



**UNIVERSITI PUTRA MALAYSIA**

***EVALUATION OF LOCAL SPICES AND HERBS AS MARINATING  
INGREDIENTS FOR THE REDUCTION OF HETEROCYCLIC AMINES  
IN GRILLED BEEF***

**SHABNAM SEPAHPOUR**

**FSTM 2018 3**



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IN GRILLED BEEF**

By

**SHABNAM SEPAHPOUR**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

**May 2017**

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## **DEDICATION**

This thesis is dedicated to

My Dear parents

Who taught me to be strong and tolerant  
&  
Whose souls are my eternal guardian angels



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

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**May 2017**

**Chairman : Professor Jinap Selamat, PhD**  
**Faculty : Food Science and Technology**

Heterocyclic aromatic amines (HCAs) are an important group of food mutagens and potential carcinogens which are formed during heat processing. Phenolic compounds, ubiquitous in plants are well known for their free radical scavenging activities which can inhibit the formation of HCAs. There is no study which have investigated the effects of different proportions of spices/herbs on the reduction of HCAs in grilled beef. This research has been conducted to determine the antioxidant property of selected herbs and spices, optimize the proportion of their combination to achieve the highest reduction of HCAs, identify the main bioactive inhibitors of HCAs formation and explore the level of mutagenicity and toxicity of marinated grilled beef. Selected herbs and spices (turmeric, curry leaf, lemon grass and torch ginger) were evaluated for their total phenolic compound (TPC), total phenolic content (TFC), DPPH radical scavenging activity and ferric reducing antioxidant power (FRAP). The bioactive compounds of the samples were identified using high performance liquid chromatography (HPLC). Nineteen combinations of selected herbs and spices (turmeric, curry leaf, lemon grass and torch ginger) based on a simplex centroid mixture design were used to marinate beef in order to determine the optimal proportion which achieve the highest reduction of HCAs. The results showed that the combination of turmeric and lemon grass exhibited the highest reduction of HCAs. The optimum mixture of crude extract of these herbs was fractionated to determine the main inhibitors of PhIP formation. Six fractions were collected and applied into PhIP model system containing glucose, phenylalanine and creatinine. Liquid chromatography-mass spectrometry quadruple time of flight (LC-MS/MS Q-TOF) and orthogonal partial least-squares (OPLS) analysis was used to correlate PhIP reduction to different metabolite profiles of each fraction. *In vivo* and *in vitro* experiments were respectively carried out using zebrafish embryo and *Salmonella* Typhimurium (Ames test), to determine whether total HCAs reduction in optimized

marinated grilled beef exhibited comparable reduction of toxicity and mutagenicity. The results of experiment showed that turmeric possessed the significantly highest TPC (172.10 mgGA/dw), TFC (380.7 mgQE/dw), DPPH antioxidant activity (47.35%) and FRAP (55.79 mgQE/gdw basis) which was followed by curry leaf, torch ginger and lemon grass. The combination of turmeric with lemon grass (50:50 w/w) showed the significantly highest reduction of total HCAs at 94.7%. Using the response optimizer, the optimal proportion of spices and herbs for reduction of total HCAs was found to be a combination of turmeric:lemon grass with the proportion of 52.42%:47.57%. OPLS results indicated that the fraction 5 exhibited remarkable inhibitory activity in the formation of PhIP. The potential inhibitor compounds in this fraction were identified as luteolin, curcumin, luteolin 6-C-Glucosyl (Isoorientin), luteolin 6-C-Pentosyl-8-C-pentosyl, Pentosyl-8-C-deoxyhexosyl luteolin, apigenin 7-rutinoside, 2"-O-Rhamnosyl isoorientin, Embigenin 2"-(2"-acetylramnoside), apigenin 7-rutinoside-4'-glucoside, luteoin 7-(6'''-acetylloosyl-(1->3)-glucosyl-(1->2)glucoside, Kaempferol 3-rutinoside-7-sophoroside. The results of experiment also showed that the mixture of turmeric and lemon grass (52.42%:47.57%) can be used in the diet of zebrafish without any detrimental effect on liver, kidney, fertility, growth rate and marinades decreased the mutagenic activity in the marinated grilled samples compared with those of the unmarinated samples. The results of the mutagenic activity demonstrated that this optimized marinade formula significantly ( $p < 0.05$ ) diminished mutagenicity of grilled beef in bacterial Ames test.

It can be concluded that all selected herbs and spices utilized in this study possess antioxidant activity which can reduce total HCAs concentration in grilled beef. It is evidenced that combination of turmeric and lemon grass (52.42%:47.57%) gave satisfactory results for the maximum reduction of total HCAs and mutagenicity without any detrimental effect on zebrafish embryo.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

## **PENILAIAN REMPAH DAN HERBA TEMPATAN KE ATAS PENURUNAN AMINA HETEROSIKLIK DALAM DAGING LEMBU PANGGANG**

Oleh

**SHABNAM SEPAHPOUR**

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Amina aromatic heterosiklik (HCAs) adalah kumpulan utama mutagen dan karsinogen makanan yang berpotensi terbentuk semasa proses pemanasan. Sebatian phenolik yang wujud di dalam tumbuhan dikenali dengan kebolehan untuk memerangkap radikal bebas yang mana boleh menghalang pembentukan HCAs. Tiada kajian dilakukan berkaitan kesan campuran rempah/herba ke atas penurunan HCAs dalam daging lembu panggang. Matlamat khusus kajian ini adalah untuk menentukan aktiviti antioksidan herba dan rempah terpilih, mengoptimumkan nisbah gabungan herba dan rempah untuk mencapai penurunan tertinggi bagi HCAs, mengenal pasti pengabalan bioaktif utama pembentukan HCAs dan meneroka tahap mutagen dan ketoksikan daging lembu panggang diperap. Kajian ini telah dijalankan untuk menentukan kesan rempah / herba terpilih pada pengurangan amina heterocyclic (HCAs) dalam daging lembu panggang. Herba terpilih dan rempah (kunyit, daun kari, serai dan bunga kantan) telah dinilai untuk kandungan phenolik, kandungan flavonoid, keupayaan memerangkap radikal bebas menggunakan asay DPPH dan kuasa penurunan antioksidan ion ferik menggunakan asay FRAP. Sebatian bioaktif dalam sampel dikenalpasti dengan menggunakan high performance liquid chromatography, (HPLC). Sembilan belas kombinasi rempah/herba telah (kunyit, daun kari, serai dan bunga kantan) digunakan berdasarkan rekabentuk simplek campuran sentroid telah digunakan untuk menentukan kadar yang optimum yang mencapai penurunan tertinggi sebanyak HCAs. Keputusan menunjukkan kombinasi kunyit dengan serai mempamerkan kadar penurunan HCAs yang tinggi. Campuran optimum ekstrak mentah herba ini telah difraksikan untuk mendapatkan penghalang utama pembentukan PhIP menggunakan HPLC. Enam fraksi telah dikumpul dan digunakan ke dalam sistem model PhIP yang mengandungi glukosa, penilalanin dan kreatinin. Liquid chromatography-mass spectrometry quadruple time of flight (LC-MS/MS Q-TOF) dan orthogonal partial least-squares (OPLS) analisis telah digunakan untuk mengaitkan penurunan pembentukan PhIP dalam fraksi yang berbeza bagi profil

metabolik berbeza untuk setiap fraksi yang diperolehi daripada campuran ekstrak kunyit dan serai. Eksperimen *in vivo* dan *in vitro* telah dilakukan untuk menentukan sama ada jumlah penurunan HCAs dalam daging yang diperap menggunakan perkadaran rempah/herba yang telah dioptimumkan mempamerkan penurunan yang setanding bagi ketoksikan dan kemutagenan dalam embrio ikan zebrafish dan dalam *Salmonella* Typhimurium (Ujian Ames) untuk menentukan sama ada jumlah pengurangan HCAs dalam dioptimumkan daging lembu panggang diperap mempamerkan pengurangan setanding ketoksikan dan mutagen. Keputusan eksperimen menunjukkan bahawa kunyit memiliki TPC tertinggi ( $172.10 \pm 1.40$  mgGA/berat kering), TFC ( $380.7 \pm 5.5$  mgQE/berat kering), aktiviti antioksidan menggunakan assay DPPH ( $47.35 \pm 2.57\%$ ) and kuasa penurunan antioksidan ion ferik menggunakan assay FRAP ( $55.79 \pm 0.39$  mgQE/g berat kering) tinggi dan diikuti oleh daun kari, bunga kantan dan serai. kombinasi kunyit dengan serai (50:50 b/b) telah menurunkan 94.7% kandungan HCAs (204.7 to 10.86 ng/g). Menggunakan pengoptimuman respon, perkadaran optimum rempah/herba untuk penurunan kandungan HCAs adalah 52.42%:47.57% (kunyit:serai). Keputusan OPLS menunjukkan fraksi 5 mempamerkan aktiviti penghalangan pembentukan PhIP yang luar biasa. Sebatian penghalang yang berpotensi dalam fraksi ini dikenalpasti sebagai luteolin, kurkumin, luteolin 6-C-Glucosyl (Isoorientin), luteolin 6-C-Pentosyl-8-C-pentosyl, Pentosyl-8-C-deoxyhexosyl luteolin, apigenin 7-rutinoside, 2"-O-Rhamnosyl isoorientin, Embigenin 2"-(2"-acetylramnoside), apigenin 7-rutinoside-4'-glucoside, luteoin 7-(6'''-acetylloosyl-(1->3)-glucosyl-(1->2)glucoside, Kaempferol 3-rutinoside-7-sophoroside. Keputusan juga menunjukkan campuran boleh digunakan dalam diet ikan zebrafish tanpa kesan yang memudaratkan ke atas hati, buah pinggang, kesuburan, kadar pertumbuhan dan perapan telah menurunkan aktiviti metagenik dalam sampel yang diperap berbanding sampel kawalan yang tidak diperap.

Ia dapat disimpulkan bahawa semua herba dan rempah terpilih yang digunakan dalam kajian ini mempunyai aktiviti antioksidan yang boleh mengurangkan kepekatan jumlah HCAs dalam daging panggang. Ia telah terbukti bahawa gabungan kunyit dan serai (52.42%:47.57%) memberi keputusan yang memuaskan bagi pengurangan maksima jumlah HCAs dan kesan mutagen tanpa sebarang kemudatan pada embrio zebrafish.



## ACKNOWLEDGEMENTS

A great thank to the ever-present God, for giving me the strength to continue and accomplish this work.

This dissertation would not be possible without the guidance and the help of several individuals who contributed their valuable assistance in the preparation and completion of this study.

At first I would like to express my deep sense of respect, gratitude and appreciation to my chief supervisor Prof. Jinap Selamat for the valuable guidance and strong support throughout my study. Working under this supervision was the most memorable and honorable experience I have ever had.

I would like to offer my special thanks to Datu Prof. Dr. Mohd Yazid Manap for constant support, helpful guidance and valuable discussions through my study. I really appreciate the support and advice you gave me during these years.

I wish to express my sincere gratitude to Associate Prof. Dr. Alfi Khatib and Associate Prof. Dr. Ahmad Faizal Abdull Razis for their comments and valuable advice and constructive suggestion and guidance throughout of the duration of this research project.

I wish to express my sincere gratitude to my beloved friends, Dr Atena Pirooz, Dr Gisla Daniali, Dr. Morvarid Akhavan, Milad Kazemi and Sahar Tamnak who believed in me pursuing this PhD as a step forward.

I am offering my sincere thanks to my friend Sharina who helped me in translation my English abstract to Bahasa Malay.

My Special appreciation to my dear friend Daniel who was a great source of motivation, encouragement throughout the period of my thesis writing. I also deeply thank him for helping me in English editing of the thesis.

Finally my special thanks go to my beloved parents and brothers, who encouraged and supported me in all my efforts of PhD Study. I cannot express strongly enough my gratitude for your support and continuous encouragement.

I certify that a Thesis Examination Committee has met on 11 May 2017 to conduct the final examination of Shabnam Sepahpour on her thesis entitled "Evaluation of Local Spices and Herbs as Marinating Ingredients for the Reduction of Heterocyclic Amines in Grilled Beef" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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## LIST OF ABBREVIATIONS

ACs	amino-carbolines
AOAC	Association of Official Analytical Chemist
A $\alpha$ C	2-Amino-9 <i>H</i> -pyrido[2,3- <i>b</i> ]indole
BaP	Benzo[a]pyrene
BHA	Butylated hydroxyanisole
C	Carbon
C-C	Carbon-Carbon
CE	Crude extract
DEG	Diethylene glycol
dG	Deoxyguanosine
DMIP	2-Amin-1, 6-dimethylimidazo[4,5- <i>b</i> ]pyridine
DNA	Deoxyribo nucleic acid
DPPH	1,1-diphenyl-2-picrylhydrazyl
EGCG	Epigallocatechin gallate
ESI	Electrospray ionization
ESI	Electro spray ionization
FID	Flame ionization detector
FRAP	Ferric reducing antioxidant power assay
g	Gram
GAE	Gallic acid equivalent
GC-MS	Gas chromatography mass spectrometry
Glu-P-1	2-Amino-6-methyl-dipyrido[1,2- <i>a</i> :3',2'- <i>d</i> ]imidazole
Glu-P-2	2-Amino-dipyrido[1,2- <i>a</i> :3',2'- <i>d</i> ]imidazole

h	Hour
H	Hydrogen
HCAs	Heterocyclic amines
HCL	Hydrochloric acid
HPLC	High-performance liquid chromatography
HSD	Tukey's honestly significant different
I.D.	Internal Diameter
IAAs	Imidazo-azaarenes
IARC	The International Agency for Research on Cancer
IQ	2-amino-3-methylimidazo[4,5-f] quinolone
IQ	2-Amino-3-methylimidazo[4,5-f]quinolone
IQx	2-Amino-3-methylimidazo[4,5-f]quinoxaline
kPa	Kilopascal
LC-MS	Liquid chromatography-mass spectrometry
LC-MS/MS	Liquid chromatography- mass spectrometry/mass spectrometry
LOD	Limit of detection
LOQ	limit of quantification
M	Molar
MeAαC	2-Amino-3-methyl-9 <i>H</i> -pyrido[2,3- <i>b</i> ]indole
MeIQ	2-Amino-3,4-dimethylimidazo[4,5-f]quinolone
MeIQx	2-Amino-3,8-dimethylimidazo[4,5-f]quinoxaline
mg	Milligram
min	Minute
ml	Milliliter

mM	Millimolar
MRM	Multiple reaction monitoring
MS <sup>2</sup>	mass spectrometry/mass spectrometry
MVDA	Multivariate data analysis
N <sub>2</sub>	Nitrogen
NaOH	Sodium hydroxide
ND	Not detected
ng	Nanogram
nm	Nanometer
OH <sup>-</sup>	Hydroxide
OPLS	Orthogonal projections to latent structures
PDA	Photodiode array detector
Phe-P-1	2-Amino-5-phenylpyridine
PhIP	2-amino-1-methyl-6-phenylimidazo[4,5-b]pyridine
ppb	Parts-per-billion
Q-TOF	Quadrupole time-of-flight
Quercetin equivalents	QE
TCA	Trichloroacetic acid
TFC	Total flavonoid compounds
TMIP	2-Amino-1,5,6-trimethylimidazo[4,5- <i>b</i> ]pyridine
TPC	Total phenolic content
Trp-P-1	3-Amino-1,4-dimethyl-5 <i>H</i> -pyrido[4,3- <i>b</i> ]indole
Trp-P-2	3-Amino-1-methyl-5 <i>H</i> -pyrido[4,3- <i>b</i> ]indole
UHPLC	ultra-high performance liquid chromatography



UV	Ultraviolet
v/v	Volume/volume
w/v	Weight/volume
w/w	Weight/ Weight
$\alpha$	Alfa
$\beta$	Beta
$\gamma$	Gama
$\delta$	Sigma
$\mu\text{g}$	Microgram
%	Percentage
$^{\circ}\text{C}$	Degree celcius
$\mu\text{g}$	Microgram
$\mu\text{l}$	Microliter
$\mu\text{m}$	Micrometer
$\mu\text{mol}$	Micromole
4,7,8-TriMeIQx	2-Amino-3,4,7,8-tetramethyl-3H-imidazo[4,5-f]quinoxaline
4,8-DiMeIQx	2-Amino-3,4,8-trimethylimidazo[4,5-f]quinoxaline
4-CH <sub>2</sub> OH-8-MeIQx	2-Amino-4-hydroxymethyl-3,8-dimethylimidazo[4,5-f]quinoxaline
4-OH-PhIP	2-Amino-1-methyl-6-(4-hydroxyphenyl)imidazo[4,5-b]pyridine
7,8-DiMeIQx	2-Amino-3,7,8-trimethylimidazo[4,5-f]quinoxaline
7,9-DiMeIgQx	2-Amino-1,7,9-trimethylimidazo[4,5-g]quinoxaline

## CHAPTER 1

### INTRODUCTION

#### 1.1 Background of study

Heterocyclic amines (HCAs) are mutagenic and/or carcinogenic compounds form during thermal processing of meat and fish products at the temperatures over 150 °C. Based on the temperature of formation (Gibis & Weiss, 2012) they are classified into two groups, polar or thermic HCAs and nonpolar or pyrolytic HCAs. Polar or thermic HCAs which are known as amino imidazo-azaarenes, generate at temperatures above 150 °C through Maillard reaction between reducing sugars, especially glucose, and different free amino acids and creatine or creatinine. Non-polar or pyrolytic HCAs which are known as amino-carbolines form by pyrolysis of free amino acids at temperatures above 300 °C (Kizil, Oz, & Besler, 2011).

The International Agency for Research on Cancer (IARC) classified HCAs as a probable human carcinogens (2-amino-3-methylimidazo[4,5-f] quinoline (IQ) (class 2A) and possible human carcinogens (class 2B) (2-amino-1-methyl-6-phenylimidazo[4,5-b]pyridine (PhIP), (2-amino-3,4-dimethyl-imidazo[4,5-f]quinoline (MeIQ), and 2-amino-3,8-dimethyl-imidazo[4,5-f]quinoxaline (MeIQx) (Gibis, Kruwinnus, & Weiss, 2015).

Several epidemiological studies on cancer provide strong evidences which show an association between high consumption of heat processed meat and high risk of cancer in breast, prostate, lung, stomach (Alaejos, Pino, & Afonso, 2008) and colon (Butler et al., 2008). Moreover, many of HCAs have been shown to be mutagenic in Ames/*salmonella* test. It is demonstrated that some HCAs are 100 fold more mutagenic than aflatoxin B1 and over 2,000-fold more than benzo[a]pyrene (BaP) in *in-vitro* Ames test (Kizil et al., 2011).

In the past years, several studies were performed to develop some strategies which could beneficially reduce or inhibit HCAs formation in real meat matrix or chemical model systems. Microwave pretreatment prior to deep-frying and grilling of chicken and beef showed significant reduction of HCAs concentration (Jinap et al., 2013). It has been reported that the formation of HCAs can be reduced by using natural or synthetic antioxidants in chemical model or real meat systems. It has been reported that synthetic or natural antioxidants such as butylated hydroxyanisole (BHA),  $\alpha$ -tocopherol, eugenol (Moon & Shin, 2013), epigallocatechin gallate (EGCG), naringenin and quercetin-3-O-glucoside (Cheng, Chen, & Wang, 2007) fresh garlic and anion (Gibis, 2007), black paper powder (Oz & Kaya, 2011), virgin olive oil (Persson, Graziani, Ferracane, Fogliano, & Skog, 2003) and extracts of hibiscus (Gibis & Weiss, 2010), green tea (Quelhas et al., 2010), rosemary and grape seed (Ahn & Grün, 2005) reduce HCAs.

PhIP (2-Amino-1-methyl -6- phenylimidazo[4,5 - b]pyridine) is the most abundant HCAs occurred in high concentration in meat product which is cooked at high temperatures. Carcinogen and mutagen PhIP is originated from the condensation of creatinine with Millard reaction product of glucose and phenylalanine (Puangsombat, Gadgil, Houser, Hunt, & Smith, 2012; Yu, Chen, & Yu, 2016). *Salmonella* mutagenicity test (Ames test) (Apostolides, Balentine, Harbowy, & Weisburger, 1996; Singla & Kaur, 2003; Zhang et al., 2011) has demonstrated that PhIP is mutagenic compound for *S. Typhimurium* strain TA98 and TA 100 and several other studies have shown that PhIP induced tumors in prostate (Cross et al., 2005), colon (Nicken, Schröder, von Keutz, Breves, & Steinberg, 2013) and colorectal tissue (Rohrmann, Hermann, & Linseisen, 2009).

Therefore, inhibition or mitigation of PhIP formation has been the goal of numerous research projects. Several studies have shown that the application of plant extracts rich in phenolic compounds in real meat system or chemical model system could suppress the PhIP formation (Ahn & Grün, 2005; Cheng et al., 2009; Moon & Shin, 2013). Conversely, Cheng et al. (2007) indicated that some polyphenols such as rutin, chlorogenic acid, hesperidin, carnosic acid and rosmarinic acid increase PhIP concentration in model system, therefore, phytochemical screening of plant extract is a good way to achieve information about the constituents which are key inhibitory factors in PhIP formation.

## **1.2 Importance of study**

Since 1939 when Widmark, the Swedish chemist observed that extracts of heat processed horse meat induced carcinogenic tumor on the mice skin, many studies on the evaluation of mutagenicity and carcinogenicity of heat processed (fried, grilled, barbequed) meat have been conducted. They can be generally categorized into three classes: (i) those studying the mechanism of HCAs formation in real meat or chemical model systems and possible mechanism of their mutagenicity in *S. Typhimurium* (Ames test) and carcinogenicity in rodents, (ii) those isolating, characterizing and quantifying HCAs from foods and (iii) those investigating the reduction of HCAs using different ways. To date, many studies have been conducted to mitigate HCAs generation in food by different strategies such as reducing cooking temperature and time (Oz, Kaban, & Kaya, 2010; Puangsombat et al., 2012), microwave pretreatment (Jinap et al., 2013; Felton, Fultz, Dolbeare, & Knize, 1994) and using synthetic or natural antioxidants (Gibis, 2007; Gibis & Weiss, 2010; Damašius et al., 2011). It has been reported that the concentration of HCAs in meat products can be reduced by adding spices and herbs as marinade ingredient prior to heat processing. There are many publications which used single synthetic antioxidants or single herbs, spices or fruit extracts for the reduction of HCAs; however, no publication has been found on evaluation of using mixture of herbs and spices.

According to Food Consumption Statistics of Malaysia (2010), consumption of beef and chicken among Malaysian has increased considerably. The intakes of beef and chicken for Malaysian were estimated to be 104.16 g/day (Jing, 2008). This level of meat consumption assures high probability of Malaysian exposure to HCAs in the harmful level that leads to cancer. Malaysian regularly employ many high temperature cooking methods such as grilling, barbequing, deep frying and roasting to prepare meat. Chicken and beef satay are good examples of popular foods among Malaysians which get pre prepared at high temperatures.

The latest study on dietary exposure to HCAs in cooked meat and fish among Malaysians was done by Jahurul et al. (2010). The results showed that the dietary intake of HCAs in foods consumed by people in Selangor, Malaysia was 553.7 ng/capita/day and the intake of PhIP was the highest (30.6 ng/g), followed by MeIQx and MeIQ. The results reveal that grilled meat and fish products were the major contributors to the exposure of HCAs. Several studies (Ahn & Grün, 2005; Gibis & Weiss, 2012; Oz & Kaya, 2011; Quelhas et al., 2010; Ruan et al., 2014; Sabally, Sleno, Jauffrit, Iskandar, & Kubow, 2016; Zöchling, Murkovic, & Pfannhauser, 2002) showed that marinating meat by some spices and herbs before grilling can reduce HCAs formation.

The reduction of HCAs using local spices and herbs and their combination have not been investigated. Hence, this study is aimed to find associations between using combination of local spices and herbs on reduction of HCAs in grilled beef.

### **1.3 Objectives**

The specific objectives of this study are as follow:

1. To determine the antioxidant properties and bioactive compounds of selected local spices and herbs.
2. To optimize the best concentration of combination of local spices and herbs and evaluate the effectiveness for the reduction of HCAs in grilled meat.
3. To identify the compounds in mixture extracts of optimized spices and herbs which responsible for the reduction of PhIP formation in model system.
4. To determine the reduction of toxicity and mutagenicity of the optimized marinated grilled beef

## REFERENCES

- Ahn, J., & Grün, I. U. (2005). Heterocyclic amines: 2. Inhibitory effects of natural extracts on the formation of polar and nonpolar heterocyclic amines in cooked beef. *Journal of Food Science*, 70(4), 263-268.
- Alaejos, M. S., Pino, V., & Afonso, A. M. (2008). Metabolism and toxicology of heterocyclic aromatic amines when consumed in diet: Influence of the genetic susceptibility to develop human cancer. A review. *Food Research International*, 41(4), 327-340.
- Alvarado, C., & McKee, S. (2007). Marination to improve functional properties and safety of poultry meat. *The Journal of Applied Poultry Research*, 16(1), 113-120.
- Álvarez-Casas, M., García-Jares, C., Llompарт, M., & Lores, M. (2014). Effect of experimental parameters in the pressurized solvent extraction of polyphenolic compounds from white grape marc. *Food Chemistry*, 157, 524-532.
- Amanuma, K., Tone, S., Saito, H., Shigeoka, T., & Aoki, Y. (2002). Mutational spectra of benzo [a] pyrene and MeIQx in rpsL transgenic zebrafish embryos. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 513(1), 83-92.
- Ames, B. N., McCann, J., & Yamasaki, E. (1975). Methods for detecting carcinogens and mutagens with the Salmonella/mammalian-microsome mutagenicity test. *Mutation Research/Environmental Mutagenesis and Related Subjects*, 31(6), 347-363.
- Anslyn, E. V., & Dougherty, D. A. (2006). *Modern Physical Organic Chemistry*: University Science Books.
- Apostolides, Z., Balentine, D. A., Harbowy, M. E., & Weisburger, J. H. (1996). Inhibition of 2-amino-1-methyl-6-phenylimidazo [4, 5-b] pyridine (PhIP) mutagenicity by black and green tea extracts and polyphenols. *Mutation Research/Environmental Mutagenesis and Related Subjects*, 359(3), 159-163.
- Arimoto-Kobayashi, S., Ishida, R., Nakai, Y., Idei, C., Takata, J., Takahashi, E., Konuma, T. (2006). Inhibitory effects of beer on mutation in the Ames test and DNA adduct formation in mouse organs induced by 2-Amino-1-methyl-6-phenylimidazo [4, 5-b] pyridine (PhIP). *Biological and Pharmaceutical Bulletin*, 29(1), 67-70.
- Arvidsson, P., Boekel, M., Skog, K., & Jägerstad, M. (1997). Kinetics of formation of polar heterocyclic amines in a meat model system. *Journal of Food Science*, 62(5), 911-916.



- Badawy, A., Morgan, C., & Turner, J. (2008). Application of the Phenomenex EZ: faast™ amino acid analysis kit for rapid gas-chromatographic determination of concentrations of plasma tryptophan and its. *Amino Acids*, 34:587–596.
- Bai, W., Zhang, Z., Tian, W., He, X., Ma, Y., Zhao, Y., & Chai, Z. (2010). Toxicity of zinc oxide nanoparticles to zebrafish embryo: a physicochemical study of toxicity mechanism. *Journal of Nanoparticle Research*, 12(5), 1645-1654.
- Barceló-Barrachina, E., Moyano, E., Galceran, M., Lliberia, J., Bagó, B., & Cortes, M. (2006). Ultra-performance liquid chromatography–tandem mass spectrometry for the analysis of heterocyclic amines in food. *Journal of Chromatography A*, 1125(2), 195-203.
- Bianchi, F., Careri, M., Corradini, C., Elviri, L., Mangia, A., & Zagnoni, I. (2005). Investigation of the separation of heterocyclic aromatic amines by reversed phase ion-pair liquid chromatography coupled with tandem mass spectrometry: The role of ion pair reagents on LC–MS/MS sensitivity. *Journal of Chromatography B*, 825(2), 193-200.
- Borgen, E., Solyakov, A., & Skog, K. (2001). Effects of precursor composition and water on the formation of heterocyclic amines in meat model systems. *Food Chemistry*, 74(1), 11-19.
- Butler, L. M., Millikan, R. C., Sinha, R., Keku, T. O., Winkel, S., Harlan, B., Sandler, R. S. (2008). Modification by N-acetyltransferase 1 genotype on the association between dietary heterocyclic amines and colon cancer in a multiethnic study. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 638(1), 162-174.
- Carpenter, C. (2010). Determination of fat content. In S. Nielson (Ed.), *Food analysis laboratory manual*, (pp. 29-37). New York. Springer
- Casal, S., Mendes, E., Fernandes, J., Oliveira, M., & Ferreira, M. (2004). Analysis of heterocyclic aromatic amines in foods by gas chromatography–mass spectrometry as their tert.-butyldimethylsilyl derivatives. *Journal of Chromatography A*, 1040(1), 105-114.
- Cheng, K. W., Chen, F., & Wang, M. (2007). Inhibitory activities of dietary phenolic compounds on heterocyclic amine formation in both chemical model system and beef patties. *Molecular Nutrition & Food Research*, 51(8), 969-976.
- Cheng, K. W., Wong, C. C., Chao, J., Lo, C., Chen, F., Chu, I. K., Wang, M. (2009). Inhibition of mutagenic PhIP formation by epigallocatechin gallate via scavenging of phenylacetaldehyde. *Molecular Nutrition & Food Research*, 53(6), 716-725.
- Cheng, K. W., Wu, Q., Zheng, Z. P., Peng, X., Simon, J. E., Chen, F., & Wang, M. (2007). Inhibitory effect of fruit extracts on the formation of heterocyclic amines. *Journal of Agricultural and Food Chemistry*, 55(25), 10359-10365.

- Cheung, C., Ma, X., Krausz, K. W., Kimura, S., Feigenbaum, L., Dalton, T. P., Gonzalez, F. J. (2005). Differential metabolism of 2-amino-1-methyl-6-phenylimidazo [4, 5-b] pyridine (PhIP) in mice humanized for CYP1A1 and CYP1A2. *Chemical Research In Toxicology*, 18(9), 1471-1478.
- Chou, C. M., Chen, Y. C., Su, S., Chen, G. D., Huang, K. Y., Lien, H. W., Cheng, C. H. (2015). Activation of MEK2 is sufficient to induce skin papilloma formation in transgenic zebrafish. *Journal of Biomedical Science*, 22(1), 1.
- Cross, A. J., Peters, U., Kirsh, V. A., Andriole, G. L., Reding, D., Hayes, R. B., & Sinha, R. (2005). A prospective study of meat and meat mutagens and prostate cancer risk. *Cancer Research*, 65(24), 11779-11784.
- Cuyckens, F., & Claeys, M. (2004). Mass spectrometry in the structural analysis of flavonoids. *Journal of Mass Spectrometry*, 39(1), 1-15.
- Damašius, J., Venskutonis, P., Ferracane, R., & Fogliano, V. (2011). Assessment of the influence of some spice extracts on the formation of heterocyclic amines in meat. *Food Chemistry*, 126(1), 149-156.
- Dandekar, D. V., & Gaikar, V. (2002). Microwave assisted extraction of curcuminoids from *Curcuma longa*. *Separation Science and Technology*, 37(11), 2669-2690.
- Del Campo, G., Gallego, B., Berregi, I., & Casado, J. A. (1998). Creatinine, creatine and protein in cooked meat products. *Food Chemistry*, 63(2), 187-190.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S., & Ju, Y.-H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatica*. *Journal of Food and Drug Analysis*, 22(3), 296-302.
- Dumont, J., Jossé, R., Lambert, C., Anthérieu, S., Le Hegarat, L., Aninat, C., Guillouzo, A. (2010). Differential toxicity of heterocyclic aromatic amines and their mixture in metabolically competent HepaRG cells. *Toxicology and Applied Pharmacology*, 245(2), 256-263.
- Dwivedi, S., Vasavada, M. N., & Cornforth, D. (2006). Evaluation of Antioxidant Effects and Sensory Attributes of Chinese 5- Spice Ingredients in Cooked Ground Beef. *Journal of food science*, 71(1), C12-C17.
- Ecobichon, D. J. (1997). *The basis of toxicity testing*, (pp.160-163). New York. CRC press
- Eriksson, L., Byrne, T., Johansson, E., Trygg, J., & Vikström, C. (2013). *Multi-and megavariable data analysis basic principles and applications*, (pp. 110-115). Sweden. Umetrics Academy.

- Farhadian, A., Jinap, S., Faridah, A., & Zaidul, I. S. M. (2012). Effects of marinating on the formation of polycyclic aromatic hydrocarbons (benzo[a]pyrene, benzo[b]fluoranthene and fluoranthene) in grilled beef meat. *Food Control*, 28(2), 420–425.
- Felton, J. S., Fultz, E., Dolbeare, F.A., & Knize, M.G. (1994). Effect of microwave pretreatment on heterocyclic aromatic amine mutagens/carcinogens in fried beef patties. *Food and Chemical Toxicology*, 32(10), 897–903.
- Figueirinha, A., Paranhos, A., Pérez-Alonso, J. J., Santos-Buelga, C., & Batista, M. T. (2008). Cymbopogon citratus leaves: Characterization of flavonoids by HPLC–PDA–ESI/MS/MS and an approach to their potential as a source of bioactive polyphenols. *Food Chemistry*, 110(3), 718–728.
- Gibis, M. (2007). Effect of oil marinades with garlic, onion, and lemon juice on the formation of heterocyclic aromatic amines in fried beef patties. *Journal of Agricultural and Food Chemistry*, 55(25), 10240–10247.
- Gibis, M. (2016). Heterocyclic Aromatic Amines in Cooked Meat Products: Causes, Formation, Occurrence, and Risk Assessment. *Comprehensive Reviews in Food Science and Food Safety*, 15(2), 269–302.
- Gibis, M., Kruwinnus, M., & Weiss, J. (2015). Impact of different pan-frying conditions on the formation of heterocyclic aromatic amines and sensory quality in fried bacon. *Food Chemistry*, 168, 383–389.
- Gibis, M., & Weiss, J. (2010). Inhibitory effect of marinades with hibiscus extract on formation of heterocyclic aromatic amines and sensory quality of fried beef patties. *Meat Science*, 85(4), 735–742.
- Gibis, M., & Weiss, J. (2012). Antioxidant capacity and inhibitory effect of grape seed and rosemary extract in marinades on the formation of heterocyclic amines in fried beef patties. *Food Chemistry*, 134(2), 766–774.
- Gironi, F., & Piemonte, V. (2011). Temperature and solvent effects on polyphenol extraction process from chestnut tree wood. *Chemical Engineering Research and Design*, 89(7), 857–862.
- Hagerman, A. E. (1988). Extraction of tannin from fresh and preserved leaves. *Journal of Chemical Ecology*, 14(2), 453–461.
- Hasnol, N., Jinap, S., & Sanny, M. (2014). Effect of different types of sugars in a marinating formulation on the formation of heterocyclic amines in grilled chicken. *Food Chemistry*, 145, 514–521.
- Heim, K. E., Tagliaferro, A. R., & Bobilya, D. J. (2002). Flavonoid antioxidants: chemistry, metabolism and structure-activity relationships. *The Journal of Nutritional Biochemistry*, 13(10), 572–584.



- Herraiz, T. (2004). Relative exposure to  $\beta$ -carbolines norharman and harman from foods and tobacco smoke. *Food Additives and Contaminants*, 21(11), 1041-1050.
- Herraiz, T., Guillén, H., & Arán, V. J. (2008). Oxidative metabolism of the bioactive and naturally occurring  $\beta$ -carboline alkaloids, norharman and harman, by human cytochrome P450 enzymes. *Chemical Research in Toxicology*, 21(11), 2172-2180.
- Hwang, H. I., Hartman, T. G., Rosen, R. T., Lech, J., & Ho, C. T. (1994). Formation of Pyrazines from the Maillard Reaction of Glucose and Lysine- $\alpha$ -amine-15N. *Journal of Agricultural and Food Chemistry*, 42(4), 1000-1004.
- Jägerstad, M., Skog, K., Grivas, S., & Olsson, K. (1991). Formation of heterocyclic amines using model systems. *Mutation Research/Genetic Toxicology*, 259(3-4), 219-233.
- Jahurul, M., Jinap, S., Ang, S., Abdul-Hamid, A., Hajeb, P., Lioe, H., & Zaidul, I. (2010). Dietary exposure to heterocyclic amines in high-temperature cooked meat and fish in Malaysia. *Food Additives and Contaminants*, 27(8), 1060-1071.
- Javadi, N., Abas, F., Hamid, A. A., Simoh, S., Shaari, K., Ismail, I. S., Khatib, A. (2014). GC- MS- Based Metabolite Profiling of *Cosmos caudatus* Leaves Possessing Alpha- Glucosidase Inhibitory Activity. *Journal of Food Science*, 79(6), C1130-C1136.
- Jinap, S., Iqbal, S. Z., & Selvam, R. M. (2015). Effect of selected local spices marinades on the reduction of heterocyclic amines in grilled beef (satay). *LWT- Food Science and Technology*, 63(2), 919-926.
- Jinap, S., Mohd-Mokhtar, M., Farhadian, A., Hasnol, N., Jaafar, S., & Hajeb, P. (2013). Effects of varying degrees of doneness on the formation of Heterocyclic Aromatic Amines in chicken and beef satay. *Meat science*, 94(2), 202-207.
- Jing, A. (2008). Exposure assessment on consumption of high temperature cooked food among Malaysians, Universiti Putra Malaysia. *Bachelor Thesis*.
- Kallithraka, S., Garcia- Viguera, C., Bridle, P., & Bakker, J. (1995). Survey of solvents for the extraction of grape seed phenolics. *Phytochemical Analysis*, 6(5), 265-267.
- Kataoka, H., Nishioka, S., Kobayashi, M., Hanaoka, T., & Tsugane, S. (2002). Analysis of mutagenic heterocyclic amines in cooked food samples by gas chromatography with nitrogen-phosphorus detector. *Bulletin of Environmental Contamination and Toxicology*, 69(5), 0682-0689.

- Kato, T., Harashima, T., Moriya, N., Kikugawa, K., & Hiramoto, K. (1996). Formation of the mutagenic/carcinogenic imidazoquinoxaline-type heterocyclic amines through the unstable free radical Maillard intermediates and its inhibition by phenolic antioxidants. *Carcinogenesis*, 17(11), 2469-2476.
- Kim, N. M., Kim, J., Chung, H. Y., & Choi, J. S. (2000). Isolation of luteolin 7-O-rutinoside and esculetin with potential antioxidant activity from the aerial parts of *Artemisia montana*. *Archives of Pharmacal Research*, 23(3), 237-239.
- Kizil, M., Oz, F., & Besler, H. T. (2011). A review on the formation of carcinogenic/mutagenic heterocyclic aromatic amines. *Journal of Food Processing & Technology*, 2011.
- Kolpe, U., Ramaswamy, V., Rao, B. S., & Nagabhushan, M. (2002). *Turmeric and curcumin prevents the formation of mutagenic Maillard reaction products*. Paper presented at the International Congress Series.
- Komoltri, P., & Pakdeechanuan, P. (2012). Effects of marinating ingredients on physicochemical, microstructural and sensory properties of golek chicken. *International Food Research Journal*, 19(4), 1449-1455.
- Kondjoyan, A., Chevolleau, S., Portanguen, S., Molina, J., Ikonic, P., Clerjon, S., & Debrauwer, L. (2016). Relation between crust development and heterocyclic aromatic amine formation when air-roasting a meat cylinder. *Food Chemistry*, 213, 641-646.
- Langouët, S., Paehler, A., Welti, D. H., Kerriguy, N., Guillouzo, A., & Turesky, R. J. (2002). Differential metabolism of 2-amino-1-methyl-6-phenylimidazo [4, 5-b] pyridine in rat and human hepatocytes. *Carcinogenesis*, 23(1), 115-122.
- Li, Y., Huang, W., Huang, S., Du, J., & Huang, C. (2012). Screening of anti-cancer agent using zebrafish: comparison with the MTT assay. *Biochemical and Biophysical Research communications*, 422(1), 85-90.
- Liu, D., Schwimer, J., Liu, Z., Woltering, E. A., & Greenway, F. L. (2008). Antiangiogenic effect of curcumin in pure versus in extract forms. *Pharmaceutical Biology*, 46(10-11), 677-682.
- Lu, Y., & Yeap Foo, L. (2001). Antioxidant activities of polyphenols from sage (*Salvia officinalis*). *Food Chemistry*, 75(11), 197-202.
- Lv, Z., Dong, J., & Zhang, B. (2012). Rapid identification and detection of flavonoids compounds from bamboo leaves by LC-(ESI)-IT-TOF/MS. *BioResources*, 7(2), 1405-1418.
- Magagnotti, C., Pastorelli, R., Pozzi, S., Andreoni, B., Fanelli, R., & Airoidi, L. (2003). Genetic polymorphisms and modulation of 2- amino- 1- methyl- 6- phenylimidazo [4, 5- b] pyridine (PhIP)- DNA adducts in human lymphocytes. *International Journal of Cancer*, 107(6), 878-884.

- Maheshwari, R. K., Singh, A. K., Gaddipati, J., & Srimal, R. C. (2006). Multiple biological activities of curcumin: a short review. *Life Sciences*, 78(18), 2081-2087.
- Majer, B. J., Mersch-Sundermann, V., Darroudi, F., Laky, B., de Wit, K., & Knasmüller, S. (2004). Genotoxic effects of dietary and lifestyle related carcinogens in human derived hepatoma (HepG2, Hep3B) cells. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 551(1), 153-166.
- Maron, D. M., & Ames, B. N. (1983). Revised methods for the Salmonella mutagenicity test. *Mutation Research/Environmental Mutagenesis and Related Subjects*, 113(3-4), 173-215.
- Miller, L. C., & Tainter, M. (1944). Estimation of the ED50 and its error by means of logarithmic-probit graph paper. *Experimental Biology and Medicine*, 57(2), 261-264.
- Moon, S. E., & Shin, H. S. (2013). Inhibition of mutagenic 2-amino-1-methyl-6-phenylimidazo [4, 5-b] pyridine (PhIP) formation using various food ingredients in a model systems. *Food Science and Biotechnology*, 22(2), 323-329.
- Murkovic, M. (2004). Formation of heterocyclic aromatic amines in model systems. *Journal of Chromatography B*, 802(1), 3-10.
- Murkovic, M., Weber, H.-J., Geiszler, S., Fröhlich, K., & Pfannhauser, W. (1999). Formation of the food associated carcinogen 2-amino-1-methyl-6-phenylimidazo [4, 5-b] pyridine (PhIP) in model systems. *Food Chemistry*, 65(2), 233-237.
- Nauwelaers, G., Bessette, E. E., Gu, D., Tang, Y., Rageul, J., Fessard, V., Turesky, R. J. (2011). DNA adduct formation of 4-aminobiphenyl and heterocyclic aromatic amines in human hepatocytes. *Chemical Research in Toxicology*, 24(6), 913-925.
- Navarro, A., Muñoz, S. E., Lantieri, M. J., del Pilar Diaz, M., Cristaldo, P. E., de Fabro, S. P., & Eynard, A. R. (2004). Meat cooking habits and risk of colorectal cancer in Cordoba, Argentina. *Nutrition*, 20(10), 873-877.
- Nerurkar, P. V., Marchand, L. L., & Cooney, R. V. (1999). Effects of marinating with Asian marinades or western barbecue sauce on PhIP and MeIQx formation in barbecued beef. *Nutrition and Cancer*, 34(2), 147-152.
- Nicken, P., Schröder, B., von Keutz, A., Breves, G., & Steinberg, P. (2013). The colon carcinogen 2-amino-1-methyl-6-phenylimidazo [4, 5-b] pyridine (PhIP) is actively secreted in the distal colon of the rat: an integrated view on the role of PhIP transport and metabolism in PhIP-induced colon carcinogenesis. *Archives of Toxicology*, 87(5), 895-904.

- Njiwa, J., Suter, M.-F., & Eggen, R. (2010). Zebrafish Embryo Toxicity Assay, Combining Molecular and Integrative Endpoints at Various Developmental Stages *Handbook of Hydrocarbon and Lipid Microbiology* (pp. 4481-4489). Springer.
- Nowell, S., Coles, B., Sinha, R., MacLeod, S., Ratnasinghe, D. L., Stotts, C., Lang, N. P. (2002). Analysis of total meat intake and exposure to individual heterocyclic amines in a case-control study of colorectal cancer: contribution of metabolic variation to risk. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 506, 175-185.
- O'Sullivan, A., O'Callaghan, Y., O'Grady, M., Hayes, M., Kerry, J., & O'Brien, N. (2013). The effect of solvents on the antioxidant activity in Caco-2 cells of Irish brown seaweed extracts prepared using accelerated solvent extraction (ASE®). *Journal of Functional Foods*, 5(2), 940-948.
- Oguri, A., Suda, M., Totsuka, Y., Sugimura, T., & Wakabayashi, K. (1998). Inhibitory effects of antioxidants on formation of heterocyclic amines. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 402(1), 237-245.
- Oz, F., Kaban, G., & Kaya, M. (2010). Effects of cooking methods and levels on formation of heterocyclic aromatic amines in chicken and fish with Oasis extraction method. *LWT-Food Science and Technology*, 43(9), 1345-1350.
- Oz, F., & Kaya, M. (2011). The inhibitory effect of black pepper on formation of heterocyclic aromatic amines in high-fat meatball. *Food Control*, 22(3), 596-600.
- Oz, F., & Kotan, G. (2016). Effects of different cooking methods and fat levels on the formation of heterocyclic aromatic amines in various fishes. *Food Control*, 67, 216-224.
- Persson, E., Graziani, G., Ferracane, R., Fogliano, V., & Skog, K. (2003). Influence of antioxidants in virgin olive oil on the formation of heterocyclic amines in fried beefburgers. *Food and Chemical Toxicology*, 41(11), 1587-1597.
- Phenomenex EZ:faast (easy fast) amino acid sample testing kit (2003) User guide, Phenomenex, 411 Madrid Avenue, Torrance, CA 90501-1430, USA (<http://www.phenomenex.com>)
- Polak, T., Andrenšek, S., Žlender, B., & Gašperlin, L. (2009). Effects of ageing and low internal temperature of grilling on the formation of heterocyclic amines in beef Longissimus dorsi muscle. *LWT-Food Science and Technology*, 42(1), 256-264.



- Polson, C., Sarkar, P., Incledon, B., Raguvanan, V., & Grant, R. (2003). Optimization of protein precipitation based upon effectiveness of protein removal and ionization effect in liquid chromatography–tandem mass spectrometry. *Journal of Chromatography B*, 785(2), 263-275.
- Puangsombat, K., Gadgil, P., Houser, T. A., Hunt, M. C., & Smith, J. S. (2012). Occurrence of heterocyclic amines in cooked meat products. *Meat science*, 90(3), 739-746.
- Quelhas, I., Petisca, C., Viegas, O., Melo, A., Pinho, O., & Ferreira, I. (2010). Effect of green tea marinades on the formation of heterocyclic aromatic amines and sensory quality of pan-fried beef. *Food Chemistry*, 122(1), 98-104.
- Randel, G., Balzer, M., Grupe, S., Drusch, S., Kaina, B., Platt, K. L., & Schwarz, K. (2007). Degradation of heterocyclic aromatic amines in oil under storage and frying conditions and reduction of their mutagenic potential. *Food and Chemical Toxicology*, 45(11), 2245-2253.
- Rohrmann, S., Hermann, S., & Linseisen, J. (2009). Heterocyclic aromatic amine intake increases colorectal adenoma risk: findings from a prospective European cohort study. *The American Journal of Clinical Nutrition*, 89(5), 1418-1424.
- Roriz, C. L., Barros, L., Carvalho, A. M., Santos-Buelga, C., & Ferreira, I. C. (2015). Scientific validation of synergistic antioxidant effects in commercialised mixtures of *Cymbopogon citratus* and *Pterospartum tridentatum* or *Gomphrena globosa* for infusions preparation. *Food Chemistry*, 185, 16-24.
- Ruan, E., Juárez, M., Thacker, R., Yang, X., Dugan, M., & Aalhus, J. (2014). Dietary vitamin E effects on the formation of heterocyclic amines in grilled lean beef. *Meat Science*, 96(2), 849-853.
- Sabally, K., Sleno, L., Jauffrit, J.-A., Iskandar, M. M., & Kubow, S. (2016). Inhibitory effects of apple peel polyphenol extract on the formation of heterocyclic amines in pan fried beef patties. *Meat Science*, 117, 57-62.
- Shaikh, J., Ankola, D., Beniwal, V., Singh, D., & Kumar, M. R. (2009). Nanoparticle encapsulation improves oral bioavailability of curcumin by at least 9-fold when compared to curcumin administered with piperine as absorption enhancer. *European Journal of Pharmaceutical Sciences*, 37(3), 223-230.
- Shen, L., & Ji, H.-F. (2012). The pharmacology of curcumin: is it the degradation products? *Trends in Molecular Medicine*, 18(3), 138-144.
- Shiau, R. J., Shih, P. C., & Wen, Y. D. (2011). Effect of silymarin on curcumin-induced mortality in zebrafish (*Danio rerio*) embryos and larvae.
- Shin, H., & Ustunol, Z. (2004). Influence of Honey- containing Marinades on Heterocyclic Aromatic Amine Formation and Overall Mutagenicity in Fried Beef Steak and Chicken Breast. *Journal of Food Science*, 69(3), 147-153.

- Shon, M.-Y., Kim, T.-H., & Sung, N.-J. (2003). Antioxidants and free radical scavenging activity of *Phellinus baumii* (*Phellinus* of *Hymenochaetaceae*) extracts. *Food Chemistry*, 82(4), 593-597.
- Singh, A. P., Wilson, T., Luthria, D., Freeman, M. R., Scott, R. M., Bilenker, D., Vorsa, N. (2011). LC-MS-MS characterisation of curry leaf flavonols and antioxidant activity. *Food Chemistry*, 127(1), 80-85.
- Singh, R., Arlt, V. M., Henderson, C. J., Phillips, D. H., Farmer, P. B., & Da Costa, G. G. (2010). Detection and quantitation of N-(deoxyguanosin-8-yl)-2-amino-1-methyl-6-phenylimidazo [4, 5-b] pyridine adducts in DNA using online column-switching liquid chromatography tandem mass spectrometry. *Journal of Chromatography B*, 878(23), 2155-2162.
- Singla, A., & Kaur, I. (2003). Inhibitory effect of dibenzoylmethane on mutagenicity of food-derived heterocyclic amine mutagens. *Phytomedicine*, 10(6), 575-582.
- Singleton, V., & Rossi, J. A. (1965). Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American journal of Enology and Viticulture*, 16(3), 144-158.
- Skog, K., & Jägerstad, M. (1993). Incorporation of carbon atoms from glucose into the food mutagens MeIQx and 4, 8-DiMeIQx using <sup>14</sup>C-labelled glucose in a model system. *Carcinogenesis*, 14(10), 2027-2031.
- Skog, K., Johansson, M., & Jägerstad, M. (1998). Carcinogenic heterocyclic amines in model systems and cooked foods: a review on formation, occurrence and intake. *Food and Chemical Toxicology*, 36(9), 879-896.
- Skog, K., Solyakov, A., & Jägerstad, M. (2000). Effects of heating conditions and additives on the formation of heterocyclic amines with reference to amino-carbolines in a meat juice model system. *Food Chemistry*, 68(3), 299-308.
- Smith, J. S., Ameri, F., & Gadgil, P. (2008). Effect of marinades on the formation of heterocyclic amines in grilled beef steaks. *Journal of Food Science*, 73(6), 100-105.
- Smith, K. E., Heringa, M. B., Uytewaal, M., & Mayer, P. (2013). The dosing determines mutagenicity of hydrophobic compounds in the Ames II assay with metabolic transformation: passive dosing versus solvent spiking. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 750(1), 12-18.
- Sugimura, T., Wakabayashi, K., Nakagama, H., & Nagao, M. (2004). Heterocyclic amines: Mutagens/carcinogens produced during cooking of meat and fish. *Cancer Science*, 95(4), 290-299.
- Suhaj, M. (2006). Spice antioxidants isolation and their antiradical activity: a review. *Journal of Food Composition and Analysis*, 19(6), 531-537.

- Sulaiman, S. F., Sajak, A. A. B., Ooi, K. L., & Seow, E. M. (2011). Effect of solvents in extracting polyphenols and antioxidants of selected raw vegetables. *Journal of Food Composition and Analysis*, 24(4), 506-515.
- Tang, D., Liu, J. J., Bock, C. H., Neslund- Dudas, C., Rundle, A., Savera, A. T., Rybicki, B. A. (2007). Racial differences in clinical and pathological associations with PhIP- DNA adducts in prostate. *International Journal of Cancer*, 121(6), 1319-1324.
- Tang, Y., Kassie, F., Qian, X., Ansha, B., & Turesky, R. J. (2013). DNA adduct formation of 2-amino-9H-pyrido [2, 3-b] indole and 2-amino-3, 4-dimethylimidazo [4, 5-f] quinoline in mouse liver and extrahepatic tissues during a subchronic feeding study. *Toxicological Sciences*, 133(2), 248-258.
- Tikkanen, L., Latva-Kala, K., & Heiniö, R.-L. (1996). Effect of commercial marinades on the mutagenic activity, sensory quality and amount of heterocyclic amines in chicken grilled under different conditions. *Food and Chemical Toxicology*, 34(8), 725-730.
- Tønnesen, H. H., Másson, M., & Loftsson, T. (2002). Studies of curcumin and curcuminoids. XXVII. Cyclodextrin complexation: solubility, chemical and photochemical stability. *International Journal of Pharmaceutics*, 244(1), 127-135.
- Turesky, R. J. (2007). Formation and biochemistry of carcinogenic heterocyclic aromatic amines in cooked meats. *Toxicology Letters*, 168(3), 219-227.
- Turesky, R. J. (2010). Heterocyclic Aromatic Amines: Potential Human Carcinogens. *Advances in Molecular Toxicology*, 4, 37-83.
- Turesky, R. J., Bendaly, J., Yasa, I., Doll, M. A., & Hein, D. W. (2009). The Impact of NAT2 Acetylator Genotype on Mutagenesis and DNA Adducts from 2-Amino-9 H-pyrido [2, 3-b] indole. *Chemical Research in Toxicology*, 22(4), 726-733.
- Turesky, R. J., & Vouros, P. (2004). Formation and analysis of heterocyclic aromatic amine-DNA adducts in vitro and in vivo. *Journal of Chromatography B*, 802(1), 155-166.
- Verma, S. C., & Jain, C. (2011). Solvent polarity based microwave-assisted extraction of curcuminoids from *Curcuma longa* L. rhizome and their quantitative determination by HPLC-PDA method. *International Journal of Pharmaceutical Research*, 3(4), 41-47.
- Vitaglione, P., & Fogliano, V. (2004). Use of antioxidants to minimize the human health risk associated to mutagenic/carcinogenic heterocyclic amines in food. *Journal of Chromatography B*, 802(1), 189-199.

- Viuda- Martos, M., Fernández- López, J., & Pérez- Álvarez, J. (2010). Pomegranate and its many functional components as related to human health: a review. *Comprehensive Reviews in Food Science and Food Safety*, 9(6), 635-654.
- Weisburger, J. H., Dolan, L., & Pittman, B. (1998). Inhibition of PhIP mutagenicity by caffeine, lycopene, daidzein, and genistein. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 416(1), 125-128.
- Wichitnithad, W., Jongaroonngamsang, N., Pummangura, S., & Rojsitthisak, P. (2009). A simple isocratic HPLC method for the simultaneous determination of curcuminoids in commercial turmeric extracts. *Phytochemical Analysis*, 20(4), 314-319.
- Widmark, E. (1939). Presence of cancer-producing substances in roasted food. *Nature*, 143(3632), 984.
- Wijekoon, M. J. O., Bhat, R., & Karim, A. A. (2011). Effect of extraction solvents on the phenolic compounds and antioxidant activities of bunga kantan (*Etlingera elatior* Jack.) inflorescence. *Journal of Food Composition and Analysis*, 24(4), 615-619.
- Winter, H. K., Ehrlich, V. A., Grusch, M., Lackner, A., Schulte-Hermann, R., Grasl-Kraupp, B., Knasmüller, S. (2008). Use of four new human-derived liver-cell lines for the detection of genotoxic compounds in the single-cell gel electrophoresis (SCGE) assay. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 657(2), 133-139.
- Wong, D., Cheng, K. W., & Wang, M. (2012). Inhibition of heterocyclic amine formation by water-soluble vitamins in Maillard reaction model systems and beef patties. *Food Chemistry*, 133(3), 760-766.
- Wu, J. Y., Lin, C. Y., Lin, T. W., Ken, C. F., & Wen, Y. D. (2007). Curcumin affects development of zebrafish embryo. *Biological and Pharmaceutical Bulletin*, 30(7), 1336-1339.
- Yu, D., Chen, M. S., & Yu, S. J. (2016). Effect of sugarcane molasses extract on the formation of 2-amino-1-methyl-6-phenylimidazo [4, 5-b] pyridine (PhIP) in a model system. *Food Chemistry*, 197, 924-929.
- Yusop, S. M., O'Sullivan, M. G., Kerry, J. F., & Kerry, J. P. (2010). Effect of marinating time and low pH on marinade performance and sensory acceptability of poultry meat. *Meat science*, 85(4), 657-663.
- Zamora, R., Alcón, E., & Hidalgo, F. J. (2013). Effect of amino acids on the formation of 2-amino-1-methyl-6-phenylimidazo [4,5-b] pyridine (PhIP) in creatinine/phenylalanine and creatinine/phenylalanine/4-oxo-2-nonenal reaction mixtures. *Food Chemistry*, 141(4), 4240-4245.



- Zamora, R., & Hidalgo, F. J. (2015). 2-Amino-1-methyl-6-phenylimidazo [4, 5-b] pyridine (PhIP) formation and fate: an example of the coordinate contribution of lipid oxidation and Maillard reaction to the production and elimination of processing-related food toxicants. *RSC Advances*, 5(13), 9709-9721.
- Zhang, X., Ishida, R., Yuhara, Y., Kamiya, T., Hatano, T., Okamoto, G., & Arimoto-Kobayashi, S. (2011). Anti-genotoxic activity of *Vitis coignetiae* Pulliat towards heterocyclic amines and isolation and identification of caftaric acid as an antimutagenic component from the juice. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 723(2), 182-189.
- Zheng, W., & Lee, S. A. (2009). Well-done meat intake, heterocyclic amine exposure, and cancer risk. *Nutrition and Cancer*, 61(4), 437-446.
- Zöchling, S., Murkovic, M., & Pfannhauser, W. (2002). Effects of industrially produced flavours with pro- and antioxidative properties on the formation of the heterocyclic amine PhIP in a model system. *Journal of Biochemical and Biophysical Methods*, 53(1), 37-44.