



UNIVERSITI PUTRA MALAYSIA

***BIOTRANSFORMATION OF RAMBUTAN (*Nephelium lappaceum* L.)
SEED INTO A COCOA POWDER-LIKE PRODUCT***

CHAI KONG FEI

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By

CHAI KONG FEI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Philosophy**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

BIOTRANSFORMATION OF RAMBUTAN (*Nephelium lappaceum* L.) SEED INTO A COCOA POWDER-LIKE PRODUCT

By

CHAI KONG FEI

November 2017

Chairman: Professor Hasanah Mohd Ghazali, PhD
Faculty : Food Science and Technology

Rambutan (*Nephelium lappaceum* L.) is an exotic fruit originally found in Southeast Asia. There is a glut of rambutan every year which leads to wastage. A novel way to reduce wastage is to convert rambutan seeds into a cocoa powder-like product. Thus, the objectives of this study were to select the best rambutan variety for fermentation by comparing the physicochemical properties of pulp and seed of eleven varieties of rambutan fruit with those of cocoa pulp and seed, to determine the physicochemical properties of rambutan fruit sweatings, seed and seed fat during solid state fermentation at different fermentation times and turning intervals, to optimize the roasting process of rambutan seed from fermented and dried rambutan fruit and determine the physicochemical properties of rambutan seed powder, and to determine the toxicology safety of the roasted rambutan seed powder using brine shrimp lethality assay.

Eleven varieties of rambutan were examined in this study, and most of the physicochemical properties of a wild type rambutan, WT1, were found to be similar to those of cocoa bean. However, due to subsequent unavailability of the variety, rambutan Clone R4 was chosen to be the best variety to be investigated on the effect of fermentation as its titratable acidity was not significantly different from that of cocoa bean and the total sugar content of the pulp is comparable to that of cocoa pulp. Besides, the seed had the highest crude fat content (39.13 %) and lowest saponin content (14.27 mg soya saponin/100 g).

Peeled rambutan fruit Clone R4 was subjected to natural fermentation in covered perforated plastic boxes for 0, 1, 3, 5, 7 and 10 days at room temperature. Fermentation time significantly affected the physicochemical properties of sweatings (liquid released naturally from the fruit), seed and seed fat. The study showed that 8 days of fermentation was regarded as sufficient to produce well fermented seeds. The effect of

turning intervals during fermentation for 8 days did not lead to significant changes in the properties of sweatings. Also, there was no significant difference on most of the physicochemical properties of the fermented seeds, between 24 and 48 hours of turning intervals. Considering practical aspects of fermentation in both small and large scale contexts, turning at 48 hours intervals for eight days is recommended. Fermentation for 8 days is also recommended as it served to eliminate microorganisms that are potentially harmful.

Roasting conditions (time and temperature) of seeds after fermentation for 8 days with a 48-hour turning interval significantly affected the physicochemical properties of the fermented seeds. GCMS analysis showed that roasted seed powder contained pyrazines, indicating that the powder has aroma and flavor similar to that of cocoa powder. Results from the brine shrimp lethality assay showed that the LC_{50} of roasted rambutan seed powder extract was $7.07 \times 10^4 \mu\text{g/mL}$ and is considered as non-toxic. The overall findings of the study indicate that it is possible to obtain a cocoa powder-like product from the seeds of rambutan that had undergone fermentation that is relatively safe for consumption.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia Sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

BIOTRANSFORMASI BIJI RAMBUTAN (*Nephelium lappaceum* L.) KEPADA PRODUK SEPERTI SERBUK KOKO

Oleh

CHAI KONG FEI

November 2017

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Rambutan (*Nephelium lappaceum* L.) adalah sejenis buah eksotik yang berasal dari Asia Tenggara. Buah rambutan yang berlebihan telah menyebabkan pembaziran setiap tahun. Salah satu cara novel untuk mengurangkan pembaziran adalah dengan menghasilkan produk seperti serbuk koko dengan menggunakan biji rambutan. Oleh itu, objektif kajian ini adalah memilih varieti rambutan yang paling sesuai untuk proses fermentasi dengan membandingkan ciri-ciri fizikokimia isi dan biji sebelas varieti rambutan dengan isi dan biji koko, menentukan ciri-ciri fizikokimia cecair fermentasi, biji dan lemak biji rambutan semasa fermentasi substrat pepejal pada masa fermentasi dan selang pengacauan yang berbeza, mengoptimumkan proses pemangangan biji rambutan daripada buah rambutan yang telah difermentasi dan dikeringkan dan menentukan ciri-ciri fizikokimia serbuk biji rambutan, serta menentukan toksikologi keselamatan serbuk biji rambutan yang telah dipanggang dengan menggunakan ujian ketosikan anak udang.

Sebelas varieti rambutan telah dikaji dan didapati bahawa kebanyakan ciri-ciri fizikokimia rambutan jenis liar, WT1, sama dengan koko. Akan tetapi, disebabkan ketidakterdapatannya WT1 pada musim tuai yang seterusnya, rambutan klon R4 telah dipilih sebagai varieti yang paling baik untuk dikaji kerana keasidan tertitratnya tidak berbeza secara signifikan dengan koko dan jumlah gula isinya sebanding dengan koko. Selain itu, biji klon R4 mempunyai kandungan lemak mentah yang tertinggi (39.13 %) dan kandungan saponin yang terendah (14.27 mg soya saponin/100 g).

Buah rambutan Klon R4 yang telah dikupas difermentasi secara semula jadi dalam bekas plastik selama 0, 1, 3, 5, 7 dan 10 hari pada suhu bilik. Masa fermentasi menjejaskan ciri-ciri fizikokimia cecair fermentasi, biji dan lemak biji secara signifikan. Keputusan menunjukkan fermentasi sebanyak 8 hari adalah memadai untuk

menghasilkan biji yang difermentasi dengan baik. Selang pengacauan tidak memberi kesan yang signifikan kepada ciri-ciri fizikokimia cecair fermentasi semasa buah difermentasi selama 8 hari. Di samping itu, tiada perbezaan signifikan antara selang pengacauan 24 dan 48 jam pada ciri-ciri fizikokimia biji yang difermentasi, maka selang pengacauan 48 jam untuk fermentasi sebanyak 8 hari adalah disyorkan. Fermentasi selama 8 hari adalah disyorkan kerana ia dapat menghapuskan mikroorganisma yang berbahaya.

Keadaan pemanggangan (masa dan suhu) menjejaskan ciri-ciri fizikokimia biji yang telah difermentasi selama 8 hari dengan selang pengacauan 48 jam secara signifikan. Analisis GCMS menunjukkan serbuk biji yang telah dipanggang mengandungi pyrazines. Ini menunjukkan serbuk biji yang telah dipanggang mempunyai aroma dan rasa serupa dengan serbuk koko. Ujian ketoksikan anak udang menunjukkan LC_{50} ekstrak serbuk biji rambutan panggang ialah $7.07 \times 10^4 \mu\text{g/mL}$ dan dianggap sebagai tidak toksik. Dapatan kajian menunjukkan produk seperti serbuk koko dapat dihasilkan daripada biji rambutan yang telah difermentasi dan ia adalah selamat untuk dimakan.

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I certify that a Thesis Examination Committee has met on 28 November 2017 to conduct the final examination of Chai Kong Fei on his thesis entitled "Biotransformation of Rambutan (*Nephelium lappaceum* L.) Seed Into a Cocoa Powder-Like Product " in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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LIST OF ABBREVIATIONS

AAB	acetic acid bacteria
ANOVA	analysis of variance
C	Celsius
CE	catechin equivalent
cm	centimeter
DSC	Differential Scanning Calorimeter
FFA	free fatty acid
FAME	fatty acid methyl ester
g	gram
GC	gas chromatography
HCl	hydrochloric acid
HPLC	high performance liquid chromatography
I.D.	internal diameter
IV	iodine value
kg	kilogram
KOH	potassium hydroxide
L	liter
LAB	lactic acid bacteria
LCMS	liquid chromatography mass-spectrometry
M	Molar
mg	milligram
min	minute
mL	milliliter
mm	millimeter
N	normality

N ₂	nitrogen
NaOH	sodium hydroxide
ND	not detected
nm	nanometer
psi	pounds per square inch
rpm	revolutions per minute
SFI	solid fat index
TAG	triacylglycerol
UV-Vis	ultra violet-visible
v/v	volume/volume
w/v	weight/volume
μL	microliter



CHAPTER 1

INTRODUCTION

Rambutan (*Nephelium lappaceum* Linn.) is a popular tropical fruit from the Sapindaceae family. It is native to Malaysia and Indonesia (Issara, Zzaman, & Yang, 2014). The rambutan fruit has a colored skin and spinterns. The peel of the fruit is red, yellow or orange yellow in color. According to Sun et al. (2011), the fruit peel contains abundant of anthocyanins, which are mainly responsible for the red color of the ripe fruit. There are more than 187 varieties of rambutan are registered to the Department of Agriculture Malaysia and additional 25 varieties are found in Singapore, Indonesia, the Philippines, and Thailand (Zee, 1993). Figure 1.1 shows two common (red and yellow) skin colors of the rambutan fruit. The fruits are large (3-6 cm long and 3-4 cm broad) (Fila et al., 2012) and spherical in shape. Rambutan has two fruiting seasons (July-September and December-February) per year. A rambutan tree may produce more than 5000-6000 fruits per fruiting season (Ahmad & Alrozi, 2011). The fruits take 3 to 6 months to mature depending on growing climate and cultivar (Diczbalis, 2002).



Figure 1.1: Rambutan fruit with red, yellow and orange yellow skin colors

Thailand, Indonesia and Malaysia account for approximately 80 % of the total world production of rambutan and only Malaysia, Thailand and Indonesia export this fruit among the ASEAN member countries (Ahmad & Chua, 2013; Tindall, 1994). Fresh and processed rambutan products are exported from Thailand to Asian and European countries. In other Asian countries, the fruits are sold in the domestic markets (Paull & Duarte, 2012; Tindall, 1994). According to the Department of Statistics Malaysia (2016), Malaysia has sufficient rambutan fruit supply for domestic demands. In 2013, the self-sufficiency ratio of the fruits to demands in Malaysia was 98.2 % while the ratio in 2014 was 100.5 %. As advocated in the *Dasar Agromakanan Negara 2011-2020* (Ministry of Agriculture and Agro-Based Industry Malaysia, 2011), the production of the domestic fruits especially star fruit, papaya, pineapple and rambutan should be increased commercially due to the increase of export market demand. Fruits production is expected to increase in line with the planting area from 9.6 metric ton per hectare in 2010 to 12.9 metric ton per hectare in 2020 (Ministry of Agriculture and Agro-Based Industry Malaysia, 2011).

The rambutan fruit is rich in carbohydrates, fiber, iron, potassium, magnesium, vitamin C and calcium as shown in Table 1.1 (P. Lee, Tan, Yu, Curran, & Liu, 2013; Sukasih & Setyadjit, 2015; Tindall, 1994; Wall, 2006a). The skin of the fruit contains flavonoids, saponin and tannin (Lestari, Djati, Rudijanto, & Fatchiyah, 2013) while the seeds have high fat content (38 %) (Manaf, Marikkar, Long, & Ghazali, 2013), antioxidant activity and high phenolic content (Thitilertdecha, Teerawutgulrag, & Rakariyatham, 2008a). All these nutrients and minerals allow this fruit to possess medicinal and therapeutic properties.

Table 1.1: Composition of rambutan per 100 g edible portion

Constituents	Value per 100 g
Energy	297 kJ ^a
Moisture	82.1 g ^a
Fat	0.9 g ^a
Protein	0.66 g ^b
Carbohydrate	19.66 g ^b
Dietary fiber	2.8 g ^a
Ash	0.3 g ^a
Starch	0.0 g ^a
Glucose	2.8 g ^a
Fructose	3.0 g ^a
Sucrose	9.9 g ^a
Malic acid	0.05 g ^a
Citric acid	0.31 g ^a
Niacin	0.5 mg ^a
Carotene	0.0 mg ^a
Vitamin C	70.0 mg ^a
Thiamine	0.01 mg ^a
Riboflavin	0.07 mg ^a
Phosphorus	8.8-18.8 mg ^c
Potassium	133.5-249.4 mg ^c
Calcium	6.8-8.7 mg ^c
Magnesium	13.3-17.2 mg ^c
Sodium	5.5-8.2 mg ^c
Iron	0.41-0.56 mg ^c
Manganese	0.07-0.38 mg ^c
Zinc	0.16-0.26 mg ^c
Copper	0.16-0.20 mg ^c
Boron	0.11-0.16 mg ^c

(Adapted from: ^a-Tindall, 1994; ^b-Fila, Itam, & Johnson, 2013; ^c-Wall, 2006)

Generally, rambutan fruits are consumed in the fresh state. The aril (flesh or pulp) is sometimes mixed with other fruits to prepare salads. Some rambutan varieties are suitable for canning, thus extending the period of consumption other than during the local fruiting seasons. In Malaysia and Thailand, rambutan is industrially produced into

juice, jam, jelly, marmalade and spread (Morton, 1987; Dapur Kayu Enterprise, 2017; Solís-Fuentes, Camey-Ortíz, Hernández-Medel, Pérez-Mendoza, & Durán-de-Bazúa, 2010). Besides, the fruits can be processed as rambutan stuffed with a chunk of pineapple and canned in syrup (Sirisompong, Jirapakkul, & Klinkesorn, 2011). Rambutan chips are also available in the market. Some popular rambutan-derived products are shown in Figure 1.2.



Figure 1.2: Some commercial rambutan products

Although rambutan is one of the most popular fruits in Malaysia as it is available throughout the country, it is seasonal and susceptible to deterioration (3-4 days at ambient temperature, or a few weeks at 12-13 °C and 90-95 % relative humidity) (Siriphanich, 2003). When supply is greater than demand, the fruits are often wasted. The massive amount of the excessive fruits is being disregarded, causing a severe problem in the community as they gradually ferment and release off-odors. This is uncommon to be seen in any other popular non-exotic fruits such as apple, carob, peach, grape and citrus as many food industries have been working on these fruits as well as their by-products in order to fully utilize them (Davidov-Pardo & McClements, 2015; Fernández-López, Sendra, Sayas-Barberá, Navarro, & Pérez-Alvarez, 2008; Loizzo et al., 2015; Raina et al., 2015; Sheng et al., 2016). However, to date, there is no large industry in Malaysia to diversify the rambutan fruit into various products. When rambutan fruit is canned, the fruits are deseeded during processing and the seeds (~ 4-9 g/100 g) become a waste by-product (Solís-Fuentes et al., 2010). Hence, in order to reduce wastage and increase the variation of food from rambutan fruit, there is a potential in converting the rambutan seeds into a cocoa powder-like product through the process of natural fermentation involving the whole fruit (Low, 2011). In this preliminary study, cocoa powder-like product was produced by fermenting and drying rambutan fruit as well as roasting and grinding the dried fermented seed.

To date, rambutan seed powder as a possible complement to cocoa powder has not been produced. It is relatively a new idea in product development. From the literature, much of the research has been confined to cocoa powder processing (Afoakwa, Paterson, & Fowler, 2007; Afoakwa, Paterson, Fowler, & Ryan, 2008; Afoakwa, 2015; Andres-Lacueva et al., 2008; Hofberger & Tanabe, 2007; Minifie, 1989; Tomas-Barberán et al.,

2007). Report has been made on rambutan fruit fermentation (Mehdizadeh, Lasekan, Muhammad, & Baharin, 2015) but not on the production of rambutan seed powder. Scientific analysis on rambutan seed powder has not been carried out and definitely there is still lacking of information on the rambutan seed powder processing. Thus, the main objective of this study was to produce a cocoa powder-like product from rambutan seeds which had undergone the process of natural fermentation followed by roasting, and to determine the toxicity of the powder. Based on this objective, the specific objectives were:

1. To select the best rambutan variety for fermentation by comparing the physicochemical properties of pulp and seed of rambutan fruit with those of cocoa pulp and seed.
2. To determine the physicochemical properties of rambutan fruit sweatings, seed and seed fat during solid state fermentation at different fermentation times and turning intervals.
3. To optimize the roasting process of rambutan seed removed from fermented and dried rambutan fruit, and to determine the physicochemical properties and toxicity of rambutan seed powder.

REFERENCES

- Abdalla, A. E. M., Darwish, S. M., Ayad, E. H. E., El-Hamahmy, R. M., 2007. Egyptian mango by-product 1. Compositional quality of mango seed kernel. *Food Chemistry* 103(4), 1134–1140.
- Abdulkarim, S. M., Long, K., Lai, O. M., Muhammad, S. K. S., & Ghazali, H. M. (2005). Some physico-chemical properties of *Moringa oleifera* seed oil extracted using solvent and aqueous enzymatic methods. *Food Chemistry*, 93(2), 253–263. doi:10.1016/j.foodchem.2004.09.023
- Achinewhu, S. C. (1986). The effect of fermentation on carbohydrate and fatty acid composition of African oil bean seed (*Pentaclethra macrophylla*). *Food Chemistry*, 19, 105–116.
- Aculey, P. C., Snitkjaer, P., Owusu, M., Bassompierre, M., Takrama, J., Nørgaard, L., ... Nielsen, D. S. (2010). Ghanaian cocoa bean fermentation characterized by spectroscopic and chromatographic methods and chemometrics. *Journal of Food Science*, 75(6), 300–307. doi:10.1111/j.1750-3841.2010.01710.x
- Adams, M. R., Dougan, J., Glossop, E. J., & Twiddy, D. R. (1982). Cocoa sweatings—an effluent of potential value. *Agricultural Wastes*, 4(3), 225–229. doi:10.1016/0141-4607(82)90014-2
- Adeniran, H. A., Farinde, E. O., & Obatolu, V. A. (2013). Effect of heat treatment and fermentation on anti-nutrients content of lima bean (*Phaseolus lunatus*) during production of daddawa analogue. *Annual Review & Research in Biology*, 3(3), 256–266.
- Adetuyi, F. O., & Ibrahim, T. A. (2014). Effect of fermentation time on the phenolic, flavonoid and Vitamin C contents and antioxidant activities of okra (*Abelmoschus esculentus*) seeds. *Nigerian Food Journal*, 32(2), 128–137. doi:10.1016/S0189-7241(15)30128-4
- ADM Cocoa. (2006). *The de zaan® cocoa manual*. Netherlands: Archer Daniels Midland Company BV.
- Afoakwa, E. O. (2015). *Cocoa production and processing technology*. Boca Raton: Taylor & Francis Group.
- Afoakwa, E. O., Kongor, J. E., Budu, A. S., Mensah-Brown, H., & Takrama, J. F. (2015). Changes in biochemical and physico-chemical qualities during drying of pulp preconditioned and fermented cocoa (*Theobroma cacao*) beans. *African Journal of Food, Agriculture, Nutrition and Development*, 1, 9651–9670. doi: http://dx.doi.org/10.15226/jnhfs.2014.00121
- Afoakwa, E. O., Kongor, J. E., Takrama, J., & Budu, A. S. (2013). Changes in nib acidification and biochemical composition during fermentation of pulp pre-

- conditioned cocoa (*Theobroma cacao*) beans. *International Food Research Journal*, 20(4), 1843–1853.
- Afoakwa, E. O., Paterson, A., & Fowler, M. (2007). Factors influencing rheological and textural qualities in chocolate – a review. *Trends in Food Science & Technology*, 18(6), 290–298. doi:10.1016/j.tifs.2007.02.002
- Afoakwa, E. O., Paterson, A., Fowler, M., & Ryan, A. (2008). Flavor formation and character in cocoa and chocolate: a critical review. *Critical Reviews in Food Science and Nutrition*, 48(9), 840–857. doi:10.1080/10408390701719272
- Afoakwa, E. O., Paterson, A., Fowler, M., & Ryan, A. (2009). Matrix effects on flavour volatiles release in dark chocolates varying in particle size distribution and fat content using GC-mass spectrometry and GC-olfactometry. *Food Chemistry*, 113(1), 208–215. doi:10.1016/j.foodchem.2008.07.088
- Afoakwa, E. O., Quao, J., Budu, A. S., Takrama, J., & Saalia, F. K. (2011). Effect of pulp preconditioning on acidification, proteolysis, sugars and free fatty acids concentration during fermentation of cocoa (*Theobroma cacao*) beans. *International Journal of Food Sciences and Nutrition*, 62(7), 755–764. doi:10.3109/09637486.2011.581224
- Afoakwa, E. O., Quao, J., Takrama, J., & Budu, A. S. (2013). Chemical composition and physical quality characteristics of Ghanaian cocoa beans as affected by pulp pre-conditioning and fermentation. *Journal of Food Science and Technology*, 50(6), 1097–105. doi:10.1007/s13197-011-0446-5
- Agapey Chocolate Factory. (2013). Types of cacao (cocoa beans). Retrieved March 7, 2013, from <http://www.agapey.com/cocoavarieties.html>
- Ahmad, I., & Chua, P. C. (2013). Trends in production and trade of tropical fruits in ASEAN countries. In: Palupi, E.R., Krisantini, & Warrington, I. J. (Eds.), *IV International Symposium on Tropical and Subtropical Fruits* (pp. 559-580). Bogor, Indonesia: ISHS Acta Horticulturae 975.
- Ahmad, M. A., & Alrozi, R. (2011). Optimization of rambutan peel based activated carbon preparation conditions for Remazol Brilliant Blue R removal. *Chemical Engineering Journal*, 168(1), 280–285. doi:10.1016/j.cej.2011.01.005
- Almeida, M. H. G. (1999). *Cacau. Tecnologia p'os-colheita. A fraccão volátilno flavor* (PhD thesis). Instituto Superior de Agronomia.
- Amano Artisan Chocolate. (2013). Roasting cocoa for flavor development in chocolate. Retrieved August 14, 2017, from <http://www.amanochocolate.com/blog/flavor-development-in-chocolate/>
- Amaral, J. S., Casal, S., Seabra, R. M., & Oliveira, B. P. P. (2006). Effects of roasting on hazelnut lipids. *Journal of Agricultural and Food Chemistry*, 54, 1315–1321.

- Amorim, G., Jr, L., Gomes, L. H., Efraim, P., Almeida, F. C. D., & Figueira, A. (2008). Fermentation of cacao (*Theobroma cacao* L.) seeds with a hybrid *Kluyveromyces marxianus* strain improved product quality attributes. *FEMS Yeast Research*, 8, 788–798. doi:10.1111/j.1567-1364.2008.00405.x
- Andres-Lacueva, C., Monagas, M., Khan, N., Izquierdo-Pulido, M., Urpi-Sarda, M., Permanyer, J., & Lamuela-Raventos, R. M. (2008). Flavanol and flavonol contents of cocoa powder products: influence of the manufacturing process. *Journal of Agricultural and Food Chemistry*, 56, 3111–3117.
- Anjum, F., Anwar, F., Jamil, A., & Iqbal, M. (2006). Microwave roasting effects on the physico-chemical composition and oxidative stability of sunflower seed oil. *Journal of the American Oil Chemists' Society*, 83(9), 777–784.
- AOAC. (1984). *Official Methods of Analysis*, 14th Ed.; Association of Official Analytical Chemists: Washington, DC.
- AOCS. (1997). *Official Methods and Recommended Practices of the American Oil Chemists' Society*, 6th Ed. American Oil Chemists' Society: Champaign, USA.
- Aprocaci. (2010). Fermentation box. Retrieved August 18, 2017, from <http://www.aprocaci.org/en/services>.
- Apu, A., Muhit, M., Tareq, S., Pathan, A., Jamaluddin, A., & Ahmed, M. (2010). Antimicrobial activity and brine shrimp lethality bioassay of the leaves extract of *Dillenia indica* Linn. *Journal of Young Pharmacists*, 2(1), 50–53. doi:10.4103/0975-1483.62213
- Ardhana, M. M., & Fleet, G. H. (2003). The microbial ecology of cocoa bean fermentations in Indonesia. *International Journal of Food Microbiology*, 86, 87–99.
- Aremu, C. Y., Agiang, M. A., & Ayatse, J. O. (1995). Nutrient and antinutrient profiles of raw and fermented cocoa beans. *Plant Foods for Human Nutrition*, 48(3), 217–23.
- Arenas, M. G. H., Angel, D. N., Damian, M. T. M., Ortiz, D. T., Díaz, C. N., & Martinez, N. B. (2010). Characterization of rambutan (*Nephelium lappaceum*) fruits from outstanding mexican selections. *Revista Brasileira de Fruticultura*, 32(4), 1098–1104.
- Arikiah, A., Tan, Y. P., Sharma, M. & Oapperton, J. F. (1994). Experiments to determine influence of primary processing parameters and planting materials on the flavour of cocoa beans in Malaysia. *Cocoa Growers Bulletin*, 48, 36–46.
- Arlorio, M., Locatelli, M., Travaglia, F., Coïsson, J. D., Grosso, E. D., Minassi, A., ... Martelli, A. (2008). Roasting impact on the contents of clovamide (N-caffeoyl-L-DOPA) and the antioxidant activity of cocoa beans (*Theobroma cacao* L.). *Food Chemistry*, 106(3), 967–975. doi:10.1016/j.foodchem.2007.07.009

- Aroyeun, S. O., Ogunbayo, J. O. and Olaiya, A. O. (2006). Effect of modified packaging and storage time of cocoa pods on the commercial quality of cocoa beans. *British Food Journal*, 108, 141-151.
- Asep, E. K., Jinap, S., Tan, T. J., Russly, A. R., Harcharan, S., & Nazimah, S. A. H. (2008). The effects of particle size, fermentation and roasting of cocoa nibs on supercritical fluid extraction of cocoa butter. *Journal of Food Engineering*, 85(3), 450–458. doi:10.1016/j.jfoodeng.2007.08.008
- Augustin, M. A., & Chua, B. C. (1988). Composition of rambutan seeds. *Pertanika*, 11(2), 211–215.
- Awua, P. K. (2002). *The success story of cocoa processing and chocolate manufacture in Ghana*. Essex: David Jamieson and Associates Press Inc.
- Barel, M. (1997). La fermentation du cacao: le moyen de l'apprécier et de la maîtriser. *Industries alimentaires et agricoles*, 114(4), 211-214.
- Bates, R., Morris, J., & Crandall, P. (2001). Principles and practices of small- and medium-scale fruit juice processing. *FAO Agri Ser Bul* 146: 177–188.
- Beckett, S. T. (2000). *The science of chocolate*. York, UK: Royal Society of Chemistry.
- Biehl, B., Brunner, E., Passern, D., Quesnel, V. C., & Adomako, D. (1985). Acidification, proteolysis and flavour potential in fermenting cocoa beans. *Journal of the Science of Food and Agriculture*, 36, 583–598. doi:10.1002/jsfa.2740360710
- Biehl, B., Meyer, B., Crone, G., Pollmann, L. and Said, M. B. (1989). Chemical and physical changes in the pulp during ripening and post-harvest storage of cocoa pods. *Journal of the Science Food and Agriculture*, 48, 189–208.
- Bonvehi, J. S. (2005). Investigation of aromatic compounds in roasted cocoa powder. *European Food Research and Technology*, 221, 19–29. doi:10.1007/s00217-005-1147-y
- Bonvehi, J. S., & Coil, F. V. (1997). Evaluation of bitterness and astringency of polyphenolic compounds in cocoa powder. *Food Chemistry*, 60(3), 365–370.
- Bracco, U. (1994). Effect of triglyceride structure on fat absorption. *The American Journal of Clinical Nutrition*, 60(6), 1002S–1009S.
- Bravo, L. (1998). Polyphenols: chemistry, dietary sources, metabolism, and nutritional significance. *Nutrition Reviews*, 56(11), 317–333.
- Broadhurst, R.B. & Jones, W. T. (1978). Analysis of condensed tannins using acidified vanillin. *Journal of the Science of Food and Agriculture*, 29, 788–794.
- Buamah, R., Dzogbe, V. P., & Oldham, J. H. (1997). Pure yeast culture fermentation of cocoa (*Theobroma cacao* L): effect on yield of sweatings and cocoa bean quality. *World Journal of Microbiology & Biotechnology*, 13, 457–462.

- Bullangpoti, V., Visetson, S., Milne, J., & Pornbanlualap, S. (2004). Effects of mangosteen's peels and rambutan's seeds on toxicity, esterase and glutathione-s-transferase in rice weevil (*Sitophilus oryzae* L.). *Kasetsart J. (Nat. Sci.)*, 38, 84–89.
- Camu, N., De Winter, T., Verbrugghe, K., Cleenwerck, I., Vandamme, P., Takrama, J. S., ... De Vuyst, L. (2007). Dynamics and biodiversity of populations of lactic acid bacteria and acetic acid bacteria involved in spontaneous heap fermentation of cocoa beans in Ghana. *Applied and Environmental Microbiology*, 73(6), 1809–1824. doi:10.1128/AEM.02189-06
- Camu, N., Gonzalez, A., De Winter, T., Van Schoor, A., De Bruyne, K., Vandamme, P., ... De Vuyst, L. (2008). Influence of turning and environmental contamination on the dynamics of populations of lactic acid and acetic acid bacteria involved in spontaneous cocoa bean heap fermentation in Ghana. *Applied and Environmental Microbiology*, 74(1), 86–98. doi:10.1128/AEM.01512-07
- Camu, N., Winter, T. De, Addo, S. K., Takrama, J. S., Bernaert, H., & Vuyst, L. D. (2008). Fermentation of cocoa beans: influence of microbial activities and polyphenol concentrations on the flavour of chocolate. *Journal of the Science of Food and Agriculture*, 88, 2288–2297. doi:10.1002/jsfa.3349
- Carr, J. G., Davies, P. A. & Dougan, J. (1979). *Cocoa fermentation in Ghana and Malaysia*. London: Tropical Products Institute.
- Casten, J., & Synder, H.E. (1985). *Understanding Pressure Extraction of Vegetable Oils*. Virginia, USA: Volunteers in Technical Assistance (VITA).
- Cheesman, E.E. (1944). Notes on the nomenclature, classification and possible relationships of cacao populations. *Tropical Agriculture (Trinidad)*, 21, 144–159.
- Cheikh-Rouhou, S., Besbes, S., Hentati, B., Blecker, C., Deroanne, C., & Attia, H. (2007). *Nigella sativa* L.: Chemical composition and physicochemical characteristics of lipid fraction. *Food Chemistry*, 101(2), 673–681. doi:10.1016/j.foodchem.2006.02.022
- Chen, C. H., Wu, Y. L., Lo, D., & Wu, M. C. (2013). Physicochemical property changes during the fermentation of longan (*Dimocarpus longan*) mead and its aroma composition using multiple yeast inoculations. *Journal of the Institute of Brewing*, 119(4), 303–308. doi:10.1002/jib.95
- Cheng, W.-Y., Akanda, J. M. H., & Nyam, K. L. (2016). Kenaf seed oil: a potential new source of edible oil. *Trends in Food Science & Technology*, 52, 57–65. doi:10.1016/j.tifs.2016.03.014
- Chomchalow, N., Somsr, S., & Songkhla, P. N. (2008). Marketing and export of major tropical fruits from Thailand. *AU Journal of Technology*, 11(3), 133–143.

Cocks, L.V. & Van Rede, C. (1966). *Laboratory handbook for oil and fats analysts*. Academic Press: London.

Cook, L. R. (1982). *Chocolate production and use*. Meursing, E. H. (Rev.). New York: Harcourt Brace Javanovich, Inc.

Cros, E. & Jeanjean, N. (1998). Formation de l'arome cacao. In: Pontillon, J. (Ed.), *Cacao et chocolat: production, utilisation, caracteristiques* (pp. 188–206). Paris, France: Technique et Documentation Lavoisier.

Crozier, S. J., Preston, A. G., Hurst, W. J., Payne, M. J., & Mann, J. (2011). Cacao seeds are a “Super Fruit”: a comparative analysis of various fruit powders and products. *Chemistry Central Journal*, 5(1), 1-5.

C-Spot. (2011). Chocolate flavor profiles. Retrieved July 3, 2013, from <https://www.c-spot.com/atlas/chocolate-flavor-profiles>

Dapur Kayu Enterprise. (2017). Resipi traditional asli. Retrieved August 24, 2017, from <http://www.neknorfoods.com/jam.html>

Davidov-Pardo, G., & McClements, D. J. (2015). Nutraceutical delivery systems: resveratrol encapsulation in grape seed oil nanoemulsions formed by spontaneous emulsification. *Food Chemistry*, 167, 205–212. doi:10.1016/j.foodchem.2014.06.082

Delabarre, Y. (1989). Synthese bibliographique sur le ramboutan ou lychee chevelu (*Nephelium lappaceum* L.). Bibliographic summary on rambutan or hairy lychee (*Nephelium lappaceum* L.). *Fruits* 44:33-44.

Department of Statistics Malaysia. (2016). Supply and utilization accounts selected agricultural commodities, Malaysia 2010-2014. Retrieved June 29, 2017, from https://www.dosm.gov.my/v1/index.php?r=column/cthemeByCat&cat=164&bul_id=ZzNBdUIWT2I4NE4xNCt6U2VNc1Q2QT09&menu_id=Z0VTZGU1UHBUT1VJMF1paXRRR0xpdz09

Dias, D. R., Schwan, R. F., Freire, E. S., & Serôdio, R. D. S. (2007). Elaboration of a fruit wine from cocoa (*Theobroma Cacao* L.) pulp. *International Journal of Food Science & Technology*, 42(3), 319–329. doi:10.1111/j.1365-2621.2006.01226.x

Dias, J. C. & Avila, M. G. M. (1993). Influence of the drying process on the acidity of cocoa beans. *Agrotropica*, 5, 19–24.

Diczbalis, Y. (2002). *Rambutan: improving yield and quality*. Queensland: Rural Industries Research and Development Corporation.

Dimick, P. S., & Hoskin, J. C. (1999). The chemistry of flavour development in chocolate. In S. T. Beckett (Ed.), *Industrial chocolate manufacture and use* (3rd edition, pp. 137–152). Oxford: Blackwell Science.

- Dimick, P. S., & Hoskin, J. M. (1981). Chemico-physical aspects of chocolate processing - a review. *Canadian Institute of Food Science and Technology Journal*, 14(4), 269–282.
- Duncan, R. J. E., Godfrey, G., Yap, T. N., Pettiphar, G. L. and Tharumarajah, T. (1989). Improvement of Malaysian cocoa bean flavor by modification of harvesting, fermentation and drying methods – the Sime Cadbury Process. *The Planter*, 65, 157-173.
- Dyer, B. (2003). Alkalized Cocoa Powders. In *57th PMCA Production Conference* (pp. 128–135).
- Dzoghbeia, V. P., Buamah, R., & Oldham, J. H. (1999). The controlled fermentation of cocoa (*Theobroma cacao* L) using yeasts: enzymatic process and associated physico-chemical changes in cocoa sweatings. *Food Biotechnology*, 13(1), 1-12. doi:10.1080/08905439609549958
- Eiamwat, J., Reungpatthanaphong, S., & Laovitthayangoon, S. (2014). Toxicity studies on rambutan (*Nephelium lappaceum*) seed fat and oil extracts using acute oral, dermal and irritation assays. *International Journal of Natural Products Research*, 4(2), 36–39.
- Ekinci, R. (2005). The effect of fermentation and drying on the water-soluble vitamin content of tarhana, a traditional Turkish cereal food. *Food Chemistry*, 90, 127–132. doi:10.1016/j.foodchem.2004.03.036
- El Castillo del Cacao. (2016). From cacao to chocolate. Retrieved August 8, 2017, from <http://www.elcastillodelcacao.com/cacao.html>
- Elfick, J. (2017). Cocoa. Retrieved August 18, 2017, from http://www.uq.edu.au/_School_Science_Lessons/55.9.GIF
- Emmanuel, O. A., Jennifer, Q., Agnes, S. B., Jemmy, S. T. and, & Firibu, K. S. (2012). Influence of pulp-preconditioning and fermentation on fermentative quality and appearance of Ghanaian cocoa (*Theobroma cacao*) beans. *International Food Research Journal*, 19(1), 127–133.
- Escudero-López, B., Cerrillo, I., Gil-Izquierdo, Á., Hornero-Méndez, D., Herrero-Martín, G., Berná, G., ... Fernández-Pachón, M. S. (2016). Effect of thermal processing on the profile of bioactive compounds and antioxidant capacity of fermented orange juice. *International Journal of Food Sciences and Nutrition*, 67(7), 779–788. doi:10.1080/09637486.2016.1204428
- Farah, D. M. H., Zaibunnisa, A. H., & Minaswi. (2012). Optimization of cocoa beans roasting process using Response Surface Methodology based on concentration of pyrazine and acrylamide. *International Food Research Journal*, 19(4), 1355–1359.
- Febrianto, N. A., Issara, U., Yang, T. A., & Abdullah, W. N. W. (2014). Thermal behavior, microstructure, and texture properties of fermented-roasted rambutan seed fat and cocoa butter mixtures. *Pelita Perkebunan*, 30(1), 65–79.

- Febrianto, N. A., Yang, T. A., & Wan Abdullah, W. A. (2016). Cocoa-like flavor compound development of rambutan seed fat as the effect of fermentation and roasting. *International Food Research Journal*, 23(5), 2166–2174.
- Fennema, O. R. (1996). Food Additives. In: Fennema, O. R. (Ed.), *Food Chemistry* (3rd edition, pp. 768-823). New York, NY: Taylor and Francis.
- Fenwick, D. E., & Oakenfull, D. (1983). Saponin content of food plants and some prepared foods. *Journal of the Science of Food and Agriculture*, 34(2), 186–191.
- Fernández-López, J., Sendra, E., Sayas-Barberá, E., Navarro, C., & Pérez-Alvarez, J. A. (2008). Physico-chemical and microbiological profiles of “salchichón” (Spanish dry-fermented sausage) enriched with orange fiber. *Meat Science*, 80(2), 410–417. doi:10.1016/j.meatsci.2008.01.010
- Fidrianny, I., Sari, P. I., & Wirasutisna, K. R. (2015). Antioxidant activities in various peel extracts of four varieties rambutan (*Nephelium lappaceum*) using DPPH, FRAP assays. *International Journal of Pharmacognosy and Phytochemical Research*, 7(2), 280–285.
- Fila, W. O., Johnson, J. T., Edem, P. N., Odey, M. O., Ekam, V. S., Ujong, U. P., & Eteng, O. E. (2012). Comparative anti-nutrients assessment of pulp, seed and rind of rambutan (*Nephelium lappaceum*). *Annals of Biological Research*, 3(11), 5151–5156.
- Fila, W., Itam, E., & Johnson, J. (2013). Comparative proximate compositions of watermelon *Citrullus Lanatus*, squash *Cucurbita Pepo*’l and rambutan *Nephelium Lappaceum*. *International Journal of Science and Technology*, 2(1), 81–88.
- Flavor & Extract Manufacturers Association. (2017). FEMA flavor ingredient library. Retrieved Jan 20, 2017, from <https://www.femaflavor.org/flavor>
- Fowler, M. S., Leheup, P., and Cordier, J. L. (1998). Cocoa, Coffee and Tea. In: Woods, W. J. B. (Ed.), *Microbiology of fermented foods* (Vol. 1, pp. 128–147). London: Blackie Academic and Professionals
- Frauendorfer, F., & Schieberle, P. (2008). Changes in key aroma compounds of Criollo cocoa beans during roasting. *Journal of Agricultural and Food Chemistry*, 56(21), 10244–10251. doi:10.1021/jf802098f
- García-Alamilla, P., Lagunes-Gálvez, L. M., Barajas-Fernández, J., & García-Alamilla, R. (2017). Physicochemical Changes of Cocoa Beans during Roasting Process. *Journal of Food Quality*, 2017, 1–11.
- Ghazali, H. M., Hamidah, S., & Man, Y. B. C. (1995). Enzymatic transesterification of palm olein with nonspecific and 1, 3-specific lipases. *Journal of the American Oil Chemists' Society*, 72(6), 633–639.

- Goenaga, R. (2011). Dry matter production and leaf elemental concentrations of rambutan grown on an acid ultisol. *Journal of Plant Nutrition*, 34(5), 753–761. doi:10.1080/01904167.2011.540690
- Gourieva, K.B. & Tserevinov, O.B. (1979). Methods of evaluating the degree fermentation of cocoa beans. USSR Patent No. 64654.
- Granvogl, M., Bugan, S., & Schieberle, P. (2006). Formation of amines and aldehydes from parent amino acids during thermal processing of cocoa and model systems: new insights into pathways of the Strecker reaction. *Journal of Agricultural and Food Chemistry*, 54, 1730-1739.
- Griffin, P. M., & Tauxe, R. V. (1991). The epidemiology of infections caused by *Escherichia coli* O157:H7, other enterohemorrhagic *E. coli*, and the associated hemolytic uremic syndrome. *Epidemiologic Reviews*, 13(1), 60-98.
- Guehi, S. T., Dabonne, S., Ban-koffi, L., Kedjebo, D. K., & Zahouli, G. I. B. (2010). Effect of turning beans and fermentation method on the acidity and physical quality of raw cocoa beans. *Advance Journal of Food Science and Technology*, 2(3), 163–171.
- Guehi, T. S., Dadie, A. T., Koffi, K. P. B., Dabonne, S., Ban-Koffi, L., Kedjebo, K. D., & Nemlin, G. J. (2010). Performance of different fermentation methods and the effect of their duration on the quality of raw cocoa beans. *International Journal of Food Science & Technology*, 45(12), 2508–2514. doi:10.1111/j.1365-2621.2010.02424.x
- Guehi, T. S., Yao, N. D. D., Manizan, N. P., Nevry, K. R., Koffi, L. B., & Konan, M. (2008). Comparison of the degree of fermentation and fungal profiles of raw cocoa beans sourced from three Ivorian main producing regions. *African Journal of Food Science*, 2(2), 112–118.
- Guehi, T. S., Zahouli, I. B., Ban-Koffi, L., Fae, M. A., & Nemlin, J. G. (2010). Performance of different drying methods and their effects on the chemical quality attributes of raw cocoa material. *International Journal of Food Science & Technology*, 45(8), 1564–1571. doi:10.1111/j.1365-2621.2010.02302.x
- Guittard Chocolate Company. (2013). Cultivate better. Retrieved March 7, 2013, from <https://www.guittard.com/cultivate-better/honorable-sourcing>
- Gunstone, F. D. (2011). *Vegetable oils in food technology: Composition, properties and uses* (2nd edition). West Sussex, UK: Blackwell Publishing Ltd.
- Hadi, M. A. M. El, Zhang, F., Wu, F., Zhou, C., & Tao, J. (2013). Advances in fruit aroma volatile research. *Molecules*, 18, 8200–8229. doi:10.3390/molecules18078200
- Hanneman, E. (2000). The complex world of cocoa butter. *The Manufacturing Confectioner*, 80:107–112.

- Hansen, C. E., Del Olmo, M., & Burri, C. (1998). Enzyme activities in cocoa beans during fermentation. *Journal of the Science of Food and Agriculture*, 77(2), 273–281.
doi:10.1002/(SICI)1097-0010(199806)77:2<273::AID-JSFA40>3.0.CO;2-M
- Harahap, S. N., Ramli, N., Vafaei, N., & Said, M. (2011). Physicochemical and nutritional composition of rambutan Anak Sekolah (*Nephelium lappaceum* L.) seed and seed oil. *Pakistan Journal of Nutrition*, 10, 1–5.
- Hardy, F. (1960). *Cacao Manual*. Turrialba, Costa Rica: Inter-American Institute of Agricultural Sciences.
- Hashim, P., & Mat Hashim, D. (2013). A review of cosmetic and personal care products: Halal perspective and detection of ingredient. *Pertanika Journal of Science and Technology*, 21(2), 281–292.
- Hashim, P., Selamat, J., Muhammad, S. K. S., & Ali, A. (1998). Effect of mass and turning time on free amino acid, peptide-N, sugar and pyrazine concentration during cocoa fermentation. *Journal of the Science of Food and Agriculture*, 78, 543–550.
- Hassan, M., Philippe, J.B., Alain, P., Gerard, G. (1995). Production of cocoa butter equivalents from prickly-pear juice fermentation by an unsaturated fatty acid auxotroph of *Cryptococcus curvatus* grown in batch culture. *Process Biochemistry*, 30 (7), 629–634.
- Heinzler, M. and Eichner, K. (1992). The role of amodori compounds during cocoa processing – formation of aroma compounds under roasting conditions. *Zeitschrift Fur Lebensmittel-Untersuchung Und –Forschung*, 21, 445–450.
- Hernandez, B., Castellote, A. I., and Permanyer, J. J. (1991). Triglyceride analysis of cocoa beans from different geographical origins. *Food Chemistry*. 41, 269–276.
- Hiai, S., Oura, H., & Nakajima, T. (1976). Color reaction of some saponins and saponins with vanillin and sulfuric. *Planta Medica*, 29, 116–122.
doi:10.1055/s-0028-1097639
- Hidalgo, F. J., & Zamora, R. (2000). The role of lipids in nonenzymatic browning. *Grasas Y Aceites*, 51(1-2), 35–49. doi:10.3989/gya.2000.v51.i1-2.405
- Hii, C. L., Abdul Rahman, R., Jinap, S., & Che Man, Y. (2006). Quality of cocoa beans dried using a direct solar dryer at different loadings. *Journal of the Science of Food and Agriculture*, 86(8), 1237-1243. doi:10.1002/jsfa.
- Hofberger, R., & Tanabe, N. A. (2007). Chocolate and cocoa. In Y. H. Hui, R. C. Chandan, S. Clark, N. Crose, J. Dobbs, E. B. Smith, ... F. Toldra (Eds.), *Handbook of food products manufacturing* (pp. 675-694). New Jersey: John Wiley & Sons.

- Hogenbrink, G. (1984). Compatibility of specialty fats with cocoa butter. In: *38th Pennsylvania Manufacturing Confectioner's Association Production Conference* (pp. 116–120).
- Hostettmann, K. & Marston, A. (1995). *Saponins. Chemistry and Pharmacology of Natural Products*, pp. 584, Cambridge University Press: Australia.
- Howat, G. R., Powell, B. D. & Wood, G. A. R. (1957). Experiments on cocoa fermentation in West Africa. *Journal of the Science of Food and Agriculture*, 8, 67-72.
- Hunt, D. C., Jackson, P. A., Mortlock, R. E., & Kirk, R. S. (1977). Quantitative determination of sugars in foodstuffs by high-performance liquid chromatography. *Analyst*, 102, 917–920.
- International Cocoa Organization. (2017). *ICCO quarterly bulletin of cocoa statistics*. Vol. XLIII, No. 1, Cocoa year 2016/17. Côte d'Ivoire.
- International Office of Cocoa, Chocolate and Sugar Confectionery (IOCCC). (1996). Determination of Free Fatty Acids (FFA) Content of Cocoa Fat as a Measure of Cocoa Nib Acidity. *Analytical Method*, 42: 130–136.
- Ioannone, F., Di Mattia, C. D., De Gregorio, M., Sergi, M., Serafini, M., & Sacchetti, G. (2015). Flavanols, proanthocyanidins and antioxidant activity changes during cocoa (*Theobroma cacao* L.) roasting as affected by temperature and time of processing. *Food Chemistry*, 174, 256–262. doi:10.1016/j.foodchem.2014.11.019
- Issara, U., Zzaman, W., & Yang, T. A. (2014). Rambutan seed fat as a potential source of cocoa butter substitute in confectionary product. *International Food Research Journal*, 21(1), 25–31.
- Izuka, C., & Mbagwu, F. (2013). Phytochemical screening on the seed of *Theobroma Cacao* L. (sterculiaceae). *Global Research Journal of Science*, 2(2), 127 – 135.
- Jatto, W. O., Yuanfa, L., Shan, L., Wang, X., Aworh, O. (2010). Liquid-Gas chromatographic analysis of fatty acid content of south-western nigerian shea butter (*Vitellera paradoxum*). *Electronic Journal of Environmental, Agricultural and Food Chemistry*, 9(2), 358–363.
- Jeevani Osadee Wijekoon, M. M., Bhat, R., & Karim, A. A. (2011). Effect of extraction solvents on the phenolic compounds and antioxidant activities of bunga kantan (*Etilingera elatior Jack*) inflorescence. *Journal of Food Composition and Analysis*, 24(4-5), 615–619. doi:10.1016/j.jfca.2010.09.018
- Jinap, S., Wan Rosli, W. I., Russly, A. R., & Nordin, L. (1998). Effect of roasting time and temperature on volatile component profile during nib roasting of cocoa beans (*Theobroma cacao*). *Journal of the Science of Food and Agriculture*, 77, 441-448. doi:10.1002/(SICI)1097-0010(199808)77

- Kader, A.A. (1993). Modified and controlled atmosphere storage of tropical fruits. In: Champ, B. R., Highley, E., & Johnson, G. I. (Eds.), *Postharvest handling of tropical fruits* (pp. 239-249). Proceedings of the International Conference in Chiang Mai. Australian Centre for International Agricultural Research.
- Kalayasiri, P., Jeyashoke, N., & Krisnangkura, K. (1996). Survey of seed oils for use as diesel fuels. *Journal of the American Oil Chemists' Society*, 73(4), 471-474. doi:10.1007/BF02523921
- Kaphueakngam, P., Flood, A., Sonwai, S., (2009). Production of cocoa butter equivalent from mango seed almond fat and palm oil mid-fraction. *Asian Journal of Food and Agro-Industry*, 2(04), 441-447.
- Kealey, K. S., Snyder, R. M., Romanczyk Jr, L. J., Geyer, H. M., Myers, M. E., Whitacre, E. J., ... & Schmitz, H. H. (2001). *U.S. Patent No. 6,312,753*. Washington, DC: U.S. Patent and Trademark Office.
- Ketsa, S., & Paull, R. E. (1998). Rambutan. Retrieved from <http://www.portalecosus.com/uploads/rambutan/rambutan.pdf>
- Kim, H. & Keeney, P. G. (1984). Epicatechin content in fermented and unfermented cocoa beans. *Journal of Food Science*, 49,1090-1092.
- Knapp, A. W. (1926) Experiments in the fermentation of cacao. *Journal of the Society of Chemical Industry*, 45, 140-2.
- Kondo, S., Posuya, P., & Kanlayanarat, S. (2001). Changes in physical characteristics and polyamines during maturation and storage of rambutans. *Scientia Horticulturae*, 91(1-2), 101-109. doi:10.1016/S0304-4238(01)00250-3
- Kostic, M.J., 1997. Cocoa alkalization. *The Manufacturing Confectioner*, 6, 128-130.
- Kotiranta, A., Lounatmaa, K., & Haapasalo, M. (2000). Epidemiology and pathogenesis of *Bacillus cereus* infections. *Microbes and Infection*, 2, 189-198.
- Krysiak, W. (2011). Effects of convective and microwave roasting on the physicochemical properties of cocoa beans and cocoa butter extracted from this material. *Grasas Y Aceites*, 62(4), 467-478.
- Krysiak, W., Adamski, R., & Zyzelewicz, D. (2013). Factors affecting the color of roasted cocoa bean. *Journal of Food Quality*, 36, 21-31.
- Kunsch, U., Scharer, H., Patrian, B., Hohn, E., Conedera, M., Sassella, A., ... Jelmini, G. (1995). Effects of roasting on chemical composition and quality of different chestnut (*Castanea sativa* Mill) varieties. *Journal of the Science of Food and Agriculture*, 81, 1106-1112. doi:10.1002/jsfa.916
- Kyi, T. M., Daud, W. R. W., Mohammad, A. B., Wahid Samsudin, M., Kadhum, A. A. H., & Talib, M. Z. M. (2005). The kinetics of polyphenol degradation during

the drying of Malaysian cocoa beans. *International Journal of Food Science and Technology*, 40(3), 323-331.
doi:10.1111/j.1365-2621.2005.00959.x

- Lagunes, S., Loiseau, G., Luis, J., Barel, M., & Guiraud, J. (2007). Study on the microflora and biochemistry of cocoa fermentation in the Dominican Republic. *International Journal of Food Microbiology*, 114, 124-130.
doi:10.1016/j.ijfoodmicro.2006.10.041
- Lai, L. R., Hsieh, S. C., Huang, H. Y., & Chou, C. C. (2013). Effect of lactic fermentation on the total phenolic, saponin and phytic acid contents as well as anti-colon cancer cell proliferation activity of soymilk. *Journal of Bioscience and Bioengineering*, 115(5), 552-6.
doi:10.1016/j.jbiosc.2012.11.022
- Lainé, K. (2001). Survey of farming practices on farms in Côte d'Ivoire. *Development*, 1, 1-28.
- Lakshminarayana, A.S. (1980). Mango. In: Nagy S., & Shaw P.E. (eds) *Tropical and subtropical fruits composition, properties and uses*, pp 185-201. Westport, Connecticut: AVI Publishing Inc.
- Laksmi, L. D. S, Lam, P. F., Mendoza, Jr. D. B., Kosiyachinda, S. and Leang, P. C., (1987). Status of the rambutan industry in ASEAN. In: Lam P. F., & Kosiyachinda, S. (Eds.), *Rambutan: Fruit development, postharvest physiology, and marketing in ASEAN* (pp. 1-8). Kuala Lumpur: ASEAN Food Handling Bureau.
- Lam, P. F., & Kosiyachinda, S. (1987). *Rambutan: fruit development, postharvest physiology, and marketing in ASEAN*. Kuala Lumpur: ASEAN Food Handling Bureau.
- Lam, P. F., & Ng, K. H. (1982). Storage of waxed and unwaxed rambutan in perforated and sealed polyethylene bags. *Food Technology Division: Report*, (251).
- Lamikanra, O. (1997). Changes in organic acid composition during fermentation and aging of Noble Muscadine wine. *Journal of Agricultural and Food Chemistry*, 45(3), 935-937. doi:10.1021/jf960447k
- Larrauri, J. A., Rupérez, P., Borroto, B., & Saura-Calixto, F. (1996). Mango peels as a new tropical fibre: preparation and characterization. *LWT-Food Science and Technology*, 29(8), 729-733.
- Lass, R. A. (1999). Cacao growing and harvesting practices. In: Knight, I. (Ed.), *Chocolate and cocoa: health and nutrition* (pp. 11-42). Oxford, Lehrian: Blackwell Science.
- Latifah, M. N., Abdullah, H., Aziz, I., Fauziah, O., & Talib, Y. (2009). Quality changes of rambutan fruit in different packaging system. *Journal of Tropical Agriculture and Food Science*, 37(2), 143-151.

- Lee, P., Tan, R., Yu, B., Curran, P., & Liu, S. (2013). Sugars, organic acids, and phenolic acids of exotic seasonable tropical fruits. *Nutrition & Food Science*, 43(3), 267–276. doi:10.1108/00346651311327927
- Lee, S. T., Radu, S., Ariffin, A., & Ghazali, H. M. (2015). Physico-chemical characterization of oils extracted from noni, spinach, lady's finger, bitter gourd and mustard seeds, and copra. *International Journal of Food Properties*, 18(11), 2508–2527. doi:10.1080/10942912.2014.986577
- Leong, L. P. & Shui, G. (2002). An investigation of antioxidant capacity of fruits in Singapore markets. *Food Chem* 76: 69–75.
- Lees, R., & Jackson, E. B. (1975). Cocoa beans. In: Lees, R., & Jackson, E. B. (Eds.), *Sugar confectionary and chocolate manufacture* (pp. 121–123). New York: Chemical Publishing Co.
- Lefeber, T., Gobert, W., Vrancken, G., Camu, N., & Vuyst, L. De. (2011). Dynamics and species diversity of communities of lactic acid bacteria and acetic acid bacteria during spontaneous cocoa bean fermentation in vessels. *Food Microbiology*, 28(3), 457–464. doi:10.1016/j.fm.2010.10.010
- Lehrian, D. W., & Patterson, G. R., 1983. Cacao fermentation. In: Rehm, H. J., & Reed, G. (Eds.), *Biotechnology, food and feed production with microorganisms*, (Vol. 5, pp. 529-575). Weinheim: VCH Verlag.
- Leong, L. P., & Shui, G. (2002). An investigation of antioxidant capacity of fruits in Singapore markets. *Food Chemistry*, 76(1), 69–75. doi:10.1016/S0308-8146(01)00251-5
- Lestari, S. R., Djati, M. S., Rudijanto, A., & Fatchiyah. (2013). Production and potency of local rambutan at East Java as a candidate phytopharmaca. *Agrivita*, 35(3), 270–276.
- Li, Y., Feng, Y., Zhu, S., Luo, C., Ma, J., & Zhong, F. (2012). The effect of alkalization on the bioactive and flavor related components in commercial cocoa powder. *Journal of Food Composition and Analysis*, 25(1), 17–23. doi:10.1016/j.jfca.2011.04.010
- Lima, L. J. R., Almeida, M. H., Nout, M. J. R., & Zwietering, M. H. (2011). *Theobroma cacao* L., “The food of the Gods”: quality determinants of commercial cocoa beans, with particular reference to the impact of fermentation. *Critical Reviews in Food Science and Nutrition*, 51(8), 731–61. doi:10.1080/10408391003799913
- Lipp, M., & Anklam, E. (1998). Review of cocoa butter and alternative fats for use in chocolate—Part B. Analytical approaches for identification and determination. *Food Chemistry*, 62(1), 99–108. doi:10.1016/S0308-8146(97)00161-1
- Lohachoompol, V., Srzednicki, G., & Craske, J. (2004). The change of total anthocyanins in blueberries and their antioxidant effect after drying and freezing. *Journal of Biomedicine and Biotechnology*, 2004(5), 248–252. doi:10.1155/S1110724304406123

- Loizzo, M. R., Pacetti, D., Lucci, P., Núñez, O., Menichini, F., Frega, N. G., & Tundis, R. (2015). *Prunus persica* var. *platycarpa* (Tabacchiera Peach): bioactive compounds and antioxidant activity of pulp, peel and seed ethanolic extracts. *Plant Foods for Human Nutrition*, 70(3), 331–337. doi:10.1007/s11130-015-0498-1
- Lopez, A. S. and Passos, F. M. L. (1984) Factors influencing cocoa beans acidity fermentation, drying and the microflora. *Proceeding of 9th International Cacao Research Conference, Thome, Togo* (pp. 701-704).
- Low, F. F. (2011). *Effect of fermentation and roasting of rambutan seeds (need to know first the sugar, organic acid, TSS, TA and pH) on rambutan powder production* (Bachelor's thesis). Universiti Putra Malaysia.
- Luma, K. H., Saadoon, F., Varastegani, B., Yang, T. A., & Zzaman, W. (2017). Physical chemical properties of fermented and roasted rambutan seed fat (RSF) as a potential source of cocoa butter replacer. *International Journal on Advanced Science Engineering Information Technology*, 7(1), 57–62.
- Luma, K. H., Yang, T. A., & Fered, S. A. (2015). Study on color and antioxidant properties of rambutan seed fat as cocoa butter alternative. *International Journal On Advanced Science Engineering Information Technology*, 5(2), 34–38.
- Lv, Z., Chen, K., Zeng, Y., & Peng, Y. (2011). Nutritional composition of *Canarium pimela* L. kernels. *Food Chemistry*, 125(2), 692–695. doi:10.1016/j.foodchem.2010.09.067
- Maheshwari, B., Reddy, S. Y. (2005). Application of kokum (*Garcinia indica*) fat as cocoa butter improver in chocolate. *Journal of the Science of Food and Agriculture* 85, 135–140.
- Maisuthisakul, P., Suttajit, M., & Pongsawatmanit, R. (2007). Assessment of phenolic content and free radical-scavenging capacity of some Thai indigenous plants. *Food Chemistry*, 100(4), 1409–1418. doi:10.1016/j.foodchem.2005.11.032
- Malaysian Cocoa Board. (2017). Cocoa cultivated area by region and sector (ha). Retrieved June 29, 2017, from <http://www.koko.gov.my/lkm/industry/statistic/cocoacultivated.cfm>
- Manaf, Y. N. A., Marikkar, J. M. N., Long, K., & Ghazali, H. M. (2013). Physico-chemical characterisation of the fat from red-skin rambutan (*Nephellium lappaceum* L.) seed. *Journal of Oleo Science*, 62(6), 335–43.
- Maximo, G. J., Costa, M. C., & Meirelles, A. J. A. (2013). Solid-liquid equilibrium of triolein with fatty alcohols. *Brazilian Journal of Chemical Engineering*, 30(1), 33–43. doi:10.1590/S0104-66322013000100005
- McGinley, L. (1991). Analysis and quality control for processing and processed fats. In J. B. Rossel & J. L. R. Pritchard (Eds.), *Analysis of oilseeds, fats and fatty foods* (pp. 441–498). London: Elsevier Applied Science.

- Meara, M. L. (1979). Confectionery fats. In: *Proceedings of the International Congress on Oilseeds and Oils*, New Dehli, India.
- Mehdizadeh, S., Lasekan, O., Muhammad, K., & Baharin, B. (2014). Variability in the fermentation index, polyphenols and amino acids of seeds of rambutan (*Nephelium lappaceum* L.) during fermentation. *Journal of Food Composition and Analysis*, 37, 128–135. doi:10.1016/j.jfca.2014.06.017
- Mendoza Jr, D. B., Ramos Jr, P. T., & del Mundo, C. R. (1982). Maturity and ripening guide for rambutan cultivars. *ASEAN Postharvest Horticulture Training and Research Center Technical Bulletin No. 5*. University of the Philippines Los Baños.
- Mendoza, D. B., Pantastico, E. B., & Javier, F. B. (1971). Storage and handling of rambutan (*Nephelium lappaceum* L.). *The Philippine Journal of Agriculture*, 55, 322-332.
- Meursing, E. H. and Zijderveld, J. A. (1999). Cocoa mass, cocoa butter and cocoa powder. In: Beckett, S. T. (Ed.), *Industrial Chocolate Manufacture and Use* (pp. 101–114). Oxford: Blackwell Science.
- Meyer, B, Biehl, B, Said M, Samarakoddy, R. J. (1989). Postharvest pod storage: a method for pulp preconditioning to impair strong nib acidification during cocoa fermentation in Malaysia. *Journal of the Science of Food and Agriculture*, 48, 285-304.
- Meyer, B. N., Ferrigni, N. R., Putnam, J. E., Jacobsen, L. B., Nichols, D. E., & McLaughlin, J. L. (1982). Brine shrimp: a convenient general bioassay for active plant constituents. *Planta Medica*, 45(5), 31–34. doi:10.1055/s-2007-971236
- Minifie, B.W. (1989). *Chocolate, Cocoa and Confectionery: Science and Technology*. Gaithersburg, Maryland: Aspen Publishers, Inc.
- Ministry of Agriculture & Agro-Based Industry Malaysia. (2011). Memperkukuhkan industri sayur-sayuran dan buah buahan serta memulihkan industri kelapa. In *Dasar Agromakanan Negara 2011-2020* (pp. 67-75). Putrajaya: Ministry of Agriculture & Agro-Based Industry Malaysia.
- Mohamed, S., & Othman, E. (1988). Effect of packaging and modified atmosphere on the shelf life of rambutan (*Nephelium lappaceum*) I. *Pertanika*, 11(2), 217–228.
- Mohos, F. (2017). *Confectionery and chocolate engineering: principles and applications*. West Sussex, UK: John Wiley & Sons.
- Montoso Garden. (2007). *Theobroma cacao* (Sterculiaceae). Retrieved August 18, 2017, from http://www.montosogardens.com/theobroma_cacao.htm
- Morales, F. J., Somoza, V., & Fogliano, V. (2012). Physiological relevance of dietary melanoidins. *Amino Acids*, 42, 1097–1109. doi:10.1007/s00726-010-0774-1

- Moreton, R.S., 1988. Single cell oil. In: Moreton, R.S. (Ed.), *Physiology of lipid accumulating yeast* (pp. 1-32). London: Longman Scientific and Technical.
- Morini, G., & Maga, J. A. (1995). Changes in the fatty acid composition of roasted and boiled Chinese (*Castanea molissima*) and Italian (*C. sativa*) chestnuts grown at the same location. *Food Flavors: Generation, Analysis and Process Influence*, 563–568.
- Morton, J. F. (1987). Rambutan. In: Morton, J. F. (Ed.), *Fruits of warm climates* (pp. 262–265). Miami: J.F.Morton.
- Muzquiz, M., Produccion, D. De, Ridout, C. L., Price, K. R., & Fenwick, G. R. (1993). The saponin content and composition of sweet and bitter lupin seed. *Journal of the Science of Food and Agriculture*, 63, 47–52.
- Nakae, T., & Elliott, J. A. (1965). Production of volatile fatty acids by some lactic acid bacteria. ii. selective formation of volatile fatty acids by degradation of amino acids. *Journal of Dairy Science*, 48(3), 293–299.
doi:10.3168/jds.S0022-0302(65)88218-2
- Nateghi, L., Roohinejad, S., Totousaus, A., Mirhosseini, H., & Shuhaimi, M. (2012). Optimization of textural properties and formulation of reduced fat Cheddar cheeses containing fat replacers. *Journal of Food, Agriculture & Environment*, 10(2), 46–54.
- National Bureau of Agricultural Commodity and Food Standards. (2006). *Thai Agricultural Standard TAS 12-2006 Rambutan*. Chatuchak, Bangkok.
- National Confectioners Association. (2012). Cocoa bean growing regions. Retrieved October 26, 2013, from <http://www.candyusa.com/FunStuff/content.cfm?ItemNumber=1028>
- Nayak, C. A., Rastogi, N. K., Raghavarao, K. S. M. S. (2010). Bioactive constituents present in *Garcinia indica Choisy* and its potential food applications – a review. *International Journal of Food Properties*, 13, 441–453.
- Nazaruddin, R., Ayub, M. Y., Mamot, S. & Heng, C. H. (2001). HPLC determination of methylxanthines and polyphenols levels in cocoa and chocolate products, *Malaysian Journal Analytical Science*, 7, 377-386.
- Nazaruddin, R., Seng, L. K., Hassan, O., & Said, M. (2006). Effect of pulp preconditioning on the content of polyphenols in cocoa beans (*Theobroma Cacao*) during fermentation. *Industrial Crops and Products*, 24(1), 87–94.
doi:10.1016/j.indcrop.2006.03.013
- Ng, S.K. and Thamboo, S. (1967). Nutrient removal studies on Malaysian fruits – durian and rambutan. *Malaysian Agricultural Journal*, 46, 164-183.
- Nieburg, O. (2014). Chocolate finds a new friend in yeast. Retrieved August 8, 2017, from <http://www.confectionerynews.com/R-D/Chocolate-finds-a-new-friend-in-yeast>

- Nielsen, D. S., Teniola, O. D., Ban-Koffi, L., Owusu, M., Andersson, T. S., & Holzapfel, W. H. (2007). The microbiology of Ghanaian cocoa fermentations analysed using culture-dependent and culture-independent methods. *International journal of food microbiology*, *114*(2), 168-186. doi:10.1016/j.ijfoodmicro.2006.09.010
- Norbert. (2017). Making chocolate like the Mayas used to. Retrieved August 8, 2017, from <https://www.globotreks.com/destinations/belize/making-chocolate-like-the-mayas-used-to/>
- O'Hare, T. J. (1995). Postharvest physiology and storage of rambutan. *Postharvest Biology and Technology*, *6*(3-4), 189-199.
- O'Hare, T. J., Prasad, A., & Cooke, A. W. (1994). Low temperature and controlled atmosphere storage of rambutan. *Postharvest Biology and Technology*, *4*(1-2), 147-157. doi:10.1016/0925-5214(94)90016-7
- Office of Agricultural Economics, Ministry of Agriculture and Cooperatives Thailand. (2014). *Agricultural Statistics of Thailand 2013*. Nonthaburi: The Agricultural Co-operative Federation of Thailand, Limited Publisher, Branch no. 4.
- Okonogi, S., Duangrat, C., Anuchpreeda, S., Tachakittirungrod, S., & Chowwanapoonpohn, S. (2007). Comparison of antioxidant capacities and cytotoxicities of certain fruit peels. *Food Chemistry*, *103*(3), 839-846. doi:10.1016/j.foodchem.2006.09.034
- Okwu, D. E. (2005). Phytochemicals, vitamins and mineral contents of two Nigerian medicinal plants. *International Journal of Molecular Medicine and Advance Sciences*, *1*(4), 375-381.
- Olajide, J. O., Ade-Omowaye, B. I. O., Otunola, E. T. (2000). Some physical properties of shea kernel. *Journal of Agricultural Engineering Research*, *76*, 419-421.
- Onwulata, C. (2005). *Encapsulated and powdered foods*. London: Taylor and Francis.
- Oracz, J., Nebesny, E., & Zyzelewicz, D. (2014). Effect of roasting conditions on the fat, tocopherol, and phytosterol content and antioxidant capacity of the lipid fraction from cocoa beans of different *Theobroma cacao* L. cultivars. *European Journal of Lipid Science and Technology*, *116*(8), 1002-1014. doi:10.1002/ejlt.201300474
- Osman, H., Nasarudin, R., & Lee, S. L. (2004). Extracts of cocoa (*Theobroma cacao* L.) leaves and their antioxidation potential. *Food Chemistry*, *86*(1), 41-46.
- Ostovar, K., & Keeney, P. G. (1973). Isolation and characterization of microorganisms involved in the fermentation of Trinidad's cocoa beans. *Journal of Food Science*, *38*, 611-617.
- Ouattara, H. G., Koffi, B. L., Karou, G. T., Sangare, A., Niamke, S. L., & Diopoh, J. K. (2008). Implication of *Bacillus sp.* in the production of pectinolytic enzymes

- during cocoa fermentation. *World Journal of Microbiology and Biotechnology*, 24, 1753–1760. doi:10.1007/s11274-008-9683-9
- Ozdemir, M., & Devres, O. (2000a). Analysis of color development during roasting of hazelnuts using response surface methodology. *Journal of Food Engineering*, 45(1), 17–24.
- Ozdemir, M., & Devres, O. (2000b). Kinetics of color changes of hazelnuts during roasting. *Journal of Food Engineering*, 44(1), 31–38.
- Ozdemir, M., & Devres, Y. O. (1999). The thin layer drying characteristics of hazelnuts during roasting. *Journal of Food Engineering*, 42(4), 225–233. doi:10.1016/S0260-8774(99)00126-0
- Palanisamy, U., Cheng, H. M., Masilamani, T., Subramaniam, T., Ling, L. T., & Radhakrishnan, A. K. (2008). Rind of the rambutan, *Nephelium lappaceum*, a potential source of natural antioxidants. *Food Chemistry*, 109(1), 54–63. doi:10.1016/j.foodchem.2007.12.018
- Panda, S. K., Behera, S. K., Qaku, X. W., Sekar, S., Ndinteh, D. T., Nanjundaswamy, H. M., ... Kayitesi, E. (2016). Quality enhancement of prickly pears (*Opuntia sp.*) juice through probiotic fermentation using *Lactobacillus fermentum* - ATCC 9338. *LWT-Food Science and Technology*, 75, 453–459. doi:10.1016/j.crvi.2005.08.004
- Pantzaris, T. P., Ahmad, M. J. (2001). Properties and utilization of palm kernel oil. *Palm Oil Developments*, 35, (11–15), 19–23.
- Pasha, C., & Reddy, G. (2005). Food chemistry nutritional and medicinal improvement of black tea by yeast fermentation. *Food Chemistry*, 89, 449–453. doi:10.1016/j.foodchem.2004.02.054
- Passos, F. M. L., Lopez, A. S., & Silva, D. O. (1984). Aeration and its influence on the microbial sequence in cacao fermentations in Bahia, with emphasis on lactic acid bacteria. *Journal of Food Science*, 49(6), 1470–1474. doi:10.1111/j.1365-2621.1984.tb12823.x
- Paul, R., Morgan, S., & Hill, C. (2002). Preservation and fermentation: past, present and future. *International Journal of Food Microbiology*, 79(1-2), 3–16. doi:10.1016/S0168-1605(02)00174-5
- Paull, R. E., & Duarte, O. (2012). Rambutan and pulasan. In Paull, R. E., & Duarte, O. *Tropical fruits* (Vol. 2, pp. 139-160) Wallingford, UK: CABI.
- Paull, R. E., & Duarte, O. (2012). *Tropical fruits: crop production science in horticulture* 24. UK: CABI.
- Pereira, G. V. D. M., Miguel, M. G. D. C. P., Ramos, C. L., & Schwan, R. F. (2012). Microbiological and physicochemical characterization of small-scale cocoa fermentations and screening of yeast and bacterial strains to develop a

- defined starter culture. *Applied and Environmental Microbiology*, 78(15), 5395–405. doi:10.1128/AEM.01144-12
- Pettipher, G. L. (1986). An improved method for the extraction and quantitation of anthocyanins in cocoa beans and its use as an index of the degree of fermentation. *Journal of the Science of Food and Agriculture*, 37, 289–96.
- Piva, A., Di Mattia, C., Neri, L., Dimitri, G., Chiarini, M., & Sacchetti, G. (2008). Heat-induced chemical, physical and functional changes during grape must cooking. *Food Chemistry*, 106(3), 1057–1065. doi:10.1016/j.foodchem.2007.07.026
- Portillo, E., Farinas, L. G. D., & Betancourt, E. (2007). Chemical analysis of Criollo Porcelana cocoa (*Theobroma cocoa* L.) in the south of Maracaibo Lake. *Faculty of Agronomy Journal University of Zulia*, 24, 522-546.
- Prabhakaran Nair, K. P. (2010). Cocoa (*Theobroma cacao* L.). In: Prabhakaran Nair, K. P. (Ed). *The agronomy and economy of important tree crops of the developing world* (pp. 131-179). Burlington: Elsevier.
- Purtle, I. C., Gusek, T. W., Jirjis, B. F., & Boddenheim, F. W. (2010). *Patent No. US7709041 B2*. Washington, DC: U.S. Patent and Trademark Office.
- Quesnel, V. C. (1968). Oxygen consumption and heat production during the fermentation of cacao. *Turrialba (IICA)*, 18(2), 110-114.
- Raina, K., Kumar, S., Kumar, D., Somasagara, R. R., Punia, R., Kant, R., ... Agarwal, C. (2015). Grape seed extract decreases visceral adiposity and impairs the pro-tumorigenic adipose tissue secretions affecting colorectal cancer growth and progression. *Cancer Research*, 75(15 Supplement), 901. doi:10.1158/1538-7445.AM2015-901
- Rajasekaran, A., Ganesan, S., Kamini, N., Lavanya, C., Lee Yoon, L., & Shian Oh, H. (2013). Anti-nociceptive, CNS, antibacterial and antifungal activities of methanol seed extracts of *Nephelium lappaceum* L. *Oriental Pharmacy and Experimental Medicine*, 13, 149–157. doi:10.1007/s13596-012-0095-x
- Ramli, N., Hassan, O., Said, M., Samsudin, W., & Idris, N. A. (2006). Influence of roasting conditions on volatile flavor of roasted Malaysian cocoa beans. *Journal of Food Processing and Preservation*, 30(3), 280–298. doi:10.1111/j.1745-4549.2006.00065.x
- Ranganna, S. (1977). *Manual of analysis of fruit and vegetable products*. New Delhi: Tata MacGraw Hill Company Ltd.
- Reddam, R.M. (2014). Cocoa fermentaion. Retrieved August 18, 2017, from <https://www.slideshare.net/ragamadhureddam/cocoa-fermentation>
- Reddy, N. R., & Pierson, M. D. (1994). Reduction in antinutritional and toxic components in plant foods by fermentation. *Food Research International*, 27(3), 281–290. doi:10.1016/0963-9969(94)90096-5

- Reddy, S. Y., Prabhakar, J. V. (1994). Cocoa butter extenders from Kokum (*Garcinia indica*) and Phulwara (*Madhuca butyracea*) butter. *Journal of the American Oil Chemists' Society*, 7, 217–219.
- Reineccius, G. A., Andersen, D. A., Kavanagh, T. E. & Keeney, P. G. (1972). Identification and quantification of the free sugars in cocoa beans. *Journal of Agriculture and Food Chemistry*, 20, 199–202.
- Reineccius, G., & Henry, B. H. (2006). *Flavor Chemistry and Technology*. New York: Taylor and Francis.
- Rodriguez-Campos, J., Escalona-Buendía, H. B., Contreras-Ramos, S. M., & Orozco-Avila, I. (2012). Effect of fermentation time and drying temperature on volatile compounds in cocoa. *Food Chemistry*, 132(1), 277–288. doi:10.1016/j.foodchem.2011.10.078
- Rodriguez-Campos, J., Escalona-Buendía, H. B., Orozco-Avila, I., Lugo-Cervantes, E., & Jaramillo-Flores, M. E. (2011). Dynamics of volatile and non-volatile compounds in cocoa (*Theobroma cacao* L.) during fermentation and drying processes using principal components analysis. *Food Research International*, 44, 250–258. doi:10.1016/j.foodres.2010.10.028
- Rohan, T. A. (1963). Processing of raw cocoa for the market. In: Food and Agriculture Organization of the United Nations, *Agricultural studies Technical Bulletin No 60* (pp. 77-95). Italy: Food and Agriculture Organization of the United Nations.
- Rohan, T. A. (1964). The Precursors of chocolate aroma: a comparative study of fermented and unfermented cocoa beans. *Journal of Food Science*. 29, 456–459.
- Rohan, T. A. (1967). The precursors of chocolate aroma: application of gas chromatography in following formation during fermentation of cocoa beans. *Journal of Food Science*. 32, 402–404.
- Rohan, T. A., & Stewart, T. (1964). The volatile and non-volatile acids of cocoa beans. *Rev. Int. Choc.*, 19(1 1), 503-505.
- Rohan, T. A., & Stewart, T. (1966). The precursors of chocolate aroma: changes in the free amino acids during the roasting of cocoa beans. *Journal of Food Science*. 31, 202–205.
- Rohan, T. A., & Stewart, T. (1967). The precursors of chocolate aroma: production of reducing sugars during fermentation of cocoa beans. *Journal of Food Science*, 32, 399–402.
- Romain, V., Ngakegni-Limbili, A. C., & Ouamba, J. (2013). Thermal properties of monoglycerides from *Nephelium Lappaceum* L. oil, as a natural source of saturated and monounsaturated fatty acids. *Industrial & Engineering Chemistry Research*, 52, 14089–14098. doi:dx.doi.org/10.1021/ie401875v

- Romero-Cortes, T. (2012). Isolation and characterization of acetic acid bacteria in cocoa fermentation. *African Journal of Microbiology Research*, 6(2), 339–347. doi:10.5897/AJMR11.986
- Romero-Cortes, T., Salgado-Cervantes, M. A., García-Alamilla, P., García-Alvarado, M. A., Rodríguez-Jimenes, G. D. C., Hidalgo-Morales, M., & Robles-Olvera, V. (2013). Relationship between fermentation index and other biochemical changes evaluated during the fermentation of Mexican cocoa (*Theobroma cacao*) beans. *Journal of the Science of Food and Agriculture*, 93(10), 2596–604. doi:10.1002/jsfa.6088
- Sacchetti, G., Ioannone, F., Gregorio, M. De, & Di, C. (2016). Non enzymatic browning during cocoa roasting as affected by processing time and temperature. *Journal of Food Engineering*, 169, 44–52. doi:10.1016/j.jfoodeng.2015.08.018
- Said, M., & Samarakhody, R. (1984). Cocoa fermentation - effect of surface area, frequency of turning and depth of cocoa masses. In *International Conference on Cocoa and Coconuts* (pp. 1–15).
- Saltini, R., Akkerman, R., & Frosch, S. (2013). Optimizing chocolate production through traceability: a review of the influence of farming practices on cocoa bean quality. *Food Control*, 29(1), 167–187. doi:10.1016/j.foodcont.2012.05.054
- Samah, A. O., Ibrahim, N., Alimon, H., & Karim, A. M. I. (1993). Fermentation studies of stored cocoa beans. *World Journal of Microbiology & Biotechnology*, 9, 603–604. doi:10.1007/BF00386306.
- Sanagi, M. M., Hung, W. P. and Yasir, S. M. (1997). Supercritical fluid extraction of pyrazines in roasted cocoa beans: effect of pod storage period. *Journal of Chromatography A*, 785, 361–367.
- Satyarthi, J. K., Srinivas, D., & Ratnasamy, P. (2011). Hydrolysis of vegetable oils and fats to fatty acids over solid acid catalysts. *Applied Catalysis A: General*, 391(1), 427–435. doi:10.1016/j.apcata.2010.03.047
- Sauer, M., Branduardi, P., Valli, M., & Porro, D. (2004). Production of L-ascorbic acid by metabolically engineered *Saccharomyces cerevisiae* and *Zygosaccharomyces bailii*. *Applied And Environmental Microbiology*, 70(10), 6086–6091. doi:10.1128/AEM.70.10.6086
- SCAC Network. (1999). Madagascar cocoa bean for export. Retrieved August 18, 2017, from http://www.export-forum.com/africa/madagascar_cocoa.htm.
- Schwan, R. F., & Wheals, A. E. (2004). The microbiology of cocoa fermentation and its role in chocolate quality. *Critical Reviews in Food Science and Nutrition*, 44(4), 205–21. doi:10.1080/10408690490464104
- Selamat, J. (1994). Organic acids in cocoa beans - A review. *ASEAN Food Journal*, 9(1), 3–12.

- Selvaraj, Y. & Pal, D. K. (1984). Changes in the chemical composition and enzyme activity of two sapodilla (*Manilkara zapota*) cultivars during development and ripening. *J Hortic Sci*, 59: 275–281
- Senanayake, M., Jansz, E. R., & Buckle, K. A. (1995). Effect of variety and location on optimum fermentation requirements of cocoa beans: An aid to fermentation on a cottage scale. *Journal of the Science of Food and Agriculture*, 69(4), 461–465. doi: 10.1002/jsfa.2740690410
- Senanayake, M., Jansz, E. R., & Buckle, K. A. (1997). Effect of different mixing intervals on the fermentation of cocoa beans. *Journal of the Science of Food and Agriculture*, 74, 42–48.
- Sengun, I. Y., & Karabiyikli, S. (2011). Importance of acetic acid bacteria in food industry. *Food Control*, 22(5), 647–656. doi:10.1016/j.foodcont.2010.11.008.
- Shao, Y., Xie, J., Chen, P., & Li, W. (2012). Changes in some chemical components and in the physiology of rambutan fruit (*Nephelium lappaceum* L.) as affected by storage temperature and packing material. *Fruits*, 68(1), 15–24. doi:10.1051/fruits/2012045
- Sharif, S. (1997). *Effect of alkalization and quality of cocoa liquor from different origins* (Master Science thesis). Pennsylvania State University, USA.
- Sharma, S. K. (2012). Cocoa: Scientific aspects of its processing – Secondary processing. Retrieved August 14, 2017, from http://www.foodpathshala.com/news_details.php?news_id=59.
- Sheng, L., Olsen, S. A., Hu, J., Yue, W., Means, W. J., & Zhu, M. J. (2016). Inhibitory effects of grape seed extract on growth, quorum sensing, and virulence factors of CDC “top-six” non-O157 Shiga toxin producing *E. coli*. *International Journal of Food Microbiology*, 229, 24–32. doi:10.1016/j.ijfoodmicro.2016.04.001
- Shepherd, R. (1976). Large scale processing of cocoa beans temperature and acidity trends. *Planters*. 52, 311–322.
- Siriphanich, J. (2002). Postharvest physiology of tropical fruit. *Acta Horticulturae*, 575, 623 – 633.
- Siriphanich, J. (2003). *Physiology and postharvest technology of fruit and vegetable*. 5th ed. Bangkok: Kasetsart University.
- Sirisompong, W., Jirapakkul, W., & Klinkeorn, U. (2011). Response surface optimization and characteristics of rambutan (*Nephelium lappaceum* L.) kernel fat by hexane extraction. *LWT - Food Science and Technology*, 44(9), 1946–1951. doi:10.1016/j.lwt.2011.04.011
- Soeng, S., Evacuasiyany, E., Widowati, W., Fauziah, N., Manik, V., & Maesaroh, M. (2015). Inhibitory potential of rambutan seeds extract and fractions on adipogenesis in 3T3-L1 cell line. *Journal of Experimental and Integrative Medicine*, 5(1), 55. doi:10.5455/jeim.200115.or.120

- Solis-Fuentes, J. A., & Durán-de-Bazúa, M. C. (2004). Mango seed uses: thermal behaviour of mango seed almond fat and its mixtures with cocoa butter. *Bioresource Technology*, 92(1), 71–78. doi:10.1016/j.biortech.2003.07.003
- Solis-Fuentes, J. A., Camey-Ortíz, G., Hernández-Medel, M. D. R., Pérez-Mendoza, F., & Durán-de-Bazúa, C. (2010). Composition, phase behavior and thermal stability of natural edible fat from rambutan (*Nephelium lappaceum* L.) seed. *Bioresource Technology*, 101(2), 799–803. doi:10.1016/j.biortech.2009.08.031
- Somboon, Y. (1984). *Effect of temperature and maturity stages on biochemical changes during storage of rambutan (Nephelium lappaceum Linn.) cv. Seechompoo and cv. Rongrien* (Master Science thesis). Kasetsart University, Thailand.
- Sonwai, S., & Ponprachanuvut, P. (2012). Characterization of physicochemical and thermal properties and crystallization behavior of krabok (*Irvingia Malayana*) and rambutan seed fats. *Journal of Oleo Science*, 61(12), 671–9. doi:10.5650/jos.61.671
- Sopie Salomé, Y.E., Laurent, K.K., Pierre Irénée, B.J., Patrice, K., & Hilaire, K.T. (2011). Comparison of pineapple fruit characteristics of plants propagated in three different ways: By suckers, micropropagation and somatic embryogenesis. *J Nutr Food Sci* DOI: 10.4172/2155-9600.1000110.
- Srilaong, V. S., Anlayanarat, S. K., & Atsumi, Y. T. (2002). Changes in commercial quality of “Rong-Rien” rambutan in modified atmosphere. *Food Science and Technology Research*, 8(4), 337–341.
- Sturm, K., Koron, D., & Stampar, F. (2003). The composition of fruit of different strawberry varieties depending on maturity stage. *Food Chemistry*, 83(3), 417–422. doi:10.1016/S0308-8146(03)00124-9
- Suazo, Y., & Davidov-pardo, G. (2014). Effect of fermentation and roasting on the phenolic concentration and antioxidant activity of cocoa from Nicaragua. *Journal of Food Quality*, 37(1), 50–56.
- Suhendi, A., & Muhtadi. (2015). Potential activity of rambutan (*Nephelium lappaceum* L.) fruit peel extract as antidiabetic and antihypercholesterolemia. In *The 2nd International Conference on Engineering Technology and Industrial Application* (pp. 20–23).Surakarta. doi:10.13140/RG.2.1.2424.9042
- Sukasih, E., & Setyadjit. (2015). Development of new product: rambutan pulpy juice. *Procedia Food Science*, 3, 413–425. doi:10.1016/j.profoo.2015.01.046
- Swe, P.Z., Che Man, Y.B., & Ghazali, H.M. (1996). Improved NARP-HPLC Method for Separating Triglycerides of Palm Olein and Its Solid Fractions Obtained at Low Temperature Storage. *Food Chemist*, 56, 181–186.
- Talbot, G. (1999). Vegetable fats. In: Beckett, S. T., (Ed.), *Industrial Chocolate Manufacture and Use* (pp. 218–230). Oxford: Blackwell Science.

- Tee, E. S., Mohd Ismail, N., Mohd Nasir, A., & Khatijah, I. (1997). *Nutrient composition of Malaysian foods*. Kuala Lumpur: Institute Medical for Research.
- Tejas Chocolate Craftory. (2017). How chocolate is made. Retrieved August, 18, 2017, from <http://tejaschocolate.com/home/howcraftchocolateismade/>
- Terés, S., Barceló-Coblijn, G., Benet, M., Alvarez, R., Bressani, R., Halver, J. E., & Escribá, P. V. (2008). Oleic acid content is responsible for the reduction in blood pressure induced by olive oil. *Proceedings of the National Academy of Sciences of the United States of America*, 105(37), 13811–13816. doi:10.1073/pnas.0807500105
- The Chocolate Revolution. (2013). The cocoa bean. Retrieved March 7, 2013, from <http://www.chocolate-revolution.com/beans.php>
- Thitilertdecha, N., Teerawutgulrag, A., & Rakariyatham, N. (2008). Antioxidant and antibacterial activities of *Nephelium lappaceum* L. extracts. *LWT - Food Science and Technology*, 41(10), 2029–2035. doi:10.1016/j.lwt.2008.01.017
- Thitilertdecha, N., Teerawutgulrag, A., & Rakariyatham, N. (2008). Antioxidant and antibacterial activities of *Nephelium lappaceum* L. extracts. *LWT - Food Science and Technology*, 41(10), 2029–2035. doi:10.1016/j.lwt.2008.01.017
- Thompson, S. S., Miller, K. B., Lopez, A. S. (2001). Cocoa and coffee. In Doyle, M. P., & Buchanan, R. L. (Eds.), *Food microbiology - fundamentals and frontiers* (pp. 721-733). Washington: ASM Press.
- Thompson, S. S, Miller, K. B, & Lopez, A. S. (2001). Cocoa and coffee. In Doyle, M. J, Beuchat, L. R, & Montville, T. J. (Eds.), *Food Microbiology - Fundamentals and Frontiers* (pp. 721-733). Washington: ASM Press.
- Timms, R. E. (2003). Interactions between fats, bloom and rancidity. In Timms, R. E. (Ed.), *Confectionery fats handbook: properties, production and applications* (pp. 255–294). Bridgwater, UK: The Oily Press.
- Tindall, H. D. (1994). Sapindaceous fruits : botany and horticulture. In J. Janick (Ed.), *Horticultural reviews* (Vol. 16, pp. 143–196). London: John Wiley & Sons.
- Tomas-Barberán, F. A., Cienfuegos-Jovellanos, E., Marín, A., Muguerza, B., Gil-Izquierdo, A., Cerdá, B., ... Espin, J. C. (2007). A new process to develop a cocoa powder with higher flavonoid monomer content and enhanced bioavailability in healthy humans. *Journal of Agricultural and Food Chemistry*, 55, 3926–3935. doi:10.1021/jf070121j
- Tomlins, K. I., Baker, D. M., Daplyn, P., & Adomako, D. (1993). Effect of fermentation and drying practices on the chemical and physical profiles of Ghana cocoa. *Food Chemistry*, 46(3), 257-263.

- Tongumpai, P. (1980). *The study on floral, fruit and pulp development of rambutan (Nephelium lappaceum L. 'Seechompoo')* (Master Science Thesis). Kasetsart University, Thailand.
- Undurraga, D., Markovits, A., Erazo, S. (2001). Cocoa butter equivalent through enzymic interesterification of palm oil mid-fraction. *Process Biochemistry* 36, 933–939.
- Van Welzen, P. C. and Verheij, E. W. M. (1991). *Nephelium lappaceum L.* In: Verheij, E.W.M., & Coronel, R.E. (Eds.), *Plant Resources of South-East Asia. No. 2. Edible Fruits and Nuts* (pp. 235-240). Wageningen: Pudoc.
- Visetbhakdi, N. (1988). Fruit exports. *Bangkok Bank Mont Review*, 29(11), 466-473.
- Voon, Y. Y., Abdul, N. S., Rusul, G., Osman, A., & Quek, S. Y. (2006). Physicochemical, microbial and sensory changes of minimally processed durian (*Durio zibethinus* cv. D24) during storage at 4 and 28 °C. *Postharvest Biology and Technology*, 42, 168–175. doi:10.1016/j.postharvbio.2006.06.006
- Voort, F. R. Van De, Memon, K. P., Sedman, I., & Ismail, A. A. (1996). Determination of solid fat index by fourier transform infrared spectroscopy. *Journal of the American Oil Chemists' Society*, 73(4), 411–416.
- Wall, M. M. (2006). Ascorbic acid and mineral composition of longan (*Dimocarpus longan*), lychee (*Litchi chinensis*) and rambutan (*Nephelium lappaceum*) cultivars grown in Hawaii. *Journal of Food Composition and Analysis*, 19(6-7), 655–663. doi:10.1016/j.jfca.2005.12.001
- Watson, B. J. (1988). Rambutan cultivars in north Queensland. *Queensland Agricultural Journal, Jan-Feb*, 37-41.
- Whitefield, R. (2005). *Making chocolates in the factory*. London: Kenedy's Publications Ltd.
- Widjanarko, S. B., Trisnawati, C. Y., & Susanto, T. (2000). Changes in respiration, composition and sensory characteristics of rambutan packed with plastic films during storage at low temperature. *Journal of Agricultural Technology*, 1(3), 1-8.
- Wijekoon, M.M.J.O., Bhat, R. & Karim, A.A. (2011). Effect of Extraction Solvents on the Phenolic Compounds and Antioxidant Activities of Bunga Kantan (*Etligeria elatior* Jack.) Inflorescence. *Journal of Food Composition and Analysis*, 24, 615–619.
- Winayanuwattikun, P., Kaewpiboon, C., Piriyananon, K., Tantong, S., Thakernkarnkit, W., Chulalaksananukul, W., & Yongvanich, T. (2008). Potential plant oil feedstock for lipase-catalyzed biodiesel production in Thailand. *Biomass and Bioenergy*, 32(12), 1279–1286. doi:10.1016/j.biombioe.2008.03.006

- Wollgast, J., & Anklam, E. (2000). Review on polyphenols in *Theobroma cacao*: changes in composition during the manufacture of chocolate and methodology for identification and quantification. *Food Research International*, 33(6), 423-447.
- Wood, G. A. R. (1975). *Cocoa* (3rd edition). London: Long Group Limited.
- Wood, G. A. R., & Lass, R. A. (2008). From harvest to store. In: Wood, G. A. R. (Ed.), *Cocoa* (4th edition pp.444-504). London: John Wiley & Sons.
- Yang, J., Bingol, G., Pan, Z., Brandl, M. T., Mchugh, T. H., & Wang, H. (2010). Infrared heating for dry-roasting and pasteurization of almonds. *Journal of Food Engineering*, 101(3), 273–280. doi:10.1016/j.jfoodeng.2010.07.007
- Yanty, B. N. A. M., Marikkar, J. M. N., & Miskandar, M. S. (2012). Comparing the thermo-physical characteristics of lard and selected plant fats. *Grasas Y Aceites*, 63(3), 328–334. doi:10.3989/gya.023712
- Zahouli, G. I. B., Guehi, S. T., Fae, A. M., Ban-Koffi, L., & Nemlin, J. G. (2010). Effect of drying methods on the chemical quality traits of cocoa raw material. *Advance Journal of Food Science and Technology*, 2, 184–190.
- Zaidul, I.S.M., Norulaini, N.A.N., Mohd Omar, A.K., Smith Jr., R.L. (2006). Supercritical carbon dioxide (SC-CO₂) extraction and fractionation of palm kernel oil from palm kernel as cocoa butter replacers blend. *Journal of Food Engineering*, 73(3), 210–216.
- Zaidul, I.S.M., Norulaini, N.A.N., Omar, A.K.M., Smith Jr., R.L. (2007). Blending of supercritical carbon dioxide (SC-CO₂) extracted palm kernel oil fractions and palm oil to obtain cocoa butter replacers. *Journal of Food Engineering*, 78(4), 1397–1409.
- Zee, F.T. (1993). Rambutan and pilinuts: potential crops of Hawaii. In: Janick, J. and Simon, J.E. (Eds.), *New Crops* (pp.461-465). New York: John Wiley and Sons Inc.
- Zzaman, W., & Yang, T. A. (2014). Moisture, color and texture changes in cocoa beans during superheated steam roasting. *Journal of Food Processing and Preservation*, 38(3), 1364–1370. doi:10.1111/jfpp.12098