

## **UNIVERSITI PUTRA MALAYSIA**

TOXICITY EVALAUTIONS OF ETHANOLIC EXTRACT OF Christia vespertilionis (L. F.) BAKH. F. IN MALE SPRAGUE DAWLEY RATS

NURUL SYAHIRAH BINTI AHMAD SAYUTI

FPV 2018 15



# **TOXICITY EVALAUTIONS OF ETHANOLIC EXTRACT OF** *Christia vespertilionis* (L. F.) BAKH. F. IN MALE SPRAGUE DAWLEY RATS



## NURUL SYAHIRAH BINTI AHMAD SAYUTI

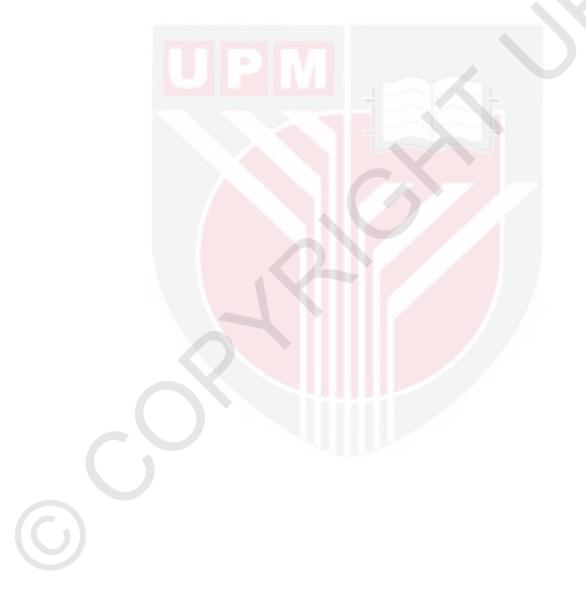
Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Science

January 2018

## COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright of Universiti Putra Malaysia unless other stated. Use may be made of any material contained within the thesis for non-commercial proposes from the copyright holder. Commercial use of material may only be made with the express, prior written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

## TOXICITY EVALUATIONS OF ETHANOLIC EXTRACT OF *Christia vespertilionis* (L.F.) BAKH. F. IN MALE SPRAGUE DAWLEY RATS

By

#### NURUL SYAHIRAH BINTI AHMAD SAYUTI

#### January 2018

## Chairman: Associate Professor Hazilawati binti Hamzah, PhDFaculty: Veterinary Medicine

The term Butterfly tea refers to the decoction of *Christia vespertilionis* (L.f.) Bakh. f. leaves which is widely consumed by cancer patients throughout Malaysia, and it has gained a huge popularity among researchers yearning to discover the real potential of this plant. Herein, this study is aimed at evaluating possible toxicity in a 14-day acute, 28-day subacute and 90-day subchronic oral toxicity of the ethanolic extract Christia vespertilionis (L.f.) Bakh. f. in male Sprague Dawley rats. The 14-day acute toxicity study was conducted to detect lethal dose 50 (LD<sub>50</sub>) Christia vespertilionis (L.f.) Bakh. f. while the 28-day subacute and 90-day subchronic toxicity studies are to detect the non-observed-adverse-effect level (NOAEL). In the acute toxicity study, rats were divided into control, 5% DMSO (vehicle) and 2000 mg/kg groups. The extract was administered orally on day 1 and observed for 14 days. Meanwhile, in the subacute and subchronic toxicity studies, a total of 30 rats were divided into control, 5% DMSO (vehicle), low dose (75 mg/kg), medium dose (125 mg/kg) and high dose (250 mg/kg) groups. The extract was administered daily from day 1 until day 28 for subacute and day 90 for subchronic. Standard toxicology parameters including mortality, behavioural changes, motor-neuronal abnormalities, feed-water consumption pattern and body weight were measured. The haematological, serum biochemical parameters and histopathological assessment of kidney and liver functions were also carried out. Results of acute oral toxicity showed that single dose (2000 mg/kg) of ethanol extract of Christia vespertilionis (L.f.) Bakh. f. leaves induced no treatment-related signs of toxicity or mortality in male Sprague Dawley rats. The haematological results also showed no changes in the control and treated groups in all 3 studies. However, serum biochemistry results for acute study, showed a significant decrease in the CK and AST level when compared with the control and treated groups. Meanwhile results for serum biochemistry in subacute and subchronic showed no changes in the control and treated groups for both studies. Organs to body weights ratio after euthanisation in all 3 studies showed no significant differences when comparing treated and control groups. On histopathological analysis, acute study showed significant differences (p<0.05) of



lesions observed on hepatic necrosis (mild to moderate) and degeneration (very mild) in the treated group (2000 mg/kg). Meanwhile, subacute study showed significant differences (p<0.05) of lesions observed on high dose, medium dose and low dose groups has mild to moderate, mild and very mild lesion of hepatic necrosis and very mild hepatic degeneration and hepatitis were scored in all three groups in subacute study. Besides, for subchronic study showed significantly differences (p<0.05) in hepatic necrosis and activated kupffer cells. Hepatic necrosis was observed mild to moderate in both high dose and medium dose groups, while low dose group only had mild lesion in subchronic study. On the other hand, the number of activated kuffer cells was significantly (p<0.05) higher in low and medium dose groups compared to the high dose group. On the other hand, all three studies, there were no significant (p>0.05) on lesion for renal toxicity were scored.

In conclusion, for the acute toxicity result, lethal dose 50  $(LD_{50})$  of *Christia vespertilionis* (L.f.) Bakh. f. is greater than 2000 mg/kg and both subacute and subchronic study showed induces dose-dependent oral hepatotoxicity in rats. As hepatic necrosis was predominantly seen compared to hepatic necrosis and hepatic degeneration in subacute toxicity study, it is suggested that subchronic toxicity study of *Christia vespertilionis* (L.f.) Bakh. f. extract induces more permanent damage to the hepatocytes.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

### ANALISIS TOKSISITI DARIPADA EKSTRAK ETANOL DAUN Christia vespertilionis (L.F.) BAKH. F. PADA TIKUS JANTAN SPRAGUE DAWLEY

Oleh

#### NURUL SYAHIRAH BINTI AHMAD SAYUTI

#### Januari 2018

## Pengerusi:Profesor MadyaHazilawati binti Hamzah, PhDFakulti:Perubatan Veterinar

Teh Rerama merujuk kepada rebusan daun Christia vespertilionis (L.f.) Bakh. f. yang di konsumi oleh pesakit kanser di Malaysia dan telah menjadi popular di Malaysia bukan sahaja dikalangan pesakit kanser tetapi juga kepada penyelidik untuk mengkaji pontensi sebenar pokok ini. Bagi kajian ini, tujuan kajian ini adalah untuk menilai aktiviti toksisiti 14-hari akut, 28-hari subakut dan 90-hari subkronik ekstrak etanol Christia vespertilionis (L.f.) Bakh. f. ke atas tikus jantan spesis Sprague Dawley. Akut 14-hari aktiviti toksisiti dijalankan untuk mengenalpasti dos Christia vespertilionis (L.f.) Bakh. f. yang menyebabkan 50% kematian (LD<sub>50</sub>) dan diikuti kajian 28-hari subakut dan subkronik toksisiti untuk mengenalpasti dos yang tidak menyebabkan kesan sampingan (NOAEL). Untuk kajian toksisiti akut, tikus dibahagikan kepada kumpulan kawalan normal, 5% DMSO (transportasi) dan kumpulan dos 2000mg/kg. Ekstrak diberi secara oral pada hari pertama (1) dan diperhati selama 14 hari. Manakala, untuk subakut dan subkronik kajian toksisiti, sebanyak 30 ekor tikus dibahagikan kepada kumpulan kawalan normal, 5% DMSO (transporatasi), dos rendah (75 mg/kg), dos sederhana (125 mg/kg) dan kumpulan dos tinggi (250 mg/kg). Ekstrak diberikan setiap hari selama 28 hari subakut dan 90 hari subkronik. Aktiviti toksisiti dinilai melalui parameter-parameter seperti kadar kematian, perubahan perangai, ketidaknormalan motor dan neuron, berat badan dan polar makanan dan air yang dikonsumi. Selain itu, analisis darah, serum biokimia dan histopatologi buah pinggang dan hati telah dijalankan. Keputusan untuk kajian akut oral toksisiti menunjukkan satu dos 2000mg/kg etanol ekstrak daun Christia vespertilionis (L.f.) Bakh. f. tidak menyebabkan kematian terhadap tikus jantan spesis Sprague Dawley. Keputusan analisis darah untuk ketiga-tiga kajian juga tidak menunjukkan perubahan yang ketara di antara kumpulan kawalan normal dan kumpulan yang diberi ekstrak. Keputusan analisis serum biokimia dalam kajian akut toksisiti menunjukkan paras CK dan AST signifikan (p<0.05) bila dibandingkan dengan kumpulan kawalan normal dan kumpulan yang diberikan ekstrak, walaubagaimanapun, keputusan analisis serum biokimia untuk subakut dan subkronik menunjukkan tiada perubahan di antara



kumpulan kawalan normal dan kumpulan yang diberikan ekstrak untuk kedua-dua kajian. Kadar perubahan berat organ ratio kepada berat badan untuk ketiga-tiga kajian juga menunjukkan tiada signifikan perubahan di antara kumpulan kawalan normal dan kumpulan yang diberikan ekstrak. Histopatologi analisis, kajian akut toksisiti menunjukkan signifikan perubahan skor nekrosis sel hepar (sedikit ke sederhana) dan degenerasi (sangat sedikit) dalam kumpulan dos 2000mg/kg. manakala kajian subakut menunjukkan signifikasi perubahan skor (p<0.05) pada kumpulan dos tinggi, dos sederhana dan dos rendah adalah sedikit ke sederhana, sedikit dan sangat sedikit pada perubahan skor nekrosis sel hepar dan skor degenerasi dan hepatitis sangat sedikit untuk ketiga-tiga dos. Disamping itu, kajian subkronik menunjukkan perubahan signifikan (p<0.05) pada skor nekrosis sel hepar dan bilangan sel kuffer yang aktif. Nekrosis sel hepar dinilai sedikit ke sederhana untuk kedua-dua dos tinggi dan dos sederhana, manakala untuk dos rendah menunjukkan sedikit perubahan skor sahaja. Bilangan sel kuffer yang aktif adalah signifikan tinggi dalam dos rendah dan dos sederhana jika dibandingkan dengan dos tinggi. Sebaliknya, ketiga-tiga kajian menunjukkan tiada signifikan (p>0.05) perubahan skor pada toksisiti buah pinggang.

Secara kesimpulannya, merujuk kepada keputusan kajian akut, dos yang menyebabkan kematian 50 peratus  $(LD_{50})$  ekstrak *Christia vespertilionis* (L.f.) Bakh. f. adalah lebih tinggi daripada 2000 mg/kg dan kedua-dua kajian subakut dan subkronik menunjukkan kebergantungan dos ke atas hepatotoksisiti tikus. Nekrosis sel hepar sebahagian besarnya dilihat dalam kajian subkronik jika dibandingkan dengan nekrosis sel hepar dan degenrasi sel hepar dalam kajian subakut, ia menunjukkan kajian subkronik toksisiti ekstrak *Christia vespertilionis* (L.f.) Bakh. f. menyebabkan lebih kerosokan kekal kepada sel hepar.

#### ACKNOWLEDGEMENTS

All praises to the Almighty Allah, the most Gracious and Merciful, Who is omnipotent and all giving, for affording me the strength and determination to complete this study. I would like to express my gratitude to my supervisor Assoc. Prof. Dr. Hazilawati Hamzah for her guidance, continued support and encouragement throughout this work. I am particularly grateful for my co-supervisors Prof. Dr. Noordin Mohamed Mustapha, Assoc. Prof Dr. Norhaizan Md. Esa, Dr. Shanmugavelu Sithambaram and Dr. Mohd. Rosly Shaari, for providing me the needed support, good comments and invaluable suggestions.

I wish to express my thanks to the Ministry of Higher Education (MOHE), Malaysia Agricultural Research and Development Institute (MARDI) and University Putra Malaysia for the scholarship, funding and allowance for supporting this work. I wish to mention and thank: Animal house unit staff (AMTREC) and all the staff in the Department of Veterinary Pathology & Microbiology, Faculty of Veterinary Medicine.

Acknowledgments are gratefully made to the following people: Farhan, Nadia, Farah, Nabila, Amira and other research mates for their contribution in varying degrees of this work.

I wish to express my thanks for my parents and family who always pray, support and encourage me both mentally and physically. Finally, I wish to acknowledge with thanks all those who, cooperated with me during this work, in all laboratory work and who read, reviewed and offered numerous helpful suggestions and corrections

The thesis was submitted to the senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

#### Hazilawati Hamzah, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

## Noordin Mohamed Mustapha, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

### Norhaizan Md. Esa, PhD

Associate Professor Faculty of Medicine and Health Science Universiti Putra Malaysia (Member)

## Shanmugavelu A/L Sithambaram, PhD

Director Strategic Livestock Research Centre Malaysia Agriculture Research and Development (Member)

## **ROBIAH BINTI YUNUS,PhD**

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

## **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

| Si           | gnature: |  |
|--------------|----------|--|
| $\mathbf{D}$ | gnature. |  |

Date:

Name and Matric No.: Nurul Syahirah Binti Ahmad Sayuti, GS44959

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

| Signature:<br>Name of<br>Chairman of<br>Supervisory<br>Committee: | Associate Professor Hazilawati Hamzah            |
|---|--|
| Signature:  |  |
| Name of<br>Member of<br>Supervisory<br>Committee:                 | Professor Noordin Mohamed Mustapha               |
| committee.  | <u>Trocssor</u> <u>Noordin Monaned Mustaplia</u> |
| Signature:<br>Name of<br>Member of                                |  |
| Supervisory<br>Committee:   | Associate Professor Norhaizan Md. Esa            |
| Signature:  |  |
| Name of<br>Member of  |  |
| Supervisory   |  |

## **TABLE OF CONTENTS**

ABSTRACT

ACKNOWLEDGEMENTS

ABSTRAK

| DEC<br>LIS<br>LIS | ROVAL<br>CLARATION<br>F OF TABLES<br>F OF FIGURES<br>F OF ABBREVIATION | vi<br>viii<br>xii<br>xv<br>xviii |
|-------------------|--|----------------------------------|
| СНА               | PTERUPIN   |                                  |
| 1                 | INTRODUCTION   | 1                                |
| -                 | 1.1 Background   | 1                                |
|                   | 1.2 Problem statement  |                                  |
|                   | 1.3 Hypotheses   | 2<br>2<br>3                      |
|                   | 1.4 Objective  | 3                                |
|                   |  |                                  |
| 2                 | LITERATURE REVIEWS   | 4                                |
|                   | 2.1 Toxicity study   | 4                                |
|                   | 2.2 Acute toxicity study   |                                  |
|                   | 2.3 Repeated dose studies  | 4<br>5<br>5                      |
|                   | 2.3.1 Subacute toxicity study  | 5                                |
|                   | 2.3.2 Subchronic toxicity study  | 6                                |
|                   | 2.4 Herbal toxicity  | 6                                |
|                   | 2.5 Christia vespertilionis (L.f.) Bakh. f.                            | 6<br>7                           |
|                   | 2.6 Solvent for extraction   | 9                                |
|                   | 2.7 Laboratory test animals  | 10                               |
|                   | 2.8 Vehicle as solvent   | 11                               |
|                   | 2.9 Dose level selection for toxicity studies                          | 12                               |
|                   | 2.10 Routes of administration used in toxicity studies                 | 12                               |
|                   | 2.11 Toxicology evaluations  | 13                               |
|                   | 2.11.1 Weekly body weight  | 13                               |
|                   | 2.11.2 Organ weight  | 13                               |
|                   | 2.11.3 Haematological and serum biochemical analyses                   | 14                               |
|                   | 2.11.4 Histopathology analyses   | 15                               |
| 3                 | METHODOLOGY  | 17                               |
|                   | 3.1 Plant material   | 17                               |
|                   | 3.2 Location of study  | 17                               |
|                   | 3.3 Animal management  | 17                               |

i iii

v

|                          | 3.4   | Acute, subacute and subchronic oral toxicity studies           | 18  |
|--------------------------|-------|--|-----|
|                          | 3.5   | Haematology and serum biochemistry analyses                    | 18  |
|                          | 3.6   | Histopathology examination                                     | 19  |
|                          | 3.7   | Statistical analyses   | 20  |
| 4                        | RESU  | LTS  | 21  |
|                          | 4.1   | Cage side observation on behavioural changes                   | 21  |
|                          | 4.2   | Body weight  | 21  |
|                          | 4.3   | Haematology results of acute, subacute and subchronic toxicity |     |
|                          |       | studies  | 23  |
|                          | 4.4   | Serum biochemistry results                                     | 27  |
|                          | 4.5   | Organ relative weight  | 31  |
|                          | 4.6   | Gross and histopathology findings                              | 34  |
| 5                        | DISCU | USSION   | 59  |
| 6                        | CONC  | CLUSION  | 65  |
| REFE                     | RENC  | ES   | 67  |
| APPE                     | NDICE |  | 79  |
|                          |       |  | 111 |
| LIST OF PUBLICATIONS 112 |       |  | 112 |
|                          |       |  |     |

 $\bigcirc$ 

## LIST OF TABLES

| Table |  | Page |
|-------|--|------|
| 1     | Acute toxicity study   | 18   |
| 2     | Subacute and subchronic toxicity study   | 18   |
| 3     | Lesions score and percentage   | 19   |
| 4     | Behavioural changes in acute, subacute and subchronic toxicity studies   | 21   |
| 5     | Body weights (Mean ±SEM) of male Sprague Dawley rats that received single dose (2000mg/kg) of ethanol extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f. leaf by oral gavage for 14-day duration  |      |
| 6     | Body weights (Mean ±SEM) of male Sprague Dawley rats that received repeated doses of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f. leaves by oral gavage for 28-day duration       |      |
| 7     | Body weights (Mean ±SEM) of male Sprague Dawley rats that received<br>repeated doses of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh.<br>f. leaves by oral gavage for 90-day duration |      |
| 8     | The haematology values (mean±SEM) of Sprague Dawley rats in acute oral toxicity study of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.   |      |
| 9     | The haematology values (mean±SEM) of Sprague Dawley rats in subacute oral toxicity study of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.  |      |
| 10    | The haematology values (mean±SEM) of Sprague Dawley rats in subchronic oral toxicity study of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.  |      |
| 11    | The serum biochemical parameters (mean±SEM) of Sprague Dawley rats in acute oral toxicity study of ethanolic extract of <i>Christian vespertilionis</i> (L.f.) Bakh. f.                                  |      |
| 12    | The serum biochemical parameters (mean±SEM) of Sprague Dawley rats in subacute oral toxicity study of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.                                |      |
| 13    | The serum biochemical parameters (mean±SEM) of Sprague Dawley rats in subchronic oral toxicity study of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.                              |      |

| 14     | The organs relative weight (mean±SEM) of Sprague Dawley rats in acute oral toxicity study of ethanol extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.   | 32 |
|--------|--|----|
| 15     | The organs relative weight (mean±SEM) of Sprague Dawley rats in subacute oral toxicity study of ethanol extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.  | 32 |
| 16     | The organs relative weight (mean±SEM) of SD rats in subchronic oral toxicity study of ethanol extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.  | 33 |
| 17     | Lesion scores of liver and kidneys of Sprague Dawley rats in acute oral toxicity study of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.  | 35 |
| 18     | Results of Man-Whitney test for comparisons between groups for the toxicity lesions in liver and kidneys of Sprague Dawley rats in acute oral toxicity of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.  | 36 |
| 19     | Lesion scores based on area affected in the liver of Sprague Dawley rats<br>in acute oral toxicity study of ethanolic extract of <i>Christia vespertilionis</i><br>(L.f.) Bakh. f.   | 37 |
| 20     | Results of Mann-Whitney test for comparison between groups based on<br>the area affected in Sprague Dawley rats of acute oral toxicity study of<br>ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.         | 37 |
| 21     | Lesion scores of liver and kidneys of Sprague Dawley rats in subacute oral toxicity study of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.   | 41 |
| 22     | Results of Man-Whitney test for comparisons between groups toxicity<br>lesions in liver and kidneys of Sprague Dawley rats in subacute oral<br>toxicity of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f. | 42 |
| 23     | Lesion scores based on area affected in the liver of Sprague Dawley rats<br>in subacute oral toxicity study of ethanolic extract of <i>M Christia</i><br><i>vespertilionis</i> (L.f.) Bakh. f.                                 | 43 |
| 24     | Results of Mann-Whitney test for comparison between groups based on<br>the area affected in Sprague Dawley rats of subacute oral toxicity study<br>of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.      | 44 |
| 25 : L | esion scores of liver and kidneys of Sprague Dawley rats in subchronic oral toxicity study of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f.  | 49 |

- 26 Results of Man-Whitney test for comparisons between groups for the toxicity lesions in liver and kidneys of Sprague Dawley rats in subchronic oral toxicity of ethanolic extract of *Christia vespertilionis* (L.f.) Bakh. f.
- 27 Lesion scores based on the area affected in the liver of Sprague Dawley rats in subchronic oral toxicity study of ethanolic extract of *Christia vespertilionis* (L.f.) Bakh. f.
- 28 Results of Mann-Whitney test for comparison between groups based on the area affected in Sprague Dawley rats in subchronic oral toxicity study of ethanolic extract of *Christia vespertilionis* (L.f.) Bakh. f.



52

51

50



## LIST OF FIGURES

| Figure |   | Page |
|--------|---|------|
| 1      | Christia vespertilionis (L.f.) Bakh. f. commonly known as the butterfly wing  | 8    |
| 2      | Photomicrograph of liver section of a control in acute toxicity study of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f. sacrified at the end of study period (H&E stain, x40)  | 38   |
| 3      | Photomicrograph of liver section of a rat in acute toxicity of a ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f. (2000mg/kg body weight) sacrified at the end of study period showing necrosis at periportal areas (encircled) H&E stain, x40   | 38   |
| 4      | Photomicrograph of liver section of a rat in acute toxicity study of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f. (2000 mg/kg body weight), showing necrosis at the periportal and centrilobular area, but midzonal area was not affected (H&E stain, x100)                              | 39   |
| 5      | Photomicrograph of liver section of a rat in acute toxicity study of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f. (2000 mg/kg body weight) showing atrophied and eosinophilic cytoplasm of hepatocyte, dense colour of nucleus (encircled) (H&E stain, x200)                             |      |
| 6      | Photomicrograph of liver section of a rat in acute toxicity study of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f. (2000 mg/kg body weight) showing sinusoid dilatation with atrophied hepatocytes and degeneration (cytoplasmic vacuolation) (arrow) (H&E stain, x200)                   | 40   |
| 7      | Photomicrograph of liver section of a rat in high dose group of subacute toxicity study of ethanolic extract of <i>Christia vespertilionis</i> (L.f.) Bakh. f. showing necrosis at periportal (1) toward midzonal (2) (H&E stain, x100)   | 45   |
| 8      | Photomicrograph of liver section of a rat in high dose group in subacute study toxicity <i>Christia vespertilionis</i> (L.f.) Bakh. f. sacrified at the end of study period showed regeneration (encircled) and karyorrlysis (arrow) at periportal area (H&E stain, x400)                                       | 45   |
| 9      | Photomicrograph of liver section of a rat in high dose group in subacute study toxicity <i>Christia vespertilionis</i> (L.f.) Bakh. f. sacrified at the end of study period showed shrunken size of hepatocyte (encircled) and presence of activated kupffer cells (arrow) at periportal area (H&E stain, x400) | 46   |
| 10     | Photomicrograph of liver section of a rat in medium dose group of subacute toxicity study of ethanolic extract of <i>Christia vespertilionis</i>  |      |

(L.f.) Bakh. f. showing atrophied of hepatocytes at periportal to midzonal (encircle), centrilobular area was not severed compare to periportal and midzonal (H&E stain, x200) 46 11 Photomicrograph of liver section of a rat in high dose group of subacute toxicity study of ethanolic extract of Christia vespertilionis (L.f.) Bakh. f. showing atrophied of hepatocytes at (encircleand karyorrlysis (arrow) at periportal area. (H&E stain, x200) 12 Photomicrograph of liver section of a rat in high dose group in subacute toxicity study of ethanolic extract of Christia vespertilionis (L.f.) Bakh. f. showing all 3 areas were necrosis (H&E stain, x200). 47 13 Photomicrograph of liver section of a rat in low dose group of subchronic toxicity study of ethanolic extract of Christia vespertilionis (L.f.) Bakh. f. showing inflammatory cells (encircled) at periportal and regeneration (arrow) at periportal-midzonal area (H&E stain, x200) 53 14 Photomicrograph of liver section of a rat in high dose group of subchronic toxicity study of ethanolic extract of Christia vespertilionis (L.f.) Bakh. f. sacrified showing atrophied of hepatocytes (encircle) and karyolysis (arrow) (H&E stain, x200) 53 15 Photomicrograph of liver section of a rat in low dose group in subchronic toxicity study of ethanolic extract of Christia vespertilionis (L.f.) Bakh. f. showing necrosis at periportal area to midzonal area (H&E stain, x100) 54 16 Photomicrograph of liver section of a rat in high dose group in subchronic toxicity study of ethanolic extract of Christia vespertilionis (L.f.) Bakh, f. showing necrosis at periportal to midzonal, centrilobular area was not severed compared to periportal and midzonal (H&E stain, x200) 54 Photomicrograph of liver section of a rat in high dose group in 17 subchronic toxicity study of ethanolic extract of Christia vespertilionis (L.f.) Bakh. f. showing necrosis at all 3 areas periportal to centrilobular (H&E stain, x100) 55 Photomicrograph of normal kidney section of a rat in acute study toxicity 18 Christia vespertilionis (L.f.) Bakh. f. (H&E stain, x100) 56 19 Photomicrograph of normal kidney section of a rat in acute study toxicity Christia vespertilionis (L.f.) Bakh. f. (H&E stain, x400) 56 20 Photomicrograph of normal kidney section of a rat in high dose subacute study toxicity Christia vespertilionis (L.f.) Bakh. f. (H&E stain, x200) 57

- 21 Photomicrograph of normal kidney section of a rat in medium dose subchronic study toxicity *Christia vespertilionis* (L.f.) Bakh. f. (H&E stain, x200)
- 22 Photomicrograph of normal kidney section of a rat in high dose subchronic study toxicity *Christia vespertilionis* (L.f.) Bakh. f. (H&E stain, x200)



58

## LIST OF ABBREVIATIONS

| ALP              | Alkaline phosphatase                       |
|------------------|--|
| ALT              | Alanine aminotransferase                   |
| AST              | Aspartate aminotransferase                 |
| СК               | Creatinine kinase                          |
| DMSO             | Dimethyl sulfoxide                         |
| DNA              | Deoxyribonucleic acid                      |
| GGT              | Gamma-glutamyl transferase                 |
| Hb               | Haemoglobin                                |
| IV               | Intravascular                              |
| LD <sub>50</sub> | Lethal dose                                |
| MCHC             | Mean corpuscular haemoglobin concentration |
| MCV              | Mean corpuscular volume                    |
| MTD              | Maximum tolerated dose                     |
| NOAEL            | Non-adverse-effect level                   |
| PCV              | Packed cell volume                         |
| RBC              | Red blood cell                             |
| WBC              | White blood cell                           |
|                  |  |

### **CHAPTER 1**

#### **INTRODUCTION**

#### 1.1 Background

Herbal remedies, particularly those used for therapeutic purposes, are widely used in many cultures for thousands of years. It is universally popular in primary healthcare, predominantly in developing countries such as Malaysia. The wide usage of these so-called "natural remedies" or "medicinal herbs" for self-medication is a result of the fact that the general public believes them to be safe and do not have any compromising health effects (Obici *et al.*, 2008). However, overtime there have been numerous warnings issued regarding the potential toxicity of these therapies, which further suggests the constant need for practitioners to keep abreast of reported incidence for renal and hepatic toxicity caused by ingestion of medicinal herbs and for investigational studies to be done on it.

In recent times, focus on plant research has increased all over the world and many evidences have been collected to show immense potential of medicinal plants used in various traditions. The wide and largely untapped field of traditional medicines still remains as an unique source for the discovery of bioactive compounds (Chen *et al.*, 2008; Kahumba *et al.*, 2014). Nearly, 80% of African and Asian population depends on traditional medicines for their primary healthcare (WHO, 2009; Karnataka Medicinal Plant Authority, 2009) and reported as high as 37% in Australia use herbal remedies as lifetime prevalence (Thomson *et al.*, 2012).

It is also estimated that about 25% of the drugs prescribed worldwide are derived from plants, with about 121 active compounds in use (Sahoo *et al.*, 2010). Between the years 2005 and 2007, 13 drugs derived from natural products were approved in the United States. More than 100 natural product-based drugs are in clinical studies (Li and Vederas 2009), and of the total 252 drugs in the World Health Organization's (WHO) essential medicine list, 11% are exclusively of plant origin (Sahoo *et al.* 2010). Besides, several clinical studies of traditional Chinese herbal medicines were undertaken, and others are still ongoing (Fu *et al.*, 2013 and Liu *et al.*, 2013).

Herbs and plants can be processed and be taken in many different forms and ways. These can be in the form of a whole herb, tea, syrup, essential oils, ointments, salves, rubs, capsules or even tablets that contains grounded or powdered form of the raw herb or its dried extract. Plants and herbs extract vary in the solvent based on its extraction, temperature extraction time, and inclusions of substances such as alcoholic extracts, vinegars extracts, hot water extract, long-term boiled extract roots or bark, and cold infusion of plants (macerates). There is no standardization, and components of an herbal extract or a product are likely to vary significantly between batches and producers (Sissi and Iris, 2011).

*Christia vespertilionis* (L.f.) Bakh. f. also known as the Rerama leaf has recently gained attention on its supposed potential to cure cancer. Various parts of this plant (mainly leaves) are widely used in traditional medicine for the treatment of numerous disorders. A decoction of the plant leaves is commonly used in remedies treating snake bites, tuberculosis, healing of bone fractures, bronchitis and cold and to increase blood circulation (Brach and Song, 2006), anti-plasmodium and high cytotoxicity against Hela and MRC54 (Nguyen *et al.*, 2007), inhibit neuroendocrine tumours (Hofer *et al.*, 2013) and inhibit growth of S180 tumour and H22 tumour cells (Wu *et al.*, 2012). Although several pharmacological studies (as antiplasmodial, anti-tumour) have been carried out on this plant, there is no experimental evidence on its toxicity. Hence, in this present study, the main aim is to evaluate its toxicity effects. This study was designed to investigate the toxicological assessment of acute, subacute and subchronic ethanolic leaf extract of *Christia vespertilionis* (L.f.) Bakh. f. on the blood, liver and kidney tissues of male Sprague Dawley rats.

### **1.2 Problem statement**

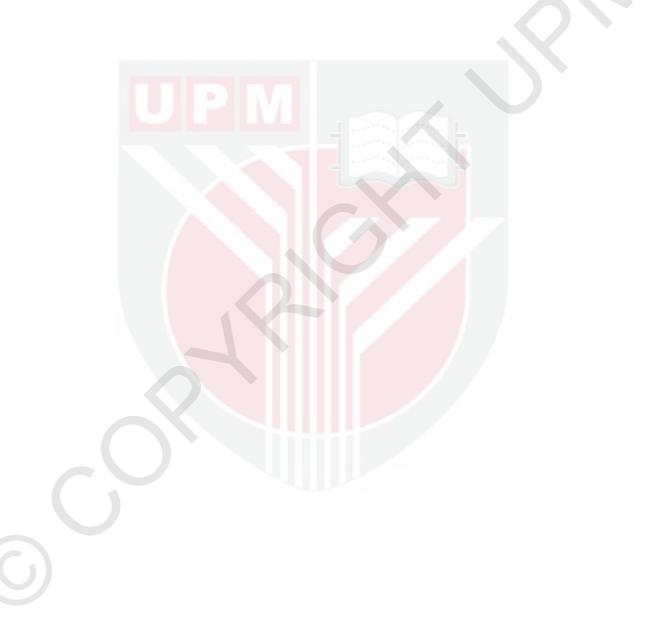
The increased interest recently in herbal medicines accelerated on the belief that these medicines are natural and have been traditionally used for centuries and are therefore assumed as safe and harmless. Nevertheless, their natural origin is not a guaranteed safety especially when there are risks associated with the use of herbal products that have been noted (Whiting *et al.*, 2002). Hence, gathering scientific information regarding the safety of consumption of this plants for use as an alternative and/or complementary medicine is very important before it is further developed into a new medicinal herbal therapy. In accord to this, a study was in need to be conducted to determine whether the use of *Christia vespertilionis* (L.f.) Bakh. f. as a plant base herbal drink that is widely distributed and consumed in Malaysia especially by cancer patients in the form of a tea bags, safe. At present time, there is no known published research on the toxicity study of this plant. Therefore, this study concentrates on the toxicity study for *M. christia verpertilionis* in male Sprague Dawley rats especially in determining the lethal dose (LD<sub>50</sub>) and the non-observe-effect level (NOEL) of the extract.

#### 1.3 Hypotheses

- a) Null hypotheses: The median lethal dose (LD50) of *Christia vespertilionis* (L.f.) Bakh. f. is lesser than 2000 mg/kg.
- b) Null hypotheses: NOAEL of *Christia vespertilionis* (L.f.) Bakh. f. for both subacute and subchronic toxicity studies is lesser than 250 mg/kg.

## 1.4 Objective

This study is conducted to evaluate acute oral toxicity study (14-day), subacute oral toxicity study (28-day) and subchronic oral toxicity study (90-day) of ethanolic extracts of *Christia vespertilionis* (L.f.) Bakh. f. in male Sprague Dawley rats.



#### REFERENCES

- Abdulrahman, F.I., Onyeyili, P.A., Sanni, S. and Ogugbuaja, V.O. (2007). Toxic effect of aqueous root-bark extract of Vitex doniana on liver and kidney functions. *International Journal of Biological Chemistry*, 1: 184-195.
- Adeneye, A.A., Ajagbonna, O.P., Adeleke, T.I and Bello, S.O. (2006). Preliminary toxicity and phytochemical studies of the stem bark aqueous extract of *Musangacecropioides* in rats. *Journal of Ethnopharmacology*, 105; 374-379.
- Ahmed, Y., Sohrab, H., Al-Reza, S.M., Shahidulla Tareq, F., Hasan, C.M., Sattar, M.A. (2010). Antimicrobial and cytotoxic constituents from leaves of Sapium baccatum. *Food Chemistry Toxicology*, 48, 549-552.
- Akanmu, M.A., Iwalewa, E.O., Elujoba, A.A. and Adelusola, K.A. (2004). <u>Toxicity</u> potentials of Cassia fistula fruits as laxative with reference to senna. <u>African</u> <u>Journal of Biomedic Research</u>, 7; 23-26.
- Alade, G.O., Akanmu, M.A., Obuotor, E.M., Osasan, S.A., Omobuwajo, O.R.(2009). <u>Acute and oral subacute toxicity of methanolic extract of *Bauhinia monandra* <u>leaf in rats. *African Journal of Pharmacy and Pharmacology*, 3:354-358.</u></u>
- Alam, N., Hossain, M.S., Khair, A., Amin, S.M., Khan, A. (2007). Comparative effects of mushroom on plasma lipid profile of hypercholestrolaemic rats. *Bangladesh Journal of Mushroom*, 1:15-22.
- Amresh, G.R., Singh, P.N. and Rao, C.V. (2008). Toxicological screening of traditional medicine Laghupatha (Cisampelos pareira) in experimental animals. Journal of Ethopharmacology, 116;454-460.
- Andrew, N. R. (2015). Ending the Use of Animals in Toxicity Testing and Risk Evaluation. Cambridge Quarterly of Healthcare Ethics, 24: 4; 448-458.
- Arslanyolu, M., Erdemgil, F. Z. (2006). Evaluation of the antibacterial activity and toxicity of isolated arctiin from the seeds of *Centaurea sclerolepis*. *Journal of Faculty Pharmacy*, 35;103-109
- Ashafa, A.O.T., Orekoya, L.O. and Yakubu, M.T. (2012). Toxicity profile ethanolic extract of *Azadirachta indica* stem bark in male Winstar rats. *Asian Pacific Journal of Tropical Biomedic*, 2(10); 811-817.
- Ashafa, A.T. and Olunu, O.O. (2011). Toxicological evaluation of ethanolic root extract of *Morinda lucida (L.) Benth. (Rubiaceae)* in male Wistar rats. *Journal of Natural pharmacology*, 2:108-114.
- Ashraf, Z., Aun, M., Muhammad, I., Ahmad, H. T.(2011). *In Vitro* antibacterial and antifungal activity of methanol, chloroform and aqueous extracts of *Origanum vulgare* and their comparative analysis. *International Journal of Organic Chemistry*, 1;257-261.

- Asyura, S.N.N., Hamzah, H., Shaari, R.M., Sithambaram, S. and Mustapha, N.M. (2016). Blood Profiles and Histopathological Changes of Liver and Kidney Tissues from Male Sprague Dawley Rats Treated with Ethanol Extracts of *Clinacanthus nutans* Leaf. *Journal of Clinical Toxicology*, 6:329.
- Balan, P., Han, K.S., Rutherfurd, S.M., Singh, H., and Moughan, P.J. (2009). Orally administered ovine serum immunoglobulins influence growth performance, organ weights and gut morphology in growing rats. *Journal of Nutrition*, 139; 244-249.
- Betancourt-Alonso, M.A., Orihuela, A., Aguirre, V., Vázquez, R., Flores-Pérez, I. (2012). Changes in behavioural and physiological parameters associated with Taenia pisiformis infection in rabbits (Oryctolagus cuniculus) that may improve early detection of sick rabbits. *World Rabbit Science*, 19:21-30.
- Bitsch, A., Jacobiah, S., Melbera, C., Wahnschaffea, U., Simetskaa, N. and Mangelsdorf, I. (2006). REPDOSE: A database on repeated dose toxicity studies of commercial chemicals—A multifunctional tool. *Regulatory Toxicology and Pharmacology*, 46(3); 202-210.
- Björnsson, E.S., Bergmann, O.M., Björnsson, H.K., Kvaran, R.B. and Olafsson, S. (2013). Incidence, presentation, and outcomes in patients with drug-induced liver injury in the general population of Iceland. *Gastroenterology*, 144;1419–1425.
- Bouwens, L., Baekeland, M., De Zanger, R. and Wisse, E.(1989). Quantitation, tissue distribution and proliferation kinetics of Kupffer cells in normal rat liver. *Hepatology*, 6:718–722.
- Brabb T. (2006). Body condition scoring in laboratory animal medicine. Presented at NWABR IACUC 101 Conference. 2 Mar 2006 Seattle, WA
- Brach AR, Song H. (2006). eFloras: New directions for online floras exemplified by the flora of china project. *Taxon*, 55(1); 188-192.
- Braeuning, A. and Schwarz, M. (2010). Zonation of heme synthesis enzymes in mouse liver and their regulation by β-catenin and Ha-ras. *Biology and Chemistry*, 391(11):1305-1313.
- Bruguera, M., Aranguibel, F., Ros, E. and Rodes, J (1978). Incidence and clinical significance of sinusoidal dilatation in liver biopsies. *Gastroenterology*, 75:474–478.
- Chan P.K, and Hayes A.W. (1994). Acute Toxicity and Eye Irritation In: Principles and Methods of Toxicology. 3rd Edition. A.W. Hayes, Editor. Raven Press, Ltd. New York, USA.
- Chen, S. T., Dou, J., Temple, R., Agarwal, R., Wu, K. M. and Walker, S. (2008), New therapies from old medicines. *National Biotechnology*, 26(10):1077-1083.

- Chan, P.K. and Hayes, A. W. (1989).Principles and Methods of Acute Toxicity and Eye Irritancy, Principles and Methods of Toxicology, Raven Press, Ltd, New York.
- Chattopadhyay, P., Agrawal, S.S. and Garg, A. (2006). Liver regenerative effect of *Phyllanthus amarus* Linn. against alcohol induced liver cell injury in partially hepatectomised albino rats. *International Journal of Pharmacology*, 2: 426-430.
- <u>Chellman, G.J., Bussiere, J.L., Makori, N., Martin, P.L., Ooshima, Y., Weinbauer, G.F.</u> (2009).Developmental and reproductive toxicology studies in nonhuman primates. *Birth Defects Research and Development of Reproductivity Toxicology*, 86(6); 446-462.
- Chitturi, S. and Farrell, G.C.(2008). Hepatotoxic slimming aids and other herbal hepatotoxins. *Journal of Gastroenterology and Hepatology*, 23:366–373.
- Claudio, P. And Alberta, A.S. (2006).Clinical chemistry and haematology historical data in control Sprague-Dawley rats from pre-clinical toxicity studies. *Experimental and Toxicologic Pathology*, 57(3); 213-219.
- Crook, M.A. (2006). Clinical Chemistry and Metabolic Medicine. 7<sup>th</sup> ed. London: Hodder Arnold.
- Cullen, J. M. (2005). Mechanistic classification of liver injury. *Toxicology and Pathology*, 33,6–8.
- Dahannukar, S. A., Kulkarni, R. A. and Rege, N.N. (2004). Pharmacology of medical plants and natural products. *Indian Journal of Pharmacology*, 32: S81-S118.
- de Arriba, S.G., Naser, B. and Nolte, K.U. (2013). Risk assessment of free hydroquinone derived from Arctostaphylos Uva-ursi folium herbal preparations. *International Journal of Toxicology*, 32(6); 442-453.
- Deng, X., Luyendyk, J.P., Ganey, P.E. and Roth, R.A. (2006). Inflammatory stress and idiosyncratic hepatotoxicity: hints from animal models. *Pharmacology reviews*, 61(3):262-282.
- Diallo, A., Gbeassor, M., Vonor, A., Eklu-Gadegbeku, K. and Aklikokou, A.(2008). Effect of *Tectona grandis* on phenylhydrazine-induced anaemia in rats. *Fitoterapia*, 79; 332-336.
- Djaldetti, M., Feller, N. (1978). Crystalline cytoplasmic inclusions in the liver cells of two mongrel dogs. *Research in Experimental Medicine*. 173(3):279–283.
- Dwivedi, S.K. and Dey, S. (2002). Medicinal herbs: a potential source of toxic metal exposure for man and animals in India. *Archives of Environmental Health*, 57(3): 229-231.

- Dybing, E., Doe, J., Groten, J., Kleiner, J. and O'Brien, J. (2002). Hazard charecterization of chemicals in food and diet: dose response, mechanism and extrapolation issues. *Food Chemical Toxicology*, 42:237-282.
- Eaton, K.A., Danon, S.J., Krakowka, S., Weisbrode, S.E. (2007). A reproducible scoring system for quantification of histologic lesions of inflammatory disease in mouse gastric epithelium. *Comparative Medicine*, 57(1):57-65.
- Eaton, D.L. and Klaasen, C.D. (1996).Principle of toxicology. The basic science of poisons, 5<sup>th</sup> ed McGrawHill; 13.
- Friedel, R., Bode, R. and Trautschold, I. (1976). Distribution of intravenously injected enzymes of heterologous, homologous and autologous origin. Distribution and transport of cell enzymes within the extracellular space. *Journal of Clinical Chemistry and Clinical Biochemistry*, 14:129–136.
- Fu, D.L., Lu, L., Zhu, W., Li, J.H., Li, H.Q., Liu, A.J., Xie, C. and Zheng, G.Q. (2013). Xiaoxuming decoction for acute ischemic stroke: a systematic review and meta-analysis. *Journal of Ethnopharmacology*, 148, 1-13.
- Gamaniel, K.S. (2000). Toxicity from medicinal plants and their products. *Nigerian Journal of Natural Products and Medicines*, 4: 4-8.
- Ghisalberti, E.L. (2007). Detection and isolation of bioactive natural products. CRC Press, Florida, 11-76.
- Greaves, P., Edwards, R., Cohen, G. M., MacFarlane, M. (2001). Diffuse hepatic apoptosis. *Toxicology and Pathology*, 29;398–400.
- Guengerich, F.P. and MacDonald, J.S. (2007). Applying mechanisms of chemical toxicity to predict drug safety. *Chemical Research and Toxicology*, 20:3; 344-369.
- Harizal, S.N., Mansor, S.M., Hasnan, J., Tharakan, J.K.J and Abdullah, J. (2010). Acute toxicity study of the standardized methanolic extract of *Mitragyna speciosa* Korth in rodent. *Journal of Ethnopharmacology*, 131; 404–409.
- Heymann, F., Trautwein, C., Tacke, F. (2009). Monocytes and macrophages as cellular targets in liver fibrosis. Inflammation Allergy Drug Targets, 8: 307-318.
  - Hickling, K. and Smith, D. (2000). The contribution of vehicles, rates of administration, and volumes to infusion studies. : Healing G, Smith D, editors. Handbook of preclinical intravenous infusion. New York (NY): Taylor & Francis.
- Hofer, D., Schwach, G., Tabrizi-Wizsy, N.G., Sadjak, A., Sturm, S., Stuppner, H., et al. (2013).Christia vespertilionis plant extracts as novel antiproliferative agent against human neuroendocrine tumour cells. *Oncology Reports*, 9(6):2219-2226.

- Holt, M.P., Cheng, L.L., Ju, C. (2008).Identification and characterization of infiltrating macrophages in acetaminophen-induced liver injury. *Journal of Leukocyte Biology*, 84 (6); 1410-1421.
- Hor, Y., Farsi. E., Lim,C.P.,Ahmad, M.,Asmawi,M.Z. and Yam, M.F. (2011). Acute and subchronic oral toxicity of *Coriolus versicolor* standardized water extract in Sprague-Dawley rats. *Journal of Ethnopharmacology*, 137; 1067–1076.
- ICH Guideline M3 (R2). (2009). Guidance on Nonclinical Safety Studies for the Conduct of Human Clinical Trials and Marketing Authorization for Pharmaceuticals.
- Irwin, S. (1968). Comprehensive observational assessment: A systematic, quantitative procedure for assessing the behavioral and physiologic state of the mouse. *Psychopharmacologia*, 3(3):222–257.
- Jain, A, Soni, M., Deb, L., Jain, A., Rout, S.P., Gupta, V.B. and Krishna, K.L. (2008). Antioxidant and hepatoprotective activity of ethanolic and aqueous extracts of Momordica dioica Roxb. Leaves. *Journal of Ethnopharmacology*, 115(1);62-66.
- Jaouad, E.H., Israilli, Z.H., Lyoussi, B. (2004). Acute toxicity and chronic toxicological studies of *Ajuga iva* in experimental animals. *Journal of Ethnopharmacology*, 91: 43-50.
- Jiro H. and Shinya S. (2012). Routes of Administration. The Laboratory Mouse (Second Edition), 2012; 709-725.
- Kage, M., Arakawa, M., Kojiro, M., and Okuda, K.(1992). Histopathology of membranous obstruction of the inferior vena cava in the Budd-Chiari syndrome. *Gastroenterology*, 102:2081–2090.
- Kahumba, J., Rasamiravaka, T., Okusa, P. N., Bakari, A., Bizumukama, L., Kalonji, J.-B., Kiendrebeogo, M., Rabemanantsoa, C., El Jaziri, M., Williamson, E. M. and Duez, P. (2014). Traditional African medicine: from ancestral knowledge to a modern integrated future, *Science*.
- Karnataka Medicinal Plants Authority. (2009). Authenticity, purity of herbal drugs critical for sustained growth in global markets: KMPA chief<u>www.</u> <u>Pharmabiz.com</u>.
- Karim C. El K., Aimee, L. A., Michael, W. D., Padade, M. V., Wujuan ,Z., Kenneth, D. R., Setchell, S. J. K nd Ronald ,J. S. (2013). Phytosterols Promote Liver Injury and Kupffer Cell Activation in Parenteral Nutrition–Associated Liver Disease. *Science Translational Medicine*, 5 (206);20-26.
- Keeffe, E.B., Sunderland, M.C., Gabourel, J.D. (1986). Serum γ-glutamyl transpeptidase activity in patients receiving chronic phenytoin therapy. *Digestive and Disease Science*, 31:1056–1061.

- Kennedy, D.O., Wightman, E.L. (2011). Herbal extracts and phytochemicals: plant secondary metabolites and the enhancement of human brain function. *Advances in Nutrition*, 2: 32-50.
- Koller, L.D. (1973). A note on eosinophilic cytoplasmic bodies in the liver of a rabbit. *Veterinary Pathology*. 10(4):295–298.
- Kullervo, H., Nathan, M.D., Greg, C., Ferenc, A. J., Eyal, Z., Ron, K., Douglas, R. (2006). Pre-clinical testing of a phased array ultrasound system for MRI-guided noninvasive surgery of the brain—A primate study. *European Journal of Radiology*, 59(2); 149-156.
- Jack, A.H., Roberts, W. and Laura, P.J.(2010). Mechanisms of Acetaminophen-Induced Liver Necrosis. *Handbook of Experimental Pharmacology*, 196;369-405.
- Laaksonen, K.S., Nevalainen, T.O., Haasio, K., Kasanen, I.H., Nieminen, P.A. and Voipio H.M. (2013). Food and water intake, growth, and adiposity of Sprague-Dawley rats with diet board for 24 months. *Laboratoty Animal*, 47(4); 245-256.
- Ladds, P.W., Strafuss, A.C. (1971). Eosinophilic cytoplasmic bodies in a bovine liver. *The Cornell Veterinarian*. 61(3):486–489.
- Lalisan, J.A., Nuñeza, O.M., Uy, M.M. (2014). Brine Shrimp (Artemia salina) Bioassay of the medicinal plant Pseudelephantopus spicatus from Iligan City, Philippines. *International Research Journal of Biological Science*, 3; 47-50.
- Larson, A.M., Polson, J., Fontana, R.J., Davern, T.J., Lalani, E., Hynan, L.S., Reisch, J.S., SchiØdt, F.V., Ostapowicz, G., Shakil, A.O. and Lee, W.M. (2005). Acute Liver Failure Study Group. Acetaminophen-induced acute liver failure: results of a United States multicenter, prospective study. *Hepatology*. 42:1364–1372
- Larrey, D. (2002). Epidemiology and individual susceptibility to adverse drug reactions affecting the liver. *Seminar Liver Disease*, 22(2):145-155.
- Leissing N, Izzo R, and Sargent H (1985). Variance estimates and individuality ratios of 25 serum constituents in beagles. *Clinical Chemistry*, 31:83-86.
- Lewinsohn, N., Dudai, Y.T. (1998). Histochemicallocalization of citral accumulation in lemongrass leaves (Cymbogoncitratus (DC.) Stapf. Poaceae). *Analysis of Botany*, 81(1); 35-39.
- Li, J. W. H. and Vederas, J. C. (2009). Drug discovery and natural products: End of an era or an endless frontier? *Science*, 325:161–165.
- Lipman, N.S. and Perkins, S.E. (2002) Factors that may influence animal research. *Laboratory Animal Medicine*, 2(29); 1143-1184.

- Lipnick, R.L., Cotruvo, J.A., Hill, R.N., Bruce, R.D., Stitzel, K.A., Walker, A.P., Chu,
   I., Goddard, M., Segal, L., Springer, J.A. and Myers, R.C. (1995).
   Comparison of the Up-and-Down, Conventional LD50, and Fixed-Dose
   Acute Toxicity Procedures. *Food and Chemical Toxicology*, 33, 223-231.
- Liu, Z.L., Xie, L.Z., Zhu, J., Li, G.Q., Grant, S.J. and Liu, J.P. (2013). Herbal medicines for fatty liver diseases. *Cochrane Database Systematic Reviews*, 24(8); CD009059.
- Lu, H., Meng, X., Li, C., Sang, S., Paten, C., Sheng, S., Hong, J., Bai, N., Winnik, B., Ho, C.T., Yang, C.S. (2003). Glucuronides of tea catechins: enzymology of biosynthesis and biological activities. American Society for Pharmacology and Experimental Therapeutics. *Drug Metabolism and Disposal*, 31(4): 452-461.
- Malomo, A.O.(2000). Toxicological implications of certriaxone administration in rats. *Nigerian Journal of Biochemistry and Molecular Biology*, 1533–1538.
- Mangelsdorf, I., Buschmann, J., and Orthen, B. (2003). Some aspects relating to the evaluation of the effects of chemicals on male fertility. Regulatory Toxicology and Pharmacology, 37; 356-369.
- Martins, A.C. (2006). Clinical chemistry and metabolic medicine. 7. UK: Edward Arnold Ltd, 7–15.
- Mendis, G.P., Gibberd, F.B., Hunt, H.A. (1993). Plasma activities of hepatic enzymes in patients on anticonvulsant therapy. *Seizure*, 2:319–323.
- Mishra, M. and Tandon, V.L. (2012).Haematological effects of aqueous extract of Ornamental plants in male Swiss albino mice. *Veterinary World*, 5(1):19-23.
- Miyaoka, Y. and Miyajima, A. (2013). To divide or not to divide: revisiting liver regeneration. Cell Division, **8** (8).
- Modesitt, S. C., and Parsons, S. J. (2010) *In vitro* and *in vivo* histone deacetylase inhibitor therapy with vorinostat and pacli- 12. *Gynecology and Oncology*, 119; 351–357.
- Mohamed, E.A.S., Lim, C.P., Ebrika, O.S., Asmawi, M.Z., Sadikun, A. and Yam, M.F. (2011). Toxicity evaluation of a standardised 50% ethanol extract of *Orthosiphon stamineus. Journal of Ethanopharmacology*, 133; 358-363.
- Mukinda, J.T. and Eagles, P.F. (2010). Acute and sub-chronic oral toxicity profile of the aqueous extract of *Polygala fruticose* in female mice and rats. *Journal of Ethanopharmacology*, 128:236-240.
- Myhre, M.J. (2000). <u>Herbal remedies, nephropathies, and renal disease</u>. *Nephrology* <u>Nursing Journal</u>, 27: 473-478.

- Naschitz, J.E., Slobodin, R.J., and Yeshurun, D. (2000). Heart diseases affecting the liver and liver diseases affecting the heart. *American Heart Journal*, 140:111–120.
- Natalie, C., Aaron, R., H., Lillian, L.and Albiruni, R. A.R. (2015). Early phase clinical trials to identify optimal dosing and safety. Molecular Oncology, 9(5): 997–1007
- Nayak, N.C., Sathar, S.A., Mughal, S., Duttagupta, S., Mathur, M. and Chopra, P.(1996). The nature and significance of liver cell vacuolation following hepatocellular injury--an analysis based on observations on rats rendered tolerant to hepatotoxic damage. *Virchows Archiv An International Journal of Pathology*, 428(6):353-65.
- Nguyen-Pouplin, J., Tran, H., Phan, T.A., Dolecek, C., Farrar, J., et al. (2007). Antimalarial and cytotoxic activities of ethnopharmacologically selected medicinal plants from South Vietnam. *Journal of Ethnopharmacology*, 109 (3):417-427.
- Nursyuhada, H., Hazilawati, H., Hutheyfa, A.H., Rosly, S.M., Shanmugavellu,S. Nordin, M.M. and Jasni, S. (2011). Detection of Bcl-2 in leukemia rats using an EvaGreen real-time RT-PCR assay. *Pertanika Journal of Tropical AgricultureScience*, 3(4); 372-380.
- Obici, S., Otobone, J.F., da Silva, V.R., Ishida, K., da Silva, J.C., Nakamura, C.V., Cortez D.A.G. and Audi, E.A. (2008). Preliminary toxicity study of dichloromethane extract of *Kielmeyera coriacea* stems in mice and rats. *Journal of Ethnopharmacology*, 115:131–139.
- Ogbonnia, S.O., Mbaka, G.O., Anyika, E.N., Osegbo, O.M. and Igbokwe, N.H. (2010). Evaluation of acute toxicity of hydro-ethanolic extract of *chromolaena* (*L.*) *king robinson (Fam. Asteracea)* in rats (2010). *Agricultural and Biology Journal of North America*, 1; 865-869.
- Ogugu, S.E., Kehinde, A.J., James, B.I., Paul, D.K. (2012). Assessment of cytotoxic effects of methanol extract of Calliandra portoricensis using Brine Shrimp (Artemia salina) Lethality Bioassay. *Global Journal of Bio-Science and Biotechnology*, 2, 257-260.
- Okokon, J.E., Iyadi, K.C., Effiong, C.O. (2004). Effect of sub chronic administration of ethanolic leaf extract of Croton zambesicus on hematological parameters of rats. *Nigerian Journal of Physiological Sciences*, 19(1-2): 10-13.
- Olorunnisola, O.S., Bradley, G. and Afolayan, A.F. (2012). Acute and sub-chronic toxicity studies of methanolic extract of Tulbaghia violacea rhizomes in Wistar rats". *African Journal of Biotechnology*, 11; 14934–14940.
- Organization for Economic Cooperation and Development (OECD) (2013). *Guidance Document on Developing and Assessing Adverse Outcome Pathways*. Series on Testing and Assessment No. 184.

- Organization for Economic Cooperation and Development (OECD) (2000). Guidance Document on Acute Oral Toxicity. Environmental Health and Safety Monograph Series on Testing and Assessment No.24.
- Organization for Economic Cooperation and Development (OECD) (2001). Guideline for testing of chemicals No. 408.
- Organization for Economic Cooperation and Development (OECD) (2008). Guidance Document on Guideline for Repeated dose-28 day Oral toxicity Study in rodents.
- Organization for Economic Cooperation and Development (OECD) (2001). Guidance Document for testing chemicals. Acute oral toxicity study-fixed dose.
- Parasuraman, S. (2011). Toxicological screening. *Journal of Pharmacology*, 2(2);74-79.
- Pari, L. and Murugan, P. (2004). Protective role of tetrahydrocurcumin against Erythromycin estolate-induced hepatotoxicity. *Pharmacology Research*, 49; 481-486.
- Pegg, R.B., Rybarczyk, A. and Amarowicz, R. (2008). <u>Chromatographic Separation</u> of Tannin Fractions from a Bearberry-leaf (Arctostaphylos Uva-ursi L. <u>Sprengel</u>). *Polish Journal of Food and Nutrition Sciences*, 58(4); 485–490
- Petterino, C. and Argentino-Storino, A. (2006). Clinical chemistry and haematology historical data in control Sprague Dawley rats from pre-clinical toxicity studies. *Experimental Toxicology Pathgology*, 57(3);213-219.
- Philip, A.B. (2002). Acute systemic toxicity. *Journal of Healthy Aging Research*, 43;27-30.
- Pimentel, M.A.B., Pizzolatti, M.G., Costa Brighente, I.M. (2002). An Application of the Brine Shrimp Bioassay for general screening of Brazilian medicinal plants. Acta Farmceutica Bonaerense, 21;175-178.
- Rasamiravaka, T., Jedrzejowski, A., Kiendrebeogo, M., Rajaonson, S., Randriamampionona, D., Rabemanantsoa, C., Andriantsimahavandy, A., Rasamindrakotroka, A., Duez, P., El, J. M. and Vandeputte, O. M., 2013, Endemic malagasy Dalbergia species inhibit quorum sensing in Pseudomonas aeruginosa PAO1, *Microbiology*, **159**(5):924-938.
- Rates, S.M.K. (2001). Herbal as source of drugs. Toxicology, 39; 603-613.
- Report of the FELASA working group on pain and distress.(1994). Laboratory Animals, 28, 97-112.
- Rhiouni, H.R., Nazari, P., Kamli-Najed, M. and Lyoussi, B. (2008). Acute and subchronic oral toxicity of an aqueous extract of leaves of *Herniaria glabra* in rodents. *Journal of Ethnopharmacology*, 118; 378-386.

- Rhiovania, H., El-Hilalya, J., Israili, Z.H., Lyoussia, B. (2008). Acute and subacute toxicity of an aqueous extract of the leaves of *Herniaria glabra* in rodents. Journal of Ethnopharmacology, 118:378-386.
- Rispin, A., Farrar, D., Margosches, E., Gupta, K., Stitzel, K., Carr, Greene, M. Mayer,
  W. And McCall, D. (2002). Alernative methods for the median lethal dose (LD50) test. Journal of Institute Animal Laboratory Research, 43(4);233-234.
- Robinson, S., Chapman, K., Hudson, S., Sparrow, S., Spencer-Briggs, D., Danks, A. et al. (2009).Guidance on dose level selection for regulatory general toxicology studies for pharmaceuticals. London, National Centre for the Replacement, Refinement and Reduction of Animals in Research Laboratory Animal Science Association (NC3Rs)/Laboratory Animal Science Association (LASA).
- Rosidah, Y., Sdikun, A., Ahmad, A., Akowuah, G.A. and Asmawi, M.Z. (2009). Toxicology evaluation of standardized methanol extract of *Gynura* procumbens, Journal of Ethnopharmacology, 123; 244-249.
- Sahoo, N., Manchikanti, P., Dey, S. (2010). Herbal drugs: Standards and regulation. *Fitoterapia*. 81(6):462–471.
- Samuel, N. (2005). Herbal remedies and anticoagulant therapy. *Tel Alvi Medical Center*, 93(1); 3-7.
- Sankpal, U. T., Abdelrahim, M., Connelly, S. F., Lee, C. M., Madero-Visbal, R., Colon, J., Smith, J., Safe, S., Maliakal, P., and Basha, R. (2012) Small molecule tolfenamic acid inhibits PC-3 cell proliferation and invasion in vitro and tumor growth in orthotopic mouse model for prostate cancer. *Prostate*, 72, 1648–1683.
- Sasidharan, S., Chen, Y., Saravanan, D., Sundram, M. and Yoga, L. (2011). Extraction, Isolation and Characterization of Bioactive Compounds from Plants' Extracts. *African Journal of Traditional and Complementary Alternative Medicine*, 8(1); 1–10.

Seeff, L.B.(2007). Herbal hepatotoxicity. Clinical Liver Diseases, 11:577-596.

- Simon B. I.A., Lidianys, M., Lewis-Luján, C. L., Lara-Espinoza, A. A., Gil-Salido, D., Fernandez-Angulo, J. L., Rubio-Pino, and David D. H. (2015). Solvent effects on phytochemical constituent profiles and antioxidant activities, using four different extraction formulations for analysis of *Bucida buceras L*. and *Phoradendron californicum*. <u>BMC Research Notes</u>, 8: 396.
- Siti, S. A., Norhaizan, M. E. and Hazilawati, H. (2014). Histopathologic Changes in Liver and Kidney Tissues from Male Sprague Dawley Rats Treated with Rhaphidophora Decursiva (Roxb.) Schott Extract. *Journal of Histology and Cytology*, S4: 001. doi:10.4172/2157-7099.S4-001.
- Sissi Wachtel-Galor and Iris F. F. B.(2011). Herbal Medicine: An Introduction to Its History, Usage, Regulation, Current Trends, and Research Needs.

- Smolensky, M.H. and Peppas, N.A. (2007). Chronobiology, drug delivery, and chronotherapeutics. *Advaced Drugs Delivery Review*, 59(9):828-851.
- Sokeng, D.S., Kamtchouing, P., Watcho, P., Jatsa, H.B., Moundipa, P.F., Ngounou, F.N., Lontsi, D. and Bopelet, M. (2001). Hypoglycémic activity of *Anacardium occidentale L.* Aqueous extract in normal and streptozotocininduced diabetic rats. *Diabetes Research and Clinical Practice*, 36: 001-009.
- Spurling, N.W. and Carey, P.F. (1992).Dose selection for toxicity studies: a protocol for determining the maximum repeatable dose. *Human Experimental Toxicology*, 11(6):449-457.
- Suresh, K.P. (2011). An overview of randomization techniques: An unbiased assessment of outcome in clinical research. *Journal of Human Reproductive Sciences*, 4(1): 8–11.
- Syed Asad, B., Iqbal, M.M., Kiranmai, M. and Ibrahim, M. (2012). Hepatoprotective Activity of Phyllanthus Amarus Seeds Extracts in CCl4 Treated Rats: In Vitro & In Vivo. *Global Journal of Medical Research*, 12 (6): 2249-4618.
- Tedong, L., Dimo, T., Dzeufiet, P.D.D., Asongalem, A.E., Sokeng, D. S., Callard, P., Flejou, J.F. and Kamtchouing, P. (2006). Antihyperglycemic and renal protective activities of *Anacardium occidentale (Anacardiaceae)* leaves in streptozotocin induced diabetic rats. *African Journal of Traditional Complementary and Alternative Medicine*, 3 (1): 23 – 35.
- Teo, S.D., Stirling, S., Thomas, A., Kiorpers, A. and Vikram, K.(2002). A 90-day oral gavage toxicity study of D-methylphenidate and D, L methylphedinate in Sprague Dawley rats. *Toxicology*, 179; 183-196.
- Timbermont, L., Lanckriet, A., Dewulf, J., Nollet, N., Schwarzer, K., Haesebrouck, F., Ducatelle, R., Van, I. F. (2010). Control of Clostridium perfringensinduced necrotic enteritis in broilers by target-released butyric acid, fatty acids and essential oils. *Avian Pathology*, 39(2);117-121.
- Thoolen, B., Maronpot, R.R., Harada, T., Nyska, A., Rousseaux, C., Nolte, T., Malarkey, D., Kaufmann, W., Kutter, K., Deschl, U., Nakae, D., Gregson, R., Winlove, M., Brix, A., Singl, B., Belpoggi, F. and Ward, J.M. (2010). Hepatobiliary lesion nomenclature and diagnostic criteria for lesions in rats and mice. *Toxicology and Pathology*, 38:5-81.
- Thomson, P., Jones, J., Evans, J., Leslie, S. J. (2012). Factors influencing the use of complementary and alternative medicine and whether patients inform their primary care physician. *Complementary Therapies in Medicine*, 20:45–53.
- Tolman, K.G. and Rej, R.(1999). Liver function. In: Tietz Text Book of Clinical Chemistry, Burtis, C.A. and E.R. Ashwood (Eds).3<sup>rd</sup> Edn., W.B. Saunders Co., Philadelphia, PA,USA.,1125-1177.
- Valla, D. C. (2002). Hepatic vein thrombosis (Budd-Chiari syndrome). *Seminar Liver Disease*, 22:5–14.

- Valatas, V., Kolios, G., Manousou, P., Xidakis, C., Notas, G., Ljumovic, D., Kouroumalis, E.A.(2004). Secretion of inflammatory mediators by isolated rat Kupffer cells: the effect of octreotide. *Regulatory Peptides*, 120:215–225.
- van Meeuwen, M.S., van Steenis, C.G.G.J., Stemmerik, J. (1961). Prelimnary revisions of some genera of Malaysian Papilionaceae II. Part 1.Herbarium Bogoriense.
- van Wijk H, Robb D. (2000). Femoral cannulation using the tail cuff model in the mouse, Healing G, Smith D, editors.Handbook of preclinical continuous intravenous infusion. New York (NY): Taylor and Francis, 61-70.
- Wall, M. E.(1998). Camptothecin and taxol: discovery to clinic. *Medicine Research Review*, **18** (5):299-314.
- Wang, D.,Zhong, K,Y,Z, Luo, X., Xiao,R., Hou,Y., Bao, W., Yang, W., Yan, H., Yao,P.and Liu,L. (2011). Acute and subchronic oral toxicities of Pu-erh black tea extract in Sprague-Dawley rats. *Journal of Ethnopharmacology*, 134; 156–164.
- Wherly P. H., Daniel, K. N. and Robert B. L. van Lier<sup>+</sup>.(2002). Analysis of Rodent Growth Data in Toxicology Studies. *Toxicology Science*, 66(2); 313-319.
- Whiting, P.W., Clouston, A. and Kerlin, P. (2002). Black cohosh and other herbal remedies associated with acute hepatitis. *Medical Journal of Australia*, 177:440–443.
- Winston D, Maimes S. (2007) Adaptogens: Herbs for strength, stamina and stress relief. Ro- chester, Vermont: Healing Arts Press.
- Wooley, A. (2003). A Guide to Practical Toxicology Evaluation, Prediction and Risk, Determination General and reproductive toxicology, Taylor and Francis, New York; 80–106.
- World Health Organization (2009). Traditional medicine <u>http://www.who.int/mediacentre/factsheets/fs134/en/</u>, Last accessed on 3rd September 2009.
- Wu Xiao-Yan, Tang Ai-Cun, Lu Qiu-Yu. (2012). Study on Antitumor Effect of the Extract from *Christia vespertilionis* in vivo. *Chinese Journal of Experimental Traditional Medical Formulae*, 8:2012.
- Xing, L and Remick, D.G (2005). . Mechanisms of Dimethyl Sulfoxide Augmentation of IL-1β Production. *The Journal of Immunology*,174: 6195-6202.
- Zhe, C. and Ji-Rong, H. (2010).Hepatic veno-occlusive disease associated with toxicity of pyrrolizidine alkaloids in herbal preparations. *The Journal of Medicine*, 256(6); 252-260.