



UNIVERSITI PUTRA MALAYSIA

***EVALUATION OF CURCUMIN DERIVATIVES AS NEW
CYCLOOXYGENASE-2 INHIBITORS VIA *In Silico* AND *In Vitro*
ANALYSES***

MOHAMMAD NAZRI BIN ABDUL BAHARI

FBSB 2016 7



**EVALUATION OF CURCUMIN DERIVATIVES AS NEW
CYCLOOXYGENASE-2 INHIBITORS VIA *In Silico* AND *In Vitro* ANALYSES**

By

MOHAMMAD NAZRI BIN ABDUL BAHARI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Master of Science**

April 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

EVALUATION OF CURCUMIN DERIVATIVES AS NEW CYCLOOXYGENASE-2 INHIBITORS VIA *In Silico* AND *In Vitro* ANALYSES

By

MOHAMMAD NAZRI BIN ABDUL BAHARI

April 2016

Chairman : Syahida Ahmad, PhD
Faculty : Biotechnology and Biomolecular Sciences

Prostaglandin E₂ (PGE₂) is one of the lipid mediators of inflammation. Chronic inflammation drives overproduction of PGE₂ that leads to development of chronic inflammatory diseases. PGE₂ is synthesized by cyclooxygenase (COX) enzyme that exists in isoforms of COX-1, which is constitutively expressed; and COX-2, which is expressed upon induction. Non-steroidal anti-inflammatory drugs (NSAIDs) inhibit COXs to control excessive production of PGE₂ during inflammation but, most of commercialized NSAIDs selectively inhibit COX-1 or being non-selective which compensate for limitations and detrimental side effects of the medicine. Hence, deciphering the mechanisms of selectively inhibiting COX-2 is of great interest. Curcumin was known as remedy to treat the inflammatory-related diseases, but suffers from poor bioavailability and instability. Synthesis of curcumin derivatives was carried out to overcome the limitations. Thus, the objectives of this study are to investigate the effects of 43 curcumin derivatives towards activated cellular PGE₂ production and COX's activity, as well as to understand its mechanism of actions *in silico* and *in vitro*. In this study, effects of curcumin derivatives on PGE₂ production in murine macrophage (RAW264.7) cells which was stimulated by combination of interferon-gamma (IFN- γ) and lipopolysaccharide (LPS), were evaluated using immunoassay procedures. Quantitative structure-activity relationship (QSAR) analysis was performed to correlate between the structure and PGE₂ inhibition activity of curcumin derivatives. Enzymatic assay and molecular docking analysis were performed to decipher the mechanism of inhibition on COX activity by curcumin derivatives. Effects of active curcumin derivatives on gene expression of COX-1 and COX-2 were also determined. Results demonstrated that 3 out of 43 compounds significantly inhibited PGE₂ production in IFN- γ /LPS-stimulated RAW264.7 cells dose-dependently which were 2,6-bis(2-fluorobenzylidene)cyclohexanone (compound 25), 2,6-bis(4-fluorobenzylidene)cyclohexanone (compound 27), and 2,5-bis(3,4,5-trimethoxybenzylidene)cyclopentanone (compound 43) with IC₅₀ values of 6.15 ± 0.48 μ M, 5.78 ± 1.67 μ M and 12.15 ± 1.88 μ M respectively which were higher than that of curcumin. Furthermore, these three compounds were not toxic to the cells (cytotoxicity IC₅₀>500 μ M). The PGE₂ inhibitory effect was contributed by the suppression of the IFN- γ /LPS-stimulated COX-2 gene expression, without affecting the phorbol myristate

acetate (PMA)-stimulated COX-1 gene expression in RAW264.7 cells by these three compounds. Arene substitution patterns and substituents of electron withdrawing groups may contribute to the PGE₂ inhibition activity of the compounds. Besides, QSAR study recommended that positive contribution of lipophilicity and numbers of rotatable bonds, and negative contribution of kappa_2 descriptor of the compounds were crucial for their anti-inflammatory properties. The enzymatic assay showed that most curcumin derivatives tested selectively inhibited COX-1 activity rather than COX-2. However, compounds 25 and 43 selectively inhibited COX-2, unlike compound 27 which favours towards COX-1 activity. Moreover, docking study revealed that compounds 25 and 43 interacted with COX's active site receptors that favour towards COX-2 inhibition, while Arg120, His90, Phe518 and Arg513 are important receptors involved in COX-2 inhibition, while Arg120 and Ser530 are important receptors in COX-1 inhibition. In conclusion, the experimental data have provided mechanistic insights into properties of compounds 25, 27, and 43 as COX-inhibitors. Compounds 25 and 43 could be potential lead compounds for development of new COX-2 selective inhibitors.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

**PENILAIAN TERHADAP TERBITAN KURKUMIN SEBAGAI PERENCAT
SIKLOOKSIGENASE-2 YANG BAHARU MELALUI ANALISIS *In Silico* DAN
*In Vitro***

Oleh

MOHAMMAD NAZRI BIN ABDUL BAHARI

April 2016

Pengerusi: Syahida Ahmad, PhD
Fakulti: Bioteknologi dan Sains Biomolekul

Prostaglandin E₂ (PGE₂) merupakan satu daripada pengantara lipid bagi radang. Keradangan kronik telah mendorong lebih pengeluaran PGE₂ yang membawa kepada perkembangan penyakit radang kronik. PGE₂ disintesis oleh enzim siklooksigenase (COX) yang wujud dalam isoform COX-1, yang diekspres sebagai penyelenggara; dan COX-2 yang terinduksi. Dadah anti-radang bukan steroid (NSAIDs) ialah dadah yang merencat COX-2 untuk mengawal lebih pengeluaran PGE₂ semasa radang, tetapi NSAIDs komersil kebanyakannya secara terpilih merencat COX-1 atau tidak berdaya memilih yang mana memampas kepada pembatasan dan kesan sampingan yang memudaratkan oleh ubat tersebut. Oleh itu, dengan mentafsirkan mekanisme yang berdaya memilih bagi merencatkan COX-2 adalah menjadi keutamaan. Kurkumin telah dikenalpasti sebagai ubat bagi merawat penyakit-penyakit berkaitan keradangan, tetapi menderita daripada bioketersediaan yang lemah dan ketidakstabilan. Sintesis terbitan kurkumin telah dilakukan bagi mengatasi keterbatasannya. Jadi, objektif kajian ini adalah bagi menyiasat kesan 43 terbitan kurkumin terhadap penghasilan PGE₂ sel teraktif dan aktiviti COX, serta bagi memahami mekanisme tindakannya *in silico* dan *in vitro*. Dalam kajian ini, kesan terbitan kurkumin terhadap penghasilan PGE₂ dalam sel makrofaj murin (RAW 264.7) yang dirangsang oleh kombinasi interferon-gama (IFN- γ) dan lipopolisakarida (LPS), dinilai melalui prosedur imunoasai. Analisis hubungan struktur-aktiviti kuantitatif (QSAR) telah dilakukan untuk mengaitkan antara struktur dan aktiviti perencatan PGE₂ oleh terbitan kurkumin. Asai enzim dan analisis mengedok molekul dilakukan bagi mentafsir mekanisme perencatan aktiviti COX oleh terbitan kurkumin. Kesan terbitan kurkumin yang aktif terhadap ekspresi gen COX-1 dan COX-2 juga ditentukan. Keputusan menunjukkan 3 daripada 43 sebatian merencat penghasilan PGE₂ dalam sel yang dirangsang IFN- γ /LPS dengan ketara mengikut dos iaitu 2,6-bis(2-fluorobenzilidena)sikloheksanon (sebatian 25), 2,6-bis(4-fluorobenzilidena)sikloheksanon (sebatian 27), dan 2,5-bis(3,4,5-trimetoksibenzilidena)siklopentanon (sebatian 43) dengan nilai IC₅₀ masing-masing 6.15 \pm 0.48 μ M, 5.78 \pm 1.67 μ M dan 12.15 \pm 1.88 μ M yang mana lebih tinggi berbanding kurkumin. Tambahan pula, tiga sebatian ini tidak toksik kepada sel

(kesitotoksian $IC_{50} > 500 \mu M$). Kesan perencatan PGE_2 disumbangkan oleh penindasan gen COX-2 yang diransang IFN- γ /LPS, tanpa memberi kesan terhadap ekspresi gen COX-1 yang diransang forbol miristat asetat dalam sel RAW264.7 oleh tiga sebatian tersebut. Pola penggantian arin dan kumpulan penarikan electron mungkin menyumbang kepada aktiviti merencat PGE_2 sebatian tersebut. Selain itu, kajian QSAR telah menganjurkan bahawa sumbangan positif lipofilik dan nombor ikatan boleh putar, dan juga sumbangan negatif penerang kappa_2 sebatian tersebut adalah penting untuk sifat anti-radangnya. Asai enzim menunjukkan kebanyakan terbitan kurkumin yang diuji dengan terpilih merencat aktiviti COX-1 berbanding COX-2. Bagaimanapun, sebatian 25 dan 43 dengan terpilih merencat COX-2, tidak seperti sebatian 27 yang cenderung kepada aktiviti COX-1. Selain itu, kajian mengedok mendedahkan bahawa sebatian 25 dan 43 berinteraksi dengan reseptor tapak aktif COX yang cenderung kepada perencatan COX-2. Arg120, His90, Phe518, dan Arg513 adalah reseptor penting yang terlibat dalam perencatan COX-2, manakala Arg120 serta Ser530 adalah reseptor penting dalam perencatan COX-1. Kesimpulannya, data eksperimen ini telah menyediakan pemahaman mekanisme bagi ciri-ciri sebatian 25, 27 dan 43 sebagai perencat COX. Sebatian 25, dan 43 boleh menjadi sebatian utama yang berpotensi untuk pembangunan perencat COX-2 yang baharu.

ACKNOWLEDGEMENTS

First and foremost, I would like to express my thankful to ALLAH s.w.t for all His blessings that empower me to complete this Master's project and thesis successfully. There are plentiful individuals towards whom I owe sincere gratitude and without their contributions, this study could not have been completed.

I would like to thank my supervisor, Dr. Syahida Ahmad for giving me the opportunities to explore the wonder of sciences. I am driven by her enthusiasm in research and I am thankful that she has always positively encouraged me to come out with my own ideas in performing this project. Her expert guidance and mentorship at all levels have made my thesis completion possible. My heartfelt gratitude is also extended to my co-supervisors, Assoc. Prof. Dr. Faridah Abas and Dr. Lam Kok Wai. With their support for the project, I have benefited greatly from their insightful knowledge and advices.

Nonetheless, I would like to express my appreciations to all my labmates especially Naimah, Amiza, Atika, Najwa, Madam Yu, Yakubu and Ibrahim in Laboratory of Drug Discovery, FBSB for their willingness to share knowledge with me. Special thanks to research officer, Dr Tan Sheau Wei from Laboratory of Vaccine and Immunotherapeutic, IBS for her expert guidance which helped me a lot in carrying out Real Time PCR from the start to its completion.

I would also like to thank Graduate Research Fellowship (UPM) and MyMaster Scholarship (Ministry of Education Malaysia) for funding me during my Master of Science programme. I am beyond grateful to my dear mother, Hasnah binti Mohamad as well as my parents in law, Mohd Sakeh bin Abd Jabar and Badariah binti Jaafar whom have always been proud of me and believe in me. May Allah grant tranquility to my late father, Abdul Bahari bin Abdul Rani whom I have admired most. My love and pray to my family members whom have always being courteous, supportive and thoughtful towards me. Their endless love, understanding, patience and sacrifices have been an enormous source of strength and inspiration for me. May Allah bless them.

Ultimately, I would like to dedicate this thesis to my other half, Nurshafika binti Mohd Sakeh. She has inspired me with great patience, responsibility and honesty in everything I do. She has never failed to lift up and polishes all the good values in me. Her intense love, support, motivation, ideas, humor and limitless encouragement from the very start of this project have made the completion of this thesis a memorable, painless one. May Allah grant her every happiness she deserves till Jannah.

I certify that a Thesis Examination Committee has met on 8 April 2016 to conduct the final examination of Mohammad Nazri bin Abdul Bahari on his thesis entitled "Evaluation of Curcumin Derivatives as New Cyclooxygenase-2 Inhibitors via *In Silico* and *In Vitro* Analyses" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Noor Azmi Shaharuddin, PhD

Senior Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Normi Mohd Yahaya, PhD

Senior Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Internal Examiner)

Malina Jasamai, PhD

Associate Professor
National University of Malaysia
Malaysia
(External Examiner)



ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 23 August 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Syahida Ahmad, PhD

Senior Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Faridah Abas, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

Lam Kok Wai, PhD

Senior Lecturer
Faculty of Pharmacy
University Kebangsaan Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Mohammad Nazri bin Abdul Bahari (GS31228)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) rules 2003 (revision 2012 – 2013) are adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee:

Dr. Syahida Ahmad

Signature: _____

Name of Member
of Supervisory
Committee:

Associate Professor Dr. Faridah Abas

Signature: _____

Name of Member
of Supervisory
Committee:

Dr. Lam Kok Wai

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF APPENDICES	xv
LIST OF ABBREVIATIONS	xvi
CHAPTER	
1. INTRODUCTION	1
2. LITERATURE REVIEW	4
2.1 Inflammation	4
2.2 Prostaglandin as mediator of inflammation	5
2.2.1 Prostaglandin	5
2.2.2 Biosynthesis of prostaglandin	6
2.2.3 Prostaglandin E ₂	8
2.3 Cyclooxygenase	10
2.3.1 Structure and mechanisms of COX enzymes	11
2.4 NSAIDs as COX inhibitors and their mechanism of action	14
2.4.1 Biological repercussions of NSAIDs	16
2.5 Curcumin	17
2.5.1 Curcumin derivatives	17
3. MATERIALS AND METHODS	19
3.1 Collection and synthesis of curcumin derivatives	19
3.1.1 General procedures for preparation of curcumin derivatives	19
3.2 Effect of curcumin derivatives towards PGE ₂ production in IFN- γ /LPS-stimulated RAW264.7 cells	21
3.2.1 Cultivation of RAW 264.7 cells	21
3.2.2 Seeding of cultivated cells, inflammatory-stimulation and treatment with curcumin derivatives	21
3.2.3 Determination of PGE ₂ concentration in culture supernatant of treated and untreated cells	22
3.2.4 Determination of cell viability of treated and untreated cells (MTT Assay)	23
3.3 2D-quantitative structure-activity relationship	23
3.3.1 Data set	23
3.3.2 Descriptors calculation	23
3.3.3 Regression analysis	24
3.3.4 Model's predictability	24
3.4 Binding orientation of curcumin derivatives in COX active sites by molecular docking approach	24

3.5 Inhibitory effect of curcumin derivatives on COX activities	25
3.5.1 COX reactions	25
3.5.2 Determination of PGE ₂ concentration in treated and untreated enzyme reactions	26
3.6 COX-1 and COX-2 Gene Expression Analysis	27
3.6.1 Stimulation and treatment of RAW264.7 cells for RNA extraction	27
3.6.2 Total RNA extraction	27
3.6.3 Total RNA quantification and integrity checking	28
3.6.4 cDNA synthesis of RNA from RAW264.7 cells	28
3.6.5 RT-qPCR Mastermix preparation and thermal cycling set up	29
3.6.6 Determination of optimum annealing temperature for <i>COX-1</i> , <i>COX-2</i> , <i>β-actin</i> , and <i>GAPDH</i> genes	30
3.6.7 Standard curve of <i>COX-1</i> , <i>COX-2</i> , <i>β-actin</i> , and <i>GAPDH</i> genes expression analysis	30
3.6.8 <i>COX-1</i> and <i>COX-2</i> genes expressions analysis	31
3.7 Statistical analysis	32
4. RESULTS	33
4.1 Effect of curcumin derivatives towards PGE ₂ production in IFN-γ/LPS-stimulated RAW264.7 cells.	33
4.2 Effect of curcumin derivatives on viability of IFN-γ/LPS-stimulated RAW264.7 cells.	33
4.3 Quantitative structure-activity relationship analysis on effect of curcumin derivatives towards PGE ₂ inhibition activity	36
4.4 Binding orientation of curcumin derivatives in COX active sites	39
4.5 Effect of curcumin derivatives on COX activity	48
4.6 Effect of different annealing temperatures on amplifications of <i>COX-1</i> , <i>COX-2</i> , <i>β-actin</i> and <i>GAPDH</i> genes	52
4.7 Effect of selected curcumin derivatives on <i>COX-1</i> and <i>COX-2</i> genes expressions in IFN-γ/LPS-stimulated RAW264.7 cells	52
5. DISCUSSION	55
6. SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	62
REFERENCES	63
APPENDICES	75
BIODATA OF STUDENT	101
LIST OF PUBLICATIONS	102

LIST OF TABLES

Table		Page
3.1	Mastermix for <i>COX-1</i> , <i>COX-2</i> , β - <i>actin</i> and <i>GAPDH</i> cDNA synthesis	28
3.2	Thermal cycler conditions of cDNA synthesis	28
3.3	Mastermix for <i>COX-1</i> , <i>COX-2</i> , β - <i>actin</i> and <i>GAPDH</i> genes amplification	29
3.4	Thermal cycler conditions for <i>COX-1</i> , <i>COX-2</i> , β - <i>actin</i> and <i>GAPDH</i> genes	29
4.1	IC ₅₀ values of PGE ₂ inhibition and cytotoxicity of curcumin derivatives on LPS/IFN- γ stimulated RAW264.7 cells	34
4.2	Observed and predicted pIC ₅₀ of curcumin derivatives from a model obtained from GFA method	37
4.3	Scoring functions of Equation 1	38
4.4	List of descriptors calculated for the QSAR model	39
4.5	Summary of molecular docking of COX-1 by compounds 25, 27, and 43	40
4.6	Summary of molecular docking of COX-2 by compounds 25, 27, and 43	40
4.7	The screening and IC ₅₀ values of COX-1 and -2 inhibition activity by curcumin derivatives	49
4.8	Annealing temperature of primers targeting <i>COX-1</i> , <i>COX-2</i> , β - <i>actin</i> and <i>GAPDH</i> genes	52

LIST OF FIGURES

Figure		Page
2.1	Biosynthetic pathway of prostanoids	7
2.2	Mechanism of arachidonate oxygenation	8
2.3	Structure of prostaglandin E ₂	9
2.4	Schematic diagram for structural differences between the binding channels of cyclooxygenase (COX)	12
2.5	Binding orientation of of arachidonic acid (AA) in the ovine COX-1 (oCOX-1) active site	14
2.6	Structure of curcumin in β -diketone and keto-enol forms	18
4.1	Effects of compound 25 on PGE ₂ production in IFN- γ /LPS-stimulated RAW264.7 cells	35
4.2	Effects of compound 25 on RAW264.7 cell viability	36
4.3	Plot of predicted pIC ₅₀ values against observed pIC ₅₀ values of training set compounds	38
4.4	Predicted binding poses retrieved from flexible docking of compound 25 in (A) 3D diagram and (B) 2D diagram of ovine COX-1 active site	42
4.5	Predicted binding poses retrieved from flexible docking of compound 25 in (A) 3D diagram and (B) 2D diagram of murine COX-2 active site	43
4.6	Predicted binding poses retrieved from flexible docking of compound 27 in (A) 3D diagram and (B) 2D diagram of ovine COX-1 active site	44
4.7	Predicted binding poses retrieved from flexible docking of compound 27 in (A) 3D diagram and (B) 2D diagram of murine COX-2 active site	45
4.8	Predicted binding poses retrieved from flexible docking of compound 43 in (A) 3D diagram and (B) 2D diagram of murine COX-1 active site	46
4.9	Predicted binding poses retrieved from flexible docking of compound 43 in (A) 3D diagram and (B) 2D diagram of murine COX-2 active site	47
4.10	Effects of compound 25 on COX-1 and COX-2 inhibition activity	50

4.11	Effects of compound 27 on COX-1 and COX-2 inhibition activity	51
4.12	Effects of compound 43 on COX-1 and COX-2 inhibition activity	51
4.13	Effect of compounds 25, 27, and 43 on COX-2 genes expressions in IFN- γ /LPS-stimulated RAW264.7 cells	53
4.14	Effect of compounds 25, 27, and 43 on COX-1 genes expressions in PMA-stimulated RAW264.7 cells	54



LIST OF APPENDICES

Appendix		Page
A	Preparation of Dulbecco's Modified Eagle's Medium With Phenol Red, Buffers, IFN- γ and LPS Stock Solution	75
B	Preparation of Diluents and Solution for PGE ₂ Inhibition Assay, and MTT Assay	77
C	Preparation of Buffers and Reagents in PGE ₂ EIA Kit	78
D	Example of PGE ₂ Standard Curve for PGE ₂ Inhibition Assay and COX Enzymatic Assay	79
E	Parameters, Models, and Validation Results in QSAR Analysis	80
F	Preparation of Buffers and Reagents in COX Enzymatic Assay	83
G	Percentage of Inhibition by Compounds 25, 27, and 43 on COX-1 and -2 Activity	85
H	Preparation of Reagents for RNA Extraction	86
I	Preparation of Agarose Gel	87
J	RNA Purity and Concentration of IFN- γ /LPS-stimulated and PMA-stimulated RAW264.7 Cells	88
K	Primers Sequences of <i>COX-1</i> , <i>COX-2</i> , <i>β-actin</i> and <i>GAPDH</i> Used in RT-qPCR	90
L	Information on Components and Thermal Cycler Conditions Used in cDNA Synthesis	91
M	Information on Components and Thermal Cycler Conditions Used in RT-qPCR	92
N	Effect of Different Annealing Temperatures on <i>COX-1</i> , <i>COX-2</i> , <i>β-actin</i> and <i>GAPDH</i> Genes Amplification	93
O	Standard Curve of <i>COX-1</i> , <i>COX-2</i> , <i>β-actin</i> and <i>GAPDH</i> Genes Expression.	97

LIST OF ABBREVIATIONS

AA	Arachidonic acid
AD	Alzheimer's disease
Adj-R ²	adjusted R ²
ANOVA	one-way analysis of variance
AP	activator protein
APC	Adenoma Prevention with Celecoxib
APC ^{min}	Adenomatous polyposis coli
APPROVe	Adenomatous Polyp Prevention on Vioxx
ARG	Arginine
ATCC	American Type Culture Collection
B ₀	maximum binding
β-actin	beta actin
BSA	Bovine serum albumin
C/EBP	CCAAT/enhancer-binding protein
cAMP	Cyclic adenosine monophosphate
CDCl ₃	deuterated chloroform
cDNA	complementary Deoxyribonucleic acid
CLASS	Celecoxib Long Term Arthritis Safety Study
CNS	Central nervous system
COX	Cyclooxygenase
CO ₂	Carbon dioxide
CRE	cAMP response element
C _T	Threshold cycle
DAMP	Damage-associated molecular patterns
DMEM	Dulbecco's Modified Eagle Medium
DMSO	Dimethyl sulfoxide
dsDNA	double strand DNA
E.C	Enzyme commission
EDTA	Ethylenediaminetetraacetic acid
EGF	epidermal growth factor
EIA	enzyme immunoassay
EIMS	Electron Ionization Mass Spectroscopy
ERK	Extracellular signal-regulated kinase
EtOH	ethanol
Ev	electronvolt
FBS	fetal bovine serum
g	gram
GAPDH	Glyceraldehyde 3-phosphate dehydrogenase
gDNA	Genomic deoxyribonucleic acid
GFA	Genetic Function Approximation
GI	gastrointestinal
Gln	Glutamine
Glu	Glutamine
GPCRs	G protein-coupled receptors
h	Hour/s
HCl	Hydrochloric acid
His	Histidine
HPLC	high-performance liquid chromatography

ICAM	Intercellular adhesion molecule 1
IC ₅₀	Inhibitory concentration 50%
IFN- γ	Interferon-gamma
IgG	immunoglobulin G
IL	Interleukin
IL- β	interleukin 1 beta
Ile	Isoleucine
iNOS	inducible nitric oxide synthase
JNK	c-Jun N-terminal kinase
kb	kilobase
KBr	Potassium bromide
Kcal	kilocalorie
LOF	Friedman's lack of fit
LOX	lipoxigenase
LPS	Lipopolisaccharide
LSE	least-squares error
M	Molar
MAPK	Mitogen-activated protein kinase
MBD	membrane binding domain
mCOX	murine cyclooxygenase
mg	miligram
MHz	megahertz
min	Minute
ml	Millilitre
mM	Millimolar
mm	millimeter
mmol	milimol
mPGES	microsomal prostaglandin E synthase
mRNA	Messenger ribonucleic acid
MS	Mass spectra
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide
NaOH	Sodium hydroxide
NF- κ B	Nuclear factor-kappa B
Nm	nanometer
NMR	Nuclear Magnetic Resonance
NSAIDs	Non-steroidal anti-inflammatory drugs
NSB	non-specific binding
NS-398	<i>N</i> -[2-(Cyclohexyloxy)-4-nitrophenyl]methanesulfonamide
NTC	Non-template control
O ₂	Oxygen
oCOX	ovine cyclooxygenase
OD	optical density
PAMP	Pathogen-associated molecular patterns
PBS	Phosphate buffer saline
PD	Parkinson's disease
PG	Prostaglandin
pg	picogram
Phe	Phenylalanine
PLA ₂	Phospholipase A ₂
PMA	Phorbol myristate acetate

ppm	parts per million
Pred-R ²	predicted R ²
QSAR	Quantitative structure-activity relationship
RNA	Ribonucleic acid
RA	Rheumatoid arthritis
RFU	relative fluorescence unit
ROS	Reactive oxygen species
rpm	Revolutions per minute
RT-qPCR	Quantitative Real-Time Polymerase Chain Reaction
R ²	Extinction of coefficient
s	second
S.E.M	Standard error of mean
Ser	Serine
SUCCESS	Successive Celecoxib Efficacy and Safety Study
TA	Total activity
TLC	thin layer chromatography
TLR	Toll-like receptor
TNF- α	Tumor necrosis factor alpha
Tyr	Tyrosine
U	Unit
UV	Ultraviolet
V	Volt
Val	Valine
VCAM	Vascular cell adhesion molecule 1
VIGOR	Vioxx Gastrointestinal Safety of Rofecoxib
VSMC	Vascular smooth muscle cells
Å	Angstrom
°C	Degree celcius
µg	Microgram
µL	Microlitre
µM	Micromolar
ηM	nanomolar

CHAPTER 1

INTRODUCTION

Inflammation is a complex biological response. As part of immune system, the inflammation response not only serves as protective frontline towards harmful stimuli, but also as convergence point towards progression of severe inflammatory defects. Inflammation is an intrinsically beneficial event for biological host in fighting against offending factors such as pathogens, toxins, injuries, and chemical irritants with aims to eliminate the injurious factors, promote healing process and tissue restoration, as well as to build up memories for a faster and more specific counteractions in future occurrence (Stables and Gilroy, 2011).

The beneficial effects of inflammation were normally implemented during acute phase which is the initial stage of inflammation. In this stage, innate immunity takes place through recruitment of activated cells like neutrophils, dendritic cells, lymphocytes and macrophages to the site of injury. They integrate to phagocytize the pathogens and promote healing process. However, persistence of pro-inflammatory stimuli will lead to sustainment of inflammation for weeks, months and even years if the stimuli cannot be eliminated or if there are problems with healing process such as auto-immunity (Stables and Gilroy, 2011).

The persistence of inflammation has been reported to be associated with pathophysiology of chronic stage. Extended period of inflammation is problematic which can harm physiological systems due to tremendous increase of multiple reactive oxygen species and pro-inflammatory mediators such as prostaglandins, cytokines, inflammatory enzymes, and protein kinases (Kellog *et al.*, 2015). Damage on cells/tissues is the initial process which may lead to several serious chronic inflammatory diseases like rheumatoid arthritis (RA), Alzheimer's disease (AD) and cancers (Ricciotti and Fitzgerald, 2011).

One of the important regulators in inflammatory response is prostaglandin. This physiologically active 20-carbon lipid compound in normal condition, engages in various homeostasis regulation such as blood pressure, gastrointestinal integrity, cardiovascular system, central nervous system (CNS) activity, and fertility (Ricciotti and Fitzgerald, 2011). Prostaglandin also involves in onset of cardinal signs of inflammation: pain, heat, redness, and swelling. Thus it has been referred as classical pro-inflammatory mediator (Funk, 2001; Harris *et al.*, 2002).

Derived from plasma membrane arachidonic acid (AA), prostaglandin exists in several isoforms, and most abundant isoform in human is PGE₂ (Serhan and Levy, 2003). PGE₂, through different binding receptors, termed EP1 to EP4, exerts multiple homeostasis and inflammatory signals (Sugimoto and Narumiya, 2007). PGE₂ can also repeatedly cooperate with cytokine and pathogen- or damage-associated molecular patterns (PAMPs and DAMPs) during multiple inflammatory events and amplifies cytokine and PAMP/DAMP signaling by boosting the expression of inflammation-related genes induced by these stimuli (Aoki and Narumiya, 2012). Alleviated PGE₂

levels in human have been linked specifically with pancreatic, colorectal, breast, and lung cancer (Wang *et al.*, 2007).

Biosynthesis of PGE₂ from AA is catalyzed by prostaglandin G/H synthase, or cyclooxygenase (COX). The enzyme exists in two isoforms. The first isoform is cyclooxygenase-1 (COX-1), that constitutively and ubiquitously expresses prostaglandin at basal level to coordinate physiological conditions. COX-1 is present in almost every cells in human body. On the other hand, the second isoform is an inducible enzyme known as cyclooxygenase-2 (COX-2). COX-2 is produced only in injured tissues relatively at high levels and subsequently produces large amount of PGE₂ upon triggered by inflammatory stimuli (Lee^a *et al.*, 2009).

Activated macrophages serve as one of the central producers of COX-2 in the course of inflammation (Bowdish *et al.*, 2007). Combination of lipopolysaccharide (LPS) and interferon-gamma (IFN- γ) synergistically stimulates the activation of macrophage (Chan and Riches, 2001). Persistence of stimulation by these pro-inflammatory mediators however may resulted in over-expression of COX-2 which in turn leads to excess production of PGE₂ and consequently ends up with various chronic inflammatory diseases and tumors (Greenhough *et al.*, 2009; Wang *et al.*, 2007).

Production of prostaglandin particularly through inflammation can be controlled by targeting COX activities. Non-steroidal anti-inflammatory drugs (NSAIDs) are a class of drugs which have been reported to deliver pharmacological effects such as analgesic, anti-pyretic, and anti-inflammatory properties (Vonkeman and Laar, 2010). The mechanisms of most NSAIDs are particularly either blocking the production of prostaglandin via competitively bind and inhibit the activity of COX enzyme, suppress the COX gene expression itself or inhibition of transcription factors (Liggett *et al.*, 2014). Although plenty of successful NSAIDs have been produced, marketed and administered for a long time, mounting of health and clinical issues are raised over time.

Epidemiological studies on several traditional NSAIDs like acetylsalicylic acid, indomethacin, ibuprofen and naproxen have revealed that these medicines are non-selective on COX isoforms, reflecting both clinical efficacy and deleterious effects, which in turn associated with gastric ulceration and renal failure (Liu *et al.*, 2001). Unfortunately, benefit-risk calculations have been biased on continuingly marketing these drugs due to their efficacy in treating chronic inflammation diseases.

Recently, NSAIDs that preferentially block COX-2, namely 'coxibs' family were considered as new generation of safe and effective drugs (Rao and Knaus, 2008). Unfortunately, clinical trials on prevention of colorectal cancer by rofecoxib and celecoxib resulted in myocardial infarction and stroke, and even death, which were far more serious than side effects of traditional NSAIDs (Bresalier *et al.*, 2005; Solomon *et al.*, 2005). The trials had led to the withdrawal of rofecoxib (Vioxx) from the market. Therefore, discovery of alternative anti-inflammatory agents is of utmost important.

On natural preference of the treatment, a wide spectrum of phytochemicals and their derivatives have been identified for development as anti-inflammatory agents. Interestingly, curcumin which is widely presents as secondary metabolite in plants has been recognized as potent inhibitor against inflammation (Bukhari *et al.*, 2013). However, curcumin suffers from major drawbacks due to poor bioavailability. Once consumed, it will go through hepatic conjugation, resulted in production of glucuronides and sulphates whereas systemic administration caused it to be eliminated (Anand *et al.*, 2007). The problem is due to unstable 7-carbon spacer of β -diketone moiety (diarylheptanoid) of curcumin, which is a specific substrate for liver aldo-keto reductases (Anand *et al.*, 2007).

Synthesizing a new class of curcumin related compounds is one of researchers' approaches (Leow *et al.*, 2014). In addition, computational analysis has become another powerful tools in modern research area which can be utilized to understand the structure-activity relationship of the potential compounds. Recently, a few studies have proved the pharmacological interest of curcumin derivatives due to its effectiveness against inflammation (Jantan *et al.*, 2012). These findings have enormously encouraged the development of better amendment of newly synthesized curcumin derivatives. Nevertheless, the effects and mechanism of actions of curcumin derivatives towards PGE_2 in normal and inflammatory conditions are poorly understood.

Here, a series of curcumin derivatives were assayed for prostaglandin E_2 inhibition in $\text{IFN-}\gamma/\text{LPS}$ -stimulated RAW264.7 (macrophage) cells as well as on pure COX-1 and -2 enzyme activities. Their potential in PGE_2 inhibition was analyzed quantitatively related to their molecular geometry. While molecular docking study was exploited to find a theoretical mechanism of inhibition of curcumin derivatives in COX active sites in comparison with their effects on the enzymatic activity. Finally, samples were tested on COX genes expression level.

Objectives of study

The general objective of this study is to elucidate the anti-inflammatory properties of curcumin derivatives in $\text{IFN-}\gamma/\text{LPS}$ -stimulated macrophage cells.

The specific objectives are:

1. To evaluate the inhibition activity of 43 curcumin derivatives towards PGE_2 production in $\text{IFN-}\gamma/\text{LPS}$ -stimulated RAW264.7 cells.
2. To measure geometric and chemical characteristics of curcumin derivatives in which related to their PGE_2 inhibition activity using quantitative structure-activity relationship (QSAR) analysis.
3. To determine the binding orientation of curcumin derivatives in COX active sites using molecular docking approach and correlate with COX enzymatic assay.
4. To determine the effect of selected curcumin derivatives on mRNA expression of COX-1 and -2 in $\text{IFN-}\gamma/\text{LPS}$ -stimulated RAW264.7 cells.

REFERENCES

- Aggarwal, B. B., Kumar, A., & Bharti, A. C. (2003). Anticancer potential of curcumin: preclinical and clinical studies. *Anticancer Research*, 23(1), 363-398.
- Aggarwal, B. B., & Sung, B. (2009). Pharmacological basis for the role of curcumin in chronic diseases: an age-old spice with modern targets. *Trends in Pharmacological Sciences*, 30(2), 85-94.
- Ahmad, W., Kumolosasi, E., Jantan, I., Bukhari, S. N., & Jasamai, M. (2014). Effects of novel diarylpentanoid analogues of curcumin on secretory phospholipase A₂, cyclooxygenase, lipo-oxygenase, and microsomal prostaglandin E synthase-1. *Chemical Biology and Drug Design*, 83(6), 670-681.
- Ahmadi, M., Emery, D. C., & Morgan, D. J. (2008). Prevention of both direct and cross-priming of antitumor CD8⁺ T-cell responses following overproduction of prostaglandin E₂ by tumor cells in vivo. *Cancer Research*, 68(18), 7520-7529.
- Alberto, M. R., Zampini, I. C., & Isla, M. I. (2009). Inhibition of cyclooxygenase activity by standardized hydroalcoholic extracts of four Asteraceae species from the Argentine Puna. *Brazilian Journal of Medical and Biological Research*, 42(9), 787-790.
- Al-Swayeh, O. A., Clifford, R. H., Del Soldato, P., & Moore, P. K. (2000). A comparison of the anti-inflammatory and anti-nociceptive activity of nitroaspirin and aspirin. *British Journal of Pharmacology*, 129(2), 343-350.
- Anand, P., Kunnumakkara, A. B., Newman, R. A., & Aggarwal, B. B. (2007). Bioavailability of curcumin: Problems and promises. *Molecular Pharmacology*, 4, 807-818.
- Anand, P., Thomas, S. G., Kunnumakkara, A. B., Sundaram, C., Harikumar, K. B., Sung, B., Tharakan, S. T., Misra, K., Priyadasini, I. K., Rajasekharan, K. N., & Aggarwal, B. B. (2008). Biological activities of curcumin and its analogues (Congeners) made by man and Mother Nature. *Biochemical Pharmacology*, 76(11), 1590-1611.
- Aoki, T. & Narumiya, S. (2012). Prostaglandins and chronic inflammation. *Trends in Pharmacological Sciences*, 33(6), 304-311.
- Awtry, E. H., & Loscalzo, J. (2000). Aspirin. *Circulation*, 101(10), 1206-1218.
- Bagga, D., Wang, L., Farias-Eisner, R., Glaspy, J. A., & Reddy, S. T. (2003). Differential effects of prostaglandin derived from ω-6 and ω-3 polyunsaturated fatty acids on COX-2 expression and IL-6 secretion. *Proceedings of The National Academy of Sciences*, 100(4), 1751-1756.
- Bao, H. H., & You, S. G. (2011). Molecular characteristics of water-soluble extracts from *Hypsizygus marmoreus* and their *in vitro* growth inhibition of various

cancer cell lines and immunomodulatory function in RAW 264.7 cells. *Bioscience, Biotechnology, and Biochemistry*, 75(5), 891-898.

- Ben-Neriah, Y., & Karin, M. (2011). Inflammation meets cancer, with NF-[kappa] B as the matchmaker. *Nature Immunology*, 12(8), 715-723.
- Blobaum, A. L., & Marnett, L. J. (2007). Structural and functional basis of cyclooxygenase inhibition. *Journal of Medicinal Chemistry*, 50(7), 1425-1441.
- Botting, R. M. (2006). Cyclooxygenase: past, present and future. A tribute to John R. Vane (1927–2004). *Journal of Thermal Biology*, 31(1), 208-219.
- Bouvard, V., Baan, R., Straif, K., Grosse, Y., Secretan, B., El Ghissassi, F., Benbrahim-Tallaa, L., Guha, N., Freeman, C., Galichet, L., & Coglianò, V. (2009). A review of human carcinogens-Part B: biological agents. *Lancet Oncology*, 10, 321–322.
- Bowdish, D. M. E., Loffredo, M. S., Mukhopadhyay, S., Mantovani, A., & Gordon, S. (2007). Macrophage receptors implicated in the “adaptive” form of innate immunity. *Microbes and Infection*, 9, 1680–1687.
- Bresalier, R. S., Sandler, R. S., Quan, H., Bolognese, J. A., Oxenius, B., Horgan, K., Lines, C., & Baron, J. A. (2005). Cardiovascular events associated with rofecoxib in a colorectal adenoma chemoprevention trial. *New England Journal of Medicine*, 352(11), 1092-1102.
- Brown, J. R., & DuBois, R. N. (2005). COX-2: a molecular target for colorectal cancer prevention. *Journal of Clinical Oncology*, 23(12), 2840-2855.
- Bukhari, S. N. A., Jantan, I. B., Jasamai, M., Ahmad, W., & Amjad, M. W. B. (2013). Synthesis and biological evaluation of curcumin analogues. *Journal of Medical Sciences*, 13(7), 501-513.
- Calder, P. C. (2006). n-3 polyunsaturated fatty acids, inflammation, and inflammatory diseases. *The American Journal of Clinical Nutrition*, 83(6), 1505-1519.
- Carey, F. A., & Sundberg, R. J. (2007). *Advanced organic chemistry: Part A: Structure and Mechanisms*. Springer Science & Business Media.
- Chan, E. D., & Riches, D. W. (2001). IFN- γ + LPS induction of iNOS is modulated by ERK, JNK/SAPK, and p38 mapk in a mouse macrophage cell line. *American Journal of Physiology-Cell Physiology*, 280(3), 441-450.
- Cheng, Y., Wang, M., Yu, Y., Lawson, J., Funk, C. D., & FitzGerald, G. A. (2006). Cyclooxygenases, microsomal prostaglandin E synthase-1, and cardiovascular function. *The Journal of Clinical Investigation*, 116(5), 1391-1399.
- Cho, J. W., Lee, K. S., & Kim, C. W. (2007). Curcumin attenuates the expression of IL-1 β , IL-6, and TNF- α as well as cyclin E in TNF- α -treated HaCaT

- cells; NF-kappaB and MAPKs as potential upstream targets. *International Journal of Molecular Medicine*, 19, 469–74.
- Crofford, L. J., Lipsky, P. E., Brooks, P., Abramson, S. B., Simon, L. S., & Van De Putte, L. B. (2000). Basic biology and clinical application of specific cyclooxygenase-2 inhibitors. *Arthritis & Rheumatism*, 43(1), 4.
- Curry, S. L., Cogar, S. M., & Cook, J. L. (2005). Nonsteroidal antiinflammatory drugs: a review. *Journal of The American Animal Hospital Association*, 41(5), 298-309.
- DeWitt, D. L., El-Harith, E. A., Kraemer, S. A., Andrews, M. J., Yao, E. F., Armstrong, R. L., & Smith, W. L. (1990). The aspirin and heme-binding sites of ovine and murine prostaglandin endoperoxide synthases. *Journal of Biological Chemistry*, 265(9), 5192-5198.
- Dannhardt, G., & Kiefer, W. (2001). Cyclooxygenase inhibitors—current status and future prospects. *European Journal of Medicinal Chemistry*, 36(2), 109-126.
- Dey, I., Lejeune, M., & Chadee, K. (2006). Prostaglandin E₂ receptor distribution and function in the gastrointestinal tract. *British Journal of Pharmacology*, 149, 611–23.
- Dhillon, N., Aggarwal, B. B., Newman, R. A., Wolff, R. A., Kunnumakkara, A. B., Abbruzzese, J. L., Ng, C. S., Badmaev, V., & Kurzrock, R. (2008). Phase II trial of curcumin in patients with advanced pancreatic cancer. *Clinical Cancer Research*, 14(14), 4491-4499.
- Dietz, R., Nastainczyk, W., & Ruf, H. H. (1988). Higher oxidation states of prostaglandin H synthase. *European Journal of Biochemistry*, 171(1-2), 321-328.
- Dobrovolskaia, M. A., & Vogel, S. N. (2002). Toll receptors, CD14, and macrophage activation and deactivation by LPS. *Microbes and Infection*, 4(9), 903-914.
- Donath, M. Y., & Shoelson, S. E. (2011). Type 2 diabetes as an inflammatory disease. *Nature Reviews Immunology*, 11(2), 98-107.
- Dubois, R. N., Abramson, S. B., Crofford, L., Gupta, R. A., Simon, L. S., Van De Putte, L. B., & Lipsky, P. E. (1998). Cyclooxygenase in biology and disease. *The FASEB Journal*, 12(12), 1063-1073.
- Fitzgerald, G. A., & Patrono, C. (2001). The coxibs, selective inhibitors of cyclooxygenase-2. *The New England Journal of Medicine*, 345, 433-442.
- Fortier, M. A., Krishnaswamy, K., Danyod, G., Boucher-Kovalik, S., & Chapdalaine, P. (2008). A postgenomic integrated view of prostaglandins in reproduction: implications for other body systems. *Journal of Physiology and Pharmacology*, 59(1), 65–89.

- Fuchs, J. R., Pandit, B., Bhasin, D., Etter, J. P., Regan, N., Abdelhamid, D., Li, C., Lin, J., & Li, P. K. (2009). Structure–activity relationship studies of curcumin analogues. *Bioorganic and Medicinal Chemistry Letters*, 19(7), 2065-2069.
- Fujihara, M., Muroi, M., Tanamoto, K. I., Suzuki, T., Azuma, H., & Ikeda, H. (2003). Molecular mechanisms of macrophage activation and deactivation by lipopolysaccharide: roles of the receptor complex. *Pharmacology and Therapeutics*, 100(2), 171-194.
- Funk, C. D. (2001). Prostaglandins and leukotrienes: advances in eicosanoid biology. *Science*, 294(5548), 1871-1875.
- Futaki, N., Takahashi, S., Yokoyama, M., Arai, I., Higuchi, S., & Otomo, S. (1994). NS-398, a new anti-inflammatory agent, selectively inhibits prostaglandin G/H synthase/cyclooxygenase (COX-2) activity *in vitro*. *Prostaglandins*, 47(1), 55-59.
- Garavito, R. M., & Mulichak, A. M. (2003). The structure of mammalian cyclooxygenases. *Annual Review of Biophysics and Biomolecular Structure*, 32(1), 183-206.
- Gautam, S. C., Gao, X., & Dulchavsky, S. (2007). Immunomodulation by curcumin. *Advances in Experimental Medicine and Biology*, 595, 321–41.
- Goel^a, A., Kunnumakkara, A. B., & Aggarwal, B. B. (2008). Curcumin as “Curecumin”: from kitchen to clinic. *Biochemical Pharmacology*, 75(4), 787-809.
- Goel^b, A., Jhurani, S., & Aggarwal, B. B. (2008). Multi-targeted therapy by curcumin: how spicy is it? *Molecular Nutrition And Food Research*, 52(9), 1010-1030.
- Greenhough, A., Smartt, H. J., Moore, A. E., Roberts, H. R., Williams, A. C., Paraskeva, C., & Kaidi, A. (2009). The COX-2/PGE₂ pathway: key roles in the hallmarks of cancer and adaptation to the tumour microenvironment. *Carcinogenesis*, 30(3), 377-386.
- Grogan, G. (2005). Emergent mechanistic diversity of enzyme-catalysed beta-diketone cleavage. *Biochemical Journal*, 388, 721-730.
- Grosser, T., Fries, S., & FitzGerald, G. A. (2006). Biological basis for the cardiovascular consequences of COX-2 inhibition: therapeutic challenges and opportunities. *Journal of Clinical Investigation*, 116(1), 4-15.
- Guan, Y., Chang, M., Cho, W., Zhang, Y., Redha, R., Davis, L., Chang, S., DuBois, R. N., Hao, C. M., & Breyer, M. (1997). Cloning, expression, and regulation of rabbit cyclooxygenase-2 in renal medullary interstitial cells. *American Journal of Physiology-Renal Physiology*, 273(1), 18-26.

- Handler, N., Jaeger, W., Puschacher, H., Leisser, K., & Erker, T. (2007). Synthesis of novel curcumin analogues and their evaluation as selective cyclooxygenase-1 (COX-1). *Chemical and Pharmaceutical Bulletin*, 55(1), 64-71.
- Hansen-Petrik, M. B., McEntee, M. F., Jull, B., Shi, H., Zemel, M. B., & Whelan, J. (2002). Prostaglandin E₂ protects intestinal tumors from nonsteroidal anti-inflammatory drug-induced regression in ApcMin/+ mice. *Cancer Research*, 62(2), 403-408.
- Harris, S. G., Padilla, J., Koumas, L., Ray, D., & Phipps, R. P. (2002). Prostaglandins as modulators of immunity. *Trends in Immunology*, 23(3), 144-150.
- Hemler, M., & Lands, W. E. M. (1976). Purification of the cyclooxygenase that forms prostaglandins. Demonstration of two forms of iron in the holoenzyme. *Journal of Biological Chemistry*, 251, 5575– 5579.
- Hochgesang, G. P., Rowlinson, S. W., & Marnett, L. J. (2000). Tyrosine-385 is critical for acetylation of cyclooxygenase-2 by aspirin. *Journal of The American Chemical Society*, 122(27), 6514-6515.
- Hosoya, T., Nakata, A., Yamasaki, F., Abas, F., Shaari, K., Lajis, N. H., & Morita, H. (2012). Curcumin-like diarylpentanoid analogues as melanogenesis inhibitors. *Journal of Natural Medicines*, 66(1), 166-176.
- Hsu, C. H., & Cheng, A. L. (2007). Clinical Studies With Curcumin. (Eds.) In *The Molecular Targets and Therapeutic Uses of Curcumin in Health and Disease* (pp. 471-480). US: Springer Science and Business Media.
- Hsu, H. Y., Chu, L. C., Hua, K. F., & Chao, L. K. (2008). Heme oxygenase-1 mediates the anti-inflammatory effect of Curcumin within LPS-stimulated human monocytes. *Journal of Cellular Physiology*, 215(3), 603-612.
- Iravani, M. M., Kashefi, K., Mander, P., Rose, S., & Jenner, P. (2001). Involvement of inducible nitric oxide synthase in inflammation-induced dopaminergic neurodegeneration. *Neuroscience*, 110, 49-58.
- Jantan, I., Bukhari, S. N. A., Lajis, N. H., Abas, F., Wai, L. K., & Jasamai, M. (2012). Effects of diarylpentanoid analogues of curcumin on chemiluminescence and chemotactic activities of phagocytes. *Journal of Pharmacy and Pharmacology*, 64(3), 404-412.
- Jiang, Y. J., Lu, B., Choy, P. C., & Hatch, G. M. (2003). Regulation of cytosolic phospholipase A₂, cyclooxygenase-1 and-2 expression by PMA, TNF α , LPS and M-CSF in human monocytes and macrophages. *Vascular Biochemistry*, 31- 38.
- Johnson, G. L., & Lapadat, R. (2002). Mitogen-activated protein kinase pathways mediated by ERK, JNK, and p38 protein kinases. *Science*, 298(5600), 1911-1912.

- Kalgutkar, A. S., Crews, B. C., Rowlinson, S. W., Marnett, A. B., Kozak, K. R., Remmel, R. P., & Marnett, L. J. (2000). Biochemically based design of cyclooxygenase-2 (COX-2) inhibitors: facile conversion of nonsteroidal antiinflammatory drugs to potent and highly selective COX-2 inhibitors. *Proceedings of the National Academy of Sciences*, 97(2), 925-930.
- Kang, B. Y., Chung, S. W., Chung, W. J., Im, S. Y., Hwang, S. Y., & Kim, T. S. (1999). Inhibition of interleukin-12 production in lipopolysaccharide-activated macrophages by curcumin. *European Journal of Pharmacology*, 384(2), 191-195.
- Kang, Y. J., Mbonye, U. R., DeLong, C. J., Wada, M., & Smith, W. L. (2007). Regulation of intracellular cyclooxygenase levels by gene transcription and protein degradation. *Progress in Lipid Research*, 46(2), 108-125.
- Kellogg, J., Esposito, D., Grace, M. H., Komarnytsky, S., & Lila, M. A. (2015). Alaskan seaweeds lower inflammation in RAW 264.7 macrophages and decrease lipid accumulation in 3T3-L1 adipocytes. *Journal of Functional Foods*, 15, 396-407.
- Kiefer, J. R., Pawlitz, J. L., Moreland, K. T., Stegeman, R. A., Hood, W. F., Gierse, J. K., Stevens, A. M., Goodwin, D. C., Rowlinson, S. W., Marnett, L. J., Stallings, W. C., & Kurumbail, R. G. (2000). Structural insights into the stereochemistry of the cyclooxygenase reaction. *Nature*, 405(6782), 97-101.
- Kier, L. B., & Hall, L. H. (1999). The Kappa Indices for Modeling Molecular Shape And Flexibility. In *Topological Indices and Related Descriptors in QSAR and QSPR* (pp. 455-490). Reading, UK : Gordon and Breach.
- Kulmacz, R. J. (2005). Regulation of cyclooxygenase catalysis by hydroperoxides. *Biochemical and Biophysical Research Communications*, 338(1), 25-33.
- Kurumbail, R. G., Stevens, A. M., Gierse, J. K., McDonald, J. J., Stegeman, R. A., Pak, J. Y., Gildehaus, D., & Stallings, W. C. (1996). Structural basis for selective inhibition of cyclooxygenase-2 by anti-inflammatory agents. *Nature*, 384(6610), 644-648.
- Laine, L. (2003). Gastrointestinal effects of NSAIDs and coxibs. *Journal of Pain and Symptom Management*, 25(2), 32-40.
- Lee, K. H., Chow, Y. L., Sharmili, V., Abas, F., Alitheen, N. B. M., Shaari, K., Israfi, D. A., Lajis, N. H., & Syahida, A. (2012). BDMC33, a curcumin derivative suppresses inflammatory responses in macrophage-like cellular system: Role of inhibition in NF- κ B and MAPK signaling pathways. *International Journal of Molecular Sciences*, 13(3), 2985-3008.
- Lee^a, H. S., Lee, C. H., Tsai, H. C., & Salter, D. M. (2009). Inhibition of cyclooxygenase 2 expression by diallyl sulfide on joint inflammation induced by urate crystal and IL-1 β . *Osteoarthritis and Cartilage*, 17(1), 91-99.

- Lee^b, K. H., Aziz, F. H. A., Syahida, A., Abas, F., Shaari, K., Israf, D. A., & Lajis, N. H. (2009). Synthesis and biological evaluation of curcumin-like diarylpentanoid analogues for anti-inflammatory, antioxidant and anti-tyrosinase activities. *European Journal of Medicinal Chemistry*, 44(8), 3195-3200.
- Lee, K. H., Abas, F., Alitheen, N. B. M., Shaari, K., Lajis, N. H., & Ahmad, S. (2011). A Curcumin Derivative, 2, 6-Bis (2, 5-dimethoxybenzylidene)-cyclohexanone (BDMC33) Attenuates Prostaglandin E2 Synthesis via Selective Suppression of Cyclooxygenase-2 in IFN- γ /LPS-Stimulated Macrophages. *Molecules*, 16(11), 9728-9738.
- Legler, D. F., Bruckner, M., Uetz-von Allmen, E., & Krause, P. (2010). Prostaglandin E2 at new glance: novel insights in functional diversity offer therapeutic chances. *The International Journal of Biochemistry and Cell Biology*, 42(2), 198-201.
- Leiroa, J., Garcí'ab, D., Arranza, J.A., Delgadob, R., Sanmarti'na, M.L., & Orallo, F., (2004). An Anacardiaceae preparation reduces the expression of inflammation-related genes in murine macrophages. *International Immunopharmacology*, 4, 991-1003.
- Leow, P. C., Bahety, P., Boon, C. P., Lee, C. Y., Tan, K. L., Yang, T., & Ee, P. L. R. (2014). Functionalized curcumin analogs as potent modulators of the Wnt/b-catenin signaling pathway. *European Journal of Medicinal Chemistry*, 71, 67-80.
- Liang, G., Li, X., Chen, L., Yang, S., Wu, X., Studer, E., Gurley, E., Hylemon, P. B., Ye, F., Li, Y., & Zhou, H. (2008). Synthesis and anti-inflammatory activities of mono-carbonyl analogues of curcumin. *Bioorganic and Medicinal Chemistry Letters*, 18(4), 1525-1529.
- Liang^a, G., Zhou, H., Wang, Y., Gurley, E. C., Feng, B., Chen, L., Jian, X., Yang, S., & Li, X. (2009). Inhibition of LPS-induced production of inflammatory factors in the macrophages by mono-carbonyl analogues of curcumin. *Journal of Cellular and Molecular Medicine*, 13(9b), 3370-3379.
- Liang^b, G., Yang, S., Zhou, H., Shao, L., Huang, K., Xiao, J., Huang, Z., & Li, X. (2009). Synthesis, crystal structure and anti-inflammatory properties of curcumin analogues. *European Journal of Medicinal Chemistry*, 44(2), 915-919.
- Liang^c, G., Shao, L., Wang, Y., Zhao, C., Chu, Y., Xiao, J., Zhao, Y., Li, X., & Yang, S. (2009). Exploration and synthesis of curcumin analogues with improved structural stability both *in vitro* and *in vivo* as cytotoxic agents. *Bioorganic and Medicinal Chemistry*, 17(6), 2623-2631.
- Liggett, J. L., Zhang, X., Eling, T. E., & Baek, S. J. (2014). Anti-tumor activity of non-steroidal anti-inflammatory drugs: cyclooxygenase-independent targets. *Cancer Letters*, 346(2), 217-224.

- Liu, C. H., Chang, S. H., Narko, K., Trifan, O. C., Wu, M. T., Smith, E., Haudenschild, C., Lane, T. F., & Hla, T. (2001). Overexpression of cyclooxygenase-2 is sufficient to induce tumorigenesis in transgenic mice. *Journal of Biological Chemistry*, 276(21), 18563-18569.
- Liu, B., Gao, H. M., Wang, J. Y., Jeohn, G. H., Cooper, C. L., & Hong, J. S. (2002). Role of nitric oxide in inflammation-mediated neurodegeneration. *Annals of the New York Academy of Sciences*, 962, 318-331.
- Livak, K. J., & Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) method. *Methods*, 25(4), 402-408.
- Loll, P. J., Sharkey, C. T., O'Connor, S. J., Dooley, C. M., O'Brien, E., Devocelle, M., Nolan, K. B., Selinsky, B. S., & Fitzgerald, D. J. (2001). O-acetylsalicylhydroxamic acid, a novel acetylating inhibitor of prostaglandin H₂ synthase: structural and functional characterization of enzyme-inhibitor interactions. *Molecular Pharmacology*, 60(6), 1407-1413.
- Lucas, S. M., Rothwell, N. J., & Gibson, R. M. (2006). The role of inflammation in CNS injury and disease. *British Journal of Pharmacology*, 147(1), 232-240.
- Malkowski, M. G., Ginell, S. L., Smith, W. L., & Garavito, R. M. (2000). The productive conformation of arachidonic acid bound to prostaglandin synthase. *Science*, 289(5486), 1933-1937.
- Marnett, L. J., & Kalgutkar, A. S. (2004). Structural Diversity of Selective COX-2 Inhibitors. In *COX-2 Inhibitors* (pp. 15-40). Boston, Massachusetts: Birkhauser-Verlag.
- Marnett, L. J., & Maddipati, K. R. (1990). Prostaglandin H synthase. In Everse, J., Everse, K., & Grisham, M. (Ed.), *Peroxidases: Chemistry and Biology* (pp. 185-205). Boca Raton, USA: CRC Press.
- Matteo, V., & Esposito, E. (2003). Biochemical and therapeutic effects of antioxidants in the treatment of Alzheimer's disease, Parkinson's disease, and amyotrophic lateral sclerosis. *Current Drug Targets-CNS & Neurological Disorders*, 2(2), 95-107.
- Merlie, J. P., Fagan, D., Mudd, J., & Needleman, P. (1988). Isolation and characterization of the complementary DNA for sheep seminal vesicle prostaglandin endoperoxide synthase (cyclooxygenase). *Journal of Biological Chemistry*, 263, 3550- 3553.
- Minghetti, L. (2004). Cyclooxygenase-2 (COX-2) in inflammatory and degenerative brain diseases. *Journal of Neuropathology & Experimental Neurology*, 63(9), 901-910.
- Murias, M., Handler, N., Erker, T., Pleban, K., Ecker, G., Saiko, P., Szekeres, T., & Jäger, W. (2004). Resveratrol analogues as selective cyclooxygenase-2

- inhibitors: synthesis and structure–activity relationship. *Bioorganic & Medicinal Chemistry*, 12(21), 5571-5578.
- Nantasenamat, C., Worachartcheewan, A., Prachayasittikul, S., Isarankura-Na-Ayudhya, C., & Prachayasittikul, V. (2013). QSAR modeling of aromatase inhibitory activity of 1-substituted 1, 2, 3-triazole analogs of letrozole. *European Journal of Medicinal Chemistry*, 69, 99-114.
- Nurfina, A. N., Reksohadiprodjo, M. S., Timmerman, H., Jenie, U. A., Sugiyanto, D., & Van der Goot, H. (1997). Synthesis of some symmetrical curcumin derivatives and their antiinflammatory activity. *European Journal of Medicinal Chemistry*, 32(4), 321-328.
- O'Brien, P. J. (2000). Peroxidases. *Chemico-biological interactions*, 129(1), 113-139.
- Park, J. Y., Pillinger, M. H., & Abramson, S. B. (2006). Prostaglandin E₂ synthesis and secretion: the role of PGE₂ synthases. *Clinical immunology*, 119(3), 229-240.
- Picot, D., Loll, P. J., & Garavito, R. M. (1994). The X-ray crystal structure of the membrane protein prostaglandin H₂ synthase-1. *Nature*, 367(6460), 243-249.
- Prusakiewicz, J. J., Felts, A. S., Mackenzie, B. S., & Marnett, L. J. (2004). Molecular basis of the time-dependent inhibition of cyclooxygenases by indomethacin. *Biochemistry*, 43(49), 15439-15445.
- Rader, D. J., & Daugherty, A. (2008). Translating molecular discoveries into new therapies for atherosclerosis. *Nature*, 451(7181), 904-913.
- Rao, P. N. P., & Knaus, E. E. (2008). Evolution of nonsteroidal anti-inflammatory drugs (NSAIDs): Cyclooxygenase (COX) inhibition and beyond. *Journal Of Pharmacy & Pharmaceutical Sciences*, 11(2), 801 -110.
- Ricciotti, E., & FitzGerald, G. A. (2011). Prostaglandins and inflammation. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 31(5), 986-1000.
- Rieke, C. J., Mulichak, A. M., Garavito, R. M., & Smith, W. L. (1999). The role of arginine 120 of human prostaglandin endoperoxide H synthase-2 in the interaction with fatty acid substrates and inhibitors. *Journal of Biological Chemistry*, 274, 17109–17114.
- Rosemond, M. J. C., John-Williams, L. S., Yamaguchi, T., Fujishita, T., & Walsh, J. S. (2004). Enzymology of a carbonyl reduction clearance pathway for the HIV integrase inhibitor, S-1360: role of human liver cytosolic aldo-keto reductases. *Chemico-biological Interactions*, 147(2), 129-139.
- Rouzer, C. A., & Marnett, L. J. (2003). Mechanism of free radical oxygenation of polyunsaturated fatty acids by cyclooxygenases. *Chemical reviews*, 103(6), 2239-2304.

- Rowlinson, S. W., Kiefer, J. R., Prusakiewicz, J. J., Pawlitz, J. L., Kozak, K. R., Kalgutkar, A. S., Stallings, W. C., Kurumbail, R. G., & Marnett, L. J. (2003). A novel mechanism of cyclooxygenase-2 inhibition involving interactions with Ser-530 and Tyr-385. *Journal of Biological Chemistry*, 278(46), 45763-45769.
- Roy, P. P., & Roy, K. (2008). On some aspects of variable selection for partial least squares regression models. *QSAR & Combinatorial Science*, 27(3), 302-313.
- Schneider, C., Boeglin, W. E., & Brash, A. R. (2004). Identification of two cyclooxygenase active site residues, leucine 384 and glycine 526, that control carbon ring cyclization in prostaglandin biosynthesis. *Journal of Biological Chemistry*, 279(6), 4404-4414.
- Schroder, K., Sweet, M. J., & Hume, D. A. (2006). Signal integration between IFN- γ and TLR signaling pathways in macrophages. *Immunobiology*, 211(6), 511-524.
- Selinsky, B. S., Gupta, K., Sharkey, C. T., & Loll, P. J. (2001). Structural analysis of NSAID binding by prostaglandin H₂ synthase: time dependent and time-independent inhibitors elicit identical enzyme conformations. *Biochemistry*, 40, 5172-5180.
- Serhan, C. N., & Levy, B. (2003). Success of prostaglandin E₂ in structure-function is a challenge for structure-based therapeutics. *Proceedings of the National Academy of Sciences of the United States of America*, 100, 8609-8611.
- Sharma, R. A., McLelland, H. R., Hill, K. A., Ireson, C. R., Euden, S. A., Manson, M. M., Pirmohamed, M., Marnett, L. J., Gescher, A. J., & Steward, W. P. (2001). Pharmacodynamic and pharmacokinetic study of oral curcuma extract in patients with colorectal cancer. *Clinical Cancer Research*, 7(7), 1894-1900.
- Sharma, R. A., Steward, W. P., & Gescher, A. J. (2007). Pharmacokinetics and Pharmacodynamics of Curcumin. In *The Molecular Targets and Therapeutic Uses of Curcumin in Health and Disease* (pp. 453-470). US: Springer.
- Sidhu, R. S., Lee, J. Y., Yuan, C., & Smith, W. L. (2010). Comparison of cyclooxygenase-1 crystal structures: cross-talk between monomers comprising cyclooxygenase-1 homodimers. *Biochemistry*, 49(33), 7069-7079.
- Simmons, D. L., Botting, R. M., & Hla, T. (2004). Cyclooxygenase isozymes: the biology of prostaglandin synthesis and inhibition. *Pharmacological reviews*, 56(3), 387-437.
- Sivakumar, P. M., Geetha Babu, S. K., & Mukesh, D. (2007). QSAR studies on chalcones and flavonoids as anti-tuberculosis agents using genetic function approximation (GFA) method. *Chemical and Pharmaceutical Bulletin*, 55(1), 44-49.
- Smith, W. L., & Marnett, L. J. (1991). Prostaglandin endoperoxide synthase: structure and catalysis. *Biochimica et Biophysica Acta (BBA)-Lipids and Lipid Metabolism*, 1083(1), 1-17.

- Smith, W. L., DeWitt, D. L., & Garavito, R. M. (2000). Cyclooxygenases: structural, cellular, and molecular biology. *Annual Review of Biochemistry*, 69(1), 145-182.
- Solomon, S. D., McMurray, J. J., Pfeffer, M. A., Wittes, J., Fowler, R., Finn, P., Anderson, W. F., Zauber, A., Hawk, E., & Bertagnolli, M. (2005). Cardiovascular risk associated with celecoxib in a clinical trial for colorectal adenoma prevention. *New England Journal of Medicine*, 352(11), 1071-1080.
- Stables, M. J., & Gilroy, D. W. (2011). Old and new generation lipid mediators in acute inflammation and resolution. *Progress in Lipid Research*, 50(1), 35-51.
- Sugimoto, Y., & Narumiya, S. (2007). Prostaglandin E receptors. *Journal of Biological Chemistry*, 282(16), 11613-11617.
- Swan, S. K., Rudy, D. W., Lasseter, K. C., Ryan, C. F., Buechel, K. L., Lambrecht, L. J., Pinto, M. B., Dilzer, S. C., Obrda, O., Sunblad, K. J., Gumbs, C. P., Elbel, D. L., Quan, H., Larson, P. J., Schwartz, J. I., Musliner, T. A., Gertz, B. J., Brater, C., & Yao, S. L. (2000). Effect of cyclooxygenase-2 inhibition on renal function in elderly persons receiving a low-salt diet: a randomized, controlled trial. *Annals of Internal Medicine*, 133(1), 1-9.
- Takeda, K., & Akira, S. (2004). TLR Signaling Pathways. In *Seminars in Immunology* (pp. 3-9). Academic Press.
- Tanabe, T., & Tohnai, N. (2002). Cyclooxygenase isozymes and their gene structures and expression. *Prostaglandins & other lipid mediators*, 68, 95-114.
- Tham, C. L., Liew, C. Y., Lam, K. W., Mohamad, A. S., Kim, M. K., Cheah, Y. K., Zakaria, Z. Sulaiman, M., Lajis, N. H., & Israf, D. A. (2010). A synthetic curcuminoid derivative inhibits nitric oxide and proinflammatory cytokine synthesis. *European Journal of Pharmacology*, 628(1), 247-254.
- Thomas, G. (2011). Medicinal Chemistry: An Introduction. (pp. 245- 246). West Sussex, UK: John Wiley & Sons.
- Trifan, O. C., & Hla, T. (2003). Cyclooxygenase-2 modulates cellular growth and promotes tumorigenesis. *Journal of Cellular and Molecular Medicine*, 7(3), 207-222.
- Tsai, A. L., & Kulmacz, R. J. (2000). Tyrosyl radicals in prostaglandin H synthase-1 and-2. *Prostaglandins and Other Lipid Mediators*, 62(3), 231-254.
- Vane, J. R. (2000). The mechanism of action of anti-inflammatory drugs. In *Advances in Eicosanoid Research* (pp. 1-23). Springer Berlin Heidelberg.
- Veber, D. F., Johnson, S. R., Cheng, H. Y., Smith, B. R., Ward, K. W., & Kopple, K. D. (2002). Molecular properties that influence the oral bioavailability of drug candidates. *Journal of Medicinal Chemistry*, 45, 2615-2623.

- Vecchio, A. J., & Malkowski, M. G. (2011). The structure of NS-398 bound to cyclooxygenase-2. *Journal of Structural Biology*, 176(2), 254-258.
- Von Euler, U. S. (1936). On the specific vasodilating and plain muscle stimulating substances from accessory genital glands in man and certain animals (prostaglandin and vesiglandin). *The Journal of Physiology*, 88(2), 213-234.
- Vonkeman, H. E., & Van de Laar, M. A. (2010). Nonsteroidal anti-inflammatory drugs: adverse effects and their prevention. *Seminars in Arthritis and Rheumatism*, 39(4), 294-312.
- Wallace, J. L., & Soldato, P. D. (2003). The therapeutic potential of NO-NSAIDs. *Fundamental & Clinical Pharmacology*, 17(1), 11-20.
- Wang, J. L., Carter, J., Kiefer, J. R., Kurumbail, R. G., Pawlitz, J. L., Brown, D., Hartmann, J., Graneto, M. J., Seibert, K., & Talley, J. J. (2010). The novel benzopyran class of selective cyclooxygenase-2 inhibitors – part I: the first clinical candidate. *Bioorganic and Medicinal Chemistry Letters*, 20, 7155–7158.
- Wang, M. T., Honn, K. V., & Nie, D. (2007). Cyclooxygenases, prostanoids, and tumor progression. *Cancer and Metastasis Reviews*, 26(3-4), 525-534.
- Wadsworth, T. L., & Koop, D. R. (2001). Effects of Ginkgo biloba extract (EGb 761) and quercetin on lipopolysaccharide-induced release of nitric oxide. *Chemico-Biological Interactions*, 137(1), 43-58.
- Yadav, V. R., Prasad, S., Sung, B., & Aggarwal, B. B. (2011). The role of chalcones in suppression of NF- κ B-mediated inflammation and cancer. *International Immunopharmacology*, 11(3), 295-309.
- Yokoyama, C., Takai, T., & Tanabe, T. (1988). Primary structure of sheep prostaglandin endoperoxide synthase deduced from cDNA sequence. *Federation of European Biochemical Societies Letters*, 23, 347– 351.
- Ye, W., Zhang, H., Hillas, E., Kohan, D. E., Miller, R. L., Nelson, R. D., Honeggar, M., & Yang, T. (2006). Expression and function of COX isoforms in renal medulla: evidence for regulation of salt sensitivity and blood pressure. *American Journal of Physiology-Renal Physiology*, 290(2), F542-F549.