

UNIVERSITI PUTRA MALAYSIA

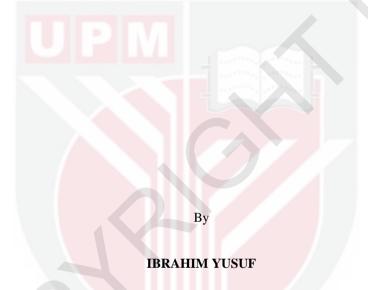
BIODEGRADATION OF RECALCITRANT CHICKEN FEATHER WASTES BY LOCALLY ISOLATED HEAVY METAL TOLERANT Alcaligenes sp. STRAIN AQ05-001

IBRAHIM YUSUF

FBSB 2016 15



BIODEGRADATION OF RECALCITRANT CHICKEN FEATHER WASTES BY LOCALLY ISOLATED HEAVY METAL TOLERANT Alcaligenes sp. STRAIN AQ05-001



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of Philosophy

May 2016

COPYRIGHT

All materials contained within the thesis including without limitation text, logos, icons, photographs and all other artworks are copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any materials contained within the thesis for non-commercial purposes from copyright holder. Commercial use of materials may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

This thesis is dedicated to my parents, family and friends for their never-ending prayers and blessings throughout my studies.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Doctor of Philosophy

BIODEGRADATION OF RECALCITRANT CHICKEN FEATHER WASTES BY LOCALLY ISOLATED HEAVY METAL TOLERANT *Alcaligenes* sp. STRAIN AQ05-001

By

IBRAHIM YUSUF

May 2016

Chairman Faculty : Siti Aqlima Binti Ahmad, PhD : Biotechnology and Biomolecular Sciences

The global increase in consumption of chicken meats as source of protein and the current use of chicken feathers as source of cheaply available biosorbent materials has led to the generation of large volume of recalcitrant melanised and non-melanised keratin containing primary and secondary feathers wastes in our environments. A need for eco-friendly method of managing these wastes became highly necessary.

A novel bacterium isolated from feather dump site located in Johor, Malaysia identified by physical, biochemical and 16S RNA sequencing techniques as Alcaligenes sp. AQ05-001 was successfully used to degrade such feather wastes to produce keratinase enzyme and protein rich hydrolysates. Initially it only degraded 1 g/L of heavy metal polluted and un polluted feather wastes, produced 8.84 U/mL keratinase in 3 days. But upon optimisation of physical and nutritional factors using one-factor-at-a-time and response surface methodology approaches, complete degradation of almost 5 g/L feather and a 10fold increase in keratinase production (88.0±1 U/mL) was recorded. The bacterium was immobilised in gellan gum gelling agent with its conditions optimised. With 0.8% gellan gum concentration, 3 mm size beads and 250 initial cell loads, immobilised cells degraded feathers at a faster rate and produced higher keratinase activities in optimised media. Both free and immobilised cells efficiently degraded recalcitrant melanised feathers faster than non-melanised feathers. They both showed a high preference for black feathers and complete degradation of 5 g/L of black feathers to dust and fibres was achieved in 18 and 24 h with immobilised and free cells, respectively. Interestingly, the strain further selectively degraded black feather completely in a flask containing 5 g/L mixture of equal amount of black, brown and white feathers, with leftover of brown feathers rachises at the end of 36 h incubation. Further, free and immobilised bacteria degraded large amount of feathers in optimised media. About 50 g/L feathers were degraded by immobilised cells while free cells degraded up to 40 g/L in 60 h with production of high amount of soluble protein hydrolysates that are rich in both essential and non-essential amino acids such as leucine, isoleucine, cysteine, aspartic acid and proline. Optimal adsorption of heavy metals by chicken feathers was carried out in broths within the pH range of 7-8.5 and

temperature between 25-35 $^{\circ}$ C. Both free and immobilised cells degraded feather-laden with single or combination of heavy metals with concurrent production of higher keratinase enzyme at varying rates. Beads were reused in biodegradation of feathers polluted with different heavy metals in semi-continuous fermentation for a number of cycles ranging from 4-10. Encapsulated cells tolerated up to 30 ppm of Ag, 20 ppm of Co, 15 ppm of Cu, 10 ppm of As, Ni, Cd and 5 ppm of Hg, Pb while free-living cells tolerated only 1 ppm of Hg and Pb. Moreover, cells in gellan gum degraded about 95% and above of feathers polluted with a mixture of 5 ppm each of Ag, As, Cd, Co, Cu, 10 and 20 ppm each of Ag, Co and Cu at 18 h. Beads that failed to degrade feathers laden with higher concentration of heavy metals like mercury, were used to degrade feathers laden with other heavy metals for many cycles without need for mineral acid desorption. Crude keratinase produced by strain AQ05-001 displayed relative activity over a temperature ranging from 30-60 °C, pH 7-10 and was highly inhibited by serine inhibitor, Tween20, Trixton-100 and sodium dodecyl sulphate. However, cell-free crude keratinase was not able to degrade feathers efficiently, but improved degradation was observed in the presence of reducing agent (2 mercaptoethanol).

The strain possesses the potential of being used in the management of melanised, nonmelanised, heavy metal free/polluted feather as well as in the production of protein-rich hydrolysates for animal feeds and also in the industrial production of valuable keratinase enzyme. Furthermore, not only its heavy metals tolerability can be harnessed in the bioremediation of heavy metal-contaminated feather wastes, but it could also use the contaminated wastes as a substrate for keratinase production. Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doctor Falsafah

PENURUNAN SISA BUANGAN BULU AYAM YANG SANGAT DEGIL OLEH PEMENCILAN BAKTERIA TEMPATAN PELBAGAI KETAHANAN LOGAM BERAT - *Alcaligenes* sp. STRAIN AQ05-001

Oleh

IBRAHIM YUSUF

Mei 2016

Pengerusi Fakulti : Siti Aqlima Binti Ahmad, PhD : Bioteknologi dan Sains Biomolekul

Peningkatan dalam pengambilan harian daging ayam dalam hidangan kita yang telah membawa kepada jumlah besar generasi multi buangan bulu ayam berwarna yang tahan protease konvensional atau serangan mikrob, dan dengan itu membentuk gangguan dalam alam sekitar. Di samping itu, penggunaan berterusan sisa bulu ini adalah semurah bahanbahan bio penyerap dalam penyingkiran logam berat dari buangan industri dan air sisa serta pelupusan haram massa besar bulu di tempat-tempat di mana ia boleh menyerap logam, telah membawa kepada pengumpulan sisa bulu yang sangat sukar dan toksik yang menimbulkan isu biodegradasi serius. Sejak cara konvensional melupuskan sisa ini tidak mesra alam, keperluan untuk mengasingkan strain logam berat toleran yang berkesan boleh merendahkan bulu adalah sangat perlu. Pengasingan, pencirian dan pengoptimaan keadaan fizikal dan kultur strain baru bakteria yang mampu menggunakan kedua-dua bahan buangan pertama dan kedua sebagai substart untuk menghasilkan hidrolisat yang kaya keratinase and protein dilaporkan. Bakteria telah dikenal pasti secara fizikal, bikimia dan jujukan 16S RNA sebagai Alcaligenes sp. AQ05-001. Ini kali pertama Alcaligenes sp. telah dugunakan dalam penurunan bahan buangan bulu. Dalam media basal dengan dan tanpa logam berat, bakteria hanya menurunkan 1 g/L bulu dan menghasilkan 8.84 U/mL keratinase dalam 3 hari. Selepas pengoptimuman faktor-faktor fizikal dan budaya mempengaruhi hasil keratinase menggunakan satu faktor-dalam-satu-masa dan kaedah gerak balas permukaan, Penurunan lengkap 5 g/L bulu dan peningkatan 10 kali ganda dalam pengeluaran keratinase (88.4 U/mL). Bakteria ini telah disekat gerak dalam agen pengelan gellan gum dengan keadaan dioptimumkan. Dengan 0.8% kepekatan gula gellan, manik saiz 3 mm dan 250 beban sel awal, sel-sel sekat gerak telah dinurunkan bulu pada kadar yang lebih cepat dan menghasilkan aktiviti keratinase lebih tinggi dalam media yang dioptimumkan. Kedua-dua sel bebas dan sekat gerak dengan cekap penunrunkan bulu melanised keras lebih cepat daripada bulu bukan melanised. Kedua-duanya menunjukkan keutamaan tinggi untuk bulu hitam dan penurunan lengkap 5 g/L bulu hitam untuk debu dan serat telah dicapai pada 18 dan 24 jam untuk sel-sel bebas dan sekat gerak, masingmasing. Menariknya, strain yang terpilih menurunkan sepenuhnya hitam dalam kelalang yang mengandungi campuran jumlah yang sama bulu hitam, coklat dan putih dengan sisa bulu rachises coklat pada akhir 36 jam pengeraman. Tambahan pula, dengan media dioptimumkan, sejumlah besar sehingga 50 g/L bulu telah diturunkan oleh sel bebabs AQ05-001 manakala sel-sel sekat gerak gerak menurunkan sehingga 40 g/L dalam 54 dan 60 jam, masing-masing dengan paduan yang sangat berjumlah protein larut hidrolisat yang kaya dengan kedua-dua asid amino penting dan tidak penting seperti leucina, isoleucina, cysteina, asid aspartik dan prolina. Kedua-dua sel bebas dan sekat gerak menunkan bulu sarat dengan tunggal atau kombinasi logam berat dengan pengeluaran serentak enzim keratinase lebih tinggi. Sel-sel yang terkandung diterima sehingga 30 ppm Ag, 20 ppm Co, 15 ppm Cu, 5 ppm Hg dan Pb manakala sel-sel hidup bebas diterima hanya 1 ppm Hg dan Pb. Selain itu, sel-sel dalam gellan gam diturunkan kira-kira 90% daripada bulu dalam media yang mengandungi campuran 5 ppm dari setiap Ag, As, Co, Cu, Ni dan 10 ppm dari setiap Ag, Co dan Cu. Manik yang sama telah digunakan dalam beberapa kitaran penurunan bulu tanpa penerapan asid mineral. Manik digunakan semula dalam biopenurunan bulu tercemar dengan logam berat yang berbeza dalam penapaian separuh berterusan sehingga tujuh kitaran. Keratinase mentah yang dihasilkan oleh strain AQ05-001 memaparkan aktiviti maksimum ke atas suhu antara 30-60 ℃, pH 7-9 dan sangat direncatkan oleh perencat serina, Tween20, Trixton-100 dan natrium dodesil sulfat. Walau bagaimanapun, sel keratinase mentah bebas tidak dapat mernurunkan bulu, tetapi penurunan hanya beberapa bulu di hadapan agen penurunan. Selain keratinase tinggi dan hasil protein larut, strain ini boleh digunakan dalam pengurusan semua sisa bulu, pengeluaran protein hidrolisat kaya untuk makanan haiwan dan juga dalam pengeluaran perindustrian enzim berharga keratinase dengan kecekapan yang tinggi dalam penunurunan kedua-dua sangat sukar melanised dan kurang bulu bukan melanised. Tambahan pula, bukan sahaja boleh diterima logam berat yang boleh dimanfaatkan dalam biopemulihan sisa logam berat tercemar bulu, tetapi ja juga boleh menggunakan sisa tercemar sebagai substrat untuk pengeluaran keratinase.

ACKNOWLEDGEMENTS

All thanks and praises are due to Allah (SWT), the Cherisher and Sustainer of the world for giving me the opportunity to successfully carry out this study. May all blessings be upon His Prophet and Messenger, Muhammad (SAW).

The completion of this work was made possible with the help, assistance and aids from many important people. First and foremost, I would like to voice out my sincere and profound gratitude to my supervisor and co-supervisors Dr. Siti Aqlima Ahmad, Assoc. Prof. Dr. Mohd Yunus Abd Shukor, Prof. Dr. Mohd Arif Syed and Assoc. Prof. Dr. Phang Lai Yee whose efforts led to the successful conduct and completion of this study. I was also indebted to the management of Bayero University Kano, Nigeria and Tetfund for selecting and sponsoring me to conduct my Ph.D. in Universiti Putra Malaysia. I also wish to express my appreciation to all my colleagues and members of Lab 204 and 115 group for the enormous contributions they rendered during the conduct of this research. No amount of thanks is comparable with their efforts.

During the conduct of this work, a lot of unexpected as well as expected bonuses were met; most of which came from friends, relatives and well-wishers. I am highly grateful to my mum for her upbringing and prayers during the course of this research. My appreciation to my beloved brothers and sisters, Saudat Yusuf, Rukayya Yusuf, Munir Yusuf, Fatima Yusuf, Abdur Razak Yusuf, Hassana and Usaina Yusuf, Abdullateef Yusuf, Maryam Yusuf, Dr. Abdulhakeem Badamasi and others. I have neither silver nor gold to reward you for your contributions to my success but I owe you prayers and continuous love.

Last and not the least is my humble wife Zainab Abdurrazak and my children Aisha Ibrahim Yusuf, Amina Ibrahim Yusuf and Yusuf Ibrahim Yusuf whose patience, endurance and prayers assisted me throughout this study and beyond.

Ibrahim Yusuf, 20-May-2016 I certify that a Thesis Examination Committee has met on 13 May 2016 to conduct the final examination of Ibrahim Yusuf on his thesis entitled "Biodegradation of Recalcitrant Chicken Feather Wastes by Locally Isolated Heavy Metal Tolerant *Alcaligenes* sp. Strain AQ05-001" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Umi Kalsom binti Md Shah, PhD

Associated Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

Raja Noor Zaliha bt Raja Abd. Rahman, PhD

Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Internal Examiner)

Janna Ong binti Abdullah, PhD

Associated Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Internal Examiner)

Hiroyuki Futamata, PhD

Professor Shizuoka University Japan (External Examiner)

ZULKARNAIN ZAINAL, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 26 July 2016

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Siti Aqlima Ahmad, PhD

Senior Lecturer Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia (Chairman)

Mohd Yunus Abd Shukor, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia (Member)

Mohd Arif Syed, PhD

Professor Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia (Member)

Phang Lai Yee, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia (Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature:__

Date:_

Name and Matric No: Ibrahim Yusuf GS 38495

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: Name of Chairman of Supervisory Committee: Dr. Siti Aqlima Ahmad

Signature:

Name of Member of Supervisory Committee: Associate Professor Dr. Mohd Yunus Abd Shukor

Signature: Name of Member of Supervisory Committee: Professor Dr. Mohd Arif Syed

Signature: Name of Member of Supervisory Committee: Professor Dr. Phang Lai Yee

TABLE OF CONTENTS

Page

ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	V
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	XX

CHAPTER

C

1	INTF	RODUCT	TION	1
2	LITE	RATUR	RE REVIEW	3
	2.1	Overvie	ew of keratin	3
		2.1.1	Composition and structure	3
		2.1.2	-	4
	2.2	Keratin	wastes and keratinase	5
		2.2.1	Types of feather	6
		2.2.2		7
		2.2.3	Feather recycling	8
		2.2.4	Applications of keratinous wastes	9
		2.2.5	Keratinase production and applications	11
		2.2.6	Keratinase properties and factors affecting their	13
			productions	
		2.2.7	Keratinase producing microorganisms	16
		2.2.8	Media composition and fermentation processes in	17
			keratinase production	
		2.2.9	Types of fermentation processes in keratinase	20
			production	
			2.2.9.1 Submerged or liquid fermentation	20
			(SmF/LF) in keratinase production	
			2.2.9.2 Solid-state fermentation (SSF)	21
			2.2.9.3 Comparing SmF and SSF	22
		2.2.10	Optimisation of culture conditions affecting keratinase	22
			production	
		2.2.11	Methods of optimisation in keratinase production	22
		2.2.12	Keratinase assay methods	26
			2.2.12.1 Azokeratin	26
			2.2.12.2 Keratin azure	27
			2.2.12.3 Soluble keratin	27
			2.2.12.4 Azocasein	27
			2.2.12.5 Folin–Ciocalteu method	27
			2.2.12.6 Keratin substrates (Noval and Nickerson method)	28

2.2.12.7 Amino acid liberation method 28

	2.3	Immobilisation studies	28
		2.3.1 Gellan gum as supportive matrix	30
	2.4	Bioremediation of heavy metals	31
3		ERIALS AND METHODS	33
	3.1	Experimental approach	33
	3.2	Chicken feather collection and processing	33
	3.3	Chemicals and equipments	33
	3.4	Preparation of media	33
		3.4.1 Skim milk agar (SMA)	33
		3.4.2 Feather meal broth (FMB) and feather meal agar (FMA)	35
	3.5	Isolation, screening and selection of heavy metals tolerant	35
		bacterium	
	3.6	Identification of selected bacterium	36
		3.6.1 Physical identification	36
		3.6.2 Biochemical identification	36
		3.6.3 Molecular identification	36
	3.7	Analytical methods	37
		3.7.1 Keratinase assay	37
		3.7.2 Synthesis of azokeratin	38
		3.7.3 Growth determination (CFU procedure)	38
		3.7.4 Determination of soluble proteins concentration	39
		3.7.5 Determination of feather degradation	39
		3.7.6 Scanning electron microscopy	40
		3.7.7 Determination of sulfhydryl group production	40
	3.8	Optimisation of factors affecting keratinase production using OFAT	40
		3.8.1 Optimisation of substrate concentration	40
		3.8.2 Optimisation of inoculum size	41
		3.8.3 Optimisation of medium pH	41
		3.8.4 Optimisation of temperature	41
		3.8.5 Optimisation of extra sources of carbon	41
		3.8.6 Optimisation of extra sources of nitrogen	42
	3.9	Optimisation using statistical approach	42
		3.9.1 Plackett-Burman factorial design (PBFD)	42
		3.9.2 Central composite design (CDD) and response surface methodology (RSM)	43
	3.10	Immobilisation studies	43
		3.10.1 Harvesting bacteria	43
		3.10.2 Immobilisation of bacteria in gellan gum	44
		3.10.3 Optimisation of gellan gum concentration	44
		3.10.4 Optimisation of bead size and number (Initial cell load)	44
		3.10.5 Quantification of bacterial concentration in beads	45
		3.10.6 Quantification of cell leakages	45
		3.10.7 Reuse of immobilised cells for repeated batch cultivation	45
	3.11	Degradation of high amount of different types of feathersd	45
	3.12	Degradation of secondary heavy metal polluted feathers	46

	3.13			rolysates for amino acid analysis	46
	3.14	Charac		f crude keratinase	46
		3.14.1		ation of optimum temperature and pH on e activities	46
		2140			47
				enzyme inhibitors	47
			Effect of a		47
				surfactants and detergents	47
		3.14.5	Degradati by crude l	on of melanised and non-melanised feathers	47
	3.15	Statistic	cal analysis		48
4	RES	ULTS AI	ND DISCU	SSION	49
	4.1			and identification of keratinase producing	49
				ing bacteria	
		4.1.1		of proteolytic bacteria from soil samples	49
				ghter and poultry farm houses	
		4.1.2		for feather-degrading and heavy metals	50
			tolerant ba		•••
		4.1.3		tion of isolate P3	53
	4.2			f Alcaligenes sp. AQ05-001 in basal feather	57
		mediun		The angenes spirit gos oor in susar reaction	57
	4.3			ratinase production by Alcaligenes sp.	59
		AQ05-		a at a time (OFAT) antimization	50
		4.3.1	4.3.1.1	or-at-a-time (OFAT) optimisation	59 59
			4.5.1.1	Optimisation of feather (substrate) concentrations	39
			4212		(1
			4.3.1.2	Optimisation of media pH	61
			4.3.1.3	Optimisation of temperature	64
			4.3.1.4	Optimisation of inoculum size	66
			4.3.1.5	Effect of additional carbon and nitrogen sources on keratinase production	67
		4.3.2	Statistical	optimisations	74
		7.3.2	4.3.2.1	Selection of most significant media	74
			7.3.2.1	components using Plackett-Burman	74
				factorial design	
			4.3.2.2	Optimisation by Response Surface	77
			4.3.2.2	Methodology (RSM)	,,
			4.3.2.3	Verification of model	90
	4.4	Immob	ilisation stu	dies	90
		4.4.1	Optimisat	ion of immobilisation protocols	90
			4.4.1.1	Optimisation of gellan gum concentration	90
			4.4.1.2	Optimisation of bead sizes	92
			4.4.1.3	Optimisation of initial cell loading (number	93
	1 5	Vant		of beads)	0.4
	4.5			tion and feather degradation in media with	94
	1.5		ed gellan g		~~
	4.6	AQ05-		lanised chicken feathers by Alcaligenes sp.	97
	4.7			sed cells for repeated batch cultivation	103
				-	

	4.8	Degradation of different feather concentration using free and immobilised cells of AQ05-001	105
	4.9	Changes in morphologies of feather degraded by strain AQ05-	108
	4.10	001 Characterisation of crude keratinase from free and immobilised cells	112
	4.11	Degradation of feathers by crude keratinase of free and immobilised cells	116
	4.12	Amino acid analysis of hydrolysates produced by degradation of black, brown and white feathers	117
	4.13	Degradation of heavy metal polluted feathers and by free and immobilised cells of Alcaligenes sp. AQ05-001	121
	4.14	Comparing Alcaligenes sp. AQ05-001 with previous keratinase-producing organisms	128
	CON	CLUSION	129
F	RENCI	ES	131

REFERENCES APPENDICES BIODATA OF STUDENT LIST OF PUBLICATIONS

5

 \bigcirc

- 155 171
- 172

LIST OF TABLES

Tab	le	Page
2.1	Table 2.1: Different basal feather medium compositions for keratinase production in g/L of distilled water	18
4.1	Sites of isolation, morphological appearance and percentage feather degradation of potential feather degrading bacteria	52
4.2	Percentage feather degradation by seven feather degrading bacteria during screening for heavy metal tolerant strain in feather meal broth with and without heavy metals	53
4.3	Physical and biochemical tests used in identification of the selected feather-degrading bacteria	54
4.4	pH kinetics during the fermentation process. pH shifts towards 8.9 from acidic region and 9.3 from high alkaline region	63
4.5	Real and coded values of Plackett-Burman factorial design experiments	74
4.6	Plackett-Burman experimental design matrix with keratinolytic enzyme production levels	75
4.7	Effects of independent factors on keratinase production and their regression coefficients	76
4.8	Coded and actual values of significant factors used in central composite design (CDD)	78
4.9	Central composite design experimental matrix generated by Design expert 6.0.6 and corresponding responses (actual and predicted)	78
4.10	Model summary statistics showing sequential model sum of squares and lack of fit tests	80
4.11	Analysis of variance (ANOVA) results for the model showing regression coefficients of the factors	81
4.12	Amino acid compositions in raw undigested black, brown and white chicken feathers	101
4.13	Keratinase production and degradation of differently coloured feathers using immobilised cells of strain AQ05-001 in semi- continuous fermentation	104

4.14	Effect of temperature on activity of crude keratinase produced by Alcaligenes sp. AQ05-001 in FMB containing 5 g/L of white feathers after 36 h of incubation	112
4.15	Effect of pH on activity of crude keratinase produced by Alcaligenes sp. AQ05-001 in FMB containing 5 g/L of white feathers after 36 h of incubation	113
4.16	Effect of enzyme inhibitors on activity of crude keratinase produced by Alcaligenes sp. AQ05-001 in FMB containing 5 g/L of white feathers after 36 h of incubation.	114
4.17	Effect of metal ions on activity of crude keratinase activity produced by Alcaligenes sp. AQ05-001 in FMB containing 5 g/L of white feathers after 36 h of incubation	115
4.18	Effect of surfactants on activity of crude keratinase produced by <i>Alcaligenes</i> sp. AQ05-001	115
4.19	Comparing degradation of 0.5 g of white feather in 20 mL cell-free crude keratinase from free and immobilised cells	116
4.20	Comparing degradation of 0.5 g of black, brown and white feather by crude keratinase with and without reducing agent	117
4.21	Amino acid profile from hydrolysates resulted from degradation of raw black, brown and white feathers by strain AQ05-001 after 12 h of incubation	118
4.22	Amino acid profile from hydrolysates resulted from degradation of raw black, brown and white feathers by strain AQ05-001 after 24 h of incubation	119
4.23	Amino acid profile from hydrolysates resulted from degradation of raw black, brown and white feathers by strain AQ05-001 after 36 h of incubation	120
4.24	Variation in net weight of 0.5 g feathers in 100 mL control feather meal broths that contains 1 ppm of each heavy metal under different media pH	122
4.25	Effect of heavy metals on keratinase activity and feather degradation by free-living cells of <i>Alcaligenes</i> sp. AQ05-001	123
4.26	Effect of different concentration of heavy metals on keratinase production and feather degradation by immobilised cells of <i>Alcaligenes</i> sp. AQ05-001	125
4.27	Concentrations of heavy metals tolerated and number of times the beads were reused	126

- 4.28 Average degradation of feathers laden with combination of heavy 127 metals and number of times the beads were reused
- 4.29 Comparing *Alcaligenes* sp. AQ05-001 with other protease/keratinase 128 producing organisms



LIST OF FIGURES

Figure		Page
2.1	Parallel sheet structure of keratin showing different bonds	3
2.2	Parts of chicken feather	6
2.3	Types of chicken feather	7
2.4	Chickens with different coloured feathers	8
2.5	Laboratory setup for keratinase production experiment	21
2.6	Step by step process of keratin fermentation experiment.	21
2.7	Structure of gellan gum	30
3.1	Flowchart of experimental approach showing stages of experiment.	34
4.1	Halo zone produced by a typical proteolytic bacterium on skim milk agar during screening	49
4.2	Pure colonies of a feather-degrading bacterium (P3) on feather meal agar indicating its ability to utilise feather as a sole source of carbon and nitrogen	51
4.3	Phylogenetic tree showing the position of Alcaligenes sp. AQ05-001 strain among Alcaligenes genera and other bacteria. The evolutionary pattern was generated by the neighbour-joining method using MEGA6. Serratia marcescens was used as an outgroup. Species names of bacteria were followed by the accession numbers. Bootstrap values were calculated based on 1000 resamplings.	56
4.4	Growth of strain AQ05-001 and keratinase production in 100 mL basal FMB containing 1% feather incubated at 30 °C for 72 h	57
4.5	Feather degradation, protein concentration and thiol concentration recorded when strain AQ05-001 was inoculated into 100 mL basal FMB with 1% feather for 72 h at 30°C	58
4.6	Feather residues in inoculated and uninoculated basal FMB after 72 hours of incubation at 30° C	59
4.7	Optimisation of feather concentration on growth of <i>Alcaligenes</i> sp. AQ05-001, keratinase production and feather degradation	60

4.8	Optimisation of pH for maximum keratinase and crude soluble protein production, cellular growth and feather degradation by Alcaligenes sp. AQ05-001. (a) Effect of pH on keratinase production and crude soluble protein concentrations. (b) Effect of pH on feather degradation and growth of Alcaligenes sp. AQ05- 001.	62
4.9	Effect of different buffers adjusted to different pH on keratinase production by <i>Alcaligenes</i> sp. AQ05-001	64
4.10	Optimisation of temperature for maximum keratinase production, cellular growth and feather degradation by Alcaligenes sp. AQ05-001. (a) Effect of temperature on keratinase production by Alcaligenes sp. AQ05-001. (b) Effect of temperature on growth of Alcaligenes sp. AQ05-001 and on its feather degradation ability.	65
4.11	Effect of different inoculum size on keratinase production by <i>Alcaligenes</i> sp. AQ05-001 in 100 mL feather meal broth pH 8 containing 5 g/L (w/v) of feathers incubated at 27°Cfor 72 h	67
4.12	Optimisation of carbon sources on keratinase production and feather degradation by Alcaligenes sp. AQ05-001 in 100 ml feather meal broth pH 8 containing 5 g/L (w/v) of feathers incubated 27oC for 48 h. (a) Effect of different extra sources of carbon on feather degradation and keratinase activities by strain AQ05-001. (b) Effect of different concentration of sucrose on keratinase production by Alcaligenes sp. AQ05-001	69
4.13	Effect of different extra sources of nitrogen on growth of AQ05-001 as well as feather degradation and keratinase production. (a) Effect of 1 g elemental nitrogen in defined inorganic nitrogen sources. (b) Effect of 1 g/L of complex organic nitrogen sources	71
4.14	Effects of different concentration of ammonium bicarbonate on keratinase production by <i>Alcaligenes</i> sp. AQ05-001 in 100 mL feather meal broth pH 8 containing 5 g/L (w/v) feathers incubated at 27° C for 48 h	72
4.15	Effects of different concentration of urea on keratinase production by <i>Alcaligenes</i> sp. AQ05-001 in 100 mL feather meal broth pH 8 containing 5 g/L (w/v) feathers incubated at 27°C for 48 h	72
4.16	Effects of different concentration of skim milk on keratinase production by <i>Alcaligenes</i> sp. AQ05-001 in 100 mL feather meal broth pH 8 containing 5 g/L (w/v) feathers incubated at 27°C for 48 h	73
4.17	Pareto-chart showing level of significance of independent factors used in Placket-burman factorial design on production of keratinase by <i>Alcaligenes</i> sp. AQ05-001	77
	xviii	

4.18	Model diagnostic plots (a) predicted versus actual (b) studentized residue versus predicted (c) normal plots of residue (d) outlier T versus run.	84
4.19	Keratinase production as a function of feather concentration and urea when ammonium bicarbonate, skim milk and sucrose are maintained at zero level in 3D and 2D surface response view	86
4.20	Keratinase production as a function of sucrose and ammonium bicarbonate when feather concentration, urea and skim milk are maintained at zero level in 3D and 2D surface response view	87
4.21	Keratinase production as a function of sucrose and skim milk when feather concentration, ammonium bicarbonate and urea are maintained at zero level in 3D and 2D surface response view	88
4.22	Keratinase production as a function of skim milk and urea when ammonium bicarbonate, feather concentration and sucrose are maintained at zero level in 3D and 2D surface response view	89
4.23	Effect of gellan gum concentration on keratinase production and feather degradation by immobilised <i>Alcaligenes</i> sp. AQ05-001 in optimised media containing 5 g/L of feather	91
4.24	Effect of bead size on keratinase production and feather degradation by immobilised Alcaligenes sp. AQ05-001 in optimised media containing 5 g/L of feather. Data represents mean \pm S.D of triplicates.	93
4.25	Effect of bead numbers on keratinase production and feather degradation by immobilised Alcaligenes sp. AQ05-001 in optimised media containing 5 g/L of feather incubated at 27oC for 24 h. Data represents mean \pm S.D of triplicates.	94
4.26	Comparing keratinase production and feather degradation by free and gellan gum immobilised cells of Alcaligenes sp. AQ05-001. Data are the means \pm standard deviations of three independent replicates.	95
4.27	Degradation of 5 g/L of feathers with 250 beads of immobilised cells of AQ05-001 after 24 hours of incubation (a) Degraded feather parts after 24 hours of incubation with 250 beads of immobilised cells of strain AQ05-001. (b) Control with intact feather after 24 hours.	96
4.28	Comparing amount of total soluble protein available during cultivation in 100 mL feather meal broth inoculated with free and immobilised cells of <i>Alcaligenes</i> sp. AQ05-001 at 6 hours intervals	97
4.29	Time course biodegradation of different coloured feathers by free cells of Alcaligenes sp. AQ05-001 in an optimised feather meal broth. Data represent mean of triplicates \pm standard deviation.	98

xix

- Time course keratinase production by free cells of Alcaligenes sp. 99
 AQ05-001 in media containing black, brown and white feathers.
 Data represent mean of triplicates ±standard deviation.
- 4.31 Time course keratinase production by immobilised cells of Alcaligenes sp. AQ05-001 in media containing black, brown and white feathers. Data represent mean of triplicates ± standard deviation.

- 4.32 Comparing rate of degradation of black, brown and white feathers by 102 strain AQ05-001 in optimised FMB. (a) the bacterium degraded black feather completely including the rachises leaving no residue (b,c) brown and white feathers were degraded but with residues.
- 4.33 Time course biodegradation of mixture of different coloured feathers 103 by Alcaligenes sp. AQ05-001 in an optimised feather medium. Data represent mean of triplicates ± standard deviation.
- 4.34 Cell leakage per cycle for each feather type. Data represent mean of 105 triplicates ±standard deviation.
- 4.35 Degradation of different amount of feathers by free and immobilised 107 Alcaligenes sp. AQ05-001 in feather meal broth containing 10, 20, 30, 40, 50 and 60 g/ L raw feathers. At indicated intervals data are means ± S.D of three independent experiments. (a) white free cells (b) white immobilised cells (c) brown free cells (d) brown immobilised cells (e) black free cells (f) black immobilised cells.
- 4.36 SEM micrographs showing bacterial colonisation and change in 111 feather structure at 12h, 24h and 36 h of incubation. (a) Feather in feather meal broth without bacteria (control) after 12 and 36 h of incubation at 27oC. (b) change in structure of white feathers and level of colonisation of Alcaligenes sp. AQ05-001 on white feather at 12, 24 and 36 h (c) change in structure of brown feathers and level of colonisation of Alcaligenes sp. AQ05-001 on brown feather at 12, 24 and 36 h (d) change in structure of black feathers and level of colonisation of Alcaligenes sp. AQ05-001 on black feather at 12, 24 and 36 h (d) change in structure of black feathers and level of colonisation of Alcaligenes sp. AQ05-001 on black feather at 12, 24 and 36 h.

LIST OF ABBREVIATIONS

CFU	Colony forming units
DTNB	Dithiobis (2-nitrobenzoic acid
EDTA	Ethylene diamine tetra acetic acid
FBA	Feather basal agar
FBB	Feather basal broth
FD	Feather degradation
FDB	Feather degrading bacteria
FDM	Feather degrading micro-organisms
g	Gram
ICL	Initial cell load
KA	Keratinase activities
kDa	Kilo Daltons
L	Litre
mL	Millilitre
ММ	Molecular mass
NCBI	National centre for biotechnology information
OFAT	One factor at a time
PBFD	Placket-Burman factorial design
PCR	Polymerase chain reaction
PMSF	Phenyl methyl sulfonyl fluoride
RSM	Response surface methodology
SDS	Sodium dodecyl sulphate
SMA	Skim milk agar
SmF/LF	Submerged or Liquid Fermentation
SSF	Solid state fermentation

- TCA Trichloroacetic acid
- TSE Transmissible spongiform encephalitis
- v/v Volume by volume
- w/v Weight by volume

 $\left(\mathbf{C} \right)$



CHAPTER I

INTRODUCTION

Chicken continue to form an important part of human meal as cheap source of protein. In the local or industrial processing of these chicken, large quantity of different types of feather are produced as waste (Korniłłowicz-Kowalska and Bohacz, 2011). These feather wastes which are made up of fibrous, insoluble, rigid structural keratins, are generally resistant to bacterial and enzymatic degradation (Anbu *et al.*, 2005). They are often disposed traditionally through processes such as burning, chemical treatments and land fillings, the methods that are not environmentally sustainable and also result in the destruction of some essential amino acids (Farag and Hassan, 2004; Rajput and Gupta, 2013). However, in order to dispose the feather wastes effectively and to harness their potentials for wealth, the need for an alternative safer method using microorganisms is highly necessary and is receiving attention from biotechnologists and related professions in the last few decades.

A number of feather degrading microorganisms (FDMs) with varying inherent capability to degrade feather and produce keratinase have been isolated and reported from different environmental sources (Anbu *et al.*, 2008; Desai *et al.*, 2010; Tatineni *et al.*, 2008). The growth of these FDMs and their ability to effectively produce keratinase and degrade feathers are grossly influenced by various physical and nutritional factors. Optimisation of these factors is therefore important in order to maximise their potential use. One-factorat-a-time (OFAT) optimisation technique is often used to achieved optimum keratinase production (Harde *et al.*, 2011), but the method has some limitations among which is lack of capability to address the interactive effects between different factors (Dutta *et al.*, 2004; Harde *et al.*, 2011; Okoroma *et al.*, 2012; Ramnani and Gupta, 2004). Statistical optimisation approaches are used to correct the limitations of the OFATs and have been used successfully in optimisation of many biological processes (Fakhfakh-zouari *et al.*, 2010).

Similarly, the continued use of chicken feathers as an economical and easy biosorbent material to remove heavy metals from surface water and effluents from industries has also resulted in generation of feather-laden heavy metals (secondary feather waste) which are also recalcitrant to degradation due to the toxic nature of heavy metals to microorganisms and also as the inhibitors of enzymatic activities (Cabrera *et al.*, 2006) thereby compounding biodegradation process. Due to tough and toxic nature of these feather wastes, there is a need for an alternative search for FDM that could withstand high concentrations of toxic metals and utilise the toxic wastes as a substrate to produce keratinase enzymes. The kind of such microorganism will be very important in the management of tonnes of secondary chemically polluted feather wastes and production of industrially important keratinase enzyme.

In the recent decades, white chicken feather wastes have become the major substrate of degradation in many studies that reported biodegradation of chicken feathers despite the

availability of other pigmented feathers (Gessesse *et al.*, 2003; Sahoo *et al.*, 2012). However, due to recent evidences which suggest that pigmented feathers are more harder to degrade by FDMs because of the presence of melanin pigments (Goldstein *et al.*, 2004; Gunderson and Frame, 2008; McGraw, 2006), melanised chicken feathers are not fully utilised to produce valuable products such as amino acids and keratinase and are still being disposed in traditional ways.

The use of immobilised cells as a microbial biocatalyst has gained attention over free cells due to higher efficiency of catalysis, operational stability, high cell density retention and reusability (Prakash *et al.*, 2010b). Production of keratinase and degradation of feather using free cells of *Bacillus*, fungi and *Streptomyces* in shake flasks using carrier such as alginate, k-carrageenan, agar and chitosan has been described. However, there is no report on the use of gellan gum as a carrier for the immobilisation of feather-degrading bacteria to produce keratinase.

In the view of the above, the study aim to isolate a native bacterium of Malaysia with a potential of withstanding toxicity of heavy metals and ability to degrade melanised feathers.

Thus, the objectives of this study are:

- 1. To isolate, identify and select heavy metal tolerant feather-degrading bacterium.
- 2. To optimise physical and nutritional conditions of the bacterium for maximum growth, keratinase production and feather degradation.
- 3. To immobilise the bacterium, optimise the immobilisation conditions and investigate degradation ability of the immobilised bacterial cells on melanised, non-melanised and mixed coloured feathers in comparison with free cells.
- 4. To determine the ability of free and immobilised bacterial cells to produce keratinase and degrade heavy metal polluted feather in batch and semi-continuous cultivation.
- 5. To characterise crude keratinase produced from free and immobilised cells and to determine their feather-degrading ability with or without reducing agents.
- 6. To analyse amino acid contents of hydrolysates produced from melanised and non-melanised by free cells of *Alcaligenes* sp. AQ05-001 at certain intervals during cultivation.

REFERENCES

- Acuña-Argüelles, M.E., Guti érez-Rojas, M., Viniegra-Gonz ález, G. and Favela-Torres, E. 1995. Production and properties of three pectinolytic activities produced by *Aspergillus niger* in submerged and solid-state fermentation. *Applied Microbiology and Biotechnology*, 43(5): 808–814.
- Adinarayana, K., Jyothi, B. and Ellaiah, P. 2005. Production of alkaline protease with immobilised cells of *Bacillus subtilis* PE-11 in various matrices by entrapment technique. AAPS PharmSciTech, 6(3): E391–397.
- Ahmad, S., Shamaan, N. and Arif, N. 2012. Enhanced phenol degradation by immobilised Acinetobacter sp. strain AQ5NOL 1. World Journal of Microbiology and Biotechnology, 28(1): 347–352.
- Akhtar, W. and Edwards, H. 1997. Fourier-transform Raman spectroscopy of mammalian and avian keratotic biopolymers. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy, 53(1): 81–90.
- Al-Asheh, S., Banat, F. and Al-Rousan, D. 2003. Beneficial reuse of chicken feathers in removal of heavy metals from wastewater. *Journal of Cleaner Production*, 11(3): 321–326.
- Al-Daghistani, H. 2012. Bio-remediation of Cu, Ni and Cr from rotogravure wastewater using immobilised, dead, and live biomass of indigenous thermophilic *Bacillus* species. *The Internet Journal of Microbiology*, 10(1).
- Alibardi, L. 2006. Cell structure of barb ridges in down feathers and juvenile wing feathers of the developing chick embryo: barb ridge modification in relation to feather evolution. *Annals of Anatomy*, 188(4): 303–318.
- Al-Musallam, A. 2013. Biodegradation of keratin in mineral-based feather medium by thermophilic strains of a new *Coprinopsis* sp. *International Biodeterioration and Biodegradation*, 79(42): 42–48.
- Altun, G.D. and Cetinus, S.A. 2007. Immobilization of pepsin on chitosan beads. Food Chemistry, 100(3): 964–971.
- Anbu, P., Gopinath, S.C.B., Hilda, A., Priya, T.L. and Annadurai, G. 2005. Purification of keratinase from poultry farm isolate-*Scopulariopsis* brevicaulis and statistical optimization of enzyme activity. *Enzyme and Microbial Technology*, 36(5–6): 639–647.
- Anbu, P., Gopinath, S. and Hilda, A. 2007. Optimization of extracellular keratinase production by poultry farm isolate *Scopulariopsis brevicaulis*. *Bioresource Technology*, 98(6): 1298–1303.

- Anbu, P., Hilda, A., Sur, H., Hur, B. and Jayanthi, S. 2008. Extracellular keratinase from *Trichophyton* sp. HA-2 isolated from feather dumping soil. *International Biodeterioration and Biodegradation*, 62: 287–292.
- Annapurna S.A., Singh, S.G.A., Kumar, A. and Kumar, H.K. 2012. Production and characterisation of thermo tolerant alkaline protease from Serratyia marcescens. *Asian Journal of Microbiology, Biotechnology and Environmental Science*, 14(4): 591–596.
- Anson, M.L. 1938. The estimation of pepsin, trypsin, papain, and cathepsin with hemoglobin. *The Journal of General Physiology*, 22(1): 79–89.
- Aquino de Souza, E., Rossi, D.M. and Z áchia Ayub, M.A. 2014. Bioconversion of residual glycerol from biodiesel synthesis into 1,3-propanediol using immobilized cells of *Klebsiella pneumoniae* BLh-1. *Renewable Energy*, 72: 253–257.
- Asahi, M., Lindquist, R. and Fukuyama, K. 1985. Purification and characterization of major extracellular proteinases from *Trichophyton rubrum*. *Biochemical Journal*, 232: 139–144.
- Awad, G.E.A., Esawy, M.A., Salam, W.A., Salama, B.M., Abdelkader, A.F. and Eldiwany, A. 2011. Keratinase production by *Bacillus pumilus* GHD in solid-state fermentation using sugar cane bagasse: optimisation of culture conditions using a Box-Behnken experimental design. *Annals of Microbiology*, 61(3): 663–672.
- Bach, E., Sant'Anna, V., Daroit, D.J., Corrêa, A.P.F., Segalin, J. and Brandelli, A. 2012. Production, one-step purification, and characterization of a keratinolytic protease from Serratia marcescens P3. Process Biochemistry, 47(12): 2455–2462.
- Baik, W., Bae, J., Cho, K. and Hartmeier, W. 2002. Biosorption of heavy metals using whole mold mycelia and parts thereof. *Bioresource Technology*, 81(3): 167–170.
- Balaji, S., Kumar, M. and Karthikeyan, R. 2008. Purification and characterization of an extracellular keratinase from a hornmeal-degrading *Bacillus subtilis* MTCC (9102). World Journal of Microbiology and Biotechnology, 24(11): 2741–2745.
- B dint, B., Bagi, Z., Tóth, A., R & Khely, G., Perei, K. and Kovács, K.L. 2005. Utilization of keratin-containing biowaste to produce biohydrogen. *Applied Microbiology and Biotechnology*, 69(4): 404–410.
- Banat, F. and Al-Asheh, S. 2000. Biosorption of phenol by chicken feathers. *Environmental Engineering and Policy*, 2(2): 85–90.
- Barbara, B. 2009. Fungi utilizing keratinous substrates. *International Biodeterioration and Biodegradation*, 63: 631–653.
- Barrowclough, G. and Sibley, F. 1980. Feather pigmentation and abrasion: test of a hypothesis. *The Auk*, 881–883.

- Baş, D. and Boyacı, İ.H. 2007. Modelling and optimization I: Usability of response surface methodology. *Journal of Food Engineering*, 78(3): 836–845.
- Ben Khedher, S., Jaoua, S. and Zouari, N. 2013. Application of statistical experimental design for optimisation of bioinsecticides production by sporeless *Bacillus thuringiensis* strain on cheap medium. *Brazilian Journal of Microbiology*, 44(3): 927–933.
- Bernal, C., Diaz, I. and Coello, N. 2006a. Response surface methodology for the optimization of keratinase production in culture medium containing feathers produced by *Kocuria rosea*. *Canadian Journal of Microbiology*, 52(5): 445–450.
- Bernal, C., Cairó, J. and Coello, N. 2006b. Purification and characterization of a novel exocellular keratinase from *Kocuria rosea*. *Enzyme and Microbial Technology*, 38(1–2): 49–54.
- Bertsch, A. and Coello, N. 2005. A biotechnological process for treatment and recycling poultry feathers as a feed ingredient. *Bioresource Technology*, 96(15): 1703–1708.
- Bezerra, M., Santelli, R. and Oliveira, E. 2008. Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 76(5): 965–977.
- Box, G.E.P. and Behnken, D.W. 1960. Some new three-level designs for the study of quantitative Variables. *Technometrics*, 2(4): 455–475.
- Box, G., Hunter, W. and Hunter, J. 1978. *Statistics for experimenters*. John Wiley and Sons, New York. pp.144.
- Box, D.B. 1960. Simplex-Sum Designs: A class of second order rotatable designs derivable from those of first order. *The Annals of Mathematical Statistics*, 31(4): 838–864.
- Bragulla, H.H. and Homberger, D.G. 2009. Structure and functions of keratin proteins in simple, stratified, keratinized and cornified epithelia. *Journal of Anatomy*, 214(4): 516–519.
- Brebu, M. and Spiridon, I. 2011. Thermal degradation of keratin waste. *Journal of Analytical and Applied Pyrolysis*, 91(2): 288–295.
- Brenner, D.J., Krieg, N., Garrity, G. and Staley, J. 2005. Bergey's manual of systematic bacteriology. Volume 2: The proteobacteria, Part B: The Gammaproteobacteria. Springer, pp 323–376.
- Bressollier, P. and Letourneau, F. 1999. Purification and characterization of a keratinolytic serine proteinase from *Streptomyces albidoflavus*. *Applied and Environmental Microbiology*, 65(6): 2570–2576.
- Burtt, E.H., Schroeder, M.R., Smith, L.A., Sroka, J.E. and McGraw, K.J. 2011. Colourful parrot feathers resist bacterial degradation. *Biology Letters*, 7(2): 214–216.

- Butler, M. 2004. Are melanized feather barbs stronger? *Journal of Experimental Biology*, 207(2): 285–293.
- Cabrera, G., Pérez, R., Gómez, J.M., Abalos, A. and Cantero, D. 2006. Toxic effects of dissolved heavy metals on *Desulfovibrio vulgaris* and *Desulfovibrio* sp. strains. *Journal of Hazardous Materials*, 135(1-3): 40–46.
- Cai, C., Lou, B. and Zheng, X. 2008. Keratinase production and keratin degradation by a mutant strain of *Bacillus subtilis*. *Journal of Zhejiang University of science B*, 9(1): 60–67.
- Cai, C. and Zheng, X. 2009. Medium optimization for keratinase production in hair substrate by a new *Bacillus subtilis* KD-N2 using response surface methodology. *Journal of Industrial Microbiology and Biotechnology*, 36(7): 875–883.
- Cameron, G., Wess, T. and Bonser, R. 2003. Young's modulus varies with differential orientation of keratin in feathers. *Journal of Structural Biology*, 143(2): 118–123.
- C ánovas, D. 2003. Heavy metal tolerance and metal homeostasis in *Pseudomonas putida* as revealed by complete genome analysis. *Environmental Microbiology*, 5(12): 1242–1256.
- Cao, Z.J., Zhang, Q., Wei, D.K., Chen, L., Wang, J., Zhang, X.Q. and Zhou, M.H. 2009. Characterization of a novel *Stenotrophomonas* isolate with high keratinase activity and purification of the enzyme. *Journal of Industrial Microbiology and Biotechnology*, 36(2): 181–188.
- Cappuccino, J.G. and Sherman, N. 1996. *Microbiology-A laboratory manual (Vol. 9)*. Benjamin Cummings Science Publishing, Carlifonia. p 471.
- Casas Lopez, J.L., Sanchez Perez, J.A., Fernandez Sevilla, J.M., Acien Fernandez, F.G. and Molina Grima, Y.C. 2004. Fermentation optimization for the production of lovastatin by *Aspergillus terreus*: use of response surface methodology. *Journal of Chemical Technology and Biotechnology*, 79(10): 1119–1126.
- Cave, M., Appana, S., Patel, M., Falkner, K.C., McClain, C.J. and Brock, G. 2010. Polychlorinated biphenyls, lead, and mercury are associated with liver disease in American adults: NHANES 2003-2004. *Environmental health perspectives*, 118(12): 1735.
- Cavello, I.A., Hours, R.A., Rojas, N.L. and Cavalitto, S.F. 2013. Purification and characterization of a keratinolytic serine protease from *Purpureocillium lilacinum* LPS # 876. *Process Biochemistry*, 48(5-6): 972–978.
- Charney, J. and Tomarelli, R. 1947. A colorimetric method for the determination of the proteolytic activity of duodenal juice. *Journal of Biological Chem*istry, 171(2): 501–505.

- Chaturvedi, V., Bhange, K., Bhatt, R. and Verma, P. 2014. Biocatalysis and Agricultural Biotechnology Production of kertinases using chicken feathers as substrate by a novel multifunctional strain of *Pseudomonas stutzeri* and its dehairing application. *Biocatalysis and Agricultural Biotechnology*, 3(2): 167–174.
- Chaudhari, P.N., Chaudhari, B.L. and Chincholkar, S.B. 2013. Iron containing keratinolytic metallo-protease produced by *Chryseobacterium gleum*. *Process Biochemistry*, 48(1): 144–151.
- Chen, P., Pickard, M.A. and Gray, M.R. 2000. Surfactant inhibition of bacterial growth on solid anthracene. *Biodegradation*, 11(5): 341–347.
- Cheng, S., Hu, H. and Shen, S. 1995. Production and characterization of keratinase of a feather-degrading *Bacillus licheniformis* PWD-1. *Bioscience Biotechnology and Biochemistry*, 59: 2239–2239.
- Chitte, R., Nalawade, V. and Dey, S. 1999. Keratinolytic activity from the broth of a feather-degrading thermophilic *Streptomyces thermoviolaceus* strain SD8. *Letters in Applied Microbiology*, 28(2): 131–136.
- Corr êa, A.P.F., Daroit, D.J. and Brandelli, A. 2010. Characterization of a keratinase produced by *Bacillus* sp. P7 isolated from an Amazonian environment. *International Biodeterioration and Biodegradation*, 64(1): 1–6.
- Das, N., Vimala, R. and Karthika, P. 2008. Biosorption of heavy metals—an overview. *Indian Journal of Biotechnology*, 7(2): 159–169.
- Davis, K.E.R., Joseph, S.J. and Janssen, P.H. 2005. Effects of growth medium, inoculum size, and incubation time on culturability and isolation of soil bacteria. *Applied and Environmental Microbiology*, 71(2): 826–834.
- De Azeredo, L.A.I., De Lima, M.B., Coelho, R.R.R. and Freire, D.M.G. 2006. Thermophilic protease production by *Streptomyces* sp. 594 in submerged and solid-state fermentations using feather meal. *Journal of Applied Microbiology*, 100(4): 641–647.
- De, J. and Ramaiah, N. 2007. Characterization of marine bacteria highly resistant to mercury exhibiting multiple resistances to toxic chemicals. *Ecological Indicators*, 7(3): 511–520.
- De, J., Ramaiah, N. and Vardanyan, L. Detoxification of toxic heavy metals by marine bacteria highly resistant to mercury. *Marine Biotechnology*, 10(4): 471–477.
- Deivasigamani, B. and Alagappan, K. 2008. Industrial application of keratinase and soluble proteins from feather keratins. *Journal of Environmental Biology*, 29(6): 933–936.

- Demir, T., Hameş, E.E., Öncel, S.S. and Vardar-Sukan, F. 2015. An optimization approach to scale up keratinase production by *Streptomyces* sp. 2M21 by utilizing chicken feather. *International Biodeterioration and Biodegradation*, 103: 134–140.
- Desai, K., Survase, S. and Saudagar, P. 2008. Comparison of artificial neural network (ANN) and response surface methodology (RSM) in fermentation media optimization: case study of fermentative production of. *Biochemical Engineering Journal*, 41(3): 266–273.
- Desai, S.S., Hegde, S., Inamdar, P., Sake, N. and Aravind, M.S. 2010. Isolation of keratinase from bacterial isolates of poultry soil for waste degradation. *Engineering in Life Sciences*, 10(4): 361–367.
- Diaz-Godinez, G. and Soriano-Santos, J. 2001. Exopectinases produced by Aspergillus niger in solid-state and submerged fermentation: a comparative study. *Journal of Industrial Microbiology and Biotechnology*, 26(5): 271–277.
- D áz-Raviña, M., B ååh, E. and Frosteg åd, A. 1994. Multiple heavy metal tolerance of soil bacterial communities and its measurement by a thymidine incorporation technique. *Applied and Environmental Microbiology*, 60(7): 2238–2247.
- Dozie, I., Okeke, C. and Unaeze, N. 1994. A thermostable, alkaline-active, keratinolytic proteinase from *Chrysosporium keratinophilum*. *World Journal of Microbiology and Biotechnology*, 10(5): 563–567.
- Dutta, J. R., Dutta, P. K. and Banerjee, R. 2004. Optimization of culture parameters for extracellular protease production from a newly isolated *Pseudomonas* sp. using response surface and artificial neural network models. *Process Biochemistry*, 39(12): 2193–2198.
- Duxbury, T. 1981. Toxicity of heavy metals to soil bacteria. *FEMS Microbiology Letters*, 11(2-3): 217–220.
- El-Metwally, M. and El-Hersh, M. 2010. Keratinase Production and Bioclegradation of Some Keratinous Wastes by *Alternaria tenuissima* and *Aspergillus nidulans*. *Research Journal of Microbiology*, 5(1): 21–35.
- El-Refai, H.A., AbdelNaby, M.A., Gaballa, A., El-Araby, M.H. and Abdel Fattah, A. F. 2005. Improvement of the newly isolated *Bacillus pumilus* FH9 keratinolytic activity. *Process Biochemistry*, 40(7): 2325–2332.
- Eslahi, N., Hemmatinejad, N. and Dadashian, F. 2014. From Feather Waste to Valuable Nanoparticles. *Particulate Science and Technology*, 32(3): 242–250.
- Fakhfakh, N., Ktari, N., Haddar, A., Mnif, I. H., Dahmen, I. and Nasri, M. 2011. Total solubilisation of the chicken feathers by fermentation with a keratinolytic bacterium, *Bacillus pumilus* A1 and the production of protein hydrolysate with high antioxidative activity. *Process Biochemistry*, 46(9): 1731–1737.

- Fakhfakh-zouari, N., Haddar, A., Hmidet, N., Frikha, F. and Nasri, M. 2010. Application of statistical experimental design for optimization of keratinases production by *Bacillus pumilus* A1 grown on chicken feather and some biochemical properties. *Process Biochemistry*, 45(5): 617–626.
- Fang, Z., Zhang, J., Liu, B., Du, G. and Chen, J. 2013. Biodegradation of wool waste and keratinase production in scale-up fermenter with different strategies by *Stenotrophomonas maltophilia. Bioresource Technology*, 140: 286–291.
- Farag, A.M. and Hassan, M.A. 2004. Purification, characterization and immobilization of a keratinase from Aspergillus oryzae. Enzyme and Microbial Technology, 34(2): 85–93.
- Farinas, C.S. 2015. Developments in solid-state fermentation for the production of biomass-degrading enzymes for the bioenergy sector. *Renewable and Sustainable Energy Reviews*, 52: 179–188.
- Fraser, R.D.B. and Parry, D.A.D. 2014. Reprint of: keratin intermediate filaments: differences in the sequences of the Type I and Type II chains explain the origin of the stability of an enzyme-resistant four-chain fragment. *Journal of Structural Biology*, 186(3): 481–490.
- Fraser, R. and MacRae, T. 1962. An investigation of the structure of β-keratin. *Journal of Molecular Biology*, 5(5): 457–515.
- Friedrich, A. and Antranikian, G. 1996. Keratin Degradation by *Fervidobacterium pennavorans*, a Novel Thermophilic Anaerobic Species of the Order Thermotogales. *Applied and Environmental Microbiology*, 62(8): 2875–2882.
- Friedrich, J., Gradisar, H., Mandin, D. and Chaumont, J. P. 1999. Screening fungi for synthesis of keratinolytic enzymes. *Letters in Applied Microbiology*, 28(2): 127– 130.
- Gadd, G.M. 2010. Metals, minerals and microbes: geomicrobiology and bioremediation. *Microbiology*, 156(3): 609–643.
- Gao, S., Wang, Y., Diao, X., Luo, G. and Dai, Y. 2010. Effect of pore diameter and crosslinking method on the immobilization efficiency of *Candida rugosa* lipase in SBA-15. *Bioresource technology*, 101(11): 3830–3837.
- Gavrilescu, M. 2004. Removal of heavy metals from the environment by biosorption. *Chemistry*, 28: 219–232.
- Geissmann, Q. (2013). OpenCFU, a new free and open-source software to count cell colonies and other circular objects. *PloS One*, 8(2): e54072.
- Gennis, R. 2013. *Biomembranes: molecular structure and function*. Springer-Verlag, New York. pp. 535.

- Gessesse, A., Hatti-Kaul, R., Gashe, B.A. and Mattiasson, B. 2003. Novel alkaline proteases from alkaliphilic bacteria grown on chicken feather. *Enzyme and Microbial Technology*, 32(5): 519–524.
- Ghasemi, Y., Shahbazi, M., Rasoul-Amini, S., Kargar, M., Safari, A., Kazemi, A. and Montazeri-Najafabady, N. 2012. Identification and characterization of featherdegrading bacteria from keratin-rich wastes. *Annals of Microbiology*, 62(2): 737– 744.
- Ghosh, A., Chakrabarti, K. and Chattopadhyay, D. 2008. Degradation of raw feather by a novel high molecular weight extracellular protease from newly isolated *Bacillus cereus* DCUW. *Journal of Industrial Microbiology Biotechnology*, 35(8): 825– 834.
- Gioppo, N., Moreira-Gasparin, F., Costa, A., Alexandrino, A., Souza, C. and Peralta, R. 2009. Influence of the carbon and nitrogen sources on keratinase production by *Myrothecium verrucaria* in submerged and solid state cultures. *Journal of Industrial Microbiology and Biotechnology*, 36(5): 705–711.
- Goldstein, G., Flory, K.R.K., Browne, B.B.A., Majid, S., Ichida, J. M. and Burtt Jr., E. H. 2004. Bacterial degradation of black and white feathers. *The Auk*, 121(3): 656–659.
- Gradisar, H., Friedrich, J., Krizaj, I. and Jerala, R. 2005. Similarities and specificities of fungal keratinolytic proteases: comparison of keratinases of *Paecilomyces marquandii* and *Doratomyces microsporus* to some known proteases. *Applied and Environmental Microbiology*, 71(7): 3420–3426.
- Grande, J.M., Negro, J.J. and Torres, M.J. 2004. The Evolution of bird plumage colouration: a role for feather-degradation bacteria? *Ardeola*, 51(2): 375–383.
- Grazziotin, A., Pimentel, F.A., Sangali, S., de Jong, E.V, and Brandelli, A. 2007. Production of feather protein hydrolysate by keratinolytic bacterium *Vibrio sp.* kr2. *Bioresource Technology*, 98(16): 3172–3175.
- Gunderson, A. 2008. Feather-degrading bacteria: a new frontier in avian and host-parasite research? *The Auk*, 124(4): 972–979.
- Gunderson, A. and Frame, A. 2008. Resistance of melanized feathers to bacterial degradation: is it really so black and white? *Journal of Avian Biology*, 39(5): 539–545.
- Guo, H., Luo, S., Chen, L., Xiao, X., Xi, Q., Wei, W. and He, Y. 2010. Bioremediation of heavy metals by growing hyperaccumulaor endophytic bacterium *Bacillus* sp. L14. *Bioresource technology*, 101(22): 8599–8605.
- Gupta, A. and Khare, S. 2006. A protease stable in organic solvents from solvent tolerant strain of *Pseudomonas aeruginosa*. *Bioresource Technology*, 97(15): 1788–1793.

- Gupta, A., Roy, I., Patel, R.K., Singh, S.P., Khare, S.K. and Gupta, M. N. 2005. One-step purification and characterization of an alkaline protease from haloalkaliphilic *Bacillus* sp. *Journal of Chromatography A*, 1075(1-2): 103–108.
- Gupta, R., Rajput, R. and Sharma, R. 2013. Biotechnological applications and prospective market of microbial keratinases. *Applied Microbiology and Biotechnology*, 97(23): 9931–9940.
- Gupta, R. and Ramnani, P. 2006. Microbial keratinases and their prospective applications: an overview. *Applied Microbiology and Biotechnology*, 70(1): 21–33.
- Gupta, R. and Sharma, R. 2013. Revisiting microbial keratinases Next generation proteases for sustainable biotechnology. *Critical Reviews in Biotechnology*, 33(2): 216–228.
- Gurav, R.G. and Jadhav, J.P. 2013. Biodegradation of keratinous waste by *Chryseobacterium* sp. RBT isolated from soil contaminated with poultry waste. *Journal of Basic Microbiology*, 53(2): 128–135.
- Habbeche, A., Saoudi, B., Jaouadi, B., Haberra, S., Kerouaz, B., Boudelaa, M., Ladjama,
 A. 2013. Purification and biochemical characterization of a detergent-stable keratinase from a newly thermophilic actinomycete *Actinomadura keratinilytica* strain Cpt29 isolated from poultry compost. *Journal of Bioscience and Bioengineering*, 117(4): 413–421.
- Han, M., Luo, W., Gu, Q. and Yu, X. 2012. Isolation and characterization of a keratinolytic protease from a feather-degrading bacterium *Pseudomonas* aeruginosa C11. African Journal of Microbiology Research, 6(9): 2211–2221.
- Hanahan, D. and Weinberg, R.A. 2011. Hallmarks of cancer: the next generation. *Cell*, 144(5): 646–674.
- Harada, T., Fujimori, K., Hirose, S. and Masada, M. 2014. Growth and β-Glucan 10C3K Production by a Mutant of Alcaligenes faecalis var. myxogenes in defined medium. Agricultural and Biological Chemistry, 30(8): 764–769.
- Harde, S., Bajaj, I. and Singhal, R. 2011. Optimization of Fermentative production of keratinase from *Bacillus subtilis* NCIM 2724. *Agriculture, Food and Analytical Bacteriology*, 1(1): 54–65.
- Hassan, M. and Haroun, B. 2013. Production and characterization of keratinolytic protease from new wool-degrading *Bacillus* species isolated from Egyptian ecosystem. *BioMed Research International*. doi.org/10.1155/2013/175012.
- Hassen, A., Saidi, N., Cherif, M. and Boudabous, A. 1998. Resistance of environmental bacteria to heavy metals. *Bioresource Technology*, 64(1): 7–15.
- Hattori, R. 1972. Growth of *Escherichia coli* on the surface of an anion-exchange resin in continuous flow system. *The Journal of General and Applied Microbiology*, 18(5): 319–330.

- Hazan, R., Que, Y.A., Maura, D. and Rahme, L.G. 2012. A method for high throughput determination of viable bacteria cell counts in 96-well plates. *BMC Microbiology*, 12(1): 259.
- Ignatova, Z. and Gousterova, A. 1999. Isolation and partial characterisation of extracellular keratinase from a wool degrading thermophilic actinomycete strain *Thermoactinomyces candidus*. *Canadian Journal of Microbiology*, 45(3): 217–222.
- Iversen, S. and Jørgensen, M. 1995. Azocasein assay for alkaline protease in complex fermentation broth. *Biotechnology Techniques*, 9(8): 573–576.
- Jaouadi, B., Abdelmalek, B. and Fodil, D. 2010. Purification and characterization of a thermostable keratinolytic serine alkaline proteinase from *Streptomyces* sp. strain AB1 with high stability in organic. *Bioresource technology*, 101(21): 8361–8369.
- Jeong, J.H., Jeon, Y.D., Lee, O.M., Kim, J.D., Lee, N.R., Park, G.T. and Son, H.J. 2010. Characterization of a multifunctional feather-degrading *Bacillus subtilis* isolated from forest soil. *Biodegradation*, 21(6): 1029–1040.
- Jiang, L. 2013. Effect of nitrogen source on curdlan production by Alcaligenes faecalis ATCC 31749. International Journal of Biological Macromolecules, 52: 218– 220.
- Jeong J.H., DongJoo Oh, D.Y.H, Hong-Sung Kim, K.H.P. and Chung-Yeol Lee, H.J.S. 2010. Keratinolytic enzyme-mediated biodegradation of recalcitrant featherby a newly isolated *Xanthomonas* sp. P5. *Polymer Degradation and Stability*, 95: 1969–1977.
- Joshi, S., Tejashwini, M. and Revati, N. 2007. Isolation, identification and characterization of a feather degrading bacterium. *International Journal of Poultry Science*, 6(9): 689–693.
- Seifert, K.F.D. 2005. Inhibiting effect of surfactants and heavy metal ions on the denitrification process. *Polish Journal of Environmental Studies*, 14(1): 87–93.
- Kar, P. and Misra, M. 2004. Use of keratin fiber for separation of heavy metals from water. *Journal of Chemical Technology and Biotechnology*, 79(11): 1313– 1319.
- Karthikeyan, R., Balaji, S. and Sehgal, P. 2007. Industrial applications of keratins-a review. *Journal of Scientific and Industrial Research*, 66(9): 710–715.
- Kaur, V. and Bera, M. 2013. Production and Characterization of Exopolysaccharide Produced by Alcaligenes faecalis B14 Isolated from Indigenous Soil. International Journal of Biotechnology Bioengineering research, 4: 365–367.
- Kazilek, C.J. 2009. Feathers | ASU Ask A Biologist. Retrieved November 23, 2015, from https://askabiologist.asu.edu/explore/feather-biology.

- Khardenavis, A.A., Kapley, A. and Purohit, H.J. 2009. Processing of poultry feathers by alkaline keratin hydrolyzing enzyme from *Serratia* sp. HPC 1383. *Waste management*, 29(4): 1409–1415.
- Kennedy, M. and Krouse, D. 1999. Strategies for improving fermentation medium performance: a review. *Journal of Industrial Microbiology and Biotechnology*, 23(6): 456–475.
- Khardenavis, A.A., Kapley, A. and Purohit, H.J. 2009. Processing of poultry feathers by alkaline keratin hydrolyzing enzyme from *Serratia* sp. HPC 1383. *Waste Management*, 29(4): 1409–1415.
- Khayat, M.E. (2011). Isolation, production and characterization of keratinase from Bacillus sp. khayat (Doctoral dissertation, Universiti Putra Malaysia).
- Khuri, A., Mukherjee, B., Sinha, B. and Ghosh, M. 2006. Design issues for generalized linear models: A review. *Statistical Science*. 21(3): 376–399.
- Kierstan, M. and Bucke, C. 2000. The immobilization of microbial cells, subcellular organelles, and enzymes in calcium alginate gels. Reprinted from Biotechnology and Bioengineering, Vol. XIX, No. 3, Pages 387–397 (1977). *Biotechnology and Bioengineering*, 67(6): 726–736.
- Kim, J., Kluskens, L., de Vos, W., Huber, R. and van der Oost, J. 2004. Crystal structure of fervidolysin from *Fervidobacterium pennivorans*, a keratinolytic enzyme related to subtilisin. *Journal of Molecular Biology*, 335: 787–797.
- Kim, J., Lim, W. and Suh, H. 2001. Feather-degrading *Bacillus* species from poultry waste. *Process Biochemistry*, 37: 287–291.
- Kitada, M. and Horikoshi, K. 1987. Bioenergetic properties of alkalophilic *Bacillus* sp. strain C-59 on an alkline medium containing K2CO3. *Journal of Bacteriology*, 169(12): 5761–5765.
- Kopsahelis, N., Panas, P., Kourkoutas, Y. and Koutinas, A.A. 2007. Evaluation of the thermally dried immobilized cells of *Lactobacillus delbrueckii* subsp. Bulgaricus on apple pieces as a potent starter culture. *Journal of agricultural and food chemistry*, 55(24): 9829–9836.
- Korkmaz, H., Hür, H. and Di, S. 2004. Characterization of alkaline keratinase of *Bacillus licheniformis* strain HK-1 from poultry waste. *Ann Microbiol*, 54(2): 201–211.
- Korniłłowicz-Kowalska, T. and Bohacz, J. 2011. Biodegradation of keratin waste: Theory and practical aspects. *Waste Management*, 31(8): 1689–1701.
- Kowata, K., Nakaoka, M., Nishio, K., Fukao, A., Satoh, A., Ogoshi, M., Takeuchi, S. 2014. Identification of a feather β-keratin gene exclusively expressed in pennaceous barbule cells of contour feathers in chicken. *Gene*, 542(1): 23–28.

- Kumar, E. 2011. Biodegradation of poultry feathers by a novel bacterial isolate Bacillus altitudinis GVC 11. Indian Journal of Biotechnology, 10(10): 502–507.
- Kumar, E., Srijana, M. and Kumar, K. 2011. A novel serine alkaline protease from Bacillus altitudinis GVC11 and its application as a dehairing agent. Bioprocess and Biosystems Engineering, 34(4): 403–409.
- Kumar, R., Balaji, S., Uma, T.S., Mandal, A.B. and Sehgal, P.K. 2010. Optimization of Influential Parameters for Extracellular Keratinase Production by *Bacillus subtilis* (MTCC9102) in Solid State Fermentation Using Horn Meal—A Biowaste Management. *Applied Biochemistry and Biotechnology*, 160(1): 30– 39.
- Kushwaha, R. 1983. The in vitro degradation of peacock feathers by some fungi/der invitro-Abbau von Pfauenfedern durch einige Pilze. *Mycoses*, 26(6): 324–326.
- Laba, W. and Rodziewicz, A. 2014. Biodegradation of hard keratins by two *Bacillus* strains. *Jundishapur Journal of Microbiology*, 7(2): e8896.
- Laba, W. and Szczekala, K.B. 2013. Keratinolytic proteases in biodegradation of pretreated feathers. *Polish Journal of Environmental Studies*, 4(22): 1101–1109.
- Lasekan, A., Abu, F. and Hashim, D. 2013. Potential of chicken by-products as sources of useful biological resources. *Waste Management*, 33(3): 552–565.
- Ledoux, M. and Lamy, F. 1986. Determination of proteins and sulfobetaine with the folinphenol reagent. *Analytical Biochemistry*, 157(1): 28–31.
- Lee, H. and Craig, J. 1991. Beak trimming effects on behavior patterns, fearfulness, feathering, and mortality among three stocks of White Leghorn pullets in cages or floor pens. *Poultry Science*, 70(2): 211–221.
- Liang, J., Han, Y. and Zhang, J. 2011. Optimal culture conditions for keratinase production by a novel thermophilic *Myceliophthora thermophila* strain GZUIFR-H49-1. *Journal of Applied Microbiology*, 110(4): 871–880.
- Lima de Silva, A.A., de Carvalho, M.A.R., de Souza, S.A.L., Dias, P.M.T., da Silva Filho, R.G., de Meirelles Saramago, C.S., Hofer, E. 2012. Heavy metal tolerance (Cr, Ag AND Hg) in bacteria isolated from sewage. *Brazilian Journal of Microbiology*, 43(4): 1620–1631.
- Lin, X., Lee, C.G., Casale, E.S. and Shih, J.C.H. 1992. Purification and Characterization of a Keratinase from a Feather-Degrading *Bacillus licheniformis* Strain. *Applied Environment and Microbiology*, 58(10): 3271–3275.
- Liu, J., Huang, X.F., Lu, L.J., Xu, J. C., Wen, Y., Yang, D.H. and Zhou, Q. 2010. Optimization of biodemulsifier production from *Alcaligenes* sp. S-XJ-1 and its application in breaking crude oil emulsion. *Journal of Hazardous Materials*, 183(1-3): 466–473.

- Lo, W.H., Too, J.R. and Wu, J.Y. 2012. Production of keratinolytic enzyme by an indigenous feather-degrading strain *Bacillus cereus* Wu2. *Journal of Bioscience* and Bioengineering, 114(6): 640–647.
- Lowry, O.C. and Rosebrough, N. 1951. Protein measurement with the Folin phenol reagent. *Journal of Biological Chemistry*, 193: 265.
- Lv, L.X., Sim, M.H., Li, Y.D. Y.Q., Min, J., Feng, W.H., Guan, W.J., Lv, Long-Xian, Ming-Hao Sim, Yu-Dong Li, Jie Min, Wei-Hong Feng, Wen-Jun Guan, and Y.-Q. L. 2010. Production, characterization and application of a keratinase from *Chryseobacterium* L99 sp. nov. *Process Biochemistry*, 45(8): 1236–1244.
- Madigan, M., Martinko, J., Parker, J. and Brock, T. (1997). *Biology of microorganisms*. Brock Biolog ú de los microorganismos (Vol. 10). Prentice hall, Barcelona. p 992.
- Malik, A. 2004. Metal bioremediation through growing cells. *Environment International*, 30(2): 261–278.
- Malviya, H.K., Rajak, R.C. and Hasija, S.K. 1992. Synthesis and regulation of extracellular keratinase in three fungi isolated from the grounds of a gelatin factory, Jabalpur, India. *Mycopathologia*, 120(1): 1–4.
- Marcondes, N.R., Taira, C.L., Vandresen, D.C., Inez, T., Svidzinski, E., Kadowaki, M.K. and Peralta, R.M. 2008. New feather degrading filamentous fungi. *Microbial Ecology*, 56(1): 13–17.
- Margesin, R. and Schinner, F. 1991. Characterization of a metalloprotease from psychrophilic Xanthomonas maltophilia. FEMS Microbiology Letters, 79(2): 257–261.
- Mazotto, A.M., Coelho, R.R.R., Cedrola, S.M.L., de Lima, M.F., Couri, S., Paraguai de Souza, E. and Vermelho, A.B. 2011. Keratinase production by three bacillus spp. Using feather meal and whole feather as substrate in a submerged fermentation. *Enzyme research*, 2011: doi.org/10.4061/2011/523780.
- Mazotto, A.M., Sonia, C., M ônica, C.T. and Damaso, A.B.V. 2013. Degradation of feather waste by *Aspergillus niger* keratinases : Comparison of submerged and solidstate fermentation. *International Biodeterioration and Biodegradation*, 85: 189– 195.
- McGraw, K. 2006. Mechanics of uncommon colors: pterins, porphyrins, and psittacofulvins. Bird coloration. 1: 354–398.
- McGraw, K.J., Mackillop, E.A., Dale, J. and Hauber, M.E. 2002. Different colors reveal different information: how nutritional stress affects the expression of melaninand structurally based ornamental plumage. *Journal of Experimental Biology*, 205(23): 3747–3755.

- McGraw, K.J., Safran, R.J. and Kakamatsu, K. 2005. How feather colour reflects its melanin content. *Functional Ecology*, 19(5): 816–821.
- Mohamedin, A. 1999. Isolation, identification and some cultural conditions of a proteaseproducing thermophilic *Streptomyces* strain grown on chicken feather as a substrate. *International Biodeterioration and Biodegradation*, 43(1-2): 13–21.
- Mohana, S. and Shrivastava, S. 2008. Response surface methodology for optimization of medium for decolorization of textile dye Direct Black 22 by a novel bacterial consortium. *Bioresource Technology*, 99(3): 562–569.
- Moorthy, I.M.G. and Baskar, R. 2013. Statistical modeling and optimization of alkaline protease production from a newly isolated alkalophilic *Bacillus* species BGS using response surface methodology and genetic algorithm. *Preparative Biochemistry and Biotechnology*, 43(3): 293–314.
- Mordasini, T., Curioni, A. and Andreoni, W. 2003. Why do divalent metal ions either promote or inhibit enzymatic reactions? The case of BamHI restriction endonuclease from combined quantum-classical simulations. *Journal of Biological Chemistry*, 278(7): 4381–4384.
- Morris, E.R., Nishinari, K. and Rinaudo, M. 2012. Gelation of gellan A review. *Food Hydrocolloids*, 28(2): 373–411.
- Moslemy, P., Neufeld, R.J. and Guiot, S.R. 2002. Biodegradation of gasoline by gellan gum-encapsulated bacterial cells. *Biotechnology and Bioengineering*, 80(2): 175–184.
- Moslemy, P., Neufeld, R.J., Millette, D. and Guiot, S.R. 2003. Transport of gellan gum microbeads through sand: an experimental evaluation for encapsulated cell bioaugmentation. *Journal of Environmental Management*, 69(3): 249–259.
- Mukherjee, A.K., Rai, S.K. and Bordoloi, N.K. 2011. Biodegradation of waste chickenfeathers by an alkaline b -keratinase (Mukartinase) purified from a mutant *Brevibacillus* sp. strain AS-S10-II. *International Biodeterioration and Biodegradation*, 65(8): 1229–1237.
- Muñoz, A.J., Ruiz, E., Abriouel, H., G álvez, A., Ezzouhri, L., Lairini, K. and Esp ńola, F.
 2012. Heavy metal tolerance of microorganisms isolated from wastewaters: Identification and evaluation of its potential for biosorption. *Chemical Engineering Journal*, 210: 325–332.
- Myers, R.H., Montgomery, D.C. and Anderson-Cook, C.M. 2009. *Response surface methodology: process and product optimization using designed experiments* (Vol. 705). John Wiley and Sons, New York. p 680.
- Naveau, A., Seidel, K. and Klein, O.D. 2014. Tooth, hair and claw: comparing epithelial stem cell niches of ectodermal appendages. *Experimental Cell Research*, 325(2): 96–103.

- Ni, H., Chen, Q., Chen, F., Fu, M., Dong, Y. and Cai, H. 2013. Improved keratinase production for feather degradation by *Bacillus licheniformis* ZJUEL31410 in submerged cultivation. *African Journal of Biotechnology*, 10(37): 7236–7244.
- Nickerson, W.J., Noval, J.J. and Robison, R.S. 1963. Keratinase: I. Properties of the enzyme conjugate elaborated by *Streptomyces fradiae*. *Biochimica et Biophysica Acta*, 77(0): 73–86.
- Noval, J. and Nickerson, W. 1959. Decomposition of native keratin by *Streptomyces fradiae*. *Journal of Bacteriology*, 77(3): 251–263.
- Nustorova, M., Braikova, D., Gousterova, A., Vasileva-Tonkova, E. and Nedkov, P. 2006. Chemical, microbiological and plant analysis of soil fertilized with alkaline hydrolysate of sheep's wool waste. *World Journal of Microbiology and Biotechnology*, 22(4): 383–390.
- Nys, R. De, Steinberg, P. and Willemsen, P. 1995. Broad spectrum effects of secondary metabolites from the red alga *Delisea pulchra* in antifouling assays. *Biofouling*, 8(4): 259–271.
- Odetallah, N., Wang, J., Garlich, J. and Shih, J. 2003. Keratinase in starter diets improves growth of broiler chicks. *Poultry Science*, 82(4): 664–670.
- Ogbonna, J., Amano, Y. and Nakamura, K. 1989. Elucidation of optimum conditions for immobilization of viable cells by using calcium alginate. *Journal of Fermentation and Bioengineering*, 67(2): 92–96.
- Oh, S., Yun, A. and Park, D. 2011. Effects of physicochemically hydrolyzed human hairs on the soil microbial community and growth of the hot pepper plant. *Biotechnology and Bioprocess Engineering*, 16(4): 746–754.
- Okoroma, E. A., Garelick, H., Abiola, O.O. and Purchase, D. 2012. Identi fi cation and characterisation of a *Bacillus licheniform*is strain with profound keratinase activity for degradation of melanised feather. *International Biodeterioration and Biodegradation*, 74: 54–60.
- Oliveira, J.T., Martins, L., Picciochi, R., Malafaya, P.B., Sousa, R.A., Neves, N.M., Reis, R.L. 2010. Gellan gum: a new biomaterial for cartilage tissue engineering applications. *Journal of Biomedical Materials Research. Part A*, 93(3): 852–863.
- Olsen, M.E., Bengtsson, C.F., Bertelsen, M.F., Willerslev, E. and Gilbert, M.T.P. 2012. DNA from keratinous tissue. Part II: feather. *Annals of Anatomy = Anatomischer Anzeiger : Official Organ of the Anatomische Gesellschaft*, 194(1): 31–35.
- Oskouie, S. and Tabandeh, F. 2007. Enhancement of alkaline protease production by *Bacillus clausii* using Taguchi experimental design. *African Journal of Biotechnology*, 6(22): 2559–2564.

- Pain, A. and Cooney, J. 1998. Characterization of organotin-resistant bacteria from Boston Harbor sediments. Archives of Environmental Contamination and Toxicology, 35(3): 412–416.
- Pal, A. and Paul, A.K. 2008. Microbial extracellular polymeric substances: central elements in heavy metal bioremediation. *Indian Journal of Microbiology*, 48(1): 49–64.
- Palanisamy, N. and Ramya, J. 2014. Diesel biodegradation capacities of indigenous bacterial species isolated from diesel contaminated soil. *Journal of Environmental Health Science and Engineering*, 12(1): 142.
- Pandey, A. 2003. Solid-state fermentation. *Biochemical Engineering Journal*, 13(2): 81–84.
- Pandey, A., Bera, D., Shukla, A. and Ray, L. 2007. Studies on Cr (VI), Pb (II) and Cu (II) adsorption–desorption using calcium alginate as biopolymer. *Chemical Speciation and Bioavailability*, 19(1): 17–24.
- Pandey, A. and Selvakumar, P. 1999. Solid state fermentation for the production of industrial enzymes. *Current Science*, 77(1): 149–162.
- Pandey, A., Soccol, C. and Mitchell, D. 2000a. New developments in solid state fermentation: I-bioprocesses and products. *Process Biochemistry*, 35(10): 1153– 1169.
- Pandey, A., Soccol, C.R., Nigam, P., Brand, D., Mohan, R. and Roussos, S. 2000b. Biotechnological potential of coffee pulp and coffee husk for bioprocesses. *Biochemical Engineering Journal*, 6(2): 153–162.
- Pandian, S., Jawahar, S. and P.P. 2012. Isolation, identification and characterization of feather degrading bacteria. *European Journal of Experimental Biology*, 2(1): 274–282.
- Park, G. and Son, H. 2009. Keratinolytic activity of *Bacillus megaterium* F7-1, a featherdegrading mesophilic bacterium. *Microbiological Research*, 164: 4478–4485.
- Park, J.H., Lamb, D., Paneerselvam, P., Choppala, G., Bolan, N. and Chung, J.W. 2011.
 Role of organic amendments on enhanced bioremediation of heavy metal (loid) contaminated soils. *Journal of hazardous materials*, 185(2): 549–574.
- Paul, T., Halder, S. K., Das, A., Bera, S., Maity, C. and Mandal, A. 2013. Biocatalysis and Agricultural Biotechnology Exploitation of chicken feather waste as a plant growth promoting agent using keratinase producing novel isolate *Paenibacillus* woosongensis TKB2. Biocatalysis and Agricultural Biotechnology, 2(1): 50–57.
- Piccirillo, C. and Pereira, S. 2013. Bacteria immobilisation on hydroxyapatite surface for heavy metals removal. *Journal of Environmental management*, 121: 87–95.

- Pillai, P. and Archana, G. 2008. Hide depilation and feather disintegration studies with keratinolytic serine protease from a novel *Bacillus subtilis* isolate. *Applied Microbiology and Biotechnology*, 78(4): 643–50.
- Pillai, P., Mandge, S. and Archana, G. 2011. Statistical optimization of production and tannery applications of a keratinolytic serine protease from *Bacillus subtilis* P13. *Process Biochemistry*, 46(5): 1110–1117.
- Pires, C., Marques, A. and Guerreiro, A. 2011. Removal of heavy metals using different polymer matrixes as support for bacterial immobilisation. *Journal of Hazardous materials*, 191(1): 277–286.
- Plackett, R. and Burman, J. 1946. The design of optimum multifactorial experiments. *Biometrika*, 33(4): 305–325.
- Prajapati, V.D., Jani, G. K., Zala, B.S. and Khutliwala, T.A. 2013. An insight into the emerging exopolysaccharide gellan gum as a novel polymer. *Carbohydrate polymers*, 93(2): 670–678.
- Prakash, P., Jayalakshmi, S. K. and Sreeramulu, K. 2010a. Production of keratinase by free and immobilized cells of *Bacillus halodurans* strain PPKS-2: partial characterization and its application in feather degradation and dehairing of the goat skin. *Applied Biochemistry and Biotechnology*, 160(7): 1909–1920.
- Prakash, P., Jayalakshmi, S. and Sreeramulu, K. 2010b. Purification and characterization of extreme alkaline, thermostable keratinase, and keratin disulfide reductase produced by *Bacillus halodurans* PPKS-2. *Applied Microbiology and Biotechnology*, 87(2): 625–633.
- Prakasham, R.S., Subba Rao, C., Sreenivas Rao, R. and Sarma, P.N. 2005. Alkaline protease production by an isolated *Bacillus circulans* under solid-state fermentation using agroindustrial waste: process parameters optimization. *Biotechnology Progress*, 21(5): 1380–1388.
- Prasad, H.V., Kumar, G., Kartik, L. and Rao, K.V.B. 2010. Screening of extracellular keratinase producing bacteria from feather processing areas in Vellore, Tamil Nadu, India. *Journal of Scientific Research*, 2(3): 559.
- Prescott, L., Harley, J. and Klein, D. 1996. Microbiology, Wm. C. Brown Publishers, Dubuque.
- Radha, S. and Gunasekaran, P. 2007. Cloning and expression of keratinase gene in *Bacillus megaterium* and optimization of fermentation conditions for the production of keratinase by recombinant strain. *Journal of Applied Microbiology*, 103(4): 1301–1310.
- Radovich, J.M. 1985. Mass transfer effects in fermentations using immobilized whole cells. *Enzyme and Microbial Technology*, 7(1): 2–10.

- Rahayu, S., Syah, D. and Thenawidjaja, M. 2012. Degradation of keratin by keratinase and disulfide reductase from *Bacillus* sp. MTS of Indonesian origin. *Biocatalysis* and Agricultural Biotechnology, 1(2): 152–158.
- Rai, S.K., Konwarh, R. and Mukherjee, A.K. 2009. Purification, characterization and biotechnological application of an alkaline β-keratinase produced by *Bacillus subtilis* RM-01 in solid-state fermentation using chicken-feather as substrate. *Biochemical Engineering Journal*, 45(3): 218–225.
- Rai, S.K. and Mukherjee, A.K. 2011. Optimization of production of an oxidant and detergent-stable alkaline keratinase from *Brevibacillus* sp. strain AS-S10-II: Application of enzyme in laundry detergent formulations and in leather industry. *Biochemical Engineering Journal*, 54(1): 47–56.
- Rajendran, A., Palanisamy, A. and Thangavelu, V. 2008. Evaluation of medium components by Plackett-Burman statistical design for lipase production by *Candida rugosa* and kinetic modeling. *Chinese Journal of Biotechnology*, 24(3): 436–444.
- Rajput, R. and Gupta, R. 2013. Thermostable keratinase from *Bacillus pumilus* KS12: production, chitin crosslinking and degradation of Sup35NM aggregates. *Bioresource Technology*, 133: 118–126.
- Rajput, R., Sharma, R. and Gupta, R. 2011. Cloning and characterization of a thermostable detergent-compatible recombinant keratinase from *Bacillus pumilus* KS12. *Biotechnology and Applied Biochemistry*, 58(2): 109–118.
- Ramarosandratana, A., Harvengt, L., Bouvet, A., Calvayrac, R. and Pâques, M. 2001. Effects of carbohydrate source, polyethylene glycol and gellan gum concentration on embryonal-suspensor mass (ESM) proliferation and maturation of maritime pine somatic embryos. *In Vitro Cellular & Developmental Biology* -*Plant*, 37(1): 29–34.
- Ramnani, P. and Gupta, R. 2004. Optimization of medium composition for keratinase production on feather by *Bacillus licheniformis* RG1 using statistical methods involving response surface methodology. *Biotechnology and Applied Biochemistry*, 40(2): 191–196.
- Ramnani, P., Singh, R. and Gupta, R. 2005. Keratinolytic potential of *Bacillus licheniformis* RG1: structural and biochemical mechanism of feather degradation. *Canadian Journal of Microbiology*, 51(3): 191–196.
- Rani, M. and Hemambika, B. 2010. Comparative assessment of heavy metal removal by immobilized and dead bacterial cells: A biosorption approach. *African Journal of Environmental Science and Technology*, 4(2).
- Rao, R. and Kumar, C. 2008. The Taguchi methodology as a statistical tool for biotechnological applications: a critical appraisal. *Biotechnology journal*, 3(4): 510–523.

- Ravi Kumar, M.N. 2000. A review of chitin and chitosan applications. *Reactive and Functional Polymers*, 46(1): 1–27.
- Rehfuss, M. and Urban, J. 2005. Alcaligenes faecalis subsp. phenolicus subsp. nov. a phenol-degrading, denitrifying bacterium isolated from a graywater bioprocessor. Systematic and Applied Microbiology, 28(5): 421–429.
- Riffel, A. and Brandelli, A. 2006. Keratinolytic bacteria isolated from feather waste. *Brazilian Journal of Microbiology*, 37(3): 395–399.
- Riffel, A., Brandelli, A. and Bellato, C. 2007. Purification and characterization of a keratinolytic metalloprotease from *Chryseobacterium* sp. kr6. *Journal of Biotechnology*, 128(3): 693–703.
- Riffel, A., Françoise, L., Philipp, H., Riffel, A., Lucas, F., Heeb, P. and Brandelli, A. 2003. Characterization of a new keratinolytic bacterium that completely degrades native feather keratin. *Archives of Microbiology*, 179(4): 258–265.
- Rouse, J. and Dyke, M.V. 2010. A review of keratin-based biomaterials for biomedical applications. *Materials*, 3(2): 999–1014
- Sahoo, D.K., Das, A., Thatoi, H., Mondal, K.C. and Mohapatra, P.D.K. 2012. Keratinase production and biodegradation of whole chicken feather keratin by a newly isolated bacterium under submerged fermentation. *Applied Biochemistry and Biotechnology*, 5: 1040–1051.
- Sanderson, G. and Clark, R. 1983. Gellan gum. Food Technology, 37(4): 62-70.
- Sandhya, C., Sumantha, A., Szakacs, G. and Pandey, A. 2005. Comparative evaluation of neutral protease production by *Aspergillus oryzae* in submerged and solid-state fermentation. *Process Biochemistry*, 40(8): 2689–2694.
- Sangali, S. and Brandelli, A. 2000. Feather keratin hydrolysis by a *Vibrio* sp. strain kr2. *Journal of Applied Microbiology*, 89(5): 735–743.
- Sawyer, R., Glenn, T. and French, J. 2000. The expression of beta (β) keratins in the epidermal appendages of reptiles and birds. *American Zoologist*, 40(4): 530–539.
- Sayed, S., Saleh, S. and Hasan, E. 2005. Removal of some polluting metals from industrial water using chicken feathers. *Desalination*, 181(1): 243–255.
- Scherbel, C. and Pichner, R. 2006. Degradation of scrapie associated prion protein (PrP by the gastrointestinal microbiota of cattle. *Veterinary research*, 37(5): 695–703.
- Scott, J. and Untereiner, W. 2004. Determination of keratin degradation by fungi using keratin azure. *Medical Mycology*, 42(3): 239–246.
- Senthilvelan, T., Kanagaraj, J. and Mandal, A.B. 2012. Application of enzymes for dehairing of skins: cleaner leather processing. *Clean Technologies and Environmental Policy*, 14(5): 889–897.

- Sharaf, E.F. and Khalil, N.M. 2011. Keratinolytic activity of purified alkaline keratinase produced by *Scopulariopsis brevicaulis* (Sacc.) and its amino acids profile. *Saudi Journal of Biological Sciences*, 18(2): 117–121.
- Sharma, R. and Gupta, R. 2010. Substrate specificity characterization of a thermostable keratinase from *Pseudomonas aeruginosa* KS-1. *Journal of Industrial Microbiology and Biotechnology*, 37(8): 785–792.
- Sharma, R., Gupta, R. and Sharma, R.G. 2010. Extracellular expression of keratinase Ker P from *Pseudomonas aeruginosa* and *E. coli. Biotechnology Letters*, 32(12): 1863–1868.
- Sharma, R. and Sharma, M. 2013. The impact of incinerators on human health and environment. *Reviews on environmental health*, 28(1): 67–72.
- Shi, S., Qu, Y., Tan, L. and Ma, F. 2015. Biosynthesis of 1,2-dihydroxydibenzofuran by magnetically immobilized cells of *Escherichia coli* expressing phenol hydroxylase in liquid-liquid biphasic systems. *Bioresource Technology*, 197: 72– 78.
- Shih, J. and Wang, J. 2006. Keratinase technology: from feather degradation and feed additive, to prion destruction. *CAB Reviews: Perspectives in Agriculture, veterinary Science, Nutrition and Natural Resources,* 1(042).
- Shih, J.C.H. and Williams, M.C. 1992. Purified *Bacillus licheniformis* PWD-1 keratinase. U.S. Patent and Trademark Office.Washinton DC.
- Shrinivas, D., Kumar, R. and Naik, G. 2012. Enhanced production of alkaline thermostable keratinolytic protease from calcium alginate immobilized cells of thermoalkalophilic *Bacillus halodurans* JB 99. *Journal of Industrial Microbiology and Biotechnology*, 39(1): 93–98.
- Singh, J.S., Abhilash, P.C., Singh, H.B., Singh, R.P. and Singh, D.P. 2011. Genetically engineered bacteria: an emerging tool for environmental remediation and future research perspectives. *Gene*, 480(1): 1–9.
- Singh, S.K., Singh, S.K., Tripathi, V.R. and Garg, S.K. 2012. Purification, characterization and secondary structure elucidation of a detergent stable, halotolerant, thermoalkaline protease from *Bacillus cereus* SIU1. *Process Biochemistry*, 47(10): 1479–1487.
- Singhania, R. and Sukumaran, R. 2010. Advancement and comparative profiles in the production technologies using solid-state and submerged fermentation for microbial cellulases. *Enzyme and Microbial technology*, 46: 7541–7549.
- Sivakumar, T. and Shankar, T. 2011. Statistical optimization of keratinase production by *Bacillus cereus. Global Journal of Biotechnology and Biochemistry*, 6(4): 197–202.

- Soares, F. and Braga, F. 2010. Optimization of medium composition for protease production by *Paecilomyces marquandii* in solid-statefermentation using response surface methodology. *African Journal of Microbiology Research*, 42: 2699–2703.
- Subramaniyam, R. and Vimala, R. 2012. Solid state and submerged fermentation for the production of bioactive substances: A comparative study. *International Journal of Sci Nat*, 3: 480–486.
- Sun, N., Wang, H., Chen, Y., Lu, S. and Xiong, Y. 2008. Effect of Surfactant SDS, Tween 80, Triton X-100 and Rhamnolipid on Biodegradation of Hydrophobic Organic Pollutants. In 2008 2nd International Conference on Bioinformatics and Biomedical Engineering. pp. 4730–4734.
- Sun, W. and Griffiths, M.W. 2000. Survival of bifidobacteria in yogurt and simulated gastric juice following immobilization in gellan-xanthan beads. *International Journal of Food Microbiology*, 61(1): 17–25.
- Suntornsuk, W. and Suntornsuk, L. 2003. Feather degradation by *Bacillus* sp. FK 46 in submerged cultivation. *Bioresource Technology*, 86(3): 239–243.
- Suvase, S.A., Annapure, U.S. and Singhal, R.S. 2010. Gellan gum as an immobilization matrix for the production of cyclosporin A. *Journal of Microbiology and Biotechnology*, 20(7): 1086–1091.
- Sworn, G., Sanderson, G. and Gibson, W. 1995. Gellan gum fluid gels. Food Hydrocolloids, 9(4): 265-271.
- Syed, D.G., Lee, J.C., Li, W.J., Kim, C.J. and Agasar, D. 2009. Production, characterization and application of keratinase from *Streptomyces gulbargensis*. *Bioresource Technology*, 100(5): 1868–1871.
- Taguchi, G. and Konishi, S. 1987. Taguchi Methods: Orthogonal Arrays and Linear Graphs-Tools for Quality Engineering. Amer Supplier Inst. p 72.
- Tamilmani, P., Umamaheswari, A. and Vinayagam, B.P. 2008. Production-of-an extracellular feather degrading enzyme by *Bacillus licheniformis* isolated from poultry farm soil in Namakkal district (Tamilnadu). *International Journal of Poultry Science*, 7(2): 184–188.
- Tanabe, T., Okitsu, N., Tachibana, A. and Yamauchi, K. 2002. Preparation and characterization of keratin–chitosan composite film. *Biomaterials*, 23(3): 817– 825.
- Tapia, D.M. and Simões, M.L.G. 2008. Production and partial characterization of keratinase produced by a microorganism isolated from poultry processing plant wastewater. *African Journal of Biotechnology*, 7(3).

- Taskin, M., Sisman, T., Erdal, S. and Kurbanoglu, E. 2011. Use of waste chicken feathers as peptone for production of carotenoids in submerged culture of *Rhodotorula* glutinis MT-5. European Food Research and Technology, 233(4): 657–665.
- Tatineni, R., Doddapaneni, K.K., Potumarthi, R.C., Vellanki, R.N., Kandathil, M.T., Kolli, N. and Mangamoori, L.N. 2008. Purification and characterization of an alkaline keratinase from *Streptomyces* sp. *Bioresource Technology*, 99(6): 1596– 1602.
- Tatineni, R., Doddapaneni, K., Potumarthi, R. and Mangamoori, L. 2007. Optimization of keratinase production and enzyme activity using response surface methodology with *Streptomyces* sp7. *Applied Biochemistry and Biotechnology*, 141(2-3): 187– 201.
- Thanapimmetha, A. 2012. Value added waste of *Jatropha curcas* residue: Optimization of protease production in solid state fermentation by Taguchi DOE methodology. *Industrial Crops and Products*, 37(1): 1–5.
- Thangam, E.B. and Rajkumar, G.S. 2000. Studies on the production of extracellular protease by *Alcaligenes faecalis*. *World Journal of Microbiology and Biotechnology*, 16(7): 663–666.
- Thys, R. C. S. and Brandelli, A. 2006. Purification and properties of a keratinolytic metalloprotease from *Microbacterium* sp. *Journal of Applied Microbiology*, 101(6): 1259–1268.
- Tiwary, E. and Gupta, R. 2010. Medium optimization for a novel 58 kDa dimeric keratinase from *Bacillus licheniformis* ER-15 : Biochemical characterization and application in feather degradation and dehairing of hides. *Bioresource Technology*, 101(15): 6103–6110.
- Tomarelli, R. 1949. The use of azoalbumin as a substrate in the colorimetric determination or peptic and tryptic activity. *The Journal of Laboratory and Clinical Medicine*, 34(3): 428.
- Tork, S., Aly, M. and Nawar, L. 2010. Biochemical and molecular characterization of a new local keratinase producing *Pseudomomanas* sp., MS21. Asian Journal of Biotechnology, 2(1): 1–13.
- Tuna, A., Okumuş, Y., Çelebi, H. and Seyhan, A.T. 2015. Thermochemical conversion of poultry chicken feather fibers of different colors into microporous fibers. *Journal* of Analytical and Applied Pyrolysis, 115: 112–124.
- Vaidya, R.J., Shah, I.M., Vyas, P.R. and Chhatpar, H.S. 2001. Production of chitinase and its optimization from a novel isolate *Alcaligenes xylosoxydans:* potential in antifungal biocontrol. *World Journal of Microbiology and Biotechnology*, 17(7): 691–696.

- Venil, C. and Lakshmanaperumalsamy, P. 2015. Taguchi experimental design for medium optimization for enhanced protease production by *Bacillus subtilis* HB04. e-Journal of Science and Technology, 4: 1–10.
- Venkatesh, C. and Pradeep, V. 2014. Metabolism of chicken feathers and concomitant electricity generation by *Pseudomonas aeruginosa* by employing microbial fuel cell (MFC). *Journal of Waste Management*, doi.org/10.1155/2014/928618.
- Venugopal, M. and Saramma, A.V. 2006. Characterization of alkaline protease from Vibrio fluvialis strain VM10 isolated from a mangrove sediment sample and its application as a laundry detergent additive. Process Biochemistry, 41(6): 1239– 1243.
- Villa, A. and Arag ão, M. 2013. Feather keratin hydrolysates obtained from microbial keratinases: effect on hair fiber. *BMC biotechnology*, 13(1): 1–11.
- Vorlop, K. and Klein, J. 1981. Formation of spherical chitosan biocatalysts by ionotropic gelation. *Biotechnology Letters*, 3(1): 9–14.
- Wang, B., Yang, W., McKittrick, J. and Meyers, M.A. 2015. Keratin: structure, mechanical properties, occurrence in biological organisms, and efforts at bioinspiration. *Progress in Materials Science*, 76: 229–318.
- Wang, S. 2003. Artificial neural network. *Interdisciplinary Computing in Java Programming*. Springer US, 2003. 81–100.
- Wawrzkiewicz, K. 1987. Intracellular keratinase of *Trichophyton gallinae*. Medical Mycology, 25(4): 261–268.
- Wawrzkiewicz, K., Wolski, T. and Łobarzewski, J. 1991. Screening the keratinolytic activity of dermatophytes in vitro. *Mycopathologia*, 114(1): 1–8.
- Wei-Hsun Lo and Jane-Yii Wu, J.R.T. 2012. Production of keratinolytic enzyme by an indigenousfeatheredegrading strain *Bacillus cereus* Wu2. *Journal of Bioscience* and Bioengineering, 114(6): 640–647.
- Williams, S.T. and Davies, F.L. 1967. Use of a Scanning Electron Microscope for the Examination of *Actinomycetes*. *Journal of General Microbiology*, 48(2): 171–177.
- Wilson, N. and Bradley, G. (1996). The effect of immobilization on rhamnolipid production by *Pseudomonas fluorescens*. *Journal of Applied Bacteriology*, 81(5): 525–530.
- Xie, F., Chao, Y., Yang, X., Yang, J., Xue, Z., Luo, Y. and Qian, S. 2010. Purification and characterization of four keratinases produced by *Streptomyces* sp. strain 16 in native human foot skin medium. *Bioresource Technology*, 101(1): 344–350.

- Yang, L., Li, X., Chu, Z., Ren, Y. and Zhang, J. 2014. Distribution and genetic diversity of the microorganisms in the biofilter for the simultaneous removal of arsenic, iron and manganese from simulated groundwater. *Bioresource technology*, 156: 384–388.
- Yoshioka, M., Miwa, T. and Horii, H. 2007. Characterization of a proteolytic enzyme derived from a *Bacillus* strain that effectively degrades prion protein. *Journal of Applied Microbiology*, 102(2): 509–515.
- Yu, M., Wu, P., Widelitz, R. and Chuong, C. 2002. The morphogenesis of feathers. *Nature*. 420(6913): 308–312.
- Yusuf, I., Shukor, M., Syed, M., Yee, P.L., Shamaan, N.A. and Ahmad, S.A. 2015. Investigation of keratinase activity and feather degradation ability of immobilised *Bacillus* sp. khayat in the presence of heavy metals in a semi continuous. *Journal of Chemical and Pharmaceutical Sciences*, 8(2): 342–347.
- Zaghloul, T.I., Embaby, A.M. and Elmahdy, A. R. 2011. Biodegradation of chicken feathers waste directed by *Bacillus subtilis* recombinant cells : Scaling up in a laboratory scale fermentor. *Bioresource Technology*, 102(3): 2387–2393.
- Zambare, V.P., Nilegaonkar, S.S., Kanekar, P.P., Zambare, V.P. and Nilegaonkar, P.P.K. 2007. Production of an alkaline protease by *Bacillus cereus* MCM B-326 and its application as a dehairing agent. *World Journal of Microbiology and Biotechnology*, 23(11): 1569–1574.
- Zduniak, P., Surmacki, A., Erciyas-Yavuz, K., Chudzińska, M. and Barałkiewicz, D. 2014. Are there different requirements for trace elements in eumelanin- and pheomelanin-based color production? A case study of two passerine species. *Comparative Biochemistry and Physiology. Part A, Molecular and Integrative Physiology*, 175: 96–101.
- Zhuang, H., Han, H., Xu, P., Hou, B., Jia, S., Wang, D. and Li, K. 2015. Biodegradation of quinoline by *Streptomyces* sp. N01 immobilized on bamboo carbon supported Fe3O4 nanoparticles. *Biochemical Engineering Journal*, 99: 44–47.