



UNIVERSITI PUTRA MALAYSIA

***IDENTIFICATION OF LIPASE INHIBITOR FROM *Orthosiphon stamineus*
Benth AND ANALYSIS OF LIPASE-INHIBITOR COMPLEX
INTERACTION***

NORSYUHADA ALIAS

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By

NORSYUHADA ALIAS

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Philosophy**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Doctor of Philosophy

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Chairman : Prof. Raja Noor Zaliha Raja Abd. Rahman, D. Eng
Faculty : Biotechnology and Biomolecular Sciences

Natural products are a vast source of potential compounds that can be developed as an anti-obesity agent. One of the mechanisms of anti-obesity agents is inhibition of pancreatic lipase. Orlistat is the only commercial pancreatic lipase inhibitor with FDA approval, but it is derived synthetically and has side effects. Hence, there is a need to find for alternative from natural resources. It is postulated that lipase inhibitor from local plants could change pancreatic lipase structure conformation and impair its function. Therefore, this study aims to screen selected plants for pancreatic lipase-inhibitory activity, to identify the lipase-inhibitory compound and to analyse the lipase-inhibitor complex interaction. Screening of 24 crude extracts for their *in vitro* activity against porcine pancreatic lipase (PPL) detected four extracts demonstrating high (>70%) inhibition, while seven extracts had medium (30-70%) inhibition and the remaining 13 extracts exhibited low (<30%) inhibition when incubated with PPL at a final concentration of 500 µg/ml for 10 min at 37°C. *P. niruri* extract displayed the most potent PPL inhibitor, followed by *O. stamineus*, *M. paniculata* and *A. bilimbi* with the IC₅₀ value of 27.7<34.7<41.5<55.2 µg/ml, respectively. The best two extracts, namely *P. niruri* and *O. stamineus*, showed noncompetitive and uncompetitive inhibition, respectively. *P. niruri* and *O. stamineus* showed total phenolic content of 431.0 ± 0.01 and 103.0 ± 0.01 mg GAE/g dry extract, while total flavonoid content of 14.8 ± 0.07 and 21.6 ± 0.03 mg QE/g dry extract, respectively. Both *P. niruri* and *O. stamineus* extracts showed high antioxidant activity, with EC₅₀ values of 8.4 and 26.3 µg/ml, respectively. Isolation of lipase-inhibitory compound from *P. niruri* and *O. stamineus* was performed via chromatographic approaches. However, the isolation process later came to focus on *O. stamineus* active fractions due to difficulty separating the *P. niruri* active fraction. A combined fraction of MK38 and MK39 from *O. stamineus* extract demonstrated the highest inhibitory activity with 50% PPL inhibition. Fractionation of combined fraction MK38 and MK39 by high-performance liquid chromatography (HPLC) yielded an active compound designated as sub-fraction P5 with 45% PPL inhibition. Sub-fraction

P5 was authenticated as rosmarinic acid by spectroscopic analyses, namely liquid chromatography-mass spectrometry-mass spectrometry (MS-MS), Fourier transform infrared spectroscopy (FTIR), and nuclear magnetic resonance (NMR). Rosmarinic acid inhibited PPL in a non-competitive manner with an IC_{50} value of 19.5 $\mu\text{g/ml}$. Circular dichroism analysis showed a conformational change of the PPL secondary structure upon binding of rosmarinic acid towards PPL. However, no diffraction data were acquired from X-ray crystallography technique. Molecular docking predicted the potential binding site of rosmarinic acid was positioned far from the active site, whereas a molecular dynamic simulation projected that the flexibility of PPL structure would be affected upon binding of rosmarinic acid towards PPL. Hence, the *In silico* results were in agreement with the inhibition mode analysis. These results have suggested that rosmarinic acid from *O. stamineus* may play a complimentary role in obesity treatment, acting as a non-competitive pancreatic lipase inhibitor.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENGENALPASTIAN PERENCAT LIPASE DARI *Orthosiphon stamineus*
Benth DAN ANALISIS TERHADAP INTERAKSI DI ANTARA KOMPLEKS
LIPASE – PERENCAT**

Oleh

NORSYUHADA ALIAS

Jun 2016

Pengerusi : Prof. Raja Noor Zaliha Raja Abd. Rahman, D. Eng
Fakulti : Bioteknologi dan Sains Biomolekul

Tumbuhan dari sumber semula jadi berpotensi untuk menghasilkan agen anti-obesiti. Salah satu mekanisme agen anti-obesiti adalah dengan merencat aktiviti lipase dari pankreas. Orlistat adalah satu-satunya perencat lipase komersial yang mendapat kelulusan FDA, tetapi ianya dihasilkan secara sintetik dan mempunyai kesan sampingan. Justeru, ada keperluan untuk mendapatkan alternatif dari sumber semulajadi. Adalah dijangkakan yang lipase perencat dari tumbuhan tempatan boleh mengubah konformasi struktur lipase dari pankreas dan menjejaskan fungsinya. Jadi, penyelidikan ini bertujuan untuk menyaring tumbuhan terpilih yang mampu merencat aktiviti lipase dari pankreas, untuk mengenalpasti sebatian perencat lipase dan untuk menganalisa interaksi di antara kompleks lipase-perencat. Saringan 24 ekstrak dari tumbuhan terpilih yang berpotensi merencat aktiviti lipase dari pankreas khinzir (PPL) secara *in vitro* mendapati empat ekstrak menunjukkan perencatan tertinggi (>70% perencatan), tujuh ekstrak menunjukkan perencatan sederhana (30–70% perencatan) dan 13 ekstrak menunjukkan perencatan yang rendah (<30% perencatan) terhadap aktiviti lipase apabila diasai dengan PPL pada kepekatan akhir sebanyak 500 µg/ml selama 10 minit pada suhu 37°C. Ekstrak dari *P. niruri* adalah perencat yang paling berkesan, diikuti oleh ekstrak dari *O. stamineus*, *M. paniculata* dan *A. bilimbi* dengan masing-masing menunjukkan nilai IC₅₀ sebanyak 27.7<34.7<41.5<55.2 µg/ml. Dua ekstrak terbaik, iaitu *P. niruri* dan *O. stamineus*, masing-masing merencat PPL secara non-kompetitif dan un-kompetitif. *P. niruri* dan *O. stamineus*, masing-masing menunjukkan kandungan fenolik berjumlah 431.0 ± 0.01 dan 103.0 ± 0.01 mg GAE/g ekstrak kering manakala kandungan flavonoid berjumlah 14.8 ± 0.07 dan 21.6 ± 0.03 mg QE/g ekstrak kering. Kedua-dua ekstrak *P. niruri* dan *O. stamineus* menunjukkan aktiviti antioksidan yang tinggi dengan nilai EC₅₀ masing-masing sebanyak 8.4 dan 26.3 µg/ml. Proses pengasingan sebatian dari kedua-dua ekstrak *O. stamineus* dan *P. niruri* telah dibuat dengan menggunakan kaedah kromatografi. Walau bagaimanapun,

proses pengasingan sebatian kemudiannya hanya tertumpu kepada pecahan aktif *O. stamineus* kerana kesukaran memisahkan pecahan aktif dari ekstrak *P. niruri*. Gabungan pecahan MK38 dan MK39 daripada ekstrak *O. stamineus* menunjukkan aktiviti perencatan tertinggi dengan 50% aktiviti lipase berjaya direncat. Pemeringkatan kedua-dua pecahan menggunakan kromatografi cecair berprestasi tinggi (HPLC) berjaya memperolehi sebatian aktif yang dinamakan sebagai sub-pecahan P5 yang merencatkan aktiviti PPL sebanyak 45%. Sub-pecahan P5 telah disahkan sebagai asid rosmarinik melalui beberapa analisis spektroskopi iaitu kromatografi cecair gandingan spektrometri jisim-spektrometri jisim (MS-MS), spektroskopi inframerah transformasi Fourier (FTIR) dan spektrometer resonans magnet nukleus (NMR). Asid rosmarinik merencat aktiviti PPL secara non-kompetitif dengan nilai IC_{50} sebanyak 19.5 $\mu\text{g/ml}$. Analisis 'circular dichroism' pula menunjukkan berlakunya perubahan konformasi di dalam struktur PPL apabila ia bergabung dengan asid rosmarinik. Walaupun begitu, tiada sebarang data pembelauan diperolehi daripada teknik kristalografi sinar-X. Dok molekul menjangkakan asid rosmarinik mengikat struktur PPL pada kedudukan yang jauh dari tapak aktif manakala simulasi molekul dinamik menunjukkan bahawa fleksibiliti struktur PPL terjejas apabila berlakunya interaksi dengan asid rosmarinik. Oleh itu, hasil kajian *In silico* adalah selari dengan analisis mod perencatan. Kesimpulannya, asid rosmarinik dari *O. stamineus* boleh memainkan peranan bagi merawat kegemukan di mana ia bertindak sebagai perencat lipase dari pankreas secara non-kompetitif.

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I certify that a Thesis Examination Committee has met on 13 June 2016 to conduct the final examination of Norsyuhada bt Alias on her thesis entitled "Identification of Lipase Inhibitor from *Orthosiphon stamineus* Benth and Analysis of Lipase-Inhibitor Complex Interaction" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Wan Zuhainis binti Saad, PhD

Senior Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Mohd. Puad bin Abdullah, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Internal Examiner)

Norazizah binti Shafee, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Internal Examiner)

Naeem Rashid, PhD

Professor
University of the Punjab
Pakistan
(External Examiner)



ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 23 August 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Raja Noor Zaliha Raja Abd. Rahman, D. Eng

Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Abu Bakar Salleh, PhD

Professor Dato'
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Mohd Shukuri Mohamad Ali, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Adam Leow Thean Chor, PhD

Senior Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

Asilah Ahmad Tajudin, PhD

Senior Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

BUJANG KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

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Name and Matric No. : Norsyuhada Alias (GS29395)

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Signature : _____
Name of Chairman of
Supervisory
Committee : Prof. Dr. Raja Noor Zaliha Raja Abd.
Rahman

Signature : _____
Name of Member of
Supervisory
Committee : Prof. Dato' Dr. Abu Bakar Salleh

Signature : _____
Name of Member of
Supervisory
Committee : Assoc. Prof. Dr. Mohd Shukuri Mohamad Ali

Signature : _____
Name of Member of
Supervisory
Committee : Dr. Adam Leow Thean Chor

Signature : _____
Name of Member of
Supervisory
Committee : Dr. Asilah Ahmad Tajudin

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LIST OF ABBREVIATIONS

Å	Angstrom
CLP	Porcine colipase
Da	Dalton
°C	Degree Celsius
EDTA	Ethylendiaminetetraacetic acid
FDA	Food and Drug Administration
FTIR	Fourier transform infrared spectroscopy
HDL	High-density lipoprotein
HPL	Human pancreatic lipase
HPLC	High performance liquid chromatography
KBr	Potassium bromide
kDa	Kilodalton
LC-MS	Liquid chromatography-mass spectrometry
LPL	Lipoprotein lipase
M	Molar
mA	Milliampere
mg	Milligram
min	Minute
ml	Milliliter
mM	Milimolar
MS-MS	Liquid chromatography-mass spectrometry-mass spectrometry
MWCO	Molecular weight cut off
nm	Nanometer
NMR	Nuclear magnetic resonance
OD	Optical density
pNPB	<i>p</i> -nitrophenyl butyrate
PPL	Porcine pancreatic lipase
PL	Pancreatic lipase
RA	Rosmarinic acid
SDS-PAGE	Sodium dodecyl sulphate polyacrylamide gel electrophoresis
SDS	Sodium dodecyl sulphate
STZ	Streptozotocin
TEMED	N, N, N, N-Tetramethylenediamide
TLC	Thin-layer chromatography
U	Unit of activity
U/mg	Unit per milligram
U/ml	Unit per milliliter
UV	Ultraviolet
V	Volt
v/v	Volume per volume
w/v	Weight per volume
µg	Microgram
µl	Microliter
µm	Micrometer

CHAPTER 1

INTRODUCTION

Obesity has been classified as an epidemic affecting both developed and developing countries worldwide. The prevalence of obesity at an alarming rate has led to a rise of medical costs. More importantly, concerns about obesity are not about 'looking good' and having a beautiful silhouette, but merely maintaining a disease-free and healthy body. Obesity is known to facilitate the development of chronic diseases such as type 2 diabetes mellitus, hypertension, heart attack, stroke, osteoarthritis, sleep apnea, and some type of cancers (Mohamed *et al.*, 2014; Wan Mohamud *et al.*, 2011). The rise of these obesity-associated complications has compelled researchers to seek out lasting solutions for weight management and control. Although reduction of caloric intake by diet and increased level of physical activity are the most recommended approaches to lose weight, these attempts do not always work. Basically, suitable weight-loss programs depend on obesity level, overall health, and the patient's inclination to take part in the weight-loss plan. The treatment tools include dietary modifications, exercise, behaviour change, prescription of weight-loss medications and weight-loss surgery as the last resort (Glazer, 2001).

1.1 Problem statement

Nowadays, the need for anti-obesity drugs and other supplements is fast gaining recognition. There are many anti-obesity drugs in the market that have received approval from the U.S Food and Drug Administration (FDA). These anti-obesity drugs have specific mode of action to treat obesity. However, practically all of them have side effects. This was proven with the withdrawal of sibutramine, rimonabant, and a few other anti-obesity drugs from the European market due to their adverse side effects. The most effective lipase inhibitor in the market right now is orlistat. Orlistat is an over-the counter medication that is also labeled as a fat blocker. However, orlistat is a synthetic drug with side effects. Orlistat may cause severe liver and kidney problems (Filippatos *et al.*, 2008). It also inhibits the absorption of certain vitamins in the body. This indicates a need to find an effective and safe lipase inhibitor. Currently, most of the commercial non-prescribed slimming preparations available in the market are derived from plants. Various side effects from the synthetic drug application have prompted interest in the use of medication derived from natural sources. This indicates that plant-based materials may be an interesting sources for the development of anti-obesity agents, especially the one targeting on pancreatic lipase inhibition.

1.2 Rationale and novelty of the study

Malaysia is a tropical country rich with plants and herbs which are yet to be explored for their benefits. Many plants have traditionally been used as slimming aids by various communities in Malaysia, for daily consumptions or external used such as ointments and creams. Although the use of plant-based materials to lose weight has been based on the knowledge handed down through generations, their scientific evidence is still lacking and not well documented. There has been no detailed study of compound(s) contributing to the lipase inhibition action or anti-obesity property of these prospective plants. Hence, this study is vital to reveal the potential ability of selected plants in Malaysia as anti-obesity agents. This study would identify local plant(s) with the lipase-inhibition action and uncover the mechanisms through which the lipase-inhibitory compound acts. This could strengthen the fundamental knowledge on the interaction between lipase and its inhibitor. Various approaches have been applied in drug-discovery technology in order to develop new drugs. In this study, a combination of computer technology with the existing instrumentation, such as X-ray crystallography and circular dichroism, was chosen to study protein-ligand interaction. This provides exposure to high-end technology and diversifies the utilization of these sophisticated instruments provided by the university.

1.3 Hypothesis

Local plants selected in this study have a potential compound(s) capable of inhibiting pancreatic lipase. Binding interaction of the inhibitory compound(s) towards pancreatic lipase could trigger conformational changes of the pancreatic lipase structure that might impair its function.

1.4 Objectives of the project

This thesis aims to gain an understanding of the mechanism of pancreatic lipase inhibition of prospective Malaysian plants. The general aim results in several objectives:

1. Screening of selected plants for pancreatic lipase-inhibitory activity.
2. Isolation and identification of a pancreatic lipase-inhibitory compound.
3. Analysis of the pancreatic lipase-inhibitor complex interaction.

1.5 Outline of the thesis

Plants and herbs are natural sources of pancreatic lipase inhibitor for obesity treatment. A review of the prevalence of obesity, natural products as a potential source of anti-obesity agent, and the technology available to study the lipase inhibition action is contained in Chapter 2. Screening of selected Malaysian plants was performed to identify the potential plant(s) with the highest inhibition activity against pancreatic lipase and related in Chapter 3. Isolation of the

active compound was carried out by several chromatographic techniques, while the identification of the active compound was performed using several spectroscopic methods, as described in Chapter 4. Chapter 5 contains analyses of the protein-ligand complex interaction between the pancreatic lipase and the active compound (the potential lipase inhibitor), conducted by incorporating experimental testing with a computational approach. Finally, a summary of the results obtained in this thesis and recommendations for future study is contained in Chapter 6.



REFERENCES

- Abdullah, S., Shaari, A. R., & Azimi, A. (2012). Effect of Drying Methods on Metabolites Composition of Misai Kucing (*Orthosiphon stamineus*) Leaves. *APCBEE Procedia*, 2, 178–182.
- Abedini, A., Roumy, V., Mahieux, S., Murielle, B., Standaert-Vitse, A., Rivière, C., Sahpaz, S., Bailleul, F., Neut, C., & Hennebelle, T. (2013). Rosmarinic acid and its methyl ester as antimicrobial components of the hydromethanolic extract of *Hyptis atrorubens* Poit. (Lamiaceae). *Evidence-Based Complementary and Alternative Medicine*, 2013, 1-11.
- Acuthan, C. R., & Padikkala, J. (1997). Hypolipidemic effect of *Alpinia galanga* (Rasna) and *Kaempferia galanga* (Kachoori). *Indian Journal of Clinical Biochemistry*, 12(1), 55–58.
- Adcock, S. A., & McCammon, J. A. (2006). Molecular Dynamics: Survey of Methods for Simulating the Activity of Proteins. *Chemical reviews*, 106(5), 1589-1615.
- Adisakwattana, S., Intrawangso, J., Hemrid, A., Chanathong, B., & Makynen, K. (2012). Extracts of Edible Plants Inhibit Pancreatic Lipase, Cholesterol Esterase and Cholesterol Micellization, and Bind Bile Acids. *Food Technology and Biotechnology*, 50(1), 11–16.
- Ado, M. A. (2010). *Screening of Malaysian Medicinal / Herbs and Aquatic Plants for Pancreatic Lipase Inhibitory Activities and Identification of Active Constituent*. Universiti Putra Malaysia, Malaysia.
- Ado, M. A., Abas, F., Mohammed, A. S., & Ghazali, H. M. (2013). Anti- and pro-lipase activity of selected medicinal, herbal and aquatic plants, and structure elucidation of an anti-lipase compound. *Molecules*, 18(12), 14651–14669.
- Agati, G., Azzarello, E., Pollastri, S., & Tattini, M. (2012). Flavonoids as antioxidants in plants: location and functional significance. *Plant Science: An International Journal of Experimental Plant Biology*, 196, 67–76.
- Ahamed, M. B. K., Aisha, A. F. A., Nassar, Z. D., Siddiqui, J. M., Ismail, Z., Omari, S. M. S., Parish C. R., & Majid, A. M. S. A. (2012). Cat's Whiskers Tea (*Orthosiphon Stamineus*) Extract Inhibits Growth of Colon Tumor in Nude Mice and Angiogenesis in Endothelial Cells via Suppressing VEGFR Phosphorylation. *Nutrition and Cancer*, 64(1), 89–99.
- Ahmad, A., Alkarkhi, A. F. M., Hena, S., & Khim, L. H. (2009). Extraction, Separation and Identification of Chemical Ingredients of *Elephantopus scaber* L. Using Factorial Design of Experiment. *International Journal of Chemistry*, 1(1), 36–49.

- Ahmed, I. M., Cao, F., Zhang, M., Chen, X., Zhang, G., & Wu, F. (2013). Difference in Yield and Physiological Features in Response to Drought and Salinity Combined Stress during Anthesis in Tibetan Wild and Cultivated Barleys. *PLoS ONE*, *8*(10), 1–14.
- Akouwah, G. A., Zhari, I., Norhayati, I., & Sadikun, A. (2005). Radical Scavenging Activity of Methanol Leaf Extracts of *Orthosiphon stamineus*. *Pharmaceutical Biology*, *42*(8), 629–635.
- Akouwah, G. A., Zhari, I., Norhayati, I., Sadikun, A., & Khamsah, S. M. (2004). Sinensetin, eupatorin, 3'-hydroxy-5, 6, 7, 4'-tetramethoxyflavone and rosmarinic acid contents and antioxidative effect of *Orthosiphon stamineus* from Malaysia. *Food Chemistry*, *87*(4), 559–566.
- Alarcon-Aguilar, F. J., Zamilpa, A., Perez-Garcia, M. D., Almanza-Perez, J. C., Romero-Nuñez, E., Campos-Sepulveda, E. A., Vazquez-Carrillo, L. I., & Roman-Ramos, R. (2007). Effect of *Hibiscus sabdariffa* on obesity in MSG mice. *Journal of Ethnopharmacology*, *114*(1), 66–71.
- Alder, B. J., & Wainwright, T. E. (1959). Studies in molecular dynamics. General method. *Journal of Chemical Physics*, *31*(2), 459–466.
- Al-Dhabi, N. A., Arasu, M. V., Park, C. H., & Park, S. U. (2014). Recent Studies on Rosmarinic acid and its Biological and Pharmacological Activities. *EXCLI Journal*, *13*, 1192–1195.
- Alonso, H., Bliznyuk, A. A., & Gready, J. E. (2006). Combining docking and molecular dynamic simulations in drug design. *Medicinal Research Reviews*, *26*(5), 531–568.
- Altaf, R., Asmawi, M. Z., & Umar, M. I. (2013). Phytochemistry and medicinal properties of *Phaleria macrocarpa* (Scheff.) Boerl. extracts. *Pharmacognosy Reviews*, *7*(13), 73–80.
- Ambili, S., Subramoniam, A., & Nagarajan, S. N. (2009). Studies on the Antihyperlipidemic Properties of *Averrhoa bilimbi* Fruit in Rats. *Planta Medica*, *75*(1), 55–58.
- Amersham, B. (2002). *Gel Filtration Principles and Methods*. Piscataway, New Jersey, USA: Amersham Biosciences.
- Angoli, M., Barilli, A., Lesma, G., Passarella, D., Riva, S., Silvani, A., & Danieli, B. (2003). Remote stereocenter discrimination in the enzymatic resolution of piperidine-2-ethanol. Short enantioselective synthesis of sedamine and allosedamine. *The Journal of Organic Chemistry*, *68*(24), 9525–7.
- Anonymous. (2015a). Blog MySetBersalin.Com. Retrieved November 19, 2015, from <http://mysetbersalin.com/blog/artikel/6-petua-mengempiskan-perut-dan-melangsingkan-badan/>

- Anonymous. (2015b). Buah Herbal Menurunkan Berat Badan. Retrieved November 19, 2015, from <http://obatpelangsingherbal.com/buah-herbal-menurunkan-berat-badan/>
- Anonymous. (2015c). Cara langsing alami dengan cepat pakai daun salam. Retrieved November 19, 2015, from <http://marilangsingalami.blogspot.my/2014/12/cara-langsing-alami-dengan-cepat-pakai-daun-salam.html>
- Anonymous. (2015d). Manfaat Dan Khasiat Daun Cermai Bagi Kesehatan. Retrieved November 21, 2015, from <http://infoherbaltradisional.blogspot.my/2013/02/manfaat-dan-khasiat-daun-cermai-bagi.html>
- Anonymous. (2015e). Mika Beauty SPA. Retrieved November 19, 2015, from <http://mikabeautyspa.blogspot.my/2012/04/herba-sauna.html>
- Anonymous. (2015f). Petua Wanita: Untuk Wanita Dalam Pantang. Retrieved November 22, 2015, from http://little-queen-qkhadeeja.blogspot.my/2013/03/petua-wanita-untuk-wanita-dalam-pantang_13.html
- Anonymous. (2015g). Ramuan Obat Pelangsing Tradisional Alami. Retrieved November 21, 2015, from <http://melangsingkantubuh.info/ramuan-obat-pelangsing-tradisional-alami>
- Anonymous. (2015h). Sulphuric acid spot test. Retrieved November 26, 2015, from <http://www.stenutz.eu/chem/spot1.html>
- Anusuya, C., & Manoharan, S. (2011). Antitumor Initiating Potential of Rosmarinic Acid in 7,12-Dimethylbenz(a)anthracene-Induced Hamster Buccal Pouch Carcinogenesis. *Journal of Environmental Pathology, Toxicology and Oncology*, 30(3), 199–211.
- Arafat, O. M., Tham, S. Y., Sadikun, A., Zhari, I., Haughton, P. J., & Asmawi, M. Z. (2008). Studies on diuretic and hypouricemic effects of *Orthosiphon stamineus* methanol extracts in rats. *Journal of Ethnopharmacology*, 118(3), 354–360.
- Arambewela, L. S. R., Arawwawala, L. D. A. M., & Ratnasooriya, W. D. (2005). Antidiabetic activities of aqueous and ethanolic extracts of *Piper betle* leaves in rats. *Journal of Ethnopharmacology*, 102(2), 239–245.
- Armand, M. (2007). Lipases and lipolysis in the human digestive tract: where do we stand?. *Current Opinion in Clinical Nutrition and Metabolic Care*, 10(2), 156–64.
- Aung, H. T., Nikai, T., Niwa, M., & Takaya, Y. (2010). Rosmarinic acid in *Argusia argentea* inhibits snake venom-induced hemorrhage. *Journal of Natural Medicines*, 64(4), 482–486.

- Awale, S., Tezuka, Y., Banskota, A. H., & Kadota, S. (2003). Siphonols A-E: Novel nitric oxide inhibitors from *Orthosiphon stamineus* of Indonesia. *Bioorganic and Medicinal Chemistry Letters*, 13(1), 31–35.
- Bachok, M. F., Yusof, B.-N. M., Ismail, A., & Hamid, A. A. (2014). Effectiveness of traditional Malaysian vegetables (ulam) in modulating blood glucose levels. *Asia Pacific Journal of Clinical Nutrition*, 23(3), 369–76.
- Bavarva, J. H., & Narasimhacharya, A. V. R. L. (2007). Comparative Antidiabetic, Hypolipidemic, and Antioxidant Properties of *Phyllanthus niruri* in Normal and Diabetic Rats. *Pharmaceutical Biology*, 45(7), 569–574.
- Benkouka, F., Guidoni, A. A., De Caro, J. D., Bonicel, J. J., Desnuelle, P. A., & Rovey, M. (1982). Porcine pancreatic lipase. The disulfide bridges and the sulfhydryl groups. *European Journal of Biochemistry*, 128(2-3), 331–41.
- Birari, R. B., & Bhutani, K. K. (2007). Pancreatic lipase inhibitors from natural sources: unexplored potential. *Drug Discovery Today*, 12(19-20), 879–89.
- Bladt, S. (1996). *Plant Drug Analysis: A Thin Layer Chromatography Atlas* (Second). New York, USA: Springer-Verlag Berlin Heidelberg.
- BMRB. (2015). Biological Magnetic Resonance Data Bank: Rosmarinic acid. Retrieved December 6, 2015, from http://www.bmrwisc.edu/metabolomics/mol_summary/show_data.php?molName=rosmarinic_acid&id=bmse000648
- Boim, M. a., Heilberg, I. P., & Schor, N. (2010). *Phyllanthus niruri* as a promising alternative treatment for nephrolithiasis. *Brazilian Journal of Urology*, 36(6), 657–664.
- Borgström, B. (1988). Mode of action of tetrahydrolipstatin: a derivative of the naturally occurring lipase inhibitor lipstatin. *Biochimica et Biophysica Acta*, 962(3), 308–316.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72, 248–254.
- Bray, G. A., & Greenway, F. L. (1999). Current and potential drugs for treatment of obesity. *Endocrine Reviews*, 20(6), 805–75.
- Bray, G. A. (2000). A concise review on the therapeutics of obesity. *Nutrition*, 16(10), 953–960.
- Buchmueller, T. C., & Johar, M. (2015). Obesity and health expenditures: Evidence from Australia. *Economics and Human Biology*, 17, 42–58.

- Bustanji, Y., Issa, A., Mohammad, M., Hudaib, M., & Tawah, K. (2010). Inhibition of hormone sensitive lipase and pancreatic lipase by *Rosmarinus officinalis* extract and selected phenolic constituents. *Journal of Medicinal Plants Research*, 4(21), 2235–2242.
- Byun, H.-G., Eom, T.-K., Jung, W.-K., & Kim, S.-K. (2007). Lipase catalyzed production of monoacylglycerols by the esterification of fish oil fatty acids with glycerol. *Biotechnology and Bioprocess Engineering*, 12(5), 491–496.
- Caballero, V., Bautista, F. M., Campelo, J. M., Luna, D., Marinas, J. M., Romero, A. A., Hidalgo, J. M., Rafael, L., Anastacia, M., & Giordano, G. (2009). Sustainable preparation of a novel glycerol-free biofuel by using pig pancreatic lipase: Partial 1,3-regiospecific alcoholysis of sunflower oil. *Process Biochemistry*, 44(3), 334–342.
- Calixto, J. B., Santos, A. R. S., Filho, V. C., & Yunes, R. A. (1998). A review of the plants of the genus *Phyllanthus*: their chemistry, pharmacology, and therapeutic potential. *Medicinal Research Reviews*, 18(4), 225–258.
- Cambillau, C., Bourne, Y., Egloff, M. P., Martinez, C., & van Tilbeurgh, H. (1997). Pancreatic lipases and their complexes with colipases and inhibitors: crystallization and crystal packing. *Methods in Enzymology*, 284, 107–19.
- Cannon, B., & Nedergaard, J. A. N. (2004). Brown Adipose Tissue: Function and Physiological Significance. *Physiological Reviews*, 84(1), 277–359.
- Cardoso, F., Soares, A. M., Silva, P., Santos, J. I., & Fontes, M. R. M. (2011). Structural and Functional Studies of a Bothropic Myotoxin Complexed to Rosmarinic Acid: New Insights into Lys49-PLA 2 Inhibition. *PloS One*, 6(12), 1–12.
- Carvalho, B. M. A., Santos, J. D. L., Xavier, B. M., Almeida, J. R., Resende, L. M., Martins, W., Marcussi, S., Marangoni, S., Stábeli, R. G., Calderon, L. A., Soares, A. M., Silva, S. L., & Marchi-Salvador, D. P. (2013). Snake Venom PLA 2 s Inhibitors Isolated from Brazilian Plants: Synthetic and Natural Molecules. *BioMed Research International*, 2013, 1-8.
- Cavalier, J. F., Buono, G., & Verger, R. (2000). Covalent inhibition of digestive lipases by chiral phosphonates. *Accounts of Chemical Research*, 33(9), 579–89.
- Champe, P. C., Harvey, R. A., & Ferrier, D. R. (2005). *Biochemistry: Lippincott's illustrated reviews*. Philadelphia, USA: Lippincott Williams & Wilkins.
- Chang, R. (2005). Enzyme kinetics. In *Physical Chemistry for the Biosciences* (pp. 1–377). California, USA: University Science Books.
- Chantre, P., & Lairon, D. (2002). Recent findings of green tea extract AR25 (Exolise) and its activity for the treatment of obesity. *Phytomedicine*, 9(1),

3–8.

- Chapus, C., Rovey, M., Sarda, L., & Verger, R. (1988). Minireview on pancreatic lipase and colipase. *Biochimie*, 70(9), 1223–1234.
- Chen, K.-L., Li, H.-X., Xu, X.-L., & Zhou, G.-H. (2014). The protective effect of rosmarinic acid on hyperthermia-induced C2C12 muscle cells damage. *Molecular Biology Reports*, 41(8), 5525–31.
- Chu, X., Ci, X., He, J., Jiang, L., Wei, M., Cao, Q., Guan, M., Xie, X., Deng, X., & He, J. (2012). Effects of a natural prolyl oligopeptidase inhibitor, rosmarinic acid, on lipopolysaccharide-induced acute lung injury in mice. *Molecules*, 17(3), 3586–3598.
- Clapham, J. C., Arch, J. R., & Tadayyon, M. (2001). Anti-obesity drugs: a critical review of current therapies and future opportunities. *Pharmacology & Therapeutics*, 89(1), 81–121.
- Colombo, R., Andrea, A. N., Teles, H. L., Silva, G. H., Bomfim, G. C. C., Burgos, R. C. R., Cavalheiro, A. J., da Silva Bolzani, V., Pelícia, C. R., Guimarães, F. M., & Heimberg, M. C. H. (2009). Validated HPLC method for the standardization of *Phyllanthus niruri* (herb and commercial extracts) using corilaginas a phytochemical marker. *Biomedical Chromatography*, 23(6), 573–580.
- Copeland, R. A., Harpel, M. R., & Tummino, P. J. (2007). Targeting enzyme inhibitors. *Expert Opinion on Therapeutic Targets*, 11(7), 967–978.
- Costa, R. S., Carneiro, T. C. B., Cerqueira-Lima, A. T., Queiroz, N. V., Alcântara-Neves, N. M., Pontes-De-Carvalho, L. C., da Silva Velozo, E., Oliveira, E. J., & Figueiredo, C. A. (2012). *Ocimum gratissimum* Linn. and rosmarinic acid, attenuate eosinophilic airway inflammation in an experimental model of respiratory allergy to *Blomia tropicalis*. *International Immunopharmacology*, 13(1), 126–134.
- Couillard, C., Mauriège, P., Imbeault, P., Prud'homme, D., Nadeau, A., Tremblay, A., Bouchard, C., & Després, J. P. (2000). Hyperleptinemia is more closely associated with adipose cell hypertrophy than with adipose tissue hyperplasia. *International Journal of Obesity and Related Metabolic Disorders*, 24(6), 782–788.
- Datta, S. P., & Ottaway, J. H. (1976). Enzymes. In *Biochemistry* (Third, pp. 128–135). London: Cassell and Collier Macmillan Publisher Limited.
- Davey, T. M., Allotey, P., & Reidpath, D. D. (2013). Is obesity an ineluctable consequence of development? A case study of Malaysia. *Public Health*, 127(12), 1057–1062.
- De Caro, J., Boudouard, M., Bonicel, J., Guidoni, A., Desnuelle, P., & Rovey, M. (1981). Porcine pancreatic lipase. Completion of the primary structure. *Biochimica et Biophysica Acta*, 671(2), 129–38.
- De La Garza, A. L., Milagro, F. I., Boque, N., Campión, J., & Martínez, J. A.

- (2011). Natural inhibitors of pancreatic lipase as new players in obesity treatment. *Planta Medica*, 77(8), 773–785.
- De Oliveira, N. C. D., Sarmiento, M. S., Nunes, E. a, Porto, C. M., Rosa, D. P., Bona, S. R., Rodrigues, G., Marroni, N. P., Pereira, P., Picada, J. N., Ferraz, A. B., Thiesen, F. V., & Da Silva, J. (2012). Rosmarinic acid as a protective agent against genotoxicity of ethanol in mice. *Food and Chemical Toxicology*, 50(5), 1208–1214.
- De Souza, R. L., Barbosa, J. M. P., Zanin, G. M., Lobão, M. W. N., Soares, C. M. F., & Lima, A. S. (2010). Partitioning of porcine pancreatic lipase in a two-phase systems of polyethylene glycol/potassium phosphate aqueous. *Applied Biochemistry and Biotechnology*, 161(1-8), 288–300.
- Dechakhamphu, A., & Wongchum, N. (2015). Screening for anti-pancreatic lipase properties of 28 traditional Thai medicinal herbs. *Asian Pacific Journal of Tropical Biomedicine*, 5(12), 1042–1045.
- Domínguez-Muñoz, J. E. (2011). Pancreatic exocrine insufficiency: diagnosis and treatment. *Journal of Gastroenterology and Hepatology*, 26 Suppl 2, 12–16.
- Domitrović, R., Potočnjak, I., Crnčević-Orlić, Z., & Škoda, M. (2014). Nephroprotective activities of rosmarinic acid against cisplatin-induced kidney injury in mice. *Food and Chemical Toxicology*, 66, 321–328.
- Duan, Y., Wu, C., Chowdhury, S., Lee, M. C., Xiong, G., Zhang, W., Yang, R., Cieplak, P., Luo, R., Lee, T., Caldwell, J., Wang, J. & Kollman, P. (2003). A point-charge force field for molecular mechanics simulations of proteins based on condensed-phase quantum mechanical calculations. *Journal of Computational Chemistry*, 24(16), 1999–2012.
- Dubois, M., Bailly, F., Mbemba, G., Mouscadet, J.-F., Debyser, Z., Witvrouw, M., & Cotellet, P. (2008). Reaction of rosmarinic acid with nitrite ions in acidic conditions: discovery of nitro- and dinitrorosmarinic acids as new anti-HIV-1 agents. *Journal of Medicinal Chemistry*, 51(8), 2575–2579.
- Dunnick, J. K., Kissling, G., Gerken, D. K., Vallant, M. A., & Nyska, A. (2007). Cardiotoxicity of Ma Huang/caffeine or ephedrine/caffeine in a rodent model system. *Toxicologic Pathology*, 35(5), 657–64.
- Dweck, A. C. (2015). A review of Asam Gelugor (*Garcinia atroviridis*) Griff. ex T. Aders. Retrieved November 19, 2015, from <http://www.dweckdata.com/Research files/Garcinia atroviridis.pdf>
- Egloff, M. P., Marquet, F., Buono, G., Verger, R., Cambillau, C., & van Tilbeurgh, H. (1995). The 2.46 Å resolution structure of the pancreatic lipase-colipase complex inhibited by a C11 alkyl phosphonate. *Biochemistry*, 34(9), 2751–2762.

- FDA. (2015). FDA Approved Drug Products. Retrieved December 3, 2015, from <http://www.accessdata.fda.gov/scripts/cder/drugsatfda/index>.
- Fernand, V. E. (2003). *Initial Characterization of Crude Extracts from Phyllanthus amarus Schum. and Thonn. and Quassia amara L. using Normal Phase Thin Layer Chromatography*. University of Suriname, Republic of Suriname.
- Filippatos, T. D., Derdemezis, C. S., Gazi, I. F., Nakou, E. S., Mikhailidis, D. P., & Elisaf, M. S. (2008). Orlistat-associated adverse effects and drug interactions: a critical review. *Drug Safety*, 31(1), 53–65.
- Finer, N. (1997). Present and future pharmacological approaches. *British Medical Bulletin*, 53(2), 409–32.
- Fried, B., & Sherma, J. (1994). *Thin-layer chromatography: Techniques and applications* (Third). New York, USA: Marcel Dekker, Inc.
- García-Lafuente, A., Guillaumon, E., Villares, A., Rostagno, M. A., & Martínez, J. A. (2009). Flavonoids as anti-inflammatory agents: Implications in cancer and cardiovascular disease. *Inflammation Research*, 58(9), 537–552.
- Garcia-Viloca, M., Gao, J., Karplus, M., & Truhlar, D. G. (2004). How enzymes work: analysis by modern rate theory and computer simulations. *Science*, 303(5655), 186–95.
- Garner, C. W., & Smith L. C. (1970). A convenient purification of porcine pancreatic lipase free of proteolytic activity. *Archives of Biochemistry and Biophysics*, 140(2), 503–507.
- Garner, C. W., & Smith, L. C. (1972). Porcine pancreatic lipase. A glycoprotein. *The Journal of Biological Chemistry*, 247(2), 561–565.
- Genest, P. W., Field, T. G., Vasudevan, P. T., & Palekar, A. A. (1998). Continuous purification of porcine lipase by rotating annular size-exclusion chromatography. *Applied Biochemistry and Biotechnology*, 73(2-3), 215–30.
- Gertsch, J. (2011). Planta Medica. *Botanical Drugs, Synergy, and Network Pharmacology: Forth and Back to Intelligent Mixtures*, 77(11), 1086–1098.
- Gholamhoseinian, A., Shahouzi, B., & Sharifi-far, F. (2010). Inhibitory effect of some plant extracts on pancreatic lipase. *International Journal of Pharmacology*, 6(1), 18–24.
- Giribabu, N., Rao, P. V., Kumar, K. P., Muniandy, S., Swapna Rekha, S., & Salleh, N. (2014). Aqueous Extract of *Phyllanthus niruri* Leaves Displays *In Vitro* Antioxidant Activity and Prevents the Elevation of Oxidative Stress in the Kidney of Streptozotocin-Induced Diabetic Male Rats. *Evidence-Based Complementary and Alternative Medicine*, 2014, 1–10.

- Glazer, G. (2001). Long-term pharmacotherapy of obesity 2000: A review of efficacy and safety. *Archives of Internal Medicine*, 161(15), 1814–24.
- Gogoi, S., Pathak, M. G., Dutta, A., & Dutta, N. N. (2008). Porcine pancreas lipase catalyzed synthesis of lauryl laurate in organic solvent media: A kinetic study. *Indian Journal of Biochemistry and Biophysics*, 45(3), 192–197.
- Goldstein, A., Aronow, L., & Kalman, S. M. (1970). Principles of Drug Action. The Basis of Pharmacology. *Journal of Medicinal Chemistry*, 13(2), 337–337.
- Gooda Sahib, N., Abdul Hamid, A., Saari, N., Abas, F., Pak Dek, M. S., & Rahim, M. (2012). Anti-Pancreatic Lipase and Antioxidant Activity of Selected Tropical Herbs. *International Journal of Food Properties*, 15(3), 569–578.
- Gooda Sahib, N., Saari, N., Ismail, A., Khatib, A., Mahomoodally, F., & Abdul Hamid, A. (2012). Plants' Metabolites as Potential Antiobesity Agents. *The Scientific World Journal*, 2012, 1–8.
- Gorinstein, S., Zachwieja, Z., Katrich, E., Pawelzik, E., Haruenkit, R., Trakhtenberg, S., & Martinbelloso, O. (2004). Comparison of the contents of the main antioxidant compounds and the antioxidant activity of white grapefruit and his new hybrid. *Lebensmittel-Wissenschaft Und-Technologie*, 37(3), 337–343.
- Gu, Y., Hurst, W. J., Stuart, D. a., & Lambert, J. D. (2011). Inhibition of key digestive enzymes by cocoa extracts and procyanidins. *Journal of Agricultural and Food Chemistry*, 59(10), 5305–5311.
- Gueto, C., Torres, J., & Vivas-Reyes, R. (2009). CoMFA, LeapFrog and blind docking studies on sulfonanilide derivatives acting as selective aromatase expression regulators. *European Journal of Medicinal Chemistry*, 44(9), 3445–3451.
- Gulati, V., Harding, I. H., & Palombo, E. A. (2012). Enzyme inhibitory and antioxidant activities of traditional medicinal plants: potential application in the management of hyperglycemia. *BMC Complementary and Alternative Medicine*, 12(77), 1-9.
- Gülçin, İ., Huyut, Z., Elmastaş, M., & Aboul-Enein, H. Y. (2010). Radical scavenging and antioxidant activity of tannic acid. *Arabian Journal of Chemistry*, 3(1), 43–53.
- Hadváry, P., Lengsfeld, H., & Wolfer, H. (1988). Inhibition of pancreatic lipase *in vitro* by the covalent inhibitor tetrahydrolipstatin. *The Biochemical Journal*, 256(2), 357–361.

- Hajian-Tilaki, K. O., & Heidari, B. (2007). Prevalence of obesity, central obesity and the associated factors in urban population aged 20-70 years, in the north of Iran: a population-based study and regression approach. *Obesity Reviews*, 8(1), 3–10.
- Hakiman, M. (2012). Total antioxidant, polyphenol, phenolic acid, and flavonoid content in *Ficus deltoidea* varieties. *Journal of Medicinal Plants Research*, 6(33), 4776–4784.
- Han, L. K., Xu, B. J., Kimura, Y., Zheng, Y. n., & Okuda, H. (2000). *Platycodi radix* affects lipid metabolism in mice with high fat diet-induced obesity. *The Journal of Nutrition*, 130(11), 2760–4.
- Han, L., Li, W., Narimatsu, S., Liu, L., Fu, H., Okuda, H., & Koike, K. (2006). Inhibitory effects of compounds isolated from fruit of *Juglans mandshurica* on pancreatic lipase. *Journal of Natural Medicines*, 61(2), 184–186.
- Han, L.-K., Zheng, Y.-N., Yoshikawa, M., Okuda, H., & Kimura, Y. (2005). Anti-obesity effects of chikusetsusaponins isolated from *Panax japonicus* rhizomes. *BMC Complementary and Alternative Medicine*, 5, 9.
- Handa, S. S. (2008). *Extraction technologies for medicinal and aromatic plants*. Trieste, Italy: International Centre for Science and High Technology.
- Hansson, T., Oostenbrink, C., & Van Gunsteren, W. (2002). Molecular dynamics simulations. *Current Opinion in Structural Biology*, 12(2), 190–196.
- Harish, R., & Shivanandappa, T. (2006). Antioxidant activity and hepatoprotective potential of *Phyllanthus niruri*. *Food Chemistry*, 95(2), 180–185.
- Hasan, F., Shah, A. A., & Hameed, A. (2006). Industrial applications of microbial lipases. *Enzyme and Microbial Technology*, 39(2), 235–251.
- Hasanein, P., & Zaheri, L. M. (2014). Effects of rosmarinic acid on an experimental model of painful diabetic neuropathy in rats. *Pharmaceutical Biology*, 52(11), 1398–1402.
- Hermoso, J., Pignol, D., Kerfelec, B., Crenon, I., Chapus, C., & Fontecilla-Camps, J. C. (1996). Lipase activation by nonionic detergents: The crystal structure of the porcine lipase-colipase-tetraethylene glycol monoethyl ether complex. *Journal of Biological Chemistry*, 271(30), 18007–18016.
- Herrero, M., Plaza, M., Cifuentes, a, & Ibáñez, E. (2010). Green processes for the extraction of bioactives from Rosemary: Chemical and functional characterization via ultra-performance liquid chromatography-tandem mass spectrometry and in-vitro assays. *Journal of Chromatography A*, 1217(16), 2512–20.

- Hill, J. O. (2006). Understanding and addressing the epidemic of obesity: An energy balance perspective. *Endocrine Reviews*, 27(7), 750–761.
- Himani, B., Seema, B., Bhole, N., Mayank, Y., Vinod, S., & Mamta, S. (2013). Misai kuching: A glimpse of maestro. *International Journal of Pharmaceutical Sciences Review and Research*, 22(2), 55–59.
- Hiratake, J. (2005). Enzyme inhibitors as chemical tools to study enzyme catalysis: rational design, synthesis, and applications. *Chemical Record*, 5(4), 209–228.
- Ho, C. H., Noryati, I., Sulaiman, S. F., & Rosma, A. (2010). *In vitro* antibacterial and antioxidant activities of *Orthosiphon stamineus* Benth. extracts against food-borne bacteria. *Food Chemistry*, 122(4), 1168–1172.
- Hollman, P. C. H., & Katan, M. B. (1999). Dietary Flavonoids: Intake, Health Effects and Bioavailability. *Food and Chemical Toxicology*, 37(9-10), 937–942.
- Hossain, M. A., & Ismail, Z. (2012). Quantification and enrichment of sinensetin in the leaves of *Orthosiphon stamineus*. *Arabian Journal of Chemistry*, (2012), 1-4.
- Hossain, M. A., Ismail, Z., Rahman, A., & Kang, S. C. (2008). Chemical composition and anti-fungal properties of the essential oils and crude extracts of *Orthosiphon stamineus* Benth. *Industrial Crops and Products*, 27(3), 328–334.
- Hossain, M. B., Rai, D. K., Brunton, N. P., Martin-Diana, A. B., & Barry-Ryan, C. (2010). Characterization of Phenolic Composition in Lamiaceae Spices by LC-ESI-MS/MS. *Journal of Agricultural and Food Chemistry*, 58(19), 10576–10581.
- Hossain, M. A., & Mizanur Rahman, S. M. (2011). Isolation and characterisation of flavonoids from the leaves of medicinal plant *Orthosiphon stamineus*. *Arabian Journal of Chemistry*, 8(2), 6–9.
- Hossain, P., Kavar, B., & El Nahas, M. (2007). Obesity and diabetes in the developing world—a growing challenge. *The New England Journal of Medicine*, 356(3), 213–5.
- Houghton, P., & Amala, R. (2012). *Laboratory Handbook for the Fractionation of Natural Extracts*. USA: Springer Science & Business Media.
- Hu, B., Cui, F., Yin, F., Zeng, X., Sun, Y., & Li, Y. (2015). Caffeoylquinic acids competitively inhibit pancreatic lipase through binding to the catalytic triad. *International Journal of Biological Macromolecules*, 80, 529–535.
- Huang, S., & Zheng, R. (2006). Rosmarinic acid inhibits angiogenesis and its mechanism of action *in vitro*. *Cancer Letters*, 239(2), 271–280.

- Huang, S.-Y., & Zou, X. (2010). Advances and Challenges in Protein-Ligand Docking. *International Journal of Molecular Sciences*, 11(8), 3016–3034.
- Hughes, J. P., Rees, S., Kalindjian, S. B., & Philpott, K. L. (2011). Principles of early drug discovery. *British Journal of Pharmacology*, 162(6), 1239–1249.
- Husain, G. M., Chatterjee, S. S., Singh, P. N., & Kumar, V. (2011). Hypolipidemic and Antiobesity-Like Activity of Standardised Extract of *Hypericum perforatum* L. in Rats. *International Scholarly Research Network*, 2011, 1–7.
- Ibrahim, H., Awadhi, A. Al, Shaban, S., & Nair, S. C. (2015). Can our residents carry the weight of the obesity crisis? A mixed methods study. *Obesity Research & Clinical Practice*, 9(3), 228–33.
- Ikeda, I., Tsuda, K., Suzuki, Y., Kobayashi, M., Unno, T., Tomoyori, H., Goto, H., Kawata, Y., Imaizumi, K., Nozawa, A. & Kakuda, T. (2005). Tea Catechins with a Galloyl Moiety Suppress Postprandial Hypertriacylglycerolemia by Delaying Lymphatic Transport of Dietary Fat in Rats. *Biochemical and Molecular Actions of Nutrients*, 135(2), 155–159.
- Insull, W. J. (2006). Clinical utility of bile acid sequestrants in the treatment of dyslipidemia: a scientific review. *Southern Medical Journal*, 99(3), 257–73.
- Iram, N. E, Khan, M. S., Jolly, R., Arshad, M., Alam, M., Alam, P., Khan, R. H., & Firdaus, F. (2015). Interaction mode of polycarbazole–titanium dioxide nanocomposite with DNA: Molecular docking simulation and in-vitro antimicrobial study. *Journal of Photochemistry and Photobiology B: Biology*, 153, 20–32.
- Ismail, M. N., Chee, S. S., Nawawi, H., Yusoff, K., Lim, T. O., & James, W. P. T. (2002). Obesity in Malaysia. *Obesity Reviews*, 3(3), 203–208.
- Iswantini, D., Silitonga, R. F., Martatilofa, E., & Darusman, L. K. (2011). *Zingiber cassumunar*, *Guazuma ulmifolia*, and *Murraya paniculata* Extracts as Antiobesity: *In Vitro* Inhibitory Effect on Pancreatic Lipase Activity. *Hayati Journal of Biosciences*, 18(1), 6–10.
- James, P. T., Leach, R., Kalamara, E., & Shayeghi, M. (2001). The worldwide obesity epidemic. *Obesity Research*, 9 Suppl 4(November), 228S–233S.
- Jayanthi, G., & Subramanian, S. (2014). Rosmarinic acid, a polyphenol, ameliorates hyperglycemia by regulating the key enzymes of carbohydrate metabolism in high fat diet – STZ induced experimental diabetes mellitus. *Biomedicine & Preventive Nutrition*, 4(3), 431–437.
- Jick, H., Vasilakis, C., Weinrauch, L. A., Meier, C. R., Jick, S. S., & Derby, L. E. (1998). A population-based study of appetite-suppressant drugs and the

risk of cardiac-valve regurgitation. *The New England Journal of Medicine*, 339(11), 719–24.

John, B., Sulaiman, C. T., George, S., & Reddy, V. R. K. (2014). Total phenolics and flavonoids in selected medicinal plants from Karala. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(1), 406–408.

Jork, H., Funk, W., Wimmer, H., & Fischer, W. R. (1990). *Thin-Layer Chromatography: Reagents and Detection Methods-Physical & Chemical Detection Methods: Fundamentals, Reagents I, Volume 1a*. New York, USA: Wiley.

Karmokar, A., Marczylo, T. H., Cai, H., Steward, W. P., Gescher, A. J., & Brown, K. (2012). Dietary intake of rosmarinic acid by ApcMin mice, a model of colorectal carcinogenesis: Levels of parent agent in the target tissue and effect on adenoma development. *Molecular Nutrition and Food Research*, 56, 775–783.

Karthik, D., Viswanathan, P., & Anuradha, C. V. (2011). Administration of Rosmarinic Acid Reduces Cardiopathology and Blood Pressure Through Inhibition of p22phox NADPH Oxidase in Fructose-Fed Hypertensive Rats. *Journal of Cardiovascular Pharmacology*, 58(5), 514–521.

Kelly, S. M., Jess, T. J., & Price, N. C. (2005). How to study proteins by circular dichroism. *Biochimica et Biophysica Acta*, 1751(2), 119–139.

Khanna, A. K., Rizvi, F., & Chander, R. (2002). Lipid lowering activity of *Phyllanthus niruri* in hyperlipemic rats. *Journal of Ethnopharmacology*, 82, 19–22.

Kim, G.-D., Park, Y. S., Jin, Y.-H., & Park, C.-S. (2015). Production and applications of rosmarinic acid and structurally related compounds. *Applied Microbiology and Biotechnology*, 99(5), 2083–2092.

Kim, H. Y., & Kang, M. H. (2005). Screening of Korean medicinal plants for lipase inhibitory activity. *Phytotherapy Research*, 19(4), 359–61.

Kim, H., Della-fera, M., Lin, J., & Baile, C. A. (2006). Docosahexaenoic Acid Inhibits Adipocyte Differentiation and Induces Apoptosis in 3T3-L1 Preadipocytes 1. *The Journal of Nutrition*, 136(12), 2965–2969.

Kim, S.-J., Um, J.-Y., Kim, S.-H., & Hong, S.-H. (2013). Protective Effect of Rosmarinic Acid is Through Regulation of Inflammatory Cytokine in Cadmium-Induced Ototoxicity. *The American Journal of Chinese Medicine*, 41(2), 391–404.

Kim, Y. M., Jeong, Y. K., Wang, M. H., Lee, W. Y., & Rhee, H. I. (2005). Inhibitory effect of pine extract on α -glucosidase activity and postprandial hyperglycemia. *Nutrition*, 21, 756–761.

- Kiran, K. R., Suresh Babu, C. V., & Divakar, S. (2001). Thermostability of porcine pancreas lipase in non-aqueous media. *Process Biochemistry*, 36(8-9), 885–892.
- Kishino, E., Ito, T., Fujita, K., & Kiuchi, Y. (2006). A Mixture of the *Salacia reticulata* (Kotala himbutu) Aqueous Extract and Cyclodextrin Reduces the Accumulation of Visceral Fat Mass in Mice and Rats with High-Fat Diet-Induced Obesity. *The Journal of Nutrition*, 136(2), 433–439.
- Kissane, N. A., & Pratt, J. S. A. (2011). Medical and surgical treatment of obesity. *Best Practice & Research Clinical Anaesthesiology*, 25(1), 11–25.
- Kotowaroo, M. I., Mahomoodally, M. F., Gurib-Fakim, A., & Subratty, A. H. (2006). Screening of traditional antidiabetic medicinal plants of Mauritius for possible α -amylase inhibitory effects *in vitro*. *Phytotherapy Research*, 20(3), 228–231.
- Krieger, E., Koraimann, G., & Vriend, G. (2002). Increasing the precision of comparative models with YASARA NOVA-A self-parameterizing force field. *Proteins*, 47(3), 393–402.
- Krishnamurthy, K. H. (2014). Medicinal plants: *Phyllanthus niruri*, Linn. Retrieved December 4, 2015, from <http://www.namahjournal.com/doc/Actual/Phyllanthus-vol-19-iss-3.html>.
- Kuru, P. (2014). *Tamarindus indica* and its health related effects. *Asian Pacific Journal of Tropical Biomedicine*, 4(9), 676–681.
- Laemmli, U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 227(5259), 680–685.
- Lee, J., Jung, E., Koh, J., Kim, Y. S., & Park, D. (2008). Effect of rosmarinic acid on atopic dermatitis. *The Journal of Dermatology*, 35(12), 768–771.
- Lee, J., Kim, Y. S., & Park, D. (2007). Rosmarinic acid induces melanogenesis through protein kinase A activation signaling. *Biochemical Pharmacology*, 74, 960–968.
- Lehner, R., & Verger, R. (1997). Purification and characterization of a porcine liver microsomal triacylglycerol hydrolase. *Biochemistry*, 36(7), 1861–8.
- Lei, F., Zhang, X. N., Wang, W., Xing, D. M., Xie, W. D., Su, H., & Du, L. J. (2007). Evidence of anti-obesity effects of the pomegranate leaf extract in high-fat diet induced obese mice. *International Journal of Obesity*, 31(6), 1023–9.
- Lewis, D. R. (2012). Direct Measurement of Lipase Inhibition by Orlistat Using a Dissolution Linked *In Vitro* Assay. *Clinical Pharmacology & Biopharmaceutics*, 1(3), 1–3.
- Lim, Y. Y., & Murtijaya, J. (2007). Antioxidant properties of *Phyllanthus amarus* extracts as affected by different drying methods. *LWT - Food Science and*

Technology, 40(9), 1664–1669.

Linné, Y., & Rössner, S. (2004). Pharmacotherapy of obesity. *Clinics in Dermatology*, 22(4), 319–24.

Little, T. J., Horowitz, M., & Feinle-Bisset, C. (2007). Modulation by high-fat diets of gastrointestinal function and hormones associated with the regulation of energy intake: Implications for the pathophysiology of obesity. *American Journal of Clinical Nutrition*, 86(3), 531–541.

Liu, F.-C., Chen, H.-L., Lin, W., Tung, Y.-T., Lai, C.-W., Hsu, A.-L., & Chen, C.-M. (2010). Application of porcine lipase secreted by *Pichia pastoris* to improve fat digestion and growth performance of postweaning piglets. *Journal of Agricultural and Food Chemistry*, 58(6), 3322–3329.

Liyana-Pathirana, C. M., & Shahidi, F. (2006). Antioxidant properties of commercial soft and hard winter wheats (*Triticum aestivum* L.) and their milling fractions. *Journal of the Science of Food and Agriculture*, 86(3), 477–485.

Loon, Y. H., Wong, J. W., Yap, S. P., & Yuen, K. H. (2005). Determination of flavonoids from *Orthosiphon stamineus* in plasma using a simple HPLC method with ultraviolet detection. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, 816(1-2), 161–166.

Lookene, A., Skottova, N., & Olivecrona, G. (1994). Interactions of lipoprotein lipase with the active-site inhibitor tetrahydrolipstatin (Orlistat). *European Journal of Biochemistry*, 222(2), 395–403.

Loret de Mola, C., Pillay, T. D., Diez-Canseco, F., Gilman, R. H., Smeeth, L., & Miranda, J. J. (2012). Body mass index and self-perception of overweight and obesity in rural, urban and rural-to-urban migrants: PERU MIGRANT study. *PloS One*, 7(11), 1-8.

Lowe, M. E. (1997). Colipase stabilizes the lid domain of pancreatic triglyceride lipase. *The Journal of Biological Chemistry*, 272(1), 9–12.

Lowe, M. E. (1997). Structure and Function of Pancreatic Lipase and Colipase. *Annual Review of Nutrition*, 17, 141–158.

Lowe, M. E. (2002). The triglyceride lipases of the pancreas. *The Journal of Lipid Research*, 43(12), 2007–2016.

Luan, H., Kan, Z., Xu, Y., Lv, C., & Jiang, W. (2013). Rosmarinic acid protects against experimental diabetes with cerebral ischemia: relation to inflammation response. *Journal of Neuroinflammation*, 10(28), 1-10.

Lucarini, R., Bernardes, W. A., Ferreira, D. S., Tozatti, M. G., Furtado, R., Bastos, J. K., Pauletti, P. M., Januário, A. H., Silva, M. L., & Cunha, W. R. (2013). *In vivo* analgesic and anti-inflammatory activities of *Rosmarinus officinalis* aqueous extracts, rosmarinic acid and its acetyl ester derivative.

Pharmaceutical Biology, 51(9), 1087–1090.

- Luft, J. R., Wolfley, J. R., & Snell, E. H. (2011). What's in a Drop? Correlating Observations and Outcomes to Guide Macromolecular Crystallization Experiments. *Crystal Growth & Design*, 11(3), 651–663.
- Lunagariya, N. A., Patel, N. K., Jagtap, S. C., & Bhutani, K. K. (2014). Inhibitors of pancreatic lipase: state of the art and clinical perspectives. *EXCLI Journal*, 13, 897–921.
- MacTavish, H. S. (1995). *Factors affecting yield and composition of floral extract from Boronia megastigma Nees*. University of Tasmania, Australia.
- Madsen, L., Petersen, R. K., & Kristiansen, K. (2005). Regulation of adipocyte differentiation and function by polyunsaturated fatty acids. *Biochimica et Biophysica Acta*, 1740(2), 266–86.
- Madureira, A. R., Campos, D. A., Fonte, P., Nunes, S., Reis, F., Gomes, A. M., Sarmiento, B., & Pintado, M. M. (2015). Characterization of solid lipid nanoparticles produced with carnauba wax for rosmarinic acid oral delivery. *Royal Society of Chemistry Advances*, 5(29), 22665–22673.
- Mayer, M. a, Höcht, C., Puyó, A., & Taira, C. A. (2009). Recent advances in obesity pharmacotherapy. *Current Clinical Pharmacology*, 4(1), 53–61.
- McDougall, G. J., Kulkarni, N. N., & Stewart, D. (2009). Berry polyphenols inhibit pancreatic lipase activity *in vitro*. *Food Chemistry*, 115(1), 193–199.
- McMaster, M. (2005). *LC/MS: A Practical User's Guide*. New Jersey, USA: John Wiley & Sons.
- Mekinić, I. G., Skroza, D., Ljubenković, I., Šimat, V., Možina, S. S., & Katalinić, V. (2014). *In vitro* antioxidant and antibacterial activity of Lamiaceae phenolic extracts: A correlation study. *Food Technology and Biotechnology*, 52(1), 119–127.
- Mendes, A. A., Oliveira, P. C., & de Castro, H. F. (2012). Properties and biotechnological applications of porcine pancreatic lipase. *Journal of Molecular Catalysis B: Enzymatic*, 78, 119–134.
- Miles, A. J., Drechsler, A., Kristan, K., Anderluh, G., Norton, R. S., Wallace, B. A., & Separovic, F. (2008). The effects of lipids on the structure of the eukaryotic cytolysin equinatoxin II: A synchrotron radiation circular dichroism spectroscopic study. *Biochimica et Biophysica Acta - Biomembranes*, 1778(10), 2091–2096.
- Mohamed, E. A. H., Mohamed, A. J., Asmawi, M. Z., Sadikun, A., Ebrika, O. S., & Yam, M. F. (2011). Antihyperglycemic Effect of *Orthosiphon stamineus* Benth Leaves Extract and Its Bioassay-Guided Fractions. *Molecules*, 16(12), 3787–3801.

- Mohamed, G. A., Ibrahim, S. R. M., Elkhayat, E. S., & El Dine, R. S. (2014). Natural anti-obesity agents. *Bulletin of Faculty of Pharmacy, Cairo University*, 52(2), 269–284.
- Mohamed, S. (2014). Functional foods against metabolic syndrome (obesity, diabetes, hypertension and dyslipidemia) and cardiovascular disease. *Trends in Food Science & Technology*, 35(2), 114–128.
- Mohammad, M., Al-Masri, I. M., Issa, A., Khdair, A., & Bustanji, Y. (2013). Inhibition of pancreatic lipase by berberine and dihydroberberine: An investigation by docking simulation and experimental validation. *Medicinal Chemistry Research*, 22(5), 2273–2278.
- Mohamed, E. A. H., Yam, M. F., Ang, L. F., Mohamed, A. J., & Asmawi, M. Z. (2013). Antidiabetic Properties and Mechanism of Action of *Orthosiphon stamineus* Benth Bioactive Sub-fraction in Streptozotocin-induced Diabetic Rats. *Journal of Acupuncture and Meridian Studies*, 6(1), 31–40.
- Mohamed, E. A. H., Siddiqui, M. J. A., Ang, L. F., Sadikun, A., Chan, S. H., Tan, S. C., Asmawi, M. Z., & Yam, M. F. (2012). Potent α -glucosidase and α -amylase inhibitory activities of standardized 50% ethanolic extracts and sinensetin from *Orthosiphon stamineus* Benth as anti-diabetic mechanism. *BMC Complementary and Alternative Medicine*, 12(176), 1-7.
- Moreno, D. A., Ilic, N., Poulev, A., & Raskin, I. (2006). Effects of *Arachis hypogaea* nutshell extract on lipid metabolic enzymes and obesity parameters. *Life Sciences*, 78(24), 2797–803.
- Moro, C. O., & Basile, G. (2000). Obesity and medicinal plants. *Fitoterapia*, 71, 73–82.
- Mukherjee, A. K. (2014). Hydrophobic-hydrophilic interaction in lipase catalytic triad and possibility of a cofactor mediated catalysis. *International Journal of Agricultural and Food Science*, 4(3), 84–89.
- Mukherjee, M. (2003). Human digestive and metabolic lipases—a brief review. *Journal of Molecular Catalysis B: Enzymatic*, 22(5-6), 369–376.
- Murray, K. K., Boyd, R. K., Eberlin, M. N., Langley, G. J., Li, L., Naito, Y., & Tabet, J. C. (2005). IUPAC standard definitions of terms relating to mass spectrometry. *Pure and Applied Chemistry*, 85(7), 1515–1609.
- Narendra, K., Swathi, J., Sowjanya, K. M., & Satya, A. K. (2012). *Phyllanthus niruri*: A Review on its Ethno Botanical, Phytochemical and Pharmacological Profile. *Journal of Pharmacy Research*, 5(9), 4681–4691.
- Ng, R. (2009). *Drugs: From Discovery to Approval* (Second). USA: John Wiley & Sons, Inc.
- Ninomiya, K., Matsuda, H., Shimoda, H., Nishida, N., Kasajima, N., Yoshino,

- T., Morikawa, T., & Yoshikawa, M. (2004). Carnosic acid, a new class of lipid absorption inhibitor from sage. *Bioorganic & Medicinal Chemistry Letters*, 14(8), 1943–1946.
- Nolandt, O. V, Walther, T. H., Roth, S., Bürck, J., & Ulrich, A. S. (2009). Structure analysis of the membrane protein TatCd from the Tat system of *B. subtilis* by circular dichroism. *Biochimica et Biophysica Acta-Biomembrane*, 1788(10), 2238–2244.
- Nussinov, R., & Tsai, C.-J. (2015). The Design of Covalent Allosteric Drugs. *Annual Review of Pharmacology and Toxicology*, 55, 249-267.
- Ohia, S. E., Opere, C. a., LeDay, A. M., Bagchi, M., Bagchi, D., & Stohs, S. J. (2002). Safety and mechanism of appetite suppression by a novel hydroxycitric acid extract (HCA-SX). *Molecular and Cellular Biochemistry*, 238(1-2), 89–103.
- Ohkoshi, E., Miyazaki, H., Kazutoshi, S., Hiroyuki, W., Aruto, Y., & Hiroaki, Y. (2007). Constituents from the Leaves of *Nelumbo nucifera* Stimulate Lipolysis in the White Adipose Tissue of Mice. *Planta Medica*, 73(12), 1255–1259.
- Okoli, C. O., Obidike, I. C., Ezike, A. C., Akah, P. A., & Salawu, O. A. (2011). Studies on the possible mechanisms of antidiabetic activity of extract of aerial parts of *Phyllanthus niruri*. *Pharmaceutical Biology*, 49(3), 248-255.
- Olah, N.-K., Radu, L., Mogosan, C., Hanganu, D., & Gocan, S. (2003). Phytochemical and pharmacological studies on *Orthosiphon stamineus* Benth. (Lamiaceae) hydroalcoholic extracts. *Journal of Pharmaceutical and Biomedical Analysis*, 33, 117–123.
- Ong, H. C., & Norzalina, J. (1999). Malay herbal medicine in Gemencheh, Negri Sembilan, Malaysia. *Fitoterapia*, 70(1), 10–14.
- Ono, Y., Hattori, E., Fukaya, Y., Imai, S., & Ohizumi, Y. (2006). Anti-obesity effect of *Nelumbo nucifera* leaves extract in mice and rats. *Journal of Ethnopharmacology*, 106(2), 238–44.
- Orlov, S., Goncharova, I., & Urbanová, M. (2014). Circular dichroism study of the interaction between mutagens and bilirubin bound to different binding sites of serum albumins. *Spectrochimica Acta - Part A: Molecular and Biomolecular Spectroscopy*, 126, 68–75.
- Özgen, U., Mavi, A., Terzi, Z., Kazaz, C., Asçi, A., Kaya, Y., & Seçen, H. (2011). Relationship between chemical structure and antioxidant activity of luteolin and its glycosides isolated from *Thymus sipyleus* subsp. *sipyleus* var. *sipyleus*. *Records of Natural Products*, 5(1), 12–21.
- Ozturk, H., Ozturk, H., Terzi, E. H., Ozgen, U., Duran, A., & Uygun, I. (2014). Protective effects of Rosmarinic acid against renal ischaemia/reperfusion injury in rats. *Journal of the Pakistan Medical Association*, 64(3), 260–265.

- Padwal, R. (2008). Cetilistat, a new lipase inhibitor for the treatment of obesity. *Current Opinion in Investigational Drugs*, 9(4), 414–21.
- Padwal, R. S., & Majumdar, S. R. (2007). Drug treatments for obesity: orlistat, sibutramine, and rimonabant. *Lancet*, 369(9555), 71–77.
- Pagan, R., Haro, C. O. de, & Sánchez, C. R. (2015). Obesity, job satisfaction and disability at older ages in Europe. *Economics & Human Biology*, 20, 42-54.
- Paithankar, V. V, Raut, K. S., Charde, R. M., & Vyas, J. V. (2011). *Phyllanthus niruri*: A magic Herb. *Research in Pharmacy*, 1(4), 1–9.
- Pan, Y., Wang, K., Huang, S., Wang, H., Mu, X., He, C., Ji, X., Zhang, J., & Huang, F. (2008). Antioxidant activity of microwave-assisted extract of longan (*Dimocarpus longan* Lour.) peel. *Food Chemistry*, 106(3), 1264–1270.
- Pandey, K. B., & Rizvi, S. I. (2009). Plant polyphenols as dietary antioxidants in human health and disease. *Oxidative Medicine and Cellular Longevity*, 2(5), 270–278.
- Panigrahi, G. T., Suthar, M. K., Verma, N., Asthana, S., Tripathi, A., Gupta, S. K., Saxena, J. K., Raisuddin, S., & Das, M. (2015). Investigation of the interaction of anthraquinones of *Cassia occidentalis* seeds with bovine serum albumin by molecular docking and spectroscopic analysis: Correlation to their *in vitro* cytotoxic potential. *Food Research International*, 77, 368–377.
- Park, Y. S., Jung, S. T., Kang, S. G., Heo, B. G., Arancibia-Avila, P., Toledo, F., Drzewiecki, J., Namiesnik, J., & Gorinstein, S. (2008). Antioxidants and proteins in ethylene-treated kiwifruits. *Food Chemistry*, 107(2), 640–648.
- Patel, D. (2015). Pharmacotherapy for the management of obesity. *Metabolism*, 64(11), 1376–1385.
- Paula, A. V, Urioste, D., Santos, J. C., & de Castro, H. F. (2007). Porcine pancreatic lipase immobilized on polysiloxane–polyvinyl alcohol hybrid matrix: catalytic properties and feasibility to mediate synthesis of surfactants and biodiesel. *Journal of Chemical Technology & Biotechnology*, 82(3), 281–288.
- Pemble, C. W., Johnson, L. C., Kridel, S. J., & Lowther, T. W. (2007). Crystal structure of the thioesterase domain of human fatty acid synthase inhibited by Orlistat. *Nature Structural and Molecular Biology*, 14, 704–709.
- Petersen, M. (2003). Rosmarinic acid. *Phytochemistry*, 62(2), 121–125.
- Pi-Sunyer, F.-X., Laferrere, B., Aronne, L. J., & Bray, G. A. (1999). Obesity — A

Modern-Day Epidemic*. *Journal of Clinical Endocrinology and Metabolism*, 84(1), 3–12.

Pitt, J. J. (2009). Principles and Applications of Liquid Chromatography-Mass Spectrometry in Clinical Biochemistry. *The Clinical Biochemist Reviews*, 30(1), 19–34.

Priya, P., Maity, A., Majumdar, S., & Ghosh Dastidar, S. (2015). Interactions between Bcl-xl and its inhibitors: Insights into ligand design from molecular dynamics simulation. *Journal of Molecular Graphics and Modelling*, 59, 1–13.

Pujiyanto, S., Lestari, Y., Suwanto, A., Budiarti, S., & Darusman, L. K. (2012). Alpha-glucosidase inhibitor activity and characterization of endophytic Actinomycetes isolated from some Indonesian diabetic medicinal plants. *International Journal of Pharmacy and Pharmaceutical Sciences*, 4, 327–333.

Pushparaj, P. N., Tan, B. K. H., & Tan, C. H. (2001). The mechanism of hypoglycemic action of the semi-purified fractions of *Averrhoa bilimbi* in streptozotocin-diabetic rats. *Life Sciences*, 70(5), 535–547.

Qi, J., Li, Y., Yokoyama, W., Majeed, H., Masamba, K. G., Zhong, F., & Ma, J. (2015). Cellulosic fraction of rice bran fibre alters the conformation and inhibits the activity of porcine pancreatic lipase. *Journal of Functional Foods*, 19, 39–48.

Raghav, P. K., Verma, Y. K., & Gangenahalli, G. U. (2012). Molecular dynamics simulations of the Bcl-2 protein to predict the structure of its unordered flexible loop domain. *Journal of Molecular Modeling*, 18(5), 1885–906.

Raina, A. P., Gupta, V., Sivaraj, N., & Dutta, M. (2013). *Andrographis paniculata* (Burm. f.) Wall. ex Nees (kalmegh), a traditional hepatoprotective drug from India. *Genetic Resources and Crop Evolution*, 60(3), 1181–1189.

Rajeshkumar, N. V., Joy, K. L., Kuttan, G., Ramsewak, R. S., Nair, M. G., & Kuttan, R. (2002). Antitumour and anticarcinogenic activity of *Phyllanthus amarus* extract. *Journal of Ethnopharmacology*, 81(1), 17–22.

Raussens, V., Ruyschaert, J.-M., & Goormaghtigh, E. (2003). Protein concentration is not an absolute prerequisite for the determination of secondary structure from circular dichroism spectra: a new scaling method. *Analytical Biochemistry*, 319(1), 114–121.

Rayalam, S., Della-Fera, M. A., & Baile, C. A. (2008). Phytochemicals and regulation of the adipocyte life cycle. *Journal of Nutritional Biochemistry*, 19(11), 717–726.

Redfern, J., Kinninmonth, M., Verran, J., & Dariel, B. (2014). Using Soxhlet

Ethanol Extraction to Produce and Test Plant Material (Essential Oils) for Their Antimicrobial Properties. *Journal of Microbiology and Biology Education*, 15(1), 45–46.

Redinger, R. N. (2009). Fat storage and the biology of energy expenditure. *Translational Research*, 154(2), 52–60.

Rethinam, P., & Sivaraman, K. (2006). Noni (*Morinda citrifolia* L) the Miracle Fruit - A Holistic Review. *International Journal of Noni Research*, 2(1-2), 1–34.

Riederer, M., & Muller, C. (2008). *Annual Plant Reviews, Biology of the Plant Cuticle*. (M. Riederer & C. Muller, Eds.) (Volume 23). USA: Wiley-Blackwell Publishing.

Rivlin, R. S. (1975). Therapy of obesity with hormones. *The New England Journal of Medicine*, 292(1), 26–29.

Robertson, K. (2006). Surgery for obesity. *Medicine*, 34(12), 521–523.

Rohman, A., & Man, Y. B. C. (2010). Fourier transform infrared (FTIR) spectroscopy for analysis of extra virgin olive oil adulterated with palm oil. *Food Research International*, 43(3), 886–892.

Rostagno, M., & Prado, J. (2013). *Natural Product Extraction: Principles and Applications*. Cambridge, UK: Royal Society of Chemistry.

Roy, A., Rv, G., & Lakshmi, T. (2011). *Averrhoa bilimbi* Linn – Nature's Drug Store-A Pharmacological Review Scientific classification. *International Journal of Drug Development and Research*, 3(3), 101–106.

Rubatzky, V., & Yamaguchi, M. (2012). *World Vegetables: Principles, Production, and Nutritive Values* (Second). USA: Springer Science & Business Media.

Saeed, N., Khan, M. R., & Shabbir, M. (2012). Antioxidant activity, total phenolic and total flavonoid contents of whole plant extracts *Torilis leptophylla* L. *BMC Complementary and Alternative Medicine*, 12(1), 221.

Sánchez-Campillo, M., Gabaldon, J. A., Castillo, J., Benavente-García, O., De/ Baño, M. J., Alcaraz, M., Vicente, V., Alvarez, N. & Lozano, J. A. (2009). Rosmarinic acid, a photo-protective agent against UV and other ionizing radiations. *Food and Chemical Toxicology*, 47(2), 386–392.

Schlichting, I. (2005). X-ray crystallography of protein-ligand interactions. In *Protein-Ligand Interactions: Methods and Applications* (Vol. 305, pp. 155–165). Tortowa, New Jersey: Humana Press Inc.

Sedighi, R., Zhao, Y., Yerke, A., & Sang, S. (2015). Preventive and protective properties of rosemary (*Rosmarinus officinalis* L.) in obesity and diabetes mellitus of metabolic disorders: a brief review. *Current Opinion in Food*

Science, 2, 58–70.

- Segel, I. H. (1976). Enzyme Inhibition. In *Biochemical Calculations – How to Solve Mathematical Problems in General Biochemistry* (Second, pp. 246–265). USA: John Wiley & Sons, Inc.
- Segura, R. L., Betancor, L., Palomo, J. M., Hidalgo, A., Fernández-Lorente, G., Terreni, M., Mateo, C., Cortés A., Fernández-Lafuente, R., & Guisán, J. M. (2006). Purification and identification of different lipases contained in PPL commercial extracts: A minor contaminant is the main responsible of most esterase activity. *Enzyme and Microbial Technology*, 39(4), 817–823.
- Selvaraj, G., Kaliyamurthi, S., & Thirugnanasambandam, R. (2015). Influence of *Rhizophora apiculata* Blume extracts on α -glucosidase: Enzyme kinetics and molecular docking studies. *Biocatalysis and Agricultural Biotechnology*, 4(4), 653–660.
- Sharma, D., Sharma, B., & Shukla, A. K. (2011). Biotechnological Approach of Microbial Lipase: A Review. *Biotechnology*, 10(1), 23–40.
- Sharma, M., Kumar, V., & Pundir, C. S. (2008). Immobilization of porcine pancreas lipase onto free and affixed arylamine glass beads and its application in removal of oil stains. *Indian Journal of Biotechnology*, 7(3), 328–332.
- Sharma, N., Sharma, V. K., & Seo, S. Y. (2005). Screening of some medicinal plants for anti-lipase activity. *Journal of Ethnopharmacology*, 97(3), 453–456.
- Sharmila, R., & Manoharan, S. (2012). Anti-tumor activity of rosmarinic acid in 7, 12-dimethylbenz (a) anthracene (DMBA) induced skin carcinogenesis in Swiss albino mice. *Indian Journal of Experimental Biology*, 50(3), 187–194.
- Shekarchi, M., Hajimehdipoor, H., Saeidnia, S., Gohari, A. R., & Hamedani, M. P. (2012). Comparative study of rosmarinic acid content in some plants of Labiatae family. *Pharmacognosy Magazine*, 8(29), 37–41.
- Shen, D., Pan, M.-H., Wu, Q.-L., Park, C.-H., Juliani, H. R., Ho, C.-T., & Simon, J. E. (2010). LC-MS Method for the Simultaneous Quantitation of the Anti-inflammatory Constituents in Oregano (*Origanum* Species). *Journal of Agricultural and Food Chemistry*, 58(12), 7119–7125.
- Sherma, J., & Bernard, F. (2003). *Handbook of Thin-Layer Chromatography* (Third). New York, USA: CRC Press.
- Shetti, A. A., Sanakal, R. D., & Kaliwal, B. B. (2012). Antidiabetic effect of ethanolic leaf extract of *Phyllanthus amarus* in alloxan induced diabetic mice. *Asian Journal of Plant Science and Research*, 2(1), 11–15.

- Shi, Y., & Burn, P. (2004). Lipid metabolic enzymes: emerging drug targets for the treatment of obesity. *Nature Reviews Drug Discovery*, 3(8), 695–710.
- Shimoda, H., Seki, E., & Aitani, M. (2006). Inhibitory effect of green coffee bean extract on fat accumulation and body weight gain in mice. *BMC Complementary and Alternative Medicine*, 6(9), 1-9.
- Sigma-Aldrich. (2015). Sigma-Aldrich technical information-Lipase from porcine pancreas. Retrieved December 3, 2015, from <http://www.sigmaaldrich.com/catalog/product/sigma/l3126?lang=en®ion=MY>
- Silalahi, M., Nisyawati, Walujo, E. B., Supriatna, J., & Mangunwardoyo, W. (2015). The Local Knowledge of Medicinal Plants Trader and Diversity of Medicinal Plants in The Kabanjahe Traditional Market, North Sumatra, Indonesia. *Journal of Ethnopharmacology*, 175, 432–443.
- Siligardi, G., Hussain, R., Patching, S. G., & Phillips-Jones, M. K. (2014). Ligand- and drug-binding studies of membrane proteins revealed through circular dichroism spectroscopy. *Biochimica et Biophysica Acta-Biomembranes*, 1838(1 Part A), 34–42.
- Siligardi, G., Panaretou, B., Meyerc, P., Singh, S., Woolfson, D. N., Piper, P. W., Pearl, L. H., & Prodromou, C. (2002). Regulation of Hsp90 ATPase activity by the Co-chaperone Cdc37p/p50 cdc37. *Journal of Biological Chemistry*, 277(23), 20151–20159.
- Silverman, R. B. (2000). Enzyme Kinetics. In *The Organic Chemistry of Enzyme-catalyzed Reactions* (p. 570). USA: Academic Press.
- Sinha, S. K., & Saiz, L. (2014). Determinants of protein–ligand complex formation in the thyroid hormone receptor α : A molecular dynamics simulation study. *Computational and Theoretical Chemistry*, 1038, 57–66.
- Sinkovits, A. F., Bryksa, B. C., Tanaka, T., & Yada, R. Y. (2007). Understanding the structure–function role of specific catalytic residues in a model food related enzyme: Pepsin. *Enzyme and Microbial Technology*, 40(5), 1175–1180.
- Sirinamarattana, A., Bavovada, R., Pengsuparp, T., & Pongthananikorn, S. (2010). Pancreatic Lipase Inhibitory Activity of Thai Medicinal Plants. *Journal of Pharmacy Research*, 3(10), 2402–2403.
- Slanc, P., Doljak, B., Kreft, S., Lunder, M., Janes, D., & Strukelj, B. (2009). Screening of selected food and medicinal plant extracts for pancreatic lipase inhibition. *Phytotherapy Research*, 23, 874–877.
- Soares, J. R., Dinis, T. C., Cunha, A. P., & Almeida, L. M. (1997). Antioxidant activities of some extracts of *Thymus zygis*. *Free Radical Research*, 26(5), 469–78.
- Sotnikova, R., Okruhlicova, L., Vlkovicova, J., Navarova, J., Gajdacova, B.,

- Pivackova, L., Fialova, S., & Krenek, P. (2013). Rosmarinic acid administration attenuates diabetes-induced vascular dysfunction of the rat aorta. *The Journal of Pharmacy and Pharmacology*, 65(5), 713–723.
- Sousa, S. F., Ribeiro, A. J. M., Coimbra, J. T. S., Neves, R. P. P., Martins, S. A., Moorthy, N. S. H. N., Fernandes, P. A., & Ramos, M. J. (2013). Protein-Ligand Docking in the New Millennium – A Retrospective of 10 Years in the Field. *Current Medicinal Chemistry*, 20(18), 2296–2314.
- Sriplang, K., Adisakwattana, S., Rungsipat, A., & Yibchok-Anun, S. (2007). Effects of *Orthosiphon stamineus* aqueous extract on plasma glucose concentration and lipid profile in normal and streptozotocin-induced diabetic rats. *Journal of Ethnopharmacology*, 109(3), 510–4.
- Stansbury, J. (2014). Rosmarinic Acid as a Novel Agent in the Treatment of Allergies and Asthma*. *Journal of Restorative Medicine*, 3, 121–126.
- Stehfest, K., Boese, M., Kerns, G., Piry, A., & Wilhelm, C. (2004). Fourier transform infrared spectroscopy as a new tool to determine rosmarinic acid in situ. *Journal of Plant Physiology*, 161(2), 151–156.
- Stunkard, A. J. (1996). Current views on obesity. *The American Journal of Medicine*, 100(2), 230–236.
- Sugiyama, H., Akazome, Y., Shoji, T., Yamaguchi, A., Yasue, M., Kanda, T., & Ohtake, Y. (2007). Oligomeric procyanidins in apple polyphenol are main active components for inhibition of pancreatic lipase and triglyceride absorption. *Journal of Agricultural and Food Chemistry*, 55(11), 4604–4609.
- Sultana, B., Anwar, F., & Przybylski, R. (2007). Antioxidant activity of phenolic components present in barks of *Azadirachta indica*, *Terminalia arjuna*, *Acacia nilotica*, and *Eugenia jambolana* Lam. trees. *Food Chemistry*, 104(3), 1106–1114.
- Svensson, F. R., Lincoln, P., Nordén, B., & Esbjörner, E. K. (2011). Tryptophan orientations in membrane-bound gramicidin and melittin—a comparative linear dichroism study on transmembrane and surface-bound peptides. *Biochimica et Biophysica Acta- Biomembranes*, 1808(1), 219–228.
- Swanson, A. (2015). Ranking of world's most obese nations may surprise you. Retrieved December 1, 2015, from <http://www.pressherald.com/2015/04/23/list-of-worlds-most-obese-nations-may-surprise-you/>
- Swarup, V., Ghosh, J., Ghosh, S., Saxena, A., & Basu, A. (2007). Antiviral and anti-inflammatory effects of rosmarinic acid in an experimental murine model of *Japanese encephalitis*. *Antimicrobial Agents and Chemotherapy*, 51(9), 3367–3370.
- Tajuddin Abd. Manap, Agricultural Senior Assistant Officer, Institute of Bioscience, Universiti Putra Malaysia, pers. comm. 9 February, 2011.

- Takeda, Y., Matsumoto, T., Terao, H., Shingu, T., Futatsuishi, Y., Nohara, T., & Kajimoto, T. (1993). Orthosiphon D and E, minor diterpenes from *Orthosiphon stamineus*. *Phytochemistry*, 33(2), 411–415.
- Tan, B. K. H., Tan, C. H., & Pushparaj, P. N. (2005). Anti-diabetic activity of the semi-purified fractions of *Averrhoa bilimbi* in high fat diet fed-streptozotocin-induced diabetic rats. *Life Sciences*, 76(24), 2827–2839.
- Terra, X., Montagut, G., Bustos, M., Llopiz, N., Ardèvol, A., Bladé, C., Fernáandez-Larrea, J., Pujadas, G., Salvadó, J., Arola, L., & Blay, M. (2009). Grape-seed procyanidins prevent low-grade inflammation by modulating cytokine expression in rats fed a high-fat diet. *Journal of Nutritional Biochemistry*, 20(3), 210–218.
- Thielecke, F., & Boschmann, M. (2009). The potential role of green tea catechins in the prevention of the metabolic syndrome - a review. *Phytochemistry*, 70(1), 11–24.
- Thompson, W. G., Cook, D. A., Clark, M. M., Bardia, A., & Levine, J. A. (2007). Treatment of obesity. *Mayo Clinic Proceedings*, 82(1), 93–101.
- Thomson, A. B. R., De Pover, A., Keelan, M., Jarocka-Cyrta, E., & Clandinin, M. T. (1997). *Lipases Part B: Enzyme Characterization and Utilization. Methods in Enzymology* (Vol. 286). Elsevier.
- Tiwari, P., Kumar, B., Mandeep, K., Kaur, G., & Kaur, H. (2011). Phytochemical screening and Extraction: A Review. *Internationale Pharmaceutica Scientia*, 1(1), 98–106.
- Valdiani, A., Kadir, M. A., Tan, S. G., Talei, D., Abdullah, M. P., & Nikzad, S. (2012). Nain-e havandi *Andrographis paniculata* present yesterday, absent today: A plenary review on underutilized herb of Iran's pharmaceutical plants. *Molecular Biology Reports*, 39(5), 5409–5424.
- van Heerden, F. R. (2008). *Hoodia gordonii*: A natural appetite suppressant. *Journal of Ethnopharmacology*, 119(3), 434–437.
- van Tilbeurgh, H., Eglhoff, M. P., Martinez, C., Rugani, N., Verger, R., & Cambillau, C. (1993). Interfacial activation of the lipase-procolipase complex by mixed micelles revealed by X-ray crystallography. *Nature*, 362(6423), 814–820.
- Vasconcelos, D. N. De, & Ximenes, V. F. (2015). Albumin-induced circular dichroism in Congo red: Applications for studies of amyloid-like fibril aggregates and binding sites. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 150, 321–330.
- Velioglu, Y. S., Mazza, G., Gao, L., & Oomah, B. D. (1998). Antioxidant Activity and Total Phenolics in Selected Fruits, Vegetables, and Grain Products. *Journal of Agricultural and Food Chemistry*, 46(10), 4113–4117.

- Venkatachalam, K., Gunasekaran, S., Jesudoss, V. A. S., & Namasivayam, N. (2013). The effect of rosmarinic acid on 1,2-dimethylhydrazine induced colon carcinogenesis. *Experimental and Toxicologic Pathology*, 65(4), 409–18.
- Venyaminov, S. Y., & Vassilenko, K. S. (1994). Determination of Protein Tertiary Structure Class from Circular Dichroism Spectra. *Analytical Biochemistry*, 222(1), 176-184.
- Verger, R., De Haas, G. H., Sarda, L., & Desnuelle, P. (1969). Purification from porcine pancreas of two molecular species with lipase activity. *Biochimica et Biophysica Acta - Protein Structure*, 88(2), 272–282.
- Walter, C. P., Bleske, B. E., & Dorsch, M. P. (2014). Pharmacotherapy for weight loss: the cardiovascular effects of the old and new agents. *Journal of Clinical Pharmacy and Therapeutics*, 39, 1–10.
- Wan Mohamud, W. N., Musa, K. I., Md Khir, A. S., & Ismail, A. A.-S. (2011). Prevalence of Overweight and Obesity Among Adult Malaysians: An Update. *Asia Pacific Journal of Clinical Nutrition*, 20(1), 35–41.
- Wang, J., Xu, H., Jiang, H., Du, X., Sun, P., & Xie, J. (2012). Neurorescue effect of rosmarinic acid on 6-hydroxydopamine-lesioned nigral dopamine neurons in rat model of Parkinson's disease. *Journal of Molecular Neuroscience*, 47(1), 113–9.
- Weibel, E. K., Hadvary, P., Hochuli, E., Kupfer, E., & Lengsfeld, H. (1987). Lipstatin, an inhibitor of pancreatic lipase, produced by *Streptomyces toxytricini*. I. Producing organism, fermentation, isolation and biological activity. *The Journal of Antibiotics*, 40(8), 1081–5.
- Weigle, D. S. (2003). Pharmacological therapy of obesity: past, present, and future. *The Journal of Clinical Endocrinology and Metabolism*, 88(6), 2462–9.
- Weintraub, M. (1992). Long-term weight control study: Conclusions. *Clinical Pharmacology and Therapeutics*, 51(5), 642–646.
- WHO. (2015a). Obesity and overweight: Fact sheet N°311. Retrieved December 3, 2015, from <http://www.who.int/mediacentre/factsheets/fs311/en/>
- WHO. (2015b). WHO, Global Health Observatory Data Repository. Retrieved November 11, 2015, from <http://www.who.int/gho/database/en/>
- WHO. (2013). *Traditional Medicine Strategy, 2014- 2023. World Health organization.*
- Widyawati, T., Yusoff, N., Asmawi, M., & Ahmad, M. (2015). Antihyperglycemic Effect of Methanol Extract of *Syzygium polyanthum* (Wight.) Leaf in

- Streptozotocin-Induced Diabetic Rats. *Nutrients*, 7(9), 7764–7780.
- Williams, M., & Daviter, T. (2013). *Protein-Ligand Interactions: Methods and Applications*. (Williams, M., & Daviter, T., Eds.) (Second). New York, USA: Humana Press.
- Wong, H., & Schotz, M. C. (2002). The lipase gene family. *Journal of Lipid Research*, 43(7), 993–999.
- Wong, B. Y., Tan, C. P., & Ho, C. W. (2013). Effect of solid-to-solvent ratio on phenolic content and antioxidant capacities of “Dukung Anak” (*Phyllanthus niruri*). *International Food Research Journal*, 20(1), 325–330.
- Wu, X., He, W., Zhang, H., Li, Y., Liu, Z., & He, Z. (2014). Acteoside: a lipase inhibitor from the Chinese tea *Ligustrum purpurascens* kudingcha. *Food Chemistry*, 142, 306–10.
- Wu, X., Wu, H., Liu, M., Liu, Z., Xu, H., & Lai, F. (2011). Analysis of binding interaction between (–)-epigallocatechin (EGC) and β -lactoglobulin by multi-spectroscopic method. *Spectrochimica Acta - Part A: Molecular and Biomolecular Spectroscopy*, 82(1), 164–168.
- Yadav, K., & Krishnan, a. (2008). Changing patterns of diet, physical activity and obesity among urban, rural and slum populations in north India. *Obesity Reviews*, 9(4), 400–408.
- Yamamoto, M., Shimura, S., Itoh, Y., Ohsaka, T., Egawa, M., & Inoue, S. (2000). Anti-obesity effects of lipase inhibitor CT-II, an extract from edible herbs, Nomame Herba, on rats fed a high-fat diet. *International Journal of Obesity*, 24(6), 758–764.
- Yang, E.-J., Ku, S.-K., Lee, W., Lee, S., Lee, T., Song, K.-S., & Bae, J.-S. (2013). Barrier protective effects of rosmarinic acid on HMGB1-induced inflammatory responses *in vitro* and *in vivo*. *Journal of Cellular Physiology*, 228(5), 975–82.
- Yang, J., Della-fera, M. A., Hartzell, D. L., Nelson-Dooley, C., Hausman, D. B., & Baile, C. A. (2006). Esculetin Induces Apoptosis and Inhibits Adipogenesis in 3T3-L1 Cells. *Obesity*, 14(10), 1691–1699.
- Yoon, S. S., Rhee, Y. H., Lee, H. J., Lee, E. O., Lee, M. H., Ahn, K. S., Lim, H. T., & Kim, S. H. (2008). Uncoupled protein 3 and p38 signal pathways are involved in anti-obesity activity of *Solanum tuberosum* L. cv. Bora Valley. *Journal of Ethnopharmacology*, 118(3), 396–404.
- Yoshikawa, M., Shimoda, H., Nishida, N., Takada, M., & Matsuda, H. (2002). *Salacia reticulata* and its polyphenolic constituents with lipase inhibitory and lipolytic activities have mild antiobesity effects in rats. *The Journal of Nutrition*, 132(7), 1819–1824.
- Yuliana, N. D., Rosa, D., & Korthout. H. V. (2009). Screening of anti-obesity

- compounds from *Orthosiphon stamineus* with the CB1 assay. *Obesity Facts*, 2, 250.
- Zakaria, Z. A., Balan, T., Suppaiah, V., Ahmad, S., & Jamaludin, F. (2014). Mechanism(s) of action involved in the gastroprotective activity of *Muntingia calabura*. *Journal of Ethnopharmacology*, 151(3), 1184–93.
- Zeng, H., Yang, R., Liang, H., & Qu, L. (2015). Molecular interactions of flavonoids to pepsin: Insights from spectroscopic and molecular docking studies. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 151, 576–590.
- Zheng, C. D., Duan, Y. Q., Gao, J. M., & Ruan, Z. G. (2010). Screening for Anti-lipase Properties of 37 Traditional Chinese Medicinal Herbs. *Journal of the Chinese Medical Association*, 73(6), 319–324.
- Zhou, Y.-J., Hu, C.-L., Wang, N., Zhang, W.-W., & Yu, X.-Q. (2013). Purification of porcine pancreatic lipase by aqueous two-phase systems of polyethylene glycol and potassium phosphate. *Journal of Chromatography. B, Analytical Technologies in the Biomedical and Life Sciences*, 926, 77–82.
- Zhu, F., Asada, T., Sato, A., Koi, Y., Nishiwaki, H., & Tamura, H. (2014). Rosmarinic acid extract for antioxidant, antiallergic, and α -glucosidase inhibitory activities, isolated by supramolecular technique and solvent extraction from *Perilla* leaves. *Journal of Agricultural and Food Chemistry*, 62(4), 885–892.