

UNIVERSITI PUTRA MALAYSIA

PREVELANCE AND PATHOGENICITY OF ROOT LESION NEMATODE, Pratylenchus spp. ON BANANA (Musa paradisiaca L.) IN PENINSULAR MALAYSIA

ADAMU SAIDU PAIKO

FP 2016 25



PREVELANCE AND PATHOGENICITY OF ROOT LESION NEMATODE, Pratylenchus spp. ON BANANA (Musa paradisiaca L.) IN PENINSULAR MALAYSIA

By

ADAMU SAIDU PAIKO

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

May 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

This work is dedicated to my beloved parents; late mother Mallama Jummai Ibrahim, my father Alhaji Adamu Saidu Jazu and my bestowed kids; Abdullah, AbdurRahman, Isah and young Mus'ab.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Master of Science

PREVELANCE AND PATHOGENICITY OF ROOT LESION NEMATODE, Pratylenchus spp. ON BANANA (Musa paradisiaca L.) IN PENINSULAR MALAYSIA

By

ADAMU SAIDU PAIKO

May 2016

Chairman Faculty : Associate Professor Kamaruzaman Bin Sijam, PhD : Agriculture

Root lesion nematode (*Pratylenchus* spp.) cause considerable damage to banana plant (Musa spp.) globally. Even though, it has been reported in Malaysia, it disease prevalence and severity of infection has not been attended to. The objective of the research therefore, was aimed at determining the disease prevalence of *Pratylenchus* spp. and the status of their damage on Musa spp. in Peninsular Malaysia. To study the prevalence and pathogenicity of root lesion nematode, Pratylenchus spp. on banana (Musa paradisiaca.) in Peninsular Malaysia, samples of soil and root were collected from banana fields nationwide in 2014. Of the 13 sampled banana fields, lesion nematodes were found in 10 fields (76% of the areas surveyed). Rating of root cortex occupied by reddish brown lesion was significant among all the states, with Johor having the highest disease severity of 36.3% followed by Selangor 20.7%, Perak 20.2% and Pahang 20.4 % respectively. The study found that all the sampled areas have exceeded threshold limit, which is 5%. Mean population density of fields having *Pratylenchus* spp. from 20 g roots and 200 g soil were significantly different, where population mean of Selangor, (root= 838 and soil= 897), (root= 18,050 and soil= 13,056) in Johor, (root= 2341 and soil= 461) Perak and (root= 11,315 and soil= 7,199) Pahang were recorded. Other plant-parasitic nematodes identified were Rotylenchulus spp., Meloidogyne spp., Helicotylenchus spp., Hoplolaimus spp. and Radophulus simili. Pathological reaction of P. coffeae against banana cultivar berangan (Musa paradisica) multiplications was observed after 12 weeks of growth. There were significant differences in vegetative growth within the various pathogens inoculation levels evaluated. Multiplication factors ranged between 1.6-4 in P. coffeae. Reduction in root length (-23.68 to -84.95), shoot length (-13.11 to -45.80), root weights (-21.30 to -99.85) and shoot weight (-9.30 to -61.62) lengths were recorded at ($P \le 0.05$) level of probability. Banana cultivar berangan showed high level of susceptibility through the activity of polyphenol oxidase and peroxidase-induced resistance at all days after inoculation with P. coffeae compared to the control, except at week 12 where it declined or non-significant with the control. Our observations from this study, revealed that P. coffeae is among the most damaging plant-parasitic nematode species associated with banana in Peninsular Malaysia replacing R. similis, which is seldom reported or localized to a particular area and in low densities, followed by *Meloidogyne* spp. That it is common on banana cultivar berangan in this country, and that its geographical distribution is not restricted. Although the morphological study of the female populations of *P. coffeae* from Peninsular Malaysia gave some little variation in morphology from the reference sources, we resolved that these variations are within the range of the previously described morphological variations in *P. coffeae* populations from other parts of the world, thus, confirming that the isolates were *P. coffeae*. In general, the glass house pathogenicity trial of *P. coffeae*, suppresses vegetative growth of the banana cultivar berangan significantly. Root lesion indexes showed higher disease severity at all inoculum levels evaluated. It can be concluded that *P. coffeae* is widely distributed and caused significant damage to banana crop in Peninsular Malaysia.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

KEJADIAN DAN PATOGENISITI NEMATODA LESI AKAR, Pratylenchus spp. PADA PISANG (Musa paradisiaca L.) DI SEMENANJUNG MALAYSIA

Oleh

ADAMU SAIDU PAIKO

Mei 2016

Pengerusi Fakulti : Profesor Madya Kamaruzaman Bin Sijam, PhD : Pertanian

Nematoda lesi akar (Pratylenchus spp.) menyebabkan kerosakan yang signifikan pada tanaman pisang (Musa spp.) di dunia. Walaupun ianya telah dilaporkan di Malaysia, namun kejadian dan keparahan jangkitan masih tidak dapat dihindari. Objektif kajian ini adalah untuk menentukan kejadian penyakit oleh *Pratylenchus* spp. dan status kerosakannya pada Musa spp. Bagi menentukan kejadian dan kepatogenan nematoda lesi akar, Pratylenchus spp. pada Musa spp. di Semenanjung Malaysia, sampel tanah dan akar telah diambil daripada kebun pisang di seluruh negara pada tahun 2014. Daripada 13 kebun pisang yang disampel, nematod lesi akar telah ditemui di 10 kebun (76% daripada jumlah kebun pisang yang disampel). Bahagian korteks akar dengan luka coklat kemerahan adalah ketara dalam kalangan semua negeri, di mana Johor mempunyai tahap keparahan jangkitan tertinggi iaitu sebanyak 36.3%, diikuti oleh Selangor, 20.7%, Perak dan Pahang, masing-masing 20.2% dan 20.4%. Kajian ini mendapati bahawa semua kawasan pensampelan melebihi had ambang, iaitu 5%. Min densiti populasi kebun yang dijangkiti dengan *Pratylenchus* spp. daripada 20 g akar dan 200 g tanah berbeza secara ketara di mana min populasi di Selangor, (akar = 838 dan tanah = 897), (akar = 18,050 dan tanah = 13,056) di Johor, (akar = 2341 dan tanah = 461) Perak dan (akar = 11,315 dan tanah = 7,199) Pahang telah direkodkan. Nematod parasit tumbuhan lain yang dikenalpasti ialah Rotylenchulus spp., Meloidogyne spp., Helicotylenchus spp., Hoplolaimus spp. dan Radophulus simili. Reaksi patologi P. coffeae terhadap pisang kultivar berangan (Musa paradisica) menunjukkan penggandaan selepas 12 minggu pertumbuhan. Terdapat perbezaan yang ketara dalam pertumbuhan vegetatif dalam patogen dan antara pelbagai peringkat inokulasi yang telah dinilai. Kadar penggandaan berada antara 1.6-4 pada P. coffeae. Pengurangan dalam panjang akar (-23.68 kepada -84.95), panjang pucuk (-13.11 kepada -45.80), berat akar (-21.30 kepada -99.85) dan berat pucuk (-9.30 kepada -61.62) telah direkodkan pada $P \le 0.05$. Pisang kultivar berangan menunjukkan kerentanan yang tinggi melalui aktiviti rintangan berasaskan 'polyphenol oxidase' dan 'peroxidase' pada semua hari selepas inokulasi berbanding kawalan, kecuali pada minggu ke-12 di mana ia telah mengalami penurunan atau tidak ketara berbanding kawalan. Pengamatan kami daripada kajian ini, mendedahkan bahawa P. coffeae adalah di antara nematoda parasit tumbuhan yang sangat merosakkan berkait dengan pisang di Semenanjung Malaysia menggantikan R. similis yang mana jarang dilaporkan atau terhad pada sesuatu kawasan dan mempunyai

kepadatan rendah seterusnya dikuti dengan *Meloidogyne* spp. Fenomena ini adalah lazim pada pisang kultivar berangan di negara ini dan taburan geografinya adalah tidak terbatas. Walaupun kajian morfologi pada populasi betina *P. coffeae* daripada Semenanjung Malaysia menunjukkan hanya sedikit variasi pada morfologinya berbanding sumber rujukan, kami memutuskan variasi ini adalah di dalam jarak variasi morfologi pada populasi *P. coffeae* sebagaimana yang telah dijelaskan sebelum ini kawasan lain di dunia, seterusnya mengesahkan bahawa solat-solat tersebut adalah *P. coffeae*. Umumnya ujian patogenisiti *P. coffeae* di rumah kaca, berupaya menekan pertumbuhan vagitatif pisang kultivar berangan dengan ketaranya. Indek lesi akar menunjukkan keparahan penyakit yang tinggi pada semua tingkat inoculum yang dinilai. Dapat disimpulkan bahawa *P. coffeae* telah luas tersebar dan menyebabkan kerosakan yang signifikan terhadap tanaman pisang di Semenanjung Malaysia.

ACKNOWLEDGEMENTS

All praise and thanks be to Allah S.W.T, the compassionate, most exalted with humility. My profound sincere thanks goes to my academic and research supervisory committee; Associate Prof. Dr. Kamaruzaman Sijam (Chairman) and Dr. Khairulmazmi B Ahmad, who by way of appreciation of ideas, encouragement, useful suggestions and constructive criticisms guided me throughout the period of the research.

My special appreciation goes to the staff of the Department of Plant Protection, Faculty of Agriculture, Universiti Putra Malaysia, especially able and capable Science Officers particularly, Mr. Shamsudin Bojang, Mrs. Junaina, Mr. Rozali, Mr. Johari and Mr. Nazri among others. Special gratitude is also extended to my parents, beloved and dear wives Habiba Muhammad and Habiba Abdul-Azeez, my kids for their patience and endurance while away for this programme. Niger State Government, Management and staff of Niger State College of Agriculture, Mokwa, Nigeria for allowing me to pursue M.Sc. program as well as my brothers, sisters and relatives for their unending moral support and understanding. Among friends and colleagues, I consider it necessary to acknowledge and appreciate the efforts of my friend and brother Magaji Usman, Dr. Yakubu Mambe, Ayoub Ahmed Ibrahim, Abdulazeez B kutawa, Osama B Zaid, Tijani Ahmad, Habu Musa and Jamilu Garba. Oladosu Adeniyi Yusuff ,Mr Ibrahim Abdulwasiu (chairman NAIJACOM) are not left out of this gratification, among others.

I certify that a Thesis Examination Committee has met on 03 May 2016 to conduct the final examination of Adamu Saidu Paiko on his thesis entitled "Prevalence and Pathogenicity of Root Lesion Nematode, *Pratylenchus* spp. on Banana (*Musa paradisiaca* L.) in Peninsular Malaysia" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Hafidzi b Mohd Noor, PhD Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Yahya bin Awang, PhD Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

Nik Ahmad Irwan Izzauddin Nik Him, PhD Associate Professor School of Biological Sciences Universiti Sains Malaysia (External Examiner)

ZULKARNAIN ZAINAL, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 26 July 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Kamaruzaman Bin Sijam, PhD Associate Professor Faculty of Agriculture

Universiti Putra Malaysia (Chairman)

Khairulmazmi Bin Ahmad, PhD Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Member)

BUJANG BIN KIM HUAT, PhD Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by Graduate Student

I hereby confirmed that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:

Date:

Name and Matric No.: Adamu Saidu Paiko, GS 40046

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:	
Name of Chairman	
Committee:	Associate Professor Dr. Kamaruzaman Bin Sijam
Signature:	
Name of Member	
of Supervisory Committee	Dr. Khairulmazmi Bin Ahmad
e chining e c	

TABLE OF CONTENTS

			Page
ABST ABST ACK APPI DEC LIST LIST LIST	FRACT FRAK NOWLI ROVAL LARAT OF TA OF FIC	EDGEMENTS ION BLES GURES ATES	i iii v vi viii xii xii xiv xv
СНА	PTER		
1	INTI 1.1 1.2 1.3	RODUCTION Background Justification Of Study General Objective 1.3.1 Objectives	1 1 2 2 2
2	LITH 2.1 2.2	 ERATURE REVIEW Banana Plant Parasitic Nnematodes 2.2.1 Introducing Plant Parasitic Nematodes (Ppn) 2.2.2 Incidence Of Root Lesion Nematodes Infestation On Banana 2.2.3 Life Cycle Of Root Lesion Nematode 2.2.4 Epidemiology Of Root Lesion Nematode 2.2.5 Effect Of Ecological Factors On Root Lesion Nematode 2.2.5 Effect Of Ecological Factors On Root Lesion Nematode 	4 5 5 6 7 10 12
	2.3 2.4	2.2.4.1 Soil Moisture 2.2.4.2 Soil Ph 2.2.4.3 Soil Temperature 2.2.4.4 Soil Texture Pathogenicity Of Root Lesion Nematode Damage Threshold Of Root Lesion Nematode	12 12 13 13 14
	2.5 2.6	Diseases Development Integrated Pest Management (Ipm) Program	15 15
3	DISE ROC BAN	EASES PREVALENCE AND SEVERITY ASSESSMENT OF)T-LESION NEMATODE PRATYLENCHUS SPP. ON IANA (MUSA SPP.)	18
	3.0 3.1	Introduction Materials And Methods 3.1.1 Sampling Collection 3.1.2 Isolation Of Nematodes 3.1.3 Morphological Identification Of Nematodes 3.1.4 Molecular Identification Of Pratylenchus 3.1.5 Root Damage Assessment On Bananas	18 19 19 21 22 22 23

C

		3.1.6 Soil Analysis	24
		3.1.6.1 Organic Carbon Content	24
		3.1.6.2 Organic Matter Content	24
		3.1.6.3 Measuring Soil Particle Seize	24
		3164 Moisture And Ph	25
		3165 Electric Conductivity	26
	32	Statistical Analysis	26
	33	Results And Discussion	26
	5.5	2.3.1 Disease Prevalence And Severity	26
		2.3.2 Morphological Identification Of Protylenchus	20
		2.2.2 Molecular Identification Of Pratylenchus	22
		2.2.4 Dravalance Of Dratulanchus Infaction On Different	27
		5.5.4 Prevalence Of Pratylenchus Infection Off Different	37
		Banana Cultivars	27
		3.3.5 Damage OI Pratylenchus Colleae. On Pisang	37
		Berangan	40
		3.3.6 Relationship Between Soil Properties And Nematode	40
		Abundance In Banana Fields In Malaysia	
	3.4	Discussion	44
	3.5	Conclusions	48
	POPU BAN 4.1 4.2 4.3 4.4 4.5 4.6 4.7 4.8 4.9	ULATION LEVELS OF PRATYLENCHUS COFFEAE ON ANA CULTIVAR BERANGAN Introduction Materials And Methods 4.2.1 Pure Culture Preparation Glasshouse Experimental Layout Assessment Of Plant Vigour Assessment Of Disease Severity Preparation Of Crude Extract For Po And Ppo Enzymes Assay 4.6.1 Nematodes Enumeration Results And Discussions 4.7.1 Plant Vigour Assessment Of Nematode Population And Root Necrosis Peroxidase (Po) And Polyphenol Oxidase (Ppo) Activity In Pisang Berangan Against P. Coffeae	49 50 50 50 51 51 52 52 52 52 57 59
	4.10	Discussion	60
	4.11	Conclusion	62
5	SUM FOR	MARY, CONCLUSION AND RECOMMENDATIONS FUTURE RESEARCH	63
REF	RENCI	FS	64
	NDICE	S.	76
BIOD		S F STUDENT	85
DIDI			05
LODI	ACAID		00

LIST OF TABLES

Table		Page
2.1	Pratylenchus spp. and their susceptible hosts	7
2.2	Pratylenchus spp., host crops and their alternative weed hosts	11
2.3	Suitable host crops and threshold limits of Pratylenchus Spp.	14
2.4	Integrated management strategies for nematode management.	17
3.1	Areas surveyed for root-lesion nematode Pratylenchus spp. in Peninsular Malaysia	21
3.2	Mean population of nematodes genera with prevalence of occurrence and percentages from both roots and soils in Peninsular Malaysia	29
3.3	Incidence and mean populations of root lesion nematodes on banana cultivar berangan in Peninsular Malaysia	30
3.4	Morphometrics of the females of the Pratylenchus coffeae populations collected in Peninsular Malaysia	32
3.5	Three representative isolates of P. coffeae (PRA-A, B and C) with their corresponding matches from the Genbank	37
3.6	Occurrence of Pratylenchus on different Banana cultivars in Peninsular Malaysia	37
3.7	Effects of soil texture towards nematodes population and genus	41
3.8	Correlations of soil physio-chemical properties on nematodes population	42
4.1	Effect of different inoculation levels of P. coffeae on plant heights (cm2) of Pisang berangan cultivar 12 weeks after inoculation	53
4.2	Effect of different inoculation levels of Pratylenchus coffeae on leaf size (cm2) of Pisang berangan cultivar, in Peninsular Malaysia 12 weeks after	53
4.3	Effect of different inoculation levels of Pratylenchus coffeae on pseudostem girth (cm2) of Pisang berangan, 12 weeks after inoculation	54
4.4	Effects of different inoculation levels of Pratylenchus coffeae on fresh root and shoot lengths (cm2) and weights (kg) of Pisang berangan, 12 weeks after inoculation	55

4.5 Reproduction and percentage root necrosis caused by different inoculation levels of Pratylenchus coffeae on Pisang berangan cultivar from Peninsular Malaysia

57



LIST OF FIGURES

Figure		Page	
2.1	Volumes and values of banana produced and exported in 2011	5	
2.2	Schematic diagram of life cycle of PPN, A) Migrotary- endoparasitic B) Ecto-parasitic C) Semi-enparasitic	9	
3.1	Areas surveyed for root-lesion nematode Pratylenchus spp. in Peninsular Malaysia	20	
3.2	Neighbour-Joining tree of three representative isolates of P .coffeae (PRA-A, B and C) populations from Malaysia with their corresponding similar isolates obtained from the Genbank .	36	
3.3	Average percentage root necrotic lesion on Pisang berangan root cortex from four states of Peninsular Malaysia	40	
3.4	Effect of organic matter content on nematodes population	42	
3.5	Effect of moisture content on nematodes population	43	
3.6	Effect of soil pH on nematodes population	43	
3.7	Effect of electrical conductivity on nematodes population	44	
4.1	Activity of PPO (Polyphenol oxidase) of Pisang Berangan roots at 1, 2, 3, 4 and 12 weeks after inoculation with 5000 P. coffeae and Un-inoculated (control).	59	
4.2	Activity of PO (Peroxidase) of Pisang berangan roots at 1, 2, 3, 4 and 12 weeks after inoculation with 5000 P. coffeae and Un- inoculated (control).	60	

 \mathbf{G}

LIST OF PLATES

Plate		Page
3.1	Nematode extraction using whitehead tray method	22
3.2	Sieves arrangement for soil particle size analysis	25
3.3	Nematodes found alongside Pratylenchus spp (A) Pratylenchus spp (B) Meloidogyne spp (C) Helicotylenchus spp	27
3.4	Plate 3.4: nematodes found alongside Pratylenchus spp D) Rotylenchulus spp (E) Hoplolaimus spp (F) Radophulus simili	28
3.5	Morpholological features of Pratylenchus coffeae. (A) Tail, (B) (a- lip region, b-stylet and c-median bulb.	31
3.6	Nucleotides comparison of local P. coffeae isolate (LOCAL) with reference isolate from Genebank, LC030392 (SBJCT) isolated from Japan.	36
3.7	Above ground symptoms of Root lesion Nematodes on Banana (A).Stunted and toppled plant (B) smaller fruits	38
3.8	Lesion scores of Pratylenchus spp on Banana from four states of Peninsular Malaysia	39
4.1	Above ground symptoms of stunted growth caused by P. coffeae (B) and control (A) 12 weeks after inoculation	56
4.2	Below ground symptoms of reddish brown lesion caused by P. coffeae (B) and control (A) 12 weeks after inoculation	58

C

CHAPTER 1

INTRODUCTION

1.1 Background

Banana is the world's most prominent fruit and also among the major staple foods, alongside, wheat, maize and rice. About a hundred and seven million metric tons of bananas were produced in 2011, across more than 130 countries covering an area of about 0.1 per cent of the agricultural area utilized in the world (Agritrade 2013). Food and Agriculture Organization of United Nations (FAO) (2013) estimated the exchange value to be around US\$9 billion and a trade value of about US\$25 billion. Over 1,000 cultivars of the bananas have been identified worldwide (Heslop-Harrison and Schwarzacher 2007). In the Malaysian scenario, it has been recognized as the second most commonly grown fruit crop after durian, with a cultivated area of about 29,000 ha and production of 530,000 metric tonnes, which amount to \$24 million (Tengku et al, 2011). Johor, Sabah and Sarawak served as major states producing banana covering 27,543 hectares in 2009. Pisang berangan and the Cavendish cultivars cover almost 50% of the banana growing land, while the remaining land are cultivated with other prominent cultivars like; Pisang Mas, Pisang Abu, Pisang Nangka, Pisang Rastali, Pisang Awak, Pisang Tanduk and Pisang Raja (Tengku et al, 2011), about 12% of the bananas produced in Malaysia, are mainly exported to Middle East, Brunei, Hong Kong, and Singapore, while remaining which are mostly cultivated by smallholders are consumed locally.

Banana suffers from many diseases caused by fungi, bacteria, viruses and nematodes (Jones, 2000). The pathogens that cause these diseases vary in the amount of damage they cause with some having very minor effects while others are extremely serious and can kill the plant. Amongst the various pathogens infecting the banana plants, the nematodes infestation is of the prime importance, causing an annual yield loss of approximately 20% (De Waele and Elsen, 2007). The parasitizing of banana by Burrowing nematode is a worldwide phenomenon. In addition, diseases like Fusarium wilt, sigatoka and blood disease have threatened banana production in Malaysia, leading to decreased in output. Another factor seen as a threat is high costs of labour and marketing problems (Abdul Rahman et al., 2014).

Nematodes with an estimated population of about a million species are diverse metazoans (Lambshead, 2004). They form part of almost all ecosystems as consumers of dissolved and particulate organic matter, parasite of plants and animals, bacterivores and herbivores. They are believed to be the most abundant in soil and aquatic sediments. McCarter (2009) put an estimated global loss of \$118 billion for 2001, resulting from their impact on crop of which almost half was linked to only two crops; maize and rice. This showed paucity study of their kind, pointing to only 26,000 (estimated < 3%) species studied to date (Hugot et al., 2001; Hallan, 2007). The crops damage from plant parasitic nematodes are agro-climatic conditions dependent, host susceptibility, pathogenicity and other climatic factors (Queneherve, 2009). The impacts of nematode infestation on banana plants are often asymptomatic, owing to the possession of thick root epidermis or recognized at very

advance stage of disease, and the farmers has no option rather than the gross removal of the plant (Abdul Rahman et al., 2014). (PPN) infection on bananas make water up take and nutrients by the host plant less, causing maturation to be delayed, stunting and reduction in bunch size. Level of the damage may differ from vegetative period slightly extended to the emergence of symptom of attack by lesion nematode, which causes the plant to topple. Geographical locations determine the effect of a community of PPN damages on crop (Quénéhervé, 2009). Damage caused by PPNs is like the damage caused by any other pests or parasites, and is influenced by the conditions of environment, host susceptibility, and nematode pathogenicity. There are only few reports on the prevalence of nematodes in Malaysia and also there is inconsistency and knowledge gap regarding the occurrence and prevalence of nematodes. In the past few decades few research work or survey have been done and a very little information is available about the infestation of nematodes and their damage on fruit crops (Sidam and Bilal Mat, 1983; Abdul Rahman et al., 2014).

The most important agriculturally group of nematodes, following cyst nematodes (*Heterodera* and *Globodera*) and root knot nematodes (*Meloidogyne* spp.) are the genus *Pratylenchus*, the root lesion nematodes (Davis and MacGuidwin 2000). Root lesion nematode (*Pratylenchus* spp.) cause considerable damage to banana plant (*Musa* spp.) globally. In Malaysia, it has been reported as one of the damaging species in a study conducted by Abdul Rahman et al., (2014) on distribution of plant parasitic nematodes in Peninsular Malaysia, however, their damaging status has not been defined.

1.2 Justification of study

There are only few reports available or work done in the past on the persistence of nematodes in general, lesion nematodes in particular on the banana cultivated areas. Therefore, it is highly desirable to conduct a research on the population distribution of root lesion nematode, *Pratylenchus* spp. and level of their infestation for management of their diseases in banana cultivated areas in Malaysia. Since condition favorable for root lesion nematode survival is found in Malaysia, and the idea of the species involved will be important to future breeding efforts, this project was undertaken.

1.3 General objective

The main objective of the present research work was to investigate the prevalence of *Pratylenchus* spp. infestation in banana (*Musa* spp.) and assessment of their disease severity in the Peninsular Malaysia.

1.3.1 Objectives

Considering the above points, the studies were conducted with the following objectives:

(1) To investigate disease prevalence and identification of root lesion nematodes among banana acreage in Peninsular Malaysia (2) To determine the pathological reaction and damaging population level of *Pratylenchus spp.* on banana cultivar berangan.



REFERENCES

- Abd-Elgawad, M.M.M. and Aboul-Eid, H.Z. (2005). Effects and prospects of phytonematode damage and control in Egypt. *Egyptian Journal of. Agricultural Research*, 2(1):439-456.
- Abdul Rahman, S. A, Moh'd S.N, Zain, M.Z. Bilal Mat, A.K. Sidam, R.Y. Othman and Z. Mohamed (2014). Population Distribution of Plant-Parasitic Nematodes of Bananas in Peninsular Malaysia, *Sains Malaysiana*. 43(2): 175–183
- Acosta, N. and Malek, R.B. (1979). Influence of temperature on population development of eight species of *Pratylenchus* on soybean. *Journal of Nematology* 11, 229-232.
- Agritrade. (2013). *India preparing a major banana export program*. Retrieved from <u>http://agritrade</u>. cta.int/Agriculture/Commodities/Bananas/ India-preparing-a-major-banana-export-programme.
- Aguilar, E.A., Turner, D.W. and Sivasithamparam, K. (2000). Fusarium oxysporum f. sp. cubense inoculation and hypoxia alter peroxidase and phenylalanine ammonia lyase activities in nodal roots of banana cultivars (*Musa* sp.) differing in their susceptibility to Fusarium wilt. *Australian Journal of Botany* 48, 589-596.
- Akhtar M, Malik A. (2000). Roles of organic soil amendments and soil organisms in the biological control of plant-parasitic nematodes: *a review. Bioresource Technology*. 74, 35-47.
- Allen, M.W. and Jensen, H.J. (1951). *Pratylenchus vulnus*, new species (Nematoda: Pratylenchidae), a parasite of trees and vines in California. *Proceedings of the Helminthological Society of Washington* 18, 47-50.
- Anwar, S.A., Rauf, C.A. & Gorsi, S.D. (1992b).Weeds as alternate hosts of phytonematodes. *Afro-Asian Journal of Nematology* 2, 41-47.
- Ardakani, A.S, Mafi, Z.T, Hesar, A.M and Goltappeh, E.M. (2014). Relationship between Soil Properties and Abundance of Tylenchulus semipenetrans in Citrus Orchards, Kohgilouyeh va Boyerahmad Province. *Journal of Agricultural. Science. Technology.* 16, 1699-1710
- Azcón-Aguilar, C., Jaizme-Vega, M.C. and Calvet, C. (2002). The contribution of arbuscular mycorrhizal fungi to the control of soilborne pathogens. In: Gianinazzi, S., Schüepp, H., Barea, J.M. & Hasselwandter, K. (Eds). Mycorrhizal technology in agriculture. Basel, Switzerland, Birkhäuser Verlag, 187-197.
- Bajaj, H.K. and Bhatti, D.S. (1984). New and known species of *Pratylenchus Filipjev*, 1936 (Nematoda: Pratylenchidae) from Haryana, India, with remarks on intraspecific variations. *Journal of Nematology*. 16, 360-367.

- Barekye A., I.N. Kashaija, W.K. Tushemereirwe and E. Adipala. (2000). Comparison of damage levels caused by *Radopholus similis* and *Helicotylenchus multicinctus* on bananas in Uganda. *Annals of Applied Biology*. 137, 273-278.
- Barker, K.R. and Koenning, S.R. (1998). Developing sustainable systems for nematode management. *Annual Review of Phytopathology* 36, 165-205.
- Boerma, H.R. and Hussey, R. (1992). Breeding plants for resistance to nematodes. *Journal of Nematology* 24, 242-252.
- Boning, C.R (2006). Florida's Best fruiting Plants: Pineapple Press Inc., Florida.
- Bridge J., Fogain R. and Speijer P. (1997). The root lesion nematodes of banana: *Pratylenchus coffeae* (Zimmermann, 1898) Filipjev and Schuurmans Stekhoven 1941. *Pratylenchus goodeyi* Sher and Allen, 1953. Musa Pest Fact Sheet No. 2.
- Broadley, R., Rigden, P., Chay-Prove, P., Daniells, J. (2004). Subtropical Banana Growers Handbook. *Queensland Department of Primary Industries*, 1-206.
- Brooks, F.E. (2004). Plant-parasitic nematodes of banana in American Samoa. *Nematropica* 34, 65-72.
- Brown, R.H. and Kerry, B.R. (1987). Principles and practice of nematode control in crops. Sydney, Australia, Academic Press, 447.
- Castillo, P., Gómez-Barcina, A. & Jiménez-Díaz, R.M. (1996).Plant parasitic nematodes associated with chickpea in southern Spain and effect of soil temperature on reproduction of *Pratylenchus thornei*. *Nematologica* 42, 211-219.
- Castillo P, Di Vito M, Voylas N, Jiménez-Díaz RM, 2001. Host-parasite relationships in root-knot disease of white mulberry. *Plant* Disease 85, 277-81.
- Castillo P. and Vovlas N. (2007). *Pratylenchus*, (Nematoda, Pratylenchidae): diagnosis, biology, pathogenicity and management. *Nematology Monographs and Perspectives* 6, 529.
- Chabrier, C., Queneherve, P., (2003). Control of the burrowing nematode (Radopholus similis Cobb) on banana: impact of the banana field destruction method on the efficiency of the following fallow. *Crop Protection*. 22, 121–127.
- Chao, C.P. (2013). R&D Strategies and Policy Making/Regulatory of Fusarium Wilt on Cavendish Banana in Taiwan. Paper presented at consultation workshop on socio economic impact of Fusarium Wilt Disease of Cavendish Banana in the Asia-Pacific region. Waterfront Insular Hotel, Davao city, City, 11-15.
- Chitamba J., P. Manjeru, C.C. Chinheya, N. Mudada, and M. Handiseni. (2013). Plantparasitic nematodes associated with banana in Rusitu Valley, Zimbabwe. *Nematropica*. 43. 113-118.

- Chitimbar, J.J. & Raski, D.J. (1985). Life History Of *Pratylenchus vulnus* on carrot discs. *Journal of Nematology* 17, 235-236.
- Choshali, A. H., Seraji, A., Rezaee, S., Shirinfekr, A., Mirghasemi, S. N. (2015).
- The effects of soil organic matter content and soil texture on the population number of *Pratylenchus* loosi, in tea plantation of Iran. *International Journal of Agronomy and Agricultural Research (IJAAR)*, p. 54-61.
- Cobb, N.A. (1917). A new parasitic nematode found infesting cotton and potatoes. Journal of Agricultural Research. 11, 27-33.
- Coomans A. 2002. Present status and future of nematode systematics. *Nematology* 4: 573-582.
- Coomans A., De Coninck L. and Heip C. 1978. Data to be considered in descriptions of new species or redescriptions of poorly known species. Annals of the Royal Zoological Society of Belgium 108: 119-122.
- Constantine, D., Rosell, G. (2001). The Musaceae: An annotated list of the species *Ensete*, *Musa* and *Musella*. (http://www.usersglobalnet.co.uk/~drc/index.htm)
- Coyne D.L, J.M. Nicol, C.B. Claudius. (2007). Practical plant nematology: a field and laboratory guide. *Green Ink Publishing Services Ltd, UK*. 82.
- Davide, R. G. (1980). Influence of cultivar, age, soil texture, and pH on *Meloidogyne incognita* and *Radophohrs similis* on banana. Plant Disease, 64 : 571-573.
- Davis, E.L., and MacGuidwin, A.E. (2000). Lesion nematode disease. *The Plant Health Instructor*. 2000-1030-02.
- Dc Man JG (1880). Die einheimischen, frei in der reinen Erde und im süssen Wasser lebenden Neniatoden monographisch bearbeitet. *Tijdschrift der Nederlandse Dieik. Vereenig.* 5:1-104.
- De Goede, R.G.M. and Bongers, T. (1998). Nematode communities of northern temperate grassland ecosystems. Giessen, Germany, Focus Verlag, 338.
- De Waele, D. and Elsen, A. 2007. Challenges in tropical plant nematology. *Annual Review* of *Phytopathology* 45: 457–485.
- De waele, D., and R. G. Davide 1998. The rootknot nematodes of banana. *Meloidogyne incognita* (Kofoid & White, 1919) Chitwood, 1949. *Meloidogyne javanica* (Treub, 1885) Chitwood, 1949. Musa Pest Factsheet No. 3
- Di Vito, M., Greco, N. and Saxena, M.C. (1992). Pathogenicity Of *Pratylenchus thornei* on chickpea in Syria. *Nematologia Mediterranea* 20, 71-73.
- Di Vito, M., Zaccheo, G. and Catalano, F. (2000). Effect of *Pratylenchu neglectus* and *P. thornei* on the growth of faba bean. *Nematologia Mediterranea* 28, 261-265.

- Doohan, F. (2005). Fungal pathogen of plants. In: Fungi: Biology and Application. Kavanagh, K. (eds.). John Wiley & Sons Ltd, England. 219-228.
- Doucet, M.E., Lax, P. and Pinochet, J. (1998). Variability of some external characters in PRATYLENCHUS vulnus Allen & Jensen, 1951 (Nematoda: Tylenchida). Fundamental and Applied Nematology 21, 205-206.
- DPI&F (2007). Summary of Plant Entry Conditions for Queensland. Plant Biosecurity Program, Queensland Department of Primary Industries & Fisheries, vailable online at <u>http://www2.dpi.qld.gov.au/extra/pdf/aphs/cond_issue.pdf</u>.
- Endo BY. 1959. Responses of root-lesion nematodes, *Pratylenchus brachyurus* and P. zeae, to various plants and soil types. *Phytopathology* 49, 417-421.
- Fallas G.A., Sarah J.L. and Fargette M. (1995). Reproductive fitness and pathogenicity of eight *Radopholus similis* populations on banana plants (Musa AAA cv. Poyo). *Nematropica* 25: 135-141.
- Ferris, H. (1981). Dynamic action thresholds for diseases induced by nematodes. *Annual Review of Phytopathology* 19, 427-436.
- Filipjev, 1936 (Nematoda: Pratylenchidae). Journal of Nematology, Morphology and Systematics 3, 165-174.
- Florini, D.A., Loria, R. and Kotcon, J.B. (1987). Influence of edaphic factors and previous crop on *Pratylenchus* spp. population densities in potato. *Journal of Nematology* 19, 85-92.
- Freckman, D.W. and Caswell, E.P. (1985). The ecology of nematodes in agroecosystems. Annual Review of Phytopathology 23, 275-296.
- Food and Agricultural Organizations. Undated. Plant quarantine. (<u>http://www.fao.org.inpho_Archive/content/documents/vlibrary/new_fao/x5417</u> e/x5417e01.htm; Accessed December, 2013).
- Gee, GW and JW Bauder. 1986. Particle-size analysis. p. 383-411. In A Klute (ed.) Methods of Soil Analysis, Part 1. Physical and Mineralogical Methods. Agronomy Monograph No. 9 (2ed). American Society of Agronomy/Soil Science Society of America, Madison, WI.
- Gnanapragasam NC, Sivapalan P. 1991. Influence of soil types and storage conditions on the recovery of *Pratylenchus* loosi from soil samples. *Afro–Asian Journal of Nematology* 1, 150-153.
- Godfrey, G.H. (1929). A destructive root disease of pineapple and other plants due to *Tylenchus brachyurus*. *Phytopathology* 19, 611-629.
- Goswami, B.K., Singh, D.V., Seth, M.L. and Gupta, J.N (1970). Studies on association of root-knot nematode Meloidogyne incognita and Sclerotium rolfsii in brinjal. *Indian Phytopathology* 3: 587-589.

- Gowen, S.R. 1995. Pests. In: S.R. Gowen (ed.), Bananas and plantains. Chapman and Hall, London, UK. p. 382–402.
- Gowen S.R. (2000). Root-lesion nematodes. Pp. 303-306 in Diseases of Banana, Abacá and Enset (D.R. Jones, ed.). CABI Publishing. Wallingford, UK.
- Gowen, S., Quénéhervé, P. & Fogain, R. (2005).Nematode parasites of bananas and plantains. In: Luc, M., Sikora, R.A. & Bridge, J. (Eds). *Plant parasitic nematodes in subtropical and tropical agriculture, 2nd edition*. Wallingford, UK, CABI Publishing, 611-643.
- Graham, T.W. (1951). Nematode root rot of tobacco and other plants. Bulletin 390. South Carolina Agricultural Experiment Station, Clemson Agricultural College, 25.
- Greco, N. and Di Vito, M. (2009). Population dynamics and damage levels. In:Perry, R.N., Moens, M. and Starr, J.L. (eds). Root-Knot Nematodes. Wallingford,Oxfordshire: CAB International, Wallingford, UK, pp. 246– 274.
- Halbrendt, J.M. and LaMondia, J.A. (2004). Crop Rotation and other cultural practices.
 In:Nematology, Advances and Perspectives, Vol 2: Nematode Management and Utilization, ed ZX Chen, SY Chen and DW Dickson, 909-930.
 Wallington/Cambridge, UK: CABI Publishers.
- Hallan, J. 2007. Synopsis of the Described Nematoda of the World. <u>http://insects.tamu.edu/research/collection/hallan/Nematoda/</u> Family/OnematodaIndex0.htm (29 April 2010).
- Hassan, N.M., (2004).Nematodes in banana in Malaysia. *Towards management of Musa nematodes in Asia and the Pacific*, 47.
- Heslop-Harrison, J.S. & Schwarzacher, T. 2007. Domestication, genomics and the future for banana. *Annals of Botany* 100: 1073-1084.
- Hugot, J., P. Baujard, and S. Morand. 2001. Biodiversity in helminths and nematodes as a field study: an overview. *Nematology* 3: 199–208.
- Hyam, R,. Pankhurst R, (1995). Plants and Their Names: A Concise Dictionary, Oxford University Press Oxford.
- International Network for the Improvement of Banana and Plantain (INIBAP), Montpellier, France, 4.
- Jamaluddin, S.H. (1999). Banana R&D thrusts in Malaysia. In Advancing Banana and Plantain R&D in Asia and the Pacific, edited by Molina, A.B. & Roa, V.N. Guangzhou: Proceedings of the 9th INIBAP-ASPNET Regional Advisory Committee Meeting. 108-111.
- Jones, D. R. (ed.). 2000. Diseases of Banana, Abacá and Enset. CABI Publishing. Wallingford, United Kingdom.

- Jones JT, Haegeman A, Etienne GJD, Hari SG, Helder J, Michael GKJ, Kikuchi T, Rosa ML, Juan EPR, Wesemael, WML, Perry RN. (2013). Top 10 plant-parasitic nematodes in molecular plant pathology. Molecular Plant Pathology 4: 946-961.
- Kable, P.F. & Mai, W.F. (1968). Influence of soil moisture on *Pratylenchus penetrans*. *Nematologica* 14, 101-122.
- Kamira M., S. Hauser, P. Van Asten, D. Coyne, and H. L. Talwana. (2013). Plant parasitic nematodes associated with banana and plantain in eastern and western Democratic Republic of Congo. *Nematropica* 43: 216-225.
- Khan, M.R. and A. Hassan, (2010). Nematode diversity in banana rhizosphere from West Bengal, India. Journal of Plant Protection Research, 50: 263-268.
- Kheiri, A., Borhani, A., Okhovat, M. & Pourjam, E. (2002). Interaction between root lesion nematode *Pratylenchus vulnus* and two species of *Fusarium* on growth and development of maple seedlings. *Mededelingen Faculteit Landbouwkundige en Toegepaste Biologische Wetenschappen, Universiteit Gent* 67, 703-706.
- Koen, H. (1967). Notes on the host range, ecology and population dynamics of *Pratylenchus brachyurus. Nematologica* 13, 118-124.
- Koike, S.T., Gaskell, M., Fouche, C., Smith, R., and Mitchell, J. (2000). Plant diseas management for organic crops. Available from URL:
- Kokkinakis, D.M and Brook, J.L, (1979). Tomato peroxidase: purification, characterization, and catalytic properties. *Journal of plant physiology*,63(1):93-9.
- Kornobis, S. and Wolny, S. (1997). Occurrence of plant parasitic nematodes on weeds in agrobiocenosis in the Wielkopolska region in Poland. *Fundamental and Applied Nematology* 20, 627-632.
- Kubo, R.K., Silva, R.A., Tomazini, M.D., Oliveira, C.M.G., Mazzafera P. & Inomoto, M.M. (2002). Patogenicidade de *PRATYLENCHUS coffeae* em plântulas de cafeeiro cv. Mundo Novo. *Fitopatologia Brasileira* 28, 41-48.
- Lambshead, P.J.D. 2004. Marine nematode biodiversity. In Z.X. Chen, S.Y. Chen, and D.W. Dickson (eds.), Nematology, Advances and Perspectives. ACSE-TUPBook Series. Pp. 436–467.
- Loof, P.A.A. (1991). The family Pratylenchidae Thorne, 1949. In: Nickle, W.R. (Ed.). Manual of agricultural nematology. New York, NY, USA, Marcel Dekker, 363-421.
- Loof, P.A.A. (1960). Taxonomic studies on the genus *Pratylenchus* (Nematoda). *Tijdschrift ober Plantenziekten* 66, 29-90.

- Mani, A. and Al-Hinai, M.S. (2003). Observations on sampling tool, vertical distribution and weed hosts of *PRATYLENCHUS jordanensis* in an alfalfa field. *Nematologia Mediterranea* 31, 155-156.
- Mansoor A. Siddiqui and M. Mashkoor Alam (2001). Integrated Management of the Root-Knot and Reniform Nematodes With Cropping Sequences And Ploughin. Arch. Phytopath. Pflanz., Vol. 33, Pp. 415-430
- Manzanilla-López, R., Evans, K. and Bridge, J. (2004). Plant diseases caused by nematodes. In: Cheng, Z.X., Chen, S.Y. & Dickson, D.W. (Eds). Nematology advances and perspectives, volume 2. Nematode management and utilization. Wallingford, UK, CABI Publishing, 637-716.
- Mateille T, Folkertsma S (1991). A survey of nematodes and fungi in roots of banana CV Poyo in the Ivory Coast. Review. *Nematol*. 14(1):3-8.
- Marin D.H., T.B. Sutton & K.R. Barker. (1998). Dissemination of banana in Latin America and the Caribbean and its relationship to the occurrence of *Radopholus similis*. *Plant Disease* 82:964-974.
- Martin, F.N. (2003). Development of alternative strategies for the management of soilborne pathogens currently controlled with methyl bromide. *Annual Review of Phytopathology*. 41: 325-50.
- McCarter, J. 2009. Molecular Approaches Toward Resistance to Plant-Parasitic Nematodes. Plant Cell Monograph, doi:10.1007/7089_2008_32. 12/24/09. (29 April 2010).
- McKeen, D.C. and Mountain, W.B. (1960). Synergism between PRATYLENCHUS penetrans and Verticillium albo-atrum in eggplant wilt. Canadian Journal of Botanical 38: 789-794.
- Mckenry, M.V. and Kretsch, J. (1987). Survey of nematodes associated with almond production in California. *Plant Disease* 71, 71-73.
- Mcsorley, R. and Parrado, L. (1981). Population fluctuations of plant-parasitic nematodes on bananas in Florida. Procedure. Florida horticultural Society, 94 : 321-323.
- Mcsorley R. and J.L. Parrado. (1986). *Helicotylenchus multicinctus* on bananas: an international problem. *Nematropica* 16: 73-91.
- Melakeberhan, H., Jones, A.L. & Bird, G.W. (2000). Effects of soil pH and *Pratylenchus penetrans* on the mortality of 'Mazzard' cherry seedlings and their susceptibility to *Pseudomonas syringae* pv. *syringae*. *Canadian Journal of Plant Pathology* 22, 131-137.
- Moens, T., M. Araya, and D. De Waele. 2001. Correlations between nematode numbers and damage to banana (*Musa* AAA) roots under commercial conditions. *Nematropica* 31:55-65.

- Morgan, G.T. & Maclean, A.A. (1968). Influence of soil pH on an introduced population of *Pratylenchus penetrans*. *Nematologica* 14, 311- 312.
- Morgan, G.T. & Maclean, A.A. (1968). Influence of soil pH on an introduced population of *Pratylenchus penetrans*. *Nematologica* 14, 311-312.
- Morgan, G.D., Stevenson, W.R., Macguidwin, A.E., Kelling, K.A., Binning, L.K. & Zhu, J. (2002). Plant pathogen population dynamics in potato fields. *Journal of Nematology* 34, 189-193.
- Nega, G. and Fetena, S. (2015).Root Necrosis Assessment of Plant Parasitic Nematodes of Banana (Musa spp.) at Arbaminch, *Ethiopia. Journal of Biology, Agriculture* and Healthcare www.iiste.org. ISSN 2224-3208 (Paper) ISSN 2225-093X (Online). Vol.5, No.15.
- Nguyen Thi, T., Elsen, A., Nhi, H., De Waele, D. (2015). Host range characterisation, in vivo reproduction and damage potential of *Pratylenchus coffeae* populations from Vietnam. *Russian Journal of Nematology*, 23 (1), 41-52.
- Nico, A.I., Rapoport, H.F., Jiménez-Díaz, R.M. & Castillo, P. (2002). Incidence and population density of plant-parasitic nematodes associated with olive planting stocks at nurseries in Southern Spain. *Plant Disease* 86, 1075-1079.
- NIPP National Institute of Plant Protection, (1997). Methods for plant protection research.Agricultural publishing house, 100.
- Noling, J.W. (2012). Nematode management in cucurbits (cucumber, melons, squash). Circular Publication No. ENY-025, IFAS Extension, University of Florida, FL, USA.
- Norton, D.C. and Burns, N. (1971). Colonization and sex ratios of *Pratylenchus alleni* in soybean roots under two soil moisture regimes. *Journal of Nematology* 3, 374-377.
- Olabiyi T.I, Olayiwolai A.O, Oyediran G.O. (2009). Influence of soil textures on distribution of phytonematodes in the south western Nigeria. *World journal of Agricultural Science* 5, 557-560.
- Oliveira, C.M.G., Inomoto, M.M., Vieira, A.M.C. & Monteiro, A.R. (1999a). Effect of initial population densities of *Pratylenchus* brachyurus on the growth of Coffea arabica cv. Mundo Novo and C. canephora cv. Apota. *Nematropica* 29, 215-221
- Olthof, T.H.A. and Wolynetz, M.S. (1991). *Pratylenchus penetrans* and *P. neglectus* in tubers of potato (*Solanum tuberosum*) in Ontario. *Canadian Journal of Plant Science* 71, 1251-1256.
- Olthof, T.H.A. and Yu, Q. (1999). Reduction of root-lesion nematodes (*Pratylenchus penetrans*) in tubers of potato (*Solanum tuberosum*) during cold storage. *Canadian Journal of Plant Pathology* 21, 154-158.

- Orton-Williams, K.J. and Siddiqi, M.R. (1973). *Radopholus similis* C.I.H Descriptions of Plant-Parasitic Nematodes. Wallingford: CAB International. Set 2 No. 27.
- Osseni, B., Sarah, J.L. and Hugon, R. (1997). Effect of soil pH on the population development of PRATYLENCHUS brachyurus (Godfrey) in pineapple roots and on the growth and yield of the plant. *Acta Horticulturae* 425, 423-433.
- Pablo Castillo and Nicola Vovlas, (2007) *Pratylenchus* (Nematoda: Pratylenchidae): Diagnosis, Biology, Pathogenicity and Management, Nematology Monographs and Perspectives. Volume 6.
- Pattison, A. B., J. M. Stanton, J. A. Cobon and V. J. Doogan. 2002. Population dynamics and economic threshold of the nematodes *Radopholus similis* and *Pratylenchus goodeyi* on banana in Australia. *International Journal of Pest Management* 48:107-111.
- Pattison, T. and Lindsay, S. (2006). Banana soil Heath user's manual. Queensland Department of Primary Industries and Fisheries.
- Pau, C.G., C.T.S. Leong, S.K. Woung, L. Eng, M. Jiwan, F.R. Kundat, Z.F.B.A.Aziz, O.H. Ahmed and N.M. Majid,2012. Isolation of indigenous strains of of *Paecilomyces lilacinus* with antagonistic activity against *Meloidogyne incognita*. *International Journal of Agriculture and Biology.*, 14: 197-203.
- Peng, Y. and Moens, M. (2002b). Tolerance of Rosa rootstocks and species to Pratylenchus penetrans. *Nematology* 4, 395-401
- Peregrine, W.T.H. and J. Bridge. (1992). The lesion nematode *Pratylenchus goodeyi*, an important pest of Enset in Ethiopia. Tropical Pest Management, 38:325-326.
- Quénéhervé, P., (2009). Integrated management of banana nematodes. In: Ciancio, A., Mukerji, K.G. (Eds.), Integrated Management in Fruit Crops and Forest Nematodes. Springer, Netherlands, 3–61.
- Ramana, K. V. and Eapen, J.S. (2000). Nematodes induced Disease of black pepper. In. Black Pepper Piper nigrum,ed. P. Ravindran, 269–295. Malaysia: Harwood academic publishers.
- Richards, L.A. 1954. Diagnosis and Improvement of Saline and Alkali Soils.
- Robinson E. 2005. Soil type guides VR nematodes applications. Farm Press, 1-2.
- Roberts P. A. and Van Gundy S. D.(1981). The Development and Influence of Meloidogyne incognito and M. javanica on Wheat. *Journal of Nematology*, Pp 346-352
- Ross, J.P. 1965. Predisposition of soybean to Fusarium wilt by Heterodera glycines and Meloidogyne incognita. *Phytopathology* 55: 361-364.

- Sarah, J. L. 1989. Banana nematodes and their control in Africa. *Nematropica* 19:199-216.
- Sarah, J.-L., Lassoudière, A. and Guérout, R. (1983). La jachère nue et l'immersion du sol: deux méthodes interessantes de lutte intégrée contre *Radopholus similis* (Cobb) dans les bananeraies de sols tourbeux de Côte d'Ivoire. *Fruits* 38, 35-42.
- Sasser J.N. and Freckman D.W. 1987. A world perspective on nematology: the role of the society. In: Veech J.A. and Dickerson D.W. (eds), Vistas on nematology. Society of Nematologists, Hyattsville, Maryland: 7-14.
- Seinhorst, J.W. (1965). The relation between nematode density and damage to plants. *Nematologica* 11, 137-154.
- Shaner, G., Stromberg, E.L. and Lacy, G.H. (1992). Nomenclature and concepts of pathogenicity and virulence. *Annual Review of Phytopathology* 30, 47-66.
- Sherbakoff, C.D. and Stanley, W.W. (1943). The more important diseases and insect pests of crops in Tennessee. *Tennessee Agriculture Experiment Station Bulletin* 186, 1-142.
- Sher, S.A. and Allen, M.W. (1953). Revision of the genus *PRATYLENCHUS* (Nematoda: Tylenchidae). University of California Publications in Zoology 57, 441-447.
- Sidam, A.K. and Bilal Mat, M. Z. (1983). Plant-parasitic nematodes in *Musa sapientum* in Peninsular Malaysia. MARDI Report 85: 1-7.
- Siddiqi M.R. (1972). *Pratylenchus coffeae*. C.I.H. Descriptions of Plant-parasitic Nematodes. Set 1, No. 6, CAB International, Wallingford, UK, 3.
- Siddiqi, M.R. (2000). Tylenchida parasites of plants and insects, 2nd edition. , UK, CABI Publishing, 833 pp.
- Sikora, R. A. Bridge, J and Starr, J. L. (2005). Overview of Integrated Nematode Management Technologies in Plant-parasitic nematodes in subtropical and tropical agriculture, 2ndedn. CAB International, Wallingford, UK..
- Speijer, P. R., C. S. Gold, E. B. Karamura, And I. N. Kashaija. (1994). Banana weevil and nematode distribution pattern in highland banana systems in Uganda: Preliminary results of a diagnostic survey. Pp. 285-289
- Speijer P. R. and D. De Waele. (1997). Screening of Musa germplasm for resistance and tolerance to nematodes. INIBAP Technical Guidelines 1.INIBAP, Montpellier, France, 42.
- Starr, J.L and Roberts P.A (2004) Resistance to plant parasitic nematodes. In: Chen, Z..X., Chen, S.Y & Dickson, D.W. (Eds). Nematology-Nematode management and utilization. Vol, 2. CABI Publishing, 879-901.

- Stekhoven Jr, J.H. (1941). *A manual of agricultural helminthology*. Leiden, The Netherlands, E.J. Brill, 878.
- Stirling, G.R. (1991). *Biological control of plant parasitic nematodes*. Brisbane, Australia, CAB International, 282.
- Sundararaju, P. and Jeyabaskaran, K.J. (2003). Evaluation of different soil types on multiplication of *PRATYLENCHUS coffeae* and growth of banana seedlings var, Nendran. *Nematologia Mediterranea* 31, 151-153.
- TengkuAb.Malik, T.M., Mohamad Roff, M.N., Rozeita, L. and Maimun, T. (2011), status of distribution, damage and mitigation of bacterial wilt in Malaysia. Malaysian Agricultural Research and Development Institute (MARDI)
- Tenkouano, A., S. Hauser, D.L. Coyne, and O. Coulibaly. (2006). Clean Planting Materials and Gnanapragasam, N.C. (1997). Plant derivatives in managing nematodes attacking tea in Sri Lanka. *International Journal of Nematology* 7, 111-112.
- Thoden TC, Korthals GW, Visser J, Van Gastel-Topper W. (2012). A field study on the host status of different crops for Meloidogyne minor and its damage potential on potatoes. *Nematology* 14, 277-284.
- Tixier, P., Risède, J.M., Dorel, M. & Malézieux, E. (2006). Modelling population dynamics of banana plant-parasitic nematodes: A contribution to the design of sustainable cropping systems. *Ecological Modelling* 198, 321-331.
- Townshend, J.L. and Webber, L.R. (1971). Movement of *Pratylenchus* penetransand the moisture characteristics of three Ontario soils. *Nematologica1* 7, 47-57.
- Townshend, J.L. (1973). Survival of Pratylenchus *penetrans* and *P. minyus* in two Ontario soils. *Nematologica* 19, 35-42
- Triantaphyllou, A.C. (1987). Genetics of nematode parasitism of plants. In: Veech, A. & Dickson, D.W. (Eds). *Vistas on nematology*. Hyattsville, MD, USA, Society of Nematologists, pp. 354-371.
- Turner, D.R. and Chapman, R.A. (1972). Infection of seedlings of alfalfa and red clover by concomitant populations of *Meloidogyne incognita* and *PRATYLENCHUS penetrans. Journal of Nematology* 4, 280-286
- Udo, I.A. and K.I. Ugwuoke, 2010. Pathogenicity of Meloidogyne incognita race 1 on turmeric (Curcuma longa L.) as influenced by inoculum density and poultry manure amendment. *Plant Pathology*. J., 9: 162-168
- Van Den Bergh, I., Nguyet, D.T.M., Tuyet, N.T., Nhi, H.H. & De Waele, D. (2006a). Influence of *PRATYLENCHUS coffeae* and *Meloidogyne* spp. on plant growth and yield of banana (*Musa* spp.) in Vietnam. *Nematology* 8, 265-271.

- Vanstone, V.A. and Russ, M.H. (2001a). Ability of weeds to host the root lesion nematodes *Pratylenchus neglectus* and *P. thornei* I. Grass weeds. *Australasian Plant Pathology* 30, 245-250.
- Vanstone, V.A. & Russ, M.H. (2001b). Ability of weeds to host the root lesion nematodes *PRATYLENCHUS neglectus* and *P. thornei* II. Broad-leaf weeds. *Australasian Plant Pathology* 30, 251-258.
- Walkley, A.; Black, I.A. 1934. An examination of Degtjareff method for determining soil organic matter, and proposed modification of the chromic acid tritation method. *Soil Science* 37:29-38.
- Walker GE. (2004). Effects of *Meloidogyne javanica* and organic amendments, inorganic fertilisers and nematicides on carrot growth and nematode abundance. *Nematologia Mediterranea* 32, 181-188.
- Wallace, H.R. (1973). Nematode ecology and plant disease. London, UK, Edward Arnold, 228.
- Winoto Suatmadji, R. (1988). *Pratylenchus penetrans* and *Rotylenchus robustus* on thirty herbaceous ornamental species. *Australasian Plant Pathology* 17, 97-98.
- Williams, B. D.; Schrank, B.; Huynh, C.; Shownkeen, R.; Waterston, R. H. (1994. A Genetic mapping system in Caenorhabditis elegans based on polymorphic sequence-tagged sites. *Genetics*, v. 131, p. 609-624.
- Willis, C.B. (1972). Effects of soil pH on reproduction of *Pratylenchus penetrans* and forage yield of alfalfa. *Journal of Nematology* 4, 291-295.
- Whitehead, A.G. and Hemming, J.R. (1965). A comparison of some quantitative methods of extracting small vermiform nematodes from soil. *Annals of Applied Biology*, 55: 25-38.
- Wuyts N, De Waele D, Swennen R (2006). Effects of plant phenylpropanoid pathway products and selected terpenoids and alkaloids on the behavior of the plant-parasitic nematodes Radopholus similis, *Pratylenchus* penetrans and Meloidogyne incognita. *Nematology* 8(1):89–101.
- Zimmermann, A.W.P. (1898). De nematoden der koffiewortels. Deel I. Mededelingen uit 's Lands Plantentuin 27, 1-64.