



UNIVERSITI PUTRA MALAYSIA

***ANTIBACTERIAL ACTIVITIES OF *Aspergillus fumigatus* SSH01
CRUDE EXTRACTS***

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CRUDE EXTRACTS**

By

MOHAMAD KHAIRIL BIN RADZALI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirement for the Degree of Master of Science**

June 2016

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Abstract of thesis presented to the Senate of Universiti of Putra Malaysia in fulfilment
of the requirement for the degree of Master of Science

ANTIBACTERIAL ACTIVITIES OF *Aspergillus fumigatus* SSH01 CRUDE EXTRACTS

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June 2016

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Antimicrobial agents are mainly derived from natural products. The scientific evidence for the various antimicrobial activities from thermotroph, as referred here, thermotolerant *Aspergillus fumigatus* SSH01 is scarce and fragmented. Additionally, bacterial resistance to antibiotics is a public health problem that is increasing around the globe which later complicated illnesses. Therefore, the main objective of this study was to determine the antibacterial activities of fungal crude extracts and to investigate the modes of action of the extracts against susceptible pathogens. Methanol was used to extract metabolites from *A. fumigatus* SSH01 by submerged fermentation. The extract was investigated for its antimicrobial properties against pathogenic Gram-positive bacteria (methicillin-resistant *Staphylococcus aureus* S547, *Staphylococcus aureus* ATCC 6538, *Bacillus subtilis* ATCC 6633 and *Listeria monocytogenes* L10) and pathogenic Gram-negative bacteria (*Escherichia coli* ATCC 8739, *Escherichia coli* 0157: H7 E187, *Salmonella enterica* serovar Thyphimurium S836 and *Pseudomonas aeruginosa* ATCC 15442) as well as yeast, *Candida albicans* ATCC 10231 using disc diffusion method. The minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) of the extract were determined by broth microdilution method. Time-killing kinetics, scanning electron microscopy (SEM), and detection of cellular leakage of proteins and 260_{nm} absorbing-compounds of susceptible Gram-positive pathogens were determined after exposure of bacterial cells to three concentrations (1/2, 1, and 2MIC) of 100% of methanolic crude extracts. The methanolic extract exhibited zone of inhibition ranged from 11.00 mm to 18.00 mm. The MIC values ranged from 0.097 mg/ml to 12.50 mg/ml whereas the MBC values ranged from 0.195 mg/ml to 25 mg/ml. The growth of all treated test microorganisms were inhibited at the concentration of twice MIC values in time-kill assay. Average log reduction of bactericidal effects (MIC and 2MIC) in viable cell count ranged from 7.75 Log₁₀ to 5.52 Log₁₀ CFU/ml and between 7.72 Log₁₀ to 6.36 Log₁₀ CFU/ml in MIC and 2MIC of the extracts, respectively. Detection of cellular leakage of proteins (595_{nm}) and 260_{nm} absorbing-compounds showed slightly increased to the time of exposure of cells to the extracts and suggesting its ability to damage the bacterial membrane. In addition, scanning electron micrograph (SEM) demonstrated disruption in morphology of the

treated cells. Liquid chromatography/time-of-flight/mass spectrophotometry (LC/TOF-MS) was used to analyze the constituents of the extracts. Six metabolites were identified included L-tyrosine, kojic acid, fumagillin, fumigaclavine B, helvolic acid and citrinin. In MTT assay, SSH01 crude extract was found to be cytotoxic against HaCat with IC_{50} of 20 mg/ml and 0.78 mg/ml after 24 and 48 h of treatment. Further studies maybe required to explain the risk of toxicity (kojic acid and citrinin) of the crude extracts and its stability for the application of human use. The constituents of the extracts should also be isolated, identified, characterized perhaps improvise for better potential as antibacterial drugs.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**AKTIVITI ANTIBAKTERIA DARIPADA EKSTRAK MENTAH
Aspergillus fumigatus SSH01**

Oleh

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Ejen antimikrob kebanyakannya diperolehi daripada produk semula jadi. Walau bagaimanapun, bukti saintifik untuk aktiviti antibakteria dari termotrof, *Aspergillus fumigatus* SSH01 yang mampu bertoleransi terhadap haba adalah terhad dan tidak cukup. Tambahan, rintangan bakteria terhadap antibiotik merupakan masalah kesihatan awam yang semakin meningkat di seluruh lapisan dunia seterusnya merumitkan lagi sesuatu penyakit. Oleh itu, objektif utama kajian ini adalah untuk menentukan aktiviti antibakteria daripada ekstrak mentah kulat dan menentukan mod tindakan ekstrak terhadap patogen. Metanol telah digunakan untuk mengekstrak metabolit daripada *A. fumigatus* SSH01 menggunakan penapaian kultur tenggelam. Sifat-sifat antimikrob ekstrak telah disiasat ke atas Gram-positif patogenik (rintang methisilin *Staphylococcus aureus* S547, *Staphylococcus aureus* ATCC 6538, *Bacillus subtilis* ATCC 6633 dan *Listeria monocytogenes* L10) dan bakteria Gram-negatif patogenik (*Escherichia coli* ATCC 8739, *Escherichia coli* 0157: H7 E187, *Salmonella enterica* serovar Thyphimurium S836 dan *Pseudomonas aeruginosa* ATCC 15442) serta yis, *Candida albicans* ATCC 10231 menggunakan kaedah sebaran cakera. Nilai kepekatan perencatan minimum (MIC) dan kepekatan bakterisidal minimum (MBC) ekstrak ditentukan dengan menggunakan kaedah mikro-pencairan. Kinetik masa-pembunuhan, imbasan mikroskop elektron (SEM), ukuran kebocoran sel terhadap protein dan penyerapan pada jarak gelombang 260_{nm} oleh patogen Gram-positif ditentukan selepas pendedahan sel-sel bakteria untuk tiga kepekatan (1/2, 1, dan 2MIC) daripada 100% ekstrak mentah metanol *A. fumigatus* SSH01. Ekstrak metanol mempamerkan zon perencatan diameter dari 11.00 mm hingga 18.00 mm dan terbatas kepada bakteria Gram-positif sahaja. Nilai MIC adalah di antara 0.097 mg/ml hingga 12.50 mg/ml manakala nilai MBC adalah antara 0.195 mg/ml hingga 25 mg/ml. Pertumbuhan semua mikroorganisma ujian direncat pada kepekatan nilai dua kali MIC dalam kinetik masa-pembunuhan. Purata pengurangan log bagi kesan bakterisidal (MIC dan 2MIC) sel yang hidup adalah dari Log₁₀ 7.75 kepada Log₁₀ 5.52 CFU/ml dan di antara Log₁₀ 7.72 kepada Log₁₀ 6.36 CFU/ml bagi MIC dan 2MIC daripada ekstrak. Analisis bagi ukuran kebocoran sel terhadap protein dan sebatian yang menyerap gelombang pada 260_{nm} menunjukkan sedikit peningkatan selepas pendedahan sel kepada ekstrak dan

disimpulkan bahasawanya ia mampu merosakkan membran bakteria. Di samping itu, imbasan elektron mikroskopi (SEM) menunjukkan perubahan morfologi terhadap sel-sel yang diuji. “Liquid chromatography/time-of-flight-mass spectrometry” (LC/TOF-MS) telah digunakan untuk menganalisis juzuk-juzuk ekstrak. Enam metabolit telah dikenal pasti termasuklah L-tirosin, asid kojik, fumagillin, fumigaclavine B, asid helvolik dan citrinin. Berdasarkan dari ujian MTT, ekstrak mentah SSH01 didapati memberi kesan sitotoksik terhadap HaCat dengan nilai IC_{50} 20 mg/ml dan 0.78 mg/ml selepas 24 dan 48 jam. Kajian lanjut mungkin diperlukan untuk menerangkan risiko ketoksikan (asid kojik dan sitrinin) daripada ekstrak mentah dan kestabilan bagi kegunaan manusia. Juzuk daripada ekstrak juga perlu diasingkan, dikenalpasti dan ditambah baik supaya berpotensi digunakan sebagai ubatan antibakteria.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

ATCC	American Type Culture Collection
ATP	Adenosine Triphosphate
BLAST	Basic Local Alignment Search Tool
CDC	Centers for Disease Control and Prevention
CFU	Colony Forming Unit
cm	Centimeter
DMSO	Dimethylsulphoxide
DNA	Deoxyribonucleic Acid
ESBL	Extended-spectrum Beta-Lactamase
g	Gram
GC-MS	Gas Chromatography-Mas Spectrometry
HIV	Human Immunodeficiency Virus
HPLC	High-Performance Liquid Chromatography
ITS	Internal transcribed spacer
L	Liter
lat.	Latitude
LC/TOF-MS	Liquid-Chromatography/Time-of-Flight Mass Spectrometry
Log	Logarithm
long.	Longitude
M	Molarity
MBC	Minimum Bactericidal Concentration
MDR	Multi-drug Resistance
mg	Milligram
MHA	Mueller-Hinton Agar
MHB	Mueller-Hinton Broth
MIC	Minimum Inhibitory Concentration
min	Minute
ml	Milliliter
mm	Millimeter
mRNA	messenger Ribonucleic Acid
MRSA	Methicillin-resistant <i>Staphylococcus aureus</i>
MTT	3-(4, 5-dimethylthiazolyl-2)-2, 5-diphenyltetrazolium Bromide
NCBI	National Center of Biotechnology Information
ND	Not Determined
NI	No Inhibition
NIST	National Institute of Standards and Technology
nm	Nanometer
OD	Optical Density
PBS	Phosphate Buffer Saline
PCR	Polymerase Chain Reaction
PDA	Potato Dextrose Agar
PDB	Potato Dextrose Broth
pH	Exponential of Hydrogen Ion
rpm	Revolution per Minute
rRNA	Ribosomal Ribonucleic Acid
SD	Standard Deviation
SEM	Scanning Electron Microscopy

sp.	Species
tRNA	transfer Ribonucleic Acid
UV	Ultraviolet
VRE	Vancomycin-resistant Enterococci
VRSA	Vancomycin-resistant <i>Staphylococcus aureus</i>
%	Percentage
μ	Micro
°C	Degree Celsius
α	Alpha
β	Beta
(v/v)	Volume per volume



CHAPTER 1

INTRODUCTION

For thousand years, natural products have played such crucial role worldwide in treating and preventing human diseases (Chin et al., 2006). The drugs production from the natural product is mainly produced by microbes. Of 23000 active compounds discovered from microorganisms (antimicrobial, antiviral, cytotoxic and immunosuppressive compounds), 42% are isolated from fungi while 32% by filamentous bacteria, the actinomycetes (Demain, 2014). In addition, 20% of antibiotics found contributed by fungi, 10-15% by non-filamentous bacteria while over half of it are from actinomycetes and these compounds are semi-synthetic derivatives from novel. Penicillin, cephalosporins, tetracycline, aminoglycosides, macrolides, ansamycins, polyenes and glycopeptides are examples of antimicrobial agents produced by chemistry and bioconversion (Demain, 2014).

However, the cost of healthcare including morbidity and mortality has increased due to the emergence of antibiotic-resistant (Deutscher and Friendman, 2010). Malaysia, the 67th largest country by total land area is facing the same issues like elsewhere in Africa, America, Europe, and Eastern Mediterranean Region (WHO, 2015). In 2005, Malaysia faced prevalence of multi-drug resistance (MDR) of nosocomial infections caused by *Pseudomonas aeruginosa* strains with 6.9% higher than Japan (2.8%) in Asia-Pacific (Tsuji et al., 2005; Raja and Singh, 2007; Giske et al., 2008). A study that was carried out by Pathmanathan et al. (2009), showed that 25 out of 97 clinical *Pseudomonas aeruginosa* isolated from Hospital Kuala Lumpur were resistant towards at least one of antimicrobial agents used (cephalosporin, carbapenem, aminoglycoside and quinolone).

To date, it is reported that majority of *Enterobacteriaceae* and *P. aeruginosa* exhibited multiple resistance to ampicillin, augmentin, cefuroxime and ceftriaxone (Kor et al., 2013). Gould et al. (2014) stated that emergence of extended-spectrum beta-lactamase (ESBL) Gram-positive bacteria, as well as multidrug-resistant *Pseudomonas* has been unchanging for years. In addition, the incidence of drug resistance in *Acinetobacter* against nearly all major antibiotics is increasing.

Fungi provide such priceless secondary metabolites values in potential biomedical applications (Gao et al., 2012). It produces a great variety of unique natural products ranging from peptides to alkaloids to terpenes and polyketides (Keller et al., 2005). Penicillin and cephalosporins are the best-known metabolites from fungi that serve as antimicrobial agents in pharmacology (Gloer, 2007). The production of fungal metabolites is renewable, modifiable and desirable. It can be produced in large scale (Gloer, 2007), improved and modified structurally by the genetic mutation, and the production can be increased by optimization of medium and culture conditions (Masurekar, 2005; Aly et al., 2011). Nonetheless, data on natural products from thermotolerant for biological activities are still fragmented. Its useful thermostable enzymes probably the major reason why it is often to be reported as prior candidates in industries.

Taken together, the research in the field of natural products isolated from thermotolerant fungal isolates perhaps could serve the humans demand to overcome the emerging and known diseases. Thus, this study was performed to investigate the antimicrobial activity of fungal crude extracts. The objectives of the study were:

1. To screen for antimicrobial properties of methanolic fungal crude extracts
2. To determine the modes of action of fungal crude extracts against susceptible test pathogens
3. To identify the fungal isolate and its metabolites as well as to perform the cytotoxicity of the crude extract



REFERENCES

- Abdel-Fattah, H. M., Moubasher, A. H., & Abdel-Hafez, S. I. (1977). Studies on mycoflora of salt-marshes in Egypt. III. Thermophilic fungi. *Bulletin of the Faculty of Science, Assiut University*, 6, 225-235.
- Adams, M. R., & Moss, M. O. (2008). Chapter 3. Factors affecting the growth and survival of microorganisms in foods. In *Food Microbiology* (pp 45-48). London: RSC Publishing.
- Ahmed, S. I., & Wangsai, J. (2007). Fermented Milk in Asia. In Y.H. Hui. *Handbook of Food Products Manufacturing: Health, Meat, Milk, Poultry, Seafood, and Vegetables* (pp. 431-448). New Jersey: Wiley.
- Alanis, A. J. (2005). Resistance to antibiotics: are we in the post-antibiotic era?. *Archives of Medical Research*, 36(6), 697-705.
- Allen, P. J., & Emerson, R. (1949). Guayule rubber: microbiological improvement by shrub retting. *Industrial & Engineering Chemistry*, 41(2), 346-365.
- Aly, A. H., Debbab, A., & Proksch. P. (2011). Fifty years of drug discovery from fungi. *Fungal Diversity*, 50(1), 3-19.
- Aminov, R. I. (2010). A brief history of the antibiotic era: lessons learned and challenges for the future. *Frontiers in Microbiology*, 134, 1-7.
- Anderson, V. E., & Osheroff, N. (2001). Type II topoisomerases as targets for quinolone antibacterials turning Dr. Jekyll into Mr. Hyde. *Current Pharmaceutical Design*, 7(5), 337-353.
- Aneiros, A., & Garateix, A. (2004). Bioactive peptides from marine sources: pharmacological properties and isolation procedures. *Journal of Chromatography B*, 803(1), 41-53.
- Apinis, A. E. (1963). Occurrence of thermophilous microfungi in certain alluvial soils near Nottingham. *Nova Hedwigia*, 5, 57-78.
- Apinis, A. E., & Pugh, G. J. F. (1967). Thermophilous fungi of birds' nests. *Mycopathologia et Mycologia Applicata*, 33(1), 1-9.
- Aqil, F., Ahmad, I., & Owais, M. (2006). Evaluation of anti-methicillin-resistant *Staphylococcus aureus* (MRSA) activity and synergy of some bioactive plant extracts. *Biotechnology Journal*, 1(10), 1093-1102.
- Aqil, M., Ahad, A., Sultana, Y., & Ali, A. (2007). Status of terpenes as skin penetration enhancers. *Drug Discovery Today*, 12(23), 1061-1067.
- Ardic, N., Sareyyupoglu, B., Ozyurt, M., Haznedaroglu, T., & Iiga, U. (2006). Investigation of aminoglycoside modifying enzyme genes in methicillin-resistant staphylococci. *Microbiological Research*, 161(1), 49-54.

- Arekemase, M. O., Kayode, R. M. O., & Ajiboye, A. E. (2011). Antimicrobial activity and phytochemical analysis of *Jatropha curcas* plant against some selected microorganisms. *International Journal of Biology*, 3(3), 52-59.
- Arivudainambi, U. S., Anand, T. D., Shanmugaiah, V., Karunakaran, C., & Rajendran, A. (2011). Novel bioactive metabolites producing endophytic fungus *Colletotrichum gloeosporioides* against multidrug-resistant *Staphylococcus aureus*. *FEMS Immunology & Medical Microbiology*, 61(3), 340-345.
- Asghari, S., Baharfar, R., Alimi, M., Ahmadipour, M., & Mohseni, M. (2014). Synthesis and antibacterial activities of pyrano [3, 2-b] pyranones from kojic acid, ethyl cyanoacetate, and benzaldehydes in aqueous K₂CO₃. *Monatshefte für Chemie-Chemical Monthly*, 145(8), 1337-1342.
- Avalos, J., & Cerda-Olmedo, E. (2004). Fungal carotenoid production: Commercial Application and Biotechnological Potential. In D.K. Arora. *Handbook of Fungal Biotechnology* (pp. 367-378). New York: Marcel Dekker.
- Bagli, J., Kluepfel, D., & St.-Jacques, M. (1973). Elucidation of structure and stereochemistry of myriocin. Novel antifungal antibiotic. *The Journal of Organic Chemistry*, 38(7), 1253-1260.
- Bai, M. P., & Rao, P. L. (1966). Thermophilic microorganisms. IV. Elaboration of malbranchins a & b by *Malbranchea pulchella*. *Indian Journal of Biochemistry*, 3(3), 187-190.
- Bandow, J. E., Brötz, H., & Hecker, M. (2002). *Bacillus subtilis* tolerance of moderate concentrations of rifampin involves the σ B-dependent general and multiple stress response. *Journal of Bacteriology*, 184(2), 459-467.
- Banerjee, D., & Rathode, B. (2013). To optimise the cultural conditions on the production of kojic acid by *Aspergillus* species. *LS: International Journal of Life Sciences*, 2(3), 168-175.
- Barchan, A., Bakkali, M., Arakrak, A., Pagán, R., & Laglaoui, A. (2014). The effects of solvents polarity on the phenolic contents and antioxidant activity of three *Mentha* species extracts. *International Journal of Current Microbiology and Applied Sciences*, 3(11), 399-412.
- Barchiesi, F., Spreghini, E., Tomassetti, S., Arzeni, D., Giannini, D., & Scalise, G. (2005). Comparison of the fungicidal activities of caspofungin and amphotericin B against *Candida glabrata*. *Antimicrobial Agents and Chemotherapy*, 49(12), 4989-4992.
- Barlow, M. (2009). What antimicrobial resistance has taught us about horizontal gene transfer. In M. B. Gogarten, J. P. Gogarten and L. C. Olendzenski. *Horizontal Gene Transfer* (pp. 397-411). New York: Humana Press.
- Beeson, W. T., Iavarone, A. T., Hausmann, C. D., Cate, J. H., & Marletta, M. A. (2011). Extracellular aldonolactonase from *Myceliophthora thermophila*. *Applied and Environmental Microbiology*, 77(2), 650-656.

- Běhal, V. (2000). Bioactive products from *Streptomyces*. *Advances in Applied Microbiology*, 47, 113-156.
- Běhal, V. (2006). Mode of action of microbial bioactive metabolites. *Folia Microbiologica*, 51(5), 359-369.
- Ben, H. K. A., Moissenet, D., Thien, H. V., & Khedher, M. (2011). Virulence factors in *Pseudomonas aeruginosa*: mechanisms and modes of regulation. *Annales de Biologie Clinique*, 69(4), 393-403.
- Berdy, J. (2005). Bioactive microbial metabolites. *The Journal of Antibiotics*, 58(1), 1-26.
- Bergeron, S., Boopathy, R., Nathaniel, R., Corbin, A., & LaFleur, G. (2015). Presence of antibiotic resistant bacteria and antibiotic resistance genes in raw source water and treated drinking water. *International Biodeterioration & Biodegradation*, 102, 370-374.
- Berrington, D. (2013). Viability reagent, prestoblue, in comparison with other available reagents, utilized in cytotoxicity and antimicrobial assays. *International Journal of Microbiology*, 2013, 1-5.
- Black, J. G. (1999). Antimicrobial therapy. In J. G. Black, *Microbiology: Principles and Explorations* (pp. 340-369). New York: John Wiley & Sons.
- Bladt, T. T., Frisvad, J. C., Knudsen, P. B., & Larsen, T. O. (2013). Anticancer and antifungal compounds from *Aspergillus*, *Penicillium* and other filamentous fungi. *Molecules*, 18(9), 11338-11376.
- Bokhari, S. A. I., Latif, F., & Rajoka, M. I. (2009). Purification and characterization of xylanases from *Thermomyces lanuginosus* and its mutant derivative possessing novel kinetic and thermodynamic properties. *World Journal of Microbiology and Biotechnology*, 25(3), 493-502.
- Bowden, S. D., Hopper-Chidlaw, A. C., Rice, C. J., Ramachandran, V. K., Kelly, D. J., & Thompson, A. (2014). Nutritional and metabolic requirements for the infection of HeLa cells by *Salmonella enterica* serovar *Typhimurium*, *PlosOne*, 9(5), 1-9.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72(1), 248-254.
- Brakhage, A. A., Bruns, S., Thywissen, A., Zipfel, P. F., & Behnsen, J. (2010). Interaction of phagocytes with filamentous fungi. *Current Opinion in Microbiology*, 13(4), 409-415.
- Braude, A. I. (1986). Infectious Diseases and Medical Microbiology, in: Braude, A. I., Davis, C. E., Fierer, J. Eds., 2nd edn (pp. 592-597)., WB Saunders Company: Philadelphia.

- Breidenstein, E. B., de la Fuente-Núñez, C., & Hancock, R. E. (2011). *Pseudomonas aeruginosa*: all roads lead to resistance. *Trends in Microbiology*, 19(8), 419-426.
- Brock, T. D. (1995). The road to Yellowstone—and beyond. *Annual Review of Microbiology*, 49, 1–28.
- Brogden, K. A. (2005). Antimicrobial peptides: pore formers or metabolic inhibitors in bacteria?. *Nature Reviews Microbiology*, 3(3), 238-250.
- Bu'lock, J.D. (1961). Intermediary metabolism and antibiotic synthesis. *Advances in Applied Microbiology*, 3, 293-342.
- Bunger, J., Westphal, G., Mönnich, A., Hinnendahl, B., Hallier, E., & Müller, M. (2004). Cytotoxicity of occupationally and environmentally relevant mycotoxins. *Toxicology*, 202(3), 199-211.
- Bush, K., Jacoby, G. A., & Medeiros, A. A. (1995). A functional classification scheme for beta-lactamases and its correlation with molecular structure. *Antimicrobial Agents and Chemotherapy*, 39(6), 1211–1233.
- Calvo, A. M., Wilson, R. A., Bok, J. W., & Keller, N. P. (2002). Relationship between secondary metabolism and fungal development. *Microbiology and Molecular Biology Reviews*, 66(3), 447-459.
- Cantas, L., Shah, S .Q. A., Cavaco, L. M., Manaia, C. M., Walsh, F., Popowska, M., Garelick, H., Burgmann, H., & Sorum, H. (2013). A brief multi-disciplinary review on antimicrobial resistance in medicine and its linkage to the global environmental microbiota. *Frontiers in Microbiology*, 4(96), 1-14.
- Cappuccino, J.G. & Sherman, N. (Eds.). (2005). *Microbiology a laboratory manual*. 7th ed. United States :Pearson Education, Inc.
- Carbonell, A., & Carrington, J. C. (2015). Antiviral roles of plant Argonautes. *Current Opinion in Plant Biology*, 27, 111-117.
- Carden, S., Okoro, C., Dougan, G., & Monack, D. (2015). Non-typhoidal *Salmonella Typhimurium* ST313 isolates that cause bacteremia in humans stimulate less inflammasome activation than ST19 isolates associated with gastroenteritis. *FEMS Pathogens and Disease*, 73(4), 1-8.
- Carlson, C. A, Pierson, L. S, Rosen, J. J., & Ingraham, J. L. (1983). *Pseudomonas stutzeri* and related species undergo natural transformation. *Journal of Bacteriology*, 153, 93–99.
- Carson, C. F., Mee, B. J., & Riley, T. V. (2002). Mechanism of action of *Melaleuca alternifolia* (tea tree) oil on *Staphylococcus aureus* determined by time-kill, lysis, leakage, and salt tolerance assays and electron microscopy. *Antimicrobial Agents and Chemotherapy*, 46(6), 1914-1920.

- CDC. (2013). Center for Disease Control and Prevention. Antibiotic Resistance Threats in the United States, 2013. Retrieved from <http://www.cdc.gov/drugresistance/pdf/ar-threats-2013-508.pdf>. (accessed on 5th March 2013).
- CDC. (2014). Center for Disease Control and Prevention. CDC 2011 Estimates of foodborne illness in the United States. Retrieved from <http://www.cdc.gov/foodborneburden/2011-foodborne-estimates.html>. (accessed on 5th March 2013).
- Centres for Disease Control and Prevention (US). (2013). Antibiotic resistance threats in the United States, 2013. Centres for Disease Control and Prevention, US Department of Health and Human Services.
- Cetin, E. T., Töreci, K., & Anđ, Ö. (1965). Encapsulated *Pseudomonas aeruginosa* (*Pseudomonas aeruginosa* mucosus) strains. *Journal of Bacteriology*, 89(5), 1432-1433.
- Chain, E., Florey, H. W., Gardner, A. D., Heatley, N. G., Jennings, M. A., Orr-Ewing, J., Sanders, A. G., & Peltier, L.F. (2005). The classic: Penicillin as a Chemotherapeutic Agent. *Clinical Orthopaedics and Related Research*, 439, 23–26.
- Chain, E., Florey, H. W., Jennings, M. A., & Williams, T. I. (1943). Helvolic acid, an antibiotic produced by *Aspergillus fumigatus*, mut. *helvola* Yuill. *British Journal of Experimental Pathology*, 24(3), 108-119.
- Chakradhar, T. (2014). An Overview of Pharmacokinetics, Pharmacodynamics, Adverse Drug Reaction of A Potent New Fifth Generation Cephalosporin: Ceftobiprole. *International Journal of Chemical and Life Sciences*, 2(10), 1237-1239.
- Chapman, S. W., Sullivan, D. C., & Cleary, J. D. (2008). In search of the holy grail of antifungal therapy. *Transactions of the American Clinical and Climatological Association*, 119, 197-216.
- Chen, M., Shao, C. L., Fu, X. M., Xu, R. F., Zheng, J. J., Zhao, D. L., She, Z.G., & Wang, C. Y. (2013). Bioactive indole alkaloids and phenyl ether derivatives from a marine-derived *Aspergillus* sp. Fungus. *Journal of Natural Products*, 76(4), 547-553.
- Chin, Y. W., Balunas, M. J., Chai, H. B., & Kinghorn, A. D. (2006). Drug discovery from natural sources. *The AAPS Journal*, 8(2), 239-253.
- Cho, H., Uehara, T., & Bernhardt, T. G. (2014). Beta-lactam antibiotics induce a lethal malfunctioning of the bacterial cell wall synthesis machinery. *Cell*, 159(6), 1300-1311.

- Chopra, I., & Roberts, M. (2001). Tetracycline antibiotics: mode of action, applications, molecular biology and epidemiology of bacterial resistance. *Microbiology and Molecular Biology Reviews*, 65(2), 232-260.
- Chu, Y. S., Niu, X. M., Wang, Y. L., Guo, J. P., Pan, W. Z., Huang, X. W., & Zhang, K. Q. (2010). Isolation of putative biosynthetic intermediates of prenylated indole alkaloids from a thermophilic fungus *Talaromyces thermophilus*. *Organic Letters*, 12(19), 4356-4359.
- Cleaves, H. J., & Miller, S. L. (1998). Oceanic protection of prebiotic organic compounds from UV radiation. *Proceedings of the National Academy of Sciences*, 95(13), 7260-7263.
- Collin, F., Karkare, S., & Maxwell, A. (2011). Exploiting bacterial DNA gyrase as a drug target: current state and perspectives. *Applied Microbiology and Biotechnology*, 92(3), 479-497.
- Collin, F., Thompson, R. E., Jolliffe, K. A., Payne, R. J., & Maxwell, A. (2013). Fragments of the bacterial toxin microcin B17 as gyrase poisons. *PloS one*, 8(4), 1-9.
- Cooney, D. G., & R. Emerson. (1964). *Thermophilic fungi. An account of their biology, activities and classification*. San Francisco: W. H. Freeman & Co.
- Cos, P., Vlietinck, A. J., Berghe, D. V., & Maes, L. (2006). Anti-infective potential of natural products: how to develop a stronger *in vitro* 'proof-of-concept'. *Journal of Ethnopharmacology*, 106(3), 290-302.
- Cowan, M. M. (1999). Plant products as antimicrobial agents. *Clinical Microbiology Reviews*, 12(4), 564-582.
- Coyle, C. M., & Panaccione, D. G. (2005). An ergot alkaloid biosynthesis gene and clustered hypothetical genes from *Aspergillus fumigatus*. *Applied and Environmental Microbiology*, 71(6), 3112-3118.
- Craig, W.A., & Vogelman, B. (1987). The Postantibiotic effect. *Annals of Internal Medicine*, 106, 900-902.
- Craveri, R., Manachini, P. L., & Aragazzini, F. (1972). Thermozytocidin new antifungal antibiotic from a thermophilic eumycete. *Experientia*, 28(7), 867-868.
- Crisan, E. V. (1959). The isolation and identification of thermophilic fungi. (M.Sc. dissertation). Purdue University, USA.
- Cuenca-Estrella, M. (2014). Antifungal drug resistance mechanisms in pathogenic fungi: from bench to bedside. *Clinical Microbiology and Infection*, 20(Supplement 6), 54-59.
- Dalhoff, A. (2012). Resistance surveillance studies: a multifaceted problem—the fluoroquinolone example. *Infection*, 40(3), 239-262.

- Dantas, G., & Sommer, M. O. (2014). How to fight back against antibiotic resistance. *American Scientist*, 102(1), 42-51.
- Darah, I., Lim, S. H., & Nithianantham, K. (2013). Effects of methanol extract of *Wedelia chinensis osbeck* (asteraceae) leaves against pathogenic bacteria with emphasise on *Bacillus cereus*. *Indian Journal of Pharmaceutical Sciences*, 75(5), 533-539.
- Dastager, S. G. (2013). Isolation, Improvement, and Preservation of Microbial Cultures. In C. R. Coccol, A. Pandey, C. Larroche. *Fermentation Processes Engineering in the Food Industry* (pp. 21-46). New York: CRC Press.
- Davies, J., Brzezinska, M., & Benveniste, R. (2006). R Factors: Biochemical Mechanisms of Resistance to Aminoglycoside Antibiotics. *Annals of the New York Academy of Sciences*, 1022(1), 226-233.
- de Barros, B. S., da Silva, J. P., de Souza Ferro, J. N., Agra, I. K. R., de Almeida Brito, F., Albuquerque, É. D., Caetano, L. C., & Barreto, E. (2011). Methanol extract from mycelium of endophytic fungus *Rhizoctonia* sp. induces antinociceptive and anti-inflammatory activities in mice. *Journal of Natural Medicines*, 65(3-4), 526-531.
- de Carvalho Peixoto-Nogueira, S., Michelin, M., Betini, J. H. A., Jorge, J. A., Terenzi, H. F., & de Moraes, M. D. L. T. (2009). Production of xylanase by *Aspergilli* using alternative carbon sources: application of the crude extract on cellulose pulp biobleaching. *Journal of Industrial Microbiology & Biotechnology*, 36(1), 149-155.
- De Kievit, T. R., Parkins, M. D., Gillis, R. J., Srikumar, R., Ceri, H., Poole, K., Iglewski, B.H., & Storey, D. G. (2001). Multidrug efflux pumps: expression patterns and contribution to antibiotic resistance in *Pseudomonas aeruginosa* biofilms. *Antimicrobial Agents and Chemotherapy*, 45(6), 1761-1770.
- Demain, A. L. (2014). Importance of microbial natural products and the need to revitalize their discovery. *Journal of Industrial Microbiology & Biotechnology*, 41(2), 185-201.
- Demanèche, S., Kay, E., Gourbière, F., & Simonet, P. (2001). Natural transformation of *Pseudomonas fluorescens* and *Agrobacterium tumefaciens* in soil. *Applied and Environmental Microbiology*, 67(6), 2617-2621.
- Deutscher, M., & Friedman, C. (2010). Antibiotic Resistance and Implications for the Appropriate Use of Antimicrobial Agents. In A.G. Mainous III and C. Pomeroy, *Management of Antimicrobials in Infectious Diseases: Impact of Antibiotic Resistance* (pp. 1-30). New York: Humana Press.
- Domagk, G. (1935). Ein Beitrag zur chemotherapie der bakteriellen infektionen. *D Med Wschr* 1935; 61: 250-3. Citat a: Classics in infectious diseases. A contribution to the chemotherapy of bacterial infections. *Reviews of Infectious Diseases*, 8, 163-166.

- Doss, A., Parivuguna, V., & Poovendran, P. (2010). Antimicrobial screening of secondary metabolites from *Phomopsis theae*. *Asian Journal of Experimental Biological Science*, 1, 370-372.
- Draenert, R., Seybold, U., Grützner, E., & Bogner, J. R. (2015). Novel antibiotics: Are we still in the pre-post-antibiotic era?. *Infection*, 43(2), 145-151.
- Drevets, D. A., & Bronze, M. S. (2008). *Listeria monocytogenes*: epidemiology, human disease, and mechanisms of brain invasion. *FEMS Immunology & Medical Microbiology*, 53(2), 151-165.
- Drews, J. (2000). Drug discovery: a historical perspective. *Science*, 287(5460), 1960-1964.
- Driks, A. (2002). Overview: development in bacteria: spore formation in *Bacillus subtilis*. *Cellular and Molecular Life Sciences CMLS*, 59(3), 389-391.
- Drlica, K., Hiasa, H., Kerns, R., Malik, M., Mustaev, A., & Zhao, X. (2009). Quinolones: action and resistance updated. *Current Topics in Medicinal Chemistry*, 9(11), 981-998.
- Dyke, K. G. H., & Gregory, P. D. (1996). Resistance to beta-lactam antibiotics: resistance mediated by beta-lactamase. In K. B. Crossley and G. L. Archer. *The Staphylococci in Human Disease. 1st edition*. (pp. 136-157). London: Churchill Livingstone.
- El-Azizi, M., Farag, N., & Khardori, N. (2015). Antifungal activity of amphotericin B and voriconazole against the biofilms and biofilm-dispersed cells of *Candida albicans* employing a newly developed *in vitro* pharmacokinetic model. *Annals of Clinical Microbiology and Antimicrobials*, 14(1), 21-30.
- El-Gendy, B. E. D. M., & Rateb, M. E. (2015). Antibacterial activity of diketopiperazines isolated from a marine fungus using t-butoxycarbonyl group as a simple tool for purification. *Bioorganic & Medicinal Chemistry Letters*, 25(16), 3125-3128.
- El-Gendy, M. M., Hawas, U. W., & Jaspars, M. (2008). Novel bioactive metabolites from a marine derived bacterium *Nocardia* sp. ALAA 2000. *The Journal of Antibiotics*, 61(6), 379-386.
- Ellis, D. H. (1980). Thermophilic fungi isolated from a heated aquatic habitat. *Mycologia*, 1030-1033.
- Ellis, D. H. (1981). Sporangiospore ornamentation of thermophilic *Rhizopus* species and some allied genera. *Mycologia*, 73, 511-523.
- European Committee on Antimicrobial Susceptibility Testing (EUCAST). (2012). Antimicrobial susceptibility testing EUCAST disk diffusion method. Version 5.0 (January 2015), 1-21.

- Fan, G., Katrolia, P., Jia, H., Yang, S., Yan, Q., & Jiang, Z. (2012). High-level expression of a xylanase gene from the thermophilic fungus *Paecilomyces thermophila* in *Pichia pastoris*. *Biotechnology Letters*, 34(11), 2043-2048.
- Finking, R., & Marahiel, M. A. (2004). Biosynthesis of nonribosomal peptides. *Annual Review of Microbiology*, 58, 453-488.
- Fischbach, M. A., & Walsh, C.T. (2009). Antibiotics for emerging pathogens. *Science*, 325(5944), 1089-1093.
- Fisher, C. R., Takai, K., & Le Bris, N. (2007). Hydrothermal vent ecosystems. *Oceanography*, 20(1), 14-23.
- Flajs, D., & Peraica, M. (2009). Toxicological properties of citrinin. *Archives of Industrial Hygiene and Toxicology*, 60(4), 457-464.
- Fleming, A. (1929). On the Antibacterial action of cultures of a *Penicillium*, with special reference to their use in the isolation of *B. influenzae*. *British Journal of Experimental Pathology*, 10, 226-236.
- Follmann, W., Behm, C., & Degen, G. H. (2014). Toxicity of the mycotoxin citrinin and its metabolite dihydrocitrinone and of mixtures of citrinin and ochratoxin A *in vitro*. *Archives of Toxicology*, 88(5), 1097-1107.
- Forbes B. A., Sahm D. F., & Weissfeld A. S. (2007). *Bailey & Scott's diagnostic microbiology*. (12th ed). St. Louis: Mosby Elsevier.
- Frandsen, G., & Pennington, S. S. (2013). The Conceptual Framework of Pharmacology: Basic Concepts and Processes. In *Abrams's Clinical Drug Therapy: Rationales for Nursing Practice* (pp. 16-34). Philadelphia: Lippincott Williams & Wilkins.
- Franklin, T.J., & Snow, G.A. (2006). Antimicrobial agents and cell membranes . In *Biochemistry and Molecular Biology of Antimicrobial Drug Action* (pp. 47-64). New York: Springer.
- Fujita, T., Inoue, K., Yamamoto, S., Ikumoto, T., Sasaki, S., Toyama, R., Chiba, K., Hoshino, Y., & Okumoto, T. (1994). Fungal metabolites. Part 11. A potent immunosuppressive activity found in *Isaria sinclairii* metabolite. *The Journal of Antibiotics*, 47(2), 208-215.
- Furtado, N. A. J. C., Said, S., Ito, I. Y., & Bastos, J. K. (2002). The antimicrobial activity of *Aspergillus fumigatus* is enhanced by a pool of bacteria. *Microbiological Research*, 157(3), 207-211.
- Galimand, M., Courvalin, P., & Lambert, T. (2003). Plasmid-mediated high-level resistance to aminoglycosides in *Enterobacteriaceae* due to 16S rRNA methylation. *Antimicrobial Agents and Chemotherapy*, 47(8), 2565-2571.

- Gallant-Behm, C. L., Yin, H. Q., Liu, S., Heggers, J. P., Langford, R. E., Olson, M. E., Hart, D. A., & Burrell, R. E. (2005). Comparison of in vitro disc diffusion and time kill-kinetic assays for the evaluation of antimicrobial wound dressing efficacy. *Wound Repair and Regeneration*, 13(4), 412-421.
- Gao, J., Radwan, M. M., Leon, F., Wang, X., Jacob, M. R., Tekwani, B. L., Khan, S. L., Lupien, S., Hill, R. A., Dugan, F. M., Cutler, H. G., & Cutler, S. J. (2012). Antimicrobial and antiprotozoal activities of secondary metabolites from the fungus *Eurotium repens*. *Medicinal Chemistry Research*, 21(10), 3080-3086.
- Garg, N., Tang, W., Goto, Y., Nair, S. K., & van der Donk, W. A. (2012). Lantibiotics from *Geobacillus thermodenitrificans*. *Proceedings of the National Academy of Sciences*, 109(14), 5241-5246.
- Gawlik-Dziki, U., Świeca, M., Sułkowski, M., Dziki, D., Baraniak, B., & Czyż, J. (2013). Antioxidant and anticancer activities of *Chenopodium quinoa* leaves extracts in vitro study. *Food and Chemical Toxicology*, 57, 154-160.
- Girfoglio, M., Rossi, M., & Cannio, R. (2012). Cellulose degradation by *Sulfolobus solfataricus* requires a cell-anchored endo- β -1-4-glucanase. *Journal of Bacteriology*, 194(18), 5091-5100.
- Giske, C. G., Monnet, D. L., Cars, O., & Carmeli, Y. (2008). Clinical and economic impact of common multidrug-resistant gram-negative bacilli. *Antimicrobial Agents and Chemotherapy*, 52(3), 813-821.
- Gloer, J. B. (2007). Applications of Fungal Ecology in the Search for New Bioactive Natural Products. In C.P. Kubicek and I.S. Druzhinina. *Mycota: Environmental and microbial relationships* (pp. 257- 286). New York: Springer.
- Gobbi, E., Falasconi, M., Zambotti, G., Sberveglieri, V., Pulvirenti, A., & Sberveglieri, G. (2015). Rapid diagnosis of *Enterobacteriaceae* in vegetable soups by a metal oxide sensor based electronic nose. *Sensors and Actuators B: Chemical*, 207, 1104-1113.
- Godowski, K. C. (1988). Antimicrobial action of sanguinarine. *The Journal of Clinical Dentistry*, 1(4), 96-101.
- Gold, N. D., Gowen, C. M., Lussier, F. X., Cautha, S. C., Mahadevan, R., & Martin, V. J. (2015). Metabolic engineering of a tyrosine-overproducing yeast platform using targeted metabolomics. *Microbial cell factories*, 14(1), 73-89.
- Gould, I. M., Wang, G. Q., Wu, J. J., Lim, V. K. E., Hutchinson, J., Walsh, T., & Turnidge, J. (2014). MDRO Beijing Consensus Meeting Report: Global burden of multidrug-resistant organisms' current antimicrobial resistance problems in Asia-Pacific. *Journal of Global Antimicrobial Resistance*, 2(1), 7-9.
- Grant, W. D., & Long, P. E. (2013). Extreme Environments. In *Environmental Microbiology* (pp. 51-67). London: Blackie.

- Grare, M., Mourer, M., Fontanay, S., Regnouf-de-Vains, J. B., Finance, C., & Duval, R. E. (2007). *In vitro* activity of para-guanidinoethylcalix [4] arene against susceptible and antibiotic-resistant Gram-negative and Gram-positive bacteria. *Journal of Antimicrobial Chemotherapy*, 60(3), 575-581.
- Griffith, F. (1928). The significance of pneumococcal types. *Journal of Hygiene*, 27(02), 113-159.
- Gröger, D., & Floss, H. G. (1997). Biochemistry of ergot alkaloids-achievements and challenges. *The Alkaloids: Chemistry and Biology*, 50, 171-218.
- Guo, J. P., Zhu, C. Y., Zhang, C. P., Chu, Y. S., Wang, Y. L., Zhang, J. X., Wu, D. K., Zhang, K. Q., & Niu, X. M. (2012). Thermolides, potent nematocidal PKS-NRPS hybrid metabolites from thermophilic fungus *Talaromyces thermophilus*. *Journal of the American Chemical Society*, 134(50), 20306-20309.
- Hadri, L. (2010). Development and application of an enzyme immunoassay for the detection of the mycotoxin fumigaclavine A, (Doctoral dissertation). Justus Liebig University Giessen, Germany.
- Han, J. H., Nachamkin, I., Zaoutis, T. E., Coffin, S. E., Linkin, D. R., Fishman, N. O., Weiner, M. G., Hu, B., Tolomeo, P., & Lautenbach, E. (2012). Risk factors for gastrointestinal tract colonization with Extended-Spectrum β -Lactamase (ESBL)-producing *Escherichia coli* and *Klebsiella* species in hospitalized patients. *Infection Control*, 33(12), 1242-1245.
- Han, P., Zhou, P., Hu, S., Yang, S., Yan, Q., & Jiang, Z. (2013). A novel multifunctional α -amylase from the thermophilic fungus *Malbranchea cinnamomea*: biochemical characterization and three-dimensional structure. *Applied Biochemistry and Biotechnology*, 170(2), 420-435.
- Haraguchi, H., Abo, T., Hashimoto, K., & Yagi, A. (1992). Action-mode of antimicrobial altersolanol A in *Pseudomonas aeruginosa*. *Bioscience, Biotechnology, and Biochemistry*, 56(8), 1221-1224.
- Harreither, W., Coman, V., Ludwig, R., Haltrich, D. & Gorton, L. (2007). Investigation of graphite electrodes modified with cellobiose dehydrogenase from the ascomycete *Myriococcum thermophilum*. *Electroanalysis*, 19(2-3), 172-180.
- Hashemi, S. R., Idrus, Z., Zakaria, Z., & Muhammad Nazrul, S. (2008). The effect of selected sterilization methods on antibacterial activity of aqueous extract of herbal plants. *Journal of Biological Sciences*, 8(6), 1072-1076.
- He, Z., Zhang, L., Mao, Y., Gu, J., Pan, Q., Zhou, S., Gao, B., & Wei, D. (2014). Cloning of a novel thermostable glucoamylase from thermophilic fungus *Rhizomucor pusillus* and high-level co-expression with α -amylase in *Pichia pastoris*. *BMC Biotechnology*, 14(1), 114-124.
- Hecker, M., & Völker, U. (2001). General stress response of *Bacillus subtilis* and other bacteria. *Advances in Microbial Physiology*, 44, 35-91.

- Hemila, H., Glode, L. M., & Palva, I. (1989). Production of diphtheria toxin CRM228 in *B. subtilis*. *FEMS Microbiology Letters*, 65(1-2), 193-198.
- Hengstler, J. G., Heimerdinger, C. K., Schiffer, I. B., Gebhard, S., Sagemuller, J., Tanner, B., Bolt, H. M., & Oesch, F. (2002). Dietary topoisomerase II-poisons: contribution of soy products to infant leukemia?. *EXCLI Journal*, 1, 8-14.
- Henry, T., Iwen, P. C., & Hinrichs, S. H. (2000). Identification of *Aspergillus* species using internal transcribed spacer regions 1 and 2. *Journal of Clinical Microbiology*, 38(4), 1510-1515.
- Heo, B. G., Park, Y. J., Park, Y. S., Bae, J. H., Cho, J. Y., Park, K., Jastrzebski, Z., & Gorinstein, S. (2014). Anticancer and antioxidant effects of extracts from different parts of indigo plant. *Industrial Crops and Products*, 56, 9-16.
- Hof, H. (2003). History and epidemiology of listeriosis. *FEMS Immunology & Medical Microbiology*, 35(3), 199-202.
- Hof, H., Nichterlein, T., & Kretschmar, M. (1997). Management of listeriosis. *Clinical Microbiology Reviews*, 10(2), 345-357.
- Höltje, J. V. (1998). Growth of the stress-bearing and shape-maintaining murein sacculus of *Escherichia coli*. *Microbiology and Molecular Biology Reviews*, 62(1), 181-203.
- Hooper, D. C. (2000). Mechanisms of action and resistance of older and newer fluoroquinolones. *Clinical Infectious Diseases*, 31(Supplement 2), S24-S28.
- Horn, W. S., Simmonds, M. S. J., Schwartz, R. E., & Blaney, W. M. (1995). Phomopsichalasin, a novel antimicrobial agent from an endophytic *Phomopsis* sp. *Tetrahedron*, 51, 3969-3978.
- Hylgaard, M., Mygind, T., & Meyer, R. L. (2012). Essential oils in food preservation: mode of action, synergies, and interactions with food matrix components. *Frontiers in Microbiology*, 12(3), 1-24.
- Iandolo, J. J., & Ordal, Z. J. (1966). Repair of thermal injury of *Staphylococcus aureus*. *Journal of Bacteriology*, 91(1), 134-142.
- IARC (International Agency for Research on Cancer) (1993) Some naturally occurring substances: heterocyclic aromatic amines and mycotoxins. *IARC Monograph on the Evaluation of Carcinogenic Risks to Human*, 56,397-444.
- Iglewski, B. H. (1996). *Pseudomonas*. In S. Baron. *Medical Microbiology*. 4th edition (Chapter 27). Galveston (TX): University of Texas Medical Branch at Galveston.
- Iossifidou, E. G., Abraham, A., Soultos, N. D., Triantafillou, E. A., & Koidis, P. A. (2012). Antimicrobial resistance profiles in *Salmonella* spp. and *Escherichia coli* isolates from turkey samples in Northern Greece. *Annals of Microbiology*, 62(2), 623-628.

- Jamal, S. (2015). Cellulytic activity of a thermophilic fungus *Aspergillus fumigatus* isolated from paper industry effluent. *Asian Journal of Bio Science*, 10(1), 15-20.
- Javed, M. M., Ikram, U. H., Irfana, M., & Farooq, L. (2011). Distribution of cellulolytic-thermophilic fungi on various substrates and geographic locations in Pakistan. *Pakistan Journal of Botany*, 43(5), 2621-2625.
- Jiang, L., Wang, F., Han, F., Prinyawiwatukul, W., No, H. K., & Ge, B. (2013). Evaluation of diffusion and dilution methods to determine the antimicrobial activity of water-soluble chitosan derivatives. *Journal of Applied Microbiology*, 114(4), 956-963.
- Johnson, J. R., Urban, C., Weissman, S. J., Jorgensen, J. H., Lewis, J. S., Hansen, G., Edelstein, P., Robicsek, A., Cleary, T., Adachi, J., Paterson, D., Lelans, K., Quinn, J., Hanson, N. D., Johnston, B. D., Clabots, C., Kuskowski, M. A., & AMERECUS Investigators. (2012). Molecular epidemiological analysis of *Escherichia coli* sequence Type ST131 (O25: H4) and blaCTX-M-15 among Extended-Spectrum β -Lactamase-producing *E. coli* from the United States (2000-2009). *Antimicrobial Agents and Chemotherapy*, AAC-05824(2), 1-35.
- Johri, B. N., Satyanarayana, T., & Olsen J. (2013). Physiology and Biochemistry of Thermophilic Moulds. In *Thermophilic Mould in Biotechnology* (pp. 85-113). Netherlands: Kluwer Academic Publishers.
- Josephine, H. R., Kumar, I., & Pratt, R. F. (2004). The perfect penicillin? Inhibition of a bacterial DD-peptidase by peptidoglycan-mimetic β -lactams. *Journal of the American Chemical Society*, 126(26), 8122-8123.
- Juayang, A. C., de los Reyes, G. B., de la Rama, A. J. G., & Gallega, C. T. (2014). Antibiotic Resistance Profiling of *Staphylococcus aureus* Isolated from Clinical Specimens in a Tertiary Hospital from 2010 to 2012. *Interdisciplinary Perspectives on Infectious Diseases*, 2014, 1-4.
- Karaman, I., Şahin, F., Güllüce, M., Ögütçü, H., Şengül, M., & Adıgüzel, A. (2003). Antimicrobial activity of aqueous and methanol extracts of *Juniperus oxycedrus* L. *Journal of Ethnopharmacology*, 85(2), 231-235.
- Keller, N. P., Turner, G., & Bennett, J. W. (2005). Fungal secondary metabolism—from biochemistry to genomics. *Nature Reviews Microbiology*, 3(12), 937-947.
- Kerzner, B., Corbelli, J., Sharp, S., Lipka, L. J., Melani, L., LeBeaut, A., Suresh, R., Mukhopadhyay, P., & Veltri, E.P. (2003). Efficacy and safety of Ezetimibe coadministered with lovastatin in primary hypercholesterolemia. *The American Journal of Cardiology*, 91(4), 418-424.
- Khodavandi, A., Alizadeh, F., Aala, F., Sekawi, Z., & Chong, P. P. (2010). *In vitro* investigation of antifungal activity of allicin alone and in combination with azoles against *Candida* species. *Mycopathologia*, 169(4), 287-295.

- Kiedrowski, M. R., & Horswill, A. R. (2011). New approaches for treating staphylococcal biofilm infections. *Annals of the New York Academy of Sciences*, 1241(1), 104-121.
- Kim, H. K., Missiakas, D., & Schneewind, O. (2014). Mouse models for infectious diseases caused by *Staphylococcus aureus*. *Journal of Immunological Methods*, 410, 88-99.
- Kirk, M. D., McKay, I., Hall, G. V., Dalton, C. B., Stafford, R., Unicomb, L., & Gregory, J., (2008). Foodborne disease in Australia: The OzFoodNet experience. *Clinical Infectious Diseases*, 47(3), 392-400.
- Klein, E. Y., Sun, L., Smith, D. L., & Laxminarayan, R. (2013). The changing epidemiology of methicillin-resistant *Staphylococcus aureus* in the United States: a national observational study. *American Journal of Epidemiology*, kws273, 1-9.
- Koizumi, Y., Arai, M., Tomoda, H., & Omura, S. (2004). Fungerin, a fungal alkaloid, arrests the cell cycle in M phase by inhibition of microtubule polymerization. *The Journal of Antibiotics*, 57(7), 415-420.
- Kojic, E. M., & Darouiche, R. O. (2004). *Candida* infections of medical devices. *Clinical Microbiology Reviews*, 17(2), 255-267.
- Kontoyiannis, D., & Bodey, G. (2002). Invasive aspergillosis in 2002: an update. *European Journal of Clinical Microbiology and Infectious Diseases*, 21(3), 161-172.
- Kor, S. B., Choo, Q. C., & Chew, C. H. (2013). New integron gene arrays from multiresistant clinical isolates of members of the *Enterobacteriaceae* and *Pseudomonas aeruginosa* from hospitals in Malaysia. *Journal of Medical Microbiology*, 62(Pt 3), 412-420.
- Korniłowicz-Kowalska, T., & Kitowski, I. (2013). *Aspergillus fumigatus* and other thermophilic fungi in nests of wetland birds. *Mycopathologia*, 175(1-2), 43-56.
- Krishnaveni, R., Rathod, V., Thakur, M. S., & Neelgund, Y. F. (2009). Transformation of L-tyrosine to L-DOPA by a novel fungus, *Acremonium rutilum*, under submerged fermentation. *Current Microbiology*, 58(2), 122-128.
- Kuan, C. H., Goh, S. G., Loo, Y. Y., Chang, W. S., Lye, Y. L., Puspanadan, S., Shahril, N., Tang, J. Y. H., Mahyudin, N. A., Nishibuchi, M., Son, R., & Nakaguchi, Y. (2015). Risk assessment of acquiring listeriosis from consumption of chicken offal in Selangor, Malaysia. *International Food Research Journal*, 22(4), 1711-1718.
- Kumar, A., Robert Antony, A., & Kannan, V. R. (2015). Exploration of endophytic microorganisms from selected medicinal plants and their control potential to multi drug resistant pathogens. *Journal of Medicinal Plants*, 3(2), 49-57.

- Kumar, S., Kaushik, N., & Proksch, P. (2013). Identification of antifungal principle in the solvent extract of an endophytic fungus *Chaetomium globosum* from *Withania somnifera*. *SpringerPlus*, 2(1), 37-47.
- Kusari, S., Hertweck, C., & Spiteller, M. (2012). Chemical ecology of endophytic fungi: origins of secondary metabolites. *Chemistry & Biology*, 19(7), 792-798.
- Kwon, H. C., Kauffman, C. A., Jensen, P. R., & Fenical, W. (2006). Marinomycins AD, antitumor-antibiotics of a new structure class from a marine actinomycete of the recently discovered genus "*Marinispora*". *Journal of the American Chemical Society*, 128(5), 1622-1632.
- Lam, K.S. (2007). New aspects of natural products in drug discovery. *Trends in Microbiology*, 15(6), 279-289.
- Landry, K. S. (2012). *Purification and Characterization of Novel Nucleases from a Thermophilic Fungus* (Doctoral dissertation). University of Massachusetts Amherst, United States.
- Langarica-Fuentes, A., Handley, P. S., Houlden, A., Fox, G., & Robson, G. D. (2014). An investigation of the biodiversity of thermophilic and thermotolerant fungal species in composts using culture-based and molecular techniques. *Fungal Ecology*, 11, 132-144.
- Latge, J. P. (1999). *Aspergillus fumigatus* and aspergillosis. *Clinical Microbiology Reviews*, 12(2), 310-350.
- Lee, J. A., & Chee, H. Y. (2010). *In vitro* antifungal activity of equol against *Candida albicans*. *Mycobiology*, 38(4), 328-330.
- Leekha, S., Terrell, C. L., & Edson, R. S. (2011). General principles of antimicrobial therapy. *Mayo Clinic Proceedings*, 86(2), 156-167.
- Leite, M. C. A., Bezerra, A. P. D. B., Sousa, J. P. D., Guerra, F. Q. S., & Lima, E. D. O. (2014). Evaluation of antifungal activity and mechanism of action of citral against *Candida albicans*. *Evidence-Based Complementary and Alternative Medicine*, 2014, 1-9.
- Levy, S.B., & Marshall, B. (2004). Antibacterial resistance worldwide: causes, challenges and responses. *Nature Medicine*, 10(2), 122-129.
- Li, D.C., Li, A.N., & Papageorgiou, A.C. (2011). Cellulases from thermophilic fungi: Recent insights and biotechnological potential. *Enzyme Research*. 2011, 1-9.
- Li, S., Su, X., Le, W., Jiang, F., Wang, B., & Rice, P. A. (2014). Antimicrobial susceptibility of *Neisseria gonorrhoeae* isolates from symptomatic men attending the Nanjing sexually transmitted diseases clinic (2011–2012): genetic characteristics of isolates with reduced sensitivity to ceftriaxone. *BMC Infectious Diseases*, 14, 622-632.

- Liapikou, A., Cilloniz, C., Mensa, J., & Torres, A. (2014). New antimicrobial approaches to Gram-positive respiratory infections. *Pulmonary Pharmacology & Therapeutics*, 1-7.
- Limsuwan, S., Kayser, O., & Voravuthikunchai, S. P. (2012). Antibacterial activity of *Rhodomyrtus tomentosa* (Aiton) Hassk. leaf extract against clinical isolates of *Streptococcus pyogenes*. *Evidence-Based Complementary and Alternative Medicine*, 2012, 1-6.
- Liu, L. F., Liu, C. C., & Alberts, B. M. (1980). Type II DNA topoisomerases: enzymes that can unknot a topologically knotted DNA molecule via a reversible double-strand break. *Cell*, 19(3), 697-707.
- Lloyd, D. H. (2007). Reservoirs of antimicrobial resistance in pet animals. *Clinical Infectious Diseases*, 45(Supplement 2), 148-152.
- Lowy, F. D. (1998). *Staphylococcus aureus* infections. *New England Journal of Medicine*, 339(8), 520-532.
- Lu, H., Zhang, H., Shi, P., Luo, H., Wang, Y., Yang, P., & Yao, B. (2013). A family 5 β -mannanase from the thermophilic fungus *Thielavia arenaria* XZ7 with typical thermophilic enzyme features. *Applied Microbiology and Biotechnology*, 97(18), 8121-8128.
- Lunde, C. S., Hartouni, S. R., Janc, J. W., Mammen, M., Humphrey, P. P., & Benton, B. M. (2009). Telavancin disrupts the functional integrity of the bacterial membrane through targeted interaction with the cell wall precursor lipid II. *Antimicrobial Agents and Chemotherapy*, 53(8), 3375-3383.
- Luzzatto, L., & Karadimitris, A. (2010). The molecular basis of anemia. In D. Provan. and J.G. Griben. *Molecular Hematology* (pp.140-164). USA: Wiley-Blackwell.
- Maalej, I., Belhaj, I., Masmoudi, N. F. & Belghith, H. (2009). Highly thermostable xylanase of the thermophilic fungus *Talaromyces thermophilus*: purification and characterization. *Applied Biochemistry and Biotechnology*, 158(1), 200-212.
- Macchione, M. M., Merheb, C. W., Gomes, E. & da Silva, R. (2008). Protease production by different thermophilic fungi. *Applied Biochemistry and Biotechnology*, 146, 223-230.
- Magda, M. A., El-Nour, M. E. M., Hassan, A. A. E. L., & Ezzdeen, L. T. (2015). Antibacterial activities of seeds, leaves and callus (hypocotyls and cotyledons) extracts of *Jatropha curcas* L. *International Journal of Biosciences (IJB)*, 6(11), 58-63.
- Magner, L. N. (2009). *A History of Infectious Disease and the Microbial World*. USA: Westport, Conn. Praeger.
- Mahesh, B., & Satish, S. (2008). Antimicrobial activity of some important medicinal plant against plant and human pathogens. *World Journal of Agricultural Sciences*, 4(5), 839-843.

- Maheshwari, R., Bharadwaj, G., & Bhat, M. K. (2000). Thermophilic fungi: their physiology and enzymes. *Microbiology and Molecular Biology Reviews*, 64(3), 461-488.
- Maheshwari, R., Kamalam, P. T., & Balasubramanyam, P. V. (1987). The biogeography of thermophilic fungi. *Current Science. Bangalore*, 56(4), 151-154.
- Mahoney, J. F., Arnold, R. C., & Harris, A. D. (1943). Penicillin Treatment of Early Syphilis—A Preliminary Report*. *American Journal of Public Health and the Nations Health*, 33(12), 1387-1391.
- Maier, R. M., Pepper, I. L., & Gerba, C. P. (2000). Environmental Determinants that Govern Extreme Environments. In *Environmental Microbiology* (pp. 141-142). San Diego: Academic Press.
- Maitra, A., & Dill, K. A. (2016). Modeling the overproduction of ribosomes when antibacterial drugs act on cells. *Biophysical Journal*, 110(3), 743-748.
- Margesin, R., & Schinner, F. (1994). Properties of cold-adapted microorganisms and their potential role in biotechnology. *Journal of Biotechnology*, 33(1), 1-14.
- Martirosyan, A., Clendening, J. W., Goard, C. A., & Penn, L. Z. (2010). Lovastatin induces apoptosis of ovarian cancer cells and synergizes with doxorubicin: potential therapeutic relevance. *BMC Cancer*. 10(1), 103-116.
- Masurekar, P. S. (2005). Strain improvement for the production of fungal secondary metabolites. In Z. An. *Handbook of Industrial Mycology* (pp. 539-561). New York: Marcel Dekker.
- Matamouros, S., & Miller, S. I. (2015). *Salmonella typhimurium* strategies to resist killing by cationic antimicrobial peptides. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1848(11), 3021-3025.
- Mathavan, I., & Beis, K. (2012). The role of bacterial membrane proteins in the internalization of microcin MccJ25 and MccB17. *Biochemical Society Transactions*, 40(6), 1539-1543.
- Mathé, L., & Van Dijck, P. (2013). Recent insights into *Candida albicans* biofilm resistance mechanisms. *Current Genetics*, 59(4), 251-264.
- Mathur, S., & Singh, R. (2005). Antibiotic resistance in food lactic acid bacteria—a review. *International Journal of Food Microbiology*, 105(3), 281-295.
- Mayer, F. L., Wilson, D., & Hube, B. (2013). *Candida albicans* pathogenicity mechanisms. *Virulence*, 4(2), 119-128.
- Mazaheri Nezhad Fard, R., Barton, M. D., & Heuzenroeder, M.W. (2011). Bacteriophage-mediated transduction of antibiotic resistance in enterococci. *Letters in Applied Microbiology*, 52(6), 559-564.

- Mead, P. S., Dunne, E. F., Graves, L., Wiedmann, M., Patrick, M., Hunter, S, Salehi, E., Fmostashari, F., Craig, A., Mshar, P., Bannerman, T., Sauders, B. D., Hayes, P., DeWitt, W., Sparling, P., Griffin, P., Morse, D., Slutsker, L., & Swaminathan, B. (2006). Nationwide outbreak of listeriosis due to contaminated meat. *Epidemiology and Infection*, 134(4), 744-751.
- Mertas, A., Garbusińska, A., Szliszka, E., Jureczko, A., Kowalska, M., & Król, W. (2015). The influence of tea tree oil (*Melaleuca alternifolia*) on fluconazole activity against fluconazole-resistant *Candida albicans* strains. *BioMed Research International*, 2015, 1-9.
- Mishra, S., Sahai, V., Bisaria, V. S., Biswas, R., Gupta, G., & Nakra, S. (2013). Xylanases from Thermophilic Fungi: Classification, Structure, and Case Study of *Melanocarpus albomyces*. In T. Satyanarayana, J. Littlechild, and Y. Kawarabayasi. *Thermophilic Microbes in Environmental and Industrial Biotechnology* (pp. 795-811). Netherlands: Springer.
- Mohamad, R., Mohamed, M. S., Suhaili, N., Salleh, M. M., & Ariff, A. B. (2010). Kojic acid: Applications and development of fermentation process for production. *Biotechnology and Molecular Biology Reviews*, 5(2), 24-37.
- Montoya, C. J., Jaimes, F., Higueta, E. A., Convers-Páez, S., Estrada, S., Gutierrez, F., Amariles, P., Giraldo, N., Penaloza, C., & Rugeles, M.T. (2009). Antiretroviral effect of lovastatin on HIV-1-infected individuals without highly active antiretroviral therapy (The LIVE study): a phase-II randomized clinical trial. *Trials*, 10(1), 41-52.
- Moran, G. J., Fang, E., Corey, G. R., Das, A. F., De Anda, C., & Prokocimer, P. (2014). Tedizolid for 6 days versus linezolid for 10 days for acute bacterial skin and skin-structure infections (ESTABLISH-2): a randomised, double-blind, phase 3, non-inferiority trial. *The Lancet Infectious Diseases*, 14(8), 696-705.
- Moretti, M., Bocchini-Martins, D. A., Silva, R. D., Rodrigues, A., Sette, L. D., & Gomes, E. (2012). Selection of thermophilic and thermotolerant fungi for the production of cellulases and xylanases under solid-state fermentation. *Brazilian Journal of Microbiology*. 43(3), 1062-1071.
- Morris, R. L., & Schmidt, T. M. (2013). Shallow breathing: bacterial life at low O₂. *Nature Reviews Microbiology*, 11(3), 205-212.
- Morvan, A., Moubareck, C., Leclercq, A., Hervé-Bazin, M., Bremont, S., Lecuit, M., Courvalin, P., & Le Monnier, A. (2010). Antimicrobial resistance of *Listeria monocytogenes* strains isolated from humans in France. *Antimicrobial Agents and Chemotherapy*, 54(6), 2728-2731.
- Moubasher, A. H., Abdel-Hafez, S. I. I. & El-Maghraby, O. M. O. (1988). Seasonal fluctuations of soil-borne and air-borne fungi of Wadi Bir-El-Ain in the Eastern Desert of Egypt. *Naturalia Monspeliensa, SeAr. Botanique*, 52, 57-69.

- Mouchacca, J. (2000). Thermotolerant fungi erroneously reported in applied research work as possessing thermophilic attributes. *World Journal of Microbiology and Biotechnology*, 16(8-9), 869-880.
- Muniesa, M., Colomer-Lluch, M., & Jofre, J. (2013). Could bacteriophages transfer antibiotic resistance genes from environmental bacteria to human-body associated bacterial populations?. *Mobile Genetic Elements*, 3(4), 1-4.
- Munita, J. M., Bayer, A. S., & Arias, C. A. (2015). Evolving resistance among gram-positive pathogens. *Clinical Infectious Diseases*, 61(Supplement 2), 48-57.
- Namuli, A., Abdullah, N., Sieo, C. C., Zuhainis, S. W., & Oskoueian, E. (2011). Phytochemical compounds and antibacterial activity of *Jatropha curcas* Linn. extracts. *Journal of Medicinal Plant Research*, 5, 3982-3990.
- Nath, A., & Joshi, S. R. (2015). Ultrastructural effect on mastitis pathogens by extract of endophytic fungi associated with ethnoveterinary plant, *Hibiscus sabdariffa* L. *Journal of Microscopy and Ultrastructure*, 3(1), 38-43.
- Navarre, W. W., & Schneewind, O. (1999). Surface proteins of Gram-positive bacteria and mechanisms of their targeting to the cell wall envelope. *Microbiology and Molecular Biology Reviews*, 63(1), 174-229.
- Neu, H. C. (1992). The Crisis in Antibiotic Resistance. *Science*, 257(5073), 1064-1073.
- Nie, H. X., Ding, X. H., Huang, Y., & Hu, S.P. (2011). Adult-onset still's disease misdiagnosed as pneumonia: two case reports. *Acta Reumatologica Portuguesa*, 36(4), 413-417.
- Nordmann, P., Cuzon, G., & Naas, T. (2009). The real threat of *Klebsiella pneumoniae* carbapenemase-producing bacteria. *The Lancet Infectious Diseases*, 9(4), 228-236.
- Norris, J.R., R.C.W. Berkeley, N.A. Logan, and A.G. O'Donnell. (1981). The genera *Bacillus* and *Sporolactobacillus*, In M. P. Starr, H. Stolp, H. G. Truper, A. Alows and H. G. Schlegel. *The Prokaryotes: A Handbook on Habitats, Isolation, and Identification of Bacteria*, Vol. 2. (pp. 1711-1742). Berlin: Springer-Verlag.
- Numata, T., Kobayashi, Y., Ito, T., Harada, K., Tsuboi, R., & Okubo, Y. (2015). Two cases of allergic contact dermatitis due to skin-whitening cosmetics. *Allergology International*, 64(2), 194-195.
- Nwagu, T. N. (2011). Growth profile and amylase hydrolytic activity of a thermophilic fungi *Aspergillus fumigatus* isolated from soil TN Nwagu and BN Okolo. *Asian Journal of Biotechnology*, 3(1), 46-57.
- O'Brien, J., & Wright, G. D. (2011). An ecological perspective of microbial secondary metabolism. *Current Opinion in Biotechnology*, 22(4), 552-558.

- Obeidat, M. (2011). Antimicrobial activity of some medicinal plants against multidrug resistant skin pathogens. *Journal of Medicinal Plants Research*, 5(16), 3856-3860.
- Okeke, I. N., Fayinka, S. T., & Lamikanra, A. (2000). Antibiotic resistance in *Escherichia coli* from Nigerian students, 1986-1998. *Emerging infectious diseases*, 6(4), 393-396.
- Oonmetta-Aree, J., Suzuki, T., Gasaluck, P., & Eumkeb, G. (2006). Antimicrobial properties and action of galangal (*Alpinia galanga* Linn.) on *Staphylococcus aureus*. *LWT-Food Science and Technology*, 39(10), 1214-1220.
- Osheroov, N. (2007). The virulence of *Aspergillus fumigatus*. In *New insights in medical mycology* (pp. 185-212). Springer Netherlands.
- Oshima, T., & Imahori, K. (1974). Physicochemical properties of deoxyribonucleic acid from an extreme thermophile. *Journal of Biochemistry*, 75(1), 179-183.
- Oskoueian, E., Abdullah, N., Ahmad, S., Saad, W. Z., Omar, A. R., & Ho, Y. W. (2011). Bioactive compounds and biological activities of *Jatropha curcas* L. kernel meal extract. *International Journal of Molecular Sciences*, 12(9), 5955-5970.
- Painter J., & Slutsker L. (2007). Listeriosis in humans .In E. T. Ryser and E. H. Marth (ed.), *Listeria, listeriosis and food safety*, 3rd edition (pp. 85–109). Boca Raton, Florida: CRC Press.
- Pan, W. Z., Huang, X. W., Wei, K. B., Zhang, C. M., Yang, D. M., Ding, J. M., & Zhang, K. Q. (2010). Diversity of thermophilic fungi in Tengchong Rehai National Park revealed by ITS nucleotide sequence analyses. *The Journal of Microbiology*, 48(2), 146-152.
- Para, G. M., & Baratti, J. C. (1984). Effect of culture conditions on the production of tyrosine phenol-lyase by *Erwinia herbicola*. *Applied and Environmental Microbiology*, 48(6), 1256-1258.
- Park, J. T., & Uehara, T. (2008). How bacteria consume their own exoskeletons (turnover and recycling of cell wall peptidoglycan). *Microbiology and Molecular Biology Reviews*, 72(2), 211-227.
- Park, S. R., Park, J. W., Ban, Y. H., Sohng, J. K., & Yoon, Y. J. (2013). 2-Deoxystreptamine-containing aminoglycoside antibiotics: recent advances in the characterization and manipulation of their biosynthetic pathways. *Natural Product Reports*, 30(1), 11-20.
- Pathmanathan, S. G., Samat, N. A., & Mohamed, R. (2009). Antimicrobial susceptibility of clinical isolates of *Pseudomonas aeruginosa* from a Malaysian hospital. *Malaysian Journal of Medical Sciences*, 16(2), 27-32.

- Peng, J., Lin, T., Wang, W., Xin, Z., Zhu, T., Gu, Q., & Li, D. (2013). Antiviral alkaloids produced by the mangrove-derived fungus *Cladosporium* sp. PJX-41. *Journal of Natural Products*, 76, 1133-1140.
- Perry, J. A., & Wright, G. D. (2013). The antibiotic resistance “mobilome”: searching for the link between environment and clinic. *Frontiers in Microbiology*, 138(4), 1-4.
- Petkova-Bocharova, T., Castegnaro, M., Michelon, J., & Maru, V. (1990). Ochratoxin A and other mycotoxins in cereals from an area of Balkan endemic nephropathy and urinary tract tumours in Bulgaria. *IARC Scientific Publications*, (115), 83-87.
- Pommerville, J. C. (2010). Disease and Resistance: Antimicrobial Drug. In *Alcamo's Fundamentals of Microbiology* (pp. 804-842). US: Jones and Bartlett Publishers.
- Priori, R., Colafrancesco, S., Gattamelata, A., Di Franco, M., Di Tondo, U., & Valesini, G. (2010). Adult-onset Still's disease: a rare disorder with a potentially fatal outcome. *Autoimmunity Highlights*, 1(1), 53-59.
- Pui, C. F., Wong, W. C., Chai, L. C., Nillian, E., Ghazali, F. M., Cheah, Y. K., Nakaguchi, Y., Nishibuchi, M., & Radu, S. (2011). Simultaneous detection of *Salmonella* spp., *Salmonella Typhi* and *Salmonella Typhimurium* in sliced fruits using multiplex PCR. *Food Control*, 22(2), 337-342.
- Raja, N. S., & Singh, N. N. (2007). Antimicrobial susceptibility pattern of clinical isolates of *Pseudomonas aeruginosa* in a tertiary care hospital. *Journal of Microbiology, Immunology, and Infection*, 40(1), 45-49.
- Ramage, G., Martínez, J. P., & López-Ribot, J. L. (2006). *Candida* biofilms on implanted biomaterials: a clinically significant problem. *FEMS Yeast Research*, 6(7), 979-986.
- Rapala-Kozik, M., Bochenska, O., Zawrotniak, M., Wolak, N., Trebacz, G., Gogol, M., Ostrowska, D., Aoki, W., Ueda, M., & Kozik, A. (2015). Inactivation of the antifungal and immunomodulatory properties of human cathelicidin LL-37 by aspartic proteases produced by the pathogenic yeast *Candida albicans*. *Infection and Immunity*, 83(6), 2518-2530.
- Rementeria, A., López-Molina, N., Ludwig, A., Vivanco, A. B., Bikandi, J., Pontón, J., & Garaizar, J. (2005). Genes and molecules involved in *Aspergillus fumigatus* virulence. *Revista Iberoamericana de Micología*, 22(1), 1-23.
- Rode, L. J., Foster, J. W., & Schuhardt, V. T. (1947). Penicillin production by a thermophilic fungus. *Journal of Bacteriology*, 53(5), 565-566.
- Romanova, N. A., Wolffs, P. F. G., Brovko, L. Y., & Griffiths, M. W. (2006). Role of efflux pumps in adaptation and resistance of *Listeria monocytogenes* to benzalkonium chloride. *Applied and Environmental Microbiology*, 72(5), 3498-3503.

- Sahoo, N. R., Kumar, P., Bhusan, B., Bhattacharya, T. K., Dayal, S., & Sahoo, M. (2012). Lysozyme in livestock: A guide to selection for disease resistance: A review. *Journal of Animal Science Advances*, 2(4), 347-360.
- Salar, R. K., & Aneja, K. R. (2007). Thermophilic fungi: taxonomy and biogeography. *Journal of Agricultural Technology*, 3(1), 77-107.
- Salman, M. T., Khan, R. A., & Shukla, I. (2008). Antimicrobial activity of *Nigella sativa* Linn. seed oil against multi-drug resistant bacteria from clinical isolates. *Natural Product Radiance*, 7(1), 10-14.
- Sampathkumar, B., Khachatourians, G. G., & Korber, D. R. (2003). High pH during trisodium phosphate treatment causes membrane damage and destruction of *Salmonella enterica* serovar *Enteritidis*. *Applied and Environmental Microbiology*, 69(1), 122-129.
- Saris, P. E., Airaksinen, U., Nurmiharju, S., Runeberg-Nyman, K., & Palva, I. (1990). Expression of *Bordetella pertussis* toxin subunits in *Bacillus subtilis*. *Biotechnology Letters*, 12(12), 873-878.
- Sarker, S. D., Nahar, L., & Kumarasamy, Y. (2007). Microtitre plate-based antibacterial assay incorporating resazurin as an indicator of cell growth, and its application in the *in vitro* antibacterial screening of phytochemicals. *Methods*, 42(4), 321-324.
- Scallan, E., Hoekstra, R. M., Angulo, F. J., Tauxe, R. V., Widdowson, M. A., Roy, S. L., Jones, J. L., & Griffin, P. M. (2011). Foodborne illness acquired in the United States—major pathogens. *Emerging Infectious Disease Journal*, 17(1), 7-15.
- Schneider, T., Kruse, T., Wimmer, R., Wiedemann, I., Sass, V., Pag, U., Jansen, A., Nielsen, A.K., Mygind, P.H., Raventos, D.S., Neve, S., Ravn, B., Bonvin, A. M. J. J., Maria, L.D., Andersen, A. S., Gammelgaard, L.K., Sahl, H. G., & Kristensen, H. H. (2010). Plectasin, a fungal defensin, targets the bacterial cell wall precursor Lipid II. *Science*, 328(5982), 1168-1172.
- Schoch, C. L., Seifert, K. A., Huhndorf, S., Robert, V., Spouge, J. L., Levesque, C. A., & Wen, C. (2012). Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for fungi. *Proceedings of the National Academy of Sciences*, 109(16), 6241-6246.
- Schuemann, J., & Hertweck, C. (2009). Biosynthesis of Fungal Polyketides. In T. Anke and D. Weber. *Physiology and Genetics* (pp. 331-351). New York: Springer Berlin Heidelberg.
- Schwarzer, D., Finking, R., & Marahiel, M. A. (2003). Nonribosomal peptides: from genes to products. *Natural Product Reports*, 20(3), 275-287.

- Seasotiya, L., Siwach, P., Bharti, P., Bai, S., Malik, A., Kumar, R., & Dalal, S. (2015). A cross sectional study on prevalence of antibiotic resistance and role of efflux pumps in fluoroquinolone resistance by using efflux pump inhibitors in isolated cultures from poultry, dairy Farms and MTCC Strains from reservoirs. *British Microbiology Research Journal*, 5(2), 107-116.
- Sebők, F., Dobolyi, C., Bobvos, J., Szoboszlay, S., Kriszt, B., & Magyar, D. (2016). Thermophilic fungi in air samples in surroundings of compost piles of municipal, agricultural and horticultural origin. *Aerobiologia*, 32(2), 255-263.
- Seitz, P., & Blokesch, M. (2013). Cues and regulatory pathways involved in natural competence and transformation in pathogenic and environmental Gram-negative bacteria. *FEMS Microbiology Reviews*, 37(3), 336-363.
- Senthilraja, P., & Kathiresan, K. (2015). In vitro cytotoxicity MTT assay in Vero, HepG2 and MCF-7 cell lines study of Marine Yeast. *Journal of Applied Pharmaceutical Science*, 5, 80-84
- Şenyuva, H. Z., Gilbert, J., & Öztürkoğlu, Ş. (2008). Rapid analysis of fungal cultures and dried figs for secondary metabolites by LC/TOF-MS. *Analytica Chimica Acta*, 617(1), 97-106.
- Shanthi, M., & Sekar, U. (2009). Multi-drug resistant *Pseudomonas aeruginosa* and *Acinetobacter baumannii* infections among hospitalized patients: risk factors and outcomes. *The Journal of the Association of Physicians of India*, 57, 636-638.
- Sharma, H. S. (1989). Economic importance of thermophilous fungi. *Applied Microbiology and Biotechnology*, 31(1), 1-10.
- Shelton, B. G., Kirkland, K. H., Flanders, W. D., & Morris, G. K. (2002). Profiles of airborne fungi in buildings and outdoor environments in the United States. *Applied and Environmental Microbiology*, 68(4), 1743-1753.
- Singh, M. P., Janso, J. E., Luckman, S. W., Brady, S. F., Clardy, J., Greenstein, M., & Maiese, W. M. (2000). Biological activity of guanacastepene, a novel diterpenoid antibiotic produced by an unidentified fungus CR115. *The Journal of Antibiotics*, 53(3), 256-261.
- Sneader, W. E. (2005). *Drug Discovery (The History)*. New York: John Wiley & Sons.
- Srivastava, S., Pathak, N., & Srivastava, P. (2011). Identification of limiting factors for the optimum growth of *Fusarium oxysporum* in liquid medium. *Toxicology International*, 18(2), 111-116.
- Stolp, H. (1988). The Natural Environments of Microorganisms. In *Microbial Ecology: Organisms, Habitats, Activities* (pp. 140-142). USA: Cambridge University Press.
- Strugnell, R. A., Scott, T. A., Wang, N., Yang, C., Peres, N., Bedoui, S., & Kupz, A. (2014). *Salmonella* vaccines: lessons from the mouse model or bad teaching?. *Current Opinion in Microbiology*, 17, 99-105.

- Su, Y. Y., & Cai, L. (2013). *Rasamsonia composticola*, a new thermophilic species isolated from compost in Yunnan, China. *Mycological Progress*, 12(2), 213-221.
- Subban, K., Subramani, R., & Johnpaul, M. (2013). A novel antibacterial and antifungal phenolic compound from the endophytic fungus *Pestalotiopsis mangiferae*. *Natural Product Research*, 27(16), 1445-1449.
- Sugita, T., Nakajima, M., Ikeda, R., Matsushima, T., & Shinoda, T. (2002). Sequence analysis of the ribosomal DNA intergenic spacer 1 regions of *Trichosporon* species. *Journal of Clinical Microbiology*, 40(5), 1826-1830.
- Svahn, K.S., Goransson, U., El-Seedi, H., Bohlin, L., Larsson, D.G.J., Olsen, B., & Chryssanthou, E. (2012). Antimicrobial activity of filamentous fungi isolated from highly antibiotic-contaminated river sediment. *Infection Ecology and Epidemiology*, 2, 1-6.
- Szweda, P., Schielmann, M., Kotlowski, R., Gorczyca, G., Zalewska, M., & Milewski, S. (2012). Peptidoglycan hydrolases-potential weapons against *Staphylococcus aureus*. *Applied Microbiology and Biotechnology*, 96(5), 1157-1174.
- Taha, M., Adetutu, E. M., Shahsavari, E., Smith, A. T., & Ball, A. S. (2014). Azo and anthraquinone dye mixture decolourization at elevated temperature and concentration by a newly isolated thermophilic fungus, *Thermomucor indicac-seudaticae*. *Journal of Environmental Chemical Engineering*, 2(1), 415-423.
- Taira, S., Jalonen, E., Paton, J. C., Sarvas, M., & Runeberg-Nyman, K. (1989). Production of pneumolysin, a pneumococcal toxin, in *Bacillus subtilis*. *Gene*, 77(2), 211-218.
- Tan, L. T. (2007). Bioactive natural products from marine cyanobacteria for drug discovery. *Phytochemistry*, 68(7), 954-979.
- Tanwar, J., Das, S., Fatima, Z., & Hameed, S. (2014). Multidrug resistance: an emerging crisis. *Interdisciplinary Perspectives on Infectious Diseases*, 2014, 1-7.
- Taylor, D.L. (2008). "Internal transcribed spacer primers and sequences for improved characterization of basidiomycetous orchid mycorrhizas". *New Phytologist*, 177, 1020-1033.
- Tekaia, F., & Latgé, J. P. (2005). *Aspergillus fumigatus*: saprophyte or pathogen?. *Current Opinion in Microbiology*, 8(4), 385-392.
- Tenover, F. C. (2006). Mechanisms of antimicrobial resistance in bacteria. *The American Journal of Medicine*, 119(6), 3-10.
- Teuber, M. (2001). Veterinary use and antibiotic resistance. *Current Opinion in Microbiology*, 4(5), 493-499.

- Thomas, A. E., Coker, H. A. B., Odukoya, O. A., Isamah, G. K., & Adepoju-Bello, A. (2003). Aflatoxin contamination of *Arachis hypogaea* (groundnuts) in Lagos area of Nigeria. *Bulletin of Environmental Contamination and Toxicology*, 71(1), 42-45.
- Thomas, C. M., & Nielsen, K. M. (2005). Mechanisms of, and barriers to, horizontal gene transfer between bacteria. *Nature Reviews Microbiology*, 3(9), 711-721.
- To, M. S., Favrin, S., Romanova, N., & Griffiths, M. W. (2002). Postadaptational resistance to benzalkonium chloride and subsequent physicochemical modifications of *Listeria monocytogenes*. *Applied and Environmental Microbiology*, 68(11), 5258-5264.
- Tobudic, S., Kratzer, C., Lassnigg, A., & Presterl, E. (2012). Antifungal susceptibility of *Candida albicans* in biofilms. *Mycoses*, 55(3), 199-204.
- Tomasz, A. (1979). The mechanism of the irreversible antimicrobial effects of penicillins: how the beta-lactam antibiotics kill and lyse bacteria. *Annual Reviews in Microbiology*, 33(1), 113-137.
- Trevor, A. J., Katzung, B. G., Masters, S. B., & Kruierring-Hall, M. (2010). Pharmacology Examination & Board Review. New York: McGraw-Hill Medical.
- Trnovsky, J., Merz, W., Della-Latta, P., Wu, F., Arendrup, M. C., & Stender, H. (2008). Rapid and accurate identification of *Candida albicans* isolates by use of PNA FISHFlow. *Journal of Clinical Microbiology*, 46(4), 1537-1540.
- Tsuji, A., Kobayashi, I., Oguri, T., Inoue, M., Yabuuchi, E., & Goto, S. (2005). An epidemiological study of the susceptibility and frequency of multiple-drug-resistant strains of *Pseudomonas aeruginosa* isolated at medical institutes nationwide in Japan. *Journal of Infection and Chemotherapy*, 11(2), 64-70.
- Turner, P.J. (2005). Extended-Spectrum B-Lactamases. *Clinical Infectious Diseases*, 41 (Supplement 4), 273-275.
- van den Bogaard, A. E., & Stobberingh, E. E. (2000). Epidemiology of resistance to antibiotics: links between animals and humans. *International Journal of Antimicrobial Agents*. 14(4), 327-335.
- van den Brink, J., van Muiswinkel, G. C., Theelen, B., Hinz, S. W., & de Vries, R. P. (2013). Efficient plant biomass degradation by thermophilic fungus *Myceliophthora heterothallica*. *Applied and Environmental Microbiology*, 79(4), 1316-1324.
- van der Klei, I. J., & Veenhuis, M. (2006). Yeast and filamentous fungi as model organisms in microbody research. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research*, 1763(12), 1364-1373.
- Vaquero, M. J. R., Alberto, M. R., & Nadra, M. C. M. (2007). Antibacterial effect of phenolic compounds from different wines. *Food Control*, 18, 93-101.

- Varga, M., Kuntova, L., Pantucek, R., Maslanova, I., Ruzickova, V., & Doskar, J. (2012). Efficient transfer of antibiotic resistance plasmid by transduction within methicillin-resistance *Staphylococcus aureus* USA300 clone. *FEMS Microbiology Letter*, 332(2), 146-152.
- Varman, R. M., & Singh, S. (2012). Investigation of Effects of Terpene Skin Penetration Enhancers on Stability and Biological Activity of Lysozyme. *AAPS PharmSciTech*, 13(4), 1084-1090.
- Vo, A. T., Van Duijkeren, E., Gaastra, W., & Fluit, A. C. (2010). Antimicrobial resistance, class 1 integrons, and genomic island 1 in *Salmonella* isolates from Vietnam. *PLoS One*, 5(2), 1-8.
- Vollmer, W., Blanot, D., & De Pedro, M. A. (2008). Peptidoglycan structure and architecture. *FEMS Microbiology Reviews*, 32(2), 149-167.
- Webber, M. A., & Piddock, L. J. V. (2003). The importance of efflux pumps in bacterial antibiotic resistance. *Journal of Antimicrobial Chemotherapy*, 51(1), 9-11.
- Wiedenbeck, J., & Cohan, F. M. (2011). Origins of bacterial diversity through horizontal genetic transfer and adaptation to new ecological niches. *FEMS Microbiology Reviews*, 35(5), 957-976.
- Wild, C. P., & Gong, Y. Y. (2010). Mycotoxins and human disease: a largely ignored global health issue. *Carcinogenesis*, 31(1), 71-82.
- Witaicenis, A., Seito, L. N., da Silveira Chagas, A., de Almeida, L. D., Luchini, A. C., Rodrigues-Orsi, P., Cestari, S. H., & Di Stasi, L. C. (2014). Antioxidant and intestinal anti-inflammatory effects of plant-derived coumarin derivatives. *Phytomedicine*, 21(3), 240-246.
- Wong, M. H. Y., Yan, M., Chan, E. W. C., Biao, K., & Chen, S. (2014). Emergence of clinical *Salmonella typhimurium* with concurrent resistant to ciprofloxacin, ceftriaxone and azithromycin. *Antimicrobial Agents and Chemotherapy*, 1-24.
- Wong-Beringer, A., Wiener-Kronish, J., Lynch, S., & Flanagan, J. (2008). Comparison of type III secretion system virulence among fluoroquinolone-susceptible and-resistant clinical isolates of *Pseudomonas aeruginosa*. *Clinical Microbiology and Infection*, 14(4), 330-336.
- World Health Organization. (2015). Worldwide country situation analysis: response to antimicrobial resistance.
- Wright, G.D. (2005). Bacterial resistance to antibiotics: Enzymatic degradation and modification. *Advanced Drug Delivery Reviews*, 57(10), 1451-1470.
- Yamagata, K., Takeda, M., Onizawa, K., & Yoshida, H. (2006). Submandibular abscess caused by *Salmonella*. *International Journal of Oral and Maxillofacial Surgery*, 35(10), 969-971.

- Yang, S., Xiong, H., Yan, Q., Yang, H., & Jiang, Z. (2014). Purification and characterization of a novel alkaline β -1, 3-1, 4-glucanase (lichenase) from thermophilic fungus *Malbranchea cinnamomea*. *Journal of Industrial Microbiology & Biotechnology*, 41(10), 1487-1495.
- Ye, Y., Gan, J., & Hu, B. (2015). Screening of Phosphorus-Accumulating Fungi and Their Potential for Phosphorus Removal from Waste Streams. *Applied Biochemistry and Biotechnology*, 1-10.
- Zakaria, Z., Sreenivasan, S., & Mohamad, M. (2007). Antimicrobial activity of *Piper ribesoides* root extract against *Staphylococcus aureus*. *Journal of Applied Biological Sciences*, 1(3), 87-90.
- Zalar, P., & Gunde-Cimerman, N. (2014). Cold-adapted yeasts in arctic habitats. In *Cold-adapted Yeasts* (pp. 49-74). Berlin, Heidelberg: Springer.
- Zarnowski, R., Westler, W. M., Lacmbouh, G. A., Marita, J. M., Bothe, J. R., Bernhardt, J., Sahraoui, A. L., Fontaine, J., Sanchez, H., Hatfield, R. D., Ntambi, J. M., Nett, J. E., Mitchell, A. P., & Andes, D. R. (2014). Novel entries in a fungal biofilm matrix encyclopedia. *MBio*, 5(4), 1-13.
- Zhanel, G. G., Karlowsky, K. A., Bohan, D. J., & Davidson, R. J. (1991). Antimicrobial activity of sub-inhibitory concentrations of aminoglycosides against *Pseudomonas aeruginosa* as determined by the postantibiotic effect. *Antimicrobials Agents Chemotherapy*, 37, 114-121.
- Zhanel, G. G., Schweizer, F., & Karlowsky, J. A. (2012). Oritavancin: mechanism of action. *Clinical Infectious Diseases*, 54(Supplement 3), 214-219.
- Zhang, Y., Wu, W. P., Hu, D. M., Su, Y. Y., & Cai, L. (2014). A new thermophilic species of *Myceliophthora* from China. *Mycological Progress*, 13(1), 165-170.
- Zhao, C., Sun, H., Wang, H., Liu, Y., Hu, B., Yu, Y., Sun, Z., Chu, Y., Cao, B., Liao, K., Lei, J., Hu, Z., Zhang, L., Zhang, X., Xu, Y., Wang, Z., & Chen, M. (2012). Antimicrobial resistance trends among 5608 clinical Gram-positive isolates in China: results from the Gram-Positive Cocci Resistance Surveillance program (2005–2010). *Diagnostic Microbiology and Infectious Disease*, 73(2), 174-181.