



UNIVERSITI PUTRA MALAYSIA

***CLINICOPATHOLOGICAL ASSESSMENT OF INTRAMAMMARY  
ADMINISTRATION OF GENETICALLY ENGINEERED *Salmonella agona*  
(4KA32) IN DOMESTIC DOGS***

SUJEY KUMAR RAJENDREN

FPV 2018 3



**CLINICOPATHOLOGICAL ASSESSMENT OF INTRAMAMMARY  
ADMINISTRATION OF GENETICALLY ENGINEERED *Salmonella*  
*agona* (4KA32) IN DOMESTIC DOGS**

By

**SUJEY KUMAR RAJENDREN**

Thesis Submitted to the School of Graduate Studies, Universiti  
Putra Malaysia, in Fulfilment of the Requirements for the Degree of  
**Master of Veterinary Science**

November 2017

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Veterinary Science

**CLINICOPATHOLOGICAL ASSESSMENT OF INTRAMAMMARY  
ADMINISTRATION OF GENETICALLY ENGINEERED *Salmonella agona*  
(4KA32) IN DOMESTIC DOGS**

By

**SUJAY KUMAR RAJENDREN**

**November 2017**

**Chair: Assoc. Prof. Gayathri Thevi Selvarajah, PhD  
Faculty: Veterinary Medicine**

Salmonella is a good bacterial candidate for anti-cancer therapy because of its characteristics such as motile, facultative anaerobiosis, and the ability to invade epithelial cells and engineered auxotrophies. *Salmonella enterica* serovar Agona is less pathogenic compared to *Salmonella enterica* serovar Typhimurium. *Salmonella enterica* serovar Agona can be genetically engineered to reduce further its pathogenic effects on hosts by eliminating the virulence genes of SopB and SopD. Genetically engineered *Salmonella enterica* serovar Agona (4KA32) was tested on mice model and showed 100% survival rate over two months period. The pathogenicity of 4KA32 in dogs are undefined. The objectives of this research are to determine the preliminary clinical and pathological effects of intra-mammary administration of 4KA32 in seven experimental domestic healthy dogs. Dogs were subjected to either single or repeated dosing over three weeks and euthanized at the end of study period for histopathology evaluation. 4KA32 was administered at various concentrations from  $1 \times 10^6$  CFU/mL to  $1 \times 10^9$  CFU/mL for both single and repeated dosing. There was significant increase ( $p < 0.05$ ) of monocyte count across all the dogs at Day 1 and persisted until the end of the study. The monocyte count of dog injected higher doses decreased significantly ( $p < 0.05$ ) when compared to monocyte count of dog injected with dose  $1 \times 10^6$  CFU/mL. There was significant increase ( $p < 0.05$ ) of neutrophils count across all the dogs at Day 1 which persisted until Day 4. The neutrophil count of dogs injected with single dose of  $1 \times 10^8$  CFU/mL and  $1 \times 10^9$  CFU/mL showed significant neutrophilia compared to single dose  $1 \times 10^6$  CFU/mL. The neutrophils count of dogs injected with repetitive dose of  $1 \times 10^7$  CFU/mL and  $1 \times 10^9$  CFU/mL significantly increased compared neutrophils count of dogs injected with dose  $1 \times 10^6$  CFU/mL. Lymphocytosis was not observed in dogs injected with repetitive dose of 4KA32. Whereas, dogs injected with single dose of 4KA32 showed significant lymphocytosis ( $p < 0.05$ ) on Day 4 only. Comparing lymphocyte count across the concentration, dogs injected with single dose of

$1 \times 10^6$  CFU/mL of 4KA32 showed lymphocytosis and lymphocyte count for higher doses significantly decreased ( $p < 0.05$ ). Serial faecal and blood samples were cultured negative for 4KA32 growth. All dogs demonstrated superficial inflammation and swelling on the injected sites which healed naturally by Day 3. The highest concentrations of 4KA32 at  $1 \times 10^9$  CFU/mL resulted in formation of superficial ulceration on Day 5 that gradually healed within seven days. None of the dogs developed fever or systemic signs including diarrhoea or inappetance during the experimental observation. Histopathology analysis revealed, mild to moderate infiltration of inflammatory cells on mammary gland of dogs injected with dose  $1 \times 10^6$  CFU/mL to  $1 \times 10^8$  CFU/mL. Severe infiltration of inflammatory cells was observed with dog injected with dose  $1 \times 10^9$  CFU/mL of 4KA32. Histopathology of intestine revealed dog injected with  $1 \times 10^7$  CFU/mL developed mild to moderate ulceration whereas dog injected with  $1 \times 10^9$  CFU/mL developed moderate to severe ulceration. This study provides evidence that 4KA32 has minimal to none systemic pathogenic effects with potential to induce local inflammation in mammary tissues. Detection of 4KA32 isolates within the lung, mesenteric lymph node and kidney without pathogenic lesions suggest that this bacteria can have potential to target metastasis of cancer cells which is common to these sites. Other tissues were negative for 4KA32. There was minimal pathogenicity observed in all the dogs injected with 4KA32. This suggest that 4KA32 is safe to be administered in dogs.

**Keyword:** *Salmonella enterica* serovar Agona, dog, mammary gland, pathogenicity

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Master Sains Veterinar

**PENILAIAN KLINIKOPATOLOGIK DALAM ANJING TEMPATAN SELEPAS  
PENYUNTIKAN *Salmonella agona* (4KA32) YANG GENETIKNYA DIUBAH  
SUAI PADA KALENJAR SUSU**

Oleh

**SUJEY KUMAR RAJENDREN**

**November 2017**

**Pengerusi: Prof. Madya. Gayathri Thevi Selvarajah, PhD**  
**Fakulti: Perubatan Veterinar**

Salmonella merupakan bakteria yang baik untuk digunakan sebagai terapi anti-kanser kerana ciri-cirinya seperti motil, anaerobiosis fakultatif, dan berupaya menyerang sel epitelium dan auxotrophies. *Salmonella enterica* serovar Agona boleh diubah secara genetik untuk mengurangkan kesan patogeniknya pada sel hos melalui perubahan pada gen virulensi SopB dan SopD. *Salmonella enterica* serovar Agona (4KA32) yang diubah suai genetiknya, telah dikaji pada modal tikus, dimana ia menunjukkan 100% kadar hidup selama dua bulan. Patogenisiti 4KA32 pada anjing belum dikaji lagi. Objektif penyelidikan ini adalah untuk menentukan kesan klinikal dan patologi awal selepas penyuntikan 4KA32, pada tujuh ekor anjing. Anjing-anjing tertakluk kepada sama ada dos tunggal atau berulang selama tiga minggu dan kemudian dimatikan untuk penilaian histopatologi pada akhir eksperimen. 4KA32 diberikan dengan pelbagai julat dos daripada  $1 \times 10^6$  CFU/mL hingga  $1 \times 10^9$  CFU/mL untuk kedua-dua rejim dos. Terdapat peningkatan ketara ( $p < 0.05$ ) bagi jumlah monosit pada kesemua anjing pada hari pertama selepas inokulasi dan ia berterusan hingga ke akhir eksperimen. Jumlah monosit untuk anjing yang disuntik dengan dos tinggi berkurang secara ketara ( $p < 0.05$ ) berbanding jumlah monosit anjing yang disuntik dos  $1 \times 10^6$  CFU/mL. Jumlah neutrofil meningkat secara ketara ( $p < 0.05$ ) pada semua anjing pada hari pertama dan ia berterusan hingga hari ke-4. Tedapat neutrofilia yang ketara ( $p < 0.05$ ) pada anjing yang disuntik dos tunggal  $1 \times 10^8$  CFU/mL dan  $1 \times 10^9$  CFU/mL berbanding dengan jumlah neutrofil pada anjing yang disuntik dos tunggal  $1 \times 10^6$  CFU/mL. Jumlah neutrofil pada anjing yang disuntik 4KA32 berulang kali dengan dos  $1 \times 10^7$  CFU/mL dan  $1 \times 10^9$  CFU/mL menunjukkan kadar peningkatan neutrofil yang ketara ( $p < 0.05$ ) berbanding jumlah neutrofil pada anjing yang disuntik 4KA32 berulang kali dengan dos  $1 \times 10^6$  CFU/mL. Limfositosis tidak dilihat pada anjing –anjing yang disuntik dos berulang, tetapi anjing yang disuntik dos tunggal 4KA32 menunjukkan limfositosis yang ketara ( $p < 0.05$ ) pada hari ke-4 sahaja. Perbandingan jumlah limfosit untuk semua dos

menunjukkan limfositosis pada anjing yang disuntik dos tunggal  $1\times10^6$  CFU/mL dan jumlah limfosit pada dos tinggi menunjukkan pengurangan yang ketara ( $p < 0.05$ ) berbanding dos  $1\times10^6$  CFU/mL. Sampel-sampel najis dan darah didapati negatif untuk kultur 4KA32. Semua anjing menunjukkan keradangan dan bengkak pada kelenjar mammari yang disuntik dimana ianya sembah secara semulajadi pada hari ke-3. Jumlah tertinggi 4KA32 iaitu pada  $1\times10^9$  CFU/mL didapati menyebabkan pembentukan ulser pada hari ke-5 suntikan yang sembah beransur-ansur dalam hari ke-7. Tiada anjing yang mengalami demam atau tanda-tanda sistemik termasuk cirir-birit atau kurang selera makan semasa pemerhatian eksperimen. Pada pemeriksaan histopatologi tisu, peyusupan sel-sel inflamasi yang sedikit hingga sederhana dilihat pada kelenjar susu anjing yang disuntik dos  $1\times10^6$  CFU/mL hingga  $1\times10^8$  CFU/mL. Peyusupan sel-sel inflamasi yang teruk dilihat pada kelenjar susu anjing yang disuntik dos  $1\times10^9$  CFU/mL 4KA32. Pemeriksaan histopatologi pada usus anjing yang disuntik dos  $1\times10^7$  CFU/mL, menunjukkan ulser yang sedikit hingga sederhana, malah usus anjing yang disuntik dos  $1\times10^9$  CFU/mL menunjukkan ulser yang sederhana hingga teruk. Kajian ini membuktikan bahawa 4KA32 tidak mempunyai sebarang kesan patogenik sistemik yang berpotensi mendorong keradangan dalam tisu-tisu mammari. Pengesanan *Salmonella* di dalam paru-paru, nodus limfa mesentrik, dan buah pinggang tanpa lesi patogen mencadangkan bahawa bakteria ini boleh berpotensi untuk menyasarkan metastasis sel-sel kanser yang biasa merebak ke organ-organ tersebut. Organ dan tisu lain tidak menunjukkan isolasi bakteria tersebut. Kajian ini menunjukkan, patogenik yang minima ditunjukkan oleh 4KA32 dan ia selamat untuk disuntik pada anjing.

**Kata Kunci:** *Salmonella enterica* serovar Agona, anjing, kelenjar susu, patogenik

## **ACKNOWLEDGEMENTS**

I would like to express my deepest gratitude to my supervisors, Assoc. Prof. Dr. Gayathri Thevi Selvarajah and Assoc Prof. Dr. Cheah Yoke Kqueen for their excellent guidance, caring, patience and providing me with an excellent atmosphere for doing research. I would like to thank Dr. Reuben Sunil Kumar Sharma for letting me use parasite laboratory for running some of my blood samples. Besides that, I would like to convey my gratitude to Dr. Suzanne Khoo, who as a friend, was always willing to help and give his best suggestions. It would have been a lonely lab without her.

I would like to thank my family and friends, who were there, cheering me up and stood by me through the good times and bad. Every challenging work needs self-efforts as well as guidance of elders especially those who were very close to our heart. My humble effort I dedicate to my loving family, whose affection, love, encouragement and prays of day and night make me able to finish my task.

I certify that a Thesis Examination Committee has met on 1<sup>st</sup> November 2017 to conduct the final examination of Sujey Kumar Rajendren on his thesis entitled "Clinicopathological Assessment of Intramammary Administration of Genetically Engineered *Salmonella Agona* (4KA32) in Domestic Dogs" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Veterinary Science.

Members of the Thesis Examination Committee were as follows:

**Faez Firdaus Jesse Abdullah, PhD**

Associate Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Chairman)

**Siti Khairani Bejo, PhD**

Associate Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Internal Examiner)

**Sailasuta Achariya, PhD**

Associate Professor  
Faculty of Veterinary Science  
Chulalongkorn University  
10330 Bangkok  
(External Examiner)

---

**NOR AINI AB. SHUKOR, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 29<sup>th</sup> January 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for Master of Veterinary Science. The members of the Supervisory Committee were as follows:

**Gayathri Thevi Selvarajah, PhD**

Associate Professor

Faculty of Veterinary Medicine

Universiti Putra Malaysia

(Chairman)

**Cheah Yoke Kqueen, PhD**

Associate Professor

Faculty of Medicines and Health Sciences

Universiti Putra Malaysia

(Member)

---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

## **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: \_\_\_\_\_

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_

Name of Chairman  
of Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_

Name of Member of  
Supervisory  
Committee: \_\_\_\_\_

## TABLE OF CONTENT

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xii
<b>LIST OF FIGURES</b>	xiv
<b>LIST OF ABBREVIATIONS</b>	xvi
CHAPTER	
<b>1 INTRODUCTION</b>	1
<b>2 LITERATURE REVIEW</b>	3
2.1 Canine Mammary Gland Tumours	3
2.2 Development of Mammary Gland	3
2.3 Incidence	4
2.4 Risk Factor for Mammary Gland Tumour Development in Dog	4
2.5 Tumour Classification	5
2.6 Diagnostic Investigation	5
2.6.1 Diagnostic cytology	5
2.6.2 Diagnostic imaging	6
2.6.3 Histopathology	6
2.7 Therapeutic Modalities	6
2.7.1 Surgical excision	7
2.7.2 Radiotherapy	7
2.7.3 Hormonal therapy	7
2.7.4 Chemotherapy	8
2.7.5 Immunotherapy	8
2.7.6 Oncolytic virotherapy	9
2.8 Using Bacterial as Anti-cancer Agent	9
2.8.1 <i>Salmonella</i> sp as anti-cancer therapy	12
2.8.2 <i>Salmonella enterica</i> serovar Typhimurium as tumour therapy	13
2.8.3 <i>Salmonella enterica</i> serovar Agona as anti-cancer agent	14
<b>3 MATERIALS AND METHODS</b>	16
3.1 Ethic Approval	16
3.2 Animal, Inclusion and Exclusion Criteria	16
3.3 Preparation of Agent	16
3.4 Experimental Design	17

3.5	Administration of Bacteria into Experimental Dogs	20
3.6	Clinical Observation During the Experimental Duration	20
3.7	Blood Sampling for Haematology Profiles	20
3.8	Post Mortem Evaluation and Histopathology	20
3.9	Faecal, Blood and Tissue Culture for 4KA32	21
3.10	Statistical Analysis	22
<b>4</b>	<b>RESULTS AND DISCUSSION</b>	<b>23</b>
4.1	Pre-experiment Clinical Observation	23
4.2	Single Dose of 4KA32 Administration in Dog	23
4.2.1	Clinical observation	21
4.2.2	Observation at the injection site after single dose of 4KA32 in Dogs	26
4.2.3	Faecal and blood culture	34
4.2.4	Haematology	34
4.3	Repetitive dose of 4KA32 administration in dogs	43
4.3.1	Clinical observation	43
4.3.2	Observation at the injection site after repetitive dose of 4KA32 in dogs	46
4.3.3	Faecal and blood culture	54
4.3.4	Haematology	54
4.3.5	Post mortem examination	62
4.3.6	Tissue culture for <i>Salmonella</i> sp	67
4.3.7	Histopathology	69
<b>5</b>	<b>GENERAL DISCUSSION AND CONCLUSION</b>	<b>81</b>
<b>REFERENCES</b>		<b>84</b>
<b>APPENDICES</b>		<b>93</b>
<b>BIODATA OF STUDENT</b>		<b>107</b>
<b>PUBLICATION</b>		<b>108</b>

## LIST OF TABLES

<b>Table</b>		<b>Page</b>
1	Three categories of bacteria that have been used in oncolytic studies and their advantages and disadvantages	11
2	Temperature values for up to 21 days after administration of single dose of 4KA32 into experimental dog's mammary gland	24
3	Faecal scoring for up to 21 days after administration of a single dosing of 4KA32 into experimental dog's mammary glands	25
4	Measurement of swelling/ lesion at the widest length on the mammary gland after a single dose administration of 4KA32 in dogs.	29
5	Neutrophil values ( $\times 10^9/L$ ) after the administration of a single dose of 4KA32 into mammary gland	36
6	Lymphocyte values ( $\times 10^9/L$ ) after the administration of single dose of 4KA32 into mammary gland	39
7	Monocyte values ( $\times 10^9/L$ ) after the administration of a single dose of 4KA32 into mammary gland	42
8	Temperature values for up to 21 days after administration of repetitive dose of 4KA32 into experimental dog's mammary glands	44
9	Faecal scoring for up to 21 days after administration of a repetitive dosing of 4KA32 into experimental dog's mammary gland	45
10	Measurement of swelling/ lesion at the widest length on the mammary gland after a repetitive dose administration of 4KA32 in dogs	49
11	Neutrophil values ( $\times 10^9/L$ ) after the administration of a repetitive dose of 4KA32 into mammary gland	56
12	Lymphocyte count ( $\times 10^9/L$ ) after the administration of repetitive dose of 4KA32 into mammary gland.	58
13	Monocyte count ( $\times 10^9/L$ ) after the administration of repetitive dose of 4KA32 into mammary gland.	61

14	Hyperemia observed during post-mortem examination after the administration of repetitive dose of 4KA32 into mammary gland	63
15	Congestion and haemorrhage observed during post-mortem examination after administration of repetitive dose of 4KA32 into mammary gland	64
16	Abscess observed during post-mortem examination after administration of repetitive dose of 4KA32 into mammary gland	65
17	Oedema observed during post-mortem examination after administration of repetitive dose of 4KA32 into mammary gland	66
18	Tissue cultures of the dogs administered with 4KA32	68
19	Histopathological alteration in dogs injected with 4KA32 into mammary gland	79

## LIST OF FIGURES

<b>Figure</b>		<b>Page</b>
1a	Experimental design for phase 1 study to evaluate recommended dose for 4KA32	18
1b	Experimental design for cross over design (pilot evaluation) study to evaluate pathogenicity at higher dose for 4KA32	19
2	Observation of injection site after administration of a single dose of 4KA32 ( $1 \times 10^6$ CFU/mL) in RS1 and RS2	30
3	Observation of injection site after administration of a single dose of 4KA32 ( $1 \times 10^7$ CFU/mL) in RS5	31
4	Observation of injection site after administration of a single dose of 4KA32 ( $1 \times 10^8$ CFU/mL) in RS6	32
5	Observation of injection site after administration of a single dose of 4KA32 ( $1 \times 10^9$ CFU/mL) in RS7	33
6	Observation of injection site after administration of a repetitive dose of 4KA32 ( $1 \times 10^6$ CFU/mL) in RS3 and RS4	50
7	Observation of injection site after administration of a repetitive dose of 4KA32 ( $1 \times 10^7$ CFU/mL) in RS5	51
8	Observation of injection site after administration of a repetitive dose of 4KA32 ( $1 \times 10^8$ CFU/mL) in RS6	52
9	Observation of injection site after administration of a repetitive dose of 4KA32 ( $1 \times 10^9$ CFU/mL) in RS7	53
10	Histopathology of mammary gland of RS3	71
11	Histopathology of mammary gland of RS5	72
12	Histopathology of mammary gland of RS6	73
13	Histopathology of mammary gland of RS7	74
14	Histopathology of intestine of RS5	75
15	Histopathology of intestine of RS7	76
16	Histopathology of lung of RS5	77



## LIST OF APPENDICES

<b>Appendix</b>		<b>Page</b>
1	Complete blood and serum biochemistry profile for RS1	93
2	Complete blood and serum biochemistry profile for RS2	94
3	Complete blood and serum biochemistry profile for RS3	95
4	Complete blood and serum biochemistry profile for RS4	96
5	Complete blood and serum biochemistry profile for RS5 (single dose)	97
6	Complete blood and serum biochemistry profile for RS6 (single dose)	98
7	Complete blood and serum biochemistry profile for RS7 (single dose)	99
8	Complete blood and serum biochemistry profile for RS5 (repetitive dose)	100
9	Complete blood and serum biochemistry profile for RS6 (repetitive dose)	101
10	Complete blood and serum biochemistry profile for RS7 (repetitive dose)	102
11	Complete blood and serum biochemistry profile for RS8 (single dose with WA32)	103
12	Fecal Scoring	104
13	IACUC approval letter	105

## LIST OF ABBREVIATIONS

TNF- $\alpha$	Tumour necrosis factor alpha
Lox	Lysyl oxidase
SPI	<i>Salmonella</i> pathogenic Island
CMT	Canine mammary gland tumour
OHE	Ovariohysterectomy
PD-1	Programmed death 1
CTLA-4	Cytotoxic T-lymphocyte-associated protein 4
T1SS	Type One Secretion System
XLD	xylose lysine deoxycholate
HE	Hektoen enteric
TSI	triple sugar iron
LIA	lysine iron agar
M cells	Microfold cells
PSA	Prostate Specific Antigen
<i>Klk1b22</i>	Kallikrein 1-related peptidase b22
TRAIL	TNF- related apoptosis-inducing ligand
DR4	Death receptor 4
DR5	Death receptor 5
FASL	Fas ligand
IL-2	Interleukin 2
IL-4	Interleukin 4
IL-18	Interleukin 18

CC chemokine-21	Chemokine (C-C motif) ligand 21
TNFSF14	Tumour necrosis factor superfamily member 14
NK	Natural killer
T3SS	Type three secretion system
CO <sub>2</sub>	Carbon dioxide
xg	Times gravity
Na <sub>2</sub> HPO <sub>4</sub> ,	Sodium phosphate
KH <sub>2</sub> PO <sub>4</sub> ,	Potassium phosphate
NaCl	Sodium chloride
NH <sub>4</sub> Cl	Ammonium chloride
MgSO <sub>4</sub>	Magnesium sulphate
CaCl <sub>2</sub>	Calcium chloride
CFU	Colony forming unit

## CHAPTER 1

### INTRODUCTION

*Salmonella entericar* serovar Agona was first discovered in the year 1952, in Ghana (Hoffmann *et al.*, 2015). It is considered a zoonotic organism, which can cause various infections in humans and animal. Transmission of this serovar from animals to humans occurs by ingestion of contaminated beef, pork, chicken, turkey meat cereals or tea (Michael *et al.*, 2006).

*Salmonella sp.* which are gram-negative, facultative anaerobes that causes intestinal infections. Surprising, *Salmonella sp.* was observed to infiltrate human tumours (Yoon *et al.*, 2017). Presence of lipopolysaccharide in *Salmonella sp.*, induces septic shock (Pawelek *et al.*, 2003). However, some mutants of *Salmonella sp.* did not show signs of virulence when Bacon and colleague injected into mice (Pawelek *et al.*, 2003). *Salmonella sp.* are facultative anaerobic which able to colonise large and small tumours. Tumour microenvironment consist of multi-potent stromal cells, fibroblast, blood vessels, endothelial cell precursors, immune cells and cytokines (Casey *et al.*, 2015). Tumours undergo angiogenesis, as tumour cells grow much faster than cells making up the blood vessel, which results in biological changes and adaptive metabolisms such a formation of defective vessels, appearance of apoptotic and necrotic cancer tissues, and emergence of hypoxic areas and heterogeneous tumour cell populations (Wei *et al.*, 2008). There are two types of hypoxia identified within tumour microenvironment. One is chronic hypoxia due to limited perfusion of oxygen through tissue and another one is cycling hypoxia due to temporary blockage of tumour blood vessels (Casey *et al.*, 2015). Hypoxia provides vast chances for bacteria to infiltrate the tumour (Wei *et al.*, 2008).

*Salmonella sp* has property to regress and inhibit tumour growth in a wide range of human and mouse tumours such as B16F10 murine melanoma and human tumour xenograft Lox, DLD-1, A549, WiDr, HTB177, and MDA-MB-231 (Luo *et al.*, 2001; Pawelek *et al.*, 2003). To take advantage of this property, hence the possibilities to genetically modify *Salmonella sp* by inserting prokaryotic expression plasmids using mammalian cells *in vitro* and *in vivo* was explored successfully (Yoon *et al.*, 2017).

Genetically modified *Salmonella enterica* serovar Typhimurium (VNP20009) demonstrated significant reduction in Tumour Necrosis Factor alpha (TNF- $\alpha$ ) production which eventually cause septic shock and deletion of *purl* gene lead to tumour specific colonization. This is the only genetically modified *Salmonella sp.* that was tested in canine thus far (Luo *et al.*, 2001). Besides that, there are A1-R strain which is leucine and arginine auxotrophic (Li *et al.*, 2013), ΔppGpp is a avirulent *Salmonella enterica* serovar Typhimurium, which was mutated at

*relA*<sup>-</sup> and *spoT*<sup>-</sup> (Na *et al.*, 2006) and LH430 which was engineered at *phoP*<sup>-</sup> and *phoQ*<sup>-</sup> which are endotoxin related genes (Li *et al.*, 2013).

In a study using mice model, *Salmonella* sp. were seen in the necrotic regions and some in the cytoplasm of the tumour cells itself. This observation leads to understanding of *Salmonella* sp. pathogenicity islands to further understand the mechanism of how *Salmonella* sp. infect and regress the tumour (Pawelek *et al.*, 2003).

It is reported that most virulent genes responsible for bacterial pathogenesis are located in *Salmonella* Pathogenicity Island (SPI) (Karasova *et al.*, 2009). The virulence factors of *Salmonella* sp. such as *sopB*, *sopD* and *pipD* important in salmonellosis causing enteropathogenicity which results in acute inflammatory cell influx, intestinal fluid secretion and enteritis causing clinical diarrhoea (Khoo *et al.*, 2015). Wild type *Salmonella enterica* serovar Agona (WA32) was genetically engineered to silence the metabolite and virulence genes to develop an attenuated strain (4KA32) which shows significant cytotoxic effects on canine mammary tumour cells *in vitro* (Lee *et al.*, 2015). Even though it is tested on canine mammary tumour cell line, the pathogenicity of 4KA32 is poorly understood in dogs. The hypothesis of the study is, 4KA32 should not be able to express any pathogenicity in term of clinical and pathological due to deletion of the virulence gene.

The main justification of this study is to determine the pathogenicity of 4KA32 on healthy dogs. The objectives of this study as follows:

1. To determine the clinical changes on dogs after administration of 4KA32.
2. To study the changes in haematology after the administration of 4KA32.
3. To determine the pathological changes in dogs after administration of 4KA32.

## REFERENCES

- Abraham, J. (2016). Hormonal therapy for cancer. *Medicine*, 44(1): 30–33.
- Alenza, P. D., Rutteman, G. R., Peña, L., Beynen, A. C., & Cuesta, P. (1998). Relation between habitual diet and canine mammary tumours in a case-control study. *Journal of Veterinary Internal Medicine / American College of Veterinary Internal Medicine*, 12: 132–139.
- Arango Duque, G., & Descoteaux, A. (2014). Macrophage Cytokines: Involvement in Immunity and Infectious Diseases. *Frontiers In Immunology*, 5: 14-19.
- Autio, K., Ruotsalainen, J., Anttila, M., Niittykoski, M., Waris, M., & Hemminki, A. et al. (2015). Attenuated Semliki Forest virus for cancer treatment in dogs: safety assessment in two laboratory Beagles. *BMC Veterinary Research*, 11(1): 46-52
- Biller, B. J. (2007). Cancer immunotherapy for the veterinary patient. *Veterinary Clinics of North America - Small Animal Practice*, 37: 1137–1149.
- Bhandal, J., Langohr, I., Degner, D., Xie, Y., Stanley, B., & Walshaw, R. (2012). Histomorphometric analysis and regional variations of full thickness skin grafts in dogs. *Veterinary Surgery*, 41(4): 448-454
- Bostock, D. (1986). Canine and feline mammary neoplasms. *British Veterinary Journal*, 142(6): 506-515.
- Burnie, A., Simpson, J., Lindsay, D., & Miles, R. (1983). The excretion of campylobacter, salmonellae and Giardia lamblia in the faeces of stray dogs. *Veterinary Research Communications*, 6(1): 133-138.
- Cakir Bayram, L., & Aydin, F. (2016). An abdominal cavity abscess associated with *Salmonella enterica* serovar Typhimurium phage type DT2 in a dog: a case report. *Veterinární Medicína*, 61(5): 272-278.
- Casals, E., Gusta, M. F., Cobaleda-Siles, M., Garcia-Sanz, A., & Puntes, V. F. (2017). Cancer resistance to treatment and antiresistance tools offered by multimodal multifunctional nanoparticles. *Cancer Nanotechnology*, 8(1): 7.
- Cassali, G. D., Gobbi, H., Malm, C., & Schmitt, F. C. (2007). Evaluation of accuracy of fine needle aspiration cytology for diagnosis of canine mammary tumours: Comparative features with human tumours. *Cytopathology*, 18(3): 191-6.
- Cheminay, C., Chakravortty, D., & Hensel, M. (2003). Role of Neutrophils in Murine Salmonellosis. *Infection And Immunity*, 72(1): 468-477.

- Cheah, Y. K., Khoo, C. H., & Taraq, N. Z. (2015). *International Patent No. P12013702237*. Geneva, Switzerland: World Intellectual Property Organisation.
- Choi, J.-W., Yoon, H.-Y., & Jeong, S.-W. (2016). Clinical outcomes of surgically managed spontaneous tumours in 114 client-owned dogs. *Immune Network*, 16(2): 116-25.
- Clarke, M., Collins, R., Darby, S., Davies, C., Elphinstone, P., Evans, V., Godwin, J., Gray, R., Hicks, C., James, S., Mackinnon, E., McGale, P., McHugh, T., Peto, R., Taylor, C., Wang, Y. & Early Breast Cancer Trialist Collaborative Group (EBCTCG). (2005). Effects of radiotherapy and of differences in the extent of surgery for early breast cancer on local recurrence and 15-year survival: an overview of the randomised trials. *The Lancet*, 366(9503): 2087–2106
- Coley, W. B. (1898). The treatment of inoperable sarcoma with the 'mixed toxins of erysipelas and bacillus prodigiosus. *Journal of the American Medical Association*, XXXI(9): 456-465.
- Conlan, J. (1997). Neutrophils and tumour necrosis factor- are important for controlling early gastrointestinal stages of experimental murine listeriosis. *Journal Of Medical Microbiology*, 46(3): 239-250
- David, Duan, W., Jozef, & Wei, M. (2011). Cancer gene therapy - Developments and future perspectives, *InTech*.
- Delepeulaire, P. (2004). Type I secretion in gram-negative bacteria. *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research*, 1694(1): 149–161.
- Dubik, D., Dembinski, T. C., & Shiu, R. P. C. (1987). Stimulation of c-myc oncogene expression associated with estrogen-induced proliferation of human breast cancer Cells1. *Cancer Research*, 47: 6517–6521.
- Edgar, R., Mazor, Y., Rinon, A., Blumenthal, J., Golan, Y., & Buzhor, E. et al. (2013). LifeMap Discovery™: The Embryonic Development, Stem Cells, and Regenerative Medicine Research Portal. *Plos ONE*, 8(7): e66629.
- Egenvall, A., Bonnett, B. N., Ohagen, P., Olson, P., Hedhammar, A., & Von Euler, H. (2005). Incidence of and survival after mammary tumors in a population of over 80,000 insured female dogs in Sweden from 1995 to 2002. *Preventive Veterinary Medicine*, 69(1–2): 109–127.
- Finlay, B., Heffron, F., & Falkow, S. (1989). Epithelial cell surfaces induce *Salmonella* proteins required for bacterial adherence and invasion. *Science*, 243(4893): 940-943.
- Fritz, S., Henson, M., Greengard, E., Winter, A., Stuebner, K., & Yoon, U. et al. (2016). A phase I clinical study to evaluate safety of orally administered, genetically engineered *Salmonella entericaserovarTyphimurium* for canine osteosarcoma. *Veterinary Medicine And Science*, 2(3): 179-190.

- Galan, J., & Curtiss, R. (1989). Cloning and molecular characterization of genes whose products allow *Salmonella typhimurium* to penetrate tissue culture cells. *Proceedings Of The National Academy Of Sciences*, 86(16): 6383-6387.
- Ganai, S., Arenas, R. B., & Forbes, N. S. (2009). Tumour-targeted delivery of TRAIL using *Salmonella typhimurium* enhances breast cancer survival in mice. *British Journal of Cancer*, 101(10): 1683–1691.
- Gamba, C. O., Dias, E. J., Ribeiro, L. G. R., Campos, L. C., Estrela-Lima, A., Ferreira, E., & Cassali, G. D. (2013). Histopathological and immunohistochemical assessment of invasive micropapillary mammary carcinoma in dogs: A retrospective study. *The Veterinary Journal*, 196(2): 241–246.
- Goldschmidt, M., Peña, L., Rasotto, R., & Zappulli, V. (2011). Classification and grading of canine mammary tumours. *Veterinary Pathology*, 48: 117–131.
- Gómez J, B., Ramírez R, M., & Maldonado E, J. (2012). Presence of lung metastases in bitches affected by malignant mammary neoplasms in Medellin (Colombia). *Revista MVZ Córdoba*, 17(2): 2983.
- Gordon, J., Machiz, S., Block, N., & Politano, V. (1974). *Salmonella typhosa* infection of kidney. *Urology*, 3(4): 470-472.
- Gray, T. T., & Fedorka-Cray, P. J. (2002). *Salmonella*. In Cliver, D. O. & Riemann, H. P. (Eds.), *Foodborne diseases* (pp. 55-68). Amsterdam: Academic Press.
- Grieves, J., Dick, E., Schlabritz-Loutsevich, N., Butler, S., Leland, M., & Price, S. et al. (2008). Barbiturate euthanasia solution-induced tissue artifact in nonhuman primates. *Journal Of Medical Primatology*, 37(3): 154-161.
- Guillonneau, X., Eandi, C., Paques, M., Sahel, J., Sapieha, P., & Sennlaub, F. (2017). On phagocytes and macular degeneration. *Progress In Retinal And Eye Research*, 61: 98-128.
- Hallstrom, K., & McCormick, B. (2011). *Salmonella* Interaction with and Passage through the Intestinal Mucosa: Through the Lens of the Organism. *Frontiers In Microbiology*, 2: 111-123.
- Heppner, F., & Möse, J. R. (1978). The liquefaction (oncolysis) of malignant gliomas by a non pathogenic clostridium. *Acta Neurochirurgica*, 42: 123–5.
- Hevia, H., Varela-Rey, M., Corrales, F., Berasain, C., Martínez-Chantar, M., & Latasa, M. et al. (2004). 5'-methylthioadenosine modulates the inflammatory response to endotoxin in mice and in rat hepatocytes. *Hepatology*, 39(4): 1088-1098.
- Hoffmann, M., Payne, J., Roberts, R. J., Allard, M. W., Brown, E. W., & Pettengill, J. B. (2015). Complete genome sequence of *Salmonella enterica* subsp.

- enterica* Serovar Agona 460004 2-1, Associated with a Multistate Outbreak in the United States. *Genome Announcements*, 3(4): e00690-15.
- Ibrahim, I. M., & Ali, H. R. (2016). Treatment of natural mammary gland tumours in canines and felines using gold nanorods-assisted plasmonic photothermal therapy to induce tumour apoptosis. *International Journal of Nanomedicine*, 11: 4849–4863.
- Igase, M., Hwang, C. C., Kambayashi, S., Kubo, M., Coffey, M., Miyama, T. S., & Mizuno, T. (2016). Oncolytic reovirus synergizes with chemotherapeutic agents to promote cell death in canine mammary gland tumour. *Canadian Journal of Veterinary Research*, 80(1): 21-31.
- Ingersoll, M., Platt, A., Potteaux, S., & Randolph, G. (2011). Monocyte trafficking in acute and chronic inflammation. *Trends In Immunology*, 32(10): 470-477.
- Karasova, D., Havlickova, H., Sisak, F., & Rychlik, I. (2009). Deletion of sodCI and spvBC in *Salmonella enterica* serovar Enteritidis reduced its virulence to the natural virulence of serovars Agona, Hadar and Infantis for mice but not for chickens early after infection. *Veterinary Microbiology*, 139: 304–309.
- Katz, D. (1988). Antigen presentation, antigen-presenting cells and antigen processing. *Current Opinion In Immunology*, 1(2): 213-219.
- Kasinskas, R. W., & Forbes, N. S. (2006). *Salmonella typhimurium* specifically chemotax and proliferate in heterogeneous tumour tissue in vitro. *Biotechnology and Bioengineering*, 94, 710–721.
- Kelsey, J. L., Moore, a S., & Glickman, L. T. (1998). Epidemiologic studies of risk factors for cancer in pet dogs. *Epidemiologic Reviews*, 20(2):204–17.
- Khoo, C. H., Sim, J. H., Salleh, N. A., & Cheah, Y. K. (2015). Pathogenicity and phenotypic analysis of sopB, sopD and pipD virulence factors in *Salmonella enterica* serovar typhimurium and *Salmonella enterica* serovar Agona. *Antonie van Leeuwenhoek, International Journal of General and Molecular Microbiology*, 107: 23–37.
- Kinsey, J. R., Gilson, S. D., Hauptman, J., Mehler, S. J., & May, L. R. (2015). Factors associated with long-term survival in dogs undergoing liver lobectomy as treatment for liver tumors. *The Canadian Veterinary Journal*, 56(6): 598–604.
- Klopfleisch, R., Kohn, B., & Gruber, A. D. (2016). Mechanisms of tumour resistance against chemotherapeutic agents in veterinary oncology. *The Veterinary Journal*, 207: 63–72.
- Ko, D., Gamazon, E., Shukla, K., Pfuetzner, R., Whittington, D., & Holden, T. et al. (2012). Functional genetic screen of human diversity reveals that a methionine salvage enzyme regulates inflammatory cell

- death. *Proceedings Of The National Academy Of Sciences*, 109(35): E2343-E2352.
- Kristiansen, V. M., Peña, L., Díez Córdova, L., Illera, J. C., Skjerve, E., Breen, A. M., & Sørenmo, K. U. (2016). Effect of ovariohysterectomy at the time of tumour removal in dogs with mammary carcinomas: A randomized controlled trial. *Journal of Veterinary Internal Medicine*, 30(1): 230-241.
- LeBlanc, A., Naik, S., Galyon, G., Jenks, N., Steele, M., & Peng, K. et al. (2013). Safety Studies on Intravenous Administration of Oncolytic Recombinant Vesicular Stomatitis Virus in Purpose-Bred Beagle Dogs. *Human Gene Therapy Clinical Development*, 24(4): 174-181.
- Lee, Y. W., Selvarajah, G. T., Rasedee, A., & Cheah, Y. K. (2015). 10<sup>th</sup> Proceedings of the Seminar in Veterinary Sciences: Anti-cancer activities of *Salmonella enterica* serovar Agona in Canine Mammary Gland Tumour *in vitro*. Universiti Putra Malaysia.
- Lichtenstein, A., & Kahle, J. (1985). Anti-tumor effect of inflammatory neutrophils: Characteristics of *in vivo* generation and *in vitro* tumor cell lysis. *International Journal Of Cancer*, 35(1): 121-127.
- Lowden, P., Wallis, C., Gee, N., & Hilton, A. (2015). Investigating the prevalence of *Salmonella* in dogs within the Midlands region of the United Kingdom. *BMC Veterinary Research*, 11(1).
- Luo, X., Li, Z., Lin, S., Le, T., Ittensohn, M., Bermudes, D., & Zheng, L. M. (2001). Antitumour effect of VNP20009, an attenuated *Salmonella*, in murine tumour models. *Oncology Research*, 12(203): 501–508.
- Macias, H., & Hinck, L. (2012). Mammary gland development. *Wiley Interdisciplinary Reviews: Developmental Biology*, 1(4): 533-557.
- Manders, W., & Vatner, S. (1976). Effects of sodium pentobarbital anesthesia on left ventricular function and distribution of cardiac output in dogs, with particular reference to the mechanism for tachycardia. *Circulation Research*, 39(4): 512-517.
- Marks, S., & Kather, E. (2003). Bacterial-associated diarrhea in the dog: a critical appraisal. *Veterinary Clinics Of North America: Small Animal Practice*, 33(5), 1029-1060.
- Michael, G. B., Cardoso, M., & Schwarz, S. (2006). Molecular analysis of *Salmonella enterica* subsp. *enterica* serovar Agona isolated from slaughter pigs. *Veterinary Microbiology*, 112(1), 43–52.
- Milstone, L. (2004). Epidermal desquamation. *Journal Of Dermatological Science*, 36(3): 131-140.
- Minohara, Y., Kato, T., Chiba, M., Doi, K., Kurihara, Y., & Kusakado, M. et al. (2002). A rare case of *Salmonella* soft-tissue abscess. *Journal Of Infection*

- And Chemotherapy*, 8(2): 185-186.
- Misdorp, W., & Weijer (1980). Animal model of human disease: breast cancer. *The American Journal of Pathology*, 98(2): 573-576.
- Mogasale, V., Ramani, E., Mogasale, V. V., & Park, J. (2016). What proportion of *Salmonella Typhi* cases are detected by blood culture? A systematic literature review. *Annals of Clinical Microbiology and Antimicrobials*, 15: 32.
- Moore, A. (2002). Radiation Therapy for the Treatment of Tumours in Small Companion Animals. *The Veterinary Journal*, 164(3): 176-187.
- Morse, E. V., Duncan, M. A., Estep, D. A., Riggs, W. A., & Blackburn, B. O. (1976). Canine salmonellosis: A review and report of dog to child transmission of *Salmonella enteritidis*. *American Journal of Public Health*, 66(1): 82-83.
- Murphy, S. (2008). Mammary tumours in dogs and cats. *In Practice*, 30(1): 334-339.
- Naess, A., Nilssen, S., Mo, R., Eide, G., & Sjursen, H. (2016). Role of neutrophil to lymphocyte and monocyte to lymphocyte ratios in the diagnosis of bacterial infection in patients with fever. *Infection*, 45(3), 299-307.
- Nakoneczna, I., & Hsu, H. S. (1980). The comparative histopathology of primary and secondary lesions in murine salmonellosis. *British Journal of Experimental Pathology*, 61(1): 76-84.
- Nallar, S. C., Xu, D.-Q., & Kalvakolanu, D. V. (2016). Bacteria and genetically modified bacteria as cancer therapeutics: Current advances and challenges. *Cytokine*, 89: 160-172.
- Nemunaitis, J., Cunningham, C., Senzer, N., Kuhn, J., Cramm, J., Litz, C., & Sznol, M. (2003). Pilot trial of genetically modified, attenuated *Salmonella* expressing the *E. coli* cytosine deaminase gene in refractory cancer patients. *Cancer Gene Therapy*, 10: 737-744.
- Netea, M., Kullberg, B., & Van der Meer, J. (2000). Circulating Cytokines as Mediators of Fever. *Clinical Infectious Diseases*, 31(5): S178-S184.
- Nishikawa, H., Sato, E., Briones, G., Chen, L.-M., Matsuo, M., Nagata, Y., & Gnjatic, S. (2006). In vivo antigen delivery by a *Salmonella typhimurium* type III secretion system for therapeutic cancer vaccines. *Journal of Clinical Investigation*, 116(7): 1946-1954.
- Pawelek, J. M., Low, K. B., & Bermudes, D. (2003). Bacteria as tumour-targeting vectors. *Lancet Oncology*, 4: 548-556.

- Payan-Carreira, R., Martins, L., Miranda, S., Olivério, P., & Silva, S. (2016). In vivo assessment of subcutaneous fat in dogs by real-time ultrasonography and image analysis. *Acta Veterinaria Scandinavica*, 58(S1).
- Pham, O. H., & McSorley, S. J. (2015). Protective host immune responses to *Salmonella* infection. *Future Microbiology*, 10: 101–110.
- Pitiriga, V., Dendrinos, J., Nikitiadis, E., Vrioni, G., & Tsakris, A. (2016). First Case of Lung Abscess due to *Salmonella enterica* Serovar Abony in an Immunocompetent Adult Patient. *Case Reports In Infectious Diseases*, 2016: 1-4.
- Rasotto, R., Zappulli, V., Castagnaro, M., & Goldschmidt, M. H. (2012). A retrospective study of those histopathologic parameters predictive of invasion of the lymphatic system by canine mammary carcinomas. *Veterinary Pathology*, 49(2): 330–40.
- Regan, D., Guth, A., Coy, J., & Dow, S. (2016). Cancer immunotherapy in veterinary medicine: Current options and new developments. *The Veterinary Journal*, 207: 20–28.
- Roberts, N. J., Zhang, L., Janku, F., Collins, A., Bai, R.-Y., Staedtke, V., & Zhou, S. (2014). Intratumoural injection of *Clostridium novyi*-NT spores induces antitumour responses. *Science Translational Medicine*, 6(249): 249ra111.
- Rongvaux, A. (2017). Innate immunity and tolerance toward mitochondria. *Mitochondrion*, (17): 30235
- Rosche, K., Aljasham, A., Kipfer, J., Piatkowski, B., & Konjufca, V. (2015). Infection with *Salmonella enterica* Serovar Typhimurium Leads to Increased Proportions of F4/80+ Red Pulp Macrophages and Decreased Proportions of B and T Lymphocytes in the Spleen. *PLOS ONE*, 10(6): e0130092.
- Rydström, A., & Wick, M. (2009). Monocyte and neutrophil recruitment during oral *Salmonella* infection is driven by MyD88-derived chemokines. *European Journal Of Immunology*, 39(11): 3019-3030.
- Saltzman, D., Heise, C., Hasz, D., Zebede, M., Kelly, S., & Curtiss, R. et al. (1996). Attenuated *Salmonella typhimurium* Containing Interleukin-2 Decreases MC-38 Hepatic Metastases: A Novel Anti-tumor Agent. *Cancer Biotherapy & Radiopharmaceuticals*, 11(2): 145-153.
- Schmitz-Winnenthal, F. H., Hohmann, N., Niethammer, A. G., Friedrich, T., Lubenau, H., Springer, M., & Beckhove, P. (2015). Anti-angiogenic activity of VXM01, an oral T-cell vaccine against VEGF receptor 2, in patients with advanced pancreatic cancer: A randomized, placebo-controlled, phase 1 trial. *Oncoimmunology*, 4(4): e1001217.
- Schneider, R., Dorn, C. R., & Taylor, D. O. (1969). Factors influencing canine mammary cancer development and postsurgical survival. *Journal of*

- National Cancer Institute, 43(6): 1249-1261.
- Shackelford, C., Long, G., Wolf, J., Okerberg, C., & Herbert, R. (2002). Qualitative and Quantitative Analysis of Nonneoplastic Lesions in Toxicology Studies. *Toxicologic Pathology*, 30(1): 93-96.
- Shafiee, R., Javanbakht, J., Atyabi, N., Kheradmand, P., Kheradmand, D., Bahrami, A., & Khadivar, F. (2013). Diagnosis, classification and grading of canine mammary tumours as a model to study human breast cancer: an Clinico-Cytohistopathological study with environmental factors influencing public health and medicine. *Cancer Cell International*, 13(1): 469.
- Shoji, K., Yoneda, M., Fujiyuki, T., Amagai, Y., Tanaka, A., Matsuda, A., & Kai, C. (2016). Development of new therapy for canine mammary cancer with recombinant measles virus. *Molecular Therapy - Oncolytics*, 3: 1-8.
- Silpa-archa, N., Kohli, I., Chaowattanapanit, S., Lim, H., & Hamzavi, I. (2017). Postinflammatory hyperpigmentation: A comprehensive overview. *Journal of the American Academy of Dermatology*, 77(4): 591-605.
- Silver, I., & Cater, D. (1964). Radiotherapy and Chemotherapy for Domestic Animals II. Treatment of Malignant Tumours in Dogs and Cats. *Acta Radiologica: Therapy, Physics, Biology*, 2(6): 457-475
- Silver, I. (1966). Symposium on mammary neoplasia in the dog and cat— The Anatomy of the Mammary Gland of the Dog and Cat. *Journal Of Small Animal Practice*, 7(11): 689-696.
- Sorenmo, K. (2003). Canine mammary gland tumours. *Veterinary Clinics of North America: Small Animal Practice*, 33(3): 573–596.
- Sorenmo, K. U., Rasotto, R., Zappulli, V., & Goldschmidt, M. H. (2011). Development, anatomy, histology, lymphatic drainage, clinical features, and cell differentiation markers of canine mammary gland neoplasms. *Veterinary Pathology*, 48(1): 85–97.
- Sousa, C. (2008). Papular, Pustular, and Subcutaneous Skin Diseases. In R. Morgan (Ed 5<sup>th</sup>), *Handbook of Small Animal Practice*, (pp. 850-857). Missouri: Saunders.
- Thamm, D. H., Kurzman, I. D., King, I., Li, Z., Sznol, M., Dubielzig, R. R., & MacEwen, E. G. (2005). Systemic administration of an attenuated, tumour-targeting *Salmonella typhimurium* to dogs with spontaneous neoplasia: Phase I evaluation. *Clinical Cancer Research*, 11(13): 4827–4834.
- Toso, J. F., Gill, V. J., Hwu, P., Marincola, F. M., Restifo, N. P., Schwartzentruber, D. J., & Rosenberg, S. a. (2002). Phase I study of the intravenous administration of attenuated *Salmonella typhimurium* to patients with metastatic melanoma. *Journal of Clinical Oncology: Official Journal of the American Society of Clinical Oncology*, 20(1), 142–152.

- Tyrkalska, S., Candel, S., Angosto, D., Gómez-Abellán, V., Martín-Sánchez, F., & García-Moreno, D. et al. (2016). Neutrophils mediate Salmonella Typhimurium clearance through the GBP4 inflammasome-dependent production of prostaglandins. *Nature Communications*, 7: 12077.
- Veena, P., Kumar, R. V. S., Raghavender, K. B. P., Srilatha, C., & Rao, T. S. C. (2011). Ultrasonographic imaging of canine mammary tumours. *Indian Veterinary Journal*, 88(11): 23–24.
- Vendrell, A., Gravisaco, M. J., Pasetti, M. F., Croci, M., Colombo, L., Rodriguez, C., & Waldner, C. I. (2011). A novel Salmonella Typhi-based immunotherapy promotes tumour killing via an antitumour Th1-type cellular immune response and neutrophil activation in a mouse model of breast cancer. *Vaccine*, 29(4): 728–736.
- Wang, X., & Lin, Y. (2008). Tumor necrosis factor and cancer, buddies or foes?. *Acta Pharmacologica Sinica*, 29(11): 1275-1288.
- Wei, M. Q., Mengesha, A., Good, D., & Anne, J. (2008). Bacterial targeted tumour therapy-dawn of a new era, 259: 16–27.
- Zulch, H., Mills, D., Lambert, R., & Kirberger, R. (2017). The use of tramadol in a Labrador retriever presenting with self-mutilation of the tail. *Journal of Veterinary Behaviour: Clinical Applications And Research*, 7(4): 252-258