhizus*) in Malaysia. *J. Phytopathol* 163: 67–71.
- Jaques, AK., Fukamizo, T., Hall, D., Barton, RC., Escott, GM., Parkinson, T., Hitchcock CA. and Adams, DJ. (2003). Disruption of the gene encoding the *ChiB1* chitinase of *Aspergillus fumigatus* and characterization of a recombinant gene product. *Microbiology* 149: 2931–2939
- Jaafar, Ruzainah Ali, Ahmad Ridhwan Bin Abdul Rahman, Nor Zaini Che Mahmud, and R. Vasudevan. (2009). “Proximate Analysis of Dragon Fruit (*Hylecereus Polyhizus*).” *American Journal of Applied Sciences* 6(7): 1341–46.
- Janisiewicz, WJ. and Peterson, DL. (2004). Susceptibility of the stem pull area of mechanically harvested apples to blue mold decay and its control with a biocontrol agent. *Plant Dis* 88: 662-666.

- Jach, G., Gornhardt, B., Mundy, J., Logemann, J., Pinsdorf, E., Leah, R., Schell, J. and Mass, C. (1995). Enhanced quantitative resistance against fungal disease by combinatorial expression of different barley antifungal proteins in transgenic tobacco. *Plant J* 8: 97-109.
- Jones, T., Federspiel, NA., Chibana, H., Dungan, J., Kalman, S., Magee BB, Newport G, Thorstenson YR, Agabian N, Magee PT, Davis RW. and Scherer, S. ( 2004). The diploid genome sequence of *Candida albicans*. *Proc. Natl Acad. Sci. USA* 101: 7329–7334.
- Junker, B. (2006). Measurement of bubble and pellet size distributions: past and current image analysis technology. *Bioprocess Biosyst Eng*.29:185–206.
- Junqueira, N.T.V. and Gasparotto, L. (1991). Controle biológico de fungos estromáticos causadores de doenças foliares em *Seringueira*, pp. 307-331. In: Anon., Controle Biológico de Doen. de Plantas. Org. W. Bettiol., Brasília, Embrapa
- Kuranda, MJ. and Robbins, PW. (1991). Chitinase is required for cell separation during growth of *Saccharomyces cerevisiae*. *J Biol Chem* 266: 19758–19767
- Kendrick, A. and Ratledge, C . (1992). Lipids of selected molds grown for production of n-3 and n-6 polyunsaturated fatty acids. *Lipids* 27:15-20
- Knogge, W. and Scheel, D. (2006). LysM receptors recognize friend and foe. *Proc Natl Acad Sci USA* 103(29):10829–10830.
- Khokhar, M. K. , Gupta, R. and Sharma, R. (2012). Biological Control of Plant Pathogens using Biotechnological Aspects:- *A Review*, 1(5): 1-6.
- Kraulis, J., Clore, GM., Nilges, M., Jones, TA., Pettersson, G., Knowles, J. and Gronenborn, AM. (1989). Determination of the three-dimensional solution structure of the C-terminal domain of cellobiohydrolase I from *Trichoderma reesei*. A study using nuclear magnetic resonance and hybrid distance geometry-dynamical simulated annealing. *Bio-chemistry* 28: 7241–7257.
- Kubicek, C.P., Herrera, E.A., Seidl, S.V. and Martinez, D.A. (2011). Comparative genome sequence analysis underscores mycoparasitism as the ancestral life style of *Trichoderma*. *Genome Biol*.12: 40-63
- Kasprzewska, A. (2003). Plant chitinases-regulation and function. *Cell Mol Biol Lett* 8(3):809–824
- Khalili, R., M.A., A.H. Norhayati, M.Y. Rokiah, R. Asmah, M.T. Mohd Nasir and M. Siti Muskinah. (2006). Proximate composition and selected mineral determination in organically grown red pitaya (*Hylocereus* sp.). *J. Trop. Agric. Food Sci.*, 34: 269-275.
- Kadokawa, Jun-ichi. (2015). “Fabrication of Nanostructured and Microstructured Chitin Materials through Gelation with Suitable Dispersion Media.” *RSC Adv.* 5(17): 12736–46.
- Karthik, Narayanan, Karthik Akanksha, Parameswaran Binod, and Ashok Pandey. (2014). “Production, Purification and Properties of Fungal Chitinases- *A Review*.” 52(November): 1025–35.
- Kloepper, JW., Leong, J., Teintze, M. and Schroth, MN. (1980) *Pseudomonas* siderophores: A mechanism explaining disease suppression in soils. *Current Microbiology* 4: 317-320
- Keen, NT. and Yoshikawa, M. (1983) b-1,3-endoglucanase from soybean releases elicitor-active carbohydrates from fungus cell wall. *Plant Physiol* 71:460–5.
- Karlsson, M. and Stenlid, J. (2008). Comparative evolutionary histories of the fungal chitinase gene family reveal non-random size expansions and contractions due to adaptive natural selection. *Evol Bioinformatics* 4:47–60

- Kulling, C., Mach, R. L., Lorito, M. and Kubicek, C. P. (2000). Enzyme diffusion from *T. atroviride* (*T. harzianum* PI) to *R. solani* is a prerequisite for triggering of *Trichoderma ech42* gene expression before mycoparasitic contact. *Applied and Environmental Microbiology*. 66: 2232-2234
- Kawachi, I., Fujieda, T., Ujita, M., Ishii, Y., Yamagishi, K. and Sato, H. (2001). Purification and properties of extracellular chitinases from the parasitic Fungus *Isaria japonica*. *Journal of Bioscience and Bioengineering* 92: 554–549.
- Kumar, PT., Srinivasan, S., Lakshmanan, VK., Tamura, H., Nair, SV. and Jayakumar, R (2011). Synthesis, characterization and cytocompatibility studies of alpha-chitin hydrogel/nano hydroxyapatite composite scaffolds. *Int J Biol Macromol* 49:20–31
- López-Cervantes, J., Adan-Bante, N.P. and Sánchez-Machado, D.I. (2010). Separation and biochemical characterization of the products from fermented shrimp wastes. In: products as real material: New ways of application. Trivandrum (India), Transworld Research Network. p. 117-132. *Waste Management.* : 135–50.
- Lugones, L.G., de Jong, J.F., de Vries, O.M., Jalving, R., Dijksterhuis, J. and Wosten, H.A. (2004). The SC15 protein of *Schizophyllum commune* mediates formation of aerial hyphae and attachment in the absence of the SC3 hydrophobin. *Mol. Microbiol.* 53: 707–716
- Lenné, J.M. and Parbery, D.G. (1976). Phyllosphere antagonists and appressoria formation in *Colletotrichum gloeosporioides*. *Transactions of the British Mycological Society*, 66:334-336.
- Le Bellec, F., F. Vaillant and E. Imbert (2006). Pitahaya (*Hylocereus* spp.): A new fruit crop, a market with a future. *Fruits* 61: 237-250.
- Liou, M. R., Hung, C. L., and Liou, R. F. (2001). First Report of Cactus virus X on *Hylocereus undatus* (Cactaceae) in Taiwan. *Plant Disease* 85(2):229-229
- Lorito, M., Woo., S.L., García Fernández, I., Colucci, G., Harman, G.E., Pintor-Toro, J.A., Filippone, E., Muccifora, S., Lawrence, C.B., Zoina, A., Tuzun, S. and Scala, F. (1998). Genes from mycoparasitic fungi as a source for improving plant resistance to fungal pathogens. *Proceedings of the National Academy of Sciences, USA* 95:7860-7865.
- Loza Cornejo, S. and Terrazas, T. (2003). Epidermal and hypodermal characteristics in North American Cactoideae (Cactaceae). *J Plant Res* 116:27–35
- Lim, S. D. (2011). “Effect of Extraction Parameters on the Yield of Betacyanins from Pitaya Fruit (*Hylocereus Polyrhizus*) Pulp.” *Journal of Food, Agriculture and Environment* 9: 158–62.
- Lazarovits, G., and Nowak, J. (1997). Rhizobacteria for improvement of plant growth and establishment. *Hortscience* 32:188-192.
- Linnakoski, R. , Puhakka-tarvainen, H. and Pappinen, A.(2012). Endophytic fungi isolated from Khaya anthotheca in Ghana. *Fungal Ecology*, 5(3): 298-308.
- López-Mondéjar, R., Ros, M. and Pascual, J.A.(2011). Mycoparasitism-related genes expression of *Trichoderma harzianum* isolates to evaluate their efficacy as biological control agent. *Biol. Control* 56: 59–66.
- Lopez-Mondejar, R., Catalano, V., Kubicek, CP. and Seidl, V. (2009). The beta-N-acetylglucosaminidases NAG1 and NAG2 are essential for growth of *Trichoderma atroviride* on chitin. *FEBS Journal* 276:5137–5148.
- Lorenz, T. C. (2012). Polymerase Chain Reaction: Basic Protocol Plus Troubleshooting and Optimization Strategies. *J. Vis. Exp.* (63)
- Lau, M. K., Arnold, A. E., and Johnson, N. C. (2013). Factors influencing communities of foliar fungal endophytes in riparian woody plants. *Fungal Ecology*, 6(5): 365-378



- Limo´n, M. C., Chaco´n, M. R., Meji´as, R., Delgado-Jarana, J., Rinco´n, A. M., Codo´n, A. C. and Beni´tez, T. (2004). Increased antifungal and chitinase specific activities of *Trichoderma harzianum* CECT 2413 by addition of a cellulose binding domain. *Appl Microbiol Biotechnol* 64: 675–685.
- Linnakoski, R., Puhakka-Tarvainen, H. and Pappinen, A. (2012). Endophytic fungi isolated from *Khaya anthotheca* in Ghana. *Fungal Ecol.* 5: 298-308.
- Masyahit, M., Sijam, K., Awang, Y. and Ghazali, M. (2013). The Occurrence of Anthracnose Disease Caused By *Colletotrichum Gloeosporioides* on Dragon Fruit (*Hylocereus* Spp.) In Peninsular Malaysia. *Acta Hortic.* (Ishs) 975:187-195
- Mohd Adzim Khalili, R. , Che Abdullah, A. B. , and Abdul Manaf, A.(2012). Total antioxidant activity, total phenolic content and radical scavenging activity both flesh and peel of red pitaya, white pitaya and papaya. *International Journal of Pharmacy and Pharmaceutical Sciences*, 4(2): 113-122.
- Mohd Adzim Khalili, R. , Che Abdullah, A. B. , and Abdul Manaf, A. (2014). Isolation and characterization of oligosaccharides composition in organically grown red pitaya, white pitaya and papaya. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(SUPPL. 2): 131-136. IJPPS.
- Madrid, H., Ruíz-Cendoya, M., Cano, J., Stchigel, A., Orofino, R. and Guarro, J. (2009). Genotyping and in vitro antifungal susceptibility of *Neoscytalidium dimidiatum* isolates from different origins. *International Journal of Antimicrobial Agents* 34:351- 354.
- Maheshwari, R., Bharadwaj, G. and Bhat, MK. (2000). Thermophilic fungi: their physiology and enzymes. *Microbiol. Mol. Biol. Rev.* 64: 461-488.
- Mizrahi, Y. and Nerd, A. (1999). *Climbing and columnar cacti: new arid land fruit crops*. In: Janick J (ed) *Perspectives on New Crops and New Uses*. Alexandria, VA, USA, ASHS Press, pp 358–366.
- Mukherjee, M., Mukherjee, PK. and Kale, SP. (2007). cAMP signalling is involved in growth, germination, mycoparasitism and secondary metabolism in *Trichoderma virens*. *Microbiology* 153:1734–1742
- Mukherjee, M. , Mukherjee, P. K. , Horwitz, B. A. , Zachow, C. , Berg, G. and Zeilinger, S.(2012). *Trichoderma*-Plant-Pathogen Interactions: Advances in Genetics of Biological Control. *Indian Journal of Microbiology*.
- Marone, M., Mozzetti, S., De Ritis, D., Pierelli, L. and Scambia, G.(2001). Semiquantitative RT-PCR analysis to assess the expression levels of multiple transcripts from the same sample. *Biological Procedures Online* 3: 19–25.
- Masyahit, M., Kamaruzaman, S., Yahya, A. and Mohd Ghazali, MS. (2009). The first report of the occurrence of anthracnose disease caused by *Colletotrichum gloeosporioides* (Penz.) Penz. & Sacc. on dragon fruit (*Hylocereus* spp.) in Peninsular Malaysia. *Am J Appl Sci* 6:902–912
- Manchester, K.L. (1996). Use of UV methods for measurement of protein and nucleic acid concentrations. *Biotechniques* 20; pp. 968–970
- Mercado, J. a, Barceló, M., Pliego, C., Rey, M., Caballero, J. L., Muñoz-Blanco, J., and Ruano-Rosa, D., (2015). Expression of the  $\beta$ -1,3-glucanase gene *bgn13.1* from *Trichoderma harzianum* in strawberry increases tolerance to crown rot diseases but interferes with plant growth. *Transgenic Research*. 24(6):979-89
- María, Cecilia Gortari and Roque, Alberto Hours (2013). Biotechnological processes for chitin recovery out of crustacean waste: A mini-review. *Electronic Journal of Biotechnology*.Vol. 16 No. 3

- Massart, S. and Jijakli, HM. (2007). Use of molecular techniques to elucidate the mechanisms of action of biocontrol agents: A review. *J. Microbiol. Methods* 69: 229-241.
- Moretto, K.C.K., Gimenes-Fernandes, N. and Dos Santos, J.M. (2001). Influence of *Trichoderma* spp. on *Colletotrichum acutatum* mycelial growth and morphology on infection of 'Tahiti' lime detached flowers. *Summa Phytopathologia*, 27(4):357-364
- Mohd, Masratul Hawa., Baharuddin Salleh, and Latiffah Zakaria. (2013). "Identification and Molecular Characterizations of *Neosectalidium Dimidiatum* Causing Stem Canker of Red Fleshed Dragon Fruit (*Hylocereus Polyrhizus*) in Malaysia." : 841–49.
- McIntyre, M., Nielsen, J. and Arnau, J. (2004). Proceedings of the 7th European Conference on Fungal Genetics. Copenhagen, Denmark
- Milgroom, MG. and Cortesi, P. (2004). Biological control of chestnut blight with hypovirulence: a critical analysis. *Annu Rev Phytopathol* 42: 311-338.
- Marcello, CM., Steindorff, AS., Silva, SP., Silva, RN. and Bataus, Lam (2010). Expression analysis of the exo- $\beta$ -1,3-glucanase from the mycoparasitic fungus *Trichoderma asperellum*. *Microbiol. Res.* 165: 75-81
- Martin, K., McDougall, BM., McIlroy, S., Jiezhong, Chen J. and Seviour, RJ. (2007). Biochemistry and molecular biology of exocellular fungal  $\beta$ -(1,3)- and  $\beta$ -(1,6)-glucanases. *FEMS Microbiol Review* 31(2):168–92
- Nerd, Avinoam, Yaron Sitrit, Ram Avtar Kaushik, and Yosef Mizrahi. 92002). "High Summer Temperatures Inhibit Flowering in Vine Pitaya Crops (*Hylocereus* Spp.)." *Scientia Horticulturae* 96: 343–50.
- Nawani, NN. and Kapadnis, BP. (2003). Chitin degrading potential of bacteria from extreme and moderate environment. *Indian J. Exp. Biol.* 41: 248-254.
- Noishiki, Y., Takami, H., Nishiyama, Y., Wada, M., Okada, S., and Kuga, S. (2003). Alkali induced conversion of  $\alpha$ -chitin to  $\beta$ -chitin. *Biomacromolecules*, 4(4): 896–899
- Nadarajah, K., Ali, H. Z., and Omar, N. S. (2014). The isolation and characterization of an endochitinase gene from a Malaysian isolate of, 8(5): 711-721.
- Ng, L. C., Ngadin, A., Azhari, M., and Zahari, N. A. (2015). Potential of *Trichoderma* spp. as Biological Control Agents Against Bakanae Pathogen (*Fusarium fujikuroi*) in Rice. *Asian Journal of Plant Pathology*, 9(2)
- Omann, MR., Lehner, S., Escobar Rodriguez, C., Brunner, K., Zeilinger, S. (2012). The seven-transmembrane receptor *Gpr1* governs processes relevant for the antagonistic interaction of *Trichoderma atroviride* with its host. *Microbiology* 158:107–118
- Ordentlich, A., Elad, Y. and Chet, I. (1988). The role of chitinase of *Serratia marcescens* in the biocontrol of *Sclerotium rolfsii*. *Phytopathology* 78:84-88.
- Palmateer, A. J., Ploetz R. C., van Santen E., and Correll, J. C. (2007). First Occurrence of Anthracnose Caused by *Colletotrichum gloeosporioides* on Pitahaya Plant Disease. *91(5)*: 631-631
- Priscila Chaverri, Fabiano Branco-Rocha, Walter Michael Jaklitsch, Romina Orietta Gazis, Thomas Degenkolb, and Gary J. Samuels (2015). Systematics of the *Trichoderma harzianum* species complex and the reidentification of commercial biocontrol strains. *Mycologia*:14-147.
- Photita, W., Taylor, P.W.J., Ford, R., Lumyong, P., McKenzie, H.C. and Hyde, K.D., (2005). Morphological and molecular characterization of *Colletotrichum* species from herbaceous plants in Thailand. *Fungal Diversity*, 18:117-133. Prameela, K.,

- Murali Mohan, C., Smitha, P.V. and Hemalatha, K.P.J. (2010). Bioremediation of shrimp biowaste by using natural probiotic for chitin and carotenoid production an alternative method to hazardous chemical method. *International Journal of Applied Biology and Pharmaceutical Technology*, vol. 1, no. 3, p. 903-910.
- Pacheco, N., Garnica-González, M., Gimeno, M., Bárzana, E., Trombotto, S., David, L. and Shirai, K. (2011). Structural characterization of chitin and chitosan obtained by biological and chemical methods. *Biomacromolecules*, vol. 12, no. 9, p. 3285-3290.
- Pavlic, D., Wingfield, M.J., Barber, P., Slippers, B., Harder, G.S. and Burgess, T.I. (2008) Seven new species of the Botryosphaeriaceae from baobab and other native trees in Western Australia. *Mycologia* 100:851–866.
- Polizzi, G., Aiello, D., Vitale, A., Giuffrida, F., Groenewald, Z. and Crous, P.W. (2009) First report of shoot blight, canker, and gummosis caused by *Neoscytalidium dimidiatum* on citrus in Italy. *Plant Dis* 93:1215
- Prasun K. Mukherjee, Benjamin A. Horwitz and Charles M. Kenerley. (2012). Secondary metabolism in *Trichoderma* – a genomic perspective *Microbiology*.158: 35-45
- Parmar, H. J. , Bodar, N. P. , Lakhani, H. N. , Patel, S. V. , Umrana, V. V. , and Hassan, M. M. (2015). Production of lytic enzymes by *Trichoderma* strains during in vitro antagonism with *Sclerotium rolfsii*, the causal agent of stem rot of groundnut. *African Journal of Microbiology Research*, 9: 365-372.
- Pathan, A.K., Bond, J. and Gaskin, R.E. (2010). Sample preparation for SEM of plant surfaces. *Materials Today* 12, *Supplement 1*(0):32–43.
- Penz, Penz. (2009). “The First Report of the Occurrence of Anthracnose Disease Caused by Department of Plant Protection, Faculty of Agriculture, Department of Crop Science, Faculty of Agriculture 6(5): 902–12.
- Polizzi, G, Aiello, D., Castello, I. and Vitale, A. (2011). “Occurrence, Molecular Characterisation, and Pathogenicity of *Neoscytalidium Dimidiatum* on Citrus in Italy.” : 237–43
- Prasanna, R., Gupta, V., Natarajan, C. and Chaudhary, V. ( 2010). Bioprospecting for genes involved in the production of chitosanases and microcystin-like compounds in the production of chitosanase and microcystin-like compounds in *Anabaena* strains. *World J Microbiol Biotechnol* 26: 717–24
- Preferences, Consumer. (2011). “Production and Marketing Reports.” 21(April).
- Paulitz, T.C. and Bélanger, R.R. (2001). Biological control in greenhouse systems. *Annu Rev Phytopathol* 39: 103-133.
- Papagianni, M. (2004). Fungal morphology and metabolite production in submerged mycelial processes. *Biotechnology Advances* 22: 189–259
- Pillay, V. J., and Nowak, J. (1997). Inoculum density, temperature and genotype effects on in vitro growth promotion and epiphytic and endophytic colonization of tomato (*Lycopersicon esculentum* L.) seedlings inoculated with a pseudomonad bacterium. *Can. J. Microbiol.* 43:354-361.
- Pratibha, Sharma. (2011). Biocontrol genes from *Trichoderma* species: A review. *African Journal of Biotechnology*, 10(86): 19898-19907.
- Polizzi, G., Aiello, D., Vitale, A., Giuffrida, F., Groenewald, Z. and Crous, P.W. (2009) First report of shoot blight, canker, and gummosis caused by *Neoscytalidium dimidiatum* on citrus in Italy. *Plant Dis* 93:1215.
- Poonpolgul, S. and Kumphai, S. (2007). *Chilli Pepper Anthracnose in Thailand*. Country Report. In: Oh DG, Kim KT, editors. Abstracts of the First International Symposium on Chilli Anthracnose. Republic of Korea: National Horticultural Research Institute, Rural Development of Administration. p. 23

- Palumbo, J. D., Yuen, G. Y., Jochum, C. C., Tatum, K., and Kobayashi, D. Y. (2005). Mutagenesis of beta-1,3-glucanase genes in *Lysobacter enzymogenes* strain C3 results in reduced biological control activity toward *Bipolaris* leaf spot of tall fescue and *Pythium* damping-off of sugar beet. *Phytopathology* 95: 701-707
- Rosenberger, RF. (1976). The Cell Wall. In: Smith JE, Berry DR (eds) *The filamentous fungi*, vol 2. Arnold, London, pp 328–344
- Rhodes, RG., Atoyan, JA. and Nelson DR. (2010). The chitobiose transporter, *chbC*, is required for chitin utilization in *Borrelia burgdorferi*. *BMC Microbiol*;10:21.
- Roiger, DJ., Jeffers, SN. and Caldewell, RW.( 1991). Occurrence of *Trichoderma* species in apple orchard and woodland soils. *Soil Biol Biochem* 23: 353-359,
- Reino, J.L., Guerrero R.F., Hernandez-Galan R. and Collado, I.G. (2008) Secondary metabolites from species of the biocontrol agent *Trichoderma*, *Phytochem. Rev.*, 7, 89– 123
- Raveh, E., J. Weiss, A. Nerd and Y. Mizrahi, (1993). Pitayas (genus *Hylocereus*): A New Fruit Crop for the Negev Desert of Israel, Janick, J. and J.E. Simon (Eds.). New Crops, New York, pp: 491-495
- Ruiz-Herrera, J. (1992). *Fungal Cell Wall. Structure, Synthesis and Assembly*. CRC Press, Boca Raton, FL
- Ruiz-Herrera, J. and Ruiz-Medrano, R. (2004) Chitin biosynthesis in fungi. Handbook of Fungal Biotechnology, 2nd edn (Arora PK, ed), pp. 315–330. *Marcel Dekker*, New York
- Reithner, B, Ibarra-Laclette E, Mach RL, Herrera-Estrella A. (2011). Identification of mycoparasitism-related genes in *Trichoderma atroviride*. *Appl. Environ. Microbiol.* 77:4361–4370
- Rojas-Avelizapa, L. I., Cruz-Camarillo, R., Guerrero, M. I., Rodriguez-Vazquez, R., and Ibarra, J. E. (1999). Selection and characterization of a proteo-chitinolytic strain of *Bacillus thuringiensis*, able to grow in shrimp waste media. *World Journal of Microbiology & Biotechnology*, 15(2): 261– 268
- Rai, Dinesh and Mehra, Prateeksha (2015). A review on interactions of *Trichoderma* with Plant and Pathogens. *Res. J. Agriculture & Forestry Sci.*, 3(2): 20-23.
- Seidl, V., Huemer, B., Seiboth, B. and Kubicek, CP. (2005). A complete survey of *Trichoderma chitinases* reveals three distinct subgroups of family 18 chitinases. *FEBS J*; 272: 5923–5939
- Sangeetha, J., and Thangadurai, D. (2013). *Staining Techniques and Biochemical Methods for the Identification of Fungi*. In V. K. Gupta, M. G. Touhy, M. Ayyachamy, K. M. Turner, and A. O'Donovan, Laboratory Protocols in Fungal Biology (pp. 237-257). Springer New York.
- Sandhya, C., Adapa, LK., Nampoothiri, KM., Binod, P., Szakacs, G. and Padney, A.(2004). Extracellular chitinase production by *Trichoderma harzianum* in submerged fermentation. *J. Basic Microbiol.*;44:49–58.
- Schirmböck, M., Lorito, M., Wang, Y. L., Hayes, C. K., Arisan-Atac, I., Scala, F. and Kubicek, C. P. (1994). Parallel formation and synergism of hydrolytic enzymes and peptaibol antibiotics, molecular mechanisms involved in the antagonistic action of *Trichoderma harzianum* against phytopathogenic fungi. *Applied and Environmental Microbiology*, 60(12): 4364–4370.
- Schmoll, M. (2008). The information highways of a biotechnological workhorse signal transduction in *Hypocrea jecorina*. *BMC Genomics* 9:430
- Sharon, E., Chet, I. and Spiegel, Y. (2011). *Trichoderma* as biological control agent. In: Davies K, Spiegel Y (eds) Biological control of plant parasitic nematodes: building



- coherence between microbial ecology and molecular mechanisms. Springer, Berlin, pp 183–202
- Seidl, V. (2008). Chitinases of filamentous fungi: a large group of diverse proteins with multiple physiological functions. *Fungal Biol Rev*;22:36–42
- Steindorff, AS., Silva, RN., Coelho, ASG., Noronha, EF. and Ulhoa, CJ. (2012): *Trichoderma harzianum* expressed sequence tags for identification of genes with putative roles in mycoparasitism against *F. solani*. *Biol Control* **61**(2):134–140.
- Steele, D. and Crane, JH. (2006). The state of the Florida tropical fruit industry and the challenges growers face. *Proc. Florida State. Hort. Soc.* 119:7–8.
- Schrempf, H. (2001). Recognition and degradation of chitin by streptomycetes. *Antonie Van Leeuwenhoek* 79(3–4):285–289.
- Samuels, G.J., Chaverri, P., Farr, D.F. and McCray, E.B. (2015). *Trichoderma* Online, Systematic Mycology and Microbiology Laboratory, ARS, USDA. Retrieved June 28, 2015, from /taxadescriptions/keys/*Trichoderma*Index.cfm
- Smith, B.J. and Black, L.L., (1990). Morphological, cultural, and pathogenic variation among *Colletotrichum* species isolated from strawberry. *Plant Disease*.74(1):69-76.
- Slippers, B., Crous, PW., Denman, S., Coutinho, TA., Wingfield, BD., Wingfield, MJ. (2004). Combined multiple gene genealogies and phenotypic characters differentiate several species previously identified as *Botryosphaeria dothidea*. *Mycologia* 96:83–101.
- Svetlana Živkovic, S., Stojanovic, Ž., Ivanovic, V., Gavrilovic, T., Popovic, J and Balaž. (2010). Screening of antagonistic activity of microorganisms against *Colletotrichum acutatum* and *Colletotrichum gloeosporioides*. *Arch. Biol. Sci.*, 62, pp. 611–623
- Stals, I., Samyn, B., Sergeant, K., White, T., Hoorelbeke, K., Coorevits, A., Devreese, B., Claeysens, M. and Piens, K. (2010). Identification of a gene coding for a deglycosylating enzyme in *Hypocrea jecorina*. *FEMS Microbiol Lett* 303(1):9–17.
- Synowiecki, J., Al-Khateeb, NAAQ. (2000). The recovery of protein hydrosylate during enzymatic isolation of chitin from shrimp *Cragon cragon* processing discards. *J Food Chem* 68:147–152
- Saitou, N. and Nei, M. (1987). The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution* 4 :406-425.
- Schuster, André and Monika, Schmoll. (2010). “Biology and Biotechnology of *Trichoderma*.” *Applied microbiology and biotechnology* 87(3): 787–99.
- Singh, Anuradha., Mohammad Shahid, Mukesh Srivastava, Sonika Pandey, Antima Sharma and Vipul Kumar. (2014). “Virology & Mycology Optimal Physical Parameters for Growth of *Trichoderma* Species at Varying pH , Temperature and Agitation.” *Virol Mycol* 3: 127.
- Shah-Smith, DA. and Burns, RG. (1997) Shelf-life of a biocontrol *Pseudomonas putida* applied to the sugar beet seeds using commercial coatings. *Biocontrol Sci Technol* 7: 65-74
- Streit, S., Michalski, C.W., Erkan, M., Kleef, J., and Friess, H. (2009). Northern blot analysis for detection and quantification of RNA in pancreatic cancer cells and tissues. *Nat Proto* 4(1): 37-43
- Shimanuki, T. (1987). Studies on the mechanisms of the infection of timothy with purple spot disease caused by *Cladosporium* (Gregory) de Vries. In *Res. Bull.* 148 Hokkaido Natl. Agric. Exp. Sta. pp 1-56.

- Sivan, A. and Chet, I. (1989). Degradation of fungal cell walls by lytic enzymes of *Trichoderma harzianum*. *Journal of General Microbiology*. 135: 675-682.
- Sanz, L., Montero, M., Redondo, J., Llobell, A. and Monte, E., (2005). Expression of an alpha-1,3-glucanase during mycoparasitic interaction of *Trichoderma asperellum*. *FEBS. J.* 272: 493-499
- Seidl, V., Druzhinina, I.S. and Kubicek, C.P., (2006). A screening system for carbon sources enhancing b-N-acetylglucosaminidase formation in *Hypocrea atroviridis* (*Trichoderma atroviride*). *Microbiology* 152: 2003-2012.
- Sietsma, H. and Wessels, J.G. (1977). Chemical Analysis of the Hyphal Walls of *Schizophyllum commune*. *Biochim. Biophys. Acta*, vol. 196, no. 2, pp. 225-239
- Sharma, P.N., Kaur, M., Sharma, O.P., Sharma, P. and Pathania, A., (2005). Morphological, pathological and molecular variability in *Colletotrichum capsici*, the cause of fruit rot of chillies in the subtropical region of north-western India. *Journal of Phytopathology* 153(4):232-237
- Samuels, GJ., Dodd, SL., Gams, W., Castlebury, LA., Petrini, O. (2002a). *Trichoderma* species associated with the green mold epidemic of commercially grown *Agaricus bisporus*. *Mycologia*.;94:146.
- Samuels, GJ., Chaverri, P., Farr, DF. and McCray, EB. (2002b). *Trichoderma* Online [Online]. Available at: <http://nt.arsgrin.gov/taxadescriptions/keys/TrichodermaIndex.cfm> [Accessed: 16 February 2013].
- Suhaila, Mamat. (2014). Isolation and characterization of antifungal activity of endophytic *Penicillium oxalicum* T3.3 for anthracnose biocontrol in dragon fruit (*Hylocereus* sp.). Master thesis, Universiti Putra Malaysia.
- Sakalidis, ML., Ray, JD., Lanoiselet, V., Hardy, GES. and Burgess, TI. (2011). Pathogenic Botryosphaeriaceae associated with *Mangifera indica* in the Kimberly Region of Western Australia. *Eur J Plant Pathol* 130:379-391.
- Souza, R. F., Gomes, R. C., Coelho, R. R. R., Alviano, C. S., and Soares, R. M. A. (2003). Purification and characterization of an endochitinase produced by *Colletotrichum gloeosporioides*. *FEMS microbiology letters*, 222(1): 45-50.
- Sambrook, J. and Russel, D.W. (2001). *Molecular Cloning: A Laboratory Manual* CSH Laboratory Press, Cold Spring Harbor, NY.
- Simkovic, M., Ditte, P., Chovanec, P., Varecka, L. Lakatos, B. (2007). Changes in growth competence of aged *Trichoderma viride* vegetative mycelia. *Antonie Van Leeuwenhoek* 91: 407-416
- Takaya, N., Yamazaki, D., Horiuchi, H., Ohta, A., Takagi, M. (1998). Cloning and characterization of a chitinase encoding gene (*chiA*) from *Aspergillus nidulans*, disruption of which decreases germination frequency and hyphal growth. *Biosci Biotechnol Biochem*; 62: 60-65
- Turner, D., Kovacs, W., Kuhls, K., Lieckfeldt, E., Peter, B., Arisan-Atac, I., Strauss, J., Samuels, G. J., Boerner, T., and Kubicek, C. P. (1997). Biogeography and phenotypic variation in *Trichoderma* sect. *Longibrachiatum* and associated *Hypocrea* species. *Mycol Res.* 101: 449-459.
- Takaya, N., Yamazaki, D., Horiuchi, H., Ohta, A. and Takagi, M. (1998). Intracellular chitinase gene from *Rhizopus oligosporus*: molecular cloning and characterization. *Microbiology*; 144: 2647-2654.
- Thirunavukkarasu, N., Dhinamala, K., and Moses Inbaraj, R. (2011). Production of chitin from two marine stomatopods *Oratosquilla* spp. (Crustacea) *J. Chem. Pharm. Res.*, 3(1):353-359

- Tamura, K. and Nei M. (1993). Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Molecular Biology and Evolution* 10:512-526.
- Thamthiankul, S., Suan-Ngay, S., Tantimavanich, S. and Panbangred, W. (2001). Chitinase from *Bacillus thuringiensis* subsp. pakistani. *Applied Microbiology and Biotechnology* 56: 395–401
- Tamura, K., Stecher, G., Peterson, D., Filipski, A., and Kumar, S. (2013). MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Molecular Biology and Evolution* 30: 2725-2729.
- Than, P.P., Jeewon, R., Hyde, K.D., Pongsupasamit, S., Mongkolporn, O. and Taylor, P.W.J., (2008). Characterization and pathogenicity of *Colletotrichum* species associated with anthracnose disease on chilli (*Capsicum* spp.) in Thailand. *Plant Pathology*, 57(3):562-572.
- Taniguchi, M., Miura, K., Iwao, H., and Yamanaka, S. (2001). Quantitative assessment of DNA microarrays comparison with Northern blot analyses. *Genomics* 71(1): 34-39
- Udvardi, M. K., Czehowski, T., and Scheible, W. R. (2008). Eleven golden rules of quantitative RT-PCR. *Plant Cell*, 20(7): 1736-1737
- Verma, V.C., Gond, S.K., Kumar, A., Kharwar, R.N. and Strobel, G.A. (2007). The endophytic mycoflora of bark, leaf and stem tissues of *Azadirachta indica* A. Juss (Neem) from Vanasi (India). *Microb Ecol* 54:119–125.
- Verma, M., Brar, S.K., Tyagi, R.D., Surampalli, R.Y. and Valero J.R. (2007). Antagonistic fungi, *Trichoderma* spp.: panoply of biological control, *Biochem. Eng. J.*, 37: 1–20
- Vecht-Lifshitz, S., Magdassi, S. and Braun, S. (1990). Pellet formation and cellular aggregation in *Streptomyces tendae*. *Biotechnol Bioeng*;35:890–6.
- Valencia-Botín, A.J., Livera-Muñoz, M. and Sandoval-Islas, J.S. (2004). Caracterización de una cepa de *Fusicoccum* sp. anamorfo de *Botryosphaeria dothidea* Moug.: Fr (Ces. and De Not.) aislada de pitahaya [*Hylocereus undatus* (Haw.) Britton y Rose] Cactaceae. *Revista Mexicana de Fitopatología* 23: 157-161. Cellular and molecular biology of filamentous fungi, ASM Press, Herndon, 56: 676–694
- Viterbo, A. and Horwitz B.A. (2010). *Mycoparasitism*. In: Borkovich KA, Ebbole DJ (eds).
- Viterbo, A., Montero, M., Ramot, O., Friesem, D., Monte, E., Llodell, A. and Chet, I. (2002). Expression regulation of the endochitinase *vhit36* from *Trichoderma asperellum* (*T. harzianum* T-203). *Curr Genet*; 42: 114–122.
- Valencia-Botín, Alberto J., Hirotaka Kokubu, and Yolanda D. Ortiz-Hernández. (2013). “A Brief Overview on Pitahaya (*Hylocereus* Spp.) Diseases.” *Australasian Plant Pathology* 42: 437–40.
- Vijaya, Suzianti Iskandar, Intan Sakinah, Mohd Anuar, and Latiffah Zakaria. (2015). “Characterization and Pathogenicity of *Colletotrichum Truncatum* Causing Stem Anthracnose of Red Fleshed Dragon Fruit (*Hylocereus Polyrhizus*) in Malaysia.” 163: 67–71.
- VanGuilder, H. D., Vrana, K. E., and Freeman, W. M. (2008). Twenty-five years of quantitative PCR for gene expression analysis. *Biotechniques*, 44(5): 619-626
- Von Arx, J.A. (1987). *Plant Pathogenic Fungi*. Berlin, J. Cramer.
- Von Arx, J.A. (1957). Die Arten der Gattung *Colletotrichum* Cda. *Phytopathologische Zeitschrift*, 29:414-468.

- Van Driesche, R.G. and Bellows, T.S. Jr. (1996). Use of host and parasitoid recruitment in quantifying losses from parasitism in insect populations with reference to *Pieris rapae* and *Cotesia glomerata*. *Ecological Entomology* 13: 215-222
- Vieira, P.M., Coelho, A.S., Steindorff, A.S., De Siqueira, S.J., Silva Rdo, N., Ulhoa, C.J. (2013): Identification of differentially expressed genes from *Trichoderma harzianum* during growth on cell wall of *Fusarium solani* as a tool for biotechnological application. *BMC Genomics*, 14:177
- Wharton, P.S. and Diéguez-Uribeondo, J. (2004). The biology of *Colletotrichum acutatum*. *Anales del Jardín Botánico de Madrid*, 61:3-22.
- Walton, J.D. (1994). Deconstructing the cell wall. *Plant Physiol.* 104:1113-1118
- White, T.J., Bruns, T., Lee, S. and Taylor, J., (1990). *Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics*. Academic Press Inc., San Diego, CA, pp. 315–322.
- Woo, S.L., Scala, F., Ruocco, M. and Lorito, M., (2006). The molecular biology of the interactions between *Trichoderma* spp., phytopathogenic fungi, and plants. *Phytopathology* 96: 181–185
- Wiwat, C., Siwayaprahm, P. and Bhumiratana (1999). Purification and Characterization of Chitinase from *Bacillus circulans* No.4.1. A. *Curr Microbiol*; 39: 134.
- Yanai, K., Takaya, N., Kojima, N., Horiuchi, H., Ohta, A. and Takagi, M. (1992). Purification of two chitinases from *Rhizopus oligosporus* and isolation and sequencing of the encoding genes. *J Bacteriol*; 174: 7398–7406.
- Yamazaki, H., Tanaka, A., Kaneko, J.-I., Ohta, A. and Horiuchi, H. (2008). *Aspergillus nidulans* ChiA is a glycosylphosphatidylinositol (GPI)-anchored chitinase specifically localized at polarized growth sites. *Fungal Genetics and Biology* 45: 963-972.
- Yoder, J.A., Glenn, B.D., Benoit, J.B., Zettler, L.W. (2008). The giant Madagascar hissing-cockroach (*Gromphadorhina portentosa*) as a source of antagonistic moulds: concerns arising from its use in a public setting. *Mycoses* 51:95–98
- Zakaria, M.H. (1989). Some Aspects of the biology and chemically assisted biological control of *Ganoderma* species in Malaysia, PhD, University of Putra, Malaysia,
- Zeilinger, S., Reithner, B., Scala, V., Peiss, I., Lorito, M., Mach, R.L. (2005) Signal transduction by *Tga3*, a novel G protein alpha subunit of *Trichoderma atroviride*. *Appl Environ Microbiol* 71:1591–1597 47.
- Zainoldin, K.H., Baba, a.S. (2009). “The Effect of *Hylocereus Polyrhizus* and *Hylocereus Undatus* on Physicochemical , Proteolysis , and Antioxidant Activity in Yogurt.” *Engineering and Technology* 60(12): 361–66.
- Zhang, Hong Bin., Ming Yan Liu, Yu Jing Tian, and Xue Qin Hu. (2011). “Comparative Characterization of Chitinases from Silkworm (*Bombyx Mori*) and Bollworm (*Helicoverpa Armigera*).” *Cell Biochemistry and Biophysics* 61: 267–75.
- Zhao, J., Xue, Q.H., Niu, G.G., Xue, L., Shen, G.H. and Du, J.Z., (2013). Extracellular enzyme production and fungal mycelia degradation of antagonistic *Streptomyces* induced by fungal mycelia preparation of cucurbit plant pathogens. *Ann. Microbiol.* 63: 809–812.