



UNIVERSITI PUTRA MALAYSIA

***ENZYMATIC SYNTHESIS OF KOJIC ACID ESTERS AND ANALYSIS OF
DEPIGMENTING ACTIVITIES USING IN VITRO AND IN VIVO MODELS***

AHMAD FIRDAUS BIN LAJIS

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By

AHMAD FIRDAUS BIN LAJIS

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Philosophy**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Doctor of Philosophy

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AHMAD FIRDAUS BIN LAJIS

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Chair: Arbakariya B. Ariff, PhD

Faculty: Biotechnology and Biomolecular Sciences

Kojic acid (KA) is a commonly known skin-whitening agent in cosmetic products. However, KA is not oil soluble that makes it difficult to be incorporated in cosmetic formulation. Therefore, KA esters are synthesized to increase its lipophilicity that allows it to be easily incorporated into cosmetic products. In previous study, kojic acid esters were synthesized via chemical processes using chemical catalysts. Chemical catalysts are not environmentally friendly and caused more harm to consumers compared to biocatalyst. The handling process and removal of hazardous chemical from products may also increase cost of production. In this study, the enzymatic synthesis of KA esters was proposed and performed in various types of bioreactor such as stirred tank reactor (STR), packed bed reactor (PBR) and fluidized reactor (FR) using three commercial immobilized lipases (TLIM, RMIM and N435) where the performances were compared. The alternative approaches and methods (solvent and solvent-free systems) for the synthesis of KA esters in different reactor systems were also investigated. Various enzymatic and bioreactor operation parameters were manipulated aimed at solving various problems in enzymatic reactor operation such as shear effects due to stirring and potential of lipase reusability. The physical, chemical and depigmenting properties of KA esters were also studied. The depigmenting activity of KA esters was evaluated using *in vitro* and *in vivo* models using B16F1 and zebrafish embryo, respectively.

For esterification using N435 lipase, very high yield (up to 50%) of KA esters were synthesized in STR compared to PBR and FR. The enzymatic esterification of KA esters in STR would currently be the best alternative as compared to other types of reactor tested, despite some of its weaknesses. Solvent-free system can be another alternative but the handling of saturated fatty acid in liquid form can be difficult because high temperature is required. Under ultraviolet (UV) light, skin color turn dark due to an increase of alpha-melanocyte stimulating hormone (α -MSH) and oxidation process. Kojic acid monopalmitate (KAMP) showed slightly higher inhibition to melanin formation compared to KA, kojic acid

monooleate (KAMO) and kojic acid monolaurate (KAML), as analysed by *in vitro* model using α -MSH stimulated B16F1 cells. The reduction of melanin formation was correlated to the reduction of mushroom tyrosinase and cellular tyrosinase activity in B16F1. KAMP also showed greater antioxidant activity than KA, KAML and KAMO as measured in anti-lipid peroxidation method and other antioxidant assay.

Evaluation of the depigmenting activity of KA esters by *in vivo* model using zebrafish embryo indicated that KAMP has better depigmenting activity than KA and KAMO. The toxicity level of KA and KA esters were also estimated where KAMP gave lower toxic effect compared to KA, KAML and KAMO, as evaluated using B16F1 cells, G361 cells and zebrafish embryo. Zebrafish embryo treated with KAMP also showed higher survival rate, high hatchability, stable heart-beat rate and no significant teratogenic effect as compared to KAML.

The results from this study have demonstrated that KA esters can be synthesized in solvent-free system as an alternative to solvent system. Solvent-free system has advantage of using chemical at low cost and environmental friendly process. Large scale production of KA esters could be performed using STR and FR, where the yield could be further improved by the improvement of mixing condition and optimization of the process variables. The absence of toxic and better depigmenting effect of KA esters as compared to KA suggest that KA esters are safe to be applied as skin-whitening agents in commercial cosmetic formulation.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENGHASILAN ASID KOJIK ESTER SECARA ENZIMATIK DAN ANALISIS NYAH-PIGMEN DENGAN MENGGUNAKAN MODEL *IN VITRO* DAN *IN VIVO*

Oleh

AHMAD FIRDAUS BIN LAJIS

Februari 2016

Pengerusi: Arbakariya B. Ariff, PhD

Fakulti: Bioteknologi dan Sains Biomolekul

Asid kojik (KA) merupakan agen pemutih yang biasa digunakan di dalam pelbagai produk kosmetik. Walaubagaimanapun, KA tidak larut minyak yang menyebabkan sukar untuk diformulasikan dalam kosmetik. Oleh itu, KA ester menjadi pilihan untuk produk kosmetik. Di dalam kajian yang terdahulu, penghasilan KA ester dilakukan dengan menggunakan pemangkin kimia. Penggunaan pemangkin kimia adalah tidak mesra alam dan lebih membahayakan pengguna berbanding pemangkin biologi. Penghasilan KA ester secara enzimatik telah dikaji dan dilakukan di dalam pelbagai jenis reaktor seperti tangki reaktor bergerak (STR), reaktor lapisan terpadat (PBR) and reaktor terangkat (FR) dengan menggunakan lipase tersekatgerak komersial, iaitu lipozim TLIM, RMIM and N435, dimana keberhasilan mereka dibandingkan. Kaedah alternatif (pelarut dan tanpa pelarut) untuk penghasilan KA ester di dalam pelbagai reaktor juga dikaji. Pelbagai parameter enzimatik dan operasi reaktor diubah-suai dengan tujuan untuk menyelesaikan pelbagai masalah di dalam operasi reaktor enzim seperti kesan ricih dan potensi penggunaan semula lipase. Sifat fizikal, kimia dan nyah-pigmen aktiviti oleh KA ester dengan menggunakan pelbagai kaedah juga dikaji. Nyah-pigmen aktiviti oleh asid kojik ester telah dianalisis dengan model *in vitro* dan *in vivo* menggunakan B16F1 dan embrio ikan zebra.

Bagi esterifikasi dengan lipase N435, penghasilan KA ester yang sangat tinggi (sehingga 50%) dapat disintesis di dalam STR berbanding PBR dan FR. Enzimatik esterifikasi KA ester di dalam STR adalah alternatif yang terbaik walaupun terdapat beberapa kelemahan berbanding dengan reaktor lain. Sistem tanpa pelarut mungkin boleh dijadikan alternatif untuk penghasilan KA ester tetapi suhu yang tinggi diperlukan untuk mencairkan asid lemak tepu (asid palmitik dan asid laurik). Apabila kulit terdedah kepada sinaran ultraungu (UV), warna kulit akan berubah menjadi gelap disebabkan meningkatnya hormon alpha-melanocyte (α -MSH) dan proses oksidasi. Kajian terhadap hiper-pigmen B16F1 yang telah dirangsang oleh α -MSH menunjukkan asid kojik palmitik (KAMP) mempunyai inhibisi yang tinggi terhadap penghasilan melanin berbanding dengan menggunakan KA, asid kojik laurik (KAML) dan

asid kojik oleik (KAMO). Pengurangan penghasilan melanin di dalam B16F1 yang telah dirawat bersama KAMP, berkadar langsung dengan pengurangan aktiviti tyrosinase cendawan dan tyrosinase seluler di dalam B16F1. Di dalam kajian ini, KAMP juga menunjukkan kesan antioksidasi yang tinggi berbanding KA, KAML and KAMO sebagaimana telah dinilai menggunakan kaedah anti-perosidasi lipid dan kaedah yang lain.

Penilaian terhadap keberkesanan nyah-pigmen oleh KA ester menggunakan model *in vivo* iaitu embrio ikan zebra menunjukkan bahawa KAMP memiliki aktiviti nyah-pigmen yang lebih baik daripada KA and KAMO. Tahap toksik KA dan KA ester juga dinilai dan KAMP menunjukkan tahap toksik yang lebih rendah daripada KA, KAML and KAMO, sepertimana telah dinilai dengan menggunakan sel B16F1, sel G361 dan embrio ikan zebra. Embrio ikan Zebra yang dirawat dengan KAMP juga menunjukkan kadar hidup yang tinggi, kadar penetasan yang tinggi, kadar denyutan jantung yang stabil dan tiada kesan teratogen berbanding KAML.

Keputusan dari kajian ini menunjukkan bahawa KA ester boleh disintesis di dalam sistem tanpa pelarut sebagai alternatif kepada sistem dengan pelarut. Sistem tanpa pelarut mempunyai kelebihan dari segi pengurangan kos dan lebih mesra alam. Produksi berskala besar boleh dilakukan dengan menggunakan sistem STR dan FR, dimana penghasilan KA ester boleh ditambah baik dengan mengoptimalkan parameter dan keadaan sebatian campuran. Ketiadaan kesan toksik dan peningkatan aktiviti nyah-pigmen oleh KA ester berbanding KA membolehkan KA ester digunakan di dalam ramuan kosmetik sebagai agen pemutih yang lebih selamat dan berkesan.

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Arbakariya B. Ariff, PhD

Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Muhajir Hamid, PhD

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

Syahida Ahmad, PhD

Senior Lecturer

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

BUJANG KIM HUAT, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

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Signature: _____

Name of Chairman
of Supervisory
Committee:

Prof. Dr. Arbakariya B. Ariff

Signature: _____

Name of Member of
Supervisory
Committee:

Assoc. Prof. Dr. Muhajir Hamid

Signature: _____

Name of Member of
Supervisory
Committee:

Dr. Syahida Ahmad

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LIST OF ABBREVIATIONS

α -MSH	alpha-melanocyte stimulating hormone
BCR	Bed column reactor
BHA	Butylhydroxyanisole
BHT	Butylhydroxytoluene
BSA	Bovine serum albumin
cAMP	Cyclic adenosine monophosphate
cGMP	Cyclic guanosine monophosphate
DHI	Dihydroxyindole
DHICA	Dihydroxyindole-2-carboxylic acid
DMEM	Dulbecco's modified Eagle's medium
DMF	Dimethylformamide
DMSO	Dimethyl sulfoxide
DOPA	L-3,4-dihydroxyphenylalanine
DOT	Dissolve oxygen tension
DPPH	2,2-diphenyl-1-picrylhydrazyl
DSC	Differential Scanning Calorimetry
EDTA	Ethylenediaminetetraacetic acid
EGF	Epidermal growth factor
ERK	Extracellular signal-regulated kinase
FBR	Fluidized bed reactor
FeCl ₃	Iron(III) chloride
FTIR	Fourier transform infrared spectroscopy
FR	Fluidized reactor
FRAP	Ferric reducing antioxidant power
GCMS	Gas Chromatography Mass Spectrometry
HEPES	4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid
HMBC	Heteronuclear multiple-bond correlation spectroscopy
HSQC	Heteronuclear single-quantum correlation spectroscopy
H ₂ O ₂	Hydrogen peroxide
IBMX	Isobutylmethylxanthine
JNK	c-Jun N-terminal kinase
KA	Kojic acid
KAMO	Kojic acid monooleate
KAML	Kojic acid monolaurate
KAMP	Kojic acid monopalmitate
MC1R	Melanocortin receptor
MITF	Microphthalmia-associated transcription factor
Msg1	melanocyte-specific gene 1
NMR	Nuclear magnetic resonance
N435	Novozyme 435
PBR	Packed bed reactor
PKA	Protein kinase A
PTU	1-phenyl-2-thiourea
RMIM	immobilized lipase from <i>Rhizomucor miehei</i>
RPA	Reducing power ability
SDS-PAGE	Sodium dodecyl sulfate- polyacrylamide gel electrophoresis
STR	Stirred tank reactor
TBA	Thiobarbituric acid

TBARS
TCA
TGA
TLIM
TPTZ
UATR

Thiobarbituric acid reactive substances
Trichloroacetic acid
Thermogravimetric Analysis
Lipozyme *Thermomyces lanuginosus* lipase
2,4,6-tri(2-pyridyl)-s-triazine
Universal Attenuated Total Reflectance



CHAPTER 1

INTRODUCTION

Kojic acid (KA) is produced by various *Aspergillus* and *Penicillium* species such as *A. flavus*, *A. parasiticus* and *P. rubrum* (Wei et al., 1991; Parrish et al., 1966). KA producing strains, *A. flavus*, can be improved by the monospore isolation and mutation methods to obtain a stable monokaryotic strain capable of producing high amount of KA (Madihah et al., 1996; Ariff et al., 1996; Rosfarizan et al., 1998). The optimal dissolved oxygen tension (DOT) control strategy for KA fermentation by *A. flavus* for KA production in STR has been proposed by Rosfarizan et al., (2002). In this DOT control strategy, DOT was controlled at high level (>80%) during active growth phase and then switched to low level (30%) during production phase in batch submerged fermentation to give the highest yield and overall productivities. Proposed pathway for KA biosynthesis is shown in Figure 1.1 (Bajpai et al., 1981).

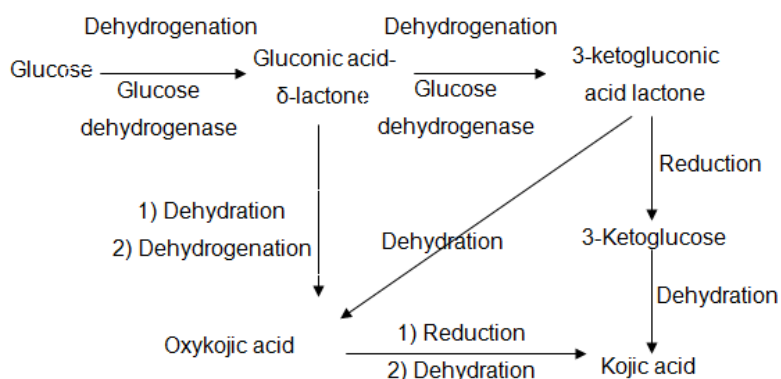


Figure 1.1. Biosynthetic pathways for kojic acid biosynthesis in *A. flavus*

KA has a wide range of industrial application such as to treat problems related to oxidation, photodamage, hyperpigmentation and skin wrinkling (Wright et al., 2014; Kotyzová et al., 2004). Hydrophilic property of KA has restricted its application in cosmetic, oily food and pharmaceutical products (Mohamad et al., 2010). KA has also been criticized for weak depigmenting effect and unstable for long storage. Moreover, there are concerns of its toxicity, carcinogenicity and hepatocarcinogenicity (Chusiri et al., 2011).

To improve KA characteristic, various KA derivatives such as KA esters have been synthesized. In industry, KA ester was developed and produced via chemical process which is not environmentally friendly (Manosroi et al., 2005; Radzi et al., 2011). A new synthesis approach using enzymatic process where cost and the use of hazardous chemical can be reduced or minimized has been developed (Chaibakhsh et al., 2009). Unlike chemical process using chemical catalyst, immobilized lipase is susceptible to inhibition, which therefore require an appropriate bioreactor design, mode of operation, substrate supply, efficient

product removal, reuse of immobilized lipase, scale-up and process control (Chaibakhsh et al., 2010). KA can be esterified into a more stable form using lipase with aims of producing better depigmenting effect. The implementation of lipase-based ester production in solvent system has been carried out using as lipase of *Candida antarctica* B lipase and lipase of *Thermomyces lanuginosus* (Noureddini et al., 2005; Adachi and Kobayashi, 2005; Noureddini et al., 2002). However, report on the production of KA esters in solvent-free system is still lacking. Solvent-free system offer several advantages over solvent system such as simplification of experimental procedure, reduce cost of chemical, reduce by-product to increase rate of reaction and reduce pollution (Singh and Chowdhury, 2011).

So far, research on enzymatic production of KA ester was only conducted in a small tube or shake flask where reports on the use of reactor system which can be scaled-up, for the production of KA esters are lacking (Ashari et al., 2009; Liu and Shaw, 1998). Stirred tank reactor (STR) is a typical mixing type reactor which is a commonly used in laboratory and industrial scales due to ease of fabrication and construction, maintenance and operation (Halim et al., 2009). Packed bed reactor (PBR) is a typical plug flow reactor. PBR provides a larger reacting surface area per unit volume than STR and is often applied in continuous process with high volumetric productivity (Balcão et al., 1996; Rahman et al., 2011; Dahlan et al., 2005).

In order for KA ester to be commercialized, their safety and efficiency as whitening agent is necessary to be analyzed and determined. *In vitro* method is the easiest way to evaluate depigmenting effect of KA esters, which can be done using simple mushroom tyrosinase assay (Kaatz et al., 1999). As auto-oxidation was involved in melanin production, it is necessary to evaluate the antioxidant assay of KA esters (Jean et al., 2013; Costin and Hearing, 2007). This can be done using various antioxidant assay such as FRAP, and DPPH assays (Saha et al., 2008). However, these evaluations are not adequate and require *in vitro* cells model assessment as supporting data. Examples of *in vitro* models include B16 cells, G361 cells and A376 cells (Kang et al., 2009; Sato and Toriyama, 2009). The *in vitro* model using B16 cells gives some advantages over simple *in vitro* mushroom tyrosinase assessment as it allow investigation at cellular and molecular levels (Kim et al., 2003). In addition to *in vitro* model, further experiment on animal *in vivo* model is often carried out to confirm the previous findings using *in vitro* model (Akhtar et al., 2014; Oliveira et al., 2010). *In vivo* models allow the assessment of organ physiology and morphology, as well as organ-specific cell-to-cell interactions (Gonçalez et al., 2013; da Silva et al., 2013). Although KA and KA esters have been produced industrially and commercialized, the question of safety is still not fully answered. Therefore, the appropriate methodology for evaluation of KA and KA esters in term of its safety for use in cosmetic and pharmaceutical formulation shall be developed.

Therefore, the objectives of this study were

1. To investigate the influence of various bioreactor designs and parameters on the performance of the enzymatic esterification of KA and fatty acid to various KA esters (KA monooleate, KA monolaurate and KA monopalmitate) using commercial lipases in solvent and non-solvent systems.
2. To evaluate the depigmenting effect of KA esters, produced by the enzymatic esterification, in hormone-stimulated hyper-pigmented B16F1 melanoma cells using *in vitro* models
3. To evaluate anti-oxidant properties of KA esters, produced by the enzymatic esterification, to free radicals involved in pigmentation process
4. To evaluate the hypopigmenting, toxic and teratogenic effect of KA esters, produced by the enzymatic esterification, using *in vivo* animal model with zebrafish embryo

BIBLIOGRAPHY

- Acosta, A., Filice, M., Gloria F.L., Palomo, J.M. and Guisan, J. M. (2011). Kinetically controlled synthesis of monoglyceryl esters from chiral and prochiral acids methyl esters catalyzed by immobilized *Rhizomucor miehei* lipase. *Bioresource Technology* 102(2): 507–512.
- Adachi, S. and Kobayashi, T. (2005). Synthesis of esters by immobilized-lipase-catalyzed condensation reaction of sugars and fatty acids in water-miscible organic solvent. *Journal of Bioscience and Bioengineering* 99(2): 87–94.
- Adlercreutz, P. (2013). Immobilisation and application of lipases in organic media. *Chemistry Society Review* 42: 6406-6436.
- Adnani, A., Basri, M., Chaibakhsh, N., Ahangar, H.A., Salleh, A. B., Abdul-Rahman, R.Z.R. and Abdul-Rahman, M. B. (2011). Chemometric analysis of lipase-catalyzed synthesis of xylitol esters in a solvent-free system. *Carbohydrate Research* 346(4): 472–479.
- Ahn, S.M., Rho, H.S., Baek, H.S., Joo, Y.H., Hong, Y.D, Shin, S.S., Park, Y.H., and Park, S.N. (2011). Inhibitory activity of novel kojic acid derivative containing trolox moiety on melanogenesis. *Bioorganic and Medicinal Chemistry Letters* 21: 7466–7469.
- Akhtar, N. (2014). *In vivo* pathogenicity studies of *Aspergilli* in lepidopteran model host *Galleria mellonella*. *APCBEE Procedia* 8: 293- 298.
- Akhtar, N., Khan, H.M.S., Ashraf, S., Mohammad, I.S. and Ali, A. (2014). Skin depigmentation activity of *Crocus sativus* extract cream. *Tropical Journal of Pharmaceutical Research* 13(11): 1803-1808.
- Al-Edresi, S. and Baie, S. (2010). *In-vitro* and *in-vivo* evaluation of a photo-protective kojic dipalmitate loaded into nano-creams. *Asian Journal of Pharmaceutical Sciences* 5(6): 251-265.
- Aleena, F., Jimbow, K. and Ito, S. (1990). Melanocytotoxicity and antimelanoma effects of phenolic amine compounds in mice *in vivo*. *Cancer Research* 50: 3743-3747.
- AlGhamdi, K.M and Kumar, A. (2010). Depigmentation therapies for normal skin in vitiligo universalis. *Journal of the European Academy of Dermatology and Venereology* 25(7):749-757.
- Al-Zuhair, S. and Taher, H. (2015). Application of lipases. In Al-zuhair (ed.), *Supercritical Fluids Technology in Lipase Catalyzed Processes* (pp. 150-176). England: CRC Press.

- Ali, T.E.S. and Legler, J. (2011). Developmental toxicity of nonylphenol in zebrafish (*Danio rerio*) embryos. *Indian Journal of Marine Sciences* 40(4): 509-515.
- An, S.M., Koh, J.S. and Boo, Y.C. (2010). p-coumaric acid not only inhibits human tyrosinase activity *in vitro* but also melanogenesis in cells exposed to UVB. *Phytotherapy Research* 24(8): 1175-1180.
- Andrade, L.N., de Lima, T.M., Curi, R. and Castrucci, A.M. (2005). Toxicity of fatty acids on murine and human melanoma cell lines. *Toxicology in Vitro* 19(4): 553-560.
- Aoki, Y., Tanigawa, T., Abe, H. and Fujiwara, Y. (2007). Melanogenesis inhibition by an oolong tea extract in B16 mouse melanoma cells and UV-induced skin pigmentation in brownish guinea pigs. *Bioscience, Biotechnology and Biochemistry* 71(8): 1879-1885.
- Arcos, J.A., Garcia, H.S. and Hill, C.G. (2000). Continuous enzymatic esterification of glycerol with (poly)unsaturated fatty acids in a packed bed reactor. *Biotechnology and Bioengineering* 68(5): 563-570.
- Ariff, A.B., Rosfarizan, M., Heng, L.S., Madihah, S. and Karim, M.I.A. (1997). Kinetics and modelling of kojic acid production by *Aspergillus flavus* Link in batch fermentation and resuspended cell mycelial. *World Journal of Microbiology and Biotechnology* 13(2):195-201.
- Ariff, A.B., Salleh, M.S., Ghani, B., Hassan, M.A., Rusul, G., Karim, M.I.A. (1996). Aeration and yeast extract requirements for kojic acid production by *Aspergillus flavus* Link. *Enzyme Microbiology and Technology* 19: 545-550.
- Asada, N., Kedamori, A., Kusano, Y. and Takeuchi, T. (2014). Pheomelanin formation and low tyrosinase activity in fading body color variant BdLR strain *Oryzias latipes*. *Journal of Life Sciences* 8(6): 517-521.
- Asghari, S., Maryam F.N. and Ahmadipour, M. (2010). Regioselective vinylation of kojic acid using acetylenic esters in the presence of triphenylphosphine or tert-butyl isocyanide. *Monatsh Chemistry* 141(7): 781-786.
- Ashari, S.E., Rosfarizan, M., Ariff, A.B., Basri, M. and Salleh, A.B. (2009). Optimization of enzymatic synthesis of palm-based kojic acid ester using response surface methodology. *Journal of Oleo Science* 58(10): 501-510.
- Ashrafuzzaman, M., Pyo, J.I and Cheong, C.S. (2014). Sucrose Derivatives Preparation using *Thermomyces lanuginosus* Lipase and Their Application. *Bulletin Korean Chemistry Society* 35(2): 477-482.

- Aytemir, M. D. and Calis, U. (2010). Anticonvulsant and neurotoxicity evaluation of some novel kojic acids and allomaltol derivatives. *Archiv der Pharmazie* 343(3): 173-181.
- Aytemir, M. D. and Özçelik B. (2010). A study of cytotoxicity of novel chlorokojic acid derivatives with their antimicrobial and antiviral activities. *European Journal of Medicinal Chemistry* 45(9): 4089-4095.
- Aytemir, M.D., Erol, D.D., Hider, R.C. and Ozalp, M. (2003). Synthesis and Evaluation of Antimicrobial Activity of New 3-Hydroxy-6-methyl-4-oxo-4H-pyran-2-carboxamide Derivatives. *Turkish Journal of Chemistry* 27:757-776.
- Aytemir, M.D., Ozçelik, B. and Karakaya, G. (2013). Evaluation of bioactivities of chlorokojic acid derivatives against dermatophytes coupled with cytotoxicity. *Bioorganic and Medicinal Chemistry Letters* 23(12): 3646-3649.
- Aytemir, M. D., Septioğlu, E. and Calış, U. (2010). Synthesis and anticonvulsant activity of new kojic acid derivatives. *Arzneimittelforschung* 60(1): 22–29.
- Balcão, V.M., Paiva, A.L. and Malcata, F.X. (1996). Bioreactors with immobilized lipases: State-of-the-art. *Enzyme and Microbial Technology* 18(6): 392-416.
- Balaguer, A., Salvador, A. and Chisvert, A. (2008). A rapid and reliable size-exclusion chromatographic method for determination of kojic dipalmitate in skin-whitening cosmetic products. *Talanta* 75(2): 407–411.
- Balasubramanian, K.A., Nalini, S. and Manohar, M. (1992). Nonesterified fatty acids and lipid peroxidation. *Molecular and Cellular Biochemistry* 111: 131-135.
- Balasubramanian, K.A., Nalini, S., Cheeseman, K.H. and Slater, T.F (1989). Nonesterified fatty acids inhibit iron-dependent lipid peroxidation. *Biochimica et Biophysica Acta* 1003(3): 232-237.
- Balaz, S., Uher, M., Brtko, J., Veverka, M., Bransova, J., Dobias, J., Podova, M. and Buchvald, J. (1993). Relationship between antifungal activity and hydrophobicity of kojic acid derivatives. *Folia Microbiologica* 38(5): 387-391.
- Barbazuk, W.B., Korf, I., Kadavi, C., Heyen, J., Tate, S., Wun, E., Bedell, J. A., McPherson, J. D. and Johnson, S. L. (2000). The Syntenic Relationship of the Zebrafish and Human Genomes. *Genome Research* 10(9): 1351-1358.
- Barret, M.C., Mahon, M.F., Molloy, K.C., Steed, J.W. and Wright, P. (2001). Synthesis and Structural Characterization of Tin(II) and Zinc(II) Derivatives of Cyclic α -Hydroxyketones, Including the Structures of

Sn(maltol)₂, Sn(tropolone)₂, Zn(tropolone)₂, and Zn(hinokitiol)₂. *Inorganic Chemistry* 40(17): 4384-4388.

Beberok, A., Otręba, M., Wrześniok, D. and Buszman, E. (2013). Cytotoxic effect of lomefloxacin in culture of human epidermal melanocytes. *Pharmacology Reports* 65(3): 689-699.

Beberok, A., Wrześniok, D., Otręba, M., Miliński, M., Rok, J. and Buszman, E. (2015). Effect of norfloxacin and moxifloxacin on melanin synthesis and antioxidant enzymes activity in normal human melanocytes. *Molecular and Cellular Biochemistry* 401 (1-2): 107–114.

Berman, J.R., Skariah, G., Maro, G.S., Mignot, E. and Mourrain, P. (2009). Characterization of two melanin-concentrating hormone genes in zebrafish reveals evolutionary and physiological links with the mammalian MCH system. *Journal of Comparative Neurology* 517(5): 695-710.

Bertolotto, C., Bille, K., Ortonne, J.P. and Ballotti, R. (1996). Regulation of tyrosinase gene expression by cAMP in B16 melanoma cells involves two CATGTG motifs surrounding the TATA box: implication of the microphthalmia gene product. *Journal of Cell Biology* 134(3): 747–755.

Boissy, R.E., Visscher, M. and DeLong, M.A. (2005). DeoxyArbutin: a novel reversible tyrosinase inhibitor with effective *in vivo* skin lightening potency. *Experimental Dermatology* 14(8): 601-608.

Botterweck, A.A.M., Verhagen, H., Goldbohm, R.A., Kleinjans, J. and Van den Bradt, P.A. (2000). Intake of butylated hydroxyanisole and butylated hydroxytoluene and stomach cancer risk: Results from analyses in the Netherlands cohort study. *Food and Chemical Toxicology* 38(7): 599-605.

Bousselsela, H., Benhouda, A., Yahia, M., Benbia, S., Ghecham, A. and Zidani, A. (2012). *In vitro* evaluation of antioxidant and antibacterial activities of extracts of *Hertia cheirifolia*'s leaves. *Natural Science* 4(11): 825-831.

Brannen, K.C., Julieta M.P.K., Danberry, T.L and Karen A.A.R (2010). Development of a zebrafish embryo teratogenicity assay and quantitative prediction model. *Birth Defects Research (Part B)* 89(1): 66–77.

Briganti, S., Cameli, N. and Picardo, M. (2007). Skin-lightening formulation: A comparative *in vivo* and *in vitro* study. *Cosmetic Dermatology* 20(4): 216-220.

Briganti, S., Camera, E. and Picardo, M. (2003). Chemical and instrumental approaches to treat hyperpigmentation. *Pigment Cell Research* 16(2):101-110.

Brígda, A.I.S., Pinheiro, Á.D.T., Ferreira, A.L.O., Pinto, G.A.S. and Gonçalves, L. R. B. (2007). Immobilization of *candida antarctica* lipase B by covalent

attachment to green coconut fiber. *Applied Biochemistry and Biotechnology* 136-140(1-12): 67-80.

Brtko, J., Rondahl, L., Ficková, M., Hudecová, D., Eybl, V. and Uher, M. (2004). Kojic acid and its derivatives: History and present state of art. *Central European Journal of Public Health* 12: S16–S18

Buranajaree, S., Donsing, P., Jeenapongsa, R. and Viyoch J. (2011). Depigmenting action of a nanoemulsion containing heartwood extract of *Artocarpus incisus* on UVB-induced hyperpigmentation in C57BL/6 mice. *Journal of Cosmetic Science* 62(1): 1–14

Burchill, S.A., Bennett, D.C., Holmes, A. and Thody, A.J. (1991). Tyrosinase expression and melanogenesis in melanotic and amelanotic B16 mouse melanoma cells. *Pathobiology* 59(5): 335–339.

Burdock, G. A., Soni, M. G. and Carabin, G. I. (2001). Evaluation of health aspects of kojic acid in food. *Regression Toxicological Pharmacology* 33(1): 80-101.

Burnett, C.L., Bergfeld, W.F., Belsito, D.V., Hill, R.A., Klaassen, C.D., Liebler, D.C., Marks, J.G. Jr, Shank, R.C., Slaga, T.J., Snyder, P.W., Andersen, F.A. (2010). Final report of the safety assessment of kojic acid as used in cosmetics. *International Journal of Toxicology* 29(6):244S-73.

Camp, E and Lardelli, M. (2001). Tyrosinase gene expression in zebrafish embryos. *Development Genes Evolution* 211(3):150-153.

Cao, X.G., Li, X.X., Bao, Y.Z., Xing, N.Z. and Chen, Y. (2007). Responses of Human Lens Epithelial Cells to Quercetin and DMSO. *Investigate Ophthalmology and Visual Science* 48(8): 3714-3718.

Carta, G., Gainer, J.L. and Benton, A.H. (1991). Enzymatic synthesis of esters using an immobilized lipase. *Biotechnology and Bioengineering* 37: 1004-1009.

Carta, G., Gainer, J.L. and Gibson, M.E. (1992). Synthesis of esters using a nylon-immobilized lipase in batch and continuous reactors. *Enzyme and Microbial Technology* 14(11): 904-910.

Cassani, J., Luna, H., Navarro, A. and Castillo, E. (2007). Comparative esterification of phenylpropanoids versus hydrophenylpropanoids acids catalyzed by lipase in organic solvent media. *Electronic Journal of Biotechnology* 10(4): 508-513.

Cassels, B.K. and Asencio, M. (2011). Anti-HIV activity of natural triterpenoids and hemisynthetic derivatives 2004–2009. *Phytochemistry Reviews* 10(4): 545-564.

- Castillo, E., Dossat, V., Marty, A., Condoret, J.S. and Combes, D. (1997). The role of silica gel in lipase-catalyzed esterification reactions of high-polar substrates. *Journal of the American Oil Chemists' Society* 74(2): 77-85.
- Castro, M.S.D, Domínguez, P. and Sinisterra J.V. (2000). Enzymatic amidation and alkoxyacylation of amines using native and immobilised lipases with different origins: a comparative study. *Tetrahedron* 56(10): 1387-1391.
- Cha, J.Y and Kim, S.Y. (2013). Anti-melanogenesis in B16F0 melanoma cells by extract of fermented *Cordyceps militaris* containing high cordycepin. *Journal of Life Science* 23(12): 1516-1524.
- Chaibakhsh, N., Abdul-Rahman, M. B., Abd-Aziz, S., Basri, M., Salleh, A. B. and Abdul-Rahman, R.N.Z. (2009). Optimized lipase-catalysed synthesis of adipase ester in a solvent-free system. *Journal of Indian Microbiology and Biotechnology* 36(9): 1149–1155.
- Chaibakhsh, N., Abdul-Rahman, M.B., Vahabzadeh, F., Abd-Aziz, S., Basri, M., and Salleh, A.B (2010). Optimization of operational conditions for adipate ester synthesis in a stirred tank reactor. *Biotechnology and Bioprocess Engineering* 15(5): 846-853.
- Chamouleau, F., Coulon, D., Girardin, M. and Ghoul, M. (2001). Influence of water activity and water content on sugar esters lipase-catalyzed synthesis in organic media. *Journal of Molecular Catalysis B: Enzymatic* 11(4): 949–954.
- Chan, C.F., Huang, C.C., Lee, M.Y. and Lin, Y.S. (2014). Fermented broth in tyrosinase and melanogenesis inhibition. *Molecules* 19(9): 13122-13135.
- Chang, M.S., Choi, M.J., Park, S.Y. and Park, S.K. (2010). Inhibitory effects of Hoelen extract on melanogenesis in B16/F1 melanoma cells. *Phytotherapy Research* 24(9): 1359-1364.
- Chang, S.W., Shaw, J.F., Shieh, C.H. and Shieh, C.J. (2006). Optimal formation of hexyl laurate by Lipozyme IM-77 in solvent-free system. *Journal of Agricultural and Food Chemistry* 54(19): 7125-7129.
- Chang, S.W., Shaw, J.F., Yang, C.K. and Shieh, C.J. (2007). Optimal continuous biosynthesis of hexyl laurate by a packed bed bioreactor. *Process Biochemistry* 42(9):1362-1366.
- Chang, T.S. (2012). Natural melanogenesis inhibitors acting through the down-regulation of tyrosinase activity. *Materials* 5(9): 1661-1685
- Chen, C.S., Liu, K. , Lou, Y.H. and Shieh, C.J. (2002). Optimisation of kojic acid monolaurate synthesis with lipase PS from *Pseudomonas cepacia*. *Journal of the Science of Food and Agriculture* 82(6): 601-605.

- Chen, H.C., Kuo, C.H., Twu, Y.K., Chen, J.H., Chang, C. M. J., Liu, Y.C. and Shieh, C.J (2011a). A continuous ultrasound-assisted packed-bed bioreactor for the lipase-catalyzed synthesis of caffeic acid phenethyl ester. *Journal of Chemical Technology and Biotechnology* 86(10): 1289-1294.
- Chen, T.H., Wang, Y.H. and Wu, Y.H. (2011b). Developmental exposures to ethanol or dimethylsulfoxide at low concentrations alter locomotor activity in larval zebrafish: Implications for behavioral toxicity bioassays, *Aquatic Toxicology* 102(3-4): 162-166.
- Chen, W.C., Tseng, T.S., Hsiao, N.W., Lin, Y.L., Wen, Z.H., Tsai, C.C., Lee, Y.C., Lin, H.H., Tsai, K.C. (2015). Discovery of highly potent tyrosinase inhibitor, T1, with significant anti-melanogenesis ability by zebrafish *in vivo* assay and computational molecular modeling. *Scientific Reports* 5: 7995.
- Chen, X., Zhang, B., Yuan, X., Yang, F., Liu, J., Zhao, H., Liu, L., Wang, Y., Wang, Z. and Zheng, Q. (2012). Isoliquiritigenin-induced differentiation in mouse melanoma B16F0 cell line. *Oxidative Medicine and Cellular Longevity* 2012: 534934
- Chen, Y.H., Lu, P.J., Hulme, C. and Shaw, A.Y. (2013). Synthesis of kojic acid-derived copper-chelating apoptosis inducing agents. *Medicinal Chemistry Research* 22(2): 995-1003.
- Cheong, L. Z., Tan, C. P., Long, K., Yusoff, M. S. A., Arifin, N., Lo, S. K. and Lai, O. M. (2007). Production of a diacylglycerol-enriched palm olein using lipase-catalyzed partial hydrolysis: Optimization using response surface methodology. *Food Chemistry* 105(4): 1614-1622.
- Cheryan, M. and Mehaia, M.A. (1986). Membrane Bioreactors. In M. Dekker (Ed.), *Membrane Separations in Biotechnology* (pp. 255-302). New York: MacGregor, W.C.
- Chluba-de Tapia, J., Bagutti, C., Cotti, R. and Eberle, A.N. (1996). Induction of constitutive melanogenesis in amelanotic mouse melanoma cells by transfection of the human melanocortin-1 receptor gene. *Journal of Cell Science* 109(8):2023-2030.
- Cho, J.C., Rho, H.S., Baek, H.S., Ahn, S.M., Woo, B.Y., Hong, Y.D., Cheon, J.W., Heo, J.M., Shin, S.S., Park, Y.H. and Suh, K.D. (2012a). Depigmenting activity of new kojic acid derivative obtained as a side product in the synthesis of cinnamate of kojic acid. *Bioorganic and Medicinal Chemistry Letters* 22(5): 2004–2007.
- Cho, J.C., Rho, H.S., Joo, Y.H., Lee, C.S., Lee, J., Ahn, S.M., Kim, J.E., Shin, S.S., Park, Y.H., Suh, K.D and Park, S.N. (2012b). Depigmenting activities of kojic acid derivatives without tyrosinase inhibitory activities. *Bioorganic Medical Chemical Letter* 22(12): 4159-4162.

- Chodurek, E., Orchel, A., Orchel, J., Kurkiewicz, S., Gawlik, N., Dzierżewicz, Z. and Stępień, K. (2012). Evaluation of melanogenesis in A-375 melanoma cells treated with 5,7-dimethoxycoumarin and valproic acid. *Cellular and Molecular Biology Letters* 17(4): 616-632.
- Choi, H., Kim, K., Han, J., Choi, H., Jin, S.H., Lee, E.K., Shin, D.W., Lee, T.R., Lee, A.Y. and Noh, M. (2012). Kojic acid-induced IL-6 production in human keratinocytes plays a role in its anti-melanogenic activity in skin. *Journal of Dermatological Science* 66(3): 207-215.
- Choi, S. W., Lee, S.K., Kim, E.O., Oh, J.H., Yoon, K.S., Parris, N., Hicks, K.B., and Moreau, R.A. (2007a). Antioxidant and antimelanogenic activities of polyamine conjugates from corn bran and related hydroxycinnamic acids. *Journal of Agricultural Food Chemistry* 55: 3920-3925.
- Choi, T.Y., Kim, J.H., Dong, H.K., Kim, C.H., Hwang, J.S., Ahn, S., Sun, Y.K., Kim, C.D., Lee, J.H. and Yoon, T.J. (2007b). Zebrafish as a new model for phenotype-based screening of melanogenic regulatory compounds. *Pigment Cell Research* 20(2): 120-127.
- Choi, Y.K., Rho, Y.K., Yoo, K.H., Lim, Y.Y., Li, K., Kim, B.J., Seo, S.J., Kim, M.N., Hong, C.K. and Kim, D.S. (2010). Effects of vitamin C vs. multivitamin on melanogenesis: comparative study *in vitro* and *in vivo*. *International Journal of Dermatology*. 49(2): 218-226.
- Choo, S.J., Ryoo, I.J., Kim, Y.H., Xu, G.H., Kim, W.G., Kim, K.H., Moon, S.J., Son, E.D., Bae, K. and Yoo, I.D. (2009). Silymarin inhibits melanin synthesis in melanocyte cells. *Journal of Pharmacy and Pharmacology* 61(5): 663-667.
- Chou, S.T., Chang, W.L., Chang, C.T., Hsu, S.L., Lin, Y.C. and Shih, Y. (2013). *Cinnamomum cassia* essential oil inhibits α -MSH-induced melanin production and oxidative stress in murine B16 melanoma cells. *International Journal of Molecular Science* 14(9): 19186–19201.
- Chowdary, G. V. and Prapulla, S. G. (2002). The influence of water activity on the lipase catalysed synthesis of butyl butyrate transesterification, *Process Biochemistry*, 38(3): 393-397.
- Chowdary, G. V., Ramesh, M. N. and Prapulla, S. G. (2000). Enzymic synthesis of isoamyl isovalerate using immobilized lipase from *Rhizomucor miehei*: a multivariate analysis. *Process Biochemistry* 36(4): 331-339.
- Chusiri, Y., Wongpoomchai, R., Kakekashi, A., Wei, M., Wanibuchi, H., Vinitketkumnuan, U. and Fukushima, S. (2011). Non-genotoxic mode of action and possible threshold for hepatocarcinogenicity of kojic acid in F344 rats. *Food and Chemical Toxicology* 49(2): 471-476.

- Cichorek, M., Wachulska, M., Stasiewicz, A. and Tyminińska, A. (2013). Skin melanocytes: biology and development. *Postepy Dermatologii Alergologii* 30(1): 30–41.
- Clendennen S. K., Boaz, N. W. and Clauson, J. M. (2012). E.P Patent No. 2,155,886 B1. London, G.B.: European Patent Office.
- Cnop, M., Hannaert, J.C., Hoorens, A., Eizirik, D.L. and Pipeleers, D.G. (2001). Inverse relationship between cytotoxicity of free fatty acids in pancreatic islet cells and cellular triglyceride accumulation. *Diabetes* 50(8): 1771-1777.
- Coda, R., Rizzello, C.G., Pinto, D. and Gobbetti, M. (2012). Fermentation of cereal flours antioxidant peptides during sourdough selected lactic acid bacteria synthesize. *Applied Environmental Microbiology* 78(4): 1087-1094.
- Colombié, S., Tweddell, R.J., Condoret, J.S. and Marty, A. (1998). Water activity control: a way to improve the efficiency of continuous lipase esterification. *Biotechnology and Bioengineering* 60(3): 362-368.
- Cosmetic Ingredient Review Expert Panel (1987). 3: Final report on the safety assessment of oleic acid, lauric acid, palmitic acid, myristic acid, and stearic acid. *International Journal of Toxicology* 6(3): 321-401.
- Costin, G.E. and Hearing, V.J. (2007). Human skin pigmentation: melanocytes modulate skin color in response to stress. *The Journal of the Federation of American Societies for Experimental Biology* 21(4): 976 -994.
- Costin G.E. and Rom H.R. (2013). Optimized *in vitro* pigmentation screening assay using a reconstructed three dimensional human skin model. *Journal of Chemistry* 50(1):15-27.
- Csanádi, Z., Katalin, B.B., Szentgyörgyi, E., Gubicza, L., Knez, E. and Habulin, M. (2010). Enzymatic esterification of glycerol and stearic acid in non-conventional media. *Acta Chimica Slovenica* 57(1): 244-249.
- da Silva, M.C., Naozuka, J., Oliveira, P.V., Vanetti, M.C., Bazzolli, D.M., Costa, N.M. and Kasuya, M.C. (2010). *In vivo* bioavailability of selenium in enriched *Pleurotus ostreatus* mushrooms. *Metallomics* 2(2): 162-126.
- Dahlan, I., Kamaruddin, A.H. and Najafpour, G.D. (2005). Citronellyl butyrate synthesis in non-conventional media using packed-bed immobilized *candida rugosa* lipase reactor. *International Journal of Engineering* 18(2): 153-164.
- Del Marmol, V., Solano, F., Sels, A., Huez, G., Libert, A., Lejeune, F. and Ghanem, G. (1993). Glutathione depletion increases tyrosinase activity in human melanoma cells. *Journal of Investigate Dermatology* 101: 871-874.

- Deng, L., Wang, X.J, Nie, K.L., Wang, F., Liu, J.F, Wang, P. and Tan, T.W. (2011). Synthesis of wax esters by lipase-catalyzed esterification with immobilized lipase from *Candida* sp. 99-125. *Chinese Journal of Chemical Engineering* 19(6): 978-982.
- Denvir, M.A., Tucker, C.S. and Mullins, J.J. (2008). Systolic and diastolic ventricular function in zebrafish embryos: Influence of norepinephrine, MS-222 and temperature. *BioMed Central Biotechnology* 8: 21.
- Deschodt-Lanckman, M., Vanneste, Y., Loir, B., Michel, A., Libert, A., Ghanem, G. and Lejeune, F. (1990). Degradation of alpha-melanocyte stimulating hormone (alpha-MSH) by CALLA/endopeptidase 24.11 expressed by human melanoma cells in culture. *International Journal of Cancer* 46(6):1124-30.
- Dhongade, H.J. and Chandewar, A.V. (2013). An *in-vitro* investigation of the antioxidant activity of *Phyllanthus amarus*. *International Journal of Biomedical And Advance Research* 4(6): 435-439.
- Ding, H.Y., Chang, T.S., Chiang, C.M., Li, S.Y. and Tseng, D.Y. (2011). Melanogenesis inhibition by crude extract of *Magnolia officinalis*. *Journal of Medicinal Plants Research* 5(2): 237-244.
- Divakar, R. K., Upadhyaya, P., Sharma, M. and Gupta, P. K. (2012). Noninvasive imaging of ethanol-induced developmental defects in zebrafish embryos using optical coherence tomography K. *Birth Defects Research B: Developmental Reproduction Toxicology* 95(1): 7–11.
- Dowd, P. F. (1990). U.S Patent No. 4,956,353. Washington, D.C.: U.S Patent and Trademark Office.
- Drozd, R., Hintermann, E., Tanner, H., Zumsteg, U. and Eberle, A.N. (1999). (D-(p-benzoylphenylalanine)¹³, tyrosine¹⁹)-melanin-concentrating hormone, a potent analogue for MCH receptor crosslinking. *Journal of Peptide Science* 5(5): 234-242.
- Drozd, R., Siegrist, W., Baker, B.I., Chluba-de Tapia, J. and Eberle, A.N. (1995). Melanin-concentrating hormone binding to mouse melanoma cells in vitro. *FEBS Letters* 359(2-3):199-202.
- Dubey, S.K., Misra, K., Tiwari, A. and Bajaj, A.K. (2006). Chemically induced pigmentary changes of human skin, interaction of some azo dyes with human DNA. *Journal of Pharmacology and Toxicology* 1(3): 234-247.
- Ebanks, J.P., Wickett, R.R. and Boissy, R.E. (2009). Mechanisms regulating skin pigmentation: The rise and fall of complexion coloration. *International Journal of Molecular Science* 10(9): 4066–4087.
- EFSA Panel on Food Additives and Nutrient Sources added to Food (ANS) (2011). Scientific opinion on the re-evaluation of butylated

hydroxyanisole–BHA (E 320) as a food additive. *European Food Safety Authority Journal* 9(10): 2392-2441.

EFSA Panel on Food Additives and Nutrient Sources added to Food (ANS) (2012). Scientific Opinion on the reevaluation of butylated hydroxytoluene BHT (E 321) as a food additive. *European Food Safety Authority Journal* 10(3): 2588-2631.

Eisenhofer, G., Tian, H., Holmes, C., Matsunaga, J., Roffler, S.T. and Hearing, V.J. (2003). Tyrosinase: a developmentally specific major determinant of peripheral dopamine. *The Federation of American Societies for Experimental Biology Journal* 1248-1255.

El-Aasar, S. A. (2006). Cultural conditions studies on kojic acid production by *Aspergillus parasiticus*. *International Journal of Agricultural Biology* 8(4): 468-473.

El-Boulifi, N., Ashari, S.E., Serrano, M., Aracil, J. and Martínez, M. (2014). Solvent-free lipase-catalyzed synthesis of a novel hydroxyl-fatty acid derivative of kojic acid. *Enzyme and Microbial Technology* 55: 128-132.

El-Kady, I.A., Zohri, A.N.A. and Hamed, S.R. (2014). Kojic acid production from agro-industrial by-products using fungi. *Biotechnology Research International* 2014: 642385.

Emami, S., Ghafouri, E., Faramarzi, M.A., Samadi, N., Irannejad, H. and Foroumadi, A. (2013). Mannich bases of 7-piperazinylquinolones and kojic acid derivatives: Synthesis, *in vitro* antibacterial activity and in silico study. *European Journal of Medicinal Chemistry* 68C: 185-191.

Fang, Y.Z., Yang, S. and Wu, G. (2002). Free radicals, antioxidants, and nutrition. *Nutrition* 18(10): 872-879.

Farard, J., Logé, C., Pfeiffer, B., Lesur, B. and Duflos, M. (2009). A convenient synthesis of 5-arylamino-4H-pyran-4-ones using palladium-catalyzed amination. *Tetrahedron Letters* 50(41): 5729–5732.

Farishian, R.A. and Wittaker, J.H. (1980). Phenylalanine lowers melanin synthesis in mammalian melanocytes by reducing tyrosinase uptake: Implications for pigment reduction in phenylketonuria. *Journal of Investigative Dermatology* 74(2): 85–89.

Ferrer, M., Soliveri, J., Plou, F.J., López-Cortés, N., Reyes-Duarte, D., Christensen, M., Copa-Patiño, J.L. and Ballesteros, A. (2005). Synthesis of sugar esters in solvent mixtures by lipases from *Thermomyces lanuginosus* and *Candida antarctica* B, and their antimicrobial properties. *Enzyme and Microbial Technology* 36: 391-398.

Fernandes, A.G.A., Maia, P.I.S., Souza, E.J., Lemos, S.S., Batista, A.A., Abram, U., Ellena, J., Castellano, E.E. and Deflon, V.M. (2008).

Rhenium chelate complexes with maltolate or kojate. *Polyhedron* 27: 2983–2989.

- Fickova, M., Pravdova, E., Rondhal, L., Uher, M. and Brtko, J. (2008). *In vitro* antiproliferative and cytotoxic activities of novel kojic acid derivatives: 5-benzyloxy-2-selenocyanatomethyl- and 5-methoxy-2-selenocyanatomethyl-4-pyranone. *Journal of Applied Toxicology* 28(4): 554-559.
- Fitzhugh, O.R., Schouboe, P.J. and Nelson, A.A. (1960). Oral toxicities of laurie acid and certain laurie acid derivatives. *Toxicology and applied pharmacology* 2(1): 59-67.
- Forouzani, M. and Hassan, G.B. (2014). Efficient one-pot pTSA-catalyzed Synthesis of 2-substituted aryl (indolyl)kojic acid and kojyl thioether derivatives under mild conditions. *The Open Organic Chemistry Journal* 8:1-5.
- Foti, M., Piattelli, M., Baratta, M.T. and Ruberto, G. (1996). Flavonoids, coumarins, and cinnamic acids as antioxidants in a micellar system. Structure-activity relationship. *Journal of Agricultural Food Chemistry* 44(2): 497-501.
- Fu, X.F., Zheng, J.Y., Ying, X.X., Yan, H.D. and Wang, Z. (2014). Investigation of Lipozyme TL IM-catalyzed transesterification using ultraviolet spectrophotometric assay. *Chinese Journal of Catalysis* 35:553-559.
- Funasaka, Y., Komoto, M. and Ichihashi, M. (2000). Depigmenting effect of alpha-tocopheryl ferulate on normal human melanocytes. *Pigment Cell Research* 13(8): 170-174.
- Gandhi, N.N., Vijayalakshmi, V., Sawant, S.B. and Joshi J.B. (1996). Immobilization of *Mucor miehei* lipase on ion exchange resins. *The chemical engineering journal* 61: 149-156.
- Garg, D., Shaikh, A., Muley, A. and Marar, T. (2012). *In-vitro* antioxidant activity and phytochemical analysis in extracts of *Hibiscus rosa-sinensis* stem and leaves. *Free Radicals and Antioxidants* 2(3): 2012.
- Gellin, G.A., Possick, P.A. and Perone, V.B. (1970). Depigmentation from 4-tertiarybutyl catechol an experimental study. *Journal of Investigate Dermatology* 55(3): 190-197.
- Gonçalez, M.L., Corrêa, M.A. and Chorilli, M. (2013). Skin delivery of kojic acid-loaded nanotechnology-based drug delivery systems for the treatment of skin aging. *BioMed Research International* 2013: 271276.
- Gu, F., Kim, M.J., Hayat, K., Xia, S., Feng, B. and Zhang, X. (2009). Characteristics and antioxidant activity of ultrafiltrated Maillard reaction products from a caseinglucose model system. *Food Chemistry* 117(1): 48-54.

- Gulati, V., Harding, I.H. and Palombo, E.A. (2012). Enzyme inhibitory and antioxidant activities of traditional medicinal plants: Potential application in the management of hyperglycemia. *BMC Complementary and Alternative Medicine* 12:77.
- Gülçin, I. (2005). The antioxidant and radical scavenging activities of black pepper (*Piper nigrum*) seeds. *International Journal of Food Sciences and Nutrition* 56(7): 491–499.
- Gülçin, I. (2006). Antioxidant activity of caffeic acid (3,4-dihydroxycinnamic acid). *Toxicology* 217(2-3): 213-220.
- Gunnarsson, L., Jauhiainen, A., Kristiansson, E., Nerman, O. and Larsson, D.G. (2008). Evolutionary conservation of human drug targets in organisms used for environmental risk assessments. *Environmental Science and Technology* 42(15): 5807-5813.
- Ha, J.H., Kang, W.H., Lee, J.O., Cho, Y.K., Park, S.K., Lee, S.K., and Cho, H.K. (2011). Clinical evaluation of the depigmenting effect of *Glechoma Hederacea* extract by topical treatment for 8 weeks on UV-induced pigmentation in Asian skin. *European Journal of Dermatology* 21(2): 218-222.
- Ha, S.K., Koketsu, M., Lee, K., Choi, S.Y., Park, J.H., Ishihara, H., and Kim, S.Y. (2005). Inhibition of tyrosinase activity by N,N-unsubstituted selenourea derivatives. *Biological Pharmaceutical Bulletin* 28(5): 838-840.
- Habulin, M., Krmelj, V. and Knez, Z. (1996). Synthesis of Oleic Acid Esters Catalyzed by Immobilized Lipase. *Journal of Agricultural and Food Chemistry* 44(1): 338–342
- Halim, S.F.A., A.H. Kamaruddin and Fernando, W.J.N. (2009). Continuous biosynthesis of biodiesel from waste cooking palm oil in a packed bed reactor: Optimization using Response Surface Methodology (RSM) and mass transfer studies. *Bioresource Technology* 100(2): 710-716.
- Hamed S.H., Sriwiriyanont, P., deLong, M.A., Visscher, M.O., Wickett, R.R. and Boissy R.E. (2006). Comparative efficacy and safety of deoxyarbutin, a new tyrosinase-inhibiting agent. *Journal of Cosmetic Science* 57(4): 291-308.
- Hamid, M.A., Sarmidi, M.R. and Park, C.S. (2012). Mangosteen leaf extract increases melanogenesis in B16F1 melanoma cells by stimulating tyrosinase activity *in vitro* and by up-regulating tyrosinase gene expression. *International Journal of Molecular Medicine* 29(2): 209-217.
- Hassan, M. A., Ismail, F., Yamamoto, S., Yamada, H. and Nakanishi, K. (1995). Enzymatic synthesis of galactosylkojic acid with immobilized β -

galactosidase from *Bacillus circulans*. *Bioscience Biotechnology and Biochemistry* 59(3): 543-545.

Hearing, V.J. (2009). The expanding role and presence of neuromelanins in the human brain—why gray matter is gray. *Pigment Cell and Melanoma Research* 22(1): 10–11.

Heo, S.J., Ko, S.C., Kang, S.M., Cha, S.H., Lee, S.H., Kang, D.H., Jung, W.K., Affan, A., Oh, C. and Jeon, Y.J. (2010). Inhibitory effect of diphlorethohydroxycarmalol on melanogenesis and its protective effect against UV-B radiation-induced cell damage. *Food and Chemical Toxicology* 48(5): 1355-1361.

Higdon, J.V and Frei, B. (2003). Tea catechins and polyphenols: health effects, metabolism, and antioxidant functions. *Critical Review Food Science Nutrition* 43(1): 89-143.

Hill, A.J., Teraoka, H., Heideman, W. and Peterson, R.E. (2005). Zebrafish as a model vertebrate for investigating chemical toxicity. *Toxicological Sciences*, 86(1): 6–19.

Hill, S.E., Buffey, J., Thody, A.J., Oliver, I., Bleehen, S.S. and MacNeil, S. (1989). Investigation of the regulation of pigmentation in alpha-melanocyte-stimulating hormone responsive and unresponsive cultured B16 melanoma cells. *Pigment Cell Research* 2(3): 161–166.

Hilterhaus, L., Minow, B., Müller, J., Berheide, M., Quitmann, H., Katzer, M., Thum, O., Antranikian, G., Zeng, A.P. and Liese, A. (2008). Practical application of different enzymes immobilized on sephabeads. *Bioprocess and Biosystems Engineering* 31(3): 163-171.

Hirata, M., Nakamura, K.I. and Kondo, S. (2005). Pigment cell distributions in different tissues of the zebrafish, with special reference to the striped pigment pattern. *Developmental Dynamic* 234(2): 293–300.

Hishida, T.O., Tomita, H. and Yamamoto, T.O. (1961). Melanin formation in color varieties of the medaka (*Oryzias latipes*). *Embryologia* 5(4): 335–346.

Hlavsov'a, K., Wimmer, Z., Xanthakis, E., Bern'ašek, P., Sovov'a, H and Zarev 'uckae, M. (2008). Lipase activity enhancement by SC-CO₂ treatment. *Z. Naturforsch* 63b: 779-784.

Hoq, M.M. Yamane, T. and Shimizu, S. (1984). Continuous synthesis of glycerides by lipase in a microporous membrane bioreactor. *Journal of American Oil Chemists' Society* 61(4): 776-781.

Hoshino, S., Nishimura, M., Fukuyama, K., Gellin, G.A and Epstein, J.H. (1981). Effects of 4-tertiary butyl catechol on melanocytes of hairless mice. *Journal of Investigative Dermatology* 76(4): 231–238.

- Hsieh, H.J., Giridhar, R. and Wu, W.T. (2007). Regioselective formation of kojic acid-7-o- α -D-glucopyranoside by whole cells of mutated *Xanthomonas campestris*. *Enzyme and Microbial Technology* 40(2): 324-328.
- Hsieh, J.F., Chen, S.T. and Cheng, S.L (2011). Molecular profiling of A375 human malignant melanoma cells treated with kojic acid and arbutin, Breakthroughs in Melanoma Research, Dr Yohei Tanaka (Ed.), InTech publisher.
- Hu, Z.M., Zhou, Q., Lei, T.C., Ding, S.F. and Xu, S.Z. (2009). Effects of hydroquinone and its glucoside derivatives on melanogenesis and antioxidation: Biosafety as skin whitening agents. *Journal of Dermatological Science* 55(3):179-184.
- Huang, H.P., Shih, Y.W., Chang, Y.C., Hung, C.N. and Wang, C.J. (2008a). Chemoinhibitory effect of mulberry anthocyanins on melanoma metastasis involved in the Ras/PI3K pathway. *Journal of Agricultural and Food Chemistry* 56(19): 9286–9293.
- Huang, Y.H., Lee, T.H., Chan, K.J., Hsu, F.L., Wu, Y.C. and Lee, M.H. (2008b). Anemonin is a natural bioactive compound that can regulate tyrosinase-related proteins and mRNA in human melanocytes. *Journal of Dermatological Science* 49(2): 115-23.
- Hudecová, D., Jantová, S., Melník, M. and Uher, M. (1996). New azidometalkojates and their biological activity. *Folia Microbiologica* 41(6): 473-476.
- Hurst, E.A., Harbour, J.W. and Cornelius, L.A. (2003). Ocular melanoma: a review and the relationship to cutaneous melanoma. *Archives of Dermatology* 139(8): 1067–1073.
- Hussein-Al-Ali, S.H., El-Zowalaty, M.E., Hussein, M.Z., Ismail, M., Dorniani, D. and Webster, T.J. (2014). Novel kojic acid-polymer-based magnetic nanocomposites for medical applications. *International Journal of Nanomedicine* 9(1): 351–362.
- Ishak, A. 2015. Enzymatic esterification of kojic acid and palmitic acid by immobilized lipase for the synthesis of kojic acid palmitate. Master thesis. Universiti Putra Malaysia.
- Ismail, M., Mariod, A., Bagalkotkara G. and Ling, H.S (2010). Fatty acid composition and antioxidant activity of oils from two cultivars of antalupe extracted by supercritical fluid extraction. *Grasas Y Aceites* 61(1): 37-44.
- Ito, Y., Jimbow, K. and Ito, S. (1987). Depigmentation of black guinea pig skin by topical application of cysteaminyphenol, cysteinylphenol, and related compounds. *Journal of Investigative Dermatology* 88(1): 77–82.

- Jakovetic, S.M., Lukovic, N.D., Boskovic-Vragolovic, N.M., Bezbradica, D.I., Picazo-Espinosa, R. and Knezevic-Jugovic, Z.D. (2013). Comparative study of batch and fluidized bed bioreactors for lipase-catalyzed ethyl cinnamate synthesis. *Industrial and Engineering Chemistry Research* 52(47): 16689-16697.
- Jean, J., Angers, L., Gendreau, I. and Pouliot, R. (2013). Pigmented Skin Models: Understand the Mechanisms of Melanocytes. In J.A. Andrades (Ed.), *Regenerative Medicine and Tissue Engineering* (pp. 759-785). Croatia: In Tech Publishers.
- Jeong, Y.M., Oh, W.K., Tran, T.L., Kim, W.K., Sung, S.H., Bae, K., Lee, S. and Sung, J.H. (2013). Aglycone of rh₄ inhibits melanin synthesis in B16 melanoma cells: Possible involvement of the protein kinase A pathway. *Bioscience, Biotechnology and Biochemistry* 77(1): 119-125.
- Jerkovic, I. and Marijanovic, Z.O. (2010). Oak (*Quercus frainetto* Ten.) Honeydew honey-approach to screening of volatile organic composition and antioxidant capacity (DPPH and FRAP Assay). *Molecules* 15(5): 3744-3756.
- Jiang, F., Guo, N. and Disting, G.J. (2008). Modulation of nicotinamide adenine dinucleotide phosphate oxidase expression and function by 3,4-dihydroxyflavonol in phagocytic and vascular cells. *Journal of pharmacology and experimental therapeutics* 324(1): 261–269.
- Jin, E.J. and Thibaudeau, G. (1999). Effects of lithium on pigmentation in the embryonic zebrafish (*Brachydanio rerio*). *Biochimica et Biophysica Acta* 1449(1): 93-99.
- Jin, Z., Ntwali, J., Han, S. Y., Zheng, S. P. and Lin, Y. (2012). Production of flavor esters catalyzed by CALB-displaying *Pichia pastoris* whole-cells in a batch reactor. *Journal of Biotechnology* 159(1-2): 108–114.
- Jonfia-Essien, W.A., West, G., Alderson, P.G. and Tucker, G. (2008). Phenolic content and antioxidant capacity of hybrid variety cocoa beans. *Food Chemistry* 108(3): 1155–1159.
- Ju, H.Y., Yang, C.K., Yen, Y.H. and Shieh, C.J. (2009). Continuous lipase-catalyzed synthesis of hexyl laurate in a packed-bed reactor: optimization of the reaction conditions in a solvent-free system. *Journal of Chemical Technology and Biotechnology* 84(1): 29–33.
- Kaatz, H., Streffer, K., Wollenberger, U. and Peter, M.G. (1999). Inhibition of mushroom tyrosinase by kojic acid octanoates. *Z. Naturforsch*, 54c:70-74.
- Kadam, R.S., Scheinman, R.I. and Kompella, U.B. (2012). Pigmented-MDCK (P-MDCK) cell line with tunable melanin expression: an *in vitro* model for the outer blood-retinal barrier. *Molecular Pharmaceutics* 9(11): 3228-35.

- Kadekaro, A.L., Andrade, L.N., Floeter-Winter, L.M., Rollag, M.D., Virador, V., Vieira, W. and Castrucci, A.M. (2004). MT-1 melatonin receptor expression increases the antiproliferative effect of melatonin on S-91 murine melanoma cells. *Journal of Pineal Research* 36(3):204-11.
- Kadokawa, J., Nishikura, T., Muraoka, R., Tagaya, H. and Fukuoka, N. (2003). Synthesis of kojic acid derivatives containing phenolic hydroxy groups. *Synthetic Communication* 33(7): 1081-1086.
- Kahl, R. and Kappus, H. (1993). Toxicology of the synthetic antioxidants BHA and BHT in comparison with the natural antioxidant vitamin E. *Z. Lebensm Unters Forsch.* 196(4):329-338.
- Kamei, Y., Otsuka, Y. and Abe, K. (2009). Comparison of the inhibitory effects of vitamin E analogues on melanogenesis in mouse B16 melanoma cells. *Cytotechnology* 59:183–190.
- Kang, S.S., Kim, H.J., Jin, C. and Lee, Y.S. (2009). Synthesis of tyrosinase inhibitory (4-oxo-4H-pyran-2-yl) acrylic acid ester derivatives. *Bioorganic and Medicinal Chemistry Letters* 19(1): 188–191.
- Kanwar, S.S., Kaushal, R.K., Verma, M.L., Kumar, Y., Chauhan, G.S., Gupta, R. and Chimni, S.S. (2005). Synthesis of ethyl laurate by hydrogel immobilized lipase of *Bacillus coagulans* MTCC-6375. *Indian Journal of Microbiology* 45(3): 187-193.
- Kapucu, N., Guvenc, A., Toglu, U. M., Alimi, A. and Kapucu, H. (2003). Lipase catalysed synthesis of oleyl oleate: Optimization by response surface methodology. *Chemical Engineering Communication* 190(5): 779-796.
- Karlsson, J., von-Hofsten, J. and Olsson, P.E. (2001). Generating transparent zebrafish: a refined method to improve detection of gene expression during embryonic development. *Marine Biotechnology* 3(6):522-527.
- Karakaya, G., Aytemir, M.D., Özçelik, B. and Çalıř, Ü. (2013). Design, synthesis and *in vivo/in vitro* screening of novel chlorokojic acid derivatives. *Journal of Enzyme Inhibition and Medicinal Chemistry* 28(3): 627-638.
- Kasraee, B., Handjani, F., Parhizgar, A., Omrani, G.R., Fallahi, M.R., Amini, M., Nikbakhsh, M., Tran, C., Hügin, A., Sorg, O. and Saurat, J.H. (2005). Topical methimazole as a new treatment for postinflammatory hyperpigmentation: Report of the First Case. *Dermatology* 211(4): 360–362.
- Kasraee, B., Nikolic, D.S., Salomon, D., Carraux, P., Fontao, L., Piguet, V., Omrani, G.R., Sorg, O. and Saurat, J.H. (2012). Ebselen is a new skin depigmenting agent that inhibits melanin biosynthesis and melanosomal transfer. *Experimental Dermatology* 21(1): 19-24.

- Kasser, J.H., Kandioller, W., Hartinger, C.G., Nazarov, A.A., Aron, V.B., Dyson, P.J. and Keppler, B.K. (2010). Mannich products of kojic acid and N-heterocycles and their Ru (II)-arene complexes: Synthesis, characterization and stability. *Journal of Organometallic Chemistry* 695: 875-881
- Kawaguchi, S., Nakamura, T., Honda, G., Yokohama, N. and Sasaki, Y.F. (2008). *In vivo* genotoxic potential of kojic acid in rodent multiple organs detected by the comet assay. *Genes and environment* 30(1): 25-32.
- Kawashima, A., Shimada, Y., Yamamoto, M., Sugihara, A., Nagao, T., Komemushi, S. and Tominaga, Y. (2001). Enzymatic synthesis of high-purity structured lipids with caprylic acid at 1,3-positions and polyunsaturated fatty acid at 2-position. *Journal of the American Oil Chemists Society* 78(6): 611-616.
- Kawashima, T., Yonemoto, K., Gellin, G.A., Epstein, W.L. and Fukuyama, K. (1984). Effects of 4-Tertiary butyl catechol on glutathione-metabolizing enzymes *in vivo* and *in vitro*. *Journal of Investigative Dermatology* 82(1): 53–56.
- Kayahara, H., Shibata, N., Tadasa, K., Maeda, H., Kotani, T. and Ichimoto, I (1990). Amino acid and peptide derivatives of kojic acid and their antifungal properties. *Agricultural and Biological Chemistry* 54(9): 2441-2442.
- Kelsh, R.N., Brand, M., Jiang, Y.J., Heisenberg, C.P., Lin, S., Haffter, P., Odenthal, J., Mullins, M.C., van Eeden, F.J., Furutani-Seiki, M., Granato, M., Hammerschmidt, M., Kane, D.A., Warga, R.M., Beuchle, D., Vogelsang, L. and Nüsslein-Volhard, C. (1996). Zebrafish pigmentation mutations and the processes of neural crest development. *Development* 123:369-389.
- Keng, P.S., Basri, M., Ariff, A., Abdul Rahman, M.B., Abdul Rahman, R. N. Z., and Salleh, A. B. (2008). Scale-up synthesis of lipase-catalyzed palm esters in stirred-tank reactor. *Bioresource Technology* 99(14): 6097–6104.
- Khamaruddin, N.H., Basri, M., Lian, G.E.C., Salleh, A.B., Abdul-Rahman, R.N.Z.R., Ariff, A., Mohamad, R., and Awang, R. (2008). Enzymatic synthesis and characterization of palm-based kojic acid ester. *Journal of Oil Palm Research* 20: 461-469.
- Khan, R.A., Khan, M.R., Sahreen, S. and Ahmed, M. (2012). Assessment of flavonoids contents and *in vitro* antioxidant activity of *Launaea procumbens*. *Chemistry Central Journal* 6: 43.
- Khopde, S.M., Priyadarsini, K.I., Guha, S.N., Satav, J.G., Venkatesan, P. and Rao, M.N. (2000). Inhibition of radiation-induced lipid peroxidation by tetrahydrocurcumin: possible mechanisms by pulse radiolysis. *Bioscience, Biotechnology and Biochemistry* 64(3): 503-509.

- Kim, D.H., Hwang, J.S., Baek, H.S., Kim, K.J., Lee, B.G., Chang, I., Kang, H.H. and Lee, O.S. (2003). Development of 5-[(3-aminopropyl) phosphinoxy]-2-(hydroxymethyl)-4H-pyran-4-one as a novel whitening agent. *Chemical Pharmaceutical Bulletin* 51(2): 113-116.
- Kim, D.S., Park, S.H., Kwon, S.B., Li, K., Youn, S.W. and Park, K.C. (2004a). (-)-Epigallocatechin-3-gallate and hinokitiol reduce melanin synthesis via decreased MITF production. *Archives of Pharmacol Research* 27(3): 334–339.
- Kim, H., Choi, J., Cho, J.K., Kim, S.Y. and Lee, Y.S. (2004b). Solid-phase synthesis of kojic acid-tripeptides and their tyrosinase inhibitory activity, storage stability, and toxicity. *Bioorganic and Medicinal Chemistry Letters* 14(11): 2843-2846.
- Kim, J.H., Chang, P.K., Chan, K.L., Faria, N.C., Mahoney, N., Kim, Y.K., Martins, Mde. L. and Campbell, B.C. (2012). Enhancement of commercial antifungal agents by kojic acid. *International Journal of Molecular Sciences* 13(11): 13867-13880.
- Kim, K.D., Song, M.H., Yum, E.K., Jeon, O.S., Ju, Y.W. and Chang, M.S., (2010). Melanogenesis Inhibition by mono-hydroxycinnamic ester derivatives in B16 melanoma cells. *Bulletin Korean Chemistry Society* 31(1): 181-184.
- Kim, S.S., Kim, M.J., Choi, Y.H., Kim, B.K., Kim, K.S., Park, K.J., Park, S.M., Lee, N.H. and Hyun, C.G. (2013). Down-regulation of tyrosinase, TRP-1, TRP-2 and MITF expressions by citrus press-cakes in murine B16F10 melanoma. *Asian Pacific Journal of Tropical Biomedicine* 3(8): 617–622.
- Kim, Y.J. (2007). Anti-melanogenic and antioxidant properties of gallic acid. *Biological Pharmacological Bulletin* 30(6):1052–1055.
- Kim, Y.J., Kang, K.S. and Yokozawa, T. (2008). The anti-melanogenic effect of pycnogenol by its anti-oxidative actions. *Food and Chemical Toxicology* 46(7): 2466–2471.
- Kimmel, C.B., Ballard, W.W., Kimmel, S.R., Ullmann, B. and Schilling, T.F. (1995). Stages of embryonic development of the zebrafish. *Development Dynamic* 203(3): 253-310.
- Kimmel, C.B. (1989). Genetics and early development of zebrafish. *Trends Genetic* 5(8):283-288.
- Kinoshita, M., Hori, N., Aida, K., Sugawara, T. and Ohnishi, M. (2007). Prevention of melanin formation by yeast cerebroside in B16 mouse melanoma cells. *Journal of Oleo Science* 56(12): 645–648.
- Kinoshita, M., Morita, T., Toyohara, H., Hirata, T., Sakaguchi, M., Ono, M., Inoue, K., Wakamatsu, Y. and Ozato, K. (2001). Transgenic medaka

overexpressing a melanin-concentrating hormone exhibit lightened body color but no remarkable abnormality. *Marine Biotechnology* 3(6): 536-543.

Kitao, S. and Serine, H. (1994). Syntheses of two kojic acid glucosides with sucrose phosphorylase from *Leuconostoc mesenteroides*. *Bioscience, Biotechnology, and Biochemistry* 58(2):419-420.

Knez, D. Ž., Leitgeb, D. M. and Lavrič, Z. B. (1990). Synthesis of oleic acid esters with immobilized lipase. *European Journal of Lipid Science and Technology* 92(4): 169–172.

Knez, Z. and Habulin, M. (1994). Lipase-catalyzed esterification at high pressure. *Biocatalysis* 9(1-4): 115-121

Kobayashi, T., Adachi, S., Nakanishi, K. and Matsuno, R. (2001). Semi-continuous production of lauroyl kojic acid through lipase-catalysed condensation in acetonitrile. *Biochemical Engineering Journal* 9(2): 85-89.

Kobayashi, Y., Kayahara, H., Tadasa, K. and Tanaka, H. (1996). Synthesis of N-kojic-amino acid and N-kojic-amino acid-kojiolate and their tyrosinase inhibitory activity. *Bioorganic and Medicinal Chemistry Letters* 6(12): 1303-1308.

Kobayashi, Y., Kayahara, H., Tadasa, K., Nakamura, T. and Tanaka, H. (1995). Synthesis of amino acid derivatives of kojic acid and their tyrosinase inhibitory activity. *Bioscience Biotechnology and Biochemistry* 59(9): 1745-1746.

Kobori, M., Sasaki, Y. and Shinohara, K. (1994). Suppressive effect of spinach extract on the formation of melanin in B16 mouse melanoma cells. *Cytotechnology* 14: 119-122.

Kolbe L, Mann T, Gerwat W, Batzer J, Ahlheit S, Scherner C, Wenck H, Stäb F. (2013). 4-n-butylresorcinol, a highly effective tyrosinase inhibitor for the topical treatment of hyperpigmentation. *Journal of Europe Academy Dermatology Venereology* 27(1): 19-23.

Korytowski, W. and Sarna, T. (1990). Bleaching of melanin pigments. Role of copper ions and hydrogen peroxide in autooxidation and photooxidation of synthetic dopa-melanin. *Journal of Biological Chemistry* 265(21): 12410-12416.

Kosugi, Y., Tanaka, H. and Tomizuka, N. (1990). Continuous hydrolysis of oil by immobilized lipase in a countercurrent reactor. *Biotechnology and Bioengineering* 36(6): 617-622.

Kotani, T., Ichimoto, I., Tatsumi, C. and Fujita, T. (1976). Bacteriostatic activities and metal chelation of kojic acid analogs. *Agricultural and Biological Chemistry* 40(4): 765-770.

- Kotyzová, D., Eybl, V., Koutenský, J., Brtko, J. and Glattre, E. (2004). Effects of kojic acid on oxidative damage and on iron and trace element level in iron-overloaded mice and rats. *Central European Journal of Public Health* 12: S41-44.
- Krishna, S.H., Sattur, A.P. and Karanth, N.G. (2001). Lipase-catalyzed synthesis of isoamyl isobutyrate-optimization using a central composite rotatable design. *Process Biochemistry* 37(1): 9-16.
- Krishnamoorthy, G., Chellappan, D.R., Joseph, J., Ravindhran, D., Shabi, M.M., Uthrapathy, S., Rajamanickam, V.G. and Dubey, G.P (2009). Antioxidant activity of *Nelumbo nucifera* (Gaertn) flowers in isolated perfused rat kidney. *Brazilian Journal of Pharmacognosy* 19(1B): 224-229.
- Krishnan, C.V., Garnett, M., Hsiao, B. and Chu, B. (2007). Electrochemical measurements of isopolyoxomolybdates: 1. pH dependent behavior of sodium molybdate. *International Journal of Electrochemical Science* 2: 29-51.
- Kubo, I., Masuoka, N., Ha, T.J. and Tsujimoto, K. (2006). Antioxidant activity of anacardic acids. *Food Chemistry* 99(3): 555–562.
- Kumar, K.J.S., Vani, M.G., Wang, S.Y., Liao, J.W., Hsu, L.S., Yang, H.L. and Hseu Y.C. (2013). *In Vitro* and *In Vivo* Studies Disclosed the Depigmenting Effects of Gallic Acid:A Novel Skin Lightening Agent for Hyperpigmentary Skin Diseases. *BioFactors* 39(3): 259–270.
- Kumar, S. and Kanwar S.S. (2011). Synthesis of ethyl ferulate in organic medium using celite-immobilized lipase. *Bioresource Technology* 102(3): 2162–2167.
- Kumar, S., Ola, R.P., Pahujani, S., Kaushal, R., Kanwar, S.S. and Gupta, R. (2006). Thermostability and esterification of a polyethylene-immobilized lipase from *Bacillus coagulans* BTS-3. *Journal of Applied Polymer Science* 102(4): 3986–3993.
- Kumar, R., Parsad, D., Kanwar, A. and Kaul, D. (2012). Development of melanocyte-keratinocyte co-culture model for controls and vitiligo to assess regulators of pigmentation and melanocytes. *Indian Journal of Dermatology, Venereology and Leprology* 78(5): 599-604.
- Kuo, C.H., Chen, H.H., Chen, J.H., Liu, Y.C. and Shieh, C.J. (2012). High yield of wax ester synthesized from cetyl alcohol and octanoic acid by lipozyme RMIM and Novozym 435. *International Journal of Molecular Sciences* 13(9): 11694-11704.
- Kwak, S.Y., Choi, H.R., Park, K.C. and Lee, Y.S. (2011a). Kojic acid-amino acid amide metal complexes and their melanogenesis inhibitory activities. *Journal of Peptide Science* 17(12): 791-797.

- Kwak, S.Y., Lee, S., Choi, H.R., Park, K.C. and Lee, Y.S. (2011b). Dual effects of caffeoyl-amino acidyl-hydroxamic acid as an antioxidant and depigmenting agent. *Bioorganic and Medicinal Chemistry Letters* 21(18): 5155-5158.
- Kwak, S.Y., Noh, J.M., Park, S.H., Byun, J.W., Choi, H.R., Park, K.C. and Lee, Y.S. (2010). Enhanced cell permeability of kojic acid-phenylalanine amide with metal complex. *Bioorganic and Medicinal Chemistry Letters* 20(2): 738-741.
- Laane, C., Boeren, S., Vos, K. and Veeger C. (1987). Rules for optimization of biocatalysis in organic solvents. *Biotechnology and Bioengineering* 30(1):81-87.
- Lai, K.L., Su, Y., Chen, R., Zhang, Z., Huang, Y. and Chen, Z.Y. (2001). Theaflavins in black tea and catechins in green tea are equally effective antioxidants. *Journal of Nutrition* 131(9): 2248-2251.
- Lam, U.D.P., Hoang, D.N., Lee, H.B., Kim, B., Lee, J.D., Shin, J.H. and Kim, E.K. (2011). Depigmenting effect of *Sterculia lynchophora* on B16F10 melanoma and C57BL/6 melan-a cells. *Korean Journal of Chemical Engineering* 28(4): 1074–1077.
- Larson, R.A. (1995). Plant defenses against oxidative stress. *Archives of Insect Biochemistry and Physiology* 29(2): 175–186.
- Laszlo, J. A., Jackson, M. and Blanco, R. M. (2011). Active-site titration analysis of surface influences on immobilized *Candida antarctica* lipase B activity. *Journal of Molecular Catalysis B: Enzymatic* 69(1-2): 60-65.
- Lawrence, C. (2007). The husbandry of zebrafish (*Danio rerio*): A review, *Aquaculture* 269: 1-20.
- Lee, H., Kim, E.H., Choi, H.R., Sohn, U.D., Yun, H.Y., Baek, K.J., Kwon, N.S., Park, K.C and Kim, D.S. (2012). Dipeptides inhibit melanin synthesis in Mel-Ab cells through down-regulation of tyrosinase. *Korean Journal of Physiology and Pharmacology* 16(4): 287–291.
- Lee, J., Jung, E., Park, J., Jung, K., Park, E., Kim, J., Hong, S., Park, J., Park, S., Lee, S. and Park, D. (2005). Glycyrrhizin induces melanogenesis by elevating a cAMP level in B16 melanoma cells. *Journal of Investigate Dermatology* 124:405-411.
- Lee, J.C., Kim, H.R., Jim, K. and Jang, Y.S. (2002a). Antioxidant property of an ethanol extract of the stem of *Opuntia ficus-indica* var *saboten*. *Journal of Agricultural and Food Chemistry* 50: 6490-6496.
- Lee, J.H., Jang, J.Y., Park, C., Kim, B.W., Choi, Y.H. and Choi, B.T. (2010). Curcumin suppresses alpha-melanocyte stimulating hormone-stimulated

melanogenesis in B16F10 cells. *International Journal of Molecular Medicine* 26(1): 101-106.

- Lee, J.H., Park, H., Chung, H., Choi, S., Kim, Y., Yoo, H., Kim, T.Y., Hann, H.J., Seong, I., Kim, J., Kang, K.G., Han, I.O. and Oh, E.S. (2009). Syndecan-2 regulates the migratory potential of melanoma cells. *Journal of Biological Chemistry* 284(40): 27167–27175.
- Lee, S.H., Choi, S.Y., Kim, H., Hwang, J.S., Lee, B.G., Gao, J.J. and Kim, S.Y. (2002b). Mulberroside F isolated from the leaves of *Morus alba* inhibits melanin. *Biosynthesis Biological Pharmaceutical Bulletin* 25(8): 1045-1048.
- Lee, Y.S., Park, J.H., Kim, M.H., Seo, S.H. and Kim, H.J. (2006). Synthesis of tyrosinase inhibitory kojic acid derivative. *Archive of Pharmacology* 339: 111-114.
- Levene, P.A. and West, C.J. (1914). Purification and melting points of saturated aliphatic acids. *Journal of Biological Chemistry* 18(3): 463-467.
- Li, C. and Wang, M.H. (2011). Antioxidant activity of peach blossom extracts. *Journal of Korean Society Applied Biology and Chemistry*. 54(1): 46-53.
- Li, H., Kim, J., Hahn, H.G., Yun, J., Jeong, H.S., Yun, H.Y., Baek, K.J., Kwon, N.S., Min, Y.S., Park, K.C. and Kim, D.S. (2014). KHG26792 inhibits melanin synthesis in Mel-Ab cells and a skin equivalent model. *Korean Journal of Physiology and Pharmacology* 18(3): 249–254.
- Li, X., Jeong, J. H., Lee, K. T., Rho, J. R., Choi, H. D., Kang, J. S. and Son, B. W. (2003). γ -pyrone derivatives, kojic acid methyl ethers from a marine-derived fungus *Altenaria* sp. *Archives of Pharmacal Research* 26(7): 532-534.
- Li, X., Lin, J., Gao, Y., Han, W. and Chen, D. (2012). Antioxidant activity and mechanism of *Rhizoma Cimicifugae*. *Chemistry Central Journal* 6:140
- Lieschke, G.J. and Currie, P.D. (2007). Animal models of human disease: zebrafish swim into view. *Nature Review Genetic* 8(5): 353-367.
- Lim, Y.J., Lee, E.H., Kang, T.H., Ha, S.K., Oh, M.S., Kim, S.M., Yoon, T.J., Kang, C., Park, J.H. and Kim, S.Y. (2009). Inhibitory effects of arbutin on melanin biosynthesis of alpha-melanocyte stimulating hormone-induced hyperpigmentation in cultured brownish guinea pig skin tissues. *Archives of Pharmacal Research* 32(3): 367-373.
- Lin, V.C.H., Ding, H.Y., Kuo, S.Y., Chin, L.W., Wu, J.Y. and Chang, T.S. (2011). Evaluation of *in vitro* and *in vivo* depigmenting activity of raspberry ketone from *Rheum officinale*. *International Journal of Molecular Sciences* 12: 4819-4835.

- Liu, K. J. and Shaw, J.F. (1998). Lipase-catalyzed synthesis of kojic acid esters in organic solvents. *Journal of American Oil Chemistry Society Press* 75(11): 1507-1511.
- Ma, H.L., Whitters, M.J., Konz, R.F., Senices, M., Young, D.A., Grusby, M.J., Collins, M. and Dunussi-Joannopoulos, K. (2003). IL-21 activates both innate and adaptive immunity to generate potent antitumor responses that require perforin but are independent of IFN- γ . *Journal of Immunology* 171(2): 608–615.
- Madihah, M.S., Ariff, A.B., Hassan, M.A., Rusul, G. and Karim, M.I.A. (1996). Enhanced kojic acid production by *Aspergillus flavus* link in growth medium containing methanol. *ASEAN Food Journal* 11(4): 158-162.
- Magalhães-Moraes, M.N.d.C., de Oliveira Poletini, M., Ribeiro Ramos, B.C., de Lima, L.H.R.G. and de Lauro Castrucci, A.M. (2014). Effect of light on expression of clock genes in *Xenopus laevis* melanophores. *Photochemistry and Photobiology* 90(3): 696–701.
- Majeed M., Pineda, M.R.T.V., Chan, G., Gabriel, M.T., Dayrit, J., Pelayo, C.A. and Prakash, L. (2010). The safety and efficacy of 0.25% tetrahydrocurcumin (turmeric) cream as depigment agent against 4% hydroquinone cream. *Colour Cosmetics* 3: 44-46.
- Makpol, S., Arifin, N.N.M., Ismail, Z., Chua K.H., Yusof Y.A.M. and Ngah, W.Z.W. (2009). Modulation of melanin synthesis and its gene expression in skin melanocytes by palm tocotrienol rich fraction. *African Journal of Biochemistry Research* 3(12): 385-392.
- Manosroi, A., Wongtrakul, P., Manosroi, J., Midorikawa, U., Hanyu, Y., Yuasa, M., Sugawara, F., Sakai, H. and Abe, M. (2005). The entrapment of kojic oleate in bilayer vesicles. *International Journal of Pharmaceutics* 298(1): 13–25.
- Mansur, J.D.S., Fukuyama, K., Gellin, G.A. and Epstein, W.L. (1978). Effects of 4-tertiary butyl catechol on tissue cultured melanocytes. *Journal of Investigative Dermatology* 70(5): 275–279.
- Marit, J.S. and Weber, L.P. (2012). Persistent effects on adult swim performance and energetics in zebrafish developmentally exposed to 2,3,7,8-tetrachlorodibenzo-p-dioxin. *Aquatic Toxicology* 106–107:131–139.
- Martins, A.B., Graebin, N.G., Lorenzoni, A.S.G., Roberto F. L., Ayub, M.A.Z. and Rodrigues, R. C. (2011). Rapid and high yields of synthesis of butyl acetate catalyzed by Novozym 435: Reaction optimization by response surface methodology. *Process Biochemistry* 46(12): 2311–2316.
- Masoud, M.S., El-Thana, S.A. and El-Enein, A. (1989). Palladium(II)-kojic acid interaction. *Transition Metal Chemistry* 14(2): 155-156.

- Masyithah, Z., Sembiring, S. B., Alfian, Z. and Herawan, T. (2011). The optimization of enzymatic synthesis for lauroyl-n-methyl glucamide surfactants. *Indonesian Journal of Chemistry* 11(3): 223-228.
- Mendesa, A.A., Giordano, R.C., Giordano, R.d.L.C., de Castroc, H.F. (2011). Immobilization and stabilization of microbial lipases by multipoint covalent attachment on aldehyde-resin affinity: Application of the biocatalysts in biodiesel synthesis. *Journal of Molecular Catalysis B: Enzymatic* 68:109–115.
- Menon, V.P. and Sudheer, A.R. (2007). Antioxidant and anti-inflammatory properties of curcumin. *Advance Experimeantal Medicine Biology* 595: 105-25.
- Menter, J.M., Etemadi, A.A., Chapman, W., Hollins, T.D. and Willis, I. (1993). In vivo depigmentation by hydroxybenzene derivatives. *Melanoma Research* 3(6): 443-449.
- Meziti, A., Meziti, H., Boudiaf, K., Mustapha, B. and Bouriche, H. (2012). Polyphenolic profile and antioxidant activities of *Nigella sativa* seed extracts *in vitro* and *in vivo*. *World Academy of Science, Engineering and Technology* 64: 24-32.
- Mikami, M., Sonoki, T., Ito, M., Funasaka, Y., Suzuki, T. and Katagata, Y. (2013). Glycosylation of tyrosinase is a determinant of melanin production in cultured melanoma cells. *Molecular Medicine Reports* 8(3): 818-822.
- Mishra, J., Yousuf, A., Singh, R.D. and Aradhana. (2009). Phytochemical investigation and *in-vitro* antioxidant potential of leaves of *Murraya koenigii*. *International Journal of Integrative Biology* 7(3): 171-174.
- Mitani, H., Koshiishi, I., Sumita, T. and Imanari, T. (2001). Prevention of the photodamage in the hairless mouse dorsal skin by kojic acid as an iron chelator. *European Journal Pharmacological* 411(1-2): 169-174.
- Mitra, D., Luo, X., Morgan, A., Wang, J., Hoang, M.P., Lo, J., Guerrero, C.R., Lennerz, J.K., Mihm, M.C., Wargo, J.A., Robinson, K.C., Devi, S.P., Vanover, J.C., D'Orazio, J.A., McMahon, M., Bosenberg, M.W., Haigis, K.M., Haber, D.A., Wang, Y. and Fisher, D.E. (2012). An ultraviolet-radiation-independent pathway to melanoma carcinogenesis in the red hair/fair skin background. *Nature* 491(7424): 449-453.
- Mitsumori K., Onodera H., Takahashi M., Funakoshi T., Tamura T., Yasuhara K., Takegawa K. and Takahashi M. (1999). Promoting effects of kojic acid due to serum TSH elevation resulting from reduced serum thyroid hormone levels on development of thyroid proliferative lesions in rats initiated with N-bis(2-hydroxypropyl)nitrosamine. *Carcinogenesis* 20(1): 173-176.

- Miyashita, Y., Moriya, T., Yokosawa, N., Hatta, S., Arai, J.-I., Kusunoki, S., Toratani, S., Yokosawa, H., Fujii, N. and Asami, K. (1996), Light-sensitive response in melanophores of *Xenopus laevis*: II. Rho is involved in light-induced melanin aggregation. *Journal of Experimental Zoology* 276: 125–131.
- Mohamad, R., Mohamed, M. S., Suhaili, N., Salleh, M. M. and Ariff, A. B (2010). Kojic acid: Applications and development of fermentation process for production. *Biotechnology and Molecular Biology Reviews* 5(2): 24-37.
- Molenda, J. J., Basinger, M. A., Hanusa, T. P. and Jones M. M. (1994). Synthesis and iron(III) binding properties of 3-hydroxypyrid-4-ones derived from kojic acid. *Journal of Inorganic Biochemistry* 55(2): 131-146.
- Momtaz, S., Mapunya, B.M., Houghton, P.J., Edgerly, C., Hussein, A., Naidoo, S., Lall, N., (2008) Tyrosinase inhibition by extracts and constituents of *Sideroxylon inerme* L. stem bark, used in South Africa for skin lightening, *Journal of Ethnopharmacology* 119(3): 507-512.
- Moore, T. (1943). Dental depigmentation in rat. *Biochemical Journal* 37(1):112-115.
- Morishita, F. (1987). Responses of the melanophores of the medaka, *Oryzias latipes*, to adrenergic drugs: evidence for involvement of alpha 2 adrenergic receptors mediating melanin aggregation. *Comparative Biochemistry and Physiology C* 88(1):69-74.
- Moto, M., Mori, T., Okamura, M., Kashida, Y. and Mitsumori, K. (2006). Absence of liver tumor-initiating activity of kojic acid in mice. *Archives of Toxicology*, 80(5): 299-304.
- Mutschler, J., Rausis, T., Bourgeois, J. M., Bastian, C., Zufferey, D., Isabelle Vanessa Mohrenz, I. V. and Fischer, F. (2009). Ionic liquid-coated immobilized lipase for the synthesis of methylglucose fatty acid esters. *Green Chemistry* 11: 1793–1800.
- Nagai, S. and Izumi, T. (1981). U.S Patent No. 4,278,656. Onojo, Japan.: U.S Patent and Trademark Office.
- Nagata, H., Takekoshi, S., Takeyama, R., Homma, T. and Yoshiyuki Osamura, R. (2004). Quercetin enhances melanogenesis by increasing the activity and synthesis of tyrosinase in human melanoma cells and in normal human melanocytes. *Pigment Cell Research* 17(1):66-73.
- Nakagawa, M., Kawai, K. and Kawai, K. (1995). Contact allergy to kojic acid in skin care products. *Contact Dermatitis* 32(1): 9-13.

- Nakajima, H., Fukazawa, K., Wakabayashi, Y., Wakamatsu, K. and Imokawa, G. (2012). Withania somnifera extract attenuates stem cell factor-stimulated pigmentation in human epidermal equivalents through interruption of ERK phosphorylation within melanocytes. *Journal of Natural Medicine* 66(3): 435-46.
- Nakajima, H., Fukazawa, K., Wakabayashi, Y., Wakamatsu, K., Senda, K. and Imokawa, G. (2012). Abrogating effect of a xanthophyll carotenoid astaxanthin on the stem cell factor-induced stimulation of human epidermal pigmentation. *Archive of Dermatological Research* 304(10): 803-16.
- Nakajima, N., Ishihara, K. and Hamada, H. (2001). Functional glucosylation of kojic acid and daidzein with the eucalyptus membrane-associated UDP – glucosyltransferase reaction system. *Journal of Bioscience and Bioengineering* 92(5): 469-471.
- Nawarak, J., Rosa, H.L., Kao, S. H., Liao, H.H., Sinchaikul, S., Chen, S.T. and Cheng, S.L. (2008). Proteomics analysis of kojic acid treated A375 human malignant melanoma cells. *Journal of Proteome Research* 7(9): 3737–3746.
- Ndhlala, A.R., Moyo, M. and Staden, J.V. (2010). Natural Antioxidants: Fascinating or Mythical Biomolecules?. *Molecules* 15: 6905-6930.
- Nishimura, T., Kometani, T., Takii, H., Terada, Y. and Okada, S. (1994). Acceptor specificity in the glucosylation reaction of *Bacillus subtilis* X-23 α -Amylase towards various phenolic compounds and the structure of kojic acid glucoside. *Journal of fermentation and Bioengineering* 78(1): 37-41.
- Nitoda, T., Fan, M.D. and Kubo, I. (2008). Effects of cuminaldehyde on melanoma cells. *Phytotherapy Research* 22(6): 809–813.
- Niwa, Y. and Akamatsu, H. (1991). Kojic acid scavenges free radicals while potentiating leukocyte functions including free radical generation. *Inflammation* 15(4): 303–315.
- Noh, J.M., Kwak, S.Y., Kim, D.H. and Lee, Y.S. (2007). Kojic acid-tripeptide amide as a new tyrosinase inhibitor. *Biopolymers* 88(2): 300-307.
- Noh, J.M., Kwak, S.Y., Seo, H.S., Seo, J.H., Kim, B.G. and Seo, J.H. (2009). Kojic acid-amino acid conjugates as tyrosinase inhibitors. *Bioorganic Medicine Chemistry Letter* 19(19): 5586-5589.
- Nohynek, G.J., Kirkland, D., Marzin, D., Toutain, H., Leclerc-Ribaud, C., Jinnai, H. (2004). An assessment of the genotoxicity and human health risk of topical use of kojic acid [5-hydroxy-2-(hydroxymethyl)-4H-pyran-4-one]. *Food Chemical Toxicology* 42(1): 93-105.

- Noureddini, H., Gao, X., Joshi, S. and Wagner, P.R. (2002). Immobilization of *Pseudomonas cepacia* lipase by sol-gel entrapment and its application in the hydrolysis of soybean oil. *Journal of the American Oil Chemists' Society* 79(1): 33-40.
- Noureddini, H., Gao, X. and Philkana R.S. (2005). Immobilized *Pseudomonas cepacia* lipase for biodiesel fuel production from soybean oil. *Bioresource Technology* 96(7): 769–777.
- Norhaizan, M.E., Ng, S.K., Norashareena, M.S. and Abdah, M.A. (2011). Antioxidant and cytotoxicity effect of rice bran phytic acid as an anticancer agent on ovarian, breast and liver cancer cell lines. *Malaysian Journal of Nutrition* 17(3): 367-375.
- Novellino, L., Napolitano, A. and Protà, G. (1999). 5,6-Dihydroxyindoles in the fenton reaction: a model study of the role of melanin precursors in oxidative stress and hyperpigmentary processes. *Chemical Research in Toxicology* 12(10): 985–992.
- Nurchi, V.M., Crisponi, G., Lachowicz, J.I., Murgia, S., Pivetta, T., Remelli, M., Rescigno, A., Niclós-Gutiérrez, J., González-Pérez, J.M., Domínguez-Martín, A., Castiñeiras, A. and Szewczuk, Z. (2010). Iron(III) and aluminum(III) complexes with hydroxypyronone ligands aimed to design kojic acid derivatives with new perspectives. *Journal of Inorganic Biochemistry* 104(5): 560-569.
- Nurchi, V.M., Lachowicz, J.I., Crisponi, G., Murgia, S., Arca, M., Pintus, M., Gans, P., Niclos-Gutierrez, J., Domínguez-Martín, A., Castiñeiras, A., Remelli, M., Szewczuk, Z. and Lisg, T. (2011). Kojic acid derivatives as powerful chelators for iron(III) and aluminium(III). *Dalton Transactions* 40(22): 5984-5998.
- Odoko, M., Yamamoto, K. and Okabe, N. (2002). Bis(5-hydroxy-2-hydroxymethyl-4-pyrone-kappa 2O4,O5)bis(2-hydroxymethyl-5-oxido-4-pyrone-kappa 2O4,O5)calcium(II) tetrahydrate. *Acta Crystallographica Section C, Crystal Structure Communications* 58(9): 469-470.
- Ohara, M., Kobayashi, M., Fujiwara, H., Kitajima, S., Mitsuoka, C. and Watanabe, H. (2004). Blue light inhibits melanin synthesis in B16 melanoma 4A5 cells and skin pigmentation induced by ultraviolet B in guinea-pigs. *Photodermatology, Photoimmunology and Photomedicine* 20(2): 86-92.
- Ohguchi, K., Banno, Y., Akao, Y. and Nozawa, Y. (2004). Involvement of phospholipase D1 in melanogenesis of mouse B16 melanoma cells. *Journal of Biological Chemistry* 279(5): 3408–3412.
- Oliveira, A., Sereno, R. and Azeredo, J. (2010). *In vivo* efficiency evaluation of a phage cocktail in controlling severe colibacillosis in confined conditions and experimental poultry houses. *Veterinary Microbiology* 146(3-4): 303–308.

- Olszewska-Słonina, D.M., Styczyński, J., Czajkowski, R., Drewa, T.A. and Musiałkiewicz, D. (2007). Cell cycle, melanin contents and apoptosis processes in B16 and Cloudman S91 mouse melanoma cells after exposure to cytostatic drugs. *Acta Poloniae Pharmaceutica* 64(5): 469-478.
- Onitsuka, S., Yong, Z.J., Shaikh, A.C., Furuno, H. and Inanaga, J. (2012). Silica gel-mediated organic reactions under organic solvent-free conditions. *Molecules* 17(10): 11469-11483.
- Ortonne, J.P. and Bissett, D. L (2008). Latest insights into skin hyperpigmentation. *Journal of Investigative Dermatology Symposium Proceedings*, 13(1): 10-14.
- Ou, B.X., Huang, D.J. Hampsch-Woodill, M. Flanagan, J.A. and Deemer, E.K. (2002). Analysis of antioxidant activities of common vegetables employing oxygen radical absorbance capacity (ORAC) and ferric reducing antioxidant power (FRAP) assays: A comparative study. *Journal of Agricultural Food Chemistry* 50: 3122-3128.
- Ozturk, G., Erol, D.D., Uzbay, T. and Aytemir, M.D. (2001). Synthesis of 4(1H)-pyridinone derivatives and investigation of analgesic and antiinflammatory activities. *Il Farmaco* 56(4): 251-256.
- ÖzYüreK, M., Kola, U., Güçlü, K., Aras, C., Altun, M., Celi, S.E., Berker, K.I., Bektaşoğlu, B. and Apa, R. (2009). Antioxidant capacities of some food plants wildly grown in Ayvalik of Turkey. *Food Science Technology Research* 15(1): 59- 64.
- Pacher, P., Nivorozhkin, A. and Szabo, C. (2006). Therapeutic effects of xanthine oxidase inhibitors: Renaissance half a century after the discovery of allopurinol. *Pharmacological reviews* 58(1): 87–114.
- Padayatty, S.J., Katz, A., Wang, Y., Eck, P., Kwon, O., Lee, J.H., Chen, S., Corpe, C., Dutta, A., Dutta, S.K. and Levine, M. (2003). Vitamin C as an antioxidant: evaluation of its role in disease prevention. *Journal of American College Nutrition* 22(1):18-35.
- Pan, T., Zhu, J., Hwu, W.J. and Jankovic, J. (2012a). The role of alpha-synuclein in melanin synthesis in melanoma and dopaminergic neuronal cells. *PLoS One* 7(9): e45183.
- Pan, X., Chen, B., Wang, J., Zhang, X., Zhul, B. and Tan, T. (2012b). Enzymatic synthesizing of phytosterol oleic esters. *Applied Biochemistry and Biotechnology* 168(1): 68–77.
- Panich, U., Kongtaphan, K., Onkoksoong T., Jaemsak, K., Phadungrakwittaya, R., Thaworn, A., Akarasereenont, P. and Wongkajornsilp, A. (2010). Modulation of antioxidant defense by *Alpinia galanga* and *Curcuma*

aromatica extracts correlates with their inhibition of UVA-induced melanogenesis. *Cell Biology and Toxicology* 26(2): 103–116.

- Panich, U., Pluemsamran, T., Tangsupa-a-nan, V., Wattanarangsang, J., Phadungrakwittaya, R., Akarasereenont, P. and Laohapand, T. (2013). Protective effect of AVS073, a polyherbal formula, against UVA-induced melanogenesis through a redox mechanism involving glutathione-related antioxidant defense. *BMC Complementary and Alternative Medicine* 13: 159.
- Panzica-Kelly, J.M., Zhang, C.X., Danberry, T.L., Flood, A., DeLan, J.W., Brannen, K.C. and Augustine-Rauch, K.A. (2010). Morphological score assignment guidelines for the dechorionated zebrafish teratogenicity assay. *Birth Defects Research. Part B: Developmental and Reproductive Toxicology* 89(5): 382-395.
- Parrish, F.W., Wiley, B.J., Simmons, E.G. and Long, J.L (1966). Production of aflatoxins and kojic acid by species of *Aspergillus* and *Penicillium*. *Applied Microbiology* 14(1): 139.
- Park, H., Song, K.H., Jung, P.M., Kim, J.E., Ro, H., Kim, M.Y and Ma, J.Y (2013). Inhibitory effect of arctigenin from *Fructus arctii* extract on melanin synthesis via repression of tyrosinase expression. *Evidence-Based Complementary and Alternative Medicine* 2013: 965312.
- Park, J.S., Kim, D.H., Lee, J.K., Lee, J.Y., Kim, D.H., Kim, H.K., Lee, H.J. and Kim, H.C. (2010). Natural ortho-dihydroxyisoflavone derivatives from aged Korean fermented soybean paste as potent tyrosinase and melanin formation inhibitors. *Bioorganic and Medicinal Chemistry Letters* 20(3): 1162-1164.
- Park, S.H., Kim, D.S., Park, S.H., Shin, J.W., Youn, S.W. and Park, K.C. (2008). Inhibitory effect of p-coumaric acid by *Rhodiola sachalinensis* on melanin synthesis in B16F10 cells. *Pharmazie* 63(4): 290-295.
- Park, Y.D., Jung, J.Y., Kim, D.W., Kim, W.S., Hahn, M.J. and Yang, J.M. (2003). Kinetic inactivation study of mushroom tyrosinase: Intermediate detection by denaturants. *Journal of Protein Chemistry* 22(5): 463-471.
- Paus, R. and Cotsarelis, G. (1999). The biology of hair follicles. *The New England Journal of Medicine* 341(7):491-497.
- Peng, H.Y., Lin, C.C., Wang, H.Y., Shih, Y., Chou, S.T. (2014). The melanogenesis alteration effects of *Achillea millefolium* L. essential oil and linalyl acetate: Involvement of oxidative stress and the JNK and ERK signaling pathways in melanoma cells. *PLoS ONE* 9(4): e95186.
- Pershina, V., Bastug, T. and Kratz, J.V. (2001). Quantum chemical predictions of properties and experimental behaviour of elements 106, 107, and 108, *Radiochimistry Acta* 89: 729-732.

- Pickering, D.S. and Niles, L.P. (1992). Expression of nanomolar-affinity binding sites for melatonin in Syrian hamster RPMI 1846 melanoma cells. *Cellular Signalling* 4(2): 201-207.
- Prazeres, D. M. and Cabral, J. M. (1994). Enzymatic membrane bioreactors and their applications. *Enzyme and Microbiology Technology* 16(9): 738-750.
- Prieto, M.A., Rodríguez-Amado, I., Vázquez J.A. and Murado M.A. (2012). β -Carotene assay revisited. Application to characterize and quantify antioxidant and prooxidant activities in a microplate. *Journal of Agricultural Food Chemistry* 60(36): 8983-8993.
- Pryor, W.A. and Godber, S.S. (1991). Noninvasive measures of oxidative stress status in humans. *Free Radical Biology Medicine* 10(3-4):177-84.
- Potten, C.S. (1970). Radiation depigmentation of mouse hair: Effect of the hair growth cycle on the sensitivity. *The Journal of Investigate Dermatology* 55(6): 410-418.
- Qi, Z., Miller, G.W. and Voit, E.O. (2008). Computational systems analysis of dopamine metabolism. *PLoS ONE* 3(6): e2444.
- Qiu, L., Zhang, M., Sturm, R.A., Gardiner, B., Tonks, I., Kay, G. and Parsons, P.G. (2000). Inhibition of melanin synthesis by cystamine in human melanoma cells. *Journal of Investigative Dermatology* 114(1): 21–27.
- Quigley, I.K. and Parichy D.M (2002). Pigment pattern formation in zebrafish: A model for developmental genetics and the evolution of form. *Microscopy Research and Technique* 58(6):442–455.
- Radzi, S.M., Basri, M., Salleh, A.B., Ariff, A., Mohammad, R., Abdul-Rahman, M. B. and Abdul-Rahman, R.N.Z. (2005). Large scale production of liquid wax ester by immobilized lipase. *Journal of Oleo Science* 54(4): 203-209.
- Radzi, S. M., Basri, M., Salleh, A. B., Ariff, A., Mohammad, R., Abdul-Rahman, M. B. and Abdul-Rahman, R. N. Z. R. (2006). Optimisation study of large-scale enzymatic synthesis of oleyl oleate, a liquid wax ester, by response surface methodology. *Journal of Chemical Technology and Biotechnology* 81(3): 374–380.
- Radzi, S.M., Mustafa, W.A.F., Othman, S.S. and Noor, H.M. (2011). Green synthesis of butyl acetate, a pineapple flavour via lipase-catalyzed reaction. *World Academy of Science, Engineering and Technology* 59: 677-680.
- Rael, L.T., Thomas, G.W., Craun, M.L., Curtis, C.G., Bar-Or, R. and Bar-Or, D. (2004). Lipid peroxidation and the thiobarbituric acid assay: Standardization of the assay when using saturated and unsaturated fatty acids. *Journal of Biochemistry and Molecular Biology* 37(6): 749-752.

- Rahman, N.K., Kamaruddin, A.H. and Uzir, M.H. (2011). Enzymatic synthesis of farnesyl laurate in organic solvent: initial water activity, kinetics mechanism, optimization of continuous operation using packed bed reactor and mass transfer studies. *Bioprocess and Biosystems Engineering* 34(6): 687-699.
- Rahman, N.Z.R.A, Salleh, A.B. and Basri, M. (2006). Lipase specificity In A.B. Salleh (Ed.), *New Lipases and Proteases* (pp. 1-159). Malayisa: Nova biomedical Nova Publishers.
- Rajendran, A., Palanisamy, A. and Thangavelu, V. (2009). Lipase catalyzed ester synthesis for food processing industries. *Brazilian Archives of Biology and Technology* 52(1): 207-219.
- Raku, T. and Tokiwa, Y., (2003). Regioselective synthesis of kojic acid esters by *Bacillus subtilis* protease. *Biotechnology Letters* 25: 969-974.
- Ramani, R., Sudini, S., Boddupalli, B.M. and Anisetti, R.N (2012). Antioxidant, free radical scavenging and *in vitro* cytotoxic studies of ethanolic extract of *Leucas indica* var *lavandulifolia* and *Leucas indica* var *nagalapuramiana*. *Asian Pacific Journal of Tropical Biomedicine* 2(3): S1637-S1642.
- Ray, S. (2012). Studies on fluidized bed column reactor for the substrate conversion using immobilised extracellular alkaline lipase of isolated strain of *Serratia* sp (C4). *Journal of Microbiology and Biotechnology Research* 2(4): 538-544.
- Reddy, B.V., Reddy, M.R., Madan, C.H., Kumar, K.P. and Rao, M.S. (2010). Indium(III) chloride catalyzed three-component coupling reaction: a novel synthesis of 2-substituted aryl(indolyl)kojic acid derivatives as potent antifungal and antibacterial agents. *Bioorganic and Medicinal Chemistry Letters* 20(24): 7507-7511.
- Reddy, B.V.S., Reddy, S.M., Swain, M., Dudem, S., Kalivendib, S.V. and C. Suresh Reddy, C.S. (2014). Enantioselective 1,4-addition of kojic acid derivatives to β -nitroolefins catalyzed by a cinchonine derived sugar thiourea. *RSC Advances* 4(18): 9107-9111.
- Rechner, A.R., Kuhnle, G., Bremner, P., Hubbard, G.P., Moore, K.P. and Rice-Evans, C.A. (2002). The metabolic fate of dietary polyphenols in humans. *Free Radical Biological Medicine* 33(2): 220-235.
- Reimers, M.J., Flockton, A.R. and Tanguay, R.L. (2004). Ethanol- and acetaldehyde-mediated developmental toxicity in zebrafish. *Neurotoxicology and Teratology* 26(6): 769-781.
- Rho, H.S., Ahn, S.M., Yoo, D.S., Kim, M.K., Cho, D.H. and Cho, J.Y. (2010a). Kojyl thioether derivatives having both tyrosinase inhibitory and anti-

inflammatory properties. *Bioorganic and Medicinal Chemistry Letters* 20(22): 6569–6571.

Rho, H.S., Baek, H.S., You, J.W., Kim, S., Lee, J.Y., Kim, D.H. and Chang, I.S. (2007). New 5-Hydroxy-2-(hydroxymethyl)-4H-pyran-4-one derivative has both tyrosinase inhibitory and antioxidant properties, *Bulletin Korean Chemistry Society* 28(3): 471-473.

Rho, H.S., Goh, M., Lee, J.Y., Ahn, S. M., Yeon, J.H., Yoo, D.S., Kim, D.H., Kim, H.G., and Cho, J.Y. (2011). Ester derivatives of kojic acid and polyphenols containing adamantane moiety with tyrosinase inhibitory and anti-inflammatory properties. *Bulletin Korean Chemistry Society* 32(4): 1411-1414.

Rho, H. S., Yoo, D. S., Ahn, S. M., Kim, M. K., Cho, D. H., and Cho, J. Y. (2010b). Inhibitory activities of kojyl thioether derivatives against nitric oxide production induced by lipopolysaccharide. *Bulletin Korean Chemistry Society* 31(11): 3463-3466.

Rho, H.S., Baek, H.S., Ann, S.M., Kim, D.H. and Chang, I.S. (2008). Synthesis of New Anti-melanogenic compounds containing two molecules of kojic acid. *Bulletin Korean Chemistry Society* 29(8): 1569-1571.

Ricchi, M., Odoardi, M.R., Carulli, L., Anzivino, C., Ballestri, S., Pinetti, A., Fantoni, L.I., Marra, F., Bertolotti, M., Banni, S., Lonardo, A., Carulli, N. and Loria, P. (2009). Differential effect of oleic and palmitic acid on lipid accumulation and apoptosis in cultured hepatocytes. *Journal of gastroenterology and hepatology* 24(5): 830–840.

Rodrigues, A. P. D., Carvalho, A. S. C., Santos, A. S., Alves, C. N., Nascimento J. L. M. d. and Silva, E. O. (2011). Kojic acid, a secondary metabolite from *Aspergillus* sp., acts as an inducer of macrophage activation. *Cell Biology International* 35(4): 335–343.

Rodrigues, R.C. and Ayub, M.A.Z. (2011). Effects of the combined use of *Thermomyces lanuginosus* and *Rhizomucor miehei* lipases for the transesterification and hydrolysis of soybean oil. *Process Biochemistry* 46: 682-688.

Rosfarizan, M. and Ariff, A.B. (2007). Biotransformation of various carbon sources to kojic acid by cell-bound enzyme system of *A. flavus* Link 44-1. *Biochemical Engineering Journal* 35(2): 203-209.

Rosfarizan, M., Ariff, A.B., Hassan, M.A., Karim, M.I.A., Shimizu, H., Shioya, S. (2002). Important of carbon source feeding and pH control strategies for maximum kojic acid production from sago starch by *A. flavus*. *Journal of Bioscience and Bioengineering* 94(2): 99-105.

Rosfarizan, M., Madihah, S. and Ariff, A.B. (1998). Isolation of a kojic acid producing fungus capable of using starches as a carbon source. *Letter of Applied Microbiology* 26(1): 27-30.

- Rusciano, D., Lorenzoni, P. and Burger, M.M. (1999). Regulation of c-met expression in B16 murine melanoma cells by melanocyte stimulating hormone. *Journal of Cell Science* 112(5): 623–630.
- Saatchi, K., Thompson, K.H., Patrick, B.O., Pink, M., Yuen, V.G., McNeill, J.H. and Orvig, C. (2005). Coordination chemistry and insulin-enhancing behavior of vanadium complexes with maltol C₆H₆O₃ structural isomers. *Inorganic Chemistry* 44(8): 2689-2697.
- Sabeder, S., Habulin, M. and Knez, Z. (2006). The lipase-catalyzed synthesis of fatty acid fructose esters in organic media and in supercritical carbon dioxide. *Chemical Industry and Chemical Engineering Quarterly* 12(3): 147-151.
- Sadeghi, B. and Shahedi, M.R. (2013). A clean, simple, and efficient synthesis of 2-substituted aryl (indolyl) kojic acid derivatives by kaolin/Ag nanocomposite as a reusable catalyst: A green protocol. *Journal of Chemistry* 2013: 418969.
- Saha, A.K., Acharya, S. and Roy, A. (2012). Antioxidant level of wild edible mushroom: *Pleurotus djamor (Fr.) Boedijn*. *Journal of Agricultural Technology* 8(4): 1343-1351.
- Saghaie, L., Pourfarzam, M., Fassihi, A. and Sartippour, B. (2013). Synthesis and tyrosinase inhibitory properties of some novel derivatives of kojic acid. *Research in Pharmaceutical Sciences* 8(4): 233–242.
- Saha, M.R., Hasan, S.M.R., Akter, R., Hossain, M.M., Alam, M.S., Alam, M.A. and Mazumder, M.E.H. (2008). *In vitro* free radical scavenging activity of methanol extract of the leaves of *Mimusops elengi linn*. *Bangla Journal of Veterinary Medicine* 6(2): 197-202.
- Salis, A., Solinas, V. and Monduzzi, M. (2003). Wax esters synthesis from heavy fraction of sheep milk fat and cetyl alcohol by immobilised lipases, *Journal of Molecular Catalysis B: Enzymatic* 21(4-6): 167–174.
- Sander, K., Kottke, T., Weizel, L. and Stark, H. (2010). Kojic acid derivatives as histamine H(3) receptor ligands. *Chemical and pharmaceutical bulletin* 58(10): 1353-1361.
- Saponjić, S., Knežević-Jugović, Z. D., Bezbradica, D. I., Zuza, M. G., Saied, O. A., Bosković-Vragolović, N. and Mijin, D. Z. (2010). Use of *Candida rugosa* lipase immobilized on sepabeads for the amyl caprylate synthesis: Batch and fluidized bed reactor study. *Electronic Journal of Biotechnology* 13(6): 1-15.
- Sarı, A., Karaipekli, A. and Alka, C. (2009). Preparation, characterization and thermal properties of lauric acid/expanded perlite as novel form-stable

composite phase change material. *Chemical Engineering Journal* 155(3): 899–904.

- Saritha, V., Anilakumar, K.R. and Khanum, F. (2010). Antioxidant and antibacterial activity of *Aloe vera* gel extracts. *International Journal of Pharmaceutical and Biological Archives* 1(4): 376-384.
- Sasaki, Y., Kobori, M., Mori, H. and Shinohara, K. (1993). Suppression of melanin production in a mouse B16 melanoma 4A5 cell line by a spinach extract. *Animal Cell Technology: Basic and Applied Aspects* 5: 183-188.
- Sato, E., Tsukimoto, M., Shimura, N., Awaya, A. and Kojima, S. (2011b). Mechanism of pigmentation by minocycline in murine B16 melanoma cells. *Yakugaku Zasshi* 131(5): 731-738.
- Sato, K. and Toriyama, M. (2009). Depigmenting effect of catechins. *Molecules* 14(11): 4425-4432
- Sato, K., Takahashi, H. and Toriyama, M. (2011a). Depigmenting mechanism of NSAIDs on B16F1 melanoma cells. *Archives of Dermatological Research* 303(3): 171-180.
- Sato, K., Takahashi, H., Ibra, R. and Toriyama, M. (2008a). Down-regulation of tyrosinase expression by acetylsalicylic acid in murine B16 melanoma. *Biological and Pharmaceutical Bulletin* 31(1): 33-37.
- Sato, T., Matsumoto, K., Okumura, T., Yokoi, W., Naito, E., Yoshida, Y., Nomoto, K., Ito, M. and Sawada, H. (2008b). Isolation of lactate-utilizing butyrate-producing bacteria from human feces and *in vivo* administration of *Anaerostipes caccae* strain L2 and galacto-oligosaccharides in a rat model. *FEMS Microbiology Ecology* 66(3): 528-536.
- Schmidt, E., Jirovetz, L., Buchbauer, G., Eller, G.A., Stoilova, I., Krastanov, A., Stoyanova, A. and Geissler, M. (2006). Composition and antioxidant activities of the essential oil of cinnamon (*Cinnamomum zeylanicum* Blume) leaves from Sri Lanka. *Journal of essential oil-bearing plants* 9(2): 170-182.
- Schwartzkopf, K.S., Stookey, J.M., Hull, P.R. and Clark, E.G. (1994). Screening of depigmenting compounds for the development of an alternate method of branding beef cattle. *Journal of Animal Science* 72(6): 1393-1398.
- Scior, T., Mack, H.G., García, J.A.G. and Koch, W. (2008). Antidiabetic Bis-Maltolato-OxoVanadium(IV): Conversion of inactive trans- to bioactive cis-BMOV for possible binding to target PTP-1B. *Drug Design, Development and Therapy* 2: 221–231.
- Scott, G., Leopardi, S., Printup, S. and Madden, B.C. (2002). Filopodia are conduits for melanosome transfer to keratinocytes. *Journal of Cell Science* 115(7):1441-51.

- Sengupta, A., Pal, M., SilRoy, S. and Ghosh, M. (2010). Comparative study of sterol ester synthesis using *Thermomyces lanuginosus* lipase in stirred tank and packed-bed bioreactors. *Journal of American Oil Chemistry Society* 87(9): 1019-1025.
- Sern, C.H., May, C.Y., Zakaria, Z. and Daik, R. (2008). Synthesis of palmitic acid-based esters and their effect on the pour point of palm oil methyl esters. *Journal of Palm Oil Research* 20: 542-547.
- Sharma C.K. and Kanwar S.S. (2012). Synthesis of methyl cinnamate using immobilized lipase from *B. Licheniformis* MTCC-10498. *Research Journal of Recent Science* 1(3): 68-71.
- Sheng, Z.W., Ma, W.H., Gao, J.H., Bi, Y., Zhang, W.N., Dou, H.T. and Jin, Z.Q. (2011). Antioxidant properties of banana flower of two cultivars in China using 2,2-diphenyl-1-picrylhydrazyl (DPPH,) reducing power, 2,2'-azinobis-(3-ethylbenzthiazoline-6-sulphonate (ABTS) and inhibition of lipid peroxidation assays. *African Journal of Biotechnology* 10(21): 4470-4477.
- Shibata, S., Okano, S., Yonemitsu, Y., Onimaru, M., Sata, S., Nagata-Takeshita, H., Inoue, M., Zhu, T., Hasegawa, M., Moroi, Y., Furue, M. and Sueishi, K. (2006). Induction of efficient antitumor immunity using dendritic cells activated by recombinant Sendai virus and its modulation by exogenous IFN- β gene. *Journal of Immunology* 177(6): 3564–3576.
- Shimbo, K., Ikuta, K., Nishijima, H., Sugimoto, K., Etoh, H., Sakata, K. and Ina, K., (2009). Synthesis of phosphatidyl kojic acid by phospholipase D and its properties. *Journal of Japan Oil Chemists Society*, 44(8): 579-585.
- Shin, S.H. and Lee, Y.M (2013). Glyceollins, a novel class of soybean phytoalexins, inhibit SCF-induced melanogenesis through attenuation of SCF/c-kit downstream signalling pathways. *Experimental and Molecular Medicine* 45: e17.
- Shin, Y.H., Seo, Y.K., Yoon, H.H., Song, K.Y. and Park, J.K. (2012). Effect of keratinocytes on regulation of melanogenesis in culture of melanocytes. *Biotechnology and Bioprocess Engineering* 17(1): 203-210.
- Shin, Y.J., Han, C.S., Lee, C.S., Kim, H.S., Ko, S.H., Hwang, S.K., Shin, J.W., Ye, S.K. and Chung, M.H., (2010). Zeolite 4A, a Synthetic Silicate, Suppresses Melanogenesis through the Degradation of Microphthalmia-Associated Transcription Factor by Extracellular Signal-Regulated Kinase Activation in B16F10 Melanoma Cells. *Biological Pharmaceutical Bulletin* 33(1): 72-76.
- Shioda, T., Fenner, M.H. and Isselbacher, K.J. (1996). msg1, a novel melanocyte-specific gene, encodes a nuclear protein and is associated with pigmentation. *Proceedings of the National Academy of Sciences of the United States of America* 93(22): 12298–12303.

- Shukor M.Y. and Syed, M.A. (2010). Microbiological reduction of hexavalent molybdenum to molybdenum blue, current research, technology and education topics. In A.Mendez-Vilas (Ed.), *Applied Microbiology and Microbial Biotechnology* (pp. 1304-1310). Spain: Formatex
- Sibi, M.P. and Zimmerman, J. (2006). Pyrones to pyrans: enantioselective radical additions to acyloxy pyrones. *Journal of the American Chemical Society* 128(41): 13346-13347.
- Šimo, F. and Šima, J. (2011). Determination of photoredox properties of individual kinetically labile complexes in equilibrium systems. *Chemical Papers* 65(5): 730-734.
- Šimo, F., Moncol, J., Šípoš, R., Padělková, Z. and Šima, J. (2011). The molecular and crystal structures of 2-iodokojic acid: Experimental and theoretical determination. *Journal of Chemical Crystallography* 41(8): 1093–1098.
- Singh, M.S. and Chowdhury, S. (2012). Recent developments in solvent-free multicomponent reactions: a perfect synergy for eco-compatible organic synthesis. *Royal Society of Chemistry Advances* 2:4547-4592.
- Singh, S., Singh, J., Gulia, S. and Kakkar, R. (2013). Metal Ion Selectivity of Kojate Complexes: A Theoretical Study. *Journal of Theoretical Chemistry* 2013: 342783.
- Sipes, N.S., Padilla, S. and Knudsen, T.B (2011). Zebrafish-As an Integrative Model for Twenty-first Century Toxicity Testing. *Birth Defects Research. Part C, Embryo today: reviews* 93(3): 256-267.
- Šípoš, R., Šima, J., Izakovič, M. and Tarapčík, P. (2011). Solution properties of azidokojatoiron(III) complexes. *Journal of Solution Chemistry* 40(7): 1200-1208.
- Skoronski, E., Padoin, N., Soares, C. and Furigo Jr., A. (2014). Stability of immobilized *Rhizomucor miehei* lipase for the synthesis of pentyl octanoate in a continuous packed bed bioreactor. *Brazilian Journal of Chemical Engineering* 31(3): 633-641.
- Slominski, A., Tobin, D.J., Shibahara S. and Wortsman, J. (2004). Melanin pigmentation in mammalian skin and its hormonal regulation. *Physiological Review* 84(4): 1155-1228.
- Smith, C.N. and Lindsay, C.D. (2001). Kojic acid reduces the cytotoxic effects of sulfur mustard on cultures containing human melanoma cells *in vitro*. *Journal of Applied Toxicology* 21(6): 435-440.
- Sochor, J., Ryvolova, M., Krystofova, O., Salas, P., Hubalek, J., Adam, V., Trnkova, L., Havel, L., Beklova, M., Zehnalek, J., Provaznik, I. and Kizek, R. (2010). Fully automated spectrometric protocols for determination of

antioxidant activity: Advantages and disadvantages. *Molecules* 15(12): 8618-8640.

Solca, F.F., Chluba-de Tapia, J., Iwata, K. and Eberle, A.N. (1993). B16-G4F mouse melanoma cells: an MSH receptor-deficient cell clone. *FEBS Letters* 322(2):177-80.

Song, T.Y., Chen, C.H., Yang, N.C., Fu, C.S., Chang, Y.T. and Chen, C.L., (2009). The correlation of *in vitro* mushroom tyrosinase activity with cellular tyrosinase activity and melanin formation in melanoma cells A2058. *Journal of Food and Drug Analysis* 17(3): 156-162.

Springer, M., Engelhart, K., and Biesalski, H.K., (2003). Effects of 3-isobutyl-1-methylxanthine and kojic acid on cocultures and skin equivalents composed of HaCaT and human melanocytes. *Archives of Dermatology Research* 295(2): 88-91.

Stamatis, H., Xenakis, A., Bornscheuer, U., Scheper, T., Menge, U. and Kolisis, F. N. (1993). *Pseudomonas cepacia* lipase: Esterification reactions in AOT microemulsion systems. *Biotechnology letters* 15(7): 703-708.

Stehr, C.M., Linbo, T.L., Incardona, J.P. and Scholz, N.L. (2006). The Developmental neurotoxicity of fipronil: Notochord degeneration and locomotor defects in zebrafish embryos and larvae. *Toxicological Sciences* 92(1): 270-278.

Stenson, A.C. and Cioffi, E.A. (2007). Speciation of M+3-hydroxypyrrone chelation complexes by electrospray ionization ion trap and fourier transform ion cyclotron resonance mass spectrometry. *Rapid Communication in Mass Spectrometry* 21(16): 2594-2600.

Stojković, D., Petrović, J., Soković, M., Glamočlija, J., Jelena K.M., and Petrović, S. (2013). *In situ* antioxidant and antimicrobial activities of naturally occurring caffeic acid, p-coumaric acid and rutin, using food systems. *Journal of the Science of Food and Agriculture* 93(13): 3205–3208.

Streffer, K., Kaatz, H., Bauer, C. G., Makower, A., Schulmeister, T., Scheller, F. W., Peter, M. G. and Wollenberger, U. (1998). Application of a sensitive catechol detector for determination of tyrosinase inhibitors. *Analytica Chimica Acta* 362(1): 81-90.

Sudhir, P.R., Wu, H.F. and Zhou, Z.C. (2005). Probing the interaction of kojic acid antibiotics with iron(III) chloride by using electrospray tandem mass spectrometry. *Rapid Communication Mass Spectrometry* 19(2):209-212.

Sun, T., Powers, J.R. and Tang, J. (2007). Evaluation of the antioxidant activity of asparagus, broccoli and their juices. *Food Chemistry* 105: 101–106.

Sung, M. S., Kimura, H. and Kusakabe, K. (2011). Esterification of oleic acid in a three-phase, fixed-bed reactor packed with a cation exchange resin catalyst. *Bioresource Technology* 102(2): 2130–2132.

- Svingen, T. and Tonissen, K.F. (2003). Altered HOX gene expression in human skin and breast cancer cells. *Cancer Biology and Therapy* 2(5): 518-23.
- Syamsul, K. M. W., Salina, M. R., Siti, S. O., Hanina, M. N., Basyaruddin, M. A. R. and Jusoff, K. (2010). Green synthesis of lauryl palmitate via lipase-catalyzed reaction. *World Applied Sciences Journal* 11(4): 401-407.
- Synytsya, A., Blafkova', P., Synytsya, A., C'opikova', J., Jir'i Spevacek, J. and Uher, M. Conjugation of kojic acid with chitosan. *Carbohydrate Polymers* 72(1): 21-31.
- Tai, H. P. and Brunner, G. (2009). Sugar fatty acid ester synthesis in high-pressure acetone-CO₂ system. *Journal of Supercritical Fluids* 48(1): 36-40.
- Takekoshi, S., Nagata, H. and Kitatani, K. (2014). Stimulation of melanogenesis by nordihydroguaiaretic acid in human melanoma cells. *Acta Histochemistry and Cytochemistry* 47(5): 203-210.
- Tamura T, Mitsumori K, Totsuka Y, Wakabayashi K, Kido R, Kasai H, Nasu M, Hirose M. (2006). Absence of *in vivo* genotoxic potential and tumor initiation activity of kojic acid in the rat thyroid. *Toxicology* 222(3): 213-224.
- Tan, T., Chen, B. Q., and Ye, H. (2006). Enzymatic synthesis of 2-ethylhexyl palmitate by lipase immobilized on fabric membranes in the batch reactor. *Biochemical Engineering Journal* 29(1-2): 41-45.
- Tan, X., Song, Y.H., Park, C., Lee, K.W., Kim, J.Y., Kim, D.W., Kim, K.D., Lee, K.W., Curtis-Long, M.J. and Park, K.H. (2015). Highly potent tyrosinase inhibitor, neorauflavane from *Campylotropis hirtella* and inhibitory mechanism with molecular docking. *Bioorganic and Medicinal Chemistry* (article in press)
- Tanaka, R., Tsujii, H., Yamada, T., Kajimoto, T., Amano, F., Hasegawa, J., Hamashima, Y., Node, M., Katoh, K. and Takebe, Y. (2009). Novel 3 α -methoxyserrat-14-en-21 β -ol (PJ-2)-curcumin, kojic acid, quercetin, and baicalein conjugates as HIV agents. *Bioorganic and Medicinal Chemistry* 17(14): 5238-5246.
- Taylor, K., Casalegno, C. and Stengel, W. (2011). A critique of the EC's expert (draft) reports on the status of alternatives for cosmetics testing to meet the 2013 deadline. *Alternatives to animal experimentation* 28(2): 131-148.
- Terabayashi, Y., Sano, M., Yamane, N., Marui, J., Tamano, K., Sagara, J., Dohmoto, M., Oda, K., Ohshima, E., Tachibana, K., Higa, Y., Ohashi, S., Koike, H., and Machida, M. (2010). Identification and characterization of genes responsible for biosynthesis of kojic acid, an industrially important

compound from *Aspergillus oryzae*. *Fungal Genetics and Biology* 47(12): 953–961.

- Thaipong, K., Boonprakob, U., Crosby, K., Cisneros-Zevallos, L. and Byrne, D.H. (2006). Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruit extracts. *Journal of Food Composition and Analysis* 19(6–7): 669–675.
- Tian, L., Zhou, J., Casimiro, M.C., Liang, B., Ojeifo, J.O., Wang, M., Hyslop, T., Wang, C. and Pestell, R.G. (2009a). Activating peroxisome proliferator-activated receptor gamma mutant promotes tumor growth *in vivo* by enhancing angiogenesis. *Cancer Research* 69(24): 9236–9244.
- Tian, S., He, P.Y., Zhang, J.Z. and Chen, Z. (2012). Effect of kappa elastin on melanogenesis in A375 human melanoma cells and its related mechanism. *Chinese Medicinal Journal* 125(22): 4088–4092.
- Tian, Y., Hoshino, T., Chen, C.J., E, Y., Yabe, S. and Liu, W. (2009b). The evaluation of whitening efficacy of cosmetic products using a human skin pigmentation spot model. *Skin Research Technology* 15(2): 218–223.
- Tobin, D.J. (2009). Aging of the Hair Follicle Pigmentation System. *International Journal of Trichology* 1(2): 83–93.
- Tobin, D.J., Slominski, A., Botchkarev, V. and Paus, R. (1999). The fate of hair follicle melanocytes during the hair growth cycle. *Journal of Investigate Dermatology Symposium Proceeding* 4(3): 323–332.
- Tokiwa, Y., Totani, T., Shimikawa, H., and Raku, T. (2003). J.P Patent No. 155,283. Japan, J.P.: Japanese Patent Office.
- Tornvall, U., Cecilia, O. C., Rajni, H. K. and Adlercreutz, D. (2007). Stability of immobilized *Candida antarctica* lipase B during chemo-enzymatic epoxidation of fatty acids. *Enzyme and Microbial Technology* 40(3): 447–451.
- Tu, C.X., Lin, M., Lu, S.S., Qi, X.Y., Zhang, R.X. and Zhang YY. (2012). Curcumin inhibits melanogenesis in human melanocytes. *Phytotherapy Research* 26(2): 174–179.
- Tuba, A.K. and Gülçin, I. (2008). Antioxidant and radical scavenging properties of curcumin. *Chemico-Biological Interactions* 174(1): 27–37.
- Uher, M., Chalabala, M. and Cizmárik, J. (2000). Kojic acid and its derivatives as potential therapeutic agents. *Ceska Slovenska Farmacie* 49(6): 288–298.
- Ujang, Z., Subramaniam, T., Diah, M.M., Wahid, H., Abdullah, B., Abd-Rashid, A.H. and Appleton, D. (2013). Bioguided fractionation and purification of natural bioactives obtained from *Alpinia conchigera* water extract with

melanin inhibition activity. *Journal of Biomaterials and Nanobiotechnology* 4: 265-272.

Unal, M.U. (1998). A study on the lipase-catalyzed esterification in organic solvent. *Turk Journal of Agriculture and Forestry* 22: 573-578.

Usenko, C.Y., Harper, S.L. and Tanguay, R.L (2007). *In vivo* evaluation of carbon fullerene toxicity using embryonic zebrafish. *PubMed Central* 45(9): 1891–1898.

Vad, N.M., Kandala, P.K., Srivastava, S.K. and Moridani, M.Y. (2010). Structure-toxicity relationship of phenolic analogs as anti-melanoma agents: an enzyme directed prodrug approach. *Chemico-Biological Interactions* 183(3): 462–471.

Vajragupta, O., Boonchoong, P., Sumanont, Y., Watanabe, H., Wongkrajang, Y. and Kammasud, N. (2003). Manganese-based complexes of radical scavengers as neuroprotective agents. *Bioorganic and Medical Chemistry* 11(10):2329-2337.

Vavricka, C.J., Christensen, B.M. and Li, J. (2010). Melanization in living organisms: a perspective of species evolution. *Protein and Cell* 1(9): 830-841.

Vegesna, V., Whithers, H.R. and Taylor, J.M. (1987). The effect on depigmentation after multifractionated irradiation of mouse resting hair follicles. *Radiation Research* 111(3):464-473.

Versonnen, J.B., Arijs, K., Verslycke, T., Lema, W. and Janssen, C.R (2003). *In vitro* and *in vivo* estrogenicity and toxicity of o-, m-, and p-dichlorobenzene. *Environmental Toxicology and Chemistry* 22(2): 329–335.

Videira, I.F.S., Moura, D.F.L. and Magina, S. (2013). Mechanisms regulating melanogenesis. *Anais Brasileiros de Dermatologi* 88(1): 76–83.

Vladimir-Knežević, S., Blažeković, B., Štefan, M.B., Alegro, A., Koszegi, T. and Petrik, J. (2011). Antioxidant activities and polyphenolic contents of three selected *Micromeria* species from Croatia. *Molecules* 16(2): 1454-1470.

Walas S.M (1999). Chemical Reactors. In D.W. Green and J.O. Maloney (Ed.), *Perrys Chemical Engineering Handbook* (pp. 1-20). New York: The McGraws Hill Companies Inc.

Wan, H. M., Chen, C. C., Giridhar, R. and Chang, T. S. (2005). Repeated-batch production of kojic acid in a cell-retention fermenter using *Aspergillus oryzae* M3B9. *Journal of Industrial Microbiology and Biotechnology* 32(6): 227-233.

Wang, K., Liu, C., Di, C.J., Ma, C., Han, C.G., Yuan, M.R., Li, P.F., Li, L. and Liu, Y.X. (2014). Kojic acid protects C57BL/6 mice from gamma-

irradiation induced damage. *Asian Pacific Journal of Cancer Prevention* 15(1): 291-297.

Wang, T.J., An, J., Chen, X.H., Deng, Q.D. and Yang, L. (2014). Assessment of *Cuscuta chinensis* seeds' effect on melanogenesis: Comparison of water and ethanol fractions *in vitro* and *in vivo*. *Journal of Ethnopharmacology* 154(1): 240-248.

Wei, C. I., Huang, T. S., Fernando, S. Y. and Chung, K. T. (1991). Mutagenicity studies of kojic acid. *Toxicology Letters* 59 (1-3): 213–220.

Wei, D., Gu, C., Song, Q. and Wu, S. (2003). Enzymatic esterification for glycoside lactate synthesis in organic solvent. *Enzyme and Microbial Technology* 33(4): 508–512.

Wei, Y., Zhang, C., Zhao, P., Yang, X. and Wang, K. (2011). A new salicylic acid-derivatized kojic acid vanadyl complex: synthesis, characterization and anti-diabetic therapeutic potential. *Journal of Inorganic Biochemistry* 105(8): 1081-1085.

Weigt, S., Huebler, N., Strecker, R., Braunbeck, T. and Broschard, T.H. (2012). Developmental effects of coumarin and the anticoagulant coumarin derivative warfarin on zebrafish (*Danio rerio*) embryos. *Reproductive Toxicology* 33(2): 133-141.

Weigt, S., Huebler, N., Strecker, R., Braunbeck, T., and Broschard, T.H. (2011). Zebrafish (*Danio rerio*) embryos as a model for testing proteratogens. *Toxicology* 281(1-3): 25-36.

Whittemore, J. and Neis (1998). U.S Patent No. 5,824,327. New York, N.Y.: U.S Patent and Trademark office.

Wicks, N.L., Chan, J.W., Najera, J.A., Ciriello, J.M. and Oancea, E. (2011). UVA phototransduction drives early melanin synthesis in human melanocytes. *Current Biology* 21(22): 1906-1911.

Won, K. and Lee, S.B. (2001). Effects of water and silica gel on enzyme agglomeration in organic solvents. *Biotechnology and Bioprocess Engineering* 6(2): 150-155.

Wright, J.A., Richards, T. and Srail, S.K.S (2014). The role of iron in the skin and cutaneous wound healing. *Frontier in Pharmacology* 5: 156

Xin, J.Y., Wang, Y., Liu, T., Lin, K., Chang, L. and Xia, C.G (2012). Biosynthesis of corn starch palmitate by lipase Novozym 435. *International Journal of Molecular Sciences* 13(6): 7226–7236.

Yamamoto, S., Nakanishi, K. and Hassan, M. A. (1997). Chromatographic separation of galactosylkojic acid. *Journal of Fermentation and Bioengineering* 84(1): 82-85.

- Yee, L. N., Akoh, C. C. and Phillips, R. S. (1997). Lipase PS-catalyzed transesterification of citronellyl butyrate and geranyl caproate: Effect of reaction parameters. *Journal of the American Oil Chemists' Society*, 74(3): 255-260.
- Yokozawa, T. and Kim, Y.J. (2007). Piceatannol inhibits melanogenesis by its antioxidative actions. *Biological Pharmaceutical Bulletin* 30(11): 2007-1011.
- Yoo, D.S., Lee, J., Choi, S.S., Rho, H.S., Cho, D.H., Shin, W.C. and Cho, J.Y. (2010). A modulatory effect of novel kojic acid derivatives on cancer cell proliferation and macrophage activation. *Pharmazie* 6:261-266.
- Yoon, I. H., Lee, Y. H., Park, C. W., Ji, H. and Lee, Y. S. (2010). Synthesis of dimers of (4-Oxo-4H-pyran-2-yl) acrylic acid as tyrosinase inhibitors. *Bulletin Korean Chemistry Society* 31(7): 2036-2038.
- Yu, J.S. and Kim, A.K. (2010). Effect of combination of taurine and azelaic acid on antimelanogenesis in murine melanoma cells. *Journal of Biomedical Science* 17(1): S45.
- Yumi, P., Jongsung, L., Junho, P., Deokhoon, P. (2003). Effects of kojic acid, arbutin and vitamin C on cell viability and melanin synthesis in B16BL6 cells. *Journal of the society of cosmetic scientists of Korea* 29(1): 151-167.
- Zang, L.Y., Cosma, G., Gardner, H., Shi, X., Castranova, V., and Vallyathan, V. (2000). Effect of antioxidant protection by p-coumaric acid on low-density lipoprotein cholesterol oxidation. *American Journal Physiology of Cell Physiology* 279(4): C954–C960.
- Zborowski, K., Grybos, R. and Proniewicz, L.M. (2005). Molecular structures of oxovanadium(IV) complexes with maltol and kojic acid: a quantum mechanical study. *Inorganic Chemistry Communications* 8(1): 76-78.
- Zborowski, K., Korenova, A., Uher, M. and Proniewicz, L.M. (2004). Quantum chemical studies on tautomeric equilibria in chlorokojic and azidokojic acids. *Journal of Molecular Structure (Theochem)* 683(1–3): 15–22.
- Zborowski, K.K., Sola, M., Poater, J. and Proniewicz, L.M. (2010). Theoretical studies on aromaticity of selected hydroxypyrones. Part 3#. Chelatoaromaticity phenomenon in metalcomplexes of hydroxypyrones. *Journal of Physical Organic Chemistry* 24(6): 499–450.
- Zhang, J. and Xu, J. (1995). Oleyl oleate synthesis by immobilized lipase from *Candida sp.* 1619. *Chinese Journal of Biotechnology* 11(4): 243-251.
- Zhao, C., Wang, X., Zhao, Y., Li, Z., Lin, S., Wei, Y. and Yang, H (2011). A novel xenograft model in zebrafish for high resolution investigating dynamics of neovascularization in tumors, *PLoS ONE* 6(7): e21768.

- Zheng, Q. (2012). Research on production method of sugar alcohol based on enzyme method synthesis. *International Proceeding of Computer Science and Information Technology Press* 25: 162-166.
- Zhu, Y.J., Qiu, L., Zhou, J.J., Guo, H.Y., Hu, Y.H., Li, Z.C., Wang, Q., Chen, Q.X. and Liu, B. (2010). Inhibitory effects of hinokitiol on tyrosinase activity and melanin biosynthesis and its antimicrobial activities. *Journal of Enzyme Inhibition and Medicinal Chemistry* 25(6): 798-803.
- Zirak, M. and Eftekhari-Si, B. (2015). Kojic acid in organic synthesis. *Turkish Journal of Chemistry* 39: 439-496.
- Zon, L. I. and Peterson, R. T. (2005). *In vivo* drug discovery in the zebrafish. *Nature Review Drug Discovery* 4(1): 35-44.
- Zong, M. H., Wu, H. and Tan, Z. Y. (2008). Substantially enhancing enzymatic regioselective acylation of 1- β -d-arabinofuranosylcytosine with vinyl caprylate by using a co-solvent mixture of hexane and pyridine. *Chemical Engineering Journal* 144(1): 75-78.
- Zoupanioti, M., Merianou, E., Karandreas, T., Stamatis, H. and Xenakis, A. (2010). Esterification of phenolic acids catalyzed by lipases immobilized in organogels. *Biotechnology Letters* 32(10): 1457-1462.
- Zuasti, A., Jiménez-Cervantes, C., García-Borrón, J.C. and Ferrer, C. (1998). The melanogenic system of *Xenopus laevis*. *Archive Histology and Cytology* 61(4): 305-316.
- Zulkefli, H.N., Mohamad, J. and Abidin, N.Y.Z (2013). Antioxidant activity of methanol extract of *Tinospora crispa* and *Tabernaemontana corymbosa*, *Sains Malaysiana* 42(6): 697-706.