



UNIVERSITI PUTRA MALAYSIA

***THE ROLE OF MEMBRANE TRANSPORTERS (NHE-1 AND AE-2) IN  
SECONDARY BONE HEALING OF TIBIA-FRACTURED  
SPRAGUEDAWLEY  
RATS***

KAREEM OBAYES HANDOOL

FPV 2018 6



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DAWLEY RATS**

KAREEM OBAYES HANDOOL

By

Thesis Submitted to the School of Graduate Studies, Universiti Putra  
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Philosophy

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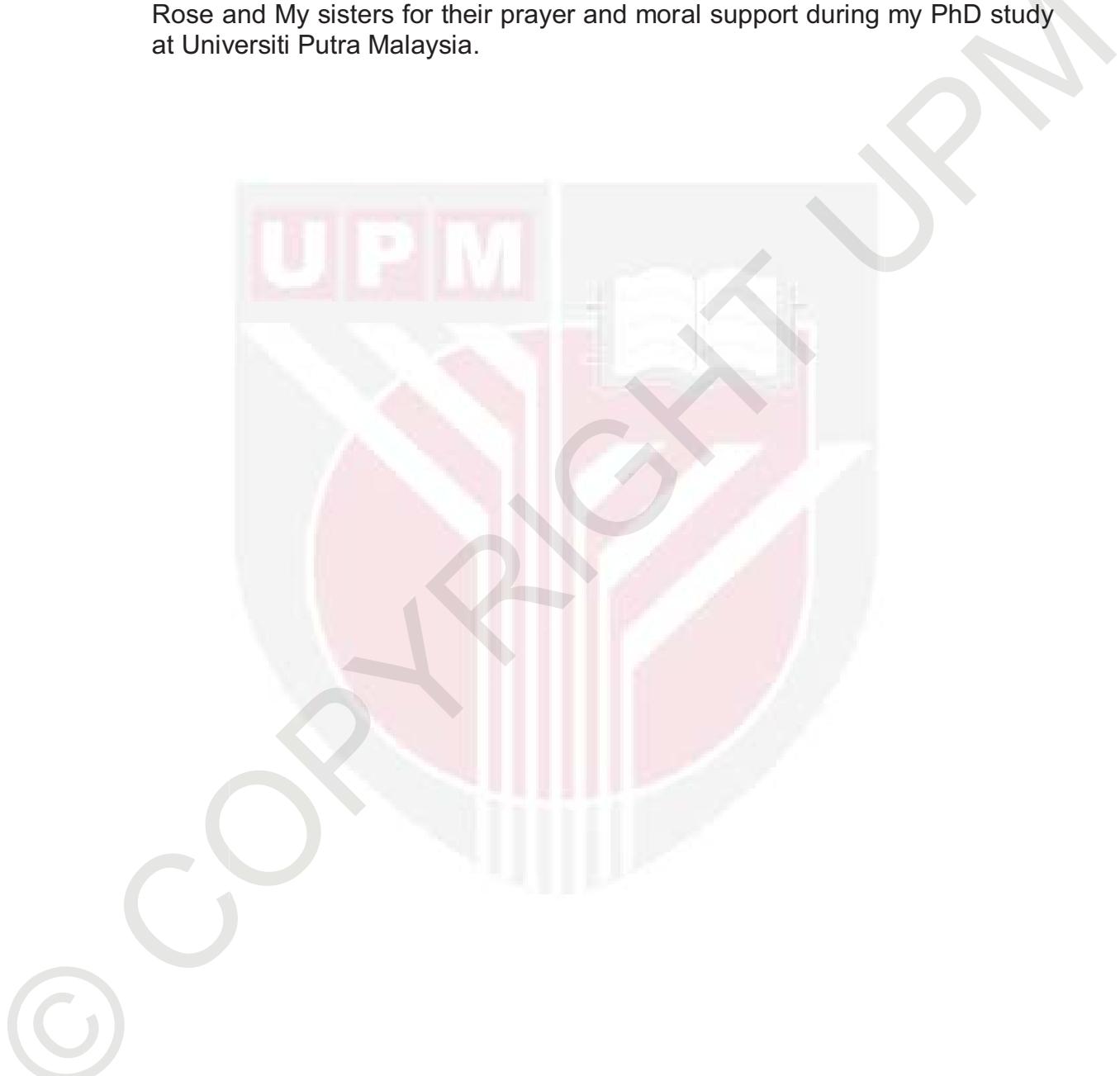
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## **DEDICATION**

Consequences of years of study, research and day and night investigation is the present thesis that I would like to dedicate to my beloved My Father and Mother, My brother Ali Obayes Handool, My Wife Halima Hander Ali, My sons Taha (God bless his soul), Tariq and Idris, My daughters Zahra, Athra and Rose and My sisters for their prayer and moral support during my PhD study at Universiti Putra Malaysia.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
fulfilment of the requirement for the degree of Doctor of Philosophy

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April 2017

**Chairman : Loqman Haji Mohamad Yusof, PhD**  
**Faculty : Veterinary Medicine**

Injuries emerging from orthopedic cases are increasingly becoming one of the major areas of attention in medicine. The development of bone has two major pathways, namely intramembranous bone formation and endochondral bone formation. Chondrocyte swelling describes the process emerging from the net movement of water into the cell which relies primarily on an osmotic gradient. It is likely that there is an important role of transporters which regulate the movement of  $\text{Na}^+$  and anions (e.g.,  $\text{HCO}_3^-$ ) across the cell membrane as these are known to be essential for the control of cell volume and pH in a wide range of cell types. This study hypothesizes that plasma membrane transporters have a role in cellular differentiation and regulation of endochondral ossification for secondary fracture healing. The objectives of this study were to evaluate the modified device to induce fracture for secondary fracture healing in a rat model, to study the different cellular stages of endochondral ossification, to evaluate the role of specific plasma membrane transporters ( $\text{Na}^+/\text{H}^+$  and  $\text{HCO}_3^-$ ) in secondary fracture healing and to evaluate the effect of EIPA (5-(N-ethyl-N-isopropyl) amiloride and DIDS (4,4'-diisothiocyanato-2,2'-stilbenedisulfonic acid) in secondary bone healing by using a rat tibial fracture model. A total of 55 female Sprague-Dawley rats of 8 weeks old were divided into three experiments: normal fracture healing ( $n=25$ , control), EIPA ( $n=15$ ) and DIDS ( $n=15$ ). Rats were sacrificed at 1, 2, 3, 4 and 6 weeks post-operative and assessed by clinical observation, radiology, histology, immunohistochemistry examination and statistical analysis. The modified device for producing fractures in the rat model is easy, cheap and reproducible, without complications. The result of gross callus area percentage and gross callus index showed significant difference at week 1 compared to the other weeks ( $P<0.05$ ); only four rats had slight comminution

and 21 rats without comminution. A radiographic examination showed clinical union at week 3 in 60% of the rats, and good clinical union (100%) with less callus formation in week 6. Histomorphometric for woven bone, lamellar bone and bone marrow fibrosis percentage area revealed significant differences ( $P<0.05$ ). Proliferative and hypertrophic chondrocyte zones percentage area showed a significant difference ( $P<0.05$ ). Immunoperoxidase staining for NHE-1 and AE-2 revealed significant differences ( $P<0.05$ ) in all weeks compared to week 6.

Following treatment with EIPA and DIDS, gross observation showed that the fracture line was clearly visible until week 4, manual fragment movements continued until week 2 and the callus area was smaller than in normal fracture healing. The X-ray callus index with DIDS treatment showed a significant difference ( $P<0.05$ ). Histomorphometric with EIPA and DIDS treatment showed that the percentage area for woven bone, lamellar bone, periosteal fibrosis and marrow fibrosis revealed a significant difference ( $P<0.05$ ); besides, the proliferative and hypertrophic chondrocyte zones percentage area showed a significant difference ( $P<0.05$ ). Immunohistochemistry density reaction for NHE-1 and AE-2 in EIPA and DIDS showed a significant difference ( $P<0.05$ ), the density reaction started a weak reaction, then declined directly to be absent in week 4 and week 6, whereas in normal fracture healing a strong reaction for NHE-1 started in the first four weeks then declined in week 6; however AE-2 began at a moderate level then increased strongly in weeks 3 and 4 and declined in week 6. The immunohistochemistry result refers to the direct effect of the inhibitors in the NHE-1 and AE-2 chondrocyte transporter proteins. These results suggest that NHE-1 and AE-2 have a role in the endochondral ossification of secondary bone healing. The inhibition of the hypertrophic chondrocyte zone following treatment with EIPA and DIDS, further strengthened the study hypothesis that NHE-1 and AE-2 inhibit fracture healing.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PERANAN PENGANGKUT MEMBRAN (NHE-1 DAN AE-2) DALAM  
PENYEMBUHAN SEKUNDER TULANG TIKUS SPRAGUE-DAWLEY  
TIBIA-TERPATAH**

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April 2017

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**Fakulti : Perubatan Veterinar**

Kecederaan yang muncul dari kes-kes ortopedik semakin menjadi salah satu bidang utama perhatian di dalam bidang perubatan. Pembangunan tulang mempunyai dua laluan utama, iaitu pembentukan tulang intramembran dan pembentukan tulang endokondral. Pembengkakan kondrosit menghuraikan proses yang muncul dari pergerakan bersih air ke dalam sel yang bergantung terutamanya kepada kecerunan osmosis. Kemungkinan terdapat peranan penting pengangkut-pengangkut yang mengawal pergerakan  $\text{Na}^+$  dan ion negatif (contohnya,  $\text{HCO}_3^-$ ) di seluruh membran sel kerana semua ini dikenali sebagai penting untuk kawalan isipadu sel dan pH bagi pelbagai jenis sel. Kajian ini menghipotesiskan bahawa pengangkut membran plasma mempunyai peranan di dalam pembezaan sel dan pengaturan osifikasi endokondral untuk penyembuhan patah sekunder. Objektif kajian ini adalah untuk menilai peranti yang diubah suai untuk mendorong fraktur untuk penyembuhan fraktur sekunder di dalam model tikus, untuk mengkaji ossifikasi endokondral di peringkat-peringkat sel yang berbeza, untuk menilai peranan pengangkut membran plasma tertentu ( $\text{Na}^+/\text{H}^+$  dan  $\text{HCO}_3^-$ ) dalam penyembuhan fraktur sekunder dan untuk menilai kesan EIPA (5-(N-etil-N-isopropyl) amiloride) dan DIDS (asid 4,4'-diisothiociano-2,2'-stilbenedisulfonic) dalam penyembuhan tulang sekunder dengan menggunakan model fraktur tibia tikus. Sebanyak 55 tikus betina Sprague-Dawley berumur 8 minggu dibahagikan kepada tiga eksperimen: penyembuhan fraktur normal ( $n=25$ , kawalan), EIPA ( $n=15$ ) dan DIDS ( $n=15$ ). Tikus-tikus tersebut dikorbankan pada 1, 2, 3, 4 dan 6 minggu selepas pembedahan dan dinilai melalui pemerhatian klinikal, radiologi, histologi, pemeriksaan imunohistokimia dan analisis statistik. Peranti yang diubahsuai untuk menghasilkan fraktur di dalam model tikus adalah mudah, murah dan boleh diulang, tanpa komplikasi. Hasil peratusan kawasan kalus kasar dan indeks kalus kasar menunjukkan

perbezaan yang signifikan pada minggu 1 berbanding minggu-minggu lain ( $P<0.05$ ); hanya empat tikus mempunyai pengecilan sedikit dan 21 tikus tanpa pengecilan. Suatu pemeriksaan radiografi menunjukkan pencantuman klinikal pada minggu 3 dalam 60% daripada tikus tersebut, dengan pencantuman klinikal yang baik (100%) dan pembentukan kalus yang kurang pada minggu 6. Histomorphometri untuk tulang tenunan, tulang lamela dan peratusan kawasan fibrosis sumsum tulang menunjukkan perbezaan yang signifikan ( $P<0.05$ ). Peratusan kawasan zon-zon pembiasaan dan hipertrofi kondrosit menunjukkan perbezaan yang signifikan ( $P<0.05$ ). Mewarnakan dengan Immunoperoxidase untuk NHE-1 dan AE-2 menunjukkan perbezaan yang signifikan ( $P<0.05$ ) di semua minggu berbanding minggu 6.

Selepas rawatan dengan EIPA dan DIDS, pemerhatian kasar menunjukkan bahawa garis fraktur itu jelas kelihatan sehingga minggu 4, pergerakan serpihan manual berterusan sehingga minggu 2 dan kawasan kalus adalah lebih kecil daripada dalam penyembuhan fraktur normal. Indeks kalus X-ray dengan rawatan DIDS menunjukkan perbezaan yang signifikan ( $P<0.05$ ). Histomorphometri dengan EIPA dan rawatan DIDS menunjukkan bahawa peratusan kawasan untuk tulang tenunan, tulang lamela, fibrosis periosteum dan fibrosis sumsum menunjukkan perbezaan yang signifikan ( $P<0.05$ ); selain itu, peratusan kawasan zon-zon pembiasaan dan hipertrofi kondrosit menunjukkan perbezaan yang signifikan ( $P<0.05$ ). Reaksi ketumpatan immunohistokimia untuk NHE-1 dan AE-2 dalam EIPA dan DIDS menunjukkan perbezaan yang signifikan ( $P<0.05$ ), tindak balas ketumpatan memulakan tindak balas yang lemah, kemudian menurun secara langsung dan tidak ada di minggu 4 dan minggu 6, sedangkan pada penyembuhan fraktur normal reaksi yang kuat untuk NHE-1 bermula pada empat minggu pertama kemudian menurun pada minggu 6; bagaimanapun AE-2 bermula pada tahap sederhana kemudian meningkat dengan kukuh pada minggu 3 dan 4 dan menurun pada minggu 6. Hasil immunohistokimia merujuk kepada kesan langsung daripada perencat dalam protein pengangkut kondrosit NHE-1 dan AE-2. Keputusan-keputusan ini menunjukkan bahawa NHE-1 dan AE-2 mempunyai peranan dalam ossifikasi endochondral penyembuhan tulang sekunder. Perencatan zon hipertrofi kondrosit selepas rawatan dengan EIPA dan DIDS, mengukuhkan lagi hipotesis kajian bahawa NHE-1 dan AE-2 merencatkan penyembuhan fraktur.

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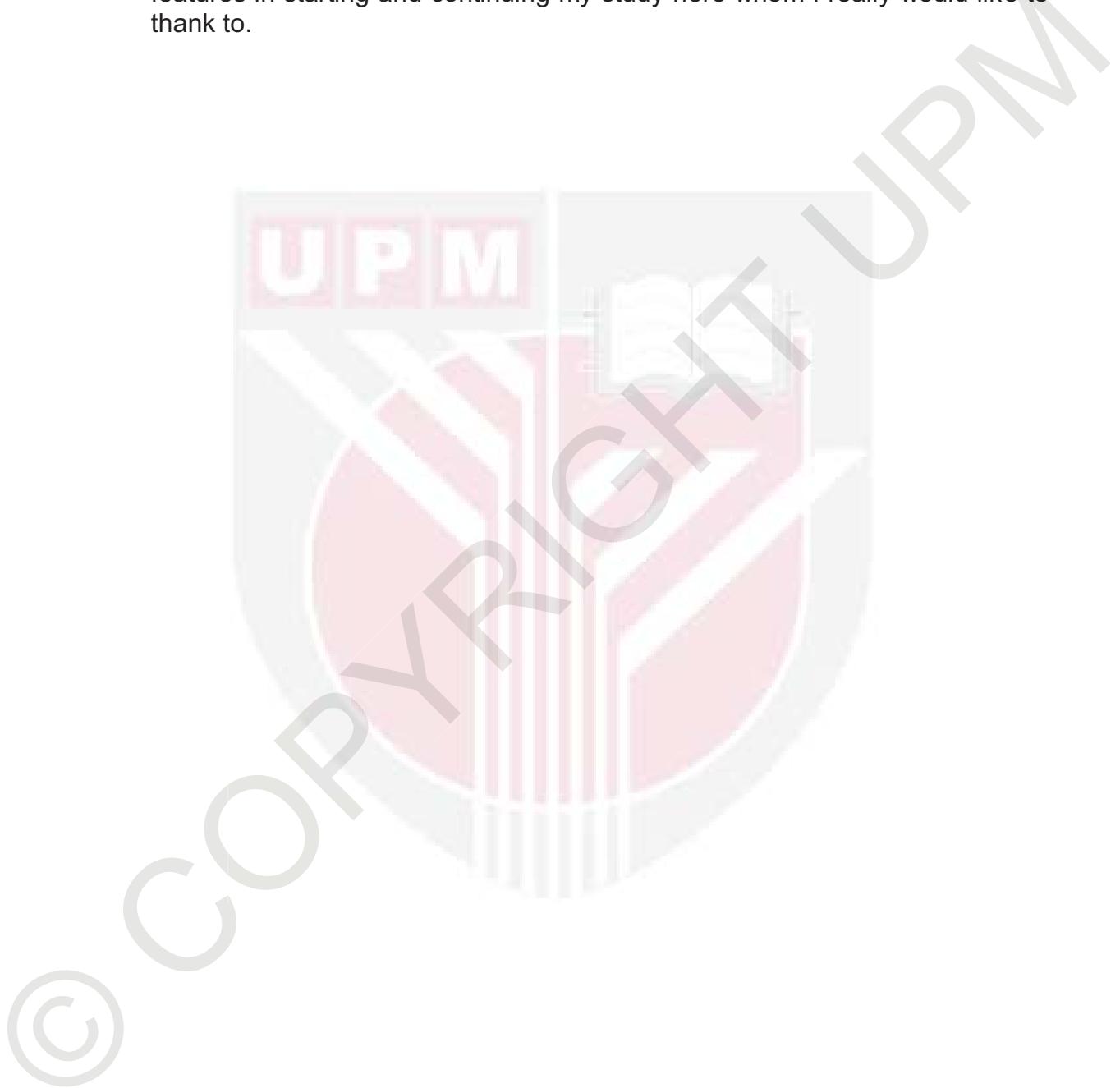
I'd mainly like to thank and appreciate Assoc. Prof. Dr. Md. Sabri Mohd Yusof. Head Department of Veterinary Pathology & Microbiology, Faculty of Veterinary Medicine, University Putra Malaysia, member of supervisory committee, who actively helped to evaluate the histological slides, for his excellent feedback and suggestions.

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This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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## LIST OF ABBREVIATIONS

Ab	Antibody
ACUC	Animal care and user committee
AE	Anion exchanger
Akt	Protein kinase B (PKB), also known as Akt
ANOVA	Analysis of Variance
ASCT	Amino acid transporters,
ASICs	Acid sensing ion channels
BAX	A gene located on chromosome 19q13.3-q13.4
BM	Bone marrow fibrosis
BMPs	Bone morphogenic proteins
BO	Bio-Oss
C	Cartilage
°C	Degree celsius
Ca.	Calcium
CA	Callus area
cAMP	Cyclic adenosine monophosphate
Ca <sub>5</sub> (PO <sub>4</sub> ) <sub>3</sub> (OH) <sub>2</sub>	Hydroxyapatite
CA	Callus area
CB	Cortical bone
CI	Callus index
Cl <sup>-</sup>	Chloride ion
cm	Centimeter
CO <sub>2</sub>	Carbon dioxide
CSD	Critical sized defect

CT	Connective tissue
DBM	Demineralized bone matrix
DEPC	Diethylpyrocarbonate
DIDS	4,4'-diisothiocyanostilbene-2,2'-disulfonic acid
DPX	Distrene, Plasticizer, Xylene
DMSO	Dimethyl Sulfoxide
eAE	Erythroid Anion Exchangers
ECG	Electrocardiography
ECM	Extracellular matrix
EIPA	5-(N-Ethyl-N-isopropyl)-Amiloride
eNOS	Endothelial Nitric Oxide Synthase
ERM	Ezrin-Radixin-Moesin.
ESF	External skeletal fixation
FGF	Fibroblast growth factor
Fig.	Figure
g	Gram
G.	Groups
GAGs	Glycosaminoglycans
GF	Growth factor
GLUT	Glucose-transporter
H <sup>+</sup>	Hydrogen ion
HA	Hydroxyapatite
HB	Host bone
HCL	Hydrochloride
H & E	Hematoxylin and Eosin
HCO <sub>3</sub> <sup>-</sup>	Bicarbonate

HMIT	Hydrogen myo-inositol transporter
HOCH <sub>2</sub> CH <sub>2</sub> NH <sub>3</sub> <sup>+</sup>	Ethanolamine
HOCH <sub>2</sub> CH(COO-) <sup>-</sup> NH <sub>3</sub> <sup>+</sup>	Phosphate serine
HCZ	Hypertrophic chondrocyte zone
IC <sub>50</sub>	Median inhibitory concentration
i.e.	Id est.
IGF	Insulin growth factor
IHC	Immunohistochemistry
IL	Interleukin
IM	Intramedullary pin
i.m	Intramuscular
IP	Intraperitoneal
IU	International unit
I.V	Intravenous
K <sup>+</sup>	Potassium ion
Ki	The inhibitory constant of drug
KO	Knockout
KVp	Kilo voltage-peaks
Lat.	Lateral
Lb	Lamellar bone
LM	Light microscope
MAs	Mill-ampere-seconds
MCSF	Macrophage colony stimulating factor
MFS	Major facilitator superfamily
MB	Mature bone
ml	Milliliter

mm	Millimeter
mM	Mill mole
MOBL	Mesenchymal osteoblasts
mRNA	Messenger Ribonucleic Acid
MV	Millivolts
MSC	Mesenchymal stem cell
no	Number
Na <sup>+</sup>	Sodium ion
NB	New bone
NCSD	Non-critical sized defect
NBCs	Sodium bicarbonate transporters
NHE	Sodium Hydrogen Exchanger
NKCC	Sodium, potassium, and chloride transporter
No	Number
NO	Nitric Oxide
NPPB	5-nitro-2-(3-phenylpropyl-amino) benzoic acid
OB	Original bone
OH	Hydroxyl group
OPGL	Osteoprogerin Ligand
OSTB	Osteoblasts
OSTP	Osteoporosis
p.	Page
p	P-Value
PBS	Phosphate buffer solution
PDGF	Platelet derived growth factor
Pf	Periosteal fibrosis

PGE	Prostaglandin E
PH	Power of hydrogen
pHe	Extracellular pH
pHi	Intracellular pH
PI3K	Phosphatidylinositol-3-OH kinase
pp.	Pages
PCZ	Proliferative chondrocyte zone
RANKL	Receptor Activator for Nuclear Factor $\kappa$ B ligand
ROI	Region of interest
S	Sample
SAU	Sindh Agriculture University
S/C	Subcutaneous
SLC	Solute carrier family
SOBL	Surface osteoblasts
SPSS	Statistical package for social science
SE	Standard error
SEM	Scanning electron microscope
SITS	Stilbene derivatives
SO <sub>4</sub>	Sulfate
STD	Standard deviation
TGF	Transforming growth factor
TGF-B	Transforming growth factors-b
TM	Transmembrane
TNFR	Tumor necrosis factor receptor
UDC	Ursodeoxycholate
UPA	Urokinase-type plasminogen activator

UPM	Universiti Putra Malaysia
Vas	Vascularization
Wk	Week
Wb	Woven bone
W/V	Weight/Volume
$\mu\text{m}$	Micromillileter



## CHAPTER 1

### INTRODUCTION

Major injuries of bone associated with multiple trauma and traffic accidents, which lead to prolonged periods of treatment with significant socioeconomic impacts, are still considered as basic health issues in advanced countries (Sfeir *et al.*, 2005; Fayaz *et al.*, 2011). Bone fracture healing can occur via two techniques endochondral bone formation and intramembranous bone formation. Secondary bone healing includes the classical phases of injury, haemorrhage, inflammation, soft callus formation, mineralization callus, and remodelling callus.

This process of secondary bone healing strictly be similar to endochondral ossification, which includes a cartilage template being substituted by bone (Shapiro, 2008). Endochondral ossification is essential processes during fetal development of the mammalian skeletal system also an essential process during the rudimentary formation of long bones, the growth of the length of long bones, (Brighton *et al.*, 1973) and the natural healing of bone fractures (Brighton and Robert, 1986; Fayaz *et al.*, 2011). Bone healing utilized the similar formation designs as bone growth by enlargement of chondrocytes, however the specific approach of healing is determined through the biomechanical environment delivered (Kim *et al.*, 2013; Sathyendra & Darowish, 2013).

The plasma membrane is the borderline that separates the living cell from its surroundings and exhibits selective permeability, which permits certain material pass more easily than others do. Phospholipid bilayer, cholesterol and protein are the fundamental structure of the plasma membrane. For cellular biological homeostasis to be maintained, molecules must be removed from and transported into organelles and cells. This crucial function is accomplished through way of transport proteins that exist in intracellular membranes and in the cytosol. These proteins control the inflow of vital ions, nutrients, environmental toxins, cellular waste, and other xenobiotics, which play vital roles in cellular homeostasis and enable the movement of vital biological molecules such as sugars, amino acids, nucleotides, and vitamins through cellular membranes against or along their electrochemical gradients (Kim, 2002; Eraly, *et al.*, 2004).

In the human genome about two thousand transporter-associated genes, underlining their biological importance and role in cellular homeostasis was discovered. The solute carrier families carry out more than three hundred transporters from these transporter genes.

The sodium/hydrogen ion exchangers (NHEs) are fundamental transporter proteins in the solute carrier family 9 (SLC-9) and perform important functions in transepithelial salt, acid and base transport, and regulation of extracellular and intracellular pH, cell volume regulation, growth, proliferation, differentiation and apoptosis (Landowski *et al.*, 2008).

The second major group of anion exchangers-bicarbonate cotransporter family is the sodium bicarbonate transporters (NBCs), which perform a crucial role in acid-based movement in most tissues and cell types including kidney, heart, liver, blood cells, intestine, stomach, pancreas, central nervous system and reproductive systems (Abduladze *et al.*, 1998).

Bush *et al.* (2010) in previous study provided an indication of the role of the Na-K-2Cl cotransporter (NKCC1) in volume increased of growth plate hypertrophic chondrocyte zone. However, another study on the role of anion exchanger (AE-2) and  $\text{Na}^+/\text{H}^+$  antiporter (NHE-1) in bone growth plate chondrocytes has been reported (Loqman *et al.*, 2013). This significance is due to the fact that indirect bone healing can occur in the similar formation design of enlargement of chondrocytes in endochondral ossification of secondary bone healing. This indicates the role of the transporters that control the movement of anions (e.g.,  $\text{HCO}_3^-$ ) and  $\text{Na}^+$  through chondrocyte cell membranes.

Many diseases for instance cystic fibrosis, non-insulin-dependent diabetes, type I cystinuria, mellitus haemolytic anaemia, epilepsy and schizophrenia acquired or inherited are produced via defective or dysregulation expression of transporter proteins (Landowski *et al.*, 2008). For example, cystic fibrosis is a common life-limiting autosomal recessive genetic disorder, with highest prevalence in Europe, North America, and Australia. The disease is caused by mutation of a gene that encodes a chloride-conducting transmembrane channel called the cystic fibrosis transmembrane conductance regulator (CFTR), which regulates the anion transport and mucociliary clearance in the airways. Functional failure of CFTR results in mucus retention and chronic infection and subsequently in local airway inflammation that is harmful to the lungs (Elborn, 2016).

Currently, studies investigating animal models of secondary fracture bone healing are still few and data on the different cellular stages of endochondral ossification in secondary bone fracture healing are limited. Insufficiency of information also exists on the role of specific plasma membrane transporters ( $\text{Na}^+/\text{H}^+$  and  $\text{HCO}_3^-$ ) in chondrocytes for secondary bone fracture healing.

Orthopedic surgery and orthopedics, similar to other specialties, have been developed through the requirement and responsibility. Orthopedic surgeons have developed the capability to avoid major losses of a bone's function and actually, they can prevent expectable death. They try to find excellence in their art, by making sure that the patient achieves optimal condition in the shortest period throughout the safest procedures and possible methods with lost economic losses.

Delayed healing results in an increased duration of immobilization in non-union or mal-union of the bone ends, increases the risk of joint stiffness, and is associated with Long-term morbidity, hospitalization, care patients and death, also increases the risk of inadequate fracture alignment. Long fracture healing period, suffering to patients and large costs for society, a pharmacological therapy, the high socioeconomic costs and the complications impaired in bone healing, bone regeneration and the targeted therapies were the major problem in bone fracture healing.

This study was intended to evaluate the role of specific cell plasma membrane transporters and cellular differentiation of endochondral ossification process in secondary fracture healing using laboratory animal models.

#### **Hypothesis:**

Specific plasma membrane transporters ( $\text{Na}^+/\text{H}^+$  and  $\text{HCO}_3^-$ ) play a significant role in cellular differentiation and regulation of endochondral ossification in secondary bone healing.

#### **Objectives:**

This study was conducted in recognition of the fact that plasma membrane transporters are important in cellular differentiation and regulation of endochondral ossification for secondary fracture healing. The specific objectives of this study were:

- 1- To evaluate the modified three-point bending pliers to induce fracture using *in vivo* animal model for secondary bone fracture healing.
- 2- To study the different cellular stages of endochondral ossification in secondary bone fracture healing.
- 3- To determine the role of specific plasma membrane transporters ( $\text{Na}^+/\text{H}^+$  and  $\text{HCO}_3^-$ ) in secondary bone healing.
- 4- To evaluate the effect of EIPA in secondary bone healing in the rat tibial fracture model.
- 5- To evaluate the effect of DIDS in secondary bone healing in the rat tibial fracture model.

## REFERENCES

- Abuladze, N., Lee, I., Newman, D., Hwang, J., Boorer, K., Pushkin, A., and Kurtz, I. (1998). Molecular Cloning, Chromosomal Localization, Tissue Distribution, and Functional Expression of the Human Pancreatic Sodium Bicarbonate Cotransporter. *Journal of Biological Chemistry*. 273(28):17689-17695.
- Adams, C.S., and Shapiro, I.M. (2002). The Fate of the Terminally Differentiated Chondrocyte: Evidence for Microenvironmental Regulation of Chondrocyte Apoptosis. Critical. *Journal of Dental Research. Oral Biology Medicine*. 13(6):465-473.
- Aerssens, J., Boonen, S., Lowet, G. and Dequeker, J. (1998). Interspecies differences in bone (3<sup>rd</sup> Ed). *Philadelphia: Mosby*. 17: 355–61.
- Al-Aql, Z.S., Alagl, A.S., Graves D.T., Gerstenfeld, L.C. and Einhorn, T.A. (2008). Molecular Mechanisms Controlling Bone Formation during Fracture Healing and Distraction Osteogenesis. *Journal of Dental Research*. 87(2): 107-118.
- Alagl, A. S. and Graves, D.T. (2007). Molecular Mechanisms Controlling Bone Formation during Fracture Healing and Distraction Osteogenesis. Critical Reviews in Oral Biology and Medicine. *Journal of Dental Research*. 87(2): 107–118.
- Alberts, B., Bray, D., Hopkin, K., Johnson, A., Lewis, J., Raff, M., Roberts, K. and Walter, P. (2004). Membrane Transport. In: Essential Cell Biology. (2<sup>nd</sup> Ed). GS Garland Science. pp. 389-426.
- Allen, H.L., Wase, Bear, W.T. (1980). Indomethacin and aspirin: effect of nonsteroidal anti-inflammatory agents on the rate of fracture repair in the rat. *Acta Orthopaedica Scandinavica*. 51(4):595-600.
- Alper, S.L. (2009). Molecular physiology and genetics of Na<sup>+</sup>-independent SLC-4 anion exchangers. *Journal of Experimental Biology*. 212 (Pt 11): 1672-83.
- Alves, C., Ma, Y., Li, X. and Fliegel, L. (2014). Characterization of human mutations in phosphorylatable amino acids of the cytosolic regulatory tail of SLC9A1. *Biochemistry and Cell Biology*. 92(6): 524-529.
- Amini, S., Veilleux, D., and Villemure, I. (2011). Three-dimensional in situ zonal morphology of viable growth plate chondrocytes: A confocal microscopy study. *Journal of Orthopedic Research*. 29 (5):710-717.
- Amith, S.R., Wilkinson, J.M., Baksh, S. and Fliegel, L. (2015). The Na<sup>+</sup>/H<sup>+</sup> exchanger (NHE-1) as a novel co-adjuvant target in paclitaxel therapy of triple-negative breast cancer cells. *Oncotarget*. Jan 20; 6(2):1262-75.
- An, Y.H., Woolf, S.K., and Friedman, R.J. (2000). Pre-clinical in vivo evaluation of orthopaedic bioabsorbable devices. *Biomaterials* 21(24):2635-2652.

- Andreassen, K.H., Pedersen, K.V., Osther, S.S., Jung, H.U., Lildal, S.K., and Osther, P.J.S. (2016). How should patients with cystine stone disease be evaluated and treated in the twenty-first century? *Urolithiasis*. Volume 44, Issue 1, pp. 65-76.
- Apelt, J., Mehlhorn, G., S and chliebs, R. (1999). Insulin-sensitive GLUT4 glucose transporters are colocalized with GLUT3-expressing cells and demonstrate a chemically distinct neuron-specific localization in rat brain. *Journal of Neuroscience Research*. 57(5):693–705.
- Aranda, V., Martinez, I., Melero, S., Lecanda, J., Banales, J. M., Prieto, J. and Medina, J. F. (2004). Shared apical sorting of anion exchanger isoforms AE-2a, AE-2b1, and AE-2b2 in primary hepatocytes. *Biochemical and Biophysical Research Communications* Volume 319, Issue 3, 2 July 2004, pp. 1040–1046
- Arenas, F., Hervias, I., Uriz, M., Joplin, R., Prieto, J. and Medina, J. F. (2008). Combination of ursodeoxycholic acid and glucocorticoids up regulates the AE-2 alternate promoter in human liver cells. *The Journal of Clinical Investigation*. 118,695 -709.
- Arriza, J.L., Kavanaugh, M.P., Fairman, W.A., Wu, Y.N., Murdoch, G.H., North, R.A., Amara, S.G., (1993). Cloning and expression of a human neutral amino acid transporter with structural similarity to the glutamate transporter gene family. *Journal of Biology and Chemistry*. 268 (21), 15329–15332.
- Attaphitaya, S., Park, K., and Melvin, J. E. (1999). Molecular cloning and functional expression of a rat  $\text{Na}^+/\text{H}^+$  exchanger (NHE5) highly expressed in brain. *The Journal of Biological Chemistry*. 274(7) 4383-4388.
- Attaphitaya, S., Park, K., and Melvin, J.E. (1999). Molecular cloning and functional expression of a rat  $\text{Na}^+/\text{H}^+$  exchanger (NHE5) highly expressed in brain. *Journal of Biology and Chemistry*. 274(7) 4383-4388.
- Augustin, R. (2010). The protein family of glucose transport facilitators: It's not only about glucose after all. Critical Review. *Journal of the International Union of Biochemistry and Molecular Biology (IUBMB)*. 62(5):315–333.
- Aurégan, J.C., Coyle, R. M., Danoff, J. R., Burky, R. E. and Rosenwasser, Y. M. P. (2013). The rat model of femur fracture for bone and mineral research. *Bone Joint Research*. 2: (8)149–54.
- Banales, J. M., Arenas, F., Rodriguez-Ortigosa, C. M., Saez, E., Uriarte, I., Doctor, R. B., Prieto, J. and Medina, J. F. (2006). Bicarbonate-rich choleresis induced by secretin in normal rat is taurocholate-dependent and involves AE-2 anion exchanger. *Hepatology*. 43(2):266-75.

- Bandyopadyay-Ghosh, S. (2008). Bone as a collagen-hydroxyapatite composite and its repair. *Journal in Trends in Biomaterials & Artificial Organs*. 22(2):112-120.
- Barrère, F., Blitterswijk, C.A., and de Groot, K. (2006). Bone regeneration: molecular and cellular interactions with calcium phosphate ceramics. *International Journal Nanomedicine*. 1(3): 317–332.
- Barry, S. (2010). Non-steroidal anti-inflammatory drugs inhibit bone healing: A review. *Veterinary and Comparative Orthopaedics and Traumatology*, 23(6):385-92
- Beltran, A.R., Ramírez, M.A., Carraro-Lacroix, L.R., Hiraki, Y., Rebouças, N.A., and Malnic, G. (2008). NHE-1, NHE-2, and NHE-4 contribute to regulation of cell pH in T84 colon cancer cells. *European Journal of Physiology*. Volume 455, Issue 5, pp. 799-810.
- Benedetti, A., Antonio Sario, A. D., Casini, A., Ridolfi, F., Bendia, E., Pigini, P., Tonnini, C., D'ambrosio, I., Feliciangeli, G., Macarri, G., and Svegliati-Baroni, G., (2001). Inhibition of the  $\text{Na}^+/\text{H}^+$  Exchanger Reduces Rat Hepatic Stellate Cell Activity and Liver Fibrosis: An In Vitro and *In Vivo* Study. *Gastroenterology*.Volume 120, Issue 2, pp. 545–556.
- Bertazzo, S. and Bertran, C. A. (2006). Morphological and dimensional characteristics of bone mineral crystals. *Bioceramics*. 309–311 (Pt. 1, 2): 3–10.
- Bertazzo, S., Bertran, C.A. and Camilli, J.A. (2006). Morphological Characterization of Femur and Parietal Bone Mineral of Rats at Different Ages. *Key Engineering Materials*. 309–311: 11–14.
- Black, J., Perdigon, P. and Brown, N. (1984). Stiffness and strength of fracture callus. Relative rates of mechanical maturation as evaluated by a uniaxial tensile test. *Clinical Orthopedic Relation Research*: (182):278-88.
- Blokhus, T.J., Bruine, J.H.D., de Bramer, J.A.M., den Boer, F.C., Bakker, F.C., Patka, P., Haarman, H.J.Th.M., and Manoliu, R.A. (2001).The reliability of plain radiography in experimental fracture healing. *Skeletal Radiology*. 30(3)151–156.
- Bolander, M. E. (1992). Regulation of fracture repair by growth factors. *Proc. Soc. Experimental Biology of Medicine*. 200(2)165–170.
- Bonewald, L. F. (2011). “The amazing osteocyte,” *Journal of Bone and Mineral Research*, vol. 26, no. 2, pp. 229–238.
- Bonnarens, F. and Einhorn, T. (1984). Production of a standard closed fracture in laboratory animal bone. *Journal Orthopedic Research*. 2: (1)97–101.

- Bozeat, N.D., Xiang, S.Y., Ye, L.L., Yao, T.Y., Duan, M.L., Burkin, D.J., Lamb, F.S., Duan, D.D (2011). Activation of volume regulated chloride channels protects myocardium from ischemia/reperfusion damage in second- window ischemic preconditioning. *Cell Physiology Biochemistry*. 28(6):1265-1278.
- Brandi, M. L. (2010). How innovations are changing our management of osteoporosis. *Medicographia*. 32, (1)1-6.
- Breur, G.J., VanEnkervort, B.A., Farnum, C.E., Wilsman, N.J. (1991). Linear relationship between the volume of hypertrophic chondrocytes and the rate of longitudinal bone growth in growth plates. *Journal Orthopaedic Research* 9:348–359.
- Brighton, C. T., and Robert, M. H. (1991). "Early histological and ultrastructural changes in medullary fracture callus", *Journal of Bone and Joint Surgery*, 73-A (6): 832-847
- Brighton, C. T., and Robert M. H. (1986). "Histochemical localization of calcium in the fracture callus with potassium pyroantimonate: possible role of chondrocyte mitochondrial calcium in callus calcification", *Journal of Bone and Joint Surgery*, 68-A (5): 703-715.
- Brighton, C. T., Yoichi, S., and Robert, M. H. (1973). "Cytoplasmic structures of epiphyseal plate chondrocytes; quantitative evaluation using electron micrographs of rat costochondral junctions with specific reference to the fate of hypertrophic cells", *Journal of Bone and Joint Surgery*, 55-A: 771-784.
- Brown, B.S. (1996). Biological Membrane. The Biological Society. pp. 1-22.
- Bruzzaniti, A. and Baron, R. (2006). Molecular regulation of osteoclast activity. *Endocrine and Metabolic Disorders*. 7(1-2):123-39.
- Buckwalter, J.A., Einhorn, T.A., and O'Keefe, R.J. (2007). American Academy of Orthopaedic Surgeons. Orthopaedic basic science: foundations of clinical practice. 3rd edition. Rosemont (IL): *American Academy of Orthopaedic Surgeons*. pp. 331–46.
- Burkitt, H. G., Stevens, A., Lowe, J. S. and Young, B. (1999). Wheatear's Basic Histopathology.3<sup>rd</sup> ed. Churchill Livingstone. Pp. 1-261.
- Bush, P.G., and Hall, A.C. (2001). The osmotic sensitivity of isolated and in situ bovine articular chondrocytes. *Journal of Orthopedic Research*. 19(5):768-778.
- Bush, P.G., Pritchard, M., Loqman, M.Y., Damron, T. a., and Hall, A.C. (2010). A key role for membrane transporter NKCC1 in mediating chondrocyte volume increase in the mammalian growth plate. *Journal of Bone Minerals Research*. 25(7):1594-603.

- Cardone, R.A., Casavola, V., and Reshkin, S.J. (2005). The role of disturbed pH dynamics and the  $\text{Na}^+/\text{H}^+$  exchanger in metastasis. *Cancer*. 5(10):786-95.
- Carruthers, A., DeZutter, J., Ganguly, A., and Devaskar, S.U. (2009). Will the original glucose transporter isoform please stand up! *American Journal Physiology Endocrinology Metabolism*. 297(4):E836–848.
- Cavendish, M. (2010). Mammal anatomy: an illustrated guide. New York: p. 129.
- Chakkalak, D.A., Strates, B.S., Mashoof, A.A., Garvin, K.L., Novak, J.R., Fritz, E.D., Moliner, T.G., and McGuire, M.H. (1999). Repair of segmental bone defects in the rat an experimental model of human fracture healing. *Bone* 25 (3) 321-332.
- Checa, S., Prendergast, P.J., Duda, G.N. (2011). Inter-species investigation of the mechano-regulation of bone healing: Comparison of secondary bone healing in sheep and rat. *Journal of Biomechanics*. 44(7):1237-1245.
- Chen, Y-X, and O'Brien, E.R. (2003). Ethyl isopropyl amiloride inhibits smooth muscle cell proliferation and migration by inducing apoptosis and antagonising urokinase plasminogen activator activity. *Canadian Journal of Physiology and Pharmacology*. 81(7): 730-739
- Chow, A., Dobbins, J. W., Aronson, P. S. and Igarashi, P. (1992). cDNA cloning and localization of a band 3-related protein from ileum. *American Journal of Physiology*. 263(3 Pt.1):G345-52.
- Cogan, M. G. (1990). Angiotensin II: a powerful controller of sodium transport in the early proximal tubule. *Hypertension*. 15:451-458.
- Compston, J. (1998). Bone Histomorphometry. In Methods in bone biology 1st edition. Edited by: Arnett TR, Henderson B. London, England: Chapman & Hall; 177-99.1.
- Cooper, G.M. and Hausman, R.E. (2009). The Plasma Membrane. In: The Cell A Molecular Approach. (5<sup>th</sup> Ed). ASM Press. pp. 529-569.
- Cruess, R. L., & Dumont, J. A. C. Q. U. E. S. (1975). Fracture healing. *Canadian journal of surgery. Journal canadien de chirurgie*, 18(5), 403-413.
- Curry, J.D. (2006). "The Structure of Bone Tissue" Bones: Structure and wellMechanics Princeton U. Press. Princeton, NJ. pp. 12–14.
- Danielli, J. F., and Davson, H. (1935). A contribution to the theory of permeability of thin films. *Journal of Cellular and Comparative Physiology*. Volume 5, Issue 4 5:495-508.

- Das, S., Steenbergen, C., and Murphy, E. (2012). Does the voltage dependent anion channel modulate cardiac ischemia reperfusion injury? *Biochimica Biophysica Acta (BBA) - Biomembranes*. Volume 1818, Issue 6, 1451-1456.
- Datta, H. K., Ng, W. F., Walker, J. A., Tuck, S. P., and Varanasi, S. S. (2008). "The cell biology of bone metabolism," *Journal of Clinical Pathology*, vol. 61, no. 5, pp. 577–587.
- Deakin, P.J., Young, B., Lowe, J.S., Stevens, A., and John, W. (2006). Wheater's functional histology: a text and colour atlas (5<sup>th</sup> Ed.). [Edinburgh?]: Churchill Livingstone/Elsevier. p. 273.
- Delmas, P., and Coste, B. (2013). Mechano-gated ion channels in sensory systems. *Cell*. 155: (2)278-284.
- Dimitriou, R., Jones, E., McGonagle, D., and Giannoudis, P.V. (2011). Bone regeneration: current concepts and future directions. (*BMC Medicine BioMed Center*. 9:66
- Dimitriou, R., Tsiridis, E. and Giannoudis, P.V. (2005). Current concepts of molecular aspects of bone healing. *Injury*. 36: (12)1392-1404.
- Downey, P. A. and Siegel, M. I., (2006). "Bone biology and the clinical implications for osteoporosis," *Physical Therapy*, vol. 86, no. 1, pp. 77–91.
- Draper, E. and Goodship A. (2003). A novel technique for four-point bending of small bone samples with semi-automatic analysis. *Journal of Biomechanics*. 36: (10)1497–502.
- Duda, G.N., and Schmidt-Bleek, K. (2014). T and B cells participate in bone repair by infiltrating the fracture callus in a two-wave fashion. *Bone*. 64:155-65.
- Eastaugh-Waring, S.J., Joslin, C.C., Hardy, J.R.W., and Cunningham, J.L. (2009). Quantification of fracture healing from radiographs using the maximum callus index. *Clinical orthopaedics and related research*. 467(8):1986-1991.
- Elborn, J.S. (2016). Cystic Fibrosis. *School of Medicine, Dentistry and Biomedical Sciences*. 388: 2519–31.
- Einhorn, T., and Lane, J.M, (1998). The Cell and Molecular Biology of Fracture Healing. *Clinical orthopaedics and related research*. 355 Supple: S7-S21.
- Eriksson, M., Taskinen, M., Leppä, S. (2006). Mitogen Activated Protein Kinase-Dependent Activation of c-Jun and c-Fos is required for Neuronal differentiation but not for Growth and Stress Response in PC12 cells. *Journal of Cell and Physiology*. 207(1):12-22.

- Estai, M. A., Suhami, F.H., Das, S., Fadzilah, F.M., Alhabshi, S.M.I., Shuid, A.N., and Soelaiman, I. (2011). Piper sarmentosum enhances fracture healing in ovariectomized osteoporotic rats: a radiological study. *Clinics*. 66(5):865-872.
- Farnum, C.E., Lee, R., O'Hara, K., and Urban, J.P.G. (2002). Volume increase in growth plate chondrocytes during hypertrophy: the contribution of organic osmolytes. *Bone*. 30:574–581.
- Fattah, H., Hambaroush, Y., and Goldfarb, D.S. (2014). Cystine nephrolithiasis. *Translational Andrology and Urology*. 3(3): 228–233.
- Fayaz, H.C., Giannoudis, P. V., Vrahas, M.S., Smith, R.M., Moran, C., Pape, H.C., Krettek, C., & Jupiter, J.B. (2011). The role of stem cells in fracture healing and non-union. *International orthopaedics*. 35(11):1587-1597.
- Fedchenko, N., and Reifenrath, J. (2014). Different approaches for interpretation and reporting of immunohistochemistry analysis results in the bone tissue. *Diagnosis Pathology*. 9(1):221.
- Fernández, K.S. and de Alarcón, P.A. (2013). Development of the hematopoietic system and disorders of hematopoiesis that present during infancy and early childhood. *Pediatric clinics of North America*. 60 (6): 1273–89.
- Fernández-Tresguerres-Hernández-Gil, I., Alobera-Gracia, M.A., Del Canto-Pingarrón, M., and Blanco-Jerez, L. (2006). Physiological bases of bone regeneration II. The remodeling process. *Medicina oral, patología oral y cirugía bucal*. 11: (2) E151–E157.
- Fliegel, L. (2008). Molecular biology of the myocardial Na<sup>+</sup>/H<sup>+</sup> exchanger. *Journal of molecular and cellular cardiology*. 44: (2) 228–237.
- Fliegel, L. (2009). Regulation of the Na<sup>+</sup>/H<sup>+</sup> exchanger in the healthy and diseased myocardium. *Expert opinion on therapeutic targets*. 13 :( 1) 55–68.
- Fliegel, L. (2014). Critical Review the Na<sup>+</sup>/H<sup>+</sup> Exchanger and pH Regulation in the Heart. *IUBMB Life. Biochemistry & Molecular Biology*. 66 (10). 67-685.
- Florencio-Silva, R., Sasso, G.R.D.S., Sasso-Cerri, E., Simões, M.J., and Cerri, P.S. (2015). Biology of Bone Tissue: Structure, Function, and Factors That Influence Bone Cells. *BioMed Research International*. Volume 2015 (2015) 1-17.
- Frymoyer, J.W. and Pope, M.H. (1977). Facture healing in the sciatically denervated rat. *Journal of trauma*. 17(5): 355–61.
- Gartner, L.P. and Hiatt, J.L. (2007). Color textbook of Histology. 3<sup>rd</sup> ed. Saunders Elsevier. pp. 1-156.

- Garty, H., and Palmer, L.G. (1997). Epithelial sodium channels: function, structure and regulation. *Physiological reviews*. 77: (2)359–96.
- Gawenis, L.R., Ledoussal, C., Judd, L.M., Prasad, V., Alper, S.L., Stuart-Tilley, A., Woo, A.L., Grisham, C., Sanford, L.P., Doetschman, T., Miller, M.L., and Shull, G.E. (2004). Mice with a targeted disruption of the AE-2 Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchanger are achlorhydric. *The Journal of biological chemistry*. 279(29):30531-9.
- Geris, L., Gerisch, A., Sloten, J.V., Weinerb, R., and Oosterwycka, H.V. (2008). Angiogenesis in bone fracture healing: a bioregulatory model. *Journal of theoretical biology*. 251(1):137-58
- Gerstenfeld, L.C., Alkhiary, Y.M., Krall, E.A., Nicholls, F.H., Stapleton, S.N., Fitch, J.L., Bauer, M., Kayal, R., Graves, D.T., Jepsen, K.J., and Einhorn, T.A. (2006). Three-dimensional reconstruction of fracture callus morphogenesis. *The journal of histochemistry and cytochemistry*. 54(11):1215-28.
- Gibson, J.S., McCartney, D., Sumpter, J., Fairfax, T.P.A., Milner, P.I., Edwards, H.L., Wilkins, R.J.(2009). Rapid effects of hypoxia on H  $\beta$  homeostasis in articular chondrocytes. *European journal of physiology*. 458:1085–1092.
- Glowacki, J. (1998). Angiogenesis in fracture repair. *Clinical orthopaedics and related research*. 355(Suppl.). S82–S89.
- Goldhahn, J., Fron, J.M., Kanis, J., Papapoulos, S., Reginster, L.Y., Rizzoli, R., Dere, W., Mitlak, B., Tsouderos, Y., and Boonen S. (2012). Implications for fracture healing of current and new osteoporosis treatments: an ESCEO consensus paper. *Calcified tissue international*. 90(5):343-53.
- Goldschlager, T., Abdelkader, A., Kerr, J., Boundy, I. and Jenkin, G. (2010). Undecalcified bone preparation for histology, histomorphometry and fluorochrome analysis. *Journal of visualized experiments: (JoVE)*. 8; (35): 13-15.
- Gomulkiewicz, J., Miekisz, J. and Miekisz, S. (2007). Ion Transport through Cell Membrane Channels. pp. 1-21.
- Greiff, J. (1978). A method for the production of an undisplaced reproducible tibial fracture in the rat. *Injury*; 9(4):278-81.
- Grundnes, O. and Reikeras, O. (1993). The importance of the hematoma for fracture healing in rats. *Acta orthopaedica Scandinavica*. 64 (3), 340–342.
- Grundnes, O. and Reikeras, O. (1993). The role of hematoma and periosteal sealing for fracture healing in rats. *Acta orthopaedica Scandinavica*. 64 (1), 47–49.

- Guissart, C., Li, X., Leheup, B., Drouot, N., Montaut-Verient, B., Raffo, E., Jonveaux, P., Roux, A.F., Claustres, M., Fliegel, L., and Koenig, M. (2015). Mutation of SLC9A1 , encoding the major Na<sup>+</sup>/H<sup>+</sup> exchanger , causes ataxia – deafness Lichtenstein–Knorr syndrome. *Human molecular genetics*: 24(2):463-470.
- Guo, L.I., Heinzinger, N.K., Stevenson, M., Schopfer, L. M., and Salhany, J. M. (1994). Microbiology Inhibition of gpl20-CD4 Interaction and Human Immunodeficiency Virus Type 1 Infection in Vitro by Pyridoxal 5'. Antimicrobial Agents and Chemotherapy, *American Society for Phosphate* Vol. 38, pp. 2483-2487.
- Hadjiargyrou, M and O'Keefe, R. J. (2014). The Convergence of Fracture Repair and Stem Cells: Interplay of Genes, Aging, Environmental Factors and Disease. *Journal of Bone Mineral Research*. 2014 Nov; 29(11): 2307–2322.
- Halıcı, M., Öner, M., Ahmet Güney, A., Canöz, Ö. Narin, F., and Canan Halıcı, C. (2010). Melatonin promotes fracture healing in the rat model. *Eklem Hastalık Cerrahisi*. 21(3):172-177.
- Hall and Susan J. (2007). *Basic Biomechanics with OLC*. (5th ed., Revised Ed.). Burr Ridge: McGraw-Hill Higher Education. p. 88.
- Hall, A.C., and Macnicol, M.F. (2008). New insights into function of the growth plate a possible role for membrane transporters. *The Journal of bone and joint surgery. British volume*. 90(12):1541-1547.
- Hall, Arthur, C., Guyton and John, E. (2005). *Textbook of medical physiology* (11th Ed.). Philadelphia: W.B. Saunders. Pp. 981.
- Hankenson, K.D., Zmmerman, G., and Marcucio, R. (2014). Biological Perspectives of Delayed Fracture Healing. *Injury*; 45(0 2): S8–S15.
- Hannan, K.M., and Little, P.J. (1998). Mechanisms regulating the vascular smooth muscle Na<sup>+</sup>/H<sup>+</sup> exchanger (NHE-1) in diabetes. *Canadian Journal of Biochemistry and Cell Biology*. 76(5):751– 759.
- Harguindey, S., Arranz, J.L., Wahl, M.L., Orive, G., and Reshkin, S.J. (2009). Proton transport inhibitors as potentially selective anticancer drugs. *Anticancer Research*; 29(6):2127-36.
- Harguindey, S., Arranz, J.L., Wahl, M.L., Orive, G., and Reshkin, S.J. (2009). Proton transport inhibitors as potentially selective anticancer drugs. *Anticancer research*. 29:2127–36.
- Harrison's principles of internal medicine. (2008). (17<sup>th</sup> Ed.). New York [etc.]: McGraw-Hill Medical. p. 2365.
- Hatt, (2008). Hard tissue surgery. In Chitty, J. and Lierz, M. (Eds.). BSAVA manual of raptors, pigeons and passerine birds.). *British Small Animal Veterinary Association*. pp. 157-175.

- Henry, G. A. (2016). Fracture Healing and Complications. <https://veteriankey.com/fracture-healing-and-complications>.
- Hiltunen, A., Euorio, E., and Aro, H.T. (1993). A standardized experimental fracture in mouse tibia. *Journal Orthopedic Research*, 11: (2)305-12.
- Hock, J.M., Centrella, M., Canalis, E., (2004). Insulin-like growth factor I (IGF-I) has independent effects on bone matrix formation and cell replication. *Endocrinology*. 122(1):254-60.
- Hoffmann, E.K., Holm, N.B., and Lambert, I.H. (2014). Functions of volume-sensitive and calcium-activated chloride channels. *The International Union of Biochemistry and Molecular Biology*. 66(4):257-67.
- Hoffmann, E.K., Lambert, I.H., and Pedersen, S.F. (2009). Physiology of cell volume regulation in vertebrates. *Physiological reviews*. 89(1):193-277.
- Hollinger, J. and Wong, M. E. (1996). The integrated processes of hard tissue regeneration with special emphasis on fracture healing. *Oral surgery, oral medicine, oral pathology, oral radiology, and endodontics*. 82 (6), 594–606.
- Hulth, A. (1989). Current concepts of fracture healing. *Clinical orthopaedics and related research*; (249):265-84.
- Hwang, S. M., Koo, N. Y., Jin, M., Davies, A. J., Chun, G., Choi, S., Kim, J., and Park, K. (2011). Intracellular acidification is associated with changes in free cytosolic calcium and inhibition of action potentials in rat trigeminal ganglion. *The Journal of Biological Chemistry*. 286(3) 1719-1729.
- Ibberson, M., Riederer, B.M., Uldry, M., Guhl, B., Roth, J., and Thorens, B. (2002). Immunolocalization of GLUTX1 in the testis and to specific brain areas and vasopressin-containing neurons. *Endocrinology*.143 (1):276–284.
- Jackson, R.W., Reed, C.A., Israel, J.A., Abou-Keer, F.K. and Garside, H. (1970). Production of a standard experimental fracture. *Canadian journal of surgery*. 13(4):415-20.
- Jansen, I. D., Mardones, P., Lecanda, F., de Vries, T. J., Recalde, S., Hoeben, K. A., & Bronckers, A. L. (2009). AE-2a, b-Deficient mice exhibit osteopetrosis of long bones but not of calvaria. *Federation of American Societies for Experimental Biology journal FASEB Journal*. 23(10):3470-81.
- Jennings, M. L. (2005). Evidence for a Second Binding/Transport Site for Chloride in Erythrocyte Anion Transporter AE-1 Modified at Glutamate 681. *Biophysical Journal* .Volume 88, Issue 4, April pp. 2681–2691.

- Josephsen, K., Praetorius, J., Frische, S., Gawanis, L. R., Kwon, T. H., Agre, P., Nielsen, S. and Fejerskov, O. (2009). Targeted disruption of the Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchanger AE-2 results in osteopetrosis in mice. *Proceedings of the National Academy of Sciences of the United States of America.* 106(5):1638-41.
- Kalish, B.T., Kieran, M.W., Puder, M., Panigrahy, D. (2013). The growing role of eicosanoids in tissue regeneration, repair, and wound healing. *Prostaglandins & other lipid mediators.* 104-105:130-8.
- Kanai, Y., Clemenccon, B., Simonin, A., Leuenberger, M., Lochner, M., Weisstanner, M., and Hediger, M. A. (2013). The SLC1 high-affinity glutamate and neutral amino acid transporter family. *Molecular aspects of medicine.* 34(2-3):108-120.
- Kidder, L.S., Chen, X., Schmidt, A.H., and Lew, W.D. (2009). Osteogenic Protein-1 Overcomes Inhibition of Fracture Healing in the Diabetic Rat: A Pilot Study. *Clinical Orthopedic Relative Research.* 467(12):3249-3256.
- Kini, U. and Nandeesh, B.N. (2012). Physiology of Bone Formation, Remodelling, and Metabolism. *Radionuclide and Hybrid Bone Imaging,* 2: 29-57.
- Klein, M., Vignaud, J.M., Hennequin, V., Toussaint, B., Bresler, L., Plénat, F., Leclère, J., Duprez, A., and Weryha, G, (2001). Increased expression of the vascular endothelial growth factor is a pejorative prognosis marker in papillary thyroid carcinoma. *The Journal of clinical endocrinology and metabolism.* 86(2):656-8.
- Kleyman, T.R., and Cragoe, E.J. (1988). Amiloride and its analogs as tools in the study of ion transport. *The Journal of Membrane Biology.* 105: (1)1–21.
- Kopito, R.R. and Lodish, H.F. (1985). Primary structure and transmembrane orientation of the murine anion exchange protein. *Nature.* 316(6025):234-8.
- Kwong, F. N., and Harris, M. B. (2008). Recent developments in the biology of fracture repair. *The Journal of the American Academy of Orthopaedic Surgeons.* 16(11):619–25.
- Landowski, C.P., Suzuki, Y. and Hediger, M.A. (2008). The Mammalian Transporter Families. *Science Open Research.* 8. Pp. 91-146.
- Lau, K-H.W., Kothari, V., Das, A., Zhang, X-B., and Baylink, D. J. (2013). Cellular and molecular mechanisms of accelerated fracture healing by COX2 gene therapy: studies in a mouse model of multiple fractures. *Bone;* 53(2): 369-81.

- Lee, N.K., Sowa, H., Hinoi, E., Ferron, M., Ahn, J.D., Confavreux, C., Dacquin, R., Mee, P.J., McKee, M.D., Jung, D.Y., Zhang, Z., Kim, J.K., Mauvais-Jarvis, F. and Ducy, P.K.G. (2007). Endocrine regulation of energy metabolism by the skeleton. *Cell.* 130 (3): 456–469.
- Lehnart, S.E. and Marks, A.D. (2007). Transport of ions and small molecules across membranes. In: Cells. Lewin, B., Cassimeris, L., Lingappa, V.S., and Plopper, G. (editors). pp. 31-88.
- Li, X. and Fliegel, L. (2014). Functional role of arginine 425 in the mammalian  $\text{Na}^+/\text{H}^+$ : *Biochemistry and Cell Biology*. 92 (6)541-546.
- Li, X., Prins, D., Michalak, M. and Fliegel, L. (2013). Calmodulin-dependent binding to the NHE-1 cytosolic tail mediates activation of the  $\text{Na}^+/\text{H}^+$  exchanger by  $\text{Ca}_2^+$  and endothelin. *American journal of physiology. Cell physiology*. 305(11):C1161-9.
- Lienau, J., Schell, H., Duda, G.N., Seebek, P., Muchow, S., and Bail, H.J., (2005). Initial vascularization and tissue differentiation are influenced by fixation stability. *Journal of Orthopaedic Research*, 23,639–645.
- Lin, C.Y., Chang, Y.H., Lin, K.J., Yen, T.C., Tai, C.L., Chen, C.Y., Lo, W.H., Hsiao, I.T., and Hu, Y.C. (2010). The healing of critical-sized femoral segmental bone defects in rabbits using baculovirus-engineered mesenchymal stem cells. *Biomaterials*. 31(12):3222-30.
- Liu, A.H., Cao, Y.N., Liu, H.T., Zhang, W.W., Liu, Y., Shi, T.W., Jia, G.L., and Wang, X.M. (2008). DIDS attenuates staurosporine-induced cardiomyocyte apoptosis by PI3K/Akt signaling pathway: activation of eNOS/NO and inhibition of Bax translocation. *Cellular physiology and biochemistry: international journal of experimental cellular physiology, biochemistry, and pharmacology*. 22(1-4):177-86.
- Liu, A.H., Cao, Y.N., Liu, H.T., Zhang, W.W., Liu, Y., Shi, T.W., Jia, G.L., and Wang, X.M. (2008). Dids attenuates staurosporine-induced cardiomyocyte apoptosis by pi3k/akt signaling pathway: Activation of enos/no and inhibition of bax translocation. *Cellular physiology and biochemistry: international journal of experimental cellular physiology, biochemistry, and pharmacology*. 22(1-4):177-86.
- Lodish, H., Berk, A., Kaiser, C.A., Krieger, M., Bretscher, A., Ploegh, H., Amon, A. and Scott, M.P. (2013). Transmembrane Transport of Ions and Small Molecules. In: Molecular Cell Biology. (7<sup>th</sup> Ed). Macmillan. pp. 473-516.
- Loffing, J., and Kaissling, B. (2003). "Sodium and calcium transport pathways along the mammalian distal nephron: from rabbit to human". *American Journal of Physiology - Renal Physiology*. 284 (4): F628–F643.
- Loo, S.Y., Chang, M.K., Chua, C.S., Kumar, A.P., Pervaiz, S and Clement, M.V. (2012). NHE-1: a promising target for novel anti-cancer therapeutics. *Current pharmaceutical designosis*. 18(10): 1372-1382.

- Loqman, M.Y., Bush, P.G., Farquharson, C., and Hall, A.C. (2013). Suppression of mammalian bone growth by membrane transport inhibitors. *Journal of cellular biochemistry*. 114 (3):658-68.
- Lyaruu, D. M., Bronckers, A. L., Mulder, L., Mardones, P., Medina, J. F., Kellokumpu, S., Oude Elferink, R. P. and Everts, V. (2008). The anion exchanger AE-2 is required for enamel maturation in mouse teeth. *Matrix Biology*. 27(2): 119–127.
- Maae, E., Nielsen, M., Steffensen, K.D., Jakobsen, E.H., Jakobsen, A., and Sørensen, F.B. (2011). Estimation of immunohistochemical expression of VEGF in ductal carcinomas of the breast. *The Journal of Histochemistry and Cytochemistry: official journal of the Histochemistry Society*. 59(8):750-760.
- Mackie, E.J., Tatarczuch, L., and Mirams, M. (2011). The skeleton: a multi-functional complex organ: the growth plate chondrocyte and endochondral ossification. *Journal Endocrinol*. 2011 Nov; 211(2):109-21.
- Madison, M. and Martin, R. B. (1993). Fracture healing, in Operative Orthopaedics (Chapman, M. W., ed.), Lippincott, Philadelphia, pp. 221–228.
- Maggiano, I. S., Maggiano, C. M., Tiesler, V., Kierdorf, H., Stout, S. D., and Schultz, M. A., (2011). Distinct Region of Microarchitectural Variation in Femoral Compact Bone: Histomorphology of the Endosteal Lamellar Pocket. *International Journal of Osteoarchaeology*. 21: (6)743–750.
- Manigrasso, M. O and Connor, J. (2004). Characterization of a closed femur fracture model in mice. *Journal of orthopaedic trauma*. 18(10):687-95.
- Manjubala, Liu, Y., Epari, D. R., Roschger, P., Schell, H., Fratzl, P. and duda, G. n. (2009). Spatial and temporal variations of mechanical properties and mineral content of the external callus during bone healing. *Bone*. 45: (2)185-192.
- Marie, Reumann, K., Nair, T., Strachna, O., Adele, L., Boskey, A.L. and Mayer-Kuckuk, P. (2010). Production of VEGF receptor 1 and 2 mRNA and protein during endochondral bone repair is differential and healing phase specific. *Journal of Applied Physiology*. 109(6): 1930–1938.
- Marsell, R., and Einhorn, T.A. (2011). The biology of fracture healing. *Injury*, 42, (6)551-5.
- Marsh, D. R., and Li, G. (1999). The biology of fracture healing: optimising outcome. *British medical bulletin*. 55(4): 856-869.
- Martinez-Anso, E., Castillo, J. E., Diez, J., Medina, J. F. and Prieto, J. (1994). Immunohistochemical detection of chloride/bicarbonate anion exchangers in human liver. *Hepatology*; 19(6):1400-6.

- Martini, L., Fini, M., Giavaresi, G. and Giardino, R. (2001). Sheep model in orthopedic research: a literature review. *Comparative medicine*. 51(4):292-9.
- Marzona, L., and Pavolini, B. (2009). Play and players in bone fracture healing match. *Clinical Cases in Mineral and Bone Metabolism*. 6(2): 159–162.
- Masereel B., Pochet, L., and Laeckmann, D. (2003). An overview of inhibitors of  $\text{Na}^+/\text{H}^+$  exchanger. *European Journal of Medicinal Chemistry*. 38(6) 547-554.
- Masereel, B., Pochet, L., and Laeckmann, D. (2003). An overview of inhibitors of  $\text{Na}^+/\text{H}^+$  exchanger. *European journal of medicinal chemistry*. 38(6) 547-554.
- Matos, M., Araújo, F.P., and Paixão, F.B. (2008). Histomorphometric evaluation of bone healing in rabbit fibular osteotomy model without fixation. *Journal of Orthopaedic Surgery and Research*. 2008;3:4.
- Matthews, H., Ranson, M., and Kelso, M.J. (2011). Anti-tumour/metastasis effects of the potassium-sparing diuretic amiloride: an orally active anti-cancer drug waiting for its call-of-duty? *International Journal of Cancer*. 129(9):2051-61.
- McKelvay, D. and Hollingshead, K.W. (2004). *Veterinary Anesthesia and Analgesia*. 3<sup>rd</sup> ed. Mosbey. pp.: 315–349.
- McKibbin, B. (1978). The biology of fracture healing in long bones. *The Journal of bone and joint surgery. British volume*. 60-B (2):150-62.
- Medina, J.F., Recalde, S., Prieto, J., Lecanda, J., Saez, E., Funk, C.D., Vecino, P., van Roon, M.A., Ottenhoff, R., Bosma, P.J., Bakker, C.T., and Elferink, R.P. (2003). Anion exchanger 2 is essential for spermiogenesis in mice. *The National Academy of Sciences of the USA*. 100: (26)15847–15852.
- Mills, L. A. and Simpson, A. H. R. W. (2012). *In vivo models of bone repair*. *The Journal of bone and joint surgery. British volume*. 94(7):865-74.
- Miraglia, E., Viariso, D., Riganti, C., Costamagna, C., Ghigo, D., and Bosia, A. (2005).  $\text{Na}^+/\text{H}^+$  exchanger activity is increased in doxorubicin-resistant human colon cancer cells and its modulation modifies the sensitivity of the cells to doxorubicin. *International Journal Cancer*. 20; 115(6):924-9.
- Model, M.A. (2014). Possible causes of apoptotic volume decrease: an attempt at quantitative review. *American journal of physiology. Cell physiology*. 306(5):C417-24.
- Mohamad, S., Shuid, A.N., Mohamed, N., Mokhtar, F.M.A., Abdullah, S., Othman, F., Suhaimi, F., Muhammad, N., and Soelaiman, I.N. (2012). The effects of alpha-tocopherol supplementation on fracture healing in a postmenopausal osteoporotic rat model. *Clinics*. 67(9):1077-1085.

- Monfoulet, L., Rabier, B., Chassande, O., and Fricain, J-C. (2010). Drilled hole defects in mouse femur as models of intramembranous cortical and cancellous bone regeneration. *Calcified tissue international*. 86(1):72-81.
- Morgan, P.E., Supuran, C.T., and Casey, J.R. (2004). Carbonic anhydrase inhibitors that directly inhibit anion transport by the human Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchanger, AE-1. *Molecular Membrane Biology*. Nov-Dec; 21(6):423-33.
- Mounien, L., Marty, N., Tarussio, D., Metref, S., Genoux, D., Preitner, F., Foretz, M., and Thorens, B. (2010). Glut2-dependent glucose-sensing controls thermoregulation by enhancing the leptin sensitivity of NPY and POMC neurons. *Official publication of the Federation of American Societies for Experimental Biology (Faseb J)*. 24(6):1747-58
- Mueckler, M. and Thorens, B. (2013). The SLC2 (GLUT) Family of Membrane Transporters. *Molecular Aspects Medicine*. 34(0): 121–138.
- Nass, R., and Rao, R. (1998). Novel Localization of a Na<sup>+</sup> /H<sup>+</sup> Exchanger in a Late Endosomal. *The Journal of Biological Chemistry*. 273(33):21054 21060.
- Neill, K. R. O., Stutz, C. M., Mignemi, N. A., Burns, M. C., Murry, M. R., Nyman, J. S. and Schoenecker, J. G. (2012). Micro-computed tomography assessment of the progression of fracture healing in mice. *Bone*. 50(6):1357-67.
- Nemeth, G. G., Bolander, M. E., and Martin, G. R. (1988). Growth factors and their role in wound and fracture healing. *Progress in clinical and biological research*. 266, 1–17.
- Netter, F. H. (1987). Musculoskeletal system: anatomy, physiology, and metabolic disorders. Summit, New Jersey: Ciba-Geigy Corporation ISBN 0-914168-88-6, p.129.
- Newman, R. L., Duthie, R. B. and Francis, M. J. O. (1985). Nuclear magnetic resonance studies of the fracture repair. *Progress in clinical and biological research*. 198: 297-303.
- Numata, M. and Orlowski J. (2001). Molecular cloning and characterization of a novel (Na<sup>+</sup>, K<sup>+</sup>)/H<sup>+</sup> exchanger localized to the trans-Golgi network. *The Journal of Biological Chemistry*. 276, 17387-17394.
- Nunamaker, D.M. (1998). Experimental models of fracture repair. *Clinical orthopaedics and related research*, (355 Suppl.):S56-65.
- Okada, Y., Shimizu, T., Maeno, E., Tanabe, S., Wang, X., and Takahashi. N. (2006). Volume-sensitive chloride channels involved in apoptotic volume decrease and cell death. *Journal of Membrane Biology*. 209(1):21-9.

- Oryan, A., Monazzah, S., and Bigham-sadegh, A. (2015). Bone Injury and Fracture Healing Biology. *Biomedical and Environmental Sciences*. 28(1):57-71.
- Otto, T.E., Patka, P., M. and Haarman, H.J.T.M. (1995). Closed Fracture Healing: A Rat Model. *European surgical research. Europäische chirurgische Forschung. Recherches chirurgicales européennes*. 27(4):277-84.
- Öztürk, A., Yetkin, H., Memis, L., Cila, E., Bolukbasi, S. and Gemalmaz, C. (2006). Demineralized bone matrix and hydroxyapatite/tri-calcium phosphate mixture for bone healing in rats. *International Orthopaedic. Springer-Verlag*. 30(3): 147–152.
- Paine, M. L., Snead, M. L., Wang, H. J., Abuladze, N., Pushkin, A., Liu, W., Kao, L. Y., Wall, S. M., Kim, Y. H. and Kurtz, I. (2008). Role of NBCe1 and AE-2 in secretory ameloblasts. *Journal of dental research*. 87(4):391-5.
- Palacin, M., Borsani, G., and Sebastio, G. (2001). The molecular bases of cystinuria and lysinuric protein intolerance. *Current Opinion in Genetics and Development*. 11:328–335.
- Pannier, S. (2011). Congenital pseudarthrosis of the tibia. *Orthopaedics & Traumatology: Surgery & Research*. Volume 97, Issue 7, pp. 750–761.
- Parfitt, M., Drezner, M.K., Glorieux, F.H., Kanis, J.A., Malluche, H., Meunier, P.J., Ott, S.M., and Recker, R.R. (1987). Bone histomorphometry: standardization of nomenclature, symbols, and units. Report of the ASBMR Histomorphometry Nomenclature Committee. *Journal of Bone Minerals Research*. 2(6):595-610.
- Pearce, A.I., Richards, R.G., Milz, S., Schneider, E. and Pearce, S.G. (2007). Animal models for implant biomaterial research in bone: A review. *European Cells and Materials*, 13: 1-10. Philadelphia, pp.: 221–228.
- Pei, Y. and Fu, Q. (2011). Yeast-incorporated gallium promotes fracture healing by increasing callus bony area and improving trabecular microstructure on ovariectomized osteopenic rats. *Biological trace element research*. 141(1-3):207-15.
- Perren, S.M. (2014). Fracture healing: fracture healing understood as the result of a fascinating cascade of physical and biological interactions. Part I. An Attempt to Integrate Observations from 30 Years AO Research. *Acta Chirurgiae orthopaedicae et Traumatologiae czechoslovaca*. 81(6):355-64.
- Peterson, T.S., Spitsbergen, J.M., Feist, S.W. and Kent, M.L. (2011). Luna stain, an improved selective stain for detection of microsporidian spores in histologic sections. *Diseases of aquatic organisms*. 95(2):175-80.

- Phillips, A.M. (2005). Overview of the fracture healing cascade. *Injury*; 36(Suppl.3): S5–7.
- Pilia, M., Guda, T. and Appleford, M. (2013). Development of composite scaffold for load-bearing segmental bone defect (Review article). *BioMed Research International*. www.hindawi.com 2013. Volume 2013 (2013), Article ID 458253, 15 pages.
- Plopper, G., Sharp, D., and Sikorski, E. (2015). Transport of ion and Small Molecules across Membranes. In: Lewin's Cells. (3rd Ed). Jones & Bartlett Learning. pp. 229-300.
- Poupon, R., Ping, C., Chretien, Y., Corpechot, C., Chazouilleres, O., Simon, T., Heath, S. C., Matsuda, F., Poupon, R. E., Housset, C. Barbu V (2008). Genetic factors of susceptibility and of severity in primary biliary cirrhosis. *Journal of Hepatoogyl*. 49(6):1038-45.
- Putney, L.K., Denker S.P., and Barber, D.L. (2002). The changing face of the  $\text{Na}^+/\text{H}^+$  exchanger, NHE-1: structure, regulation, and cellular actions. *The Annual Review of Pharmacology and Toxicology*. 42: 527–552.
- Rahn, B. A., Gallinaro, P., Baltensperger, A., Perren, S.M. (1971). Primary bone healing. An experimental study in the rabbit. *Journal of Bone Joint Surgery. American*. 53(4), 783–786.
- Recalde, S., Muruzabal, F., Looije, N., Kunne, C., Burrell, M. A., Saez, E., Martinez-Anso, E., Salas, J. T., Mardones, P., Prieto, J., Medina, J.F., and Elferink, R.P. (2006). Inefficient chronic activation of parietal cells in AE-2a, b ( $^{-/-}$ ) mice. *The American journal of pathology*. 169(1):165-76.
- Renshaw S. (2007). Immunochemical Staining Techniques. Chapter 4.Pp. 45-96.
- Reumann, M.K., Nair, T., Strachna, O., Boskey, A.L., and Mayer-Kuckuk, P. (2010). Production of VEGF receptor 1 and 2 mRNA and protein during endochondral bone repair is differential and healing phase specific. *Journal of applied physiology*. 109(6):1930-1938.
- Rever, L.J., Manson, P.N., Randolph, M.A., Yaremchuk, M.J., Weiland, A., and Siegel, J.H. (1991). The healing of facial bone fractures by the process of secondary union. *Plastic and reconstructive surgery*. 87(3):451-8.
- Ritchie, R.O., Koester, K.J., Ionova, S., Yao, W., Lane, N.E., and Ager, J.W. (2008). Measurement of the toughness of bone: a tutorial with special reference to small animal studies. *Bone*. 43(5):798-812.
- Robert, B., and Satter, M.D. (1999). Fractures and Joints Injuries In: Text Book of disorders and injuries of the muculoskeletal system. 3<sup>rd</sup>. Pp. 415-437.

- Rokkanen, P., and Slätis, P. (1964). The repair of experimental fractures during long-term anticoagulant treatment. *Acta orthopaedica Scandinavica*. 35:21-38.
- Romero, M. F., Chen A., Parker, M. D., and Boron W. F. (2013). The SLC4 Family of Bicarbonate ( $\text{HCO}_3^-$ ) Transporters. *Molecular aspects of medicine*. April; 34(2-3): 159–182.
- Ross, M.H. and Pawlina, W. (2006). Connective Tissue in: Histology a Text and Atlas with Correlated Cell and Molecular Biology 5<sup>th</sup> Ed. Pp. 146-236.
- Rotin, D., Steele-Norwood, D., Grinstein, S., and Tannock, I. (1989). Requirement of the  $\text{Na}^+/\text{H}^+$  exchanger for tumor growth. *Cancer research*. 1; 49(1):205-11.
- Rundle, C.H., Strong, D.D., Chen, S.T., Linkhart, T.A., Sheng, M.H., Wergedal, J.E., Lau, K.H., and Baylink, D.J. (2008). Retroviral-based gene therapy with cyclooxygenase-2 promotes the union of bony callus tissues and accelerates fracture healing in the rat. *The journal of gene medicine*. 10(3):229-41.
- Sagalovsky, S. (2013). Bone remodeling: cellular-molecular biology and cytokine RANK-RANKL-Osteoprotegerin (OPG) system and growth factors. *Crimean Journal of Experimental and Clinical Medicine*, 3(1-2), 36-44.
- Salas, J. T., Banales, J. M., Sarvide, S., Recalde, S., Ferrer, A., Uriarte, I., Oude Elferink, R. P., Prieto, J. and Medina, J. F. (2008). AE-2a, b-deficient mice develop antimitochondrial antibodies and other features resembling primary biliary cirrhosis. *Gastroenterology*. 134(5):1482-93.
- Saraf, S. and Kumaraswamy, V. (2013). Basic research: Issues with animal experimentations. *Indian Journal Orthopedic*. 47 (1): 6-9.
- Saris, D.B., Vanlauwe, J., Victor, J., Haspl, M., Bohnsack, M., Fortems, Y., Vandekerckhove, B., Almqvist, K.F., Claes, T., Handelberg, F., Lagae, K., van der Bauwheide, J., Vandenneucker, H., Yang, K.G., Jelic, M., Verdonk, R., Veulemans, N., Bellemans, J., and Luyten, F.P. (2008). Characterized chondrocyte implantation results in better structural repair when treating symptomatic cartilage defects of the knee in a randomized controlled trial versus microfracture. *The American journal of sports medicine*. 36(2):235-46.
- Sarmiento, A., & Latta, L. (1995). Functional fracture bracing: tibia, humerus, and ulna. Springer Science & Business Media.
- Sathyendra, V. and Darowish, M. (2013). Basic science of bone healing. *Hand Clinical*. 29 (4): 473-81.
- Schenk, R. and Willenegger, H. (1967). Morphological findings in primary fracture healing: callus formation. *Simple Biology Hungar*. 7, 75–80.

- Schindeler, A., Morse, A., Harry, L. (2008). Models of tibial fracture healing in normal composition, density, and quality: potential implications for in vivo bone research and Nf1-deficient mice. *Journal of orthopaedic research*. 26(8):1053-60.
- Schneider, B.P., Wang, M., Radovich, M., Sledge, G.W., Badve, S., Thor, A., Flockhart, D.A., Hancock, B., Davidson, N., Gralow, J., Dickler, M., Perez, E.A., Cobleigh, M., Shenkier, T., Edgerton, S., Miller, K.D. (2008). Association of vascular endothelial growth factor and vascular endothelial growth factor receptor-2 genetic polymorphisms with outcome in a trial of paclitaxel compared with paclitaxel plus bevacizumab in advanced breast cancer: ECOG 2100. *Journal of Clinical Oncology*. 26(28):4672-8.
- Schwarz, C., Peters, A., Schmidt-Bleek, K., Ellinghaus, A., Duda, G.N., Schell, H., and Lienau, J., (2011). Histological analysis of the processes underlying non-healing of a segmental bone defect in a rat model. In:Proceedings of the Transactions of the 57<sup>th</sup> Annual Meeting of the Orthopaedic Research Society, California, USA, vol. 36, Long Beach, CA, 2011.
- Sengupta, P. (2013). The Laboratory Rat: Relating Its Age With Human's. *International Journal of Preventive Medicine*. 4(6): 624–630
- Sfeir, C., Ho, L., Doll, B. A., Azari, K. and Hollinger, J.O. (2005). Fracture Repair. *Bone Regen. Repair Biology Clinical Application*. (11): 21-44.
- Shapiro, F. (2008). Bone Development and its Relation to Fracture Repair. The Role of Mesenchymal Osteoblasts and Osteoblasts. *European Cells and Materials*. 15: 53-76.
- Shetty, B., Udyavar, A., Rao, C.P., Bidarkotimath, S., and Santhosh, V.S. Kuriakose (2014). Effects of ofloxacin on experimental fracture healing in Wister rats. *Journal of Drug Delivery & Therapeutics*. 4(1), 53-58.
- Singer, S. J., and Nicolson, G. L. (1972). The fluid mosaic model of the structure of cell membranes. *Science*. 175(4023):720-31.
- Sisask, G., Marsell, R., Sundgren-Andersson, A., Larsson, S., Ljunggren, O.N.O., Kenneth, B. and Jonsson. (2013). Rats treated with AZD2858, a GSK3 inhibitor, heal fractures rapidly without endochondral bone formation. *Bone*. 54(1):126-32.
- Slätis, P. and Rokkanen, P. (1965). The normal repair of experimental fractures. *Acta Orthopaedica Scandinavica*. 36:3, 221-229.
- Slätis, P., and Rokkanen, P. (1967). The mineral phase in the repair of experimental fractures. *Annales chirurgiae gynaecologiae Fenniae*. 56(2):193-201.
- Slepkov, E.R., Rainey, J.K., Sykes, B.D., and Fliegel, L. (2007). Structural and functional analysis of the Na<sup>+</sup>/H<sup>+</sup> exchanger. *Biochemical Journal*. 401(Pt. 3): 623–633.

- Sowash J.R. (2009). Rat Dissection. pp. 4-28.
- Sykaras, N. and Opperman, L. A. (2003). Bone morphogenetic proteins (BMPs): how do they function and what can they offer the clinician? *Journal of oral science*. 45(2): 57-74.
- Tanner, M.J. (1993). Molecular and cellular biology of the erythrocyte anion exchanger (AE-1). *Seminars in haematology*. 30(1):34-57.
- Tattersall AL, Meredith D, Furla P, Shen MR, Ellory JC, Wilkins RJ. 2003. Molecular and functional identification of the  $\text{Na}^+/\text{H}^+$  exchange isoforms NHE1 and NHE3 in isolated bovine articular chondrocytes. *Cellular Physiology, Biochemistry*. 13(4):215–222.
- Tattersall, A.L., and Wilkins, R.J. (2008). Modulation of  $\text{Na}^+/\text{H}^+$  exchange isoforms NHE1 and NHE3 by insulin-like growth factor-1 in isolated bovine articular chondrocytes. *Journal of Orthopaedic Surgery and Research*. 26:1428–1433.
- Thomas, T. A. (1998). The cell and molecular biology of fracture healing. *Clinical Orthopaedics & Related Research*. Fracture Healing Enhancement. 1(355 Suppl.), S7–S21.
- Thorens, B., Cheng, Z.Q., Brown, D., and Lodish, H.F. (1990). Liver glucose transporter: a basolateral protein in hepatocytes and intestine and kidney cells. *American Journal Physiology*. 259(6 Pt. 1):C279–285.
- Thorens, B., Mueckler, M. (2010). Glucose transporters in the 21<sup>st</sup> Century. *American Journal Physiology Endocrinology Metabolisms*. 298(2):E141–145.
- Tietz, P. S., Marinelli, R. A., Chen, X. M., Huang, B., Cohn, J., Kole, J., McNiven, M. A., Alper, S. and LaRusso, N. F. (2003). Agonist-induced coordinated trafficking of functionally related transport proteins for water and ions in cholangiocytes. *Journal Biology Chemistry*. 278, 20413 -20419.
- Tsiridis, E., Upadhyay, N. and Giannoudis, P. (2007). Molecular aspects of fracture healing: which are the important molecules? *Injury*. 38 Suppl. 1:S11-25.
- Urner, F., and Sakkas, D. (1999). A possible role for the pentose phosphate pathway of spermatozoa in gamete fusion in the mouse. *Biology of Reproduction*. 60(3):733–739.
- Uusitalo, H., Rantakokko, J., Ahonen, M., Jamsa, T., Tuukkanen, J., Kahari, V.M., Vuorio, E. and Aro, H. T. (2001). A metaphyseal defect model of the femur for studies of murine bone healing. *Bone*. 28(4):423-429.
- Von, R.B. (2015). Animal models in bone repair. *Disease Models*, 13, 23-27.

- Wang, X., Cao, Y., Shen, M., Wanga, B., Zhang, W., Liua, Y., Hea, X., Wanga, L., Xiaa, Y., Dinga, M., Xuc, X., and Rena, J. (2015). DIDS Reduces Ischemia/Reperfusion-Induced Myocardial Injury in Rats. *Cellular Physiology and Biochemistry*. 35(2):676-688.
- Wells, R.G., and Hediger, M.A. (1992). Cloning of a rat kidney cDNA that stimulates dibasic and neutral amino acid transport and has sequence similarity to glucosidases. *Proceedings of the National Academy of Sciences U S A*. 89:5596–5600.
- Wildemann, B., Lange, K., Strobel, C., Fassbender, M., Willie B. and Schmidmaier, G. (2011). Local BMP-2 application can rescue the delayed osteotomy healing in a rat model. *Injury*. 42(8), 746-752.
- Wittenburg, G., Volkel, C., Mai, R., Lauer, G. (2009). Immunohistochemical comparison of differentiation markers on paraffin and plastic embedded human bone samples. *Journal Physiology Pharmacology*, 60 (Suppl. 8):43–49.
- Wu, J., Glimcher, L. H. and Aliprantis, A. O. (2008).  $\text{HCO}_3^-/\text{Cl}^-$  anion exchanger SLC-4A2 is required for proper osteoclast differentiation and function. *Proceedings of the National Academy of Sciences of the United States of America*. 105(44):16934-9.
- Xu, X., Pacheco, B.D., Leng, L., Bucala, R., Ren, J. (2013). Macrophage migration inhibitory factor plays a permissive role in the maintenance of cardiac contractile function under starvation through regulation of autophagy. *Cardiovascular Research*. 99(3):412-21.
- Xue, L. Aihara, E. Wang, T. C. and Montrose, M. H. (2011). Trefoil factor 2 requires  $\text{Na}^+/\text{H}^+$  exchanger 2 activity to enhance mouse gastric epithelial repair. *The Journal of Biological Chemistry*. 286(44) 38375-38382.
- Yang, X., Wang, D., Dong, W., Song, Z., and Dou, K. (2010). Inhibition of  $\text{Na}^+/\text{H}^+$ exchanger 1 by 5- (N ethyl-N-isopropyl) amiloride reduces hypoxia-induced hepatocellular carcinoma invasion and motility. *Cancer letters*. 295(2):198-204.
- Yegengil, C. (2011). Fracture Healing in a Denervation and/or Nerve Ending Interpositioning Model in the Rat. *International Journal in Clinical Medicine*. Vol.2 No.3, 301-306.
- Yeh, W.L., Lin, C.J., and Fu, W.M. (2008). Enhancement of glucose transporter expression of brain endothelial cells by vascular endothelial growth factor derived from glioma exposed to hypoxia. *Molecular Pharmacology*. 73(1):170–177.
- Young, B., Lowe, J.S., Stevens, A., Heath, J. W., and Deakin, P.J. (2006). Wheater's functional histology: a text and colour atlas (5th Ed.). [Edinburgh?]: Churchill Livingstone/Elsevier. ISBN 978-0-443-068-508. Drawings by Philip J.

- Yu, L., and Hales, C.A. (2011). Silencing of sodium-hydrogen exchanger 1 attenuates the proliferation, hypertrophy, and migration of pulmonary artery smooth muscle cells via E2F1. *American Journal of Respiratory Cell and Molecular Biology* 45(5):923– 930.
- Yu, Y.Y, Lieu, S., Lu, C., and Colnot, C. (2010). Bone morphogenetic protein 2 stimulates endochondral ossification by regulating periosteal cell fate during bone repair. *Bone*. 47(1):65-73.
- Yukata, K., Xie, C., Li, T-F., Takahata, M., Hoak, D., Kondabolu, S., Zhang, X., Awad, H. A., Schwarz, E. M., Beck, C. A., Jonason, J. H. and O'Keefe, R. J. (2014). Aging periosteal progenitor cells have reduced regenerative responsiveness to bone injury and to the anabolic actions of PTH 1-34 treatment. *Bone*. 62: 79-89.
- Zerangue, N., Kavanaugh, M.P. (1996). ASCT-1 is a neutral amino acid exchanger with chloride channel activity. *The Journal of biological chemistry*. 271 (45), 27991–27994.
- Zheng, X.B., Wang, R., Yang, H.L., and Sun, X.L. (2013). Effects of chloride ion channel and its blockers on myocardial ischemia reperfusion arrhythmias in rabbits. *Zhonghua Yi Xue Za Zhi*. 93(15):1168-73.