



UNIVERSITI PUTRA MALAYSIA

***CHARACTERIZATION AND ANTIFUNGAL PROPERTIES OF
OIL PALM MESOCARP FIBER SUPERHEATED STEAM CONDENSATE***

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By

NUR SHARMILA BINTI SHARIP

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

June 2016

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

CHARACTERIZATION AND ANTIFUNGAL PROPERTIES OF OIL PALM MESOCARP FIBER SUPERHEATED STEAM CONDENSATE

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Degradation of hemicellulose, low molecular weight lignin and some amount of cellulose during pretreatment of lignocellulose by steam treatment produces acids, furans and phenolic compounds. These compounds are known to have antimicrobial properties. It has been recently reported that superheated steam (SHS) can be used as pretreatment method to alter the structure of lignocellulose prior to bioconversion or biocomposite production. Since SHS is a dry steam, the mechanism of lignocellulose degradation is unclear and hence its degradation products are unknown. Hence this study was conducted, with the aim to characterize the condensate from SHS treatment of oil palm mesocarp fiber (OPMF) and subsequently evaluating the effect of SHS treatment temperature on antifungal properties of the condensate. Results showed that OPMF condensate obtained (OS sample) was light yellowish-brown in colour with average pH of 3.04 ± 0.02 . Detailed chemical compositional study by GCMS showed the presence of two different groups of compounds, with phenolic groups showed the most intense composition compared to the others. HPLC analysis showed that four types of carboxylic acids were generated during SHS treatment and formic acid was found as the most concentrated acid in the condensate (657 mg/L). Meanwhile, the concentrated OS sample (labeled as RF fraction) contained higher number of compounds (62 compounds), which were classified into ten different groups. Furthermore, the chemical composition of the condensates were exponentially increased by increases of SHS temperature; from 200°C to 240°C, while no significant increment was seen from 190°C to 200°C. This is in consistent with lignocellulose thermal degradation temperature. RF fraction of samples obtained from SHS treatment at 240°C showed 5.5 ± 0.17 , 5.2 ± 0.1 and 8.0 ± 0.4 inhibition ratios on *Bacillus cereus*, *Escherichia coli* and *Staphylococcus aureus*, respectively. For fungal species, growth inhibitory test by agar dilution method showed that the growth of *Aspergillus fumigatus* UPM2 and *Trichoderma asperellum* UPM1 were suppressed by 51.8%, and 45.5%, correspondingly. This is supported by spore germination inhibition whereby no germination of spore was seen for both fungi tested. It is interesting to note that the OPMF condensate managed to fully inhibit the growth of *Ganoderma boninense* UPM13, which is a type of fungus causing basal stem rot (BSR) disease at the oil palm plantation. Results obtained herewith are very

interesting as at present there is lack of information on the chemical composition of steam condensate obtained from SHS treatment of lignocellulose. The inhibitory effect of the OPMF SHS condensate on the growth of *G. boninense* UPM13 is indeed a new insight and should be used as a basis for future study on the development of antifungal agent for combating BSR disease.



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**PENCIRIAN DAN SIFAT ANTIKULAT KONDENSAT GENTIAN SABUT
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Degradasi hemiselulosa, lignin yang mempunyai berat molekul rendah dan sebahagian selulosa semasa prarawatan lignoselulosa oleh rawatan wap menghasilkan sebatian asid, furan dan fenolik. Sebatian ini diketahui mempunyai ciri anti-mikrob. Baru-baru ini telah dilaporkan bahawa stim panas lampau (SHS) boleh digunakan sebagai kaedah prarawatan untuk mengubah struktur lignoselulosa sebelum penukaran bio atau pengeluaran biokomposit. Berikutan SHS adalah wap kering, mekanisme degradasi lignoselulosanya tidak jelas dan maka, produk degradasinya tidak diketahui. Maka kajian ini telah dijalankan, bertujuan untuk mencirikan kondensat daripada rawatan SHS gentian sabut kelapa sawit (OPMF) dan kemudiannya menilai kesan suhu rawatan SHS pada ciri antimikrob kondensat. Hasil kajian menunjukkan bahawa kondensat OPMF yang diperolehi (sampel OS) adalah berwarna kuning-keperangan dengan purata pH 3.04 ± 0.02 . Kajian kimia terperinci oleh GCMS menunjukkan terdapat kehadiran dua kumpulan yang berbeza daripada sebatian tersebut, dengan kumpulan fenolik menunjukkan komposisi yang paling banyak berbanding yang lain. Analisis HPLC menunjukkan empat jenis asid karboksilik dijana semasa rawatan SHS dan asid formik merupakan asid yang paling pekat di dalam kondensat (657 mg/L). Sementara itu, sampel OS pekat (dilabelkan sebagai sampel RF) mengandungi bilangan sebatian yang lebih tinggi (62 sebatian), yang kemudiannya dikelaskan kepada sepuluh kumpulan yang berbeza. Tambahan lagi, komposisi kimia kondensat semakin meningkat dengan kenaikan suhu SHS; dari 200°C ke 240°C, manakala tiada kenaikan ketara dilihat dari 190°C ke 200°C. Ini adalah selari dengan suhu degradasi lignoselulosa. Sampel RF kondensat yang diperolehi daripada rawatan SHS 240°C masing-masing menunjukkan nisbah perencatan 5.5 ± 0.17 , 5.2 ± 0.1 dan 8.0 ± 0.4 bagi *Bacillus cereus*, *Escherichia coli* dan *Staphylococcus aureus*. Bagi spesies kulat, ujian perencatan pertumbuhan dengan kaedah pencairan agar menunjukkan bahawa pertumbuhan *Aspergillus fumigatus* UPM2 dan *Trichoderma asperellum* UPM1 dibantutkan sebanyak 51.8%, dan 45.5%. Ini disokong oleh perencatan spora di mana tiada percambahan spora dilihat bagi semua spora kulat yang diuji. Menariknya, kondensat OPMF berjaya menghalang sepenuhnya pertumbuhan *Ganoderma boninense* UPM13, iaitu sejenis kulat yang menyebabkan penyakit reput pangkal batang (BSR) di ladang kelapa sawit. Keputusan yang diperolehi dari

kajian ini adalah sangat menarik kerana pada masa kini terdapat kekurangan maklumat mengenai komposisi kimia wap kondensat daripada rawatan SHS lignoselulosa. Kesan rencatan daripada kondensat OPMF SHS kepada pertumbuhan *G. boninense* UPM13 merupakan satu penemuan baharu yang patut digunakan sebagai asas untuk kajian masa depan kepada pembangunan ejen antikulat bagi memerangi penyakit BSR.



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I certify that a Thesis Examination Committee has met on June 30, 2016 to conduct the final examination of Nur Sharmila Binti Sharip on her thesis entitled "Characterization and Antifungal Properties of Oil Palm Mesocarp Fiber Superheated Steam Condensate" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATION

Abs	Absorbance
DF 1	Distilled fraction 1
DF 2	Distilled fraction 2
GCMS	Gas performance mass spectroscopy
H ⁺	Hydrogen ion
H ₃ O ⁺	Hydronium ion
HPLC	High performance liquid chromatography
NA	Nutrient agar
OPEFB	Oil palm empty fruit bunch
OPF	Oil palm frond
OPMF	Oil palm mesocarp fiber
OPT	Oil palm trunk
OS	Original sample
PDA	Potato dextrose agar
PKS	Palm kernel shell
POME	Palm oil mill effluent
RF	Residue fraction
SHS	Superheated steam
UPM	Universiti Putra Malaysia
UV	Ultraviolet
UV-Vis	Ultraviolet-visible

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CHAPTER 1

INTRODUCTION

The concept of 'Waste to Wealth' focuses on utilization of unused resources to produce useful and high-end product. It has gained a lot of interest especially involving lignocellulose biomass. This biomass which came from agriculture and forestry industries, grasses and woody materials are largely available resources around the world. In Malaysia, largest biomass is contributed by palm oil industries which produce about 80 million dry tones of lignocellulose biomass annually (Agensi Inovasi Malaysia, 2011). Six types of biomass from oil palm industries are oil palm frond (OPF), oil palm trunk (OPT), oil palm empty fruit bunch (OPEFB), palm kernel shell (PKS), palm oil mill effluent (POME) and oil palm mesocarp fiber (OPMF) (Agensi Inovasi Malaysia, 2011).

To begin with, the utilization of these vast amounts of biomass as alternative sources to produce purposive products such as fermentable sugars, biofuel and biocomposite filler in various applications requires pretreatment in order to modify and alter its complex lignocellulose materials structure. This could be achieved by biological, chemical and physical pretreatments. For instance, it has been reported that acid and alkaline pretreatment of OPEFB managed to improve bioconversion of OPEFB as substrate for cellulase production due to the alteration of lignin content by the pretreatment (Umikalsom *et al.*, 1997). Hydrothermal pretreatment through hot water extraction has been reported to reduce hemicellulose and lignin content which in turn caused structural changes on the cellulose–hemicellulose–lignin matrix, resulting in the opening and expansion of specific surface area and pore volume and hence, provided better condition for hydrolysis of sugars (Zakaria *et al.*, 2015a; Zakaria *et al.*, 2015b).

SHS pretreatment has been recently reported as one of the pretreatment methods for oil palm biomass meant for sugar (Mahmud *et al.*, 2013; Bahrin *et al.*, 2012) and biocomposite (Then *et al.*, 2014a; Then *et al.*, 2014b; Nordin *et al.*, 2013) production. SHS can be defined as a type of unsaturated steam with temperature higher than its boiling point at given pressure (Nordin *et al.*, 2013; Bahrin *et al.*, 2012). It was found that hemicellulose was removed during SHS treatment of OPMF, which eventually provided better surface for polymer-fiber interaction in biocomposite making (Nordin *et al.*, 2013). In the same report, it was mentioned that lignin, hemicellulose, and cellulose degraded within temperature ranges of 160°C to 900°C, 220°C to 315°C and 315°C to 400°C, respectively. This indicates the possibility of having lignocellulose components degradation products as by-products during SHS pretreatment. Degradation products of lignocellulose varied from sugar degradation product of cellulose and hemicellulose such as sugar oligomers, furfural, acetic acid and formic acid, as well as phenol and extractive from lignin degradation (Garrote *et al.*, 2004). These products are toxic to microbial growth, thus it could be potentially used as antifungal agent (Cruz *et al.*, 2001; Garrote *et al.*, 2004). In relation to microorganism's inhibitory potential of lignocellulose degradation products, the

study on its application as antifungal agent could be very beneficial. The control of fungal growth is important in various areas of industrial, medical, pharmaceutical, food processing and environment.

Studies on pretreatment of oil palm biomass by SHS have been carried out by several researchers (Then *et al.*, 2014a; Then *et al.*, 2014b; Nordin *et al.*, 2013; Mahmud *et al.*, 2013; Bahrin *et al.*, 2012). It has been postulated that application of SHS for pretreatment had partially degraded the lignocellulose structure of the biomass into smaller compounds which can be easily volatile at high temperature. In regard with high temperature used during SHS, it is expected that the lignocellulose degradation products were vaporized and released out to the environment along with the steam. As it is known that lignocellulose degradation products have microbial growth inhibitory properties, therefore this research will be conducted, mainly to characterize and identify the degradation products obtained from SHS pretreatment of oil palm biomass, and eventually to determine its potential as an antifungal agent.

CHAPTER 1 of this thesis briefly introduces about the palm oil biomass, its utilization as alternative sources and the pretreatment methods; the SHS pretreatment; the degradation products from lignocellulose materials and its inhibition potential. Problem statements and objectives of this research were also included in this chapter. Details explanation and other's findings on topics related to this study were reviewed in CHAPTER 2. For the next chapter (CHAPTER 3), overview of the experimental procedure, materials and methods were explained. It covers the growth and maintenance of the bacteria (*Bacillus cereus*, *Escherichia coli*, *Staphylococcus aureus*) and fungi (*Trichoderma asperellum* UPM1, *Aspergillus fumigatus* UPM2, *Ganoderma boninense* UPM13) strain; the SHS treatment and condensation process of OPMF; fractionation of the OPMF condensate; analytical methods; antimicrobial screening and antifungal tests using agar dilution method and spore germination method.

CHAPTER 4 discusses on the findings of this research. Physical properties of OPMF condensate from 240°C SHS were characterized by means of its appearance and acidity while chemical properties were determined using HPLC and GCMS analysis. The effect of SHS treatment temperature on OPMF condensate composition and characteristics were also discussed. The selection of fraction with strongest inhibitory potential using antibacterial screening test was explained in this chapter. The antifungal properties of chosen OPMF condensate fraction on *Trichoderma asperellum* UPM1, *Aspergillus fumigatus* UPM2, and *Ganoderma boninense* UPM13 were also discussed in this chapter. Last chapter covers summary of the research and few recommendations for future works. It followed by references, appendices, biodata and list of publications.

The study aims to characterize condensate from SHS pretreatment of OPMF and subsequently identify the potential use of the steam condensate as antifungal agent. The specific objectives of this research are:

1. To characterize the condensates obtained from superheated steam (SHS) pretreatment of oil palm mesocarp fiber (OPMF) at various pretreatment temperatures.
2. To evaluate antifungal properties of OPMF SHS condensate in relation to the chemical composition of the condensate.



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REFERENCES

- Abdul Hamid, Z. A., Arai, T., Sitti Fatimah, M. R., Kosugi, A., Sulaiman, O., Hashim, R., ... & Murata, Y. (2014). Analysis of free sugar and starch in oil palm trunks (*Elaeis Guineensis Jacq.*) from various cultivars as a feedstock for bioethanol production. *International Journal of Green Energy*, (just-accepted).
- Abdul Khalil, H. P. S., Siti Alwani, M., Ridzuan, R., Kamarudin, H., & Khairul, a. (2008). Chemical Composition, Morphological Characteristics, and Cell Wall Structure of Malaysian Oil Palm Fibers. *Polymer-Plastics Technology and Engineering*, 47(3), 273–280.
- Abdullah, S. S. S., Shirai, Y., Bahrin, E. K., & Hassan, M. A. (2015). Fresh oil palm frond juice as a renewable, non-food, non-cellulosic and complete medium for direct bioethanol production. *Industrial Crops and Products*, 63, 357–361.
- Abe, T., & Miyashita, K. (2006). Surface sterilization of dried fishery produce in superheated steam and hot air. *Journal of the Japanese Society for Food Science and Technology (Japan)*.
- Abnisa, F., Arami-Niya, A., Daud, W. W., & Sahu, J. N. (2013). Characterization of bio-oil and bio-char from pyrolysis of palm oil wastes. *BioEnergy Research*, 6(2), 830-840.
- Agbor, V. B., Cicek, N., Sparling, R., Berlin, A., & Levin, D. B. (2011). Biomass pretreatment: fundamentals toward application. *Biotechnology Advances*, 29(6), 675–85.
- Agensi Inovasi Malaysia. (2011). National Biomass Strategy 2020 : New wealth creation for Malaysia' s palm oil industry. Agensi Inovasi Malaysia, Kuala Lumpur.
- Ali, N., Aziz, C. A. C., & Hassan, O. (2015). Alkali pretreatment and acid hydrolysis of coconut pulp and empty fruit bunch to produce glucose. *Jurnal Teknologi*, 74(7).
- Aliyu, A. S., Aziz, A. A., Yahya, A., & Latiff, Z. A. (2015). Characterization of oil palm frond juice and fiber as feedstock for biobutanol production. *Jurnal Teknos-2k*.
- Alizadeh, A., Alizadeh, O., Amari, G., & Zare, M. (2013). Essential oil composition, total phenolic content, antioxidant activity and antifungal properties of Iranian *Thymus daenensis* subsp. *daenensis* Celak. as in influenced by ontogenetical variation. *Journal of Essential Oil Bearing Plants*, 16(1), 59-70.
- Almeida, J. R., Modig, T., Petersson, A., Hähn-Hägerdal, B., Lidén, G., & Gorwa-Grauslund, M. F. (2007). Increased tolerance and conversion of inhibitors in lignocellulosic hydrolysates by *Saccharomyces cerevisiae*. *Journal of Chemical Technology and Biotechnology*, 82(4), 340-349.
- Ariols, M. G., Tejado, A., Blanco, M. A., Mondragon, I., & Labidi, J. (2009). Agricultural palm oil tree residues as raw material for cellulose, lignin and hemicelluloses production by ethylene glycol pulping process. *Chemical Engineering Journal*, 148(1), 106-114.

- Al-Samarrai, G., Singh, H., & Syarhabil, M. (2012). Evaluating eco-friendly botanicals (natural plant extracts) as alternatives to synthetic fungicides. *Annals of Agricultural and Environmental Medicine*, 19(4).
- Amborabé, B. E., Fleurat-Lessard, P., Chollet, J. F., & Roblin, G. (2002). Antifungal effects of salicylic acid and other benzoic acid derivatives towards *Eutypa lata*: structure–activity relationship. *Plant Physiology and Biochemistry*, 40(12), 1051-1060.
- Ang, S. K., Adibah, Y., Suraini, A. A., & Madihah, M. S. (2015). Potential Uses of Xylanase-Rich Lignocellulolytic Enzymes Cocktail for Oil Palm Trunk (OPT) Degradation and Lignocellulosic Ethanol Production. *Energy & Fuels*, 29(8), 5103-5116.
- Ang, S. K., Shaza, E. M., Adibah, Y., Suraini, A. A., & Madihah, M. S. (2013). Production of cellulases and xylanase by *Aspergillus fumigatus* SK1 using untreated oil palm trunk through solid state fermentation. *Process Biochemistry*, 48(9), 1293-1302.
- Anwar, Z., Gulfraz, M., & Irshad, M. (2014). Agro-industrial lignocellulosic biomass a key to unlock the future bio-energy: a brief review. *Journal Of Radiation Research and Applied Sciences*, 7(2), 163-173.
- Ariffin, H., Hassan, M. A., Kalsom, M. U., Abdullah, N., & Shirai, Y. (2008). Effect of physical, chemical and thermal pretreatments on the enzymatic hydrolysis of oil palm empty fruit bunch (OPEFB). *Journal of Tropical Agriculture and Food Science*, 36(2), 259-268.
- Asadullah, M., Ab Rasid, N. S., Kadir, S. A. S. A., & Azdarpour, A. (2013). Production and detailed characterization of bio-oil from fast pyrolysis of palm kernel shell. *Biomass and Bioenergy*, 59, 316-324.
- Asadullah, M., Adi, A. M., Suhada, N., Malek, N. H., Saringat, M. I., & Azdarpour, A. (2014). Optimization of palm kernel shell torrefaction to produce energy densified bio-coal. *Energy Conversion and Management*, 88, 1086-1093.
- Bahrin, E. K., Baharuddin, A. S., Ibrahim, M. F., Razak, M. N. A., Sulaiman, A., Aziz, S. A., Hassan, M.A., Shirai, Y., & Nishida, H. (2012). Physicochemical property changes and enzymatic hydrolysis enhancement of oil palm empty fruit bunches treated with superheated steam. *BioResources*, 7(2), 1784-1801.
- Bakar, A., Kartini, N., Hassan, M. A., & Mohamad Ghazali, F. (2010). Isolation and selection of appropriate cellulolytic mixed microbial cultures for cellulases production from oil palm empty fruit bunch. *Biotechnology*, 9(1), 73-78.
- Balat, M. (2011). Production of bioethanol from lignocellulosic materials via the biochemical pathway: a review. *Energy Conversion and Management*, 52(2), 858-875.
- Barchyn, D., & Cenkowski, S. (2014). Process analysis of superheated steam pre-treatment of wheat straw and its relative effect on ethanol selling price. *Biofuel Research Journal*, 1(4), 123-128.

- Baskaran, M., Hashim, R., Said, N., Raffi, S. M., Balakrishnan, K., Sudesh, K., & Sugimoto, T. (2012). Properties of binderless particleboard from oil palm trunk with addition of polyhydroxyalkanoates. *Composites Part B: Engineering*, 43(3), 1109-1116.
- Baskaran, M., Hashim, R., Sulaiman, O., Hiziroglu, S., Sato, M., & Sugimoto, T. (2015). Optimization of press temperature and time for binderless particleboard manufactured from oil palm trunk biomass at different thickness levels. *Materials Today Communications*, 3, 87-95.
- Bazargan, A., Kostić, M. D., Stamenković, O. S., Veljković, V. B., & McKay, G. (2015). A calcium oxide-based catalyst derived from palm kernel shell gasification residues for biodiesel production. *Fuel*, 150, 519-525.
- Bazargan, A., Rough, S. L., & McKay, G. (2014). Compaction of palm kernel shell biochars for application as solid fuel. *Biomass and Bioenergy*, 70, 489-497.
- Behera, S., Arora, R., Nandhagopal, N., & Kumar, S. (2014). Importance of chemical pretreatment for bioconversion of lignocellulosic biomass. *Renewable and Sustainable Energy Reviews*, 36, 91-106.
- Bennett, J. W., & Klich, M. A. (Eds.). (1992). *Aspergillus: Biology And Industrial Applications* (pp. 21-23). Boston: Butterworth-Heinemann.
- Bernardo, A. M., Dumoulin, E. D., Lebert, A. M., & Bimbenet, J. J. (1990). Drying of sugar beet fiber with hot air or superheated steam. *Drying Technology*, 8(4), 767-779.
- Björk, H., & Rasmuson, A. (1995). Moisture equilibrium of wood and bark chips in superheated steam. *Fuel*, 74(12), 1887-1890.
- Black, J. G. (2002). *Microbiology. Principles and Explorations* Jonh Wiley & Sons. Inc., 5th Ed.
- Blasco, R., & Alvarez, P. I. (1999). Flash drying of fish meals with superheated steam: isothermal process. *Drying Technology*, 17(4-5), 775-790.
- Bond, J. F., Mujumdar, A. S., Van Heiningen, A. R. P., & Douglas, W. J. M. (1994). Drying paper by impinging jets of superheated steam. Part 2: Comparison of steam and air as drying fluids. *The Canadian Journal of Chemical Engineering*, 72(3), 452-456.
- Borquez, R. M., Canales, E. R., & Quezada, H. R. (2008). Drying of fish press-cake with superheated steam in a pilot plant impingement system. *Drying Technology*, 26(3), 290-298.
- Bracey, D., Holyoak, C. D., & Coote, P. J. (1998). Comparison of the inhibitory effect of sorbic acid and amphotericin B on *Saccharomyces cerevisiae*: is growth inhibition dependent on reduced intracellular pH?. *Journal of Applied Microbiology*, 85(6), 1056-1066.
- Brady, J. E., Humiston, G. E., & Heikkinen, H. (1990). *General chemistry: Principles and Structure* (p. 425). Wiley.
- Branca, C., Giudicianni, P., & Di Blasi, C. (2003). GC/MS characterization of liquids generated from low-temperature pyrolysis of wood. *Industrial & Engineering Chemistry Research*, 42(14), 3190-3202.

- Brul, S., & Coote, P. (1999). Preservative agents in foods: mode of action and microbial resistance mechanisms. *International Journal of Food Microbiology*, 50(1), 1-17.
- Brunner, G. (2009). Near critical and supercritical water. Part I. Hydrolytic and hydrothermal processes. *The Journal of Supercritical Fluids*, 47(3), 373-381.
- Carvalho, W. S., Cunha, I. F., Pereira, M. S., & Ataíde, C. H. (2015). Thermal decomposition profile and product selectivity of analytical pyrolysis of sweet sorghum bagasse: Effect of addition of inorganic salts. *Industrial Crops and Products*, 74, 372-380.
- Cavaliere, S. J. (2005). *Manual of antimicrobial susceptibility testing*. Retrieved July 11, 2014, from <http://dspace.cus.ac.in/jspui/handle/1/1251>
- Cavendish, M. (2003). How it works: Science and technology. *SIJ: Marshall Cavendish*, 11.
- Chandel, A. K., Da Silva, S. S., & Singh, O. V. (2013). Detoxification of lignocellulose hydrolysates: biochemical and metabolic engineering toward white biotechnology. *BioEnergy Research*, 6(1), 388-401.
- Chandler, K., Deng, F., Dillow, A. K., Liotta, C. L., & Eckert, C. A. (1997). Alkylation reactions in near-critical water in the absence of acid catalysts. *Industrial & Engineering Chemistry Research*, 36(12), 5175-5179.
- Chandra, R. P., Bura, R., Mabee, W. E., Berlin, D. A., Pan, X., & Saddler, J. N. (2007). Substrate pretreatment: The key to effective enzymatic hydrolysis of lignocellulosics?. In *Biofuels* (pp. 67-93). Springer Berlin Heidelberg.
- Chang, S. H. (2014). An overview of empty fruit bunch from oil palm as feedstock for bio-oil production. *Biomass and Bioenergy*, 62, 174-181.
- Chang, V. S., & Holtzapfel, M. T. (2000). Fundamental factors affecting biomass enzymatic reactivity. In *Twenty-first Symposium on Biotechnology for Fuels and Chemicals* (pp. 5-37). Humana Press.
- Che Maail, C. M. H., Ariffin, H., Hassan, M. A., Shah, U. K. M., & Shirai, Y. (2014). Oil palm frond juice as future fermentation substrate: A feasibility study. *BioMed Research International*, 2014.
- Chen, H. Z. (2005). *Biotechnology of Lignocellulose*. Beijing: Chemical Industry Press.
- Chen, R., Wang, Y. Z., Liao, Q., Zhu, X., & Xu, T. F. (2013). Hydrolysates of lignocellulosic materials for biohydrogen production. *Biochemistry and Molecular Biology Reports*, 46(5), 244-251.
- Cherubini, F., & Strømman, A. H. (2011). Chemicals from lignocellulosic biomass: opportunities, perspectives, and potential of biorefinery systems. *Biofuels, Bioproducts and Biorefining*, 5(5), 548-561.
- Chin, M. J., Poh, P. E., Tey, B. T., Chan, E. S., & Chin, K. L. (2013). Biogas from palm oil mill effluent (POME): Opportunities and challenges from Malaysia's perspective. *Renewable and Sustainable Energy Reviews*, 26, 717-726.

- Chong, P. S., Jahim, J. M., Harun, S., Lim, S. S., Mutalib, S. A., Hassan, O., & Nor, M. T. M. (2013). Enhancement of batch biohydrogen production from prehydrolysate of acid treated oil palm empty fruit bunch. *International Journal of Hydrogen Energy*, 38(22), 9592-9599.
- Cowan, M. M. (1999). Plant products as antimicrobial agents. *Clinical Microbiology Reviews*, 12(4), 564-582.
- Cruz, J. M., Domínguez, J. M., Domínguez, H., & Parajó, J. C. (2001). Antioxidant and antimicrobial effects of extracts from hydrolysates of lignocellulosic materials. *Journal of Agricultural and Food Chemistry*, 49(5), 2459-2464.
- Dahlan, I. (2000). Oil palm frond, a feed for herbivores. *Asian Australasian Journal of Animal Sciences*, 13, 300-303.
- Demirbas, A. (2007). The influence of temperature on the yields of compounds existing in bio-oils obtained from biomass samples via pyrolysis. *Fuel Processing Technology*, 88(6), 591-597.
- Demirbas, A. (2009). Pyrolysis of biomass for fuels and chemicals. *Energy Sources, Part A*, 31(12), 1028-1037.
- Din, M. M., Ponraj, M., Van Loosdrecht, M., Ujang, Z., Chelliapan, S., & Zambare, V. (2014). Utilization of palm oil mill effluent for polyhydroxyalkanoate production and nutrient removal using statistical design. *International Journal of Environmental Science and Technology*, 11(3), 671-684.
- Douglas, W. J. M. (1994). Drying paper in superheated steam. *Drying Technology*, 12(6), 1341-1355.
- Duarte, C. L., Ribeiro, M. A., Oikawa, H., & Mori, M. N. (2013). Study of thermal treatment combined with radiation on the decomposition of polysaccharides in sugarcane bagasse. *Radiation Physics and Chemistry*, 84, 191-195.
- Dube, H. C. (2009). *Introduction To Fungi*, 3E. Vikas Publishing House Pvt Ltd.
- Ecolab. (2016). *B. cereus*. Retrieved September 25, 2015, from <http://www.ecolab.com/innovation/microbial-risks/b-cereus/>
- Eklund, T. (1985). The effect of sorbic acid and esters of p-hydroxybenzoic acid on the protonmotive force in *Escherichia coli* membrane vesicles. *Microbiology*, 131(1), 73-76.
- Elliott, G. M. (1992). *U.S. Patent No. 5,108,691*. Washington, DC: U.S. Patent and Trademark Office.
- Ellis, D. (2016). *Mycology Online*. Retrieved November 17, 2014, from http://www.mycology.adelaide.edu.au/Laboratory_Methods/Microscopy_Techniques_and_Stains/lactophenol.html
- Elustondo, D., Ahmed, S., & Oliveira, L. (2014). Drying western red cedar with superheated steam. *Drying Technology*, 32(5), 550-556.
- Eng, C. C., Ibrahim, N. A., Zainuddin, N., Ariffin, H., & Yunus, W. M. Z. W. (2014). Impact strength and flexural properties enhancement of methacrylate silane treated oil palm mesocarp fiber reinforced biodegradable hybrid composites. *The Scientific World Journal*, 2014.

- Eom, I. Y., Yu, J. H., Jung, C. D., & Hong, K. S. (2015). Efficient ethanol production from dried oil palm trunk treated by hydrothermolysis and subsequent enzymatic hydrolysis. *Biotechnology for Biofuels*, 8(1), 1.
- Esfahani, R. M., Wan Ab Karim Ghani, W. A., Mohd Salleh, M. A., & Ali, S. (2012). Hydrogen-rich gas production from palm kernel shell by applying air gasification in fluidized bed reactor. *Energy & Fuels*, 26(2), 1185-1191.
- Ezebor, F., Khairuddean, M., Abdullah, A. Z., & Boey, P. L. (2014). Oil palm trunk and sugarcane bagasse derived heterogeneous acid catalysts for production of fatty acid methyl esters. *Energy*, 70, 493-503.
- Fee, C. G. (2011). Management of *Ganoderma* Diseases in Oil Palm Plantations. *Sustainable Agriculture - an insight into Ganoderma*, Kuala Lumpur, 325-339.
- Fitzgerald, D. J., Stratford, M., & Narbad, A. (2003). Analysis of the inhibition of food spoilage yeasts by vanillin. *International Journal of Food Microbiology*, 86(1), 113-122.
- Freese, E., Sheu, C. W., & Galliers, E. (1973). Function of lipophilic acids as antimicrobial food additives. *Nature*, 241, 321-325.
- Friedman, M., Henika, P. R., & Mandrell, R. E. (2003). Antibacterial activities of phenolic benzaldehydes and benzoic acids against *Campylobacter jejuni*, *Escherichia coli*, *Listeria monocytogenes*, and *Salmonella enterica*. *Journal of Food Protection*, 66(10), 1811-1821.
- Fuente-Hernández, A., Lavoie, J. M., Corcos, P. O., & Beauchet, R. (2013). *Biofuels and Co-products Out of Hemicelluloses*. INTECH Open Access Publisher.
- Fyhr, C., & Rasmuson, A. (1997). Some aspects of the modelling of wood chips drying in superheated steam. *International Journal of Heat and Mass Transfer*, 40(12), 2825-2842.
- Galbe, M., & Zacchi, G. (2012). Pretreatment: the key to efficient utilization of lignocellulosic materials. *Biomass and Bioenergy*, 46, 70-78.
- Gan, P. P., Ng, S. H., Huang, Y., & Li, S. F. Y. (2012). Green synthesis of gold nanoparticles using palm oil mill effluent (POME): A low-cost and eco-friendly viable approach. *Bioresource Technology*, 113, 132-135.
- Garrote, G., Cruz, J. M., Moure, A., Dominguez, H., & Parajó, J. C. (2004). Antioxidant activity of byproducts from the hydrolytic processing of selected lignocellulosic materials. *Trends in Food Science & Technology*, 15(3), 191-200.
- Garrote, G., Dominguez, H., & Parajo, J. C. (1999). Hydrothermal processing of lignocellulosic materials. *European Journal of Wood and Wood Products*, 57(3), 191-202.
- Geddes, C. C., Peterson, J. J., Roslander, C., Zacchi, G., Mullinnix, M. T., Shanmugam, K. T., & Ingram, L. O. (2010). Optimizing the saccharification of sugar cane bagasse using dilute phosphoric acid followed by fungal cellulases. *Bioresource Technology*, 101(6), 1851-1857.
- Gelmini, L., Hilts, R. W., Petrucci, R. H., & Wismer, R. K. (2002). *General Chemistry Principles and Modern Applications: Selected Solutions Manual*. Prentice Hall.

- Goh, C. S., Tan, H. T., Lee, K. T., & Brosse, N. (2011). Evaluation and optimization of organosolv pretreatment using combined severity factors and response surface methodology. *Biomass and Bioenergy*, 35(9), 4025-4033.
- Halim, H.A. (2005). Recovery of useful products from pyroligneous acid (Bachelor Degree thesis, Universiti Putra Malaysia).
- Hamilton, G. (2006). *Kingdom of Life - Fungi*. Lorenz Educational Press.
- Hammel, K. E. (1997). Fungal degradation of lignin. In G. Cadisch & K. E. Giller (Eds.), *Driven by Nature: Plant Litter Quality and Decomposition*. (pp. 33–46). CAB International, Wallingford.
- Hamzah, F., Idris, A., & Shuan, T. K. (2011). Preliminary study on enzymatic hydrolysis of treated oil palm (Elaeis) empty fruit bunches fibre (EFB) by using combination of cellulase and β 1-4 glucosidase. *Biomass and Bioenergy*, 35(3), 1055-1059.
- Hanipah, S. H., Mohammed, M. A. P., & Baharuddin, A. S. (2016). Non-linear mechanical behaviour and bio-composite modelling of oil palm mesocarp fibres. *Composite Interfaces*, 23(1), 37-49.
- Harmini, S., Hutomo, M. D., Bahruddin, L., Millati, R., Cahyanto, M. N., Niklasson, C., & Taherzadeh, M. J. (2013). Fungal pretreatment of oil palm empty fruit bunch: effect of manganese and nitrogen. *Cellulose Chemical Technology*, 47(9-10), 751-757.
- Hasibuan, R., & Daud, W. R. W. (2009). Quality changes of superheated steam-dried fibers from oil palm empty fruit bunches. *Drying Technology*, 27(2), 194-200.
- Health Council of the Netherlands. (2001). *Disinfectants in consumer products*. Retrieved February 2, 2015, from U368/HvD/HB/679-E.
- Helm, R. (2000). *Reactions of Polysaccharides*. Retrieved December 17, 2015, from <http://dwb4.unl.edu/Chem/CHEM869E/CHEM869ELinks/www.chem.vt.edu/chem-dept/helm/3434WOOD/notes1/polyrxn.html>
- Hendriks, A. T. W. M., & Zeeman, G. (2009). Pretreatments to enhance the digestibility of lignocellulosic biomass. *Bioresource Technology*, 100(1), 10-18.
- Ho, A. L., Carneiro, F., Duarte, L. C., Roseiro, L. B., Charalampopoulos, D., & Rastall, R. A. (2014). Production and purification of xylooligosaccharides from oil palm empty fruit bunch fibre by a non-isothermal process. *Bioresource Technology*, 152, 526-529.
- Hongzhang, C., & Liying, L. (2007). Unpolluted fractionation of wheat straw by steam explosion and ethanol extraction. *Bioresource Technology*, 98(3), 666-676.
- Hörmeyer, H. F., Tailliez, P., Millet, J., Girard, H., Bonn, G., Bobleter, O., & Aubert, J. P. (1988). Ethanol production by *Clostridium thermocellum* grown on hydrothermally and organosolv-pretreated lignocellulosic materials. *Applied Microbiology and Biotechnology*, 29(6), 528-535.
- Hossain, N., & Jalil, R. (2015). sugar and bioethanol production from oil palm trunk (OPT). *Asia Pacific Journal of Energy and Environment*, 2(2), 2-2.

- Howe, R. A., & Andrews, J. M. (2012). BSAC standardized disc susceptibility testing method (version 11). *Journal of Antimicrobial Chemotherapy*, 67(12), 2783-2784.
- Hsu, T. C., Guo, G. L., Chen, W. H., & Hwang, W. S. (2010). Effect of dilute acid pretreatment of rice straw on structural properties and enzymatic hydrolysis. *Bioresource Technology*, 101(13), 4907-4913.
- Hu, F., & Ragauskas, A. (2012). Pretreatment and lignocellulosic chemistry. *Bioenergy Research*, 5(4), 1043-1066.
- Husseinsyah, S., Seong Chun, K., Hadi, A., & Ahmad, R. (2014). Effect of filler loading and coconut oil coupling agent on properties of low-density polyethylene and palm kernel shell eco-composites. *Journal of Vinyl and Additive Technology*.
- Ibrahim, M. F., Abd-Aziz, S., Razak, M. N. A., Phang, L. Y., & Hassan, M. A. (2012). Oil palm empty fruit bunch as alternative substrate for acetone–butanol–ethanol production by *Clostridium butyricum* EB6. *Applied Biochemistry and Biotechnology*, 166(7), 1615-1625.
- Ibrahim, M. F., Abd-Aziz, S., Yusoff, M. E. M., Phang, L. Y., & Hassan, M. A. (2015). Simultaneous enzymatic saccharification and ABE fermentation using pretreated oil palm empty fruit bunch as substrate to produce butanol and hydrogen as biofuel. *Renewable Energy*, 77, 447-455.
- Inayat, A., Ahmad, M. M., Mutalib, M. A., & Yusup, S. (2012). Process modeling for parametric study on oil palm empty fruit bunch steam gasification for hydrogen production. *Fuel Processing Technology*, 93(1), 26-34.
- Iyota, H., Nishimura, N., Onuma, T., & Nomura, T. (2001). Drying of sliced raw potatoes in superheated steam and hot air. *Drying Technology*, 19(7), 1411-1424.
- Jalč, D. (2002). Straw enrichment for fodder production by fungi. In *Agricultural Applications* (pp. 19-38). Springer Berlin Heidelberg.
- Jamradloedluk, J., Nathakaranakule, A., Soponronnarit, S., & Prachayawarakorn, S. (2007). Influences of drying medium and temperature on drying kinetics and quality attributes of durian chip. *Journal of Food Engineering*, 78(1), 198-205.
- Jeon, H., Kang, K. E., Jeong, J. S., Gong, G., Choi, J. W., Abimanyu, H., & Choi, G. W. (2014). Production of anhydrous ethanol using oil palm empty fruit bunch in a pilot plant. *Biomass and Bioenergy*, 67, 99-107.
- Joanne, M. W., Linda, M. S., & Christopher, J. W. (2008). Prescott, Harley and Klein's microbiology.
- Johansson, A., Fyhr, C., & Rasmuson, A. (1997). High temperature convective drying of wood chips with air and superheated steam. *International Journal of Heat and Mass Transfer*, 40(12), 2843-2858.
- Jönsson, L. J., & Martín, C. (2016). Pretreatment of lignocellulose: formation of inhibitory by-products and strategies for minimizing their effects. *Bioresource Technology*, 199, 103-112.
- Jönsson, L. J., Alriksson, B., & Nilvebrant, N. O. (2013). Bioconversion of lignocellulose: inhibitors and detoxification. *Biotechnology for Biofuels*, 6(1), 1.

- Joseph, C. G., Daud, W. M. A. W., Shane, Q. K., & Sanmugam, K. (2015). Parametric and adsorption kinetic studies of reactive black 5 removal from textile simulated wastewater using oil palm (*Elais guineensis*) empty fruit bunch. *Journal of Applied Sciences*, 15(8), 1103.
- Kang, R., Helms, R., Stout, M. J., Jaber, H., Chen, Z., & Nakatsu, T. (1992). Antimicrobial activity of the volatile constituents of *Perilla frutescens* and its synergistic effects with polygodial. *Journal of Agricultural and Food Chemistry*, 40(11), 2328-2330.
- Kamcharoen, A., Champreda, V., Eurwilaichitr, L., & Boonsawang, P. (2014). Screening and optimization of parameters affecting fungal pretreatment of oil palm empty fruit bunch (EFB) by experimental design. *International Journal of Energy and Environmental Engineering*, 5(4), 303-312.
- Kartal, S. N., Imamura, Y., Tsuchiya, F., & Ohsato, K. (2004). Preliminary evaluation of fungicidal and termiticidal activities of filtrates from biomass slurry fuel production. *Bioresource technology*, 95(1), 41-47.
- Kassim, M. A., Kheang, L. S., Bakar, N. A., Aziz, A. A., & Som, R. M. (2011). Bioethanol production from enzymatically saccharified empty fruit bunches hydrolysate using *Saccharomyces cerevisiae*. *Research Journal of Environmental Sciences*, 5(6), 573.
- Katritzky, A. R., Allin, S. M., & Siskin, M. (1996). Aquathermolysis: reactions of organic compounds with superheated water. *Accounts of Chemical Research*, 29(8), 399-406.
- Kelly-Yong, T. L., Lee, K. T., Mohamed, A. R., & Bhatia, S. (2007). Potential of hydrogen from oil palm biomass as a source of renewable energy worldwide. *Energy Policy*, 35(11), 5692-5701.
- Khalid, I., Sulaiman, O., Hashim, R., Razak, W., Jumhuri, N., & Rasat, M. S. M. (2015). Evaluation on layering effects and adhesive rates of laminated compressed composite panels made from oil palm (*Elaeis guineensis*) fronds. *Materials & Design*, 68, 24-28.
- Khairunniza-Bejo, S., & Vong, C. N. (2014). Detection of Basal Stem Rot (BSR) Infected Oil Palm Tree Using Laser Scanning Data. *Agriculture and Agricultural Science Procedia*, 2, 156-164.
- Khan, Z., Yusup, S., Ahmad, M. M., & Chin, B. L. F. (2014). Hydrogen production from palm kernel shell via integrated catalytic adsorption (ICA) steam gasification. *Energy Conversion and Management*, 87, 1224-1230.
- Kim, J. S., Lee, Y. Y., & Kim, T. H. (2016). A review on alkaline pretreatment technology for bioconversion of lignocellulosic biomass. *Bioresource Technology*, 199, 42-48.
- Kiriyama, T., Sasaki, H., Hashimoto, A., Kaneko, S., & Maeda, M. (2014). Size dependence of the drying characteristics of single lignite particles in superheated steam. *Metallurgical and Materials Transactions E*, 1(4), 349-363.
- Klinke, H. B., Ahring, B. K., Schmidt, A. S., & Thomsen, A. B. (2002). Characterization of degradation products from alkaline wet oxidation of wheat straw. *Bioresource Technology*, 82(1), 15-26.

- Komonkiat, I., & Cheirsilp, B. (2013). Felled oil palm trunk as a renewable source for biobutanol production by *Clostridium* spp. *Bioresource Technology*, 146, 200-207.
- Kostić, M. D., Bazargan, A., Stamenković, O. S., Veljković, V. B., & McKay, G. (2016). Optimization and kinetics of sunflower oil methanolysis catalyzed by calcium oxide-based catalyst derived from palm kernel shell biochar. *Fuel*, 163, 304-313.
- Kuhn, D. M., & Ghannoum, M. A. (2003). Indoor mold, toxigenic fungi, and *Stachybotrys chartarum*: infectious disease perspective. *Clinical Microbiology Reviews*, 16(1), 144-172.
- Kumar, P., Barrett, D. M., Delwiche, M. J., & Stroeve, P. (2009). Methods for pretreatment of lignocellulosic biomass for efficient hydrolysis and biofuel production. *Industrial & Engineering Chemistry Research*, 48(8), 3713-3729.
- Kumneadklang, S., Larpiattaworn, S., Niyasom, C., & Sompong, O. (2015). Bioethanol production from oil palm frond by simultaneous saccharification and fermentation. *Energy Procedia*, 79, 784-790.
- Kundu, A., Gupta, B. S., Hashim, M. A., & Redzwan, G. (2015). Taguchi optimization approach for production of activated carbon from phosphoric acid impregnated palm kernel shell by microwave heating. *Journal of Cleaner Production*, 105, 420-427.
- Lai, Z. Y., & Goh, S. M. (2015). Mechanical strength of binary oil palm kernel shell and HZSM-5 zeolite fuel pellets. *Fuel*, 150, 378-385.
- Laidler, K. J. (1984). The development of the Arrhenius equation. *Journal of Chemical Education*, 61(6), 494-498.
- Lalak, J., Kasprzycka, A., Martyniak, D., & Tys, J. (2016). Effect of biological pretreatment of *Agropyron elongatum* 'BAMAR' on biogas production by anaerobic digestion. *Bioresource Technology*, 200, 194-200.
- Lalitha, M.K. (2005). Manual on antimicrobial susceptibility testing. *Indiana Assoc. of Med. Microbiol.*, pp: 46.
- Lamaming, J., Hashim, R., Leh, C. P., Sulaiman, O., Sugimoto, T., & Nasir, M. (2015). Isolation and characterization of cellulose nanocrystals from parenchyma and vascular bundle of oil palm trunk (*Elaeis guineensis*). *Carbohydrate Polymers*, 134, 534-540.
- Lara-Ortiz, T., Riveros-Rosas, H., & Aguirre, J. (2003). Reactive oxygen species generated by microbial NADPH oxidase NoxA regulate sexual development in *Aspergillus nidulans*. *Molecular Microbiology*, 50(4), 1241-1255.
- Larsson, S., Palmqvist, E., Hahn-Hägerdal, B., Tengborg, C., Stenberg, K., Zacchi, G., & Nilvebrant, N. O. (1999). The generation of fermentation inhibitors during dilute acid hydrolysis of softwood. *Enzyme and Microbial Technology*, 24(3), 151-159.
- Larsson, S., Reimann, A., Nilvebrant, N. O., & Jönsson, L. J. (1999). Comparison of different methods for the detoxification of lignocellulose hydrolyzates of spruce. *Applied Biochemistry and Biotechnology*, 77(1-3), 91-103.

- Lasaga, A. (1998). *Kinetic Theory in The Earth Sciences*. Princeton University Press. Retrieved March 6, 2015, from <http://www.amazon.com/Kinetic-Theory-Sciences-Antonio-Lasaga/dp/0691037485>
- Lee, H. V., Hamid, S. B. A., & Zain, S. K. (2014). Conversion of lignocellulosic biomass to nanocellulose: structure and chemical process. *The Scientific World Journal*, 2014.
- Li, H., Qu, Y., Yang, Y., Chang, S., & Xu, J. (2016). Microwave irradiation—A green and efficient way to pretreat biomass. *Bioresource Technology*, 199, 34-41.
- Lin, R., Cheng, J., Ding, L., Song, W., Zhou, J., & Cen, K. (2015). Inhibitory effects of furan derivatives and phenolic compounds on dark hydrogen fermentation. *Bioresource Technology*, 196, 250-255.
- Lu, Y., Wei, X. Y., Cao, J. P., Li, P., Liu, F. J., Zhao, Y. P., & Wang, S. Z. (2012). Characterization of a bio-oil from pyrolysis of rice husk by detailed compositional analysis and structural investigation of lignin. *Bioresource Technology*, 116, 114-119.
- Mahmud, N. A. N., Baharuddin, A. S., Bahrin, E. K., Sulaiman, A., Naim, M. N., Zakaria, R., Hassan, M.A., Nishida, H., & Shirai, Y. (2013). Enzymatic saccharification of oil palm mesocarp fiber (OPMF) treated with superheated steam. *Bioresources*, 8(1), 1320-1331.
- Malaysian Palm Oil Board. (2014). Oil palm planted area as at December 2013. *Malaysian Palm Oil Board*, 2013 (December), 2014.
- Mansor, H., Khoo, K. C., & Harris, E. M. (1991). Chemical components of pyrolygneous acid liquors from the pyrolysis of rubberwood. *Pertanika*, 14(2), 153-158.
- Marcotullio, G., Krisanti, E., Giuntoli, J., & De Jong, W. (2011). Selective production of hemicellulose-derived carbohydrates from wheat straw using dilute HCl or FeCl₃ solutions under mild conditions. X-ray and thermogravimetric analysis of the solid residues. *Bioresource Technology*, 102(10), 5917-5923.
- Markowski, M., Cenkowski, S., Hatcher, D. W., Dexter, J. E., & Edwards, N. M. (2003). The effect of superheated-steam dehydration kinetics on textural properties of asian noodles. *Transactions of the American Society of Agricultural and Biological Engineers*, 46(2), 389.
- Martin, C., Klinke, H. B., & Thomsen, A. B. (2007). Wet oxidation as a pretreatment method for enhancing the enzymatic convertibility of sugarcane bagasse. *Enzyme and Microbial Technology*, 40(3), 426-432.
- Matsushita, Y. I., Sugamoto, K., Hidaka, K. I., & Matsui, T. (2002). Analysis of wood-vinegars prepared from sugi (*Cryptomeria japonica* D. Don) sapwood and its components. *Nippon Kagaku Kaishi*, (3), 385-392.
- Mayachiew, P., & Devahastin, S. (2008). Comparative evaluation of physical properties of edible chitosan films prepared by different drying methods. *Drying Technology*, 26(2), 176-185.
- McCall, J. M., & Douglas, W. J. M. (2006). Use of superheated steam drying to increase strength and bulk of papers produced from diverse commercial furnishes. *Drying Technology*, 24(2), 233-238.

- McDonnell, G. E. (2007). *Antisepsis, Disinfection, and Sterilization: Types, Action, and Resistance*. ASM Press. Washington, DC.
- McKendry, P. (2002). Energy production from biomass (part 1): overview of biomass. *Bioresource Technology*, 83(1), 37-46.
- MedicineNet. (2016). *Definition of Antimicrobial Agent*. Retrieved February 19, 2016, from <http://www.medicinenet.com/script/main/art.asp?articlekey=10204>.
- Meklin, T., Haugland, R. A., Reponen, T., Varma, M., Lummus, Z., Bernstein, D., Wymer, L.J., & Vesper, S. J. (2004). Quantitative PCR analysis of house dust can reveal abnormal mold conditions. *Journal of Environmental Monitoring*, 6(7), 615-620.
- Misono, I. I., Zain, N. K. M., Aziz, R. A., Vidyadharan, B., & Jose, R. (2015). Electrochemical properties of carbon from oil palm kernel shell for high performance supercapacitors. *Electrochimica Acta*, 174, 78-86.
- Moghadam, R. A., Yusup, S., Uemura, Y., Chin, B. L. F., Lam, H. L., & Al Shoaibi, A. (2014). Syngas production from palm kernel shell and polyethylene waste blend in fluidized bed catalytic steam co-gasification process. *Energy*, 75, 40-44.
- Mogk, D. (2012). *Phase Rule*. Retrieved February 13, 2016, from http://serc.carleton.edu/research_education/equilibria/phaserule.html.
- Mohaiyiddin, M. S., Ong, L. H., & Akil, H. M. (2013). Preparation and characterization of palm kernel shell/polypropylene biocomposites and their hybrid composites with nanosilica. *BioResources*, 8(2), 1539-1550.
- Mohamed, M., Yusup, S., Wahyudiono, W., Machmudah, S., & Goto, M. (2014). Effect of temperature on the extraction of bio-oil from oil palm mesocarp fiber using supercritical CO₂. *Jurnal Teknologi*, 69(4).
- Mosier, N., Wyman, C., Dale, B., Elander, R., Lee, Y. Y., Holtzapple, M., & Ladisch, M. (2005). Features of promising technologies for pretreatment of lignocellulosic biomass. *Bioresource Technology*, 96(6), 673-686.
- Moulthrop, J. S., Swatloski, R. P., Moyna, G., & Rogers, R. D. (2005). High-resolution ¹³C NMR studies of cellulose and cellulose oligomers in ionic liquid solutions. *Chemical Communications*, (12), 1557-1559.
- Mujumdar, A. S. (1990). Superheated Steam Drying—Principles, Practice and Potential for Use of Electricity, Canadian Electrical Association Report 817U671. *Montreal, Canada*.
- Mumtaz, T., Yahaya, N. A., Abd-Aziz, S., Yee, P. L., Shirai, Y., & Hassan, M. A. (2010). Turning waste to wealth-biodegradable plastics polyhydroxyalkanoates from palm oil mill effluent – a Malaysian perspective. *Journal of Cleaner Production*, 18(14), 1393-1402.
- Mun, S. P., & Ku, C. S. (2010). Pyrolysis GC-MS analysis of tars formed during the aging of wood and bamboo crude vinegars. *Journal of wood Science*, 56(1), 47-52.
- Mussatto, S. I., & Roberto, I. C. (2004). Alternatives for detoxification of diluted-acid lignocellulosic hydrolyzates for use in fermentative processes: a review. *Bioresource Technology*, 93(1), 1-10.

- Na, B. I., Kim, Y. H., Lim, W. S., Lee, S. M., Lee, H. W., & Lee, J. W. (2013). Torrefaction of oil palm mesocarp fiber and their effect on pelletizing. *Biomass and Bioenergy*, 52, 159-165.
- Nakatsu, T., Van Loveren, A. G., Kang, R. K., & Cilia, A. T. (1999). *U.S. Patent No. 5,965,518*. Washington, DC: U.S. Patent and Trademark Office.
- Naomoto Corporation (2015). Retrieved August 20, 2015, from http://www.naomoto.com/english/food/oven/dc_quto.html#
- Narendranath, N. V., Thomas, K. C., & Ingledew, W. M. (2001). Effects of acetic acid and lactic acid on the growth of *Saccharomyces cerevisiae* in a minimal medium. *Journal of Industrial Microbiology and Biotechnology*, 26(3), 171-177.
- Nikaido, H. (1994). Prevention of drug access to bacterial targets: permeability barriers and active efflux. *Science*, 264(5157), 382-388.
- Nitipan, S., Mamimin, C., Intrasingkha, N., Birkeland, N. K., & Sompong, O. (2014). Microbial community analysis of thermophilic mixed culture sludge for biohydrogen production from palm oil mill effluent. *International Journal of Hydrogen Energy*, 39(33), 19285-19293.
- Njoku, V. O., Islam, M. A., Asif, M., & Hameed, B. H. (2015). Adsorption of 2, 4-dichlorophenoxyacetic acid by mesoporous activated carbon prepared from H 3 PO 4-activated langsung empty fruit bunch. *Journal of Environmental Management*, 154, 138-144.
- Nor-Azemi, S. N. I., Fuadi, F. A., & Syed-Hassan, S. S. A. (2014). Effect of pretreatment on adsorption of nickel by oil palm mesocarp fiber. *Advanced Materials Research*, 906, 131-136.
- Nordin, N. I. A. A., Ariffin, H., Andou, Y., Hassan, M. A., Shirai, Y., Nishida, H., Yunus, W.M.Z.W., Karuppuchamy, S., & Ibrahim, N. A. (2013). Modification of oil palm mesocarp fiber characteristics using superheated steam treatment. *Molecules*, 18(8), 9132-9146.
- Öhgren, K., Bura, R., Saddler, J., & Zacchi, G. (2007). Effect of hemicellulose and lignin removal on enzymatic hydrolysis of steam pretreated corn stover. *Bioresource technology*, 98(13), 2503-2510.
- Olsson, L., & Hahn-Hägerdal, B. (1996). Fermentation of lignocellulosic hydrolysates for ethanol production. *Enzyme and Microbial Technology*, 18(5), 312-331.
- Pakowski, Z., & Adamski, R. (2011). On prediction of the drying rate in superheated steam drying process. *Drying Technology*, 29(13), 1492-1498.
- Palakawong, C., Sophanodora, P., Toivonen, P., & Delaquis, P. (2013). Optimized extraction and characterization of antimicrobial phenolic compounds from mangosteen (*Garcinia mangostana* L.) cultivation and processing waste. *Journal of the Science of Food and Agriculture*, 93(15), 3792-3800.
- Palmer, J. M., & Evans, C. S. (1983). The enzymic degradation of lignin by white-rot fungi. *Philosophical Transactions of the Royal Society of London B: Biological Sciences*, 300(1100), 293-303.

- Palmqvist, E., & Hahn-Hägerdal, B. (2000a). Fermentation of lignocellulosic hydrolysates. I: Inhibition and detoxification. *Bioresource Technology*, 74(1), 17–24.
- Palmqvist, E., & Hahn-Hägerdal, B. (2000b). Fermentation of lignocellulosic hydrolysates. II: Inhibitors and mechanisms of inhibition. *Bioresource Technology*, 74(1), 25–33.
- Pan, X., Xie, D., Yu, R. W., & Saddler, J. N. (2008). The bioconversion of mountain pine beetle-killed lodgepole pine to fuel ethanol using the organosolv process. *Biotechnology and Bioengineering*, 101(1), 39-48.
- Peng, F., Peng, P., Xu, F., & Sun, R. C. (2012). Fractional purification and bioconversion of hemicelluloses. *Biotechnology Advances*, 30(4), 879-903.
- Pimpaporn, P., Devahastin, S., & Chiewchan, N. (2007). Effects of combined pretreatments on drying kinetics and quality of potato chips undergoing low-pressure superheated steam drying. *Journal of Food Engineering*, 81(2), 318-329.
- Piñeros-Castro, Y., & Velásquez-Lozano, M. (2014). Biodegradation kinetics of oil palm empty fruit bunches by white rot fungi. *International Biodeterioration & Biodegradation*, 91, 24-28.
- Pogaku, R., Bono, A., & Chu, C. (Eds.). (2013). *Developments in Sustainable Chemical and Bioprocess Technology*. Springer US.
- Prachayawarakorn, S., Soponronnarit, S., Wetchacama, S., & Jaisut, D. (2002). Desorption isotherms and drying characteristics of shrimp in superheated steam and hot air. *Drying Technology*, 20(3), 669-684.
- Prawitwong, P., Kosugi, A., Arai, T., Deng, L., Lee, K. C., Ibrahim, D., Murata, Y., Sulaiman, O., Hashim, R., Sudesh, K., & Ibrahim, W. A. B. (2012). Efficient ethanol production from separated parenchyma and vascular bundle of oil palm trunk. *Bioresource Technology*, 125, 37-42.
- Pronyk, C., Cenkowski, S., & Muir, W. E. (2010). Drying kinetics of instant Asian noodles processed in superheated steam. *Drying Technology*, 28(2), 304-314.
- Pronyk, C., Cenkowski, S., Muir, W. E., & Lukow, O. M. (2008). Effects of superheated steam processing on the textural and physical properties of Asian noodles. *Drying Technology*, 26(2), 192-203.
- Purwandari, F. A., Sanjaya, A. P., Millati, R., Cahyanto, M. N., Horváth, I. S., Niklasson, C., & Taherzadeh, M. J. (2013). Pretreatment of oil palm empty fruit bunch (OPEFB) by N-methylmorpholine-N-oxide (NMMO) for biogas production: structural changes and digestion improvement. *Bioresource Technology*, 128, 461-466.
- Quiroz-Castañeda, R. E., & Folch-Mallol, J. L. (2011). Plant cell wall degrading and remodeling proteins: current perspectives. *Biotechnología Aplicada*, 28, 205-15.
- Rahmat, B., Pangesti, D., Natawijaya, D., & Sufyadi, D. (2014). Generation of wood-waste vinegar and its effectiveness as a plant growth regulator and pest insect repellent. *BioResources*, 9(4), 6350-6360.

- Raman, J. K., & Gnansounou, E. (2014). Ethanol and lignin production from Brazilian empty fruit bunch biomass. *Bioresource Technology*, 172, 241-248.
- Ramos, L. P. (2003). The chemistry involved in the steam treatment of lignocellulosic materials. *Química Nova*, 26(6), 863-871.
- Rashid, R. Z. A., Salleh, H. M., Ani, M. H., Yunus, N. A., Akiyama, T., & Purwanto, H. (2014). Reduction of low grade iron ore pellet using palm kernel shell. *Renewable Energy*, 63, 617-623.
- Rasmussen, H., Sørensen, H. R., & Meyer, A. S. (2014). Formation of degradation compounds from lignocellulosic biomass in the biorefinery: sugar reaction mechanisms. *Carbohydrate Research*, 385, 45-57.
- Razali, W. A. W., Baharuddin, A. S., Zaini, L. A., Mokhtar, M. N., Taip, F. S., & Zakaria, R. (2014). Effect of seed sludge quality using oil palm empty fruit bunch (OPEFB) bio-char for composting. *BioResources*, 9(2), 2739-2756.
- Rozman, H. D., Tay, G. S., Abubakar, A., & Kumar, R. N. (2001). Tensile properties of oil palm empty fruit bunch-polyurethane composites. *European Polymer Journal*, 37(9), 1759-1765.
- Sabri, M., Zubaidah, A., Samsudin, H., Yaacob, S. S., Yuzir, M., & Ali, M. (2015). Production of biogas from anaerobic co-digestion of oil palm mesocarp fibre and cattle manure: in batch. *Applied Mechanics and Materials* (Vol. 735, pp. 200-204).
- Sagehashi, M., Miyasaka, N., Shishido, H., & Sakoda, A. (2006). Superheated steam pyrolysis of biomass elemental components and Sugi (Japanese cedar) for fuels and chemicals. *Bioresource Technology*, 97(11), 1272-1283.
- Saidu, M., Yuzir, A., Salim, M. R., Azman, S., & Abdullah, N. (2014). Biological pre-treated oil palm mesocarp fibre with cattle manure for biogas production by anaerobic digestion during acclimatization phase. *International Biodeterioration & Biodegradation*, 95, 189-194.
- Saini, J. K., Saini, R., & Tewari, L. (2015). Lignocellulosic agriculture wastes as biomass feedstocks for second-generation bioethanol production: concepts and recent developments. *3 Biotech*, 5(4), 337-353.
- Salihu, A., Alam, Z., AbdulKarim, M. I., & Salleh, H. M. (2013). Suitability of using palm oil mill effluent as a medium for lipase production. *African Journal of Biotechnology*, 10(11), 2044-2052.
- Salmond, C. V., Kroll, R. G., & Booth, I. R. (1984). The effect of food preservatives on pH homeostasis in *Escherichia coli*. *Microbiology*, 130(11), 2845-2850.
- Satish, S., Mohana, D. C., Ranhavendra, M. P., & Raveesha, K. A. (2007). Antifungal activity of some plant extracts against important seed borne pathogens of *Aspergillus* sp. *An International Journal of Agricultural Technology*, 3(1), 109-119.
- Shafiei, M., Kumar, R., & Karimi, K. (2015). Pretreatment of lignocellulosic biomass. In *Lignocellulose-Based Bioproducts* (pp. 85-154). Springer International Publishing..

- Shafizadeh, F., Chin, P. P. S., & Goldstein, I. S. (1977). Wood technology, chemical aspects. In *American Chemical Society Symposium Series* (Vol. 43, pp. 57-81).
- Shahirah, M. N. N., Gimbutun, J., Pang, S. F., Zakria, R. M., Cheng, C. K., Chua, G. K., & Asras, M. F. F. (2015). Influence of nutrient addition on the bioethanol yield from oil palm trunk sap fermented by *Saccharomyces cerevisiae*. *Journal of Industrial and Engineering Chemistry*, 23, 213-217.
- Shallom, D., & Shoham, Y. (2003). Microbial hemicellulases. *Current Opinion in Microbiology*, 6(3), 219-228.
- Shamsudin, S., Shah, U. K. M., Zainudin, H., Abd-Aziz, S., Kamal, S. M. M., Shirai, Y., & Hassan, M. A. (2012). Effect of steam pretreatment on oil palm empty fruit bunch for the production of sugars. *Biomass and Bioenergy*, 36, 280-288.
- Shen, D. K., & Gu, S. (2009). The mechanism for thermal decomposition of cellulose and its main products. *Bioresource Technology*, 100(24), 6496-6504.
- Shinoj, S., Visvanathan, R., Panigrahi, S., & Kochubabu, M. (2011). Oil palm fiber (OPF) and its composites: a review. *Industrial Crops and Products*, 33(1), 7-22.
- Sinclair, J. B., & Dhingra, O. D. (1995). *Basic Plant Pathology Methods*. CRC press.
- Sindhu, R., Binod, P., & Pandey, A. (2016). Biological pretreatment of lignocellulosic biomass – an overview. *Bioresource Technology*, 199, 76-82.
- Singh, J., Suhag, M., & Dhaka, A. (2015). Augmented digestion of lignocellulose by steam explosion, acid and alkaline pretreatment methods: a review. *Carbohydrate Polymers*, 117, 624-631.
- Singh, L., Siddiqui, M. F., Ahmad, A., Rahim, M. H. A., Sakinah, M., & Wahid, Z. A. (2013). Application of polyethylene glycol immobilized *Clostridium* sp. LS2 for continuous hydrogen production from palm oil mill effluent in upflow anaerobic sludge blanket reactor. *Biochemical Engineering Journal*, 70, 158-165.
- Singh, R., Shukla, A., Tiwari, S., & Srivastava, M. (2014). A review on delignification of lignocellulosic biomass for enhancement of ethanol production potential. *Renewable and Sustainable Energy Reviews*, 32, 713-728.
- Sinha, A. P., Singh, K., & Mukhopadhyay, A. N. (1988). *Soil fungicides. Volume I*. CRC Press, Inc.
- Sjostrom, E. (2013). *Wood chemistry: Fundamentals and Applications*. Elsevier.
- Sölken, W. (2008). *Steam and Condensate systems - What is Steam?*. Retrieved January 2, 2016, from http://www.wermac.org/steam/steam_part3.html
- Somjai, T., Achariyaviriya, S., Achariyaviriya, A., & Namsanguan, K. (2009). Strategy for longan drying in two-stage superheated steam and hot air. *Journal of Food Engineering*, 95(2), 313-321.

- Sompong, O., Boe, K., & Angelidaki, I. (2012). Thermophilic anaerobic co-digestion of oil palm empty fruit bunches with palm oil mill effluent for efficient biogas production. *Applied Energy*, 93, 648-654.
- Sotome, I., Takenaka, M., Koseki, S., Ogasawara, Y., Nadachi, Y., Okadome, H., & Isobe, S. (2009). Blanching of potato with superheated steam and hot water spray. *LWT-Food Science and Technology*, 42(6), 1035-1040.
- Souza, N. F., Pinheiro, J. A., Silva, P., Morais, J. P. S., de Souza, F., de Sá Moreira, M., Brígida, A.I.S., Muniz, C.R., & de Freitas Rosa, M. (2015). Development of chlorine-free pulping method to extract cellulose nanocrystals from pressed oil palm mesocarp fibers. *Journal of Biobased Materials and Bioenergy*, 9(3), 372-379.
- Speckhahn, A., Srzednicki, G., & Desai, D. K. (2010). Drying of beef in superheated steam. *Drying Technology*, 28(9), 1072-1082.
- Sreekala, M. S., Kumaran, M. G., & Thomas, S. (1997). Oil palm fibers: Morphology, chemical composition, surface modification, and mechanical properties. *Journal of Applied Polymer Science*, 66(5), 821-835.
- Sun, F., & Chen, H. (2008). Organosolv pretreatment by crude glycerol from oleochemicals industry for enzymatic hydrolysis of wheat straw. *Bioresource Technology*, 99(13), 5474-5479.
- Sun, Y., & Cheng, J. (2002). Hydrolysis of lignocellulosic materials for ethanol production: a review. *Bioresource Technology*, 83(1), 1-11.
- Syaima, M. T. S., Ong, K. H., Noor, I. M., Zamratul, M. I. M., Brahim, S. A., & Hafizul, M. M. (2015). The synthesis of bio-lubricant based oil by hydrolysis and non-catalytic of palm oil mill effluent (POME) using lipase. *Renewable and Sustainable Energy Reviews*, 44, 669-675.
- Taechapairoj, C., Prachayawarakorn, S., & Soponronnarit, S. (2004). Characteristics of rice dried in superheated-steam fluidized-bed. *Drying Technology*, 22(4), 719-743.
- Taherzadeh, M. J., & Karimi, K. (2007a). Acid-based hydrolysis processes for ethanol from lignocellulosic materials: a review. *BioResources*, 2(3), 472-499.
- Taherzadeh, M. J., & Karimi, K. (2007b). Enzymatic-based hydrolysis processes for ethanol from lignocellulosic materials: A review. *BioResources*, 2(4), 707-738.
- Takacs, E., Wojnarovits, L., Földváry, C., Hargittai, P., Borsa, J., & Sajo, I. (2000). Effect of combined gamma-irradiation and alkali treatment on cotton-cellulose. *Radiation Physics and Chemistry*, 57(3), 399-403.
- Tang, Z., & Cenkowski, S. (2001). Equilibrium moisture content of spent grains in superheated steam under atmospheric pressure. *Transactions of the American Society of Agricultural and Biological Engineers*, 44(5), 1261.
- Tang, Z., Cenkowski, S., & Muir, W. E. (2000). Dehydration of sugar-beet pulp in superheated steam and hot air. *Transactions of the American Society of Agricultural and Biological Engineers*, 43(3), 685.

- Then, Y. Y., Ibrahim, N. A., Zainuddin, N., Ariffin, H., & Wan Yunus, W. M. Z. (2013). Oil palm mesocarp fiber as new lignocellulosic material for fabrication of polymer/fiber biocomposites. *International Journal of Polymer Science*, 2013.
- Then, Y. Y., Ibrahim, N. A., Zainuddin, N., Ariffin, H., Chieng, B. W., & Yunus, W. M. Z. W. (2015). influence of fiber content on properties of oil palm mesocarp fiber/poly (butylene succinate) biocomposites. *BioResources*, 10(2), 2949-2968.
- Then, Y. Y., Ibrahim, N. A., Zainuddin, N., Ariffin, H., Yunus, W. M. Z. W., & Chieng, B. W. (2014a). Surface modifications of oil palm mesocarp fiber by superheated steam, alkali, and superheated steam-alkali for biocomposite applications. *BioResources*, 9(4), 7467-7483.
- Then, Y. Y., Ibrahim, N. A., Zainuddin, N., Ariffin, H., Yunus, W. M. Z. W., & Chieng, B. W. (2014b). The influence of green surface modification of oil palm mesocarp fiber by superheated steam on the mechanical properties and dimensional stability of oil palm mesocarp fiber/poly (butylene succinate) biocomposite. *International Journal of Molecular Sciences*, 15(9), 15344-15357.
- Tian, X F., Fang, Z., & Guo, F. (2012). Impact and prospective of fungal pre-treatment of lignocellulosic biomass for enzymatic hydrolysis. *Biofuels, Bioproducts and Biorefining*, 6(3), 335-350.
- Todar, K. (2012). *Staphylococcus aureus* and Staphylococcal Disease. Retrieved September 25, 2015, from <http://textbookofbacteriology.net/staph.html>
- Traub, W. H., & Leonhard, B. (1994). Agar Disk Diffusion (Bauer-Kirby) Tests with Various Fastidious and Nonfastidious Reference (ATCC) Strains: Comparison of Several Agar Media. *Chemotherapy*, 40(6), 374-383.
- Umikalsom, M. S., Ariff, A. B., Shamsuddin, Z. H., Tong, C. C., Hassan, M. A., & Karim, M. I. A. (1997). Production of cellulase by a wild strain of *Chaetomium globosum* using delignified oil palm empty-fruit-bunch fibre as substrate. *Applied Microbiology and Biotechnology*, 47(5), 590-595.
- United Nations Environment Programme. (2012). Converting Waste Oil Palm Trees Into a Resource. *United Nations Environment Programme*.
- Urbaniec, K., & Malczewski, J. (1997). Experimental investigations of beet pulp drying in superheated steam under pressure. *Drying Technology*, 15(6-8), 2004-2013.
- van Deventer, H. C. (1997). Feasibility of energy efficient steam drying of paper and textile including process integration. *Applied Thermal Engineering*, 17(8), 1035-1041.
- van Kuijk, S. J. A., Sonnenberg, A. S. M., Baars, J. J. P., Hendriks, W. H., & Cone, J. W. (2015). Fungal treated lignocellulosic biomass as ruminant feed ingredient: a review. *Biotechnology Advances*, 33(1), 191-202.
- Wang, W. D., & Xu, S. Y. (2007). Degradation kinetics of anthocyanins in blackberry juice and concentrate. *Journal of Food Engineering*, 82(3), 271-275.

- Wang, X., Hu, J., & Zeng, J. (2012). Steam explosion pulping of oil palm empty fruit bunch fiber. *BioResources*, 7(1), 1008-1015.
- Wasserscheid, P., & Keim, W. (2000). Ionic liquids—new “solutions” for transition metal catalysis. *Angewandte Chemie International Edition*, 39(21), 3772-3789.
- Werner, K., Pommer, L., & Broström, M. (2014). Thermal decomposition of hemicelluloses. *Journal of Analytical and Applied Pyrolysis*, 110, 130-137.
- Wimmerstedt, R. (1995). Steam drying—history and future. *Drying Technology*, 13(5-7), 1059-1076.
- Wirjosentono, B., Guritno, P., & Ismail, H. (2004). Oil palm empty fruit bunch filled polypropylene composites. *International Journal of Polymeric Materials*, 53(4), 295-306.
- Wititsiri, S. (2011). Production of wood vinegars from coconut shells and additional materials for control of termite workers, *Odontotermes* sp. and striped mealy bugs, *Ferrisia virgata*. *Sonklanakarin Journal of Science and Technology*, 33(3), 349.
- Yamada, H., Tanaka, R., Sulaiman, O., Hashim, R., Hamid, Z. A. A., Yahya, M. K. A., Kosugi, A., Arai, T., Murata, Y., Nirasawa, S., & Yamamoto, K. (2010). Old oil palm trunk: a promising source of sugars for bioethanol production. *Biomass and Bioenergy*, 34(11), 1608-1613.
- Yamaguchi, H., Hiratani, T., Iwata, K., & Yamamoto, Y. (1982). Studies on the mechanism of antifungal action of aculeacin A. *The Journal of Antibiotics*, 35(2), 210-219.
- Yan, W., Acharjee, T. C., Coronella, C. J., & Vásquez, V. R. (2009). Thermal pretreatment of lignocellulosic biomass. *Environmental Progress & Sustainable Energy*, 28(3), 435-440.
- Yang, H., Yan, R., Chen, H., Lee, D. H., & Zheng, C. (2007). Characteristics of hemicellulose, cellulose and lignin pyrolysis. *Fuel*, 86(12), 1781-1788.
- Yasin, N. H. M., Fukuzaki, M., Maeda, T., Miyazaki, T., Maail, C. M. H. C., Ariffin, H., & Wood, T. K. (2013). Biohydrogen production from oil palm frond juice and sewage sludge by a metabolically engineered *Escherichia coli* strain. *International Journal of Hydrogen Energy*, 38(25), 10277-10283.
- Yulia, E. (2005). Antifungal activity of plant extracts and oils against fungal pathogens of pepper (*Piper nigrum* L.), cinnamon (*Cinnamomum zeylanicum* Blume), and turmeric (*Curcuma domestica* Val.) (Doctoral dissertation, James Cook University).
- Yunos, N. S. H. M., Baharuddin, A. S., Yunos, K. F. M., Naim, M. N., & Nishida, H. (2012). Physicochemical property changes of oil palm mesocarp fibers treated with high-pressure steam. *BioResources*, 7(4), 5983-5994.
- Zahari, M. A. K. M., Abdullah, S. S. S., Roslan, A. M., Ariffin, H., Shirai, Y., & Hassan, M. A. (2014). Efficient utilization of oil palm frond for bio-based products and biorefinery. *Journal of Cleaner Production*, 65, 252-260.
- Zahari, M. A. K. M., Zakaria, M. R., Ariffin, H., Mokhtar, M. N., Salihon, J., Shirai, Y., & Hassan, M. A. (2012). Renewable sugars from oil palm frond juice as an alternative novel fermentation feedstock for value-added products. *Bioresource technology*, 110, 566-571.

- Zaidan, M. R., Noor Rain, A., Badrul, A. R., Adlin, A., Norazah, A., & Zakiah, I. (2005). In vitro screening of five local medicinal plants for antibacterial activity using disc diffusion method. *Tropical Biomedicine*, 22(2), 165-170.
- Zainudin, M. H. M., Hassan, M. A., Tokura, M., & Shirai, Y. (2013). Indigenous cellulolytic and hemicellulolytic bacteria enhanced rapid co-composting of lignocellulose oil palm empty fruit bunch with palm oil mill effluent anaerobic sludge. *Bioresource Technology*, 147, 632-635.
- Zakaria, M. R., Ariffin, H., Johar, N. A. M., Abd-Aziz, S., Nishida, H., Shirai, Y., & Hassan, M. A. (2010). Biosynthesis and characterization of poly (3-hydroxybutyrate-co-3-hydroxyvalerate) copolymer from wild-type *Comamonas* sp. EB172. *Polymer Degradation and Stability*, 95(8), 1382-1386.
- Zakaria, M. R., Hirata, S., & Hassan, M. A. (2014). Combined pretreatment using alkaline hydrothermal and ball milling to enhance enzymatic hydrolysis of oil palm mesocarp fiber. *Bioresource Technology*, 169, 236-243.
- Zakaria, M. R., Hirata, S., & Hassan, M. A. (2015a). Hydrothermal pretreatment enhanced enzymatic hydrolysis and glucose production from oil palm biomass. *Bioresource Technology*, 176, 142-8.
- Zakaria, M. R., Norrahim, M. N. F., Hirata, S., & Hassan, M. A. (2015b). Hydrothermal and wet disk milling pretreatment for high conversion of biosugars from oil palm mesocarp fiber. *Bioresource Technology*, 181, 263-269..
- Zakaria, M. R., Tabatabaei, M., Ghazali, F. M., Abd-Aziz, S., Shirai, Y., & Hassan, M. A. (2010). Polyhydroxyalkanoate production from anaerobically treated palm oil mill effluent by new bacterial strain *Comamonas* sp. EB172. *World Journal of Microbiology and Biotechnology*, 26(5), 767-774.
- Zanirun, Z., Bahrin, E. K., Lai-Yee, P., Hassan, M. A., & Abd-Aziz, S. (2015). Enhancement of fermentable sugars production from oil palm empty fruit bunch by ligninolytic enzymes mediator system. *International Biodeterioration & Biodegradation*, 105, 13-20.
- Zavel, M., Bross, D., Funke, M., Büchs, J., & Spiess, A. C. (2009). High-throughput screening for ionic liquids dissolving (ligno-) cellulose. *Bioresource Technology*, 100(9), 2580-2587.
- Zhang, D., Ong, Y. L., Li, Z., & Wu, J. C. (2012). Optimization of dilute acid-catalyzed hydrolysis of oil palm empty fruit bunch for high yield production of xylose. *Chemical Engineering Journal*, 181, 636-642.
- Zhang, K., Pei, Z., & Wang, D. (2016). Organic solvent pretreatment of lignocellulosic biomass for biofuels and biochemicals: A review. *Bioresource Technology*, 199, 21-33.
- Zhao, W., Yang, R., Zhang, Y., & Wu, L. (2012). Sustainable and practical utilization of feather keratin by an innovative physicochemical pretreatment: high density steam flash-explosion. *Green Chemistry*, 14(12), 3352-3360.
- Zhao, X., Cheng, K., & Liu, D. (2009). Organosolv pretreatment of lignocellulosic biomass for enzymatic hydrolysis. *Applied Microbiology and Biotechnology*, 82(5), 815-827.

- Zhu, S., Wu, Y., Chen, Q., Yu, Z., Wang, C., Jin, S., Ding, Y., & Wu, G. (2006). Dissolution of cellulose with ionic liquids and its application: a mini-review. *Green Chemistry*, 8(4), 325-327.
- Zhuang, X., Wang, W., Yu, Q., Qi, W., Wang, Q., Tan, X., Zhou, G., & Yuan, Z. (2016). Liquid hot water pretreatment of lignocellulosic biomass for bioethanol production accompanying with high valuable products. *Bioresource Technology*, 199, 68-75.



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